

UNIVERSITI PUTRA MALAYSIA

PROTOCOL DEVELOPMENT FOR MICROPROPAGATION OF Phyllanthus niruri linn. (DUKUNG ANAK) VIA IN VITRO CULTURE

ANIS SURAYA BINTI AZAL

FP 2021 46



PROTOCOL DEVELOPMENT FOR MICROPROPAGATION OF Phyllanthus niruri Linn. (DUKUNG ANAK) VIA IN VITRO CULTURE



By

ANIS SURAYA BINTI AZAL

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

June 2021

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

PROTOCOL DEVELOPMENT FOR MICROPROPAGATION OF Phyllanthus niruri Linn. (DUKUNG ANAK) VIA IN VITRO CULTURE

By

ANIS SURAYA BINTI AZAL

June 2021

Chairman : Mohd Hakiman Mansor, PhD Faculty : Agriculture

Phyllanthus niruri, commonly known as Dukung Anak or Gale of Stone, is an herbaceous plant belonging to the family Phyllanthaceae that has commonly been distributed in tropical countries, including Malaysia. Even though this plant is considered a problematic weed for farmers, it has been used traditionally to treat various ailments such as diabetes, flu, and cough. Overexploitation in Malaysia, on the other hand, has resulted in wide destruction of this plant from its natural habitat. Besides, there is no information on the production of *P. niruri* in Malaysia. This study has provided a solution to this problem through *in vitro* technology approach. Specifically, this study's objectives were to study the growth efficiency of *P. niruri* with different surface sterilants, to study in vitro growth responses of *P. niruri* towards different growth factors, and to determine the factors affecting in vitro rooting and acclimatization of P. niruri. P. niruri explant's nodal segment was prepared for surface sterilization using Clorox® and nanosilver with 10, 20, 30 (% and ppm, respectively). The aseptic culture was established with the lowest percentage of contamination, 4.44% using 30% Clorox[®]. The explant from the previous experiment was multiplied by different plant growth factors, which were different types of basal media and its strength, different cytokinin and its concentrations, different carbon sources and its strength, and different initial pH of basal media. Murashige and Skoog (MS) media and Gamborg B5 (B5) with the strength of half, full and double were used in the first experiment. Full-strength MS media produced a decent number of shoots with 2.33 shoots, 3.11 cm length of shoots and 27.91 number of leaves. In a subsequent experiment, MS media was supplemented with different types of cytokinin, which were 6-benzylaminopurine (BAP), kinetin (Kn), zeatin (Zn), and 2-isopentenyl adenine (2iP) with different concentrations of 2.5, 5.0, 7.5, and 10μ M. The best treatment was obtained from MS media without cytokinin (control), which produced 5.00 number of shoots, 3.68 cm length of shoot, and 27.33 number of leaves. In the next experiment, basal media were manipulated with different carbon sources like sucrose, fructose, and glucose at different concentrations of 20, 25, 30, 35, and 40 g/L. The result showed the explant in media enriched with 30 g/L of sucrose has the highest number of shoots with an average of 3.2 shoots and shoot elongation of 2.27 cm. Lastly, the initial pH media was tested at 5.2, 5.4, 5.6, 5.8, 6, and 6.2. The highest number of shoots was recorded at a pH level of 5.6 (2.93). In the last experiment, the explant was used for root induction using different auxins at different concentrations. There were indole-3-butyric acid (IBA), 1-naphthalene acetic acid (NAA), and indole-3acetic acid (IAA) at 1.25, 2.5, and 5 µM, respectively. Media with 2.5 µM IBA produced the highest number of the shoot and the most extended length of the root with 17.92 shoots and 1.29 cm root length. Lastly, the explant was acclimatized ex vitro in different potting media; peat moss: cocopeat, peat moss: vermiculite, peat moss: perlite at a ratio of 1:1 w/w and mixed of all media peat moss: cocopeat: vermiculite: perlite at a ratio of 1:1:1:1 w/w. The plantlet in potting media consisting of peat moss: cocopeat (1:1 ratio) showed the best performance with 88% plant survival. In conclusion, protocol for micropropagation of *P. niruri* has been developed in this study.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

PEMBANGUNAN PROTOKOL BAGI MIKROPROPAGASI Phyllanthus niruri Linn. (DUKUNG ANAK) MELALUI KULTUR IN VITRO

Oleh

ANIS SURAYA BINTI AZAL

Jun 2021

Pengerusi : Mohd Hakiman Mansor, PhD Fakulti : Pertanian

Phyllanthus niruri, dikenali sebagai Dukung Anak atau Gale of Stone ialah satu pokok herba dari keluarga Phyllanthaceae yang kebanyakan tumbuh di negara tropika termasuk Malaysia. Walaupun pokok ini dikenali sebagai rumpai yang mengundang masalah di kalangan petani, pokok ini telah digunakan secara tradisional untuk merawat pelbagai penyakit seperti kencing manis, selesema dan batuk. Oleh kerana pokok ini telah dieksploitasi secara berlebihan, ia menyebabkan kemusnahan pokok ini di alam semulajadi. Tambahan pula, tiada sebarang maklumat mengenai pengeluaran herba ini di Malaysia. Justeru, kajian ini dicadangkan untuk menyelesaikan masalah berkenaan dengan menggunakan pendekatan kaedah teknologi in vitro. Secara khasnya, matlamat kajian ini adalah untuk mengkaji kecekapan pertumbuhan P. niruri dengan pensteril permukaan yang berbeza, untuk mengkaji tindak balas pertumbuhan secara in vitro P. niruri terhadap faktor pertumbuhan yang berbeza dan untuk menentukan faktor yang memberi kesan terhadap pengakaran in vitro dan aklimatisasi pokok P. niruri. Segmen nod pokok P. niruri telah digunakan untuk pensterilan permukaan menggunakan Clorox[®] dan nanosilver dengan kepekatan 10, 20, 30 (% dan ppm, masing-masing). Kultur aseptik telah dihasilkan dengan peratusan kontaminasi yang terendah, 4.44% menggunakan 30% Clorox®. Eksplan daripada eksperimen sebelumnya digandakan dengan faktor tumbesaran tumbuhan yang berbeza iaitu jenis media basal dan kepekatannya yang berbeza, sitokinin dan kepekatannya yang berbeza, sumber karbon dan kepekatannya yang berbeza dan nilai pH awal media basal yang berbeza. Di dalam eksperimen pertama, media Murashige and Skoog (MS) dan Gamborg B5 (B5) dengan kepekatan separuh, penuh dan seganda telah digunakan. Hasil kajian menunjukkan media MS dengan kepekatan penuh menghasilkan bilangan pucuk iaitu 2.33 pucuk, 3.11 sm kepanjangan pucuk dan 27.91 jumlah daun. Dalam eksperimen seterusnya, media MS telah dibekalkan

dengan jenis sitokinin yang berbeza iaitu 6-benzilaminopurina (BAP), kinetin (Kn), zeatin (Zn) dan 2-isopentenil adenina (2iP) dengan kepekatan yang berbeza iaitu 2.5, 5.0, 7.5 dan 10 μM. Hasil terbaik yang didapati ialah daripada media MS yang dibekalkan tanpa sebarang sitokinin (kawalan), iaitu menghasilkan 5.00 bilangan pucuk dan menghasilkan 3.68 sm panjang pucuk bersama 27.33 bilangan daun. Dalam eksperimen berikutnya, media basal dimanipulasi dengan sumber karbon yang berbeza seperti suckrosa, fruktosa dan glukosa pada kepekatan yang berbeza iaitu 20, 25, 30, 35 dan 40 g/L. Keputusan mendapati bahawa eksplan di dalam media yang dibekalkan dengan 30 g/L sukrosa mempunyai bilangan pucuk tertinggi dengan nilai purata 3.2 pucuk dengan kepanjangan pucuk yang tertinggi iaitu 2.27 sm. Akhirnya, nilai awal pH bagi media basal telah dikaji pada nilai 5.2, 5.4, 5.6, 5.8, 6 dan 6.2. Bilangan pucuk yang tertinggi dicatatkan pada nilai pH 5.6 iaitu 2.93 pucuk. Pada eksperimen terakhir, eksplan digunakan untuk induksi akar menggunakan jenis auxin dan kepekatannya yang berbeza. Terdapat asid indol-3-butirik (IBA), asid 1-naftalenaasetik (NAA) dan asid indol-3-asetik (IAA) pada 1.25, 2.5 dan 5 μM, masing-masing. Media yang mengandungi 2.5 μM IBA menghasilkan bilangan pucuk dan kepanjangan pucuk yang tertinggi iaitu 17.92 dan 1.29 sm, masing-masing. Seterusnya, eksplan diaklimatisasi secara ex vitro di dalam media pengerasan berbeza; lumut gambut: gambut coco, lumut gambut: vermikulit, lumut gambut: perlit pada nisbah 1:1 dan campuran ke semua media lumut gambut: gambut coco: vermikulit: perlit pada ratio 1:1:1:1. Pokok di dalam media yang mengandungi lumut gambut: gambut coco (ratio 1:1) menunjukkan prestasi yang terbaik dengan 88% peratus hidup. Kesimpulannya, protokol untuk membiakkan P. niruri secara mikropropagasi telah dibangunkan dalam kajian ini.

ACKNOWLEDGEMENTS

All praises and thanks due to Allah Almighty for his Mercy and Grace.

I would like to express my gratitude to the following people who helped me a lot in order for me to complete my master's study, especially my supervisor Dr. Mohd Hakiman Mansor, for his continuous support and kindness, criticism and encouragement, patience, and guidance throughout my journey and also Dr. Azizah Misran for her invaluable support and advice.

My most profound appreciation goes to my family, especially my parents, Mr. Azal Manas and Mrs. Foziyah Ismail, and my siblings, Anis Baidura Azal and Mohd Herwan Azal, for their prayers and support. They have been showering me with tenderness, love, and care.

I am wholly indebted to my dear friends who were always there for me in times of hardship during my postgrad journey, Nurul Syafiqah Zulkifli, Nor Elmira Sherryna Gazali, Muhammad Ammar Abdul Rahman, my tissue culture lab mates, and my Sumandak girl gang. I also would like to thank all staff in the Department of Crop Science, Faculty of Agriculture, especially Mr. Helmy, who always help when needed. Finally, I thank Universiti Putra Malaysia for providing the scholarship and grant for this study.

Last but not least, I want to thank me. I want to thank me for believing in me. I want to thank me for doing all of this hard work. I want to thank me for having no days off. I want to thank me for never quitting. I want to thank me for always being a giver and try to give more than i receive. I want to thank me for try to do more right than wrong. I want to thank me for just being me at all times.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Mohd Hakiman Mansor, PhD

Senior Lecturer Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Azizah Misran, PhD Senior Lecturer Faculty of Agriculture Universiti Putra Malaysia (Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date: 14 October 2021

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _	Date:
0	

Name and Matric No.: Anis Suraya binti Azal (GS48630)

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: Name of Chairman of Supervisory Committee: Dr. Mohd Hakiman Mansor Signature: Name of Member of Supervisory Committee: Dr. Azizah Misran

TABLE OF CONTENTS

			Page
ABSTRACT			i
ABSTRAK			iii
ACKNOWL	EDGEMI	ENTS	v
APPROVAL			vi
DECLARAT	ION		viii
LIST OF TA	BLES		xiv
LIST OF FIG	JURES		xv
LIST OF AB	BREVIA	TIONS	xvi
CHAPTER			
1	INTR	ODUCTION	1
	1.1	Background of study	1
	1.2	Problem statements	2
	1.3	Objectives of study	3
2		RATURE REVIEW	4
	2.1	Genus Phyllanthus	4
	2.2	Phyllanthus niruri	4
	2.3	Medicinal benefits of <i>P. niruri</i>	5
	2.4	Bioactive compounds of <i>P. niruri</i>	6
	2.5	Cultivation of <i>P. niruri</i>	7
	2.6	Micropropagation	8
		2.6.1 Culture initiation and multiplication	10
		2.6.2 Surface sterilization	11
		 2.6.2 Surface sterilization 2.6.3 Basal medium 2.6.4 Carbon sources 	13 14
			14 15
		2.6.5 pH value2.6.6 Plant growth regulators	15 15
		2.6.6.1 Cytokinin	15 16
		2.6.6.2 Auxin	10
		2.6.7 <i>In vitro</i> rooting	17
		2.6.6 Acclimatization	18
			10
3	GENI	ERAL METHODOLOGY	19
	3.1	Location	19
	3.2	Materials and methods	19
		3.2.1 Plant materials	19
		3.2.2 Preparation of culture medium	19
	3.3	Preparation of plant growth regulators (PGRs)	20

	3. 3.			erilization of explants on of aseptic explant	20 20
4	EF			ERENT CHEMICAL STERILANTS	21
	0	N SUI	RFACE S	TERILIZATION OF <i>Phyllanthus niruri</i>	
	4.		Introduct		21
	4.	2	Materials	and methods	22
	4.	3	Treatmen		22
			4.3.1	Data collection	23
			4.3.2	Experimental design and statistical analysis	23
	4.	4	Results ar	nd discussion	24
	4.	5	Conclusio	on	26
5	Π	N VITF	RO GROV	VTH RESPONSES OF Phyllanthus	27
	n	iruri T	' <mark>OWAR</mark> D	S DIFFERENT PLANT GROWTH	
	F	ACTO	-		
	5.		Introduct		27
	5.	2	Materials	and methods	28
			5.2.1	Experiment 1: Effect of different basal	28
				media and its strength on Phyllanthus	
				niruri	
			5.2.2	Treatments of experiment 1	28
			5.2.3	Data collections	29
	5.			ent 2: Effect of different cytokinin and its	29
			concentra	tions on <i>Phyllanthus niruri</i>	
			5.3.1	Treatments of experiment 2	29
			5.3.2	Data collections	30
	5.	4	Experime	ent 3: Effect of different carbon sources	30
			and its str	cengths on Phyllanthus niruri	
			5.4.1	Treatments of experiment 3	30
			5.4.2	Data collections	30
	5.			ent 4: Effect of different initial pH values	30
			of basal n	nedia on Phyllanthus niruri	
			5.5.1	Treatments of experiment 4	30
			5.5.2	Data collections	31
	5.		Experime	ntal design and statistical analysis	31
	5.	7	Results a	nd discussion	31
			5.7.1	Growth responses of <i>P. niruri</i> as	31
				influenced by different basal media	
				and its strengths	
			5.7.2	Growth responses of <i>P. niruri</i> as	33
				influenced by cytokinin and its	
				concentrations	
		ļ	5.7.3	Growth responses of P. niruri	36
				as influenced by different	
				carbon sources and its	

xi

			strengths	
		5.7.4	Growth responses of <i>P. niruri</i> as	38
			influenced by different initial pH	
			values of basal media	
	5.8	Conclusion		40
	0.0	conclusion		10
6	EVALU	UATION O	N EFFECT OF DIFFERENT TYPES	41
	AND (CONCENTI	RATION OF AUXINS ON IN VITRO	
	ROOT	ING AND	ACCLIMATIZATION OF Phyllanthus	
	niruri			
	6.1	Introducti	on	41
	6.2	Materials	and methods	42
		6.2.1	Experiment 1: Effect of different	42
			auxin and its concentrations on in	
			<i>vitro</i> rooting of <i>P. niruri</i>	
		6.2.2	Treatments of experiment 1	42
		6.2.3	Data collections	42
	6.3	Materials	and methods	42
		6.3.1	Experiment 2: Effect of different	42
			potting media on acclimatization of P.	
			niruri	
		6.3.2 Treat	tments	43
			collections	43
	6.4	Experimen	ntal design and statistical analysis	43
	6.5		d discussion	43
		6.5.1	In vitro rooting of P. niruri influenced	43
			by different auxins and its	
			concentrations	
		6.5.2	Acclimatization of <i>P. niruri</i>	46
	6.6	Conclusio	n	49
-	CONC		ND RECONCIEND A TIONS FOR	50
7		LUSION AI RE RESEAR	ND RECOMMENDATIONS FOR	50
	7.1	Conclusio		50
	7.1 7.2			
	1.2	Recomme	ndation	50
REFEREN	NCES			52
APPEND				68
BIODAT		UDENT		89
LIST OF				90
				20

xii

6

LIST OF TABLES

Table		Page
2.1	Botanical classification of Phyllanthus niruri	5
4.1	Different chemical sterilants and its concentrations used in surface sterilization experiment	27
4.2	Layout of experimental unit	31
4.3	Effect of different chemical sterilant and concentrations towards the percentage of contamination (%) of <i>P. niruri</i>	33
5.1	Different types of basal media and its strengths used in experiment 1	35
5.2	Effect of different basal media and its strength towards the number of shoots, length of the shoot (cm), and number of leaves of <i>P. niruri</i>	38
5.3	Effect of different cytokinin and its concentrations towards the number of shoots, length of the shoot (cm), and number of leaves of <i>P. niruri</i>	41
5.4	Effect of different carbon sources and its strengths towards the number of shoots, length of the shoot (cm), and number of leaves of <i>P. niruri</i>	43
5.5	Effect of different initial pH values of basal media towards the number of shoots, length of the shoot (cm), and number of leaves of <i>P. niruri</i>	45
6.1	Different potting media mixture used in acclimatization stage	50
6.2	Effect of different auxins and its concentrations towards the number of roots and length of root (cm) of <i>P. niruri</i>	52
6.3	Effect of different potting media mixture on the percentage of survived plantlet (%) during acclimatization of <i>P. niruri</i>	56

LIST OF FIGURES

Figure		Page
2.1	A) <i>P. niruri</i> plant, B) fruits and, C) flowers beneath the leaves	6
2.2	Example of <i>P. niruri</i> end products available in market	7
2.3	Chemical structure of phyllanthin and hypophyllanthin of <i>P. niruri</i>	8
2.4	Flow chart of plant micropropagation procedure	12
4.2	A) Inoculated nodal segment of <i>P. niruri</i> after surface sterilization treatment, B) Fungus contamination on <i>P. niruri</i> , C) Bacterial contamination on <i>P. niruri</i> , and D) clean culture of <i>P. niruri</i> .	28
5.1	A) Axillary bud proliferation of <i>P. niruri</i> and B) Multiplied shoot of <i>P. niruri</i> .	40
6.1	A) Root development of <i>P. niruri</i> on week 2 and, B) Root development and elongation of <i>P. niruri</i> on week 4	51
6.2	Acclimatization process of <i>P. niruri</i> plantlet A) the <i>P. niruri</i> explants were covered with plastic and B) Survived <i>P. niruri</i> plantlet on week 4 of acclimatization	55

G

LIST OF ABBREVIATIONS

2,4-D	2,4-dichlorophenoxy-acetic acid
2iP	6-(γ,γ-Dimethylallylamino) purine
ANOVA	Analysis of variance
B5	Gamborg B5
BA	Benzyladenine
BAP	6-benzylaminopurine
CRD	completely randomized design
HCl	hydrochloric acid
IAA	indole-3-acetic acid
IBA	indole-3-butyric acid
Kinetin	6-Furfurylaminopurine
LSD	least significant difference
MS	Murashige and Skoog
NAA	naphthalene-acetic acid
NaOC1	sodium hypochlorite
NaOH	sodium hydroxide
NS	Nano silver
PGRs	Plant growth regulators
RAPD	random amplified polymorphic DNA
RH	relative humidity
UPM	Universiti Putra Malaysia
Zeatin	6-(4-hydroxy-3-methylbut-2-enylamino) purine

 \bigcirc

CHAPTER 1

INTRODUCTION

1.1 Background of study

Over the past several years, medicinal plants have gained greater attention and recognition in Malaysia, particularly as demand for alternative medicine and natural health products increases. A total of 351 hectares of Malaysian land has yielded 1317 metric tonnes of herbs in 2007. Since then, Malaysia's cultivated herbaceous area has grown from 1,198 hectares in 2011 to 2317 hectares in 2017, an annual increase of roughly 14% (Department of Agriculture, 2017). The Malaysia Economic Transformation Program, through the National Key Economic Areas Agriculture sector, has identified numerous high potential herbs that can be commercially utilized as a new economic growth source in the herbal industry. One of these herbs is *P. niruri*, identified locally as Dukung Anak (Farizah et al., 2015).

Phyllanthus niruri (Phyllanthaceae) has traditionally been used to treat several ailments such as jaundice, kidney stone, flu, fever, and diabetes (Unander and Blumberg, 1991; Calixto et al., 1998; Karthikeyan et al., 2008). Several researchers discovered that *P. niruri* possesses anti-tumor, anti-oxidant, anti-carcinogenic, and hepatoprotective properties (Chatterjee and Sil, 2006; Harish and Shivanandappa, 2006; Rajeshkumar et al., 2002). In addition, certain phenolic compounds such as gallic acid, epicatechin, gallocatechin, epigallocatechin, epicatechin 3-O-gallate, epigallocatechin 3-O-gallate have gained significant interest from the researcher (Ishimaru et al., 1992). Furthermore, a novel compound named niruriside was isolated from methanol extract of *P. niruri* that are capable of inhibiting the binding of human immunodeficiency virus Rev protein (HIV REV) (Qian-Cutrone et al., 1996). These secondary metabolites have a wide range of therapeutic activities that have essential pharmacological effects on humans. Hence, many products were made from this medicinal plant have been successfully commercialized.

The main issue for the commercialization of herbal-based products is the uniformity and consistency of the material. In contrast, the growing demand for medicinal plants would undoubtedly decrease the sustainable supply of raw materials in the future. This might occur due to overharvesting and unsustainable agriculture practices, which lead to many restrictions, such as a limited supply of herbs. Moreover, to extract elevated amounts of secondary metabolites, large quantities of raw materials are expected.

Plant micropropagation may also serve as an alternative solution to ensuring the sustainable supply of plant materials as it is an excellent technique to produce plants in a large amount in a short period. For example, 10000 *Ornithogalum* plants could be grown from a single leaf in eight months using micropropagation, instead of the conventional method that takes ten years to collect the same number (Thiart, 2003). Micropropagation is the *in vitro* aseptic culture of cells, tissues, or even organs of plants grown under a controlled environment and nutrient media for growth and multiplication. This technology can be used to eliminate diseases, secondary metabolite production, plant improvement, and conservation of threatened and endangered species (Hussain et al., 2012). Plant micropropagation consists of four major stages 1) surface sterilization of explant, 2) induction and multiplication of shoot, 3) *in vitro* root induction, and 4) acclimatization. The correct protocol should therefore be used to perform the plant micropropagation technique.

Appropriate selection of chemical sterilization is essential in the first stage of plant micropropagation. All microorganisms that potentially could contaminate the culture should be removed and eliminated. This step will establish an aseptic culture to be carried out in the following phase of micropropagation. On the other hand, numerous factors affect the explant's development in the growth and multiplication phase (Read, 2007). For instance, the type of basal media and its concentrations (Abobkar and Elshahed, 2016), type of carbon source (Sul and Korban, 2004), and its concentrations and initial pH value of culture medium also play an essential role in influencing the shoot induction of *P. niruri* explant. Culture medium supplemented with plant growth regulators; cytokinin and auxin were used in many plants to propagate via in vitro techniques (Preil, 2003). Commonly, cytokinin contributes to shoot induction (Schaller et al., 2014), whereas auxin helps in root induction (Woodward and Bartel, 2005), both functioning by promoting cell division and differentiation (Trigiano and Gray, 2011). In vitro plantlets will then be transferred to potting media during acclimatization, the final stage. In this stage, the survival rate of plantlets is also influenced by the type of potting media.

1.2 Problem statement

Due to its high medicinal values and properties, *P. niruri* has captured the attention of people all over the world. This plant is harvested with minimal commercialized production, and no sanitary consideration is taken on the sample collected. Besides, the Herbs and Spices Statistic in Malaysia stated that the total production of local herbs was 2312.42 Mt however, there is no information on the production of *P. niruri* (Jabatan Pertanian Malaysia, 2016). This suggests that the plant has not been thoroughly utilized for commercialization, especially in Malaysia. Herbs, particularly for medicinal

purposes, are in high demand right now. However, the herbal sector faced many obstacles due to lack of raw materials and knowledge in associated herbs. Thus, this study may propagate *P. niruri* rapidly and produce quality and safe *P. niruri* for human consumption. Hence, plant micropropagation is the most suitable approach to solve these problems.

1.3 Objectives of study

The general objective was to develop protocol for *in vitro* culture of *P. niruri* via micropropagation technique.

Besides, the specific objectives of this study were:

- 1. To determine the efficiency of different types and concentrations of chemical sterilants on surface sterilization of *P. niruri* explants.
- 2. To examine the growth responses on multiplication of *P. niruri* explant towards different growth factors.
- 3. To evaluate the effect of different types and concentrations of auxins on *in vitro* rooting and determine the suitable potting media for acclimatization of *P. niruri*.

REFERENCES

- Abdi, G., Salehi, H., and Khosh-Khui, M. (2008). Nano silver: A novel nanomaterial for removal of bacterial contaminants in valerian (*Valeriana officinalis* L.) tissue culture. *Acta Physiologiae Plantarum*, 30(5), 709–714.
- Abhyankar, G., and Reddy, V. D. (2007). Rapid micropropagation via axillary bud proliferation of *Adhatoda vasica* Nees from nodal segments. *Indian Journal of Experimental Biology*, 45(3), 268–271.
- Abobkar, I. M., and Elshahed, A. M. (2016). Plant Tissue Culture Media. *Intech*, 5(13), 29–40.
- Ahmad, N., Shahid, A., Javed, S. B., Khan, M. I., and Anis, M. (2015). Micropropagation of Vitex spp. through in vitro manipulation: Current status and future prospectives. *Journal of Applied Research on Medicinal and Aromatic Plants*, 2(4), 114–123.
- Ahmad, T., Abbasi, N. A., Hafiz, I. A., and Ali, A. (2007). Comparison of sucrose and sorbitol as main carbon energy sources in micropropagation of peach rootstock GF-677. *Pakistan Journal of Botany*, 39(4), 1269–1275.
- Alam, I., Sharmin, S. A., Naher, M. K., Alam, M. J., Anisuzzaman, M., and Alam, M. F. (2010). Effect of growth regulators on meristem culture and plantlet establishment in sweet potato [*Ipomoea batatas* (L.) Lam.]. *Plant OMICS*, 3(2), 35–39.
- Ali, A., Sajid, A., Naveed, N. H., Majid, A., Saleem, A., and Khan, U. A. (2011). Initiation, proliferation and development of micro-propagation system for mass scale production of banana through meristem culture. *African Journal* of *Biotechnology*, 10(70), 15731–15738.
- Ali, A., Ahmad, T., Abbasi, N. A., and Hafiz, I. A. (2009). Effect of different concentrations of auxins on In vitro rooting of olive cultivar "Moraiolo." *Pakistan Journal of Botany*, 41(3), 1223–1231.
- Ali, H., Houghton, P. J., and Soumyanath, A. (2006). α-Amylase inhibitory activity of some Malaysian plants used to treat diabetes; with particular reference to *Phyllanthus amarus*. *Journal of Ethnopharmacology*, 107(3), 449–455.
- Ana, K. P., Ana, P. da S., Joselita, C. de S., Silvio, L. T., Juliana, M. R., Ana, R. P., and Cristiane, D. da P. (2016). Sodium hypochlorite sterilization of culture

medium in micropropagation of *Gerbera hybrida* cv. Essandre. *African Journal of Biotechnology*, 15(36), 1995–1998.

- Aremu, A. O., Plačková, L., Bairu, M. W., Novák, O., Plíhalová, L., Doležal, K., and Van Staden, J. (2014). How does exogenously applied cytokinin type affect growth and endogenous cytokinins in micropropagated *Merwilla plumbea? Plant Cell, Tissue and Organ Culture, 118*(2), 245–256.
- Bagalkotkar, G., Sagineedu, S. R., Saad, M. S., and Stanslas, J. (2006). Phytochemicals from *Phyllanthus niruri* Linn. and their pharmacological properties: a review. *Journal of Pharmacy and Pharmacology*, 58(12), 1559– 1570.
- Bais, H. P., Green, J. B., Walker, T. S., Okemo, P. O., and Vivanco, J. M. (2002). In Vitro propagation of Spilanthes mauritiana DC., an endangered medicinal herb through axillary bud cultures. Vitro Cellular & Developmental Biology. Plant, 35(919), 598–601.
- Bajpai, A., Kalim, S., Chandra, R., and Kamle, M. (2016). Recurrent somatic embryogenesis and plantlet regeneration in *Psidium guajava* L. *Brazilian Archives of Biology and Technology*, 59(0), 1–11.
- Banerjee, N., and de Langhe, E. (1985). A tissue culture technique for rapid clonal propagation and storage under minimal growth conditions of Musa (Banana and plantain). *Plant Cell Reports*, 4(6), 351–354.
- Baskaran, P., and Jayabalan, N. (2016). Role of basal media, carbon sources and growth regulators in micropropagation of two valuable medicinal orchids of Bangladesh. *International Journal of Science and Research*, 5(6), 1022–1026.
- Bavarva, J. H., and Narasimhacharya, A. V. R. L. (2007). Comparative antidiabetic, hypolipidemic, and antioxidant properties of *Phyllanthus niruri* in normal and diabetic Rats. *Pharmaceutical Biology*, 45(7), 569–574.
- Belete, K., and Balcha, A. (2015). Micropropagation of *Plectranthus edulis* (Vatke) Agnew from shoot tip and nodal explants. *African Journal of Agricultural Research*, 10(1), 6–13.
- Bello, I., Usman, N. S., Dewa, A., Abubakar, K., Aminu, N., Asmawi, M. Z., and Mahmud, R. (2020). Blood pressure lowering effect and vascular activity of *Phyllanthus niruri* extract: The role of NO/cGMP signaling pathway and βadrenoceptor mediated relaxation of isolated aortic rings. *Journal of Ethnopharmacology*, 250-278.

Bhatia, P., and Ashwath, N. (2005). Effect of medium pH on shoot regeneration

from the cotyledonary explants of tomato. *Biotechnology*, 4(1), 7–10.

- Bhattacharjee, R., and Sil, P. C. (2007). Protein isolate from the herb, *Phyllanthus niruri L*. (Euphorbiaceae), plays hepatoprotective role against carbon tetrachloride induced liver damage via its antioxidant properties. *Food and Chemical Toxicology*, *45*, 817–826.
- Bhattacharyya, R., and Bhattacharya, S. (2001). High frequency in vitro propagation of *Phyllanthus amarus* Schum. & Thom. by shoot tip culture. *Indian Journal of Experimental Biology*, 39(11), 1184–1187.
- Bilderback, T. E., Warren, S. L., Owen, J. S., and Albano, J. P. (2005). Healthy substrates need physicals too! *HortTechnology*, *15*(4), 747–751.
- Bouman, R. W., Keßler, P. J. A., Telford, I. R. H., Bruhl, J. J., and Van Welzen, P. C. (2018). Subgeneric delimitation of the plant genus *Phyllanthus* (Phyllanthaceae). *Blumea: Journal of Plant Taxonomy and Plant Geography*, 63(2), 167–198.
- Bozena, B., and Szczerba, J. (1991). Influence of different carbon sources on invertase activity and growth of sour cherry (*Prunus cerasus* L.) shoot cultures. *Journal of Experimental Botany*, 42(7), 911–915.
- Brown, D. C., and Thorpe, T. (1980). Crop improvement through tissue culture. *World Journal of Microbiology and Biotechnology*, 11, 409–415.
- Bunn, E. (2002). Microbial contaminants in plant tissue culture propagation. In K. Sivasithamparam, K. W. Dixon, and R. L. Barret (Eds.), *Microorganisms in Plant Conservation and Biodiversity* (pp. 307–335). Kluwer Academic Publisher.
- Butenko, R. G., Lipsky, A. K., Chernyak, N. D., and Arya, H. C. (1984). Changes in culture medium pH by cell suspension cultures of *Dioscorea deltoidea*. *Plant Science Letters*, 35(3), 207–212.
- Calixto, J. B., Santos, A. R. S., Cechinel Filho, V., and Yunes, R. A. (1998). A review of the plants of the genus *Phyllanthus*: Their chemistry, pharmacology, and therapeutic potential. *Medicinal Research Reviews*, *18*(4), 225–258.
- Carlile, W. R., Cattivello, C., and Zaccheo, P. (2015). Organic growing media: constituents and properties. *Vadose Zone Journal*, 14(6), 2–13.
- Castellano, J. J., Shafii, S. M., Ko, F., Donate, G., Wright, T. E., Mannari, R. J. and Robson, M. C. (2007). Comparative evaluation of silver-containing

antimicrobial dressings and drugs. *International Wound Journal*, 4(2), 114–122.

- Catapan, E., Luís, M., Da Silva, B., Netto Moreno, F., and Maria Viana, A. (2002). Micropropagation, callus and root culture of *Phyllanthus urinaria* (Euphorbiaceae). *Plant Cell, Tissue and Organ Culture*, 70(3), 301–309.
- Cha-um, S., Ulziibat, B., and Kirdmanee, C. (2010). Effect of temperature and relative humidity during in vitro acclimatization, on physiological changes and growth characters of Phalaenopsis adapted to in vivo. *Australian Journal of Crop Science*, 4(9), 313–343.
- Chatterjee, M., and Sil, P. C. (2006). Hepatoprotective effect of aqueous extract of *Phyllanthus niruri* on nimesulide-induced oxidative stress in vivo. *Indian Journal of Biochemistry and Biophysics*, 43(5), 299–305.
- Chee, B., and Saad, A. hayat. (2019). Dukung Anak. In *Herba Emas Negara* (pp. 21–40). Forest Research Institute Malaysia (FRIM).
- Daud, N. H., Jayaraman, S., and Mohamed, R. (2012). An improved surface sterilization technique for introducing leaf, nodal and seed explants of *Aquilaria malaccensis* from field sources into tissue culture. *Asia-Pacific Journal of Molecular Biology and Biotechnology*, 20(2), 55–58.
- De Klerk, G. J., Ter Brugge, J., and Marinova, S. (1997). Effectiveness of indoleacetic acid, indolebutyric acid and naphthaleneacetic acid during adventitious root formation *in vitro* in *Malus* "Jork 9." *Plant Cell, Tissue and Organ Culture*, 49(1), 39–44.
- De Padua, L. S., Bunyapraphatsara, N., and Lemmens, R. H. M. J. (1999). Plant resource of South-East Asia. *Medicinal and Poisonous Plants*, 12(1), 210-218.
- Debnath, S. C. (2003). Improved shoot organogenesis from hypocotyl segments of lingonberry (*Vaccinium vitis-idaea* L.). In Vitro Cellular and Developmental Biology - Plant, 39(5), 490–495.
- Department of Agriculture. (2014). Buletin SPBT Jabatan Pertanian. In *Putrajaya: Jabatan Pertanian*.
- Devi, C. S., and Srinivasan, V. M. (2006). Studies on various atmospheric microorganisms affecting the plant tissue culture explant. *American Journal* of *Plant Physiology*, 1(2), 205–209.
- Dube, P., Gangopadhyay, M., Dewanjee, S., and Ali, M. N. (2011). Establishment of a rapid multiplication protocol of *Coleus forskohlii* Briq. and *in vitro*

conservation by reduced growth. *Indian Journal of Biotechnology*, 10(2), 228–231.

- Durak, D., Kalender, S., Uzun, F. G., Demir, F., and Kalender, Y. (2010). Mercury chloride-induced oxidative stress in human erythrocytes and the effect of vitamins C and E *in vitro*. *African Journal of Biotechnology*, *9*(4), 488–495.
- Ebrahim, M. K. H. (2004). Comparison, determination and optimizing the conditions required for rhizome and shoot formation, and flowering of in vitro cultured calla explants. *Scientia Horticulturae*, 101(3), 305–313.
- El-Hawaz, R., Park, D., Bridges, W. C., and Adelberg, J. (2016). Optimizing *in vitro* mineral nutrition and plant density increases greenhouse growth of *Curcuma longa* L. during acclimatization. *Plant Cell, Tissue and Organ Culture,* 126(1), 33–42.
- Emmanuel, E., Keck, G., Blanchard, J. M., Vermande, P., and Perrodin, Y. (2004). Toxicological effects of disinfections using sodium hypochlorite on aquatic organisms and its contribution to AOX formation in hospital wastewater. *Environment International*, 30(7), 891–900.
- Fakhrfeshani, M. (2014). Disinfecting effects of nano silver fluids in gerbera (Gerbera jamesonii) capitulum tissue culture. Journal of Biological and Environmental Science, 6(17), 121–127.
- Farizah, A., Azlan S. Z., M., Noorasiah, S., Adibah A., Majid, and F., Ahmad. (2015). Issues and Challenges in the Development of the Herbal Industry in Malaysia. *Prosiding Persidangan Kebangsaan Ekonomi Malaysia (Perkem)*, 10, 227–238.
- Feng, J. C., Yu, X. M., Shang, X. L., Li, J. D., and Wu, Y. X. (2010). Factors influencing efficiency of shoot regeneration in *Ziziphus jujuba* Mill. "Huizao." *Plant Cell, Tissue and Organ Culture*, 101(1), 111–117.
- Gaba, V. P. (2005). Plant growth regulators in plant tissue culture and development. Boca Raton, FL: CRC Press ,87–99.
- Gallavotti, A. (2013). The role of auxin in shaping shoot architecture. *Journal of Experimental Botany*, 64(9), 2593–2608.
- Gamborg, O. L., Miller, R. A., and Ojima, K. (1968). Nutrient requirements of suspension cultures of soybean root cells. *Experimental Cell Research*, 50(1), 151–158.

George, E. F., Hall, M. A., and Klerk, G. . (2008). Plant Propagation by Tissue

Culture 3rd Edition.

- Ghosh, S., Ghosh, B., and Jha, S. (2007). *In vitro* tuberisation of *Gloriosa superba* L. on basal medium. *Scientia Horticulturae*, 114(3), 220–223.
- González, A., and Ayerbe, L. (2010). Effect of terminal water stress on leaf epicuticular wax load, residual transpiration and grain yield in barley. *Euphytica*, 172(3), 341–349.
- Haida, Z., Nakasha, J. J., and Hakiman, M. (2020). *In vitro* responses of plant growth factors on growth, yield, phenolics content and antioxidant activities of *Clinacanthus nutans* (Sabah snake grass). *Plants*, *9*(8), 1–17.
- Harish, R., and Shivanandappa, T. (2006). Antioxidant activity and hepatoprotective potential of *Phyllanthus niruri*. *Food Chemistry*, 95(2), 180– 185.
- Hartman, H. T., Kester, D. E., Davies, F. T., and Geneve, R. L. (1975). *Plant propagation: principle and practices*. Prentice-Hall.
- Hassan, M. M., Ibrahim, I. A., Fathy, N. M., Ebrahim, M. K. H., and Komor, E. (2014). Protocol for micropropagated date palm acclimatization: Effect of micropropagated plantlet type, soil composition, and acclimatization season. *International Journal of Fruit Science*, 14(2), 225–233.
- Hassan, N. H., Ali, N. A. M., Zainudin, F., and Ismail, H. (2011). Effect of 6benzylaminopurine (BAP) in different basal media on shoot multiplication of *Aquilaria hirta* and detection of essential oils in the *in vitro* shoots. *African Journal of Biotechnology*, 10(51), 10500–10503.
- Hicks, G. S. (1980). Patterns of organ development in plant tissue culture and the problem of organ determination. *The Botanical Review*, 46(1), 1–23.
- Hilae, A., and Te-chato, S. (2005). Effects of carbon sources and strength of MS medium on germination of somatic embryos of oil palm (*Elaeis quineensis* Jacq.). *Songklanakarin Journal Science Technology*, 27, 629–635.
- Hussain, A., Ahmed, I., Nazir, H., and Ullah, I. (2012). Plant Tissue Culture: Current Status and Opportunities. In *Recent Advances in Plant in vitro Culture*, 1–28.
- Ilan, A., and Khayat, E. (1997). An overview of commercial and technological limitations to marketing of micropropagated plants. *Acta Horticulturae*, 447, 643–648.

- Iliev, I., Rubos, A., Scaltsoyiannes, A., Nellas, C., and Kitin, P. (2003). Anatomical study of *in vitro* obtained fasciated shoots from *Betula pendula* Roth. *Acta Horticulturae*, 616, 481–484.
- Ishimaru, K., Yoshimatsu, K., Yamakawa, T., Kamada, H., and Shimomura, K. (1992). Phenolic constituents in tissue cultures of *Phyllanthus niruri*. *Phytochemistry Reviews*, *31*(6), 2015–2018.
- Ismail, H., Muniandi, S. K., Yusoff, A. M., Hassan, N. H., Aini, N., and Shukor, A. (2016). *In vitro* micropropagation of *Acacia auriculiformis* form selected juvenile sources. *Dendrobiology Journal*, 75, 157–165.

Jabatan Pertanian Malaysia, D. of A. (2016). Herbs and Spices Statistic.

- Kadhimi, A. A., Alhasnawi, A. N., Mohamad, A., Wan Yusoff, W. M., and Che Mohd. Zain, C. R. (2014). Tissue culture and some of the factors affecting them and the micropropagation of strawberry. *Life Science Journal*, 11(8), 484–493.
- Kalidass, C., and Mohan, V. R. (2009). *In vitro* clonal propagation of *Phyllanthus urinaria* (Euphorbiaceae) A medicinal plant. *Researcher*, 1(4), 56–61.
- Karjee, S., Singh, K. P., and Panwar, S. (2020). Development of *in-vitro* protocol for direct regeneration from thalamus ex-plant of *Tagetes patula* L. var. Pusa Deep. *Journal of Pharmacognosy and Phytochemistry*, 9(4), 373–377.
- Karthikeyan, K., Chandran, C., and Kulothungan, S. (2008). In vitro propagation of Phyllanthus niruri L.- A medicinal plant. Journal of Science Transaction in Environment and Technovation, 1(3), 131–133.
- Kashyap, B., and Dhiman, S. R. (2011). Effect of media on hardening of *in vitro* multiplied plantlets of *Gloxinia* and *Saintpaulia* under low cost polytunnels. *International Journal of Farm Science*, 1(2), 63–67.
- Khanna, A. K., Rizvi, F., and Chander, R. (2002). Lipid lowering activity of *Phyllanthus niruri* in hyperlipemic rats. *Journal of Ethnopharmacology*, 82(1), 19–22.
- Khuri, S., and Moorby, J. (1995). Investigations into the role of sucrose in potato cv. Estima microtuber production in vitro. *Annals of Botany*, 75, 295–303.
- Kirdmanee, C., Kitaya, Y., and Kozai, T. (1995). Effect of CO₂ enrichment and supporting material *in vitro* on photoautotrophic growth of eucalyptus plantlets *in vitro* and *ex vitro*. *In Vitro Cellular & Developmental Biology Plant*, 31, 144–149.

- Kollmeier, M., Felle, H. H., and Horst, W. J. (2000). Is basipetal auxin flow involed in inhibition of root elongation? *Plant Physiology*, 122, 945–956.
- Kumaran, A., and Joel Karunakaran, R. (2007). In vitro antioxidant activities of methanol extracts of five Phyllanthus species from India. Food Science and Technology, 40(2), 344–352.
- Kuttan, R., and Harikumar, K. B. (2012). Taxonomy of genus *Phyllanthus*. In *Phyllantus Species- scientific evaluation and medicinal application* (pp. 1–22). Boca Raton: CRC Press.
- Lamhamedi, M. S., Chamberland, H., and Tremblay, F. M. (2003). Epidermal transpiration, ultrastructural characteristics and net photosynthesis of white spruce somatic seedlings in response to *in vitro* acclimatization. *Physiologia Plantarum*, 118(4), 554–561.
- Lavanya, M., Venkateshwarlu, B., and Poornasri Devi, B. (2009). Acclimatization of neem microshoots adaptable to semi-sterile conditions. *Indian Journal of Biotechnology*, 8(2), 218–222.
- Leifert, C., and Waites, W. M. (1992). Bacterial growth in plant tissue culture media. *Journal of Applied Bacteriology*, 72(6), 460–466.
- Liang, O., and Keng, C. (2006). *In vitro* plant regeneration, flowering and fruiting of *Phyllanthus niruri* L. (Euphorbiaceae). *International Journal of Botany*, 2(4), 409–414.
- Lipavská, H., and Konrádová, H. (2004). Somatic embryogenesis in conifers: The role of carbohydrate metabolism. *In Vitro Cellular and Developmental Biology Plant*, 40(1), 23–30.
- Lloyd, G., and McCown, B. (1980). Commercially feasible micropropagation of Mountain Laurel, *Kalmia Latifolia*, by use of shoot-tip culture. *International Plant Propagators' Society*, 421–427.
- Loria, R., Coombs, J., Yoshida, M., Kers, J., and Bukhalid, R. (2003). A paucity of bacterial root diseases: Streptomyces succeeds where others fail. *Physiological and Molecular Plant Pathology*, *62*(2), 65–72.
- Ludwig-Müller, J. (2000). Indole-3-butyric acid in plant growth and development. *Plant Growth Regulation*, *32*, 219–230.
- Lund, B. O., Miller, D. M., and Woods, J. S. (1993). Studies on Hg(II)-induced H₂O₂ formation and oxidative stress *in vivo* and *in vitro* in rat kidney mitochondria. *Biochemical Pharmacology*, 45(10), 2017–2024.

- Madhulatha, P., Kirubakaran, S. I., and Sakthivel, N. (2006). Effects of carbon sources and auxins on *in vitro* propagation of banana. *Biologia Plantarum*, 50(4), 782–784.
- Maina, S. M., Emongor, Q., Sharma, K. K., Gichuki, S. T., Gathaara, M., and de Villiers, S. M. (2010). Surface sterilant effect on the regeneration efficiency from cotyledon explants of groundnut (*Arachis hypogea* L.) varieties adapted to Eastern and Southern Africa. *African Journal of Biotechnology*, 9(20), 2866–2871.
- Malik, S., Zia, M., Riaz-ur-Rehman, and Chaudhary, M. F. (2007). *In vitro* plant regeneration from direct and indirect organogenesis of *Memordica charantia*. *Pakistan Journal of Biological Sciences*, 10, 4118–4122.
- Manjusha, A. V. M., and Sathyanarayana, B. N. (2010). Acclimatization studies in stevia (*Stevia rebaudiana* Bert.). *Acta Horticulturae*, 865, 129–134.
- Martin, S. M., and Rose, D. (1976). Growth of plant cell (Ipomoea) suspension cultures at controlled pH levels. *Canadian Journal of Botany*, 54(11), 1264–1270.
- Micali, S., Sighinolfi, M. C., Celia, A., De Stefani, S., Grande, M., Cicero, A. F., and Bianchi, G. (2006). Can *Phyllanthus niruri* affect the efficacy of extracorporeal shock wave lithotripsy for renal stones? A randomized, prospective, long-term study. *Journal of Urology*, 176(3), 1020–1022.
- Mihaljevic, I., Dugalic, K., Tomas, V., Viljevac, M., Pranjic, A., Cmelik, Z. and Jurkovic, Z. (2013). *In vitro* sterilization procedures for micropropagation of 'oblacinska' sour cherry. *Journal of Agricultural Sciences, Belgrade*, 58(2), 117–126.
- Mng'omba, S.A., du Toit, E. S., Akinnifesi K., F. and Sileshi, G. (2012). Efficacy and utilization of fungicides and other antibiotics for aseptic plant cultures. *Fungicides for Plant and Animal Diseases*. In tech Open Acces Publisher.
- Modgil, M., Sharma, T., and Thakur, M. (2009). Commercially feasible protocol for rooting and acclimatization of micropropagated apple rootstocks. *Acta Horticulturae*, 839, 209–214.
- Moharir, K. K., Sharma, M. K., Mishra, A., and Joshi, C. K. (2020). Effect of different colour shade nets and substrates during acclimatization on micropropagated banana plantlet (*Musa* spp.) in different seasons. *Plant Cell Biotechnology and Molecular Biology*, 21(71–72), 280–290.

Monfort, L. E. F., Bertolucci, S. K. V., Lima, A. F., de Carvalho, A. A., Mohammed,

A., Blank, A. F., and Pinto, J. E. B. P. (2018). Effects of plant growth regulators, different culture media and strength MS on production of volatile fraction composition in shoot cultures of *Ocimum basilicum*. *Industrial Crops and Products*, *116*, 231–239.

- Morard, P., and Henry, M. (1998). Optimization of the mineral composition of *in vitro* culture media. *Journal of Plant Nutrition*, 21(8), 1565–1576.
- Moreira, J., Klein-Júnior, L. C., Filho, V. C., and Buzzi, F. D. C. (2013). Antihyperalgesic activity of corilagin, a tannin isolated from *Phyllanthus niruri* L. (Euphorbiaceae). *Journal of Ethnopharmacology*, 146(1), 318–323.
- Murashige, T., and Skoog, F. (1962). A Revised medium for rapid grwoth and bio assays with tobacco tissue cultures. *Physiologia Plantarum*, 15, 473–497.
- Murugaiyah, V., and Chan, K. L. (2007). Determination of four lignans in *Phyllanthus niruri* L. by a simple high-performance liquid chromatography method with fluorescence detection. *Journal of Chromatography A*, 1154(1–2), 198–204.
- Nadha, H. K., Salwan, R., Kasana, R. C., Anand, M., and Sood, A. (2012). Identification and elimination of bacterial contamination during *in vitro* propagation of *Guadua angustifolia* Kunth. *Pharmacognosy Magazine*, 8(30), 93–97.
- Naik, A. D., and Juvekar, A. R. (2003). Effect of alkaloidal extract of *Phyllanthus niruri* on HIV replication. *Indian Journal of Medical Sciences*, 57(9), 387–393.
- Narendra, K., Swathi, J., Sowjanya, K., and Satya Savithri, A. (2012). *Phyllanthus niruri*: A review on its ethno botanical, phytochemical and pharmacological profile. *Journal of Pharmacy Research*, 5(9), 4681–4691.
- Nejatzadeh-Barandozi, F., Darvishzadeh, F., and Aminkhani, A. (2014). Effect of nano silver and silver nitrate on seed yield of *Ocimum basilicum* L. *Organic and Medicinal Chemistry Letters*, 4(1), 1–6.
- Nguyen, Q. T., Xiao, Y., and Kozai, T. (2019). Photoautotrophic micropropagation. *Plant Factory: An Indoor Vertical Farming System for Efficient Quality Food Production: Second Edition*, 27, 333–346.
- Nowak, B., Miczyński, K., and Hudy, L. (2004). Sugar uptake and utilisation during adventitious bud differentiation on *in vitro* leaf explants of "Węgierka Zwykła" plum (*Prunus domestica*). *Plant Cell, Tissue and Organ Culture*, 76(3), 255–260.

- Ogunsanwo, Y. R. (2007). Sources of microbial contamination in tissue culture laboratories in southwestern Nigeria. *African Journal of Agricultural Research*, 2(3), 067–072.
- Orlikowska, T., Nowak, K., and Reed, B. (2017). Bacteria in the plant tissue culture environment. *Plant Cell, Tissue and Organ Culture*, 128(3), 487–508.
- Parveen, S., Mir, H., Ranjan, T., Pal, A. K., and Kundu, M. (2019). Effect of surface sterilants on *in vitro* establishment of pineapple (*Ananas comosus* (L.) Merill.) cv. Kew. *Current Journal of Applied Science and Technology*, 33(2), 1– 6.
- Patel, A., Singh, P., and Khan, S. (2018). Standardization of protocol for *in vitro* micropropagation of *Phyllanthus niruri*: An important medicinal plant. UK *Journal of Pharmaceutical Biosciences*, 6(4), 42.
- Patel, P., Harde, P., Pillai, J., Darji, N., and Patel, B. (2012). Antidiabetic herbal drugs: A review. *Pharmacophore*, *3*(1), 18–29.
- Pence, V. C. (2005). In vitro collecting (IVC). I. The effect of collecting method and antimicrobial agents on contamination in temperate and tropical collections. *In Vitro Cellular and Developmental Biology - Plant*, 41(3), 324–332.
- Poobathy, R., Zakaria, R., Murugaiyah, V., and Subramaniam, S. (2019). Surface sterilization and micropropagation of *Ludisia discolor*. *Biocatalysis and Agricultural Biotechnology*, 22, 101380.
- Prajapati, N. D. (2003). Handbook of medicinal plants. Agrobis.
- Preece, J. E., and Sutter, E. G. (1991). Acclimatization of micropropagated plants to the greenhouse and field. *Micropropagation*, 71–93.
- Preil, W. (2003). Micropropagation of ornamental plants. In *Plant Tissue Culture* 100 years since Gottlied Haberlandt, 115–133.
- Qian-Cutrone, J., Huang, S., Trimble, J., Li, H., Lin, P. F., Alam, M., and Kadow, K. F. (1996). Niruriside, a new HIV REV/RRE binding inhibitor from *Phyllanthus niruri. Journal of Natural Products*, 59(2), 196–199.
- Rajasubramaniam, S., and Saradhi, P. P. (1997). Rapid multiplication of *Phyllanthus fraternus*: A plant with anti-hepatitis viral activity. *Industrial Crops and Products*, 6(1), 35–40.

Rajeshkumar, N. V., Joy, K. L., Kuttan, G., Ramsewak, R. S., Nair, M. G., and

Kuttan, R. (2002). Antitumour and anticarcinogenic activity of *Phyllanthus amarus* extract. *Journal of Ethnopharmacology*, *81*(1), 17–22.

- Raviv, M., Chen, Y., and Inbar, Y. (1986). Peat and peat substitutes as growth media for container-grown plants. *The Role of Organic Matter in Modern Agriculture*, 257–287.
- Read, P. E. (2007). Micropropagation: Past, present and future. *Acta Horticulturae*, 748, 17–27.
- Reed, B. M., Sarasan, V., Kane, M., Bunn, E., and Pence, V. C. (2011). Biodiversity conservation and conservation biotechnology tools. *In Vitro Cellular and Developmental Biology Plant*, 47(1), 1–4.
- Rezali, N. I., Jaafar Sidik, N., Saleh, A., Osman, N. I., and Mohd Adam, N. A. (2017). The effects of different strength of MS media in solid and liquid media on *in vitro* growth of *Typhonium flagelliforme*. *Asian Pacific Journal of Tropical Biomedicine*, 7(2), 151–156.
- Ribeiro, J. M., Teixeira, S. L., and Bastos, D. C. (2011). *In vitro* culture of *Sequoia sempervirens* L. on nutritive media sterilized with sodium hypochlorite. *Ciência Florestal*, 21(1), 77–82.
- Rohr, R., Iliev, I., Scaltsoyiannes, A., and Tsoulpha, P. (2003). Acclimatization of micropropagated forest trees. *Acta Horticulturae*, *616*, 59–69.
- Saad, A. I., and Elshahed, A. M. (2012). Plant tissue culture media. *Recent Advances in Plant in vitro Culture*, 17(10), 30–40.
- Sadik, K., Arinaitwe, G., Rubaihayo, P., Kiggundu, A., and Mukasa, S. (2014). TDZ AND 4-CPPU in Gamborg B5 salts with MS vitamins doubles embryogenic 191 response from male flowers of EA-AAA banana. *African Crop Science Journal*, 22(3), 191–204.
- Sahrawat, A. K., and Chand, S. (1999). Stimulatory effect of copper on plant regeneration in indica rice (*Oryza sativa* L.). *Journal of Plant Physiology*, 154(4), 517–522.
- Samali A. (2012). Evaluation of chemical constituents of *Phyllanthus niruri*. *African Journal of Pharmacy and Pharmacology*, 6(3), 125–128.
- Schaller, G. E., Street, I. H., and Kieber, J. J. (2014). Cytokinin and the cell cycle. *Current Opinion in Plant Biology*, 21, 7–15.

- Schubert, S., Schubert, E., and Mengel, K. (1990). Effect of low pH of the root medium on proton release, growth, and nutrient uptake of field bean (*Vicia faba*). Kluwer Acadamic Publiser, 443-448.
- Shakil, N. A., Pankaj, Kumar, J., Pandey, R. K., and Saxena, D. B. (2008). Nematicidal prenylated flavanones from *Phyllanthus niruri*. *Phytochemistry*, 69(3), 759–764.
- Sharma, J., Gairola, S., Gaur, R. D., and Painuli, R. M. (2012). The treatment of jaundice with medicinal plants in indigenous communities of the Sub-Himalayan region of Uttarakhand, India. *Journal of Ethnopharmacology*, 143(1), 262–291.
- Shekhawat, M. S., Kannan, N., Manokari, M., and Ravindran, C. P. (2015). *In vitro* regeneration of shoots and *ex vitro* rooting of an important medicinal plant *Passiflora foetida* L. through nodal segment cultures. *Journal of Genetic Engineering and Biotechnology*, 13(2), 209–214.
- Singh, A., Jani, K., Kumari, P., and Agarwal, P. K. (2014). Effect of MgCl₂ and double concentration of Murashige and Skoog medium on *in vitro* plantlet and root cultures generation in halophytic grasswort *Salicornia brachiata*. *Plant Cell, Tissue and Organ Culture*, 120(2), 563–570.
- Singh, M. K., and Ahmad, M. (2020). Phytochemcial profile of *Phyllanthus niruri* L and of its potent bioactive compounds. *Bioscience Biotechnology Research Communication*, 13(3), 1545–1551.
- Singha, S., Oberly, G. H., and Townsend, E. C. (1987). Changes in nutrient composition and pH of the culture medium during *in vitro* shoot proliferation of crabapple and pear. *Plant Cell, Tissue and Organ Culture*, 11(3), 209–220.
- Skirvin, R. M., Chu, M. C., Mann, M. L., Young, H., Sullivan, J., and Fermanian, T. (1986). Stability of tissue culture medium pH as a function of autoclaving, time, and cultured plant material. *Plant Cell Reports*, 5(4), 292– 294.
- Sondi, I., and Salopek-Sondi, B. (2004). Silver nanoparticles as antimicrobial agent: A case study on *E. coli* as a model for Gram-negative bacteria. *Journal of Colloid and Interface Science*, 275(1), 177–182.
- Sridhar, T. M., and Naidu, C. V. (2011). Effect of different carbon sources on *in vitro* shoot regeneration of *Solanum nigrum* (Linn.) - An important antiulcer medicinal plant. *Journal of Phytology*, 3(2), 78–82.

- Staswick, P. E., Serban, B., Rowe, M., Tiryaki, I., Maldonado, M. T., Maldonado, M. C., and Suza, W. (2005). Characterization of an arabidopsis enzyme family that conjugates amino acids to indole-3-acetic acid. *Plant Cell*, 17(2), 616–627.
- Sul, I. I. W., and Korban, S. S. (2004). Effects of salt formulations, carbon sources, cytokinins, and auxin on shoot organogenesis from cotyledons of *Pinus pinea* L. *Plant Growth Regulation*, 43(3), 197–205.
- Sutter, E., and Langhans, R. W. (1982). Formation of epicuticular wax and its effect on water loss in cabbage plants regenerated from shoot-tip culture. *Canadian Journal of Botany*, *60*(12), 2896–2902.
- Tanimoto, E. (2005). Regulation of root growth by plant hormones Roles for auxin and gibberellin. *Critical Reviews in Plant Sciences*, 24(4), 249–265.
- Teixeira, S. L., Ribeiro, J. M., and Teixeira, M. T. (2006). Influence of NaClO on nutrient medium sterilization and on pineapple (*Ananas comosus* cv Smooth cayenne) behavior. *Plant Cell, Tissue and Organ Culture*, *86*(3), 375–378.
- Tesfa, M., and Admassu, B. (2016). In vitro rooting and acclimatization of micropropagated elite sugarcane (Saccharum officinarum L.) genotypes -N52 and N53. Journal of Tissue Science & Engineering, 07(01).
- Thiart, S. (2004). Manipulation of growth by using tissue culture techniques. *Proceeding of The International Plant Propagator's Society*, 53, 61-66.
- Tisarum, R., Theerawitaya, C., Samphumphuang, T., and Cha-um, S. (2018). Regulation of anthocyanin accumulation in rice (*Oryza sativa* L. subsp. indica) using MgSO₄ spraying and low temperature. *Archives of Agronomy and Soil Science*, *64*(12), 1663–1677.
- Torres, K. C. (2012). *Tissue culture techniques for horticultural crops*. Springer Science & Business Media.
- Trigiano, R., and Gray, D.J. (2011). Plant Tissue Culture, Development and Biotechnology. CRC Press.
- Trigiano, R. N., and Gray, D. J. (1999). *Plant Tissue Culture Concept and Laboratory Exercise*. CRC Press.
- Tripathi, A. K., Verma, R. K., Gupta, A. K., Gupta, M. M., and Khanuja, S. P. S. (2006). Quantitative determination of phyllanthin and hypophyllanthin in *Phyllanthus* species by high-performance thin layer chromatography. *Phytochemical Analysis*, *17*(6), 394–397.

- Unander, D. W., and Blumberg, B. S. (1991). *In vitro* activity of *Phyllanthus* (Euphorbiaceae) species against the DNA polymerase of hepatitis viruses: Effects of growing environment and inter- and intra-specific differences. *Economic Botany*, 45(2), 225–242.
- Van der Krieken, W. M., Breteler, H., Visser, M. H. M., and Mavridou, D. (1993). The role of the conversion of IBA into IAA on root regeneration in apple: Introduction of a test system. *Plant Cell Reports*, *12*(4), 203–206.
- Vasil, I. K., and Thorpe, T. (2013). *Plant cell and tissue culture*. Springer Science & Business Media, 215-246.
- Vasudevan, R., and van Staden, J. (2011). Cytokinin and explant types influence *in vitro* plant regeneration of Leopard Orchid (*Ansellia africana* Lindl.). *Plant Cell, Tissue and Organ Culture,* 107(1), 123–129.
- Venkatachalam, P., Kalaiarasi, K., and Sreeramanan, S. (2015). Influence of plant growth regulators (PGRs) and various additives on *in vitro* plant propagation of *Bambusa arundinacea* (Retz.) Wild: A recalcitrant bamboo species. *Journal of Genetic Engineering and Biotechnology*, 13(2), 193–200.
- Vuylsteke, D. R. (1998). Shoot-tip culture for the propagation, conservation and distribution of *Musa* germaplasm. International Institue of Tropical Agriculture, Ibadan, Nigeria, 82.
- Wada, S., Tanimoto, E., and Masuda, Y. (1968). Cell elongation and metabolic turnover of the cell wall as affected by auxin and cell wall degrading enzymes. *Plant and Cell Physiology*, 9(2), 369–376.
- Warakagoda, P. S., and Subasinghe, S. (2009). *In vitro* culture establishment and shoot proliferation of *Jatropha curcas* L. *Exposure*, 12(2), 77-80.
- Webster, S., and Mitchell, S. (2003). A novel surface sterilization method for reducing microbial contamination of field grown medicinal explants intended for *in vitro* culture. *In Vitro Cellular & Developmental Biology Plant*, 12,1–8.
- Wetherell, D. F., and Dougall, D. K. (1976). Sources of nitrogen supporting growth and embryogenesis in cultured wild carrot tissue. *Physiologia Plantarum*, *37*(2), 97–103.
- White, P. (1963). The Cultivation of Animal and Plant Cells. Philip R. White. Ronald, New York, ed. 2, In *Science* ,141, 515–516.

Williams, R. R., Taji, A. M., & Winney, K. A. (1990). The effect of Ptilotus plant

tissue on pH of *in vitro* media. *Plant Cell, Tissue and Organ Culture,* 22(3), 153–158.

- Woodward, A. W., and Bartel, B. (2005). Auxin: Regulation, action, and interaction. *Annals of Botany*, 95(5), 707–735.
- Yaacob, M., Mohammad, A. A., Ariffin, Z., and Ariffin, A. (2002). Manual teknologi penanaman dukung anak. *Institut Penyelidikan dan Kemajuan Pertanian Malaysia (MARDI)*.
- Yaseen, M., Ahmad, T., Sablok, G., Standardi, A., and Hafiz, I. A. (2013). Review: Role of carbon sources for *in vitro* plant growth and development. *Molecular Biology Reports*, 40(4), 2837–2849.
- Yoshihara, T., and Hanyu, H. (1992). pH changes in culture medium with progress of growing stages, callus, multiple shoot and intact plant of strawberry. *Acta Horticulturae*, 319, 291–296.
- Zazimalová, E., Brezinova, A., Holík, J., and Opatrny, Z. (1996). Partial auxin deprivation affects endogenous cytokinins in an auxin-dependent, cytokinin-independent tobacco cell strain. *Plant Cell Reports*, 16(1-2), 76–79.
- Ziv, M. (2010). Silicon effects on growth acclimatization and stress tolerance of bioreactor cultured *Ornithogalum dubium* plants. *Acta Horticulturae*, 865, 29–36.

BIODATA OF STUDENT

Anis Suraya binti Azal was born on April 12th of 1993, in Petaling Jaya, Selangor. She had her primary education at Sekolah Kebangsaan Taman Medan and Sekolah Rendah Agama Kg. Medan. She then continues her secondary schooling at Sekolah Agama Menengah Sg. Merab Luar (SAMSMEL) for only until PMR (lower secondary assessment). She left SAMSMEL to further her study in Sekolah Menengah Kebangsaan Seksyen 18 Shah Alam and sat for SPM (Malaysian Certificate of Education Examination) in 2010. She then started her pre-university study at the age of 18 at Selangor Matriculation College and started her freshman in Universiti Malaysia Terengganu (UMT) two years later in Bachelor of Science in Agrotechnology (Postharvest Technology) and graduated in 2016.



PUBLICATION

Suraya, A. A., Misran, A., and Hakiman, M. (2021). The efficient and easy micropropagation protocol of *Phyllanthus Niruri*. *Plants*, *10*(10), 2141.



6



UNIVERSITI PUTRA MALAYSIA STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

ACADEMIC SESSION : ____

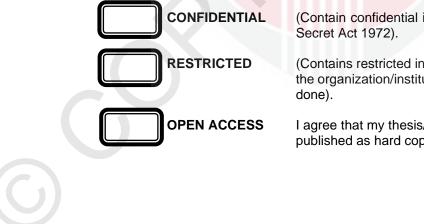
TITLE OF THESIS / PROJECT REPORT :

NAME OF STUDENT :

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

- 1. This thesis/project report is the property of Universiti Putra Malaysia.
- 2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
- 3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as: *Please tick ($\sqrt{}$)



(Contain confidential information under Official Secret Act 1972).

(Contains restricted information as specified by the organization/institution where research was done).

I agree that my thesis/project report to be published as hard copy or online open access.

This thesis is submitted for: PATENT	Embargo from (date)	(date)	_ until
		Approved by:	
(Signature of Student) New IC No/ Passport No.:		(Signature of C of Supervisory Name:	
Date :		Date :	
[Note : If the thesis is CONFI the letter from the organizati confidentially or restricted.]		STRICTED, plea	

G