



UNIVERSITI PUTRA MALAYSIA

UNTARGETED TANDEM MASS SPECTROMETRY-BASED MOLECULAR NETWORK AND ZEBRAFISH EMBRYOTOXICITY AND TERATOGENIC EFFECTS OF *Christia vespertilionis* (L. F.) Bakh. F. LEAF EXTRACT

ANIS IRFAN BINTI NORAZHAR

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By

ANIS IRFAN BINTI NORAZHAR

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirements for the Degree of Master of Science**

August 2020

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

UNTARGETED TANDEM MASS SPECTROMETRY-BASED MOLECULAR NETWORK AND ZEBRAFISH EMBRYOTOXICITY AND TERATOGENIC EFFECTS OF *Christia vespertilionis* (L. F.) Bakh. F. LEAF EXTRACT

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August 2020

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Christia vespertilionis (L.f.) Bakh. f., is a non-climbing ornamental plant with unique butterfly-shaped leaves, hence its vernacular name ‘butterfly wing’ or ‘pokok rerama’ in Malay. In Malaysia, the green-leaved variety gained popularity in recent years due to testimonial reports by local users and distributors for its medicinal uses, which included among others, as a cure for cancer. Despite these popular uses, there is very limited information on the chemical constituents of the species, presenting a significant gap in the cheminformatics of the species. Additionally, more information on the safety profile of the plant is required, towards safeguarding consumer safety. Therefore, the primary objectives of the present study were to establish the chemical profile of the green-leaved variety of the plant, specifically the leaf extract and to evaluate the extract for potential toxicity effects. The metabolite profile of the leaf methanolic extract was established by deploying untargeted tandem mass spectrometry-based molecular networking approach. The toxic effects of the extract involving mortality rate, heartbeat rate, hatchability rate, and spontaneous tail coliling as well as the teratogenic effects were determined on zebrafish (*Danio rerio*) embryos, as the *in-vivo* assay model. Zebrafish embryos at 5 hour post-fertilization (hpf) were exposed to 50, 100, 200, 400, and 800 µg/mL of the extract up to 120 hpf. The multi-informative molecular map generated for the leaf metabolome permitted the putative identification of 62 metabolites, comprising 10 C-glycosylflavones, 2 mono- and 2 di-hydroxyflavones, 1 flavone-C,O-diglycoside, 3 flavonol-3-O-glycosides, 15 phenolic acids, 1 phenethyl glycoside and 1 its derivative, 4 hydroxyjasmonic acid derivatives, 4 carotenoids, 2 chlorophylls, 3 monoacylglycerols, 1 sphingolipids, 4 amino acids, 1 nucleoside, 3 organic acids, 1 coumarin derivative and 4 fatty acid amides. As a means of structural validation, two unknown chemical constituents were targeted for isolation which subsequently resulted in their characterization as apigenin-6-C-β-D-glucoside 4'-O-α-D-apiofuranoside [2] and apigenin-6-C-β-D-[(4'',6''-O-dimalonyl)-glucoside] 4'-O-α-D-apiofuranoside [10], which were newly reported in the plant kingdom as the

new derivatives of apigenin-6-*C*- β -D-glucoside. In the toxicity assay, the median lethal concentration (LC_{50}) value of the extract was determined to be 419.84 $\mu\text{g}/\text{mL}$, which was classified as safe. Nevertheless, from evaluation of possible teratogenic effects on zebrafish embryos, it was further revealed that the extract was toxic at higher concentrations starting from 200 $\mu\text{g}/\text{mL}$ onwards with multiple signs of developmental defects. The magnitude of these defects was observed to be concentration-dependent. Moreover, no hatching and spontaneous movement of tail coiling were observed at concentrations of 400 and 800 $\mu\text{g}/\text{mL}$ due to the delayed growth and early mortality, respectively. A significant reduction in heartbeat rate was also recorded in all surviving embryos at 400 $\mu\text{g}/\text{mL}$. The present study has provided some important insights on the plant's chemical and biological characteristics which are a pre-requisite to further research towards its valorization as a candidate in phytotherapy. Further extensive studies of the leaf extract using other animal models as well as the *in-vitro* assay are required for the establishment of its safe dose for human consumption.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

SPEKTROMETRI JISIM TANDEM TAK BERTUMPU BERASASKAN RANGKAIAN MOLEKULAR DAN KESAN TOKSIK DAN TERATOGENIK EMBRIO ZEBRAFISH EKSTRAK DAUN *Christia vespertilionis* (L. F.) Bakh. F.

Oleh

ANIS IRFAN BINTI NORAZHAR

Ogos 2020

Pengerusi : Profesor Khozirah Shaari, PhD
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Christia vespertilionis (L. f.) Bakh. f., adalah tanaman hiasan yang tidak memanjat dengan daun berbentuk rama-rama yang unik, oleh itu nama vernakularnya 'sayap rama-rama' atau 'pokok rerama' dalam bahasa Melayu. Di Malaysia, varieti daun hijau memperoleh populariti dalam beberapa tahun kebelakangan ini kerana laporan testimoni oleh pengguna dan pengedar tempatan mengenai kegunaan perubatannya, yang antara lainya termasuklah sebagai penawar penyakit barah. Walaupun penggunaannya popular, terdapat maklumat yang sangat terhad tentang unsur kimia spesies ini, yang menunjukkan jurang yang ketara. Sebagai tambahan, lebih banyak maklumat mengenai profil keselamatan pokok ini diperlukan, untuk menjamin keselamatan pengguna. Oleh itu, objektif utama kajian ini adalah untuk menentukan profil sebatian kimia dari varieti daun hijau, khususnya ekstrak daun dan untuk menilai kemungkinan kesan toksik ekstrak ini. Profil sebatian kimia ekstrak metanol daun ditentukan dengan menggunakan pendekatan rangkaian molekul berasaskan spektrometri jisim tandem tak bertumpu. Kesan-kesan toksik dari ekstrak yang melibatkan kadar kematian, degupan jantung, kadar penetasan, dan gelungan spontan ekor serta kesan teratogenik ditentukan ke atas embrio zebrafish (*Danio rerio*), sebagai model uji *in-vivo*. Embrio zebrafish pada 5 jam pasca pensenyawaan (hpf) didedahkan kepada 50, 100, 200, 400, dan 800 µg/mL ekstrak sehingga 120 hpf. Peta molekul pelbagai maklumat yang dihasilkan daripada metabolom daun membolehkan pengenalpastian sebanyak 62 sebatian kimia yang terdiri daripada 10 C-glikosilflavon, 2 mono- dan 2 di-hidroksiflavon, 1 flavon C,O-glikosida, 3 flavonol-3-O-glikosida, 15 fenolik asid, 1 fenil glikosida dan 1 deravatifnya, 4 deravatif asid hidroksijasmonik, 4 karotenoid, 2 klorofil, 3 monoasilgliserol, 1 sphingolipid, 4 asid amino, 1 nukleosida, 3 asid organik, 1 derivatif kumarin dan 4 asid lemak amida. Untuk pengesahan struktur kimia, dua komponen kimia yang tidak diketahui telah disasarkan untuk pengasingan yang kemudiannya dicirikan sebagai apigenin-6-C-β-D-glukosida

4'-O- α -D-apiofuranosida [2] and apigenin-6-C- β -D-[(4'',6''-O-dimalonil)-glukosida] 4'-O- α -D-apiofuranosida [10]. Dalam ujian toksik, nilai kepekatan median maut (LC₅₀) ekstrak ditentukan sebagai 419.84 $\mu\text{g}/\text{mL}$, yang diklasifikasikan sebagai selamat. Walaupun begitu, berdasarkan penilaian kemungkinan kesan teratogenik ke atas embrio zebrafish, didapati bahawa ekstrak ini sangat toksik pada kepekatan yang tinggi bermula dari 200 $\mu\text{g}/\text{mL}$ dan seterusnya dengan pelbagai tanda kecacatan perkembangan. Magnitud kecacatan ini dilihat bergantung kepada kepekatan. Selain itu, tiada penetasan dan pergerakan spontan gegelung ekor diperhatikan pada kepekatan 400 dan 800 $\mu\text{g}/\text{mL}$ kerana pertumbuhan yang terbantut dan kematian awal, masing-masing. Kajian ini telah memberikan beberapa pandangan penting mengenai ciri-ciri kimia dan biologi pokok ini yang menjadi prasyarat untuk penyelidikan selanjutnya ke arah membangunkannya sebagai calon fitoterapi. Kajian luas yang lebih lanjut mengenai ekstrak daun ini menggunakan model haiwan lain dan juga ujian *in-vitro* diperlukan untuk menentukan dos yang selamat untuk penggunaan manusia.

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This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS

1D	One-Dimensional
2D	Two-Dimensional
¹ H NMR	Proton Nuclear Magnetic Resonance
¹³ C-NMR	Carbon -13 Nuclear Magnetic Resonance
ACN	Acetonitrile
CD ₃ OD	Deuterated Methanol
CID	Collision-Induced Dissociation
COSY	Correlation Spectroscopy
d	Doublet
DAD	Diode Array Detector
dd	Doublet of Doublet
ESI	Electrospray Ionization
FA	Formic Acid
HESI	Heated Electrospray Ionization
HMBC	Heteronuclear Multiple Bond Coherence
HPLC	High Performance Liquid Chromatography
HSQC	Heteronuclear Single Quantum Correlation
Hz	Hertz
<i>J</i>	Coupling Constant
LC-MS	Liquid Chromatography Mass Spectrometry
mg	Milligram
μg	Microgram
mL	Milliliter
min	Minute
<i>m/z</i>	Mass-to-charge ratio
nm	Nanometer
ppm	Parts per million

Prep-HPLC	Preparative High Performance Liquid Chromatography
PTFE	Polytetrafluoroethylene
RT	Retention time
s	Singlet
t	Triplet
TIC	Total Ion Chromatogram
TMS	Tetramethylsilane
UPLC-MS/MS	Ultra-Performance Liquid Chromatography-Tandem Mass Spectrometry
UV	Ultraviolet



CHAPTER 1

INTRODUCTION

1.1 Background

Since time immemorial, humans have relied on the plants as a main source of therapeutic agents for the treatment of various kinds of diseases and ailments, as well as for maintaining their health. The precious ethnomedicinal knowledge of the diverse plant resources used by the different cultures around the world has been passed on over many generations. Despite the exponential growth of the utilization of synthetic drugs in modern treatment, the practice of using herbs and herbal formulations as alternative medicine continues to be important among communities across the globe with the belief that natural forms are safer than the synthetic ones. Statistically, it has been estimated that four billion people living in developing countries (80% of the world population) still depend on herbal medicine as their primary healthcare source (Hudson *et al.*, 2018). In recent years, multiple product forms derived from plant materials such as supplements, health drinks, tonics, and teas have been marketed in order to meet consumer demands and preferences. Nevertheless, the safety of these particular plant materials as well as their derived products is not guaranteed due to insufficient scientific data concerning their safety and toxicological profiles. This is particularly important since herbal medicines are more often consumed under self-medication without a medical supervision (Pariyani *et al.*, 2015). Surprisingly, it has been reported that the consumption of herbals may also cause adverse effects such as nausea, vomiting, weakness, dizziness, hypotension, paraesthesia of the mouth and tongue, arrhythmia, and ventricular fibrillation (Hussin *et al.*, 2001). These adverse effects are strongly attributed to the phytochemical composition of the plants, in which some phytochemical constituents could also act as toxin-like substances (Chandra *et al.*, 2012). Hence, it is crucial to have proper scientific information regarding the phytochemical composition and toxicological data of the plants in order to prove their safety as an alternative medicine. In the toxicological study, the use of zebrafish (*Danio rerio*) embryos as an alternative animal model to the classical rodent model has gained attention in recent years. The new model is increasingly being applied to evaluate the toxicity profiles of chemical compounds (Jayasinghe & Jayawardena, 2019). The optical transparency of the embryos which allows direct visualization of the internal organs for assessment of toxicity endpoint as well as by having 70% homology with the human's genes have made zebrafish embryos one of the most promising animal model for studying toxicity (Hill *et al.*, 2005; Howe *et al.*, 2013).

Christia vespertilionis (L. F.) Bakh. F., (Family: Fabaceae) is popularly known as 'butterfly wing', 'mariposa' (meaning butterfly in Spanish) or pokok rerama in the Malay language. Several varieties of the small plant species are commonly grown for ornamental purposes but there has been records of its use in traditional medicine for treating diverse ailments such as tuberculosis, bronchitis, cold, muscle weakness, poor blood circulation and snake bites (Dash, 2016). The health use of the species has recently risen in popularity in Malaysia, following several testimonials from

consumers on the effectiveness of the water decoction of the green-leafed variety as an anticancer herbal remedy. These reports have raised concerns on the safety issues of the plant use and directed interests among researchers with a view to ascertain the health claims made. To date, a number of biological investigations have been carried out and the plant species has been reported to possess antioxidant (Abd Mutalib & Abd Latip, 2019), anti-proliferative (Hofer *et al.*, 2013), anti-malarial (Upadhyay *et al.*, 2013; Nguyen-Pouplin *et al.*, 2007), and cytotoxic properties (Nguyen-pouplin *et al.*, 2007; Abd Mutalib & Abd Latip, 2019). However, there still remains a huge gap on the phytochemical composition of the plant, especially on its leaf metabolome. Although initial results from the bioactivity studies indicated that the plant could prove to have therapeutic utility, the safety profile of the plant is still largely unknown. These knowledge and information gaps need to be filled and supplemented for a better understanding of its safety implications and potential applications in healthcare.

1.2 Objectives of the present study

The present study was undertaken with the primary objective of mapping the chemical space within the leaf metabolome of the plant. To achieve this goal, a comprehensive metabolite profiling of the leaf methanolic extract of the green-leafed variety of *C. vespertilionis* was thus performed using ultra-high performance liquid chromatography coupled with tandem mass spectrometry (UHPLC-MS/MS)-based molecular networking approach. From the metabolite profile obtained, MS targeted isolation was also conducted in an attempt to validate the proposed structures of the unknown chemical constituents. Furthermore, the toxicity and teratogenic effect of the plant extract also was evaluated through an *in-vivo* study using zebrafish (*Danio rerio*) embryos. The objectives are summarized as follows:

1. To profile the chemical constituents of the leaf methanolic extract of the *C. vespertilionis* (green-leafed variety) via untargeted tandem mass spectrometry-based molecular networking approach.
2. To isolate and elucidate the structures of the targeted compounds using spectroscopic techniques.
3. To evaluate the toxicity and teratogenic effects of the leaf methanolic extract towards zebrafish (*Danio rerio*) embryos.

REFERENCES

- Abad-García, B., Garmón-Lobato, S., Berrueta, L. A., Gallo, B., & Vicente, F. (2008). New features on the fragmentation and differentiation of C-glycosidic flavone isomers by positive electrospray ionization and triple quadrupole mass spectrometry. *Rapid Communications in Mass Spectrometry*, 22(12), 1834–1842.
- Abd Mutalib, N., & Abd Latip, N. (2019). Synergistic Interactions Between *Christia Vespertilionis* Leaves Extract and Chemotherapy Drug Cyclophosphamide on WRL-68 Cell Line. *Asian Journal of Pharmaceutical Research and Development*, 7(3), 109–113.
- Aguilar-mogas, A., Sales-pardo, M., Navarro, M., Guimerà, R., Yanes, O. (2017). iMet: a network-based computational tool to assist in the annotation of metabolites from tandem mass spectra. *Analytical Chemistry*, 1-20.
- Ahmad, F., Anwar, F., & Hira, S. (2016). Review on medicinal importance of fabaceae family. *Pharmacologyonline*, 3, 151–156.
- Alafiatayo, A. A., Lai, K., Syahida, A., Mahmood, M., & Shaharuddin, N. A. (2019). Phytochemical Evaluation, Embryotoxicity, and Teratogenic Effects of *Curcuma longa* Extract on Zebrafish (*Danio rerio*). *Evidence-Based Complementary and Alternative Medicine*, 1-10.
- Allard, P., Péresse, T., Bisson, J., Gindro, K., Marcourt, L., Pham, V. C., Roussi, F., Litaïdon, M., & Wolfender, J. (2016). Integration of Molecular Networking and In-Silico MS/MS Fragmentation for Natural Products Dereplication. *Analytical Chemistry*, 1-10.
- Aron, A. T., Gentry, E., McPhail, K. L., Nothias, L. F., Nothias-Esposito, M., Bouslimani, A., Petras, D. (2019). Reproducible Molecular Networking Of Untargeted Mass Spectrometry Data Using GNPS. Retrieved February 7, 2020, from https://chemrxiv.org/articles/Reproducible_Molecular_Networking_Of_Untargeted_Mass_Spectrometry_Data_Using_GNPS_/9333212/1
- Audoïn, C., Zampalégré, A., Blanchet, N., Giuliani, A., Roulland, E., Laprêvotte, O., & Genta-Jouve, G. (2018). MS/MS-Guided Isolation of Clarinoside, a New Anti-Inflammatory Pentalogin Derivative. *Molecules*, 23, 1-9.
- Bajpai, V.K., Alam, B., Quan, K.T., Choi, H., An, H., Ju, M., Lee, S., Huh, Y.S., Han, Y., Na, M. (2018). Cytotoxic properties of the anthraquinone derivatives isolated from the roots of *Rubia philippinensis*. *BMC Complementary and Alternative Medicine*, 18(1), 1-6.
- Balls, M. (2002). Future Improvements: Replacement In Vitro Methods. *ILAR Journal*, 43, S69–S73.

- Bambino, K., & Chu, J. (2017). Zebrafish in Toxicology and Environmental Health. *Curr Top Dev Biol*, 124, 331–367.
- Barham, J. M. (1996). *Christia vespertilionis* var. *vespertilionis* Leguminosae. *Curtis's Bot. Mag*. 13 (1), 19-21.
- Bezerra, A. G., Negri, G., Duarte-Almeida, J. M., Smaili, S. S., & Carlini, E. A. (2016). Phytochemical analysis of hydroethanolic extract of *Turnera diffusa* Willd and evaluation of its effects on astrocyte cell death. *Einstein (São Paulo)*, 14(1), 56–63.
- Caballero M. V., Candiracci M. (2018). Zebrafish as screening model for detecting toxicity and drugs efficacy. *J Unexplored Med Data*, 1-14.
- Chadburn, H. (2012). *Christia vespertilionis*. *The IUCN Red List of Threatened Species*2012: e.T19892028A20042444.
- Chandra, S.J, Sandhya, S., K, V., Banji, D., Sudhakar, K., & Rsnakk, C. (2012). Plant toxins-useful and harmful effects. *Hygeia Journal for Drug and Medicines*, 4(1),79-90.
- Chen, Y., Yu, H., Wu, H., Pan, Y., Wang, K., Jin, Y., & Zhang, C. (2015). Characterization and Quantification by LC-MS/MS of the Chemical Components of the Heating Products of the Flavonoids Extract in Pollen *Typhae* for Transformation Rule Exploration. *Molecules*, 20(10), 18352–18366.
- Chua, Y. G., Bloodworth, B. C., Leong, L. P., & Li, S. F. Y. (2014). Metabolite profiling of edible bird's nest using gas chromatography/mass spectrometry and liquid chromatography/mass spectrometry. *Rapid Communications in Mass Spectrometry*, 28(12), 1387–1400.
- Clark, T. S., Pandolfo, L. M., Marshall, C. M., Mitra, A. K., & Schech, J. M. (2018). Body Condition Scoring for Adult Zebrafish (*Danio rerio*). *Journal of the American Association for Laboratory Animal Science*, 57 (6), 698-702.
- Cuyckens, F., & Claeys, M. (2004). Mass spectrometry in the structural analysis of flavonoids. *Journal of Mass Spectrometry*, 39(1), 1–15.
- Da Silva, R. R., Wang, M., Nothias, L.-F., van der Hoof, J. J. J., Caraballo-Rodríguez, A. M., Fox, E., Balunas, M.J., Klassen, J. L., Lopes, N. P., & Dorrestein, P. C. (2018). Propagating annotations of molecular networks using in silico fragmentation. *PLOS Computational Biology*, 14(4), 1-26.
- Dash, G. (2016). An appraisal of *Christia vespertilionis* (L. F.) bakh. F.: A promising medicinal plant. *International Journal of Pharmacognosy and Phytochemical Research*, 8 (6), 1037-1039.

- De Oliveira, G., Neto, F. C., Demarque, D. P., Pereira-Junior, J. A., Filho, R. C. S. P., R., de Melo, Almeida, J. R. G., Lopes, J. L. C., Lopes, N. P. (2016). Dereplication of Flavonoid Glycoconjugates from *Adenocalymma imperatoris-maximiliani* by Untargeted Tandem Mass Spectrometry-Based Molecular Networking. *Planta Medica*, 83(07), 636–646.
- Delgado-Povedano, M. del M., Sánchez de Medina, V., Bautista, J., Priego-Capote, F., & Luque de Castro, M. D. (2016). Tentative identification of the composition of *Agaricus bisporus* aqueous enzymatic extracts with antiviral activity against HCV: A study by liquid chromatography–tandem mass spectrometry in high resolution mode. *Journal of Functional Foods*, 24, 403–419.
- Doerge, D. R., Divi, R. L., Nation, D. I., & Sed, T. A. (1995). Porphyrin n-cation and protein radicals in peroxidase catalysis and inhibition by anti-thyroid chemicals. *Xenobiotica*, 2(7), 761–767.
- Doke, S. K., & Dhawale, S. C. (2015). Alternatives to animal testing: A review. *Saudi Pharmaceutical Journal*, 23(3), 223–229.
- Du, W. Y., Xiao, Y., Yao, J. J., Hao, Z., & Zhao, Y. B. (2017). Involvement of NADPH oxidase in high-dose phenolic acid-induced pro-oxidant activity on rat mesenteric venules. *Experimental and therapeutic medicine*, 13(1), 17–22.
- Ducharme, N. A., Reif, D. M., Gustafsson, J.-A., & Bondesson, M. (2015). Comparison of toxicity values across zebrafish early life stages and mammalian studies: Implications for chemical testing. *Reproductive Toxicology*, 55, 3–10.
- El Sayed, A. M., Ezzat, S. M., El Naggar, M. M., & El Hawary, S. S. (2016). *In-vivo* diabetic wound healing effect and HPLC–DAD–ESI–MS/MS profiling of the methanol extracts of eight Aloe species. *Revista Brasileira de Farmacognosia*, 26(3), 352–362.
- Emami, S., & Dadashpour, S. (2015). Current developments of coumarin-based anti-cancer agents in medicinal chemistry. *Eur. J. Med. Chem*, 102, 611–630.
- EU. 2010. Directive 2010/63/EU of the European Parliament and of the Council of 22 September 2010 on the protection of animals used for scientific purposes *Official Journal of the European Union*.
- Frank, A. M., Bandeira, N., Shen, Z., Tanner, S., Briggs, S. P., Smith, R. D., & Pevzner, P. A. (2008). Clustering millions of tandem mass spectra. *Journal of proteome research*, 7(1), 113–122.
- Finney, D.J. (1971). *Probit Analysis*, Cambridge University Press, London.

Figure 2.2. Some commercial products made from *Christia vespertilionis* available on the Malaysian market. Downloaded from Tok Guru Pondok, by Ustaz Nik Ab Rahim bin Tuan Guru Dato' Nik Abdul Aziz, 2018, retrieved from <http://www.tokgurupondok.com.my>.

- Flora of China. (2010). *CHRISTIA Moench, Suppl. Meth. 39. 1802, 10*, 289-290. Retrieved from <http://www.eFloras.org>
- Gao, T., Yao, H., Song, J., Liu, C., Zhu, Y., Ma, X., Pang, X., Xu, H., & Chen, S. (2010). Identification of medicinal plants in the family Fabaceae using a potential DNA barcode ITS2. *Journal of Ethnopharmacology*, 130(1), 116–121.
- Gao, X.-P., Feng, F., Zhang, X.-Q., Liu, X.-X., Wang, Y.-B., She, J.-X., He, Z.-H., & He, M.-F. (2014). Toxicity Assessment of 7 Anticancer Compounds in Zebrafish. *International Journal of Toxicology*, 33(2), 98–105.
- Garnock-Jones, P.J. (1983). Note on some folklore drugs and remedies of the Lau Group, Fiji. *South Pac J Nat Sci*, 4, 4-8.
- Gaitan, E., Lindsay, R. H., Reichert, R. D., Ingbar, S. H., Cooksey, R. C., Legan, J., Meydrech, E. F., Hill, J., Kubota, K. (1989). Antithyroid and goitrogenic effects of millet: role of C-glycosylflavones. *J. Clin. Endocrinol. Metabol.* 68(4), 707–714.
- Halili, J. F. & Quilang, J. (2011). The zebrafish embryo toxicity and teratogenicity assay. *The Philippine Biota*, 63-71.
- Hill, A. J., Teraoka, H., Heideman, W., & Peterson, R. E. (2005). Zebrafish as a Model Vertebrate for Investigating Chemical Toxicity. *Toxicological Sciences*, 86(1), 6–19.
- Hirota, B. C. K., Miyazaki, C. M. S., Mercali, C. A., Verdan, M. C., Kalegari, M., Gemin, C., Lordello, A. L. L, Miguel, M. D., Miguel, O. G. (2012). C-glycosyl flavones and a comparative study of the antioxidant, hemolytic and toxic potential of *Jatropha multifida* leaves and bark. *International Journal of Phytomedicine* 4 (1), 1-5.
- Hofer, D., Schwach, G., Tabrizi-Wizsy, N. G., Sadjak, A., Sturm, S., Stuppner, H., & Pfragner, R. (2013). *Christia vespertilionis* plant extracts as novel antiproliferative agent against human neuroendocrine tumor cells. *Oncology Reports*, 29(6), 2219–2226.
- Howe, K., Clark, M. D., Torroja, C. F., Torrance, J., Berthelot, C., Muffato, M., Matthews, L. (2013). The zebrafish reference genome sequence and its relationship to the human genome. *Nature*, 498–503.
- Hudson, A., Lopez, E., Almalki, A. J., Roe, A. L., & Calderón, A. I. (2018). A Review of the Toxicity of Compounds Found in Herbal Dietary Supplements. *Planta Medica*, 84(09/10), 613–626.
- Hussin, A. H. (2001). Adverse Effects Of Herbs And Drug-Herbal Interactions. *Malysian Journal of pharmacy*, 1(2), 39–44.
- Ismail, H. F., Hashim, Z., Soon, W. T., Rahman, N. S. A., Zainudin, A. N., & Majid, F. A. A. (2017). Comparative study of herbal plants on the phenolic and

- flavonoid content, antioxidant activities and toxicity on cells and zebrafish embryo. *Journal of Traditional and Complementary Medicine*, 7(4), 452–465.
- Jayasinghe, C. D., & Jayawardena, U. A. (2019). Toxicity Assessment of Herbal Medicine Using Zebrafish Embryos: A Systematic Review. *Evidence-Based Complementary and Alternative Medicine*, 1–17.
- Jones, W. P., & Kinghorn, A. D. (2012). Extraction of Plant Secondary Metabolites. *Natural Products Isolation*, 341–366.
- Kachlicki, P., Piasecka, A., Stobiecki, M., & Marczak, L. (2016). Structural Characterization of Flavonoid Glycoconjugates and Their Derivatives with Mass Spectrometric Techniques. *Molecules*, 21(11), 1-21.
- Kinna, G., Kolle, G., Carter, A., Key, B., Lieschke, G. J., Perkins, A., & Little, M. H. (2008). Knockdown of zebrafish *crim1* results in a bent tail phenotype with defects in somite and vascular development. *Mechanisms of Development*, 123, 277–287.
- Kumar, S., Chandra, P., Bajpai, V., Singh, A., Srivastava, M., Mishra, D. K., & Kumar, B. (2015). Rapid qualitative and quantitative analysis of bioactive compounds from *Phyllanthus amarus* using LC/MS/MS techniques. *Industrial Crops and Products*, 69, 143–152.
- Lee, H. (2005). Pharmaceutical Applications of LiquidChromatography Coupled with Mass Spectrometry (LC/MS). *Journal of LiquidChromatography & Related Technologies*, 28, 1161-1202.
- Lee, E. M., Lee, S. S., Chung, B. Y., Cho, J.-Y., Lee, I. C., Ahn, S. R., Jang, S. J., & Kim, T. H. (2010). Pancreatic Lipase Inhibition by C-Glycosidic Flavones Isolated from *Eremochloa ophiuroides*. *Molecules*, 15(11), 8251–8259.
- Lee, J. J., Saiful Yazan, L., Kassim, N. K., Che Abdullah, C. A., Esa, N., Lim, P. C., & Tan, D. C. (2020). Cytotoxic Activity of *Christia vespertilionis* Root and Leaf Extracts and Fractions against Breast Cancer Cell Lines. *Molecules*, 25(11), 1-18.
- Li, F., Janussen, D., Peifer, C., Pérez-Victoria, I., & Tasdemir, D. (2018). Targeted Isolation of Tsitsikammamines from the Antarctic Deep-Sea Sponge *Latrunculia biformis* by Molecular Networking and Anticancer Activity. *Marine Drugs*, 16(8), 1-17.
- Li, X., Lu, M., Tang, D., & Shi, Y. (2015). Composition of Carotenoids and Flavonoids in *Narcissus* Cultivars and their Relationship with Flower Color. *PLOS ONE*, 10(11), 1-14.
- Li, S., Lin, Z., Jiang, H., Tong, L., Wang, H., & Chen, S. (2016). Rapid Identification and Assignment of the Active Ingredients in Fufang Banbianlian Injection Using HPLC-DAD-ESI-IT-TOF-MS. *Journal of Chromatographic Science*, 54(7), 1225–1237.

- Li, Y.-L., Li, J., Wang, N.-L., & Yao, X.-S. (2008). *Flavonoids and a New Polyacetylene from Bidens parviflora Willd. Molecules, 13(8), 1931–1941.*
- Maes, J., Verlooy, L., Buenafe, O. E., de Witte, P. A. M., Esguerra, C. V., & Crawford, A. D. (2012). Evaluation of 14 Organic Solvents and Carriers for Screening Applications in Zebrafish Embryos and Larvae. *PLoS ONE, 7(10), e43850.*
- Martin, K.R., & Appel, C.L. (2009). Polyphenols as dietary supplements: A double-edged sword. *Dove Press Journal: Nutrition and Dietary Supplements, 2, 1-12.*
- Mekky, R. H., Contreras, M. del M., El-Gindi, M. R., Abdel-Monem, A. R., Abdel-Sattar, E., & Segura-Carretero, A. (2015). Profiling of phenolic and other compounds from Egyptian cultivars of chickpea (*Cicer arietinum* L.) and antioxidant activity: a comparative study. *RSC Advances, 5(23), 17751–17767.*
- Moser, V. C. (2011). Functional assays for neurotoxicity testing. *Toxicologic Pathology, 39(1), 36-45.*
- Muhammad, G., Hussain, M. A., Jantan, I., & Bukhari, S. N. A. (2015). Mimosa pudica L., a High-Value Medicinal Plant as a Source of Bioactives for Pharmaceuticals. *Comprehensive Reviews in Food Science and Food Safety, 15(2), 303–315.*
- Mukherjee, P. K., Kumar, V., Kumar, N. S., & Heinrich, M. (2008). The Ayurvedic medicine *Clitoria ternatea*—From traditional use to scientific assessment. *Journal of Ethnopharmacology, 120(3), 291–301.*
- Murakami, T., Kohno, K., Ninomiya, K., Matsuda, H., & Yoshikawa, M. (2001). Medicinal Foodstuffs. XXV. Hepatoprotective Principle and Structures of Ionone Glucoside, Phenethyl Glycoside, and Flavonol Oligoglycosides from Young Seedpods of Garden Peas, *Pisum sativum* L. *Chem. Pharm. Bull, 49(8) 1003—1008.*
- Murugesu, S., Uddin, Q., Ibrahim, Z., Fathamah, B., Benchoula, K., Idris, N., El-seedi, H. R. (2019). Toxicity study on *Clinacanthus nutans* leaf hexane fraction using *Danio rerio* embryos. *Toxicology Reports, 6, 1148–1154.*
- Negri, G., Santi, D. d., & Tabach, R. (2012). Chemical composition of hydroethanolic extracts from *Siparuna guianensis*, medicinal plant used as anxiolytics in Amazon region. *Revista Brasileira de Farmacognosia, 22(5), 1024-1034.*
- Neto, F. C., Guaratini, T., Costa-Lotuflo, L., Colepicolo, P., Gates, P. J., & Lopes, N. P. (2016). Re-investigation of the fragmentation of protonated carotenoids by electrospray ionization and nanospray tandem mass spectrometry. *Rapid Communications in Mass Spectrometry, 30(13), 1540–1548.*
- Nikolić, D., Gödecke, T., Chen, S.-N., White, J., Lankin, D. C., Pauli, G. F., & van Breemen, R. B. (2012). Mass spectrometric dereplication of nitrogen-containing constituents of black cohosh (*Cimicifuga racemosa* L.). *Fitoterapia, 83(3), 441–460.*

- Nishimura, Y., Murakami, S., Ashikawa, Y., Sasagawa, S., Umemoto, N., Shimada, Y., & Tanaka, T. (2015). Zebrafish as a systems toxicology model for developmental neurotoxicity testing. *Congenital Anomalies*, 55(1), 1–16.
- Nguyen-Pouplin, J., Tran, H., Tran, H., Phan, T. A., Dolecek, C., Farrar, J., Tran, T. H., Caron, P., Bodo, B., Grellier, P. (2007). Antimalarial and cytotoxic activities of ethnopharmacologically selected medicinal plants from South Vietnam. *Journal of Ethnopharmacology*, 109(3), 417–427.
- OECD, Test No. 236: Fish Embryo Acute Toxicity (FET) Test, OECD Guidelines for the Testing of Chemicals, Section 2, OECD Publishing, Paris, 2013.
- Osman, M. S., Ghani, Z. A., Ismail, N. F., Razak, N. A. A., Jaapar, J., & Ariff, M. A. M. (2017). Qualitative comparison of active compounds between red and green Mariposa *Christia Vespertillonis* leaves extracts. AIP Conference Proceedings 1885, 020282.
- Ouyang, H., Li, T., He, M., Li, Z., Tan, T., Zhang, W., Li, Y., Feng, Y., & Yang, S. (2016). Identification and Quantification Analysis on the Chemical Constituents from Traditional Mongolian Medicine Flos Scabiosae Using UHPLC–DAD–Q-TOF-MS Combined with UHPLC–QqQ-MS. *Journal of Chromatographic Science*, 54(6), 1028–1036.
- Pamanji, R., Yashwanth, B., Bethu, M. S., Leelavathi, S., Ravinder, K., & Rao, J. V. (2015). Toxicity effects of profenofos on embryonic and larval development of Zebrafish (*Danio rerio*). *Environmental Toxicology and Pharmacology*, 39(2), 887–897.
- Pariyani, R., Safinar Ismail, I., Azam, A. A., Abas, F., Shaari, K., & Sulaiman, M. R. (2015). Phytochemical Screening and Acute Oral Toxicity Study of Java Tea Leaf Extracts. *BioMed Research International*, 2015, 1–8.
- Parrot, D., Blümel, M., Utermann, C., Chianese, G., Krause, S., Kovalev, A., Gorb, S. N., & Tasdemir, D. (2019). Mapping the Surface Microbiome and Metabolome of Brown Seaweed *Fucus vesiculosus* by Amplicon Sequencing Integrated Metabolomics and Imaging Techniques. *Scientific Reports*, 1–17.
- Peng, J., Fan, G., Hong, Z., Chai, Y., & Wu, Y. (2005). Preparative separation of isovitexin and isoorientin from *Patrinia villosa* Juss by high-speed counter-current chromatography. *Journal of Chromatography A*, 1074(1-2), 111–115.
- Piasecka, A., Kachlicki, P., & Stobiecki, M. (2019). Analytical Methods for Detection of Plant Metabolomes Changes in Response to Biotic and Abiotic Stresses. *International Journal of Molecular Sciences Review*, 20, 1-22.
- Piraud, M., Vianey-Saban, C., Petritis, K., Elfakir, C., Steghens, J.-P., Morla, A., & Bouchu, D. (2003). ESI-MS/MS analysis of underivatized amino acids: a new tool for the diagnosis of inherited disorders of amino acid metabolism. Fragmentation study of 79 molecules of biological interest in positive and negative ionisation mode. *Rapid Communications in Mass Spectrometry*, 17(12), 1297–1311.

- Quinn, R. A., Nothias, L. F., Vining, O., Meehan, M., Esquenazi, E., & Dorrestein, P. C. (2016). Molecular Networking As a Drug Discovery, Drug Metabolism, and Precision Medicine Strategy. *Trends in Pharmacological Sciences*, 38(2), 143–154.
- Ram, A., Balachandar, S., Vijayananth, P., & Singh, V. P. (2011). Medicinal plants useful for treating chronic obstructive pulmonary disease (COPD): Current status and future perspectives. *Fitoterapia*, 82(2), 141–151.
- Rahman, A. H. M., and Parvin, M. I. A. (2014). Study of Medicinal Uses on Fabaceae Family at Rajshahi, Bangladesh. *Research in Plant Sciences*, 2, 6-8.
- Ren, Z., Nie, B., Liu, T., Yuan, F., Feng, F., Zhang, Y., Zhou, W., Xu, X., Yao, M., & Zhang, F. (2016). Simultaneous Determination of Coumarin and Its Derivatives in Tobacco Products by Liquid Chromatography-Tandem Mass Spectrometry. *Molecules*, 21(11), 1511.
- Ridder, L., van der Hooft, J. J. J., & Verhoeven, S. (2014). Automatic Compound Annotation from Mass Spectrometry Data Using MAGMa. *Mass Spectrometry*, 3(2), 1-7.
- Ridder, L., Hooft, J. J. J. Van Der, Verhoeven, S., Vos, R. C. H. De, Schaik, R. Van, & Vervoort, J. (2012). Substructure-based annotation of high-resolution multistage MSⁿ spectral trees, (July), 2461–2471.
- Rivera, S. M., Christou, P., & Canela-Garayoa, R. (2013). Identification of carotenoids using mass spectrometry. *Mass Spectrometry Reviews*, 33(5), 353–372.
- Russell, W. M. S. & Burch, R. L. (1959). The principles of humane experimental technique, London, UK.
- Saint-Amant, L., & Drapeau, P. (1998). Time course of the development of motor behaviors in the zebrafish embryo. *Journal of Neurobiology*, 37(4), 622–632.
- Sarmah, S., & Marrs, J. (2016). Zebrafish as a Vertebrate Model System to Evaluate Effects of Environmental Toxicants on Cardiac Development and Function. *International Journal of Molecular Sciences*, 17(12), 1-16.
- Scheubert, K., Hufsky, F., & Böcker, S. (2013). Computational mass spectrometry for small molecules. *Journal of cheminformatics*, 5(1), 1-24.
- Seraglio, S. K. T., Valese, A. C., Daguer, H., Bergamo, G., Azevedo, M. S., Gonzaga, L. V., Fett, R., & Costa, A. C. O. (2016). Development and validation of a LC-ESI-MS/MS method for the determination of phenolic compounds in honeydew honeys with the diluted-and-shoot approach. *Food Research International*, 87, 60–67.
- Shaikh, A., Kohale, K., Ibrahim, M., & Khan, M. (2019). Teratogenic effects of aqueous extract of Ficus glomerata leaf during embryonic development in

- zebrafish (*Danio rerio*). *Journal of Applied Pharmaceutical Science*, 9(5), 107–111.
- Sharma, K., Amandeep, A., & Chauhan, E.S. (2019). Antidiabetic and lipid lowering extenuating impact of *Glycine max* leaves (soyabean) in type II diabetes mellitus subjects. *Int J Pharm Sci & Res* 2019, 10(5), 2280-2284.
- Shioi, Y., Watanabe, K., & Takamiya, K. -i. (1996). Enzymatic Conversion of Pheophorbide a to the Precursor of Pyropheophorbide a in Leaves of *Chenopodium album*. *Plant and Cell Physiology*, 37(8), 1143–1149.
- Smitha, S., & Reshma, J. (2019). Anatomical Profiling and Phytochemical Analysis of *Christia Vespertilionis* (L.F.). *International Journal of Pharmacy and Biological Sciences*, 9(1), 40–50.
- Simirgiotis, M. J., Schmeda-Hirschmann, G., Bórquez, J., & Kennelly, E. J. (2013). The *Passiflora tripartita* (Banana Passion) Fruit: A Source of Bioactive Flavonoid C-Glycosides Isolated by HSCCC and Characterized by HPLC–DAD–ESI/MS/MS. *Molecules*, 18(2), 1672–1692.
- Sun, G., & Liu, K. (2017). Developmental toxicity and cardiac effects of butyl benzyl phthalate in zebrafish embryos. *Aquatic Toxicology*, 192, 165–170.
- Thakur, R. S., & Ahirwar, B. (2018). A steroidal derivative from *Trigonella foenum graecum* L. that induces apoptosis in vitro and in vivo. *Journal of Food and Drug Analysis*, 1-9.
- The Plant List. (2013). Angiosperms, Leguminosae, *Christia*. Retrieved from <http://www.theplantlist.org>
- Thiagarajan, S. K., Rama Krishnan, K., Ei, T., Husna Shafie, N., Arapoc, D. J., & Bahari, H. (2019). Evaluation of the Effect of Aqueous *Momordica charantia* Linn. Extract on Zebrafish Embryo Model through Acute Toxicity Assay Assessment. *Evidence-Based Complementary and Alternative Medicine*, 2019, 1–9.
- Tsimogiannis, D., Samiotaki, M., Panayotou, G., & Oreopoulou, V. (2007). Characterization of Flavonoid Subgroups and Hydroxy Substitution by HPLC-MS/MS. *Molecules*, 12(3), 593–606.
- Turi, C. E., Finley, J., Shipley, P. R., Murch, S. J., & Brown, P. N. (2015). Metabolomics for Phytochemical Discovery: Development of Statistical Approaches Using a Cranberry Model System. *Journal of Natural Products*, 78, 953–966.
- Upadhyay, H. C., Sisodia, B. S., Cheema, H. S., Agrawal, J., Pal, A., Darokar, M. P., & Srivastava, S. K. (2013). Novel Antiplasmodial Agents from *Christia vespertilionis*. *Natural product communications*, 8, 1591-1594.
- Van Breemen, R. B., Dong, L., & Pajkovic, N. D. (2012). Atmospheric pressure chemical ionization tandem mass spectrometry of carotenoids. *International Journal of Mass Spectrometry*, 312, 163–172.

- Vera, J. S. De, Castro, M. E. G. De, Dulay, R. M. R., Milton, R., & Dulay, R. (2016). Phytochemical Constituents and Teratogenic Effect of Lyophilized Extracts of *Bixa orellana* L. (Achuete) and *Piper betle* L. (Ikmo) Leaves in Danio rerio Embryos. *Der Pharma Chemica*, 8(18), 432–437.
- Wang, M., Carver, J. J., Phelan, V. V., Sanchez, L. M., Garg, N., Peng, Y., Luzzatto-Knaan, T. (2016). Sharing and community curation of mass spectrometry data with Global Natural Products Social Molecular Networking. *Nature Biotechnology*, 34(8), 828–837.
- Wang, S., Tu, H., Wan, J., Chen, W., Liu, X., Luo, J., Xu, J., & Zhang, H. (2016). Spatio-temporal distribution and natural variation of metabolites in citrus fruits. *Food Chemistry*, 199, 8–17.
- Watrous, J., Roach, P., Alexandrov, T., Heath, B. S., Yang, J. Y., Kersten, R. D., der Voort, M.V., Pogliani, K., Gross, H., Raaijmakers, J. M., Moore, B.S., Laskin, J., Bandeira, N., & Dorrestein, P. C. (2012). Mass spectral molecular networking of living microbial colonies. *Proceedings of the National Academy of Sciences*, 109(26), 1-40.
- Whiting, P. A. (2007). Commercial production of *Christia subcordata* Moench by establishing cultural practices and by applying plant growth regulators [dissertation]. The University of Georgia.
- Wink, M. (2013). Evolution of secondary metabolites in legumes (Fabaceae). *South African Journal of Botany*, 89, 164–175.
- Wolfender, J., Marti, G., Thomas, A., & Bertrand, S. (2015). Current approaches and challenges for the metabolite profiling of complex natural extracts. *Journal of Chromatography A*, 1382, 136–164.
- Zhang, C., Willett, C., & Fremgen, T. (2003). Zebrafish: An Animal Model for Toxicological Studies. *Current Protocols in Toxicology*, 1.7.1-1.7.18.
- Zilani, M. N. H., Sultana, T., Asabur Rahman, S. M., Anisuzzman, M., Islam, M. A., Shilpi, J. A., & Hossain, M. G. (2017). Chemical composition and pharmacological activities of *Pisum sativum*. *BMC Complementary and Alternative Medicine*, 17(1), 1-9.

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