



UNIVERSITI PUTRA MALAYSIA

***GENETIC DIVERSITY AND EXPRESSION ANALYSES OF SECRETED IN
XYLEM EFFECTOR GENES OF *Fusarium oxysporum* f. sp. cubense
ISOLATED FROM PENINSULAR MALAYSIA***

SUHANNA BINTI AHMAD

FP 2021 7



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By

SUHANNA BINTI AHMAD

**Thesis Submitted to School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Degree of Master of
Science**

July 2021

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Abstract of the thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirements for the degree of Master of Science

GENETIC DIVERSITY AND EXPRESSION ANALYSES OF *SECRETED IN XYLEM* EFFECTOR GENES OF *Fusarium oxysporum* f. sp. *cubense* ISOLATED FROM PENINSULAR MALAYSIA

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SUHANNA BINTI AHMAD

July 2021

Chair : Dzarifah binti Mohamed Zulperi, PhD
Faculty : Agriculture

Banana is a commercially important fruit in the majority of tropical and subtropical countries. Banana plantations cover nearly 35,000 hectares of land, accounting for approximately 24% of Malaysia's total fruit production, with Johor plantations accounting for approximately 32% of the nation's banana production. The *Fusarium* wilt pathogen, *Fusarium oxysporum* f. sp. *cubense* (Foc) is a major cause of disease in the banana industry, limiting banana production in Malaysia and causing significant economic losses. Currently, the majority of locally grown bananas (Berangan, Rasthali, Mas) and plantains used for cooking or processing (Raja, Abu, and Awak) have been found to be susceptible to Foc-TR4. To date, AFLP, RAPD, ISSR and SSR markers are extensively used in PCR-based molecular characterization of Foc. Although these markers are effective in characterizing Foc, they all have their own limitations. An effector gene known as *Secreted in Xylem* (*SIX*) has been explored to broaden the molecular diagnostic toolbox for Foc. The main objective of this study was to identify expression of *SIX* genes in banana plantlet during infection with the Foc-TR4. This will be an important step in understanding the pathogenicity response of the plant and will help to develop any disease management strategies. In this study, the virulence potential of 27 isolates of Foc of banana based on pathogenicity test and the presence of *SIX1*, *SIX7* and *SIX8* genes were determined. All of 27 Foc 'Tropical Race 4' (TR4) (VCG 01213/160) isolates were obtained from Biological Control Laboratory, Department of Plant Protection, University Putra Malaysia (UPM). All isolates were pathogenic towards *Musa acuminata* Berangan AAA at different severity levels under greenhouse condition. Pathogenicity assays indicated that the level of aggressiveness varies between isolates. The Foc-TR4 isolates were screened for the presence of three *SIX* genes (*SIX1*, *SIX7* and *SIX8*) and the genetic differentiation of Foc-TR4 was evaluated by PCR analysis using three specific primers (*SIX1*, *SIX7* and *SIX8*). Among these three *SIX* genes, *SIX1* and

SIX8 genes were detected in all 27 Foc-TR4 isolates from Peninsular Malaysia. The presence of *SIX1* and *SIX8* in Foc-TR4 suggested these genes were likely involved in pathogenicity on banana. Phylogenetic analysis using *SIX1* and *SIX8* sequences showed that they were related to TR4 isolates (VCG 01213/16) from Australia and Indonesian with bootstrap values of 94% and 100%, respectively. There was no variation observed in the *SIX1* and *SIX8* sequences since all 27 isolates clustered in the same clade. Virulence level do not correlate with the existence or lack of these genes. Highest and lowest disease severity values (80.73% and 10.42%) on banana plantlet were selected. The T30 and T36 isolates were found to be highly aggressive and weakly aggressive, respectively. Both T30 and T36 isolates were analyzed by qRT-PCR analyses for the expression of *SIX1* and *SIX8* genes to elucidate whether these genes play a role in the pathogenicity response of Foc-TR4 on *Musa acuminata* cv. Berangan plantlets during early and late of infection stages. Expression analyses of *SIX1* and *SIX8* were constructed from banana roots collected after 0, 2, 4, 8, 12, 15, 20- and 30-day post-infection (dpi). The *SIX1* and *SIX8* genes were expressed after being induced by the host and showed higher expression at 8 dpi and 12 dpi respectively in both T30 and T36 isolates. From these results, it can be concluded that highly expressed virulence-associated genes, *SIX1* and *SIX8* can be used as markers for the assessment of Foc-TR4 virulence.

Abstrak tesis yang dikemukakan kepada Senat Universiti Universiti Putra
Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

**KEPELBAGAIAN GENETIK DAN ANALISIS EKSPRESI EFEKTOR GEN
SECRETED IN XYLEM *Fusarium oxysporum* f. sp. *cubense* DIISOLASI
DARI SEMENANJUNG MALAYSIA**

Oleh

SUHANNA BINTI AHMAD

Julai 2021

Pengerusi : Dzarifah binti Mohamed Zulperi, PhD
Fakulti : Pertanian

Pisang adalah salah satu buah yang penting di kebanyakan negara tropika dan subtropika. Ladang pisang meliputi hampir 35,000 hektar tanah, merangkumi sekitar 24% dari keseluruhan pengeluaran buah di Malaysia dengan ladang di Johor menyumbang sekitar 32% daripada pengeluaran pisang negara. Layu *Fusarium* yang di sebabkan oleh patogen *Fusarium oxysporum* f. sp. *cubense* (Foc) adalah salah satu penyakit utama dalam industri pisang, yang mengehadkan pengeluaran pisang di Malaysia dan menyebabkan kerugian ekonomi yang ketara. Pada masa ini, sebilangan besar pisang yang ditanam (Berangan, Rastali, Mas) dan pisang yang digunakan untuk memasak atau memproses (Raja, Abu, Awak) didapati rentan terhadap Foc-TR4. Sehingga kini, penanda SSR, ISSR, AFLP dan RAPD digunakan secara meluas dalam pencirian Foc di peringkat molekul berasaskan PCR. Walaupun penanda ini berkesan dalam pencirian Foc, tetapi penggunaannya masih terbatas. Gen efektor yang dikenali sebagai *Secreted in Xylem (SIX)* telah diterokai untuk perluaskan diagnostik molekul Foc. Objektif utama kajian ini adalah untuk mengenal pasti pengekspresan gen *SIX* pada tanaman pisang selepas dijangkiti dengan Foc-TR4. Ini akan menjadi langkah penting dalam memahami tindak balas kepatogenan tanaman dan dapat membantu membangunkan strategi pengurusan penyakit. Dalam kajian ini, potensi kevirulenan bagi 27 pencilan Foc pisang berdasarkan ujian kepatogenan dan kehadiran gen *SIX1*, *SIX7* dan *SIX8* telah ditentukan. Kesemua 27 pencilan Foc 'Tropical Race 4' (TR4) (VCG 01213/160) diperolehi dari Makmal Kawalan Biologi, Jabatan Perlindungan Tanaman, Universiti Putra Malaysia (UPM). Kesemua pencilan adalah patogenik terhadap *Musa acuminata* Berangan AAA pada tahap keterukan penyakit yang berbeza di dalam rumah hijau. Ujian kepatogenan menunjukkan bahawa tahap keagresifan antara pencilan adalah berbeza. Pencilan Foc-TR4 disaring untuk mengetahui kehadiran tiga gen *SIX* (*SIX1*, *SIX7* dan *SIX8*) dan pembezaan genetik Foc-TR4 dinilai melalui analisis

PCR menggunakan tiga primer khusus (*SIX1*, *SIX7* dan *SIX8*). Di antara ketiganya gen *SIX* ini, gen *SIX1* dan *SIX8* dikesan pada kesemua 27 pencilan Foc-TR4 dari Semenanjung Malaysia. Kehadiran *SIX1* dan *SIX8* dalam Foc-TR4 menunjukkan bahawa gen ini berkemungkinan terlibat dalam kepatogenesis pada tanaman pisang. Analisis filogenetik pada kawasan jujukan *SIX1* dan *SIX8* menunjukkan bahawa mereka berkait rapat dengan pencilan TR4 (VCG 01213/16) dari Australia dan Indonesia dengan nilai bootstrap masing-masing 94% dan 100%. Tiada variasi yang diperhatikan dalam jujukan *SIX1* dan *SIX8* kerana kesemua 27 pencilan berada di dalam kelompok yang sama. Tahap kevirulenan tidak berkait dengan kewujudan atau kekurangan gen ini. Nilai keterukan penyakit yang paling tinggi dan paling rendah (80.73% dan 10.42%) dipilih untuk kajian seterusnya. Pencilan T30 dan T36 masing-masing didapati paling agresif dan paling lemah. Kedua-dua pencilan T30 dan T36 dianalisa berdasarkan analisis qRT-PCR untuk mengetahui tahap pengekspresan gen *SIX1* dan *SIX8* bagi melihat sama ada gen ini berperanan dalam tindak balas kepatogenesis Foc-TR4 pada *Musa acuminata* cv. Berangan semasa peringkat awal dan akhirjangkitan. Analisis pengekspresan gen *SIX1* dan *SIX8* diambil daripada akar pisang selepas diinokulasi dengan Foc-TR4 pada 0, 2, 4, 8, 12, 15, 20- dan 30 hari (dpi). Gen *SIX1* dan *SIX8* terekspres selepas diaruh oleh perumah dan menunjukkan aras pengekspresan yang tinggi pada hari ke lapan selepas inokulasi bagi pencilan T30 dan hari ke-12 selepas inokulasi bagi pencilan T36. Daripada hasil kajian ini, dapat disimpulkan bahawa gen *SIX1* dan *SIX8* berkait dengan kevirulenan dan dapat digunakan sebagai penanda untuk penilaian tahap kevirulenan Foc-TR4.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirements for the Degree of Master of Science. The members of the Supervisory Committee were as follows:

Dzarifah binti Mohamed Zulperi, PhD

Senior Lecturer
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Ganesan Vadamalai, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

Mohd Aswad bin Adul Wahab, PhD

Senior Lecturer
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

Noor Baity Saidi, PhD

Associate Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 11 November 2021

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Signature: _____

Name of Chairman
of Supervisory
Committee:

Dzarifah binti Mohamed Zulperi, PhD

Signature: _____

Name of Member of
Supervisory
Committee:

Ganesan Vadamalai, PhD

Signature: _____

Name of Member of
Supervisory
Committee:

Mohd Aswad bin Abdul Wahab, PhD

Signature: _____

Name of Member of
Supervisory
Committee:

Noor Baity binti Saidi, PhD

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LIST OF ABBREVIATIONS

x g	Gravitational Constant
ANOVA	Analysis of variance
BLAST	Basic local alignment system tool
cDNA	Complimentary deoxyribonucleic acid
CFU	Colony forming unit
DNA	deoxyribonucleic acid
dNTP	Deoxyribonucleotide triphosphate
EDTA	Ethylenediaminetetraacetic acid
EtBr	Ethidium bromide
ITS	Intergenic space
MEGA	Molecular evolutionary genetics analysis
MgCl ₂	Magnesium chloride
NCBI	National Centre for Biotechnology Information
PDA	Potato Dextrose Agar
PCR	Polimerase Chain Reaction
qRT-PCR	Real-time reverse transcription polymerase chain reaction
RNA	Ribonucleic acid
RNase	Ribonuclease
Rpm	Nanogram
nm	Rotation per minute
SIX	Secreted in xylem
rpm	Rotation per minute
TAE	Tris-acetate EDTA
TBE	Tris HCL-boric acid-EDTA

TEF1- α	Translation elongation factor one alpha
<i>Taq</i>	<i>Thermas aquaticus</i>
TM	Melting temperature
<i>TUB2</i>	Beta Tubulin
USDA	United States Department of Agriculture





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CHAPTER 1

INTRODUCTION

1.1 Background of study

Bananas are an extremely important crop in Asia and the Pacific, both as a staple food and as a major export commodity for many developing countries. In Malaysia, banana is ranked second, after durian, as the most cultivated fruit crop in Malaysia, accounting for about 24% of total fruit production, primarily in Johor, Pahang, Sarawak and Sabah (MOA, 2019). Banana production in Malaysia is a lucrative business account for more than 330,000 metric tons of banana production and such enormous scale has generated revenue of approximately USD8 million of export value (FAO, 2014). Approximately 50% of locally cultivated bananas are AAA subgroups, Pisang Berangan and Cavendish type (Mohamad Roff et al., 2012). This crop has become an important commodity in Malaysia, with Cavendish types and Pisang Berangan planted on a commercial scale for both the domestic and export markets.

However, like many other crops, banana is also subjected to the attack of the fungal pathogen. Indeed, one of the main problems threatening the banana industry today both locally and globally is the banana *Fusarium* wilt disease. Most edible banana cultivars in Malaysia were reported to be susceptible to *Fusarium* wilt, caused by the soil inhabiting fungus, *Fusarium oxysporum* f. sp. *cubense* Tropical Race 4 (Foc-TR4) which has caused an estimated total loss of US\$ 14.1 million in Malaysia alone (Aquino et al., 2013; Ploetz, 2015). The retail price of bananas has risen from RM1.53 per kg in 2000 to RM 4.25 per kg in 2018 because of the Foc-TR4 outbreak (Tumin and Shahrudin, 2019). There is also a low genetic diversity between plantations due to selective planting of common cultivars (Fourie et al., 2011). This monoculture practice is always a predisposing factor for pest and disease outbreaks (Ploetz, 2015).

Currently there are no effective measures to control this disease as the nature of the pathogen is a soil-borne, mutate easily and a vascular inhabitant. Foc-TR4 can be easily spread from one farm to another by moving infected planting materials between farms, transporting human fungal propagules, farm tools, vehicles, irrigation as well as flood waters (Dita et al., 2018). Before suitable management strategies are implemented to control *Fusarium* wilt, a thorough understanding of their genetic diversity and virulence is essential.

1.2 Problem statements and significance of study

Understanding the biology of Foc-TR4 isolates is necessary prior to disease management (Czislowski et al., 2018). Generally, traditional pathogenicity test was used to distinguish pathogenic and non-pathogenic isolates under greenhouse condition and through molecular methods based on standard approaches. To date, most genetic diversity studies of Foc, using common SSR, ISSR, AFLP or RAPD markers did not show correlation between the phylogenetic data and virulence level of Foc. Since aggressiveness components are defined by quantitative traits, markers derived from fungal virulence or effector genes might present an attractive alternative as genetic marker. Hence, one of the necessary preparations in overcoming the problem is to discover more fungal effectors genes from pathogenicity traits. Advances in diversity evaluation methods have shown that the currently defined race structure does not sufficiently reflect the extent of genetic and phenotypic variation in Foc (Ploetz and Pegg, 2000a; Dita et al., 2010; Fourie et al., 2011).

Secreted in Xylem (SIX) genes were the first effector identified in *Fusarium oxysporum* f. sp. *lycopersici* (Fol), a causal agent of *Fusarium* wilt of tomato (Ma et al, 2010). The molecular characteristics and presence of *SIX* genes in Foc-TR4 isolates from Malaysia have not been studied in detail. To date, PCR assays and subsequent amplicon sequences are performed using a housekeeping gene primer such as the *Translation Elongation Factor (TEF1- α)* to investigate the phylogenetic relationship among locally isolated Foc and other isolates. The ability of *SIX* genes to be used as genetic markers in the phylogenetic analysis has not yet been evaluated. Reliable pathogenicity-based markers are required to distinguish between the phylogenetic data and the virulence levels of Foc isolates. Pathogenicity testing is therefore needed to understand the biology of local isolates prior to the implementation of any disease management strategies. It is therefore important to examine whether *SIX1*, *SIX7* and *SIX8* genes can clearly determine the genetic relationship between Foc-TR4 isolates in Peninsular Malaysia. These three genes were chosen based on previous reports that the presence or absence of these genes is capable of recognizing Foc in bananas. Numerous effector genes which express small, secreted proteins, such as *SIX1*, are specifically enhanced during infection (Rep et al., 2005). The expression of *SIX1* and *SIX8* was evaluated using qRT-PCR in order to study the circumstances requiring the use of Foc effectors and to identify the host signal that induces the development of these proteins. These expression patterns can be used as standards in pathogenicity studies, which are normally used to assess a plant's response to infection. These *SIX* gene could be used as a diagnostic tool to detect the presence of *Fusarium* wilt in farms. It can assist farmers in protecting their crops from *Fusarium* wilt and eliminating it at an early stage of infection. Bananas of high quality will open up more market opportunities for our agricultural products. Aside from that, investigating the role of known pathogenicity-related genes involved is a necessary step toward understanding how resistant plants are produced. Data derived from this study will serve as an invaluable genomic reference that will further our understanding of the molecular activities that occur specifically in banana plants during their early and late response to Foc-TR4. Potential effector genes can also be used

as DNA markers in the development of durable disease resistance strategies (Gibriel et al., 2016). The invention identifies gaps in current knowledge about the ability of *SIX* genes to function at the early and late stages of infection and provide insights into future research that can be pursued to improve our understanding of the functions of *SIX* genes.

1.3 Research objectives

The aim of this research was to address the identified research problems with the following research objectives:

1. To identify pathogenic variation and detect the presence of *SIX* genes of *F.oxysporum* f. sp. *cubense* Tropical Race 4 (Foc-TR4).
2. To determine the phylogenetic relationships between Foc-TR4 isolates using *SIX* genes.
3. To investigate the expression profile of *SIX* genes in Foc-TR4 isolates at different time-point through quantitative real-time PCR analyses (qRT-PCR).

REFERENCES

- Ab Halim, N. (2016). Policy intervention for the development of the pineapple industry in Malaysia. Taipei, Taiwan: FFTC Agricultural Policy Articles.
- Alfano, J.R., and Collmer, A (2004). Type III secretion system effector proteins: double agents in bacterial disease and plant defense. *Annual Review of Phytopathology*, 42(1): 385 – 414.
- Agrios, G.N. (2005) *Plant Pathology*, 5th edn. San Francisco, CA: Elsevier Academic Press.
- Ammar, M.I. (2007). *Fusarium* species associated with corm rots and wilt of banana (*Musa* sp.) under Egyptian conditions. *Egyptian Journal of Phytopathology*, 35: 81-98.
- An, B., Hou, X., Guo, Y., Zhao, S., Luo, H., He, C., and Wang, Q. (2019). The effector *SIX8* is required for virulence of *Fusarium oxysporum* f.sp. *cubense* tropical race 4 to Cavendish banana. *Fungal Biology*, 123: 423-430
- Andersen, C.L., Jensen, J.L. and Orntoft, T.F. (2004). Normalization of real-time quantitative reverse transcription-PCR data: A model-based variance estimation approach to identify genes suited for normalization, applied to bladder and colon cancer data sets. *Cancer Research*, 64:5245-5250.
- Aquino, A.P., Bandoles, G.G., and Lim, V.A.A. (2013). R&D and policy directions for effective control of *Fusarium* wilt disease of Cavendish banana in the Asia-Pacific region. Taipei, Taiwan: FFTC Agricultural Policy Articles.
- Araújo, N.A.F., Pasqual, M., Pio, L.A.S., Alves, E., de Matos Moura, N., and da Silva Costa, S. (2017). Identification and aggressiveness of four isolates of *Fusarium oxysporum* f. sp. *cubense* from Latundan banana in Brazil. *Journal of Phytopathology*, 165: 257-264.
- Bang, A., Xingrong, H., Yunfeng, G., Shixue, Z., Hongli, L., Chaozu, H., and Qiannan, W. (2019). The effector *SIX8* is required for virulence of *Fusarium oxysporum* f.sp. *cubense* tropical race 4 to Cavendish banana. *Fungal Biology*, 123(5):423–430.
- Bentley, S., Pegg, K.G., Moore, N.Y., Davis, R.D., and Buddenhagen, I.W. (1998). Genetic variation among vegetative compatibility groups of *Fusarium oxysporum* f. sp. *cubense* analysed by DNA fingerprinting. *Phytopathology*, 88: 1283-1293.

- Bhadauria, V., MacLachlan, R., Pozniak, C., and Banniza, S. (2015). Candidate effectors contribute to race differentiation and virulence of the lentil anthracnose pathogen *Colletotrichum lentis*. *BMC Genomics*, 16(1).
- Buddenhagen, I.W. (2009). Understanding strain diversity in *Fusarium oxysporum* f. sp. *cubense* and history of introduction of 'tropical race 4' to better manage banana production. *Acta Horticulturae*, 828: 193-204.
- Bustin, S. A., Benes, V., Garson, J. A., Hellemans, J., Huggett, J., Kubista, M., and Wittwer, C.T. (2009). The MIQE guidelines: Minimum information for publication of quantitative real-time PCR experiments. *Clinical Chemistry*, 55(4), 611–622.
- Butler, D. (2013). Fungus threatens top banana. *Nature*, 12: 195-196.
- Cao, S., Yang, N., Zhao, C., Liu, J., Han, C., and Wu, X. (2018). Diversity of *Fusarium* species associated with root rot of sugar beet in China. *Journal of General Plant Pathology*, 84(5):321–329.
- Carvalhais, L.C., Henderson, J., Rincon-Florez, V.A., O'Dwyer, C., Czislawski, E., Aitken, E.A.B., and Drenth, A. (2019). Molecular diagnostics of banana *Fusarium* wilt targeting secreted-in xylem genes. *Frontiers in Plant Science*, 10:547.
- Casadevall, A. and Pirofski, L. (1999). Host–pathogen interactions: redefining the basic concepts of virulence and pathogenicity. *Infection and Immunity*, 67, 3703–3713.
- Catanzariti, A.M., Lim, G.T.T. and Jones, D.A. (2015) The tomato I-3 gene: a novel gene for resistance to *Fusarium* wilt disease. *New Phytologist*, 207(1),106–118.
- Catanzariti, A.M., Do, H.T.T., Bru, P., de Sain, M., Thatcher, L.F., Rep, M. and Jones, D.A. (2017). The tomato I gene for *Fusarium* wilt resistance encodes an atypical leucine-rich repeat receptor-like protein whose function is nevertheless dependent on SOBIR1 and SERK3/BAK1. *The Plant Journal*, 89(6), 1195–1209.
- CCAFS (2017). Annual report 2016: Power of partnerships. Wageningen, The Netherlands: CGIAR Research Program on Climate Change, Agriculture and Food Security (CCAFS). Available online at: bitly.com/ccafs2016

- Chakrabarti, A., Rep, M., Wang, B., Ashton, A., Dodds, P., and Ellis, J., (2011). Variation in potential effector genes distinguishing Australian and non-Australian isolates of the cotton wilt pathogen *Fusarium oxysporum* f. sp. *vasinfectum*. *Plant Pathology*, 60, 232–43.
- Covey, P.A., Kuwitzky, B., Hanson, M., and Webb, K.M. (2014). Multilocus analysis using putative fungal effectors to describe a population of *Fusarium oxysporum* from sugar beet. *Phytopathology*, 104(8):886–896.
- Cunha, C.M.S., Hinz, R.H., Pereira, A., Tcacenco, F.A., and Stadnik, M.J. (2015). Aggressiveness and genetic diversity of *Fusarium oxysporum* f. sp. *cubense* from Santa Catarina, southern Brazil. *Tropical Plant Pathology*, 40(5):326–334.
- Czislowski, E., Fraser-Smith, S., Zander, M., O'Neill, W.T., Meldrum, R.A., Tran-Nguyen, L.T.T., Batley, J., and Aitken, E.A.B. (2018). Investigation of the diversity of effector genes in the banana pathogen, *Fusarium oxysporum* f. sp. *cubense*, reveals evidence of horizontal gene transfer. *Molecular Plant Pathology*, 19(5):1155–1171. 1155.
- Dale, J., James, A., Paul, J.Y., Khanna, H., Smith, M., Peraza-Echeverria, S. and Harding, R. (2017). Transgenic Cavendish bananas with resistance to *Fusarium* wilt tropical race 4. *Nature Communications*, 8(1).
- Damodaran, T., Rajan, S., Mishra, V.K., Jha, S.K., Ahmad, I., and Gopal, R. (2018). First report of *Fusarium* wilt in banana caused by *Fusarium oxysporum* f. sp. *cubense* Tropical Race 4 in India. *Plant Disease*, DOI: 10.1094/PDIS-07-18-1263-PDN.
- Davidson, J., and Ammann, K. (2017). New GMO regulations for old: Determining a new future for EU crop biotechnology. *GM Crops & Food*, 8(1): 13-34.
- de Guillen, K., Ortiz-Vallejo, D., Gracy, J., Fournier, E., Kroj, T., and Padilla, A. (2015). Structure analysis uncovers a highly diverse but structurally conserved effector family in phytopathogenic fungi. *PLoS Pathogen*, 11 (10): e1005228.
- de Jonge, R., Bolton, M.D., and Thomma, B.P. (2011). How filamentous pathogens co-opt plants: the ins and outs of fungal effectors. *Current Opinion in Plant Biology*, 14(4), pp. 400-406.
- DeLaughter, D. M. (2018). The Use of the Fluidigm C1 for RNA Expression Analyses of Single Cells. *Current Protocols in Molecular Biology*, 122(1), e55.

- de Sain, M., and Rep, M. (2015). The Role of Pathogen-Secreted Proteins in Fungal Vascular Wilt Diseases. *International Journal of Molecular Sciences*, 16(10), 23970–23993.
- D'Hont, A., Denoeud, F., Aury, J.-M., Baurens, F.-C., Carreel, F., Garsmeur, O., and Rouard, M. (2012). The banana (*Musa acuminata*) genome and the evolution of monocotyledonous plants. *Nature*, 488:213.
- Di Pietro, A., Madrid, M.P., Caracuel, Z., Delgado-Jarana, J., and Roncero, M.I.G. (2003). *Fusarium oxysporum*: exploring the molecular arsenal of a vascular wilt fungus. *Molecular Plant Pathology*, 4:315-325.
- Dita, M.A., Waalwijk, C.I., Buddenhagen, I.W., Souza, M.T., and Kema, G.H.J. (2010). A molecular diagnostic for tropical race 4 of the banana *Fusarium* wilt pathogen. *Plant Pathology*, 59(2):348–357.
- Dita, M.A., Waalwijk, C., Paiva, L.V., Souza, J.R.M.T., and Kema, G.H.J. (2011). A greenhouse bioassay for the *Fusarium oxysporum* f. sp. *cubense* x 'Grand Naine' (Musa, AAA, Cavendish subgroup) interaction. *Acta Horticulturae*, 897(897):377–380.
- Dita, M.A.R., Vicente, L.P., and Martínez, E. (2014). Inoculation of *Fusarium oxysporum* f. sp. *cubense* causal agent of *Fusarium* wilt in banana. In: Vicente, L.P., Dita, M.A.R. and Martínez, E. (eds.) *Technical Manual: Prevention and Diagnostic of Fusarium Wilt (Panama Disease) of Banana caused by Fusarium oxysporum f. sp. cubense Tropical Race 4 (TR4)*. FAO, United Nations pp. 55-58.
- Dita, M., Barquero, M., Heck, D., Mizubuti, E.S.G., and Staver, C.P. (2018). *Fusarium wilt* of banana: knowledge on epidemiology and research needs toward sustainable disease management. *Frontiers in Plant Science*, 9: 1468.
- Ellis, M.L., Lanubile, A., Garcia, C., and Munkvold, G.P. (2016). Association of putative fungal effectors in *Fusarium oxysporum* with wilt symptoms in soybean. *Phytopathology*, 106: 762-773.
- FAO, (2014). *Banana market review and banana statistics 2012–2013*. In: Intergovernmental group on bananas and tropical fruits. Rome: FAO publications.
- FAOSTAT (2017). *Banana market review 2015-2016*. Rome: Food and Agriculture Organization of the United Nations pp. 1-7.
- FAOSTAT (2018a). *Banana market review 2017*. Rome: Food and Agriculture Organization of the United Nations pp. 1-5.

- FAOSTAT (2018b). Banana market review: Preliminary results for 2018. Rome: Food and Agriculture Organization of the United Nations pp. 1-12.
- Flood, J. and Day, R. (2015). Managing risks from pests in global commodity networks – policy perspectives. *Food Security*, 8(1): 89–101.
- Fourie, G., Steenkamp, E.T., Gordon, T.R., and Viljoen, A. (2009). Evolutionary relationships among the *Fusarium oxysporum* f. sp. *ubense* vegetative compatibility groups. *Applied and Environmental Microbiology*, 75(14):4770–4781.
- Fourie, G., Steenkamp, E.T., Ploetz, R.C., Gordon, T.R., and Viljoen, A. (2011). Current status of the taxonomic position of *Fusarium oxysporum* formae specialis *ubense* within the *Fusarium oxysporum* complex. *Infection, Genetics and Evolution*, 11(3):533–542.
- Fraser-Smith, S., Czislawski, E., Meldrum, R.A., Zander, M., O'Neill, W., Balali, G.R., and Aitken, E.A.B. (2014). Sequence variation in the putative effector gene SIX8 facilitates molecular differentiation of *Fusarium oxysporum* f. sp. *ubense*. *Plant Pathology*, 63(5):1044–1052.
- Gan, P., Ikeda, K., Irieda, H., Narusaka, M., O'Connell, R.J., Narusaka, Y., Takano, Y., Kubo, Y., and Shirasu, K. (2013). Comparative genomic and transcriptomic analyses reveal the hemibiotrophic stage shift of *Colletotrichum* fungi. *New Phytologist*, 197(4): 1236–1249.
- García-Bastidas, F., Ordóñez, N., Konkol, J., Al-Qasim, M., Naser, Z., Abdelwali, M., Salem, N., Waalwijk, C., Ploetz, R.C., and Kema, G.H.J. (2014). First report of *Fusarium oxysporum* f. sp. *ubense* tropical race 4 associated with Panama disease of banana outside Southeast Asia. *Plant Disease*, 98: 694.
- Gawehns, F., Houterman, P.M., Ichou, F.A., Michielse, C.B., Hijdra, M., Cornelissen, B.J., Rep, M., and Takken, F.L. (2014). The *Fusarium oxysporum* effector SIX6 contributes to virulence and suppresses I-2 mediated cell death. *Molecular Plant-Microbe Interactions*, 27:336-348.
- Gerlach, W. and Nirenberg, H. (1982). *The Genus Fusarium; A pictorial atlas*. Paul Parey, Berlin.
- Ghag, S.B., Shekhawat, U.K.S., and Ganapathi, T.R. (2015). *Fusarium* wilt of banana: biology, epidemiology and management. *International Journal of Pest Management*, 61: 250-263.
- Gibriel, H.A.Y., Thomma, B.P.H.J., and Seidl, M.F. (2016). The Age of Effectors: Genome-Based Discovery and Applications. *Phytopathology*, 106(10), 1206–1212.

- Ginzinger, D. G. (2002). Gene quantification using real-time quantitative PCR. *Experimental Hematology*, 30(6), 503–512.
- Giraldo, M.C. and Valent, B. (2013). Filamentous plant pathogen effectors in action. *Nature Reviews Microbiology*, 11(11), 800–814.
- Guo, L., Han, L., Yang, L., Zeng, H., Fan, D., Zhu, Y., and Huang, J. (2014). Genome and Transcriptome Analysis of the Fungal Pathogen *Fusarium oxysporum* f. sp. *cubense* Causing Banana Vascular Wilt Disease. *PLoS ONE*, 9(4), e95543.
- Harringt.C., Thorpe, D.J., and Alfenas, A.C. (2011). Genetic variation in aggressiveness to native and exotic hosts among Brazilian populations of *Ceratocystis fimbriata*. *Phytopathology*, 101(5):555–566.
- Hermanto, C. (2013). Socio-economic impact, research and development and policy making/regulatory of *Fusarium* wilt on banana in Indonesia. Paper presented at the Consultation-Workshop on the Socio- Economic Impacts of *Fusarium* Wilt Disease of Cavendish Banana in the Asia Pacific Region, Waterfront Insular Hotel, Davao City, Philippines, 11-15 November 2013.
- Heslop-Harrison, J. S., and Schwarzacher, T. (2007). Domestication Genomics and the Future for Banana. *Annals of Botany*, 100(5), 1073–1084.
- Hilton, A., Zhang, H., Yu, W., and Shim, W.B. (2017). Identification and characterisation of pathogenic and endophytic fungal species associated with Pokkah Boeng disease of sugarcane. *The Plant Pathology Journal*, 33(3):238–248.
- Hogenhout, S.A., van der Hoorn, R.A., Terauchi, R., and Kamoun, S. (2009). Emerging concepts in effector biology of plant associated organisms. *Molecular Plant Microbe Interactions*, 22: 1157–122.
- Houterman, P.M., Speijer, D., Dekker, H.L., DE Koster, C.G., Cornelissen, B.J.C., and Rep, M. (2007). The mixed xylem sap proteome of *Fusarium oxysporum* infected tomato plants. *Molecular Plant Pathology*, 8(2):215–221.
- Houterman, P.M., Ma, L., van Ooijen, G., de Vroomen, M.J., Cornelissen, B.J.C., Takken, F.L.W. and Rep, M. (2009) The effector protein Avr2 of the xylem-colonizing fungus *Fusarium oxysporum* activates the tomato resistance protein I-2 intracellularly. *The Plant Journal*, 58, 970–978.

- Hwang, S.C. (2013). Unconventional banana breeding through somaclonal variation selection in Taiwan. Retrieved from <http://banananetworks.org/Bapnet/files/2013/02/SCHwang.pdf> on January 3, 2019.
- Inami, K., Yoshioka-Akiyama, C., Morita, Y., Yamasaki, M., Teraoka, T., and Arie, T. (2012). A Genetic Mechanism for Emergence of Races in *Fusarium oxysporum* f. sp. *lycopersici*: Inactivation of Avirulence Gene AVR1 by Transposon Insertion. PLoS ONE, 7(8),
- Jarozuk-Scisel, J., Kurek, E., Winiarczyk, K., Baturo, A. and Lukanowski, A. (2008). Colonization of root tissues and protection against *Fusarium* wilt of rye (*Secale cereale*) by non pathogenic rhizosphere strains of *Fusarium culmorum*. Biological Control 45(3), 297–307.
- Jeger, M. J., Waller, J. M., Johanson, A., and Gowen, S. R. (1996). Monitoring in banana pest management. Crop Protection, 15(4), 391–397.
- Jelinski, N. A., Broz, K., Jonkers, W., Ma, L. J., and Kistler, H. C. (2017). Effector gene suites in some soil isolates of *Fusarium oxysporum* are not sufficient predictors of vascular wilt in tomato. Phytopathology, 107, 842–851.
- Jenkins, S., Taylor, A., Jackson, A.C, Armitage, A.D., Bates, H.J, Mead, A., Harrison R.J and Clarkson J.P. (2021) Identification and Expression of Secreted In Xylem Pathogenicity Genes in *Fusarium oxysporum* f. sp. *lisi*. Frontiers in Microbiology. 12:593140
- Kamoun, S. (2007). Groovy times: Filamentous pathogen effectors revealed. Current Opinion in Plant Biology, 10:358-365.
- Kashiwa, T., Suzuki, T., Sato, A., Akai, K., Teraoka, T., Komatsu, K., and Arie, T. (2016). A new biotype of *Fusarium oxysporum* f. sp. *lycopersici* race 2 emerged by a transposon-driven mutation of avirulence gene AVR1. FEMS Microbiology Letters, 363.
- Kazan, K. and Gardiner, D.M. (2018). *Fusarium* crown rot caused by *Fusarium pseudograminearum* in cereal crops: recent progress and future prospects. Molecular Plant Pathology, 19(7): 1547–1562.
- Kleemann, J., Rincon-Rivera, L.J., Takahara, H., Neumann, U., Ver Loren van Themaat, E., van der Does, H.C., Hacquard, S., Stüber, K., Will, I., Schmalenbach, W., Schmelzer, E., and O'Connell, R.J. (2012). Sequential delivery of host-induced virulence effectors by appressoria and intracellular hyphae of the phytopathogen *Colletotrichum higginsianum*. PLoS Pathogen, 8: e1002643.
- Kumar, S., Stecher, G., and Tamura, K. (2016). MEGA7: Molecular Evolutionary Genetics Analysis Version 7.0 for bigger datasets. Molecular Biology and Evolution, 33(7):1870–1874.

- Lanubile, A., Ellis, M.L., Marocco, A., and Munkvold, G.P. (2016). Association of effector SIX6 with vascular wilt symptoms caused by *Fusarium oxysporum* on soybean. *Phytopathology*, 106:1404-1412.
- Laurence, M.H., Summerell, B.A. and Liew, E.C.Y. (2015) *Fusarium oxysporum* f. sp. *canariensis*: evidence for horizontal gene transfer of putative pathogenicity genes. *Plant Pathology*, 64, 1068–1075.
- Leong, S.K., Latiffah, Z., and Baharuddin, S. (2010). Genetic diversity of *Fusarium oxysporum* f. sp. *cubense* isolates from Malaysia. *African Journal of Microbiology Research*, 4(11):1026–1037.
- Leslie, J.F., and Summerell, B.A. (2006). *The Fusarium Laboratory Workshop Manual*. Blackwell Publishing, USA.
- Leslie, J.F. (1993). Vegetative compatibility in fungi. *Annual Reviews of Phytopathology*, 31: 127-151.
- Li, C.Y., Mostert, G., Zuo, C.W., Beukes, I., Yang, Q.S., Sheng, O., Kuang R.B., Wei, Y.R., Hu, C.H., Rose, L., Karangwa, P., Yang, J., Deng, G.M., Liu, S.W., Gao, J., Viljoen, A., and Yi, G.J. (2013). Diversity and distribution of the banana wilt pathogen *Fusarium oxysporum* f.sp. *cubense* in China. *Fungal Genomics and Biology*, 3: 2.
- Li, C., Shen, S., Zuo, C., Sun, Q., Ye, Q., Yi, G., and Huang, B. (2011). The use of GFP transformed isolates to study the infection of banana with *Fusarium oxysporum* f. sp. *cubense* race 4. *European Journal of Plant Pathology*, 131: 327-340.
- Li, E., Wang, G., Xiao, J., Ling, J., Yang, Y., and Xie, B. (2016). A SIX1 Homolog in *Fusarium oxysporum* f. sp. *conglutinans* Is Required for Full Virulence on Cabbage. *PLOS ONE*, 11(3), e0152273.
- Lievens, B., Houterman, P.M., and Rep, M. (2009). Effector gene screening allows unambiguous identification of *Fusarium oxysporum* f. sp. *lycopersici* races and discrimination from other *formae speciales*. *FEMS Microbiology Letters*, 300(2):201-215.
- Livak, K. J., and Schmittgen, T. D. (2001). Analysis of Relative Gene Expression Data Using Real-Time Quantitative PCR and the 2- $\Delta\Delta$ CT Method. *Methods*, 25(4), 402–408.
- Lo Presti, L., Lanver, D., Schweizer, G., Tanaka, S., Liang, L., Tollot, M., Zuccaro, A., Reissmann, S., and Kahmann, R. (2015). Fungal effectors and plant susceptibility. *Annual Review of Plant Biology* 66: 513– 545.

- Lucas, J. A. (1998). Plant Pathogens and Plant Diseases. *Plant Pathology*, 47(4), 542–542.
- Ma, L.J., van der Does, H.C., Borkovich, K.A., Coleman, J.J., Daboussi, M.J., Di Pietro, A., Dufresne, M., Freitag, M., Grabherr, M., and Henrissat, B. (2010). Comparative genomics reveals mobile pathogenicity chromosomes in *Fusarium*. *Nature*. 464(7287):367–373.
- Ma, L., Houterman, P.M., Gawehns, F., Cao, L., Sillo, F., Richter, H., Clavijo-Ortiz, M.J., Schmidt, S.M., Boeren, S., Vervoort, J., Cornelissen, B.J., Rep, M., and Takken, F.L. (2015). The AVR2-SIX5 gene pair is required to activate I-2-mediated immunity in tomato. *New Phytologist*, 208(2): 507-518.
- Maldonado, B.L.D., Villarruel, O.J.L., Calderon, O.M.A., and Sanchez Espinosa, A.C. (2018). Secreted in xylem (SIX) genes in *Fusarium oxysporum* f. sp. *ubense* and their potential acquisition by horizontal transfer. *Advances in Biotechnology and Microbiology*, 10: 555779.
- Maryani, N., Lombard, L. Poerba, Y.S., Subandiyah, S., Crous, P.W., and Kema, G.H.J. (2019). Phylogeny and genetic diversity of the banana *Fusarium* wilt pathogen *Fusarium oxysporum* f. sp. *ubense* in the Indonesian centre of origin. *Studies in Mycology*, 92: 155-194.
- Maymon, M., Shapitz, U., Harel, Y., Levy, E., Elkind, G., Teverovsky, E., Gofman, R., Haberman, A., Zemorski, R., Nadav, E., Levi, Y., Or, G., Galpaz, N., Israeli, Y., and Freeman, S. (2018). First report of *Fusarium oxysporum* f. sp. *ubense* Tropical Race 4 causing *Fusarium* wilt of Cavendish bananas in Israel. *Plant Disease*, 59: 348.
- McCarthy, D.J., and Smyth, G.K. (2009). Testing significance relative to a fold-change threshold is a TREAT. *Bioinformatics*, 25(6),765–771.
- Meldrum, R.A., Fraser-Smith, S., Tran-Nguyen, L.T.T., Daly, A.M., and Aitken, E.A.B. (2012). Presence of putative pathogenicity genes in isolates of *Fusarium oxysporum* f. sp. *ubense* from Australia. *Australasian Plant Pathology*, 41(5):551–557.
- Michielse, C.B. and Rep, M. (2009). Pathogen profile update: *Fusarium oxysporum*. *Molecular Plant Pathology*, 10, 311–324.
- MOA (2019). Booklet statistik tanaman: Sub-sektor tanaman makanan. Malaysia: Statistical Unit, Planning, Information Technology and Communication Division, Department of Agriculture.
- Mohamad Roff, M.N., Tengku Abdul Malik, T.M., and Sharif, H. (2012). "Challenges to banana production in Malaysia: A threat to food security," *The Planter Kuala Lumpur* 88(1030):20-30.

- Molina, A.B., Williams, R.C., Hermanto, C., Suwanda, B., Komolong, and Kokoa, P. (2010). Final report: Mitigating the threat of banana *Fusarium* wilt: Understanding the agroecological distribution of pathogenic forms and developing disease management strategies. Cranberra, Australia: ACIAR Publication.
- Moore, N.Y., Pegg, K.G., Allen, R.N., and Irwin, J.A.G. (1993). Vegetative compatibility and distribution of *Fusarium oxysporum* f. sp. *ubense* in Australia. Australian Journal of Experimental Agriculture. 33(6): 797–802.
- Mostert, D., Molina, A.B., Daniells, J., Fourie, G., Hermanto, C., Chao, C.P., Fabregar, E., Sinohin, V.G., Masdek, N., and Thangavelu, R. (2017). The distribution and host range of the banana *Fusarium* wilt fungus, *Fusarium oxysporum* f. sp. *ubense*, in Asia. PLoS One. 12(7): e0181630.
- Neely, D., and Beckman, C. H. (1990). The Nature of Wilt Diseases in Plants. Mycologia, 82(5).
- Nik Masdek, N.H. (1991). Response of banana cultivars to *Fusarium oxysporum* f. sp. *ubense*. Proceedings of the National IRPA seminar (Agriculture Sector) Vol 1. Edited by Y.H. Ho et. al.
- Nik Rozana, N.M.M., Suntharalingam, C., and Othman, M.F. (2017). Competitiveness of Malaysia's Fruits in the Global Market: Revealed Comparative Advantage Analysis. Malaysian Journal of Mathematical Sciences 11(S) February: 143–157.
- Niu, X., Zhao, X., Ling, K.-S., Levi, A., Sun, Y., and Fan, M. (2016). The FonSIX6 gene acts as an avirulence effector in the *Fusarium oxysporum* f. sp. *niveum* - watermelon pathosystem. Scientific Reports, 6(1).
- Nitani, T., Akai, K., Hasegawa, R., Ayukawa, Y., Garcia, R.R., Chitose, A., Komatsu, K., Kikuno, H., Natsuaki, K.T., and Arie, T. (2018). Panama disease of banana occurred in Miyakojima Island, Okinawa, Japan. Journal of General Plant Pathology, 84: 165-168
- O'Connell, R. J., Thon, M. R., Hacquard, S., Amyotte, S. G., Kleemann, J., Torres, M. F., and Alkan, N. (2012). Lifestyle transitions in plant pathogenic *Colletotrichum* fungi deciphered by genome and transcriptome analyses. Nature Genetics, 44(9), 1060–1065.
- O'Neill, W.T., Pattison, A.B., Daniells, J.W., Hermanto, C., and Molina, A.B. (2011). Vegetative compatibility group analysis of Indonesian *Fusarium oxysporum* f. sp. *ubense* isolates. Acta Horticulturae, 897: 345-351.

- Ordoñez, N., García-Bastidas, F., Laghari, H.B., Akkary, M.Y., Harfouche, E.N., al Awar, B.N., and Kema, G.H.J. (2016). First report of *Fusarium oxysporum* f. sp. *cubense* Tropical Race 4 causing Panama disease in Cavendish bananas in Pakistan and Lebanon. *Plant Disease*, 100: 209.
- Ortiz, R., and Swennen, R. (2014). From crossbreeding to biotechnology facilitated improvement of banana and plantain. *Biotechnology Advances*, 32(1), 158-169.
- Passos, M. A. N., de Cruz, V. O., Emediato, F. L., de Teixeira, C. C., Azevedo, V. C. R., Brasileiro, A. C. M., and Miller, R. N. (2013). Analysis of the leaf transcriptome of *Musa acuminata* during interaction with *Mycosphaerella musicola*: gene assembly, annotation and marker development. *BMC Genomics*, 14(1), 78
- Pauziah, M., Suhana, O., Rozeita, L., and Maimun, T. (2017). Status of *Fusarium* wilt disease in Malaysia. In: Sinohin, V.G., Molina, A., Johnson, V., & Ganjun, Y. (Eds.) *Proceedings of the 10th banana Asia Pacific network steering committee meeting*. Guangzhou, China: INIBAP.
- Pegg, K.G., Moore, N.Y., and Sorensen, S. (1994). Variability in populations of *Fusarium oxysporum* f. sp. *cubense* from the Asia Pacific region. In: Jones, D.R. (ed.) *The Improvement and Testing of Musa: A Global Partnership*, pp. 70-82. Montpellier: INIBAP.
- Peng, K.C., Tai, C.F., and Chao, C.P. (2013). Socio-economic impact of *Fusarium* wilt on Cavendish banana in Taiwan. Paper presented at the Consultation-Workshop on the Socio-Economic Impacts of *Fusarium* Wilt Disease of Cavendish Banana in the Asia-Pacific Region, Waterfront Insular Hotel, Davao City, Philippines, 11-15 November 2013.
- Peraza-Echeverria, S., J. L. Dale, R. M. Harding, M. K. Smith and C. Collet. (2008). Characterization of disease resistance gene candidates of the nucleotide binding site (NBS) type from banana and correlation of a transcriptional polymorphism with resistance to *Fusarium oxysporum* f. sp. *cubense* race 4. *Mol. Breed.* 22(4): 565-579.
- Ploetz, R.C. (1990). *Fusarium* wilt of banana. APS Press, St. Paul, Minnesota
- Ploetz, R.C. (1994). *Fusarium* wilt and IMTP Phase II. The improvement and testing of Musa: a global partnership, pp 57-69. INIBAP.
- Ploetz, R.C. (2005). Panama disease, an old nemesis rears its ugly head: Part 1 and 2. In: *Plant Health Progress*. DOI: 10.1094/PHP-2005-1221-01-RV.

- Ploetz, R.C. (2006a). Panama disease, an old nemesis rears its ugly head: Part 2. The Cavendish era and beyond. *Plant Health Progress*.
- Ploetz, R.C. (2006b). *Fusarium* wilt of banana is caused by several pathogens referred to as *Fusarium oxysporum* f. sp. *cabense*. *Phytopathology*, 96:653.
- Ploetz, R.C. (2015). *Fusarium* wilt of banana. *Phytopathology*, 105: 1512- 1521.
- Ploetz, R.C., and Pegg, K. (1997). *Fusarium* wilt of banana and Wallace's line: Was the disease originally restricted to his Indo-Malayan region? *Australasian Plant Pathology*, 26: 239-249.
- Ploetz, R.C., and Pegg, K.G. (2000). Fungal diseases of root, corm and pseudostem. 2000. In: Jones DR, editor, *Diseases of banana, abaca and enset*. Wallingford, UK: CAB International; p. 143–172.
- Ploetz, R.C., and Evans, E.A. (2015). Banana diseases and the future of the industry. *Horticulture Review*, 43: 311-351
- Poon, N. K., Teo, C. H., and Othman, R. Y. (2019). Differential gene expression analysis of Secreted in Xylem (SIX) genes from *Fusarium oxysporum* f. sp. *cabense* tropical race 4 in *Musa acuminata* cv. Berangan and potential application for early detection of infection. *Journal of General Plant Pathology*, 86: 13–23.
- Porter, L.D., Pasche, J.S., Chen, W., and Harveson, R.M. (2015). Isolation, identification, storage, pathogenicity tests, hosts and geographic range of *Fusarium solani* f. sp. *pisi* causing *Fusarium* root rot of pea. *Plant Health Progress*, 16: 136-145.
- ProMusa (2018). Mediawatch: TR4 present in the UK. Retrieved from <http://www.promusa.org/blogpost580-TR4-present-in-the-UK>, assessed on December 31, 2018.
- Puhalla, J.E. (1985). Classification of strains of *Fusarium oxysporum* on the basis of vegetative compatibility. *Canadian Journal of Botany*, 63: 179-183.
- Purwati, R.D., Hidayah, N., Sudindro and Sudarsono, (2008). Inoculation methods and conidial densities of *Fusarium oxysporum* f. sp. *cabense* in Abaca. *Hayati Journal of Biosciences*, 15:1–7.
- Rep, M., Van Der Does, H.C., Meijer, M., Van Wijk, R., Houterman, P.M., Dekker, H.L., De Koster, C.G., and Cornelissen, B.J.C. (2004). A small, cysteine-rich protein secreted by *Fusarium oxysporum* during colonization of xylem vessels is required for I-3-mediated resistance in tomato. *Molecular Microbiology*, 53(5):1373–1383.

- Rep, M., Meijer, M., Houterman, P.M., Van Der Does, H.C. and Cornelissen, B.J.C. (2005). *Fusarium oxysporum* evades I-3-mediated resistance without altering the matching avirulence gene. *Molecular Plant-Microbe Interactions*, 18, 15–23.
- Rocha, L.O., Laurence, M.H., Ludowici, V.A., Puno, V.I., Lim, C.C., Tesoriero, L.A., Summerell, B.A. and Liew, E.C.Y. (2016). Putative effector genes detected in *Fusarium oxysporum* from natural ecosystems of Australia. *Plant Pathology*. 65, 914–929.
- Rovenich, H., Boshoven, J.C. and Thomma, B.P.H.J. (2014). Filamentous pathogen effector functions: of pathogens, hosts and microbiomes. *Current Opinion in Plant Biology*, 20, 96–103.
- Scheerer, L., Pemsil, D., Dita, M., Perez Vicente, L., and Staver, C. (2018). A quantified approach to projecting losses caused by *Fusarium* wilt Tropical race 4. *Acta Horticulturae*, 1196: 211-2018.
- Schmidt, S.M., Houterman, P.M., Schreiver, I., Ma, L., Amyotte, S.G., Chellappan, B., Boeren, S., Takken, F.L.W., and Rep, M. (2013). MITEs in the promoters of effector genes allow prediction of novel virulence genes in *Fusarium oxysporum*. *BMC Genomics*, 14(1):119.
- Shekhawat, U.K. S., and Ganapathi, T.R. (2013). *MusaWRKY71* Overexpression in Banana Plants Leads to Altered Abiotic and Biotic Stress Responses. *PLoS ONE*, 8(10), e75506.
- Siamak, S.B., and Zheng, S. (2018). Banana *Fusarium* wilt (*Fusarium oxysporum* f. sp. *cubense*) control and resistance, in the context of developing wilt-resistant bananas within sustainable production systems. *Horticultural Plant Journal*, 4: 208-218.
- Simbaqueba, J., Catanzariti, A.M., González, C., and Jones, D.A. (2018). Evidence for horizontal gene transfer and separation of effector recognition from effector function revealed by analysis of effector genes shared between cape gooseberry- and tomato-infecting *formae speciales* of *Fusarium oxysporum*. *Molecular Plant Pathology*, 19, 2302–2318.
- Srinivasan, K., Spadaro, D., Poli, A., Gilardi, G., Gullino, M.L., and Garibaldi, A. (2012). Genetic diversity and pathogenicity of *Fusarium oxysporum* isolated from wilted rocket plants in Italy. *Phytoparasitica*. 40(2):157–170.
- Stergiopoulos, I. and de Wit, P.J. (2009). Fungal effector proteins. *Annual Review of Phytopathology*. 47:233-263.
- Stergiopoulos, I., Kourmpetis, Y.A., Slot, J.C., Bakker, F.T., De Wit, P.J.G.M. and Rokas, A. (2012). In silico characterization and molecular evolutionary analysis of a novel superfamily of fungal effector proteins. *Molecular Biology and Evolution*, 29, 3371–3384.

- Stover, R.H. (1962). *Fusarium* wilt (Panama disease) of bananas and other *Musa* species. *Phytopathology Paper No. 4. CMI, Kew, Surrey, England. 177pp.*
- Sun, E.J., Su, H.J. and Ko, W.H. (1978). Identification of *Fusarium oxysporum* on the basis of vegetative compatibility. *Canadian Journal of Botanical, 63:179-191.*
- Sutherland, R., Viljoen, A., Myburg, A.A., and van der Bergh, N. (2013). Pathogenicity associated genes in *Fusarium oxysporum* f. sp. *cubense*. *South African Journal of Science, 109: 1-10.*
- Swarupa, V., Ravishankar, K. V., and Rekha, A. (2014). Plant defense response against *Fusarium oxysporum* and strategies to develop tolerant genotypes in banana. *Planta, 239(4), 735–751.*
- Synder, W.C., and Hansen, H.N. (1940). Culture methods in relation to *Fusarium* identification. *Phytopathology, 29: 827.*
- Talas, F., Wurschum, T., Reif, J.C., Parzies, H.K. and Miedaner, T. (2012). Association of single nucleotide polymorphic sites in candidate genes with aggressiveness and deoxynivalenol production in *Fusarium graminearum* causing wheat head blight. *BMC Genetics, 13: 14.*
- Tamietti, G. and Valentino, D. (2006). Soil solarization as an ecological method for the control of *Fusarium* wilt of melon in Italy. *Crop Protection, 25: 389- 397.*
- Tamura, K., Peterson, D., Peterson, N., Stecher, G., Nei, M., and Kumar, S. (2011). MEGA5: molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Molecular Biology and Evolution, 28(10): 2731–2739.*
- Tan, J., Tao, Q., Niu, H., Zhang, Z., Li, D., and Gong, Z. (2015). A novel allele of *monoecious* (*m*) locus is responsible for elongated fruit shape and perfect flowers in cucumber (*Cucumis sativus* L). *Theoretical and Applied Genetics, 128(12): 2483–2493.*
- Taylor, S., Wakem, M., Dijkman, G., Alsarraj, M., and Nguyen, M. (2010). A practical approach to RT-qPCR publishing data according to the MIQE guidelines. *Methods 50: S1–S5.*
- Taylor, A., Vagany, V., Jackson, A.C., Harrison, R.J., Rainoni, A., and Clarkson, J.P. (2016). Identification of pathogenicity-related genes in *Fusarium oxysporum* f. sp. *cepae*. *Molecular Plant Pathology, 17(7):1032–1047.*

- Thangavelu, R., Kumar, K.M., Devi, P.G., and Mustaffa, M.M. (2012). Genetic diversity of *Fusarium oxysporum* f. sp. *ubense* isolates (Foc) of India by inter simple sequence repeats (ISSR) analysis. *Molecular Biotechnology*, 51(3):203–211.
- Thatcher, L.F., Gardiner, D.M., Kazan, K., and Manners, J.M. (2012). A highly conserved effector in *Fusarium oxysporum* is required for full virulence on *Arabidopsis*. *Molecular Plant-Microbe Interactions*, 25: 180-190.
- Thompson, A. and Johnston, A. (1953). A host list of plant diseases in Malaya. In: *Mycological Papers*. Commonwealth Mycological Institute, CMI, Kew, Surrey, England. vol. 52. p. 38.
- Tumin, S.A., and Shaharudin, A.A.A. (2019). Banana: The world's most popular fruit. Retrieved from: [@-http://www.krinstitute.org/What_We_Are_Reading](http://www.krinstitute.org/What_We_Are_Reading) @-Banana;_The_Worlds_Most_Popular_Fruit.aspx on July 31, 2019.
- UNCTAD INFOCOMM (2016). *Banana*. New York and Geneva: United Nations Conference on Trade and Development pp. 1-21.
- Van Dam, P., Fokkens, L., Schmidt, S.M., Linmans, J.H.J., Kistler, H.C., Ma, L.J., and Rep, M. (2016). Effector profiles distinguish formae speciales of *Fusarium oxysporum*. *Environmental Microbiology*, 18(11): 4087–4102.
- Van Den Berg, N., Berger, D. K., Hein, I., Birch, P. R. J., Wingfield, M. J., and Viljoen, A. (2007). Tolerance in banana to *Fusarium* wilt is associated with early up-regulation of cell wall-strengthening genes in the roots. *Molecular Plant Pathology*, 8(3), 333–341.
- van der Does, H.C., and Rep, M. (2007). Virulence genes and the evolution of host specificity in plant-pathogenic fungi. *Molecular Plant-Microbe Interactions*, 20: 1175-1182.
- van der Does, H.C., Duyvesteyn, R.G.E., Goltstein, P.M., van Schie, C.C.N., Manders, E.M.M., Cornelissen, B.J.C. and Rep, M. (2008). Expression of effector gene SIX1 of *Fusarium oxysporum* requires living plant cells. *Fungal Genetics and Biology*, 45(9), 1257–1264.
- Vandesompele, J., De Preter, K., and Pattyn, F. (2002). Accurate normalization of real-time quantitative RT-PCR data by geometric averaging of multiple internal control genes. *Genome Biology*, 3(7): RESEARCH0034.
- Van de Wouw, A. P., and Howlett, B. J. (2010). Fungal pathogenicity genes in the age of “omics.” *Molecular Plant Pathology*, 12(5), 507–514.
- Vézina, A. (2018). *Fusarium* wilt of banana. Montpellier, France: ProMusa Biodiversity International.

- Viljoen, A. (2002). The status of *Fusarium* wilt (Panama disease) of banana in South Africa. *South African Journal of Science*, 98: 1-4.
- Waller, J.M., Lenne, J.M., and Waller, S.J. (2002). *Plant Pathologist's Pocketbook*. 3rd edn. CABI Publishing, New York. pp. 27.
- Warman, N.M., and Aitken, E.A.B. (2018). The movement of *Fusarium oxysporum* f. sp. *cubense* (sub-tropical race 4) in susceptible cultivars of Banana. *Frontiers in Plant Science*, 9:1748.
- Waite, B.H., Stover, R.H. (1960). Studies on *Fusarium* wilt of bananas. VI. Variability and the cultivars concept in *Fusarium oxysporum* f. sp. *cubense*. *Canadian Journal of Botany*, 38: 985–994.
- Whetzel, H. H. (1929). The terminology of plant pathology. *Proceedings of the International Congress of Plant Sciences*, Ithaca, NY, 1926:1204-1215.
- Wibowo, A., Santosa, A.T., Subandiyah, S., Hermanto, C., and Taylor, M.F.P. (2013). Control of *Fusarium* wilt of banana by using *Trichoderma harzianum* and resistant banana cultivars. *Acta Horticulturae*, 975: 173 177.
- Widinugraheni, S., Niño-Sánchez, J., van der Does, H.C., van Dam, P., García Bastidas, F.A., Subandiyah, S., Meijer, H.J.G., Kistler, H.C., Kema, G.H.J., and Rep, M. (2018). A SIX1 homolog in *Fusarium oxysporum* f. sp. *cubense* tropical race 4 contributes to virulence towards Cavendish banana. *PLoS ONE*, 13: e0205896.
- Williams, A.H., Sharma, M., Thatcher, L.F., Azam, S., Hane, J.K., Sperschneider, J., Kidd, B.N., Anderson, J.P., Ghosh, R., Garg, G., Lichtenzweig, J., Kistler, H.C., Shea, T., Young, S., Buck, S.-A.G., Kamphuis, L.G., Saxena, R., Pande, S., Ma, L.-J., Varshney, R.K. and Singh, K.B. (2016) Comparative genomics and prediction of conditionally dispensable sequences in legume-infecting *Fusarium oxysporum* formae speciales facilitates identification of candidate effectors. *BMC Genom.* 17, 191.
- Win, J., Krasileva, K. V., Kamoun, S., Shirasu, K., Staskawicz, B. J., and Banfield, M. J. (2012). Sequence Divergent RXLR Effectors Share a Structural Fold Conserved across Plant Pathogenic Oomycete Species. *PLoS Pathogens*, 8(1), e1002400.
- Yaakob, D., K.G., Pegg and Siti Hawa, J. (1994). *Fusarium* wilt of banana: Cultivar susceptibility and characterisation of isolates. 4th International Conference on Plant Protection in the Tropics.
- Yadeta, K.A., and Thomma, B.P.H.J. (2013). The xylem as battle ground for plant hosts and vascular wilt pathogens. *Frontiers in Plant Science*, 4: 97.

Zhang, Y., and Ma, L.J. (2017). Deciphering Pathogenicity of *Fusarium oxysporum* From a Phylogenomics Perspective. *Advances in Genetics*, 179–209.

Zheng, F., García-Bastidas, F.A., Li, X., Zeng, L., Bai, T., Xu, S., Yin, K., Li, H., Fu, G., Yu, Y., Yang, L., Nguyen, H.C., Douangboupha, B., Khaing, A.A., Drenth, A., Seidi, M.F., Meijer, H.J.G., and Kema, G.H.J. (2018). New geographical insights of the latest expansion of *Fusarium oxysporum* f. sp. *ubense* tropical race 4 into the Greater Mekong subregion. *Frontiers in Plant Science*, 9: 457.



BIODATA OF STUDENT

Suhanna binti Ahmad was born on 28th September 1985 in Alor Setar, Kedah. She completed her formal education with Sijil Pelajaran Malaysia in Sekolah Menengah Kebangsaan Convent Kajang, Selangor. In 2003, she pursued her study in Science Matriculation program at Kolej Mara Seremban. A year after that, she continued her undergraduate degree in Bachelor of Applied Science (Agrobiology) at Universiti Sains Malaysia (USM) and graduated in November 2007. She was employed as a research officer in the Pest and Disease Management department at MARDI from 2008 until present. She enrolled as a Master candidate in the field of Plant Pathology in 2017 with her study fully funded by MARDI.

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