



**UNIVERSITI PUTRA MALAYSIA**

***IN VITRO REGENERATION AND EVALUATION OF PHYTOCHEMICAL  
AND ANTIMICROBIAL ACTIVITY OF KUNYIT PUTIH  
[Curcuma zedoaria (Christm.) Roscoe]***

**KHALID ABUBAKER ALGHANNAY**

**FP 2021 53**



***IN VITRO* REGENERATION AND EVALUATION OF PHYTOCHEMICAL  
AND ANTIMICROBIAL ACTIVITY OF *KUNYIT PUTIH*  
[*Curcuma zedoaria* (Christm.) Roscoe]**

By

**KHALID ABUBAKER ALGHANNAY**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra  
Malaysia, in Fulfilment of the Requirements for Degree of  
Doctor of Philosophy**

**March 2019**

## **COPYRIGHT**

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



## DEDICATION

To those who have spent their lives to be always happy, **My Parents**

To those who stood with me morally and physically, **My Brothers and Sisters**

To which endure my circumstances and supported my suffering, **My Wife**

To those who I aspire and look forward to get benefit from my career and be better than me, **My Children**



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

**IN VITRO REGENERATION AND EVALUATION OF PHYTOCHEMICAL AND ANTIMICROBIAL ACTIVITY OF KUNYIT PUTIH**  
**[*Curcuma zedoaria* (Christm.) Roscoe]**

By

**KHALID ABUBAKER ALGHANNAY**

**March 2019**

**Chairman : Associate Professor Halimi Moh bin Saud, PhD**  
**Faculty : Agriculture**

Today's healthcare system is hampered with numerous health problems such as degenerative disorder, chronic diseases, resistant infection etc. Plants are source of precursors of many natural products and secondary metabolites with pharmacological and therapeutic potentials but some major set-back in plant natural product medicines are non-availability of medicinal plant material and in-ability to extract the bioactive compounds with the appropriate solvent. Therefore, there is highly need to develop an in vitro micro propagation technique for rapid multiplication to produce high quality planting materials of *C. zedoaria*, which is suffering from persistent endophytic and epiphytic microbial contamination and from low response to media, and to evaluate the phytochemicals screening method to explore their antioxidant compounds and antimicrobial properties. Some procedures do exist but generally do not address well the initial stage of culture establishment and phytochemical screening as well. Hence, the objectives of this study is to establish an in vitro regeneration protocol for *C.zedoaria* using tissue culture techniques. Secondly to optimize ideal solvent and concentrations suitable for screening the phytochemicals of *C. zedoaria* leaves and rhizome for Total Phenolic Content (TPC), Total Flavonoids Content (TFC) and Antioxidant activity (AO). The most suitable solvents (isopropanol for TPC and AOC/ methanol for TFC), from last experiment results, were applied as the third objective, as comparative study, to examine the extent of the difference between (NFGP) and (TCPP). Eventually, the evaluation of the antimicrobial activity of the different solvents and oils extracted from the leaves and rhizomes of *C. zedoaria* against pathogenic bacteria was carried out. Surface sterilization were assessed on explants (rhizome with apical buds), with 3 different sterilizing agents (NaOCl, HgCl<sub>2</sub> and Nano Silver) at different concentrations and immersion times, in attempt to determining the most suitable method of reducing explant contamination. We detect the ability of 6-benzylaminopurine (BAP) alone for shoot induction, while (BAP),

kinetin, and thidiazuron (TDZ) were individually evaluated for shoot formation, however IBA and NAA was used separately to stimulate root formation. To screen for phytochemicals constituents, the leaf and rhizome of *C. zedoaria* were extracted with different polar solvents including ethanol, methanol, dimethyl sulfoxide, acetonitrile, acetone, isopropanol, and glycerol with different concentrations (0.0, 10, 30, 50, 70, 90, and 100%), We used Folin Ciocalteu's reagent, DPPH scavenging assay and colorimetric method using Aluminium Chloride, to determined TPC, AOX and TFC successively, while Disk Diffusion Test and also Minimum and Bactericidal Inhibitory Concentration (MIC and MBC) tests, was used to evaluate the antimicrobial activities. The results revealed that NaOCl was the most suitable sterilizing agent with 77.77% of sterilized success and survived explants compared to HgCl<sub>2</sub> and NS after decontamination stage. The shoot induction and formation results showed that 3 mg/L of BAP strongly stimulated the shoot formation of *C. zedoaria* by producing 4.3 shoots per explants when compared to kinetin and TDZ. In evaluating the effects of different auxins on root induction of *C. zedoaria* revealed that 2.0 mg/L of NAA produced the highest root number by the mean of 6.3 roots per explant in *C. zedoaria* compared to other auxin hormonal treatments. The shoot induction and formation results showed that 3 mg/L of BAP strongly stimulated the shoot formation of *C. zedoaria* by producing 4.3 shoots per explants when compared to kinetin and TDZ. In evaluating the effects of different auxins on root induction of *C. zedoaria* revealed that 2.0 mg/L of NAA produced the highest root number by the mean of 6.3 roots per explant in *C. zedoaria* compared to other auxin hormonal treatments. The result of the phytochemical assay revealed that the highest flavonoid content in leaf tissue (3.309 mg) was achieved using dimethyl sulfoxide solvent (70%), methanol at 90% (3.01 mg), followed by 90% ethanol with a value of 2.7 mg, while in the rhizome, methanol displayed the highest concentration at 100 and 90 % by 4.8 and 4.5 mg respectively. Our finding showed that the Isopropyl alcohol was promising option for evaluating (AOC) and (TPC) due to its extractability potential on both Leaves & Rhizome tissues, while the Methanol is most suitable solvent for (TFC) in both cases too, despite competition and convergence of solvent impact. However, there was no significant difference in leaf and rhizome content for phytochemicals measured (TFC, TPC and AOC) in both NFGP and TCPP, hence there were no real differences using this propagation method (tissue culture) in certain phytochemical content. The antimicrobial inhibitory effect of *C. zedoaria* against certain pathogenic gram positive and gram negative revealed that these positive effects were present at all levels of all extracts (solvents, concentrations and type of tissues) and same is true for the effect of oil samples. This effect is incremental by increasing concentration each time. The effect of rhizome extract is relatively higher than that of leaf extract at all concentrations whereas, the solvent type did not have any obvious effect on inhibition, although minor effect was observed with isopropanol. The most MIC & MBC results is within the range of medium-impact rates, which has positive connotations and distinguished implications. In conclusion, *C. zedoaria* can be effective to treat diseases caused by bacterial infections.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

**PENJANAAN *IN VITRO* DAN PENILAIAN FITOKIMIA DAN AKTIVITI  
ANTIMIKROBIAL BAGI KUNYIT PUTIH  
[*Curcuma zedoaria* (Christm.) Roscoe]**

Oleh

**KHALID ABUBAKER ALGHANNAY**

Mac 2019

**Pengerusi : Profesor Madya Halimi Moh bin Saud, PhD**  
**Fakulti : Pertanian**

Sistem penjagaan kesihatan hari ini terjejas dengan banyak masalah kesihatan seperti gangguan degeneratif, penyakit kronik, jangkitan tahan dan lain-lain. Tumbuh-tumbuhan adalah sumber utama produk-produk semulajadi dan metabolit sekunder dengan potensi farmakologi dan terapeutik namun halangan utama dalam ubat berasaskan produk tumbuhan semula jadi adalah ketidakmampuan untuk mengeluarkan sebatian bioaktif dengan pelarut yang sepatutnya. Justeru, ianya sangat diperlukan untuk membangunkan teknik penanaman mikro *in vitro* untuk penggandaan yang cepat untuk menghasilkan bahan penanaman *C. zedoaria* yang berkualiti tinggi, yang mana mengalami pencemaran mikrob endophytic dan epiphytic yang berterusan dan mengalami tindak balas yang rendah terhadap media, dan juga untuk menilai kaedah pemeriksaan fitokimia untuk meneroka sifat sebatian antioksidan dan antimikrob. Terdapat beberapa prosedur yang ada tetapi pada amnya tidak sesuai untuk tahap awal pembentukan kultur dan juga pemeriksaan fitokimia. Oleh itu, objektif kajian ini adalah untuk mewujudkan protokol regenerasi *in vitro* untuk *C. zedoaria* menggunakan teknik kultur tisu. Kedua adalah untuk mengoptimumkan pelarut dan kepekatan yang sesuai untuk pemeriksaan fitokimia daun *C. zedoaria* dan rizom untuk Total Kandungan Fenolik (TPC), Total Kandungan Flavonoid (TFC) dan aktiviti Antioksidan (AO). Pelarut yang paling sesuai (isopropanol untuk TPC dan AOC / methanol untuk TFC), dari hasil eksperimen lepas, digunakan sebagai objektif ketiga, sebagai kajian perbandingan, untuk melihat sejauh mana perbezaan antara (NFGP) dan (TCPP). Akhirnya, penilaian aktiviti antimikrob bagi pelarut dan minyak yang berbeza yang diekstrak dari daun dan rizom *C. zedoaria* terhadap bakteria patogen telah dijalankan. Pensterilan awal dinilai pada eksplan (rizom dengan tunas apikal), dengan 3 agen steril yang berbeza (NaOCl, HgCl<sub>2</sub> dan Nano Silver) pada kepekatan dan masa penyerapan yang berlainan, dalam usaha untuk menentukan kaedah yang paling sesuai untuk mengurangkan pencemaran pada eksplan. Kami

mengesan keupayaan 6-benzylaminopurine (BAP) sendirian untuk induksi pucuk, sementara (BAP), kinetin, dan thidizuron (TDZ) secara individu dinilai untuk pembentukan pucuk, namun IBA dan NAA digunakan secara berasingan untuk merangsang pembentukan akar. Untuk memaparkan unsur-unsur fitokimia, daun dan rizom *C. zedoaria* diekstraks dengan pelarut polariti yang berbeza termasuk etanol, metanol, dimetil sulfoksida, asetonitril, aseton, isopropanol, dan gliserol dengan kepekatan yang berbeza (0.0, 10, 30, 50, 70, 90, dan 100%), kami menggunakan reagen Folin Ciocalteu, kaedah DPPH pengarkian dan kaedah kolorimetrik menggunakan Aluminium Chloride, untuk menentukan TPC, AOX dan TFC secara berturut-turut, manakala ujian Penyebaran Cakera dan ujian Kepekatan Minimum dan Bakterisidal (MIC dan MBC) digunakan untuk menilai aktiviti antimikrob. Keputusan menunjukkan bahawa NaOCl adalah agen penyerap yang paling sesuai dengan 77.77% kejayaan steriliti dan eksplan yang terselamat berbanding dengan HgCl<sub>2</sub> dan NS selepas peringkat dekontaminasi. Keputusan induksi dan pembentukan pucuk menunjukkan bahawa 3 mg/L BAP sangat merangsang pembentukan tembakau *C. zedoaria* dengan menghasilkan 4.3 pucuk per eksplan apabila dibandingkan dengan kinetin dan TDZ. Dalam menilai kesan-kesan auksin yang berbeza pada induksi akar *C. zedoaria* mendedahkan bahawa 2.0 mg / L NAA menghasilkan bilangan akar tertinggi dengan purata 6.3 akar per eksplan *C. zedoaria* berbanding dengan rawatan hormon auxin yang lain. Hasil ujian fitokimia menunjukkan bahawa kandungan flavonoid tertinggi dalam tisu daun (3.309 mg) dicapai menggunakan pelarut dimetil sulfoksida (70%), metanol pada 90% (3.01 mg), diikuti oleh 90% etanol dengan nilai 2.7 mg, manakala dalam rizom, metanol menunjukkan kepekatan tertinggi pada 100 dan 90% masing-masing sebanyak 4.8 dan 4.5 mg. Penemuan kami menunjukkan bahawa alkohol Isopropil adalah pilihan yang bagus untuk menilai (AOC) dan (TPC) kerana potensi kebolehan ekstraksi pada kedua-dua Daun & Rhizome tisu, sementara Metanol adalah pelarut paling sesuai untuk (TFC) dalam kedua-dua kes, walaupun terdapat persaingan dan pertindihan kesan pelarut. Walau bagaimanapun, tidak terdapat perbezaan yang signifikan dalam kandungan daun dan rizom untuk ujian fitokimia yang direkod (TFC, TPC dan AOC) pada kedua-dua NFGP dan TCPP, oleh itu tidak terdapat perbezaan yang ketara menggunakan kaedah penanaman (kultur tisu) dalam kandungan fitokimia tertentu. Kesan penghalang antimikrob daripada *C. zedoaria* terhadap sebahagian patogen gram positif dan gram negatif mendedahkan bahawa kesan positif ini berlaku pada semua peringkat ekstrak (pelarut, kepekatan dan jenis tisu) dan sama juga berlaku untuk sampel minyak. Kesan ini semakin meningkat dengan setiap kali peningkatan dalam kepekatan. Kesan ekstrak rizom adalah lebih tinggi daripada ekstrak daun pada semua kepekatan sedangkan jenis pelarut tidak mempunyai sebarang kesan jelas pada perencatan, walaupun kesan kecil diperhatikan dengan isopropanol. Hasil MIC & MBC yang paling banyak adalah dalam lingkungan kadar kesan sederhana, yang mempunyai impak positif dan implikasi yang berbeza. Kesimpulannya, *C. zedoaria* berkesan untuk merawat penyakit yang disebabkan oleh jangkitan bakteria.



## ACKNOWLEDGEMENTS

First of all, I am thankful to Allah for giving me the strength, guidance, and perseverance to complete this thesis. May blessing and peace be upon Prophet Muhammad (S. A. W.) who was sent to the world as a mercy. Journey undoubtedly long, will not say it is arduous, perhaps this word does not give it its right and can not describe its scenes, but all this vanishes, with reaching the goal. A scientific achievement, requires a clear plan, deliberate stages and continuous assessment, otherwise, you will be tired and the shocks will frustrate you. Creating a spirit of cooperation and perseverance makes many of these obstacles more flexible, more realistic and easier to deal with.

Thank you very much, UPM. Thanks to everyone I have met and he assisted me or advised me or benefited me by whatever method. I will not call most of them by name, they really are unknown soldiers. But I will pray to God for all of them to guide them.

Thank you to the Supervisory Committee; Assoc. Prof. Dr. Halimi Moh Bin Saud, Assoc. Prof. Dr. Uma Rani Sinniah and Assoc. Prof. Dr. Intan Safinar Ismail. Thank you, my ex supervisor Assoc. Prof. Dr Maheran Abdul Aziz. Thank you Mr Nasrul Amri.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

**Halimi bin Mohd Saud, PhD**

Associate Professor  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Chairman)

**Uma Rani a/p Sinniah, PhD**

Professor  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Member)

**Intan Safinar binti Ismail, PhD**

Associate Professor  
Faculty of Science  
Universiti Putra Malaysia  
(Member)

---

**ZALILAH MOHD SHARIFF, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 12 August 2021

## Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No: Khalid Abubaker Alghannay, GS21560

## Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: \_\_\_\_\_  
Name of Chairman  
of Supervisory  
Committee: Associate Professor Dr. Halimi bin Mohd Saud

Signature: \_\_\_\_\_  
Name of Member  
of Supervisory  
Committee: Professor Dr. Uma Rani a/p Sinniah

Signature: \_\_\_\_\_  
Name of Member  
of Supervisory  
Committee: Associate Professor Dr. Intan Safinar binti Ismail

## TABLE OF CONTENTS

	<b>Page</b>
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	iii
<b>ACKNOWLEDGEMENTS</b>	v
<b>APPROVAL</b>	vi
<b>DECLARATION</b>	viii
<b>LIST OF TABLES</b>	xiv
<b>LIST OF FIGURES</b>	xv
<b>LIST OF ABBREVIATIONS</b>	xviii
<b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	<b>1</b>
1.1 Background of Study	1
1.2 Problem Statement and Justification	2
<b>2 LITERATURE REVIEW</b>	<b>4</b>
2.1 History of herbal medicine	4
2.2 Herbal Medicine	4
2.3 Phytochemical processing	5
2.4 The Malaysian Herbal Industry Scheme	8
2.5 Zingiberaceae family	9
2.5.1 Characteristics, habitats, and distribution of Zingiberaceae	9
2.6 Curcuma genus and its medicinal potentials	11
2.7 Curcuma zedoaria epitome	12
2.7.1 Classification and distribution	12
2.7.2 Botanical description of Curcuma	12
2.7.3 Significance and ethnomedicinal uses	13
2.7.4 Curcuma zedoaria and its pharmacological effects	15
2.7.5 Curcuma zedoaria and its Antioxidants activity	15
2.7.6 Phenolic compound of <i>C. zedoaria</i>	17
2.7.7 Flavonoids compound of <i>C. zedoaria</i>	18
2.7.8 Crude extract and dry powder of <i>C. zedoaria</i>	19
2.7.9 <i>C. zedoaria</i> essential oil	19
2.7.10 Application of tissue culture in plants for micropropagation	21
2.7.11 Micropropagation of <i>C. zedoaria</i>	22
2.8 Limitations and Motives	23
2.9 Culture media, conditions, and regeneration	24
2.10 Acclimatization to tissue culture conditions	25
2.11 Summary of Literature	26

<b>3</b>	<b><i>IN VITRO</i> REGENERATION OF KUNYIT PUTIH</b>	29
3.1	Introduction	29
3.2	Materials and methods	30
3.2.1	Study location and plant material	30
3.2.2	Glassware and chemicals	30
3.2.3	Media preparation and incubation	30
3.2.4	Sterilization	31
3.2.5	BAP concentration and shoot induction	31
3.2.6	BAP concentration and shoot multiplication	32
3.2.7	Kinetin concentration and shoot multiplication	32
3.2.8	TDZ concentration and shoot multiplication	32
3.2.9	NAA concentration and root induction	33
3.2.10	IBA concentration and root induction	33
3.2.11	Acclimatization	33
3.2.12	Statistical analysis	34
3.3	Results and discussion	34
3.3.1	Effect of surface sterilization on explants disinfection using different concentrations of NaOCl	34
3.3.2	Effect of surface sterilization on explants disinfection using different concentrations of mercuric chloride	36
3.3.3	Effect of surface sterilization on explants disinfection using different concentrations of Nano Silver	38
3.3.4	Effects of different BAP concentration on shoot induction of kunyit putih rhizomes	42
3.3.5	Effects of different BAP concentration on shoot multiplication	45
3.3.6	Effects of different Kinetin concentration on shoot multiplication	47
3.3.7	Effects of different TDZ concentration on shoot multiplication	50
3.3.8	Effects of different IBA concentration on root induction	52
3.3.9	Effects of different NAA concentration on root induction	55
3.4	Conclusion	56
<b>4</b>	<b>DETERMINATION OF PHYTOCHEMICAL CONTENTS USING DIFFERENT SOLVENTS IN LEAF AND RHIZOME OF KUNYIT PUTIH GROWN NATURALLY AND IN VITRO CULTIVATED</b>	58
4.1	Introduction	58
4.2	Materials and methods	60
4.2.1	Preparation of various solvents concentrations	60
4.2.2	Samples collection and preparation	60
4.2.3	Phytochemicals screening	60
4.2.4	Statistical analysis	62

4.3	Results and discussions	62
4.3.1	Screening of Various Solvents Concentrations and Determination of Phytochemical Contents in Leaf and Rhizome Extracts of Kunyit Putih	62
4.3.2	Comparative Phytochemicals Analysis of Kunyit putih Leaves and Rhizomes in Both Tissue Cultured Produced Plants (TCPP) and Naturally Field- Grown Plant (NFGP)	81
4.4	Conclusion	85
<b>5</b>	<b>ANTIMICROBIAL ACTIVITY OF THE DIFFERENT SOLVENT EXTRACTS OF LEAVES, RHIZOMES, AND OILS OF <i>Curcuma zedoaria</i></b>	<b>86</b>
5.1	Introduction	86
5.2	Materials and methods	87
5.2.1	Sample collection and preparation of plant extracts	87
5.2.2	Oil extraction from <i>C. zedoaria</i>	88
5.2.3	Microorganism analysis	88
5.2.4	Media preparation	88
5.2.5	Standardization of inoculum	89
5.2.6	Determination of sensitivity discs concentrations bioassay (antimicrobial susceptibility test)	89
5.2.7	Determination of minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC)	89
5.3	Results and discussion	90
5.3.1	Determining antimicrobial activity of the methanol, ethanol, and isopropanol extracts of <i>C. zedoaria</i> rhizomes and leaves by using sensitivity Discs Iffusion bioassay	90
5.3.2	Measurement the Minimum Inhibitory and Bactericidal Concentration (MIC and MBC)	94
5.3.3	Determining antimicrobial activity of the essential oil of <i>C. zedoaria</i> dry and fresh rhizomes extracts by using sensitivity Discs Iffusion bioassay	97
5.3.4	Measurement the Minimum Inhibitory and Bactericidal Concentration (MIC and MBC)	100
5.4	Conclusion	102
<b>6</b>	<b>GENERAL DISCUSSION, CONCLUSION AND RECOMMENDATION FOR FUTURE</b>	<b>103</b>
6.1	General discussion	103
6.2	Conclusion	107
6.3	Recommendations for future research	109

<b>REFERENCES</b>	110
<b>APPENDICES</b>	133
<b>BIODATA OF STUDENT</b>	143



© COPYRIGHT UPM



## LIST OF TABLES

Table	Page
2.1 Lists of common forms of herbal related products and their commonly accepted definitions (Ismail, 2003)	7
2.2 Zingiberaceae in Asian Countries	10
2.3 Scientific classification of Zingiberaceae	10
2.4 Taxonomic classification of <i>Curcuma zedoaria</i>	12
2.5 Display different parts of <i>C. zedoaria</i> used in traditional medicine (Lobo et al., 2009b)	14
2.6 Phytochemical analysis of methanol extract of rhizomes of <i>C. zedoaria</i> (Sumathi et al., 2013)	19
4.1 Total flavonoids content in the dry matter of Kunyit putih leaves (mg eq rutin gr <sup>-1</sup> ) at different solvents concentrations	64
4.2 Total flavonoids content of the dry matter of Kunyit hitam rhizome (mg eq rutin gr <sup>-1</sup> ) at different solvent concentrations	67
4.3 Total antioxidant content in the dry matter of Kunyit putih leaf at different solvents concentrations	71
4.4 Total antioxidant content (%) in the dry matter of Kunyit Putih rhizome at different solvent concentrations	73
4.5 Total phenolic content (mg eq rutin gr <sup>-1</sup> ) in the dry matter of Kunyit putih leaves at different solvent concentrations	76
4.6 Total phenolic content (mg eq rutin gr <sup>-1</sup> ) in the dry matter of Kunyit putih rhizome at different solvent concentrations	79
5.1 Antimicrobial activity of the methanol, ethanol, and isopropanol extracts of <i>C. zedoaria</i> rhizomes and leaves.	92
5.2 Minimum Inhibition Concentration (MIC; mg/mL) and Minimum Bacterial Concentration (MBC; mg/mL) of different solvents extracts	96
5.3 Antimicrobial activity of the essential oil of <i>C. zedoaria</i> rhizomes	98
5.4 Minimum Inhibition Concentration (MIC; Mg/L) and Minimum Bacterial Concentration (MBC; Mg/L) of essential oil samples of <i>C. zedoaria</i>	101

## LIST OF FIGURES

Figure	Page
1.1 Workflow of the research objectives carried out in this thesis	133
2.1 Increasing value of herbal products with processing and standardization (Ismail, 2003)	8
2.2 <i>C.zedoaria</i> plant. (A): one of the adaptation stages for plants resulting from tissue culture; (B): whole plant after reaching full maturity	13
3.1 Effect of surface sterilization using different concentrations of NaOCl on disinfection of kunyit putih explants. Means followed by a same letter (s) are not significantly different based on DMRT ( $P \leq 0.05$ )	35
3.2 Effect of surface sterilization using different mercuric chloride concentrations on disinfection of kunyit putih explants. Means followed by a same letter (s) are not significantly different based on DMRT ( $P \leq 0.05$ ) on explants disinfection	37
3.3 Effect of surface sterilization using different concentrations of nano silver on explants disinfection of kunyit putih explants. Means followed by a same letter (s) are not significantly different based on DMRT ( $P \leq 0.05$ )	39
3.4 Surface sterilization of kunyit putih explants using different sterilizing agents. (A) and (B): natural state of rhizomes shows the position of the apical buds; (C), (D) and (E): sterilization of excised explants; (F), (G), (H), (I), (J) and (K): <i>in vitro</i> emergence of Kunyit putih buds; and (L) <i>in vitro</i> shoot emergence. (Bar=1 cm)	41
3.5 Effects of different concentration of BAP on (A) percentage of shoot formation of kunyit putih; and (B) a mean number of shoot produced per rhizome explants. Means followed by a same letter (s) are not significantly different based on DMRT ( $P \leq 0.05$ )	43
3.6 Effects of BAP on shoot induction of kunyit putih explants after eight weeks of culture. (A), (B) shoot induction, (C) shoot and leaf development (Bar = 0.6 cm)	44
3.7 Effects of different concentration of BAP on the mean number and mean length of shoots produced per explants of kunyit putih. Means followed by a same letter (s) are not significantly different based on DMRT ( $P \leq 0.05$ )	46

3.8	Effects of BAP on shoot multiplication of kunyit putih explants after eight weeks of culture. (A) Shoot induction, (B) and (C) well-formed multiple shoot regenerants. (Bar = 1 cm)	47
3.9	Effects of different kinetin concentration on shoot multiplication Means followed by a same letter (s) are not significantly different based on DM ( $P \leq 0.05$ )	48
3.10	Effects of kinetin on shoot multiplication of kunyit putih explants after eight weeks of culture. (A) Shoot induction, (B) and (C) well-formed	49
3.11	Effects of TDZ on shoot multiplication of kunyit putih explants after eight weeks of culture. Means followed by a same letter (s) are not	50
3.12	Effects of TDZ on shoot multiplication of kunyit putih explants	52
3.13	Effects of different IBA concentration on root induction of kunyit putih explants after eight weeks of culture. Means followed by a same letter (s) are not significantly different based on DMRT ( $P \leq 0.05$ )	53
3.14	Effects of IBA on root induction of kunyit putih explants after eight weeks of culture. (A) and (B) well-formed root developments. (Bar = 1	54
3.15	Effects of different NAA concentration on root induction of kunyit putih explants after eight weeks of culture. Means followed by a same letter (s) are not significantly different based on DMRT ( $P \leq 0.05$ ).	55
3.16	Effects of NAA on root induction of kunyit putih explants after eight weeks of culture. (A) and (B) root inductions. (Bar = 1 cm)	56
4.1	Total flavonoids contents in the dry matter of Kunyit Putih leaves	65
4.2	Total flavonoids contents of the dry matter of Kunyit Putih rhizome obtained with different solvents concentrations	69
4.3	Total Antioxidant content in the dry matter of Kunyit Putih Leaves	72
4.4	Total antioxidant content in the dry matter of Kunyit Putih rhizome obtained using different solvents concentrations	74
4.5	Total phenolic contents in the dry matter of Kunyit Putih leaves obtained using different solvents concentrations	77
4.6	Total phenolic content in the dry matter of Kunyit Putih rhizome obtained using different solvents concentrations	80
4.7	Total flavonoids content of TCPP and NFGP leaves and rhizome of kunyit putih (Methanol 90%)	82

4.8	The Total phenolic content of NFGP and TCPP leaves and rhizome of kunyit putih(Isopropanol 30%)	83
4.9	The Total antioxidant activity of NFGP and TCPP leaves and rhizome of kunyit putih (Isopropanol 30%)	84
5.1	Inhibition zone of of rhizome crude extracts against s.m bacteria. (A), (B) and (C) 10, 25 and 50 mg/mL of rhizome crude extracts	93
5.2	Minimum bactericidal concentration of <i>C. zedoaria</i> extract against <i>E. Coli</i> strain	97
5.3	Disc-diffusion test of <i>C. zedoaria</i> oil extract against B.S Bacteria	99
5.4	Minimum bactericidal concentration of <i>C. zedoaria</i> extract against <i>Streptococcus mutans</i>	102

## LIST OF ABBREVIATIONS

<i>A. anitratus</i>	Acinetobacter anitratus
AD	Air Drying
ATCC	American Type Culture Collection
<i>B. cereus</i>	<i>Bacillus cereus</i>
<i>B. subtilis</i>	<i>Bacillus subtilis</i>
CFU	Colony forming unit
DMSO	Dimethylsulfoxide
<i>E. coli</i>	<i>Escherichia coli</i>
FAO	Food and Agriculture Organization
IBS	Institute of Bioscience
<i>K. pneumonia</i>	<i>Klebsiella pneumonie</i>
<i>L. monocytogenes</i>	<i>Listeria monocytogenes</i>
MBC	Minimal bactericidal concentration
MHA	Mueller Hinton agar
MIC	Minimal inhibitory concentration
NFGP	Naturally Field-Grown Plant
<i>P. acane</i>	<i>Propionibacterium acne</i>
<i>S. aureus</i>	<i>Staphylococcus aureus</i>
<i>S. mutans</i>	<i>Streptococcus mutans</i>
TCPP	<i>Tissue Cultured Produced Plants</i>
UPM	Universiti Putra Malaysia
WHO	World Health Organization
DMSO	Dimethyl Sulfoxide
DPPH	DPPH radical scavenging assay
ISOPRO	IsoPropanol

OD	Oven Drying
GAE	Gallic Acid Equivalent
MeOH	Methanol
EtOH	Ethanol
Aq	Water
mL	MilliL
°C	Degree in Celsius
ppm	Part Per Million
TAC	Total Antioxidant Contents
TFC	Total Flavonoid Content
TPC	Total Phenolic Contents

# CHAPTER 1

## INTRODUCTION

### 1.1 Background of Study

Major challenges facing today's healthcare system are issues such as chronic diseases, resistant infections, autoimmune disorder, and degenerative disease of aging and insecure medicines, despite the advancement in medical sciences (Tholkappiyavathi et al., 2013). The rhizomes of *Curcuma zedoaria* have been extensively used in the treatment of ulcers, wounds, tumors, atherosclerosis, and inflammation as well as various traditional preparations used for the treatment of Ohyl syndrome, a major caused of blood stagnation (Seo et al., 2005; Tariq et al., 2016). Bharalee et al, (2005), confirms that their rhizomes, are traditionally used as appetizer and antipyretic and its useful in bronchitis, asthma, tuberculosis and also against enlargement of spleen. Rhizomes are also used as tonic to the brain and heart, expectorant and in treatment of pains, inflammation, toothache, bruises and sprains.

*Curcuma zedoaria* is a perennial herb and member of the genus *Curcuma*, family Zingiberaceae (Ullah et al., 2014; Tiphara et al, 2007). It is a potential source of therapeutic molecules for the treatment of a range of ailments as the genus is credited with anti-inflammatory, antibiotic, antiviral, anticancerous, hypocholestraemic, choleric, antidiabetic, antihepatotoxic, antivenomous, and antirheumatic properties (Parthasarathy et al., 2006; Safitri et al., 2017; Xiong et al., 2017). The *Curcuma* has attracted attention since last 3 decades.

The *C. zedoaria* is associated with phenolic and flavonoid compounds possessing strong antioxidant activities (Avanço et al., 2017; Tiphara et al, 2007). However, there are conflicting reports on the phenolic content and contribution of phenolic compound to the total antioxidant activities of *C. zedoaria* (Kim et al., 2011; Ghasemzadeh et al., 2011; Hossain et al., 2011). It is also showed that the extraction yield of phenolic and flavonoid content is greatly depending on the solvent polarity (Senathilake et al., 2016). The antioxidant activity of polyphenols is due to their ability to scavenge free radicals, donate hydrogen atoms or electron, or chelate metal cations (Złotek et al., 2016). Presently, the focus has been shifted to naturally occurring antioxidant. The use of natural antioxidant is considered to be safe rather than synthetic as latter has the potential for carcinogenic. The antioxidants present in diet can reduce attacks and risks of diseases by reducing the free radicals.

Tissue culture technique has been successfully used to grow wild plants that are difficult to propagate through the conventional ways or plants with certain defects (Lima et al., 2012). Especially, when the populations have decreased as a result of over exploitation by destructive harvesting, and when there is a growing demand

for clonally uniform elite plants and, also, when species have been over collected by hobbyists for medicine, food or fragrance, then, in vitro propagation can provide an alternate source of plants and alleviate the pressures on wild populations (Kapai et al., 2010). Plant tissue culture is an enabling technology from which many novel tools have been developed to assist plant breeders to increase the speed or efficiency of the breeding process and to create a new variation for crop improvement (Pikulthong et al., 2016). However, the establishment of a good axenic culture is another inherent problem with *Curcuma* rhizome (Das et al., 2010a).

## 1.2 Problem Statement and Justification

The screening of phytochemicals has been utilized for exploring antioxidant compounds in plants (Tiwari et al., 2011; Wadood et al., 2013; Ribeiro et al., 2016). However, the yield of extraction and antioxidant activity does not only depend on the extraction method, and also on the solvent used for extraction. The current increase in demand for *C. zedoaria* is the motivating factor to investigate its biochemical properties and the actual content in various tissue, due to lack of studied screening methods for solvent to be used. The solubility of antioxidant compounds in *C. zedoaria* with different polarities and chemical characteristics is influenced by the solvent of extraction. (Do et al., 2014). The increase in antibiotic resistance and failure of chemotherapeutics exhibited by pathogenic microbial infectious agents has led to the screening of several medicinal plants for their potential antimicrobial activity (Du Toit and Rautenbach, 2000; Wilson et al., 2005; Wayne et al., 2012; Medini et al., 2014). However, the test systems ought to be simple, rapid, reproducible, and inexpensive and maximize high sample throughput in order to cope with a varied number of extracts and fractions (Dias et al., 2012).

The methods of propagation are slow, inconsistent, and it suffers frequent crop loss. Hence, it's imperative to provide more attractive, flexible and innovative way to accelerate the acquisition of healthy plants in reasonable quantities and these drawbacks is also coupled with high rate of contamination of tissue culture of *C. zedoaria* plant. Where the stages of in vitro propagation of *C. zedoaria* comprises of selection explants, aseptic culture establishment, multiplication of propagules, rooting, and acclimatization (Anisuzzaman et al., 2008). Sterilization is the most challenging step of explants aseptic culture establishment (Bhattacharya et al., 2014). Among the various explants previously tested, rhizome buds have been found to be very suitable material for in vitro propagation (tissue culture) of *Curcuma* (Islam et al., 2004; Roy and Raychaudhuri, 2004; Tiphara et al, 2007). Among the common problems of explants from wild plants include; diseased specimens, or plant parts located close to or below the soil that make it difficult or impossible to disinfect due to both endophytic and epiphytic microbes (Reed et al., 1995).



Thus, a tissue culture technique will play an important role in the study of *C. zedoaria* plants (Loc et al., 2005). It is deemed important to develop a Micropropagation technique to make commercially available of pathogen-free germplasm. The tissue culture offers an alternative tool for rapid multiplication and conservation of disease-free propagules, which will further enable uninterrupted supply of raw materials. In this project, *C. zedoaria* leaves and rhizome from Malaysian traditional plants which has been used in folkloric practices for the management of diseases was selected for this study.

Thus, the hypotheses of this study states that, the selected cytokines and auxins have distinctive potential to stimulate shoot growth and root induction respectively by micro propagation techniques and extract from in vitro culture of *C. zedoaria* with high antioxidative and flavonoid content will have antibacterial effect against pathogenic bacterial. Therefore, the main objectives of this study are:

1. To establish in vitro regeneration protocol for Kunyit Putih (*C. zedoaria*) using tissue culture techniques.
2. To optimize ideal solvents and its concentrations require for the selection and screening of various phytochemical extracts (secondary metabolite) of *C. zedoaria* in leaf and rhizome for total phenolic content (TPC) and total flavonoids content (TFC), and antioxidant activity (AO).
3. To compare extractability of phytochemicals contents using methanol and isopropanol on total flavonoids and phenolic/antioxidant contents of Kunyit Putih leaves and rhizomes before and after tissue culture for in vitro and in-vivo study.
4. To evaluate the antimicrobial activity of the different solvent extracts of leaves, rhizomes, and oils of *C. zedoaria* against gram-positive and negative pathogenic bacteria.

## REFERENCES

- Abdi, G., Salehi, H., and Khosh-Khui, M. (2008). Nano silver: a novel nanomaterial for removal of bacterial contaminants in valerian (*Valeriana officinalis* L.) tissue culture. *Acta Physiologiae Plantarum*, 30(5), 709–714.
- Adenan, M. I. (2003). Malaysian herbs and herbal products. A two and half day course of herbal and phytochemical processing. CEPP short course notes. *Chemical Engineering Pilot Plant*, Universiti Teknologi Malaysia, Malaysia).
- Aggarwal, B. B., Sundaram, C., Malani, N., Ichikawa, H. (2007). Curcumin: the Indian solid gold, In: *The molecular targets and therapeutic uses of curcumin in health and disease* (pp. 1-75). Springer, Boston, MA.
- Ahmed, S., & Hasan, M. M. (2017). Exploring globally used antiurolithiatic plants of S to Z families: including Saxifragaceae, Scrophulariaceae, Solanaceae, Urticaceae, Vitaceae, Zingiberaceae and Zygophyllaceae. *Journal of Pharmacognosy and Phytochemistry*, 6(4), 1430-1436.
- Ahmad, I., Mahmood, Z., Mohammad, F. (1998). Screening of some Indian medicinal plants for their antimicrobial properties. *Journal of Ethnopharmacology* 62, 183–193.
- Akter, N. (2017). Antibacterial Sensitivity Test of Crude Extract of (*Curcuma zedoaria*, *Solanum virginianum* and *Stephania japonica*) and Resistant Pattern of Clinically Isolated Bacteria against Conventionally Used Antibiotics (Doctoral dissertation, East West University).
- Al Abdallat, A. M., Sawwan, J. S., & Al Zoubi, B. (2011). Agrobacterium tumefaciens-mediated transformation of callus cells of *Crataegus aronia*. *Plant Cell, Tissue and Organ Culture (PCTOC)*, 104(1), 31-39.
- Alamgir, A. N. M. (2017). Cultivation of herbal drugs, biotechnology, and In Vitro production of secondary metabolites, high-value medicinal plants, herbal wealth, and herbal trade. In *Therapeutic Use of Medicinal Plants and Their Extracts: Volume 1* (pp. 379-452). Springer, Cham.
- Ali, B. H., Blunden, G., Tanira, M. O., and Nemmar, A. (2008). Some phytochemical, pharmacological and toxicological properties of ginger (*Zingiber officinale* Roscoe): a review of recent research. *Food and Chemical Toxicology. An International Journal Published for the British Industrial Biological Research Association*, 46(2), 409–20.
- Ali, B., and Hasnain, S. (2007). Potential of bacterial indoleacetic acid to induce adventitious shoots in plant tissue culture. *Letters in applied microbiology*, 45(2), 128-133.

- Almeida, M. C. D. (2012). Effect of the radiation processing in species of Zingiberaceae family: turmeric (*Curcuma longa* L.), ginger (*Zingiber officinale* Roscoe) and zedoaria (*Curcuma zedoaria* (Christm.) Roscoe).
- Almodaifer, S., Alsibaie, N., Alhoumedan, G., Alammari, G., & Kavita, M. S. (2017). Role of Phytochemicals in Health and Nutrition. *BAOJ Nutrition*, 3(2), 028.
- Andriyani, M. (2017). Zingiberaceae of the Ternate Island: Almost a Hundread Years after Beguin's Collection. *Jurnal Biologi Indonesia*, 6(3).
- Anisuzzaman, M., Sharmin, S. A., Mondal, S. C., Sultana, R., Khalekuzzamzan, M., Alam, I., and Alam, M., F. (2008). *In vitro* microrhizome induction in *Curcuma zedoaria* (Christm.) Roscoe—a conservation prioritized medicinal plant. *J. Biol. Sci*, 8(7), 1216- 1220.
- Anonymous (1985). The Wealth of India—Raw Materials IA (Revised): A Dictionary of Indian Raw Materials and Industrial Products, vol. 69. CSIR, New Delhi, pp. 401–406 *Journal of Biological Sciences*, 8(7), 1216–1220.
- Apisariyakul, A., Vanittanakom, N., and Buddhasukh, D. (1995). Antifungal activity of turmeric oil extracted from *Curcuma longa* (Zingiberaceae). *Journal of ethnopharmacology*, 49(3), 163-169.
- Archana, C. P. (2015). Zingiberaceae members excluding ginger and turmeric- a review. *International Journal of Scientific Research Today*, 1(January), 101–107.
- Arif, M. T. (2002). Keynote address: Traditional Complimentary Medicine in the Malaysian Healthcare System. 4th International Conference for Traditional Complimentary Medicine, Sunway Convention Centre, Kuala Lumpur, 14th October.
- Aschwanden, C. (2001). Herbs for health, but how safe are they? *Bulletin of the World Health Organization*, 79, 691-692.
- Ashebir, M., and Ashenafi, M. (2017). Assessment of the antibacterial activity of some traditional medicinal plants on some food-borne pathogens. *The Ethiopian Journal of Health Development (EJHD)*, 13(3).
- Audu, J. A. (1995). Studies on the effectiveness of medicinal herbs used as anthelmintics by traditional medical practitioners in South of Bauchi State II. *Journal of Economic and Taxonomic Botany* 19, 653–661.
- Avanço, G. B., Ferreira, F. D., Bomfim, N. S., Peralta, R. M., Brugnari, T., Mallmann, C. A., and Machinski Jr, M. (2017). *Curcuma longa* L. essential oil composition, antioxidant effect, and effect on *Fusarium verticillioides* and fumonisin production. *Food Control*, 73, 806-813.

- Awin, T., Mediani, A., Shaari, K., Faudzi, S. M. M., Sukari, M. A. H., Lajis, N. H., and Abas, F. (2016). Phytochemical profiles and biological activities of *Curcuma* species subjected to different drying methods and solvent systems: NMR-based metabolomics approach. *Industrial Crops and Products*, 94, 342-352.
- Azahar, N. F., Gani, S. S. A., and Mokhtar, N. F. M. (2017). Optimization of phenolics and flavonoids extraction conditions of *Curcuma zedoaria* leaves using response surface methodology. *Chemistry Central Journal*, 11(1), 96.
- Azam, M. G., Noman, M. S., and Pavel, M. A. M. (2017). Evaluation of anti-diarrhoeal activity of *Curcuma zedoaria* rhizome. *Journal of Pharmacognosy and Phytochemistry*, 6(3), 171-173.
- Aziz, R. A., Sarmidi, M. R., Kumaresan, S., Taher, Z. M., and Foo, D. C. Y. (2003). Phytochemical Processing: The Next Emerging Field in Chemical Engineering—Aspects and Opportunities. *Jurnal Kejuruteraan Kimia Malaysia*, 3, 45-60.
- Azwanida, N. N. (2015). A review on the extraction methods use in medicinal plants, principle, strength and limitation. *Med Aromat Plants*, 4(196), 2167-0412.
- Bai, L., Leong-Škorničková, J., Li, D., and Xia, N. (2017). Taxonomic studies on Zingiber (Zingiberaceae) in China IV: *Z. pauciflorum*, a new species from Yunnan. *Nordic Journal of Botany*, 36(3), njb-01534.
- Bajaj, Y. P. S., Furmanowa, M., and Olszowska, O. (1988). Biotechnology of the Micropropagation of medicinal and aromatic plants. In *Medicinal and aromatic plants I* (pp. 60-103). Springer Berlin Heidelberg.
- Bakkali, F., Averbeck, S., Averbeck, D., and Idaomar, M. (2008). Biological effects of essential oils - A review. *Food and Chemical Toxicology*, 46(2), 446-475.
- Balachandran, S. M., Bhat, S. R., and Chandel, K. P. (1990). *In vitro* clonal multiplication of turmeric (*Curcuma* spp.) and ginger (*Zingiber officinale* Rosc.). *Plant Cell Reports*, 8(9), 521-4.
- Balick, M. J., and Mendelsohn, R. (1992). Assessing the economic value of traditional medicines from tropical rain forests. *Conservation biology*, 6(1), 128-130.
- Bank, E. (2003). *Exporting Indian Healthcare*.
- Banisalam, B., Sani, W., Philip, K., Imdadul, H., and Khorasani, A. (2011). Comparison between *in vitro* and *in vivo* antibacterial activity of *Curcuma zedoaria* from Malaysia. *African Journal of Biotechnology*, 10(55), 11676-11681.

- Barbieri, R., Coppo, E., Marchese, A., Daglia, M., Sobarzo-Sánchez, E., Nabavi, S. F., & Nabavi, S. M. (2017). Phytochemicals for human disease: An update on plant-derived compounds antibacterial activity. *Microbiological research*, 196, 44-68.
- Basak, S., Sarma, G. C., & Rangan, L. (2010). Ethnomedical uses of Zingiberaceous plants of Northeast India. *Journal of ethnopharmacology*, 132(1), 286-296.
- Bharalee, R., Das, A., and Kalita, M. C. (2005). *In vitro* clonal propagation of *Curcuma caesia* Roxb and *Curcuma zedoaria* Rosc from rhizome bud explants. *Journal of plant biochemistry and biotechnology*, 14(1), 61-63.
- Bhattacharya, M., Goyal, A., and Mishra, T. (2014). *In vitro* regeneration of some lesser known medicinal Zingibers: a review. *Biology of Useful Plants and Microbes*, (January 2014), 167–186.
- Bhojwani, S. S., & Razdan, M. K. (1983). Clonal propagation. *Plant tissue culture: Theory and practice*, 5, 313-72.
- Bhowmik, S. S. D., Kumaria, S., & Tandon, P. (2013). Rhizome derived callogenic plantlet production of *Mantisia wengeri* (Zingiberaceae), a rare and endemic medicinal plant of Mizoram, North-East India.
- Boonmee, A., Srisomsap, C., Chokchaichamnankit, D., Karnchanatat, A., and Sangvanich, P. (2011). A proteomic analysis of *Curcuma comosa* Roxb. rhizomes. *Proteome science*, 9(1), 43.
- Borthakur, M., Hazarika, J., and Singh, R. S. (1999). A protocol for Micropropagation of *Alpinia galanga*. *Plant Cell, Tissue and Organ Culture*, 55(3), 231–233.
- Bouldin, A. S., Smith, M. C., Garner, D. D., Szeinbach, S. L., Frate, D. A., & Croom, E. M. (1999). Pharmacy and herbal medicine in the US. *Social Science & Medicine*, 49(2), 279-289.
- Braga, M. E., & Meireles, M. A. A. (2007). Accelerated solvent extraction and fractioned extraction to obtain the *Curcuma longa* volatile oil and oleoresin. *Journal of food process engineering*, 30(4), 501-521.
- Budziszewski, G. J., Croft, K. P. C., and Hildebrand, D. F. (1996). Uses of biotechnology in modifying plant lipids. *Lipids*, 31(6), 557–569.
- Calixto, J. B., Otuki, M. F., and Santos, A. R. (2003). Anti-inflammatory compounds of plant origin. Part I. Action on arachidonic acid pathway, nitric oxide and nuclear factor kappa B (NF-kappaB). *Planta Medica*, 69(11), 973–83.

- Carvalho, F. R., Vassao, R. C., Nicoletti, M. A., and Maria, D. A. (2010). Effect of *Curcuma zedoaria* crude extract against tumor progression and immunomodulation. *Journal of Venomous Animals and Toxins including Tropical Diseases*, 16(2), 324-341.
- Chadwick, D. J., and Marsh, J. (Eds.). (2008). *Ethnobotany and the search for new drugs* (Vol. 185). John Wiley and Sons.
- Chattopadhyay, I. K., Biswas, U., Bandyopadhyay and R.K. Banerjee (2004). Turmeric and curcumin: Biological actions and medicinal applications. *Curr. Sci.*, 87: 44-53.
- Chauhan, P., Keni, K., and Patel, R. (2017). Investigation of phytochemical screening and antimicrobial activity of *Curcuma longa*. *Int. J. Adv. Res. Biol. Sci.*, 4(4), 153-163.
- Chen, I. N., Chang, C. C., Ng, C. C., Wang, C. Y., Shyu, Y. T., and Chang, T. L. (2008). Antioxidant and antimicrobial activity of Zingiberaceae plants in Taiwan. *Plant foods for human Nutrition*, 63(1), 15-20.
- Chen, X., and Schluesener, H. (2008). Nanosilver: a nanoproduct in medical application. *Toxicology Letters*.
- Chen, W., Lu, Y., Gao, M., Wu, J., Wang, A., and Shi, R. (2011). Anti-angiogenesis effect of essential oil from *Curcuma zedoaria* *in vitro* and *in vivo*. *Journal of Ethnopharmacology*, 133(1), 220–226.
- Chen, C. C., Chen, Y., Hsi, Y. T., Chang, C. S., Huang, L. F., Ho, C. T., ... & Kao, J.Y. (2013). Chemical constituents and anticancer activity of *Curcuma zedoaria* roscoe essential oil against non-small cell lung carcinoma cells *in vitro* and *in vivo*. *Journal of agricultural and food chemistry*, 61(47), 11418-11427.
- Chiang, M., Kurmoo, Y., and Khoo, T. J. (2017). Chemical-and Cell-based Antioxidant Capacity of Methanolic Extracts of Three Commonly Edible Plants from Zingiberaceae Family. *Free Radicals and Antioxidants*, 7(1).
- Chuenwittaya, S. (2008). Effect of growing media on growth of *Curcuma* seedling (*Curcuma* hybrid) from different methods of tissue culture. Thai National AGRIS Centre, Food and Agriculture Organization of the United Nations.
- Colombo, R., de L. B., Andrea, N., Teles, H. L., Silva, G. H., Bomfim, G. C., Burgos, R. C., Cavalheiro, A. J., da Silva Bolzani, V., and Silva, D. H. S. (2009). Validated HPLC method for the standardization of *Phyllanthus niruri* (herb and commercial extracts) using corilagin as a phytochemical marker. *Biomedical Chromatography*, 23(6), 573- 580.

- Cowan, M. (1999). Plant Products as Antimicrobial Agents, *Clinical Microbiology Reviews*, 565, 568, 570. Department of Microbiology, Miami University: Oxford, Ohio.
- Cragg, G. M., & Newman, D. J. (2002). Drugs from nature: past achievements, future prospects. In *Advances in Phytomedicine* (Vol. 1, pp. 23-37). Elsevier.
- Das, A., Kesari, V., and Rangan, L. (2010a). Plant regeneration in *Curcuma* species and assessment of genetic stability of regenerated plants. *Biologia Plantarum*, 54(3), 423– 429.
- Das, K., Tiwari, R., and Shrivastava, D. (2010b). Techniques for evaluation of medicinal plant products as antimicrobial agents: current methods and future trends. *Journal of medicinal plants research*, 4(2), 104-111.
- Debnath, S. C. (2014). Bioreactor-induced adventitious shoot regeneration affects genotype-dependent morphology but maintains clonal fidelity in red raspberry. *In vitro Cellular and Developmental Biology-Plant*, 50(6), 777-788.
- Decker, A. (2009). Phenolics: Prooxidants or Antioxidants? *Nutrition Reviews*, 55(11), 396– 398.
- DellaPenna, D. (2001). Plant metabolic engineering. *Plant Physiology*, 125(1), 160-163.
- Desmiaty, Y. (2017). Antioxidant and Inhibitor Elastase Activity of *Kaempferia Rotunda* L. and *Curcuma zedoaria* (Christm.) Roscoe. 2nd ISEJ 2017 *Abstract and Full Paper Submission System*, 1(1), 125-125.
- Dhami, N., & Mishra, A. D. (2015). Phytochemical variation: how to resolve the quality controversies of herbal medicinal products?. *Journal of herbal medicine*, 5(2), 118-127.
- Diamond, S. (2017). In search of the primitive: *A critique of civilization*. Routledge.
- Dias, D. A., Urban, S., and Roessner, U. (2012). A historical overview of natural products in drug discovery. *Metabolites*, 2(2), 303-336.
- Do, Q. D., Angkawijaya, A. E., Tran-Nguyen, P. L., Huynh, L. H., Soetaredjo, F. E., Ismadji, S., and Ju, Y. H. (2014). Effect of extraction solvent on total phenol content, total flavonoid content, and antioxidant activity of *Limnophila aromatica*. *Journal of food and drug analysis*, 22(3), 296-302.
- Du Toit, E. A., and Rautenbach, M. (2000). A sensitive standardised micro-gel well diffusion assay for the determination of antimicrobial activity. *Journal of microbiological methods*, 42(2), 159-165.

- El-Hawaz, R., Park, D., Bridges, W. C., and Adelberg, J. (2016). Optimizing *in vitro* mineral nutrition and plant density increases greenhouse growth of *Curcuma longa* L. during acclimatization. *Plant Cell, Tissue and Organ Culture*, 126(1), 33-42.
- Elias, L., Pradeep, A. H., and Hegde, P. L. (2015). *Curcuma angustifolia* Roxb (Tavaksheeri): A Review. *Journal of Pharmacognosy and Phytochemistry*, 4(2), 241–243.
- Ficker, C. E., Smith, M. L., Susiarti, S., Leaman, D. J., Irawati, C., and Arnason, J. T. (2003). Inhibition of human pathogenic fungi by members of Zingiberaceae used by the Kenyah (Indonesian Borneo). *Journal of Ethnopharmacology* 85, 289–293.
- Finch, J. (2011). The ancient origins of prosthetic medicine. *The Lancet*, 377(9765), 548-549. Gantait, S., and Kundu, S. (2017). Neoteric trends in tissue culture-mediated biotechnology of Indian ipecac [*Tylophora indica* (Burm. f.) Merrill]. *3 Biotech*, 7(3), 231.
- Garcia-Viguera, C., Zafrilla, P., & Tomás-Barberán, F. A. (1998). The use of acetone as an extraction solvent for anthocyanins from strawberry fruit. *Phytochemical Analysis: An International Journal of Plant Chemical and Biochemical Techniques*, 9(6), 274- 277.
- Ghasemzadeh, A., Jaafar, H. Z., and Rahmat, A. (2011). Effects of solvent type on phenolics and flavonoids content and antioxidant activities in two varieties of young ginger (*Zingiber officinale* Roscoe) extracts. *Journal of Medicinal Plants Research*, 5(7), 1147-1154.
- Goldman, E. L. (2001). Industry self-regulation in the manufacture of dietary supplements and botanical medicines. *Clinical obstetrics and gynecology*, 44(4), 789-800.
- Grąbkowska, R., Sitarek, P., and Wysokińska, H. (2014). Influence of thidiazuron (TDZ) pretreatment of shoot tips on shoot multiplication and ex vitro acclimatization of *Harpagophytum procumbens*. *Acta physiologiae plantarum*, 36(7), 1661-1672.
- Grell, O. P., & Cunningham, A. (2017). Health care and poor relief in 18th and 19th century Northern Europe. In *Health care and poor relief in 18th and 19th century Northern Europe* (pp. 3-14). Routledge.
- Gupta, A., Mahajan, S., and Sharma, R. (2015). Evaluation of antimicrobial activity of *Curcuma longa* rhizome extract against *Staphylococcus aureus*. *Biotechnology reports*, 6, 51-55.
- Haddad, M., Sauvain, M., and Deharo, E. (2011). *Curcuma* as a parasiticidal agent: a review. *Planta medica*, 77(06), 672-678.



- Hadisaputri, Y. E., Miyazaki, T., Suzuki, S., Kubo, N., Zuhrotun, A., Yokobori, T. Kuwano, H. (2015). Molecular characterization of antitumor effects of the rhizome extract from *Curcuma zedoaria* on human esophageal carcinoma cells. *International Journal of Oncology*, 47(6), 2255–2263.
- Hagos, R., & Gebremdhin, H. (2015). Effects of cytokinin types and their concentration on in vitro shoot induction and multiplication of korarima. *International Journal of Genetics and Molecular Biology*, 7(2), 8-14.
- Hamdi, O. A. A., Satti, R., and Jacknoon, A. (2017). Sesquiterpenes from the Rhizomes of *Curcuma zedoaria* and their Cytotoxicity against Leukemic Cell Lines.
- Hamirah, M. N., Sani, H. B., Boyce, P. C., & Sim, S. L. (2007, June). Micropropagation of red ginger (*Zingiber montanum* Koenig), a medicinal plant. In *Proceedings Asia Pacific Conference on Plant Tissue and Agribiotechnology (APaCPA)* (Vol. 17, p. 21).
- Harborne, A. (1998). *Phytochemical methods a guide to modern techniques of plant analysis: springer science and business media*.
- Hashemy, T., Maki, H., Yamada, Y., Kaneko, T. S., & Syono, K. (2009). Effects of light and cytokinin on in vitro micropropagation and microrhizome production in turmeric (*Curcuma longa* L.). *Plant biotechnology*, 26(2), 237-242.
- Hatano, T., Yasuhara, T., Yoshihara, R., Agata, I., Noro, T., and Okuda, T. (1990). Effects of Interaction of Tannins with Co-existing Substances. VII: Inhibitory Effects of Tannins and Related Polyphenols on Xanthine Oxidase. *Chemical and Pharmaceutical Bulletin*, 38(5), 1224-1229.
- Herdt, R. H. (2006). Biotechnology in Agriculture. *Annual Review of Environment and Resources*, 31(1), 265–295.
- Holowchak, M. (2001). Interpreting dreams for corrective regimen: Diagnostic dreams in Greco-Roman medicine. *Journal of the history of medicine and allied sciences*, 56(4), 382-399.
- Hossain, M. A., AL-Raqmi, K. A. S., AL-Mijizy, Z. H., Weli, A. M., and Al-Riyami, Q. (2013). Study of total phenol, flavonoids contents and phytochemical screening of various leaves crude extracts of locally grown *Thymus vulgaris*. *Asian Pacific journal of tropical biomedicine*, 3(9), 705-710.
- Hossain, M. A., Shah, M. D., Gnanaraj, C., and Iqbal, M. (2011). *In vitro* total phenolics, flavonoids contents and antioxidant activity of essential oil, various organic extracts from the leaves of tropical medicinal plant *Tetragonia* from Sabah. *Asian Pacific journal of tropical medicine*, 4(9), 717-721.

- Hou, X. L., Takahashi, K., Kinoshita, N., Qiu, F., Tanaka, K., Komatsu, K., and Azuma, J. (2007). Possible inhibitory mechanism of *Curcuma* drugs on CYP3A4 in  $1\alpha, 25$  dihydroxyvitamin D 3 treated Caco-2 cells. *International journal of pharmaceutics*, 337(1), 169-177.
- Hsouna, A. B., Halima, N. B., Smaoui, S., & Hamdi, N. (2017). Citrus lemon essential oil: chemical composition, antioxidant and antimicrobial activities with its preservative effect against *Listeria monocytogenes* inoculated in minced beef meat. *Lipids in health and disease*, 16(1), 146.
- Hu, G. X., Pan, J. Q., Zhao, Z. E., and Huang, S. (2015). *Zingiber zhuxiense* (Zingiberaceae), a new species from Hubei, central China. In *Annales Botanici Fennici* (Vol. 52, No. 3–4, pp. 145-148). Finnish Zoological and Botanical Publishing Board.
- Huang, S. J., Chyau, C. C., Tsai, C. H., Chen, C. C., Mau, J. L., and Tsai, S. Y. (2015). Antioxidant Properties of Extracts from *Curcuma zedoaria* Rhizome. In *Advanced Materials Research* (Vol. 1120, pp. 920-925). Trans Tech Publications.
- Huetteman, C. A., & Preece, J. E. (1993). Thidiazuron: a potent cytokinin for woody plant tissue culture. *Plant cell, tissue and organ culture*, 33(2), 105-119.
- Hussain, J., Rehman, N. U., Al-Harrasi, A., Ali, L., Khan, A. L., & Albroumi, M. A. (2013). Essential oil composition and nutrient analysis of selected medicinal plants in Sultanate of Oman. *Asian Pacific Journal of Tropical Disease*, 3(6), 421-428.
- Hussey, G. (1986). Problems and prospects in the in vitro propagation of herbaceous plants. *Plant Tissue Culture and its Agricultural Application*, 113-123.
- Irshad, M., He, B., Liu, S., Mitra, S., Debnath, B., Li, M., and Qiu, D. (2017). In vitro regeneration of *Abelmoschus esculentus* L. cv. Wufu: Influence of anti-browning additives on phenolic secretion and callus formation frequency in explants. *Horticulture, Environment, and Biotechnology*, 58(5), 503-513.
- Isah, T. (2015). Adjustments to in vitro culture conditions and associated anomalies in plants. *Acta Biologica Cracoviensia s. Botanica*, 57(2), 9-28.
- Islam, M. A., Klopstech, K., and Jacobsen, H. J. (2004). Efficient Procedure for In vitro Microrhizome Induction in *Curcuma longa* L. (Zingiberaceae) – A Medicinal Plant of Tropical Asia. *Plant Tissue Cult.* 14(2), 123–134.
- Ismail, Z. (2003). Standardization of herbal products: A case study. Two and Half Day Course of Herbal and Phytochemical Processing, CEPP Short Course Notes. UTM Skudai, Malaysia, Johor, 7-9.

- Jain, S. M. (2001). Tissue culture-derived variation in crop improvement. *Euphytica*, 118(2), 153–166.
- Jantan, I. B., Yassin, M. S. M., Chin, C. B., Chen, L. L., and Sim, N. L. (2003). Antifungal activity of the essential oils of nine Zingiberaceae species. *Pharmaceutical biology*, 41(5), 392-397.
- Jitsopakul, N., Sangyojarn, P., Homchan, P., and Thammasiri, K. (2016, March). Micropropagation for conservation of Zingiberaceae in Surin province, Thailand. In *I International Symposium on Tropical and Subtropical Ornamentals 1167* (pp. 75-80).
- Joy, P. P., Thomas, J., Mathew, S., and Skaria, B. P. (1998). Zingiberaceous medicinal and aromatic plants. *Aromatic and Medicinal Plants Research Station, Odakkali, Asamanoor PO, Kerala, India*, 236-238.
- Kambaska, K. B., and Santilata, S. (2009). Effect of plant growth regulator on Micropropagation of ginger (*Zingiber officinale* Rosc.) cv-Suprava and Suruchi. *Journal of Agricultural Technology*, 5(2), 271-280.
- Kamboj, V. P. (2000). Herbal medicine. *Current science*, 78(1), 35-39.
- Kapai, V. Y., Kapoor, P., and Rao, I. U. (2010). *In vitro* Propagation for Conservation of Rare and Threatened Plants of India – A Review. *International Journal of Biological Technology*, 1(2), 1–14.
- Kapoor, L. D. (1990). *Handbook of Ayurvedic medicinal plants CRC Press Inc.*
- Karp, A. (1995). Somaclonal variation as a tool for crop improvement. *Euphytica*, 85(1-3), 295–302.
- Khamanashis D, and Muhammad, A.R. (2012). Analgesic and antimicrobial activities of *Curcuma zedoaria*. *International Journal of Pharmacy and Pharmaceutical Sciences*. Vol 4, Suppl 5.
- Khare, C. P. (2007). *Indian Medicinal Plants-An Illustrated Dictionary*. 1st Indian Reprint Springer (India) Pvt. Ltd., New Delhi, India, 28.
- Kim, H. ., Lee, J.W, and Kim, Y. (2011). Antimicrobial activity and antioxidant effect of *Curcuma longa*, *Curcuma aromatica* and *Curcuma zedoaria*. *Korean Journal of Food Preservation*, 18(2), 219-225.
- Kim, M., Cobbin, D., and Zaslowski, C. (2008). Traditional Chinese medicine tongue inspection: an examination of the inter- and intrapractitioner reliability for specific tongue characteristics. *The Journal of Alternative and Complementary Medicine*, 14(5), 527-536.

- Koarapatchaikol, K., Kanmarangkol, S., & Kaewraksa, J. K. (2017). In vitro plant regeneration via callus culture in turmeric (*Curcuma longa* L.). *Burapha Science Journal*, 22(1), 1-13.
- Kochuthressia, K. P., Britto, S. J., Raj, L. J. M., Jaseentha, M. O., & Senthilkumar, S. R. (2010). Efficient regeneration of *Alpinia purpurata* (Vieill.) K. Schum. plantlets from rhizome bud explants. *Int Res J Plant Sci*, 1(2), 43-47.
- Kozai, T., Xiao, Y., Nguyen, Q. T., Afreen, F., and Zobayed, S. M. (2005). Photoautotrophic (sugar-free medium) micropropagation systems for large-scale commercialization. *Propagation of ornamental plants*, 5(1), 23-34.
- Kumaran, A., and Karunakaran, R. J. (2007). *In vitro* antioxidant activities of methanol extracts of five *Phyllanthus* species from India. *LWT-Food Science and Technology*, 40(2), 344-352.
- Kusuma, S. A. F., Moelyono, M. W., and Hamka, R. (2017). Antifungal effect of *Curcuma zedoaria* ethanol extract and fractions against *aspergillus Niger*.
- Kusumastuti, M. Y., Bhatt, A., Indrayanto, G., and Keng, C. L. (2014). Effect of sucrose, benzylaminopurine and culture condition on *in vitro* propagation of *Curcuma xanthorrhiza* Roxb. and *Zingiber aromaticum* val. *Pakistan Journal of Botany*, 46(1), 280-289.
- Lai, E. Y., Chyau, C. C., Mau, J. L., Chen, C. C., Lai, Y. J., Shih, C. F., & Lin, L. L. (2004). Antimicrobial activity and cytotoxicity of the essential oil of *Curcuma zedoaria*. *The American journal of Chinese medicine*, 32(02), 281-290.
- Lal, J. (2012). Turmeric, Curcumin and Our Life: A Review. *Bulletin of Environment, Pharmacology and Life Sciences*, 1(7), 11-17.
- Lapornik, B., Prošek, M., and Wondra, A. G. (2005). Comparison of extracts prepared from plant by-products using different solvents and extraction time. *Journal of food engineering*, 71(2), 214-222.
- Lara, H. H., Ayala-Núñez, N. V., Turrent, L. D. C. I., and Padilla, C. R. (2010). Bactericidal effect of silver nanoparticles against multidrug-resistant bacteria. *World Journal of Microbiology and Biotechnology*, 26(4), 615-621.
- Larsen, K., Ibrahim, H., Khaw, S. H., & Saw, L. G. (1999). Gingers of peninsular Malaysia and Singapore. *Natural History Publications* (Borneo).
- Li, Y. M., Jia, M., Li, H. Q., Zhang, N. D., Wen, X., Rahman, K. and Qin, L. P. (2015). *Cnidium monnieri*: a review of traditional uses, phytochemical and ethnopharmacological properties. *The American journal of Chinese medicine*, 43(05), 835-877.

- Lim, Y.Y., and Murtijaya, J. (2007). Antioxidant properties of *Phyllanthus amarus* extracts as affected by different drying methods. *LWT-Food Science and Technology*, 40(9), 1664-1669.
- Lima, G. P. P., da Silva Campos, R. A., Willadino, L. G., Câmara, T. J., and Vianello, F. (2012). Polyamines, gelling agents in tissue culture, micropropagation of medicinal plants and bioreactors. *In Recent Advances in Plant in vitro Culture. InTech*.
- Lincy, A., and Sasikumar, B. (2010). Enhanced adventitious shoot regeneration from aerial stem explants of ginger using TDZ and its histological studies. *Turkish Journal of Botany*, 34, 21–29.
- Liu, Y., Roy, S. S., Nebie, R. H., Zhang, Y., and Nair, M. G. (2013). Functional food quality of *Curcuma caesia*, *Curcuma zedoaria* and *Curcuma aeruginosa* endemic to Northeastern India. *Plant foods for human nutrition*, 68(1), 72-77.
- Lobo, R., Prabhu, K. S., Shirwaikar, A., and Shirwaikar, A. (2009). *Curcuma zedoaria* Rosc. (white turmeric): a review of its chemical, pharmacological and ethnomedicinal properties. *Journal of Pharmacy and Pharmacology*, 61(1), 13-21.
- Loc, N. H., Duc, D. T., Kwon, T. H., and Yang, M. S. (2005). Micropropagation of zedoary (*Curcuma zedoaria* Roscoe) - a valuable medicinal plant. *Plant Cell, Tissue and Organ Culture*, 81(1), 119–122.
- Luthra, P. M., Singh, R., and Chandra, R. (2001). Therapeutic uses of *Curcuma longa* (turmeric). *Indian Journal of Clinical Biochemistry*, 16(2), 153-160.
- Maciel, N., and Criley, R. A. (2002). Morphology, growth and flowering behavior of *Curcuma zedoaria*. In *XXVI International Horticultural Congress: Elegant Science in Floriculture* 624 (pp. 111-116).
- Magner, L. N., & Kim, O. J. (2017). *A history of medicine*. CRC Press.
- Mahna, N., Vahed, S. Z., and Khani, S. (2013). Plant *in vitro* culture goes nano: nanosilver- mediated decontamination of ex vitro explants. *J Nanomed Nanotechol*, 4(161), 1.
- Makabe, H., Maru, N., and Kuwabara, A. (2006). Anti-inflammatory sesquiterpenes from *Curcuma zedoaria*. *Natural product research*, 20(7), 680-685.
- Maobe, M. A., Gitu, L., Gatebe, E., Rotich, H., and Box, P. O. (2012). Phytochemical analysis of phenol and flavonoid in eight selected medicinal herbs used for the treatment of diabetes, malaria and pneumonia in Kisii, Kenya. *Academic Journal of Cancer Research*, 5(2), 31-39.

- Marliani, L., Budiana, W., and Anandari, Y. (2017). The effect of extraction condition on the polyphenol content and antioxidant activity of *Curcuma zedoaria* (Christm.) Roscoe Rhizome. *Indonesian Journal of Pharmaceutical Science and Technology*, 4(2), 57-63.
- Marret, F. (2003). Origin and evolution of tropical rain forests, RJ Morley. Publisher John Wiley and Sons Ltd, Chichester 2000 (362 pp) ISBN 0-471-98326-8. *Journal of Quaternary Science*, 18(5), 466-467.
- Martin, K. W., and Ernst, E. (2003). Herbal medicines for treatment of bacterial infections: a review of controlled clinical trials. *Journal of Antimicrobial Chemotherapy*, 51(2), 241-246.
- Masoumian, M., Arbakariya, A., Syahida, A., and Maziah, M. (2011). Flavonoids production in *Hydrocotyle bonariensis* callus tissues. *Journal of MedicinPlants Research*, 5(9), 1564-1574.
- Matkowski, A. (2008). Plant *in vitro* culture for the production of antioxidants—a review. *Biotechnology advances*, 26(6), 548-560.
- Matsuda, H., Ninomiya, K., Morikawa, T., and Yoshikawa, M. (1998). Inhibitory effect and action mechanism of sesquiterpenes from zedoariae rhizoma on d-galactosamine/lipopolysaccharide-induced liver injury. *Bioorganic and Medicinal Chemistry Letters*, 8(4), 339-344.
- Matsuda, H., Morikawa, T., Ninomiya, K., and Yoshikawa, M. (2001). Hepatoprotective constituents from zedoariae rhizoma: absolute stereostructures of three new carabrane-type sesquiterpenes, curcumenolactones A, B, and C. *Bioorganic and medicinal chemistry*, 9(4), 909-916.
- Matsuda, H., Tewtrakul, S., Morikawa, T., Nakamura, A., and Yoshikawa, M. (2004). Anti-allergic principles from Thai zedoary: structural requirements of curcuminoids for inhibition of degranulation and effect on the release of TNF- $\alpha$  and IL-4 in RBL-2H3 cells. *Bioorganic and medicinal chemistry*, 12(22), 5891-5898.
- Mau, J. L., Lai, E. Y., Wang, N. P., Chen, C. C., Chang, C. H., and Chyau, C. C. (2003). Composition and antioxidant activity of the essential oil from *Curcuma zedoaria*. *Food Chemistry*, 82(4), 583-591.
- Medini, F., Fellah, H., Ksouri, R., and Abdelly, C. (2014). Total phenolic, flavonoid and tannin contents and antioxidant and antimicrobial activities of organic extracts of shoots of the plant *Limonium delicatulum*. *Journal of Taibah University for Science*, 8(3), 216-224.
- Merican, I. (2002). Traditional/complementary medicine: the way ahead. *Medical Journal of Malaysia*, 57(3), 261-265.

- Miachir, J. I., Romani, V. L. M., Amaral, A. F. D. C., Mello, M. O., Crocomo, O. J., and Melo, M. (2004). Micropropagation and callogenesis of *Curcuma zedoaria* Roscoe. *Scientia Agricola*, 61(4), 427–432.
- Miguel, M. G. (2010). Antioxidant activity of medicinal and aromatic plants. A review. *Flavour and Fragrance Journal*, 25(5), 291–312.
- Mishra, K., Ojha, H., and Chaudhury, N. K. (2012). Estimation of antiradical properties of antioxidants using DPPH assay: A critical review and results. *Food Chemistry*, 130(4), 1036–1043.
- Moghadamtousi, S. Z., Abdul Kadir, H., Hassandarvish, P., Tajik, H., Abubakar, S., and Zandi, K. (2014). A review on antibacterial, antiviral, and antifungal activity of curcumin. *BioMed Research International*, 2014, Article ID 186864.
- Mohanty, S., Panda, M. K., Subudhi, E., and Nayak, S. (2008). Plant regeneration from callus culture of *Curcuma aromatica* and *in vitro* detection of somaclonal variation through cytophotometric analysis. *Biologia Plantarum*, 52(4), 783- 786.
- Moradpour, M., Aziz, M. A., and Abdullah, S. N. A. (2016). Establishment of *invitro* culture of rubber (*Hevea brasiliensis*) from field-derived explants: effective role of silver nanoparticles in reducing contamination and browning. *J Nanomed Nanotechnol*, 7(375), 2.
- Mujumdar, A. M., Naik, D. G., Dandge, C. N., and Puntambekar, H. M. (2000). Antiinflammatory activity of *Curcuma amada* Roxb. in albino rats. *Indian journal of Pharmacology*, 32(6), 375-377.
- Mukunthan, K. S., Satyan, R. S., and Patel, T. N. (2017). Pharmacological evaluation of phytochemicals from South Indian Black Turmeric (*Curcuma caesia* Roxb.) to target cancer apoptosis. *Journal of Ethnopharmacology*, 209, 82-90.
- Müller, W. E., Batel, R., Schröder, H. C., and Müller, I. M. (2004a). Traditional and modern biomedical prospecting: Part I- the history. *Evidence-Based Complementary and Alternative Medicine*, 1(1), 71-82.
- Müller, W. E. G., Schröder, H. C., Wiens, M., Perović-Ottstadt, S., Batel, R., and Müller, I. M. (2004b). Traditional and Modern Biomedical Prospecting: Part II—the Benefits. *Evidence-Based Complementary and Alternative Medicine*, 1(2), 133–144.
- Muthu kumar, T., Christy, A. M. V., Mangadu, A., Malaisamy, M., Sivaraj, C., Arjun, P., Raaman, N., and Balasubramanian, K. (2012). Anticancer and antioxidant activity of *Curcuma zedoaria* and *Curcuma amada* rhizome extracts. *Journal of Academia and Industrial Research (JAIR)* 1(2), 91–96.

- Nadkarni, A. K. (1996). [Indian materia medica]; Dr. KM Nadkarni's Indian materia medica: with Ayurvedic, Unani-Tibbi, Siddha, allopathic, homeopathic, naturopathic and home remedies, *appendices and indexes*. 1 (Vol. 1). Popular Prakashan.
- Nascimento, G. G., Locatelli, J., Freitas, P. C., and Silva, G. L. (2000). Antibacterial activity of plant extracts and phytochemicals on antibiotic-resistant bacteria. *Brazilian journal of microbiology*, 31(4), 247-256.
- Nasirujjaman, K., Uddin, M. S., Zaman, S., & Reza, M. A. (2005). Micropropagation of turmeric (*Curcuma longa* Linn.) through in vitro rhizome bud culture. *Journal of Biological Sciences*, 5(4), 490-492.
- Navarro, D. D. F., de Souza, M. M., Neto, R. A., Golin, V., Niero, R., Yunes, R. A., and Cechinel Filho, V. (2002). Phytochemical analysis and analgesic properties of *Curcuma zedoaria* grown in Brazil. *Phytomedicine* 9(5), 427– 432.
- Nayak, S. (2000). In vitro multiplication and microrhizome induction in *Curcuma aromatica* Salisb. *Plant Growth Regulation*, 32(1), 41-47.
- Naz, S. H. A. G. U. F. T. A., Ilyas, S., Javad, S. U. M. E. R. A., & Ali, A. (2009). In vitro clonal multiplication and acclimatization of different varieties of turmeric (*Curcuma longa* L.). *Pak. J. Bot*, 41(6), 2807-2816.
- Naz, S., Jabeen, S., Ilyas, S., Manzoor, F., Aslam, F., and Ali, A. (2010). Antibacterial activity of *Curcuma longa* varieties against different strains of bacteria. *Pak J Bot*, 42(1), 455-62.
- Negi, P. S., Jayaprakasha, G. K., Jagan Mohan Rao, L., Sakariah, K. K. (1999). Antibacterial activity of turmeric oil: a byproduct from curcumin manufacture. *Journal of Agricultural and Food Chemistry* 47 (10), 4297–4300.
- Nejatzadeh-Barandozi, Fatemeh, Fariborz Darvishzadeh, and Ali Aminkhani. "Effect of nano silver and silver nitrate on seed yield of (*Ocimum basilicum* L.)." *Organic and medicinal chemistry letters* 4, no. 1 (2014): 11.
- Nghia, L. T., Tung, H. T., Huy, N. P., Luan, V. Q., and Nhut, D. T. (2017). THE Effects of Silver Nanoparticles on Growth of *Chrysanthemum orifolium* Ramat. cv."JIMBA" in different cultural systems. *Vietnam Journal of Science and Technology*, 55(4), 503.
- Niamsa, N., and Sittiwet, C. (2009). Antibacterial activity of *Curcuma longa* aqueous extract. *Journal of Pharmacology and Toxicology*, 4(4), 173-177.
- Palai, S. K., Rout, G. R., & Das, P. (1997). Micropropagation of ginger (*Zingiber officinale* Rosc.)-interaction of growth regulators and culture conditions. *Biotechnology of spices, medicinal and aromatic plants. India: Indian Society for Spices, Kerala*, 20-24.



- Panda, M. K., Mohanty, S., Subudhi, E., Acharya, L., & Nayak, S. (2007). Assessment of genetic stability of micropropagated plants of *Curcuma longa* L. by cytophotometry and RAPD analyses. *Int J Integr Biol*, 1(3), 189-195.
- Papas, A. M. (Ed.). (1998). *Antioxidant status, diet, nutrition, and health* (Vol. 9). CRC press.
- Parthasarathy, V., and Sasikumar, B. (2006). Biotechnology of *Curcuma*. CAB Reviews: Perspectives in Agriculture, *Veterinary Science, Nutrition and Natural Resources*, 1(020), 1–9.
- Pavallekoodi, G., & Sreeramanan, S. (2016). Micropropagation of ginger (*Zingiber officinale* var. *Rubrum*) using buds from microshoots. *Pak J Bot*, 48, 1153-1158.
- Perdana, F. (2017). Effect of benzil amino purin and sucrose on development of microrhizome and xanthorrhizol in tissue culture temulawak (*Curcuma xanthorrhiza* roxb.). *2nd ISEJ 2017 Abstract and Full Paper Submission System*, 1(1), 153-153.
- Pikulthong, V., Teerakathiti, T., Thamchaipenet, A., and Peyachoknagul, S. (2016). Development of somatic embryos for genetic transformation in *Curcuma longa* L. and *Curcuma mangga* Valetton and Zijp. *Agriculture and Natural Resources*, 50(4), 276-285.
- Pilzer, P. Z. (2010). *The New Wellness Revolution: How to Make a Fortune in the Next Trillion Dollar Industry*. John Wiley and Sons.
- Prajapati, N. D., Purohit, S. S., Sharma, A. K., and Kumar, T. (2003). A Handbook of Medicinal Plants. *Agrobios published company, 3rd edition, India, 353*.
- Prakosa, D. G., and Andayani, S. (2015). *In vitro* phytochemical and antibacterial activity test on temu putih extract *Curcuma zedoaria* against aeromonas hydrophila. *International Journal of Scientific and Technology Research*, 4(8), 36-37.
- Prathanturarug, S., Soonthornchareonnon, N., Chuakul, W., Phaidee, Y., and Saralamp, P. (2005). Rapid micropropagation of *Curcuma longa* using bud explants pre-cultured in thidiazuron-supplemented liquid medium. *Plant cell, tissue and organ culture*, 80(3), 347-351.
- Purkayastha, J., Nath, S. C., and Klinkby, N. (2006). Essential oil of the rhizome of *Curcuma zedoaria* (Christm.) Rosc. Native to northeast India. *Journal of Essential Oil Research*, 18(2), 154–155.
- Purseglove, J. W. (1975). *Persea Americana*. Tropical Crops: Dicotyledons. *John Wiley*, New York, 192-198.

- Putri, M. S. (2014). White Turmeric (*Curcuma Zedoaria*): Its Chemical Substance And The Pharmacological Benefits. *Jurnal Majority*, 3(7).
- Rahman, A. H. M. M., Jahan-E-Gulsan, S. M., & Naderuzzaman, A. T. M. (2014). Ethno-Gynecological Disorders of Folk Medicinal Plants Used by Santhals of Dinajpur District, Bangladesh. *Frontiers of Biological & Life Sciences*, 2(3), 62-66.
- Raihana, R., Faridah, Q. Z., Julia, A. A., Abdelmageed, A. H. A., & Kadir, M. A. (2011). In vitro culture of *Curcuma mangga* from rhizome bud. *Journal of Medicinal Plants Research*, 5(28), 6418-6422.
- Rao, C. H., & Rao, C. H. (2005). Agriculture, food security, poverty, and environment: essays on post-reform India. New Delhi, India: *Oxford University Press*.
- Rates, S. M. K. (2001). Plants as source of drugs. *Toxicon*, 39(5), 603-613.
- Ravindran, P. N., Babu, K. N., & Sivaraman, K. (2007). Turmeric: the genus *Curcuma*. *CRC press*.
- Reed, B. M., and Tanprasert, P. (1995). Detection and control of bacterial contaminants of plant tissue cultures. A review of recent literature. *Plant tissue culture and Biotechnology*, 1(3), 137-142.
- Ribeiro, I. G., De Castro, T. C., Gayer, C. R. M., Coelho, M. G. P., and Albarello, N. (2016). Phytochemical screening of field-grown plants and *in vitro* tissue culture of *Hovenia dulcis* Thunb. *Plant Cell Culture and Micro-propagation*, 6(2), 57-64.
- Roy, S., & Raychaudhuri, S. S. (2004). In vitro regeneration and estimation of curcumin content in four species of *Curcuma*. *Plant Biotechnology*, 21(4), 299-302.
- Ruffoni, B., Pistelli, L., Bertoli, A., & Pistelli, L. (2010). Plant cell cultures: bioreactors for industrial production. In *Bio-farms for nutraceuticals* (pp. 203-221). *Springer, Boston, MA*.
- Safitri, A., Batubara, I., and Khumaida, N. (2017). Thin layer chromatography fingerprint, antioxidant, and antibacterial activities of rhizomes, stems, and leaves of *Curcuma aeruginosa* Roxb. In *Journal of Physics: Conference Series* (Vol. 835, No. 1, p. 012014). IOP Publishing.
- Sahu, R., and Saxena, J. (2013). Screening of Total Phenolic and Flavonoid Content in Conventional and Non-Conventional Species of *Curcuma*. *Journal of Pharmacognosy and Phytochemistry*, 2(1).

- Saipriya, G., Kumaresan, R., Nayak, P. K., Venkatesan, K. A., Antony, M. P., & Kumar, T. (2017). Studies on the adsorption behavior of americium and europium on radiolytically degraded solvent impregnated resin containing neutral and acidic extractants. *Journal of Radioanalytical and Nuclear Chemistry*, 314(3), 2557-2568.
- Sakihama, Y., Cohen, M. F., Grace, S. C., and Yamasaki, H. (2002). Plant phenolic antioxidant and prooxidant activities: phenolics-induced oxidative damage mediated by metals in plants. *Toxicology*, 177(1), 67–80.
- Salisu, I. B., Abubakar, A. S., Sharma, M., and Pudake, R. N. (2014). Study of antimicrobial activity of nano silver (NS) in tissue culture media. *International Journal of Current Research and Review*, 6(13), 1.
- Salvi, N. D., George, L., & Eapen, S. (2002). Micropropagation and field evaluation of micropropagated plants of turmeric. *Plant Cell, Tissue and Organ Culture*, 68(2), 143-151.
- Sasikumar, B. (2005). Genetic resources of *Curcuma*: diversity, characterization and utilization. *Plant Genetic Resources: Characterization and Utilization*, 3(2), 230–251.
- Sathyagowri, S., and Seran, T. H. (2011). *In vitro* plant regeneration of ginger (*Zingiber officinale* Rosc.) with emphasis on initial culture establishment. *International Journal of Medicinal and Aromatic Plants*, 1(3), 195-202.
- Savitt, T. L. (2007). Race and Medicine in Nineteenth-and Early-twentieth-century America. *Kent University Press*, 2007. 453 p.
- Scalbert, A., Johnson, I. T., and Saltmarsh, M. (2005). Polyphenols: antioxidants and beyond. *The American journal of clinical nutrition* 81(1), 215S–217.
- Selvakkumar, C., Balakrishnan, A., & Lakshmi, B. S. (2007). Rapid *in vitro* micropropagation of *Alpinia officinarum* Hance., an important medicinal plant, through rhizome bud explants. *Asian J Plant Sci*, 6, 1251-1255.
- Senathilake, K. S., Karunanayake, E. H., Samarakoon, S. R., Tennekoon, K. H., and de Silva, E. D. (2016). Rhizome extracts of *Curcuma zedoaria* Rosc induce caspase dependant apoptosis via generation of reactive oxygen species in filarial parasite *Setaria digitata* *in vitro*. *Experimental parasitology*, 167, 50-60.
- Seo, W. G., Hwang, J. C., Kang, S. K., Jin, U. H., Suh, S. J., Moon, S. K., and Kim, C. H. (2005). Suppressive effect of *Zedoariae* rhizoma on pulmonary metastasis of B16 melanoma cells. *Journal of Ethnopharmacology*, 101(1-3), 249–257.

- Seran, T. H. (2013). *In vitro* propagation of ginger (*Zingiber officinale* Rosc.) through direct organogenesis: A review. *Pakistan Journal of Biological Sciences*, 16(24), 1826-1835.
- Shagufta, B., Sivakumar, M., Kumar, S., Agarwal, R. K., Bhilegaonkar, K. N., Kumar, A., & Dubal, Z. B. (2017). Antimicrobial resistance and typing of *Salmonella* isolated from street vended foods and associated environment. *Journal of food science and technology*, 54(8), 2532-2539.
- Shah, R., Patel, A., Shah, M., and Peethambaran, B. (2015). Anti-acne activity of *Achillea* "Moonshine" petroleum ether extract. *Journal of Medicinal Plants Research*, 9(27), 755–763.
- Shahzad, A., Sharma, S., Parveen, S., Saeed, T., Shaheen, A., Akhtar, R., ... & Ahmad, Z. (2017). Historical perspective and basic principles of plant tissue culture. In *Plant biotechnology: principles and applications* (pp. 1-36). Springer, Singapore.
- Sharma, T. R., and Singh, B. M. (1995). *In vitro* microrhizome production in *Zingiber officinale* Rosc. *Plant cell reports*, 15(3-4), 274-277.
- Sharma R. A, Gescher A. J, and Steward, W.P. (2005). Curcumin: The story so far, *European journal of cancer*, 41(13), 1955-1968.
- Sharmin, S. A., Alam, M. J., Sheikh, M. M. I., Zaman, R., Khalekuzzaman, M., Mondal, S. C., & Alam, I. (2013). Micropropagation and antimicrobial activity of *Curcuma aromatica* Salisb., a threatened aromatic medicinal plant. *Turkish Journal of Biology*, 37(6), 698-708.
- Shirgurkar, M. V., John, C. K., & Nadgauda, R. S. (2001). Factors affecting *in vitro* microrhizome production in turmeric. *Plant Cell, Tissue and Organ Culture*, 64(1), 5-11.
- Shiva, V. (1997). The turmeric patent is just the first step in stopping biopiracy. *Third World Resurgence*, 2-4.
- Shretha, I., Joshi, N. (1993). Medicinal plants of the Lele Village of Latipur District, Nepal. *International Journal of Pharmacognosy* 31(2), 130–134.
- Silva, G. L., Lee, I. S., and Kinghorn, A. D. (1998). Special problems with the extraction of plants. In *Natural Products Isolation*. (Vol. 4, pp. 343–363). Humana Press.
- Simoh, S., and Zainal, A. (2015). Chemical Profiling of *Curcuma aeruginosa* Roxb. Rhizome using different Techniques of Solvent Extraction. *Asian Pacific Journal of Tropical Biomedicine*, 5(5), 412–417.

- Singh, J., Singh, V., Shukla, S., & Rai, A. K. (2016). Phenolic content and antioxidant capacity of selected cucurbit fruits extracted with different solvents. *J Nutr Food Sci*, 6(565), 2.
- Sirirugsa, P., (1999). Thai Zingiberaceae: Species Diversity and Their Uses. *Pure and Applied Chemistry*, 70(11), 23–27.
- Skendi, A., Irakli, M., and Chatzopoulou, P. (2017). Analysis of phenolic compounds in Greek plants of Lamiaceae family by HPLC. *Journal of Applied Research on Medicinal and Aromatic Plants*, 6, 62-69.
- Souri, E., Amin, G., Farsam, H., Jalalizadeh, H., and Barezi, S. (2010). Screening of Thirteen Medicinal Plant Extracts for Antioxidant Activity. *Iranian Journal of Pharmaceutical Research*, 149–154.
- Srivastava, N., Sharma, V., Kamal, B., Dobriyal, A. K., & Jadon, V. S. (2010). Advancement in research on Aconitum sp.(Ranunculaceae) under different area: a review. *Biotechnology*, 9(4), 411-427.
- Stanly, C., Bhatt, A., and Keng, C. L. (2010). A comparative study of *Curcuma zedoaria* and Zingiber zerumbet plantlet production using different Micropropagation systems. *Journal of Biotechnology*, 9(28), 4326–4333.
- Sumathi, S., Iswariya, G. T., Sivaprabha, B., Dharani, B., Radha, P., and Padma, P.R. (2013). Comparative Study of Radical Scavenging Activity and Phytochemical Analysis of Fresh and. *International Journal of Pharmaceutical Sciences and Research*, 4(3), 1069–1073.
- Sun, W., Wang, S., Zhao, W., Wu, C., Guo, S., Gao, H., and Chen, X. (2017). Chemical constituents and biological research on plants in the genus *Curcuma*. *Critical reviews in food science and nutrition*, 57(7), 1451-1523.
- Sunitibala, H., Damayanti, M., & Sharma, G. J. (2001). In vitro propagation and rhizome formation in *Curcuma longa* Linn. *CYTOBIOS-CAMBRIDGE-*, 71-82.
- Syamsir, D. R., Sivasothy, Y., Hazni, H., Abdul Malek, S. N., Nagoor, N. H., Ibrahim, H., and Awang, K. (2017). Chemical Constituents and Evaluation of Cytotoxic Activities of *Curcuma zedoaria* (Christm.) Roscoe Oils from Malaysia and Indonesia. *Journal of Essential Oil Bearing Plants*, 20(4), 972-982.
- Tang, J. L., Zhan, S. Y., and Ernst, E. (1999). Review of randomised controlled trials of traditional Chinese medicine. *Bmj*, 319(7203), 160-161.
- Tariq, S., Imran, M., Mushtaq, Z., and Asghar, N. (2016). Phytopreventive antihypercholesterolemic and antilipidemic perspectives of zedoary (*Curcuma zedoaria* Roscoe.) herbal tea. *Lipids in health and disease*, 15(1), 39.

- Tholkappiyavathi, K., Selvan, K. M., Nayanila, S. K., Yoganandam, G. P., and opal, V. (2013). A Concise Review on *Curcuma zedoaria*. *International Journal of Phytotherapy*, 3(1), 1–4.
- Thomas, T. D. (2007). Pretreatment in thidiazuron improves the *in vitro* shoot induction from leaves in *Curculigo orchioides* Gaertn an endangered medicinal plant. *Acta Physiologiae Plantarum*, 29(5), 455-461.
- Tiphara, P., Sangvanich, P., Macth, M., and Petsom, A. (2007). Mannose-binding Lectin from *Curcuma zedoaria* Rosc. *Journal of Plant Biology*, 50(2): 167-173.
- Tiwari, P., Kumar, B., Kaur, M., Kaur, G., and Kaur, H. (2011). Phytochemical screening and extraction: a review. *Internationale pharmaceutica sciencia*, 1(1), 98-106.
- Tschanz, D. W. (2003). Arab roots of European medicine. *Heart Views*, 4(2), 9.
- Tuntiwachwuttikul, P. I. T. T. A. Y. A., Pancharoen, O. R. A. S. A., Kanjanapothi, D., Panthong, A., Taylor, W. C., & Reutrakul, V. (1986). *Zingiberaceous plants*.
- Turkmen, N., Sari, F., and Velioglu, Y. S. (2006). Effects of extraction solvents on concentration and antioxidant activity of black and black mate tea polyphenols determined by ferrous tartrate and Folin–Ciocalteu. *Food chemistry*, 99(4), 835–841.
- Tushar, Basak, S., Sarma, G. C., and Rangan, L. (2010). Ethnomedical uses of Zingiberaceous plants of Northeast India. *Journal of Ethnopharmacology*, 132(1), 286–96.
- Tu, Y. (2011). The discovery of artemisinin (qinghaosu) and gifts from Chinese medicine. *Nature medicine*, 17(10), 1217.
- Tyagi, R. K., Agrawal, A., Mahalakshmi, C., Hussain, Z., and Tyagi, H. (2007). Low- cost media for *in vitro* conservation of turmeric (*Curcuma longa* L.) and genetic stability assessment using RAPD markers. *In vitro Cellular and Developmental Biology-Plant*, 43(1), 51-58.
- Tyagi, R. K., Yusuf, A., Dua, P., and Agrawal, A. (2004). *In vitro* Plant Regeneration and Genotype Conservation of Eight Wild Species of *Curcuma*. *Biologia Plantarum*, 48(1), 129–132.
- Ullah, H. A., Zaman, S., Juhara, F., Akter, L., Tareq, S. M., Masum, E. H., and Bhattacharjee, R. (2014). Evaluation of antinociceptive, in-vivo and *in vitro* anti-inflammatory activity of ethanolic extract of *Curcuma zedoaria* rhizome. *BMC complementary and alternative medicine*, 14(1), 346.
- Van Wyk, B. E. (2008). A broad review of commercially important southern African medicinal plants. *Journal of ethnopharmacology*, 119(3), 342-355.

- Van Wyk, B. E., & Wink, M. (2017). Medicinal plants of the world (No. Ed. 2). *CABI*.
- Victório, C. P. (2011). Therapeutic value of the genus *Alpinia*, Zingiberaceae. *Revista Brasileira de Farmacognosia*, 21(1), 194–201.
- Victório, C. P., Kuster, R. M., and Lage, C. L. S. (2009). Detection of flavonoids in *Alpinia purpurata* (Vieill.) K. Schum. leaves using high-performance liquid chromatography. *Revista Brasileira de Plantas Mediciniais*, 11(2), 147–153.
- Wadood, A., Ghufran, M., Jamal, S. B., Naeem, M., Khan, A., Ghaffar, R., and Asnad, C. (2013). Phytochemical analysis of medicinal plants occurring in local area of Mardan. *Biochem Anal Biochem*, 2(4), 1-4.
- Wayne, P. A. (2011). Clinical and laboratory standards institute. Performance standards for antimicrobial susceptibility testing. *Twenty-Second Informational Supplement. CLSI Document M100-S122*.
- WHO (1993). Summar 9 WHO guidelines for the assessment of herbal medicines. *Herbal Grom* 28, 13–14.
- Wilson, B., Abraham, G., Manju, V. S., Mathew, M., Vimala, B., Sundaresan, S., and Nambisan, B. (2005). Antimicrobial activity of *Curcuma zedoaria* and *Curcuma malabarica* tubers. *Journal of Ethnopharmacology*, 99(1), 147–151.
- Xiong, C., Li, Q., Li, S., Chen, C., Chen, Z., and Huang, W. (2017). *In vitro* antimicrobial activities and mechanism of 1-octen-3-ol against food-related bacteria and pathogenic fungi. *Journal of Oleo Science*, 66(9), 1041-1049.
- Yoshioka, T., Fujii, E., Endo, M., Wada, K., Tokunaga, Y., Shiba, N., Hohsho, H., Shibuya, H., and Muraki, T. (1998). Anti-inflammatory potency of dehydrocurdione, a zedoary-derived sesquiterpene. *Inflammation Research* 47 (12), 476–481.
- Yosuf, N., A., Khalid, N., and Ibrahim, H. (2007). Establishment of tissue culture for selected medicinal *Curcuma*. *Malaysian Journal of Science*, 26(1), 85–91.
- Zhang, A., Sun, H., and Wang, X. (2014). Potentiating therapeutic effects by enhancing synergism based on active constituents from traditional medicine. *Phytotherapy research*, 28(4), 526-533.
- Zhang, H. Y., Yang, D. P., and Tang, G. Y. (2006). Multipotent antioxidants: from screening to design. *Drug Discovery Today*, 11(15-16), 749–754.
- Zhang, S., Liu, N., Sheng, A., Ma, G., and Wu, G. (2011). Direct and callus-mediated regeneration of *Curcuma soloensis* Valetton (Zingiberaceae) and ex vitro performance of regenerated plants. *Scientia horticultruae*, 130(4), 899-905.

Zhao, X., Hong, Y., & Drlica, K. (2014). Moving forward with reactive oxygen species involvement in antimicrobial lethality. *Journal of Antimicrobial Chemotherapy*, 70(3), 639-642.

Zieliński, H., and Kozłowska, H. (2000). Antioxidant activity and total phenolics in selected cereal grains and their different morphological fractions. *Journal of Agricultural and Food Chemistry*, 48(6), 2008-2016.

Złotek, U., Mikulska, S., Nagajek, M., and Świeca, M. (2016). The effect of different solvents and number of extraction steps on the polyphenol content and antioxidant capacity of basil leaves (*Ocimum basilicum* L.) extracts. *Saudi journal of biological sciences*, 23(5), 628-633.

Zuraida, A. R., Sabrina, E., Shukri, M., Razali, M., and Norma, H. (2015). In vitro Micropropagation of a Valuable Medicinal Plant, *Piper crocatum*. *Int. J. Pure App. Biosci*, 3(3), 10-16.



## BIODATA OF STUDENT

Khalid Abubaker Alghannay, was born on August 15, 1969 in Sebha city located in the southern province of Libya. He had obtained a high school certificate in basic sciences, Biology Division in 1988, from Brak Secondary School, and then obtained a Bachelor of Agricultural Sciences, Department of Plant Production, Sebha University in the 1993. He completed his specialization in the field of horticulture by being able to obtain a master's in agricultural sciences, by studying the nutrition of evergreen Orchids (fruit trees), specifically in 2001, from University of Tripoli, where his primary interests focused on foliar fertilization and its impact on the productivity of grape vines and their natural characteristics. He derived his experiences through his work as a faculty staff member at a university of Sabha, and his skills expanded by supervising some graduation research at that time. Currently, he is about to complete his PhD from the esteemed UBM University, Faculty of Agriculture, Department of Agriculture Technology, which focused on devising a sterilization protocol for one of the commonly used ginger varieties in Malaysia and developing a method for its propagation through tissue culture technology. His study also included surveying the methods of extracting some phytochemicals and exploring the extent of their effect on inhibiting the growth of some pathogenic bacteria.



## UNIVERSITI PUTRA MALAYSIA

### STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

ACADEMIC SESSION : Second Semester 2020/2021

**TITLE OF THESIS / PROJECT REPORT :**

IN VITRO REGENERATION AND EVALUATION OF PHYTOCHEMICAL AND ANTIMICROBIAL  
ACTIVITY OF KUNYIT PUTIH [*Curcuma zedoaria* (Christm.) Roscoe]

**NAME OF STUDENT:** KHALID ABUBAKER ALGHANNAY

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

1. This thesis/project report is the property of Universiti Putra Malaysia.
2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as :

\*Please tick (✓)

**CONFIDENTIAL**

(Contain confidential information under Official Secret Act 1972).

**RESTRICTED**

(Contains restricted information as specified by the organization/institution where research was done).

**OPEN ACCESS**

I agree that my thesis/project report to be published as hard copy or online open access.

This thesis is submitted for :

**PATENT**

Embargo from \_\_\_\_\_ until \_\_\_\_\_  
(date) (date)

**Approved by:**

\_\_\_\_\_  
(Signature of Student)  
New IC No/ Passport No.:

Date :

\_\_\_\_\_  
(Signature of Chairman of Supervisory Committee)  
Name:

Date :

**[Note : If the thesis is CONFIDENTIAL or RESTRICTED, please attach with the letter from the organization/institution with period and reasons for confidentiality or restricted. ]**