



**UNIVERSITI PUTRA MALAYSIA**

***ISOLATION AND EVALUATION OF PROBIOTIC  
POTENTIAL OF LACTIC ACID BACTERIA IN VILLAGE  
CHICKEN GASTROINTESTINAL TRACT***

**ARLENE DEBBIE LINGOH**

**FSPM 2020 1**



**ISOLATION AND EVALUATION OF PROBIOTIC POTENTIAL OF LACTIC  
ACID BACTERIA IN VILLAGE CHICKEN GASTROINTESTINAL TRACT**

**By**

**ARLENE DEBBIE ANAK LINGOH**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfilment of the Requirements for the Degree of Master of Science**

**May 2020**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

## **ISOLATION AND EVALUATION OF PROBIOTIC POTENTIAL OF LACTIC ACID BACTERIA IN VILLAGE CHICKEN GASTROINTESTINAL TRACT**

By

**ARLENE DEBBIE ANAK LINGOH**

May 2020

**Chair : Leong Sui Sien, PhD**  
**Faculty : Agriculture and Food Science (Bintulu Campus)**

Currently, small-scale poultry industries are still utilising feed-additive antibiotics in livestock growth improvement and feed efficiency although it has been banned due to the increased incidences of antibiotic resistance among zoonotic bacteria and the loss of therapeutic efficacy in both veterinary and human medicine. A group of lactic acid bacteria in probiotic, which is a live beneficial microbial feed supplement has been used in commercial animal feed as substitution to the antibiotic. However, the continuous increase in demand for organic farming in food-producing animals clearly necessitate probiotic strains with more effective properties to reduce diseases and improve productivity. This study was conducted purposely to identify and characterize potential probiotic properties and diversity of lactic acid bacteria isolated from the intestinal tract of village chicken. Village chickens that have not been fed any commercial feed have been used as a source of potential probiotic strains to prevent a source of antibiotic-resistant lactic acid bacteria. Five village chickens were caught randomly from every four locations around Bintulu to collect the crop, gizzard, small intestine, large intestine and cecum. Samples were processed and plated for bacterial colony count. 800 bacteria were isolated, clustered and analysed using (GTG)<sub>5</sub> fingerprinting and probiotic analysis test. Phenotypic characterization resulted in only 21 isolates possess the lactic acid bacteria characteristics such as gram-positive, catalase-negative, non-motile and able to ferment glucose. Genotypic characterization has identified that the isolated lactic acid bacteria was *Enterococcus*, *Lactobacillus* and *Bacillus*. Only isolates CR1AM6, L1M10, C4M8, L4AM2 and L4AM5 has shown resistant towards bile salt, pepsin and acidic condition which will enable these strains to survive the harsh condition in the gut. These isolates also able to ferment inulin which is beneficial towards the host as it helps the live microbiota to be implemented easily in the gut.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

## **PENGASINGAN DAN PENILAIAN POTENSI PROBIOTIK BAKTERIA ASID LAKTIK DALAM SALURAN GASTROUSUS AYAM KAMPUNG**

Oleh

**ARLENE DEBBIE ANAK LINGOH**

**Mei 2020**

**Pengerusi : Leong Sui Sien, PhD**  
**Fakulti : Sains Pertanian dan Makanan (Kampus Bintulu)**

Pada masa ini, industri unggas berskala kecil masih menggunakan antibiotik aditif makanan dalam peningkatan pertumbuhan ternakan dan kecekapan pakan walaupun telah dilarang kerana peningkatan kejadian ketahanan terhadap antibiotik di kalangan bakteria zoonotik dan hilangnya keberkesanan terapi dalam perubatan veterinar dan manusia. Sekumpulan bakteria asid laktik dalam probiotik, yang merupakan makanan tambahan mikroba bermanfaat hidup telah digunakan dalam makanan haiwan komersial sebagai pengganti antibiotik. Namun, peningkatan permintaan pertanian organik pada haiwan penghasil makanan secara berterusan memerlukan strain probiotik dengan sifat yang lebih berkesan untuk mengurangkan penyakit dan meningkatkan produktiviti. Kajian ini dilakukan dengan sengaja untuk mengenal pasti dan mencirikan potensi probiotik dan kepelbagaian bakteria asid laktik yang diasingkan dari saluran usus ayam kampung. Ayam kampung yang tidak diberi makanan komersial telah digunakan sebagai sumber strain probiotik yang berpotensi untuk mencegah sumber bakteria asid laktik tahan antibiotik. Lima ekor ayam kampung ditangkap secara rawak dari setiap empat lokasi di sekitar Bintulu untuk mengumpulkan tanaman, gizzard, usus kecil, usus besar dan cecum. Sampel diproses dan dilapisi untuk penghitungan koloni bakteria. 800 bakteria diasingkan, dikelompokkan dan dianalisis menggunakan (GTG)<sub>5</sub> cap jari dan ujian analisis probiotik. Pencirian fenotipik mengakibatkan hanya 21 isolat yang mempunyai ciri bakteria asid laktik seperti gram positif, katalase-negatif, tidak bergerak dan mampu menetap glukosa. Pencirian genotip telah mengenal pasti bahawa bakteria asid laktik terpencil adalah *Enterococcus*, *Lactobacillus* dan *Bacillus*. Hanya pencilan CR1AM6, L1M10, C4M8, L4AM2 dan L4AM5 yang menunjukkan rintangan terhadap garam hempedu, pepsin dan keadaan berasid yang akan membolehkan pencilan ini bertahan dalam keadaan yang ekstrem di usus. Pencilan ini juga dapat memfermentasi inulin yang bermanfaat bagi mikrobiota kerana ia membantu mikrobiota hidup dengan mudah di dalam usus.

## ACKNOWLEDGEMENTS

Firstly, I would like to express my sincere gratitude to my advisor Dr. Leong Sui Sien for the continuous support of my study and related research, for her patience, motivation, and immense knowledge. Her guidance helped me in all the time of research and writing of this thesis. I could not have imagined having a better advisor and mentor for my Master study.

In addition, I would like to express appreciation to my co-advisor, Assoc. Prof. Dr. Shahrul Razid Sarbini for his kind support, and guidance throughout my project; especially for allowing me to begin the project in his lab, sharing chemicals, equipment and other materials.

The physical and technical contribution of Grant Putra IPM (GP-IPM/2017/9533000) which allowed me to undertake this research is truly appreciated. Without their support and funding, this project could not have reached its goal.

Next, I am also very grateful towards the laboratory technicians and staff of UPMKB especially Assistance Science Officer of Analysis Biocompound Laboratory, Miss Georgina Sylvia Niwin for being kind and supportive during my research period.

A special warm thanks you towards the undergraduate students (Nur Hidayah, and Lai Jia Yi) and internship students (Wafri Wasli, Jasmine Eva, Careen Moses, Nur Bahriah and Siti Aisyah) for their assistance in my data collection.

I would like to thank all my colleagues especially Nur Bazilah, Calvin Tan Zhe Kai, Fidelia Johny, Chai Lee Ling, Kathleen Michelle Mikal and Erra Fazira, with whom I have shared moments of deep anxiety but also of big excitement. Their presence was very important in a process that is often felt as tremendously solitaire.

Finally, I must express my very profound gratitude to my parents (Mr. Lingoh Masam and Mrs. Rebecca Seri) and family for providing me with unfailing support and continuous encouragement throughout my years of study and through the process of researching and writing this thesis. This accomplishment would not have been possible without them. Thank you.

I certify that a Thesis Examination Committee has met on 19 May 2020 to conduct the final examination of Arlene Debbie anak Lingoh on her thesis entitled "Isolation and Evaluation of Probiotic Potential of Lactic Acid Bacteria in Village Chicken Gastrointestinal Tract" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

**Phebe Ding, PhD**

Associate Professor  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Chairman)

**Patricia King Jie Hung, PhD**

Associate Professor  
Faculty of Agriculture and Food Science (Bintulu Campus)  
Universiti Putra Malaysia  
(Internal Examiner)

**Samuel Lihan, PhD**

Senior Lecturer  
Faculty of Resource Science and Technology  
Universiti Malaysia Sarawak  
Malaysia  
(External Examiner)

---

**ZURIATI AHMAD ZUKARNAIN, PhD**

Professor Ts. and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 07 August 2020

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

**Leong Sui Sien, PhD**

Senior Lecturer  
Faculty of Agriculture and Food Science  
Universiti Putra Malaysia Bintulu Sarawak Campus  
(Chairman)

**Shahrul Razid Sarbini, PhD**

Associate Professor  
Faculty of Agriculture and Food Science  
Universiti Putra Malaysia Bintulu Sarawak Campus  
(Member)

---

**ZALILAH MOHD SHARIFF, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 13 August 2020



## Declaration by graduate student

- I hereby confirm that:
- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_ Date: 08/10/2021

Name and Matric No.: Arlene Debbie anak Lingoh, GS51717

## Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature:

Name of Chairman of  
Supervisory Committee:

\_\_\_\_\_  
Dr. Leong Sui Sien

Signature:

Name of Member of  
Supervisory Committee:

\_\_\_\_\_  
Assoc. Prof. Dr. Shahrul Razid Sarbini

## TABLE OF CONTENTS

	<b>Page</b>
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	ii
<b>ACKNOWLEDGEMENTS</b>	iii
<b>APPROVAL</b>	iv
<b>DECLARATION</b>	vi
<b>LIST OF TABLES</b>	xi
<b>LIST OF FIGURES</b>	xii
<b>LIST OF ABBREVIATIONS</b>	xiv

### CHAPTER

<b>1</b>	<b>INTRODUCTION</b>	1
<b>2</b>	<b>LITERATURE REVIEW</b>	4
	2.1 Poultry industry	4
	2.2 Poultry industry in Malaysia	5
	2.3 Challenges faced by poultry industry	5
	2.4 Antibiotics	6
	2.5 Application of antibiotics in poultry	6
	2.6 Antibiotic resistance	7
	2.7 Probiotics	8
	2.8 Probiotic market	9
	2.9 Lactic acid bacteria (LAB)	10
	2.10 Probiotic properties	11
	2.11 Malaysian village chicken	11
	2.12 Gastrointestinal tract of poultry	12
	2.13 Gastrointestinal microflora of chicken	12
<b>3</b>	<b>GENERAL MATERIALS AND METHODS</b>	14
	3.1 Sampling sites	14
	3.2 Sample collection and processing	14
	3.3 Total viable colony count	15
	3.4 Preparation of stock culture	15
	3.5 Bacteria identification	15
	3.5.1 Phenotypic characterization	15
	3.5.2 Genomic DNA extraction	16
	3.5.3 Genotypic characterization of lactic acid bacteria	16
	3.6 Probiotic analysis	17
	3.7 Antimicrobial test	17
	3.8 <i>In vitro</i> digestion	18
	3.8.1 Oral phase	18
	3.8.2 Gastric phase	18
	3.8.3 Intestinal phase	19
	3.9 <i>In-vitro</i> pure culture Fermentation	19
	3.9.1 Inoculum preparation	19
	3.9.2 Fermentation medium	19
	3.9.3 <i>In-vitro</i> pure culture fermentation condition	19

	3.10	Statistical analysis	20
<b>4</b>		<b>MICROFLORA DIVERSITY OF VILLAGE CHICKEN GASTROINTESTINAL TRACT</b>	<b>21</b>
	4.1	Introduction	21
	4.2	Materials and methods	22
		4.2.1 Sampling sites	22
		4.2.2 Sample collection and processing	22
		4.2.3 Total viable colony count	22
		4.2.4 Preparation of stock culture	23
		4.2.5 Bacteria identification	23
		4.2.6 Statistical analysis	25
	4.3	Results	25
		4.3.1 Total viable count	25
		4.3.2 Biochemical analysis of LAB isolates	30
		4.3.3 Molecular clustering of LAB isolates	33
		4.3.4 Molecular identification of bacteria using 16S rRNA sequencing	38
	4.4	Discussion	39
	4.5	Conclusion	41
<b>5</b>		<b>PROBIOTIC SCREENING OF LAB ISOLATED FROM VILLAGE CHICKEN GUT</b>	<b>42</b>
	5.1	Introduction	42
	5.2	Materials and methods	43
		5.2.1 Bacterial strains	43
		5.2.2 Carbohydrate fermentation test	44
		5.2.3 Acid tolerance test	44
		5.2.4 Bile tolerance	44
		5.2.5 Pepsin tolerance	45
		5.2.6 Antimicrobial test	45
		5.2.7 Statistical analysis	46
	5.3	Results	46
		5.3.1 Carbohydrate fermentation test	46
		5.3.2 Acid tolerance test	47
		5.3.3 Pepsin tolerance test	52
		5.3.4 Bile tolerance test	54
		5.3.5 Selected isolates	56
		5.3.6 Antimicrobial test	58
	5.4	Discussions	59
	5.5	Conclusion	61
<b>6</b>		<b><i>IN-VITRO</i> PURE CULTURE FERMENTATION OF INULIN BY LACTIC ACID BACTERIA ISOLATED FROM VILLAGE CHICKEN GASTROINTESTINAL TRACT</b>	<b>62</b>
	6.1	Introduction	62

6.2	Materials and methods	63
6.2.1	Bacterial strains	63
6.2.2	<i>In vitro</i> digestion of inulin	63
6.2.3	<i>In vitro</i> pure culture fermentation	65
6.2.4	Statistical analysis	65
6.3	Results	66
6.4	Discussions	69
6.5	Conclusion	69
<b>7</b>	<b>SUMMARY, GENERAL CONCLUSION AND RECOMMENDATION FOR FUTURE RESEARCH</b>	<b>70</b>
	<b>REFERENCES</b>	<b>72</b>
	<b>APPENDICES</b>	<b>85</b>
	<b>BIODATA OF STUDENT</b>	<b>89</b>
	<b>LIST OF PUBLICATIONS</b>	<b>90</b>

## LIST OF TABLES

Table		Page
3.1	Assessment of probiotic abilities of lactic acid bacteria	17
3.2	Composition of digestion fluids	18
4.1	The mean total bacteria count ( $\log_{10}$ cfu mL <sup>-1</sup> ) isolated from gastrointestinal tract of village chickens at Kampung Jepang on multiple medias	26
4.2	The mean total bacteria count ( $\log_{10}$ cfu mL <sup>-1</sup> ) isolated from gastrointestinal tract of village chicken at Batu 12 on multiple medias	27
4.3	The mean total bacteria count ( $\log_{10}$ cfu mL <sup>-1</sup> ) isolated from gastrointestinal tract of village chickens at Wet Market on multiple medias	28
4.4	The mean total bacteria count ( $\log_{10}$ cfu/mL) isolated from gastrointestinal tract of village chickens at UPMKB on multiple media	29
4.5	Biochemical characterization of some isolated lactic acid bacteria isolates	32
4.6	The 16S rRNA Sequencing results of LAB isolated from village chickens' gastrointestinal tract	38
5.1	Isolation sources of lactic acid bacteria	43
5.2	Carbohydrate fermentation test of LAB	46
5.3	Acid resistance of 25 isolates	48
5.4	Pepsin resistance of 25 isolates	52
5.5	Bile resistance of 25 isolates	54
5.6	Antimicrobial activity of LAB isolates	58
6.1	Bacterial strains	63
6.2	Composition of digestion fluids	63

## LIST OF FIGURES

Figure		Page
3.1	The sampling sites of mix breed village chicken in Bintulu Sarawak	14
4.1	The percentage (%) of Gram positive and Gram-negative bacteria distribution isolated from gastrointestinal tract of village chickens at Kampung Jepak, Bintulu	30
4.2	The percentage (%) of Gram positive and Gram-negative bacteria distribution isolated from gastrointestinal tract of village chickens at Batu12, Bintulu	31
4.3	The percentage (%) of Gram positive and Gram-negative bacteria distribution isolated from gastrointestinal tract of village chickens at Wet Market, Bintulu	31
4.4	The percentage (%) of Gram positive and Gram-negative bacteria distribution isolated from gastrointestinal tract of village chickens at UPMKB, Bintulu	32
4.5	Dendrogram showing the lactic acid bacteria distribution isolated from gastrointestinal tracts of village chickens at Kampung Jepak	34
4.6	Dendrogram showing the lactic acid bacteria distribution isolated from gastrointestinal tracts of village chickens at Batu 12	35
4.7	Dendrogram showing the lactic acid bacteria distribution isolated from gastrointestinal tracts of village chickens at Wet Market	36
4.8	Dendrogram showing the lactic acid bacteria isolated from gastrointestinal tracts of village chickens at UPMKB	37
5.1	Fermentation test (isolates able to ferment glucose through the changes color of medium from red to yellow and produced no gas inside the Durham tubes)	47
5.2	Tolerance of isolated LAB at pH 2 (OD 620nm values) tested for 0 and 4 hours	49
5.3	Tolerance of isolated LAB at pH 2.5 (OD 620nm values) tested for 0 and 4 hours	50
5.4	Tolerance of isolated LAB at pH 3 (OD 620nm values) tested for 0 and 4 hours	51

5.5	Tolerance of isolated LAB against 0.3% of pepsin (OD 620nm values) tested for 0 and 4 hours	53
5.6	Tolerance of isolated LAB against 0.3% of bile (OD 620nm values) tested for 0 and 4 hours	55
5.7	Selected isolates in acid tolerance test (pH2)	56
5.8	Selected isolates in 0.3% pepsin tolerance test	56
5.9	Selected isolates in 0.3% bile tolerance test	57
6.1	Absorbance (nm) of LAB isolates throughout 24 hours <i>in vitro</i> pure culture fermentation of inulin	67
6.2	Absorbance (nm) of LAB isolates for 0, 6, 12, and 24 hours <i>in vitro</i> pure culture fermentation of inulin	68



## LIST OF ABBREVIATIONS

HPAI	Highly Pathogenic Avian Influenza
US	United States
EU	European Union
FAO	Food and Agriculture Organisation
AGP	Antibiotic Growth Promoters
LAB	Lactic Acid Bacteria
FAOSTAT	Food and Agriculture Organization Corporate Statistical Database
FDA	Food and Drug Administration
USA	United States of America
WHO	World Health Organization
FAO/WHO	Food and Agriculture Organization of the United Nations and the World Health Organisation
GIT	Gastrointestinal Tract
GB	Goblet cells
IELs	Intraepithelial lymphocytes
PCR	Polymerase Chain Reaction
DNA	Deoxyribonucleic Acid
dNTPs	deoxyribonucleotide triphosphates
rDNA	Recombinant Deoxyribonucleic Acid
UPMKB	Universiti Putera Malaysia Kampus Bintulu
PBS	Phosphate Buffer Saline
MRS	de-Mann Rogosa Sharpe
NB	Nutrient broth
NA	Nutrient Agar
EMB	Eosin Methylene Blue

CFU	Colony Forming Unit
NCBI	National Center for Biotechnology Information
UK	United Kingdom
OD	Optical density
SSF	Simulated Salivary Fluid
SIF	Simulated Intestinal Fluid
SGF	Simulated Gastric Fluid
TAE	Tris acetate EDTA
KCl	Potassium Chloride
$\text{KH}_2\text{PO}_4$	potassium dihydrogenphosphate
$\text{NaHCO}_3$	Sodium bicarbonate
$\text{NaCl}$	Sodium Chloride
$\text{MgCl}_2 \cdot 6\text{H}_2\text{O}$	Magnesium Chloride, Hexahydrate
$(\text{NH}_4)_2\text{CO}_3$	Ammonium carbonate
$\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$	Calcium Chloride Dihydrate
$\text{HCl}$	Hydrochloric Acid
$\text{NaOH}$	Sodium Hydroxide
C <sub>inh</sub>	Coefficient of inhibition
C	Cecum
CR	Crop
S	Small intestine
L	Large intestine
G	Gizzard
$\text{CO}_2$	Carbon dioxide
BSH	bile salt hydrolase enzyme
$\text{H}_2\text{O}_2$	Hydrogen peroxide

SCFA            Short chain fatty acid  
DP              Degree of polymerization



© COPYRIGHT UPM

## CHAPTER 1

### INTRODUCTION

Recently, domesticated bird's flesh and eggs are the foremost consumed food derived from animal amongst different sources world widely and it became vital for food security and nutrition. Because of its natural origin, ability to provide the essential macronutrient and meet the global needs, the industry arises as to the foremost competent sub-sector (Mottet & Tempio, 2017). The most widespread in poultry meat such as turkey, duck, hens and chicken is 70 to 80 percent of world consumption, as stated by Skarp *et al.*, (2016). The world's preferred source of protein can be considered chicken meat because it's cheap and easy to get. For the last 50 years, compared to other livestock products, the average annual growth for poultry was 5% while it was only 1.5%, 3.1% and 1.7% for beef, pork and ruminants respectively (Alexandratos & Bruinsma, 2012). World production of poultry meat in 2017 is forecast at 120.5 million, up 1.1 per cent from 2016. Following regular outbreaks of Highly Pathogenic Avian Influenza (HPAI) in Africa, Asia and Europe, production increased in almost all major poultry growing areas. Most of those increased production in 2017 originated from the US (+2.4%), the Russian Federation (+7.2%), Brazil (+1.9%), Turkey (+12.5%), India (+4.8%), Thailand (+6.7%), Mexico (+3.8%), and the EU (+0.8%). World poultry meat demand increased by 21.3 million tonnes between 2010 and 2017 (Food and Agriculture Organization, 2018). According to Mottet & Tempio, (2017), due to rising population and consumer incomes, global demand for cheaper protein sources will continue to intensify with the increase in global chicken consumption. This will lead to an increase in the production of commercial chicken to meet global demand.

In Malaysia, the establishment of a reliable and safe poultry-specialized system has allowed entrepreneurs to enhance their poultry business. According to the 2018 Department of Veterinary Services Malaysia survey, the production of poultry meat rises from 1,664.9('000 M. Tonne) in 2017 to 1,653.7('000 M. Tonne) in 2018. In addition, the number is expected to rise into 1,716.7('000 M. Tonne) in 2019 (Department of Veterinary Services, 2018). This improvement has modified the Malaysian poultry industry from the smaller grounds production system to extremely modernised, commercialised and economical production systems. The high consumption has helped the Malaysian poultry industry to support the high production output throughout the years compared to other livestock industries. Besides being the most cost-effective supply of meat protein, chicken meat has no prohibitions or restrictions in dietary and religion consumption in contrast with pork and beef that are restricted to Malays and Hindus, correspondingly. Therefore, the availability consumption of the meat to any or all races conjointly contributed to the rapid climb of the industry (Abdurofi *et al.*, 2017). However, the increasing demand has raised concern for the long-run sustainability of the poultry industry. As a result, most poultry farmers resulted within the use of antibiotics to increase growth rates, manufacture more eggs, and reduce death rate and diseases (Samsuddin *et al.*, 2015).

One of the foremost exceptional medical uncovering of 20<sup>th</sup> century is the discovery of antibiotics while their functions in treating the animal and human diseases are still crucial during this 21<sup>st</sup> century. Between the early 1940s to 1950s, antibiotics have been widely used in the animal food industry (Allen *et al.*, 2013). In animal production, antibiotics are being used for medicine treatments, prophylactics preventatives, and growth starters in food animal production (Cheng *et al.*, 2014). They are even have been enforced in animal feed as a growth promoter in expanding feed potency of conventional farm animal and poultry production (Allen *et al.*, 2013; Mungroo & Neethirajan, 2014). Growth promoters is defined as the substances that are deliberately supplemented to a diet to trigger the utmost response of the genomic potential of the host in terms of development and enhancement in feed effectiveness (Dhama *et al.*, 2014). However, the long use and the misuse of antibiotics as a growth promoter have increased antibiotic-resistant microorganism and this has turned the meat chicken as the medium for the transmission of multidrug-resistant microorganism to the consumers (Salim *et al.*, 2013; Aliyu *et al.*, 2013). As a result, this could cause the failure of antibiotic treatment (Salim *et al.*, 2013; Mungroo & Neethirajan, 2014) and may be impossible to beat infections (Mungroo & Neethirajan, 2014). All the reported cases show that the continual of exploitation antibiotics in feed animals have led to serious issues as they can threaten public health and reduces drug effectiveness. Therefore, the use of antibiotics in animal feed has been forbidden by many countries including South Korea, which has explicitly restricted the use of antibiotic growth promoters (AGP) in the animal diet (Salim *et al.*, 2013). With the elimination of antibiotics as a growth driver for livestock, the poultry industry is now facing a major challenge in sustaining output efficiency, as feed prices have risen and the prohibition on antimicrobial usage in feed causes high morbidity and mortality in poorly managed flocks (Salim *et al.*, 2013; Dhama *et al.*, 2014). Efficient substitution of growth promoters is therefore urgently needed to ensure that contemporary levels of animal production can be maintained without intimidating public health (Seal *et al.* 2013).

One of the solutions that are accustomed overcome the antibiotic-resistant in animal feed is by substituting antibiotics with probiotics. Recently, probiotics have gained a lot of interest from the broiler chicken industry as a substitution for the prophylactic usage of antibiotics in animal feeds (Musikasang *et al.*, 2009; Salim *et al.*, 2013 ). According to Hill *et al.*, (2014), the definition of probiotic is “live microorganisms that, when administered in adequate amounts, confer a health benefit on the host”. Due to historical belief that lactic acid bacteria are necessary members of the colonic microflora and consequently commonly regarded as safe, it often used in most probiotics preparation (Jadhav *et al.*, 2018). The feeding of chicken with diet supplemented with probiotics will benefit the birds by having a balance ecological microflora inside the gut and minimizing the growth of potential pathogens like enterobacteria and *Campylobacter* species in poultry gut which will cause damage to the host by enhancing their immunity system (Aliakbarpour *et al.*, 2012; Ritzi, *et al.*, 2018).

Although numerous publications show improvement in the performance of broilers, layers and turkeys, there are also reports that probiotics have a limited and variable growth-promoting effect and, in some cases, none (Karaoglu and Durdag, 2005; O'Dea *et al.*, 2006; Lee *et al.*, 2010; Waititu *et al.*, 2014). This inconsistency in results can be attributed to differences in the type and dose of strain used, variations in treatment, duration and duration of administration, diet and the environment. Various commercial

probiotic products on the global market are currently available for poultry. Nonetheless, due to inadequate analysis of the unique beneficial properties of the probiotic strains formulated in the drug, some of them may not be highly potent (Binek, 2016). Moreover, most manufacturers lack the patience to carry out an in-depth study of each strain in order to ascertain its full probiotic potential before commercialization, as most industries maximize profits at minimal cost. (Reuben *et al.*, 2019). While several LAB strains have probiotic characteristics and have been established as suitable probiotic supplements for different hosts, new probiotic strains with better health promotion characteristics are still being searched for than are currently available in the market. Consequently, this research was designed to isolate, identify and test strains of lactic acid bacteria with sufficient probiotic properties from the village chicken gut for use as poultry probiotics.

The objectives of the study are:

1. To isolate lactic acid bacteria from different part of the gastrointestinal tract of village chicken.
2. To evaluate the probiotic potential of lactic acid bacteria isolated from different part of gastrointestinal tract of village chicken.
3. To investigate the effect of inulin on the growth of lactic acid bacteria.

## REFERENCES

- Aazami, N., Kalantar, E., Poormazaheri, H., Pour, N. S. vali, & Jouzan, G. S. (2016). Selection and characterization of potential probiotic *Lactobacilli* spp isolated from chicken feces may be used as a potent antibacterial agent. *Asian Journal of Dairy and Food Research*, 35(1), 50–57. <https://doi.org/10.18805/ajdfr.v35i1.9252>
- Abdurofi, I., Ismail, M. M., Kamal, H. a. W., & Gabdo, B. H. (2017). Economic analysis of broiler production in Peninsular Malaysia. *International Food Research Journal*, 24(4), 1387–1392.
- Albazaz, R. I., & Byukunal Bal, E. B. (2014). Microflora of Digestive Tract in Poultry. *Kahramanmaraş Sütçü İmam Üniversitesi Doğa Bilimleri Dergisi*, 17(1), 39. <https://doi.org/10.18016/ksujns.40137>
- Alexandratos, N., & Bruinsma, J. (2012). *World agriculture towards 2030/2050: the 2012 revision* (No. 12–03). *ESA Working paper*. Rome. <https://doi.org/10.1002/jso.2930300113>
- Aliakbarpour, H. R., Chamani, M., Rahimi, G., Sadeghi, A. A., & Quejeq, D. (2012). The *Bacillus subtilis* and lactic acid bacteria probiotics influences intestinal mucin gene expression, histomorphology and growth performance in broilers. *Asian-Australasian Journal of Animal Sciences*, 25(9), 1285–1293. <https://doi.org/10.5713/ajas.2012.12110>
- Aliyu, A. B., Saleha, A. A., Jalila, A., & Zunita, Z. (2016). Risk factors and spatial distribution of extended spectrum  $\beta$ -lactamase-producing- *Escherichia coli* at retail poultry meat markets in Malaysia: A cross-sectional study. *BMC Public Health*, 16(1), 1–9. <https://doi.org/10.1186/s12889-016-3377-2>
- Allen, H. K., Levine, U. Y., Looft, T., Bandrick, M., & Casey, T. A. (2013). Treatment, promotion, commotion: Antibiotic alternatives in food-producing animals. *Trends in Microbiology*, 21(3), 114–119. <https://doi.org/10.1016/j.tim.2012.11.001>
- Alloui, M. N., Szczurek, W., & Świątkiewicz, S. (2013). The Usefulness of Prebiotics and Probiotics in Modern Poultry Nutrition: A Review. *Annals of Animal Science*, 13(1), 17–32. <https://doi.org/10.2478/v10220-012-0055-x>
- Apajalahti, J., Kettunen, A., & Graham, H. (2004). Characteristics of the gastrointestinal microbial communities , with special reference to the chicken, (April 2015). <https://doi.org/10.1079/WPS200415>
- Ashraf, S., Zaneb, H., Yousaf, M. S., Ijaz, A., Sohail, M. U., Muti, S., ... Rehman, H. (2013). Effect of dietary supplementation of prebiotics and probiotics on intestinal microarchitecture in broilers reared under cyclic heat stress. *Journal of Animal Physiology and Animal Nutrition*, 97(SUPPL.1), 68–73. <https://doi.org/10.1111/jpn.12041>

- Astuti, I. (2016). Isolation , Characterization , and Identification Lactic Acid Bacteria from Chicken Waste Faeces that Potential as Probiotics, 6(5), 180–191.
- Awais, M. M., Akhtar, M., Iqbal, Z., Muhammad, F., & Anwar, M. I. (2012). Seasonal prevalence of coccidiosis in industrial broiler chickens in Faisalabad, Punjab, Pakistan. *Tropical Animal Health and Production*, 44(2), 323–328. <https://doi.org/10.1007/s11250-011-0024-x>
- Bai, S. P., Wu, A. M., Ding, X. M., Lei, Y., Bai, J., Zhang, K. Y., & Chio, J. S. (2013). Effects of probiotic-supplemented diets on growth performance and intestinal immune characteristics of broiler chickens. *Poultry Science*, 92(3), 663–670. <https://doi.org/10.3382/ps.2012-02813>
- Bakari, D., Tatsadjieu, N. L., Mbawala, A., & Mbofung, C. M. (2011). Assessment of Physiological Properties of Some Lactic Acid Bacteria Isolated From the Intestine of Chickens Use As Probiotics and Antimicrobial Agents Against Enteropathogenic Bacteria. *Innovative Romanian Food Biotechnology*, 8, 33–40.
- Baldi. (2013). Comparative Insight of Regulatory Guidelines for Probiotics in USA, India and Malaysia: A Critical Review. *International Journal of Biotechnology for Wellness Industries*, 51–64. <https://doi.org/10.6000/1927-3037.2013.02.02.1>
- Barbut, S. (2016). *Poultry Products Processing; An Industry Guide*. CRC Press.
- Bednarczyk, M., Stadnicka, K., Kozłowska, I., Abiuso, C., Tavaniello, S., Dankowiakowska, A., ... Maiorano, G. (2016). Influence of different prebiotics and mode of their administration on broiler chicken performance. *Animal*, 10(8), 1271–1279. <https://doi.org/10.1017/S1751731116000173>
- Binek, M. (2016). Assessment of potentially probiotic properties of Lactobacillus strains isolated from chickens, 19(1), 15–20. <https://doi.org/10.1515/pjvs-2016-0003>
- Birmani, M. W., Nawab, A., Ghani, M. W., Li, G., Xiao, M., & An, L. (2019). A Review: Role of Inulin in Animal Nutrition. *Journal of Food Technology Research*, 6(1), 18–27. <https://doi.org/10.18488/journal.58.2019.61.18.27>
- Borda-Molina, D., Seifert, J., & Camarinha-Silva, A. (2018). Current Perspectives of the Chicken Gastrointestinal Tract and Its Microbiome. *Computational and Structural Biotechnology Journal*, 16, 131–139. <https://doi.org/10.1016/j.csbj.2018.03.002>
- Braykov, N. P., Eisenberg, J. N. S., Grossman, M., Zhang, L., Vasco, K., Cevallos, W., ... Levy, K. (2016). Antibiotic Resistance in Animal and Environmental Samples Associated with Small-Scale Poultry Farming in Northwestern Ecuador. *MSphere*, 1(1), 1–15. <https://doi.org/10.1128/msphere.00021-15>
- Broom, L. J., & Kogut, M. H. (2018). The role of the gut microbiome in shaping the immune system of chickens. *Veterinary Immunology and Immunopathology*, 204(October), 44–51. <https://doi.org/10.1016/j.vetimm.2018.10.002>



- Carr, F. J., Chill, D., & Maida, N. (2002). The lactic acid bacteria: A literature survey. *Critical Reviews in Microbiology*, 28(4), 281–370. <https://doi.org/10.1080/1040-840291046759>
- Chen, C. Y., Chen, S. W., & Wang, H. T. (2017). Effect of supplementation of yeast with bacteriocin and *Lactobacillus* culture on growth performance, cecal fermentation, microbiota composition, and blood characteristics in broiler chickens. *Asian-Australasian Journal of Animal Sciences*, 30(2), 211–220. <https://doi.org/10.5713/ajas.16.0203>
- Chen, Y.-S., Yanagida, F., & Shinohara, T. (2005). Isolation and identification of lactic acid bacteria from soil using an enrichment procedure. *Letters in Applied Microbiology*, 40(3), 195–200. <https://doi.org/10.1111/j.1472-765X.2005.01653.x>
- Cheng, G., Hao, H., Xie, S., Wang, X., Dai, M., Huang, L., & Yuan, Z. (2014). Antibiotic alternatives: The substitution of antibiotics in animal husbandry? *Frontiers in Microbiology*, 5(MAY), 1–15. <https://doi.org/10.3389/fmicb.2014.00217>
- Chin, R. P., Empel, P. C. M. van, & Hafez, H. M. (2008). *Ornithobacterium rhinotracheale* Infection. In Y. M. Saif, A. M. Fadly, J. R. Glisson, L. R. McDougald, L. K. Nolan, & D. E. Swayne (Eds.), *Diseases of Poultry* (12th ed., pp. 765–774). Blackwell Publishing. <https://doi.org/10.1016/B978-1-4557-0297-8.00402-X>
- Dakpogan, H. B., & Salifou, S. (2013). Coccidiosis prevalence and intensity in litter-based high stocking density layer rearing system of Benin. *Journal of Animal & Plant Sciences*, 17(2), 2522–2526.
- Dankowiakowska, A., Kozłowska, I., & Bednarczyk, M. (2013). Probiotyki, prebiotyki i synbiotyki - Sposób działania, ograniczenia i osia{ogonek}gnie{ogonek}cia. *Journal of Central European Agriculture*, 14(1), 467–478. <https://doi.org/10.5513/JCEA01/14.1.1222>
- Danzeisen, J. L., Kim, H. B., Isaacson, R. E., Tu, Z. J., & Johnson, T. J. (2011). Modulations of the chicken cecal microbiome and metagenome in response to anticoccidial and growth promoter treatment. *PLoS ONE*, 6(11). <https://doi.org/10.1371/journal.pone.0027949>
- De Vuyst, L., Camu, N., De Winter, T., Vandemeulebroecke, K., Van de Perre, V., Vancanneyt, M., ... Cleenwerck, I. (2008). Validation of the (GTG)<sub>5</sub>-rep-PCR fingerprinting technique for rapid classification and identification of acetic acid bacteria, with a focus on isolates from Ghanaian fermented cocoa beans. *International Journal of Food Microbiology*, 125(1), 79–90. <https://doi.org/10.1016/j.ijfoodmicro.2007.02.030>
- Denbow, D. M. (2015). Gastrointestinal Anatomy and Physiology. In *Sturkie's Avian Physiology: Sixth Edition* (Sixth Edit, pp. 337–366). Elsevier. <https://doi.org/10.1016/B978-0-12-407160-5.00014-2>

- Department of Veterinary Services. (2018). Jabatan Perkhidmatan Veterinar Perangkaan Ternakan 2017/2018. Retrieved November 25, 2019, from <http://www.dvs.gov.my/index.php/pages/view/2758?mid=42>
- Dhama, K., Verma, V., Sawant, P. M., Tiwari, R., Vaid, R. K., & Chauhan, R. S. (2011). Applications of Probiotics in Poultry: Enhancing Immunity and Beneficial Effects on Production Performances and Health - A Review. *Journal of Immunology and Immunopathology*, 13(1), 1–19.
- Dhama, Kuldeep, Tiwari, R., Ullah Khan, R., Chakraborty, S., Gopi, M., Karthik, K., ... Tulasi Sunkara, L. (2014). Growth Promoters and Novel Feed Additives Improving Poultry Production and Health, Bioactive Principles and Beneficial Applications: The Trends and Advances- A Review. *International Journal of Pharmacology*, 10(3), 129–159. <https://doi.org/10.3923/ijp.2014.129.159>
- Diarra, Moussa S., & Malouin, F. (2014). Antibiotics in Canadian poultry productions and anticipated alternatives. *Frontiers in Microbiology*, 5(JUN), 1–15. <https://doi.org/10.3389/fmicb.2014.00282>
- Diarra, Moussa Sory, Delaquis, P., Rempel, H., Bach, S., Harlton, C., Aslam, M., ... Topp, E. (2014). Antibiotic resistance and diversity of Salmonella enterica serovars associated with broiler chickens. *Journal of Food Protection*, 77(1), 40–49. <https://doi.org/10.4315/0362-028.JFP-13-251>
- Dittoe, D. K., Ricke, S. C., & Kiess, A. S. (2018). Organic acids and potential for modifying the avian gastrointestinal tract and reducing pathogens and disease. *Frontiers in Veterinary Science*. <https://doi.org/10.3389/fvets.2018.00216>
- Farrell, D., Ventura da Silva, M., Glatz, P., Pym, R., Micheal Williams, C., Ravindran, V., ... Davies, A. (2013). *Poultry Development Review. The role of poultry in human nutrition*. FAO.
- Flint, H. J., Scott, K. P., Duncan, S. H., Louis, P., & Forano, E. (2012). Microbial degradation of complex carbohydrates in the gut © 2012 Landes Bioscience . Do not distribute © 2012 Landes Bioscience . Do not distribute. *Gut Microbes*, (August), 289–306.
- Food and Agriculture Organisation. (2018). Meat Market Review. *Meat Market Review*, (April), 1–11. Retrieved from <http://www.fao.org/3/I9286EN/i9286en.pdf>
- Fuller, R. (2001). The Chicken Gut Microflora and Probiotic Supplements. *Journal of Poultry Science*, 38(3), 189–196. <https://doi.org/10.2141/jpsa.38.189>
- Gabriel, I., Lessire, M., Mallet, S., & Guillot, J. . (2006). Microflora of the digestive tract: critical factors and consequences for poultry. *World's Poultry Science Journal*, 62(03), 499–511. <https://doi.org/10.1079/wps2006111>
- García, Y., Boucourt, R., Albelo, N., & Núñez, O. (2007). Inulin fermentation by lactic acid bacteria with probiotic characteristics. *Cuban Journal of Agricultural Science*, 41(3), 251–254.

- Gevers, D., Huys, G., & Swings, J. (2001). Applicability of rep-PCR fingerprinting for identification of *Lactobacillus* species. *FEMS Microbiology Letters*, 205(1), 31–36. [https://doi.org/10.1016/S0378-1097\(01\)00439-6](https://doi.org/10.1016/S0378-1097(01)00439-6)
- Gong, J, Yin, F., Hou, Y., & Yin, Y. (2014). Review: Chinese herbs as alternatives to antibiotics in feed for swine and poultry production: Potential and challenges in application. *Canadian Journal of Animal Science*, 94(2), 223–241. <https://doi.org/10.4141/CJAS2013-144>
- Gong, Jianhua, Si, W., Forster, R. J., Huang, R., Yu, H., Yin, Y., ... Han, Y. (2007). 16S rRNA gene-based analysis of mucosa-associated bacterial community and phylogeny in the chicken gastrointestinal tracts: From crops to ceca. *FEMS Microbiology Ecology*, 59(1), 147–157. <https://doi.org/10.1111/j.1574-6941.2006.00193.x>
- Gopal, P. K., Prasad, J., Smart, J., & Gill, H. S. (2001). In vitro adherence properties of *Lactobacillus rhamnosus* DR20 and *Bifidobacterium lactis* DR10 strains and their antagonistic activity against an enterotoxigenic *Escherichia coli*. *International Journal of Food Microbiology*, 67(3), 207–216. [https://doi.org/10.1016/S0168-1605\(01\)00440-8](https://doi.org/10.1016/S0168-1605(01)00440-8)
- Gotcheva, V., Hristozova, E., Hristozova, T., Guo, M., Roshkova, Z., & Angelov, A. (2013). Assessment of Potential Probiotic Properties of Lactic Acid Bacteria and Yeast Strains. *Food Biotechnology*, 16(3), 211–225.
- Hawaz, E. (2014). Isolation and identification of probiotic lactic acid bacteria from curd and in vitro evaluation of its growth inhibition activities against pathogenic bacteria, 8(13), 1419–1425. <https://doi.org/10.5897/AJMR2014.6639>
- Hill, C., Guarner, F., Reid, G., Gibson, G. R., Merenstein, D. J., Pot, B., ... Sanders, M. E. (2014). Expert consensus document: The international scientific association for probiotics and prebiotics consensus statement on the scope and appropriate use of the term probiotic. *Nature Reviews Gastroenterology and Hepatology*, 11(8), 506–514. <https://doi.org/10.1038/nrgastro.2014.66>
- Hosseindoust, A., Park, J. W., & Kim, I. H. (2016). Effects of *Bacillus subtilis*, Kefir and  $\beta$ -Glucan Supplementation on Growth Performance, Blood Characteristics, Meat Quality and Intestine Microbiota in Broilers. *Korean Journal of Poultry Science*, 43(3), 159–167. <https://doi.org/10.5536/kjps.2016.43.3.159>
- Iraporda, C., Rubel, I. A., Manrique, G. D., & Abraham, A. G. (2019). Influence of inulin rich carbohydrates from Jerusalem artichoke (*Helianthus tuberosus* L.) tubers on probiotic properties of *Lactobacillus* strains. *LWT*, 101, 738–746. <https://doi.org/10.1016/j.lwt.2018.11.074>
- Jadhav, K., Sharma, K. S., Katoch, S., Sharma, V. K., & Mane, B. G. (2018). Probiotics in Broiler Poultry Feeds : A Review Probiotics in Broiler Poultry Feeds : A Review. *International Journal of Animal and Veterinary Sciences*, (January).

- Jakaria, A. T. M., Islam, A., & Khatun, M. M. (2012). Prevalence, Characteristics and Antibiogram Profiles of *Escherichia coli* Isolated from Apparently Healthy Chickens in Mymensingh, Bangladesh. *Microbes and Health*, *0153*(June), 27–29.
- Jeong, J. S., & Kim, I. H. (2014). Effect of *Bacillus subtilis* C-3102 spores as a probiotic feed supplement on growth performance, noxious gas emission, and intestinal microflora in broilers. *Poultry Science*, *93*(12), 3097–3103. <https://doi.org/10.3382/ps.2014-04086>
- Kaplan, H., & Hutkins, R. W. (2000). Fermentation of fructooligosaccharides by lactic acid bacteria and bifidobacteria. *Applied and Environmental Microbiology*, *66*(6), 2682–2684. <https://doi.org/10.1128/AEM.66.6.2682-2684.2000>
- Kers, J. G., Velkers, F. C., Fischer, E. A. J., Hermes, G. D. A., Stegeman, J. A., & Smidt, H. (2018). Host and environmental factors affecting the intestinal microbiota in chickens. *Frontiers in Microbiology*, *9*(FEB), 1–14. <https://doi.org/10.3389/fmicb.2018.00235>
- Khalil, M. I., & Anwar, M. N. (2016). Isolation, Identification and Characterization of Lactic Acid Bacteria from Milk and Yoghurts. *Research & Reviews: Journal of Food and Dairy Technology*, *4*(3), 17–26.
- Khedid, K., Faid, M., Mokhtari, A., Soulaymani, A., & Zinedine, A. (2009). Characterization of lactic acid bacteria isolated from the one humped camel milk produced in Morocco. *Microbiological Research*, *164*(1), 81–91. <https://doi.org/10.1016/j.micres.2006.10.008>
- Khosravi, A., Boldaji, F., Dastar, B., & Karimi Torshizi, M. A. (2017). Symbiosis between *Enterococcus faecium* DSM 3530 and fructan compounds of different degree of polymerization: A preliminary in vitro assay in a condition simulated chicken caecum. *Journal of Agricultural Science and Technology*, *19*(3), 603–611.
- Landsman, A., St-Pierre, B., Gibbons, W., Rosales-Leija, M., & Brown, M. (2019). Investigation of the potential effects of host genetics and probiotic treatment on the gut bacterial community composition of aquaculture-raised pacific whiteleg shrimp, *litopenaeus vannamei*. *Microorganisms*, *7*(8). <https://doi.org/10.3390/microorganisms7080217>
- Langata, L. M., Maingi, J. M., Musonye, H. A., Kiiru, J., & Nyamache, A. K. (2019). Antimicrobial resistance genes in *Salmonella* and *Escherichia coli* isolates from chicken droppings in Nairobi, Kenya. *BMC Research Notes*, *12*(1), 1–6. <https://doi.org/10.1186/s13104-019-4068-8>
- Latha, S., Vinothini, G., John Dickson Calvin, D., & Dhanasekaran, D. (2016). In vitro probiotic profile based selection of indigenous actinobacterial probiont *Streptomyces* sp. JD9 for enhanced broiler production. *Journal of Bioscience and Bioengineering*, *121*(1), 124–131. <https://doi.org/10.1016/j.jbiosc.2015.04.019>
- Lei, X., Piao, X., Ru, Y., Zhang, H., Péron, A., & Zhang, H. (2015). Effect of *Bacillus amyloliquefaciens*-based direct-fed microbial on performance, nutrient utilization, intestinal morphology and cecal microflora in broiler chickens. *Asian-Australasian Journal of Animal Sciences*, *28*(2), 239–246. <https://doi.org/10.5713/ajas.14.0330>

- Leong, S. S., Lihan, S., Yee, L. T., Chuan, C. H., & Koon, L. C. (2013). Isolation and Characterization of Antibiotic Resistant Bacteria from Swiftlet Feces in Swiftlet Farm Houses in Sarawak , Malaysia Isolation and Characterization of Antibiotic Resistant Bacteria from Swiftlet Feces in Swiftlet Farm Houses in Sarawak , Mala. *Microbiology Indonesia*, 7(4), 137–143. <https://doi.org/10.5454/mi.7.4.1>
- Lescat, M., Clermont, O., Woerther, P. L., Glodt, J., Dion, S., Skurnik, D., ... Denamur, E. (2013). Commensal *Escherichia coli* strains in Guiana reveal a high genetic diversity with host-dependant population structure. *Environmental Microbiology Reports*, 5(1), 49–57. <https://doi.org/10.1111/j.1758-2229.2012.00374.x>
- Lin, W. H., Yu, B., Jang, S. H., & Tsen, H. Y. (2007). Different probiotic properties for *Lactobacillus fermentum* strains isolated from swine and poultry. *Anaerobe*, 13(3–4), 107–113. <https://doi.org/10.1016/j.anaerobe.2007.04.006>
- Loh, T. C. (2002). Livestock production and the feed industry in Malaysia. In *Protein Sources for the Animal Feed Industry* (pp. 329–339).
- M.R., M., Ainy M., N., Fatimah U.Z.A, U., L.C., C., S., M., & M., K. (2018). Occurrence of Tetracyclines , Sulphonamides and Quinolones Residues in Chicken Meat Sample from Selected Chicken Slaughterhouses in Peninsular Malaysia. *Malaysian Journal of Verterinary Research*, 9(2), 88–101.
- Magnusson, J., Ström, K., Roos, S., Sjögren, J., & Schnürer, J. (2003). Broad and complex antifungal activity among environmental isolates of lactic acid bacteria. *FEMS Microbiology Letters*, 219(1), 129–135. [https://doi.org/10.1016/S0378-1097\(02\)01207-7](https://doi.org/10.1016/S0378-1097(02)01207-7)
- Mahfuz, S. U., Nahar, M. J., Mo, C., Ganfu, Z., Zhongjun, L., & Hui, S. (2017). Inclusion of probiotic on chicken performance and immunity: A review. *International Journal of Poultry Science*. <https://doi.org/10.3923/ijps.2017.328.335>
- Majidi-Mosleh, A., Sadeghi, A. A., Mousavi, S. N., Chamani, M., & Zarei, A. (2017). Performance Parameters , Jejunal Bacterial Population and Mucin Gene Expression in Broiler Chicken. *Brazilian Journal of Poultry Science*, 97–102.
- Maldonado, N. C., Ficoesco, C. A., Mansilla, F. I., Melián, C., Hébert, E. M., Vignolo, G. M., & Nader-Macías, M. E. F. (2018). Identification, characterization and selection of autochthonous lactic acid bacteria as probiotic for feedlot cattle. *Livestock Science*, 212(April), 99–110. <https://doi.org/10.1016/j.livsci.2018.04.003>
- Manning, J., Gole, V., & Chousalkar, K. (2015). Screening for *Salmonella* in backyard chickens. *Preventive Veterinary Medicine*, 120(2), 241–245. <https://doi.org/10.1016/j.prevetmed.2015.03.019>
- Mao, X. F., Piao, X. S., Lai, C. H., Li, D. F., Xing, J. J., & Shi, B. L. (2005). Effects of  $\beta$ -glucan obtained from the Chinese herb *Astragalus membranaceus* and lipopolysaccharide challenge on performance, immunological, adrenal, and somatotropic responses of weanling pigs. *Journal of Animal Science*, 83(12), 2775–2782. <https://doi.org/10.2527/2005.83122775x>

- Maria Tufail. (2011). Isolation and evaluation of antibacterial activity of bacteriocin produced by *Lactobacillus bulgaricus* from yogurt. *African Journal of Microbiology Research*, 5(22). <https://doi.org/10.5897/ajmr11.846>
- Mehdi, Y., Létourneau-Montminy, M. P., Gaucher, M. Lou, Chorfi, Y., Suresh, G., Rouissi, T., ... Godbout, S. (2018). Use of antibiotics in broiler production: Global impacts and alternatives. *Animal Nutrition*. <https://doi.org/10.1016/j.aninu.2018.03.002>
- Menconi, A., Morgan, M. J., Pumford, N. R., Hargis, B. M., & Tellez, G. (2013). Physiological Properties and Salmonella Growth Inhibition of Probiotic Bacillus Strains Isolated from Environmental and Poultry Sources, 2013.
- Mensink, M. A., Frijlink, H. W., Van Der Voort Maarschalk, K., & Hinrichs, W. L. J. (2015). Inulin, a flexible oligosaccharide I: Review of its physicochemical characteristics. *Carbohydrate Polymers*. Elsevier Ltd. <https://doi.org/10.1016/j.carbpol.2015.05.026>
- Minekus, M., Alminger, M., Alvito, P., Ballance, S., Bohn, T., Bourlieu, C., ... Brodtkorb, A. (2014). A standardised static in vitro digestion method suitable for food-an international consensus. *Food and Function*, 5(6), 1113–1124. <https://doi.org/10.1039/c3fo60702j>
- Mojgani, N., Hussaini, F., & Vaseji, N. (2015). Characterization of Indigenous *Lactobacillus* Strains for Probiotic Properties. *Jundishapur Journal of Microbiology*, 8(2), 2–8. <https://doi.org/10.5812/jjm.17523>
- Mottet, A., & Tempio, G. (2017). Reviews Global poultry production : current state and future outlook and challenges. *World's Poultry Science Journal*, 73(June), 1–12. <https://doi.org/10.1017/S0043933917000071>
- Mungroo, N. A., & Neethirajan, S. (2014). Biosensors for the detection of antibiotics in poultry industry-A Review. *Biosensors*, 4(4), 472–493. <https://doi.org/10.3390/bios4040472>
- Musikasang, H., Sohsomboon, N., Tani, A., & Maneerat, S. (2012). Bacteriocin-producing lactic acid bacteria as a probiotic potential from Thai indigenous chickens. *Czech Journal of Animal Science*, 57, 137–149. <https://doi.org/10.17221/5568-CJAS>
- Musikasang, H., Tani, A., H-kittikun, A., & Maneerat, S. (2009). Probiotic potential of lactic acid bacteria isolated from chicken gastrointestinal digestive tract. *World J Microbiol Biotechnol* (2009), 1337–1345. <https://doi.org/10.1007/s11274-009-0020-8>
- Nami, Y., Haghshenas, B., & Khosroushahi, A. Y. (2018). Molecular identification and probiotic potential characterization of lactic acid bacteria isolated from human vaginal microbiota. *Advanced Pharmaceutical Bulletin*, 8(4), 683–695. <https://doi.org/10.15171/apb.2018.077>

- Nematollahi, A., Moghaddam, G., & Pourabad, F. (2009). Prevalence of Eimeria Species Among Broiler Chicks in Tabriz (Northwest of Iran). *Munis Entomology & Zoology*, 4(1), 53–58.
- Noohi, N., Ebrahimipour, G., Rohani, M., Talebi, M., & Pourshafie, M. R. (2014). Phenotypic characteristics and probiotic potentials of Lactobacillus spp. isolated from poultry. *Jundishapur Journal of Microbiology*, 7(9), 1–6. <https://doi.org/10.5812/jjm.17824>
- Ohimain, E. I., & Ofongo, R. T. S. (2012). The Effect of Probiotic and Prebiotic Feed Supplementation on Chicken Health and Gut Microflora : A Review. *International Journal of Animal and Veterinary Advances*, 4(2), 135–143.
- Onishi, J. C., Campbell, S., Moreau, M., Patel, F., Brooks, A. I., Xiuzhou, Y., ... Storch, J. (2017). Bacterial communities in the small intestine respond differently to those in the caecum and colon in mice fed low- and high-fat diets. *Microbiology (United Kingdom)*, 163(8), 1189–1197. <https://doi.org/10.1099/mic.0.000496>
- Pakpinyo, S., & Sasipreeyajan, J. (2007). Molecular characterization and determination of antimicrobial resistance of Mycoplasma gallisepticum isolated from chickens. *Veterinary Microbiology*, 125(1–2), 59–65. <https://doi.org/10.1016/j.vetmic.2007.05.011>
- Pan, D., & Yu, Z. (2013). Intestinal microbiome of poultry and its interaction with host and diet. *Gut Microbes*, 5(1). <https://doi.org/10.4161/gmic.26945>
- Pham, T. N. L., Le, T. B., & Yoshimi, B. (2003). Impact of two probiotic Lactobacillus strains feeding on fecal lactobacilli and weight gains in chicken. *J. Gen. Appl. Microbiol*, 29–36.
- Pineda-Quiroga, C., Camarinha-Silva, A., Atxaerandio, R., Ruiz, R., & García-Rodríguez, A. (2017). Changes in broiler performance, duodenal histomorphometry, and caeca microbiota composition in response to wheat-barley based diets supplemented with non-antibiotic additives. *Animal Feed Science and Technology*, 234, 1–9. <https://doi.org/10.1016/j.anifeedsci.2017.09.002>
- Posch, J., Feierl, G., Wuest, G., Sixl, W., Schmidt, S., Haas, D., ... Marth, E. (2006). Transmission of Campylobacter spp. in a poultry slaughterhouse and genetic characterisation of the isolates by pulsed-field gel electrophoresis. *British Poultry Science*, 47(3), 286–293. <https://doi.org/10.1080/00071660600753763>
- Pourabedin, M., & Zhao, X. (2015). Prebiotics and gut microbiota in chickens. *FEMS Microbiology Letters*, 362(15), 1–8. <https://doi.org/10.1093/femsle/fnv122>
- Pradipta, M. S. I., Harimurti, S., & Hadisaputro, W. (2017). Microencapsulation of Indigenous Poultry Lactic Acid Bacteria Probiotic on the Competitive Exclusion against Salmonella Enteritidis and Escherichia Coli in vitro. *Buletin Peternakan*, 41(2), 134. <https://doi.org/10.21059/buletinpeternak.v41i2.17017>

- Putra, T. F., Suprpto, H., Tjahjaningsih, W., & Pramono, H. (2018). The antagonistic activity of lactic acid bacteria isolated from peda, an Indonesian traditional fermented fish. In *IOP Conference Series: Earth and Environmental Science* (Vol. 137). <https://doi.org/10.1088/1755-1315/137/1/012060>
- R. Majidzadeh Heravi. (2011). Screening of lactobacilli bacteria isolated from gastrointestinal tract of broiler chickens for their use as probiotic. *African Journal of Microbiology Research*, 5(14), 1858–1868. <https://doi.org/10.5897/ajmr11.416>
- Rajoka, M. S. R., Hayat, H. F., Sarwar, S., Mehwish, H. M., Ahmad, F., Hussain, N., ... Shi, J. (2018). Isolation and evaluation of probiotic potential of lactic acid bacteria isolated from poultry intestine. *Microbiology (Russian Federation)*, 87(1), 116–126. <https://doi.org/10.1134/S0026261718010150>
- Rebolé, A., Ortiz, L. T., Rodríguez, M. L., Alzueta, C., Treviño, J., & Velasco, S. (2010). Effects of inulin and enzyme complex, individually or in combination, on growth performance, intestinal microflora, cecal fermentation characteristics, and jejunal histomorphology in broiler chickens fed a wheat- and barley-based diet. *Poultry Science*, 89, 276–286. <https://doi.org/10.3382/ps.2009-00336>
- Réhault-Godbert, S., Guyot, N., & Nys, Y. (2019). The golden egg: Nutritional value, bioactivities, and emerging benefits for human health. *Nutrients*, 11(3), 1–26. <https://doi.org/10.3390/nu11030684>
- Rehman, H. U., Vahjen, W., Awad, W. A., & Zentek, J. (2007). Indigenous bacteria and bacterial metabolic products in the gastrointestinal tract of broiler chickens. *Archives of Animal Nutrition*, 61(5), 319–335. <https://doi.org/10.1080/17450390701556817>
- Rehman, H. U., Vahjen, W., Awad, W. A., & Zentek, P. J. (2016). Archives of Animal Nutrition Indigenous bacteria and bacterial metabolic products in the gastrointestinal tract of broiler chickens. *Archives of Animal Nutrition*, 2817(October). <https://doi.org/10.1080/17450390701556817>
- Rehman, T. U., Khan, M. N., Sajid, M. S., Abbas, R. Z., Arshad, M., Iqbal, Z., & Iqbal, A. (2011). Epidemiology of Eimeria and associated risk factors in cattle of district Toba Tek Singh, Pakistan. *Parasitology Research*, 108(5), 1171–1177. <https://doi.org/10.1007/s00436-010-2159-5>
- Reuben, R. C., Roy, P. C., Sarkar, S. L., Alam, R. U., & Jahid, I. K. (2019). Isolation, characterization, and assessment of lactic acid bacteria toward their selection as poultry probiotics. *BMC Microbiology*, 19(1), 253. <https://doi.org/10.1186/s12866-019-1626-0>
- Ríos-Covián, D., Ruas-Madiedo, P., Margolles, A., Gueimonde, M., De los Reyes-Gavilán, C. G., & Salazar, N. (2016). Intestinal short chain fatty acids and their link with diet and human health. *Frontiers in Microbiology*. <https://doi.org/10.3389/fmicb.2016.00185>



- Rioux, K. P., Madsen, K. L., & Fedorak, R. N. (2005). The role of enteric microflora in inflammatory bowel disease: Human and animal studies with probiotics and prebiotics. *Gastroenterology Clinics of North America*. <https://doi.org/10.1016/j.gtc.2005.05.005>
- Ritzi, M. M., Abdelrahman, W., Mohnl, M., & Dalloul, R. A. (2014). Effects of probiotics and application methods on performance and response of broiler chickens to an *Eimeria* challenge. *Poultry Science*, *93*, 2772–2778. <https://doi.org/10.3920/BM2016.0080>
- Roy, P. C., & Sarkar, S. L. (2020). Characterization and evaluation of lactic acid bacteria from indigenous raw milk for potential probiotic properties. *Journal of Dairy Science*, *2022*. <https://doi.org/10.3168/jds.2019-17092>
- Salim, H. M., Kang, H. K., Akter, N., Kim, D. W., Kim, J. H., Kim, M. J., ... Kim, W. K. (2013). Supplementation of direct-fed microbials as an alternative to antibiotic on growth performance, immune response, cecal microbial population, and ileal morphology of broiler chickens. *Poultry Science*, *92*(8), 2084–2090. <https://doi.org/10.3382/ps.2012-02947>
- Samanta, A. K., Jayapal, N., Senani, S., Kolte, A. P., & Sridhar, M. (2013). Prebiotic inulin: Useful dietary adjuncts to manipulate the livestock gut microflora. *Brazilian Journal of Microbiology*. <https://doi.org/10.1590/S1517-83822013005000023>
- Samsuddin, N. S., Sharaai, A. H., & Ismail, M. M. (2015). Sustainability of Chicken Meat Production in achieving Food Security in Malaysia. *Advances in Environmental Biology*, *9*(23), 1–6. Retrieved from <http://www.aensiweb.com/AEB/>
- Seal, B. S., Lillehoj, H. S., Donovan, D. M., & Gay, C. G. (2013). Alternatives to antibiotics: a symposium on the challenges and solutions for animal production. *Animal Health Research Reviews / Conference of Research Workers in Animal Diseases*, *14*(1), 78–87. <https://doi.org/10.1017/S1466252313000030>
- Sergeant, M. J., Constantinidou, C., Cogan, T. A., Bedford, M. R., Penn, C. W., & Pallen, M. J. (2014). Extensive microbial and functional diversity within the chicken cecal microbiome. *PLoS ONE*, *9*(3). <https://doi.org/10.1371/journal.pone.0091941>
- Shafer, A. L., Hovick, C. a, Gatewood, D. M., Leader, S., Hyde, R. L. W., & Management, Q. (2015). Standard Operating Policy/Procedure Standard Bacterial Plate Count. *United States Department of Agriculture Center for Veterinary Biologics*, 1–7.
- Shakoor, G., Akbar, A., Samad, A., Khan, S. A., Ur, F., Shakoor, M., & Ahmad, D. (2017). Isolation of lactic acid bacteria from chicken gut and its probiotic potential characterization. *International Journal of Biosciences*, *6655*, 1–9.

- Sharma, R., Sharma, S., Yadav, V., Shukla, P. C., Sharma, V., Baghel, R., ... Pradhan, S. (2018). Microbial and functional feed supplement to improve livestock and poultry productivity with special reference to synbiotics: A review. *The Pharma Innovation Journal*, 7(7), 62–68. Retrieved from [www.thepharmajournal.com](http://www.thepharmajournal.com)
- Shirzad, M. R., Seifi, S., Gheisari, H. R., Hachesoo, B. A., Habibi, H., & Bujmehrani, H. (2011). Prevalence and risk factors for subclinical coccidiosis in broiler chicken farms in Mazandaran province, Iran. *Tropical Animal Health and Production*, 43(8), 1601–1604. <https://doi.org/10.1007/s11250-011-9876-3>
- Silbergeld, E. K., Graham, J., & Price, L. B. (2008). Industrial Food Animal Production, Antimicrobial Resistance, and Human Health. *Annual Review of Public Health*, 29(1), 151–169. <https://doi.org/10.1146/annurev.publhealth.29.020907.090904>
- Skarp, C. P. A., Hänninen, M. L., & Rautelin, H. I. K. (2016). Campylobacteriosis: The role of poultry meat. *Clinical Microbiology and Infection*, 22(2), 103–109. <https://doi.org/10.1016/j.cmi.2015.11.019>
- Stanley, D., Geier, M. S., Chen, H., Hughes, R. J., & Moore, R. J. (2015). Comparison of fecal and cecal microbiotas reveals qualitative similarities but quantitative differences. *BMC Microbiology*, 15(1), 1–11. <https://doi.org/10.1186/s12866-015-0388-6>
- Stanley, D., Hughes, R. J., & Moore, R. J. (2014). Microbiota of the chicken gastrointestinal tract: Influence on health, productivity and disease. *Applied Microbiology and Biotechnology*. <https://doi.org/10.1007/s00253-014-5646-2>
- Švec, P., Vancanneyt, M., Seman, M., Snauwaert, C., Lefebvre, K., Sedláček, I., & Swings, J. (2005). Evaluation of (GTG)5-PCR for identification of *Enterococcus* spp. *FEMS Microbiology Letters*, 247(1), 59–63. <https://doi.org/10.1016/j.femsle.2005.04.030>
- Svihus, B. (2014). Function of the digestive system. *Journal of Applied Poultry Research*, 23(2), 306–314. <https://doi.org/10.3382/japr.2014-00937>
- Taheri, H. R., Moravej, H., Tabandeh, F., Zaghari, M., & Shivazad, M. (2009). Screening of lactic acid bacteria toward their selection as a source of chicken probiotic. *Poultry Science*, 88(8), 1586–1593. <https://doi.org/10.3382/ps.2009-00041>
- Tang, C., Hoo, P. C. X., Tan, L. T. H., Pusparajah, P., Khan, T. M., Lee, L. H., ... Chan, K. G. (2016). Golden needle mushroom: A culinary medicine with evidenced-based biological activities and health promoting properties. *Frontiers in Pharmacology*, 7(DEC). <https://doi.org/10.3389/fphar.2016.00474>
- Tengku Abdul Hamid, T. H., & Ezani, E. (2015). Isolation of Lactic Acid Bacteria from Malaysian Non-Broiler Chicken (*Gallus Gallus*) Intestine with Potential Probiotic for Broiler Feeding. *IJUM Engineering Journal, Special Issue on Biotechnology*, (April).

- Torok, V. A., Allison, G. E., Percy, N. J., Ophel-Keller, K., & Hughes, R. J. (2011). Influence of antimicrobial feed additives on broiler commensal posthatch gut microbiota development and performance. *Applied and Environmental Microbiology*, 77(10), 3380–3390. <https://doi.org/10.1128/AEM.02300-10>
- Tserovska, L., Stefanova, S., & Yordanova, T. (2002). Identification of Lactic Acid Bacteria Isolated From Katyk , Goat ' S Milk and Cheese. *Journal of Culture Collections*, 3, 48–52.
- Waite, D. W., & Taylor, M. W. (2014). Characterizing the avian gut microbiota: Membership, driving influences, and potential function. *Frontiers in Microbiology*, 5(MAY), 1–12. <https://doi.org/10.3389/fmicb.2014.00223>
- Wallace, R. J., Oleszek, W., Franz, C., Hahn, I., Baser, K. H. C., Mathe, A., & Teichmann, K. (2010). Dietary plant bioactives for poultry health and productivity. *British Poultry Science*, 51(4), 461–487. <https://doi.org/10.1080/00071668.2010.506908>
- Walter, J. (2008). Ecological role of lactobacilli in the gastrointestinal tract: Implications for fundamental and biomedical research. *Applied and Environmental Microbiology*, 74(16), 4985–4996. <https://doi.org/10.1128/AEM.00753-08>
- Whaley, S. E., Sigman, M., Neumann, C., Bwibo, N., Guthrie, D., Weiss, R. E., ... Murphy, S. P. (2003). Animal Source Foods to Improve Micronutrient Nutrition and Human Function in Developing Countries. *J. Nutr*, 133, 3965–3971.
- Wichienchot, S., Thammarutwasik, P., Jongjareonrak, A., Chansuwan, W., Hmadhlu, P., Hongpattarakere, T., ... Ooraikul, B. (2011). Extraction and analysis of prebiotics from selected plants from southern Thailand. *Songklanakarinn Journal of Science and Technology*, 33(5), 517–522.
- Yegani, M., & Korver, D. R. (2008). Factors affecting intestinal health in poultry. *Poultry Science*, 87(10), 2052–2063. <https://doi.org/10.3382/ps.2008-00091>
- Yeoman, C. J., Chia, N., Jeraldo, P., Sipos, M., Goldenfeld, N. D., & White, B. A. (2012). The microbiome of the chicken gastrointestinal tract. *Animal Health Research Reviews / Conference of Research Workers in Animal Diseases*, 13(1), 89–99. <https://doi.org/10.1017/S1466252312000138>
- ZaidiZaidi, N. A., Abdul Hamid, A. A., & Tengku Abdul Hamid, T. H. (2017). Lactic acid bacteria with antimicrobial properties isolated from the intestines of Japanese quail (*Coturnix Coturnix Japonica*). *Science Heritage Journal*, 1(1), 10–12. <https://doi.org/10.26480/gws.01.2017.10.12>
- Zakaria, Z., Afandi, A. A., Mohd Noor, S. N., Hussin, N., & Shahidan, N. (2018). Prebiotic Activity Score Of Breadfruit Resistant Starch (*Artocarpus Altilis*), Breadfruit Flour, And Inulin during In-Vitro Fermentation By Pure Cultures (*Lactobacillus Plantarum*, And *Bifidobacterium Bifidum*). *Journal Of Agrobiotechnology*, 9(1S), 122–131.

## BIODATA OF STUDENT

Arlene Debbie anak Lingoh was born and raised in Tatau, Sarawak. She is currently residing at Tatau, Bintulu, Sarawak. Previously, she was an undergraduate student of Universiti Malaysia Sarawak (UNIMAS) and obtained Bachelor of Science in Resource Chemistry. Currently, she is a graduate student in Master of Science in Universiti Putra Malaysia.

