



**UNIVERSITI PUTRA MALAYSIA**

***DEVELOPMENT OF GOLD NANOPARTICLES-DNA-BASED SENSOR  
FOR DETECTION OF *Ganoderma boninense****

**SITI AKHTAR MOHSHIM**

**ITMA 2022 4**



**DEVELOPMENT OF GOLD NANOPARTICLES-DNA-BASED SENSOR FOR  
DETECTION OF *Ganoderma boninense***

**By**

**SITI AKHTAR MOHSHIM**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfilment of the Requirements for the Degree of Master of Science**

**November 2021**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



*To my beloved late father, Haji Mohshim Mahamud,*

*my lovely mother, Hajjah Che Long Hashim,*

*my wonderful husband, Mohamad Irwan Ahmad,*

*my love of my life,*

*Alesya Irdina Mohamad Irwan,*

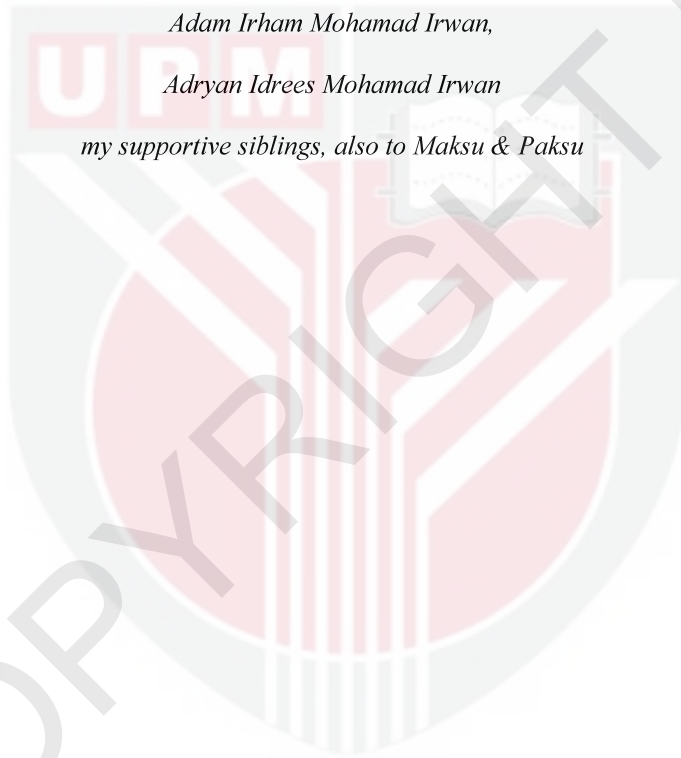
*Aaron Iskandar Mohamad Irwan,*

*Aurora Ilyssa Mohamad Irwan,*

*Adam Irham Mohamad Irwan,*

*Adryan Idrees Mohamad Irwan*

*my supportive siblings, also to Maksu & Paksu*





Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

## **DEVELOPMENT OF GOLD-NANOPARTICLES-DNA-BASED SENSOR FOR DETECTION OF *Ganoderma boninense***

By

**SITI AKHTAR MOHSHIM**

**November 2021**

**Chair : Associate Professor Shahrul Ainliah Alang Ahmad, PhD**  
**Institute : Nanoscience and Nanotechnology**

Basal stem rot (BSR) disease caused by the pathogenic fungus known as *Ganoderma boninense* (*G. boninense*) is now an endemic threat to palm health and thus plantation productivity. Currently, there are no effective treatment for BSR, early detection of *G. boninense* is crucial for the control of action. Researchers have developed many scientific discoveries towards detection of the killer fungus. However, the methods are time consuming and required expensive instrumentation and reagents, which some of them need an expertise to run the experiments. These limits bring to the aim of the study, to develop an early detection method employing genomic deoxyribonucleic acid (DNA) without amplification process for the detection of *G. boninense*. The detection of *G. boninense* was conducted based on oligonucleotide probes conjugated to AuNPs. Hybridization of the complementary target DNA was operated in two modes, i.e., (i) colorimetric assay, and (ii) observable features of DNA nano array. In this work, hybridization of targeted DNA was carried out in the presence of two DNA probes conjugated to AuNPs (Probe 1-AuNPs and Probe 2-AuNPs), yielding color changes in solution and visual microscopic changes on nano array. The developed colorimetric sensor utilizing DNA-AuNPs conjugates have a high specificity and ultrasensitive to detect unamplified genomic *G. boninense* DNA as low as 500 zeptogram (zg) which found to be more sensitive compared to the previous research. Besides, the developed method also can detect the stem rot disease as early as stage 1 infection, where there is no visual symptomatic observed on the infected plant. Another mode of detection was performed employing unamplified genomic DNA utilizing the developed Probe 1-AuNPs conjugate with immobilized DNA Probe 2 nano array. The method was able to differentiate and gave a clear illustration under simple light microscope in the positive and negative tested samples with a sensitivity obtained for *G. boninense* DNA of 30 ng. In connection to that, both developed sensors shall be the best early detection methods where it gave better simple understanding to all ages and also suitable for people without scientific background. Therefore, it may consider to be a new alternative early detection method in combating the oil palm pathogen.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

## **PEMBANGUNAN SENSOR BERASASKAN DNA-NANOZARAH EMAS BAGI PENGESANAN *Ganoderma boninense***

Oleh

**SITI AKHTAR MOHSHIM**

**November 2021**

**Pengerusi : Professor Madya Shahrul Ainliah Alang Ahmad, PhD**  
**Institut : Nanosains dan Nanoteknologi**

Penyakit reput pangkal batang (RPB) yang disebabkan oleh sejenis kulat patogen yang dikenali sebagai *Ganoderma boninense* (*G. boninense*) kini menjadi ancaman endemik terhadap kesihatan dan juga produktiviti tanaman kelapa sawit. Pada ketika ini, masih tiada rawatan yang berkesan untuk membasmi RPB dan oleh sebab itu pengesanan awal *G. boninense* sangat penting untuk tindakan kawalan. Para penyelidik telah membangunkan pelbagai penemuan saintifik bagi mengesan kulat pembunuh ini. Walau bagaimanapun, kaedah-kaedah yang telah dibangunkan agak memakan masa dan memerlukan instrumentasi dan bahan uji yang mahal, yang mana sesetengah daripadanya memerlukan tenaga pakar untuk menjalankan eksperimen yang dibangunkan. Kekangan ini telah mendorong kepada tujuan kajian ini iaitu untuk menghasilkan kaedah pengesanan awal yang baharu melalui penggunaan asid deoksiribonukleik (DNA) genom tanpa melalui proses memperbanyakkan salinan DNA bagi mengesan *G. boninense*. Pengesanan *G. boninense* dilakukan melalui penggunaan prob oligonukleotida yang dihubungkan ke AuNPs. Hibridisasi DNA sasaran pelengkap dikendalikan dalam dua mod, iaitu, (i) ujian kolorimetrik, dan (ii) melalui pemerhatian tatasusunan nano DNA di atas substrat emas. Dalam kajian ini, hibridisasi DNA yang didasarkan dijalankan dengan kehadiran dua prob DNA yang disambungkan ke AuNPs (Prob 1-AuNPs dan Prob 2-AuNPs), yang menghasilkan perubahan warna dalam larutan dan perubahan visual mikroskopik pada tatasusunan nano. Teknik pengesanan melalui perubahan warna yang dibangunkan ini mempunyai tahap pengkhususan dan sensitivity yang tinggi di mana dapat mengesan DNA genomik *G. boninense* tanpa memerlukan proses amplifikasi pada kadar serendah 500 zeptogram (zg) yang mana ianya didapati lebih sensitif berbanding penyelidikan sebelumnya. Selain itu, kaedah yang dibangunkan ini juga dapat mengesan penyakit reput pangkal batang seawal jangkitan pada tahap 1, di mana tiada gejala yang dikenalpasti pada kelapa sawit yang telah dijangkiti. Bagi kaedah pengesanan kedua, hibridisasi DNA genomik yang tidak diamplifikasikan dilaksanakan menggunakan konjugat Prob 1-nanozarah emas dan tatasusunan nano DNA Prob 2 takgerak. Kaedah ini menghasilkan ilustrasi yang jelas apabila dilihat di bawah mikroskop cahaya biasa dalam membezakan sampel positif dan negatif yang diuji di mana kepekaan pengesanan susunan DNA sasaran *G. boninense* dapat dikesan pada

kadar serendah 30 nanogram (ng). Sehubungan itu, penderia yang dibangunkan ini merupakan kaedah pengesanan awal yang terbaik di mana ia memberikan pemahaman mudah kepada semua peringkat umur pengguna dan juga sesuai digunakan oleh individu yang tidak mempunyai latar belakang saintifik. Oleh itu, kaedah pengesanan ini boleh dipertimbangkan sebagai alternatif baharu kepada kaedah pengesanan awal dalam memerangi patogen kelapa sawit.



## ACKNOWLEDGEMENTS

First of all, I would like to convey my sincere grateful to Almighty Allah S. W. T. for His bounty and grace towards the completion of this research project.

I would like to thank and express my deepest gratitude to my husband, Mohamad Irwan Ahmad, and my mom, Hajjah Che Long Hashim for the faithfulness, sacrifice, care, and endless support during my journey pursuing this master's degree. Bunches of thanks to my childrens, Alesya, Aaron, Aurora, Adam, and Adryan, for inspired me every day to grow and be better mom while studying and working.

I owe my acknowledgement to the person who offered me to this opportunity, Dr. Wong Lu Shin. I would like to convey my gratitude to the chairman of my supervisor committee, Associate Professor Dr. Shahrul Ainliah Alang Ahmad for accepting me as your student in her research group. She is good in guiding me and also very supportive. Also, I owe my sincere grateful acknowledgement to Dr. Muhammad Zamharir Ahmad, the one who was very supportive and guiding me a lot from the beginning until the end of my master's journey.

I would like to thank Newton Fund of British Council and The University of Manchester for the knowledge exchange opportunity for 2 months. Through the exchange period, I have best learned with the western work culture, the research activities and environment in Manchester, United Kingdom. My heartiest thanks towards Manchester Institute of Biotechnology for providing me a good hands-on experience and knowledge.

My research works would have been success without the help of the people at my workplace, Biotechnology & Nanotechnology Research Centre (BN Research Centre), a department of Malaysian Agricultural Research & Development Institute (MARDI). I am indebted to the entire staff of Biosensor Programme and Agri-Nanotechnology Programme at Malaysian Agriculture Research and Development Institute (MARDI) for their kind help during this project. Special thank goes to Dr. Lau Han Yih (MARDI) for helping me the most in various stages of molecular biology and experimental works. Her advice and support are also boosting my passion throughout this master's research works. I also would like to express my gratitude to my former supervisor, Professor Dr. Vijay Kumar (Biotechnology Research Institute, Universiti Malaysia Sabah), for sharing his knowledge in molecular biology and for being very supportive throughout my master's journey. Also not forgetting to my colleagues and my fellow friends for your support and somehow give me psychological advice that really do motivate and encourage me towards finishing this research works.

Finally, I wish to express my gratefulness, indebtedness and very special thanks to all my family members especially my mom, for unconditionally showing their prayers, moral and financial support during the tenure of my study. Last but not least, sending the Al-Fatihah and my gratitude to my beloved late father, who's the first person had encouraged me to pursue this master's degree.

*“Our fate lives within us...”*

Siti Akhtar Mohshim, 2021



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

**Shahrul Ainliah binti Alang Ahmad, PhD**

Associate Professor  
Department of Chemistry  
Faculty of Sciences  
Universiti Putra Malaysia  
(Chairman)

**Nor Azah binti Yusof, PhD**

Professor  
Department of Chemistry  
Faculty of Sciences  
Universiti Putra Malaysia  
(Member)

**Muhammad Zamharir bin Ahmad, PhD**

Director of MARDI Perlis  
Malaysian Agricultural Research and Development Institute (MARDI)  
(Member)

---

**ZALILAH MOHD SHARIFF, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 14 April 2022

## Declaration by Graduate Student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and the copyright of the thesis are fully-owned by Universiti Putra Malaysia, as stipulated in the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from the supervisor and the office of the Deputy Vice-Chancellor (Research and innovation) before the thesis is published in any written, printed or electronic form (including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials) as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld in accordance with the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2015-2016) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No.: \_\_\_\_\_ Siti Akhtar Mohshim, GS47578

### **Declaration by Members of Supervisory Committee**

This is to confirm that:

- the research and the writing of this thesis were done under our supervision;
- supervisory responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2015-2016) are adhered to.

Signature: \_\_\_\_\_

Shahrul Ainliah binti Alang Ahmad, PhD

Associate Professor

Committee: \_\_\_\_\_

Chairman

Signature: \_\_\_\_\_

Nor Azah binti Yusof, PhD

Professor

Committee: \_\_\_\_\_

Member

Signature: \_\_\_\_\_

Muhammad Zamharir bin Ahmad, PhD

Committee: \_\_\_\_\_

Member



## TABLE OF CONTENTS

<b>ABSTRACT</b>	<b>Page</b>
<i>ABSTRAK</i>	i
<b>ACKNOWLEDGEMENTS</b>	ii
<b>APPROVAL SHEET</b>	iv
<b>DECLARATION</b>	vi
<b>LIST OF TABLES</b>	viii
<b>LIST OF FIGURES</b>	xiv
<b>LIST OF ABBREVIATIONS</b>	xv
<b>LIST OF SYMBOLS AND UNITS</b>	xx
	xxvi

## CHAPTER

<b>1</b>	<b>INTRODUCTION</b>	<b>1</b>
1.1	Background	1
1.2	Motivations	2
1.2.1	Problem statements	2
1.2.2	Objectives of the study	3
1.2.3	Practical relevance of the study	3
1.2.4	Importance and significant contribution	4
1.3	Thesis structure overview	4
<b>2</b>	<b>LITERATURE REVIEW</b>	<b>6</b>
2.1	Oil palm	6
2.1.1	Oil palm cultivation and productions	6
2.1.2	Economic impact	8
2.1.3	Effect of diseases and pests in yield	10
2.2	Stem rot disease	14
2.2.1	Pathogen	14
2.3	Symptoms and classification of stem rot disease infection	15
2.4	<i>G. boninense</i> detection methods	23
2.4.1	Physical methods	23
2.4.1.1	Morphological observation	23
2.4.1.2	<i>Ganoderma</i> Selective Medium (GSM)	24
2.4.1.3	Tomography	24
2.4.2	Serological method	26
2.4.2.1	Enzyme-linked immunosorbent assay (ELISA)	26
2.4.3	DNA-based biosensor	27
2.4.3.1	Polymerase chain reaction (PCR)	27
2.4.3.2	Loop-mediated isothermal amplification (LAMP)	28
2.4.3.3	Multiplex polymerase chain reaction	29
2.4.4	Biosensor method	29
2.4.4.1	Electrochemical biosensor	29
2.4.4.2	DNA-based electrochemical sensor	31

2.4.5	Colorimetric-based sensor	31
2.4.5.1	Colorimetric DNA-based sensor	31
2.5	Gold nanoparticles (AuNPs) in sensing applications	32
2.6	DNA-gold nanoparticles (AuNPs) in sensing applications	32
2.7	DNA sequence selection	34
2.7.1	Internal-transcribed spacer (ITS) region	35
<b>3</b>	<b>MATERIALS AND METHODS</b>	<b>36</b>
3.1	Experimental overview	36
3.2	Materials and chemicals	38
3.3	Preparation of buffers and biochemical solutions	38
3.3.1	100 $\mu$ M synthetic DNA and primer stock solution	38
3.3.2	0.01 M tris(carboxyethyl)phosphine (TCEP)	38
3.3.3	5.0 M sodium chloride (NaCl)	39
3.3.4	0.01 M phosphate buffered saline (1 $\times$ PBS), pH 7.4	39
3.3.5	0.5 $\times$ phosphate buffered saline-tween 0.01% (0.5 $\times$ PBST 0.01%)	39
3.3.6	Potato dextrose agar (PDA)	39
3.3.7	Peptone glucose yeast (PGY) broth	39
3.3.8	PDA-PGY plate preparation	39
3.3.9	0.3 M Tris(hydroxymethyl)aminomethane hydrochloride (Tris-HCl), pH 8.0	40
3.3.10	0.25 M ethylenediaminetetraacetic acid (EDTA), pH 8.0	40
3.3.11	DNA extraction buffer	40
3.3.12	3.0 M sodium acetate ( $C_2H_3NaO_2$ ), pH 5.2	41
3.3.13	70% ethanol (EtOH)	41
3.3.14	1 $\times$ tris-acetate-EDTA (TAE)	41
3.3.15	0.5 $\times$ tris-acetate-EDTA (0.5 $\times$ TAE)	41
3.3.16	1 $\times$ tris-borate-EDTA (TBE)	41
3.3.17	Agarose gel (0.8%/1%/2.5%)	41
3.3.18	1.0 M potassium chloride (KCl)	42
3.3.19	0.1 M magnesium chloride ( $MgCl_2$ )	42
3.3.20	0.1 M tris-hydrochloride (Tris-HCl), pH 9.0	42
3.3.21	0.1 M phosphate buffer (PB), pH 7.0	42
3.3.22	Hybridization buffer A	42
3.3.23	Hybridization buffer B	42
3.3.24	1 M ammonium acetate ( $CH_3CO_2NH_4$ )	43
3.3.25	Saturated NaCl	43
3.4	Equipment and instrumentations	43
3.5	Selection and verification of target DNA sequence	44
3.6	Material characterizations	46
3.7	Functionalization of DNA probes with AuNPs	47
3.7.1	Optimization of DNA-AuNPs conjugates	47
3.7.1.1	Optimization of salt concentration	48
3.7.1.2	Optimization of pH value of AuNPs	49
3.8	Characterization of DNA-AuNPs conjugates	49

3.8.1	UV-vis spectroscopy	49
3.8.2	Agarose gel electrophoresis	49
3.8.3	Zeta potential ( $\zeta$ P)	50
3.8.4	Dynamic light scattering (DLS)	51
3.8.5	Transmission electron microscopy	52
3.9	Preparation of biological samples	52
3.9.1	Sampling of <i>Ganoderma</i> biological samples	52
3.9.2	Negative control sample	53
3.9.3	Isolation of <i>Ganoderma</i> pure cultures	54
3.10	Pure culture identification	57
3.11	Genomic DNA extraction and quantification	57
3.12	Verifications of extracted genomic DNA	59
3.13	Genomic DNA fragmentation	59
3.14	DNA hybridization	60
3.14.1	Study on effect of NaCl concentration in DNA hybridization	60
3.14.2	DNA hybridization with synthetic ssDNA	61
3.14.3	Hybridization assay with genomic DNA	61
3.14.4	Sensitivity assay for genomic DNA	62
3.15	Integration of Probe 1-AuNPs conjugate with DNA Probe 2 nano array and hybridization on the array	62
<b>4</b>	<b>RESULTS AND DISCUSSION</b>	<b>64</b>
4.1	Target DNA sequence selection, verification, and generation	64
4.2	Characterizations of AuNPs	66
4.3	Optimizations of DNA-AuNPs conjugates	67
4.3.1	Salt concentration	67
4.3.2	pH	69
4.4	Optimized DNA-AuNPs conjugates characterizations	71
4.5	Self-aggregation of DNA-AuNPs conjugates	73
4.6	Biological samples collection	74
4.6.1	Pure mycelial <i>Ganoderma</i> isolated from fruiting bodies	74
4.6.2	Genomic DNA extraction	77
4.6.3	Polymerase chain reaction (PCR)	77
4.6.4	Fragmented DNA	78
4.7	Hybridization of DNA-AuNPs with synthetic ssDNAs	79
4.8	Hybridization of DNA-AuNPs with biological samples	80
4.9	Sensitivity study of <i>G. boninense</i> with the DNA-AuNPs conjugates	81
4.10	Hybridization of DNA-AuNPs conjugate on DNA Probe 2 nano array	82
<b>5</b>	<b>SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH</b>	<b>85</b>
5.1	Summary	85
5.2	Conclusions	85

5.3	Recommendations for future research	86
<b>REFERENCES</b>		87
<b>APPENDICES</b>		113
<b>BIODATA OF STUDENT</b>		115
<b>LIST OF PUBLICATIONS</b>		116



## LIST OF TABLES

Table	Page
2.1 List of diseases that infect the oil palm trees.	11
2.2 Disease severity classification and the symptoms.	18
3.1 Selected DNA sequences. The underlined base (highlighted in red color) is a mismatch base from Target DNA.	45
3.2 Information of 20 nm AuNPs provided by respective commercial brands.	47
3.3 Series of NaCl addition in optimizing the optimum NaCl concentration for conjugation of DNA-AuNPs.	48
3.4 List of biological samples used in this study.	53
3.5 Primers for <i>Ganoderma</i> verification.	59
3.6 NaCl and water volume for self-aggregation study.	60
4.1 Information of DNA code in generating the complementary sequence to the TDNA.	65
4.2 $\zeta$ P and DLS measurements of unfunctionalized AuNPs and DNA-AuNPs conjugate.	73
4.3 Concentration and purity of DNA extracted from all biological samples for this study.	77

## LIST OF FIGURES

Figure		Page
2.1	Oil palm fruit structure showing the palm pulp (mesocarp) and the palm kernel (endosperm).	6
2.2	The increase in oil palm planted areas in Malaysia from 1975 to 2019. The chart shows that the oil palm cultivation area increased to nearly 6.0 MHa in 2019.	7
2.3	Production trends of Malaysian palm oil from 2009 - 2019.	8
2.4	Graph showing the average of monthly price for Malaysian CPO from 2018 to 2019.	9
2.5	The export trends for Malaysian oil-palm based products in 10 years (2009 to 2019).	9
2.6	Prevalence of stem rot symptoms in oil palms where red arrows pointing to the signs of infection while the blue arrows showing the basidiomata. (A) Multiple unopen spear leaves (red arrows), (B) skirt-like formation on the canopy of infected palms, (C) stem rot at the tree base with the presence of <i>Ganoderma</i> sp. fungal basidiomata and, (D) Large fruiting bodies of <i>Ganoderma</i> sp. which commonly associated with the disease. Photo (A) – (C) were courtesy from Mr. Abdullah Bakri, Unico Desa Plantation, Lahad Datu, Sabah, and photo (D) was taken by the author at the idle oil palm plantation in Dengkil, Selangor.	16
2.7	Photograph shows the fungal basidiomata abundance at the base of rotten dead palm due to the disease infection. Photo by Mr. Abdullah Bakri, Desa Unico Plantation, Lahad Datu, Sabah.	17
2.8	<i>Ganoderma</i> isolate on GSM which shown morphological characteristic of brown halo formation around the white colony. Photos are adapted from Naidu et al. (2016).	24
2.9	(A) <i>In situ</i> sonic tomographic scan, (B) in situ gamma-ray tomographic scan using GammaScorpion, (C) sonic tomogram images for healthy (C1), mild (C2), and severe (C3) trees, (D) gamma-ray tomogram images for healthy (D1), and infected (D2 and D3) trees. Photographs and images adapted from Ishaq et al. (2014), and Abdullah et al. (2013).	25
2.10	The processes in production of polyclonal antibody (pAb) before ELISA.	27
2.11	Schematic drawing showing the principal steps of PCR.	28

2.12	Schematic diagram of (A) screen printed electrode, (B) principle of electrochemical sensor.	30
2.13	Schematic diagram of illustration for functionalization of citrate-capped AuNP with thiol-modified ssDNA strand.	33
2.14	(A) Schematic diagram of Mirkin's and his colleagues' colorimetric concepts employing the DNA-nanoparticles conjugates. (B) The result of colorimetric changes where which obviously shown cuvette with red solution contains non-target DNA, while cuvette with purple-bluish color contains target DNA. (C) Result of kinetic study of the absorbance value at 260 nm corresponding to the temperature for the modified DNA and unmodified DNA. All images were adapted from Mirkin et al. (1996) and Saha et al. (2012).	34
2.15	Schematic diagram of the internal transcribe spacer (ITS) region.	35
3.1	Experimental overview showing the flowchart in developing the colorimetric DNA sensor and PPL nano array utilizing gold nanoparticles for the detection of <i>G. boninense</i> .	37
3.2	Photo of spreading PGY broth onto PDA medium to prepare two-layered solid (PDA) and liquid medium (PGY) for <i>Ganoderma</i> mycelial culture isolation.	40
3.3	The nucleotide BLAST® upper form (in the left box) under the "Enter Query Sequence" is a section to enter sequences by accession number, or in FASTA format. The following form (highlighted in yellow color) is the section for user to write titles or information regarding queries made.	44
3.4	Setting of desired organisms ( <i>Ganoderma</i> ) selection was filled in with its taxid number (5314) or can be chosen from the drop-down list to enable the BLAST® system to identify the selected specific organism. The "BLAST" button was then pressed to start the search.	45
3.5	DNA probes with thiol linker showing the organosulfur (thiol linker) where Probe 1 represents conjugates at 5' location while Probe 2 represents thiol linker at 3' location.	46
3.6	Functionalization of DNA probes on AuNPs.	47
3.7	An illustration of typical agarose gel electrophoresis apparatus.	50
3.8	Schematic diagram illustrating a negatively charged nanoparticle suspended in dispersion ionic solution.	51



3.9	Locations of sampling. The red pins indicate the original locations where the <i>Ganoderma</i> and oil palm biological samples were collected.	53
3.10	Photographs showing two groups of <i>Ganoderma</i> fruiting bodies that were dried after being cleaned with tap water. (A) samples collected from Lahad Datu and (B) sample collected from an idle plantation in Dengkil.	54
3.11	Diagram of <i>Ganoderma</i> pure cultures isolation process.	56
3.12	The experimental scheme showing the process of genomic DNA extraction for all biological samples in this study. The concentration of the extracted DNA was then measured by NanoDrop ND-1000 Spectrophotometer.	58
3.13	Schematic diagrams represent of the colorimetric ssDNA detection using the developed DNA-AuNPs conjugates (Probe 1-AuNPs and Probe 2-AuNPs conjugates).	61
3.14	The illustration of conjugated DNA-AuNPs interaction during DNA hybridization with the uneven length of fragmented <i>G. boninense</i> positive DNA and non- <i>Ganoderma</i> DNA.	62
3.15	Schematic diagram of immobilization of Probe 2 on gold surface using polymer pen lithography (PPL), and the integration with Probe 1-AuNPs conjugate followed by DNA hybridization.	63
4.1	Information of <i>G. boninense</i> strain FA5035 taken from NCBI GenBank for TDNA investigation.	64
4.2	TDNA BLAST® result showing graphic overview of the database sequences aligned to the query sequence.	65
4.3	Result of manually generating the complementary sequence to TDNA bases (orange color) in reverse mode.	66
4.4	Characterizations of AuNPs from different suppliers. (A) Photographs of the original color of citrate-stabilized AuNPs from Aldrich, Alfa Aesar, and Kestrel BioSciences, (B) UV-vis spectra of the AuNPs where black line indicates the Aldrich AuNPs, red line indicates the Alfa Aesar AuNPs, and blue line indicates the Kestrel BioSciences AuNPs. (C) TEM images of AuNPs from (1) Aldrich AuNPs, (2) Alfa Aesar AuNPs, and (3) Kestrel BioSciences AuNPs.	67
4.5	Photograph of DNA-AuNPs conjugates after the salt aging step employing NaCl concentrations (0.1 – 0.25 M) where (A) DNA-AuNPs conjugates with Aldrich AuNPs, (B) DNA-AuNPs	68



	conjugates with Alfa Aesar AuNPs, and (C) DNA-AuNPs conjugates with Kestrel BioSciences AuNPs.	
4.6	UV-vis absorption spectra of Probe 1- and Probe 2-AuNPs conjugates using different NaCl concentrations in salt-aging step. (A) and (B) represent conjugation using Aldrich AuNPs, (C) and (D) represent conjugation using Alfa Aesar AuNPs, (D) and (E) represent conjugation using Kestrel BioSciences AuNPs.	69
4.7	Photograph and UV-vis spectra of DNA-AuNPs conjugates at different pH (4.5 – 7.0) of (A) Probe 1-AuNPs, and (B) Probe 2-AuNPs.	70
4.8	Characterization of DNA-functionalized AuNPs. (A) UV-vis spectra of unfunctionalized AuNPs (black line), conjugated with Probe 1 (red line) and Probe 2 (blue line). (B) Agarose gel electrophoresis of unfunctionalized AuNPs (1), Probe 1-AuNP conjugates (2) and Probe 2-AuNP conjugates (3). (C) TEM micrographs of 20 nm AuNPs (1), Probe 1-AuNP conjugates (2) and Probe 2-AuNP conjugates (3). The red arrows indicate halo-like formation surrounding the AuNPs surface after conjugation.	72
4.9	Photographs (A) and absorption spectra (B) of DNA-AuNPs during hybridization assays (without DNA template) in the presence of different concentrations of NaCl (0.0 – 2.5 M).	73
4.10	Morphologies of fungal (photo A, B, and C) and bacteria (photo D) colony growth in the earlier stage of pure culture isolation process. From these isolates (except for the bacteria colony), a single white fungal colony was carefully selected for further isolations.	75
4.11	Fungal colonies from sample M1 (Lahad Datu, Sabah) and M2 (Dengkil, Selangor) showing a similar characteristic as the pure <i>G. boninense</i> isolates received from Faculty of Agriculture, UPM.	76
4.12	Screenshot image contains M1 sample data that deposited to NCBI GenBank by the Faculty of Agriculture, UPM.	76
4.13	Agarose gel electrophoresis showing the PCR results of all genomic DNA using ITS3 and GanET.	78
4.14	Agarose gel electrophoresis analysis of fragmented DNA obtained from all genomic DNA samples. The size of molecular markers, 100 bp as a reference to the DNA fragments.	79
4.15	(A) Photographs, (B) UV-vis spectra and (C) TEM micrographs of the DNA hybridization assays with (C1) Water, (C2) TDNA,	80

(C3) MDNA or (C4) NDNA. Each assay was performed with 500 ng of ssDNA respectively.

- |      |  |    |
|------|--|----|
| 4.16 | (A) Photographs of the hybridization assay mixtures from 500 ng genomic DNA of each biological sample before (left) and after centrifugation and resuspension (right). Both photos were taken under the white light screen. (B) UV-vis absorption spectra of the hybridization assay mixtures. (C) Graph of UV-vis $\lambda_{\text{max}}$ shifts for the assay mixtures. | 81 |
| 4.17 | (A) Photograph of the hybridization assay mixtures showing the color change is proportional to the mass of DNA. (B) Graph of UV-vis $\lambda_{\text{max}}$ of assay mixture against amount of DNA from M2 sample.  | 82 |
| 4.18 | Light microscope images of specificity results for hybridization of DNA Probe 2 nano array and Probe 1-AuNPs where (A) with 240 ngTDNA, (B) 120 ng TDNA, (C) 400 ng of M1 DNA, and (D) 225 ng of S1 sample respectively.   | 83 |
| 4.19 | Hybridization on nano array with series of different concentration ranging from (A) 400 ng, (B) 160 ng, (C) 30 ng, and (D) 3 ng of M1 DNA samples are shown.   | 84 |

## LIST OF ABBREVIATIONS

2D	2-dimensional
3D	3-dimensional
5.8S	5.8 subunits
8-MOP	8-methoxypsoralen
18S	18 subunits
28S	28 subunits
A	Adenine
A <sub>280</sub>	Absorbance value at 280 nm
A <sub>260</sub>	Absorbance value at 260 nm
AG	Aktiengesellschaft or stock corporation
ANN	Artificial neural network
<i>Apo E</i>	<i>Apolipoprotein E</i> gene
Au	Gold
AuNP(s)	Gold nanoparticle(s)
Au-S	Gold-Sulfide
BLAST <sup>®</sup>	Basic Local Alignment Search Tool <sup>®</sup>
bp	Base pair
BSR	Basal stem rot
C	Cytosine
CCCVd	Coconut cadang-cadang viroid
cDNA	Complementary DNA
CNV	Copy number variant(s)
Co.	Company
Corp.	Corporation

CPO	Crude palm oil
CT	Computer tomography
Cy5	Cyanine 5 fluorescent labeled
Dev	Deviation
DI	Deionized
DLS	Dynamic light scattering
DNA(s)	Deoxyribonucleic acid(s)
DNA-AuNPs	DNA-gold nanoparticles
DOSM	Department of Statistics Malaysia
DPO	Dual priming oligonucleotide
dsDNA	Double-stranded DNA
EDTA	Ethylenediaminetetraacetic acid
e.g.	Example
ELISA	Enzyme-linked immunosorbent assay
E-Nose	Electronic nose
ERM	European Reference Material
Err	Error
et al.	And others
etc.	And others
FASTA	A text-based format for representing either nucleotide sequences or amino acid (protein) sequences, in which nucleotides or amino acids are represented using single-letter codes
FFB	Fresh fruit bunch
FRET	Fluorescence resonance energy transfer
G	Guanine
<i>G.</i>	<i>Ganoderma</i>

G4	Guanine-quadruplex
G-C	Guanine-cytosine content
GanET	Reverse primer for identification of <i>Ganoderma boninense</i>
GSM	<i>Ganoderma</i> selective medium
HPLC	High performance liquid chromatography
i.e.	That is
IDT	Integrated DNA Technologies
Inc.	Incorporated
<i>in situ</i>	On site/on position/in place where the event take place
<i>in vitro</i>	In “test tube” experiment
IP	Isoelectric point
IRMM	Institute for Reference Materials and Measurements
ITS	Internal transcribed spacer
ITS1	Internal transcribed spacer 1
ITS2	Internal transcribed spacer 2
ITS3	Forward primer for identification of <i>Ganoderma boninense</i>
Jacq.	Jacquin
JRC	Joint Research Centre
LAMP	Loop-mediated isothermal amplification
LC	Limited company
Llc.	Limited liability company
LOD	Limit of detection
LSPR	Localized surface plasmon resonance
Ltd.	Limited
MARDI	Malaysian Agricultural Research and Development Institute
MEA	Malt extract agar

MDNA	Mismatch DNA
MMP	Magnetic microparticles
MPOB	Malaysian Palm Oil Board
Mr.	Mister
NCBI	National Center for Biotechnology Information
NDNA	Non-complementary DNA
NIFOR	The Nigerian Institute for Oil Palm Research
Nil	Nothing/Unavailable
NNLS	Non-negative least squares
pAb(s)	Polyclonal antibody(s)
PALS	Phase-analysis light scattering
PB	Phosphate buffer
PBS	Phosphate-buffered saline
PCR	Polymerase chain reaction
PCR-RFLP	PCR-Restriction length polymorphism
PDA	Potato dextrose agar
PGY	Peptone yeast glucose
PGY-PDA	Peptone glucose yeast-potato dextrose agar
pH	Numerical/logarithmic scale to specify acidity or alkalinity
PKO	Palm kernel oil
POC	Point-of-care
PPL	Polymer pen lithography
PPO	Processed palm oil
Probe 1- AuNPs	Probe 1-gold nanoparticles
Probe 2- AuNPs	Probe 2-gold nanoparticles

Pte.	Private
Q1	Quarter 1
QD	Quantum dot
RFLP	Restriction fragment length polymorphisms
RNA	Ribonucleic acid
rRNA	Ribosomal ribonucleic acid
RT	Room temperature
RT-PCR	Real-time PCR
S1	Round-up Ready™ soybean sample
SCMV	Sugar cane mosaic virus
SEM	Scanning electron microscope
<i>sp.</i>	Species
<i>spp.</i>	Several species
SPPR	Surface plasmon polariton resonance
SPR	Surface plasmon resonance
ssDNA	Single-stranded DNA
Std	Standard
T	Thymine
TAE	Tris-acetate-EDTA
taxid	Taxonomy identifier
TBE	Tris-borate-EDTA
TCEP	Tris(2-carboxyethyl)phosphine
TDNA	Target DNA
TEM	Transmission electron microscope/microscopy
UPM	Universiti Putra Malaysia

USA	United States of America
USR	Upper stem rot
UV	Ultraviolet
UV-vis	Ultraviolet-visible





## LIST OF SYMBOLS AND UNITS

°C	Degree Celsius
$\lambda$	Lambda
$\lambda$ max	Maximum peak absorption
%	Percent
$\geq$	Greater or equal to
$\leq$	Less or equal to
$\pm$	More or less
+	Positive charge
-	Negative charge, or minus, or to
/	Or
$\sim$	Approximately
:	Ratio
™	Trademark
®	Registered trademark
×	Time(s)
→	To
$\mu$ L	Microliter(s)
$\mu$ M	Micromolar
$\mu$ m	Micrometer(s) or micron
$\zeta$	Zeta
$\zeta$ P	Zeta potential
cm	Centimeter(s)
g	Gram(s)

h	Hour(s)
M	Molar
MΩ cm-1	Megaohms per centimeter resistivity unit
M-lcm-1	Molar absorptivity coefficient unit
mg	Milligram(s)
Ha	Hectares
L	Litre
MHa	Million hectares
min	Minute(s)
mL	Milliliter(s)
mm	Millimeter(s)
mM	Millimolar
MMt	Million metric tons
MT	Metric tons
Mt	Million tons
MV	Millivolt(s)
ng	Nanogram(s)
ng/μL	Nanogram(s) per microlitre(s)
nm	Nanometer(s)
nM	Nanomolar
pmol	Picomol
RM	Ringgit Malaysia
RPM	Revolution per minute
s	Second(s)
t	Ton(s)

V	Volt(s)
v/v	Volume per volume
yg/ $\mu$ L	Yoctogram(s) per microlitre
zg	Zeptogram



## CHAPTER 1

### INTRODUCTION

#### 1.1 Background

Palm oil is an edible vegetable oil that has been found in almost all of our daily products such as consumer foods, livestock's foods, confectioneries and cosmetics (Voora et al., 2019). A few years ago, palm oil has also been used as biofuels in response to the sustainable agricultural campaign and towards global concern for a greener world (Mykhaylova, 2018; Supramaniam, 2016). Consumption of the palm oil is widely used around the world especially in Malaysia and is claimed as one of the cheapest edible and versatile oil compared to other edible oils such as olive oil, sunflower oil, canola oil, etc. (Hirschmann, 2020). Oil palm (*Elaeis guineensis* Jacq.) is one of the major agricultural commodity for Malaysian trade sector (Din, 2018; Ministry of Plantation Industries and Commodities, 2020). A statistic report provided by Statistica Inc., showed that Malaysia dominates the world's palm oil industry (Hirschmann, 2020) where the export volume of Malaysian palm oil for quarter 1 (Q1) in 2020 shows a reduction to 12.13 Mt compared to Q1 in 2019 (13.83 Mt). This situation shows that the export volume of Malaysian palm oil within a year has decreased by 12.3% (Ministry of Plantation Industries and Commodities, 2020). Due to the increasing of global demand on the palm oil, Malaysia is also experiencing an inadequate for domestic supply.

The reductions in palm oil export as well as domestic needs is attributable to the stem rot diseases (basal stem rot and upper stem rot) on the oil palm trees, which are infected by a fungus named *Ganoderma boninense* (*G. boninense*) (A. Roslan and A. S. Idris, 2012; Flood et al., 2000). The industries and also the palm oil producing countries especially Malaysia, suffer a huge losses due to the shrinkage of the export and foreign exchanges (Hushiarian, 2015). The value of losses could be greater in the future due to the difficulties in identifying the diseases at early stage. The damages caused serious impact to the smallholders as well as to the oil-palm based industries (A. Roslan and A. S. Idris, 2012). It is necessity to overcome these problems where hitherto been no cure for the diseases. To date, there are no reported solution in eradicating this stem rot diseases. To overcome this issue, an early diagnostics method which is specific to the main causal of stem rot diseases yet also sensitive must be developed. It is required to have an early detection to identify suffered oil palm trees as early as possible in a way to save the entire palms of the plantation thus able to rescue the smallholders and the oil palm industries.

Several studies either by biological, chemical, and physical methods were attempted to overcome the absence of early detection method on *Ganoderma* sp. Nevertheless, some of the techniques that previously reported are not based on analytical methods but only by traditional symptoms observation. These methods require very frequent monitoring by workers to avoid infection in the entire plantation area. Another observation method is through fungal cultural studies on *Ganoderma* selective medium (GSM) (Darus & Abu Seman, 1992). Due to slow progress, both methods are not suitable to be labeled as early detection methods and the difficulties to cover a large-scale applications became a matter

of course (L. Rajendran et al., 2009). With rapid and advances in the research area, various types of biosensors have been developed through signal transduction such as, by colorimetric (Natarajan et al., 1986), electrochemical (Dutse et al., 2012; Hushiarian, 2015), and optical (Ahmadi et al., 2017; Alexander et al., 2014) methods which applied to detect *G. boninense*. The combination of substances such as antibodies, cells, enzymes, and deoxyribonucleic acid (DNA) together with the developed technologies can result to a more selective biosensor method in identifying *G. boninense*. Molecular DNA- and immunological-based detection methods such as, polymerase chain reaction (PCR) (Abu Seman et al., 2003, 2010; Mandal et al., 2014; Panchal & Bridge, 2005) and enzyme-linked immunosorbent assay (ELISA) (Madiah et al., 2014; Utomo et al., 1997) were reported as early detection methods. But on the other hand, most of the methods require highly skilled labor and must be operated in the laboratory.

The availability of nucleic acids through synthetic synthesis which is able to specifically target analytes and produce specific binding that exhibit a simple helical ribbon formation to a complex structures such as, hairpin and guanine-quadruplex (G4) together with their relative chemical properties, making them more preferred than antibodies and other biological analytes (Mondal et al., 2018). Nucleic acid which is known as DNA has become an interesting biomolecule due to these advantages. In addition, the easiness to operate, high in target affinity, easy to label, as well as low-cost production, makes it an attractive analyte for the development of biodiagnostic methods (Famulok & Mayer, 2011). Besides the DNA, gold nanoparticle (AuNP) has been known as a metal nanomaterial that offers as colorimetric-based sensing in foods (Draz et al., 2018), clinical (S.-H. Chen et al., 2009), environmental (Amanulla et al., 2017), and agricultural sectors (Baetsen-Young et al., 2018). Gold nanoparticles (AuNPs) with combination of biological substances are an ideal pair in biosensor developments. The stability of AuNPs increases after functionalization with nucleic acids and thus makes them suitable in producing sensing tools. Therefore, development of colorimetric methods using DNA functionalized with AuNPs could be an interesting new alternative detection system in targeting the *G. boninense*. Additionally, the high target selectivity and ultrasensitivity from the deployment of DNA-AuNPs conjugates in many biosensor applications has resulted in extraordinary and spectacular potential for early detection of *G. boninense*.

## **1.2 Motivations**

### **1.2.1 Problem statements**

*G. boninense* is a pathogen that causes to basal stem rot (BSR) and upper stem rot (USR) of oil palm, eventually reducing the yield of the crop. Until now, there is no effective treatment for this fungus. While waiting for the emergence of the fungal exterminator to exist, early detection method is needed to save the oil palm plantations and industries as well as the economy of the producing countries. Researchers around the world come with innovative ideas in developing sensors to detect the major cause of the deadly disease for oil palm, which is known as *G. boninense* fungus. However, most of the published or commercial techniques such as, GSM which involves a time-consuming traditional culturing method and observation (Darus & Abu Seman, 1992), a sensitive ELISA method (Madiah et al., 2014; Utomo et al., 1997) which might create a cross-reactivity

results due to unspecific antibodies developed from animals source (Lau, 2016), PCR (Abu Seman et al., 2003; Mandal et al., 2014; Panchal & Bridge, 2005) that famously known as very selective and sensitive method but yet the method is shadowed by the utilization of sophisticated equipment and the needed of expertise. These methods clearly show that there are limitations as well as the need of high-trained personnel with expensive instrumentation end up led to expensive costing in order to detect the fungus. Since 1996, 5 – 6% healthy-looking oil palms were reported infected with the fungus (Ariffin et al., 1996). The early stage (level 1) *Ganoderma* infected oil palms are asymptomatic, thus made it difficult to define the oil palm condition (Ibrahim et al., 2020). Previous studies indicate that the earliest symptoms can be differentiated between the healthy and level 1 infected oil palm trees using a high resolution hyperspectral remote sensing (Shafri et al., 2011). However, the sensitivity detection through the method was unreported. The advanced biosensors such as, tomography (J. Abdullah et al., 2013; Ishaq et al., 2014), electronic nose (E-Nose) (Kresnawaty et al., 2020; Markom et al., 2009), ergosterol analysis (Muniroh et al., 2014), infrared spectroscopy (Alexander et al., 2014; Noor Azmi et al., 2020), etc. can provide rapid and robust in pathogen detection but not a simple yet sensitive detection methods.

### 1.2.2 Objectives of the study

The aim of this thesis was to develop a simple, specific, and sensitive gold nanoparticles-DNA-based sensors for the determination of *G. boninense*. The proposed sensing methods were aimed to be conducted in two modes, which are:

- colorimetric assay in combination with UV-visible (UV-vis) readouts
- detection based on dot features of nano array appearance with optical microscope.

In order to achieve the goal, the specific technical objectives are:

1. To design two DNA probes (Probe 1 and Probe 2) by selecting specific sequence from ITS gene of *G. boninense* genome.
2. To produce, optimize and characterize DNA-AuNPs conjugates (Probe 1-AuNPs and Probe 2-AuNPs) as hybridization platform for colorimetric sensor.
3. To investigate hybridization systems in assay solution and on nano array employing the produced DNA-AuNPs conjugates with synthetic DNA and fragmented genomic DNA samples.

### 1.2.3 Practical relevance of the study

This study is motivated by the need in designing a novel selective and sensitive DNA biosensor at point-of-care (POC) without amplification of DNA where complex protocol, equipment and biological sample purification are eliminated. Selected DNA probes conjugated to AuNPs were employed to detect DNA target sequence (35 bases) which exist in *G. boninense*. The color of the assay is expected to change (from reddish to purplish color) through DNA hybridization. A localized surface plasmon resonance (LSPR) readout from the assay could be carried out to show quantitative results due to aggregation of AuNPs in the presence of complementary sequence. The hybridization of target sequence in synthetic and genomic DNA on the nano array surface could be observed under a simple light microscope, which offers a simple, easy-to-use and portable sensor for on-site monitoring.

#### **1.2.4 Importance and significant contribution**

The arose problem statements specify to the question; what is important or influencing actions to be taken in order to eradicate the disease? Herein, the author will discuss the findings of the study which will contribute to the benefit of consumers and industries related to the oil palm products. The necessity of this study is to develop a new early detection method in detecting *G. boninense* with a simple application but yet still selective and sensitive. The study is also to create a POC diagnostic concerning to the loss of crops due to basal stem rot (BSR) disease which affecting the economics of palm oil producing countries. Thus, the developed sensor will lead towards eliminating the disease faster than the current available approaches.

#### **1.3 Thesis structure overview**

This thesis is composed of five chapters with references and appendices. Each chapter will be discussed in detail as follows:

1. Chapter 1 presents general overview of the oil palm and BSR disease. Research motivation, objectives and significant relevance of this study were also discussed in this chapter.
2. Chapter 2 presents a literature review of the oil palm, economic impact, BSR disease outbreak and the pathogens that trigger the BSR disease and infection to the oil palm. This chapter also discussed about current available detection methods and disease management strategies. The chapter also includes the development of colorimetric detection targeting the complementary DNA utilizing DNA probes functionalized AuNPs and hybridization mechanism. In addition, the properties and functionalization of DNA probes to AuNPs were also reviewed.
3. Chapter 3 describes the experimental details from selecting and analyzing the DNA probe sequences, real sample collections, pure culture isolation and identification. The conjugation of DNA-AuNPs, optimizations and characterizations of the functionalized nanoparticles properties are presented. Details of hybridization technique based on color changes and the quantitative LSPR study are described.



4. Chapter 4 presents the experimental results obtained from the study. The specificity and sensitivity level depicting the limit of detection (LOD) of the developed sensor are discussed in this chapter employing two methods i. e., hybridization in assay solution, as well as on nano array.
5. Chapter 5 presents a conclusion and summary of the present study. Some recommendations for possible future works are also included.





## REFERENCES

- A. Roslan and A. S. Idris. (2012). Economic Impact of *Ganoderma* Incidence on Malaysian Oil Palm Plantation – A Case Study in Johor. *Oil Palm Industry Economic Journal*, 12(1), 24–30.
- Abdullah, F. (2000). Spatial and Sequential Mapping of the Incidence of Basal Stem Rot of Oil Palms (*Elaeis guineensis*) on a Former Coconut (*Cocos nucifera*) Plantation. In J. Flood, P. D. Bridge, & M. Holderness (Eds.), *Ganoderma Diseases of Perennial Crops* (pp. 183–194). CABI.  
<https://doi.org/10.1079/9780851993881.0183>
- Abdullah, F., Husrita, H., & Siddiquee, S. (2008, June 2). *Ganoderma boninense* Strain FA5035 18S Ribosomal RNA Gene, Partial Sequence; Internal Transcribed Spacer 1, Complete Sequence; and 5.8S Ribosomal RNA Gene, Partial Sequence (MN148580.1). Nucleotide [Internet]; Bethesda (MD): National Library of Medicine (US), National Center for Biotechnology Information.  
<https://www.ncbi.nlm.nih.gov/nuccore/189098236>
- Abdullah, J., Hassan, H., Shari, M. R., Mohd, S., Mustapha, M., Mahmood, A. A., Jamaludin, S., Ngah, M. R., & Hamid, N. H. (2013). GammaScorpion: Mobile Gamma-Ray Tomography System for Early Detection of Basal Stem Rot in Oil Palm Plantations. *Optical Engineering*, 52(3), 036502.  
<https://doi.org/10.1117/1.oe.52.3.036502>
- Abdullah, R., & Wahid, M. B. (2010). World Palm Oil Supply, Demand, Price and Prospects: Focus on Malaysian and Indonesian Palm Oil Industry. In *Malaysian Palm Oil Board MPOB*.  
[http://mpoc.org.my/upload/WorldPalmOil\\_SupplyDemandPriceProspects\\_MalaysianIndonesianIndustry\\_FullReport.pdf](http://mpoc.org.my/upload/WorldPalmOil_SupplyDemandPriceProspects_MalaysianIndonesianIndustry_FullReport.pdf)
- Abu Seman, I., & Abdul Rahman, R. (2008). Polyclonal Antibody for Detection of *Ganoderma*. *MPOB Information Series*, 405(405).  
<http://www.sciencedirect.com/science/article/pii/S0032063311003485>
- Abu Seman, I., Darus, A., Swinburne, T. R., & Watt, T. A. (2000). *The Identity of Ganoderma Species Responsible for Basal Stem Rot Disease of Oil Palm in Malaysia – Pathogenicity Test*. Malaysian Palm Oil Board (MPOB).  
<http://palmoilis.mpob.gov.my/TOTV3/tt-no-77b-the-identity-of-ganoderma-species-responsible-for-basal-stem-rot-disease-of-oil-palm-in-malaysia-pathogenicity-test/>
- Abu Seman, I., & Kamarudin, N. (2016). *Control and Management of Ganoderma Disease in Peat Areas*. [http://soppoa.org.my/wp-content/uploads/2016/12/1.6\\_Ganoderma.pdf](http://soppoa.org.my/wp-content/uploads/2016/12/1.6_Ganoderma.pdf)
- Abu Seman, I., S., R., Madihah, A. Z., & Wahid, M. B. (2010). Multiplex PCR-DNA Kit for Early Detection and Identification of *Ganoderma* Species in Oil Palm. *MPOB TS*, 73(531). [www.mpob.gov.my](http://www.mpob.gov.my)

- Abu Seman, I., Yamaoka, M., Hayakawa, S., Basri, M. W., Noorhasimah, I., & Ariffin, D. (2003). PCR Technique for Detection *Ganoderma*. *MPOB Information Series*, June, 1–4.
- Adaskaveg, J. E., & Gilbertson, R. L. (1986). Cultural Studies and Genetics of Sexuality of *Ganoderma lucidum* and *G. tsugae* in Relation to the Taxonomy of the *G. lucidum* Complex. *Mycologia*, 78(5), 694. <https://doi.org/10.2307/3807513>
- Adaskaveg, J. E., & Gilbertson, R. L. (1988). Basidiospores, Pilocystidia, and Other Basidiocarp Characters in Several Species of the *Ganoderma lucidum* Complex. *Mycologia*, 80(4), 493. <https://doi.org/10.2307/3807851>
- Aderungboye, F. O. (1977). Diseases of the Oil Palm. *PANS*, 23(3), 305–326. <https://doi.org/10.1080/09670877709412457>
- Ahmadi, P., Muharam, F. M., Ahmad, K., Mansor, S., & Abu Seman, I. (2017). Early Detection of *Ganoderma* Basal Stem Rot of Oil Palms Using Artificial Neural Network Spectral Analysis. *Plant Disease*, 101(6), 1009–1016. <https://doi.org/10.1094/PDIS-12-16-1699-RE>
- Ahn, S., Jung, S., & Lee, S. (2013). Gold Nanoparticle Contrast Agents in Advanced X-ray Imaging Technologies. *Molecules*, 18(5), 5858–5890. <https://doi.org/10.3390/molecules18055858>
- Akul, Y., Kumar, V. S., & Chong, K. P. (2018). Designing Primers for Loop-Mediated Isothermal Amplification (LAMP) for Detection of *Ganoderma boninense*. *Bulgarian Journal of Agricultural Science*, 24(5), 854–859. [https://journal.agrojournal.org/page/en/details.php?article\\_id=1250](https://journal.agrojournal.org/page/en/details.php?article_id=1250)
- Alexander, A., Dayou, J., Sipaut, C. S., Phin, C. K., & Chin, L. P. (2014). On the possibility of using FTIR for detection of *Ganoderma boninense* in infected oil palm tree. *International Journal of Advances in Agricultural and Environmental Engineering*, 1(2), 161–163. <https://doi.org/10.15242/IJAAEE.C514559>
- Ali, M. A., Dong, L., Dhau, J., Khosla, A., & Kaushik, A. (2020). Perspective—Electrochemical Sensors for Soil Quality Assessment. *Journal of The Electrochemical Society*, 167(3), 037550. <https://doi.org/10.1149/1945-7111/ab69fe>
- Alvarez, E., Mejía, J. F., Contaldo, N., Paltrinieri, S., Duduk, B., & Bertaccini, A. (2014). ‘*Candidatus Phytoplasma asteris*’ Strains Associated with Oil Palm Lethal Wilt in Colombia. *Plant Disease*, 98(3), 311–318. <https://doi.org/10.1094/PDIS-12-12-1182-RE>
- Amanda, W. I., & Prakoso, H. T. (2018). Modified *Ganoderma* selective medium to meet Indonesia’s government regulation. *IOP Conference Series: Earth and Environmental Science*, 183, 012020. <https://doi.org/10.1088/1755-1315/183/1/012020>
- Amanulla, B., Palanisamy, S., Chen, S.-M., Chiu, T.-W., Velusamy, V., Hall, J. M.,

- Chen, T.-W., & Ramaraj, S. K. (2017). Selective Colorimetric Detection of Nitrite in Water using Chitosan Stabilized Gold Nanoparticles Decorated Reduced Graphene oxide. *Scientific Reports*, 7(1), 14182. <https://doi.org/10.1038/s41598-017-14584-6>
- Ariffin, D., Idris, A. B., & Marzuki, A. (1996). Spread of *Ganoderma boninense* and vegetative compatibility studies of a single field palm isolates. In D. Ariffin, M. W. Basri, N. Rajanaidu, D. Mohd Tayeb, K. Paranjothy, S. C. Cheah, K. C. Chang, & S. Ravigadevi (Eds.), *Proceedings of the 1996 PORIM International Palm Oil Congress: Competitiveness for the 21st Century (Agriculture Conference)* (pp. 508–515).
- Assis, K., Chong, K. P., Idris, A. S., & Ho, C. M. (2016). Economic Loss Due to *Ganoderma* Disease in Oil Palm. *International Journal of Economics and Management Engineering*, 10(2), 631–635. <https://doi.org/10.5281/zenodo.1111999>
- Azuan, N. H., Khairunniza-Bejo, S., Abdullah, A. F., Kassim, M. S. M., & Ahmad, D. (2019). Analysis of Changes in Oil Palm Canopy Architecture From Basal Stem Rot Using Terrestrial Laser Scanner. *Plant Disease*, 103(12), 3218–3225. <https://doi.org/10.1094/PDIS-10-18-1721-RE>
- Badalyan, S., Gharibyan, N., & Zambonelli, A. (2012). Morphological and Genetic Characteristics of Different Collections of *Ganoderma* P . Karst . Species. *Proceedings of the 18th Congress of the International Society for Mushroom Science*, 247–255.
- Baetsen-Young, A. M., Vasher, M., Matta, L. L., Colgan, P., Alocilja, E. C., & Day, B. (2018). Direct colorimetric detection of unamplified pathogen DNA by dextrin-capped gold nanoparticles. *Biosensors and Bioelectronics*, 101(September 2017), 29–36. <https://doi.org/10.1016/j.bios.2017.10.011>
- Bakhori, N., Yusof, N., Abdullah, A., & Hussein, M. (2013). Development of a Fluorescence Resonance Energy Transfer (FRET)-Based DNA Biosensor for Detection of Synthetic Oligonucleotide of *Ganoderma boninense*. *Biosensors*, 3(4), 419–428. <https://doi.org/10.3390/bios3040419>
- Baptista, P. V., McCusker, M. P., Carvalho, A., Ferreira, D. A., Mohan, N. M., Martins, M., & Fernandes, A. R. (2018). Nano-Strategies to Fight Multidrug Resistant Bacteria - “A Battle of the Titans.” *Frontiers in Microbiology*, 9(JUL), 1–26. <https://doi.org/10.3389/fmicb.2018.01441>
- Barcelos, E., Rios, S. de A., Cunha, R. N. V., Lopes, R., Motoike, S. Y., Babiychuk, E., Skirycz, A., & Kushnir, S. (2015). Oil palm natural diversity and the potential for yield improvement. *Frontiers in Plant Science*, 6, 1–16. <https://doi.org/10.3389/fpls.2015.00190>
- Barthel, M., Jennings, S., Schreiber, W., Sheane, R., Royston, S., Llp, K., Fry, J., Leng Khor, Y., & McGill, J. (2018). *Study on the environmental impact of palm oil consumption and on existing sustainability standards For European Commission*,

- Bartlett, J. M. S., & Stirling, D. (2003). A Short History of the Polymerase Chain Reaction. In J. M. S. Bartlett & D. Stirling (Eds.), *PCR Protocols. Methods in Molecular Biology<sup>TM</sup>* (pp. 3–6). Humana Press.
- Bhalla, N., Jolly, P., Formisano, N., & Estrela, P. (2016). Introduction to biosensors. *Essays in Biochemistry*, 60(1), 1–8. <https://doi.org/10.1042/EBC20150001>
- Bhatia, D., Mehtab, S., Krishnan, R., Indi, S. S., Basu, A., & Krishnan, Y. (2009). Icosahedral DNA Nanocapsules by Modular Assembly. *Angewandte Chemie International Edition*, 48(23), 4134–4137. <https://doi.org/10.1002/anie.200806000>
- Bivi, M. R., Farhana, M. S., Khairulmazmi, A., & Idris, A. (2010). Control of *ganoderma boninense*: A causal agent of basal stem rot disease in oil palm with endophyte bacteria in vitro. *International Journal of Agriculture and Biology*, 12(6), 833–839.
- Blundell, E. L. C. J., Vogel, R., & Platt, M. (2016). Particle-by-Particle Charge Analysis of DNA-Modified Nanoparticles Using Tunable Resistive Pulse Sensing. *Langmuir: The ACS Journal of Surfaces and Colloids*, 32(4), 1082–1090. <https://doi.org/10.1021/acs.langmuir.5b03024>
- Brann, T., Patel, D., Chauhan, R., James, K. T., Bates, P. J., Malik, M. T., Keynton, R. S., & O'Toole, M. G. (2016). Gold Nanoplates as Cancer-Targeted Photothermal Actuators for Drug Delivery and Triggered Release. *Journal of Nanomaterials*, 2016, 1–11. <https://doi.org/10.1155/2016/2036029>
- Breure, C. J., & Soebagjo, F. X. (1991). Factors associated with occurrence of crown disease in oil palm (*Elaeis guineensis* Jacq.) and its effect on growth and yield. *Euphytica*, 54(1), 55–64. <https://doi.org/10.1007/BF00145631>
- Bridge, P. D., O'Grady, E. B., Pilotti, C. A., & Sanderson, F. R. (2009). Development of molecular diagnostics for the detection of *Ganoderma* isolates pathogenic to oil palm. In J. Flood, P. D. Bridge, & M. Holderness (Eds.), *Ganoderma diseases of perennial crops* (Issue November, pp. 225–234). CABI. <https://doi.org/10.1079/9780851993881.0225>
- Brown, C. (1998). *In Situ* Hybridization with Riboprobes- An Overview. *Veterinary Pathologists*, 35(3), 159–167. <https://journals.sagepub.com/doi/pdf/10.1177/030098589803500301>
- Bui, C. T., Lambrinakos, A., & Cotton, R. G. H. (2003). Spectroscopic study of permanganate oxidation reactions of oligonucleotides containing single base mismatches. *Biopolymers*, 70(4), 628–636. <https://doi.org/10.1002/bip.10543>
- Burton, K. (1956). A study of the conditions and mechanism of the diphenylamine reaction for the colorimetric estimation of deoxyribonucleic acid. *Biochemical*

*Journal*, 62(2), 315–323. <https://doi.org/10.1042/bj0620315>

CABI. (2021). *Ganoderma boninense*. Knowledge Bank. <https://www.plantwise.org/knowledgebank/datasheet/24924#SymptomsSection>

Cabrera, F. C., Melo, A. F. A. A., de Souza, J. C. P., Job, A. E., & Crespilho, F. N. (2015). A flexible lab-on-a-chip for the synthesis and magnetic separation of magnetite decorated with gold nanoparticles. *Lab on a Chip*, 15(8), 1835–1841. <https://doi.org/10.1039/C4LC01483A>

Cantsilieris, S., Baird, P. N., & White, S. J. (2013). Molecular methods for genotyping complex copy number polymorphisms. *Genomics*, 101(2), 86–93. <https://doi.org/10.1016/j.ygeno.2012.10.004>

Cao, Y. C., Jin, R., Thaxton, C. S., & Mirkin, C. A. (2005). A two-color-change, nanoparticle-based method for DNA detection. *Talanta*, 67(3), 449–455. <https://doi.org/10.1016/j.talanta.2005.06.063>

Cao, Y., Wu, S.-H., & Dai, Y.-C. (2012). Species Clarification of The Prize Medicinal *Ganoderma mushroom* “Lingzhi.” *Fungal Diversity*, 56(1), 49–62. <https://doi.org/10.1007/s13225-012-0178-5>

Carneiro, M. C. C. G., Moreira, F. T. C., Dutra, R. A. F., Fernandes, R., & Sales, M. G. F. (2018). Homemade 3-carbon electrode system for electrochemical sensing: Application to microRNA detection. *Microchemical Journal*, 138, 35–44. <https://doi.org/10.1016/j.microc.2017.12.026>

Carpenter, D. K. (1977). Dynamic Light Scattering with Applications to Chemistry, Biology, and Physics (Berne, Bruce J.; Pecora, Robert). *Journal of Chemical Education*, 54(10), A430. <https://doi.org/10.1021/ed054pA430.1>

Chah, S., Hammond, M. R., & Zare, R. N. (2005). Gold Nanoparticles as a Colorimetric Sensor for Protein Conformational Changes. *Chemistry & Biology*, 12(3), 323–328. <https://doi.org/10.1016/j.chembiol.2005.01.013>

Chen, S.-H., Lin, K.-I., Tang, C.-Y., Peng, S.-L., Chuang, Y.-C., Lin, Y.-R., Wang, J.-P., & Lin, C.-S. (2009). Optical Detection of Human Papillomavirus Type 16 and Type 18 by Sequence Sandwich Hybridization with Oligonucleotide-Functionalized Au nanoparticles. *IEEE Transactions on Nanobioscience*, 8(2), 120–131. <https://doi.org/10.1109/TNB.2008.2011733>

Chen, X., Zhu, L., Huang, M., & Yang, C. (2019). Synthesis of Gold Nanoparticles and Functionalization With DNA for Bioanalytical Applications. In X. Wang & X. Chen (Eds.), *Novel Nanomaterials for Biomedical, Environmental and Energy Applications* (pp. 111–136). Elsevier. <https://doi.org/10.1016/B978-0-12-814497-8.00004-7>

Chong, K. P., Dayou, J., & Alexander, A. (2017a). Current Detection Methods of *G. boninense* Infection in Oil Palm. In K. P. Chong, J. Dayou, & A. Alexander (Eds.), *Detection and Control of Ganoderma boninense in Oil Palm Crop* (pp. 13–20).



Springer, Cham. [https://doi.org/10.1007/978-3-319-54969-9\\_3](https://doi.org/10.1007/978-3-319-54969-9_3)

- Chong, K. P., Dayou, J., & Alexander, A. (2017b). Pathogenic Nature of *Ganoderma boninense* and Basal Stem Rot Disease. In K. P. Chong, J. Dayou, & A. Alexander (Eds.), *Detection and Control of Ganoderma boninense in Oil Palm Crop* (1st ed., pp. 5–12). Springer, Cham. [https://doi.org/10.1007/978-3-319-54969-9\\_2](https://doi.org/10.1007/978-3-319-54969-9_2)
- Choo Yuen May, & Malaysian Palm Oil Board. (2015). *Overview of the Malaysian Oil Palm Industry 2015* (Issue January). [www.mpob.gov](http://www.mpob.gov)
- Chung, G. F. (2012). Effect of Pests and Diseases on Oil Palm Yield. In O.-M. Lai, C.-P. Tan, & C. C. Akoh (Eds.), *Palm Oil* (pp. 163–210). Elsevier. <https://doi.org/10.1016/B978-0-9818936-9-3.50009-5>
- Clogston, J. D., & Patri, A. K. (2011). Zeta Potential Measurement. In S. E. McNeil (Ed.), *Methods in Molecular Biology (Methods and Protocols): Characterization of Nanoparticles Intended for Drug Delivery* (pp. 63–70). Humana Press. [https://doi.org/10.1007/978-1-60327-198-1\\_6](https://doi.org/10.1007/978-1-60327-198-1_6)
- Corley, R. H. V., & Tinker, P. B. (2015a). Diseases of the Oil Palm. In R. H. V. Corley & P. B. Tinker (Eds.), *The Oil Palm* (Fifth Edit, pp. 399–436). John Wiley & Sons, Ltd. <https://doi.org/10.1002/9781118953297.ch13>
- Corley, R. H. V., & Tinker, P. B. (2015b). Oil Palm and Climate Change. In R. H. V. Corley & P. B. Tinker (Eds.), *The Oil Palm* (Fifth Edit, pp. 495–506). John Wiley & Sons, Ltd. <https://doi.org/10.1002/9781118953297.ch17>
- Corley, R. H. V., & Tinker, P. B. (2015c). Pests of the Oil Palm. In R. H. V. Corley & P. B. Tinker (Eds.), *The Oil Palm* (Fifth Edit, pp. 437–459). John Wiley & Sons, Ltd. <https://doi.org/10.1002/9781118953297.ch14>
- Corley, R. H. V., & Tinker, P. B. (2015d). The Products of the Oil Palm and Their Extraction. In R. H. V. Corley & P. B. Tinker (Eds.), *The Oil Palm* (Fifth Edit, pp. 460–482). John Wiley & Sons, Ltd. <https://doi.org/10.1002/9781118953297.ch15>
- Dade, H. A. (1928). *Ceratostomella paradoxa*, the perfect stage of *Thielaviopsis paradoxa* (de Seynes) von Höhnelt. *Transactions of the British Mycological Society*, 13(3–4), 184–IN7. [https://doi.org/10.1016/S0007-1536\(28\)80017-9](https://doi.org/10.1016/S0007-1536(28)80017-9)
- Daima, H. K., Selvakannan, P. R., Shukla, R., Bhargava, S. K., & Bansal, V. (2013). Fine-Tuning the Antimicrobial Profile of Biocompatible Gold Nanoparticles by Sequential Surface Functionalization Using Polyoxometalates and Lysine. *PLoS ONE*, 8(10), e79676. <https://doi.org/10.1371/journal.pone.0079676>
- Daniel, M.-C., & Astruc, D. (2004). Gold nanoparticles: assembly, supramolecular chemistry, quantum-size-related properties, and applications toward biology, catalysis, and nanotechnology. *Chemical Reviews*, 104(1), 293–346. <https://doi.org/10.1021/cr030698+>
- Darus, A., & Abu Seman, I. (1992). The *Ganoderma* selective medium (GSM). *PORIM*

- Darus, A., Abu Seman, I., & Hassan, A. H. (1989). Significance of the Black Line Within Oil Palm Tissue Decayed by *Ganoderma boninense*. *International Journal of Oil Palm Research and Development*, 1, 11–16.
- Darus, A., Abu Seman, I., & Singh, G. (2009). Status of *Ganoderma* in Oil Palm. In J. Flood, P. D. Bridge, & M. Holderness (Eds.), *Ganoderma Diseases of Perennial Crops* (pp. 49–68). CABI. <https://doi.org/10.1079/9780851993881.0049>
- Darus, A., & Wahid, M. B. (2000). Intensive IPM for Management of Oil Palm Pests. *Oil Palm Bulletin*, 41. file:///Users/samakhtarmohshim/Documents/Mendeley Desktop/Ariffin Darus and Mohd Basri Wahid (2000) - Intensive IPM for Management of Oil Palm Pests.pdf
- Denijn, M., De Weger, R. A., Berends, M. J., Compier-Spies, P. I., Jansz, H., Van Unnik, J. A., & Lips, C. J. (1990). Detection of calcitonin-encoding mRNA by radioactive and non-radioactive in situ hybridization: improved colorimetric detection and cellular localization of mRNA in thyroid sections. *Journal of Histochemistry & Cytochemistry*, 38(3), 351–358. <https://doi.org/10.1177/38.3.2406337>
- Dharanivasan, G., Mohammed Riyaz, S. U., Michael Immanuel Jesse, D., Raja Muthuramalingam, T., Rajendran, G., & Kathiravan, K. (2016). DNA templated self-assembly of gold nanoparticle clusters in the colorimetric detection of plant viral DNA using a gold nanoparticle conjugated bifunctional oligonucleotide probe. *RSC Advances*, 6(14), 11773–11785. <https://doi.org/10.1039/c5ra25559g>
- Din, A. K. (2018). Oil Palm Economic Performance in Malaysia and R&D Progress in 2017. *Journal of Oil Palm Research*, 30(2), 163–195. <https://doi.org/10.21894/jopr.2018.0030>
- Dorsey, J. F., Sun, L., Joh, D. Y., Witztum, A., Kao, G. D., Alonso-Basanta, M., Avery, S., Hahn, S. M., Al Zaki, A., & Tsourkas, A. (2013). Gold nanoparticles in radiation research: potential applications for imaging and radiosensitization. *Translational Cancer Research*, 2(4), 280–291. <https://doi.org/10.3978/j.issn.2218-676X.2013.08.09>
- Draz, M. S., Ma, L., & Lu, X. (2018). Ultrasensitive DNA Detection with Hydrodynamic Separation of Plasmonic Nanoparticles and Isothermal Amplification. *Journal of Biomedical Nanotechnology*, 14(6), 1025–1038. <https://doi.org/10.1166/jbn.2018.2507>
- Draz, M. S., & Shafiee, H. (2018). Applications of gold nanoparticles in virus detection. *Theranostics*, 8(7), 1985–2017. <https://doi.org/10.7150/thno.23856>
- Dutse, S. W., Yusof, N. A., Ahmad, H., Hussein, Mohd Zobir DNA-based biosensor for detection of *Ganoderma boninense*, an oil palm pathogen utilizing newly synthesized ruthenium complex [Ru(phen)<sub>2</sub>(qtpy)]<sup>2+</sup> based on a PEDOT-, Zainal, Z., & Hushiarian, R. (2013). DNA-Based Biosensor for Detection of *Ganoderma boninense*, an Oil Palm Pathogen Utilizing Newly Synthesized

Ruthenium Complex [Ru(phen)<sub>2</sub>(qtpy)]<sup>2+</sup> Based on a PEDOT-PSS/Ag Nanoparticles Modified Electrode. *International Journal of Electrochemical Science*, 8(9), 11048–11057. <http://www.electrochemsci.org/papers/vol8/80911048.pdf>

- Dutse, S. W., Yusof, N. A., Ahmad, H., Hussein, M. Z., & Zainal, Z. (2012). An Electrochemical DNA Biosensor for *Ganoderma boninense* Pathogen of the Oil Palm Utilizing a New Ruthenium Complex, [Ru(dppz)<sub>2</sub>(qtpy)]Cl<sub>2</sub>. *International Journal of Electrochemical Science*, 7(9), 8105–8115. <http://www.electrochemsci.org/papers/vol7/70981105.pdf>
- Ebralidze, I. I., Laschuk, N. O., Poisson, J., & Zenkina, O. V. (2019). Colorimetric Sensors and Sensor Arrays. In *Nanomaterials Design for Sensing Applications* (pp. 1–39). <https://doi.org/10.1016/B978-0-12-814505-0.00001-1>
- Elghanian, R., Storhoff, J. J., Mucic, R. C., Letsinger, R. L., & Mirkin, C. A. (1997). Selective Colorimetric Detection of Polynucleotides Based on the Distance-Dependent Optical Properties of Gold Nanoparticles. *Science*, 277(5329), 1078–1081. <https://doi.org/10.1126/science.277.5329.1078>
- Esashika, K., & Saiki, T. (2018). DNA Hybridization Assay Using Gold Nanoparticles and Electrophoresis Separation Provides 1-pM Sensitivity. *Bioconjugate Chemistry*, 29(1), 182–189. <https://doi.org/10.1021/acs.bioconjchem.7b00682>
- Evanoff, D. D., & Chumanov, G. (2005). Synthesis and Optical Properties of Silver Nanoparticles and Arrays. *ChemPhysChem*, 6(7), 1221–1231. <https://doi.org/10.1002/cphc.200500113>
- Famulok, M., & Mayer, G. (2011). Aptamer Modules as Sensors and Detectors. *Accounts of Chemical Research*, 44(12), 1349–1358. <https://doi.org/10.1021/ar2000293>
- Fang, Y. I. (2017). *Electrochemical Biosensors for Plant Volatile Organic Compounds*. University of Georgia.
- Faraday, M. (1857). The Bakerian Lecture: Experimental Relations of Gold (and Other Metals) to Light. *Philosophical Transactions of the Royal Society of London*, 147(1857), 145–181. <https://doi.org/10.1098/rstl.1857.0011>
- Flood, J., Hasan, Y., Turner, P. D., & O'Grady, E. B. (2000). The spread of *Ganoderma* from infective sources in the field and its implications for management of the disease in oil palm. In J. Flood, P. D. Bridge, & M. Holderness (Eds.), *Ganoderma diseases of perennial crops* (pp. 101–112). CABI. <https://doi.org/10.1079/9780851993881.0101>
- Fowotade, S. A., Yusof, N. A., Abdullah, J., Sulaiman, Y., & Abd Rahman, S. F. (2019). Enhanced electrochemical sensing of secondary metabolites in oil palms for early detection of *Ganoderma boninense* based on novel nanoparticle-chitosan functionalized multi-walled carbon nanotube platform. *Sensing and Bio-Sensing Research*, 23(March), 100274. <https://doi.org/10.1016/j.sbsr.2019.100274>



- Fritzsche, W. (2001). DNA-gold conjugates for the detection of specific molecular interactions. *Reviews in Molecular Biotechnology*, 82(1), 37–46. [https://doi.org/10.1016/S1389-0352\(01\)00028-9](https://doi.org/10.1016/S1389-0352(01)00028-9)
- Gannon, F. (1994). DNA probes for the identification of microorganisms. *Journal of Industrial Microbiology*, 13(2), 71–76. <https://doi.org/10.1007/BF01584101>
- Ghulam Khadir, A. P. (2020). *Overview Of The Malaysian Oil Palm Industry 2020*. [https://bepi.mpob.gov.my/images/overview/Overview\\_of\\_Industry\\_2020.pdf](https://bepi.mpob.gov.my/images/overview/Overview_of_Industry_2020.pdf)
- Ghulam Khadir, A. P., Hishamuddin, E., Loh, S. K., Ong-Abdullah, M., Mohamed Salleh, K., Zanal Bidin, M. N. I., Sundram, S., Hasan, Z. A. A., & Idris, Z. (2020). Oil Palm Economic Performance in Malaysia and R&D Progress in 2019. *Journal of Oil Palm Research*, 32(June), 159–190. <https://doi.org/10.21894/jopr.2020.0032>
- Ghulam Khadir, A. P., & Malaysian Palm Oil Board. (2019). *Overview of the Malaysian Oil Palm Industry 2019*. [www.mpob.gov](http://www.mpob.gov)
- Gilbert, W. (1986). Origin of life: The RNA world. *Nature*, 319(6055), 618–618. <https://doi.org/10.1038/319618a0>
- Giorgi-Coll, S., Marin, M. J., Sule, O., Hutchinson, P. J., & Carpenter, K. L. H. (2020). Aptamer-modified gold nanoparticles for rapid aggregation-based detection of inflammation: an optical assay for interleukin-6. *Microchimica Acta*, 187(1), 13. <https://doi.org/10.1007/s00604-019-3975-7>
- Gorea, E. A. (2016). *Microscopic Analysis of Oil Palm (elaeis guineensis) Infection by Ganoderma boninense* [University of Queensland]. [https://espace.library.uq.edu.au/data/UQ\\_566688/s4361884\\_mphil\\_finalthesis.pdf?Expires=1604336633&Key-Pair-Id=APKAJKNB4MJBNC6NLQ&Signature=C~uX87yn8zjwKRouqEuXoRc6qmCyKfld67LJnWfV8107DIAG4jFJX0-k-oFSXtXNbELW5q6FtU5jq4nzk1nKY8zeMHov5u0ed-z~4RAw6OG6abg-3](https://espace.library.uq.edu.au/data/UQ_566688/s4361884_mphil_finalthesis.pdf?Expires=1604336633&Key-Pair-Id=APKAJKNB4MJBNC6NLQ&Signature=C~uX87yn8zjwKRouqEuXoRc6qmCyKfld67LJnWfV8107DIAG4jFJX0-k-oFSXtXNbELW5q6FtU5jq4nzk1nKY8zeMHov5u0ed-z~4RAw6OG6abg-3)
- Guillén, G. A. Z. (2011). *Ultrasensitive Detection of Pathogens in Real-Time Potentiometric Biosensors Based on Single-Walled Carbon Nanotubes and Aptamers* [Universitat Rovira I Virgili]. [https://www.tesisenred.net/bitstream/handle/10803/51768/Gustavo\\_A\\_Zelada-Guillen\\_%28PHD\\_THESIS%29.pdf?sequence=1&isAllowed=y](https://www.tesisenred.net/bitstream/handle/10803/51768/Gustavo_A_Zelada-Guillen_%28PHD_THESIS%29.pdf?sequence=1&isAllowed=y)
- Gur, M. (1986). The physiological basis of wavelength discrimination: Evidence from dichoptic and ganzfeld viewing. *Vision Research*, 26(8), 1257–1262. [https://doi.org/10.1016/0042-6989\(86\)90106-9](https://doi.org/10.1016/0042-6989(86)90106-9)
- Hara, T. O., & Singh, B. (2021). Electrochemical Biosensors for Detection of Pesticides and Heavy Metal Toxicants in Water: Recent Trends and Progress. *ACS ES&T Water*, 1(3), 462–478. <https://doi.org/10.1021/acsestwater.0c00125>
- Hasenoehl, C., Alexander, C. M., Azzarelli, N. N., & Dabrowiak, J. C. (2012). Enhanced

detection of gold nanoparticles in agarose gel electrophoresis. *Electrophoresis*, 33(8), 1251–1254. <https://doi.org/10.1002/elps.201100556>

Hashim, K. (1990). *Basal Stem Rot of Oil Palm : Incidence, Etiology and Control* [Universiti Putra Malaysia]. [http://ethesis.upm.edu.my.ezadmin.upm.edu.my/id/eprint/4865/1/FP\\_1990\\_5\\_F.pdf](http://ethesis.upm.edu.my.ezadmin.upm.edu.my/id/eprint/4865/1/FP_1990_5_F.pdf)

Hirschmann, R. (2020). *Palm oil industry in Malaysia - Statistics & Facts*.

Hittinger, J. P. (2015). Optimization and Characterization of Au Nanoparticle-DNA Conjugate Devices. In *NNIN REU Research Accomplishments*.

Ho, Y. W., & Nawawi, A. (1985). *Ganoderma boninense* Pat . from Basal Stem Rot of Oil Palm ( *Elaeis guineensis* ) in Peninsular Malaysia. *Pertanika*, 8(3), 425–428.

Hoang, M., Huang, P.-J. J., & Liu, J. (2016). G-Quadruplex DNA for Fluorescent and Colorimetric Detection of Thallium(I). *ACS Sensors*, 1(2), 137–143. <https://doi.org/10.1021/acssensors.5b00147>

Hřibová, E., Čížková, J., Christelová, P., Taudien, S., de Langhe, E., & Doležal, J. (2011). The ITS1-5.8S-ITS2 Sequence Region in the Musaceae: Structure, Diversity and Use in Molecular Phylogeny. *PLoS ONE*, 6(3), e17863. <https://doi.org/10.1371/journal.pone.0017863>

Hurst, S. J., Lytton-Jean, A. K. R., & Mirkin, C. A. (2006). Maximizing DNA Loading on a Range of Gold Nanoparticle Sizes. *Analytical Chemistry*, 78(24), 8313–8318. <https://doi.org/10.1021/ac0613582>

Hushiarian, R. (2015). *Development of A DNA Biosensor Based on Magnetic Nanoparticles for The Detection of Ganoderma boninense*. Universiti Putra Malaysia.

Hushiarian, R., Yusof, N. A., & Dutse, S. W. (2013). Detection and Control of *Ganoderma boninense* : Strategies and Perspectives. *SpringerPlus*, 2(1), 1–12. <https://doi.org/10.1186/2193-1801-2-555>

Husin, N. A., Khairunniza-Bejo, S., Abdullah, A. F., Kassim, M. S. M., Ahmad, D., & Aziz, M. H. A. (2020). Classification of Basal Stem Rot Disease in Oil Palm Plantations Using Terrestrial Laser Scanning Data and Machine Learning. *Agronomy*, 10(11), 1624. <https://doi.org/10.3390/agronomy10111624>

Hutter, E., & Maysinger, D. (2013). Gold-nanoparticle-based biosensors for detection of enzyme activity. *Trends in Pharmacological Sciences*, 34(9), 497–507. <https://doi.org/10.1016/j.tips.2013.07.002>

Ibrahim, M. S., Abu Seman, I., Rusli, M. H., Izzuddin, M. A., Kamarudin, N., Hashim, K., & Abd Manaf, Z. (2020). Surveillance of *Ganoderma* Disease in Oil Palm Planted by Participants of The Smallholders Replanting Incentive Scheme in Malaysia. *Journal of Oil Palm Research*, 32(June), 237–244.

- Idris, A.S, Ismail, S., & Ariffin, D. (2014). *Update R&D on Ganoderma and Other Field Diseases in Oil Palm* (Issue August). <http://soppoa.org.my/wp-content/uploads/2014/08/Workshop-Paper-2-Ganoderma-major-diseases.pdf>
- Idris, Abu Seman, Mazliham, M. S., Loonis, P., & Mohd Basri, W. (2010). GanoSken for early detection of *Ganoderma* infection in oil palm. *MPOB Information Series, June 2010*(MPOB TT No. 442), 3–6.
- Ishaq, I., Alias, M. S., Kadir, J., & Kasawani, I. (2014). Detection of Basal Stem Rot Disease at Oil Palm Plantations Using Sonic Tomography. *Journal of Sustainability Science and Management*, 9(2), 52–57. <http://jssm.umt.edu.my/wp-content/uploads/sites/51/2015/02/5.pdf>
- Ismail, A. (2013). The Effect of Labour Shortage in the Supply and Demand of Palm Oil in Malaysia. *Oil Palm Industry Economic Journal*, 3, 15–26. <http://palmoilis.mpob.gov.my/publications/OPIEJ/opiejv13n2-azman.pdf>
- Jang, N. H. (2002). The Coordination Chemistry of DNA Nucleosides on Gold Nanoparticles as a Probe by SERS. *Bulletin of the Korean Chemical Society*, 23(12), 1790–1800. <https://doi.org/10.5012/bkcs.2002.23.12.1790>
- Jans, H., Liu, X., Austin, L., Maes, G., & Huo, Q. (2009). Dynamic light scattering as a powerful tool for gold nanoparticle bioconjugation and biomolecular binding studies. *Analytical Chemistry*, 81(22), 9425–9432. <https://doi.org/10.1021/ac901822w>
- Jargalmaa, S., Eimes, J. A., Park, M. S., Park, J. Y., Oh, S.-Y., & Lim, Y. W. (2017). Taxonomic evaluation of selected *Ganoderma* species and database sequence validation. *PeerJ*, 5(7), e3596. <https://doi.org/10.7717/peerj.3596>
- Jeffery Daim, L. D. (2017). *Proteomic analyses of Elaeis guineensis Jacq.(African Oil Palm) basal stem rot disease related to Ganoderma boninense* (Vol. 1) [University of Malaya]. <http://studentsrepo.um.edu.my/id/eprint/7531>
- Ji, M., Hou, P., Li, S., He, N., & Lu, Z. (2004). Colorimetric silver detection of methylation using DNA microarray coupled with linker-PCR. *Clinica Chimica Acta*, 342(1–2), 145–153. <https://doi.org/10.1016/j.cccn.2003.12.017>
- Kamarudin, N., Abu Seman, I., Mohd Masri, M. M., & Rusli, H. (2017). Biosecurity planning and mitigation of devastating pests and diseases for oil palm in Malaysia.pdf. *Planter*, 93, 489–497. <http://www.isp.org.my>
- Kamarudin, N., Seman, I. A. B. U., Mazmira, M., Masri, M., & Rusli, H. (2018). *Safeguarding The Malaysian Oil Palm Industry from Devastating Pests and Diseases Malaysian Oil Palm Industry*. <http://intranet.mpob.gov.my/wp-content/uploads/2019/03/Biosecurity-Plan-Seminar-2018.pdf>
- Karthikeyan, M., Radhika, K., Bhaskaran, R., Mathiyazhagan, S., Samiyappan, R., &

- Velazhahan, R. (2010). Rapid detection of *Ganoderma* disease of coconut and assessment of inhibition effect of various control measures by immunoassay and PCR. *Plant Protection Science*, 42(No. 2), 49–57. <https://doi.org/10.17221/2771-PPS>
- Kim, H. K., Shim, M. Y., Seo, G. S., & Kim, H. G. (2002). Comparison of Characteristics of *Ganoderma lucidum* According to Geographical Origins (III): Classification between Species of Genus *Ganoderma* Using Dikaryon-Monokaryon Mating. *Mycobiology*, 30(2), 61. <https://doi.org/10.4489/MYCO.2002.30.2.061>
- Kinge, T. R., Mih, A. M., & Coetzee, M. P. A. (2012). Phylogenetic Relationships Among Species of *Ganoderma* (*Ganodermataceae*, Basidiomycota) from Cameroon. *Australian Journal of Botany*, 60(6), 526. <https://doi.org/10.1071/BT12011>
- Kleppe, K., Ohtsuka, E., Kleppe, R., Molineux, I., & Khorana, H. G. (1971). Studies on Polynucleotides: XCVI. Repair Replication of Short Synthetic DNA's as Catalyzed by DNA Polymerases. *Journal of Molecular Biology*, 56(2), 341–361. [https://doi.org/10.1016/0022-2836\(71\)90469-4](https://doi.org/10.1016/0022-2836(71)90469-4)
- Kok, S. M., Goh, Y. K., Tung, H. J., GoH, K. J., Wong, W. C., & Goh, Y. K. (2013). In vitro growth of *Ganoderma boninense* isolates on novel palm extract medium and virulence on oil palm (*Elaeis guineensis*) seedlings. *Malaysian Journal of Microbiology*, 9(2), 1–30. <https://doi.org/10.21161/mjm.45212>
- Kong, F.-Y., Zhang, J.-W., Li, R.-F., Wang, Z.-X., Wang, W.-J., & Wang, W. (2017). Unique Roles of Gold Nanoparticles in Drug Delivery, Targeting and Imaging Applications. *Molecules (Basel, Switzerland)*, 22(9), 1445. <https://doi.org/10.3390/molecules22091445>
- Koppel, D. E. (1972). Analysis of Macromolecular Polydispersity in Intensity Correlation Spectroscopy: The Method of Cumulants. *The Journal of Chemical Physics*, 57(11), 4814–4820. <https://doi.org/10.1063/1.1678153>
- Korlapati, S., Sushil, S. N., Jeyakumar, P., Shankar, G., Sharma, O. P., Raj Boina, D., Sain, S. K., Reddy, M. N., Asre, R., Murali, R., Arya, S., & Kumar, S. (2015). *AESA BASED IPM PACKAGE - Oil Palm*. <https://niphm.gov.in/IPMPackages/Oilpalm.pdf>
- Kresnawaty, I., Mulyatni, A. S., Eris, D. D., Prakoso, H. T., Tri-Panji, Triyana, K., & Widiastuti, H. (2020). Electronic Nose for Early Detection of Basal Stem Rot Caused by *Ganoderma* in Oil Palm. *IOP Conference Series: Earth and Environmental Science*, 468(1), 012029. <https://doi.org/10.1088/1755-1315/468/1/012029>
- Kumunda, C., Adekunle, A. S., Mamba, B. B., Hlongwa, N. W., & Nkambule, T. T. I. (2021). Electrochemical Detection of Environmental Pollutants Based on Graphene Derivatives: A Review. *Frontiers in Materials*, 7(February). <https://doi.org/10.3389/fmats.2020.616787>

- Kwan, Y.-M., Meon, S., Ho, C.-L., & Wong, M.-Y. (2016). Selection of reference genes for quantitative real-time PCR normalization in *Ganoderma*-infected oil palm (*Elaeis guineensis*) seedlings. *Australasian Plant Pathology*, 45(3), 261–268. <https://doi.org/10.1007/s13313-016-0417-4>
- Land, M. F., & Osorio, D. (2014). Extraordinary Color Vision. *Science*, 343(6169), 381–382. <https://doi.org/10.1126/science.1249614>
- Lau, H. Y. (2016). *Development of point-of-care and multiplex diagnostic methods for the detection of plant pathogens*. The University of Queensland.
- Lee, J.-S., Ulmann, P. A., Han, M. S., & Mirkin, C. A. (2008). A DNA–Gold Nanoparticle-Based Colorimetric Competition Assay for the Detection of Cysteine. *Nano Letters*, 8(2), 529–533. <https://doi.org/10.1021/nl0727563>
- Lee, K. W. (2015, February 4). *Ganoderma boninense* Isolate UPMGB001 Internal Transcribed Spacer 1, Partial Sequence; 5.8S Ribosomal RNA Gene, Complete Sequence; and Internal Transcribed Spacer 2, Partial Sequence (KM220584.1). Nucleotide [Internet]; Bethesda (MD): National Library of Medicine (US), National Center for Biotechnology Information. <https://www.ncbi.nlm.nih.gov/nuccore/KM220584>
- Lelong, C. C., Roger, J.-M., Brégand, S., Dubertret, F., Lanore, M., Sitorus, N., Raharjo, D., & Caliman, J.-P. (2010). Evaluation of Oil-Palm Fungal Disease Infestation with Canopy Hyperspectral Reflectance Data. *Sensors*, 10(1), 734–747. <https://doi.org/10.3390/s100100734>
- Leslie A. Pray. (2008). *Discovery of DNA Double Helix: Watson and Crick | Learn Science at Scitable*. Nature Education. <https://www.nature.com/scitable/topicpage/discovery-of-dna-structure-and-function-watson-397/>
- Leung, C.-H., Zhong, H.-J., Lu, L., Chan, D. S.-H., & Ma, D.-L. (2013). Luminescent and colorimetric strategies for the label-free DNA-based detection of enzyme activity. *Briefings in Functional Genomics*, 12(6), 525–535. <https://doi.org/10.1093/bfpg/elt004>
- Lévesque, C. A., Vrain, T. C., & Boer, S. H. De. (1994). Development of a Species-Specific Probe for *Pythium ultimum* Using Amplified Ribosomal DNA. *Phytopathology*, 84(5), 474. <https://doi.org/10.1094/Phyto-84-474>
- Li, F.-H., Sun, X.-D., Niu, X.-Q., Cao, H.-X., & Yu, F.-Y. (2018). First Report of Basal Stem Rot on Oil Palm Caused by *Thielaviopsis paradoxa* in Hainan, China. *Plant Disease*, 102(10), 2029. <https://doi.org/10.1094/PDIS-01-18-0009-PDN>
- Li, F., Zhang, H., Dever, B., Li, X.-F., & Le, X. C. (2013). Thermal Stability of DNA Functionalized Gold Nanoparticles. *Bioconjugate Chemistry*, 24(11), 1790–1797. <https://doi.org/10.1021/bc300687z>
- Li, & Rothberg, L. J. (2004). Label-Free Colorimetric Detection of Specific Sequences



in Genomic DNA Amplified by the Polymerase Chain Reaction. *Journal of the American Chemical Society*, 126(35), 10958–10961. <https://doi.org/10.1021/ja048749n>

- Liaghat, S., Mansor, S., Ehsani, R., Shafri, H. Z. M., Meon, S., & Sankaran, S. (2014). Mid-infrared spectroscopy for early detection of basal stem rot disease in oil palm. *Computers and Electronics in Agriculture*, 101, 48–54. <https://doi.org/10.1016/j.compag.2013.12.012>
- Lim, F., Nor Fakhra, I., Abdul Rasid, O., Abu Seman, I., Parveez Ghulam, K. A., Azmi Shaharuddin, N., Shaharuddin, N. A., Lim, F., Nor Fakhra, I., Abdul Rasid, O., Seman Idris, A., Kadir Ahmad Parveez, G., & Azmi Shaharuddin, N. (2014). Isolation and Selection of Reference Genes for *Ganoderma boninense* Gene Expression Study Using Quantitative Real-Time PCR (qPCR). *Journal of Oil Palm Research*, 26(262), 170–181. <http://jopr.mpob.gov.my.ezadmin.upm.edu.my/isolation-and-selection-of-reference-genes-for-ganoderma-boninense-gene-expression-study-using-quantitative-real-time-pcr-qpcr/>
- Ling-Chie Wong, Bong, C.-F. J., & Abu Seman, I. (2012). *Ganoderma* Species Associated with Basal Stem Rot Disease of Oil Palm. *American Journal of Applied Sciences*, 9(6), 879–885. <https://doi.org/10.3844/ajassp.2012.879.885>
- Liu, J. (2012). Adsorption of DNA onto gold nanoparticles and graphene oxide: surface science and applications. *Physical Chemistry Chemical Physics*, 14(30), 10485. <https://doi.org/10.1039/c2cp41186e>
- Long, Z. (2006). DNA Molecules, Properties and Detection of Single. In *Encyclopedia of Analytical Chemistry* (pp. 1–9). John Wiley & Sons, Ltd. <https://doi.org/10.1002/9780470027318.a1407>
- Madiah, A. Z., Idris, A. S., & Rafidah, A. R. (2014). Polyclonal Antibodies of *Ganoderma boninense* Isolated From Malaysian Oil Palm for Detection of Basal Stem Rot Disease. *African Journal of Biotechnology*, 13(34), 3455–3463. <https://doi.org/10.5897/AJB2013.13604>
- Madiah, A. Z., Maizatun-Suriza, M., Idris, A. S., Bakar, M. F. A., Kamaruddin, S., Bharudin, I., Abu Bakar, F. D., & Murad, A. M. A. (2018). Comparison of DNA Extraction and Detection of *Ganoderma*, Causal of Basal Stem Rot Disease in Oil Palm Using Loop-Mediated Isothermal Amplification. *Malaysian Applied Biology*, 47(5), 119–127.
- Malatesta, M. (2016). Transmission electron microscopy for nanomedicine: novel applications for long-established techniques. *European Journal of Histochemistry*, 60(4), 8–12. <https://doi.org/10.4081/ejh.2016.2751>
- Malaysian Palm Oil Board. (n.d.). *Factsheets and Pictorial Guide Lincus lethifer - Pentatomid*. <http://sawitsecure.mpob.gov.my/pentatomid-lincus-lobuliger/>
- Malaysian Palm Oil Board. (2018). *MALAYSIA PRICES OF CRUDE PALM OIL*.

[https://bepi.mpob.gov.my/admin2/price\\_local\\_daily\\_view\\_cpo\\_msia.php?more=Y&jenis=1Y&tahun=2018](https://bepi.mpob.gov.my/admin2/price_local_daily_view_cpo_msia.php?more=Y&jenis=1Y&tahun=2018)

- Maluin, F. N., Hussein, M. Z., & Idris, A. S. (2020). An Overview of the Oil Palm Industry: Challenges and Some Emerging Opportunities for Nanotechnology Development. *Agronomy*, 10(3), 356. <https://doi.org/10.3390/agronomy10030356>
- Mandal, P. K., Babu, M. K., Jayanthi, M., & Satyavani, V. (2014). PCR based early detection of *Ganoderma* sp. causing basal stem rot of oil palm in India. *Journal of Plantation Crops*, 42(3), 392–394. <https://updatepublishing.com/journal/index.php/JPC/article/view/5639>
- Markom, M. A., Shakaff, A. Y. M., Adom, A. H., Ahmad, M. N., Hidayat, W., Abdullah, A. H., & Fikri, N. A. (2009). Intelligent electronic nose system for basal stem rot disease detection. *Computers and Electronics in Agriculture*, 66(2), 140–146. <https://doi.org/10.1016/j.compag.2009.01.006>
- Mawar, R., Ram, L., Deepesh, & Mathur, T. (2020). *Ganoderma*. In N. Amaresan, K. Annapurna, A. Sankaranarayanan, M. S. Kumar, & K. Kumar (Eds.), *Beneficial Microbes in Agro-Ecology* (pp. 625–649). Academic Press. <https://doi.org/10.1016/B978-0-12-823414-3.00031-9>
- May, C. Y. (2012). Malaysia: economic transformation advances oil palm industry. *INFORM Magazine*. <https://www.aocs.org/stay-informed/inform-magazine/featured-articles/malaysia-economic-transformation-advances-oil-palm-industry-september-2012?SSO=True>
- Mehrvar, M., & Abdi, M. (2004). Recent Developments, Characteristics, and Potential Applications of Electrochemical Biosensors. *Analytical Sciences*, 20(8), 1113–1126. <https://doi.org/10.2116/analsci.20.1113>
- Miao, X.-M., Xiong, C., Wang, W.-W., Ling, L.-S., & Shuai, X.-T. (2011). Dynamic-Light-Scattering-Based Sequence-Specific Recognition of Double-Stranded DNA with Oligonucleotide-Functionalized Gold Nanoparticles. *Chemistry - A European Journal*, 17(40), 11230–11236. <https://doi.org/10.1002/chem.201003010>
- Ming, K. K., & Chandramohan, D. (2002). Malaysian Palm Oil Industry at Crossroads and its Future Direction. *Oil Palm Industry Economic Journal*, 2(2), 2–7. <http://palmoilis.mpob.gov.my/publications/opiejv2n2-10.pdf>
- Ministry of Plantation Industries and Commodities. (2020). *Agricommodity PocketStats Q2/2020*. <https://www.mpic.gov.my>
- Mirkin, C. A., Letsinger, R. L., Mucic, R. C., & Storhoff, J. J. (1996). A DNA-based method for rationally assembling nanoparticles into macroscopic materials. *Nature*, 382(6592), 607–609. <https://doi.org/10.1038/382607a0>
- Mishra, G., Barfidokht, A., Tehrani, F., & Mishra, R. (2018). Food Safety Analysis Using Electrochemical Biosensors. *Foods*, 7(9), 141. <https://doi.org/10.3390/foods7090141>

- Mitra, D., Dimove, I. K., & Waldeisen, J. R. (2017). *Colorimetric Detection of Nucleic Acid Amplification* (Patent No. US20170044599A1). United States Patent Application Publication.  
<https://patentimages.storage.googleapis.com/57/c2/85/2fb3abff68fca2/US20170044599A1.pdf>
- Mohammed, C. L., Rimbawanto, A., & Page, D. E. (2014). Management of Basidiomycete Root- and Stem-Rot Diseases in Oil Palm, Rubber and Tropical Hardwood Plantation Crops. *Forest Pathology*, 44(6), 428–446.  
<https://doi.org/10.1111/efp.12140>
- Mohd.Rashid, M. R., Bong, C. F. J., Khairulmazmi, A., & Idris, A. S. (2015). Aggressiveness of *Ganoderma boninense* and *G. zonatum* Isolated From Upper- and Basal Stem Rot of Oil Palm (*Elaeis guineensis*) in Malaysia. *Journal of Oil Palm Research*, 27(3), 229–240.  
[https://www.researchgate.net/publication/283428503\\_Aggressiveness\\_of\\_Ganoderma\\_boninense\\_and\\_G\\_zonatum\\_isolated\\_from\\_upper\\_and\\_basal\\_stem\\_rot\\_of\\_oil\\_palm\\_Elaeis\\_guineensis\\_in\\_Malaysia](https://www.researchgate.net/publication/283428503_Aggressiveness_of_Ganoderma_boninense_and_G_zonatum_isolated_from_upper_and_basal_stem_rot_of_oil_palm_Elaeis_guineensis_in_Malaysia)
- Mohd Rashid, M. R., Khairulmazmi, A., Idris, A. S., Jalloh, M. B., & Wahida, N. H. (2017). *Ganoderma* Species of Basal and Upper Stem Rots in Oil Palm ( *Elaeis Guineensis* ) in Sarawak. *Journal of Academia UiTM Negeri Sembilan*, 5(December), 27–35.
- Mohshim, S. A., Alang Ahmad, S. A., Ahmad, M. Z., Sathyapriya, H., & Wong, M. Y. (2019, July 12). *Ganoderma boninense* isolate UPMLD1806 small subunit ribosomal RNA gene (MN148580.1). Nucleotide [Internet]; Bethesda (MD): National Library of Medicine (US), National Center for Biotechnology Information. <https://www.ncbi.nlm.nih.gov/nuccore/1698454397>
- Moncalvo, J.-M., & Buchanan, P. K. (2008). Molecular Evidence for Long Distance Dispersal Across the Southern Hemisphere in the *Ganoderma applanatum-australe* Species Complex (Basidiomycota). *Mycological Research*, 112(4), 425–436. <https://doi.org/10.1016/j.mycres.2007.12.001>
- Moncalvo, J.-M., & Ryvarden, L. (1997). *A Nomenclatural Study of the Ganodermataceae Donk (Volume 11 of Synopsis fungorum)* (Synopsis f). Fungiflora.
- Moncalvo, J.-M., Wang, H., & Hseu, R.-S. (1995). Phylogenetic Relationships in *Ganoderma* Inferred from the Internal Transcribed Spacers and 25S Ribosomal DNA Sequences. *Mycologia*, 87(2), 223. <https://doi.org/10.2307/3760908>
- Mondal, B., Ramlal, S., Lavu, P. S., & Kingston, J. (2018). Highly Sensitive Colorimetric Biosensor for Staphylococcal Enterotoxin B by a Label-Free Aptamer and Gold Nanoparticles. *Frontiers in Microbiology*, 9(179).  
<https://doi.org/10.3389/fmicb.2018.00179>
- Monge-Pérez, J. E., & Chinchilla, C. M. (1994). Common spear rot / crown disease in oil palm (*Elaeis guineensis* Jacq.): anatomy of the affected tissue. *Journal of Oil*



- Monošík, R., Stred'anský, M., & Šturdík, E. (2012). Application of Electrochemical Biosensors in Clinical Diagnosis. *Journal of Clinical Laboratory Analysis*, 26(1), 22–34. <https://doi.org/10.1002/jcla.20500>
- Morales, F. J., Lozano, I., Sedano, R., Castano, M., & Arroyave, J. (2002). Partial Characterization of a Potyvirus Infecting African Oil Palm in South America. *Journal of Phytopathology*, 150(4–5), 297–301. <https://doi.org/10.1046/j.1439-0434.2002.00749.x>
- Mullis, K. B., & Faloona, F. A. (1987). Specific Synthesis of DNA *in vitro* via a Polymerase-Catalyzed Chain Reaction. In R. Wu (Ed.), *Methods in Enzymology* (Vol. 155, pp. 335–350). Academic Press. [https://doi.org/10.1016/0076-6879\(87\)55023-6](https://doi.org/10.1016/0076-6879(87)55023-6)
- Mullis, K. B., Faloona, F. A., Scharf, S., Saiki, R., Horn, G., & Erlich, H. (1992). Specific enzymatic amplification of DNA *in vitro*: the polymerase chain reaction. 1986. *Biotechnology (Reading, Mass.)*, 24(Table 1), 17–27. <http://www.ncbi.nlm.nih.gov/pubmed/1422010>
- Muniroh, M. S., Sariah, M., Zainal Abidin, M. A., Lima, N., & Paterson, R. R. M. (2014). Rapid detection of *Ganoderma*-infected oil palms by microwave ergosterol extraction with HPLC and TLC. *Journal of Microbiological Methods*, 100(1), 143–147. <https://doi.org/10.1016/j.mimet.2014.03.005>
- Muro, M. A. (2005). Probe Design, Production, and Applications. In W. J.M. & R. R. (Eds.), *Medical Biomethods Handbook* (pp. 13–23). Humana Press. <https://doi.org/10.1385/1-59259-870-6:013>
- Murphy, D. J. (2014). The future of oil palm as a major global crop: Opportunities and challenges. *Journal of Oil Palm Research*, 26(1), 1–24. <http://jopr.mpob.gov.my/the-future-of-oil-palm-as-a-major-global-crop-opportunities-and-challenges/?v=true>
- Murphy, D. J., Goggin, K., & Paterson, R. R. M. (2020). *Oil palm in the 2020s and beyond: challenges and solutions*. 19(April), 1–37. <https://doi.org/10.20944/preprints202012.0129.v1>
- Mykhaylova, N. (2018). *Low-cost Sensor Array Devices as a Method for Reliable Assessment of Exposure to Traffic-related Air Pollution Mixtures Research Objectives*. University of Toronto.
- Naidu, Y., Idris, A. S., Madihah, A. Z., & Kamarudin, N. (2016). In vitro Antagonistic Interactions Between Endophytic Basidiomycetes of Oil Palm (*Elaeis guineensis*) and *Ganoderma boninense*. *Journal of Phytopathology*, 164(10), 779–790. <https://doi.org/10.1111/jph.12498>
- Natarajan, S., Bhaskaran, R., & Shanmugam, N. (1986). Preliminary studies to develop techniques for early detection of Thanjavur wilt in Coconut. *Indian Coconut*

- Nigerian Institute For Oil Palm Research. (1977). *Thirteenth Annual Report of the Nigerian Institute For Oil Palm Research*. [https://drive.google.com/file/d/1\\_Fig8BLl8V6W4PsPx-wflpwUpsoZirDc/view](https://drive.google.com/file/d/1_Fig8BLl8V6W4PsPx-wflpwUpsoZirDc/view)
- Noor Azmi, A. N., Bejo, S. K., Jahari, M., Muharam, F. M., Yule, I., & Husin, N. A. (2020). Early Detection of *Ganoderma boninense* in Oil Palm Seedlings Using Support Vector Machines. *Remote Sensing*, 12(23), 3920. <https://doi.org/10.3390/rs12233920>
- Nordin, A. B. A., Shariff, F. M., Balu, N., & Nik Idris, N. A. (2012). Supply and Demand Performance for the Oils and Fats Industry in Malaysia. *Oil Palm Industry Economic Journal*, 12, 14–21. <http://palmoilis.mpob.gov.my/publications/OPIEJ/opiejv12n2-ahmad.pdf>
- Notomi, T., Okayama, H., Masubuchi, H., Yonekawa, T., Watanabe, K., Amino, N., & Hase, T. (2000). Loop-mediated isothermal amplification of DNA. In *Nucleic Acids Research* (Vol. 28, Issue 12).
- Nur, Y. (2013). *Gold Nanoparticles: Synthesis, Characterisation and their Effect on Pseudomonas Fluorensceus* (Issue April). University of Birmingham.
- Ohi, M., Li, Y., Cheng, Y., & Walz, T. (2004). Negative staining and image classification — powerful tools in modern electron microscopy. *Biological Procedures Online*, 6(1), 23–34. <https://doi.org/10.1251/bpo70>
- Osama, E. A., Mohamed, A. M., Abdel Rahim, M. A. E. S., & Shaban, R. M. S. (2011). Non Liquid Nitrogen-Based-Method for Isolation of DNA from Filamentous Fungi. *African Journal of Biotechnology*, 10(65), 14337–14341. <https://doi.org/10.5897/AJB11.1401>
- Pamies, R., Cifre, J. G. H., Espín, V. F., Collado-González, M., Baños, F. G. D., & de la Torre, J. G. (2014). Aggregation behaviour of gold nanoparticles in saline aqueous media. *Journal of Nanoparticle Research*, 16(4), 2376. <https://doi.org/10.1007/s11051-014-2376-4>
- Panchal, G., & Bridge, P. D. (2005). Following Basal Stem Rot in Young Oil Palm Plantings. *Mycopathologia*, 159(1), 123–127. <https://doi.org/10.1007/s11046-004-4434-4>
- Paterson, R. (2019). *Ganoderma boninense* Disease of Oil Palm to Significantly Reduce Production After 2050 in Sumatra if Projected Climate Change Occurs. *Microorganisms*, 7(1), 24. <https://doi.org/10.3390/microorganisms7010024>
- Paterson, R. R. M., Kumar, L., Shabani, F., & Lima, N. (2017). World climate suitability projections to 2050 and 2100 for growing oil palm. *The Journal of Agricultural Science*, 155(5), 689–702. <https://doi.org/10.1017/S0021859616000605>
- Paterson, R. Russell M. (2020). Future scenarios for oil palm mortality and infection by

- Phytophthora palmivora in Colombia, Ecuador and Brazil, extrapolated to Malaysia and Indonesia. *Phytoparasitica*, 48(4), 513–523. <https://doi.org/10.1007/s12600-020-00815-6>
- Paterson, R.R.M. (2007). *Ganoderma* disease of oil palm—A white rot perspective necessary for integrated control. *Crop Protection*, 26(9), 1369–1376. <https://doi.org/10.1016/j.cropro.2006.11.009>
- Pellegrino, T., Sperling, R. A., Alivisatos, A. P., & Parak, W. J. (2007). Gel Electrophoresis of Gold-DNA Nanoconjugates. *Journal of Biomedicine and Biotechnology*, 2007, 1–9. <https://doi.org/10.1155/2007/26796>
- Pilotti, C. A., Sanderson, F. R., & Aitken, E. A. B. (2003). Genetic structure of a population of *Ganoderma boninense* on oil palm. *Plant Pathology*, 52(4), 455–463. <https://doi.org/10.1046/j.1365-3059.2003.00870.x>
- Pilotti, Carmel A. (2005). Stem rots of oil palm caused by *Ganoderma boninense*: pathogen biology and epidemiology. *Mycopathologia*, 159(1), 129–137. <https://doi.org/10.1007/s11046-004-4435-3>
- Pilotti, Carmel A., Sanderson, F. R., & Aitken, E. A. B. (2002). Sexuality and Interactions of Monokaryotic and Dikaryotic Mycelia of *Ganoderma boninense*. *Mycological Research*, 106(11), 1315–1322. <https://doi.org/10.1017/S0953756202006755>
- Pirker, J., & Mosnier, A. (2015). *Global oil palm suitability assessment*. <http://pure.iiasa.ac.at/id/eprint/11682/1/IR-15-006.pdf>
- Priyadarshini, E., & Pradhan, N. (2017). Gold nanoparticles as efficient sensors in colorimetric detection of toxic metal ions: A review. *Sensors and Actuators B: Chemical*, 238, 888–902. <https://doi.org/10.1016/j.snb.2016.06.081>
- Rajendran, L., Kandan, A., Karthikeyan, G., Raguchander, T., & Samiyappan, R. (2009). Early Detection of *Ganoderma* Causing Basal Stem Rot Disease in Coconut Plantations. *Journal of Oil Palm Research*, 21(June), 627–635.
- Rajendran, P. (2015). *Plasmonic nanoparticles with tailored attinebility for direct oligonucleotide sensing - Optimization & Application* [ETH Zurich]. <https://doi.org/10.3929/ethz-a-010470339>
- Raju, J., Naik, S. T., Priti, S., Suryanarayana, V., Benagi, V. I., Nirmalanath, J., & Giri, M. S. (2015). Rapid Detection of *Ganoderma* Disease of Coconut by using ITS-PCR and Assessment of Inhibition Effect of Various Control Measures by Fungicides and Bioagents. *Journal of Pure and Applied Microbiology*, 9(4), 3325–3331. [https://krishi.icar.gov.in/jspui/bitstream/123456789/23409/1/JPAM\\_Vol\\_9\\_N\\_4\\_3325-3331.pdf](https://krishi.icar.gov.in/jspui/bitstream/123456789/23409/1/JPAM_Vol_9_N_4_3325-3331.pdf)
- Rani, E., Mohshim, S. A., Ahmad, M. Z., Goodacre, R., Ahmad, S. A. A., & Wong, L. S. (2019). Polymer pen lithography-fabricated DNA arrays for highly sensitive and

- selective detection of unamplified *Ganoderma boninense* DNA. *Polymers*, 11(3). <https://doi.org/10.3390/polym11030561>
- Rani, Ekta, Mohshim, S. A., Ahmad, M. Z., Goodacre, R., Alang Ahmad, S. A., & Wong, L. S. (2019). Polymer Pen Lithography-Fabricated DNA Arrays for Highly Sensitive and Selective Detection of Unamplified *Ganoderma boninense* DNA. *Polymers*, 11(3), 561. <https://doi.org/10.3390/polym11030561>
- Rani, Ekta, Mohshim, S. A., Yusof, N. H., Ahmad, M. Z., Goodacre, R., Alang Ahmad, S. A., & Wong, L. S. (2020). Sensitive and Selective Detection of DNA Fragments Associated with *Ganoderma boninense* by DNA-Nanoparticle Conjugate Hybridisation. *Journal of Materials Science*, 55(30), 14965–14979. <https://doi.org/10.1007/s10853-020-05058-8>
- Rani, M., Moudgil, L., Singh, B., Kaushal, A., Mittal, A., Saini, G. S. S., Tripathi, S. K., Singh, G., & Kaura, A. (2016). Understanding the mechanism of replacement of citrate from the surface of gold nanoparticles by amino acids: a theoretical and experimental investigation and their biological application. *RSC Advances*, 6(21), 17373–17383. <https://doi.org/10.1039/C5RA26502A>
- Rees, R. W., Flood, J., Hasan, Y., Potter, U., & Cooper, R. M. (2009). Basal stem rot of oil palm (*Elaeis guineensis*); mode of root infection and lower stem invasion by *Ganoderma boninense*. *Plant Pathology*, 58(5), 982–989. <https://doi.org/10.1111/j.1365-3059.2009.02100.x>
- Robertson, J. S. (1962). Dry basal rot, a new disease of oil palms caused by *Ceratocystis paradoxa* (dade) moreau. *Transactions of the British Mycological Society*, 45(4), 475–IN3. [https://doi.org/10.1016/S0007-1536\(62\)80007-2](https://doi.org/10.1016/S0007-1536(62)80007-2)
- Rodriguez-Silva, A. A. (2016). *Graphene Oxide-based Novel Supercapacitor Immunosensors for Physiological Biomarkers Detection* (Issue April). Ohio University.
- Ropital, F. (2011). Environmental degradation in hydrocarbon fuel processing plant: issues and mitigation. In *Advances in Clean Hydrocarbon Fuel Processing* (pp. 437–462). Elsevier. <https://doi.org/10.1533/9780857093783.5.437>
- Rosi, N. L., & Mirkin, C. A. (2005). Nanostructures in Biodiagnostics. *Chemical Reviews*, 105(4), 1547–1562. <https://doi.org/10.1021/cr030067f>
- Ryvarden, L. R. (1995). Can We Trust Morphology in *Ganoderma*? In P. K. Buchanan & J. M. Monclavo (Eds.), *Ganoderma: Systematics Phytopathology and Pharmacology* (pp. 19–24). Hseu Ruey-Shyang, Applied Microbiology Laboratory, Agricultural Chemistry Department, National Taiwan University, Taipei.
- Saha, K., Agasti, S. S., Kim, C., Li, X., & Rotello, V. M. (2012). Gold Nanoparticles in Chemical and Biological Sensing. *Chemical Reviews*, 112(5), 2739–2779. <https://doi.org/10.1021/cr2001178>

- Sahoo, P. R., Sethy, K., Mohapatra, S., & Panda, D. (2016). Loop mediated isothermal amplification: An innovative gene amplification technique for animal diseases. *Veterinary World*, 9(5), 465–469. <https://doi.org/10.14202/vetworld.2016.465-469>
- Salam, F. (2010). *Development of immunosensors for Salmonella typhimurium* [Cranfield University]. <http://dspace.lib.cranfield.ac.uk/handle/1826/8193>
- Samseemoung, G., Jayasuriya, H. P. W., & Soni, P. (2011). Oil palm pest infestation monitoring and evaluation by helicopter-mounted, low altitude remote sensing platform. *Journal of Applied Remote Sensing*, 5(1), 053540. <https://doi.org/10.1117/1.3609843>
- Sarkar, S., Maity, A., Sarma Phukon, A., Ghosh, S., & Chakrabarti, R. (2019). Salt Induced Structural Collapse, Swelling, and Signature of Aggregation of Two ssDNA Strands: Insights from Molecular Dynamics Simulation. *The Journal of Physical Chemistry B*, 123(1), 47–56. <https://doi.org/10.1021/acs.jpcb.8b09098>
- Sassolas, A., Leca-Bouvier, B. D., & Blum, L. J. (2008). DNA Biosensors and Microarrays. *Chemical Reviews*, 108(1), 109–139. <https://doi.org/10.1021/cr0684467>
- Sato, K., Hosokawa, K., & Maeda, M. (2003). Rapid aggregation of gold nanoparticles induced by non-cross-linking DNA hybridization. *Journal of the American Chemical Society*, 125(27), 8102–8103. <https://doi.org/10.1021/ja034876s>
- Scarff, C. A., Fuller, M. J. G., Thompson, R. F., & Iadaza, M. G. (2018). Variations on Negative Stain Electron Microscopy Methods: Tools for Tackling Challenging Systems. *Journal of Visualized Experiments*, 132. <https://doi.org/10.3791/57199>
- Schoch, C. L., Seifert, K. A., Huhndorf, S., Robert, V., Spouge, J. L., Levesque, C. A., Chen, W., Bolchacova, E., Voigt, K., Crous, P. W., Miller, A. N., Wingfield, M. J., Aime, M. C., An, K.-D., Bai, F.-Y., Barreto, R. W., Begerow, D., Bergeron, M.-J., Blackwell, M., ... Schindel, D. (2012). Nuclear ribosomal internal transcribed spacer (ITS) region as a universal DNA barcode marker for Fungi. *Proceedings of the National Academy of Sciences*, 109(16), 6241–6246. <https://doi.org/10.1073/pnas.1117018109>
- Seo, G. S., & Kirk, P. M. (2000). *Ganodermataceae: nomenclature and classification*. In J. Flood, P. D. Bridge, & M. Holderness (Eds.), *Ganoderma diseases of perennial crops* (pp. 3–22). CABI. <https://doi.org/10.1079/9780851993881.0003>
- Shafri, H. Z. M., Anuar, M. I., Seman, I. A., & Noor, N. M. (2011). Spectral Discrimination of Healthy and *Ganoderma*-Infected Oil Palms from Hyperspectral Data. *International Journal of Remote Sensing*, 32(22), 7111–7129. <https://doi.org/10.1080/01431161.2010.519003>
- Shamaila, S., Zafar, N., Riaz, S., Sharif, R., Nazir, J., & Naseem, S. (2016). Gold Nanoparticles: An Efficient Antimicrobial Agent against Enteric Bacterial Human Pathogen. *Nanomaterials*, 6(4), 71. <https://doi.org/10.3390/nano6040071>



- Shevade, V. S., & Loboda, T. V. (2019). Oil palm plantations in Peninsular Malaysia: Determinants and constraints on expansion. *PLOS ONE*, 14(2), e0210628. <https://doi.org/10.1371/journal.pone.0210628>
- Singh, A., Chaudhary, S., Agarwal, A., & Verma, A. S. (2014). Antibodies. In A. S. Verma & A. Singh (Eds.), *Animal Biotechnology* (pp. 265–287). Academic Press. <https://doi.org/10.1016/B978-0-12-416002-6.00015-8>
- Singh, G. (1991). *Ganoderma* - The scourge of oil palms in the coastal areas. *The Planter*, 67(786), 421–444.
- Smith, B. J., & Sivasithamparam, K. (2000). Isozymes of *Ganoderma* Species From Australia. *Mycological Research*, 104(8), 952–961. <https://doi.org/10.1017/S0953756200002446>
- Sohn, J. S., Kwon, Y. W., Jin, J. II, & Jo, B. W. (2011). DNA-Templated Preparation of Gold Nanoparticles. *Molecules*, 16(10), 8143–8151. <https://doi.org/10.3390/molecules16108143>
- Song, J., Li, Z., Cheng, Y., & Liu, C. (2010). Self-aggregation of oligonucleotide-functionalized gold nanoparticles and its applications for highly sensitive detection of DNA. *Chemical Communications*, 46(30), 5548. <https://doi.org/10.1039/c0cc00308e>
- Storhoff, J. J., Elghanian, R., Mirkin, C. A., & Letsinger, R. L. (2002). Sequence-Dependent Stability of DNA-Modified Gold Nanoparticles. *Langmuir*, 18(17), 6666–6670. <https://doi.org/10.1021/la0202428>
- Su, X., Teh, H. F., Lieu, X., & Gao, Z. (2007). Enzyme-Based Colorimetric Detection of Nucleic Acids Using Peptide Nucleic Acid-Immobilized Microwell Plates. *Analytical Chemistry*, 79(18), 7192–7197. <https://doi.org/10.1021/ac0709403>
- Sun, L., Zhang, Z., Wang, S., Zhang, J., Li, H., Ren, L., Weng, J., & Zhang, Q. (2009). Effect of pH on the Interaction of Gold Nanoparticles with DNA and Application in the Detection of Human p53 Gene Mutation. *Nanoscale Research Letters*, 4(3), 216–220. <https://doi.org/10.1007/s11671-008-9228-z>
- Sun, S.-J., Gao, W., Lin, S.-Q., Zhu, J., Xie, B.-G., & Lin, Z.-B. (2006). Analysis of Genetic Diversity in *Ganoderma* Population With a Novel Molecular Marker SRAP. *Applied Microbiology and Biotechnology*, 72(3), 537–543. <https://doi.org/10.1007/s00253-005-0299-9>
- Supramaniam, C. V. (2016). *Molecular interaction between Ganoderma boninense and young oil palm*. University of Nottingham.
- Suwandi, Akino, S., & Kondo, N. (2012). Common Spear Rot of Oil Palm in Indonesia. *Plant Disease*, 96(4), 537–543. <https://doi.org/10.1094/PDIS-08-10-0569>
- Sze, A., Erickson, D., Ren, L., & Li, D. (2003). Zeta-potential measurement using the Smoluchowski equation and the slope of the current–time relationship in

- electroosmotic flow. *Journal of Colloid and Interface Science*, 261(2), 402–410. [https://doi.org/10.1016/S0021-9797\(03\)00142-5](https://doi.org/10.1016/S0021-9797(03)00142-5)
- Tan, K. (1968). Oil Palm Diseases in Malaya. *International Journal of Pest Management: Part B*, 14(1), 46–52. <https://doi.org/10.1080/05331846809432282>
- Taton, T. A., Mirkin, C. A., & Letsinger, R. L. (2000). Scanometric DNA Array Detection with Nanoparticle Probes. *Science*, 289(5485), 1757–1760. <https://doi.org/10.1126/science.289.5485.1757>
- Tchotet Tchoumi, J. M., Coetzee, M. P. A., Rajchenberg, M., & Roux, J. (2019). Taxonomy and species diversity of *Ganoderma* species in the Garden Route National Park of South Africa inferred from morphology and multilocus phylogenies. *Mycologia*, 111(5), 730–747. <https://doi.org/10.1080/00275514.2019.1635387>
- Teengam, P., Siangproh, W., Tuantranont, A., Vilaivan, T., Chailapakul, O., & Henry, C. S. (2017). Multiplex Paper-Based Colorimetric DNA Sensor Using PyrrolidinyI Peptide Nucleic Acid-Induced AgNPs Aggregation for Detecting MERS-CoV, MTB, and HPV Oligonucleotides. *Analytical Chemistry*, 89(10), 5428–5435. <https://doi.org/10.1021/acs.analchem.7b00255>
- Thavanathan, J., Huang, N. M., & Thong, K.-L. (2015). Colorimetric biosensing of targeted gene sequence using dual nanoparticle platforms. *International Journal of Nanomedicine*, 10, 2711. <https://doi.org/10.2147/IJN.S74753>
- Thomas, E. M., & Testa, S. M. (2017). The colorimetric determination of selectively cleaved adenosines and guanosines in DNA oligomers using bichinchonic acid and copper. *JBIC Journal of Biological Inorganic Chemistry*, 22(1), 31–46. <https://doi.org/10.1007/s00775-016-1405-4>
- Thompson, A. (1931). Stem Rot of the Oil Palm in Malaya. *Bulletin Department of Agriculture, Straits Settlements and F.M.S., Science Series*, 6, 23.
- Tian, W., Wang, L., Lei, H., Sun, Y., & Xiao, Z. (2018). Antibody production and application for immunoassay development of environmental hormones: a review. *Chemical and Biological Technologies in Agriculture*, 5(1), 5. <https://doi.org/10.1186/s40538-018-0117-0>
- Tomaszewska, E., Soliwoda, K., Kadziola, K., Tkacz-Szczesna, B., Celichowski, G., Cichomski, M., Szmaja, W., Grobelny, J., & Yu, W. W. (2013). Detection Limits of DLS and UV-Vis Spectroscopy in Characterization of Polydisperse Nanoparticles Colloids. *Journal of Nanomaterials*, 2013. <https://doi.org/10.1155/2013/313081>
- Tong, P., Shao, Y., Chen, J., He, Y., & Zhang, L. (2015). A sensitive electrochemical DNA biosensor for *Microcystis* spp. sequence detection based on an Ag@Au NP composite film. *Analytical Methods*, 7(7), 2993–2999. <https://doi.org/10.1039/C4AY02482F>

- Torres-Torres, M. G., & Guzmán-Dávalos, L. (2012). The Morphology of *Ganoderma* Species With a Laccate Surface. *Mycotaxon*, 119(1), 201–216. <https://doi.org/10.5248/119.201>
- Tsai, T.-T., Shen, S.-W., Cheng, C.-M., & Chen, C.-F. (2013). Paper-based tuberculosis diagnostic devices with colorimetric gold nanoparticles. *Science and Technology of Advanced Materials*, 14(4), 044404. <https://doi.org/10.1088/1468-6996/14/4/044404>
- Turner, P. D. (1965). The incidence of *Ganoderma* disease of oil palms in Malaya and its relation to previous crop. *Annals of Applied Biology*, 55(3), 417–423. <https://doi.org/10.1111/j.1744-7348.1965.tb07954.x>
- Utomo, C., & Niepold, F. (2000a). The Development of Diagnostic Tools for *Ganoderma* in oil palm. In J. Flood, P. D. Bridge, & M. Holderness (Eds.), *Ganoderma Diseases of Perennial Crops* (pp. 235–247). CABI. <https://doi.org/10.1079/9780851993881.0235>
- Utomo, C., & Niepold, F. (2000b). Development of Diagnostic Methods for Detecting *Ganoderma*-infected Oil Palms. *Journal of Phytopathology*, 148(9–10), 507–514. <https://doi.org/10.1046/j.1439-0434.2000.00478.x>
- Utomo, C., Pamin, K., & Niepold, F. (1997). Early detection of *Ganoderma* in oil palms by Elisa technique. *Jurnal Penelitian Kelapa Sawit (Indonesia)*, 5(2), 79–92.
- Utomo, C., Werner, S., Niepold, F., & Deising, H. B. (2005). Identification of *Ganoderma*, the Causal Agent of Basal Stem Rot Disease in Oil Palm Using a Molecular Method. *Mycopathologia*, 159(1), 159–170. <https://doi.org/10.1007/s11046-004-4439-z>
- Valentini, P., Fiammengo, R., Sabella, S., Gariboldi, M., Maiorano, G., Cingolani, R., & Pompa, P. P. (2013). Gold-Nanoparticle-Based Colorimetric Discrimination of Cancer-Related Point Mutations with Picomolar Sensitivity. *ACS Nano*, 7(6), 5530–5538. <https://doi.org/10.1021/nn401757w>
- van der Velden, V. H. J., Szczepański, T., & van Dongen, J. J. M. (2001). Polymerase Chain Reaction, Real-Time Quantitative. In *Encyclopedia of Genetics* (pp. 1503–1506). Academic Press. <https://doi.org/10.1006/rwgn.2001.1726>
- Verma, H. N., Singh, P., & Chavan, R. M. (2014). Gold nanoparticle: synthesis and characterization. *Veterinary World*, 7(2), 72–77. <https://doi.org/10.14202/vetworld.2014.72-77>
- Verma, M. S., Rogowski, J. L., Jones, L., & Gu, F. X. (2015). Colorimetric Biosensing of Pathogens Using Gold Nanoparticles. *Biotechnology Advances*, 33(6), 666–680. <https://doi.org/10.1016/j.biotechadv.2015.03.003>
- Vo-Dinh, T. (2017). Nanotechnology in Biology and Medicine. In T. Vo-Dinh (Ed.), *Nanotechnology in Biology and Medicine: Methods, Devices, and Applications* (Second). CRC Press (Taylor & Francis Group).



<https://doi.org/10.4324/9781315374581>

- Voora, V., Larrea, C., Bermudez, S., & Baliño, S. (2019). Global Market Report : Palm Oil. In *Sustainable Commodities Marketplace*. <https://www.iisd.org/system/files/publications/ssi-global-market-report-palm-oil.pdf>
- Wakefield, E. M. (1920). Diseases of the Oil Palm in West Africa. *Bulletin of Miscellaneous Information (Royal Gardens, Kew)*, 1920(9), 306. <https://doi.org/10.2307/4107530>
- Walker, J. M. (2011). *Characterization of Nanoparticles Intended for Drug Delivery, Methods in Molecular Biology* (S. E. McNeil (ed.)). Humana Press.
- Waller, J. M., Lenné, J. M., & Waller, S. J. (2001). *Plant Pathologist's Pocketbook* (J. M. Waller, J. M. Lenné, & S. J. Waller (eds.); Third Edit). [http://www.agrifs.ir/sites/default/files/Plant\\_Pathologists%27\\_Pocketbook\\_%7BJM\\_Waller%7D\\_%5B9780851994598%5D\\_%28CABI-2001%29\\_0.pdf](http://www.agrifs.ir/sites/default/files/Plant_Pathologists%27_Pocketbook_%7BJM_Waller%7D_%5B9780851994598%5D_%28CABI-2001%29_0.pdf)
- Wang, G., & Maier, R. J. (2009). A RecB-Like Helicase in *Helicobacter pylori* Is Important for DNA Repair and Host Colonization. *Infection and Immunity*, 77(1), 286–291. <https://doi.org/10.1128/IAI.00970-08>
- Wang, Y., Kong, S. L., & Su, X. Di. (2020). A Centrifugation-Assisted Visual Detection of SNP in Circulating Tumor DNA Using Gold Nanoparticles Coupled With Isothermal Amplification. *RSC Advances*, 10(3), 1476–1483. <https://doi.org/10.1039/C9RA09029K>
- Warnon, S., Zammattéo, N., Alexandre, I., Hans, C., & Remacle, J. (2000). Colorimetric Detection of the Tuberculosis Complex Using Cycling Probe Technology and Hybridization in Microplates. *BioTechniques*, 28(6), 1152–1160. <https://doi.org/10.2144/00286st05>
- Weissleder, R., Kelly, K., Sun, E. Y., Shtatland, T., & Josephson, L. (2005). Cell-specific targeting of nanoparticles by multivalent attachment of small molecules. *Nature Biotechnology*, 23(11), 1418–1423. <https://doi.org/10.1038/nbt1159>
- Wong, Y.-P., Othman, S., Lau, Y., Radu, S., & Chee, H. (2018). Loop-mediated isothermal amplification (LAMP): a versatile technique for detection of microorganisms. *Journal of Applied Microbiology*, 124(3), 626–643. <https://doi.org/10.1111/jam.13647>
- Yazid, S. N. E., Jinap, S., Ismail, S. I., Magan, N., & Samsudin, N. I. P. (2020). Phytopathogenic organisms and mycotoxigenic fungi: Why do we control one and neglect the other? A biological control perspective in Malaysia. *Comprehensive Reviews in Food Science and Food Safety*, 19(2), 643–669. <https://doi.org/10.1111/1541-4337.12541>
- Zakaria, L., Ali, N. S., Salleh, B., & Zakaria, M. (2009). Molecular Analysis of *Ganoderma* species from Different Hosts in Peninsula Malaysia. *Journal of*

*Biological Sciences*, 9(1), 12–20. <https://doi.org/10.3923/jbs.2009.12.20>

Zeng, S., Yong, K.-T., Roy, I., Dinh, X.-Q., Yu, X., & Luan, F. (2011). A Review on Functionalized Gold Nanoparticles for Biosensing Applications. *Plasmonics*, 6(3), 491–506. <https://doi.org/10.1007/s11468-011-9228-1>

Zeven, A. C. (1967). *The Semi-Wild Oil Palm and its Industry in Africa*. <https://doi.org/10.2307/2258269>

Zhaoping, L., Geisler, W. S., & May, K. A. (2011). Human Wavelength Discrimination of Monochromatic Light Explained by Optimal Wavelength Decoding of Light of Unknown Intensity. *PLoS ONE*, 6(5), e19248. <https://doi.org/10.1371/journal.pone.0019248>

Zhou, L.-W., Cao, Y., Wu, S.-H., Vlasák, J., Li, D.-W., Li, M.-J., & Dai, Y.-C. (2015). Global Diversity of the *Ganoderma lucidum* complex (*Ganodermataceae*, *Polyporales*) Inferred From Morphology and Multilocus Phylogeny. *Phytochemistry*, 114, 7–15. <https://doi.org/10.1016/j.phytochem.2014.09.023>

Zhou, X., Li, Q., Zhao, J., Tang, K., Lin, J., & Yin, Y. (2007). Comparison of Rapid DNA Extraction Methods Applied to PCR Identification of Medicinal Mushroom *Ganoderma* spp. *Preparative Biochemistry and Biotechnology*, 37(4), 369–380. <https://doi.org/10.1080/10826060701593282>

Zwart, D. C. (2021). *Sonic Tomography Advanced Decay Detection in Trees*. <https://www.bartlett.com/resources/sonic-tomography.pdf>