



UNIVERSITI PUTRA MALAYSIA

***EFFICACY OF HERBAL EXTRACT FORMULATIONS IN INHIBITING α -
GLUCOSIDASE AND NITRIC OXIDE ACTIVITY USING METABOLOMICS
APPROACH***

CHANDRADEVAN A/L MACHAP

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METABOLOMICS APPROACH**

By

CHANDRADEVAN A/L MACHAP

**Thesis Submitted to School of Graduate Studies,
Universiti Putra Malaysia, in Fulfilment of the Requirements for the
Degree of Master of Science**

April 2021

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia
in fulfillment of the requirement for the degree of Master of Science

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Chairman : Professor Faridah Abas, PhD
Institute : Bioscience

Development of poly-herbal products is at the pinnacle in today's herbal market. The idea of mixing multiple herbs as a single product has shown promising results in fighting non-communicable diseases. Two commonly grown herbs in Malaysia, *Gynura procumbens* (GP) and *Cleome gynandra* (CG) have been previously reported to possess health benefiting activities especially in treating diabetes and inflammation. However, the idea of mixing both herbs as a single product and testing the formulation for the aforementioned illness has never been reported before. Thus, this study was proposed to investigate the anti-diabetic and anti-inflammation properties of herbal extract formulations between GP and CG using metabolomics approach. Prior to the preparation of the extract formulations, freeze dried GP and CG leaves were extracted with different ethanol concentrations (0%, 20%, 50%, 70% and 100%). The individual extracts were tested for their total phenolic content (TPC), 2,2-diphenyl-1-picrylhydrazyl (DPPH) scavenging, α -Glucosidase and nitric oxide (NO) inhibitory activities. The results indicated that 100% ethanolic extract of GP had the highest TPC (5.91 mg GAE/100mgde) and the lowest IC₅₀ values for DPPH scavenging (63.73 μ g/mL) and NO inhibition (25.20 μ g/mL) activity. For CG, the 20% and 100% ethanolic extracts had insignificant high TPC at 3.46 and 3.71 mg GAE/100mgde respectively. However, 20% ethanolic extract of CG has the lowest IC₅₀ value (40.36 μ g/mL) in DPPH scavenging activity and in returns, 100% ethanolic extract of CG has the lowest IC₅₀ value (60.75 μ g/mL) in NO inhibition activity. However, none of the extracts have any inhibition

activity against α -Glucosidase enzyme; and subsequently not tested for the extract formulations. Extracts with best activities overall (100% GP, 100% and 20% CG) were then analysed via UHPLC-ESI-Orbitrap-MS/MS. A total of 58 metabolites from the groups of hydroxycinnamic acids, hydroxybenzoic acids, and flavonoid derivatives were tentatively identified from the extracts in which 24 of them were assigned for the first time from both herb species. In the second part of the study, ^1H NMR based metabolomics approach was applied to all extracts from both herbs. Principal component analysis (PCA) and hierarchical cluster analysis (HCA) revealed a good separation between the extracts and the corresponding metabolites identified via ^1H NMR spectroscopy. Phenolic compounds such as flavonoids (kaempferol, quercetin), hydroxycinnamic acids (caffeoylquinic, dicaffeoylquinic acids), hydroxybenzoic acid (gallic acid), organic acids (mallic acid, citric acid) and amino acids (phenylalanine, choline) are among the metabolites that contributed to the correlation with the extract bioactivities. The PLS model for both herbs has an overall acceptable goodness of fit and predictive power, which further strengthens the validity of this study. In the final part of the study, six formulations were designed from the best extracts of both herbs in certain ratios and their ability to scavenge DPPH, NO and inhibiting NO were re-tested. The results showed that formulation of 100% GP + 20% CG at 3:1 ratio has the lowest IC_{50} at 76.72 $\mu\text{g}/\text{mL}$ but not significant compared to 20% CG extract alone (71.12 $\mu\text{g}/\text{mL}$) in DPPH scavenging assay. In returns, NO scavenging assay indicated a significant inhibition ($p < 0.05$) in 100% GP + 100% CG at 1:3 ratio with IC_{50} registered at 85.01 $\mu\text{g}/\text{mL}$. On contrary, formulation of 100% GP + 100% CG at 3:1 ratio has the lowest IC_{50} (94.56 $\mu\text{g}/\text{mL}$) in NO inhibition activity compared to other formulation extracts. The interaction patterns between the extract formulations were studied using the combination index method. In general, the formulations showed a synergistic>additive>antagonistic interaction from low to high fraction affect. The present study offers the idea on how metabolomics approach can be utilized in developing poly-herbal based products. Integrating the data obtained from bioactivities studies, UHPLC-ESI-Orbitrap-MS/MS and ^1H NMR spectroscopy, a proper and standardised extract formulations can be developed. In conclusion, this study provides a strong justification of GP/CG herbal extract formulations as a potential anti-inflammatory agent in functional food prospect.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
Sebagai memenuhi keperluan untuk ijazah Master Sains

**KEBERKESANAN FORMULASI EKSTRAK HERBA DALAM
MERENCAT AKTIVITI α -GLUCOSIDASE DAN NITRIK OKSIDA
DENGAN MENGGUNAKAN PENDEKATAN METABOLOMIKS**

Oleh

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Pembangunan produk berasaskan poli-herba semakin mendapat perhatian di pasaran kini. Idea mencampurkan perlbagai herba sebagai produk tunggal telah menunjukkan hasil yang memberangsangkan dalam menangani penyakit tidak berjangkit. Dua herba yang biasa didapati di Malaysia, *Gynura procumbens* (GP) dan *Cleome gynandra* (CG) telah dilaporkan mempunyai aktiviti kesihatan terutamanya dalam merawat diabetik dan keradangan. Walaubagaimanapun, idea untuk mencampurkan kedua-dua herba tersebut sebagai produk tunggal dan kemudiannya diuji keberkesanan dalam merawat diabetik dan keradangan belum pernah dilaporkan lagi. Maka, satu kajian menggunakan formulasi ekstrak herba antara GP dan CG dalam menentukan aktiviti anti-diabetik dan anti-keradangan melalui pendekatan metabolomiks telah dicadangkan. Sebelum formulasi ekstrak dibuat, daun GP dan CG yang telah disejuk beku telah diekstrak menggunakan etanol pada beberapa kepekatan berbeza (0%, 20%, 50% dan 100%). Ekstrak individu disaring untuk mendapatkan jumlah kandungan asid fenolik (TPC), aktiviti pemerangkap radikal bebas 2,2-difenil-1-pikrilhidrazil (DPPH), perencatan enzim α -glucosidase dan nitrik oksida (NO). Analisis menunjukkan ekstrak 100% etanol GP mempunyai kandungan asid fenolik tertinggi (5.91 mg GAE/100mgde) dan nilai IC_{50} terendah untuk aktiviti pemerangkap radikal bebas DPPH (63.73 μ g/mL) dan perencatan NO (25.20 μ g/mL). Untuk CG, didapati ekstrak etanol pada 20% dan 100% mempunyai kandungan asid fenolik tertinggi tetapi tidak signifikan pada bacaan 3.46 and 3.71 mg GAE/100mgde masing-masing. Walaubagaimanapun, ekstrak 20% etanol CG mempunyai nilai

IC₅₀ terendah untuk aktiviti pemerangkap radikal bebas DPPH (40.36 µg/mL) dan sebaliknya, ekstrak 100% etanol mencatatkan nilai IC₅₀ terendah untuk perencatan NO (60.75 µg/mL). Keputusan juga menunjukkan tiada satu ekstrak daripada kedua-dua herba dapat merencatkan tindakbalas enzim α-glucosidase, maka asai ini tidak diuji kepada formulasi ekstrak. Seterusnya, ekstrak yang mempunyai aktiviti terbaik keseluruhannya (100% GP, 100% and 20% CG) dianalisa menggunakan UHPLC-ESI-Orbitrap-MS/MS. Sejumlah 58 metabolit daripada kumpulan asid hidroksisinamik, asid hidroksibenzoik dan flavanoid terbitan telah dikenalpasti secara tentatif dan 24 daripadanya kenalpasti buat pertama kali dalam ekstrak kedua-dua herba tersebut. Pada bahagian kedua kajian, pendekatan metabolomiks menggunakan ¹H NMR telah dilaksanakan pada ekstrak kedua-dua herba. Analisis komponen principal (PCA) dan analisis kluster hierarki (HCA) menunjukkan pemisahan yang bagus antara ekstrak dan metabolit sepadan yang dikenalpasti melalui ¹H NMR. Asid fenolik seperti flavonoid (kaempferol, kuersetin), asid hidroksisinamik (asid kafeolkuinik, asid dikafeolkuinik), asid hidroksibenzoik (asid galik), asid organik (asid malik, asid sitrik) dan asid amino (fenilalanin, kolin) adalah antara metabolit yang dikenalpasti menyumbang kepada bioaktiviti ekstrak. Modal PLS untuk kedua-dua herba juga mempunyai uji kesesuaian dan kuasa jangkaan yang sesuai, dimana ianya mengukuhkan lagi validiti kajian ini. Enam formulasi herba telah direka daripada ekstrak terbaik GP dan CG pada nisbah tertentu di bahagian terakhir kajian ini. Kebolehan formulasi ekstrak untuk memerangkap radikal bebas DPPH dan merencat NO telah diuji semula. Analisis menunjukkan formulasi 100% GP + 20% CG pada nisbah 3:1 mempunyai IC₅₀ 76.72 µg/mL tetapi tidak signifikan berbanding ekstrak 20% CG sahaja (71.12 µg/mL) untuk asai pemerangkap radikal bebas DPPH. Manakala, dalam asai pemerangkap radikal bebas NO, formulasi 100% GP + 100% CG pada nisbah 1:3 mempunyai nilai IC₅₀ terendah yakni 85.01 µg/mL (p<0.05). Sebaliknya, formulasi 100% GP + 100% CG pada nisbah 3:1 mempunyai nilai IC₅₀ terendah iaitu 94.56 µg/mL untuk aktiviti perencatan NO berbanding formulasi lain. Corak interaksi antara ekstrak di dalam formulasi juga dikaji menggunakan kaedah indeks kombinasi. Secara am, formulasi-formulasi tersebut menunjukkan interaksi sinergi> penambahan> antagonistik dari kesan fraksi rendah ke tinggi. Kajian ini memberikan idea bagaimana pendekatan metabolomiks dapat digunakan dalam pembangunan produk poli-herba. Satu formulasi ekstrak yang piawai dapat dibangunkan apabila maklumat daripada kajian bioaktiviti, UHPLC-ESI-Orbitrap-MS/MS dan spektroskopi ¹H NMR diintegrasikan. Secara kesimpulannya, kajian ini menunjukkan potensi formulasi ekstrak GP/CG sebagai agen anti-keradangan dalam prospek makanan berfungsi.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS

^{13}C NMR	Carbon-13 nuclear magnetic resonance
^1H NMR	Protonated nuclear magnetic resonance
ACD	Advanced chemistry development
ANOVA	Analysis of variance
ATCC	American type culture collection
C18	Carbon 18
CD_3OD	Deuterated methanol
CG	<i>Cleome gynandra</i>
CI	Combination index
CMC	Carboxymethyl cellulose
D_2O	Deuterated oxide
de	Dried extract
DMEM	Dulbecco's modified eagle's medium
DMSO	Dimethyl sulfoxide
DPPH	2,2-diphenyl-1-picrylhydrazyl
eNOS	Epithelial nitric oxide synthase
ETOH	Ethanol
Fa	Fraction affect
<i>Fasn</i>	Fatty acid synthase
FBS	Fetal bovine serum
GAE	Gallic acid equivalent
GCMS	Gas chromatography mass spectrometry
GLP-1	Glucagon-like peptide-1
GP	<i>Gynura procumbens</i>

HCA	Hierarchical clustering analysis
HDL	High density lipoprotein
HPLC	High performance liquid chromatography
iNOS	Inducible nitric oxide synthase
IFN- γ	Interferon gamma
KH ₂ PO ₄	Potassium dihydrogen phosphate
LCMS	Liquid chromatography mass spectrometry
LDL	Low density lipoprotein
LPO	Lipid peroxide
LPS	Lipopolysaccharide
MARDI	Malaysia Agricultural, Research and Development Institute
MOLGEN	Molecular structure generation
MTT	3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide
MVDA	Multivariate data analysis
NaNO ₂	Sodium nitrite
NHMS	National health and Morbidity Survey
NaOD	Sodium deuterium oxide
nNOS	Neuronal nitric oxide synthase
NO	Nitric oxide
NOS	Nitric oxide synthase
NSAIDs	Non-steroidal anti-inflammatory drugs
PBS	Phosphate-buffered saline
PCA	Principal component analysis

PLS	Partial least Square
PNPG	4-nitrophenyl α -D-glucopyranoside
R _f	Retention factor
SGLT2	Sodium glucose cotransporter 2
T1DM	Type 1 diabetes mellitus
T2DM	Type 2 diabetes mellitus
TLC	Thin layer chromatography
TLRs	Toll-like receptors
TPC	Total phenolic content
TSP	Trimethylsilylpropanoic acid
UHPLC-ESI-Orbitrap-MS/MS	Ultra high performance liquid chromatography-electron spray ionisation-orbitrap-mass spectrometer/mass spectrometer
UV	Ultraviolet
VIP	Variable importance in projections

CHAPTER 1

GENERAL INTRODUCTION

The rising incidence of non-communicable diseases is in the spotlight. The occurrence of diabetes, cardiovascular disease, cancer and some diseases has sparked the development of modern medicines. However, what is at stake is that these modern medicines exhibit some unwanted side effects on internal organs after a prolonged usage. As such, finding a cure for these diseases has always been a challenging task. Natural products, in turn, have been stealing the limelight as an alternative solution in combating against these diseases.

Inflammation in the cells has been associated with the development of diabetes in past studies (Wellen and Hotamisligil, 2005; Sjöholm and Nystrom, 2006). The accumulation of certain hormones and anti-inflammatory factors in damaged cells somehow inhibits the insulin signaling pathway, which in turn increases blood glucose levels. Previous studies using *Gynura procumbens* extract have shown that it has anti-diabetic and anti-inflammatory effects (Iskander et al., 2002; Siti Pauliena, 2006; Hassan et al., 2010, Kaewseejan et al., 2012, Wong et al., 2015). In addition, *Cleome gynandra* has been studied extensively for its anti-inflammation and various health properties (Sivanesan & Hazeena, 2007, Mishra et al., 2011, Ethadi et al., 2013, Msika et al., 2013). In-vivo studies also have indicated that none of the extracts possessed toxicity when they were tested at 5000 mg/kg and 3000 mg/kg for *G. procumbens* and *C. gynandra* respectively (Narendhirakannan et al., 2005, Zahra et al., 2011). However, none of the past studies have shown the efficacy of these herbs to act as anti-diabetic or anti-inflammatory agents when they are formulated as herbal mixtures.

The emergence of products based on herbal formulation has had a major impact on the development of the herbal industry in Malaysia. As the saying goes, „two is better than one“, herbal practitioners and users tend to believe that mixed herbals have a more synergistic effect in fighting against non-communicable diseases rather than their individual herbs. Their dogma is in line with previously reported data. Kaur and Velecha (2014) in their review paper reported the efficacy of various herbal formulations products such as *Dihar*, *Diabet*, *Diasol*, and *Dianex* in treating diabetes. Herbal formulation between *Zingiber officinalis*, *Piper nigrum* and *Terminalia bellerica* were found to reduce serum triglyceride, total cholesterol and systolic blood pressure in high fat-diet fed rats compared to the individual herbal extract (Aziz et al., 2013).

To date, studies on the efficacy of herbal formulations consisting of *G. procumbens* and *C. gynandra* in treating diabetes and inflammation have not been reported. It was hypothesized that the formulated herbal extract between

G. procumbens and *C. gynandra* will have better efficacy in treating diabetes and inflammation compared to their individual extracts. The hypothesis was postulated based on the fact that both herbs have been studied extensively for their anti-diabetic and anti-inflammatory activities. A significant activity from the herbal extract mixture will definitely boost the herbal industry in Malaysia. As such, the present study was proposed to investigate the efficacy of *G. procumbens* and *C. gynandra*, outlining a few objectives that reflect in-depth studies:

1. To determine the effect of solvent extraction on the biological activities of *G. procumbens* and *C. gynandra* extracts,
2. To identify the secondary metabolites in the active extracts by means of the UHPLC-Orbitrap-MS/MS approach,
3. To differentiate the *G. procumbens* and *C. gynandra* extracts and correlate them with their biological activities by using ¹H NMR-based metabolomics, and
4. To derive herbal extract formulations from active extracts of *G. procumbens* and *C. gynandra* and re-analyse their bioactivity in comparison to the individual extracts.

REFERENCES

- Abdul Hamid, N.A., Abas, F., Ismail, I.S., Shaari, K., & Lajis, N.H. (2015). Influence of different drying treatments and extraction solvents on the metabolite profile and nitric oxide inhibitory activity of Ajwa Dates. *Journal of Food Science* 80: 2603-2610.
- Abdul Hamid, N.A., Maulidiani, M., Mediani, A., Yahya, U.I.I., Ismail, I.S., Tham, C.L., Shadid, K., & Abas, F. (2018). Physicochemical characteristics, nutritional composition, and phytochemical profiles of nine Algerian date palm fruit (*Phoenix dactylifera* L.) varieties. *Journal of Food Chemistry* 42: 1-13.
- Aggarwal, N., & Shishu. (2011). A review of recent investigation on medicinal herbs possessing anti-diabetic properties. *Journal of Nutritional Disorders and Therapy* 1: 1-10.
- Aharoni, A., & Galili, G. (2011). Metabolic engineering of the plant primary-secondary metabolism interface. *Current Opinion in Biotechnology* 22: 239-244.
- Ahmad, F., Zaidi, M.A.S., Sulaiman, N., & Majid, F.A.A. (2015). Proceedings from Persidangan Kebangsaan Ekonomi Malaysia Ke-10, Melaka, 18th-20th September 2015. *Issues and challenges in the development of the herbal industry in Malaysia*.
- Ahmad, N.S., Ramli, A., Islahudin, F., & Paraidathahu, T. (2013). Medication adherence in patients with type 2 diabetes mellitus treated at primary health clinics in Malaysia. *Patient Preference and Adherence* 7: 525-530.
- Ahmed, F.A., Abd El-Wahab Khamis, I.M., & Desoukey, S.Y. (2011). Flavonoids of *Neotorularia aculeolata* plant. *Journal of Pharmaceutical and Nutritional Science* 1: 134-139.
- Aksoy, L., Kolay, E., Agilonu, Y., Aslan, Z., & Kargioglu, M. (2010). Free radical scavenging activity, total phenolic content, total antioxidant status, and total oxidant status of endemic *Thermopsis turcica*. *Saudi Journal of Biological Sciences* 3: 235-239.
- Algariri, K., Meng, K.Y., Atangwho, I.J., Asmawi, M.Z., Sadikum, A., Murugayah, V., & Ismail, N. (2013). Hypoglycemic and anti-hyperglycemic study of *Gynura procumbens* leaf extract. *Asian Pacific Journal of Tropical Biomedicine* 3: 358-366.
- American Diabetes Association (2001). *Postprandial blood glucose*. *Diabetes Care* 24: 775-778.

- Amin, K.A., Safwat, GM., & Srirajaskanthan, R. (2012). Dietary sugars: chemistry, analysis, function and effects. In *High sucrose diet and antioxidant defense*, ed. Preedy VR, pp. 770-778. 3rd edn. Royal Society of Chemistry.
- Anbazhagi, T., Kadavul, K., Suguna, G., & Petrus, A.J.A. (2009). Studies on the pharmacognostical and in vitro antioxidant potential of *Cleome gynandra* Linn. leaves. *Natural Product Radiance* 8: 151-157.
- Aziz, N., Mehmood, M.H., & Gilani, A.H. (2013). Studies on two polyherbal formulations (ZPTO and ZTO) for comparison of their antidyslipidemic, antihypertensive and endothelial modulating activities. *BMC complementary and Alternative Medicine* 13: 371-380.
- Bag, A., & Chattopadhyay, R.R. (2015). Evaluation of synergistic antibacterial and antioxidant efficacy of essential oils of spices and herbs in combination. *PLoS ONE* 10: 1-17.
- Bala, A., Indrajit, K., & Pallab, K.H. (2010). Isolation and HPLC characterization of the flavonoid fractions from *Cleome gynandra* and comparative antioxidant activity. *Ethnomedicine and Therapeutic Validation* 32: 245-259.
- Balachandran, C., Emi, N., Arun, Y., Yamamoto, Y., Ahilan, B., Sangeetha, B., Duraipandiyar, V., Inaguma, Y., Okamoto, A., Ignacimuthu, S., Al-Dhabi, N.A., & Peruma, P.T. (2015). In vitro anticancer activity of methyl caffeate isolated from *Solanum torvum* Sawrtz. fruit. *Chemico-Biological Interactions* 242: 81-90.
- Barros, L., Duenas, M., Dias, M.I., Sousa, M.J., Buelga, C.S., & Ferreira, I.C.F.R. (2012). Phenolic profiles of in vivo and in vitro grown *Coriandrum sativum* L. *Food Chemistry* 132: 841-848.
- Bhope, S.G., Nagore, D.H., Kuber, V.V., Gupta, P.K., & Patil, M.J. (2011). Design and development of a stable polyherbal formulation based on the results of compatibility studies. *Pharmacognosy Research* 3: 122-129.
- Boora, F., Chirisa, E., & Mukanganyama, S. (2014). Evaluation of nitrite radical scavenging properties of selected Zimbabwean plant extracts and their phytoconstituents. *Journal of Food Processing* 2014: 1-7.
- Chamara, A.M.R., Kuganesan, A., Dolawatta, K.D., Amarathunga, I.M., Wickramasinghe, W.Y.H., Madushini, Y.M.P.K., & Thiripuranathar, G. (2018). Evaluation of bioactivities of two polyherbal formulations found in Sri Lankan ayurvedic treatments. *International Journal of Pharmaceutical Sciences and Research* 9: 2073-2079.

- Chan, L.K., Lim, S.Y., & Pan, L.P. (2009). Micropropagation of *Gynura procumbens* (Lour.) Merr. an important medicinal plant. *Journal of Medicinal Plants Research* 3: 105-111.
- Chang, J.B., Lane, M.E., Yang, M., & Heinrich, M. (2016). A hexa-herbal TCM decoction used to treat skin inflammation: an LC-MS-based phytochemical analysis. *Planta Medicine* 82: 1134-1141.
- Chaudhury, A., Duvoor, C., Dendi, V.S.R., Kraleti, S., Chada, A., Ravilla, R., Marco, A., Shekhawat, N.S., Montales, M.T., Kuriakose, K., Sasapu, A., Beebe, A., Patil, N., Musham, C.K., Lohani, G.P., & Mirza, W. (2017). Clinical review of antidiabetic drugs: implications for type 2 diabetes mellitus management. *Frontiers in Endocrinology* 8: 1-12.
- Che, C.T., Wang, Z.J., Chow, M.S.S., & Lam, W.K. (2013). Herb-herb combination for therapeutic enhancement and advancement: theory, practice and future perspectives. *Molecules* 18: 5125-5141.
- Chen, F., Long, X., Liu, Z., Shao, H., & Liu, L. (2014). Analysis of phenolic acids of Jerusalem artichoke (*Helianthus tuberosus* L.) responding to salt-stress by liquid chromatography/tandem mass spectrometry. *The Scientific World Journal* 2014: 1-8.
- Chen, J., Mangelinckx, S., Ma, L., Wang, z., Li, W., & De Kimpe, N. (2014). Caffeoylquinic acid derivatives isolated from the aerial parts of *Gynura divaricata* and their yeast α -glucosidase and PTP1B inhibitory activity. *Fitoterapia* 99: 1-6.
- Chen, L., Li, H., Song, H., & Zhang, G. (2009). A new cerebroside from *Gynura divaricata*. *Fitoterapia* 80: 517-520.
- Chen, L., Wang, J.J., Song, H.T., Zhang, G.G. & Qin, L.P. (2009). New cytotoxic cerebroside from *Gynura divaricata*. *Chinese Chemical Letters* 20: 1091-1093.
- Choi, S.I., Park, M.H., & Han, J.S. (2016). *Gynura procumbens* extract alleviates postprandial hyperglycemia in diabetic mice. *Preventive Nutrition and Food Science* 21: 181-186.
- Chou, T.C. (2006). Theoretical basis, experimental design, and computerized simulation of synergism and antagonism in drug combination studies. *Pharmacological Reviews* 58: 621-681.
- Chung, J.Y., Kim, Y.S., & Yoo, S.H. (2017). Regulation of inflammation by sucrose isomer, turanose, in raw 264.7 cells. *Journal of Cancer Prevention* 22: 195-201.
- Chweya, J.A., & Nameus, A.M. (1997). *Cat's whiskers: Cleome gynandra L. promoting the conservation and use of underutilized and neglected*

crops. Institute of Plant Genetics and Crop Plant Research, Gatersleben, Rome, Italy.

- Claridge, T.D.W. (1999). *Practical aspects of high-resolution NMR*. High-Resolution NMR Techniques in Organic Chemistry. Oxford, UK: Elsevier Press.
- Clifford, M.N., Knight, S., & Kunhert, N. (2014). Discriminating between the six isomers of dicaffeoylquinic acid by LC-MSⁿ. *Journal of Agricultural and Food Chemistry* 53: 3821-3832.
- Cui, Q., Lewis, i.A., Hegeman, A.D., Anderson, M.E., Li, J., Schulte, C.F., Westler, W.M., Eghbalnia, H.R., Sussman, M.R & Markley, J.L. (2008). Metabolite identification via the Madison Metabolomics Consortium Database. *Nature Biotechnology* 26: 162.
- Dean, P.J., Youngja, P., & Thomas, R.Z. (2012). Nutritional metabolomics: progress in addressing complexity in diet and health. *Annual Review of Nutrition* 32: 183-202.
- DeepaShree, C.L., & Shubha, G. (2014). Evaluation of *Cleome gynandra* for its chemical constituents, antioxidant potential and detection of flavonoids using Thin Layer Chromatography. *International Journal of Scientific Research* 3: 58-60.
- Dehkharghanian, M., Adenier, H., & Vijayalakshmi, M.A (2010). Analytical methods study of flavonoids in aqueous spinach extract using positive electrospray ionisation tandem quadrupole mass spectrometry. *Food Chemistry* 121: 863-870.
- Deng, L., Gu, H., Zhu, J., Nagana Gowda, G.A., Djukovic, D., Chiorean, G.E., & Raftery, D. (2016). Combining NMR and LC-MS using backward variable elimination: metabolomics analysis of colorectal cancer, polyps and health controls. *Analytical Chemistry* 16: 7975-7983.
- Devaraj, S., Dasu, M.R., & Jialal, I. (2010). Diabetes is a proinflammatory state: a translational perspective. *Expert Reviews of Endocrinal Metabolism* 5: 19-28.
- Devi, P.B., Vijayabharathi, R., Sathyabama, S., Malleshi, N.G., & Priyadarisini, V.B. (2014). Health benefits of finger millets (*Eleusine coracana* L.) polyphenols and dietary fibre: a review. *Journal of Food Science and Technology* 51: 1021-1040.
- Dias, D.A., Urban, S., & Roessner, U. (2012). A historical overview of natural product in drug discovey. *Molecule* 2: 303-336.

- Dominique, P.F. (2014). Practical aspects of NMR signal assignment in larger and challenging proteins. *Progress in Nuclear Magnetic Resonance Spectroscopy* 1: 47-75.
- Doshi, G.M., & Vanmali, B.V. (2016). Development and evaluation of herbal formulation from *Poyalthia longifolia*, *Tabernaemontana alternifolia*, *Benincasa hispida* plant extracts. *Der Pharmacia Lettre* 8: 170-183.
- Dufour, N, & Rao, R.P. (2011). Secondary metabolites and other small molecules as intercellular pathogenic signals. *FEMS Microbiology Letters* 314: 10-17.
- Elya, B., Handayani, R., Sauriasi, R., Azizahwati, Hasyati, U.S., Permana, I.T., & Permatasari, Y.I. (2015). Antidiabetic activity and phytochemical screening of extract from Indonesian plants by inhibition of alpha amylase, alpha glucosidase and dipeptidyl peptidase IV. *Pakistan Journal of Biological Sciences* 18: 279-284.
- Emma, L.S., Markus, M., & Werner, B. (2009). Matching structures to mass spectra using fragmentation patterns: are the results as good as they look? *Analytical Chemistry* 81: 3608-3617.
- Ethadi, S.R., Pragada, R.R., Battu, G.R., & Talluri, M.R. (2013). Antiinflammatory and antibacterial activities of different extracts of *Gynandropsis gynandra*. *International Journal of Pharmacy and Pharmaceutical Sciences* 5: 162-165.
- Esser, N., Poels, S.L., Piette, J., Scheen, A.J., & Paquot, N. (2014). Inflammation as a link between obesity, metabolic syndrome and type 2 diabetes. *Diabetes Research and Clinical Practice* 105: 141-150.
- Fang, N., Yu, S., & Prior, R.L. (2002). LC/MS/MS characterization of phenolic constituents in dried plums. *Journal of Agricultural and Food Chemistry* 50: 3579-3585.
- Felhi, S., Daoud, A., Hajlaoui, H., Mnafigui, K., Gharsallah, N., & Kadri, A. (2017). Solvent extraction effects on phytochemical constituents profiles, antioxidant and antimicrobial activities and functional group analysis of *Ecballium elaterium* seeds and peels fruits. *Food and Science Technology* 37: 483-492.
- Gan, S.H., Kamal, M.A., Lima, M.M.S., Khalil, M.I., Pasupuleti, V.R., & Aliev, G. (2015). Medicinal plants in management of type 2 diabetes and neurodegenerative disorders. *Evidence-Based Complementary and Alternative Medicine* 2015: 1-2.
- Galanakis, C.M., Goulas, V., Tsakona, S., Manganaris, G.A., & Gekas, V. (2010). A knowledge base for the recovery of natural phenols with

- different solvents. *International Journal of Food Properties* 16: 382-396.
- Gumustas, M., Kurbanoglu, S., Uslu, B., & Ozkan, S.A. (2013). UPLC versus HPLC on drug analysis: advantageous, applications and their validation parameters. *Chromatographia* 76: 1365-1427.
- Hakim, P., Abdullah, S.H., & Noor, M.M. (2008). Effects of *Gynura procumbens* extract and glibenclamide on sperm quality and specific activity of testicular lactate dehydrogenase in streptozotocin-induced diabetic rats. *Malaysian Journal of Biochemistry and Molecular Biology* 16: 10-14.
- Haminiuk, C.W.I., Plata-Oviedo, M.S.V., Mattos, G.D., Carpes, S.T., & Branco, I.G. (2014). Extraction and quantification of phenolic acids and flavanols from *Eugenia pyriformis* using different solvents. *Journal of Food Science and Technology* 51: 2862-2866.
- Hassan, Z., Yam, M.F., Ahmad, M., & Yusof, A.P.M (2010). Antidiabetic properties and mechanism of action of *Gynura procumbens* water extract in streptozotocin-induced diabetic rats. *Molecules* 15: 9008-9023.
- HighChem LLC (2020). mzCloud – Advanced Mass Spectral Database. Available at <https://mzcloud.org>. Accessed on 1st January 2020.
- Hottinger, D.G., Beebe, D.S., Kozhimanni, T., Prielipp, R., & Belani, K.G. (2014). Sodium nitroprusside in 2014: A clinical concepts review. *Journal of Anaesthesiology Clinical Pharmacology* 30: 462-471.
- Huang, D., Ou, B., & Prior, R. (2005). The chemistry behind antioxidant capacity assays. *Journal of Agricultural and Food Chemistry* 53: 1841-1856.
- Hussin, A.H. (2001). Adverse effect of herbs and drug-herbal interaction. *Malaysian Journal of Pharmacy* 1: 39-44.
- Iskander, M.N., Song, Y., & Coupar, I.M. (2002). Anti-inflammatory screening of the medicinal plant *Gynura procumbens*. *Plant Food for Human Nutrition* 57: 233-244.
- Institute for Public Health. *Non-Communicable Diseases, Risk Factors & Other Health Problems*. National Health and Morbidity Survey (NHMS) Vol. II (2015).
- Jae Yong, K., Heesook, K., Hakjoon, C., Ara, J., Huwon, K., Hyojeong, Y, Sojeong, Im., & Chulyung, C. (2018). Anti-inflammatory effects of *Stauntonia hexaphylla* fruit extract in lipopolysaccharide-activate RAW 264.7 macrophages and rats by Carrageenan-induced hind paw swelling. *Nutrients* 10: 1-12.

- James, J.P (2009). Principles and applications of liquid chromatography-mass spectrometry in clinical biochemistry. *Clinical Biochemistry Review* 30: 19-34.
- Jialal, I., & Kaur, H. (2012). The role of Toll-Like Receptors in diabetes-induced inflammation: implications for vascular complications. *Current Diabetes Reports* 12: 172-179.
- Jianwei, Z., Shan, Z., Peipei, Y., Linlin, Y., Jin, H., Lingling, S., Xiaojing, Z., Yujun, L., & Chao, M. (2014). α -glucosidase inhibitory activity of polyphenols from the burs of *Castanea mollissima* blume. *Molecules* 19: 8373-8386.
- Jinazali, H., Mtimuni, B., & Chilembwe, E. (2017). Nutrient composition of cat's whickers (*Cleome gynandra* L.) from different agro ecological zones in Malawi. *African Journal of Food Science* 11: 24-29.
- Jong, T.T., & Chou-Hwang, J.Y. (1997). An optically active chromanone from *Gynura formosana*. *Phytochemistry* 40: 553-554.
- Joshi, V.K., Joshi, A., & Dhiman, K.S. (2017). The Ayurvedic Pharmacopoeia of India, development and perspectives. *Journal of Ethnopharmacology* 197: 32-38
- Jupitara, D., & Kalita, J.C. (2016). Effects of *Cleome gynandra* Linn: leaf extract on ovarian folliculogenesis of albino mice. *Journal of Traditional Medicine and Clinical Naturopathy* 5: 2-4.
- Kadioglu, O., Nass, J., Saeed, M.E., Schuler, B., & Efferth, T. (2015). Kaempferol is an anti-inflammatory compound with activity towards NF- κ B pathway proteins. *Anticancer Research* 35: 2645-2650.
- Kaewseejan, N., Puangpronpitag, D., & Nakornriab, M. (2012). Evaluation of phytochemical composition and antibacterial property of *Gynura procumbens* extract. *Asian Journal of Plant Science* 1-7.
- Kang, M.G., Lee, H.J., Cho, J.Y., Yang, S.J., & Kim, D.J. (2016). Anti-inflammatory effects of sucrose-derived oligosaccharides produced by a constitutive mutant *L. mesenteroids* B-512FMCM dextranucrase in high fat diet-fed mice. *Biochemical and Biophysical Research Communications* 26: 350-355.
- Karar, M.G.E. & Kuhnert, N. (2015). UPLC-ESI-Q-TOF-MS/MS characterization of phenolics from *Crataegus monogyna* and *Crataegus laevigata* (Howthorn) leaves, fruits and their herbs derived drops (*Crataegutt Trofen*). *Journal of Chemical Biology and Therapeutics* 1: 1-23.

- Kasali, F.M., Kadima, J.N., Mpiana, P.T., Ngbolua, K.N., & Tshibangu, D.S.T. (2013). Assessment of anti-diabetic activity and acute toxicity of leaf extracts from *Physalis peruvian* L. in guinea pig. *Asian Pacific Journal of Tropical Biomedicine* 3: 841-846.
- Kasem, W.T., & Fathy, S. (2016). Flavanoids and isoenzymes as chemotaxonomic markers in *Cleome* L. (Cleomaceae Bercht and J. Presl). *Current Botany* 7: 11-16.
- Kaur, M., & Valecha, V. (2014). Diabetics and antidiabetic herbal formulations: an alternative to allopathy. *European Journal of Medicine* 6: 226-240.
- Kazeem, M.I., Adamson, J.O., & Ogunwande, I.A. (2013). Modes of inhibition of α -amylase and α -glucosidase by aqueous extract of *Morinda lucida* Benth leaf. *BioMed Research International* 2013: 1-6.
- Khafif, A., Schantz, S.P., Chou, T.C., Edelstein, D., & Sacks, P.G. (1998). Quantitation of chemopreventive synergism between (-)-epigallocatechin-3-gallate and curcumin in normal, premalignant and malignant human oral epithelial cells. *Carcinogenesis* 19: 419-424.
- Khatib, A., Perumal, V., Ahmed, Q.U., Uzir, B.F., & Murugesu, S. (2018). Low inhibition of alpha-glucosidase and xanthine oxidase activities of ethanol extract of *Momordica charantia* fruit. *Journal of Pharmaceutical Negative Results* 8: 20-24.
- Khoo, L.W., Mediani, A., Zolkeflee, N.K.Z., Leong, S.W., Ismail, I.S., Khatib, A., Shaari, K., & Abas, F. (2015). Phytochemical diversity of *Clinacanthus nutans* extracts and their bioactivity correlations elucidated by NMR based metabolomics. *Phytochemistry Letters* 14: 123-133.
- Kim, H.K., Choi, Y.H., & Verpoorte, R. (2010). NMR-based metabolomics analysis of plants. *Nature protocols* 5: 536-549.
- Kim, J., Kim, H., Choi, H., Jo, A., Kang, H., Yun, H., Im, S., & Choi, C. (2018). Anti-inflammatory effects of a *Stauntonia hexaphylla* fruit extract in lipopolysaccharide activated RAW-264.7 macrophages and rats by Carrageenan-induced hind paw swelling. *Nutrients* 10: 1-12.
- Kiyohara, H., Matsumoto, T., & Yamada, H. (2014). Combination effects of herbs in a multi-herbal formula: expression of Juzen-taiho-to's immune-modulatory activity on the intestinal immune system. *Evidence-Based Complementary and Alternative Medicine* 1: 83-91.
- Klonoff, D.C. (2010). Incretin therapy for type 2 diabetes mellitus. *Advances in Therapy* 27: 881-894.

- Kumbhare, M.R., Sivakumar, T., Udavant, P.B., Dhake, A.S., & Surana, A.R. (2012). Antioxidant activity, phytochemical screening, cytotoxicity and total phenolic content in extracts of *Caesalpinia pulcherrima* (Caesalpinaceae) pods. *Pakistan Journal of Biological Science* 15: 325-332.
- Lawal, U., Maulidiani, M., Shaari, K., Ismail, I.S., Khatib, A., & Abas, F. (2017). Discrimination of *Ipomoea aquatica* cultivars and bioactivity correlations using NMR-based metabolomics approach. *Plant Biosystems* 151: 833-843.
- Latha, S., Vijayakumar, R., Senthil Kumar, B.R., Bupesh, G., Shiva Vijayakumar, T., Manikandan, E., Maazaa, M., Sri Kumar, R., & Deepika, V. (2016). Acute and repeated oral toxicity of antidiabetic polyherbal formulation flax seed, fenugreek and jamun seeds in Wistar albino rat. *Journal of Diabetes and Metabolism* 7: 1-7.
- Lee, H.W., Hakim, P., Rabu, A., & Sani, H.A. (2012). Antidiabetic effect of *Gynura procumbens* leaves extracts involve modulation of hepatic carbohydrate metabolism in streptozotocin-induced diabetic rats. *Journal of Medicinal Plants Research* 6: 796-812.
- Lee, J.H., Park, E., Jin, H.J., Lee, Y., Choi, S.J., Lee, G.W., Chang, P.S., & Paik, H.D. (2017). Anti-inflammatory and anti-genotoxic activity of branched chain amino acids (BCAA) in lipopolysaccharide (LPS) stimulated RAW 264.7 macrophages. *Food Science and Biotechnology* 26: 1371-1377.
- Lee, S.Y., Mediani, A., Nur Ashikin, A.H., Azliana, A.B.S, & Abas, F. (2014). Antioxidant and α -glucosidase inhibitory activities of the leaf and stem of selected traditional medicinal plants. *International Food research Journal* 21: 165-172.
- Li, S., Lin, Z., Jiang, H., Tong, L., Wang, H., & Chen, S. (2016). Rapid identification and assignation of the active ingredients in Fufang Banbianlian injection using HPLC-DAD-ESI-IT-TOF-MS. *Journal of Chromatography Science* 54: 1225-1237.
- Lin, W.Y., Yen, M.H., Teng, C.M., Tsai, I.L., & Chen, I.S. (2004). Cerebrosids from rhizomes of *Gynura japonica*. *Journal of the Chinese Chemical Society* 51: 1429-1434.
- Linda, I.S., & Anthony, J.A. (2015). Phytochemical constituents and antioxidant properties of acetone extract of *Cleome gynandra* L. growing in the Eastern Cape, South Africa. *African Journal of Traditional, Complementary and Alternative Medicines* 12: 1-8.
- Liu, H., Zhang, X., Wu, C., Wu, H., Guo, P., & Xu, X. (2013). Anti-hyperlipidemic caffeoylquinic acids from the fruits of *Pandanus*

- tectorius* Soland. *Journal of Applied Pharmaceutical Science* 3: 016-019.
- Liu, W., Yu, Y., Yang, R., Wan, C., Xu, B., and Cao, S. (2010). Optimisation of total flavonoid compound extraction from *Gynura medica* leaf using response surface methodology and chemical composition analysis. *International Journal of Molecular Sciences* 11: 4750-4763.
- Lubtow, M.M., Haider, M.S., Kirsch, M., Klisch, S., & Luhenxor, R. (2019). Like dissolves like? A comprehensive evaluation of partial solubility parameters to predict polymer-drug compatibility in ultrahigh drug-loaded polymer micelles. *Biomacromolecules* 20: 3041-3056.
- Lynch, K.L. (2017). Mass Spectrometry for Clinical Laboratory. In Hari Nair & William Clarke (Ed), *Toxicology: liquid chromatography mass spectrometry* (pp. 109-130). Elsevier Publication.
- Ma, T., Liaset, B., Hao, Q., Petersen, R.K., Fjære, E., Ngo, H.T., Lillefosse, H.H., Ringholm, S., Sonne, S.B., Treebak, J.T., Pilegaard, H., Frøyland, L., Kristiansen, K., & Madsen, L. (2011). Sucrose counteracts the anti-inflammatory effect of fish oil in adipose tissue and increases obesity development in mice. *PLoS ONE* 6: 1-7.
- Machap, C., & Jaganath, I.B. (2014). High throughput analysis on selected polyphenol production and principal component analysis (PCA) in *Phyllanthus watsonii* grown under different environmental conditions. *Journal of Tropical Agriculture and Food Science* 42: 157-167.
- Mahmood, A.A., Mariod, A.A., Al-Bayaty, F., & Abdel-Wahab, S.I. (2010). Anti-ulcerogenic activity of *Gynura procumbens* leaf extract against experimentally-induced gastric lesions in rats. *Journal of Medicinal Plants Research* 48: 658-691.
- Marsha, J.L. (2012). *15.3B Natural product screening: anti-oxidant screen for extracts*. Retrieved on 2nd June 2018 from <https://www.scribd.com/document/271054618/15-3b-Natural-Product-Screening-Anti-Oxidant-Screen-DPPH-of-Extract-Crude-Extract1>.
- Martucci, M.E.O., De Vos, R.C.H., Corollo, C.A., & Neto, L.G. (2014). Metabolomics as a potential chemotaxonomical tool: application in the genus *Vernonia* Schreb. *PLoS ONE* 9: 1-8.
- Masek, A., Chrzescijanska, E., & Zaborski, M. (2014). Estimation of the antioxidative properties of amino acids - an electrochemical approach. *International Journal of Electrochemical Science* 9: 7904-7915.
- Mediani, A., Abas, F., Khatib, A., Maulidiani, M., Shaari, K., Choi, Y.H., & Lajis, N. (2012). ¹H NMR-based metabolomics approach to

understanding the drying effects on the phytochemicals in *Cosmos caudatus*. *Food Research International* 49: 763-770.

- Mediani, A., Abas, F., Khatib, A., Tan, C.P., Intan, S.I., Shaari, K., Ismail, A., & Lajis, N.H. (2015). Phytochemical and biological features of *Phyllanthus niruri* and *Phyllanthus urinaria* harvested at different growth stages revealed by ¹H NMR-based metabolomics. *Industrial Crops and Products* 77: 602-613.
- Meltem, Y.M., Anneka, M.G., Alexander, J.M., Erk, S., & Balz, F. (2012). Inhibition of α -amylase and α -glucosidase activity by tea and grape seed extracts and their constituents catechins. *Journal of Agricultural, Food and Chemistry* 60: 8924-8929.
- Mishra, S.S., Moharana, S.K., & Dash, M.R. (2011). Review on *Cleome gynandra*. *International Journal of Research in Pharmacy and Chemistry* 1: 681-689.
- Motaal, A.A., Ezzat, S.H., Tadros, M.G., & El-Askary, H.I. (2016). In vivo anti-inflammatory activity of caffeoylquinic acid derivatives from *Solidago virgaurea* in rats. *Pharmaceutical Biology* 54: 2864-2870.
- Moyo, M., Amoo, S.O., Aremu, A.O., Gruz, J., Subrtova, M., Jarosova, M., Tarkowski, P., & Dolezal, K. (2018). Determination of mineral constituents, phytochemicals and antioxidant qualities of *Cleome gynandra*, compared to *Brassica oleracea* and *Beta vulgaris*. *Frontier in Chemistry* 6: 1-9.
- Msika, P., Saunois, A., Baudouin, C., Leclere-Bienfait, S., & Debrock, S. (2013). Extract of the above-ground portions of *Gynandropsis gynandra* or *Cleome gynandra*, and cosmetic, dermatological or pharmaceutical compositions including same. Patent ID: US2013/0171082 A1.
- Muchuweti, M., Mupure, C., Ndhala, A., Murenje, T., & Benhura, M.A.N. (2007). Screening of antioxidant and radical scavenging activity of *Vigna unguiculata*, *Bidens pilosa*, and *Cleome gynandra*. *American Journal of Food Technology* 2: 161-168.
- Mustaffa, F., Indukar, J., Ali, N.I.M., Hanapi, A., Shah, M., Ismail, S., & Mansor, S.M. (2011). A review of Malaysian medicinal plants with potential antidiabetic activity. *Journal of Pharmacy Research* 4: 4217-4224.
- Narendhirakannan, R.T., Kandaswamy, M., & Subramanian, S. (2005). Anti-inflammatory activity of *Cleome gynandra* L. on haematological and cellular constituents in adjuvant-induced arthritic rats. *Journal of Medicinal Food* 8: 93-99.

- Narendhirakannan, R.T., Subramanian, S. & Kandaswamy, M. (2007). Anti-inflammatory and lysosomal stability actions of *Cleome gynandra* L. studied in adjuvant induced arthritic rats. *Food and Chemical Toxicology* 45: 1001-1012.
- National Center for Biotechnology Information, U.S. National Library of Medicine. Rockville Pike, MD 2018. Available at <https://www.ncbi.nlm.nih.gov>. Accessed on 1st September 2018.
- Ncube, E.N., Mhlongo, M., Piater, L.A., Steenkamp, P.A., Dubery, I.A., & Madala, N.E. (2014). Analyses of chlorogenic acids and related cinnamic acid derivatives from *Nicotiana tabacum* tissues with the aid of UPLC-QTOF-MS/MS based on the in-source collision-induced dissociation method. *Chemistry Central Journal* 8: 1-10.
- Ngwe, H., Win, K.C., Kyaw, M.M., Zin, T.T., Thu, K., & Maw, S.S. (2012). Investigation of the bioactive principles and α -glucosidase inhibitory effect of some Myanmar traditional medicinal plants. Paper presented at the 4th AUN/SEED-Net Regional Conference on Biotechnology, Faculty of Engineering, Chulalongkorn University and Burapha University, Thailand, January 26th-27th.
- Niroso, T., Sravanthi, K., & Brahma, R. R. (2016). Evaluation of antidiabetic activity of *Cleome gynandra* leaves. *Indo American Journal of Pharmaceutical Sciences* 3: 482-486.
- Nurusyhadah, I., & Furzani, P. (2021). Formulation of capsulated herbal mixture using *Zingiber officinale* and *Labisa pumila* for postnatal care. *Enhanced Knowledge in Science and Technology* 1: 1-7.
- Ogbonna, J.D.N., Kenechukwu, F.C., Attama, A.A., & Chime, S.A. (2012). Different approaches to formulation of herbal extracts/ phytopharmaceuticals/ bioactive phytoconstituents- a review. *International Journal of Pharmaceutical Sciences Review and Research* 16: 1-8.
- Ogunkunle, A.T.J., Oyelakin, T.M., Enitan, A.O., & Oyewole, F.E. (2014). A quantitative documentation of the composition of two powdered herbal formulations (antimalarial and haematinic) using ethnomedicinal information from Ogbomoso, Nigeria. *Evidence-Based Complementary and Alternative Medicine* 1-8.
- Ojiako, O.K., Chikezie, P.C., & Ogbuji, A.C. (2016). Radical scavenging potentials of single and combinatorial herbal formulations *in vitro*. *Journal of Traditional and Complementary Medicine* 6: 153-159.
- Pandya, U., Doshi, A., & Sahay, N.S. (2017). Development of herbal disinfectants formulation for mopping households and its antibacterial activities. *Natural Products Research* 31: 2665-2668.

- Paterson, I., & Lam, N.Y.S. (2017). Challenges and discoveries in the total synthesis of complex polyketide natural products. *The Journal of Antibiotics* 1-19.
- Parasuraman, S., Gan, S.T., & Dhanaraj, S.A. (2014). Polyherbal formulation: concept of ayurveda. *Pharmacognosy Reviews* 8: 73-80.
- Petchi, R.R., Vijaya, C., and Parasuraman, S. (2014). Antidiabetic activity of polyherbal formulation in streptozotocin-nicotinamide induced diabetic wistar rats. *Journal of Traditional and Complementary Medicine* 4: 108-117.
- Purwal, L., Gupta, S.P.N., & Pande, M.S. (2007). Development and evaluation of herbal formulation for hair growth. *E-Journal of Chemistry* 5: 34-38.
- Rahman, A.F.M.M., & Asad, M.S.A. (2013). Chemical and biological investigation of the leaves *Gynura procumbens*. *International Journal of Biosciences* 3: 36-43.
- Ranjeet, K., Satyanshu, K., Arnab, C., & Sunil, K.C. (2010). High-performance liquid chromatographic method for identification and quantification of three potent liver protective coumarinlignosides-cleomiscosin A, cleomiscosin B and cleomiscosin C-in extracts of *Cleome viscosa*. *Biomedical Chromatography* 24: 1000-1005.
- Ranjitha, J., Madonna, S., Michael, D., & Vijayalakshmi, S. (2014). Isolation of novel phytoconstituents from the stem part of *Cleome gynandra* Linn and their antimicrobial activity. *International Journal of Phytomedicine* 6: 341-345.
- Ravichandra, B., Saketh Ram, P., Saritha, Ch., & Shakaraiah, P. (2014). Anti-diabetic and anti-dyslipidemia activities of *Cleome gynandra* in alloxan induced diabetic rats. *Journal of Pharmacology and Toxicology* 9: 55-61.
- Resat, A., Kubilay, G., Birsen, D., Mustafa, O., Saliha, E.C., Burcu, B., Isil, K.B., & Dilek, O. (2007). Comparative evaluation of various total antioxidant capacity assays applied to phenolic compounds with the CUPRAC assays. *Molecules* 12: 1496-1547.
- Roell, K.R., Reif, D.M., & Motsinger-Reif, A.A (2017). An introduction of terminology and methodology of chemical synergy- perspectives from across disciplines. *Frontiers in Pharmacology* 8: 1-11.
- Rosidah, Yam, M.F., Sadikum, A., & Asmawi, M.Z. (2008). Antioxidant potential of *Gynura procumbens*. *Pharmaceutical Biology* 46: 616-625.
- Rubio, L., Motilva, M.J., & Romero, M.P. (2013). Recent advances in biologically active compounds in herbs and spices: a review of the

most effective antioxidant and anti-inflammatory active principles. *Critical Reviews in Food Science and Nutrition* 53: 943-953.

- Saengsai, J. (2003). *Isolation, identification and HPLC analysis of the lead compound in Gynura procumbens extract and juice*. Master of Science thesis, Mahidol University.
- Sanz, M., Simon, B.F., Cadahia, E., Esterules, E., Munoz, A.M., Hernandez, T., Estrella, I., & Pinto, E. (2012). LC-DAD/ESI-MS/MS study of phenolic compounds in ash (*Fraxinus excelsior* L. and *F. americana* L.) heartwood: effect of toasting intensity at cooperage. *Journal of Mass Spectrometry* 47: 905-918.
- Sari, K.R.P., Sudarsono, & Nugroho, A.E. (2015). Effect of herbal combination of *Andrographis paniculata* (Burm.f) Ness and *Gynura procumbens* (Lour.) Merr ethanolic extracts in alloxan-induced hyperglycemic rats. *International Food Research Journal* 22: 1332-1337.
- Sarot, C., Eun-Jung, P., Bahman, R., John, M.P., & Leng, C.C. (2010). Inhibition of nitric oxide production in lipopolysaccharide (LPS)-activated Murine Macrophage RAW 264.7 cells by the norsesterterpene peroxide, epimuquibilin A. *Murine Drugs* 8: 429-437.
- Schulze, T., René, M., Nikiforos, A., Emma, S., Eric, B., Li D.H., Raalizadeh., Hoffmann, N., Tanaka, S., Michael, W., Treutler, H., & Kohlhoff. (2020). MassBank/MassBank-data: Release version 2020.11. *Zenedo*.
- Seow, L.J., Beh, H.K., Amin Malik, S.A.M, Murugaiyah, V., Ismail, N., & Asmawi, M.Z. (2011). Anti-angiogenic activity of *Gynura segetum* leaf extracts and its fraction. *Journal of Ethnopharmacology* 134: 221-227.
- Shimizu, Y., Imayoshi, Y., Kato, M., Maeda, K., Iwabuchi, H., & Shimomura, K. (2011). New eudesmane-type sesquiterpenoids and other volatile constituents from the roots of *Gynura bicolor* DC. *Flavour and Fragrance Journal* 26: 55-64.
- Simirgiotis, M.J., Benites, J., Areche, C., & Sepulveda, B. (2015). Antioxidant capacities and analysis of phenolic compounds in three endemic *Nolana* species by HPLC-PDA-ESI-MS. *Molecules* 20: 11490-11507.
- Siti Pauliena, M.B. (2006). *Determination of anti-diabetic activity of Gynura procumbens using bioassay-guided fractionation*. Master of Science thesis, University Putra Malaysia.
- Sivanesan, D., & Hazeena, B.V. (2007). Preventive role of *Gynandropsis gynandra* L., against aflatoxin B1 induced lipid peroxidation and antioxidant defense mechanism in rat. *Indian Journal of Experimental Biology* 45: 299-303.

- Sjoholm, A., & Nystrom, T. (2006). Inflammation and the etiology of type 2 diabetes. *Diabetes Metabolism Research and Reviews* 22: 4-10.
- Spinola, V., Pinto, J., & Catilho, P.C. (2014). Identification and quantification of phenolic compounds of selected fruits from Madeira island by HPLC-DAD-ESI-MSⁿ and screening for their antioxidant activity. *Food Chemistry* 173: 14-30.
- Spyros, A., & Dais, P. (2012). *NMR spectroscopy in food analysis*. UK: RSC Publication.
- Sunmonu, T.O., & Afolayan, A.J. (2013). Evaluation of antidiabetic activity and associated toxicity of *Artemisia afra* aqueous extract in Wistar rats. *Evidence-Based Complementary Alternative Medicine* 2013: 1-8.
- Taewoo, J., Kandhasamy, S., Sunghyun, H., Jaehak, L., Sun-Yung, P., Songmum, K., & Jin-Woo, J. (2014). Inhibition of nitric oxide production in LPS-stimulated RAW 264.7 cells by stem bark of *Ulmus pumila* L. *Saudi Journal of Biological Sciences* 21: 427-435.
- Tan, H.L., Chan, K.G., Pusparajah, P., Lee, L.H., & Goh, B.H. (2016). *Gynura procumbens*: an overview of biological activities. *Frontiers in Pharmacology* 7: 1-14.
- Tolonen, A., Joutsamo, T., Mattila, S., Kamarainen, T., & Jalonen, J. (2002). Identification of isomeric dicaffeoylquinic acids from *Eleutherococcus senticosus* using HPLC-ESI/TOF/MS and ¹H-NMR methods. *Phytochemical Analysis* 13: 316-328.
- Trilocana, Y., Dasi Jeeva, M.B., & Rajeswara, R.P. (2017). The study of anti-diabetic activity of aqueous extract of root of *Gynandropis gynandra* in diabetic rats. *Indian Journal of Research in Pharmacy and Biotechnology* 5: 13-18.
- Vidushi, K., Shilpa, G., Neha, S., Ashok, K., Nazir, A.L., Mowkshi, K., Prabhu, D., Parduman, R.S., Asha, B., & Zabeer, A. (2017). Anti-inflammatory potential of hentriacontane in LPS stimulated RAW 264.7 cells and mice model. *Biomedicine & Pharmacotherapy* 92: 175-186.
- Vladimir, T. (2016). Metabolomics: bridging the gap between pharmaceutical development and population health. *Metabolites* 6: 1-13.
- Wang, J., Fang, X., Ge, L., Cao, F., Zhao, L., Wang, Z., & Xiao, W. (2018). Antitumor, antioxidant and anti-inflammatory activities of kaempferol and its corresponding glycosides and the enzymatic preparation of kaempferol. *PLoS ONE* 13: 1-12.

- Wei, X., Song, H., Yin, L., Rizzo, M.G., Sidhu, R., Covey, D.F., Ory, D.S., & Semenkovich, C.F. (2016). Fatty acid synthesis configures the plasma membrane for inflammation in diabetes. *Nature* 539: 294-298.
- Wellen, K. & Hotamisligil, G.S (2005). Inflammation, stress, and diabetes. *Journal of Clinical Investigation* 115: 1111-1119.
- Wen-Fei C., Chieh-Fu, C., & Jin-Jung, L. (2000). Mechanism of suppression of inducible nitric oxide synthase (iNOS) expression in RAW 264.7 cells by andrographolide. *British Journal of Pharmacology* 29: 1553-1560.
- Widyawati, P.S., Budianta, T.D.W., Kusuma, F.A., & Wijaya, E.L. (2014). Difference of solvent polarity to phytochemical content and antioxidant activity of *Pluchea indica* Less leaves extracts. *International Journal of Pharmacognosy and Phytochemical Research* 15: 850-855.
- Wilson, L.T. Statistical correlation (2009). Available at <https://explorable.com/statistical-correlation>. Accessed January 31, 2019.
- Windono, T., Jenie, U.A., & Kardono, B.S. (2012). Isolation and elucidation of Pyrrolizidine alkaloid from tuber *Gynura pseud-china* (L) DC. *Journal of Applied Pharmaceutical Science* 05: 2012.
- Wishart, D.S., Feunang, Y.D., Marcu, A., Guo, A.C., Liang, K., et al., (2018). HMDB 4.0-The Human Metabolome Database for 2018. *Nucleic Acids Research* 46: 608-617.
- World Health Organisation. *Attaining the nine global non-communicable diseases targets: a shared responsibility*. Global Status Report on non-communicable disease (2014).
- Wong, S.K., Lee, M.S.J., Sudi, S., Hassan, R.M., Lee, P.C., Embi, N. & Sidek, H. (2015). Anti-malarial and anti-inflammatory effects of *Gynura procumbens* are mediated by kaempferol via inhibition of glycogen synthase kinase-3 β (GSK3 β). *Sains Malaysiana* 44: 1489-1500.
- Yang, Y., Zhang, Z., Li, S., Ye, X., Li, X., & He, K. (2014). Synergy effects of herb extracts: pharmacokinetics and pharmacodynamics basis. *Fitoterapia* 92: 133-147.
- Yuan, H., Ma, Q., Ye, L., & Piao, G. (2016). The traditional medicine and modern medicine from natural products. *Molecule* 21: 1-18.
- Yoo, H., Ku, S.K., Baek, Y.D., & Bae, J.S. (2014). Anti-inflammatory effects of rutin on HMGB1-induced inflammatory responses in vitro and in vivo. *Inflammatory response* 63: 197-206.

- Yun, E.S., Park, S.K., Kim, B.S., Chae, Y.Z., Cho, S.M., Yi, H., Cho, H.J., & Shin, H.C. (2011). Determination of the esculetin contents of medicinal plants by liquid chromatography-tandem mass spectrometry. *Biomedical Chromatography* 26: 1247-1251.
- Yue, L., Zhanying, H., Guangguo, T., Xin, D., Genjin, Y., Liang, Z., Xiaofei, C., Zhenyu, Z., Ziyang, L., Baohua, Q., Guoqing, Z., & Yifeng, C. (2014). NMR and LC/MS- based global metabolomics to identify serum biomarkers differentiating hepatocellular carcinoma from liver cirrhosis. *International Journal of Cancer* 135: 658-668.
- Zahra, A.A., Kadir, F.A., Mahmood, A.A., Al Hadi, A.A., Suzy, S.M., Sabri, S.Z., Latif, I.I., & Ketuly, K.A. (2011). Acute toxicity study and wound healing potential of *Gynura procumbens* leaf extracts in rats. *Journal of Medicinal Plants Research* 5: 2551-2558.
- Zhang, J., Chen, M., Ju, W., Liu, S., Xu, M., Chu, J., & Wu, T. (2009). Liquid chromatography/tandem mass spectrometry assay for the simultaneous determination of chlorogenic acid and cinnamic acid in plasma and its application to a pharmacokinetic study. *Journal of Pharmaceutical and Biomedical Analysis* 51: 685-690.
- Zhang, X.F. & Tan, B.K.H. (2000). Effects of an ethanolic extract of *Gynura procumbens* on serum glucose, cholesterol and triglyceride levels in normal and streptozotocin-induced diabetic rats. *Singapore Medical Journal* 41.
- Zhou, X., Seto, S.W., Chang, D., Kiat, H., Razmovski-Naumovski, V., Chan, K., & Bensoussan, A. (2016). Synergistic effects of Chinese herbal medicine: a comprehensive review of methodology and current research. *Frontiers in Pharmacology* 7: 1-16.
- Zwyrzykowska, A., Kupczynski, R., Jarosz, B., Szumny, A., & Kucharska, A.Z. (2015). Qualitative and quantitative analysis of polyphenolic compounds in *Ilex* Sp. *Open Chemistry* 13: 1303-1312.