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CHARACTERIZATION OF PHYSICO-CHEMICAL FEATURES OF TERUNG ASAM SARAWAK (Solanum lasiocarpum Dunal) FRUIT DURING GROWTH, MATURATION AND RIPENING

ALBERT TING KOON SOON

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By

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Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

May 2021

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

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May 2021

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A study on characterizing the changes in physico-chemical, antioxidant activities and cellular structure of "terung asam Sarawak" fruit was conducted on weekly intervals from anthesis until fruit senescence. In this study, fully bloomed flowers of "terung asam" plant were tagged and the fruits were thinned to allow only one per cluster. The experiment was conducted by using a completely randomized design with twelve fruit replications for each week after anthesis (WAA). Logistic regression analysis was used to analyse morphological traits (fruit length, diameter, fresh weight and volume). Data from measurements of respiration rate and ethylene production, colour, moisture content, firmness, soluble solids concentration (SSC), pH, titratable acidity (TA), total phenolic content, total carotenoid content and antioxidant activities were analysed by using analysis of variance whereas means were separated by Duncan's multiple range test (DMRT). Light microscopy (LM) and scanning electron microscopy (SEM) were utilized to document "terung asam" fruit's growth at the cellular level. Results illustrated that "terung asam" fruit took about 115 days or 16 weeks from anthesis until ripening. The fruit exhibits a single sigmoid growth pattern where three physiological stages (S1, S2 and S3) were identified. At S1, cells of "terung asam" fruit went through rapid division, followed by cell expansion at S2 before reaching physiological maturity at S3. These changes were in accordance with cellular structure as observed under LM and SEM. "Terung asam" fruit appeared green in colour at the initial growth stage before it became yellow then orange at later weeks. The visual colour changes of "terung asam" fruit were tally with L^* , a^* and b^* values as fruit matured and ripened. Fruit firmness increased initially and then decreased during ripening whereas SSC and TA increased gradually as WAA progressed. In contrast, fruit pH decreased gradually as WAA progressed. As "terung asam" fruit matured and ripened, citric acid increased but succinic acid decreased. On the other hand, malic acid increased as fruit matured but

decreased when fruit ripened. Respiration and ethylene production rate of "terung asam" fruit were high at the initial stage of growth then reduced and remained constant at later stages. As for total carotenoid content, it was initially low in concentration and the carotenoid pigments started to accumulate as the fruit reached maturity and achieved the highest amount when fruit reached 16 WAA. Total phenolic content decreased at the early growth stage then started to increase as the fruit reached maturation and dropped again as the fruit reached 16 WAA. "Terung asam" fruit at 13 until 15 WAA displayed the highest amount of antioxidant activities, total phenolic content and vitamin C. "Terung asam" fruit's vitamin C at ripening stage can reach about 3 to 4 folds more than that of guava (Psidium guajava L.) fruit. Results from the current study concluded that "terung asam" fruit harvested at 14 WAA could be determined as optimum harvest maturity when the fruit has developed good organoleptic (firmness, moisture, SSC, TA, organic acids, etc.) and quality attributes (vitamin C, carotenoid content, phenolic content and antioxidant activities) during ripening. Since "terung asam" fruit is a promising fruit by having a high amount of vitamin C and total phenolic content (TPC), this fruit need to be intensively commercialized for fruit and food industries to explore and utilize "terung asam" fruit to its full potential. All in all, the physico-chemical, antioxidant activities and cellular structure of "terung asam" fruit changed as it grew, matured and ripened.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

PENCIRIAN FIZIKO-KIMIA BUAH TERUNG ASAM SARAWAK (Solanum lasiocarpum Dunal) SEMASA PERTUMBUHAN, KEMATANGAN DAN PERANUMAN

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Suatu kajian tentang pencirian perubahan fiziko-kimia, aktiviti antioksida dan struktur sel buah terung asam Sarawak telah dijalankan pada selang seminggu dari antesis sehingga penuaan buah. Dalam penelitian ini, bunga terung asam yang mekar sepenuhnya telah ditandakan dan proses penjarangan dilakukan selepas berputik untuk memastikan hanya satu buah yang membesar di setiap jambak. Eksperimen ini dijalankan dengan menggunakan reka bentuk rawak lengkap dengan dua belas replikasi buah untuk setiap minggu selepas antesis (WAA). Ciri-ciri morfologi buah (panjang, diameter, berat dan isipadu) tertakluk kepada analisis regresi logistik. Data dari ukuran kadar pernafasan dan penghasilan etilena, warna, kandungan kelembapan, kekerasan, kepekatan pepejal terlarut (SSC), pH, keasidan titrat (TA), kandungan fenolik, kandungan karotenoid dan aktiviti antioksida telah dianalisa dengan menggunakan kaedah analisis varians manakala min dibezakan dengan menggunakan kaedah ujian pelbagai jarak Duncan (DMRT). Pertumbuhan buah terung asam di peringkat sel telah direkod dengan menggunakan mikroskop cahaya (LM) dan mikroskop pengimbas elektron (SEM). Keputusan kajian ini menunjukkan bahawa buah terung asam mengambil masa kira-kira 115 hari atau 16 minggu dari antesis sehingga ranum. Buah ini telah mempamerkan jenis pola pertumbuhan sigmoid di mana terdapat tiga peringkat fisiologi iaitu S1, S2 dan S3 telah dikenalpasti. Pada peringkat S1, pertumbuhan secara perlahan diperhatikan di mana pembahagian sel-sel giat berlaku. Apabila buah memasuki peringkat S2, pengembangan sel banyak berlaku sebelum buah tersebut mencapai kematangan fisiologi di peringkat S3. Perubahan ini adalah sejajar dengan perubahan struktur selular yang diperhatikan di bawah LM dan SEM. Pada peringkat pertumbuhan yang awal, buah terung asam adalah berwarna hijau sebelum ia berubah menjadi kuning pada peringkat matang dan oren pada

peringkat akhir pertumbuhan. Di sepanjang pertumbuhan, perubahan warna visual buah terung asam adalah sejajar dengan nilai-nilai L*, a* dan b*. Kekerasan buah meningkat pada awal penilaian, kemudian ia menurun di sepanjang proses kematangan manakala SSC dan TA meningkat secara beransur-ansur ketika WAA berkembang. Sebaliknya, pH buah telah menurun secara beransur-ansur ketika WAA berkembang. Asid-asid organik seperti asid askorbik dan asid sitrik telah meningkat secara beransur-ansur ketika kematangan buah meningkat tetapi asid sukinik telah menurun secara beransur-ansur ketika buah matang dan ranum. Sebaliknya, asid malik telah meningkat semasa buah memcapai kematangan tetapi menurun sedikit ketika buah ranum. Peningkatan kedua-dua asid-asid organik dan SSC dalam buah ketika buah mencapai kematangan dan ranum telah membantu buah "terung asam" memiliki rasa masam yang unik. Kadar respirasi dan pengeluaran etilena buah terung asam adalah tinggi pada peringkat awal pertumbuhan sebelum kedua-duanya menurun kekal stabil pada peringkat akhir pertumbuhan. Untuk kandungan karotenoid dalam buah terung asam, ianya didapati hanya pada kadar yang rendah pada peringkat awal pertumbuhan dan pigmen karotenoid mula terkumpul ketika buah semakin matang dan berada pada jumlah yang tertinggi ketika buah mencapai 16 WAA. Jumlah kandungan fenolik menurun pada peringkat awal pertumbuhan dan kemudian meningkat ketika buah telah mencapai kematangan dan turun lagi ketika buah mencapai 16 WAA. Oleh itu, buah terung asam pada 13 WAA hingga 15 WAA telah menunjukkan jumlah aktiviti antioksidan, kandungan fenolik total dan vitamin C yang tertinggi. Kandungan vitamin C dalam buah terung asam apabila mencapai tahap keranuman adalah sekitar 3 hingga 4 kali lebih tinggi daripada buah jambu (Psidium guajava L.). Hasil dari kajian ini juga menunjukkan bahawa buah terung asam yang dituai pada 14 WAA dapat dianggap sebagai kematangan penuaian yang optimum disebabkan buah tersebut telah mempunyai perkembangan organoleptik yang baik (ketegasan, kelembapan, SSC, TA, asid-asid organik dan lain-lain) serta sifat-sifat kualiti (vitamin C, kandungan karotenoid, kandungan fenolik dan aktiviti antioksidan) semasa buah tersebut ranum. Memandangkan buah terung asam mempunyai potensi jumlah vitamin C dan kandungan fenolik (TPC) yang tinggi, buah ini perlu dikomersialkan secara intensif untuk industri buah dan makanan untuk meneroka dan memanfaatkan buah terung asam semaksimum yang mungkin. Secara keseluruhannya, adalah jelas bahawa kualiti buah terung asam berubah- ubah sepanjang ia berkembang, matang dan ranum.

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- 4.25 Changes of total phenolic content in "terung asam" fruit as weeks after anthesis progressed. Means separation (n=12) followed by different letters are significantly different at $P \le 0.05$ according to DMRT.
- 4.26 Changes of antioxidant activities (DPPH, ABTS and FRAP) in "terung asam" fruit as weeks after anthesis progressed. Means separation (n=12) followed by different letters are significantly different at $P \le 0.05$ according to DMRT.

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LIST OF ABBREVIATIONS

	(C ₂ H ₃)NaO ₂ .3H ₂ O	Sodium acetate
	А	Absorbance
	A ^{1%} _{1 cm}	Absorption coefficient
	ABTS	2,2'-Azino-bis(3-ethylbenzothiazoline-6-sulfonic acid)
	ABTS [.] ⁺	ABTS radical cation
	ACC	1-Aminocyclopropane-1-carboxylic acid
	ACO	ACC oxidase
	ACS	ACC synthase
	ANOVA	Analysis of variance
	C_2H_4	Ethylene
	C ₂ H ₄ O ₂	Glacial acetic acid
	CO ₂	Carbon dioxide
	CRD	Completely randomized design
	DPPH	Diphenyl-2-picrylhydrazyl
	DPX	Dibutylphthalate polystyrene xylene
	ΔE_{ab}	Total colour differences
	FeCl ₃ .6H ₂ O	Ferric chloride solution
	FeSO ₄ .7H ₂ O	Ferrous sulphate
	FRAP	Ferric ion reducing antioxidant power
	GAE	Gallic acid equivalent
	H ₂ SO ₄	Sulphuric acid
U	L* LM	Lightness Light microscopy
	DMRT	Duncan's Multiple Ranged Test

Ν	Newton
NaCl	Sodium chloride
NaOH	Sodium hydroxide
O• ²⁻	Superoxide anions
ORAC	Oxygen radical absorbance capacity
PE	Pectinesterase
PG	Polygalacturonase
PL	Pectate lyase
PPO	Polyphenol oxidases
RI	Refractive index
ROO•	Peroxyl radicals
ROS	Reactive oxygen species
SAM	S-adenosyl-L-methionine
SEM	Scanning electron microscope
SSC	Soluble solids concentration
ТА	Titratable acidity
ТЕ	Trolox equivalent
TPC	Total phenolic content
TPTZ	2,4,6-Tris (1-pyridyl)-5-triazine
USDA	United State Department of Agriculture
UV	Ultraviolet
WAA	Week after anthesis
β-GAL	β-Glactosidase

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CHAPTER 1

INTRODUCTION

Terung Asam Sarawak (Solanum lasiocarpum) is one of the indigenous plant or wild fruit vegetable in the state of Sarawak, Malaysia, that belongs to the Solanaceae family of eggplant. It has a unique sour taste and is served as vegetables and flavouring in local dishes which is popular among locals (Dayod and Lim, 2015). Most eggplant, especially this indigenous "terung asam" Sarawak fruit contains a high amount of vitamin C with ample concentration of calcium, fiber, phosphorus and potassium (Jansen et al., 1996). However, it can be considered as underutilized crops due to insufficient genetic materials, germplasm collection, traditional knowledge, knowledge on uses as well as research and development activities (Diengngan and Hasan, 2015). Up to date, there are only a number of "terung asam" accessions in Sarawak where they were assigned by the collector at different origin. For example, "Terung Mas" was collected in Semongok, Hairy in Rampangi, AC. No. 148 in Miri, AC. No. 155 in Mamdi and AC. No. 167 in Ensebang, Balai Ringin (Missnan, 2004). According to Shariah et al. (2013), growing of "terung asam" on intensively used land has been almost impossible due to the presence of bacterial wilt disease.

Nowadays, with the upsurge in demand, high market size and price especially in Sarawak state, "terung asam" plant has been planted as one of the local specialty fruit vegetables throughout the Sarawak. Shariah (2013) stated that the production of "terung asam" fruit was about 16 to 20 tonnes per hectare on average and with market price of about USD 1.43 - 2.39 per kilogram depending on quality and size. The "terung asam" fruit price is on par with other fruit and vegetables such as tomato (USD 2.06) and eggplant (USD 2.11) as reported in the latest average prices of daily commodity at the retail level by the Federal Agriculture Marketing Authority (FAMA, 2021). Hence, fruit growth which refers to a change in fruit physical measurements (weight, length, width, and volume) is of great importance (Valero and Serrano, 2010). According to Opara (2000), fruit growth also involves various physico-chemical and phytochemical changes that take place at both physical and cellular aspects which can be utilized as indices of fruit growth. These changes are associated with the chronological fruit development starting from flowering to maturity and senescence. The obtained cumulative data of the increment in physical measurements can then be used to show a simple or double-sigmoid curve depending on fruit species (Valero and Serrano, 2010).

On the other hand, fruit growth and development are also closely associated with increased cell number, cell size and intercellular air spaces at the cellular level (Valero and Serrano, 2010). Thus, fruit cellular characteristics can also provide additional information to support fruit growth and development. Tadesse et al. (2002) stated that understanding these changes is crucial in determining optimum harvest time which directly affects fruit postharvest life

and quality. The authors further clarified that fruits harvested immature are more susceptible to mechanical damage, sagging and probably lose the ability to ripen properly. On the other hand, harvesting overripe fruits are often not applicable for commercial use due to their shorter shelf life. Therefore, there is a need to understand the process of fruit development and their maturity stages to help growers cope with fruit quality standards of market demand for further exploitation (Fawole and Opara, 2013a).

As to consumer acceptability, consumers tend to choose a better quality of fruit in term of fruit colour, taste and visual aspect (Fawole and Opara, 2013a). Fruit imports and accumulates water, minerals, sugar, amino acids, organic acids during growth and at the same time initiate production of flavour and aroma compounds (Conde et al., 2007). These changes are also closely related to fruit ripening, which can be defined as a process where fruits reach their final maturity and then undergo numerous physical and chemical changes (Thompson, 2003). Therefore, the physico-chemical changes in fruit during growth, such as firmness and colour (physical qualities); sugar, pH and organic acids (chemical qualities); respiration rate and ethylene gas production (physiological qualities) are indeed very essential in deciding the optimum fruit stage for consumer consumption or utilization (Fawole and Opara, 2013a). The group of phytochemicals such as phenolic compounds, flavonoids and vitamin C can also be used as indices of fruit growth since the amount of these phytochemicals is very much associated with fruit growth (Fawole and Opara, 2013b).

Various growth prediction and physico-chemical profiles have been created for crops, including brinjal (*Solanum melongena* L.) (Singh and Sharma, 1990) and tomato (*Lycopersicon esculentum*) (Kaur et al., 2006) vegetable fruit, but there is no such work being done on "terung asam" fruit yet. Most of the researchers working on "terung asam" only focus on cultural practices, fruit chemical composition and potential applications or importance without clearly describing which fruit growth stages that they used in their studies (Voon and Kueh, 1999; Shariah et al., 2013). Therefore, a comprehensive study including physico-chemical, cellular structure and phytochemical changes in "terung asam" fruit from fruit set until senescence was carried out. Understanding these changes will lead to appropriate identification of harvest time in order to maintain fruit quality and finally improve their productivity and profitability. Hence, the objectives of this study were:

- 1. To characterize the growth and development of "terung asam" fruit from fruit set to ripening
- 2. To investigate physico-chemical and phytochemical changes as well as cellular structure from fruit set to ripening.

REFERENCES

- Abbas, M. E. F. and Fandi, B. S. (2002). Respiration rate, ethylene production and biochemical changes during fruit development and maturation of jujube (*Ziziphus mauritiana* Lamk). *Journal of the Science of Food and Agriculture*, 82: 1472-1476.
- Adams-Phillips, L., Barry, C. and Giovannoni, J. (2004). Signal transduction systems regulating fruit ripening. *Trends in Plant Science*, 9: 331-338.
- Addadi, L. and Geva, M. (2003). Molecular recognition at the interface between crystals and biology: generation, manifestation and detection of chirality at crystal surfaces. *Royal Society of Chemistry (CrystEngComm)*, 5(26): 140-146.
- Alexander, L. and Grierson, D. (2002). Ethylene biosynthesis and action in tomato: a model for climacteric fruit ripening. *Journal of experimental botany*, 53(377): 2039-2055.
- Ali, Z. M., Chin, L. H. and Lazan, H. (2004). A comparative study on wall degrading enzymes, pectin modifications and softening during ripening of selected tropical fruits. *Plant Science*, 167: 317-327.
- Almeida, M. M. B., de Sousa, P. H. M., Arriaga, Â. M. C., do Prado, G. M., de Carvalho Magalhães, C. E., Maia, G. A. and de Lemos, T. L. G. (2011). Bioactive compounds and antioxidant activity of fresh exotic fruits from Northeastern Brazil. *Food Research International*, 44(7): 2155-2159.
- Alothman, M., Bhat, R. and Karim, A. A. (2009). Antioxidant capacity and phenolic content of selected tropical fruits from Malaysia, extracted with different solvents. *Food Chemistry*, 115(3): 785-788.
- Alzamora, S. M., Castro, M. A., Vidales, S. L, Nieto, A. B. and Salvatori, D. (2000). The role of tissue microstructure in the textural characteristics of minimally processed fruits. In Alzamora S.M., Tapia M. S. and Lopez-Malo A. (Eds.), *Minimally Processed Fruits and Vegetables* (pp. 153-171). Maryland: Aspen Publishers Inc.
- Anthon, G. E., LeStrange, M. and Barrett, D. M. (2011). Changes in pH, acids, sugars and other quality parameters during extended vine holding of ripe processing tomatoes. *Journal of the Science of Food and Agriculture*, 91(7): 1175-1181.
- Anttonen, M. J., Hoppula, K. I., Nestby, R., Verheul, M. J. and Karjalainen, R. O. (2006). Influence of fertilization, mulch color, early forcing, fruit order, planting date, shading, growing environment, and genotype on the

contents of selected phenolics in strawberry (*Fragaria × ananassa* Duch.) fruits. *Journal of Agricultural and Food Chemistry*, 54(7): 2614-2620.

- Arah, I. K., Amaglo, H., Kumah, E. K. and Ofori, H. (2015). Preharvest and postharvest factors affecting the quality and shelf life of harvested tomatoes: a mini review. *International Journal of Agronomy*, 2015: 10-16.
- Arena, M. E., Postemsky, P. and Curvetto, N. R. (2012). Accumulation patterns of phenolic compounds during fruit growth and ripening of *Berberis buxifolia*, a native Patagonian species. *New Zealand Journal of Botany*, 50(1): 15-28.
- Artés, F., Gómez, P.A. and Artés-Hernández, F. (2006). Modified atmosphere packaging of fruits and vegetables. *Stewart Postharvest Review*, 5: 1-13.
- Bakar, M. F. A., Mohamed, M., Rahmat, A. and Fry, J. (2009). Phytochemicals and antioxidant activity of different parts of bambangan (*Mangifera pajang*) and tarap (*Artocarpus odoratissimus*). *Food Chemistry*, 113(2): 479-483.
- Baldwin, E. A. (2002). Fruit flavor, volatile metabolism and consumer perceptions. In Knee, M. (Ed.), *Fruit quality and its biological basis* (pp. 89-106). United Kingdom: Sheffield Academic Press Limited.
- Balibrea, M. E., Martínez-Andújar, C., Cuartero, J., Bolarín, M. C. and Pérez-Alfocea, F. (2006). The high fruit soluble sugar content in wild *Lycopersicon* species and their hybrids with cultivars depends on sucrose import during ripening rather than on sucrose metabolism. *Functional Plant Biology*, 33(3): 279-288.
- Barragán-Iglesias, J., Méndez-Lagunas, L. L. and Rodríguez-Ramírez, J. (2018). Ripeness indexes and physicochemical changes of papaya (*Carica papaya* L. cv. Maradol) during ripening on-tree. *Scientia Horticulturae*, 236: 272-278.
- Barry, C. and Giovannoni, J. (2007). Ethylene and fruit ripening. *Journal of Plant Growth Regulation*, 26: 143–159.
- Bashir, H. A. and Abu-Goukh, A. B. A. (2003). Compositional changes during guava fruit ripening. *Food Chemistry*, 80(4): 557-563.
- Batista-Silva, W., Nascimento, V. L., Medeiros, D. B., Nunes-Nesi, A., Ribeiro, D. M., Zsögön, A. and Araújo, W. L. (2018). Modifications in organic acid profiles during fruit development and ripening: Correlation or causation? *Frontiers in Plant Science*, 9:1689.

- Beck, C. B. (2010). *An introduction to plant structure and development*. United Kingdom: Cambridge University Press, Cambridge.
- Berüter, J. (2004). Carbohydrate metabolism in two apple genotypes that differ in malate accumulation. *Journal of Plant Physiology*, 161: 1011–1029.
- Biondi, M. (2007). *Dynamics of grape berry volume change during ripening* (Unpublished Doctoral dissertation). Washington State University.
- Blanke, M. M. and Lenz, F. (1989). Fruit photosynthesis. *Plant, Cell and Environment*, 12(1): 31-46.
- Bouaziz, M., Chamkha, M. and Sayadi, S. (2004). Comparative study on phenolic content and antioxidant activity during maturation of the olive cultivar Chemlali from Tunisia. *Journal of Agricultural and Food Chemistry*, 52(17): 5476-5481.
- Bouzayen, M., Latché, A., Nath, P. and Pech, J. C. (2010). Mechanism of fruit ripening – Chapter 16. In E. C. Pua, M. R. Davey (Eds.), *Plant developmental biology-Biotechnological perspectives*, 1: 319-339. Springer Berlin Heidelberg.
- Bunea, A., Rugină, D., Sconţa, Z., Pop, R. M., Pintea, A., Socaciu, C., Tabaran, F., Grootaert, C., Struijs, K. and VanCamp, J. (2013). Anthocyanin determination in blueberry extracts from various cultivars and their antiproliferative and apoptotic properties in B16-F10 metastatic murine melanoma cells. *Phytochemistry*, 95:436-444.
- Cara, B. and Giovannoni, J. J. (2008). Molecular biology of ethylene during tomato fruit development and maturation. *Plant Science*, 175(1-2): 106-113.
- Castaneda-Ovando, A., de Lourdes Pacheco-Hernández, M., Páez-Hernández, M. E., Rodríguez, J. A. and Galán-Vidal, C. A. (2009). Chemical studies of anthocyanins: A review. *Food Chemistry*, 113(4): 859-871.
- Cerretani, L. and Bendini, A. (2010). Rapid assays to evaluate the antioxidant capacity of phenols in virgin olive oil. *In Olives and olive oil in health and disease prevention* (pp. 625-635). Academic Press.
- Cetin, A. and Geçibesler, I. H. (2015). Evaluation as antioxidant agents of 1, 2, 4-triazole derivatives: effects of essential functional groups. *Journal of Applied Pharmaceutical Science*, 5(6): 120-126.
- Chai, X. F. and Ding, P. (2012). *Postharvest quality and antioxidant activity of Karanda (Carissa carandas) fruit.* In VII International Postharvest Symposium 1012 (pp. 177-182).

- Chander, M. P., Kartick, C., Gangadhar, J. and Vijayachari, P. (2014). Ethno medicine and healthcare practices among Nicobarese of Car Nicobar– An indigenous tribe of Andaman and Nicobar Islands. *Journal of Ethnopharmacology*, 158:18-24.
- Conde, C., Silva, P., Fontes, N., Dias, A. C. P., Tavares, R. M., Sousa, M. J., Agasse, A., Delrot, S. and Gerós, H. (2007). Biochemical changes throughout grape berry development and fruit and wine quality. *Food*, 1: 1-22.
- Contreras-Calderón, J., Calderón-Jaimes, L., Guerra-Hernández, E. and García-Villanova, B. (2011). Antioxidant capacity, phenolic content and vitamin C in pulp, peel and seed from 24 exotic fruits from Colombia. *Food Research International*, 44(7): 2047-2053.
- Corner, E. J. H. (1976). *The Seeds of Dicotyledons: Volume 1*. Cambridge University Press, Cambridge.
- Cortez, P. A. and Carmello-Guerreiro, S. M. (2008). Ontogeny and structure of the pericarp and the seed coat of *Miconia albicans* (Sw.) Triana (Melastomataceae) from" cerrado", Brazil. *Brazilian Journal of Botany*, 31(1): 71-79.
- Crisosto, G. M., Crisosto, C. H., Echeverría, G. and Puy, J. (2007). Segregation of plum and pluot cultivars according to their organoleptic characteristics. *Postharvest Biology and Technology*, 44(3): 271–276.
- Dai, J. and Mumper, R. J. (2010). Plant phenolics: extraction, analysis and their antioxidant and anticancer properties. *Molecules*, 15(10): 7313-7352.
- Dayod, M. and Lim, L., 2015. *Graft compatibility between Solanum lasiocarpum and Solanum torvum: As a potential tool to mitigate soil-borne diseases.* 4th International Conference on Agriculture & Horticulture, 13-15.
- De Jong, M., Wolters-Arts, M., Feron, R., Mariani, C. and Vriezen, W. H. (2009). The *Solanum lycopersicum* auxin response factor 7 (SIARF7) regulates auxin signaling during tomato fruit set and development. *The Plant Journal*, 57(1): 160-170.
- Dennis, J. F. G. (1988). Fruit development. In Tesar, M. B. (Ed.), *Physiological Basis of Crop Growth and Development* (pp. 273). Madison: American Society of Agronomy.
- Desouky, I. M., Shaltout, A. D., Haggag, L. F. and Farahat, E. M. M. (2016). Changes in some constituents of Kalamata and Picual olive fruits during development. *International Journal of ChemTech Research*, 9(6): 17-22.

- Diakou, P., Svanella, L., Raymond, P., Gaudillère, J. P. and Moing, A. (2000). Phosphoenolpyruvate carboxylase during grape berry development: protein level, enzyme activity and regulation. *Functional Plant Biology*, 27(3): 221-229.
- Díaz-Mula, H. M., Zapata, P. J., Guillén, F., Castillo, S., Martínez-Romero, D., Valero, D. and Serrano, M. (2008). Changes in physicochemical and nutritive parameters and bioactive compounds during development and on-tree ripening of eight plum cultivars: A comparative study. *Journal of the Science of Food and Agriculture*, 88: 2499–2507.
- Diengngan, S. and Hasan, M. A. (2015). Genetic diversity of underutilized fruits in India for environmental sustainability. *Advances in Plants and Agriculture Research*, 2(7): 76-83.
- Ding, P. and Mashah, N. C. (2016). Growth, maturation and ripening of underutilized *Carissa congesta* fruit. *Fruits*, 71(3): 171-176.
- Ding, P. and Syazwani, S. (2016). Physicochemical quality, antioxidant compounds and activity of MD-2 pineapple fruit at five ripening stages. *International Food Research Journal*, 23(2): 549.
- Ding, P., Ahmad, S. H. and Ghazali, H. M. (2007). Changes in selected quality characteristics of minimally processed carambola (*Averrhoa carambola* L.) when treated with ascorbic acid. *Journal of the Science of Food and Agriculture*, 87(4): 702-709.
- Ding, P., Ahmad, S. H., Razak, A. R. A., Saari, N. and Mohamed, M. T. M. (2010). Plastid ultrastructure, chlorophyll contents, and colour expression during ripening of cavendish banana (*Musa acuminate* 'Williams') at 18° C and 27° C. *New Zealand Journal of Crop and Horticultural Science*, 35(2): 201-210.
- Djeridane, A., Yousfi, M., Nadjemi, B., Boutassouna, D., Stocker, P. and Vidal,
 N. (2006). Antioxidant activity of some Algerian medicinal plants extracts containing phenolic compounds. *Food Chemistry*, 97(4): 654-660.
- Døving, A. and Måge, F. (2002). Methods of testing strawberry fruit firmness. Acta Agriculturae Scandinavica, Section B, Soil and Plant Science, 52(1): 43-51.
- Dragovic-uzelac, V., Levaj, B., Mrkic, V., Bursac, D. and Boras, M. (2007). The content of polyphenols and carotenoids in three apricot cultivars depending on stage of maturity and geographical region. *Food Chemistry*. 102: 966–975.

- Elkington, B. G., Sydara, K., Newsome, A., Hwang, C. H., Lankin, D. C., Simmler, C., Napolitano, J. G., Ree, R., Graham, J. G., Gyllenhaal, C., Bouamanivong, S., Souliya, O., Pauli, G. F., Franzblau, S. G. and Soejarto, D. D. (2014). New finding of an anti-TB compound in the genus *Marsypopetalum* (Annonaceae) from a traditional herbal remedy of Laos. *Journal of Ethnopharmacology*, 151(2): 903-911.
- Erel, O. (2004). A novel automated direct measurement method for total antioxidant capacity using a new generation, more stable ABTS radical cation. *Clinical Biochemistry*, 37(4): 277-285.
- Ezura, H. and Hiwasa-Tanase, K. (2010). Fruit development. In Pua, E. C. and Davey, M. R. (Eds.), *Plant Developmental Biology-Biotechnological Perspectives*, (pp. 301-318). Berlin Heidelberg: Springer-Verlag.
- Fadda, A. and Mulas, M. (2010). Chemical changes during myrtle (*Myrtus communis* L.) fruit development and ripening. *Scientia Horticulturae*, 125(3): 477-485.
- FAMA, Federal Agriculture Marketing Authority (2021). Report on latest average prices of daily commodity at retail levels. Retrieved on January 9, 2021 from FAMA Website: https://sdvi2.fama.gov.my/price/direct/price/daily_commodityRpt.asp?Pri cing=A&LevelCd=03&PricingDt=2021/1/7&PricingDtPrev=2021/1/7
- Farahzety, A. M. (2016). Current status and future prospects of vegetable grafting in Malaysia. Conference: International Workshop on Grafting to Improve Fruit-Vegetable Production, 121-124.
- Farinati, S., Rasori, A., Varotto, S. and Bonghi, C. (2017). Rosaceae fruit development, ripening and post-harvest: An epigenetic perspective. *Frontiers In Plant Science*, 8: 1247.
- Fawole, O. A. and Opara, U. L. (2013a). Fruit growth dynamics, respiration rate and physico-textural properties during pomegranate development and ripening. *Scientia Horticulturae*, 157: 90-98.
- Fawole, O. A. and Opara, U. L. (2013b). Changes in physical properties, chemical and elemental composition and antioxidant capacity of pomegranate (cv. Ruby) fruit at five maturity stages. *Scientia Horticulturae*, 150: 37-46.
- Fawole, O. A. and Opara, U. L. (2013c). Developmental changes in maturity indices of pomegranate fruit: A descriptive review. *Scientia Horticulturae*, 159: 152-161.

- Fernández-Panchón, M. S., Villano, D., Troncoso, A. M. and García-Parrilla, M. C. (2008). Antioxidant activity of phenolic compounds: From *in vivo* results to *in vitro* evidence. *Critical Reviews in Food Science and Nutrition*, 24: 649-671.
- Ferrer, A., Remón, S., Negueruela, A. I. and Oria, R. (2005). Changes during the ripening of the very late season Spanish peach cultivar Calanda: feasibility of using CIELAB coordinates as maturity indices. *Scientia Horticulturae*, 105(4): 435-446.
- Findlay, N., Oliver, K. J., Nil, N. and Coombe, B. G. (1987). Solute accumulation by grape pericarp cells: IV. Perfusion of pericarp apoplast via the pedicel and evidence for xylem malfunction in ripening berries. *Journal of Experimental Botany*, 38(4): 668-679.
- Fischer, G., Almanza-Merchán, P. J. and Ramírez, F. (2012). Source-sink relationships in fruit species: A review. *Revista Colombiana de Ciencias Hortícolas*, 6(2): 238-253.
- Fischer, U. A., Carle, R. and Kammerer, D. R. (2011). Identification and quantification of phenolic compounds from pomegranate (*Punica* granatum L.) peel, mesocarp, aril and differently produced juices by HPLC-DAD–ESI/MSn. Food Chemistry, 127(2): 807-821.
- Frenkel, C. and Hartman, T. G. (2012). Decrease in fruit moisture content heralds and might launch the onset of ripening processes. *Journal of Food Science*, 77(10): S365-76.
- Gapper, N. E., McQuinn, R. P. and Giovannoni, J. J. (2013). Molecular and genetic regulation of fruit ripening. *Plant Molecular Biology*, 82(6), 575-591.
- Gardner, P. T., White, T. A., McPhail, D. B. and Duthie, G. G. (2000). The relative contributions of vitamin C, carotenoids and phenolics to the antioxidant potential of fruit juices. *Food Chemistry*, 68(4): 471-474.
- Ghasemnezhad, M., Sherafati, M. and Payvast, G. A. (2011). Variation in phenolic compounds, ascorbic acid and antioxidant activity of five coloured bell pepper (*Capsicum annum*) fruits at two different harvest times. *Journal of Functional Foods*, 3(1): 44-49.
- Gil, M. I., Tomas-Barberan, F. A., Hess-Pierce, B., Holeroft, D. M. and Kader, A. A. (2000). Antioxidant activity of pomegranate juice and its relationship with phenolic composition and processing. *Journal of Agriculture and Food Chemistry*, 48: 4581–4589.

- Giovannoni, J. (2001). Molecular biology of fruit maturation and ripening. Annual Review of Plant Physiology and Plant Molecular Biology, 52: 725–749.
- Giovannoni, J. (2004). Genetic regulation of fruit development and ripening. *The Plant Cell*, 16: 170-180.
- Glew, R. H., Ayaz, F. A., Sanz, C., VanderJagt, D. J., Huang, H. S., Chuang, L. T. and Strnad, M. (2003). Changes in sugars, organic acids and amino acids in medlar (*Mespilus germanica* L.) during fruit development and maturation. *Food Chemistry*, 83(3): 363-369.
- Goiris, K., Muylaert, K., Fraeye, I., Foubert, I., De Brabanter, J. and De Cooman, L. (2012). Antioxidant potential of microalgae in relation to their phenolic and carotenoid content. *Journal of Applied Phycology*, 24(6): 1477-1486.
- Goulao, L. F. and Oliveira, C. M. (2008). Cell wall modifications during fruit ripening: When a fruit is not the fruit. *Trends in Food Science and Technology*, 19(1): 4-25.
- Guillén, F., Castillo, S., Zapata, P.J., Martínez-Romero, D., Valero, D. and Serrano, M. (2006). Efficacy of 1-MCP treatment in tomato fruit. 2. Effect of cultivar and ripening stage at harvest. *Postharvest Biology and Technology*, 42(3): 235-242.
- Gundogdu, M., Ozrenk, K., Ercisli, S., Kan, T., Kodad, O. and Hegedus, A. (2014). Organic acids, sugars, vitamin C content and some pomological characteristics of eleven hawthorn species (*Crataegus* spp.) from Turkey. *Biological Research*, 47: 1-5.
- Guo, C., Yang, J., Wei, J., Li, Y., Xu, J. and Jiang, Y. (2003). Antioxidant activities of peel, pulp and seed fractions of common fruits as determined by FRAP assay. *Nutrition Research*, 23: 1719–1726.
- Gupta, D. and Garg, S. (1966). Isolation of solanine from the fruits of *Solanum ferox*. *Naturwissenschaften*, 53: 108-109.
- Haggag, L., Shahin, M. F. M., Genaidy, E. A. E. and Fouad, A. A. (2013). Changes in Fruit weight, dry matter, moisture content and oil percentage during fruit development stages of two olive cultivars. *Middle East Journal*, 2(1): 21-27.
- Halliwell, B. (2007). Dietary polyphenols: Good, bad, or indifferent for your health? *Cardiovascular Research*, 73: 341–347.

- Harada, T., Kurahashi, W., Yanai, M., Wakasa, Y. and Satoh, T. (2005). Involvement of cell proliferation and cell enlargement in increasing the fruit size of Malus species. *Scientia Horticulturae*, 105(4): 447-456.
- Harrill, R. (1998). Using a refractometer to test the quality of fruits and vegetables. Pineknoll Publishing, Keedysville, Maryland, United States.
- Hernández, Y., Lobo, M. G. and González, M. (2006). Determination of vitamin C in tropical fruits: A comparative evaluation of methods. *Food Chemistry*, 96(4): 654-664.
- Holcroft, D. (2015). Water relations in harvested fresh produce. *The Postharvest Education Foundation (PEF),* 15: 01.
- Hossan, M. S., Roy, P., Seraj, S., Mou, S. M., Monalisa, M. N., Jahan, S., Khan, T., Swarna, A., Jahan, R., and Rahmatullah, M. (2012).
 Ethnomedicinal knowledge among the Tongchongya tribal community of Roangchaari Upazila of Bandarban district, Bangladesh. *American-Eurasian Journal of Sustainable Agriculture*, 6(4): 349-359.
- Huang, D., Ou, B. and Prior, R. L. (2005). The chemistry behind antioxidant capacity assays. *Journal of Agricultural and Food Chemistry*, 53(6): 1841-1856.
- Ignat, T., Schmilovitch, Z., Feföldi, J., Bernstein, N., Steiner, B., Egozi, H. and Hoffman, A. (2013). Nonlinear methods for estimation of maturity stage, total chlorophyll, and carotenoid content in intact bell peppers. *Biosystems Engineering*, 114(4): 414-425.
- Ikoma, Y., Komatsu, A., Kita, M., Ogawa, K., Omura, M., Yano, M. and Moriguchi, T. (2001). Expression of a phytoene synthase gene and characteristic carotenoid accumulation during citrus fruit development. *Physiologia Plantarum*, 111(2): 232-238.
- Ilarslan, H., Palmer, R. G., Imsande, J. and Horner, H. T. (1997). Quantitative determination of calcium oxalate and oxalate in developing seeds of soybean (Leguminosae). *American Journal of Botany*, 84: 1042-1046.
- Irawan, D., Wijaya, C. H., Limin, S. H., Hashidoko, Y., Osaki, M. and Kulu, I. P. (2006). Ethnobotanical study and nutrient potency of local traditional vegetables in Central Kalimantan. *Tropics*, 15(4): 441-448.
- Ishikawa, T., Dowdle, J. and Smirnoff, N. (2006). Progress in manipulating ascorbic acid biosynthesis and accumulation in plants. *Physiologia Plantarum*, 126: 343-355.

- Jamaludin, N. A., Ding, P. and Hamid, A. A. (2011). Physico-chemical and structural changes of red-fleshed dragon fruit (*Hylocereus polyrhizus*) during fruit development. *Journal of the Science of Food and Agriculture*, 91(2): 278-285.
- Jansen, P. C. M., Wilk, C. V. D. and Hetterscheid, W. L. A. (1996). *Plants yielding non-seed carbohydrates.* Leiden: Backhuys Publisher, PROSEA Foundation.
- Judd, W. S., Campbell, C. S., Kellogg, E. A. and Stevens, P. F. (1999). Plant systematics: A phylogenetic approach. *Ecologia Mediterranea*, 25(2): 215.
- Kader, A. A. (2002). *Postharvest technology of horticultural crops* (Vol. 3311). University of California Agriculture and Natural Resources, United States.
- Kala, A. and Prakash, J. (2006). The comparative evaluation of the nutrient composition and sensory attributs of four vegetables cooked by different methods. *International Journal of Food Science and Technology*, 41: 163-171.
- Kamis, A. B., Modu, A. S. and Mwajim, B. (2004). Effect of ripening on the proximate and some biochemical composition of a local tomato cultivar (Nadaffreta) grown at Lake Alau region of Borno state. *Journal of Applied Sciences*, 4: 424-426.
- Kaur, D., Sharma, R., Abas Wani, A., Singh Gill, B. and Sogi, D. S. (2006). Physicochemical changes in seven tomato (*Lycopersicon esculentum*) cultivars during ripening. *International Journal of Food Properties*, 9(4): 747-757.
- Kawabata, S., Sasaki, H. and Sakiyama, R. (2005). Role of transpiration from fruits in phloem transport and fruit growth in tomato fruits. *Physiologia Plantarum*, 124(3):371-380.
- Kays, S. J. and Paull, R. E. (2004). *Metabolic process in harvested products.* Athens, GA: Exon Press.
- Kennedy, D. O. (2014). Polyphenols and the human brain: Plant "secondary metabolite" ecologic roles and endogenous signaling functions drive benefits. *Advances in Nutrition*, 5(5): 515-533.
- Kheng, T. Y., Ding, P. and Abdul Rahman, N. A. (2012). Determination of optimum harvest maturity and physico-chemical quality of Rastali banana (*Musa* AAB Rastali) during fruit ripening. *Journal of the Science* of Food and Agriculture, 92(1): 171-176.

- Khoo, H. E., Prasad, K. N., Kong, K. W., Jiang, Y. and Ismail, A. (2011). Carotenoids and their isomers: Color pigments in fruits and vegetables. *Molecules*, 16(2): 1710-1738.
- Kim, S. K., Bae, R. N., Na, H., Dal Ko, K. and Chun, C. (2013). Changes in physicochemical characteristics during fruit development in June-bearing strawberry cultivars. *Horticulture*, *Environment*, and *Biotechnology*, 54(1): 44-51.
- Klatt, B. K., Holzschuh, A., Westphal, C., Clough, Y., Smit, I., Pawelzik, E. and Tscharntke, T. (2014). Bee pollination improves crop quality, shelf life and commercial value. *Proceedings of the Royal Society B: Biological Sciences*, 281(1775): 2013-2440.
- Kris-Etherton, P. M., Hecker, K. D., Bonanome, A., Koval, S. M., Binkoski, A. M., Hilpert, K. F., Griel, A. E. and Etherton, T. D. (2002). Bioactive compounds in foods: Their role in the prevention of cardiovascular disease and cancer. *The American Journal of Medicine*, 113(9): 71–88.
- Kruawan, K. and Kangsadalampai, K. (2006). Antioxidant activity, phenolic compound contents and antimutagenic activity of some water extract of herbs. *Thai Journal of Pharmaceutical Sciences*, 30(2): 28-35.
- Ku, V. V. V., Shohet, D., Wills, R. B. H. and Kim, G. H. (1999). Importance of low ethylene levels to delay senescence of non-climacteric fruit and vegetables. *Australian Journal of Experimental Agriculture*, 39(2): 221-224.
- Lara, I., Belge, B. and Goulao, L. F. (2014). The fruit cuticle as a modulator of postharvest quality. *Postharvest Biology and Technology*, 87: 103-112.
- Lasbrook, C. C. (2005). New insights into cell disassembly during fruit ripening. *Stewart Postharvest Review*, 3(2): 1-18.
- Lee, S. H., Jeong, Y. S., Song, J., Hwang, K. A., Noh, G. M. and Hwang, I. G. (2017). Phenolic acid, carotenoid composition, and antioxidant activity of bitter melon (*Momordica charantia* L.) at different maturation stages. *International Journal of Food Properties*, 20(3): 3078-3087.
- Lee, S. K. and Kader, A. A. (2000). Preharvest and postharvest factors influencing vitamin C content of horticultural crops. *Postharvest Biology and Technology*, 20(3): 207–220.
- Leonardi, C., Baille, A. and Guichard, S. (2000). Predicting transpiration of shaded and non-shaded tomato fruits under greenhouse environments. *Scientia Horticulturae*, 84: 297-307.

- Lester, G. E. (2008). Antioxidant, sugar, mineral, and phytonutrient concentrations across edible fruit tissues of orange-fleshed Honeydew melon (*Cucumis melo* L.). *Journal of Agricultural and Food Chemistry*, 56: 3694-3698.
- Lim, T. K. (2013). Solanum lasiocarpum. Edible Medicinal and Non-medicinal *Plants*. Netherlands: Springer.
- Ling, Y. S. (2012). *Study on the antioxidant constituents of the leaves of Solanum lasiocarpum*. (Unpublished Doctoral dissertation). Nanfang University of Technology.
- Lobit, P., Génard, M., Wu, B. H., Soing, P. and Habib, R. (2003). Modelling citrate metabolism in fruits: Responses to growth and temperature. *Journal of Experimental Botany*, 54: 2489-2501.
- Løkke, M. M., Seefeldt, H. F., Edwards, G. and Green, O. (2011). Novel wireless sensor system for monitoring oxygen, temperature and respiration rate of horticultural crops post-harvest. *Sensors*, 11(9): 8456-8468.
- Luthria, D. L. and Mukhopadhyay, S. (2006). Influence of sample preparation on assay of phenolic acids from eggplant. *Journal of Agricultural and Food Chemistry*, 54(2006): 41-47.
- Mahmood, T., Anwar, F., Abbas, M. and Saari, N. (2012). Effect of maturity on phenolics (phenolic acids and flavonoids) profile of strawberry cultivars and mulberry species from Pakistan. *International Journal of Molecular Sciences*, 13(4): 4591-4607.
- Marina, I. H., Velumuttu, O. and Pekka, E. K. (1989). Carotenoids in finish foods: Vegetables, fruits and berries. *Journal of Agricultural and Food Chemistry*, 37:655-659.
- Martínez, S., López, M., González-Raurich, M. and Bernardo Alvarez, A. (2005). The effects of ripening stage and processing systems on vitamin C content in sweet peppers (*Capsicum annuum* L.). *International Journal of Food Sciences and Nutrition*, 56(1): 45-51.
- Matthews, M. A. and Shackel, K. A. (2005). Growth and water transport in fleshy fruit. *In Vascular Transports in Plants, Academic press,* pp. 181-197.
- May, J. M. (1999). Is ascorbic acid an antioxidant for the plasma membrane? Federation of American Societies Experimental Biology Journal, 13: 995-1006.

- Mditshwa, A., Magwaza, L. S., Tesfay, S. Z. and Opara, U. L. (2017). Postharvest factors affecting vitamin C content of citrus fruits: A review. *Scientia Horticulturae*, 218: 95-104.
- Medlicott, A. P., Bhogal, M. and Reynolds, S. B. (1986). Changes in peel pigmentation during ripening of mango fruit (*Mangifera indica* var. Tommy Atkins). *Annals of Applied Biology*, 109(3): 651-656.
- Mena, P., García-Viguera, C., Navarro-Rico, J., Moreno, D. A., Bartual, J., Saura, D. and Martí, N. (2011). Phytochemical characterisation for industrial use of pomegranate (*Punica granatum* L.) cultivars grown in Spain. *Journal of Science and Food Agriculture*, 91(10): 1893-1906.
- Mercado-Silva, E., Benito-Bautista, P. and de los Angeles García-Velasco, M. (1998). Fruit development, harvest index and ripening changes of guavas produced in central Mexico. *Postharvest Biology and Technology*, 13(2): 143-150.
- Metcalfe, C. R. and Chalk, L. (1985). *Anatomy of the Dicotyledons: Wood Structure and Conclusion of the General Introduction.* Great Britain: The Clarendon.
- Meyer, R. S., Bamshad, M., Fuller, D. Q. and Litt, A. (2014). Comparing medicinal uses of eggplant and related Solanaceae in China, India, and the Philippines suggests the independent development of uses, cultural diffusion, and recent species substitutions. *Economic Botany*, 68(2): 137-152.
- Miccolis, V. and Saltveit, M. E. (1991). Morphological and physiological changes during fruit growth and maturation of seven melon cultivars. *Journal of the American Society for Horticultural Science*, 116(6): 1025-1029.
- Missnan, R. (2004). Morphological Characterization of Local Accessions of Terung Dayak (*Solanum lasiocarpum* Dunal). Doctoral dissertation, Universiti Malaysia Sarawak.
- Mohammad, M. and Ding, P. (2019). Physico-textural and cellular structure changes of *Carissa congesta* fruit during growth and development. *Scientia Horticulturae*, 246: 380-389.
- Mohanan, N. and George, K.V. (2004). *Cultivated crops and their wild relatives of Kerala State.* Conference: National Seminar on Biodiversity Conservation and Environment Management, At Catholicate College, Pathanamthitta, Kerala.

- Moing, A., Rothan, C., Svanella, L., Just, D., Diakou, P., Raymond, P., Gaudillère, J. P. and Monet, R. (2000). Role of phosphoenolpyruvate carboxylase in organic acid accumulation during peach fruit development. *Physiologia Plantarum*, 108(1): 1-10.
- Moulehi, I., Bourgou, S., Ourghemmi, I. and Tounsi, M. S. (2012). Variety and ripening impact on phenolic composition and antioxidant activity of mandarin (*Citrus reticulate* Blanco) and bitter orange (*Citrus aurantium* L.) seeds extracts. *Industrial Crops and Products*, 39: 74-80.
- Mphahlele, R. R., Stander, M. A., Fawole, O. A. and Opara, U. L. (2014). Effect of fruit maturity and growing location on the postharvest contents of flavonoids, phenolic acids, vitamin C and antioxidant activity of pomegranate juice (cv. Wonderful). *Scientia Horticulturae*, 179: 36-45.
- Muhammad, A., Sallem, U., Sharmsur, R., Zia, U. and Muhammad, A. (2014). Comparison of different types of watermelon and their important nutrients. *Journal of Biology Agriculture and Healthcare*, 4(14): 59-65.
- Muhammad, H. and Ding, P. (2007). Cellular changes during fruit ripening of Harumanis mango. *Malaysian Journal of Microscopy*, 3: 19-24.
- Oms-Oliu, G., Aguiló-Aguayo, I., Soliva-Fortuny, R. and Martín-Belloso, O. (2009). Effect of ripeness at processing on fresh-cut 'Flor de Invierno'pears packaged under modified atmosphere conditions. *International Journal of Food Science and Technology*, 44(5): 900-909.
- Opara, L. U. (2000). Fruit growth measurement and analysis. *Horticultural Reviews*, 24: 373-431.
- Ozgen, M., Drugac, C., Serce, S. and Kaya, C. (2008). Chemical and antioxidant properties of pomegranate cultivars grown in Mediterranean region of Turkey. *Food Chemistry*, 111(3): 703-706.
- Paiva, E. A. S. and Machado, S. R. (2005). Role of intermediary cells in *Peltodon radicans* (Lamiaceae) in the transfer of calcium and formation of calcium oxalate crystals. *Brazilian Archives of Biology and Technology*, 48(1):147-153.
- Paliyath, G. and Murr, D. P. (2006). *Biochemistry of Fruits.* In Paliyath, G., Murr, D. P., Handa, A. K. and Lurie, S. (Eds.) Postharvest Biology and Technology of Fruits, Vegetables, and Flowers (pp. 19-50). USA: Wiley-Blackwell Publishing.
- Paniagua, A. C., East, A. R., Hindmarsh, J. P. and Heyes, J. (2013). Moisture loss is the major cause of firmness change during postharvest storage of blueberry. *Postharvest Biology and Technology*, 79: 13-19.

- Pateiro, M., Barba, F. J., Domínguez, R., Sant'Ana, A. S., Khaneghah, A. M., Gavahian, M., Gómez, B. and Lorenzo, J. M. (2018). Essential oils as natural additives to prevent oxidation reactions in meat and meat products: A review. *Food Research International*, 113: 156-166.
- Paula, M. P. (2003). Cryopreservation of Solanum lasiocarpum Dunal using vitrification technique. Ph.D. Thesis. Universiti Malaysia Sarawak (Unpublished).
- Pérez-Jiménez, J., Arranz, S., Tabernero, M., Díaz-Rubio, M. E., Serrano, J., Goñi, I. and Saura-Calixto, F. (2008). Updated methodology to determine antioxidant capacity in plant foods, oils and beverages: Extraction, measurement and expression of results. *Food Research International*, 41(3): 274-285.
- Perkins-Veazie, P. (1995). Growth and ripening of strawberry fruit. *Horticultural Reviews*, 17: 267-297.
- Perkins-Venzie, P. and Collins, J. K. (2001). Contributions of non-volatile phytochemicals to nutrition and flavour. *HortTechnology*, 11: 539-546.
- Perotti, V. E., Moreno, A. S. and Podestá, F. E. (2014). Physiological aspects of fruit ripening: the mitochondrial connection. *Mitochondrion*, 17: 1-6.
- Prasanna, V., Prabha, T. N. and Tharanathan, R. N. (2007). Fruit ripening phenomena–An overview. *Critical Reviews in Food Science and Nutrition*, 47(1): 1-19.
- Rahman, Z. A., Zaidan, M. W. A. M., Othman, A. N., Ahmad, M. A., Simoh, S. and Ismail, M. A. H. (2019). Optimizing extraction of phenolics and flavonoids from *Solanum ferox* fruit. *Natural Science*, 11(4): 99-105.
- Rančić, D., Quarrie, S. P. and Pećinar, I. (2010). Anatomy of tomato fruit and fruit pedicel during fruit development. *Microscopy: Science, Technology, Applications and Education*, 2: 851-861.
- Ranganna, S. (1997). *Manual of Analysis of Fruit and Vegetable Products* (pp. 9-82). New Delhi: Tata McGraw-hill Publishing Company Limited.
- Reddy, V., Urooj, A. and Kumar, A. (2005). Evaluation of antioxidant activity of some plant extracts and their application in biscuits. *Food Chemistry*, 90(1-2): 317-321.
- Richardson, A. C., Boldingh, H. L., McAtee, P. A., Gunaseelan, K., Luo, Z., Atkinson, R. G., David, K. M., Burdon, J. N. and Schaffer, R. J. (2011).

Fruit development of the diploid kiwifruit, *Actinidia chinensis* 'Hort16A'. *BMC Plant Biology*, 11(1): 1-14.

- Rieder, H. (1899). *Atlas of Urinary Sediments*. Glasgow, Scotland: Charles Griffin and Company.
- Roch, L., Prigent, S., Klose, H., Cakpo, C. B., Beauvoit, B., Deborde, C., Fouillen, L., Delft, P.V., Jacob, D., Usadel, B., Dai, Z., Genard, M., Vercambre, G., Colombie, S., Moing, A., & Gibon, Y. (2020). Biomass composition explains fruit relative growth rate and discriminates climacteric from non-climacteric species. *Journal of Experimental Botany*, 71(19): 5823-5836.
- Rodrigo, M. J., Marcos, J. F. and Zacarías, L. (2004). Biochemical and molecular analysis of carotenoid biosynthesis in flavedo of orange (*Citrus sinensis* L.) during fruit development and maturation. *Journal of Agricultural and Food Chemistry*, 52(22): 6724-6731.
- Rodriguez, S. D. C., López, B. and Chaves, A. R. (1999). Changes in polyamines and ethylene during the development and ripening of eggplant fruits (*Solanum melongena*). *Journal of Agricultural and Food Chemistry*, **47**: 1431-1434.
- Ruffner, H. P., Possner, D., Brem, S. and Rast, D. M. (1984). The physiological role of malic enzyme in grape ripening. *Planta*, 160(5): 444-448.
- Rymbai, H., Srivastav, M., Sharma, R. R. and Singh, S. K. (2012). Lenticels on mango fruit: Origin, development, discoloration and prevention of their discoloration. *Scientia Horticulturae*, 135: 164-170.
- Sablani, S. S., Opara, L. U. and Al-Balushi, K. (2006). Influence of bruising and storage temperature on vitamin C content of tomato fruit. *Journal of Food Agriculture and Environment*, 4(1): 54.
- Safafar, H., Van Wagenen, J., Møller, P. and Jacobsen, C. (2015). Carotenoids, phenolic compounds and tocopherols contribute to the antioxidative properties of some microalgae species grown on industrial wastewater. *Marine Drugs*, 13(12): 7339-7356.
- Saikia, B., Borthakur, S. and Saikia, N. (2010). Medico-ethnobotany of Bodo tribals in Gohpur of Sonitpur district, Assam. *Indian Journal of Tranditional Knowledge*, 9: 52-54.
- Salvador, A., Arnal, L., Besada, C., Larrea, V., Quiles, A. and Pérez-Munuera, I. (2007). Physiological and structural changes during ripening and deastringency treatment of persimmon fruit cv.'Rojo Brillante'. *Postharvest Biology and Technology*, 46(2): 181-188.

- Samuels, J. (2012). *Genetically engineered Bt. brinjal and the implications for plant biodiversity.* Cornwall, United Kingdom: Novel Solanaceae Crops Project.
- Sarkar, N., Srivastava, P. K. and Dubey, V. K. (2009). Understanding the language of vitamin C. *Current Nutrition and Food Science*, 5(1): 53-55.
- Sass, J. E. (1958). *Botanical Microtechnique*. Ames: Iowa State University Press.
- Scalbert, A., Manach, C., Morand, C., Rémésy, C. and Jiménez, L. (2005). Dietary polyphenols and the prevention of diseases. *Critical Reviews in Food Science and Nutrition*, 45: 287-306.
- Schechter, I., Proctor, J. T. A. and Elfving, D. C. (1993). Characterization of seasonal fruit growth of 'Idared' apple. *Scientia Horticulturae*, 54(3): 203-210.
- Schroeder, C. A. (1961). Some morphological aspects of fruit tissues grown *in vitro*. *Botanical Gazette*, 122(3): 198-204.
- Seeram, N. P. (2008). Berry fruits for cancer prevention: Current status and future prospects. *Journal of Agricultural and Food Chemistry*, 56: 630–635.
- Serrano, M., Díaz-Mula, H. M., Zapata, P. J., Castillo, S., Guillén, F., Martínez-Romero, D., Valverde, J. M. and Valero, D. (2009). Maturity stage at harvest determines the fruit quality and antioxidant potential after storage of sweet cherry cultivars. *Journal of Agricultural and Food Chemistry*, 57: 3240–3246.
- Sha, S., Li, J., Wu, J. and Zhang, S. (2011). Characteristics of organic acids in the fruit of different pear species. *African Journal of Agricultural Research*, 6(10): 2403-2410.
- Shamsudin, R., Daud, W. R. W., Takriff, M. S. and Hassan, O. (2007). Physicochemical properties of the Josapine variety of pineapple fruit. *International Journal of Food Engineering*, 3(5).
- Shariah, U. (2013). "Terung Asam Sarawak", a geographical indications (GI)registered product of Sarawak. New Sunday Tribune. Retrieved August 12, 2019, from https://data.sarawak.gov.my/home/data/resource_download/a28db33a-862f-465d-927a-367d9ad84445/

- Shariah, U., Rajmah, M. R., Wong, M. H. and Nur, N. H. (2013). *Terung Asam Sarawak Technology Package*. Sarawak, Malaysia: Department of Agriculture Sarawak.
- Shin, Y., Ryu, J. A., Liu, R. H., Nock, J. F. and Watkins, C. B. (2008). Harvest maturity, storage temperature and relative humidity affect fruit quality, antioxidant contents and activity, and inhibition of cell proliferation of strawberry fruit. *Postharvest Biology and Technology*, 49(2): 201-209.
- Shiow, C. H., Miao, C. L., Yi, Y. M., Ya, D. C., Chin, W. C., Jr, Y. Z., Ching, W. Y. and Chiu, L. C. (2013). The study of *Rubus formosensis* and *Solanum lasiocarpum* on skin-whitening activity. *Chia-Nan Annual Bulletin*, 39: 97-106.
- Shiratake, K. and Martinola, E. (2007). Transporters in fruit vacuoles. *Plant Biotechnology*, 24: 127-133.
- Sinclair, W. B. and Ramsey, R. C. (1944). Changes in the organic-acid content of Valencia oranges during development. *Botanical Gazette*, 106(2): 140-148.
- Sindhu, E. R., Preethi, K. C. and Kuttan, R. (2010). Antioxidant activity of carotenoid lutein *in vitro* and *in vivo*. Indian Journal of Experimental Biology, 48(8): 843-848.
- Singh, B. P. and Sharma, N. K. (1990). Physico-chemical changes with maturity in some promising varieties of brinjal (*Solanum melongena* L.). *Haryana Journal of Horticultural Sciences*, 19(3-4): 318-325.
- Siriamornpun, S. and Kaewseejan, N. (2017). Quality, bioactive compounds and antioxidant capacity of selected climacteric fruits with relation to their maturity. *Scientia Horticulturae*, 221: 33-42.
- Sirisomboon, P., Tanaka, M., Fujita, S. and Kojima, T. (2000). Relationship between the texture and pectin constituents of Japanese pear. *Journal of Texture Studies*, 31(6): 679-690.
- Smirnoff, N. (1995). Antioxidant systems and plant response to the environment. In Smirnoff, N. (Eds). Environment and Plant Metabolism: Flexibility and Acclimation (pp. 217-243). Oxford: Bios Scientific.
- Soares, F. D., Pereira, T., Marques, M. O. M. and Monteiro, A. R. (2007). Volatile and non-volatile chemical composition of the white guava fruit (*Psidium guajava*) at different stages of maturity. *Food Chemistry*, 100(1): 15-21.

- Sureshkumar, V. B., George, D., Prakash, N. S., Prakasan, C. B. and Mohanan, K. V. (2013). A study on the pattern of fruit development in robusta coffee (*Coffea canephora* var. robusta). *International Journal of Plant, Animal and Environmental Sciences*, 3(2): 248-257.
- Szeto, Y. T., Tomlinson, B. and Benzie, I. F. F. (2002). Total antioxidant and ascorbic acid content of fresh fruits and vegetables: implications for dietary planning and food preservation. *The British Journal of Nutrition*, 87(1): 55-59.
- Tadesse, T., Hewett, E. W., Nichols, M. A., and Fisher, K. J. (2002). Changes in physicochemical attributes of sweet pepper cv. Domino during fruit growth and development. *Scientia Horticulturae*, 93(2): 91–103.
- Tee, Y. K. and Ding, P. (2010). Changes in tannin concentration of Rastali banana (Musa AAB Rastali) during growth and development. Conference: 21st Malaysian Society of Plant Physiology, Malaysia, 18-20.
- Tee, Y. K., Ding, P. and Abdul Rahman, N. A. (2012). Determination of optimum harvest maturity and physico-chemical quality of Rastali banana (*Musa* AAB Rastali) during fruit ripening. *Journal of the Science of Food and Agriculture*, 92(1): 171-176.
- Tee, Y. K., Ding, P. and Rahman, N. A. A. (2011). Physical and cellular structure changes of Rastali banana (*Musa* AAB) during growth and development. *Scientia Horticulturae*, 129(3): 382-389.
- Terao, A., Hyodo, H., Satoh, S. and Iwai, H. (2013). Changes in the distribution of cell wall polysaccharides in early fruit pericarp and ovule, from fruit set to early fruit development, in tomato (*Solanum lycopersicum*). *Journal of Plant Research*, 126(5): 719-728.
- Terefe, N. S. and Versteeg, C. (2011). Texture and microstructure of fruits and vegetables. *Practical Food and Research*, 1: 89-115.
- Thaipong, K., Boonprakob, U., Crosby, K., Cisneros-Zevallos, L. and Byrne, D. H. (2006). Comparison of ABTS, DPPH, FRAP, and ORAC assays for estimating antioxidant activity from guava fruit extracts. *Journal of Food Composition and Analysis*, 19(6-7): 669-675.
- Thompson, A. K. (2003). Fruit ripening conditions. In Thompson, A. K. (Eds.), *Fruit and Vegetables: Harvesting, Handling and Storage* (pp. 86-97). United Kingdom: Blackwell Publishing Limited.

- Ting, H., Tan, S. R., John, A. N. (2017). Consumption intention toward ethnic food: determinants of Dayak food choice by Malaysians. *Journal of Ethnic Foods*, 1: 21-27.
- Toivonen, P. M. and Brummell, D. A. (2008). Biochemical bases of appearance and texture changes in fresh-cut fruit and vegetables. *Postharvest Biology and Technology*, 48(1): 1-14.
- Trong, L. V., Tuong, L. Q., Thinh, B. B., Khoi, N. T. and Trong, V. T. (2019). Physiological and biochemical changes in tomato fruit (*Solanum lycopersicum* L.) during growth and ripening cultivated in Vietnam. *Bioscience Research*, 16(2): 1736-1744.
- Tsao, R. and Akhtar, M. H. (2005). Nutraceuticals and functional foods: Current trend in phytochemical antioxidant research. *Journal of Food Agriculture and Environment*, 3: 10-17.
- Tucker, G. and Robards, K. (2008). Bioactivity and structure of biophenols as mediators of chronic diseases. *Critical Reviews in Food Science and Nutrition*, 48: 929-966.
- Usenik V., Fabc^{*}ic^{*} J. and Štampar F. (2008). Sugars, organic acids, phenolic composition and antioxidant activity of sweet cherry (*Prunus avium* L.). *Food Chemistry*, 107: 185-192.
- Valero, D. and Serrano, M. (2010). *Postharvest Biology and Technology for Preserving Fruit Quality.* Boca-Raton: CRC –Taylor and Francis.
- Volz, R. K., Harker, F. R. and Lang, S. (2003). Firmness decline in Gala apple during fruit development. *Journal of the American Society for Horticultural Science*, 128(6): 797-802.
- Voon, B. H. and Kueh, H. S. (1999). The nutritional value of indigenous fruits and vegetables in Sarawak. Asia Pacific Journal of Clinical Nutrition, 8(1): 24-31.
- Wang, Y., Zhu, J., Meng, X., Liu, S., Mu, J. and Ning, C. (2016). Comparison of polyphenol, anthocyanin and antioxidant capacity in four varieties of *Lonicera caerulea* berry extracts. *Food Chemistry*, 197: 522-529.
- Webb, M. A. (1999). Cell-mediated crystallization of calcium oxalate in plants. *The Plant Cell*, 11: 751-761.
- Webb, M. A. and Arnott, H. J. (1982). A survey of calcium oxalate crystals and other mineral inclusions in seeds. *Scanning Electron Microscopy*, 3: 1109-1131.

- Weigend, M., Mustafa, A. and Ensikat, H. J. (2018). Calcium phosphate in plant trichomes: the overlooked biomineral. *Planta*, 247(1): 277-285.
- Whiley, A. W., Schaffer, B. and Lara, S. P. (1992). Carbon dioxide exchange of developing avocado (*Persea americana* Mill.) fruit. *Tree Physiology*, 11(1): 85-94.
- Willcox, J. K., Ash, S. L. and Catignani, G. L. (2004). Antioxidants and prevention of chronic disease. *Critical Reviews in Food Science and Nutrition*, 44: 275-295.
- Wills, R. B. H., McGlasson, W. B., Graham, D. and Joyce, D. C. (2007). Physiology and Biochemistry. In Wills, R. B. H., McGlasson, W. B., Graham, D. and Joyce, D. C. (Eds.), *Postharvest: An Introduction to Physiology and Handling Fruits, Vegetables and Ornamentals* (pp. 29-51). Australia: University New South Wales Press Limited.
- Wongmetha, O., Ke, L. S. and Liang, Y. S. (2015). The changes in physical, bio-chemical, physiological characteristics and enzyme activities of mango cv. Jinhwang during fruit growth and development. *Wageningen Journal of Life Sciences*, 72: 7-12.
- Yahia, E. M. and Carrillo-Lopez, A. (2018). (Eds.) *Postharvest physiology and biochemistry of fruits and vegetables*. UK: Woodhead Publishing.
- Yahia, E. M., Contreras-Padilla, M. and Gonzalez-Aguilar, G. (2001). Ascorbic acid content in relation to ascorbic acid oxidase activity and polyamine content in tomato and bell pepper fruits during development, maturation and senescence. *Food Science and Technology*, 34(7): 452-457.
- Yeats, T. H. and Rose, J. K. (2013). The formation and function of plant cuticles. *Plant Physiology*, 163(1): 5-20.
- Yeum, K. J., Russle, R. M., Krinsky, N. I. and Aldini, G. (2004). Biomarkers of antioxidant capacity in the hydrophilic and lipophilic compartments of human plasma. *Archives of Biochemistry and Biophysics*, 430: 97–103.
- Youssef, M. and Ibrahim, R. (2016). Molecular markers associated with high vitamin-C content in guava. *Journal of Agricultural Chemistry and Biotechnology*, 7(3): 49-55.
- Zaidan, M. W. A. M., Subri, N. F. D. A., Othman, S. M., Rahman, M. A. B. A., Ahmad, M. A., Pin, C. H., Rahman, Z. A. and Simoh, S. (2019). Evaluation of total polyphenol content and antioxidant properties in yellow and purple fruit colour phenotypes of terung asam (*Solanum ferox* Linn). *Journal of Tropical Plant Physiology*, 11(2): 31-38.

- Zapata, P. J., Guillén, F., Martínez-Romero, D., Castillo, S., Valero, D. and Serrano, M. (2008). Use of alginate or zein as edible coatings to delay postharvest ripening process and to maintain tomato (*Solanum lycopersicon* Mill) quality. *Journal of the Science of Food and Agriculture*, 88(7): 1287–1293.
- Zhong, R., Taylor, J. J. and Ye, Z. H. (1999). Transformation of the collateral vascular bundles into amphivasal vascular bundles in an *Arabidopsis* mutant. *Plant Physiology*, 120(1): 53-64.
- Zhou, H. C., Lin, Y. M., Wei, S. D. and Tam, N. F. Y. (2011). Structural diversity and antioxidant activity of condensed tannins fractionated from mangosteen pericarp. *Food Chemistry*, 129(4): 1710-1720.
- Zulkhairi, M. A., Umikalsum, M. B., Razali, M., Siti Aisyah, M. N., Nur Daliana, Y., Rosali, H., Noralienatul Azlia, A. A., Nur Syafini, G. and Aimi Athirah, A. (2019). Discovery of antioxidant activities from traditional eggplant (*Terung telunjuk*) at different maturity stages. In Southeast Asia Vegetable Symposium (SEAVEG) (pp.131).
- Zuzunaga, M., Serrano, M., Martínez-Romero, D., Valero, D. and Riquelme, F. (2001). Comparative study of two plum (*Prunus salicina* Lindl.) cultivars during growth and ripening. *Food Science and Technology International*, 7: 123–130.

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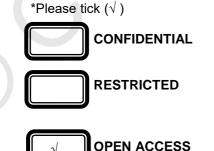
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