



**UNIVERSITI PUTRA MALAYSIA**

***DETERMINATION OF FUNCTIONALLY IMPORTANT SEQUENCE  
VARIANTS IN PROMOTER SEQUENCE OF HOMOGENISATE  
GERANYLGERANYL TRANSFERASE FROM *Elaeis guineensis* Jacq.  
GERMPLASM MATERIALS***

**MOHD SHAHRUL NIZWANSHAH BIN KARIM**

**IPTSM 2021 16**



**DETERMINATION OF FUNCTIONALLY IMPORTANT SEQUENCE  
VARIANTS IN PROMOTER SEQUENCE OF HOMOGENISATE  
GERANYLGERANYL TRANSFERASE FROM *Elaeis guineensis* Jacq.  
GERMPLASM MATERIALS**

**By**

**MOHD SHAHRUL NIZWANSHAH BIN KARIM**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfilment of the Requirement for the Degree of Master of Science**

**October 2020**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



## DEDICATION

*This thesis is dedicated to my wife, Tuan Syaripah Najihah, who instilled in me the virtues of perseverance, determination and relentlessly encouraged me to strive for excellence. She's been a constant source of support and encouragement during the challenges of graduate school and life. This work is also dedicated to my mother Norminah Atim who has always loved me unconditionally and whose good examples have taught me to work hard for the things that I aspire to achieve.*



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

**DETERMINATION OF FUNCTIONALLY IMPORTANT SEQUENCE  
VARIANTS IN PROMOTER SEQUENCE OF HOMOGENISATE  
GERANYLGERANYL TRANSFERASE FROM *Elaeis guineensis* Jacq.  
GERMPLASM MATERIALS**

By

**MOHD SHAHRUL NIZWANSHAH BIN KARIM**

**October 2020**

**Chairman: Professor Datin Siti Nor Akmar Abdullah, PhD  
Faculty : Institute of Tropical Agriculture & Food Security**

Vitamin E consists of tocopherols and tocotrienols which are lipid-soluble compounds produced by plants with various beneficial medicinal properties. Tocotrienols have cholesterol-lowering and anticancer properties and alpha-tocotrienols can prevent inducible neurodegeneration. There is high variability in the level of vitamin E in the *E. guineensis* germplasm materials from Angola and Tanzania ranging from 300-1600 ppm, while the level is 500-1000 ppm in the commercial variety. However, the development of markers for tocotrienols rich oil palm is still very scarce and the understanding of the regulation of vitamin E biosynthesis is important for genetic improvement specifically nutritionally vitamin E rich oil palm. Therefore, this study aimed to identify the SNPs in the promoter of HGGT that associate with the high vitamin E in *E. guineensis* germplasm materials. *Homogentisate geranylgeranyl transferase (HGGT)* is an important vitamin E biosynthetic enzyme that catalyses the first committed step for tocotrienols biosynthesis. Sequence alignment of 14 accessions from Angola and Tanzania with varying levels of vitamin E content showed the presence of an SNP (-454) that associate with high vitamin E Angolan (AH) and Tanzanian (TH) palms and two SNPs, -781 and -113 unique to AH. Functional characterisation to determine the roles of SNPs in influencing HGGT promoter activity was carried out by reporter gene assay in mesocarp tissues bombarded with four different HGGT promoter constructs. The pBGWFS7 vector used for cloning the HGGT promoter fragments contains two reporter genes, a green fluorescent protein (GFP) and GUS. The promoters were commercial DXP (COM), AH1, TH96 and a mutated derivative of COM (CM, g.-454A>G) produced by introducing the variant found in common in AH1 and TH96. The quantitative fluorometric GUS assay on GFP positive bombarded tissues shows that the lowest expression level was obtained from COM with GUS expression of 0.27 pmol MU min<sup>-1</sup> mg protein<sup>-1</sup> per copy numbers. The expression level obtained for COM is about the same as the constitutive CaMV 35S promoter that was used for comparison. There was no significant difference in the GUS expression level between the COM with the CM (g.-454A>G). The results showed that AH1 gave the highest GUS expression of 0.82 pmol MU min<sup>-1</sup> mg protein<sup>-1</sup> per copy numbers. It

suggested that the CAAT-box unique to AH at the SNP (-113), a well-known proximal promoter element may enhance HGGT promoter activity and the tocotrienols content in AH1. Taken together, these studies have provided valuable information on the vitamin E biosynthetic regulatory mechanism in oil palm germplasm as these results suggest that the HGGT promoter is potentially useful for engineering of high vitamin E markers through SNPs association and it is essential for the future genetic improvement effort to produce high vitamin E oil palm.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

**PENENTUAN KEPELBAGAIAN JUJUKAN YANG PENTING DALAM  
JUJUKAN PROMOTER KEPADA HOMOGENTISATE GERANYLGERANYL  
TRANSFERASE GEN DARIPADA KUMPULAN GERMPLASMA *Elaeis*  
*guineensis* Jacq.**

Oleh

**MOHD SHAHRUL NIZWANSHAH BIN KARIM**

**Oktober 2020**

**Pengerusi: Professor Datin Siti Nor Akmar Abdullah, PhD**  
**Fakulti : Institut Pertanian Tropika & Sekuriti Makanan**

Vitamin E terdiri daripada tokoferol dan tokotrienol yang merupakan sebatian terlarut lipid yang dihasilkan oleh tumbuh-tumbuhan dengan ianya mempunyai pelbagai ciri-ciri perubahan yang bermanfaat. Tokotrienol mempunyai kebolehan menurunkan kolesterol dan bersifat anti-kanser manakala alfa-tokotrienol mampu mencegah degenerasi-saraf. Germplasma *E. guineensis* seperti Angola dan Tanzania mempunyai variasi kandungan vitamin E yang tinggi dan pelbagai iaitu bermula dari 300 sehingga 1600 ppm. Manakala dapat dilihat pula pada kadar 500 sehingga 1000 ppm dalam germplasma *E. guineensis* komersil. Walau bagaimanapun, pembangunan penanda kelapa sawit yang kaya kandungan tokotrienol masih terhad dan pemahaman mengenai pengaturan biosintesis vitamin E adalah penting demi penambahbaikan genetik khususnya kelapa sawit yang kaya vitamin E. Oleh itu, kajian ini dijalankan dengan tujuan untuk mengenal pasti SNP yang terdapat pada jujukan promoter HGGT yang mempunyai hubung kait dengan kandungan vitamin E yang tinggi dalam bahan germplasma *E. guineensis*. *Homogentisate geranylgeranyl transferase (HGGT)* adalah enzim biosintetik vitamin E yang penting dalam memangkinkan langkah pertama dalam proses biosintesis tokotrienol. Justeru itu, kajian ini bertujuan untuk mengenal pasti SNP dalam promoter HGGT yang menjadi faktor penyumbang kadar kandungan vitamin E yang tinggi. Urutan penjarangan daripada 14 aksesori Angola dan Tanzania dengan kadar kandungan vitamin E yang berbeza-beza menunjukkan terdapat kehadiran SNP pada kedudukan (-454) yang berpotensi menyumbang kepada kandungan vitamin E yang tinggi pada populasi germplasma kelapa sawit Angolan (AH) dan Tanzanian (TH) dan dua SNP lagi iaitu pada kedudukan (-781) dan (-113) adalah unik dan hanya terdapat pada germplasma kelapa sawit AH. Analisa pencirian fungsi untuk mengenal pasti peranan SNP tersebut dalam mempengaruhi aktiviti promoter HGGT dilakukan dengan menggunakan assai gen pelapor yang dibombardir ke dalam tisu mesokarp yang terdiri daripada empat binaan promoter HGGT yang berbeza. Vektor pBGWFS7 telah digunakan untuk pengklonan promoter HGGT dimana ia mengandungi dua gen pelapor, iaitu protein berpendarfluor

hijau (GFP) dan GUS. Para promoter itu adalah DXP (COM), AH1, TH96 dan mutan terbitan COM (CM, g.-454A>G) yang dihasilkan dengan memasukkan varian yang biasa ditemukan dalam AH1 dan TH96. Analisa kuantitatif fluorometrik GUS pada tisu positif GFP menunjukkan bahawa tahap ekspresi terendah adalah pada COM dengan ekspresi GUS sebanyak 0.27 pmol MU  $\text{minit}^{-1}$  mg protein $^{-1}$  per nombor salinan. Manakala, Tahap ekspresi yang diperoleh untuk COM adalah hampir sama dengan promoter konstitutif CaMV 35S yang digunakan sebagai perbandingan. Dapat dilihat, tidak terdapat perbezaan yang signifikan dalam tahap ekspresi GUS antara COM dengan versi bermutasi, CM (g.-454A>G). Tahap ekspresi tertinggi adalah dari promoter AH1 dan diikuti oleh TH96. Keputusan menunjukkan bahawa AH1 memberikan aktiviti GUS tertinggi dengan kadar ekspresi GUS sebanyak 0.82 pmol MU  $\text{minit}^{-1}$  mg protein $^{-1}$  per nombor salinan. Justeru itu, kami mencadangkan bahawa kotak CAAT yang terdapat pada AH di SNP (-113) adalah unik kepada AH sahaja di mana ia merupakan unsur promoter proksimal yang telah dikenal pasti mampu meningkatkan aktiviti promoter HGGT dan kandungan tocotrienol dalam AH Secara keseluruhan, kajian ini telah memberikan maklumat yang bernilai mengenai mekanisme pengawalseliaan biosintetik vitamin E dalam germplasma kelapa sawit dan hasil kajian telah menunjukkan bahawa promoter HGGT berpotensi untuk digunakan dalam kejuruteraan penanda bagi kandungan vitamin E yang tinggi melalui hubung kait SNP dimana ia juga penting dalam pembangunan genetik pada masa hadapan dalam usaha menghasilkan kelapa sawit yang mempunyai kandungan vitamin E yang tinggi.



## ACKNOWLEDGEMENT

First and foremost, Alhamdulillah praises Allah for His showers of blessings that I have completed my master study successfully with such a long journey filled with challenges and sweet memories along the way to finish it. I would like to express my deep and sincere gratitude to my research supervisor Prof. Datin Dr. Siti Nor Akmar from the Institute of Plantation Studies, Universiti Putra Malaysia for giving me the opportunity to do research and providing invaluable guidance throughout this research. Her continued assistance and guidance as my project supervisor throughout my studies benefit me a lot in a way to understand my work clearly and deeply. It was a great privilege and honor to work and study under her supervision. I would also like to thank for her generosity and understanding. Not forgotten, my appreciation to my co-supervisor, Assoc. Prof. Dr. Noor Azmi Shahrudin for his support and knowledge guidance in this project.

I am extending my heartfelt acknowledgement and appreciation to my beloved wife, Tuan Syaripah Najihah Tuan Mohd Razali, who has always stood right by me throughout all the time, who endlessly supported and motivated me whenever I felt down or when I felt to give up my project. She is the love of my life and my strength pill. She has made countless sacrifices to help me to get to this point, without her my master studies seem far and impossible to accomplish. I can't thank you enough for encouraging me throughout this experience my love.

To my lab mates, Luqman, Redzyque, Nazri, Engku, Hazwan, Mehdi, Sulaiman, Isiaka, Syafiqah, Sze Ling and Hanan, I thanked them for particularly spent a great deal of time helping me through my lab works and makes the lab as our second home full of laughter and happiness for more than three years. Sincere thanks to my friends Imran and Alfian, for their help during the time when I first arrived at UPM.

Last but not least, my deepest gratitude goes to my family. To my beloved mother Norminah binti Atim for her constant love, prayers and support. Your prayer for me was what sustained me this far and without you, I doubt that I would be in this place today. Also, I express my thanks to my sisters, brothers, sister in law and brothers-in-law for their support and valuable prayers. I am also grateful and would also express my appreciation to my parents in law for their support, encouragement and understanding. This list of acknowledgements can only capture a small fraction of the people who supported me to complete my work directly and indirectly. I send my deep thanks to all.

This thesis was submitted to the senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

**Siti Nor Akmar binti Abdullah, PhD**

Professor  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Chairman)

**Noor Azmi bin Shaharuddin, PhD**

Associate Professor  
Faculty of Biotechnology and Biomolecular Sciences  
Universiti Putra Malaysia  
(Member)

---

**ZALILAH MOHD SHARIFF, PhD**  
Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 14 January 2021

## Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No: Mohd Shahrul Nizwanshah bin Karim, GS43748

## Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: \_\_\_\_\_

Name of  
Chairman of  
Supervisory  
Committee:

Siti Nor Akmar Abdullah, PhD

Signature: \_\_\_\_\_

Name of  
Member of  
Supervisory  
Committee:

Noor Azmi Shahrudin, PhD

## TABLE OF CONTENTS

	<b>Page</b>
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	iii
<b>ACKNOWLEDGEMENT</b>	v
<b>APPROVAL</b>	vi
<b>DECLARATION</b>	viii
<b>LIST OF TABLES</b>	xiii
<b>LIST OF FIGURES</b>	xiv
<b>LIST OF ABBREVIATION</b>	xv
<b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	<b>1</b>
<b>2 LITERATURE REVIEW</b>	<b>3</b>
2.1 The Malaysian Palm Oil Industry	3
2.2 Oil Palm	3
2.2.1 Introductory to oil palm	3
2.2.2 Oil Palm Morphology	4
2.2.3 Oil Palm fruit development	4
2.2.4 Oil palm breeding	4
2.2.5 Composition of the Palm oil	5
2.2.6 Palm oil and human health	5
2.3 Metabolic Engineering for Tocotrienols and Improvement	6
2.4 Vitamin E	6
2.5 Tocotrienols	7
2.5.1 Tocotrienol Biosynthesis	7
2.5.2 Metabolite Function of Tocotrienol	7
2.5.3 Homogentisate Geranylgeranyl Transferase ( <i>HGGT</i> ) Gene	8
2.6 Gene Promoter	8
2.6.1 Plant Gene Promoters	8
2.6.2 Core Promoter & Proximal Promoter	8
2.6.3 The Distal Promoter Region	9
2.6.4 Homogentisate Geranylgeranyl Transferase ( <i>HGGT</i> ) Promoter	9
2.7 Transient Expression Assay by Biolistic Method	10
2.8 Single Nucleotides Polymorphism (SNPs)	10
2.9 GUS fluorometric Assay	11
2.10 Mutagenesis by PCR driven overlap extension	11
<b>3 MATERIALS AND METHODS</b>	<b>13</b>
3.1 Plant Collection Samples	13
3.2 Extraction of Oil Palm Genomic DNA	14
3.2.1 Genomic DNA Extraction	14
3.2.2 Genomic DNA Integrity and Quantification	14

3.2.3 Analysis of HGGT promoter sequence	15
3.3 Isolation of the HGGT promoter from Commercial (DXP), Angola high Vitamin E, and Tanzania high vitamin E by Polymerase Chain Reaction	15
3.3.1 Gene Specific Primer design	15
3.3.2 KAPA HiFi Hotstart ReadyMix PCR reaction	15
3.3.3 Gel Extraction of the PCR Product and DNA Purification	16
3.4 Formation of Mutagenic HGGT promoter from Commercial (DXP) germplasm by PCR Overlaps Extension	16
3.4.1 Site-directed Mutagenesis Primers Design	16
3.4.2 (1st) Primary PCR Reaction Mutagenesis	17
3.4.3 Secondary (2nd) PCR Reaction Mutagenesis and Purification	17
3.4.4 Sequencing and Mutagenesis Screening	17
3.5 Construction of the Commercial DXP (COM), Commercial Mutated DXP (CM), Angola High vitamin E (AH1) and Tanzania High vitamin E (TH96) HGGT promoter construct	18
3.6 BP and LR Recombination Reaction via Gateway® Technology	18
3.6.1 Cloning and Transformation of <i>HGGT</i> gene promoter into Gateway Vector	18
3.6.2 Preparation of Competent Cells for Transformation	19
3.7 Gateway Cloning: BP Reaction	19
3.7.1 Ligation of PCR product into the Entry vector	19
3.7.2 Transforming Competent cells	19
3.7.3 Plasmid Purification of Entry Vector	20
3.8 Gateway Cloning: LR Reaction	20
3.8.1 Ligation of PCR product into the Expression vector	20
3.8.2 Transforming Competent cells	20
3.8.3 Plasmid Purification of Destination Vector	21
3.9 Sequence Analysis HGGT Promoter	21
3.10 Transient transformation of the Oil Palm Mesocarp Tissue	21
3.10.1 Preparation of Target Materials for Bombardment	21
3.10.2 Preparation of DNA-Coated Microcarriers	22
3.10.3 Particle Bombardment	22
3.10.4 Bombarded Tissue Selection	22
3.10.5 DNA Extraction of Bombarded Tissues	23
3.11 Absolute Quantification for Normalization	23
3.12 GUS Fluorometric Assay	24
3.13 Determination of Sequence Variations in <i>E. guineensis</i> High Vitamin E accessions.	24
3.14 Experimental Design and Data Analysis	25
<b>4 RESULTS AND DISCUSSION</b>	<b>26</b>
4.1 Sequence Characterization of the Oil Palm HGGT Promoter	26
4.2 Isolation of HGGT promoter from Commercial DXP (COM), Angola (AH1) and Tanzania (TH96) Germplasm	28
4.3 Site-directed Mutagenesis by PCR-driven overlap extension	32
4.4 Determination of the Amount of Recombinant Plasmids Bombarded into Oil Palm Tissue Slices	35

4.5 Analysis of Oil palm germplasm of HGGT Promoter in Mesocarp Tissues	36
4.6 SNP Germplasm Specific at the Angola HGGT promoter region	38
4.7 Discussion	42
<b>5 SUMMARY, CONCLUSION AND RECOMMENDATION FOR FUTURE STUDIES</b>	<b>45</b>
5.1. Summary and Conclusion	45
5.2. Recommendation for Future Research	46
<b>REFERENCES</b>	<b>47</b>
<b>APPENDICES</b>	<b>56</b>
<b>BIODATA OF STUDENT</b>	<b>77</b>
<b>PUBLICATIONS</b>	<b>78</b>



## LIST OF TABLES

Table		Page
3.1	List of plant materials used from different <i>E. guineensis</i> germplasm population and their respective vitamin E scoring level.	13
3.2	Gene specific primers used in PCR.	15
3.3	List of primers used in site-directed mutagenesis.	17
3.4	<i>E. guineensis</i> accessions HPLC data and their vitamin E composition (ppm) collected from MPOB.	25
4.1	Putative <i>cis</i> -regulatory elements enriched in HGGT promoter.	27
4.2	Genomic DNA of <i>E. guineensis</i> , accession, concentration (ng/ $\mu$ l) and purity (A260/A280).	28
4.3	SNPs variant from different <i>E. guineensis</i> accession.	33
4.4	The copy number for all promoter construct.	36
4.5	Nucleotide variants at the two SNPs found in the HGGT promoter region in the <i>E. guineensis</i> Angolan and Tanzanian accessions with variable vitamin E content.	39



## LIST OF FIGURES

Figure		Page
4.1	Agarose gel electrophoresis analysis of the genomic DNA extracted from different <i>E. guineensis</i> accession spear leaves.	29
4.2	Agarose gel electrophoresis analysis of PCR product of HGGT promoter amplified from different accession of <i>E. guineensis</i> germplasm materials.	30
4.3	Alignment of the isolated full length HGGT promoter sequence from commercial DXP (COM), Angola (AH1) and Tanzania (TH96).	32
4.4	Primers involved in nucleotide substitution of the COM HGGT Promoter.	33
4.5	A schematic diagram for site-directed mutagenesis of the HGGT promoter at position -454.	34
4.6	Agarose gel electrophoresis analysis of the PCR products for nucleotide substitution at position -454 in HGGT promoter by site-directed mutagenesis.	35
4.7	Transient expression of GUS driven by HGGT promoter from different oil palm germplasm in oil palm mesocarp tissues.	37
4.8	Multiple sequence alignment of HGGT promoter sequences of <i>E. guineensis</i> individuals from different germplasm.	38
4.9	SNP at the -113 position in the promoter sequences of the Angolan and Tanzanian accessions with high or low vitamin E content.	40
4.10	Phylogenetic relationship between the amplified promoter of HGGT genes from Commercial DXP, Angola and Tanzania germplasm of different accessions of <i>Elaeis guineensis</i> .	41

## LIST OF ABBREVIATION

AH	Angola High Vitamin E
AL	Angola Low Vitamin E
Bp	base pair
CaCl <sub>2</sub>	calcium chloride
CM	Commercial Mutated DXP
COM	Commercial DXP
Ct	threshold cycle
CTAB	hexadecyl trimethyl ammonium bromide
DNA	deoxyribonucleic acid
EDTA	ethylene diamine tetraacetate
<i>E. coli</i>	<i>Escherichia coli</i>
<i>E. guineensis</i>	<i>Elaeis guineensis</i>
GUS	β-glucuronidase
GGDP	geranylgeranyldiphosphate
HCL	hydrochloric acid
HGGT	homogentisate geranylgeranyl transferase
HPT	homogentisate phytyltransferase
IPTG	isopropyl β-D-I-thiogalactopyranoside
Kb	kilobase
LB	Luria-Bertani
M	molar
4-MU	4-methylumbelliferone
4-MUG	4-methylumbelliferyl-β-D-glucuronide
MgCl <sub>2</sub>	magnesium chloride

Min	minutes
NaCl	sodium chloride
Ng	nanogram
OD	optical density
PCR	polymerase chain reaction
RT-PCR	real time PCR
SDS	sodium dodecyl sulfate
SNP	Single nucleotide polymorphism
TAE	Tris-acetate-EDTA
TE	Tris-EDTA
TH	Tanzania High Vitamin E
TL	Tanzania Low Vitamin E
TSS	transcription start site
UTR	untranslated region
v/v	volume per volume
V	volts
w/v	weight per volume
X-gal	5-bromo-4-chloro-indoyl- $\beta$ -D-galactopyranoside
$\mu$ g	microgram
$\mu$ M	micromolar
$\mu$ l	microliter
g	relative centrifugal force
$^{\circ}$ C	degree celcius

## CHAPTER 1

### INTRODUCTION

Oil Palm (*Elaeis guineensis*) or locally known as “kelapa sawit” originated from Africa. It is planted in almost 43 countries today in the tropical belt ranging from South East Asia, Africa and South America. Oil palm was introduced to Malaya Peninsular (now known as Malaysia) by the British Government in early 1875 as an ornamental plant for landscaping (Cramb & McCarthy, 2016), laying the foundation for the industry in Malaysia with the first commercial planting in Tennamaran Estate in Selangor in 1917 (Nambiappan et al., 2018). Today, Malaysia has now become one of the major oil palm producers in the world accounting for 28% of oil palm production (Malaysian Palm Oil Counsel [MPOC], 2020a). The industry has emerged as one of the most highly organized sectors in the Malaysian agricultural system and has contributed significantly in sustaining the Malaysian economy (Kushairi et al., 2017).

Red palm oil extracted from oil palm mesocarp contain a mixture of different antioxidants and phytonutrients such as Vitamin E (tocopherol/tocotrienols), pro-vitamin A (alpha and beta carotene), phytosterol complex and coenzymes. It is found, red oil palm contains high levels of vitamin E especially tocotrienols, with crude palm oil containing up to 800 mg/kg of total vitamin E (Zainal et al., 2019). The nanomolar concentration of  $\alpha$ -tocotrienol prevents inducible neurodegeneration (Sen et al., 2010; Khanna et al., 2003). Moreover, there is increasing interest in using tocotrienols as nutraceuticals in cancer treatment within the last few years due to their anticancer properties. Accumulating evidence on the anti-cancer potency of vitamin E showed that  $\gamma$ -tocotrienols and  $\delta$ -tocotrienols have the highest anti-cancer activities (Constantinou et al., 2019). Malaysian Palm Oil Board (MPOB) located in Kluang, Johor has assembled the largest *E. guineensis* germplasm materials in the world (Zaki et al., 2012). Among the *E. guineensis* germplasm materials, the Angolan and Tanzanian materials were observed to have high variability in the level of vitamin E content ranging from 300 – 1600 ppm, while the level is 500 – 1000 ppm in the commercial variety (Wahid et al., 2005; Rajanaidu et al., 2000). This variation may arise due to the molecular setup of the *Homogentisate geranylgeranyl transferase* gene which catalyses the first committed step of tocotrienols biosynthesis in plants.

Single nucleotide polymorphisms (SNP) are increasingly becoming the marker of choice in genetic analysis and agricultural breeding programs (Pootakham et al. 2015; Babura et al. 2017). They are widely used due to their abundance and possibilities for automation. SNPs in genes encoding proteins are of interest as they may be directly associated with the traits being studied. The SNPs can be in the coding or the noncoding regions such as the promoters and introns (Uppu et al., 2018). Genetic characterization of diverse rice germplasm was performed based on SNPs at the promoter region from a rice sucrose synthase 3 (RSUS3) gene involves in starch biosynthesis (Lee et al., 2009). SNPs have also been used to assess interspecific differences between *E. guineensis* and

*E. oleifera*, the two most important oil palm species (Riju & Arunachalam 2009). Crop intervention through genetic modification and engineering of the oil palm vitamin E biosynthesis will provide a crucial understanding of how the expression of a key enzyme involved in the oil palm vitamin E biosynthesis is regulated. Moreover, further understanding of the association of SNP in the promoter of the *HGGT* gene with high vitamin E content is important. The identification of the sources of vitamin E content variation among the oil palm germplasm accessions would become a valuable study and provide useful strategies for oil palm genetic improvement through marker-assisted selection.

Therefore, it would be interesting to study functional nucleotide variants in the vitamin E *HGGT* biosynthetic gene as the regulatory sequence or the promoter for controlling expressions of the downstream gene will be the key factor in the success of oil palm genetic engineering programme (Sambanthamurthi et al., 2002).

The objectives of this study are:

1. To identify variant nucleotide in the promoter sequence of *homogentisate geranylgeranyl transferase (HGGT)* gene from oil palm germplasm materials associating with the high vitamin E trait.
2. To produce promoter reporter constructs designed to determine the effects of identified sequence variant on HGGT promoter activity.
3. To identify the effect of Single Nucleotide Polymorphism (SNPs) on the activity of the HGGT promoter through transient reporter gene assay of bombarded mesocarp tissues slices.

It was hypothesised that the variation in vitamin E content found among *E. guineensis* germplasm materials could be associated with functionally variant nucleotides (SNPs) in the promoter sequence of *homogentisate geranylgeranyl transferase (HGGT)* gene which catalyses the first committed step of tocotrienols biosynthesis in plants.

## REFERENCES

- Abdullah, N., Yusop, M. R., Ithnin, M., Saleh, G., & Latif, M. A. (2011). Genetic variability of oil palm parental genotypes and performance of its' progenies as revealed by molecular markers and quantitative traits. *Comptes Rendus Biologies*, 334(4), 290-299.
- Abdullah, S. N. A., Cheah, S. C., & Murphy, D. J. (2002). Isolation and characterisation of two divergent type 3 metallothioneins from oil palm, *Elaeis guineensis*. *Plant Physiology and Biochemistry*, 40(3), 255-263.
- Ab Gapor, M. T., Ong, A. S. H., Kato, A., Watanabe, H., & Kawada, T. (1989). Antioxidant activities of palm vitamin E with special reference to tocotrienols. *Elaeis*, 1(1), 63-67.
- Agarwal, P., Kumar, R., Pareek, A., & Sharma, A. K. (2017). Fruit preferential activity of the tomato RIP1 gene promoter in transgenic tomato and Arabidopsis. *Molecular Genetics and Genomics*, 292(1), 145-156.
- Aggarwal, B. B., Sundaram, C., Prasad, S., & Kannappan, R. (2010). Tocotrienols, the vitamin E of the 21st century: Its potential against cancer and other chronic diseases. *Biochemical Pharmacology*, 80(11), 1613-1631.
- Agius, F., Amaya, I., Botella, M. A., & Valpuesta, V. (2004). Functional analysis of heterologous and homologous promoters in strawberry fruits using transient expression. *Journal of Experimental Botany*, 56(409), 37-46.
- Alexander, E., Lipka, M.A., Gore, M.M., Alex, M., Haining, L., Tyler, T., Charles, C., C. Robin B., Edward S. B., Torbert, R., and Dean DellaPenna, (2013). Genome-Wide Association Study and Pathway-Level Analysis of Tocochromanol Levels in Maize Grain. G3: GENES, *Genomes, Genetics* 3: 8 1287-1299.
- Babura, S. R., Abdullah, S. N. A., & Khaaza, H. (2017). Advances in genetic improvement for tocotrienol production: A Review. *Journal of Nutritional Science and Vitaminology*, 63, 215-221.
- Babura, S. R. (2018). *Identification of sequence variants in key vitamin E genes from Elaeis guineensis Jacq. germplasm for development of DNA-based markers* (Unpublished doctoral thesis), Universiti Putra Malaysia, Malaysia.
- Barrett, L. W., Fletcher, S., & Wilton, S. D. (2012). Regulation of eukaryotic gene expression by the untranslated gene regions and other non-coding elements. *Cellular and Molecular Life Sciences*, 69(21), 3613-3634.
- Batley, J., Barker, G., O'Sullivan, H., Edwards, K. J., & Edwards, D. (2003). Mining for single nucleotide polymorphisms and insertions/deletions in maize expressed sequence tag data. *Plant physiology*, 132(1), 84-91.
- Biłas, R., Szafran, K., Hnatuszko-Konka, K., & Kononowicz, A. K. (2016). Cis-regulatory elements used to control gene expression in plants. *Plant Cell, Tissue*

and *Organ Culture*, 127(2), 269–287.

- Brookes, A. J. (1999). The essence of SNPs. *Gene*, 234(2), 177-186.
- Cahoon, E. B., Hall, S. E., Ripp, K. G., Ganzke, T. S., Hitz, W. D., & Coughlan, S. J. (2003). Metabolic redesign of vitamin E biosynthesis in plants for tocotrienol production and increased antioxidant content. *Nature Biotechnology*, 21(9), 1082–1087.
- Cardenas, E., & Ghosh, R. (2013). Vitamin E: A dark horse at the crossroad of cancer management. *Biochemical Pharmacology*, 86(7), 845–852.
- Chen, S., Li, H., & Liu, G. (2006). Progress of vitamin E metabolic engineering in plants. *Transgenic Research*, 15(6), 655–665.
- Clegg, S., & de Matos, J.A. (2017). Palm Oil and Development in Malaysia and Indonesia. In *Sustainability and Organizational Change Management* (pp. 78–79). Routledge.
- Clemente, T. E., & Cahoon, E. B. (2009). Soybean Oil: Genetic Approaches for Modification of Functionality and Total Content. *Plant Physiology*, 151(3), 1030–1040.
- Collakova, E., & Dellapenna, D. (2003). Homogentisate Phytyltransferase Activity Is Limiting for Tocopherol Biosynthesis in Arabidopsis. *Plant Physiology*, 131(2), 632-642.
- Constantinou, C., Charalambous, C., & Kanakis, D. (2020). Vitamin E and cancer: an update on the emerging role of  $\gamma$  and  $\delta$  tocotrienols. *European journal of nutrition*, 59(3), 845-857.
- Corley, R. H. V., & Tinker, P. B. H. (2015). The Classification and Morphology of the Oil Palm. *The Oil Palm*. Chichester: John Wiley & Sons.
- Cramb, R., & McCarthy, J. F. (2016). Characterising oil palm production in Indonesia and Malaysia. *The Oil Palm Complex: Smallholders, Agribusiness and the State in Indonesia and Malaysia*, 27.
- Cummins, J. E. (1994). The use of Cauliflower Mosaic Virus (CaMV) in genetic engineering. *Plant Sciences*.
- Doyle, J. J., & Doyle, J. L. (1990). Isolation of plant DNA from fresh tissue. *Focus*, 12, 13-15.
- Dudley, J. W., & Moll, R. H. (1969). Interpretation and Use of Estimates of Heritability and Genetic Variances in Plant Breeding 1. *Crop science*, 9(3), 257-262.
- Ebong, P. E., Owu, D. U., & Isong, E. U. (1999). Influence of palm oil (*Elaeis guineensis*) on health. *Plant Foods for Human Nutrition*, 53(3), 209–222.
- Ebongue, G. F. N., Dhouib, R., Carrière, F., Zollo, P. A., & Arondel, V. (2006). Assaying

- lipase activity from oil palm fruit ( *Elaeis guineensis* Jacq .) mesocarp. *Plant Physiology and Biochemistry*, 44(10), 611-617.
- Edem, D.O. (2002). Palm oil: Biochemical,physiological,nutritional, hematological and toxicological aspects: A review. *Plant Foods for Human Nutrition*, 57(3-4), 319-341.
- Erb, I., & Van Nimwegen, E. (2011). Transcription factor binding site positioning in yeast: Proximal promoter motifs characterize Tata-Less promoters. *PLoS ONE*, 6(9).
- Fior, S., & Gerola, P. D. (2009). Impact of ubiquitous inhibitors on the GUS gene reporter system: evidence from the model plants Arabidopsis, tobacco and rice and correction methods for quantitative assays of transgenic and endogenous GUS. *Plant Methods*, 5, 19.
- Gupta, S. K. (Ed.). (2015). *Breeding Oilseed Crops for Sustainable Production: Opportunities and Constraints*. Academic Press.
- Gupta, P. K., Roy, J. K., & Prasad, M. (2001). Single nucleotide polymorphisms: a new paradigm for molecular marker technology and DNA polymorphism detection with emphasis on their use in plants. *Curr Sci*, 80(4), 524-535.
- Guthrie, N., Gapor, A., Chambers, A. F., & Carroll, K. K. (1997). Inhibition of proliferation of estrogen receptor-negative MDA-MB-435 and -positive MCF-7 human breast cancer cells by palm oil tocotrienols and tamoxifen, alone and in combination. *The Journal of Nutrition*, 127(3), 544S-548S.
- Hanifah, F. H. A., Abdullah, S. N. A., Othman, A., Shaharuddin, N. A., Saud, H. M., Hasnulhadi, H. A. H., & Munusamy, U. (2018). GCTTCA as a novel motif for regulating mesocarp-specific expression of the oil palm (*Elaeis guineensis* Jacq.) stearyl-ACP desaturase gene. *Plant cell reports*, 37(8), 1127-1143.
- Hartley, J., Temple, G., & Brasch, M. (2000). DNA cloning using in vitro site-specific recombination. *Genome Research*, 10(11), 1788-95.
- Heckman, K. L., & Pease, L. R. (2007). Gene splicing and mutagenesis by PCR-driven overlap extension. *Nature Protocols*, 2(4), 924-932.
- Hernandez-Garcia, C. M., & Finer, J. J. (2014). Identification and validation of promoters and cis-acting regulatory elements. *Plant Science*, 217, 109-119.
- Ho, S. N., Hunt, H. D., Horton, R. M., Pullen, J. K., & Pease, L. R. (1989). Site-directed mutagenesis by overlap extension using the polymerase chain reaction. *Gene*, 77(1), 51-59.
- Huminięcki, Ł., & Horbańczuk, J. (2017). Can We Predict Gene Expression by Understanding Proximal Promoter Architecture? *Trends in Biotechnology*, 35(6), 530-546.
- Hunter, S. C., & Cahoon, E. B. (2007). Enhancing vitamin E in oilseeds: Unraveling



tocopherol and tocotrienol biosynthesis. *Lipids*, 42(2), 97–108.

- Jalani, B. S., Cheah, S. C., Rajanaidu, N., & Darus, A. (1997). Improvement of palm oil through breeding and biotechnology. *Journal of the American Oil Chemists' Society*, 74(11), 1451-1455.
- Janssen, B., & Gardner, R. C. (1989). cocultivation with *Agrobacterium*. *Young*, 61–72.
- Jefferson, R. A., Bevan, M., & Kavanagh, T. (1987a). The use of the *Escherichia coli*  $\beta$ -glucuronidase gene as a gene fusion marker for studies of gene expression in higher plants. *Biochemical Society Transactions*, 15(1), 17–18.
- Jefferson, R. A., Kavanagh, T. A., & Bevan, M. W. (1987b). GUS fusions: beta-glucuronidase as a sensitive and versatile gene fusion marker in higher plants. *The EMBO Journal*, 6(13), 3901–3907.
- Jeong, M. J., & Shih, M. C. (2003). Interaction of a GATA factor with cis-acting elements involved in light regulation of nuclear genes encoding chloroplast glyceraldehyde-3-phosphate dehydrogenase in *Arabidopsis*. *Biochemical and Biophysical Research Communications*, 300(2), 555–562.
- Jiang, Q. (2014). Natural forms of vitamin E: Metabolism, antioxidant, and anti-inflammatory activities and their role in disease prevention and therapy. *Free Radical Biology and Medicine*, 72, 76–90.
- Joseph, S., & David, W. R. (2001). Molecular cloning: a laboratory manual. *The Quarterly Review of Biology*, 76(3), 348-349.
- Khanna, S., Roy, S., Ryu, H., Bahadduri, P., Swaan, P. W., Ratan, R. R., & Sent, C. K. (2003). Molecular Basis of Vitamin E Action: Tocotrienol modulates 12-lipoxygenase, a key mediator of glutamate-induced neurodegeneration. *Journal of Biological Chemistry*, 278(44), 43508–43515.
- Klein, T. M., Wolf, E. D., Wu, R., & Sanford, J. C. (1987). High-velocity microprojectiles for delivering nucleic acids into living cells. *Nature*, 327(6117), 70.
- Kong, S. L., Abdullah, S. N. A., Ho, C. L., & Amiruddin, M. D. (2016). Molecular cloning, gene expression profiling and in silico sequence analysis of Vitamin E biosynthetic genes from the oil palm. *Plant Gene*, 5, 100–108.
- Kushairi, A., Singh, R., & Ong-Abdullah, M. (2017). The oil palm industry in Malaysia: Thriving with transformative technologies. *Journal of Oil Palm Research*, 29(4), 431–439.
- Lee, T. I., & Young, R. A. (2000). Transcription of eukaryotic protein-coding genes. *Annual review of genetics*, 34(1), 77-137.
- Lee, D., Ezhkova, E., Li, B., Pattenden, S. G., Tansey, W. P., & Workman, J. L. (2005). The proteasome regulatory particle alters the SAGA coactivator to enhance its interactions with transcriptional activators. *Cell*, 123(3), 423-436.

- Lee, G. A., Koh, H. J., Chung, H. K., Dixit, A., Chung, J. W., Ma, K. H., ... & Kim, T. S. (2009). Development of SNP-based CAPS and dCAPS markers in eight different genes involved in starch biosynthesis in rice. *Molecular breeding*, 24(1), 93-101.
- Levine, M., Tjian, R., Howard, T., Medical, H., & Hall, B. (2003). Transcription regulation and animal diversity. *Nature*, 424, 147-51.
- Luo, T., Xia, W., Gong, S., Mason, A. S., Li, Z., Liu, R., Dou, Y., Tang, W., Fan, H., Zhang, C., & Xiao, Y. (2020). Identifying Vitamin E Biosynthesis Genes in *Elaeis guineensis* by Genome-Wide Association Study. *J. Agric. Food Chem.* 68, 678-685.
- Lohi, J., Lehti, K., Valtanen, H., Parks, W. C., & Keski-Oja, J. (2000). Structural analysis and promoter characterization of the human membrane-type matrix metalloproteinase-1 (MT1-MMP) gene. *Gene*, 242(1-2), 75-86.
- Lorito, M., Hayes, C. K., Di Pietro, A., & Harman, G. E. (1993). Biolistic transformation of *Trichoderma harzianum* and *Gliocladium virens* using plasmid and genomic DNA. *Current genetics*, 24(4), 349-356.
- McIntyre, B. S., Briski, K. P., Gapor, A., & Sylvester, P. W. (2000). Antiproliferative and apoptotic effects of tocopherols and tocotrienols on preneoplastic and neoplastic mouse mammary epithelial cells (44544). *Proceedings of the Society for Experimental Biology and Medicine*, 224(4), 292-301.
- MPOB (2020). Monthly Production Of Oil Palm Products Summary For The Month Of December 2019. Retrieved 20 October 2020 from <http://bepi.mpob.gov.my/index.php/en/production/production-2019/production-of-oil-palm-products-2019.html>
- MPOC (2020a). Malaysian Palm Oil Industry. Retrieved 20 October 2020 from <http://mpoc.org.my/malaysian-palm-oil-industry/>
- MPOC (2020b). The Oil Palm Tree. Retrieved 20 October 2020 from <http://mpoc.org.my/the-oil-palm-tree/>
- Mukherjee, S., & Mitra, A. (2009). Health Effects of Palm Oil. *Journal of Human Ecology*, 26(3), 197-203.
- Munusamy, U., Abdullah, S. N. A., Aziz, M. A., & Khazaai, H. (2015). Metabolic Engineering of A-Tocotrienol through Ptg Mechanisms and Isoprenoid/Non-Mevalonate Pathways in Perennial Crops. *Plant Cell Biotechnology and Molecular Biology*, 119-129.
- Murashige, T., & Skoog, F. (1962). A revised method for rapid growth and bioassays with tobacco tissue cultures. *Physiol Plant*, 15,473-497.
- Nambiappan, B., Ismail, A., Hashim, N., Ismail, N., Shahari, D. N., Idris, N. A. N., ... & Kushairi, A. (2018). Malaysia: 100 years of resilient palm oil economic performance. *Journal of Oil Palm Research*, 3 (1), 13-25.

- Nasu, S., Suzuki, J., Ohta, R., Hasegawa, K., Yui, R., Kitazawa, N., ... & Minobe, Y. (2002). Search for and analysis of single nucleotide polymorphisms (SNPs) in rice (*Oryza sativa*, *Oryza rufipogon*) and establishment of SNP markers. *DNA research*, 9(5), 163-171.
- Neuzil, P. (2006). *Transgenic Plant Method & Protocols*. *Nucleic Acids Research* (Vol. 34).
- Neves-Borges, A. C., Guimarães-Dias, F., Cruz, F., Mesquita, R. O., Nepomuceno, A. L., Romano, E., ... & Alves-Ferreira, M. (2012). Expression pattern of drought stress marker genes in soybean roots under two water deficit systems. *Genetics and molecular biology*, 35(1), 212-221.
- Omidvar, V., Abdullah, S. N. A., Izadfar, A., Ho, C. L., & Mahmood, M. (2010). The oil palm metallothionein promoter contains a novel AGTTAGG motif conferring its fruit-specific expression and is inducible by abiotic factors. *Planta*, 232(4), 925-936.
- Oo, K. C., Teh, S. K., Khor, H. T., & Ong, A. S. H. (1985). Fatty acid synthesis in the oil palm (*Elaeis guineensis*): Incorporation of acetate by tissue slices of the developing fruit. *Lipids*, 20(4), 205-210.
- Packer, L. (1994). Vitamin E is nature's master antioxidant. *Science & Medicine*, 1(1), 54.
- Piechulla, B., Merforth, N., & Rudolph, B. (1998). Identification of tomato Lhc promoter regions necessary for circadian expression. *Plant molecular biology*, 38(4), 655-662.
- Pootakham, W., Jomchai, N., Ruang-areerate, P., Shearman, J. R., Sonthirod, C., Sangsrakru, D., ... & Tangphatsornruang, S. (2015). Genome-wide SNP discovery and identification of QTL associated with agronomic traits in oil palm using genotyping-by-sequencing (GBS). *Genomics*, 105(5-6), 288-295.
- Porto, M. S., Pinheiro, M. P. N., Batista, V. G. L., Dos Santos, R. C., De Albuquerque Melo Filho, P., & De Lima, L. M. (2014). Plant promoters: An approach of structure and function. *Molecular Biotechnology*, 56(1), 38-49.
- Rafalski, A. (2002). Applications of single nucleotide polymorphisms in crop genetics. *Current opinion in plant biology*, 5(2), 94-100.
- Ramli, Z., & Abdullah, S. N. A. (2003). Development of a transient promoter assay system for oil palm. *Journal of Oil Palm Research*, 15(2), 62-69.
- Ramli, Z., & Abdullah, S. N. A. (2010). Functional characterisation of the oil palm type 3 metallothionein-like gene (MT3-B) promoter. *Plant molecular biology reporter*, 28(3), 531-541.
- Rajanaidu, N., Kushairi, A., Rafii, M., Mohd, D. A., Maizura, I., & Jalani, B. S. (2000). Oil palm breeding and genetic resources. *Advances in oil palm research*, 1, 171-237.

- Rees, A. R. (1965). Some factors affecting the viability of oil palm seed in storage. *J Nigerian Inst Oil Palm Res*, 4, 317-324.
- Riju, A., & Arunachalam, V. (2009). Interspecific differences in single nucleotide polymorphisms (SNPs) and indels in expressed sequence tag libraries of oil palm *Elaeis guineensis* and *E. oleifera*. *Nature Precedings*, 1-1.
- Risch, N., & Merikangas, K. (1996). The future of genetic studies of complex human diseases. *Science*, 273(5281), 1516-1517.
- Roy, A. L., & Singer, D. S. (2015). Core promoters in transcription: Old problem, new insights. *Trends in Biochemical Sciences*, 40(3), 165–171.
- Sailo, B. L., Banik, K., Padmavathi, G., Javadi, M., Bordoloi, D., & Kunnumakkara, A. B. (2018). Tocotrienols: The promising analogues of vitamin E for cancer therapeutics. *Pharmacological Research*, 130, 259–272.
- Sambanthamurthi, R., Siti, N. A., & Parveez, G. A. (2002). Genetic manipulation of the oil palm-challenges and prospects. *Planter*, 78(919), 547-562.
- Sambanthamurthi, R., Sundram, K., & Tan, Y. A. (2000). Chemistry and biochemistry of palm oil. *Progress in lipid research*, 39(6), 507-558.
- Santi, C., Svistoonoff, S., Constans, L., Auguy, F., Duhoux, E., Bogusz, D., & Franche, C. (2003). Choosing a reporter for gene expression studies in transgenic actinorhizal plants of the Casuarinaceae family. *Plant and Soil*, 254(1), 229–237.
- Sattler, S. E., Gilliland, L. U., Magallanes-Lundback, M., Pollard, M., & DellaPenna, D. (2004). Vitamin E is essential for seed longevity and for preventing lipid peroxidation during germination. *The plant cell*, 16(6), 1419-1432.
- Sen, C. K., Khanna, S., Rink, C., & Roy, S. (2007). Tocotrienols: The Emerging Face of Natural Vitamin E. *Vitamins and Hormones*, 76(07), 203–261.
- Sen, C. K., Khanna, S., & Roy, S. (2006). Tocotrienols: Vitamin E beyond tocopherols. *Life Sciences*, 78(18), 2088–2098.
- Sen, C. K., Rink, C., & Khanna, S. (2010). Palm oil-derived natural vitamin E  $\alpha$ -tocotrienol in brain health and disease. *Journal of the American College of Nutrition*, 29(sup3), 314S-323S.
- Šmídková, M., Hola, M., & Angelis, K. J. (2010). Efficient biolistic transformation of the moss *Physcomitrella patens*. *Biologia plantarum*, 54(4), 777-780.
- Sundram, K., Sambanthamurthi, R., & Tan, Y. (2003). Palm fruit chemistry and nutrition Palm fruit chemistry and nutrition. *Asia Pacific Journal Clinic Nutrition*, 12(3), 355–362.
- Terada, R., & Shimamoto, K. (1990). Expression of CaMV35S-GUS gene in transgenic rice plants. *MGG Molecular & General Genetics*, 220(3), 389–392.

- Theriault, A., Chao, J.-T., Wang, Q., Gapor, A., & Adeli, K. (1999). Tocotrienol: a review of its therapeutic potential. *Clinical Biochemistry*, 32(5), 309–319.
- Tranbarger, T. J., Kluabmongkol, W., Sangsakru, D., Morcillo, F., Tregear, W. J., Tragoonrung, S., & Billotte, N. (2012). SSR markers in transcripts of genes linked to post-transcriptional and transcriptional regulatory functions during vegetative and reproductive development of *Elaeis guineensis*. *BMC plant biology*, 12(1), 1.
- Uppu, S., Krishna, A., & Gopalan, R. P. (2016). A review on methods for detecting SNP interactions in high-dimensional genomic data. *IEEE/ACM transactions on computational biology and bioinformatics*, 15(2), 599-612.
- Urban, A., Neukirchen, S., & Jaeger, K. E. (1997). A rapid and efficient method for site-directed mutagenesis using one-step overlap extension PCR. *Nucleic Acids Research*, 25(11), 2227–2228.
- Venkatesh, T. V., Karunanandaa, B., Free, D. L., Rottnek, J. M., Baszis, S. R., & Valentin, H. E. (2006). Identification and characterization of an Arabidopsis homogentisate phytyltransferase paralog. *Planta*, 223(6), 1134–1144.
- Vijay, V., Pimm, S. L., Jenkins, C. N., & Smith, S. J. (2016). The impacts of oil palm on recent deforestation and biodiversity loss. *PloS one*, 11(7), e0159668.
- Wahid, M. B., Abdullah, S. N. A., & IE, H. (2005). Oil Palm. *Plant Production Science*, 8(3), 288-297.
- Xu, M., Gonzalez-Hurtado, E., & Martinez, E. (2016). Core promoter-specific gene regulation: TATA box selectivity and Initiator-dependent bi-directionality of serum response factor-activated transcription. *Biochimica et Biophysica Acta - Gene Regulatory Mechanisms*, 1859(4), 553–563.
- Yang, Z., Lee, M. J., Zhao, Y., & Yang, C. S. (2012). Metabolism of tocotrienols in animals and synergistic inhibitory actions of tocotrienols with atorvastatin in cancer cells. *Genes and Nutrition*, 7(1), 11–18.
- Yu, Y., Lee, C., Kim, J., & Hwang, S. (2005). Group-specific primer and probe sets to detect methanogenic communities using quantitative real-time polymerase chain reaction. *Biotechnology and Bioengineering*, 89(6), 670-9.
- Zainal, Z., Abdul Rahim, A., Khaza'ai, H., & Chang, S. K. (2019). Effects of Palm Oil Tocotrienol-Rich Fraction (TRF) and Carotenes in Ovalbumin (OVA)-Challenged Asthmatic Brown Norway Rats. *International journal of molecular sciences*, 20(7), 1764.
- Zaki, N. M., Singh, R., Rosli, R., & Ismail, I. (2012). *Elaeis oleifera* genomic-SSR markers: exploitation in oil palm germplasm diversity and cross-amplification in Arecaceae. *International journal of molecular sciences*, 13(4), 4069-4088.
- Zhang, C., Cahoon, R. E., Hunter, S. C., Chen, M., Han, J., & Cahoon, E. B. (2013a). Genetic and biochemical basis for alternative routes of tocotrienol biosynthesis for enhanced vitamin e antioxidant production. *Plant Journal*, 73(4), 628–639.

Zhang, G. Y., Liu, R. R., Xu, G., Zhang, P., Li, Y., Tang, K. X., ...& Liu, Q. Q. (2013b). Increased  $\alpha$ -tocotrienol content in seeds of transgenic rice overexpressing Arabidopsis  $\gamma$ -tocopherol methyltransferase. *Transgenic Research*, 22(1), 89–99.



## BIODATA OF STUDENT

The student, Mohd Shahrul Nizwanshah Bin Karim was born in Sandakan, Sabah on June 4th, 1991. He received his primary education in Sekolah Kebangsaan Rancangan Suan Lamba from 1998 to 2003. He then continued his secondary school in SMK Elopura (Bestari) from 2004 to 2006 (PMR) and in SM Sains Sabah from 2007 to 2008 (SPM). In 2009, he enrolled at Labuan Matriculation College for his matriculation and completed his bachelor's degree in Agricultural Science at Universiti Putra Malaysia, Serdang in November 2014. After a year working as an Assistant Estate Manager with IOI Group, he pursued his studies in Master of Science at the Institute of Tropical Agriculture and Food Security, UPM in 2015 under MyBrain15 sponsorship program. In October 2019, he then decided to join the Malaysian Agricultural Research and Development Institute (MARDI) as a Research Officer to develop his passion for research in agriculture.

Experiences gained when preparing the thesis made him a better person, he learned patience, friendship, courage, and discipline. He is very thankful for the whole period of his study that he has been bestowed by supportive and encouraging persons from his family and friends. The success of his studies is above all attributed to his wife's love, prayers, and care.

## PUBLICATIONS

### Journal

Karim, M. S. N., Abdullah, S. N. A., Nadzir, M. M. M., Moradpour, M., Shahrudin, N. A., & Abdullah, M. O. (2020). Single nucleotide polymorphisms in oil palm HOMOGENTISATE GERANYL-GERANYL TRANSFERASE promoter for species differentiation and TOCOTRIENOL improvement. *Meta Gene*, 100818.

### International Conference

Karim, Mohd Shahrul Nizwanshah and Abdullah, Siti Nor Akmar (2017) Mutagenesis of the promoter of the oil palm homogentisate geranylgeranyl transferase gene (*HGGT*) by PCR-driven overlap extension. In: International Conference on Big Data Applications in Agriculture (ICBAA2017), 5-6 Dec. 2017, Auditorium Putra, TNCPI Building, Universiti Putra Malaysia. (pp. 191-195).

Karim, Mohd Shahrul Nizwanshah, Abdullah, Siti Nor Akmar and Shahrudin, Noor Azmi (2018) Quantification of the amount of recombinant plasmid bombarded into oil palm mesocarp tissue. In: International Conference on Comparative & Interactomics for Agriculture (ICCGIA2018), 1<sup>st</sup>-2<sup>nd</sup> Nov. 2018, Mini Auditorium 1, TNCPI Building, Universiti Putra Malaysia (pp. 40-54).





**UNIVERSITI PUTRA MALAYSIA**

**STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT**

**ACADEMIC SESSION : First Semester 2020/2021**

**TITLE OF THESIS / PROJECT REPORT :**

DETERMINATION OF FUNCTIONALLY IMPORTANT SEQUENCE VARIANTS IN  
PROMOTER SEQUENCE OF HOMOGENISATE GERANYLGERANYL TRANSFERASE  
FROM *Elaeis guineensis* Jacq. GERMPLASM MATERIALS

**NAME OF STUDENT: MOHD SHAHRUL NIZWANSHAH BIN KARIM**

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

1. This thesis/project report is the property of Universiti Putra Malaysia.
2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as :

\*Please tick (✓)

**CONFIDENTIAL**

(Contain confidential information under Official Secret Act 1972).

**RESTRICTED**

(Contains restricted information as specified by the organization/institution where research was done).

**OPEN ACCESS**

I agree that my thesis/project report to be published as hard copy or online open access.

This thesis is submitted for :

**PATENT**

Embargo from \_\_\_\_\_ until \_\_\_\_\_  
(date) (date)

**Approved by:**

\_\_\_\_\_  
(Signature of Student)  
New IC No/ Passport No.:

Date :

\_\_\_\_\_  
(Signature of Chairman of Supervisory Committee)  
Name:

Date :

**[Note : If the thesis is CONFIDENTIAL or RESTRICTED, please attach with the letter from the organization/institution with period and reasons for confidentiality or restricted. ]**