



UNIVERSITI PUTRA MALAYSIA

***PHYSICOCHEMICAL, MICROBIOLOGICAL AND BIOLOGICAL
PROPERTIES OF FERMENTED KEPAYANG (*Pangium edule Reinw.*)
SEEDS***

NUR HAFIZAH BINTI MUSTAFFER

FSTM 2021 9



UPM
UNIVERSITI PUTRA MALAYSIA
BERILMU BERBAKTI

**PHYSICOCHEMICAL, MICROBIOLOGICAL AND BIOLOGICAL
PROPERTIES OF FERMENTED KEPAYANG (*Pangium edule* Reinw.) SEEDS**

By

NUR HAFIZAH BINTI MUSTAFFER

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia in
Fulfilment of the Requirement for the Degree of Master of Science**

July 2020

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

**PHYSICOCHEMICAL, MICROBIOLOGICAL AND BIOLOGICAL
PROPERTIES OF FERMENTED KEPAYANG (*Pangium edule* Reinw.) SEEDS**

By

NUR HAFIZAH BINTI MUSTAFFER

July 2020

Chairman : Yaya Rukayadi, PhD
Faculty : Food Science and Technology

Kepayang (*Pangium edule* Reinw.) is one of the indigenous fruits in Malaysia, Indonesia, Papua New Guinea, Vanuatu, and the Philippines. This *P. edule* Reinw. seeds can be developed as a functional food through fermentation. Unfortunately, the properties of fermented *P. edule* Reinw. seeds are not well reported yet. Therefore, the aims of this study were to analyse the physicochemical and microbiological characteristics of *P. edule* Reinw. seeds during fermentation process. Besides that, the objective of this study was to determine the antimicrobial and antioxidant activities of *P. edule* Reinw. seeds extract and to identify the presence of bioactive compounds in raw and fermented *P. edule* Reinw. seeds extract. These *P. edule* Reinw. seeds were obtained from Jerantut, Pahang in July 2018. These *P. edule* Reinw. seeds were undergone the spontaneous fermentation process which covered with charcoal for 40 days. The physicochemical properties were evaluated according to the Association of Analytical Communities (AOAC) methods while the microbiological analysis was determined using microbial selective media. Then, the raw and fermented *P. edule* Reinw. seeds were extracted using ethanol and water. The antimicrobial activity was following the Clinical and Laboratory Standard Institute (CLSI) methods. The antioxidant activity was measured using 2,2-diphenyl-1-dipicrylhydrazyl (DPPH) radical scavenging assay while total phenolic content was measured by following Folin-Ciocalteu methods. Other than that, the bioactive compounds present in extracts were identified using gas chromatography-mass spectrometry (GC-MS) profile and liquid chromatography-mass spectrometry (LC-MS) profile. In this study, the results showed that there were no significant differences in water activity while the pH value was decreased along fermentation days. The TSS was significantly increased from day 0 (11.00) to day 30 (19.50), however, it drastically decreased after 40 days fermentation which is 11.50. The value of lightness (L^*) parameter was decreased from 66.44 on day 0 to 25.28 on day 40. There were no significant differences for a^* and b^* parameter. For proximate analysis, the percentage of ash, crude protein, crude fat, and crude fibre were significantly increased while moisture content and carbohydrate were significantly decreased along the fermentation day. The microbiological analysis showed the reducing number of *B. cereus* from 2.15 Log_{10} CFU/g to not detected on day 40, while there were slightly increase in the number

of TPC, coliform, mould and yeast, *S. aureus*, *P. aeruginosa* and LAB. Interestingly, there was no *Salmonella* spp. and *E. coli* detected along fermentation process. The antimicrobial activity of raw and fermented seeds extract was evaluated against 16 types of pathogenic microorganisms. The disc diffusion assay (DDA) showed that the microorganisms were inhibited by extracts in the range of 6.25 mm and 9.88 mm. The lowest MIC and MBC values were at concentration 6.25 mg/mL and 12.50 mg/mL extract, respectively. Generally, the time-kill curve study showed that *B. cereus*, *K. pneumoniae*, *P. aeruginosa*, *S. aureus*, and *C. albicans* can be killed using raw and fermented *P. edule* Reinw. seeds extract. Besides, the highest DPPH activity was observed in ferment-water extract at concentration 10 mg/mL which about 95.61% while the highest total phenolic content was obtained from the ferment-ethanol extract (173.79 mg GAE/100 mg). The GC-MS analysis revealed the volatile compounds in raw and fermented *P. edule* Reinw. seeds extract while LC-MS analysis was used to identify the non-volatile compounds in both positive and negative ion modes. In conclusion, there were some changes in the physicochemical properties of *P. edule* Reinw. seeds during fermentation, nonetheless, the fermented seeds showed a low level of microbial counts and potentially has the antimicrobial and antioxidant activities. The results of this study might be used as basic information to develop the fermented seeds as a functional food.

Keywords: Antimicrobial activity, antioxidant activity, bioactive compounds, fermented *Pangium edule* Reinw. seed, microbiological analysis, physicochemical properties

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk Ijazah Master Sains

**KARAKTERISTIK FISIKOKIMIA, ANALISA MIKROBIOLOGI DAN
AKTIVITI BIOLOGI BIJI BUAH KEPAYANG (*Pangium edule* Reinw.) YANG
DIPERAM**

Oleh

NUR HAFIZAH BINTI MUSTAFFER

Julai 2020

Pengerusi : Yaya Rukayadi, PhD
Fakulti : Sains dan Teknologi Makanan

Kepayang (*Pangium edule* Reinw.) adalah salah satu daripada buah-buahan asli yang terdapat di Malaysia, Indonesia, Papua New Guinea, Vanuatu dan Filipina. Biji *P. edule* Reinw. ini boleh dikembangkan sebagai makanan fungsional melalui pemeraman. Malangnya, sifat biji *P. edule* Reinw. yang telah diperam ini masih belum diketahui sepenuhnya. Oleh itu, matlamat kajian ini adalah untuk menganalisis ciri-ciri fisikokimia dan mikrobiologi biji buah *P. edule* Reinw. semasa proses pemeraman. Selain itu, objektif kajian ini adalah untuk menentukan aktiviti antimikrobial dan antioksidant bagi ekstrak biji *P. edule* Reinw. dan untuk mengenalpasti sebatian aktif yang terdapat dalam ekstrak biji *P. edule* Reinw. ini. Buah *P. edule* Reinw. ini diperolehi dari Jerantut, Pahang pada Julai 2018. Biji buah *P. edule* Reinw. ini telah menjalani proses pemeraman spontan yang ditutupi dengan arang selama 40 hari. Ciri-ciri fizikokimia telah dinilai menggunakan kaedah Persatuan Komuniti Analitik (AOAC) manakala analisis mikrobiologi telah ditentukan menggunakan media selektif mikroba. Kemudian, biji *P. edule* Reinw. yang mentah dan sudah diperam ini diekstrak menggunakan etanol dan air suling. Aktiviti antimikrobial ditentukan mengikut kaedah Institut Standard Klinikal dan Makmal (CLSI). Kegiatan antioksidan diukur dengan menggunakan ujian memerangkap radikal DPPH manakala jumlah kandungan fenolik diukur mengikut kaedah Folin-Ciocalteu. Selain itu, sebatian bioaktif yang terdapat dalam ekstrak telah dikenalpasti menggunakan kaedah analisa GC-MS dan LC-MS profil. Dalam kajian ini, hasil menunjukkan bahawa tidak ada perbezaan yang ketara untuk aktiviti air manakala nilai pH menurun sepanjang hari pemeraman. TSS meningkat dengan ketara dari hari 0 (11.00) hingga hari 30 (19.50), tetapi ia menurun secara drastik selepas pemeraman selama 40 hari (11.50). Nilai parameter L^* menurun dari 66.44 pada hari ke 0 hingga 25.28 pada hari ke-40. Tidak ada perbezaan yang ketara untuk parameter a^* dan b^* . Untuk analisis jarak dekat, peratusan abu, protein mentah, lemak mentah, dan serat kasar meningkat dengan ketara manakala kandungan kelembapan dan karbohidrat menurun sepanjang hari pemeraman. Analisis mikrobiologi menunjukkan penurunan jumlah *B. cereus* dari 2.15 Log₁₀ CFU/g kepada tidak ditemui pada hari ke-40, manakala, terdapat sedikit peningkatan dalam jumlah TPC, *P. aeruginosa*, *S. aureus*, LAB, koliform, kulat

dan yis. Menariknya, tiada *E. coli*, dan *Salmonella* spp. ditemui sepanjang proses pemeraman. Kegiatan antimikrobia ekstrak biji yang mentah dan yang diperam telah dinilai terhadap 16 jenis mikroorganisma patogen. Ujian DDA pula menunjukkan bahawa mikroorganisma berjaya direncatkan oleh ekstrak dengan lingkungan antara 6.25 mm dan 9.88 mm. Nilai MIC dan MBC terendah masing-masing berada pada kepekatan 6.25 mg/mL dan 12.50 mg/mL ekstrak. Secara amnya, analisis keluk-masa pembunuhan menunjukkan bahawa, *B. cereus*, *K. pneumoniae*, *P. aeruginosa*, *S. aureus* dan *C. albicans* boleh dibunuh dengan menggunakan ekstrak biji *P. edule* Reinw. yang mentah dan yang telah diperam. Selain itu, aktiviti DPPH tertinggi dikenalpasti dalam ekstrak yang diperam dengan air pada kepekatan 10 mg/mL yang mana kira-kira 95.61% manakala jumlah kandungan fenolik tertinggi diperolehi dari ekstrak biji diperam-etanol (173.79 mg GAE/100 mg). Analisis GC-MS menunjukkan sebatian yang mudah meruap dalam ekstrak biji *P. edule* Reinw. yang mentah dan diperam manakala analisis LC-MS telah digunakan untuk mengenal pasti sebatian tidak mudah menguap dalam mod ion positif dan negatif. Sebagai kesimpulan, terdapat beberapa perubahan sifat fisikokimia biji *P. edule* Reinw. semasa proses pemeraman, walaubagaimanapun, biji yang diperam ini menunjukkan tahap kiraan mikrob yang rendah dan berpotensi mempunyai aktiviti antimikrob dan antioksidan. Hasil kajian ini mungkin dapat digunakan sebagai maklumat asas untuk menjadikan biji yang diperam ini sebagai makanan yang berfungsi.

Kata kunci: Aktiviti antimikroba, aktiviti antioksidan, sebatian bioaktif, pemeraman biji buah *Pangium edule* Reinw., keselamatan mikrobiologi, ciri-ciri fisikokimia,

ACKNOWLEDGEMENT

“In the name of Allah, the most Gracious and the most Merciful”

Thanks to Almighty God for giving me the strength, wisdom, and endurance in health until I am able to pursue and completed my master’s dual degree UPM-KU. Upon success, I would like to express my gratitude toward people who have helped me in all aspects throughout this period. My greatest gratitude was expressed to my supervisor in KU, Dr. Kullanart Tongkhao and in UPM, Assoc. Prof Dr. Yaya Rukayadi for their invaluable guidance and constructive suggestions during this entire study. I am really appreciated their patient and willingness to share knowledge and experience. A million thanks to my co-supervisors, Dr. Kanithaporn Vangnai and Dr. Nurul Shazini Binti Ramli for their endless support and offered their expertise in this research.

Never in my life imagined would be able to conduct my research at two different universities at the same time. Thus, I would like to send special thanks to Mr. Saiful Mohd Maskan who helps me throughout my admission to KU. I have gained so many new beneficial knowledges throughout my research and I would extend sincere thanks to all staff in Microbiology and Biochemistry Laboratory, Department of Food Science and Technology, Faculty of Agro-Industry for their assistance during my laboratory work.

My deepest appreciation goes to my parents, Mustaffer Bin Abdullah and Zubaidah Binti Yusoff, and all my family members for their endless love, encouragement, support and advice. It is a great pleasure to acknowledge my colleagues for helping me and support throughout my study. Last but not least, I would like to thank Yayasan Bank Rakyat and SEARCA for providing me adequate funding and allowances along my master's journey in Universiti Putra Malaysia (UPM), Malaysia and Kasetsart University (KU), Thailand.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Yaya Rukayadi, PhD

Associate Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Chairman)

Nurul Shazini Binti Ramli, PhD

Lecturer
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Member)

Kullanart Tongkhao, PhD

Assistant Professor
Faculty of Agro-Industry
Kasetsart University, Thailand
(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work; quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: Nur Hafizah Binti Mustaffer; GS 49891

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	vii
LIST OF TABLES	xiv
LIST OF FIGURES	xvi
LIST OF ABBREVIATIONS	Xvii
 CHAPTER	
1 INTRODUCTION	
1.1 Background	1
1.2 Problem Statements	2
1.3 Research Objectives	3
 2 LITERATURE REVIEW	
2.1 <i>Pangium edule</i> Reinw.	4
2.1.1 Origin and background of <i>Pangium edule</i> Reinw.	4
2.1.2 Classification of <i>Pangium edule</i> Reinw.	4
2.1.3 Morphology of <i>Pangium edule</i> Reinw.	5
2.1.4 Traditional uses of <i>Pangium edule</i> Reinw.	7
2.1.5 Common product of <i>Pangium edule</i> Reinw.	7
2.2 Fermentation Process	10
2.2.1 Factor affect microbial growth in fermentation process	10
2.2.2 Types of fermentation	12
2.2.3 Fermented foods	13
2.2.4 Fermentation of <i>Pangium edule</i> Reinw.	13
2.3 Physicochemical Properties	14
2.4 Microbiological Analysis	16
2.5 Plant Extraction	17
2.5.1 Method for extraction	17
2.5.2 Solvent for extraction	18
2.6 Biological Activity	19
2.7 Antimicrobial Activity	19
2.7.1 Agar diffusion assay	20
2.7.2 Minimum inhibition concentration (MIC)	20
2.7.3 Minimum bactericidal concentration (MBC) or minimum fungicidal concentration (MFC)	21
2.7.4 Time-kill curve assay	22
2.8 Antioxidants Activity	22
2.8.1 Antioxidant activity of DPPH radical scavenging assay	23
2.8.2 Total phenolic content	24
2.9 Detection of Bioactive Compounds	25
2.9.1 Gas chromatography-mass spectrometry (GC-MS)	25

2.9.2	Liquid chromatography-mass spectrometry (LC-MS)	26
-------	---	----

3 MATERIALS AND METHODS

3.1	<i>Pangium edule</i> Reinw. Seeds Collection	27
3.2	Preparation for Fermentation of <i>Pangium edule</i> Reinw. Seeds	27
3.3	Preparation of Media and Chemical	28
3.3.1	Preparation of microbiological agar/broth	28
3.3.2	Preparation of dimethyl sulfoxide (DMSO)	30
3.3.3	Preparation of chlorhexidine	30
3.3.4	Preparation of phosphate buffer saline	30
3.4	Physicochemical Properties	30
3.4.1	pH value	30
3.4.2	Total soluble solid (TSS)	31
3.4.3	Water activity (a_w)	31
3.4.4	Colour determination	31
3.4.5	Proximate analysis	31
3.4.5.1	Determination of moisture content	32
3.4.5.2	Determination of ash content	32
3.4.5.3	Determination of crude protein content	32
3.4.5.4	Determination of crude fat content	33
3.4.5.5	Determination of crude fibre content	33
3.4.5.6	Determination of carbohydrate content	34
3.5	Microbiological Analysis	34
3.5.1	Sample preparation	35
3.5.2	Total plate count (TPC)	35
3.5.3	Coliform bacteria	35
3.5.4	Mould and yeast count	36
3.5.5	<i>Bacillus cereus</i>	36
3.5.6	<i>Staphylococcus aureus</i>	36
3.5.7	<i>Salmonella</i> spp.	36
3.5.8	<i>Escherichia coli</i>	36
3.5.9	<i>Pseudomonas aeruginosa</i>	36
3.5.10	Lactic acid bacteria (LAB)	37
3.6	Extraction Method	37
3.7	Biological Activity	38
3.7.1	Antimicrobial activity analysis	38
3.7.1.1	Preparation of inoculum and microbial strains	38
3.7.1.2	Preparation of sample concentration	39
3.7.1.3	Disc diffusion assay (DDA)	39
3.7.1.4	Minimum inhibitory concentration (MIC)	40
3.7.1.5	Minimum bactericidal concentration (MBC) or Minimum Fungicidal concentration (MFC)	40
3.7.1.6	Time-kill curve assay	40
3.7.2	Antioxidant activity and total phenolic content	41
3.7.2.1	Antioxidant Activity of DPPH Radical Scavenging Assay	41

	3.7.2.2	Total Phenolic Content	41
3.8		Identification of Bioactive Compounds	42
	3.8.1	Gas chromatography-mass spectrometry (GC-MS)	42
	3.8.2	Liquid chromatography-mass spectrometry (LC-MS)	42
3.9		Statistical Analysis	43
4		RESULTS AND DISCUSSION	
4.1		Physicochemical Properties of <i>Pangium edule</i> Reinw. Seeds during Fermentation	44
	4.1.1	pH value	45
	4.1.2	Total soluble solids (TSS)	45
	4.1.3	Water activity (a_w)	46
	4.1.4	Colour determination	46
	4.1.5	Proximate analysis	47
4.2		Microbiological Analysis of <i>Pangium edule</i> Reinw. Seeds during Fermentation	50
	4.2.1	Total plate count (TPC)	50
	4.2.2	Coliform bacteria	51
	4.2.3	Mould and yeast count	52
	4.2.4	<i>Bacillus cereus</i>	53
	4.2.5	<i>Staphylococcus aureus</i>	54
	4.2.6	<i>Salmonella</i> spp.	55
	4.2.7	<i>Escherichia coli</i>	55
	4.2.8	<i>Pseudomonas aeruginosa</i>	55
	4.2.9	Lactic acid bacteria (LAB)	56
4.3		Yield of <i>Pangium edule</i> Reinw. Seeds Extract	57
4.4		Antimicrobial Activity of <i>Pangium edule</i> Reinw. Seeds Extract	59
	4.4.1	Disc Diffusion Assay (DDA)	59
	4.4.2	Minimum Inhibition Concentration (MIC) and Minimum Bactericidal Concentration (MBC) or Minimum Fungicidal Concentration (MFC)	63
	4.4.3	Time-kill curve analysis	67
	4.4.3.1	Time-kill curve of <i>Pangium edule</i> Reinw. seeds extract on <i>Klebsiella pneumoniae</i> ATCC13773	68
	4.4.3.2	Time-kill curve of <i>Pangium edule</i> Reinw. seeds extract on <i>Pseudomonas aeruginosa</i> ATCC9027	70
	4.4.3.3	Time-kill curve of <i>Pangium edule</i> Reinw. seeds extract on <i>Staphylococcus aureus</i> ATCC29737	72
	4.4.3.4	Time-kill curve of <i>Pangium edule</i> Reinw. seeds extract on <i>Bacillus cereus</i> ATCC33019	74
	4.4.3.5	Time-kill curve of <i>Pangium edule</i> Reinw. seeds extract on <i>Candida albicans</i> ATCC10231	75
4.5		Antioxidant Activities and Total Phenolic Content of <i>Pangium edule</i> Reinw. Seeds Extracts	78

4.5.1	DPPH scavenging activity in <i>Pangium edule</i> Reinw. seeds extract	78
4.5.2	Total phenolic content in <i>Pangium edule</i> Reinw. seeds extract	80
4.6	Identification of Bioactive Compounds in <i>Pangium edule</i> Reinw. Seeds Extracts	81
4.6.1	Gas Chromatography-Mass Spectrometry (GC-MS) based profile of bioactive compounds in <i>Pangium edule</i> Reinw. seeds extract	81
4.6.2	Liquid Chromatography-Mass Spectrometry (LC-MS) based profile of bioactive compounds in <i>Pangium edule</i> Reinw. seeds extract	89
5	SUMMARY, CONCLUSION AND RECOMMENDATION FOR FUTURE RESEARCH	
5.1	Summary	100
5.2	Conclusion	101
5.3	Recommendation for Future Research	102
	REFERENCES	103
	APPENDICES	126
	BIODATA OF STUDENT	131
	LIST OF PUBLICATIONS	132

LIST OF TABLES

Table		Page
2.1	Classification of <i>Pangium edule</i> Reinw. based on rank, scientific and common name	5
3.1	Preparation of media for enumeration of microorganisms according to manufacturer	29
3.2	Pathogenic bacteria and fungi strains.	39
4.1	Physicochemical properties of <i>Pangium edule</i> Reinw. seeds during fermentation process	44
4.2	Proximate analysis of <i>Pangium edule</i> Reinw. seeds during fermentation process	47
4.3	Total and percentage yield of raw and fermented <i>Pangium edule</i> Reinw. seeds extracts.	57
4.4	Inhibition zone of raw and fermented <i>Pangium edule</i> Reinw. seeds extract against pathogenic bacteria and fungi	60
4.5	Minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC) of raw and fermented <i>Pangium edule</i> Reinw. seeds extract against pathogenic bacteria.	64
4.6	Minimum inhibitory concentration (MIC) and minimum fungicidal concentration (MFC) of raw and fermented <i>Pangium edule</i> Reinw. seeds extract against <i>Candida</i> species	66
4.7	Concentration of raw and fermented <i>Pangium edule</i> Reinw. seeds extract used for $\frac{1}{2}\times$ MIC, $1\times$ MIC, $2\times$ MIC and $4\times$ MIC	68
4.8	Summary of time-kill curve analysis for selected pathogenic bacteria and <i>Candida</i> species.	77
4.9	The percentage inhibition of DPPH scavenging activities (%) of raw and fermented <i>Pangium edule</i> Reinw. seeds extracts.	79
4.10	Compounds present in ethanol extract of raw <i>Pangium edule</i> Reinw. seeds identified through GC-MS analysis.	84
4.11	Compounds present in ethanol extract of fermented <i>Pangium edule</i> Reinw. seeds identified through GC-MS analysis.	87
4.12	Tentative identification of compounds presents in raw <i>Pangium edule</i> Reinw. seeds extract identified through LC-MS analysis.	93
4.13	Tentative identification of compounds presents in fermented <i>Pangium edule</i> Reinw. seeds extract identified through LC-MS analysis.	96

LIST OF FIGURES

Figure		Page
2.1	Morphology of <i>Pangium edule</i> Reinw. tree, leaves, flowers, fruits and seeds	6
2.2	Whole plant parts of <i>Pangium edule</i> Reinw. including twig (1), female flower (2), male flower (3) and detail of fruiting twig (4)	7
2.3	Raw and fermented <i>Pangium edule</i> Reinw. Seeds	8
2.4	<i>Pangium edule</i> Reinw. seeds oil and some of the products using this oil	9
2.5	DPPH radical and its reduction by an antioxidant.	24
3.1	The fermentation process of <i>Pangium edule</i> Reinw. Seeds	28
4.1	Total plate count of <i>Pangium edule</i> Reinw. seeds during fermentation process.	51
4.2	Coliform bacteria count of <i>Pangium edule</i> Reinw. seeds during fermentation process.	52
4.3	Mould and yeast count of <i>Pangium edule</i> Reinw. seeds during fermentation process	53
4.4	<i>Bacillus cereus</i> count of <i>Pangium edule</i> Reinw. seeds during fermentation process.	54
4.5	<i>Staphylococcus aureus</i> count of <i>Pangium edule</i> Reinw. seeds during fermentation process.	54
4.6	<i>Pseudomonas aeruginosa</i> count of <i>Pangium edule</i> Reinw. seeds during fermentation process.	56
4.7	Lactic acid bacteria count of <i>Pangium edule</i> Reinw. seeds during fermentation process	57
4.8	Example of the inhibition zone of raw <i>Pangium edule</i> Reinw. seeds extract at concentration 100 mg/mL (10%) on <i>Bacillus megaterium</i> ATCC14581	59
4.9	Time-kill curve plots for <i>K. pneumoniae</i> following exposure to raw <i>Pangium edule</i> Reinw. seeds extracted using ethanol at concentrations 0, 6.25, 12.50, 25.00, 50.00 mg/mL. Values given in the brackets are 0× MIC, 0.5× MIC, 1× MIC, 2× MIC, and 4× MIC, respectively.	69
4.10	Time-kill curve plots for <i>K. pneumoniae</i> following exposure to fermented <i>Pangium edule</i> Reinw. seeds extracted using ethanol at concentrations 0, 6.25, 12.50, 25.00, 50.00 mg/mL. Values given in the brackets are 0× MIC, 0.5× MIC, 1× MIC, 2× MIC, and 4× MIC, respectively.	69
4.11	Time-kill curve plots for <i>P. aeruginosa</i> following exposure to raw <i>Pangium edule</i> Reinw. seeds extracted using ethanol at concentrations 0, 3.13, 6.25, 12.50, 25.00 mg/mL. Values given in the brackets are 0× MIC, 0.5× MIC, 1× MIC, 2× MIC and 4× MIC, respectively.	71
4.12	Time-kill curve plots for <i>P. aeruginosa</i> following exposure to fermented <i>Pangium edule</i> Reinw. seeds extracted using ethanol at concentrations 0, 3.13, 6.25, 12.50, 25.00 mg/mL. Values given in the brackets are 0× MIC, 0.5× MIC, 1× MIC, 2× MIC and 4× MIC, respectively.	71

4.13	Time-kill curve plots for <i>S. aureus</i> following exposure to raw <i>Pangium edule</i> Reinw. seeds extracted using ethanol at concentrations 0, 6.25, 12.50, 25.00, 50.00 mg/mL. Values given in the brackets are 0× MIC, 0.5× MIC, 1× MIC, 2× MIC, and 4× MIC, respectively.	73
4.14	Time-kill curve plots for <i>S. aureus</i> following exposure to fermented <i>Pangium edule</i> Reinw. seeds extracted using ethanol at concentrations 0, 6.25, 12.50, 25.00, 50.00 mg/mL. Values given in the brackets are 0× MIC, 0.5× MIC, 1× MIC, 2× MIC, and 4× MIC, respectively.	73
4.15	Time-kill curve plots for <i>B. cereus</i> following exposure to raw <i>Pangium edule</i> Reinw. seeds extracted using ethanol at concentrations 0, 6.25, 12.50, 25.00, 50.00 mg/mL. Values given in the brackets are 0× MIC, 0.5× MIC, 1× MIC, 2× MIC, and 4× MIC, respectively.	74
4.16	Time-kill curve plots for <i>B. cereus</i> following exposure to fermented <i>Pangium edule</i> Reinw. seeds extracted using ethanol at concentrations 0, 6.25, 12.50, 25.00, 50.00 mg/mL. Values given in the brackets are 0× MIC, 0.5× MIC, 1× MIC, 2× MIC, and 4× MIC, respectively.	75
4.17	Time-kill curve plots for <i>C. albicans</i> following exposure to raw <i>Pangium edule</i> Reinw. seeds extracted using ethanol at concentrations 0, 3.13, 6.25, 12.50, 25.00 mg/mL. Values given in the brackets are 0× MIC, 0.5× MIC, 1× MIC, 2× MIC, and 4× MIC, respectively.	76
4.18	Time-kill curve plots for <i>C. albicans</i> following exposure to fermented <i>Pangium edule</i> Reinw. seeds extracted using ethanol at concentrations 0, 3.13, 6.25, 12.50, 25.00 mg/mL. Values given in the brackets are 0× MIC, 0.5× MIC, 1× MIC, 2× MIC, and 4× MIC, respectively.	76
4.19	Total phenolic content (mg GAE/g extract) of water and ethanol of raw and fermented <i>Pangium edule</i> Reinw. seeds extracts.	80
4.20	Chromatogram of ethanol extract of raw <i>Pangium edule</i> Reinw. seeds by GC-MS.	83
4.21	Chromatogram of ethanol extract of fermented <i>Pangium edule</i> Reinw. seeds by GC-MS.	86
4.22	Chromatogram of compounds in ethanol extract of raw <i>Pangium edule</i> Reinw. seeds in positive ion mode.	91
4.23	Chromatogram of compounds in ethanol extract of raw <i>Pangium edule</i> Reinw. seeds in negative ion mode.	92
4.24	Chromatogram of compounds in ethanol extract of fermented <i>Pangium edule</i> Reinw. seeds in positive ion mode.	94
4.25	Chromatogram of compounds in ethanol extract of fermented <i>Pangium edule</i> Reinw. seeds in negative ion mode.	95

LIST OF ABBREVIATIONS

ANOVA	Analysis of variance
AOAC	Association of Official Analytical Chemists
ASE	Accelerated solvent extraction
ATCC	American Type Culture Collection
ATP	Adenosine triphosphate
a_w	Water Activity
a^*	Red/greenness
b^*	Yellow/blueness
BHA	Butylated hydroxyanisole
BHT	Butylated hydroxytoluene
BSA	<i>Bacillus cereus</i> Selective Agar
CFU	Colony forming unit
CHX	Chlorhexidine
CLSI	Clinical and Laboratory Standards Institute
DDA	Disc Diffusion Assay
DMSO	Dimethylsulfoxide
DPPH	1-diphenyl-2-dipicrylhydrazyl
EMB	Eosin-methylene blue
EUCAST	European Committee on Antimicrobial Susceptibility Testing
FAO	Food and Agriculture Organization
GAE	Gallic acid equivalent
GC-MS	Gas Chromatography – Mass Spectrometry
GRAS	Generally Regarded as Safe
h	Hour
H ₂ SO ₄	Sulphuric acids
HPLC	High-Performance Liquid chromatography
ICMSF	International Commission on Microbiological Specifications for Foods
L^*	Lightness
LAB	Lactic Acid Bacteria
LC-MS	Liquid Chromatography-Mass Spectrometry
MAE	Microwave-assisted extraction
MBC	Minimum Bactericidal Concentration
MF	Molecular Formula
MFC	Minimum Fungicidal Concentration
MHA	Mueller-Hinton agar
MHB	Mueller Hinton broth
MIC	Minimum Inhibitory Concentration
MRS	De Man, Rogosa and Sharpe
MSA	Mannitol Salt Agar
MW	Molecular weight
m/z	Mass/charge ratio
N.A	No activity
NaOH	Sodium hydroxide
NAD	Nicotinamide Adenine Dinucleotide
NIST	National Institute of Standards and Technology
PBS	Phosphate buffered saline

PCA	Plate Count Agar
PDA	Potato Dextrose Agar
PEME	<i>P. edule</i> methyl ester
PROSEA	Plant Resources of South-East Asia
Ppm	Parts per million
Rpm	Revolutions per minute
Rt	Retention time
SD	Standard deviation
SFE	Supercritical fluid extraction
spp.	Species
TPC	Total Plate count
TSS	Total soluble solid
UAE	Ultrasound-assisted extraction
UPM	Universiti Putra Malaysia
XLD	Xylose lysine deoxylate

CHAPTER 1

INTRODUCTION

1.1 Background

The tropical rainforest offers excellent sources of indigenous fruits and vegetables that have a valuable effect and one of them is *kepayang* fruit or the scientific name is *Pangium edule* Reinw. This *P. edule* Reinw. tree is mainly grown in South-East Asia region including Malaysia, Indonesia, Papua New Guinea, Vanuatu, and the Philippines. The fruit is commonly consumed due to its taste and it is widely used as a preservative agent. *P. edule* Reinw. seeds have been used as an alternative approach to preserve and maintain the quality of raw fish, shrimp, and meat when the supply of ice and cooling supplies were limited (Kasim and David, 2013; Heruwati et al., 2009; Heruwati et al., 2007). Traditionally, this fruit can be used to treat the infections and it also has an anthelmintic antiseptic and antibacterial property.

Interestingly, *P. edule* Reinw. seeds are edible in raw after undergoing some treatment such as boiling or soaking in water. These seeds usually called as “*dage*” which can be eaten and commonly being utilized as a vegetable especially in West Java, Indonesia. Other than that, these *P. edule* Reinw. seeds also can undergo the fermentation process and it commercially called as “*keluwak*” (Hoe and Siong, 1999). According to Andarwulan et al. (1999a), fermentation process can make these seeds edible as it can get rid of the cyanide content and at the same time can increase their nutritional value and flavour. Besides that, this process also can change the colour of the seeds from milky white to brown or black and the texture became softer.

In general, fermentation process is one of the common methods that can be used to process and preserved the foods. Fermentation is a metabolic process that produces chemical changes in organic substrates through the action of enzymes. It can convert the complex compound of carbohydrates, such as starch or a sugar, into a simpler compound such as alcohol or acid (Chojnacka, 2006). This fermentation process enhances the freeing up water molecules and indirectly produced edible food products by removing or destroying the undesirable or anti-nutritional factors that might present in the food to ensure their food safety (Steinkraus, 2018). Other than that, fermentation also can improve their taste, flavour and appearance thereby increasing their acceptability (Ejoh, 2007). Based on previous studies, it is confirmed that fermentation process can enhance the nutrition content while increasing the food digestibility (Hasan et al., 2014; Tamang et al., 2016).

Traditionally, the process to ferment the *P. edule* Reinw. seeds were done manually using spontaneous fermentation. Hence, good hygiene practices should be emphasized during fermentation to lower or discards especially from microorganisms that dangerous for health. Previous studies had shown major microorganisms usually involved in this fermented food are bacilli family-like *Bacillus subtilis* and *B. cereus* (Steinkraus, 2018).

Besides, as studied by Gadaga et al. (2007) and Capozzi et al. (2017), *Bacillus cereus*, *Escherichia coli*, *Salmonella* spp., *Staphylococcus aureus* and *Aeromonas* are the most commonly encountered pathogens in African fermented foods. Steinkraus (2018) also stated that *tempe bongkretek*, which is also one of fermented dregs as it is produced from coconut dregs, has been reported to be contaminated by *Pseudomonas* species such as *Pseudomonas aeruginosa* and *P. cocovenenans* if no proper handling during process of fermentation. Other than that, the processing of foods such as fermentation processes can give some effect on the chemistry of these foods. During fermentation process, the microorganism might be present and can change the physical characteristics and biological of the food product. The evaporation and metabolism of the microorganism cause the water activity, a_w might vary (Tsao, 1999) while the pH value may drop along the with fermentation day.

From the best knowledge, there are still scares of studies regarding physicochemical characteristics and microbiological safety of *P. edule* Reinw. seeds during fermentation and these properties are not well reported yet. Furthermore, there are also lacks studies on the determination of bioactivities and the identification of bioactive compounds of the raw and fermented *P. edule* Reinw. seeds extracts. Therefore, this present study was conducted to analyse physicochemical characteristics and microbiological safety of *P. edule* Reinw. seeds during fermentation, to determine the antimicrobial and antioxidant activities of the raw and fermented seed extracts and to identify the bioactive compounds present in the extracts using Liquid Chromatography Mass-Spectrometry (LC-MS) and Gas Chromatography-Mass Spectrometry (GC-MS).

1.2 Problem Statements

There were some problems that were faced in this study. According to Steinkraus (2002), the physicochemical properties of the food product might be changed during the fermentation process. Besides that, fermentation process has also involved the microorganisms which may be caused by contamination (Caplice and Fitzgerald, 1999). Generally, spontaneous fermentation is an uncontrolled process, involving the succession of different types of microorganisms including pathogenic and non-pathogenic bacteria that results in a variety of end products of fermented food. Besides that, fermentation process also involved some chemical reactions which could give effects on food properties and bioactive compounds including the compound that can enhance the antioxidant and antimicrobial activity inside food product (Mehta et al., 2012). Antioxidants are one of the essential components that can be found in plant extracts that have the ability to scavenging free radical inside the biological system (Anjaneyulu et al., 2003). Moreover, the biological products derived from plant sources exhibited a wide range of antimicrobial properties against disease and food spoilage causing microorganisms. which including the pathogenic microbes and candida species. Several recent studies on antimicrobial elements in food products were able to purge the microbes responsible for causing food spoilage, thereby extending the expiry date of food products (Tajkarimi *et al.*, 2010).

In general, there are a lot of studies were conducted on the characteristic of raw and fermented *P. edule* Reinw. seeds. However, no systematic study had been reported

regarding the changes in physicochemical properties and microbiological population of *P. edule* Reinw. seeds during fermentation process. Besides that, the properties of antimicrobial and antioxidant activities had potentially present in water and ethanol extract of raw and fermented *P. edule* Reinw. seeds extracts. Although the effect of these plant extracts against some microorganisms had been studied, the information on the effectiveness of the plant extracts against certain microorganisms was scarce. Thus, the physicochemical properties and microbiological analysis of *P. edule* Reinw. seeds during fermentation need to be analysed. Besides that, the antimicrobial and antioxidant of raw and fermented *P. edule* Reinw. seeds extracts need to be determined. Other than that, the volatile and non-volatile bioactive compounds that contributed to the antimicrobial and antioxidant activities might be present in the raw and fermented *P. edule* Reinw. seeds extracts. These bioactive compounds also can be identified using Gas Chromatography Mass-Spectrometry (GC-MS) and Liquid Chromatography-Mass Spectrometry (LC-MS).

1.3 Research Objectives

The objectives of this study are:

1. To analyse the physicochemical and microbiological characteristics of *Pangium edule* Reinw. seeds during fermentation process.
2. To determine the antimicrobial and antioxidant activities of raw and fermented *Pangium edule* Reinw. seeds extract.
3. To identify the presence of bioactive compounds of raw and fermented *Pangium edule* Reinw. seeds extract.

REFERENCES

- Abdalla, A. E., Darwish, S. M., Ayad, E. H. and El-Hamahmy, R. M. (2007). Egyptian mango by-product 2: Antioxidant and antimicrobial activities of extract and oil from mango seeds kernel. *Food Chemistry*, 103(4): 1141-1152.
- Abee, T., Krockel, L. and Hill, C. (1995). Bacteriocins: Modes of action and potentials in food preservation and control of food poisoning. *International Journal of Food Microbiology*, 28(2): 169-185.
- Abraham, R., Riemersma, R. A., Wood, D., Elton, R., and Oliver, M. F. (1989). Adipose fatty acid composition and the risk of serious ventricular arrhythmias in acute myocardial infarction. *The American Journal of Cardiology*, 63(5): 269-272.
- Abramovič, H., Grobin, B., Poklar Ulrih, N. and Cigić, B. (2018). Relevance and standardization of in vitro antioxidant assays: ABTS, DPPH, and folin–ciocalteu. *Journal of Chemistry*, 2018: 1-9.
- Achi, O. K. (2005). Traditional fermented protein condiments in Nigeria. *African Journal of Biotechnology*, 4 (13): 375-380.
- Afoakwa, E. O., Quao, J., Takrama, J., Budu, A. S. and Saalia, F. K. (2013). Chemical composition and physical quality characteristics of Ghanaian cocoa beans as affected by pulp pre-conditioning and fermentation. *Journal of Food Science and Technology*, 50(6): 1097-105.
- Agushara-Bena, L. M. A., Achmad and Falah, S. (2017). Antifungal activity of *picung* leaf extracts against *Botryodiplodia theobromae* a dieback fungus of *jabon merah* seedling. *Jurnal Silvikultur Tropika*, 6(3): 110-115.
- Ajila, C. M., Naidu, K. A., Bhat, S. G. and Rao, U. P. (2007). Bioactive compounds and antioxidant potential of mango peel extract. *Food Chemistry*, 105(3): 982-988.
- Akindumila, F. and Glatz, B. A. (1998). Growth and oil production of *Apiotrichum curvatum* in tomato juice. *Journal of Food Protection*, 61(11): 1515-1517.
- Akinrele, I. A. (1970). Fermentation studies on maize during the preparation of a traditional African starch-cake food. *Journal of the Science of Food and Agriculture*, 21(12): 619-625.
- Alamin, M. A., Samia, M. A. E. B., Alqurashi, A. M., & Elsheikh, A. S. (2016). Bactericidal activity of *Psidium guajava* leaves against some pathogenic microbes. *IOSR Journal of Dental and Medical Sciences*, 15(3): 61-70.
- Alasalvar, C., Grigor, J. M., Zhang, D., Quantick, P. C. and Shahidi, F. (2001). Comparison of volatiles, phenolics, sugars, antioxidant vitamins, and sensory quality of different coloured carrot varieties. *Journal of Agricultural and Food Chemistry*, 49(3): 1410-1416.
- Al-Farsi, M., Alasalvar, C., Morris, A., Baron, M. and Shahidi, F. (2005). Comparison of antioxidant activity, anthocyanins, carotenoids, and phenolics of three native fresh and sun-dried date (*Phoenix dactylifera* L.) varieties grown in Oman. *Journal of Agricultural and Food Chemistry*, 53(19): 7592-7599.

- Alina (2019). *Basic Principles of LC, HPLC, MS, and MS*. Chemyx Inc. Retrieved September 9, 2019 from <https://www.chemyx.com/support/knowledge-base/applications/basic-principles-hplc-ms-lc-ms/>
- Al-Marzoqi, Ali & Hameed, Imad & Idan, Salah. (2015). Analysis of bioactive chemical components of two medicinal plants (*Coriandrum sativum* and *Melia azedarach*) leaves using gas chromatography-mass spectrometry (GC-MS). *African Journal of Biotechnology*, 14: 2812-2830.
- Allothman, M., Bhat, R. and Karim, A. A. (2009). Antioxidant capacity and phenolic content of selected tropical fruits from Malaysia, extracted with different solvents. *Food Chemistry*, 115(3): 785-788.
- Al-Shammari, L. A., Hassan, W. H. B. and Al-Youssef, H. M. (2012). Chemical composition and antimicrobial activity of the essential oil and lipid content of *Carduus pycnocephalus* L. growing in Saudi Arabia. *Journal of Chemical and Pharmaceutical Research*, 4(2): 1281-1287.
- Alwash, M. S., Ibrahim, N. and Ahmad, W. Y. (2013). Identification and mode of action of antibacterial components from *Melastoma malabathricum* Linn. leaves. *American Journal of Infectious Diseases*, 9(2): 46-58.
- Alzoreky, N. S. and Nakahara, K. (2003). Antibacterial activity of extracts from some edible plants commonly consumed in Asia. *International Journal of Food Microbiology*, 80(3): 223-230.
- Amoa-Awua, W. K., Terlabie, N. N. and Sakyi-Dawson, E. (2006). Screening of 42 *Bacillus* isolates for ability to ferment soybeans into dawadawa. *International Journal of Food Microbiology*, 106(3): 343-347.
- Ananchaipattana, C., Hosotani, Y., Kawasaki, S., Pongsawat, S., MdLatiful, B., Isobe, S. and Inatsu, Y. (2012). Prevalence of foodborne pathogens in retailed foods in Thailand. *Foodborne Pathogens and Disease*, 9(9): 835-840.
- Andarwulan, N., Fardiaz, S., Apriyantono, A., Hariyadi, P. and Shetty, K. (1999a). Mobilization of primary metabolites and phenolics during natural fermentation in seeds of *Pangium edule* Reinw. *Process Biochemistry*, 35(1-2): 197-204.
- Andarwulan, N., Fardiaz, D., Wattimena, G. A. and Shetty, K. (1999b). Antioxidant activity associated with lipid and phenolic mobilization during seeds germination of *Pangium edule* Reinw. *Journal of Agricultural and Food Chemistry*, 47(8): 3158-3163.
- Angelini, R., Argueta, D. A., Piomelli, D., & DiPatrizio, N. V. (2017). Identification of a widespread palmitoylethanolamide contamination in standard laboratory glassware. *Cannabis and Cannabinoid Research*, 2(1): 123-132.
- Anjaneyulu, M., Tirkey, N. and Chopra, K. (2003). Attenuation of cyclosporine-induced renal dysfunction by catechin: possible antioxidant mechanism. *Renal Failure*, 25(5): 691-707.
- Anna, M., Dara, K., Hickey, I., Mercedes, A. G. and Martin, W. (2013). Evaluation of antimicrobial activities of commercial herb and spice extracts against selected food-borne bacteria. *Journal of Food Research*, 2: 431-437.

- AOAC (Association Official Analytical Chemists). (2012). Official methods of analysis, 19th Edition, *Association Official Analytical Chemists*. Washington DC, USA.
- APHA. (1998). Standard Methods for the examination of water and wastewater. 20th Edition, *American Public Health Association, American Water Works Association and Water Environmental Federation*, Washington DC.
- Aqil, F., Ahmad, I. and Mehmood, Z. (2006). Antioxidant and free radical scavenging properties of twelve traditionally used Indian medicinal plants. *Turkish Journal of Biology*, 30(3): 177-183.
- Arikan, S. (2007). Current status of antifungal susceptibility testing methods. *Medical Mycology*, 45(7): 569-587.
- Aryal, S. (2018). Xylose Lysine Deoxycholate (XLD) agar-principle, uses, composition, preparation and colony characteristics. Microbiology Info.com, Retrieved from <https://microbiologyinfo.com/xylose-lysine-deoxycholate-xld-agar-principle-uses-composition-preparation-and-colony-characteristics/>
- Aryal, S. (2019). Mannitol Salt Agar for the isolation of *Staphylococcus aureus*. Microbiology info.com. Retrieved from <https://microbiologyinfo.com/mannitol-salt-agar-for-the-isolation-of-staphylococcus-aureus/>
- Asrori, S., Achmad, A. I., Herliyana, E. N. and Rijal, S. (2012). Efektivitas penghambatan ekstrak daging biji *picung* (*Pangium edule* Reinw.) terhadap pertumbuhan *Rhizoctonia* sp. secara in vitro. *Journal of Horticulture*, 22(3): 268-75.
- Atabani, A. E., Badruddin, I. A., Masjuki, H. H., Chong, W. T. and Lee, K. T. (2015). *Pangium edule* Reinw.: a promising non-edible oil feedstock for biodiesel production. *Arabian Journal for Science and Engineering*, 40(2): 583-594.
- Ayu, D. F., Man, Y. C. and Rohman, A. (2017). Chemical properties, fatty acid composition, and lipid profiles of *picung* (*Pangium edule* Reinw.) kernel oil from Riau province. *Applied Science and Technology*, 1(1): 41-46.
- Azlim, A. A., Ahmed, J. K., Syed, Z. I., Mustapha, S. K., Aisyah, M. R. and Kamarul, R. K. (2010). Total phenolic content and primary antioxidant activity of methanolic and ethanolic extracts of aromatic plants' leaves. *International Food Research Journal*, 17(4): 1077-1084.
- Azmir, J., Zaidul, I. S. M., Rahman, M. M., Sharif, K. M., Mohamed, A., Sahena, F., Jahurul, M. H. A., Ghafoor, K., Norulainid, N. A. N. and Omar, A. K. M. (2013). Techniques for extraction of bioactive compounds from plant materials: A review. *Journal of Food Engineering*, 117(4): 426-436.
- Azwanida, N. N. (2015). A review on the extraction methods use in medicinal plants, principle, strength and limitation. *Medicinal and Aromatic Plants Research Journals*, 4(196): 2167-0412.
- Bacteriological Analytical Manual (BAM). (1998). U. S. Food and Drug Administration. 8th Edition, Revision A.

- Badenes, M. L. and Byrne, D. H. (Eds.). (2012). *Fruit Breeding* (Vol. 8). New York: Springer Science and Business Media.
- Bahera, B. C., Verma, N. Sonone, A. and Makhija. U. (2008). Antioxidant and antibacterial properties of some cultured lichens. *Bioresource Technology*, 99: 7424-7424.
- Balouiri, M., Sadiki, M. and Ibsouda, S. K. (2016). Methods for in vitro evaluating antimicrobial activity: A review. *Journal of Pharmaceutical Analysis*, 6(2): 71-79.
- Bamforth, C. W. (2005). *Food, Fermentation and Micro-organisms*. Oxford, UK: Blackwell Science, 143-153.
- Belitz, H. D., Grosch, W. and Schieberle, P. (2004). Carbohydrates. In “Food Chemistry” 3rd ed., Translated by Burghagen, M. M. Springer-Verlag, Berlin: 245-341
- Bhargav, S., Panda, B. P., Ali, M. and Javed, S. (2008). Solid-state fermentation: An overview. *Chemical and Biochemical Engineering Quarterly*, 22(1): 49-70.
- Biswas, D. and Roymon, M. G. (2013). LC/ TOF/ ESI/ MS based detection of bioactive compounds present in leaf and bark extract of *Acacia Arabica*. *Recent Research in Science and Technology*, 5(2): 37-40.
- Bonev, B., Hooper, J., and Parisot, J. (2008). Principles of assessing bacterial susceptibility to antibiotics using the agar diffusion method. *Journal of Antimicrobial Chemotherapy*, 61(6): 1295-1301.
- Böttcher, J., Margraf, M. and Monks, K. (1997). *HPLC Basics- Principle and parameters*. Science Together, Knauer Wissenschaftliche Geräte GmbH: Berlin.
- Bravo, L. (1998). Polyphenols: chemistry, dietary sources, metabolism, and nutritional significance. *Nutrition Reviews*, 56(11): 317-333.
- Bucar, F., Wube, A. and Schmid, M. (2013). Natural product isolation—how to get from biological material to pure compounds. *Natural Product Reports*, 30(4): 525-545.
- Burt, S. (2004). Essential oils: Their antibacterial properties and potential applications in foods—a review. *International Journal of Food Microbiology*, 94(3): 223-253.
- Burtner, C. R., Murakami, C. J., Kennedy, B. K. and Kaeberlein, M. (2009). A molecular mechanism of chronological aging in yeast. *Cell Cycle*, 8(8): 1256–1270.
- Byaruhanga, Y. B., Bester, B. H. and Watson, T. G. (1999). Growth and survival of *Bacillus cereus* in *mageu*, a sour maize beverage. *World Journal of Microbiology and Biotechnology*, 15(3): 329-333.
- Campbell-Platt, G. (1987). *Fermented foods of the world*. A dictionary and guide. London: Butterworths.
- Cantón, E., Pemán, J., Viudes, A., Quindós, G., Gobernado, M. and Espinel-Ingroff, A. (2003). Minimum fungicidal concentrations of amphotericin B for bloodstream *Candida* species. *Diagnostic Microbiology and Infectious Disease*, 45(3): 203-206.

- Caplice, E. and Fitzgerald, G. F. (1999). Food fermentations: Role of microorganisms in food production and preservation. *International Journal of Food Microbiology*, 50(1-2): 131-149.
- Capozzi, V., Fragasso, M., Romaniello, R., Russo, P. and Spano, G. (2017). Spontaneous food fermentations and potential risks for human health. *Fermentation*, 3(4): 49.
- Centre for Food Safety Food and Environmental Hygiene Department. (2014). *Microbiological Guidelines for Food (For ready-to-eat food in general and specific food items)*. Queensway: Food and Environmental Hygiene Department. *ICMSF Publications*, 1-38.
- Cerretani, L. and Bendini, A. (2010). Rapid assays to evaluate the antioxidant capacity of phenols in virgin olive oil. *Olives and Olive Oil in Health and Disease Prevention*, 625–635. Academic Press.
- Chai, K. F., Adzahan, N. M., Karim, R., Rukayadi, Y. and Ghazali, H. M. (2018). Effects of fermentation time and turning intervals on the physicochemical properties of rambutan (*Nephelium lappaceum* L.) fruit Sweetings. *Sains Malaysiana*, 47(10): 2311-2318.
- Chelule, P. K., Mbongwa, H. P., Carries, S. and Gqaleni, N. (2010). Lactic acid fermentation improves the quality of *amahewu*, a traditional South African maize-based porridge. *Food Chemistry*, 122(3): 656-661.
- Chen, L., Madl, R. L., Vadlani, P. V., Li, L. and Wang, W. (2013). Value added products from soybean: removal of anti-nutritional factors via bioprocessing. *Soybean: A review*, 161-179.
- Chin, S. Y., Chye, F. Y. and Anton, A. (2016). Microbiological diversity of spontaneous fermentation of *bambangan* (*Mangifera pajang*), a traditional fermented fruit from northern Borneo. *Short Communication Biotechnology*, 2, 1-9.
- Chiu, C. M. and Muddiman, D. C. (2008). What is Mass Spectrometry? *American Society for Mass Spectrometry*.
- Chojnacka, K. (2006). *Fermentation products*. Chemical Engineering and Chemical Process Technology. 12
- Choudhary, A. (2008). Importance of negative and positive control in microbial analysis. *Pharmaceutical Guidelines*.
- Christie, W. W., & Han, X. (2010). *Isolation, Separation, Identification and Lipidomic Analysis*. UK: Oily Press, Bridgwater.
- Chrun, R., Hosotani, Y., Kawasaki, S. and Inatsu, Y. (2017). Microbiological hazard contamination in fermented vegetables sold in local markets in Cambodia. *Biocontrol Science*, 22(3): 181-185.
- Chukeatirote, E., Chainun, C., Siengsubchart, A., Moukamnerd, C., Chantawannakul, P., Lumyong, S., Boontim, N. and Thakang, P. (2010). Microbiological and biochemical changes in *thua nao* fermentation. *Research Journal of Microbiology*, 5(7): 644-650.

- Chye, F. Y. and Sim, K. Y. (2009). Antioxidant and antibacterial activities of *Pangium edule* seeds extracts. *International Journal of Pharmacology*, 5(5): 285-297
- CLSI (Clinical and Laboratory Standards Institute). (2012). Reference Method for Performance Standards for Antimicrobial Disk Susceptibility Tests; Approved Standard-Eleventh Edition, CLSI document M02-A11. USA: *CLSI*.
- Corry, J. E., Curtis, G. D. W. and Baird, R. M. (Eds.). (1996). *Culture media for food microbiology*, Elsevier. 34.
- Cowan, M.M. (1999). Plant products as antimicrobial agents. *Clinical Microbiology Reviews*, 12(4): 564-582.
- Csoma, H., Zakany, N., Capece, A., Romano, P. and Sipiczki, M. (2010). Biological diversity of *Saccharomyces* yeasts of spontaneously fermenting wines in four wine regions: comparative genotypic and phenotypic analysis. *International Journal of Food Microbiology*, 140(2-3): 239-248.
- Cunningham, J. and Sobolewski, R. (2011). Food composition databases for nutrition labelling: Experience from Australia. *Journal of Food Composition and Analysis*, 24(4-5): 682-685.
- Curutiu, C., Iordache, F., Gurban, P., Lazar, V. and Chifiriuc, M. C. (2019). Main microbiological pollutants of bottled waters and beverages. *Bottled and Packaged Water*, 403-422.
- Dai, J. and Mumper, R. J. (2010). Plant phenolics: extraction, analysis and their antioxidant and anticancer properties. *Molecules*, 15(10): 7313-7352.
- David, W., Darwin, C. and Kasim, A. (2014) Picung (*Pangium Edule* Reinw.) as traditional food preservative. *IFOAM Organic World Congress*, 1-3.
- Deshpande, S. S., Salunkhe, D. K., Oyewole, O. B., Azam-Ali, S., Battcock, M. and Bressani, R. (2000). *Fermented grain legumes, seeds, and nuts. A global perspective*. Food and Agriculture Organization of the United Nations, Rome. 20-21.
- Do, Q. D., Angkawijaya, A. E., Tran-Nguyen, P. L., Huynh, L. H., Soetaredjo, F. E., Ismadji, S. and Ju, Y. H. (2014). Effect of extraction solvent on total phenol content, total flavonoid content, and antioxidant activity of *Limnophila aromatica*. *Journal of Food and Drug Analysis*, 22(3): 296-302.
- Dong, L., Zhou, H.-W., Sonogo, L., Lers, A. and Lurie, S. (2001). Ripening of "Red Rosa" plums: effect of ethylene and 1-methylcyclopropene. *Australian Journal of Plant Physiology*, 8: 1039-1045.
- Doriya, K., Jose, N., Gowda, M., & Kumar, D. S. (2016). Solid-State Fermentation vs Submerged Fermentation for the Production of l-Asparaginase. *Advances in Food and Nutrition Research*, 115-135.
- Dzogbefia, V. P., Buamah, R. and Oldham, J. H. (1999). The controlled fermentation of cocoa (*Theobroma cacao* L.) using yeasts: Enzymatic process and associated physico-chemical changes in cocoa sweatings. *Food Biotechnology*, 13: 1-12.

- Earle, M. D. (1997). Innovation in the food industry. *Trends in Food Science and Technology*, 8(5), 166-175.
- Ejoh, R. A., Djuikwo, V. N., Gouado, I. and Mbofung, C. M. (2007). Effect of the method of processing and preservation on some quality parameters of three non-conventional leafy vegetables. *Pakistan Journal of Nutrition*, 6(2): 128-133.
- Eka, O. U. (1980). Effect of fermentation on the nutrient status of locust beans. *Food Chemistry*, 5:303-308
- El-Farmawi., E. Olama, D. Z. and Holail, H. (2014). The antibacterial effect of some natural bioactive materials against *Klebsiella pneumoniae* and MRSA. *International Journal of Current Microbiology and Applied Science*. 3: 576-588.
- Elias, M., Wiczorek, G., Rosenne, S. and Tawfik, D. S. (2014). The universality of enzymatic rate-temperature dependency. *Trends in Biochemical Sciences*, 39(1): 1-7.
- Elliott, T., Casey, A., Lambert, P. A. and Sandoe, J. (2012). Lecture Notes: *Medical Microbiology and Infection*. John Wiley and Sons.
- Ensminger, M. E. and Ensminger, A. H. (1993). *Foods and Nutrition Encyclopedia*, Two Volume Set. CRC press.
- Espinel-Ingroff, A., Chaturvedi, V., Fothergill, A. and Rinaldi, M. G. (2002a). Optimal testing conditions for determining MICs and minimum fungicidal concentrations of new and established antifungal agents for uncommon molds: NCCLS collaborative study. *Journal of Clinical Microbiology*, 40(10): 3776-3781.
- Espinel-Ingroff, A., Fothergill, A., Peter, J., Rinaldi, M. G. and Walsh, T. J. (2002b). Testing conditions for determination of minimum fungicidal concentrations of new and established antifungal agents for *Aspergillus* spp.: NCCLS collaborative study. *Journal of Clinical Microbiology*, 40(9): 3204-3208.
- Fagerquist, C. K., Neese, R. A., & Hellerstein, M. K. (1999). Molecular ion fragmentation and its effects on mass isotopomer abundances of fatty acid methyl esters ionized by electron impact. *Journal of the American Society for Mass Spectrometry*, 10(5): 430-439.
- Faridah-Hanum, I. (2007). Morphological variation of *Pangium edule* Reinw. fruits in Malaysia, Faculty of Forestry *Universiti Pertanian Malaysia (UPM), Malaysia*.
- Farkas, J. (2007). Physical methods of food preservation. In Food Microbiology: Fundamentals and Frontiers, *American Society of Microbiology*, 3: 685-712.
- Fatoki, O. A. and Onifade, D. A. (2013). Use of plant antimicrobials for food preservation. *International Journal of Veterinary and Agricultural Engineering*, 7(12): 1110-1113.
- Fernández, V., Sancho-Knapik, D., Guzmán, P., Peguero-Pina, J.J., Gil, L., Karabourniotis, G., Khayet, M., Fasseas, C., Heredia-Guerrero, J.A., Heredia, A. and Gil-Pelegrín, E. (2014). Wettability, polarity, and water absorption of holm oak leaves: effect of leaf side and age. *Plant Physiology*, 166(1): 168-180.

- Fernandez-Lopez, Zhi, N., Aleson-Carbonell, L., Perez-Alvarez, J. A. and Kuri, V. (2005). Antioxidant and antibacterial activities of natural extract: Application in beef meatball. *Meat Science*, 69: 371-380.
- Foerster, S., Unemo, M., Hathway, L. J., Low, N. and Althus, C. L. (2016). Time-kill curve analysis and pharmacodynamics modelling for *in vitro* evaluation of antimicrobials against *Neisseria gonorrhoeae*. *BMC Microbiology*, 16(216): 1-11.
- Food and Agriculture Organization (FAO). (2002). The State of Food Insecurity in the World. Rome.
- Food and Drug Administration (FDA). (2001). Bacteriological Analytical Manual (BAM) Media M127: Potato Dextrose Agar. Retrieved from <https://www.fda.gov/food/laboratory-methods-food/bam-media-m127-potato-dextrose-agar>
- Frankel, E. N. (2007). Antioxidants in food and biology: Facts and fiction. *Antioxidants in Food and Biology: Facts and Fiction*. 1-254.
- Gadaga, T. H., Nyanga, L. K. and Mutukumira, A. N. (2007). The occurrence, growth and control of pathogens in African fermented foods. *African Journal of Food, Agriculture Nutrition and Development*, 4.
- Gangoué-Piéboji, J., Eze, N., Djintchui, A. N., Ngameni, B., Tsabang, N., Pegnyemb, D. E., Biyiti, L., Ngassam, P., Koulla-Shiro, S. and Galleni, M. (2009). The in-vitro antimicrobial activity of some medicinal plants against β -lactam-resistant bacteria. *The Journal of Infection in Developing Countries*, 3(09): 671-680.
- Gargaud, M. (2011). *Encyclopedia of Astrobiology*. Berlin, Heidelberg: Springer Science and Business Media.
- Gehrt, A., Peter, J., Pizzo, P. A. and Walsh, T. J. (1995). Effect of increasing inoculum sizes of pathogenic filamentous fungi on MICs of antifungal agents by broth microdilution method. *Journal of Clinical Microbiology*, 33(5): 1302-1307.
- Ghosh, J. S. (2015). Solid state fermentation and food processing: A short review. *Journal of Nutrition and Food Science*, 6(1): 1-7.
- Giraffa, G. (2004). Studying the dynamics of microbial populations during food fermentation. *FEMS Microbiology Reviews*, 28(2): 251-260.
- Gomez, F. (2011). Aerobe. In Gargaud, M. (Ed.), *Encyclopedia of Astrobiology*, 21-21. Berlin, Heidelberg: Springer Science and Business Media.
- Griffin, D. H. (1994). Fungal physiology: Spore dormancy and germination. 2nd edition. 468. New York: John Wiley and Sons.
- Gulcin I. (2006). Antioxidant activity of caffeic acid (3,4-dihydroxycinnamic acid). *Toxicology*, 217: 213-220.
- Gulcin, I. (2012). Antioxidant activity of food constituents: an overview. *Archives of Toxicology*, 86(3): 345-391.

- Hailat, I. A. (2014). *Mass spectrometric methods for analysis of sterols and steryl esters in biological samples* (Doctoral dissertation, Memorial University of Newfoundland).
- Hallgren, B., Ryhage, R., & Stenhagen, E. (1959). The mass spectra of methyl oleate, methyl linoleate, and methyl linolenate. *Acta Chemica Scandinavica*, 13(4): 845-847
- Hamad, A., Mahardika, M. G. P., Istifah, I. and Hartanti, D. (2016). Antimicrobial and volatile compounds study of four spices commonly used in Indonesian culinary. *Journal of Food and Pharmaceutical Sciences*, 4(1): 1-5.
- Handa, S. S., Khanuja, S. P. S., Longo, G. and Rakesh, D. D. (2008). Extraction technologies for medicinal and aromatic plants, (1st edition), no. 66. Italy: *United Nations Industrial Development Organization and the International Centre for Science and High Technology*.
- Hangesti, R. A. 2006. Pengaruh Pengawetan Penggunaan Daging biji picung (*Pangium edule* Reinw) Terhadap Kesegaran dan Keamanan Ikan Kembung Segar (*Rastrelliger brachysoma*). *Tesis Institut Pertanian Bogor*. Bogor.
- Hansen, C. E., del Olmo, M. and Burri, C. (1998). Enzyme activities in cocoa beans during fermentation. *Journal of the Science of Food and Agriculture*, 77: 273-281.
- Hasan, M. N., Sultan, M. Z. and Mar-E-Um, M. (2014). Significance of fermented food in nutrition and food science. *Journal of Scientific Research*, 6(2): 373-386.
- Hase, J., Desmukh, K. K., Pokharkar, R. D., Gaje, T. R., & Phatanagre, N. D. (2017). Phytochemical Studies on Nerium oleander L. using GC-MS. *International Journal of Pharmacognosy and Phytochemical Research*, 9(6): 885-891.
- Hatami, T., Emami, S. A., Miraghaee, S. S. and Mojjarrab, M. (2014). Total phenolic contents and antioxidant activities of different extracts and fractions from the aerial parts of *Artemisia biennis* Willd. *Iranian Journal of Pharmaceutical Research (IJPR)*, 13(2): 551.
- Hawashi, M., Widjaja, T. and Gunawan, S. (2019). Solid-state fermentation of cassava products for degradation of anti-nutritional value and enrichment of nutritional value. *New Advances on Fermentation Processes*, 1-18.
- Hayek, S. A. and Ibrahim, S. A. (2013). Current limitations and challenges with lactic acid bacteria: A review. *Food and Nutrition Sciences*, 4(11): 73-87.
- Hernández, Y., Lobo, M. G. and González, M. (2009). Factors affecting sample extraction in the liquid chromatographic determination of organic acids in papaya and pineapple. *Food Chemistry*, 114(2): 734-741.
- Heruwati, E. S. and Mangunwardoyo, W. (2009). Antibacterial test of *Pangium (Pangium edule* Reinw.) extract against the growth of fish spoilage bacteria. *Indonesian Fisheries Research Journal*, 15(1): 65-73.

- Heruwati, E. S., Widyasari, H. E. and Haluan, J. (2007). Pengawetan ikan segar menggunakan biji *picung* (*Pangium edule* Reinw.). *Jurnal Pascapanen dan Bioteknologi Kelautan dan Perikanan*, 2(1): 9-18.
- Hewlett-Packard Company. (1998). *Basics of LC/MS. A primer*. USA: Agilent
- Ho, V. T. T., Zhao, J. and Fleet, G. (2014). Yeasts are essential for cocoa bean fermentation. *International Journal of Food Microbiology*, 174: 72-87.
- Hoe, V. B. and Siong, K. H. (1999). The nutritional value of indigenous fruits and vegetables in Sarawak. *Asia Pacific Journal of Clinical Nutrition*, 8(1): 24-31.
- Holbrook, R. and Anderson, J. M. (1980). An improved selective and diagnostic medium for the isolation and enumeration of *Bacillus cereus* in foods. *Canadian Journal of Microbiology*, 26(7): 753-759.
- Holzapfel, W. H. and Schillinger, U. (2002). Introduction to pre-and probiotics. *Food Research International*, 35(2-3): 109-116.
- Honn, K. V., & Zeldin, D. C. (Eds.). (2019). *The Role of Bioactive Lipids in Cancer, Inflammation and Related Diseases*. Springer. 179.
- Howell, K. S., Cozzolino, D., Bartowsky, E. J., Fleet, G. H. and Henschke, P. A. (2006). Metabolic profiling as a tool for revealing *Saccharomyces* interactions during wine fermentation. *FEMS Yeast Research*, 6(1): 91-101.
- Huang, S. L., Wu, L. C., Liang, H. K., Pan, K. T., Horng, J. T. and Ko, M. T. (2004). PGTdb: a database providing growth temperatures of prokaryotes. *Bioinformatics*, 20(2): 276-278.
- Hur, S. J., Lee, S. Y., Kim, Y. C., Choi, I. and Kim, G. B. (2014). Effect of fermentation on the antioxidant activity in plant-based foods. *Food Chemistry*, 160: 346-356.
- Hussain, S. Z. and Maqbool, K. (2014). GC-MS: Principle, technique and its application in food science. *International Journal of Current Science*, 13: 116-126.
- ICMSF (International Commission on Microbiological Specifications for Foods). (1996). *Microorganisms in food 5: Microbiological specifications of food pathogens*. London: Blackie Academic and Professional.
- ICMSF (International Commission on Microbiological Specifications for Foods) (2005). *2nd Edition. Microbial ecology of foods. Volume 2, Factors affecting life and death of microorganisms*. Orlando. Orlando : Academic Press, Vol. 2. ISBN: 0-306-48675-X.
- Jay, J. M., Loessner, M. J. and Golden, D. A. (2008). *Modern Food Microbiology*. Springer Science & Business Media.
- Jobim, M. L., Santos, R. C. V., Alves, C. F. D. S., Oliveira, R. M., Mostardeiro, C. P., Sagrillo, M. R., Filho, O. C. S., Garcia, L. F. M., Manica-Cattani, M. F., Ribeiro, E. E. and Manica da Cruz, I. B. (2014). Antimicrobial activity of amazon *Astrocaryum aculeatum* extracts and its association to oxidative metabolism. *Microbiological Research*, 169: 314-323.

- Kandola, A. (2018). *Benefits of castor oil for the face and skin*. Medical News Today. Retrieved from <https://www.medicalnewstoday.com/articles/319844.php>
- Kang, J. S. (2012). Principles and applications of LC-MS/MS for the quantitative bioanalysis of analytes in various biological samples. In *Tandem Mass Spectrometry-Applications and Principles*. IntechOpen. Europe China: 20: 441-477.
- Kasim, A. and David, W. (2013). Characteristic of *Pangium edule* Reinw. as food preservative from different geographical sites. *Asia Pacific Journal of Sustainable Agriculture, Food and Energy*, 1(1): 6-9.
- Kayser, F. H., Bienz, K. A., Eckert, J. and Zingernagel, R. M. (2001). *Medical Microbiology* (10th edition). Stuttgart, Germany: Georg Thieme Verlag.
- Khameneh, B., Iranshahy, M., Soheili, V. and Fazly Bazzaz, B. S. (2019). Review on plant antimicrobials: a mechanistic viewpoint. *Antimicrobial Resistance and Infection Control*, 8: 118.
- Khoddami, A., Wilkes, M. A. and Roberts, T. H. (2013). Techniques for analysis of plant phenolic compounds. *Molecules*, 18(2): 2328-2375.
- King, E. O., Ward, M. K. and Raney, D. E. (1954). Two simple media for the demonstration of pyocyanin and fluorescin. *The Journal of Laboratory and Clinical Medicine*, 44(2): 301-307.
- Ko, G. A. and Cho, S. K. (2018). Ethyl linoleate inhibits α -MSH-induced melanogenesis through Akt/GSK3 β / β -catenin signal pathway. *The Korean Journal of Physiology and Pharmacology*, 22(1): 53.
- Kothe, E. and Köhler, J. M. (2015). Changing growth behavior of heavy-metal tolerant bacteria: media optimization using droplet-based microfluidics. *Engineering in Life Sciences*, 15(3): 327–332.
- Kotzekidou, P., Giannakidis, P. and Boulamatsis, A. (2008). Antimicrobial activity of some plant extracts and essential oils against foodborne pathogens *in vitro* and on the fate of inoculated pathogens in chocolate. *Food Science and Technology*, 41(1): 119-127.
- Krimmel, B., Swoboda, F., Solar, S. and Reznicek, G. (2010). OH-radical induced degradation of hydroxybenzoic-and hydroxycinnamic acids and formation of aromatic products-a gamma radiolysis study. *Radiation Physics and Chemistry*, 79(12): 1247-1254.
- Krishnan, K. R., James, F. and Mohan, A. (2016). Isolation and characterization of n-hexadecanoic acid from *Canthium parviflorum* leaves. *Journal of Chemical and Pharmaceutical Research*, 8(8): 614-617.
- Kumar, V., Sharma, A., Bhardwaj, R., & Thukral, A. K. (2017). Analysis of organic acids of tricarboxylic acid cycle in plants using GC-MS, and system modeling. *Journal of Analytical Science and Technology*, 8(1): 20.
- Lazos, E. S., Aggelousis, G. and Bratakos, M. (1993): The fermentation of Trahanas: a milk-wheat flour combination. *Plant Foods for Human Nutrition*, 44: 45-62.

- Lederberg, J. (2000). *Pseudomonas*. In *Encyclopedia of Microbiology*, 2(3): 876-891. San Diego, California U.A.: Academic Press.
- Lemmer, A. (2012). Mineral substances and macronutrients in the anaerobic conversion of biomass: An impact evaluation. *Engineering in Life Sciences*, 12(3): 287-294.
- Leininger, D. J., Roberson, J. R. and Elvinger, F. (2001). Use of eosin methylene blue agar to differentiate *Escherichia coli* from other gram-negative mastitis pathogens. *Journal of Veterinary Diagnostic Investigation*, 13(3): 273-275.
- Liam, J., Faridah-Hanum, I. and Hakeem, K. R. (2013). Phytochemical compounds of *Pangium edule* Reinw. Seeds. *Proceedings of the International Forestry Graduate Students' Conference*, 116-120.
- Lim, T. K. (2013). *Pangium edule*. In *Edible Medicinal and Non-Medicinal Plants*, 780-784. Springer, Dordrecht.
- Liptáková, D., Matejčková, Z. and Valík, E. (2017). Lactic acid bacteria and fermentation of cereals and *pseudocereals*. *Fermentation Processes*, 10: 65459.
- Liu, R. H. (2004). Potential synergy of phytochemicals in cancer prevention: mechanism of action. *The Journal of Nutrition*, 134(12): 3479S-3485S.
- Liu, R. H. and Felice, D. L. (2007). Antioxidants and whole food phytochemicals for cancer prevention. In *ACS Symposium Series*, 956: 15-34. Oxford University Press.
- Liu, X., Jia, B., Sun, X., Ai, J., Wang, L., Wang, C., Zhao, F., Zhan, J. and Huang, W. (2015). Effect of initial pH on growth characteristics and fermentation properties of *Saccharomyces cerevisiae*. *Journal of Food Science*, 80(4): 800-808.
- Loganayaki, N., Siddhuraju, P. and Manian, S. (2013). Antioxidant activity and free radical scavenging capacity of phenolic extracts from *Helicteres isora* L. and *Ceiba pentandra* L. *Journal of Food Science and Technology*, 50(4): 687-695.
- Lopez, S., Bermudez, B., Pacheco, Y. M., Ortega, A., Varela, L. M., Abia, R. and Muriana, F. J. (2010). Oleic acid: The main component of olive oil on postprandial metabolic processes. In *Olives and Olive Oil in Health and Disease Prevention*, 1385-1393. Academic Press.
- Mahesh, B. and Satish, S. (2008). Antimicrobial activity of some important medicinal plant against plant and human pathogens. *World Journal of Agricultural Sciences*, 4(5): 839-843.
- Majhenič, L., Škerget, M. and Knez, Ž. (2007). Antioxidant and antimicrobial activity of *guarana* seeds extracts. *Food Chemistry*, 104(3): 1258-1268.
- Makinde, F. M. and Akinoso, R. (2014). Comparison between the nutritional quality of flour obtained from raw, roasted and fermented sesame (*Sesamum indicum* L.) seeds grown in Nigeria. *Acta Scientiarum Polonorum Technologia Alimentaria*, 13(3): 309-319.
- Mangunwardoyo, W., Ismaini, L., and Heruwati, E. S. (2008). Analisis senyawa bio aktif dari ekstrak biji *picung* (*Pangium edule* Reinw.) segar [Analysis of bioactive

- compounds in fresh seeds extract of *picung* (*Pangium edule* Reinw.)). *Berita Biologi*, 9(3): 259-264.
- McDonnell, G. and Russell, A.D. (1999). Antiseptics and disinfectants: activity, action and resistance. *Clinical Microbiology Reviews*, 12(1): 147-79.
- McGovern, P. E., Zhang, J., Tang, J., Zhang, Z., Hall, G. R., Moreau, R. A., Nunez, A., Butrym, E. D., Richards, M. P., Wang, C., Cheng, G., Zhao, Z. and Wang, C. (2004). Fermented beverages of pre-and proto-historic China. *Proceedings of the National Academy of Sciences of the United States of America*, 101(51): 17593-17598.
- Mehdizadeh, S., Lasekan, O., Muhammad, K. and Baharin, B. (2015). Variability in the fermentation index, polyphenols and amino acids of seeds of rambutan (*Nephelium lappaceum* L.) during fermentation. *Journal of Food Composition and Analysis*, 37: 128-135.
- Mehta, B. M., Kamal-Eldin, A. and Iwanski, R. Z. (2012). *Fermentation: effects on food properties*. CRC Press.
- Messens, W. and De Vuyst, L. (2002). Inhibitory substances produced by *Lactobacilli* isolated from sourdoughs—a review. *International Journal of Food Microbiology*, 72(1-2): 31-43.
- Miller, A. L. (1996). Antioxidant flavonoids: structure, function and clinical usage. *Alternative Medicine Review*, 1(2): 103-111.
- Mira, N. P., Teixeira, M. C. and S´a-Correia, I. (2010). Adaptive response and tolerance to weak acids in *Saccharomyces cerevisiae*: a genome-wide view. *Omic*s, 14(5): 525–40.
- Mohamed, N. and Abdullah, A. (2016). Comparison of total phenolic content and antioxidant activity of *Kappaphycus alvarezii* from Langkawi and Semporna. *In AIP Conference Proceedings*, 1784(1): 030040. AIP Publishing.
- Mohammed, Y. H., Ghaidaa, J. M., & Imad, H. H. (2016). Analysis of bioactive chemical compounds of *Nigella sativa* using gas chromatography-mass spectrometry. *Journal of Pharmacognosy and Phytotherapy*, 8(2): 8–24.
- Molyneux, P. (2004). The use of the stable free radical diphenylpicrylhydrazyl (DPPH) for estimating antioxidant activity. *Songklanakarin Journal of Science Technology*, 26(2): 211-219.
- Montagnac, J. A., Davis, C. R., & Tanumihardjo, S. A. (2009). Processing Techniques to Reduce Toxicity and Antinutrients of Cassava for Use as a Staple Food. *Comprehensive Reviews in Food Science and Food Safety*, 8(1): 17–27.
- Moritsuka, N., Kawamura, K., Tsujimoto, Y., Rabenarivo, M., Andriamananjara, A., Rakotoson, T., & Razafimbelo, T. (2019). Comparison of visual and instrumental measurements of soil color with different low-cost colorimeters. *Soil Science and Plant Nutrition*, 1–11.

- Morse, N. (2015). Are some health benefits of palmitoleic acid supplementation due to its effects on 5' adenosine monophosphate-activated protein kinase (AMPK)? *Lipid Technology*, 27(12): 278-281.
- Mueller, M., Pena, A. and Derendorf, H. (2004). Issues in pharmacokinetics and pharmacodynamics of anti-infective agents: kill curves versus MIC. *Antimicrobial Agents and Chemotherapy*, 48(2): 369-377.
- Muth, N. D. and Zive, M. M. (2019). *Sports nutrition for health professionals*. FA Davis Company.
- Naczki, M. and Shahidi, F. (2006). Phenolics in cereals, fruits and vegetables: Occurrence, extraction and analysis. *Journal of Pharmaceutical and Biomedical Analysis*. 41(5): 1523–1542.
- Nazarni, R., Purnama, D., Umar, S., Eni, H. (2016). The effect of fermentation on total phenolic, flavonoid and tannin content and its relation to antibacterial activity in jaruk tigarun (*Crataeva nurvala*, Buch HAM). *International Food Research Journal*, 23(1): 309-315.
- Nielsen, S. S. (2010). United States government regulations and international standards related to food analysis. In *Food Analysis*, 15-33. Springer, Boston, MA.
- Nielsen, S. S. (Ed.). (2010). *Food Analysis*, 139-141. New York: Springer.
- Noui, Y., Lombarkia, O. A., Bekrar, A., Chibane, H. A., Lekbir, A., Abdeddaim, M., Fahloul, D. and Bacha, A. (2014). Comparative study of the physicochemical characteristics and antioxidant activity of three dates varieties (*Phoenix Dactylifera* L.) grown in Algeria. *Annals Food Science and Technology*, 15(2): 276-283.
- Oboh, G., Alabi, K. B. and Akindahunsi, A. A. (2008). Fermentation changes the nutritive value, polyphenol distribution, and antioxidant properties of *Parkia biglobosa* seeds (African locust beans). *Food Biotechnology*, 22(4): 363–376.
- Ogunmoyole, T., Rocha, J. B. T., Okoronkwo, A. E. and Kade, I. J., 2009. Altered pH homeostasis modulates the glutathione peroxidase mimics and other antioxidant properties of diphenyl diselenide. *Chemico-biological interactions*, 182(2-3): 106-111.
- Onyango, C., Noetzold, H., Ziemis, A., Hofmann, T., Bley, T. and Henle, T. (2005). Digestibility and antinutrient properties of acidified and extruded maize–finger millet blend in the production of Uji. *Food Science and Technology*, 38(7): 697-707
- Oregon State University. (2012). GC-MS: How does it work? Environmental Health Sciences Center Corvallis. Retrieved from http://www.unsolvedmysteries.oregonstate.edu/MS_05
- Orij, R., Brul, S. and Smits, G. J. (2011). Intracellular pH is a tightly controlled signal in yeast. *Biochimica et Biophysica Acta (BBA)-General Subjects*, 1810(10): 933-944.

- Oseni, O. A. and Akindahunsi, A. A. (2011). Some phytochemical properties and effect of fermentation on the seeds of *Jatropha curcas* L. *American Journal of Food Technology*, 6(2): 158-165.
- Oseni, O. A., and Ekperigin, M. (2007). Studies on biochemical changes in maize wastes fermented with *Aspergillus niger*. *Biokemistri*, 19(2):75-79
- Othman, A., Mukhtar, N. J., Ismail, N. S. and Chang, S. K. (2014). Phenolics, flavonoids content and antioxidant activities of 4 Malaysian herbal plants. *International Food Research Journal*, 21(2): 759.
- Othman, M., Wiart, H. S., Khoo, C., Lim, T. J. and Ting, K. N. (2011). Optimal methods for evaluating antimicrobial activities from plant extracts. *Journal of Microbiological Methods*, 84: 161-166.
- Othman, N. B., Roblain, D., Chammen, N., Thonart, P. and Hamdi, M. (2009). Antioxidant phenolic compounds loss during the fermentation of *Chétoui olives*. *Food Chemistry*, 116(3): 662-669.
- Oxoid Manual (2019). *Bacillus cereus* selective agar base. *Thermo Fisher Scientific Inc.* 8th Edition. United Kingdom: Oxoid.
- Pankey, G. A. and Ashcraft, D. S. (2009). *In vitro* antibacterial activity of tigecycline against resistant Gram-negative *bacilli* and *enterococci* by time-kill assay. *Diagnostic Microbiology and Infectious Disease*, 64(3): 300–304.
- Papalexandratou, Z., Vrancken, G., De Bruyne, K., Vandamme, P. and De Vuyst, L. (2011). Spontaneous organic cocoa bean box fermentations in Brazil are characterized by a restricted species diversity of lactic acid bacteria and acetic acid bacteria. *Food Microbiology*, 28(7): 1326–38.
- Papoti, V. T. and Tsimidou, M. Z. (2009). Impact of sampling parameters on the radical scavenging potential of olive (*Olea europaea* L.) leaves. *Journal of Agricultural and Food Chemistry*, 57(9): 3470-3477.
- Parekh, J., Jadeja, D. and Chanda, S. (2005). Efficacy of aqueous and methanol extracts of some medicinal plants for potential antibacterial activity. *Turkish Journal of Biology*, 29: 203-210.
- Parkouda, C., Nielsen, D. S., Azokpota, P., Ivette Irène Ouoba, L., Amoa-Awua, W. K., Thorsen, L., Hounhouigan, J. D., Jensen, J. S., Tano-Debrah, K., Diawara, B. and Jakobsen, M. (2009). The microbiology of alkaline fermentation of indigenous seeds used as food condiments in Africa and Asia. *Critical Reviews in Microbiology*, 35(2): 139-156.
- Parvin, S., Easmin, D., Sheikh, A., Biswas, M., Sharma, S. C., Jahan, M. G., Islam, M.A., Roy, N. and Shovon, M.S. (2015). Nutritional analysis of date fruits (*Phoenix dactylifera* L.) in perspective of Bangladesh. *American Journal of Life Sciences*, 3(4): 274-278.
- Pay, H. and Scopes, E. (2017). Thermo scientific chromogenic coliform agar (ISO) demonstrates superior detection of coliforms from waters with low bacterial numbers. Application note. 7 *Thermo Fisher Scientific Inc.* Merck: Darmstadt, Germany.

- Pepe, O., Blaiotta, G., Bucci, F., Anastasio, M., Aponte, M. and Villani, F. (2006). *Staphylococcus aureus* and *staphylococcal enterotoxin A* in breaded chicken products: detection and behaviour during the cooking process. *Applied and Environmental Microbiology*, 72(11): 7057-7062.
- Pfaller, M. A., Sheehan, D. J. and Rex, J. H. (2004). Determination of fungicidal activities against yeasts and molds: Lessons learned from bactericidal testing and the need for standardization. *Clinical Microbiology Reviews*, 17: 268-280.
- Pichini, S., Pellegrini, M., Gareri, J., Koren, G., Garcia-Algar, O., Vall, O., Vagnarelli, F., Zuccaro, P. and Marchei, E. (2008). Liquid chromatography–tandem mass spectrometry for fatty acid ethyl esters in meconium: Assessment of prenatal exposure to alcohol in two European cohorts. *Journal of pharmaceutical and biomedical analysis*, 48(3): 927-933.
- Pitt, J. J. (2009). Principles and applications of liquid chromatography-mass spectrometry in clinical biochemistry. *The Clinical Biochemist Reviews. The Australian Association of Clinical Biochemistry*, 30(1): 19-34.
- Prakash, A. 2001. *Antioxidant Activity*. Medallion Laboratories Analytical Progress.19(2):1-6
- Pranoto, Y., Anggrahini, S. and Efendi, Z. (2013). Effect of natural and *Lactobacillus plantarum* fermentation on in-vitro protein and starch digestibility of sorghum flour. *Food Bioscience*, 2: 46-52.
- Prieto, J. M. (2012). Procedure: Preparation of DPPH Radical, and antioxidant scavenging assay. *DPPH Microplate Protocol*, 7-9.
- Prior, R. L., Wu, X. and Schaich, K. (2005). Standardized methods for the determination of antioxidant capacity and phenolics in foods and dietary supplements. *Journal of Agricultural and Food Chemistry*, 53: 4290-4302.
- Quinto, E. J., Jiménez, P., Caro, I., Tejero, J., Mateo, J. and Girbés, T. (2014). Probiotic lactic acid bacteria: A review. *Food and Nutrition Sciences*, 5(18): 1765.
- Rajeh, M. A. B., Zuraini, Z., Sasidharan, S., Latha, L. Y. and Amutha, S. (2010). Assessment of *Euphorbia hirta* L. leaf, flower, stem and root extracts for their antibacterial and antifungal activity and brine shrimp lethality. *Molecules*, 15(9): 6008-6018.
- Rajendran, P., Bharathidasan, R., & Sureshkumar, K. (2017). GC-MS analysis of phyto-components in raw and treated sugarcane juice. *International Journal of Current Microbiology and Applied Sciences*, 6(7): 51-61.
- Ramli, S., Radu, S., Shaari, K. and Rukayadi, Y. (2017). Antibacterial activity of ethanolic extract of *Syzygium polyanthum* L. (*Salam*) leaves against foodborne pathogens and application as food sanitizer. *Biomed Research International*, 2017.
- Randhir, R., Lin, Y. T. and Shetty, K. (2004). Phenolics, their antioxidant and antimicrobial activity in dark germinated *fenugreek* sprouts in response to peptide and phytochemical elicitors. *Asia Pacific Journal of Clinical Nutrition*, 13(3): 295-307.

- Ranney, A. P., & Ziemann, P. J. (2016). Identification and quantification of oxidized organic aerosol compounds using derivatization, liquid chromatography, and chemical ionization mass spectrometry. *Aerosol Science and Technology*, 51(3): 342–353.
- Ravichandran, Subramaniyam & R, Vimala. (2012). Solid state and submerged fermentation for the production of bioactive substances: a comparative study. *International Journal of Science and Nature*, 3: 480-486.
- Reller, L. B., Weinstein, M., Jorgensen, J. H. and Ferraro, M. J. (2009). Antimicrobial susceptibility testing: A review of general principles and contemporary practices. *Clinical Infectious Diseases*, 49(11): 1749-1755.
- Rios, J. L., Recio, M. C. and Villar, A. (1988). Screening methods for natural products with antimicrobial activity: A review of the literature. *Journal of Ethnopharmacology*, 23: 127-149.
- Rockland, L. B. and Stewart, G. F. (Eds.). (2013). *Water activity: influences on food quality: a treatise on the influence of bound and free water on the quality and stability of foods and other natural products*. Academic Press.
- Roemantyo. and Zuhud, E. A. M. (2001). *Pangium edule* Reinw. In: Van Valkenburg, J. L. C. H. and Bunyapraphatsara, N. (Editors): Plant Resources of South-East Asia (PROSEA), No 12(2): *Medicinal and poisonous plants*. (PROSEA) Foundation, Bogor, Indonesia. Database record: prota4u.org/prosea/view.aspx?id=1218
- Romlah, E. (1992). Mempelajari perubahan aktivitas antioksidan dan lemak selama fermentasi daging biji *picung* (*Pangium edule* Reinw.). *UT-Food Science and Technology*.
- Rukayadi, Y. and Hwang, J. K. (2006). *In vitro* activity of *xanthorrhizol* against *Streptococcus mutans* biofilms. *Letters in Applied Microbiology*, 42: 400-404.
- Rukayadi, Y., Lau, K. Y., Zainin, N. S., Zakaria, M. and Abas, F. (2013). Screening of antimicrobial activity of tropical edible medicinal plant extracts against five standard microorganisms for natural food preservatives. *International Food Research Journal*, 20(5): 2905-2910.
- Rukayadi, Y., Shim, J. S. and Hwang, J. K. (2008). Screening of Thai medicinal plants for anticandidal activity. *Mycoses*, 51: 308-312.
- Safefood 360. (2014). Water activity in foods. Retrieved from www.safefood360.com
- Sahlin, P. and Nair, B. M. (2012). Production of organic acids, titratable acidity and pH-development during fermentation of cereal flours. *International Journal of Fermented Foods*, 1(1): 15-32.
- Samudry, E. G., Sukainah, A. and Mustarin, A. (2018). Analysis of the quality of *kluwek* (*Pangium edule* Reinw.) fermented using soil and ash husk. *Jurnal Pendidikan Teknologi Pertanian*, 3(1): 25-33.

- Sánchez-Moreno, C., Larrauri, J. A. and Saura-Calixto, F. (1998). A procedure to measure the antiradical efficiency of polyphenols. *Journal of the Science of Food and Agriculture*, 76(2): 270-276.
- Sanchez-Rabaneda, F., Juregui, O., Lamuela-Raventos, R. M., Viladomat, F., Bastida, J. and Codina, C. (2004). Qualitative analysis of phenolic compounds in apple pomace using liquid chromatography coupled to mass spectrometry in tandem mode. *Rapid Communications in Mass Spectrometry. RCM*, 18(5): 553-563.
- Santoso, M., Yamaguchi, T., Matoba, T. and Takamura, H. (2014). Effects of pit fermentation and thermal cooking process on the antioxidant activity and components of *Pangium edule* seeds. *Journal of Cookery Science Japan*, 47(4): 202-213.
- Santos-Sánchez, N. F., Salas-Coronado, R., Villanueva-Cañongo, C. and Hernández-Carlos, B. (2019). Antioxidant compounds and their antioxidant mechanism. *In Antioxidants*. IntechOpen. 1-29
- Sarkar, P. K., and Nout, M. R. (Eds.). (2014). Handbook of indigenous foods Involving alkaline fermentation. CRC Press.
- Sarker, S. D. and Nahar, L. (2012). Natural products isolation: methods in molecular biology. *Springer Science and Business Media*, 864: 33-34.
- Sasidharan, S., Chen, Y., Saravanan, D., Sundram, K. M. and Latha, L. Y. (2011). Extraction, isolation and characterization of bioactive compounds from plants' extracts. *African Journal of Traditional, Complementary and Alternative Medicines*, 8(1): 1– 10
- Sathe, A. Y. (1999). *A first course in food analysis*. New Age International (P) Limited Publisher. New Delhi: 1-19
- Schwan, R. F. and Wheals, A. E. (2004). The microbiology of cocoa fermentation and its role in chocolate quality. *Critical Reviews in Food Science and Nutrition*, 44(4): 205-221.
- Sen, A., Dhavan, P., Shukla, K. K., Singh, S. and Tejavathi, G. (2012). Analysis of IR, NMR and antimicrobial activity of β -sitosterol isolated from *Momordica charantia*. *Science Secure Journal of Biotechnology*, 1(1): 9-13.
- Sendra, J. M., Sentandreu, E. and Navarro, J. L. (2006). Reduction kinetics of the free stable radical 2,2-diphenyl-1-picrylhydrazyl (DPPH•) for determination of the antiradical activity of citrus juices. *European Food Research and Technology*, 223:615-624.
- Shahidi, F. and Ambigaipalan, P. (2015). Phenolics and polyphenolics in foods, beverages and spices: Antioxidant activity and health effects – A review. *Journal of Functional Foods*, 18: 820-897
- Shahidi, F. and Zhong, Y. (2005). *Antioxidants: Regulatory status. Bailey's industrial oil and fat products*. Grice HC. 1986
- Shapton, D. A. and Shapton, N. F. (1993). *Principles and Practices for the Safe Processing of Foods*. Chapter 8. Woodhead Publishing. 220-253.

- Shimadzu Corporation. (2019). *Fundamental guide to liquid chromatography mass spectrometry (LCMS)*. Shimadzu Excellence in Science. 1-66. Retrieved from <https://www.shimadzu.com/an/lcms/support/fundamental/index.html>
- Skoog, D. A., Holler, F. J. and Crouch, S. R. (2017). *Principles of instrumental analysis*. 6th Edition. Cengage Learning.
- Smith-Palmer, A., Stewart, J. and Fyfe, L. (1998). Antimicrobial properties of plant essential oils and essences against five important food-borne pathogens. *Letters in Applied Microbiology*, 26(2): 118-122.
- Sneddon, J., Masuram, S. and Richert, J. C. (2007). Gas chromatography-mass spectrometry-basic principles, instrumentation and selected applications for detection of organic compounds. *Analytical Letters*, 40(6): 1003-1012.
- Soni, S. and Dey, G. (2014). Perspectives on global fermented foods. *British Food Journal*, 116(11): 1767-1787.
- Soobrattee, M. A., Neergheen, V. S., Luximon-Ramma, A., Aruoma, O. I. and Bahorun, T. (2005). Phenolics as potential antioxidant therapeutic agents: mechanism and actions. *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis*, 579(1): 200-213.
- Souza, E. L., Stamford, T. L. M. and Lima, E. D. O. (2006). Sensitivity of spoiling and pathogen food-related bacteria to *Origanum vulgare* L. (Lamiaceae) essential oil. *Brazilian Journal of Microbiology*, 37(4): 527-532.
- Spacil, Z., Novakova, L. and Solich, P. (2010). Comparison of positive and negative ion detection of tea catechins using tandem mass spectrometry and ultrahigh performance liquid chromatography. *Food Chemistry*, 123: 535-541.
- Spanova, M., & Daum, G. (2011). Squalene–biochemistry, molecular biology, process biotechnology, and applications. *European Journal Of Lipid Science And Technology*, 113(11): 1299-1320.
- Steinkraus, K. H. (1993). Comparison of fermented foods of the East and West. In *Fish Fermentation Technology*. Edited by Lee, C. H. and Steinkraus, K. H., Reilly PJ. Tokyo: *United Nations University Press*, 1–12.
- Steinkraus, K. H. (2002). Fermentations in world food processing. *Comprehensive Reviews in Food Science and Food Safety*, 1(1): 23-32.
- Steinkraus, K. H. (2018). Handbook of Indigenous Fermented Foods. Revised and expanded. CRC Press. *Food Science and Technology (USA)*, 73: 76-79.
- Sultana, B., Anwar, F. and Ashraf, M. (2009). Effect of extraction solvent/technique on the antioxidant activity of selected medicinal plant extracts. *Molecules*, 14(6): 2167-2180.
- Sun, C., Wu, Z., Wang, Z. and Zhang, H. (2015). Effect of ethanol/water solvents on phenolic profiles and antioxidant properties of Beijing propolis extracts. *Evidence-Based Complementary and Alternative Medicine*, 2015: 1-9.
- Supardy, R., Afifah, N., Ibrahim, D., Sulaiman, S. F. and Zakaria, N. Z. (2012). Inhibition of *Klebsiella pneumoniae* ATCC 13883 by hexane extract of *Halimeda*

discoidea (Decaisne) and the identification of its potential bioactive compounds. *Journal of Microbiology and Biotechnology*, 22(6): 872-881.

- Surono, I. S. (2016). Ethnic fermented foods and beverages of Indonesia. In *Ethnic Fermented Foods and Alcoholic Beverages of Asia*. Springer, New Delhi. 341-382
- Suyanto, P. (2012). The antioxidant activity of the extract of *Pangium edule* Reinw. (Keluak) seeds in cooked ground turkey. *University of Illinois*
- Swamy, M. K., Sinniah, U. R. and Akhtar, M. S. (2015). *In vitro* pharmacological activities and GC-MS analysis of different solvent extracts of *Lantana camara* leaves collected from tropical region of Malaysia. *Evidence-Based Complementary and Alternative Medicine*, 2015: 1-9.
- Swamy, T. A., Obey, J. and Mutuku, N. C. (2013). Phytochemical analysis of *Vernonia adoensis* leaves and roots used as a traditional medicinal plant in Kenya. *International Journal of Pharmaceutical and Biological*, 3: 46-52.
- Tahmouzi, S. (2014). Optimization of polysaccharides from *Zagros oak* leaf using RSM: antioxidant and antimicrobial activities. *Carbohydrate Polymers*, 106: 238-246.
- Tajkarimi, M. M., Ibrahim, S. A., & Cliver, D. O. (2010). Antimicrobial herb and spice compounds in food. *Food Control*, 21(9), 1199–1218.
- Tamang, J. P., Watanabe, K. and Holzapfel, W. H. (2016). Review: Diversity of microorganisms in global fermented foods and beverages. *Frontiers in Microbiology*, 7.
- Tamang, J. P. and Kailasapathy, K. (2010). *Fermented Foods and Beverages of The World*. Florida: CRC Press.
- Taylor, R. S., Manandhar, N. P. and Towers, G. H. N. (1995). Screening of selected medicinal plants of Nepal for antimicrobial activities. *Journal of Ethnopharmacology*, 46(3): 153-159.
- Terefe, N. S. (2016). *Food Fermentation*. CSIRO Food and Nutrition, Werribee, VIC, Australia: Reference Module in Food Science. Werribee, Australia: Elsevier Inc.
- Terpinc, P., Polak, T., Šegatin, N., Hanzlowsky, A., Ulrih, N. P. and Abramovič, H. (2011). Antioxidant properties of 4-vinyl derivatives of hydroxycinnamic acids. *Food Chemistry*, 128(1): 62-69.
- Tille, P. M. (2014). *Bailey and Scott's Diagnostic Microbiology*, 13: 88. Elsevier Publication.
- Tortora, G. J., Funke B. R. and Case C. L. (2015). *Microbiology: An Introduction*. San Francisco: Pearson Benjamin Cummings.
- Tripoli, E., La Guardia, M., Giammanco, S., Di Majo, D. and Giammanco, M. (2007). Citrus flavonoids: Molecular structure, biological activity and nutritional properties: A review. *Food Chemistry*, 104(2): 466-479.
- Troller, J. A. and Christian, J. H. B. (1978). Microbial growth. *Water activity and Food*, 86-102.

- Trusheva, B., Trunkova, D. and Bankova, V. (2007). Different extraction methods of biologically active components from propolis: A preliminary study. *Chemistry Central Journal*, 1(1): 13.
- Tsao, G. T. (1999). *Recent Progress in Bioconversion of Lignocellulosics 75* (pp. 78). Germany: Springer Science & Business Media.
- Tsuzuki, T., Tokuyama, Y., Igarashi, M. and Miyazawa, T. (2004). Tumor growth suppression by α -eleostearic acid, a linolenic acid isomer with a conjugated triene system, via lipid peroxidation. *Carcinogenesis*, 25(8): 1417-1425.
- Tung, Y. T., Cheng, K. C., Ho, S. T., Chen, Y. L., Wu, T. L., Hung, K. C. and Wu, J. H. (2011). Comparison and characterization of the antioxidant potential of 3 wild grapes—*Vitis thunbergii*, *V. flexuosa*, and *V. kelungeusis*. *Journal of Food Science*, 76:701-706.
- Uba, A., Abdullahi, M. I., Yusuf, A. J., Ibrahim, Z. Y., Lawal, M., Nasir, I. and Abdullahi, F.T. (2015). Mineral profile, proximate and amino acid composition of three dates. *Der Pharma Chemica Journal*, 7(5): 48-53.
- Ulusoy, S., Bolgelmez-Tinaz, G. and Secilmis-Canbay, H. (2009). Tocopherol, carotene, phenolic contents and antibacterial properties of rose essential oil, hydrosol and absolute. *Current Microbiology*, 59(5): 554-8.
- Uranga, C. C., Beld, J., Mrse, A., Córdova-Guerrero, I., Burkart, M. D., & Hernández-Martínez, R. (2016). Data from mass spectrometry, NMR spectra, GC-MS of fatty acid esters produced by *Lasiodiplodia theobromae*. *Data in Brief*, 8: 31–39.
- Useful Tropical Plants. (2019). *Pangium edule*. *Tropical Plants Database*, Ken Fern. tropical.theferns.info. June 16, 2019. Retrieved from: <tropical.theferns.info/viewtropical.php?id=Pangium+edule>
- Valgas, C., Souza, S. M. D., Smânia, E. F. and Smânia Jr, A. (2007). Screening methods to determine antibacterial activity of natural products. *Brazilian Journal of Microbiology*, 38(2): 369-380.
- Van Boekel, M. A. J. S. (2001). Kinetic aspects of the Maillard reaction: A critical review. *Food Nahrung*, 45(3): 150-159.
- Van Bogaert, I. N., Groeneboer, S., Saelens, K. and Soetaert, W. (2011). The role of cytochrome P450 monooxygenases in microbial fatty acid metabolism. *The FEBS Journal*, 278(2): 206-221
- Van Gadow, A., Joubert, E. and Hannsman, C. T. (1997). Comparison of the antioxidant activity of aspalathin with that of other plant phenols of rooibos tea (*Aspalathus linearis*), tocoferol, BHT, and BHA. *Journal of Agricultural and Food Chemistry*, 45: 632-638.
- Ventaka raman, B. Samuel, L. A., Pardha Saradhi, M., Narashimha Rao B., Naga Vashmi Krishna A., Sudhakar, M. and Radhakrishnan, T. M. (2012). Antibacterial, antioxidant activity and GCMS analysis *Eupatorium odoratum*. *Asian Journal of Pharmaceutical and Clinical Research*, 5(2): 99-106.

- Verzelloni, E., Tagliacucchi, D. and Conte, A. (2007). Relationship between the antioxidant properties and the phenolic and flavonoid content in traditional balsamic vinegar. *Food Chemistry*, 105: 564-571.
- Wang, D., Zhang, L., Huang, X., Wang, X., Yang, R., Mao, J., Wang, X., Wang, X., Zhang, Q. and Li, P. (2018). Identification of nutritional components in black sesame determined by widely targeted metabolomics and traditional Chinese medicines. *Molecules*, 23(5): 1180.
- Welch, R. A. (2006). The genus *Escherichia*. Proteobacteria: Gamma Subclass. *The Prokaryotes*, 6: 60-71
- Westmann, A. (2018). Xylose Lysine Deoxycholate (XLD) agar- principle, composition, preparation and colony characteristics. In Biology Medical Laboratory Geeks. Retrieved from <https://mltgeeks.com/xylose-lysine-deoxycholate-xld-agar-principle-composition-preparation-and-colony-characteristics/>
- White, R. L., Burgess, D. S., Manduru, M. and Bosso, J. A. (1996). Comparison of three different in vitro methods of detecting synergy: time-kill, checkerboard, and E test. *Antimicrobial Agents and Chemotherapy* (AAC), 40: 1914-1918.
- Wiley, J. M., Sherwood, L. M. and Woolverton, C. J. (2008). *Prescott, Harley and Klein's Microbiology*, 134-135. New York: McGraw Hill Inc.
- Wirthensohn, M. G., Chin, W. L., Franks, T. K., Baldock, G. A., Ford, C. M. and Sedgley, M. (2008). Characterising the flavour phenotypes of almond (*Prunus Dulcis* Mill.) kernels. *Journal of Horticultural Science and Biotechnology*. 83: 462-468.
- Witkowska, A. M., Hickey, D. K., Alonso-Gomez, M. and Wilkinson, M. (2013). Antimicrobial activity of commercial herb and spices extracts against selected foodborne bacteria. *Journal of Food Research*, 2(4): 37-54.
- Wolfender, J. L., Ndjoko, K. and Hostettmann, K. (2003). Liquid chromatography with ultraviolet absorbance–mass spectrometric detection and with nuclear magnetic resonance spectrometry: a powerful combination for the on-line structural investigation of plant metabolites. *Journal of Chromatography A*, 1000 (1-2):437-455.
- Wongwiwat, P. and Wattanachant, S. (2015). Quality changes of chicken meat jerky with different sweeteners during storage. *Journal of Food Science and Technology*, 52(12): 8329-8335.
- Xu, J., Tang, J., Jin, Y., Song, J., Yang, R., Sablani, S. S., & Zhu, M.-J. (2018). High temperature water activity as a key factor influencing survival of *Salmonella* Enteritidis PT30 in thermal processing. *Food Control*. 11: 54.
- Yang, M., Shen, Q., Li, L., Huang, Y. and Cheung, H. (2015). Phytochemical profiles, antioxidant activities of functional herb *Abrus Cantoniensis* and *Abrus Mollis*. *Food Chemistry*, 177(0): 304-312.
- Yu, L. (2001). Free radical scavenging properties of conjugated linoleic acids. *Journal of Agricultural and Food Chemistry*, 49(7): 3452-3456.

- Yusoff, N. A. H., Noor, N. F. and Rukayadi, Y. (2015). Effects of *Cosmos caudatus* Kunth. (*Ulam raja*) extract on microflora in raw chicken meat. *International Journal of Current Microbiology and Applied Sciences*, 4(2): 426-435.
- Zainin, N. S., Lau, K. Y., Zakaria, M., Son, R., Abdull Razis, A. F. and Rukayadi, Y. (2013). Antibacterial activity of *Boesenbergia rotunda* (L.) Mansf. A. extract against *Escherichia coli*. *International Food Research Journal*, 20(6): 3319-3323.
- Zapotoczny, P., Kawałko, T. and Bakier, S. (2010). Determination of the physical characteristics of food raw materials by spectrophotometry-the example of honey. *Technical Sciences*, 13: 40-52.
- Zarinah, Z., Maaruf, A. G., Nazaruddin, R., Wong, W. W. W. and Xuebing, X. (2014). Antioxidant, antimicrobial activity, and *in-vitro* cytotoxicity screening study of pili nut oil. *International Food Research Journal*, 21(1): 309-316.
- Zhang, Q. W., Lin, L. G. and Ye, W. C. (2018). Techniques for extraction and isolation of natural products: A comprehensive review. *Chinese Medicine*, 13(1): 20.
- Zhang, Q., Zhang, J., Shen, J., Silva, A., Dennis, D. A. and Barrow, C. J. (2006). A simple 96-well microplate method for estimation of total polyphenol content in seaweeds. *Journal of Applied Phycology*, 18(3-5): 445-450.
- Zhu, Y. P., Fan, J. F., Cheng, Y. Q. and Li, L. T. (2007). Improvement of antioxidant activity of Chinese traditional fermented Okara (*Meitauza*) using *Bacillus subtilis* B2. *Food Control*, 19: 654-661.
- Zinder, S. H. and Dworkin, M. (2013). Morphological and Physiological Diversity. In Rosenberg, E. (Ed.). *The Prokaryotes*, 89-122. Berlin, Heidelberg: Springer.
- Zulkarnay, Z., Shazwani, S., Ibrahim, B., Jurimah, A. J., Ruzairi, A. R. and Zaridah, S. (2015). An overview on pH measurement technique and application in biomedical and industrial process. In *2015 2nd International Conference on Biomedical Engineering (ICoBE)*, 1-6. IEEE.

BIODATA OF STUDENT

The student, Nur Hafizah Binti Mustaffer was born on 22nd March 1994 at Alor Setar, Kedah. She is the daughter of Mustaffer Bin Abdullah and Zubaidah Binti Mohd Yusoff. She started her primary education in Sekolah Kebangsaan Langgar, Kedah and continued her secondary education at Sekolah Menengah Kebangsaan Agama Yan. During form 4, she changed to Sekolah Menengah Sains Pokok Sena, Kedah. Then, she continued her foundation study at UiTM Puncak Alam study in the Science stream. After that, she furthered her study at Universiti Putra Malaysia (UPM) by taking Bachelor of Food Science and Technology and graduated in 2017. Currently, she is continued her study in Master of Science under the Dual-Degree program between UPM and Kasetsart University (KU), Thailand. She is taking master in the field of Food Security and Climate Change under the supervision of Assoc. Prof. Dr. Yaya Rukayadi at Faculty of Food Science and Technology, UPM and master in the Food Science and Technology under the supervision of Dr. Kullanart Tongkhao in Department Food Science and Technology at Faculty of Agro-Industry, KU. The following are the list of publication and thesis involved during her course of study.

Thesis:

1. Bachelor of Food Science and Technology:

Quality Evaluation of Physical Properties, Nutritional Composition and Antioxidant Activity of Six Different Palm Dates (*Phoenix Dactylifera* L.) Varieties from Saudi Arabia

2. Master of Food Security and Climate Change:

Physicochemical, Microbiological and Biological Properties of Fermented Kepayang (*Pangium Edule* Reinw.) Seeds

LIST OF PUBLICATIONS

Abdul-Hamid, N. A., Mustaffer, N. H., Maulidiani, M., Mediani, A., Ismail, I. S., Tham, C. L., Shadid, K. and Abas, F. (2018). Quality evaluation of the physical properties, phytochemicals, biological activities and proximate analysis of nine Saudi date palm fruit varieties. *Journal of the Saudi Society of Agricultural Sciences*.

Mustaffer, N. H., Rosni, N. K., Ramli, N. S., Rukayadi, Y. (2020). Changes in physicochemical properties and antioxidant activity of fermented Kepayang (*Pangium edule* R.) seeds. *Food Research*. (Manuscript in preparation for publication).





UNIVERSITI PUTRA MALAYSIA

STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

ACADEMIC SESSION : _____

TITLE OF THESIS / PROJECT REPORT :

PHYSICOCHEMICAL, MICROBIOLOGICAL AND BIOLOGICAL PROPERTIES OF
FERMENTED KEPAYANG (*Pangium edule* Reinw.) SEEDS

NAME OF STUDENT : NUR HAFIZAH BINTI MUSTAFFER

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

1. This thesis/project report is the property of Universiti Putra Malaysia.
2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as :

*Please tick (✓)

CONFIDENTIAL

(Contain confidential information under Official Secret Act 1972).

RESTRICTED

(Contains restricted information as specified by the organization/institution where research was done).

OPEN ACCESS

I agree that my thesis/project report to be published as hard copy or online open access.

This thesis is submitted for :

PATENT

Embargo from _____ until _____
(date) (date)

Approved by:

(Signature of Student)
New IC No/ Passport No.:

Date :

(Signature of Chairman of Supervisory Committee)
Name:

Date :

[Note : If the thesis is CONFIDENTIAL or RESTRICTED, please attach with the letter from the organization/institution with period and reasons for confidentially or restricted.]