



UNIVERSITI PUTRA MALAYSIA

***CHARACTERIZATION OF MICROBIAL FUNCTIONAL DIVERSITY AND
BACTERIAL COMMUNITY STRUCTURE AFFECTED BY FERTILISER,
SOIL AND PLANT GROWTH STAGES IN AEROBIC RICE***

NOR AYSHAH ALIA BINTI ALI HASSAN

IPTSM 2021 19



**CHARACTERIZATION OF MICROBIAL FUNCTIONAL DIVERSITY AND
BACTERIAL COMMUNITY STRUCTURE AFFECTED BY FERTILISER, SOIL
AND PLANT GROWTH STAGES IN AEROBIC RICE**

By

NOR AYSHAH ALIA BINTI ALI HASSAN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Degree of
Doctor of Philosophy**

December 2020

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

I dedicate this thesis to my family and to all the people that contributed toward the successful completion of my PhD programme.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

CHARACTERIZATION OF MICROBIAL FUNCTIONAL DIVERSITY AND BACTERIAL COMMUNITY STRUCTURE AFFECTED BY FERTILISER, SOIL AND PLANT GROWTH STAGES IN AEROBIC RICE

By

NOR AYSHAH ALIA BINTI ALI HASSAN

December 2020

Chairman : Associate Professor Halimi bin Mohd Saud, PhD
Institute : Tropical Agriculture and Food Security

Aerobic rice was introduced to increase water use efficiency. The rhizosphere of aerobic rice differs from that of conventional flooded rice due to their habitat differences. Recognizing the importance of microorganism to ecosystem function, bacterial community in aerobic rice rhizosphere was studied as a fundamental step towards the improvement of aerobic rice. Inorganic fertiliser could affect the microbial functional diversity and bacterial community structure of rhizosphere microorganism, which play a vital role in plant growth and soil fertility. Hence, the aim of this study was to determine the effect of fertiliser and soil type on microbial functional diversity bacterial community in aerobic rice rhizosphere. To achieve this, a total of 3 specific objectives was conducted. The first objective is to assess the effect of fertiliser rate and soil type on microbial functional diversity and bacterial community structure. The second objective is to determine the influence of fertiliser and soil type on the bacterial community composition and nitrogen cycling functional potential in the rhizosphere of aerobic rice. The third objective for this study is to evaluate the effect of plant growth stages on the microbial community in the rhizosphere of aerobic rice. Aerobic rice variety MR1A 1 was chosen for this study. A total of 4 different fertiliser rate was selected and tested with 2 different soil types. Non-fertilised, 100 kg/ha, 200 kg/ha, and 400 kg/ha of NPK compound fertiliser were applied to peat and sandy clay loam soil. The soil properties tested for this study were pH, soil moisture, cation exchange capacity, soluble phosphorus, total carbon, total nitrogen, and mechanical analysis. Plant growth characteristic recorded were plant length, numbers of tillers per hill, panicle length and straw nitrogen concentration. Microbial functional diversity was characterized by assessing the microbial activity and community level physiological profile using Biolog Ecoplate™ system. 16s amplicon sequencing was used to determined bacterial community structure, bacterial community composition, and nitrogen cycling functional potential. RDP FunGene was used to predict the functional potential of bacterial communities. Aerobic culturable microbial abundance was

performed using nutrient agar, Burk's agar, potato dextrose agar, Pikovskaya's agar, and starch casein agar was used to quantify the abundance of bacteria, nitrogen-fixing microbes, fungi, phosphate-solubilizing microbes and actinomycetes, respectively. For the plant growth stages, samples were taken during the tillering, booting, grain filling, and mature stage. Microbial community were estimated using physiological fingerprints and microbial culturable abundance. Although peat and sandy clay loam were impacted differently when applied with different fertiliser rate, microbial functional diversity responded positively for both soils. The distinct microbial evenness between peat and sandy clay loam suggested that microbial functional diversity is dependent on soil properties. Carboxylic acid and acetic acid as well as carbohydrates were sensitive carbon sources that create dissimilar metabolic function of rhizosphere bacterial community in aerobic rice. L-arginine, L-asparagine, L-serine, α -cyclodextrin, N-acetylglucosamine and α -D-lactose were good indicators for distinguishing rhizosphere that added with fertiliser. 16s amplicon sequencing analysis demonstrated that *Alphaproteobacteria*, *Nitrospirae* and *Bacteroidetes* were identified as critical phyla when fertiliser added in peat while *Gammaproteobacteria* was the crucial phylum in sandy clay loam. *Acidobacteria-6* and *Chloracidobacteria* were positive indicator showing that excessive fertiliser was applied in both soils. The addition of all fertiliser rate tested influenced the bacterial community composition by shifting the communities. This was shown through the increase of abundance by *Candidatus Nitrososphaera* and the decrease of abundance by *Dok59*, *Olivibacter*, *Thermomonas* and *Dactylosporangium* in peat. The bacterial communities in the sandy clay loam were shifted and shown by the increase of *Clostridia* and *Solirubrobacter*. The changes in bacterial communities such as increase and decrease of type of genera were dependent on soil chemical and physical properties. Although, rhizosphere bacterial communities were different in peat and sandy clay loam, the function of the genera in the communities was the same after fertiliser was introduced. The selection of genera during the bacterial communities' shift were dedicated for plant development but depended on soil type particularly soil properties. *Desulfovibrio* was identified as a genus that presence only in nonfertilised rhizosphere and can be used as an indicator in natural rhizosphere. *Chitinophaga*, *Desulfosporosinus*, *Geobacillus*, *Nannocystis*, *Nitrosovibrio*, and *Perlucidibaca* were the genus that solely presence in fertilised rhizosphere and can be used as an indicator in fertilised rhizosphere. Genera that solely presence in the fertilised rhizosphere were mainly contributed to nitrogen cycling. The growth stage of aerobic rice has an effect on determining the response and composition of microbial community in soil. Microbial community in rhizosphere responded according to plant effect produced during plant development which is primarily determined by soil type. Soil type particularly the soil physical and chemical properties are important factors in shaping the microbial community by directly influencing the environment of rhizosphere. This study concluded that microbial functional diversity and bacterial community in aerobic rice rhizosphere were influenced by fertiliser, soil type and growth stage. The use of fertiliser in the recommended rate (200 kg/ha of NPK compound fertiliser) is suitable in maintaining the function of bacterial community and thus positively influence the growth of aerobic rice.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

PENCIRIAN KEPELBAGAIAN BERFUNGSI MIKROORGANISMA DAN STRUKTUR KOMUNITI BAKTERIA YANG TERKESAN OLEH BAJA, TANAH DAN PERINGKAT PERTUMBUHAN PADI AEROB

Oleh

NOR AYSHAH ALIA BINTI ALI HASSAN

Disember 2020

Pengerusi : Profesor Madya Halimi bin Mohd Saud, PhD
Institut : Pertanian Tropika dan Sekuriti Makanan

Padi aerob diperkenalkan bagi meningkatkan kecekapan penggunaan air. Menyedari kepentingan mikroorganisma di dalam fungsi ekosistem, komuniti mikroorganisma di rhizosphere padi aerob dikaji sebagai langkah asas bagi menambahbaik padi aerob. Baja inorganik boleh mempengaruhi kepelbagaian berfungsi mikroorganisma dan struktur komuniti mikroorganisma di rhizosphere di mana mikroorganisma ini memainkan peranan penting kepada pertumbuhan tanaman dan kesuburan tanah. Oleh itu, kajian ini bertujuan untuk mengenalpasti kesan baja dan jenis tanah terhadap komuniti mikroorganisma di rhizosphere padi aerob. Untuk mencapai matlamat ini, sebanyak 3 objektif spesifik telah ditentukan. Objektif pertama ialah untuk menilai kesan kadar baja dan jenis tanah kepada kepelbagaian berfungsi mikroorganisma dan struktur komuniti mikroorganisma. Varieti padi aerob MRIA 1 telah dipilih bagi kajian ini. Sebanyak empat kadar baja berbeza dipilih dan diuji dengan 2 jenis tanah. Tiada baja, 100 kg/ha, 200 kg/ha, dan 400 kg/ha baja kompaun diaplikasi kepada tanah gambut dan tanah lom liat berpasir. Sifat tanah yang diuji adalah pH, kelembapan, kapasiti pertukaran kation, fosforus larut, jumlah karbon, jumlah nitrogen, dan analisis mekanikal. Kepelbagaian berfungsi mikroorganisma dicirikan dengan menilai aktiviti mikroorganisma dan profil fisiologi tahap komuniti dengan menggunakan sistem Biolog Ecoplate™. Penjujukan 16s amplikon digunakan bagi mengenalpasti struktur komuniti mikroorganisma. Kelimpahan mikroorganisma aerobik yang boleh dikultur diuji menggunakan agar nutrien, agar Burk's, agar dekstros kentang, agar Pikovskaya's, dan agar kanji kasein digunakan bagi mengukur kelimpahan bakteria, mikroorganisma pengikat nitrogen, kulat, mikroorganisma pelarut fosforus, dan aktinomisit, masing-masing. Kepelbagaian berfungsi mikroorganisma meningkat dengan pertambahan baja di tanah gambut dan tanah lom liat berpasir. Walaupun tanah gambut dan tanah lom liat berpasir dipengaruhi secara berbeza apabila diaplikasi dengan kadar baja yang berbeza, terdapat kesan positif bagi kedua-

dua jenis tanah ini. Terdapat beberapa spesies tertentu meningkat di dalam komuniti mikroorganisma namun terdapat juga spesies yang mengurangkan bagi adaptasi kepada perubahan dan ini telah mengekalkan kepelbagaian berfungsi mikroorganisma. Terdapat perubahan pada penggunaan sumber karbon selepas penambahan baja. Asid karboksilik dan asid asetik serta karbohidrat merupakan sumber karbon yang sensitif di dalam menghasilkan fungsi metabolik yang berbeza di dalam komuniti mikroorganisma rhizosphere padi aerob. L-arginine, L-asparagine, L-serine, α -cyclodextrin, N-acetylglucosamine dan α -D-lactose merupakan penunjuk yang bagus bagi membezakan rhizosphere yang ditambah dengan baja. *Alphaproteobacteria*, *Nitrospirae* and *Bacteroidetes* dikenalpasti sebagai phyla yang kritikal apabila baja ditambah di dalam tanah gambut manakala *Gammaproteobacteria* merupakan phyla penting bagi tanah lom liat berpasir. *Acidobacteria-6* dan *Chloracidobacteria* merupakan penunjuk positif yang menunjukkan baja berlebihan digunakan di dalam kedua-dua jenis tanah. Objektif kedua ialah bagi mengenalpasti pengaruh baja dan jenis tanah kepada komposisi komuniti mikroorganisma dan potensi kitaran nitrogen berfungsi di rhizosphere padi aerob. Pencirian pertumbuhan padi yang diukur adalah panjang pokok, bilangan anak pokok serumpun, panjang panikel, dan kepekatan nitrogen di jerami. Penjujukan 16s ampikon digunakan bagi mengenalpasti komposisi komuniti mikroorganisma, dan potensi kitaran nitrogen berfungsi. Potensi berfungsi komuniti mikroorganisma di ramal dengan menggunakan RDP FunGene. Komposisi komuniti mikroorganisma dipengaruhi oleh penambahan baja melalui pengalihan komuniti. Penambahan baja meningkatkan kelimpahan arkaea terutamanya *Candidatus Nitrososphaera* di tanah gambut. Penambahan baja meningkatkan kelimpahan bakteria di tanah lom liat berpasir terutamanya *Clostridia*, *Gammaproteobacteria* dan *Acidobacteria-6*. Perubahan komuniti mikroorganisma seperti peningkatan dan penurunan jenis genera bergantung kepada sifat kimia dan fizik tanah. Walaupun komuniti mikroorganisma rhizosphere adalah berbeza pada tanah yang berlainan, fungsi genera di dalam komuniti adalah sama selepas baja diperkenalkan. Pemilihan genera semasa peralihan komuniti mikroorganisma ditumpukan untuk pertumbuhan tumbuhan tetapi bergantung kepada jenis tanah terutamanya sifat tanah. *Desulfovibrio* dikenalpasti sebagai genus yang hanya hadir di rhizosphere yang tidak dibaja dan boleh digunakan sebagai penunjuk di rhizosphere semulajadi. *Chitinophaga*, *Desulfosporosinus*, *Geobacillus*, *Nannocystis*, *Nitrosovibrio*, dan *Perlucidibaca* merupakan genus yang hanya hadir di rhizosphere yang dibaja dan boleh digunakan sebagai penunjuk di rhizosphere yang dibaja. Genera yang hanya hadir di rhizosphere yang dibaja menyumbang terutamanya kepada kitaran nitrogen. Objektif ketiga bagi kajian ini ialah menilai kesan peringkat pertumbuhan padi aerob kepada komuniti mikroorganisma di rhizosphere. Bagi peringkat pertumbuhan padi aerob, sampel diambil semasa peringkat vegetatif, pembiakan, masak, dan matang. Pencirian pertumbuhan padi yang diukur adalah panjang pokok, bilangan anak pokok serumpun, panjang panikel, dan kepekatan nitrogen di jerami. Komuniti mikroorganisma di ukur menggunakan cap jari fisiologi dan kelimpahan mikroorganisma yang boleh dikultur. Komuniti mikroorganisma di rhizosphere bertindakbalas mengikut kesan pokok yang dihasilkan semasa pertumbuhan pokok yang mana ditentukan oleh jenis tanah. Jenis tanah terutamanya sifat fizikal dan kimia tanah merupakan faktor penting di dalam membentuk komuniti mikroorganisma dengan mempengaruhi secara langsung persekitaran

rhizosphere. Daripada kajian ini, dapat disimpulkan bahawa penambahan baja dan jenis tanah yang berbeza mampu memberi kesan dan mempengaruhi kepelbagaian berfungsi mikroorganisma dan komuniti mikroorganisma di rhizosphere padi aerob.



ACKNOWLEDGEMENTS

All praises belong to Allah. First and foremost, I would like to express my utmost and humble gratitude to the Almighty Allah for blessing and giving me the patience and making it possible for me to produce this work.

I am deeply grateful to my former and current supervisor associate professor Dr. Radziah Othman and associate professor Dr Halimi Mohd Saud for their constant support and tirelessly advice that inspired me throughout the whole journey. I would also like to thank my committee member Dr Roslan Ismail and Dr Ali Tan Kee Zuan for assisting me through their constructive ideas and expertise.

I wish to express my heartfelt indebtedness to my beloved husband for his continuous support given throughout this study. Thank you for being considerate, patience and kind to me. Though only a toddler, I would like to thank my son who being mature beyond his age and understand the work that need to done by his mother in order to accomplish this study. I sincerely express my gratitude to my parents and my sibling for the prayers, motivation and assistance given to me to complete this study. They been really helpful and eased my insecurity whenever I have one.

I would like to thank my colleagues for being supportive and gave me an insightful comment on my work. I wish to express my gratefulness to my working organization for providing me with the scholarship to pursue my graduate study.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Halimi bin Mohd Saud, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Roslan bin Ismail, PhD

Senior Lecturer
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

Ali Tan Kee Zuan, PhD

Senior Lecturer
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 12 August 2021

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature: _____ Date: _____

Name and Matric No: Nor Ayshah Alia binti Ali Hassan, GS42024

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: _____

Name of Chairman
of Supervisory
Committee:

Associate Professor Dr. Halimi bin Mohd Saud

Signature: _____

Name of Member
of Supervisory
Committee:

Dr. Roslan bin Ismail

Signature: _____

Name of Member
of Supervisory
Committee:

Dr. Ali Tan Kee Zuan

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	vi
APPROVAL	vii
DECLARATION	ix
LIST OF TABLES	xiv
LIST OF FIGURES	xvi
LIST OF ABBREVIATIONS	xix
CHAPTER	
1 INTRODUCTION	1
1.1 Background	1
1.2 Problem Statement	2
1.3 Objectives	4
2 LITERATURE REVIEW	5
2.1 Aerobic rice	5
2.1.1 Research on aerobic rice	6
2.2 Microbial community	7
2.2.1 The response of bacterial community to fertiliser	8
2.2.2 The response of bacterial community to soil properties	10
2.3 Microbial functional diversity	12
2.3.1 The response of microbial functional diversity to fertiliser	13
2.3.2 The response of microbial functional diversity to soil properties	14
3 CHARACTERIZATION OF MICROBIAL FUNCTIONAL DIVERSITY AND BACTERIAL COMMUNITY STRUCTURE IN AEROBIC RICE RHIZOSPHERE	15
3.1 Introduction	15
3.2 Materials and Method	16
3.2.1 Experimental design	16
3.2.2 Rhizosphere sampling	17
3.2.3 Soil properties	17
3.2.4 Microbial functional diversity analysis	17
3.2.5 Microbial activity	18
3.2.6 Community-level physiological profile	18
3.2.7 Carbon source utilization	18
3.2.8 Enumeration of culturable microorganism	19
3.2.9 Statistical analysis	20

	3.2.10	16s amplicon sequencing	20
	3.2.11	Data analysis	21
3.3		Result	21
	3.3.1	Soil properties	21
	3.3.2	Microbial activity	23
	3.3.3	Community-level physiological profile	25
	3.3.4	Carbon source utilization	27
	3.3.5	Population of culturable microorganism	30
	3.3.6	Sequencing data processing	32
	3.3.7	OTU analysis	32
	3.3.8	Species annotation	34
		3.3.8.1 Species relative abundance	34
		3.3.8.2 Krona taxonomy visualization	35
	3.3.9	Alpha diversity	44
	3.3.10	Beta diversity	47
	3.3.11	Correlation between soil physicochemical properties, microbial functional diversity and bacterial community structure	48
3.4		Discussion	50
3.5		Conclusion	60
4		THE INFLUENCE OF FERTILISER AND SOIL TYPE ON THE BACTERIAL COMMUNITY COMPOSITION AND NITROGEN CYCLING FUNCTIONAL POTENTIAL IN AEROBIC RICE RHIZOSPHERE	
			61
	4.1	Introduction	61
	4.2	Materials and Method	62
		4.2.1 Experimental design	62
		4.2.2 Rhizosphere sampling	63
		4.2.3 Plant growth characteristics	63
		4.2.4 16s amplicon sequencing	63
		4.2.5 Data analysis	63
		4.2.6 Statistical analysis	64
		4.2.7 Functional potential	64
	4.3	Results	64
		4.3.1 Plant growth characteristic	64
		4.3.2 Bacterial community composition	66
		4.3.3 Determination of potential keystone species	72
		4.3.4 Functional potential of bacterial community	77
	4.4	Discussion	81
	4.5	Conclusion	87
5		THE EFFECT OF PLANT GROWTH STAGES ON THE MICROBIAL COMMUNITY OF AEROBIC RICE RHIZOSPHERE	
			89
	5.1	Introduction	89
	5.2	Materials and method	89
		5.2.1 Experimental design	89
		5.2.2 Rhizosphere sampling	90

5.2.3	Soil properties	90
5.2.4	Determination of nitrogen in plant tissue	90
5.2.5	Microbial functional diversity analysis	90
5.2.6	Microbial activity	90
5.2.7	Community-level physiological profile	90
5.2.8	Carbon source utilization	91
5.2.9	Enumeration of culturable microorganism	91
5.2.10	Statistical analysis	91
5.3	Result	91
5.3.1	Soil properties	91
5.3.2	Microbial activity	93
5.3.3	Community-level physiological profile	93
5.3.4	Relationship between microbial functional diversity and soil properties	95
5.3.5	Carbon source utilization at different growth stages	99
5.3.6	Population of culturable aerobic microorganism	107
5.4	Discussion	109
5.5	Conclusion	113
6	SUMMARY, CONCLUSIONS AND RECOMMENDATIONS FOR FUTURE RESEARCH	114
6.1	Summary	114
6.2	Conclusion	115
6.3	Recommendations for future research	115
	REFERENCES	117
	BIODATA OF STUDENT	147
	LIST OF PUBLICATIONS	148

LIST OF TABLES

Table	Page
3.1 Substrates in Biolog Ecoplate™ categorized into five functional guilds as according to Weber and Legge (2009)	19
3.2 Soil chemical properties taken at 90 days after sowing of aerobic rice	22
3.3 Soil mechanical analysis taken after 90 days at sowing of aerobic rice	23
3.4 Principal component 1, 2, and 3. Eigenvalues (including percentage of variation), substrates loaded in each PC, substrate type and eigenvector for substrate in each PC obtained from Ecoplates in peat	29
3.5 Principal component 1, 2, and 3. Eigenvalues (including percentage of variation), substrates loaded in each PC, substrate type and eigenvector for substrate in each PC obtained from Ecoplates in sandy clay loam soil	30
3.6 Population of culturable aerobic microorganism	31
3.7 Sequencing data for peat and sandy clay loam soil. Paired-end reads were merged using FLASH. Sequences with 97% similarity were assigned to the same Operational Taxonomy Unit	32
3.8 Alpha indices in peat and sandy clay loam soil calculated based on 97% identity during sequence-clustering by default	45
3.9 Coefficient Correlation using Pearson between selected soil properties, microbial functional diversity and bacterial community structure in peat	49
3.10 Coefficient Correlation using Pearson between selected soil properties, microbial functional diversity and bacterial community structure in sandy clay loam soil	50
4.1 Plant growth characteristics according to different fertiliser rate and soil type. The plant growth was measured at 90 days after sowing of aerobic rice	65

4.2	Principal component 1, 2, and 3. Eigenvalues (including percentage of variation), genus loaded in each PC, phylum and eigenvector for genus in each PC obtained from 16s amplicon sequencing in peat	68
4.3	Principal component 1, 2, and 3. Eigenvalues (including percentage of variation), genus loaded in each PC, phylum and eigenvector for genus in each PC obtained from 16s amplicon sequencing in sandy clay loam	70
4.4	Functional annotation of genera in peat and sandy clay loam predicted using RDP FunGene. Genes related to nitrogen cycling was used for annotation	78
5.1	Soil physical and chemical properties of peat and sandy clay loam taken during the aerobic rice growth	92
5.2	Coefficient Correlation using Pearson between selected soil properties and microbial functional diversity in peat soil	96
5.3	Coefficient Correlation using Pearson between selected soil properties and microbial functional diversity in sandy clay loam	98
5.4	The main carbon source utilized according to the guild at every growth stage after 48 hours incubation. The highest value of optical density for each guild were selected as the main carbon source representing the guild	102
5.5	Coefficient Correlation using Pearson between carbon source guilds and microbial functional diversity in peat soil	104
5.6	Coefficient Correlation using Pearson between carbon source guilds and microbial functional diversity in sandy clay loam soil	106
5.7	Population of culturable aerobic microorganism according to aerobic rice growth stage in peat and sandy clay loam	108

LIST OF FIGURES

Figure	Page
3.1 Microbial activity in peat (a) and sandy clay loam soil (b)	24
3.2 Community-level physiological profile in peat is evaluated by measuring species richness (a), Shannon Weaver Index (b) and Shannon Evenness (c)	26
3.3 Community-level physiological profile in sandy clay loam soil is evaluated by measuring species richness (a), Shannon Weaver Index (b) and Shannon Evenness (c)	27
3.4 Carbon source utilization in peat (a) and sandy clay loam soil (b)	28
3.5 Shared and unique OTU in peat (a) and sandy clay loam soil (b)	33
3.6 Species relative abundance in peat and sandy clay loam soil	34
3.7 Taxonomic rank distribution of <i>Proteobacteria</i> in peat treated with non-fertilised peat (a), 100 kg/ha NPK compound fertiliser (b), 200 kg/ha NPK compound fertiliser (c), and 400 kg/ha NPK compound fertiliser. Circles from inside to outside represent different classification level. Area of sector indicates respective proportion of different OTU annotation	35
3.8 Taxonomic rank distribution of <i>Proteobacteria</i> in sandy clay loam soil treated with non-fertilised sandy clay loam soil (a), 100 kg/ha NPK compound fertiliser (b), 200 kg/ha NPK compound fertiliser (c), and 400 kg/ha NPK compound fertiliser. Circles from inside to outside represent different classification level. Area of sector indicates respective proportion of different OTU annotation	36
3.9 Taxonomic rank distribution of <i>Acidobacteria</i> in peat treated with non-fertilised peat (a), 100 kg/ha NPK compound fertiliser (b), 200 kg/ha NPK compound fertiliser (c), and 400 kg/ha NPK compound fertiliser. Circles from inside to outside represent different classification level. Area of sector indicates respective proportion of different OTU annotation	38
3.10 Taxonomic rank distribution of <i>Acidobacteria</i> in sandy clay loam soil treated with non-fertilised sandy clay loam soil (a), 100 kg/ha NPK compound fertiliser (b), 200 kg/ha NPK compound fertiliser (c), and 400 kg/ha NPK compound fertiliser. Circles	

	from inside to outside represent different classification level. Area of sector indicates respective proportion of different OTU annotation	39
3.11	Taxonomic rank distribution of <i>Actinobacteria</i> in peat treated with non-fertilised peat (a), 100 kg/ha NPK compound fertiliser (b), 200 kg/ha NPK compound fertiliser (c), and 400 kg/ha NPK compound fertiliser. Circles from inside to outside represent different classification level. Area of sector indicates respective proportion of different OTU annotation	40
3.12	Taxonomic rank distribution of <i>Actinobacteria</i> in sandy clay loam soil treated with non-fertilised sandy clay loam soil (a), 100 kg/ha NPK compound fertiliser (b), 200 kg/ha NPK compound fertiliser (c), and 400 kg/ha NPK compound fertiliser. Circles from inside to outside represent different classification level. Area of sector indicates respective proportion of different OTU annotation	41
3.13	Taxonomic rank distribution of <i>Firmicutes</i> in peat treated with non-fertilised peat (a), 100 kg/ha NPK compound fertiliser (b), 200 kg/ha NPK compound fertiliser (c), and 400 kg/ha NPK compound fertiliser. Circles from inside to outside represent different classification level. Area of sector indicates respective proportion of different OTU annotation	42
3.14	Taxonomic rank distribution of <i>Firmicutes</i> in sandy clay loam soil treated with non-fertilised sandy clay loam soil (a), 100 kg/ha NPK compound fertiliser (b), 200 kg/ha NPK compound fertiliser (c), and 400 kg/ha NPK compound fertiliser. Circles from inside to outside represent different classification level. Area of sector indicates respective proportion of different OTU annotation	43
3.15	Rarefaction curves for peat and sandy clay loam soil	44
3.16	Rank abundance curve for peat and sandy clay loam soil	46
3.17	Beta diversity index heatmap. Pairwise dissimilarity coefficient between pairwise samples was represented in each grid. Weighted Unifrac distance is shown in the above grid while Unweighted Unifrac distance was displayed conversely	47
3.18	UPGMA cluster tree based on Weighted Unifrac distance conducted using QIIME software	48
4.1	Species relative abundance in the classification level of class	66
4.2	Genera identified based on top ten phylum in peat	73

4.3	Genera identified based on top ten phylum in sandy clay loam	76
5.1	Mean separation of microbial functional diversity performed using Fisher's Least Significance Difference	94
5.2	Carbon source utilization based on 5 guilds	100



LIST OF ABBREVIATIONS

%	Percentage
°C	Degree Celsius
ul	Microliter
um	Micrometer
(NH ₄) ₂ SO ₄	Ammonium sulfate
AWCD	Average Well Color Development
C/N	Carbon/Nitrogen ratio
Ca	Calcium
Ca ₃ (PO ₄) ₂	Calcium phosphate
CaCl ₂	Calcium chloride
CaCO ₃	Calcium carbonate
CEC	Cation Exchange Capacity
CFU	Colony Forming Unit
CLLP	Community Level Physiological Profile
cm	Centimetre
DGGE	Denaturing Gradient Gel Electrophoresis
DNA	Deoxyribonucleic acid
E	Shannon Evenness
EC	Electrical Conductivity
FeSO ₄	Iron (II) Sulfate
FeSO ₄ .7H ₂ O	Ferrous sulfate heptahydrate
FLASH	Fast Length Adjustment of Short Reads
g	Gram

H	Shannon Diversity Index
ha	Hectare
IRRI	International Rice Research Institute
K	Potassium
K ₂ O	Potassium oxide
KCl	Potassium chloride
kg	Kilogram
KH ₂ PO ₄	Monopotassium phosphate
KNO ₃	Potassium nitrate
l	liter
LEfSe	Linear discriminant analysis effect size
LSD	Fisher's Least Significance Difference
M	Moisture
Mg	Magnesium
MgSO ₄	Magnesium sulfate
MgSO ₄ .7H ₂ O	Magnesium heptahydrate
min	Minutes
ml	Milliliter
mm	Millimeter
MnSO ₄	Manganese Sulfate
N	Nitrogen
NA	Nutrient Agar
Na	Sodium
Na ₂ MoO ₄ .2H ₂ O	Sodium molybdate

NaCl	Sodium chloride
NaSO ₄	Sodium sulfate
NF	Burk's Agar
NGS	Next Generation Sequencing
nm	Nanometer
OD	Optical Density
OTU	Operational Taxonomy Unit
P ₂ O ₅	Phosphorus Pentoxide
PC1	Principal Component 1
PC2	Principal Component 2
PC3	Principal Component 3
PCR	Polymerase Chain Reaction
PDA	Potato Dextrose Agar
PLFA	Phospholipid Fatty Acid Analysis
PS	Pikovskaya's agar
PTN	Plant total nitrogen
Qiime	Quantitative insights into microbial ecology
q-PCR	Quantitative Polymerase Chain Reaction
R	Richness
RCBD	Randomized Complete Block Design
rRNA	Ribosomal Ribonucleic Acid
SCA	Starch casein Agar
SN	Soil total nitrogen
SP	Soil Phosphorus

ssu	Small subunit
TC	Total carbon
TN	Total nitrogen
TP	Total phosphorus
T-RFLP	Terminal restriction fragment length polymorphism
UPGMA	Unweighted Pair Group with Arithmetic Mean
VWC	Volumetric Water Content



CHAPTER 1

INTRODUCTION

1.1 Background

Nitrogen and phosphorus are important components in fertiliser for agricultural crops. It has been observed that the leaching of nitrogen had increased in the past 100 years because of the excessive use of chemical fertilisers (Stocker et al., 2013). The acidification of soil and nutrients differences were among the negative effects due to high amount of fertiliser in soil. Nitrogen and phosphorus in fertiliser could negatively affect the functional diversity and community structure of rhizosphere microorganisms, which play a vital role in plant growth and soil fertility (Dotaniya & Meena, 2015).

Rhizosphere is the narrow zone surrounding and effected by plant roots (Mendes et al., 2013). The rhizosphere has been regarded as a remarkable ecosystem because microorganisms inhabiting this area showed a variety of metabolic activities. Hence, rhizosphere ecosystem offers numerous services such as structuring below and above multitrophic interactions and nutrient cycling across different trophic levels and a decrease in bacterial community could potentially limit these services. Therefore, it is important to study how bacterial community in the rhizosphere responds towards environmental changes. For paddy yield and rice nutrition, microorganism can contribute significantly and thus justified the evaluation of microbial communities and their responses against agricultural practices.

Because microbes can contribute strongly to rice nutrition and production, there is much interest in surveying the microbial communities in this flooded agroecosystem and determining how agricultural practices influence the composition and structure of these communities. The rhizosphere of aerobic rice differs from that of conventional flooded rice. The oxic and anoxic conditions in normal flooded rice have different chemical requirements, while the rhizosphere of aerobic rice is similar to that of other non-flooded crops like maize. Therefore, the bacterial community in these two crops might be different due to the difference in their habitat and soil properties around the rhizosphere.

The influence of fertiliser against microbial functional diversity and bacterial community was numerously conducted. Earlier studies have identified the role of general and specific microorganisms in nutrient input in paddy fields. These include the function of microbial biomass in supplying phosphorus and nitrogen in low fertile paddy fields (Gijnsman et al., 1997) and also the contribution of nitrogen-fixing microorganisms for nitrogen input. However, the capacity of these

microorganisms to produce nutrient is influenced by land management for example the addition of rice straw that enhanced microbial activity (Roper et al., 1995), while nitrogen fertiliser reduces the growth of nitrogen-fixing and increase their population in phosphorus (Cassman, 1998). Thus, the growth of microorganisms with specific function can be influenced by the addition of introducing input.

For example, the addition of organic and chemical fertiliser improved microbial functional diversity (Ding et al., 2018) and increases bacterial community in paddy fields (Liu et al., 2018 and Kumar et al., 2018). Soil physicochemical properties also reported to have an influence on microbial functional diversity. The combination of inorganic fertiliser with organic material elevated microbial activity, species richness, catabolic diversity and carbon source utilization with the relation on soil organic carbon availability (Li et al., 2015). By using molecular techniques such as terminal restriction fragment polymorphism and denaturing gradient gel electrophoresis, microorganism that involved in nitrogen cycling such as ammonia-oxidizing bacteria and nitrogen-fixing bacteria were reported to increase when applied with fertiliser (Gu et al., 2008; Wang et al., 2009; Wartainen et al., 2008). However, there's also contrary findings showing that chemical fertiliser negatively affected the bacterial communities in paddy fields (Guanghua et al., 2008, Yoshida et al., 2009). In the recent years, next generation sequencing has been able to identify the bacterial communities in paddy fields (Huang et al., 2019; Luo et al., 2019; Kumar et al., 2018).

Economic water insufficiency is expected to hit 22 hectares million irrigated dry-season rice in Southeast Asia by 2025 (Tuong and Bouman, 2003). Conventional lowland rice required continuous flooding and elevated water inputs. Therefore, aerobic rice was introduced with the aim to reduce water inputs and increase water use efficiency. Aerobic rice is cultivated in non-saturated and non-puddled conditions with additional irrigation and high external inputs (Bouman et al., 2005). The aerobic rice cultivation in Malaysia is around 66.3 hectares in 2014 to 2015 (Rosli, 2015). This is only 0.02% from the total rice cultivation in Malaysia. This is considered small and could be explained by the newly released local aerobic rice variety along with its planting technology. Most of microbial research on aerobic rice was on the use of beneficial microbes to improve plant growth as well as for soil enrichment (Nahi et al., 2016 and Nasarudin et al., 2018).

1.2 Problem Statement

As mentioned earlier, the bacterial community in flooded paddy and aerobic rice might be different due to the difference in their habitat and soil properties around the rhizosphere. Hence, there's a need in studying the microbial functional diversity and bacterial community in aerobic rice. Aerobic rice is a potential crop for future food production. The ability of aerobic rice to grow in non-saturated condition is suitable to address water scarcity which is beneficial for food

security. A decrease in bacterial community due to inorganic disturbance such as fertiliser could potentially limit the services that they're provided in the soil and plant. Microbial functional diversity reflects genetic diversity (Carolina, 2018) and it is important to determine the analytical level of diversity to sustain ecosystem function from disturbances.

There are very few studies on the microbial functional diversity and community in aerobic rice. These studies examined the rhizosphere microbial diversity between aerobic rice growth stages (Sutradhar, 2015) and the diversity of endophytic bacteria (Vishwakarma and Dubey, 2019). High bacterial richness and diversity was identified under aerobic rice compared to flooded paddy which could benefited the aerobic rice productivity (Vishwakarma and Dubey, 2019).

To date, there is a lack of information on the effect of fertiliser and soil type on microbial functional diversity and bacterial community particularly in the local aerobic rice variety. Prior studies mentioned (Vishwakarma and Dubey, 2019) use restriction fragment length polymorphism and there is no research of bacterial community using the 16s amplicon sequencing.

It is important to know whether there are effects of fertiliser and soil type on microbial functional diversity and bacterial community structure. If there is an effect on the bacterial community structure, then how fertiliser and soil type affected the bacterial community composition and their nitrogen cycling potential. Since microorganism directly involved in ecosystem function, the addition on external input such as fertiliser may have influenced microorganism which could benefit or detriment the growth of aerobic rice. Information on microbial functional diversity and bacterial community structure could aid in the development of better aerobic rice growing strategies in drought or water scarcity area. In addition, the response of microbial community towards aerobic rice plant growth stages is important to determine whether plant physiology have an influence on the microbial community. Hence, the effect of fertiliser and soil type on bacterial community in aerobic rice rhizosphere was investigated.

1.3 Objectives

The objective of this study was to determine the effect of fertiliser and soil type on bacterial community in aerobic rice rhizosphere. The specific objective of this study is as followed:-

1. to assess the effect of fertiliser rate and soil type on microbial functional diversity and bacterial community structure in the rhizosphere of aerobic rice.
2. to determine the influence of fertiliser and soil type on the bacterial community composition and nitrogen cycling functional potential in the rhizosphere of aerobic rice.
3. to evaluate the effect of plant growth stages on the microbial community in the rhizosphere of aerobic rice.

Based on the objectives, 3 hypotheses were deduced as followed: -

1. H1 - there are fertiliser and soil type effect on microbial functional diversity and bacterial community structure.
2. H2 - there are influence of fertiliser and soil type on the bacterial community composition and nitrogen cycling functional potential in aerobic rice rhizosphere.
3. H3 - there is an effect of plant growth stages on microbial community.

REFERENCES

- Adams, T.C., Brye, K.R., Savin, M.C., Lee, J.A., & Gbur, E.E. (2017). Microbial Carbon Substrate Utilization Differences among High- and Average-Yield Soybean Areas. *Agriculture*, 7, 48. <http://doi:10.3390/agriculture7060048>
- Agnello, A.C., Potysz, A., Fourdrin, C., Huguenot, D., & Chauhan, P.S. (2018). Impact of pyrometallurgical slags on sunflower growth, metal accumulation and rhizosphere microbial communities. *Chemosphere*, 208, 626-639. <https://doi.org/10.1016/j.chemosphere.2018.06.038>
- Ahn, J.H., Song, J., Kim, B.Y., Kim, M.S., Joa, J.H., & Weon, H.Y. (2012). Characterization of the bacterial and archaeal communities in rice field soils subjected to long-term fertilization practices. *Journal of Microbiology*, 50(5), 754-765. <https://doi.org/10.1007/s12275-012-2409-6>
- Ai, C., Liang, G., Sun, J., Wang, X., He, P., Zhou, W., & He, X. (2015). Reduced dependence of rhizosphere microbiome on plant-derived carbon in 32-year long-term inorganic and organic fertilized soils. *Soil Biology & Biochemistry*, 80, 70-78. <https://doi.org/10.1016/j.soilbio.2014.09.028>
- Al-Saedi, S.A., Razaq, I.B., & Ali, N.A. (2016). Effect of Soil Textural Classes on the Biological Nitrogen Fixation by *Bradyrhizobium* Measured by ¹⁵N Dilution Analysis. *Baghdad Science Journal*, 13(4), 734-744. <https://DOI:10.21123/bsj.2016.13.4.0734>
- Alenezi, F.N., Rekik, I., Chenari Bouket, A., Luptakova, L., Weitz, H.J., Rateb, M. E., Jaspars, M., Woodward, S., & Belbahri, L. (2017). Increased Biological Activity of *Aneurinibacillus migulanus* Strains Correlates with the Production of New Gramicidin Secondary Metabolites. *Frontiers in Microbiology*, 8, 517. <https://doi:10.3389/fmicb.2017.00517>
- Babin, D., Deubel, A., Jacquiod, S., Sørensen, S.J., Geistlinger, J., Grosch, R., & Smalla, K. (2019). Impact of long-term agricultural management practices on soil prokaryotic communities. *Soil Biology and Biochemistry*, 129, 17–28. <https://doi.org/10.1016/j.soilbio.2018.11.002>
- Badoei-Dalfard, A. (2016). L-asparaginase production in the *Pseudomonas pseudoalcaligenes* strain JHS-71 isolated from Jooshan Hot-spring. *Molecular Biology Research Communications*, 5(1), 1-10.
- Bai, R., Wang, J.T., Deng, Y., He, J.Z., Feng, K., & Zhang, L.M. (2017). Microbial Community and Functional Structure Significantly Varied among Distinct Types of Paddy Soils But Responded Differently along Gradients of Soil Depth Layers. *Frontiers in Microbiology*, 8, 945. <https://doi:10.3389/fmicb.2017.00945>

- Balasoorya, W. K., Huygens, D., Rajapaksha, R. M. C. P., & Boeckx, P. (2016). Effect of rice variety and fertilizer type on the active microbial community structure in tropical paddy fields in Sri Lanka. *Geoderma*, 265, 87–95. <https://doi.org/10.1016/j.geoderma.2015.11.007>
- Basha, S.J., Basavarajappa, R., & Babalad, H. (2016). Influence of organic and inorganic nutrient management practices on yield, economics and quality parameters of aerobic rice. *Research on Crop*, 17(2), 178. DOI: 10.5958/2348-7542.2016.00032.2
- Bebber, D.P., & Richards, V.R. (2020). A meta-analysis of the effect of organic and mineral fertilizers on soil microbial diversity. *bioRxiv*. doi: <https://doi.org/10.1101/2020.10.04.325373>
- Bei, S., Zhang, Y., Li, T., Christie, P., Li, X., Zhang, J. (2018). Response of the soil microbial community to different fertilizer inputs in a wheat-maize rotation on a calcareous soil. *Agriculture, Ecosystems and Environment*, 260, 58–69. <https://doi.org/10.1016/j.agee.2018.03.014>
- Bergkemper, F., Schöler, A., Engel, M., Lang, F., Krüger, J., Schloter, M., & Schulz, S. (2016). Phosphorus depletion in forest soils shapes bacterial communities towards phosphorus recycling systems. *Environmental Microbiology*, 18(6), 1988-2000. <https://doi.org/10.1111/1462-2920.13188>
- Bier, R.L., Bernhardt, E.S., Boot, C.M., Graham, E.B., Hall, E.K., Lennon, J.T., Nemergut, D.R., Osborne, B.B., Ruiz-González, C., Schimel, J.P., Waldrop, M.P., & Wallenstein, M.D. (2015). Linking microbial community structure and microbial processes: an empirical and conceptual overview. *FEMS Microbiology Ecology*, 91(10). <https://doi.org/10.1093/femsec/fiv113>
- Blunt, S.M., Sackett, J.D., Rosen, M.R., Benotti, M.J., Trenholm, R.A., Vanderford, B.J., Hedlund, B.P., & Moser, D.P. (2018). Association between degradation of pharmaceuticals and endocrine-disrupting compounds and microbial communities along a treated wastewater effluent gradient in Lake Mead. *Science of the Total Environment*, 622–623, 1640–1648. <https://doi.org/10.1016/j.scitotenv.2017.10.052>
- Bobuľská, L., Fazekašová, D. & Angelovičová, L. (2015) Vertical Profiles of Soil Properties and Microbial Activities in Peatbog Soils in Slovakia. *Environmental Processes*, 2, 411. <https://doi.org/10.1007/s40710-015-0073-7>
- Bonk, F., Popp, D., Harms, H., & Centler, F. (2018). PCR-based quantification of taxa-specific abundances in microbial communities: Quantifying and avoiding common pitfalls. *Journal of Microbiological Methods*, 153, 139–147. <https://doi.org/10.1016/j.mimet.2018.09.015>

- Borie, F., Rubio, R., Morales, A., Curaqueo, G. & Cornejo, P. (2010). Arbuscular mycorrhizae in agricultural and forest ecosystems in Chile. *Journal of Soil Science and Plant Nutrition*, 10(3), 185-206. <http://dx.doi.org/10.4067/S0718-95162010000100001>
- Borja Reis, A.F., de Almeida, R.E.M., Lago, B.C., Trivelin, P.C., Linqvist, B., & Favarin, J.L. (2018). Aerobic rice system improves water productivity, nitrogen recovery and crop performance in Brazilian weathered lowland soil. *Field Crops Research*, 218, 59–68. <https://doi.org/10.1016/j.fcr.2018.01.002>
- Bouman, B.A.M., Peng, S., Castaneda, A.R., & Visperas, R.M. (2005). Yield and water use of irrigated tropical aerobic rice systems. *Agricultural Water Management*, 74, 87–105. <https://doi.org/10.1016/j.agwat.2004.11.007>
- Bouman, B.A.M., Humphreys, E., Tuong, T.P., & Barker, R. (2007). Rice and water. *Advances in Agronomy*, 92, 187–23.
- Braga, R.M., Dourado, M.N., & Araújo, W.L. (2016). Microbial interactions: ecology in a molecular perspective. *Brazilian Journal of Microbiology*, 47(1), 86-98. <https://doi.org/10.1016/j.bjm.2016.10.005>
- Breidenbach, B., Pump, J., & Dumont, M. G. (2016). Microbial community structure in the rhizosphere of rice plants. *Frontiers in Microbiology*, 6(JAN), 1–12. <https://doi.org/10.3389/fmicb.2015.01537>
- Browne, P., Rice, O., Miller, S.H., Burke, J., Dowling, D.N., Morrissey, J.P., & O’Gara, F. (2009). Superior inorganic phosphate solubilization is linked to phylogeny within the *Pseudomonas fluorescens* complex. *Applied Soil Ecology*, 43(1), 131–138. DOI 10.1016/j.apsoil.2009.06.010
- Bryant, D.A., Costas, A.M., Maresca, J.A., Chew, A.G.M., Klatt, C.G., Bateson, M.M., Tallon, L.J., Hostetler, J., Nelson, W.C., Heidelberg, J.F., & Ward, D.M. (2007). *Candidatus Chloracidobacterium thermophilum*: An Aerobic Phototrophic Acidobacterium. *Science*, 317(5837), 523-526. <https://DOI:10.1126/science.1143236>
- Cai, Z., Zhang, Y., Yang, C., & Wang, S. (2018). Land-use type strongly shapes community composition, but not always diversity of soil microbes in tropical China. *Catena*, 165, 69–380. <https://doi.org/10.1016/j.catena.2018.02.018>
- Campbell, C.R., & Plank, C.O. (1997). Preparation of Plant Tissue for Laboratory Analysis. In Y.P. Kalra (Ed.), *Handbook of Reference Methods for Plant Analysis* (pp 37). CRC Press
- Cao, Y., Fu, D., Liu, T., Guo, G., & Hu, Z. (2018). Phosphorus Solubilizing and Releasing Bacteria Screening from the Rhizosphere in a Natural Wetland. *Water*, 10 (2), 195. <https://doi:10.3390/w10020195>

- Caporaso, J.G., Kuczynski, J., Stombaugh, J., Bittinger, K., Bushman, F.D., Costello, E.K., Fierer, N., Peña, A.G., Goodrich, J.K., Gordon, J.I., Huttley, G.A., Kelley, S.T., Knights, D., Koenig, J.E., Ley, R.E., Lozupone, C.A., McDonald, D., Muegge, B.D., Pirrung, M., ... Knight, R. (2010). QIIME allows analysis of high-throughput community sequencing data. *Nature methods*, 7.5, 335-336. <https://doi.org/10.1038/nmeth.f.303>
- Carolina, Z. F. A. (2018). Microbial Diversity: Genetic Diversity Vs Functional Diversity. *Journal of Microbiology & Experimentation*, 6 (1), 00814. <https://DOI:10.15406/jmen.2018.06.00184>
- Castro-Silva, C., Ruíz-Valdiviezo, V.M., Valenzuela-Encinas, C., Alcántara-Hernández, R.J., Navarro-Noya, Y.E., Vázquez-Núñez, E., Luna-Guido, M., Marsch, R., & Dendooven, L. (2013). The bacterial community structure in an alkaline saline soil spiked with anthracene. *Electronic Journal of Biotechnology*, 16(5). <https://DOI: 10.2225/vol16-issue5-fulltext-14>
- Cassman. (1998). Opportunities for increased nitrogen-use efficiency from improved resource management in irrigated rice systems. *Field Crops Research* 56(1):7-39. [https://DOI: 10.1016/S0378-4290\(97\)00140-8](https://DOI: 10.1016/S0378-4290(97)00140-8)
- Chan, C.S., Zainudin, H., Saad, A., & Azmi, M. (2012). Productive water use in aerobic rice cultivation. *Journal of Tropical Agriculture and Food Science*, 49(1), 117–126
- Chapman, H.D., & Pratt, P.F., (1978). *Methods of analysis for soils, plants and waters*. Oakland: University of California Division of Agriculture and Science. Priced Publication.
- Chavarria, D.N., Pérez-Brandan, C., Serri, D.L., Meriles, J.M., Restovich, S.B., Andriulo, A.E., Jacquelin, L., & Vargas-Gil, S. (2018). Response of soil microbial communities to agroecological versus conventional systems of extensive agriculture. *Agriculture, Ecosystems and Environment*, 264, 1–8. <https://DOI: 10.1016/j.agee.2018.05.008>
- Chávez-Romero, Y., Navarro-Noya, Y.E., Reynoso-Martinez, S.C., Sarria-Guzmán, Y., Govaerts, B., Verhulst, N., Dendooven, L., & Luna-Guido, M. (2016). 16S metagenomics reveals changes in the soil bacterial community driven by soil organic C, N-fertilizer and tillage-crop residue management. *Soil & Tillage Research*, 159, 1-8. <https://doi.org/10.1016/j.still.2016.01.007>
- Chen, K., Tang, S., Wang, G., Nie, G., Li, Q., Zhang, J., Li, W., & Li, S. (2013). *Olivibacter jilunii* sp. nov., isolated from DDT-contaminated soil. *International Journal of Systematic and Evolutionary Microbiology*, 63, 1083-1088. <https://doi: 10.1099/ijs.0.042416-0>

- Chen, H., Mothapo, N.V. & Shi, W. Soil Moisture and pH Control Relative Contributions of Fungi and Bacteria to N₂O Production. (2015). *Microbial Ecology*, 69, 180–191. <https://doi.org/10.1007/s00248-014-0488->
- Cheng, X., Hui, X., White, A.P., Guo, Z., Hu, Y., & Wang, Y. (2016). Identification of new bacterial type III secreted effectors with a recursive Hidden Markov Model profile-alignment strategy. *bioRxiv*. [https://doi: https://doi.org/10.1101/081265](https://doi.org/10.1101/081265)
- Clark, B., DeFries, R.S., & Krishnaswamy, J. (2016). Intra-annual dynamics of water stress in the central Indian Highlands from 2002 to 2012. *Regional Environmental Change*, 16(1), 83–95. <https://doi.org/10.1007/s10113-016-1017-0>
- Clerget, B., Bueno, C., Domingo, A.J., Layaoen, H.L., & Vial, L. (2016). Leaf emergence, tillering, plant growth, and yield in response to plant density in a high-yielding aerobic rice crop. *Field Crops Research*, 199, 52–64. <https://doi.org/10.1016/j.fcr.2016.09.018>
- Cui, S., Suzaki, T., Tominaga-Wada, R., & Yoshida, S. (2018). Regulation and functional diversification of root hairs. *Seminars in Cell and Developmental Biology*, 83, 115-122. <https://DOI: 10.1016/j.semcdb.2017.10.003>
- Curd, E.E., Martiny, J.B.H., Li, H. & Smith, T.B. (2018). Bacterial diversity is positively correlated with soil heterogeneity. *Ecosphere* 9(1), e02079. <https://doi.10.1002/ecs2.2079>
- da Mota, F.F, Vollú, R.E., Jurelevicius, D., & Seldin, L. (2016). Whole-genome sequence of *Rummeliibacillus stabekisii* strain PP9 isolated from Antarctic soil. *Genome Announcements*, 4(3), e00416-16. <https://doi:10.1128/genomeA.00416-16>
- Dai, Y., Qiao, X., & Wang, X. (2018). Study on cation exchange capacity for agricultural soils. *IOP Conference Series: Material Science and Engineering*, 392(3), 042039. <https://doi:10.1088/1757-899X/392/4/042039>
- DeSantis, T.Z., Hugenholtz, P., Larsen, N., Rojas, M., Brodie, E.L., Keller, K., Huber, T., Dalevi, D., Hu, P. & Andersen, G.L. (2006). Greengenes, a chimera-checked 16S rRNA gene database and workbench compatible with ARB. *Applied and Environmental Microbiology*, 72(7), 5069-5072. <https://doi: 10.1128/AEM.03006-05>
- Ding, G.C., Piceno, Y.M., Heuer, H., Weinert, N., Dohrmann, A.B., Carrillo, A., Andersen, G.L., Castellanos, T., Tebbe, C.C., & Smalla, K. (2013). Changes of Soil Bacterial Diversity as a Consequence of Agricultural Land Use in a Semi-Arid Ecosystem. *PLoS ONE*, 8(3), e59497. <https://doi:10.1371/journal.pone.0059497>

- Ding, L.J., Su, J.Q., Sun, G.X., Wu, J.S., & Wei, W.X. (2018). Increased microbial functional diversity under long-term organic and integrated fertilization in a paddy soil. *Applied Microbiology and Biotechnology*, 102(4), 1969-1982. <https://doi.org/10.1007/s00253-017-8704-8>
- Docherty, K.M., Borton, H.M., Espinosa, N., Gebhardt, M., Gil-Loaiza, J., Gutknecht, J.L., Maes, P.W., Mott, B.M., Parnell, J.J., Purdy, G., Rodrigues, P.A., Stanish, L.F., Walser, O.N., & Gallery, R.E. (2015). Key Edaphic Properties Largely Explain Temporal and Geographic Variation in Soil Microbial Communities across Four Biomes. *PLoS One*, 10(11), e0135352. <https://doi.org/10.1371/journal.pone.0135352>
- Dotaniya, M.L., & Meena, V.D. (2015). Rhizosphere effect on nutrient availability in soil and its uptake by plants: A review. *The Proceedings of the National Academy of Sciences, India, Section B: Biological Sciences*, 85 (1), 1-12. <https://doi.org/10.1007/s40011-013-0297-0>
- Dubey, G., Kollah, B., Gour, V.K., Shukla, A.K., & Mohanty, S.R. (2016). Diversity of bacteria and archaea in the rhizosphere of bioenergy crop *Jatropha curcas*. *3 Biotech* 6, 257. <https://doi.org/10.1007/s13205-016-0546-z>
- Edgar, R.C. (2013). UPARSE: highly accurate OTU sequences from microbial amplicon reads. *Nature methods*, 10(10), 996-998. <https://doi.org/10.1038/nmeth.2604>
- Edgar, R.C., Haas, B.J., Clemente, J.C., Quince, C. & Knight, R. (2011). UCHIME improves sensitivity and speed of chimera detection. *Bioinformatics*, 27(16), 2194-2000. <https://doi.org/10.1093/bioinformatics/btr381>
- El Bayoumy, M.A., Bewtra, J.K., Ali, H.I. & Biswas, N. (1999). Sulfide Production by Sulfate Reducing Bacteria with Lactate as Feed in an Upflow Anaerobic Fixed Film Reactor. *Water, Air, & Soil Pollution*, 112(1-2), 67-84. <https://doi.org/10.1023/A:100501640670>
- Elkheir, H.A., Yunus, M., Muslimin, M., Sjahril, R., Kasim, N., & Riadi, M. (2016). Seed Germination Behaviors Of Some Aerobic Rice Cultivars (*Oryza Sativa* L) After Priming With Polyethylene Glycol-8000 (Peg-8000). *International Journal Of Scientific & Technology Research*, 5(2), 227-234
- Espenberg, M., Truu, M., Mander, U., Kasak, K., Nõlvak, H., Ligi, T., Oopkaup, K., Maddison, M., & Truu, J. (2018). Differences in microbial community structure and nitrogen cycling in natural and drained tropical peatland soils. *Scientific Reports*, 8, Article number: 4742. <https://doi.org/10.1038/s41598-018-23032-y>

- Esson, K.C., Lin, X., Kumaresan, D., Chanton, J.P., Murrell, J.C., & Kostka, J.E. (2016). Alpha- and gammaproteobacterial methanotrophs codominate the active methane-oxidizing communities in an acidic boreal peat bog. *Applied Environmental Microbiology*, 82, 2363–2371. <https://doi.org/10.1128/AEM.03640-15murdi>
- Etesami, H., & Beattie, G.A. (2018). Mining Halophytes for Plant Growth-Promoting Halotolerant Bacteria to Enhance the Salinity Tolerance of Non-halophytic Crops. *Frontiers in Microbiology*, 9, 148. <https://doi.org/10.3389/fmicb.2018.00148>
- Falkowski, P.G., Fenchel, T., & Delong, E.F. (2008). The microbial engines that drive earth's biogeochemical cycles. *Science*, 320(5879), 1034–1039. <https://DOI: 10.1126/science.1153213>
- Fan, K., Weisenhorn, P., Gilbert, J.A., Shi, Y., Bai, Y., & Chu, H. (2018). Soil pH correlates with the co-occurrence and assemblage process of diazotrophic communities in rhizosphere and bulk soils of wheat fields. *Soil Biology and Biochemistry*, 121, 185–192. <https://doi.org/10.1016/j.soilbio.2018.03.017>
- Farooq, M., Nawaz, A., Ahmad, E., Nadeem, F., Hussain, M., & Siddique, K.H.M. (2017). Using Sorghum to suppress weeds in dry seeded aerobic and puddled transplanted rice. *Field Crops Research*, 214, 211–218. <https://doi.org/10.1016/j.fcr.2017.09.017>
- Felix, K.C.S., Souza, E.B., Michereff, S.J., & Mariano, R.L.R. (2012). Survival of *Ralstonia solanacearum* in infected tissues of *Capsicum annuum* and in soils of the state of Pernambuco, Brazil. *Phytoparasitica*, 40, 53-62. <https://DOI 10.1007/s12600-011-0200-6>
- Fish, J.A., Chai, B., Wang, Q., Sun, Y., Brown, C.T., Tiedje, J.M., & Cole, J.R. (2013). FunGene: The Functional Gene Pipeline and Repository. *Frontiers in Microbiology*, 4, 291. <https://doi: 10.3389/fmicb.2013.00291>
- Fornasero, L.V., Del Papa, M.F., López, J.L., Albicoro, F.J., Zabala, J.M., Toniutti, M.A., Pensiero, J.F., & Lagares, A. (2014). Phenotypic, Molecular and Symbiotic Characterization of the Rhizobial Symbionts of *Desmanthus paspalaceus* (Lindm.) Burkart That Grow in the Province of Santa Fe, Argentina. *PLoS ONE*, 9(8), e104636. <https://doi.org/10.1371/journal.pone.0104636>
- Franchia, O., Boviob, P., Ortega-Martínez, E., Rosenkranz, F., & Chamy, R. (2018). Active and total microbial community dynamics and the role of functional genes *bamA* and *mcrA* during anaerobic digestion of phenol and p-cresol. *Bioresource Technology*, 264, 290–297. <https://doi.org/10.1016/j.biortech.2018.05.060>

- Fröhling, A., Rademacher, A., Rumpold, B., Klocke, M., & Schlüter, O. (2018). Screening of microbial communities associated with endive lettuce during postharvest processing on industrial scale. *Heliyon*, *4*(7), e00671. <https://doi.org/10.1016/j.heliyon.2018.e00671>
- Fujita, K., Kunito, T., Matsushita, J., Nakamura, K., Moro, H., Yoshida, S., Toda, H., Otsuka, S., & Nagaoka, K. (2018). Nitrogen supply rate regulates microbial resource allocation for synthesis of nitrogen-acquiring enzymes. *PLOS ONE*, *13*(8), e0202086. <https://doi.org/10.1371/journal.pone.0202086>
- Galán, J.E., & Wolf-Watz, H. (2006). Protein delivery into eukaryotic cells by type III secretion machines. *Nature*, *444* (7119), 567–573. <https://doi.org/10.1038/nature05272>
- Gao, P., Sun, X., Xiao, E., Xu, Z., Li, B., & Sun, W. (2019). Characterization of iron-metabolizing communities in soils contaminated by acid mine drainage from an abandoned coal mine in Southwest China. *Environmental Science and Pollution Research International*, *26*, 9585-9598. <https://doi.org/10.1007/s11356-019-04336-6>
- Gao, S., Cao, W., Zou, C., Gao, J., Huang, J., Bai, J., Zeng, N., Shimizu, K.Y., Wright, A., & Dou, F. (2018). Ammonia-oxidizing archaea are more sensitive than ammonia-oxidizing bacteria to long-term application of green manure in red paddy soil. *Applied Soil Ecology*, *124*, 185–193. <https://doi.org/10.1016/j.apsoil.2017.09.041>
- Garland, J.L., & Mills, A.L. (1991). Classification and Characterization of Heterotrophic Microbial Communities on the Basis of Patterns of Community-Level Sole-Carbon-Source Utilization. *Applied and Environmental Microbiology*, *57*(8), 2351-2359.
- Garland, J. (1997). Analysis and interpretation of community-level physiological profiles in microbial ecology. *FEMS Microbiology Ecology*, *24*, 289-300. <https://doi.org/10.1111/j.1574-6941.1997.tb00446.x>
- Giaramida, L., Manage, P.M., Edwards, C., Singh, B.K., & Lawton, L.A. (2013). Bacterial communities' response to microcystins exposure and nutrient availability: Linking degradation capacity to community structure. *International Biodeterioration & Biodegradation*, *84*, 111-117. <https://doi.org/10.1016/j.ibiod.2012.05.036>
- Gijsman, A.J., Oberson, A., Friesen, D.K., Sanz, J.I., & Thomas, R. J. (1997). Nutrient cycling through microbial biomass under rice-pasture rotations replacing native savanna. *Soil Biology and Biochemistry*, *29*(9–10), 1433–1441. [https://doi.org/10.1016/S0038-0717\(97\)00045-X](https://doi.org/10.1016/S0038-0717(97)00045-X)
- Gee, G.W., & J.W. Bauder. (1986). Particle-size analysis. In A., Klute (Ed.) *Methods of soil analysis. Part 1. 2nd edition.* (pp. 383–411).. ASA and SSSA, Madison, WI.

- Godini, K., Samarghandi, M.R., Tahmasebi, H., Zarei, O., Karimitabar, Z., Yarahmadi, Z., & Arabestani, M.R. (2019). Biochemical and molecular characterization of novel PAH-degrading bacteria isolated from polluted soil and sludge. *Petroleum Science and Technology*, 37(15), 1763 – 1769. <https://doi.org/10.1080/10916466.2019.1575864>
- Golińska, P., Wypij, M., Rathod, D., Tikar, S., Dahm, H., & Rai, M. (2015). Synthesis of silver nanoparticles from two acidophilic strains of *Pilimelia columellifera* subsp. *pallida* and their antibacterial activities. *Journal of Basic Microbiology*, 56(5), 541-556. <https://doi.org/10.1002/jobm.201500516>
- Gomez, E., Garland, J. & Conti, M. (2004). Reproducibility in the response of soil bacterial community-level physiological profiles from a land use intensification gradient. *Applied Soil Ecology*, 26(1), 21-30. <https://doi.org/10.1016/j.apsoil.2003.10.007>
- Gomez, E., Ferreras, L. & Toresani, S. (2006). Soil bacterial functional diversity as influenced by organic amendment application. *Bioresource Technology*, 97(13), 1484-1489. <https://doi.org/10.1016/j.biortech.2005.06.021>
- González-Abradelo, D., Pérez-Llano, Y., Peidro-Guzmán, H., Sánchez-Carbente, M.D.R., Folch-Mallol, J. L., Aranda, E., Vaidyanathan, V.K., Cabana, H., Gunde-Cimerman, N., & Batista-García, R. A. (2019). First demonstration that ascomycetous halophilic fungi (*Aspergillus sydowii* and *Aspergillus destruens*) are useful in xenobiotic mycoremediation under high salinity conditions. *Bioresource Technology*, 279, 287–296. <https://doi.org/10.1016/J.BIORTECH.2019.02.002>
- Grodzinsky, A.M. (2006). Soil sickness: History. In: A.M. Grodzinsky (Ed). *Allelopathy in Soil Sickness* (pp. 1-10). Scientific Publishers.
- Grządziel, J., & Gałazka, A. (2018). Microplot long-term experiment reveals strong soil type influence on bacteria composition and its functional diversity. *Applied Soil Ecology*, 124, 117–123. <https://doi.org/10.1016/j.apsoil.2017.10.033>
- Gschwendtner, S., Engel, M., Lueders, T., Buegger, F. & Scholter, M. (2016). Nitrogen fertilization affects bacteria utilizing plant-derived carbon in the rhizosphere of beech seedlings. *Plant and Soil*, 407 (1-2), 203-215. <https://doi.org/10.1007/s11104-016-2888-z>
- Gu, Y., Yun, X., Zhang, X. Ping, T.U, SUN, X., & Lindström, K. (2008). Effect of Different Fertilizer Treatments on Quantity of Soil Microbes and Structure of Ammonium Oxidizing Bacterial Community in a Calcareous Purple Paddy Soil. *Agricultural Sciences in China*, 7(12), 1481–1489. [https://doi.org/10.1016/S1671-2927\(08\)60406-X](https://doi.org/10.1016/S1671-2927(08)60406-X)

- Gu, Y., Bai, Y., Xiang, Q., Yu, X., Zhao, K., Zhang, X., Li, C., Liu, S., & Chen, Q. (2018). Degradation shaped bacterial and archaeal communities with predictable taxa and their association patterns in Zoige wetland at Tibet plateau. *Scientific Reports*, 8, Article number: 3884. <https://doi.org/10.1038/s41598-018-21874-0>
- Gu, Y., Wang, Y., Lu, S., Xiang, Q., Yu, X., Zhao, K., Zou, L., Chen, Q., Shihua Tu, S., & Zhang, X. (2017). Long-term Fertilization Structures Bacterial and Archaeal Communities along Soil Depth Gradient in a Paddy Soil. *Frontiers in Microbiology*, 8, 1516. <https://doi.org/10.3389/fmicb.2017.01516>
- Guanghua, W., Junjie, L., Xiaoning, Q., Jian, J., Yang, W., & Xiaobing, L. (2008). Effects of fertilization on bacterial community structure and function in a black soil of Dehui region estimated by Biolog and PCR-DGGE methods. *Acta Ecologica Sinica*, 28(1), 220–226. [https://doi.org/10.1016/S1872-2032\(08\)60023-2](https://doi.org/10.1016/S1872-2032(08)60023-2)
- Gumiere, T., Rousseau, A.N., da Costa, D.P., Cassetari, A., Cotta, S.R., Andreote, F.D., Gumiere, S.J., & Pavinato, P.S. (2019). Phosphorus source driving the soil microbial interactions and improving sugarcane development. *Scientific Report*, 9, 4400. <https://doi.org/10.1038/s41598-019-40910-1>
- Hamer, U., Makeschin, F., Stadler, J., & Klotz, S. (2008). Soil organic matter and microbial community structure in set-aside and intensively managed arable soils in NE-Saxony, Germany. *Applied Soil Ecology*, 40, 465–475. <https://doi.org/10.1016/j.apsoil.2008.07.001>
- Han, C., Zhao, J., Shi, H., Tian, Y., Zhang, C., Guo, X., Xiang, W., & Wang, X. (2019). *Jiangella rhizosphaerae* sp. nov., an actinomycete isolated from the rhizosphere soil of wheat (*Triticum aestivum* L.). *International Journal of Systematic Evolutionary Microbiology*, 69(5), 1320–1326. <https://doi.org/10.1099/ijsem.0.003314>
- Hartmann, M., Frey, B., Mayer, J., Mader, P., & Widmer, F. (2015). Distinct soil microbial diversity under long-term organic and conventional farming. *The ISME Journal*, 9, 1177–1194. [10.1038/ismej.2014.210](https://doi.org/10.1038/ismej.2014.210)
- Hemkemeyer, M., Dohrmann, A.B., Christensen, B.T., & Tebbe, C.C. (2018). Bacterial Preferences for Specific Soil Particle Size Fractions Revealed by Community Analyses. *Frontiers in Microbiology*, 9, 149. <https://doi.org/10.3389/fmicb.2018.00149>
- Ho, A., Angel, R., Veraart, A.J., Daebeler, A., Jia, Z., Kim, S.Y., Kerckhof, F.M., Boon, N., & Bodelier, P.L.E. (2016). Biotic interactions in microbial communities as modulators of biogeochemical processes: Methanotrophy as a model system. *Frontiers in Microbiology*. <https://doi.org/10.3389/fmicb.2016.01285>

- Huang, L., Bai, J., Wen, X., Zhang, G., Zhang, C., Cui, B., & Liu, X. (2020). Microbial resistance and resilience in response to environmental changes under the higher intensity of human activities than global average level. *Global Change Biology*, 26, (4), 2377-2389. <https://doi.org/10.1111/gcb.14995>
- Huang, Q., Wang, J., Wang, C., & Wang, Q. (2019). The 19-years inorganic fertilization increased bacterial diversity and altered bacterial community composition and potential functions in a paddy soil. *Applied Soil Ecology*, 144, 60–67. <https://doi.org/10.1016/j.apsoil.2019.07.009>
- Huang, N., Wang, W., Yao, Y., Zhu, F., Wang, W., Chang, X. (2017) The influence of different concentrations of bio-organic fertilizer on cucumber Fusarium wilt and soil microflora alterations. *PLoS ONE*, 12(2), e0171490. <https://doi:10.1371/journal.pone.0171490>
- Huang, W.C, Tsai, H.C, Tao, C.W., Chen, J.S., Shih, Y.J., Kao, P.M., Huang, T.Y., & Hsu, B.M. (2017). Approach to determine the diversity of *Legionella* species by nested PCR-DGGE in aquatic environments. *PLoS ONE*, 12(2), e0170992. <https://doi.org/10.1371/journal.pone.0170992>
- Huang, X.F., Chaparro, J. M., Reardon, K. F., Zhang, R., Shen, Q. & Vivanco, J. M. (2014). Rhizosphere interactions: root exudates, microbes, and microbial communities. *Botany*, 92(4), 267-275. <https://doi.org/10.1139/cjb-2013-0225>
- Igwe, A.N. (2015). *Phytoremediation of hydrocarbon-contaminated soil using phenolic-exuding horticultural plants*. [Master's thesis, Texas A&M University]. Texas A&M Repository.
- Jabran, K., & Chauhan, B.S. (2015). Weed management in aerobic rice systems. *Crop Protection*, 78, 151-163. <https://doi.org/10.1016/j.cropro.2015.09.005>
- Jangid, K., Williams, M.A., Franzluebbers, A.J., Sanderlin, J.S., Reeves, J.H., Jenkins, M.B., Endale, D.M., Coleman, D.C. & Whitman, W.B. (2008). Relative impacts of land-use, management intensity and fertilization upon soil microbial community structure in agricultural systems. *Soil Biology & Biochemistry*, 40, 2843–2853. <https://doi.org/10.1016/j.soilbio.2008.07.030>
- Jia, F., Guo, S., Shen, Y., Gao, M., Liu, C., Zhou, S., Li, J., Guan, X., Wang, X., & Xiang, W. (2016). *Catellatospora vulcania* sp. nov. and *Catellatosporaparisidis* sp. nov., two novel actinobacteria isolated from volcanic sediment and the rhizosphere of *Paris polyphylla*. *Antonie van Leeuwenhoek*, 109(1), 43-50. <https://doi.org/10.1007/s10482-015-0608-y>

- Jingnan, Z., Mengjie, L., Peize, L., Weihua, K., Jiabao, S., Lin, Y., & Shuaifeng, W. (2018). Effect of pig manure amendments on microbial biomass and functional diversity in three agricultural soils. *IOP Conference Series: Earth and Environmental Science*, 012061. [https://doi :10.1088/1755-1315/186/3/012061](https://doi.org/10.1088/1755-1315/186/3/012061)
- Jiao, J.Y., Carro, L., Liu, L., Gao, X.Y., Zhang, X.T., Hozzein, W.N., Lapidus, A., Huntemann, M., Reddy, T.B.K., Varghese, N., Hadjithomas, M., Ivanova, N.N., Göker, M., Pillay, M., Eisen, J.A., Woyke, T., Klenk, H.P., Kyrpides, N.C., & Li, W.J. (2017). Complete genome sequence of *Jiangella gansuensis* strain YIM 002^T (DSM 44835^T), the type species of the genus *Jiangella* and source of new antibiotic compounds. *Standards in genomic sciences*, 12, 21. [https://doi:10.1186/s40793-017-0226-6](https://doi.org/10.1186/s40793-017-0226-6)
- Kandeler, E., Tschерko, D., & Spiegel, H. (1999) Long-term monitoring of microbial biomass, N mineralization and enzyme activities of a Chernozem under different tillage management. *Biology and Fertility of Soils*, 28, 343–351.
- Karam, D.S., Jamil, M.N.C., Jaafar, N., Rajoo, K.S., & Arifin, A. (2015). Effects of different incubation periods on microbial biomass carbon (MBC) in two soil series. *Journal of Environmental Science and Pollution Research*, 1(1), 20–22.
- Kavamura, V.N., Hayat, R., Clark, I.M., Rossmann, M., Mendes, R., Hirsch, P.R., & Mauchline, T.H. (2018). Inorganic Nitrogen Application Affects Both Taxonomical and Predicted Functional Structure of Wheat Rhizosphere Bacterial Communities. *Frontiers in Microbiology*, 9, 1074. [https://doi:10.3389/fmicb.2018.01074](https://doi.org/10.3389/fmicb.2018.01074)
- Keller, A.H., Kleinstеuber, S. & Vogt, C. (2018). Anaerobic Benzene Mineralization by Nitrate-Reducing and Sulfate-Reducing Microbial Consortia Enriched from the Same Site: Comparison of Community Composition and Degradation Characteristics. *Microbial Ecology*, 75(4), 941-953. <https://doi.org/10.1007/s00248-017-1100-1>
- Khazanah Research Institute. (2019). *The Status of the Paddy and Rice Industry in Malaysia*. http://www.krinstitute.org/assets/contentMS/img/template/editor/20190409_RiceReport_Full%20Report_Final.pdf
- Kim, B.Y. (2010). *Biosystematics of the Genus Dactylosporangium and Some Other Filamentous Actinomycetes*. [Unpublished doctoral dissertation]. Newcastle University.
- Kitir, N., Yildirim, E., Sahin, Ü, Turan, M., Ekinci, M., Ors, S., Kul, R., Ünlü, H., & Ünlü, H. (2018). *Peat Use in Horticulture in Peat book*. (B. Topcuoğlu, & M., Turan, Eds). IntechOpen. <https://DOI: 10.5772/intechopen.79171>

- Knoche, K.L., Renner, J.N., Gellett, W., Ayers, K.E., & Minter, S.D. (2016). A Self-Sufficient Nitrate Groundwater Remediation System: *Geobacter Sulfurreducens* Microbial Fuel Cell Fed by Hydrogen from a Water Electrolyzer. *Journal of The Electrochemical Society*, 163(7), 651-656. <https://doi.org/10.1149/2.0821607jes>
- Konopka, A. (2009). What is microbial community ecology?. *The ISME Journal*, 3, 1223-1230. <https://doi.org/10.1038/ismej.2009.88>
- Kumar, U., Nayak, A.K., Shahid, M., Gupta, V.V.S.R, Panneerselvam, P., Mohanty, S., Kaviraj, M., Kumar, A., Chatterjee, D., Lal, B., Gautam, P., Tripathi, R., & Panda, B.B. (2018). Continuous application of inorganic and organic fertilizers over 47 years in paddy soil alters the bacterial community structure and its influence on rice production. *Agriculture, Ecosystems and Environment*, 262, 65–75. <https://doi.org/10.1016/j.agee.2018.04.016>
- Kumar, U., Shahid, M., Tripathi, R., Mohanty, S., Kumar, A., Bhattacharyya, P., ... Nayak, A. K. (2017). Variation of functional diversity of soil microbial community in sub-humid tropical rice-rice cropping system under long-term organic and inorganic fertilization. *Ecological Indicators*, 73, 536–543. <https://doi.org/10.1016/j.ecolind.2016.10.014>
- Kuramae, E.E., Yergeau, E., Wong, L.C., Pijl, A.S., van Veen, J.A., & Kowalchuk, G.A. (2012). Soil characteristics more strongly influence soil bacterial communities than land-use type. *FEMS Microbiology Ecology*, 79(1), 12–24. <https://doi.org/10.1111/j.1574-6941.2011.01192.x>
- Kreye, C., Bouman, B., Reversat, G., Fernandez, L., Cruz, C.V., Elazegui, F., Faronilo, J., & Llorca, L. (2009). Biotic and abiotic causes of yield failure in tropical aerobic rice. *Field Crop Research*, 117(1), 97-106. <https://DOI:10.1016/j.fcr.2009.02.2005>
- Lagerlöf, J., Adolfsson, L., Börjesson, G., Ehlers, K., Vinyoles, G.P., & Sundh, I. Land-use intensification and agroforestry in the Kenyan highland: Impacts on soil microbial community composition and functional capacity. *Applied Soil Ecology*, 82, 93 – 99. <https://doi.org/10.1016/j.apsoil.2014.05.015>
- Lakshmi, M.B., Muthukumar, K., & Velan, M. (2012). Immobilization of *Mycoplana* sp MVMB2 isolated from petroleum contaminated soil onto papaya stem (*Carica papaya* L) and its application on degradation of phenanthrene. *Clean Soil, Air, Water*, 40(8), 870-877. <https://doi.org/10.1002/clen.201100639>
- Langille, M.G., Zaneveld, J., Caporaso, J.G., McDonald, D., Knights, D., Reyes, J.A., Clemente, J.C., Burkepile, D.E., Thurber, R.L.V., Knight, R., Beiko, R.G., & Huttenhower, C. (2013) Predictive functional profiling of microbial communities using 16S rRNA marker gene sequences. *Nature Biotechnology*, 31, 814–821. <https://doi:10.1038/nbt.2676>

- Legrand, F., Picota, A., Cobo-Díaz, J.F., Carof, M., Chen, W., & Gaétan Le Floch, G. (2018). Effect of tillage and static abiotic soil properties on microbial diversity. *Applied Soil Ecology*, 132, 135–145. <https://doi.org/10.1016/j.apsoil.2018.08.016>
- Lei, S., Xu, X., Cheng, Z., Xiong, J., Ma, R., Zhang, L., Yang, X., Zhu, Y., Zhang, B., & Tian, B. (2018). Analysis of the community composition and bacterial diversity of the rhizosphere microbiome across different plant taxa. *Microbiology Open*. <https://DOI: 10.1002/mbo3.762>
- Li, X., Zhang, H., Sun, M., Xu, N., Sun, G., & Zhao, M. (2020). Land use change from upland to paddy field in Mollisols drives soil aggregation and associated microbial communities. *Applied Soil Ecology*, 146, 103351. <https://doi.org/10.1016/j.apsoil.2019.09.001>
- Li, F., Xue, C., Qiu, P., Liu, Y., Shi, J., Shen, B., Yang, X., & Shen, Q. (2018). Soil aggregate size mediates the responses of microbial communities to crop rotation. *European Journal of Soil Biology*, 88, 48–56. <https://doi.org/10.1016/j.ejsobi.2018.06.004>
- Li, H., Su, J.Q., Yang, X.R., & Zhu, Y.G. (2019). Distinct rhizosphere effect on active and total bacterial communities in paddy soils. *Science of the Total Environment*, 649, 422–430. <https://doi.org/10.1016/j.scitotenv.2018.08.373>
- Li, J., Li, Y.T., Yang, X.D., Zhang, J.J., Lin, Z.A., & Zhao, B.Q. (2015). Microbial community structure and functional metabolic diversity are associated with organic carbon availability in an agricultural soil. *Journal of Integrative Agriculture*, 14(12), 2500–2511. [https://doi.org/10.1016/S2095-3119\(15\)61229-1](https://doi.org/10.1016/S2095-3119(15)61229-1)
- Li, J.G., Shen, M.C, Hou, J.F., Li, L., Wu, J.X., & Dong, Y.H. (2016). Effect of different levels of nitrogen on rhizosphere bacterial community structure in intensive monoculture of greenhouse lettuce. *Scientific Reports*, 6, Article number: 25305. <https://doi.org/10.1038/srep25305>
- Li, L., Xu, M., Eyakub Ali, M., Zhang, W., Duan, Y., and Li, D. (2018) Factors affecting soil microbial biomass and functional diversity with the application of organic amendments in three contrasting cropland soils during a field experiment. *PLoS ONE*, 13(9), e0203812. <https://doi.org/10.1371/journal.pone.0203812>
- Li, R., Khafipour, E., Krause, D.O., Entz, M.H., de Kievit, T.R., & Fernando, W.G. (2012). Pyrosequencing reveals the influence of organic and conventional farming systems on bacterial communities. *PloS one*, 7(12), e51897. <https://doi:10.1371/journal.pone.0051897>

- Li, W., Wu, M., Liu, M., Jiang, C., Chen, X., Kuzyakov, Y., Rinklebe, J., & Li, Z. (2018). Responses of Soil Enzyme Activities and Microbial Community Composition to Moisture Regimes in Paddy Soils Under Long-Term Fertilization Practices. *Pedosphere*, 28(2), 323-331. [https://doi.org/10.1016/S1002-0160\(18\)60010-4](https://doi.org/10.1016/S1002-0160(18)60010-4)
- Li, X., Meng, D., Li, J., Yin, H., Liu, H., Liu, X., Cheng, C., Xiao, Y., Liu, Z., & Yan, M. (2017). Response of soil microbial communities and microbial interactions to long-term heavy metal contamination. *Environmental Pollution*, 231, 908-917. <https://doi.org/10.1016/j.envpol.2017.08.057>
- Li, Y. (2014). *Phenotypic Diversity in Lysobacter enzymogenes in Relations to Biological Control* [Unpublished master's thesis]. University of Nebraska.
- Liao, J., Xu, Q., Xu, H., & Huang, D. (2019). Natural Farming Improves Soil Quality and Alters Microbial Diversity in a Cabbage Field in Japan. *Sustainability*, 11, 3131. <https://doi.org/10.3390/su11113131>
- Liao, J., Ye, J., Liang, Y., Khalid, M., & Huang, D. (2019). Pakchoi Antioxidant Improvement and Differential Rhizobacterial Community Composition under Organic Fertilization. *Sustainability*, 11(8), 2424. <https://doi.org/10.3390/su11082424>
- Lim, J.W., Ge, T., & Tong, Y.W. (2018). Monitoring of microbial communities in anaerobic digestion sludge for biogas optimization. *Waste Management*, 71, 334–341. <https://doi.org/10.1016/j.wasman.2017.10.007>
- Liu, C., Lin, H., Yingbo Dong, Y., Bing Li, B., & Liu, Y. (2018). Investigation on microbial community in remediation of lead-contaminated soil by *Trifolium repens* L. *Ecotoxicology and Environmental Safety*, 165, 52-60. <https://doi.org/10.1016/j.ecoenv.2018.08.054>
- Liu, K.L., Li, Y.Z., Zhou, L.J., Chen, Y., Huang, Q.H., Yu, X.C., & Li, D.M. (2018). Comparison of crop productivity and soil microbial activity among different fertilization patterns in red upland and paddy soils. *Acta Ecologica Sinica*, 38, 262–267. <https://doi.org/10.1016/j.chnaes.2017.08.003>
- Liu, Z., Beskrovnaya, P., Melnyk, R.A., Hossain, S.S., Khorasani, S., O'Sullivan, L.R., Wiesmann, C.L., Bush, J., Richard, J.D., & Haney, C.H. (2018). A genome-wide screen identifies genes in rhizosphere-associated *Pseudomonas* required to evade plant defenses. *mBio*, 9: e00433-18. <https://doi.org/10.1128/mBio.00433-18>
- Locey, K.J., & Lennon, J.T. (2016). Scaling laws predict global microbial diversity. *PNAS*, 113 (21), 5970-5975. <https://doi.org/10.1073/pnas.1521291113>

- Lopes, A.R., Manaia, C.M., & Nunes, O.C. (2014). Bacterial community variations in an alfalfa-rice rotation system revealed by 16S rRNA gene 454-pyrosequencing. *FEMS Microbiology Ecology*, 87, 3, 650–663. <https://doi.org/10.1111/1574-6941.12253>
- Lopes, L.D., Weisberg, A. J., Davis, E. W., Varize, C.D.S., Silva, M.D.P.E., Chang, J. H., Loper, J.E., & Andreote, F.D. (2019). Genomic and metabolic differences between *Pseudomonas putida* populations inhabiting sugarcane rhizosphere or bulk soil. *PLoS ONE*, 14(10), 1–18. <https://doi.org/10.1371/journal.pone.0223269>
- Lu, X. (2018). Distribution and Function of Soil *Thaumarchaeota*. [Doctoral dissertation, University of Waterloo]. UWSPACE.
- Luo, X., Han, S., Fu, X., Li, X., Wang, L., Peng, S., ... Huang, Q. (2019). The microbial network in naturally fertile paddy soil possibly facilitates functional recruitment in the rice mature stage. *Applied Soil Ecology*, 135, 174–181. <https://doi.org/10.1016/j.apsoil.2018.12.008>
- Luo, S., Wang, S., Tian, L., Shi, S., Xu, S., Yang, F., Li, X., Wang, Z., & Tian, C. (2018). Aggregate-related changes in soil microbial communities under different ameliorant applications in saline-sodic soils. *Geoderma*, 329, 108–117. <https://doi.org/10.1016/j.geoderma.2018.05.023>
- Luo, X., Fu, X., Yang, Y., Cai, P., Peng, S., Chen, W., & Huang, Q. (2016). Microbial communities play important roles in modulating paddy soil fertility. *Scientific Report*, 6, 20326. <https://doi:10.1038/srep20326>
- Luo, X., Han, S., Fu, X., Li, X., Wang, L., Peng, S., ... Huang, Q. (2019). The microbial network in naturally fertile paddy soil possibly facilitates functional recruitment in the rice mature stage. *Applied Soil Ecology*, 135, 174–181. <https://doi.org/10.1016/j.apsoil.2018.12.008>
- Ma, M., Zhou, J., Ongena, M., Liu, W., Wei, D., Zhao, B., Guan, D., Jiang, X., & Li, J. (2018). Effect of long-term fertilization strategies on bacterial community composition in a 35-year field experiment of Chinese Mollisols. *AMB Express* 8(1), 20. <https://doi:10.1186/s13568-018-0549>
- Magoč, T. & Salzberg, S.L. (2011). FLASH: fast length adjustment of short reads to improve genome assemblies. *Bioinformatics*, 27.21. 2957-2963. <https://doi: 10.1093/bioinformatics/btr507>
- Manici, L.M., Saccà, M.L., Caputo, F., Zanzotto, A., Gardiman, M., & Fila, G. (2017). Long-term grapevine cultivation and agro-environment affect rhizosphere microbiome rather than plant age. *Applied Soil Ecology* 119, 214–225. <https://doi.org/10.1016/j.apsoil.2017.06.027>
- Manikandan, R., Prabhu, H.J., & Sivashanmugam, P. (2008). Biodegradation of 2, 4-dichlorophenol using *Mycoplana dimorpha* extracts and evaluation of kinetic parameters. *African Journal of Biotechnology*, 7 (12), 2038-2048

- Martirani-Von Abercron, S.M., Marín, P., Solsona-Ferraz, M., Castañeda-Cataña, M. A., & Marqués, S. (2017). Naphthalene biodegradation under oxygen-limiting conditions: community dynamics and the relevance of biofilm-forming capacity. *Microbial Biotechnology*, 10(6), 1781–1796. <https://doi.org/10.1111/1751-7915.12842>
- Matthews, A., Pierce, S., Hipperson, H., & Raymond, B. (2019). Rhizobacterial community assembly patterns vary between crop species. *Frontiers in Microbiology*. <https://doi.org/10.3389/fmicb.2019.00581>
- McNear Jr., D. H. (2013) The Rhizosphere - Roots, Soil and Everything In Between. *Nature Education Knowledge*, 4(3), 1
- Mendes, R., Garbeva, P., & Raaijmakers, J.M. (2013). The rhizosphere microbiome: significance of plant beneficial, plant pathogenic, and human pathogenic microorganisms. *FEMS Microbiology Reviews*, 37(5), 634-663. <https://doi.org/10.1111/1574-6976.12028>
- Miller, J.M. & Rhoden, D.L. (1991). Preliminary Evaluation of Biolog, a Carbon Source Utilization Method for Bacterial Identification. *Journal of Clinical Microbiology*, 29(6), 1143-1147
- Minamisawa, K., Nishioka, K., Miyaki, T., Ye, B., Miyamoto, T., You, M., Saito, A., Saito, M., Barraquio, W.L., Teaumroong, N., Sein, T., & Sato, T. (2004). Anaerobic nitrogen-fixing consortia consisting of clostridia isolated from gramineous plants. *Applied and Environmental Microbiology*, 70(5), 3096–3102. <https://doi:10.1128/aem.70.5.3096-3102.2004>
- Mohr, K., Moradi, A., Glaeser, S., Kämpfer, P., Gemperlein, K., Nübel, U., Schumann, P., Müller, R., & Wink, J. (2018). *Nannocystis konarekensis* sp. nov., a novel myxobacterium from an Iranian desert. *International Journal of Systematic Evolutionary Microbiology*, 68(3), 721-729. <https://doi:10.1099/ijsem.0.002569>
- Mukhtar, S., Mehnaz, S., Mirza, M.S., Mirza, B.S., & Malik, K.A. (2018). Diversity of *Bacillus*-like bacterial community in the rhizospheric and non-rhizospheric soil of halophytes (*Salsola stocksii* and *Atriplex amnicola*), and characterization of osmoregulatory genes in halophilic *Bacilli*. *Canadian Journal of Microbiology*, 64(8), 567-579. <https://doi.org/10.1139/cjm-2017-0544>
- Muleta, A., & Assefa, F. (2018). Isolation and screening of antibiotic producing actinomycetes from rhizosphere and agricultural soils. *African Journal of Biotechnology*, 17(22), 700-714. <https://DOI: 10.5897/AJB2017.16080>
- Munda, S., Saha, S., Adak, T., & Jambhulkar, N. (2017). Weed Management In Aerobic Rice: Role Of Establishment Methods And Herbicides. *Experimental Agriculture*, 1-17. <https://doi:10.1017/S0014479717000576>

- Murdiyarso, D., Hergoualc'h, K., & Verchot, L. V. (2010). Opportunities for reducing greenhouse gas emissions in tropical peatlands. *Proceedings of the National Academy of Sciences*, 107(46), 19655-19660. <https://doi.org/10.1073/pnas.0911966107>
- Navarrete, A.A., Venturini, A.M., Meyer, K.M., Klein, A.M., Tiedje, J.M., Bohannon, B.J., Nüsslein, K., Tsai, S.M., & Rodrigues, J.L. (2015). Differential Response of *Acidobacteria* Subgroups to Forest-to-Pasture Conversion and Their Biogeographic Patterns in the Western Brazilian Amazon. *Frontiers in Microbiology*, 6, 1443. <https://doi.org/10.3389/fmicb.2015.01443>
- Nahi, A., Othman, R., & Omar, D. (2016). Effects of Sb16 bacterial strain and herbicides on endophytic bacterial populations and growth of aerobic rice. *Plant Soil and Environment*, 62 (10), 453-459. <https://doi.org/10.17221/289/2016-PSE>
- Nasarudin, N.A., Mohamad, J., Ismail, S., & Mispan, M.S. (2018). Effect of nitrogen, phosphorus and potassium (NPK) and bacterial bio-fertilizer on the antioxidant activity and chlorophyll content of aerobic rice. *Molecules*, 23, 55. <https://doi.org/10.3390/molecules23010055>
- Ng, L.C., Sariah, M., Sariam, O., Radziah, O., & Zainal Abidin, M.A. (2012). Bio-efficacy of microbial-fortified rice straw compost on rice blast disease severity, growth and yield of aerobic rice. *Australasian Plant Pathology*, 41(5), 541-549. <https://doi.org/10.1007/s13313-012-0145-3>
- Nie, L., Peng, S., Bouman, B.A.M., Huang, J., Cui, K., Visperas, R.M., & Xiang, J. (2009). Alleviating soil sickness caused by aerobic monocropping: Responses of aerobic rice to various nitrogen sources. *Soil Science and Plant Nutrition*, 55, 150–159. <https://doi.org/10.1111/j.1747-0765.2008.00338.x>
- O'Brien, F., Dumont, M.G., Webb, J.S., & Poppy, G.M. (2018). Rhizosphere Bacterial Communities Differ According to Fertilizer Regimes and Cabbage (*Brassica oleracea* var. *134 capitata* L.) Harvest Time, but Not Aphid Herbivory. *Frontiers in Microbiology*, 9, 1620. <https://doi.org/10.3389/fmicb.2018.01620>
- Othman, S., Hussain, Z. P. M. D., Chan, C. S., Man, A., Ahmad, R., & Amzah, B. (2014). Padi Aerob untuk mengatasi masalah kekurangan air [Aerobic rice to solve water shortage problem]. *Jurnal Teknologi*, 70(6), 65-68.
- Otlewska, A., Adamiak, J., Stryszewska, T., Kańka, S., & Gutarowska, B. (2017). Factors Determining the Biodiversity of Halophilic Microorganisms on Historic Masonry Buildings. *Microbes and Environments*, 32(2), 164–173. <https://doi.org/10.1264/jsme2.ME16159>

- Pajares, S., & Brendan J. M. Bohannon, B.J.M. (2016). Ecology of Nitrogen Fixing, Nitrifying, and Denitrifying Microorganisms in Tropical Forest Soils. *Frontiers in Microbiology*. <https://doi.org/10.3389/fmicb.2016.01045>
- Pal, L., Stres, B., Danevčič, T., Leskovec, S., & Mandic-Mulec, I. (2010). Transformations of mineral nitrogen applied to peat soil during sequential oxic/anoxic cycling. *Soil Biology and Biochemistry*, 42, 1338–1346. <https://doi.org/10.1016/j.soilbio.2010.03.013>
- Palacios, O.A., Bashan, Y., & de-Bashan, L.E. (2014). Proven and potential involvement of vitamins in interactions of plants with plant growth-promoting bacteria—an overview. *Biology and Fertility of Soils*, 50, 415–432. <https://doi.org/10.1007/s00374-013-0894-3>
- Peng, S., Bouman, B.A.M., Visperas, R.M., Castaneda, A.R., Nie, L., & Park, H.K. (2006). Comparison between aerobic and flooded rice in the tropics: Agronomic performance in an eight-season experiment. *Field Crops Research*, 96, 252–259. <https://doi.org/10.1016/j.fcr.2005.07.007>
- Pershina, E.V., Ivanova, E.A., Korvigo, I.O., Chirak, E.L., Sergaliev, N.H., Abakumov, E.V., Provorov, N.A., & Andronov, E.E. (2018). Investigation of the core microbiome in main soil types from the East European plain. *Science of the Total Environment*, 631–632, 1421–1430. <https://doi.org/10.1016/j.scitotenv.2018.03.136>
- Phillippot, L., Raaijmakers, J.M., Lemanceau, P., & van der Putten, W.H. (2013). Going back to the roots: the microbial ecology of the rhizosphere. *Nature Reviews Microbiology*, 11, 789–799. <https://doi.org/10.1038/nrmicro3109>
- Pittol, M., Scully, E., Miller, D., Durso, L., Mariana Fiuza, L., & Valiati, V. H. (2018). Bacterial Community of the Rice Floodwater Using Cultivation-Independent Approaches. *International Journal of Microbiology*, 6280484. <https://doi.org/10.1155/2018/6280484>
- Powell, J.R., Welsh, A., & Hallin, S. (2015). Microbial functional diversity enhances predictive models linking environmental parameters to ecosystem properties. *Ecology*, 96(7), 1985–1993. <https://doi.org/10.1890/14-1127.1>
- Praeg, N., Pauli, H., & Illmer, P. (2019). Microbial diversity in bulk and rhizosphere soil of *ranunculus glacialis* along a high-alpine altitudinal gradient. *Frontiers in Microbiology*, 10(JULY). <https://doi.org/10.3389/fmicb.2019.01429>
- Purkayastha, G.D., Mangar, P., Saha, A., & Saha, D. (2018). Evaluation of the biocontrol efficacy of a *Serratia marcescens* strain indigenous to tea rhizosphere for the management of root rot disease in tea. *PLoS one*, 13(2), e0191761. <https://doi.org/10.1371/journal.pone.0191761>

- Quideau, S.A., McIntosh, A.C.S., Norris, C.E., Lloret, E., Swallow, M.J.B., & Hannam, K. (2016). Extraction and Analysis of Microbial Phospholipid Fatty Acids in Soils. *Journal of Visualized Experiments: JoVE*, 114, 54360. <https://doi.org/10.3791/54360>
- Ren, C., Wang, T., Xu, Y., Deng, J., Zhao, F., Yang, G., Hana, X., Feng, Y., & Ren, G. (2018). Differential soil microbial community responses to the linkage of soil organic carbon fractions with respiration across land-use changes. *Forest Ecology and Management*, 409, 170–178. <https://doi.org/10.1016/j.foreco.2017.11.011>
- Ren, Q., Song, H., Yuan, Z., Ni, X., & Li, C. (2018). Changes in Soil Enzyme Activities and Microbial Biomass after Revegetation in the Three Gorges Reservoir, China. *Forests* 9(5), 249. <https://doi:10.3390/f9050249>
- Roper, M., Ladha, J.K., Peoples, M.B. (1995). Biological nitrogen fixation by heterotrophic and phototrophic bacteria in association with straw. *Plant and Soil*, 174, 211-224.
- Rosberg, A.K., Gruyer, N., Hultberg, M., Wohanka, W., & Alsanius, B.W. (2014). Monitoring rhizosphere microbial communities in healthy and *Pythium ultimum* inoculated tomato plants in soilless growing systems. *Scientia Horticulturae*, 173, 106-113. <https://doi.org/10.1016/j.scienta.2014.04.036>
- Rose, T.J., Quin, P., Morris, S.G., Kearney, L.J., Kimber, S., Rose, M.T., & Van Zwieten, L. (2018). No evidence for higher agronomic N use efficiency or lower nitrous oxide emissions from enhanced efficiency fertilisers in aerobic subtropical rice. *Field Crops Research*, 225, 47–54. <https://doi.org/10.1016/j.fcr.2018.06.001>
- Sadet-Bourgeteau, S., Houot, S., Dequiedt, S., Nowak, V., Tardy, V., Terrat, S., Montenach, D., Mercier, V., Karimi, B., Chemidlin Prévost-Bouré, N., & Maron, P.A. (2018). Lasting effect of repeated application of organic waste products on microbial communities in arable soils. *Applied Soil Ecology*, 125, 278–287. <https://doi.org/10.1016/j.apsoil.2018.02.006>
- Sathya, A., Vijayabharathi, R., & Gopalakrishnan, S. (2017). Plant growth-promoting actinobacteria: a new strategy for enhancing sustainable production and protection of grain legumes. *3 Biotech*, 7, 102. <https://Doi:10.1007/s13205-017-0736-3>
- Schellenberger, S., Drake, H.L., & Kolb, S. (2011). Functionally redundant cellobiose-degrading soil bacteria respond differentially to oxygen. *Applied and environmental microbiology*, 77(17), 6043-8. DOI: 10.1128/AEM.00564-11
- Schmidt, O. (2016). *Hydrogen Metabolizers: Drivers of Anaerobic Degradation Processes in Peatlands and Earthworm Guts*. [Doctoral dissertation, University of Bayreuth]. ERef Bayreuth.

- Schreiter, S., Ding, G.C., Heuer, H., Neumann, G., Sandmann, M., Grosch, R., Kropf, S., & Smalla, K. (2014). Effect of the soil type on the microbiome in the rhizosphere of field-grown lettuce. *Frontiers in Microbiology*, 5, 144. <https://doi.org/10.3389/fmicb.2014.00144>
- Selim, S., Hassan, S., Hagagy, N., Kraková, L., Grivalský, T., & Pangallo, D. (2017). Assessment of Microbial Diversity in Saudi Springs by Culture-Dependent and Culture-Independent Methods. *Geomicrobiology Journal*, 34 (5), 443-453. <https://doi.org/10.1080/01490451.2016.1219430>
- Sessitsch, A., Weilharter, A., Gerzabek, M.H., Kirchmann, H., Kandeler, E. (2001). Microbial population structures in soil particle size fractions of a long-term fertilizer field experiment. *Applied Environmental Microbiology*, 67, 4215–4222. <https://doi: 10.1128/AEM.67.9.4215-4224.2001>
- Setia, R., Marschner, P., Baldock, J., Chittleborough, D., Smith, P., & Smith, J. U. (2011). Salinity effects on carbon mineralization in soils of varying texture. *Soil Biology and Biochemistry*, 43(9), 1908-1916. <https://doi.org/10.1016/j.soilbio.2011.05.013>
- Shang, Y., Zhao, Y., Shi, H., Wang, L., & Wang, J. (2015). Analysis of water use trend and its impact factors in Tianjing: South to North Water Transfers. *Water Science and Technology*, 13(1), 33-38. <https://DOI:10.13476/j.cnki.nsbdkq.2015.01.035>
- Shanmugam, S.G., & Kingery, W.L. (2018). Changes in soil microbial community structure in relation to plant succession and soil properties during 4000 years of pedogenesis. *European Journal of Soil Biology*, 88, 80–88. <https://doi.org/10.1016/j.ejsobi.2018.07.003>
- Shen, Y., Chen, Y., & Li, S. (2016). Microbial Functional Diversity, Biomass and Activity as Affected by Soil Surface Mulching in a Semiarid Farmland. *PLoS ONE* 11(7), e0159144. <https://doi.org/10.1371/journal.pone.0159144>
- Shi, S., Nuccio, E., Herman, D.J., Rijkers, R., Estera, K., Li, J., Rocha, U.N., He, Z., Pett-Ridge, J., Brodie, E.L., Zhou, J., & Firestone, M. (2015). Successional Trajectories of Rhizosphere Bacterial Communities over Consecutive Seasons. *mBio*, 6 (4), e00746-15. <https://DOI: 10.1128/mBio.00746-15>
- Siddiqi, M.Z., Kim, S.B., Cho, J.C., Yoon, J.H., Joh, K.S., Seong, C.H., Bae, J.W., Jahng, K.Y., Jeon, C.O. & Im, W.T. (2017). Description of 39 unrecorded bacterial species in Korea, belonging to the class *Alphaproteobacteria*. *Journal of Species Research*, 6(2), 141-153. <https://DOI:10.12651/JSR.2017.6.2.141>

- Singh, A.V., Prasad, B., & Goel, R. (2018). Plant Growth Promoting Efficiency of Phosphate Solubilizing *Chryseobacterium* sp. PSR 10 with Different Doses of N and P Fertilizers on Lentil (*Lens culinaris* var. PL-5) Growth and Yield. *International Journal of Current Microbiology and Applied Sciences*, 7(5), 2280-2289. [https://DOI: 10.20546/ijcmas.2018.705.265](https://doi.org/10.20546/ijcmas.2018.705.265)
- Singh, M., Bhullar, M.S., & Chauhan, B.S. (2017). Relative time of weed and crop emergence is crucial for managing weed seed production: A study under an aerobic rice system. *Crop Protection*, 99, 33-38. <https://doi.org/10.1016/j.cropro.2017.05.013>
- Sinsabaugh, R. L. (2010). Phenol oxidase, peroxidase and organic matter dynamics of soil. *Soil Biology and Biochemistry*, 42(3), 391-404. <https://doi.org/10.1016/j.soilbio.2009.10.014>
- Song, Y., & Lin, Z. (2014). Abundance and Community Composition of Ammonia-Oxidizers in Paddy Soil at Different Nitrogen Fertilizer Rates. *Journal of Integrative Agriculture*, 13(4), 870-880. [https://doi.org/10.1016/S2095-3119\(13\)60426-8](https://doi.org/10.1016/S2095-3119(13)60426-8)
- Spang, A., Poehlein, A., Offre, P., Zumbrägel, S., Haider, S., Rychlik, N., Nowka, B., Schmeisser, C., Lebedeva, E.V., Rattei, T., Böhm, C., Schmid, M., Galushko, A., Hatzepichler, R., Weinmaier, T., Daniel, R., Schleper, C., Spieck, E., Streit, W., & Wagner, M. (2012). The genome of the ammonia-oxidizing *Candidatus Nitrososphaera gargensis*: insights into metabolic versatility and environmental adaptations. *Environmental Microbiology*, 14(12), 3122-3145. [https://doi: 10.1111/j.1462-2920.2012.02893.x](https://doi:10.1111/j.1462-2920.2012.02893.x)
- Sripreechusak, P., Phongsopitanun, W., Supong, K., Pittayakhajonwut, P., Kudo, T., Ohkuma, M., & Tanasupawat, S. (2017). *Nonomuraea rhodomycinica* sp. nov., isolated from peat swamp forest soil. *International Journal of Systematic and Evolutionary Microbiology* 67, 1683-1687. <https://doi:10.1099/ijsem.0.001843>
- Stieglmeier, M., Alves, R.J.E., & Schleper, C. (2014). The Phylum *Thaumarchaeota*. In: Rosenberg E., DeLong E.F., Lory S., Stackebrandt E., & Thompson F. (Eds.), *The Prokaryotes*. Springer
- Stocker, T.F., Qin, D., Plattner, G.K., Tignor, M., Allen, S.K., Boschung, J., Nauels, A., Xia, Y., Bex, V., & P.M. Midgley, P.M. (Eds.). (2013). *IPCC, 2013: Climate Change 2013: The Physical Science Basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change*. Cambridge University Press
- Sugiyama, A., Ueda, Y., Zushi, T., Takase, H., & Yazaki, K. (2014). Changes in the bacterial community of soybean rhizospheres during growth in the field. *PLoS ONE*, 9(6), 1-9. <https://doi.org/10.1371/journal.pone.0100709>

- Suhadolnik, M.L.S., Salgado, A.P.C, Scholte, L.L.S., Bleicher, L., Costa, P.S., Reis, M.P., Dias, M.F., Ávila, M.P., Barbosa, F.A.R., Chartone-Souza, E., & Nascimento, A.M.A. (2017). Novel arsenic-transforming bacteria and the diversity of their arsenic-related genes and enzymes arising from arsenic-polluted freshwater sediment. *Scientific Reports*, 7, Article number: 11231. <https://DOI: 10.1038/s41598-017-11548-8>
- Sun, W., Dong, Y., Gao, P., Fu, M., Ta, K., & Li, J. (2015). Microbial communities inhabiting oil-contaminated soils from two major oilfields in Northern China: Implications for active petroleum-degrading capacity. *Journal of Microbiology*, 53(6), 371-378. <https://Doi.org/10.1007/s12275-015-5023-6>
- Supong, K., Sriprechasak, P., Phongsopitanun, W., Tanasupawat, S., Danwisetkanjana, K., Bunbamrung, N., & Pittayakhajonwut, P. (2018). Antimicrobial substances from the rare actinomycete *Nonomuraea rhodomycinica* NR4-ASC07^T. *Natural Product Research*, 33(16), 2285-2291. <https://Doi.org/10.1080/14786419.2018.1440223>
- Sutradhar, M. (2015). Metagenomic analysis of rhizosphere microbial diversity in rice grown under irrigated and aerobic condition. *Dissertation Abstracts International*, <http://krishikosh.egranth.ac.in/handle/1/5810027635>
- Suzuki, K., Takemura, M., Miki, T., Nonaka, M., & Harada, N. (2019). Differences in Soil Bacterial Community Compositions in Paddy Fields under Organic and Conventional Farming Conditions. *Microbes and Environments*, 34(1), 108–111. <https://Doi:10.1264/j sme2.ME18101>
- I, S.K. (1998). *Modernization of irrigation management*. Paper presented at 6th MANCID annual conference, Sustainable rice production, Alor Setar, Kedah
- Tang, Y.F., Zhang, M.M., Chen, A.L., Zhang, W.Z., Wei, W. X. and Sheng, R. (2017). Impact of fertilization regimes on diazotroph community compositions and N₂-fixation activity in paddy soil. *Agriculture, Ecosystems and Environment*, 247, 1-8. <https://doi.org/10.1016/j.agee.2017.06.009>
- Tian, J., Wang, J., Dippold, M., Gao, Y., Blagodatskaya, E., & Kuzyakov, Y. (2016). Biochar affects soil organic matter cycling and microbial functional functions but does not alter microbial community structure in a paddy soil. *Science of the Total Environment*, 556, 89-97. <http://dx.doi.org/10.1016/j.scitotenv.2016.03.010>
- Tilg, H., Adolph, T.E., Gerner, R.R., & Moschen, A.R. (2018). The intestinal microbiota in colorectal cancer. *Cancer Cell*, 33(6), 954-964. <https://DOI: 10.1016/j.ccell.2018.03.004>

- Too, C.C., Keller, A., Sickel, W., Lee, S.M., & Yule, C.M. (2018). Microbial community structure in a Malaysian Tropical Peat Swamp Forest: The Influence of Tree Species and Depth. *Frontiers in Microbiology*. <https://doi.org/10.3389/fmicb.2018.02859>
- Tuong, T.P., & Bouman, B.A.M. (2003). Rice production in water-scarce environments. In: J.W. Kijne, R. Barker, & D. Molden (Eds.), *Water Productivity in Agriculture: Limits and Opportunities for Improvement* (pp. 53-67). CABI Publishing.
- Utermann, C., Parrot, D., Breusing, C., Stuckas, H., Staufenberger, T., Blümel, M., Labes, A., & Tasdemi, D. (2018). Combined genotyping, microbial diversity and metabolite profiling studies on farmed *Mytilus* spp. from Kiel Fjord. *Scientific Reports*, 8, Article number: 7983. <https://doi.org/10.1038/s41598-018-26177-y>
- Van Elsas, J.D., Jansson, J.K., & Trevors, J.T. (2007). *Modern Soil Microbiology II*. CRC Press.
- Vincent, A.T., Derome, N., Boyle, B., Culley, A.I., & Charette, S.J. (2017). Next-generation sequencing (NGS) in the microbiological world: How to make the most of your money. *Journal of Microbiological Methods*, 138, 60-71. <https://doi.org/10.1016/j.mimet.2016.02.016>
- Vishwakarma, P., & Dubey, S.K. (2019). Diversity of endophytic bacterial community inhabiting in tropical aerobic rice under aerobic and flooded condition. *Archives of Microbiology*, 1-13. <https://doi.org/10.1007/s00203-019-01715-y>
- Visioli, G., Sanangelantoni, A.M., Vameralli, T., Dal Cortivo, C., & Blandino, M. (2018). 16S rDNA Profiling to Reveal the Influence of Seed-Applied Biostimulants on the Rhizosphere of Young Maize Plants. *Molecules*, 23(6), 1461. <https://doi.org/10.3390/molecules23061461>
- Vranova, V., Rejsek, K., Skene, K.R., & Formanek, P. (2011). Non-protein amino acids: Plant, soil and ecosystem interactions. *Plant and Soil*, 342, 31–48. <https://doi.org/10.1007/s11104-010-0673-y>
- Wagner, S.C. (2011). Biological Nitrogen Fixation. *Nature Education Knowledge*, 3(10), 15. <https://www.nature.com/scitable/knowledge/library/biological-nitrogen-fixation-23570419/>
- Wakelin, S.A., Macdonald, L.M., Rogers, S.L., Gregg, A.L., Bolgerd, T.P., & Baldock, J.A. (2008). Habitat selective factors influencing the structural composition and functional capacity of microbial communities in agricultural soils. *Soil Biology & Biochemistry*, 40, 803–813. <https://doi.org/10.1016/j.soilbio.2007.10.015>

- Wang, H., Bouman, B.A.M., Zhao, D., Wang, C., & Moya, P. F. (2002). Aerobic rice in northern China—Opportunities and challenges. In: B.A.M. Bouman, H. Hengsdijk, B. Hardy, P.S. Bindraban, T.P. Tuong, & J. K. Ladha (Eds.) *Water-Wise Rice Production, Proceedings of the International Workshop on Water-Wise Rice Production* (pp. 143–154)., Los Banos, Philippines. International Rice Research Institute.
- Wang, Y., Ke, X., Wu, L., & Lu, Y. (2009). Community composition of ammonia-oxidizing bacteria and archaea in rice field soil as affected by nitrogen fertilization. *Systematic and Applied Microbiology*, 32(1), 27–36. <https://doi.org/10.1016/j.syapm.2008.09.007>
- Wang, H., Li, X., Li, X., Li, X., Wang, J., Zhang, H. (2017). Changes of microbial population and N-cycling function genes with depth in three Chinese paddy soils. *PloS ONE*, 12(12), e0189506. <https://doi.org/10.1371/journal.pone.0189506>
- Wang, Ji., Sui, X., Zhang, R., Xu, N., Yang, L., Liu, Y., Fu, X., Chai, C., Xu, M., Xing, J., Zhong, H., Ni, H., & Li, M. (2017). Effects of Different Land Use on Soil Bacterial Functional Diversity in Sanjiang Plain, Northeast China. *Journal of Residuals Science & Technology*, 14(1), 91-98. <https://www.dora.lib4ri.ch/wsl/islandora/object/wsl:15691>
- Wang, M., Li, S., Chen, S., Meng, N., Li, X., Zheng, H., Zhao, C., & Wang, D. (2019).
- Manipulation of the rhizosphere bacterial community by biofertilizers is associated with mitigation of cadmium phytotoxicity. *Science of the Total Environment*, 649, 413–421. <https://doi.org/10.1016/j.scitotenv.2018.08.174>
- Wang, R., Zhang, H., Sun, L., Qi, G., Chen, S., & Zhao, X. (2017). Microbial community composition is related to soil biological and chemical properties and bacterial wilt outbreak. *Scientific Reports*, 7, Article number: 343. <https://DOI: 10.1038/s41598-017-00472-6>
- Wang, Q., Garrity, G.M., Tiedje, J.M. & Cole, J.R. (2007). A Bayesian classifier for rapid assignment of rRNA sequences into the new bacterial taxonomy. *Applied and Environmental Microbiology* 73(16), 5261-5267. <https://DOI: 10.1128/AEM.00062-07>
- Wang, S.G, Hou, Y.L., & Wei Guo, W. (2010). Responses of nitrogen transformation and microbial community composition to nitrogen enrichment patch. *Pedobiologia*, 54, 9–17. <https://doi.org/10.1016/j.pedobi.2010.08.002>
- Wang, X., Wang, Q., Wang, S., Li, F., & Guo, G. (2012). Effect of biostimulation on community level physiological profiles of microorganisms in field-scale biopiles composed of aged oil sludge. *Bioresource Technology*, 111, 308–315. <https://doi.org/10.1016/j.biortech.2012.01.158>

- Wang, Y., Luo, X., & Zhang, L. (2017). Draft genome sequence of *Glycomyces fuscus* TRM 49117, isolated from a hypersaline soil sample. *Genome Announcements*, 5, e01258-17. <https://doi.org/10.1128/genomeA.01258-17>.
- Wang, Y., Wang, Z.L., Zhang, Q., Hu, N., Li, Z., Lou, Y., Li, Y., Xue, D., Chen, Y., Wu, C., Zou, C.B., & Kuzyakov, Y. (2018). Long-term effects of nitrogen fertilization on aggregation and localization of carbon, nitrogen and microbial activities in soil. *Science of the Total Environment*, 624, 1131–1139. <https://doi.org/10.1016/j.scitotenv.2017.12.113>
- Wang, M., Chen, S., Zheng, H., Li, S., Chen, L., & Wang, D. (2020). The responses of cadmium phytotoxicity in rice and the microbial community in contaminated paddy soils for the application of different long-term N fertilizers. *Chemosphere*, 238, 124700. <https://doi.org/10.1016/j.chemosphere.2019.124700>
- Wartiainen, I., Eriksson, T., Zheng, W., & Rasmussen, U. (2008). Variation in the active diazotrophic community in rice paddy-nifH PCR-DGGE analysis of rhizosphere and bulk soil. *Applied Soil Ecology*, 39, 65–75. <https://doi.org/10.1016/j.apsoil.2007.11.008>
- Weber, K.P. & Legge, R.L. (2009). One-dimensional metric for tracking bacterial community divergence using sole carbon source utilization patterns. *Journal of Microbiological Methods*, 79, 55-61. <https://doi.org/10.1016/j.mimet.2009.07.020>
- Weber, E.B., Lehtovirta-Morley, L.E., Prosser, J.I., & Gubry-Rangin, C. (2015). Ammonia oxidation is not required for growth of Group 1.1c soil Thaumarchaeota. *FEMS Microbiology Ecology*, 91(3), <https://doi.org/10.1093/femsec/fiv001>
- Wei, D., Yang, Q., Zhang, J.Z., Wang, S., Chen, X.L., Zhang, X.L., & Li, W.Q. (2008). Bacterial Community Structure and Diversity in a Black Soil as Affected by Long-Term Fertilization. *Pedosphere*, 18(5), 582–592. [https://doi.org/10.1016/S1002-0160\(08\)60052-1](https://doi.org/10.1016/S1002-0160(08)60052-1)
- Winter, G., Todd, C.D., Trovato, M., Forlani, G. & Funck, D. (2015). Physiological implications of arginine metabolism in plants. *Frontiers in Plant Science*, 6(534), 1-14. <https://doi.org/10.3389/fpls.2015.00534>
- Wolińska, A., Szafranek-Nakonieczna, A., Banach, A., Błaszczuk, M., & Stępniewska, Z. (2016). The impact of agricultural soil usage on activity and abundance of ammonifying bacteria in selected soils from Poland. *SpringerPlus*, 5, 565. <https://doi.org/10.1186/s40064-016-2264-8>
- Wu, Y., & Conrad, R. (2014). Ammonia oxidation-dependent growth of group 1.1b Thaumarchaeota in acidic red soil microcosms. *FEMS Microbiology Ecology*, 89(1), 127–134. <https://doi.org/10.1111/1574-6941.12340>

- Wu, Y., Zeng, J., Qi, Zhu, Q., Zhang, Z., & Lin, X. (2017). pH is the primary determinant of the bacterial community structure in agricultural soils impacted by polycyclic aromatic hydrocarbon pollution. *Scientific Reports*, 7, Article number:40093. <https://DOI: 10.1038/srep40093>
- Xu, L. L., Wang, Q. B., Zhang, X. Y., Sun, X. M., Dai, X. Q., Yang, F. T., Bu, J. F., & Wang, H. M. (2013). Effects of applying kind fertilizers on enzyme activities related to carbon, nitrogen, and phosphorus cycles in reddish paddy soil. *Ying Yong Sheng Tai Xue Bao = The Journal of Applied Ecology*, 24(4), 909–914.
- Xue, P.P., Carrillo, Y., Pino, V., Minasny, B., & McBratney, A.B. (2018). Soil Properties Drive Microbial Community Structure in a Large Scale Transect in South Eastern Australia. *Scientific Reports*, 8, Article number: 11725. <https://doi.org/10.1038/s41598-018-30005-8>
- Yan, Y., Kuramae, E.E., Hollander, M.D., Klinkhamer, P.G.L., & Veen, J.A.V.V. (2017). Functional traits dominate the diversity-related selection of bacterial communities in the rhizosphere. *The ISME Journal*, 11, 56–66. <https://doi:10.1038/ismej.2016.108>
- Yang, Y., Wang, P., & Zeng, Z. (2019). Dynamics of Bacterial Communities in a 30-Year Fertilized Paddy Field under Different Organic-Inorganic Fertilization Strategies. *Agronomy*, 9, 14. <https://Doi:10.3390/agronomy9010014>
- Yao, L., Wang, D., Kang, L., Wang, D., Zhang, Y., Hou, X., & Guo, Y. (2018). Effects of fertilizations on soil bacteria and fungi communities in a degraded arid steppe revealed by high through-put sequencing. *PeerJ*, 6, e4623. <https://doi:10.7717/peerj.4623>
- Yoshida, M., Ishii, S., Otsuka, S., & Senoo, K. (2009). Temporal shifts in diversity and quantity of *nirS* and *nirK* in a rice paddy field soil. *Soil Biology and Biochemistry*, 41(10), 2044–2051. <https://doi.org/10.1016/j.soilbio.2009.07.012>
- Yu, C., Hu, X.M., Deng, W., Li, Y., Xiong, C., C.H., Ye, Han, G.M. & Li, X. (2015). Changes in soil microbial community structure and functional diversity in the rhizosphere surrounding mulberry subjected to long-term fertilization. *Applied Soil Ecology*, 86, 30–40. <https://doi.org/10.1016/j.apsoil.2014.09.013>
- Yu, H., Gao, Q., Shao, Z., Ying, A., Sun, Y., Liu, J., Mao, W., & Zhang, B. (2016). Decreasing Nitrogen Fertilizer Input Had Little Effect on Microbial Communities in Three Types of Soils. *PLOS ONE*, 11(3), e0151622. <https://doi.org/10.1371/journal.pone.0151622>

- Yu, Z., Hu, Y., Dzakpasu, M., Wang, X.C., & Huu Hao Ngo, H.H. (2018). Dynamic membrane bioreactor performance enhancement by powdered activated carbon addition: Evaluation of sludge morphological, aggregative and microbial properties. *Journal of Environmental Sciences*, 75, 73-83. <https://doi.org/10.1016/j.jes.2018.03.003>
- Yuan, H., Zhu, Z., Wei, X., Liu, S., Peng, P., Gunina, A., ... Wang, J. (2019). Straw and biochar strongly affect functional diversity of microbial metabolism in paddy soils. *Journal of Integrative Agriculture*, 18(7), 1474–1485. [https://doi.org/10.1016/S2095-3119\(18\)62102-1](https://doi.org/10.1016/S2095-3119(18)62102-1)
- Yuan, J., Yuan, Y., Zhu, Y., & Cao, L. (2018). Effects of different fertilizers on methane emissions and methanogenic community structures in paddy rhizosphere soil. *Science of the Total Environment*, 627, 770–781. <https://doi.org/10.1016/j.scitotenv.2018.01.233>
- Zak, J.C., Willig, M.R., Moorhead, D.L. & Wildman, H.G. (1994). Functional diversity of microbial communities: A quantitative approach. *Soil Biology and Biochemistry*, 26(9),1101-1108. [https://doi.org/10.1016/0038-0717\(94\)90131-7](https://doi.org/10.1016/0038-0717(94)90131-7)
- Zakaria, L., Yaakop, A. S., Salleh, B. and Zakaria M. (2010). Endophytic fungi from paddy. *Tropical Life Sciences Research*, 21(1), 101-107.
- Zaman, N.K., Abdullah, M.Y., Othman, S., & Zaman, N.K. (2018). Growth and physiological performance of aerobic and lowland rice as affected by water stress at selected growth stages. *Rice Science*, 25(2), 82-93. <https://doi.org/10.1016/j.rsci.2018.02.001>
- Zeiller, M., Rothballer, M., Iwobi, A.N., Böhnelt, H., Gessler, F., Hartmann, A., & Schmid, M. (2015). Systemic colonization of clover (*Trifolium repens*) by *Clostridium botulinum* strain 2301. *Frontiers in Microbiology*, 6, 1207. <https://doi.org/10.3389/fmicb.2015.01207>
- Zeng, M., Zhong, Y., Cai, S., & Diao, Y. (2018). Deciphering the bacterial composition in the rhizosphere of *Baphicacanthus cusia* (Nees) Bremek. *Scientific Reports*, 8, Article number: 15831. <https://doi.org/10.1038/s41598-018-34177-1>
- Zenova, G. M., Gryadunova, A. A., Pozdnyakov, A. I. and Zvyagintsev, D. G. (2008). Aerobic and microaerophilic actinomycetes of typical agropeat and peat soils. *Eurasian Soil Science*, 41(2), 210-214. <https://doi.org/10.1134/S1064229308020129>
- Zhalnina, K., Dias, R., de Quadros, P.D., Davis-Richardson, A., Camargo, F.A., Clark, I.M., McGrath, S.P., Hirsch, P.R., & Triplett, E.W. (2015). Soil pH determines microbial diversity and composition in the park grass experiment. *Microbial Ecology*, 69(2), 395–406. <https://doi.org/10.1007/s00248-014-0530-2>

- Zhang, B., Zhang, J., Liu, Y., Shi, P., & Wei, G. (2018). Co-occurrence patterns of soybean rhizosphere microbiome at a continental scale. *Soil Biology and Biochemistry*, 118, 178–186. <https://doi.org/10.1016/j.soilbio.2017.12.011>
- Zhang, J., Li, M., Li, P., Kang, W., Shi, J., Yang, L., & Wang, S. (2018). Effect of pig manure amendments on microbial biomass and functional diversity in three agricultural soils. *IOP Conference Series: Earth and Environmental Science*, 012061. [https://doi :10.1088/1755-1315/186/3/012061](https://doi.org/10.1088/1755-1315/186/3/012061)
- Zhang, X., Wang, X., Tang, Q., Li, N., Liu, P., Dong, Y., Pang, W., Yang, J. & Wang, Z. (2015). Effects of cultivation of OsrHSA transgenic rice on functional diversity of microbial communities in the soil rhizosphere. *The Crop Journal*, 3, 163-167. <https://doi.org/10.1016/j.cj.2014.11.001>
- Zhang, Y., Shen, H., He, X., Thomas, B.W., Lupwayi, N.Z., Hao, X., Thomas, M.C., & Shi, X. (2017). Fertilization Shapes Bacterial Community Structure by Alteration of Soil pH. *Frontiers in Microbiology*, 8, 1325. <https://doi.org/10.3389/fmicb.2017.01325>
- Zhao, J., Liu, J., Liang, H., Huang, J., Chen, Z., Nie, Y., Wang, C., & Wang, Y. (2018). Manipulation of the rhizosphere microbial community through application of a new bio-organic fertilizer improves watermelon quality and health. *PLoS ONE*, 13(2), e0192967. <https://doi.org/10.1371/journal.pone.0192967>
- Zhao, Y., Chen, Y., Li, Z., & Peng, W. (2018). Environmental factors have a strong impact on the composition and diversity of the gut bacterial community of Chinese black honeybees. *Journal of Asia-Pacific Entomology*, 21, 261–267. <https://DOI:10.1016/J.ASPEN.2018.01.002>
- Zhao, Y.N., Zhang, Y.Q., Du, H.X., Wang, Y.H., Zhang, L.M., & Shi, X.J. (2015). Carbon sequestration and soil microbes in purple paddy soil as affected by long-term fertilization. *Toxicological & Environmental Chemistry*, 97(3-4), 464-476. <https://doi.org/10.1080/02772248.2015.1050200>
- Zheng, B.X., Ibrahim, M., Zhang, D.P., Bi, Q.F., Li, H.Z., Zhou, G.W., Ding, K., Peñuelas, J., Zhu, Y.G., & Yang, X.R. (2018). Identification and characterization of inorganic phosphate-solubilizing bacteria from agricultural fields with a rapid isolation method. *AMB Express*, 8, Article Number: 47. <https://doi.org/10.1186/s13568-018-0575-6>
- Zheng, X.B., Hao, X.L., Ding, K., Zhou, G.W., Chen, Q.L., Zhang J.B., & Zhu, Y.G. (2017). Long-term nitrogen fertilization decreased the abundance of inorganic phosphate solubilizing bacteria in an alkaline soil. *Scientific Reports*, 7, Article number: 42284. [https://doi: 10.1038/srep42284](https://doi.org/10.1038/srep42284)

Zhou, J., Jiang, X., Wei, D., Zhao, B., Ma, M., Chen, S., Cao, F., Shen, D., Guan, D., & Li, J. (2017). Consistent effects of nitrogen fertilization on soil bacterial communities in black soils for two crop seasons in China. *Scientific Reports*, 7, Article number: 3267. [https://doi: 10.1038/s41598-017-03539-6](https://doi.org/10.1038/s41598-017-03539-6)

Zhou, X., Zhang, Z., Tian, L., Li, X., & Tian, C. (2017). Microbial communities in peatlands along a chronosequence on the Sanjiang Plain, China. *Scientific Reports*, 7, 9567. [https://doi:10.1038/s41598-017-10436-5](https://doi.org/10.1038/s41598-017-10436-5)

Zhu, J., Peng, H., Ji, X., Li, C., & Li, S. (2019). Effects of reduced inorganic fertilization and rice straw recovery on soil enzyme activities and bacterial community in double-rice paddy soils. *European Journal of Soil Biology*, 94, 103116. <https://doi.org/10.1016/j.ejsobi.2019.103116>

BIODATA OF STUDENT

Nor Ayshah Alia Binti Ali Hassan was born in Batu Pahat, Johor in 1980. Mainly raised in Negeri Sembilan, she attended her primary school at Sekolah Kebangsaan Palong 5 and later attended Sekolah Menengah King George V in Seremban. She attained her Bachelor Degree of Science particularly in microbiology at Universiti Putra Malaysia in 2003 and later obtained her Master of Science (Environmental Microbiology) in 2007 at the same university. On 2006, she joined Malaysian Agricultural Research and Development Institute (MARDI) as a research officer. She has been a project leader and also a collaborator for various project. Her previous research projects were focusing on bioprospecting of beneficial microbes for agriculture, management of microbial culture collection and evaluating microbial diversity in soil.

LIST OF PUBLICATIONS

Ali Hassan, N.A.A., Othman, R., & Sadi, T. (2019). The response of microbial community in aerobic rice rhizosphere that affected by various plant growth stages and soil types. *Malaysian Journal of Microbiology*, 15(4) Special Issue 2019, 178-189

Ali Hassan, N.A.A., Mohd Saud, H. (2020). Characterization of Peat Microbial Functional Diversity in Aerobic Rice Rhizosphere. *Pertanika Journal of Tropical Agricultural Science*, 43 (4): 677 – 692.





UNIVERSITI PUTRA MALAYSIA

STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

ACADEMIC SESSION : Second Semester 2020/2021

TITLE OF THESIS / PROJECT REPORT :

CHARACTERIZATION OF MICROBIAL FUNCTIONAL DIVERSITY AND BACTERIAL
COMMUNITY STRUCTURE AFFECTED BY FERTILISER, SOIL AND PLANT GROWTH
STAGES IN AEROBIC RICE

NAME OF STUDENT: NOR AYSHAH ALIA BINTI ALI HASSAN

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

1. This thesis/project report is the property of Universiti Putra Malaysia.
2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as :

*Please tick (✓)

CONFIDENTIAL

(Contain confidential information under Official Secret Act 1972).

RESTRICTED

(Contains restricted information as specified by the organization/institution where research was done).

OPEN ACCESS

I agree that my thesis/project report to be published as hard copy or online open access.

This thesis is submitted for :

PATENT

Embargo from _____ until _____
(date) (date)

Approved by:

(Signature of Student)
New IC No/ Passport No.:

Date :

(Signature of Chairman of Supervisory Committee)
Name:

Date :

[Note : If the thesis is CONFIDENTIAL or RESTRICTED, please attach with the letter from the organization/institution with period and reasons for confidentiality or restricted.]