

UNIVERSITI PUTRA MALAYSIA

MICROBIAL DECOLORIZATION OF TRIAZO DYE, DIRECT BLUE 71 BY MIXED BACTERIAL CULTURE ISOLATED FROM MALAYSIAN SOIL

KHAIRUNNISA' BINTI MOHD ZIN

FP 2021 56



MICROBIAL DECOLORIZATION OF TRIAZO DYE, DIRECT BLUE 71 BY MIXED BACTERIAL CULTURE ISOLATED FROM MALAYSIAN SOIL

By

KHAIRUNNISA' BINTI MOHD ZIN

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

August 2021

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

MICROBIAL DECOLORIZATION OF TRIAZO DYE, DIRECT BLUE 71 BY MIXED BACTERIAL CULTURE ISOLATED FROM MALAYSIAN SOIL

By

KHAIRUNNISA' BINTI MOHD ZIN

September 2020

Chair : Mohd Izuan Effendi bin Halmi, PhD Faculty : Agriculture

Polluted wastewater from textile dyeing industrial sectors has causes severe effect towards human, plant and marine creatures and the treatment process is truly challenging. In this study, a mixed bacterial culture was isolated from Malaysian soil, screened and identified to successfully decolorize Triazo-bond Direct Blue 71 dye and act as the sole source of carbon and nitrogen. The free cells were also immobilized for better decolorization at higher concentration of dye by using sodium alginate. Response surface methodology (RSM) and Artificial Neural Network (ANN) were used to optimize the decolorization efficiency for both free and immobilized cells. Significant effect on DB71 dye decolorization percentage by free cells is denoted by the experimental variables of dye concentration, yeast extract, and pH. The optimum conditions for dye decolorization by immobilized mixed culture were determined by four variables which were dye concentration, alginate concentration, number of beads and beads size. GCMS and FTIR analysis were used to characterize the metabolites after the decolorization. Other than that, kinetics modelling study of DB71 dye decolorization allowed the estimation of decolorization rate of free and immobilized cells. Major bacterial group found from the metagenomics analysis consist of Acinetobacter (30%), Comamonas (11%), Aeromonadaceae (10%), Pseudomonas (10%), Flavobacterium (8%), Porphyromonadaceae (6%). and Enterobacteriaceae (4%). Proteobacteria (78.61%), then Bacteroidetes (14.48%) and Firmicutes (3.08%) were among the richest phylum in the mixed bacterial culture. The optimum condition for free cells predicted by RSM is at 150 mg/L of dye concentration, 3 g/L of yeast extract and pH of 6.645. ANN predicted the optimum condition at 150 mg/L, 2.9 g/L of yeast extract and pH of 6.7. Higher prediction and accuracy in the fitness was found in ANN model as proved by R^2 and AAD values of 0.99 and 0.04 subsequently fitness compared to the RSM. ANN model for immobilized cells offered a better prediction than RSM with R² of 0.99. The ANN model predict the decolorization by immobilized cell is optimum at 200 mg/L, 0.966 % of alginate concentration, 50 number of beads and 0.599 cm of beads size. Moreover, the result from GCMS and FTIR analysis of the metabolites from the decolorization of dye shows that the reduction of dye caused the absence of the untreated sample and emergence of new peaks in the treated sample in FTIR spectrum. In addition, GCMS result from the treated sample shows no toxic secondary metabolites were formed.

Luong model predicted the rate of decolorization by free cell at 10 %hr⁻¹ by using the kinetic modelling and dye concentration at 159.5 mg/L completely inhibited the decolorization based on the S_m value. Aiba model predicted the rate of decolorization by immobilized cell is at 4.645 %hr⁻¹. The use of mixed bacterial culture was found to be efficient for the decolorization of DB71 dye in this study. The optimization of immobilized cell by using RSM and ANN using sodium alginate resulted to better decolorization of dye at higher concentration which is up to 200 mg/L. Moreover, the effect of metal ions towards the decolorization shows that gel beads through immobilization were able to protect against toxic substance. It is reflected by a great tolerance result towards metal ions over free cells during DB71 dye decolorization where occurrence of metal ions may disrupt the decolorization process. The decolorization of Direct Blue 71 dye by immobilized cell was still higher than 90 % even with the presence of 1 mg/L of mercury, nickel, copper, lead, arsenic, chromium, cadmium and silver in the solution and slight decrease of decolorization was observed for both free and immobilized cell compared to the control samples. In conclusion, all the objectives of this study were achieved accordingly.

Environmental pollution caused by the released of industrial effluent containing dye has been affecting the water quality in Malaysia. Biological practice using microorganisms provides a complete degradation with no secondary pollutant besides the cost effective advantage compared to the physical and chemical wastewater treatment. There are few reported works on azo dye decolorization by mixed bacterial culture without the aid of carbon and nitrogen source. Therefore, this study found and optimized a potent mixed bacterial culture that could degrade Direct Blue 71 dye in facultative anaerobic condition and no added carbon and nitrogen sources are needed to completely decolorize the dye with no introduction of secondary toxic metabolites based on the metabolites analysis result.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia Sebagai memenuhi keperluan untuk ijazah Master Sains

DEKOLORASI MIKROB UNTUK PEWARNA "TRIAZO", "DIRECT BLUE 71" OLEH KULTUR BAKTERIA CAMPURAN YANG DIASINGKAN DARIPADA TANAH MALAYSIA

Oleh

KHAIRUNNISA' BINTI MOHD ZIN

September 2020

Pengerusi : Mohd Izuan Effendi bin Halmi, PhD Fakulti : Pertanian

Air buangan yang tercemar dari sektor industri pencelupan tekstil menyebabkan kesan yang teruk terhadap makhluk hidup manusia, tumbuhan dan laut serta proses rawatannya sangat mencabar. Dalam kajian ini, kultur bakteria campuran diasingkan dari tanah Malaysia, disaring dan dikenal pasti untuk berjaya menyahwarna pewarna Triazo Direct Blue 71 (DB71) dan bertindak sebagai satu-satunya sumber karbon dan nitrogen. Sel bebas juga telah menjadi tidak bergerak untuk penyahwarnaan yang lebih baik pada kepekatan pewarna yang kebih tinggi dengan menggunakan natrium alginat. Response Surface Methodology (RSM) dan Artificial Neural Network (ANN) digunakan untuk mengoptimumkan kecekapan dekolorasi. Kesan yang signifikan terhdadap peratusan dekolorasi pewarna DB71 adalah daripada pemboleh ubah eksperimen kepekatan pewarna, ekstrak ragi, dan pH. Keadaan optimum untuk penyahwarnaan pewarna oleh kultur campuran tidak bergerak ditentukan oleh empat pemboleh ubah jaitu kepekatan pewarna, kepekatan alginat, bilangan manik dan ukuran manik. Analisis GCMS dan FTIR digunakan untuk mencirikan metabolit setelah penyahwarnaan. Selain itu, memodelkan kajian kinetik dekolorisasi pewarna DB71 memungkinkan pengiraan kadar dekolorasi sel bebas dan tidak bergerak. Kumpulan bakteria utama yang didapati dari analisis metagenomik terdiri daripada Acinetobacter (30%), Comamonas (11%), Aeromonadaceae (10%), Pseudomonas (10%), Flavobacterium (8%), Porphyromonadaceae (6%), dan Enterobacteriaceae (4%). Proteobacteria (78.61%), kemudian Bacteroidetes (14.48%) dan Firmicutes (3.08%) adalah antara filum yang terkaya di dalam bakteria campuran itu. Keadaan optimum untuk sel bebas yang diramalkan oleh RSM ialah kepekatan pewarna 150 mg/L, 3 g/L ekstrak ragi dan pH 6.645. ANN meramalkan keadaan optimum pada 150 mg/L, 2.9 g/L ekstrak ragi dan pH 6.7. Model ANN mempunyai ramalan dan ketepatan yang lebih tinggi dalam kecekapan berbanding dengan model RSM seperti yang dibuktikan oleh nilai R² dan AAD yang masing-masingnya adalah 0.99 dan 0.04 oleh model ANN. Model ANN untuk sel tidak bergerak memberikan ramalan yang lebih baik daripada RSM dengan R² 0.99. ANN meramalkan dekolorasi oleh sel tidak bergerak adalah optimum pada 200 mg/L, kepekatan alginate 0.966%, 50 bilangan manik gel dan ukuran manik gel 0.599 cm. Lebih-lebih lagi, hasil analisis metabolit dari dekolorisasi pewarna daripada GCMS dan FTIR menunjukkan bahawa dekolorasi pewarna menyebabkan kemunculan puncak baru pada sampel yang telah dirawat dan ketiadaan puncak itu pada sampel yang tidak dirawat di dalam spektrum FTIR. Sebagai tambahan, hasil GCMS dari sampel yang dirawat menunjukkan tidak ada metabolit sekunder beracun yang terbentuk.

Model Luong meramalkan kadar dekolorasi oleh sel bebas pada 10% jam⁻¹ dan kepekatan pewarna pada 159.5 mg/L adalah penghalang dekolorasi berdasarkan nilai Sm. Model Aiba meramalkan kadar dekolorasi oleh sel yang tidak bergerak adalah pada 4.645% jam⁻¹. Hasil ini menunjukkan dekolorasi pewarna DB71 oleh kultur bakteria campuran yang efisien. Pengoptimuman sel yang tidak bergerak dengan menggunakan RSM dan ANN menggunakan natrium alginat menghasilkan penyahwarnaan pewarna yang lebih baik pada kepekatan yang lebih tinggi sehingga 200 mg/L. Lebih-lebih lagi, kesan ion logam terhadap dekolorasi menunjukkan bahawa manik gel melalui imobilisasi dapat melindungi daripada bahan toksik. Ini dicerminkan oleh hasil toleransi yang tinggi terhadap ion logam berbanding sel bebas semasa penyahwarnaan pewarna DB71 kerana ion logam boleh mengganggu proses dekolorisasi. Dekolorasi pewarna DB71 oleh sel yang tidak bergerak adalah masih lebih tinggi daripada 90% walaupun dengan adanya 1 mg/L merkuri, nikel, tembaga, plumbum, arsenic, kromium, cadmium dan perak dalam larutan dan sedikit penurunan dekolorisasi diperhatikan untuk sel bebas dan tidak bergerak berbanding dengan sampel kawalan. Kesimpulannya, semua objektif kajian ini dicapai dengan sewajarnya.

Pencemaran alam sekitar yang disebabkan oleh pelepasan bahan buangan industri yang mengandungi pewarna telah mempengaruhi kualiti air di Malaysia. Amalan biologi menggunakan mikroorganisma memberikan degradasi lengkap tanpa pencemaran sekunder selain kelebihan kos efektif berbanding dengan rawatan air sisa fizikal dan kimia. Terdapat hanya sedikit kajian yang dilaporkan mengenai dekolorasi pewarna azo oleh kultur bakteria campuran tanpa bantuan sumber karbon dan nitrogen. Oleh itu, kajian ini mendapatkan dan mengoptimumkan kultur bakteria campuran yang kuat yang dapat menurunkan pewarna Direct Blue 71 dalam keadaan anaerob fakultatif dan tidak memerlukan sumber karbon dan nitrogen untuk dekolorasi sepenuhnya pewarna tanpa pengenalan metabolit toksik sekunder berdasarkan hasil analisis metabolit.

ACKNOWLEDGEMENTS

First and foremost, I would like to thank Allah SWT for His bless in every single day from the beginning until this Master was completed. Upon completion of this study, I would like to express my gratitude to many parties. My heartfelt thanks go to my supervisor, Dr Mohd Izuan Effendi bin Halmi who gave a tremendous guidance for this project until successfully ended. Her mentorship and kindness are greatly appreciated. I would also like to deliver my thanks to my co supervisors Dr Mohd Yunus Abd Shukor and Dr Uswatun Hasanah Zaidan who have been giving encouragement and sharing knowledge about this Master's study, and my friend, Fatin, Adibah, Syu and Asraf for lending their helping hands in completing my study and research. In addition, special thanks to my beloved family members especially my parents for their strongest supports and advices for me to complete my study. Last but not least, I wish to thank the lecturers, lab assistant and staffs at Faculty of Agriculture and Universiti Putra Malaysia as a whole for the opportunity to complete my journey in Master's study.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Mohd Izuan Effendi bin Halmi, PhD

Senior Lecturer Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Mohd. Yunus Abd. Shukor, PhD

Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Member)

Uswatun Hasanah binti Zaidan, PhD

Associate Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date: 12 August 2021

Declaration by graduate student

I hereby confirm that:

- This thesis is my original work;
- Quotations, illustrations and citations have been duly referenced;
- This thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- Intellectual property from the thesis and copyright of thesis are fully owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- Written permission must be obtained from supervisor and the office if Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- There is no plagiarism or data falsification/Fabrication in the thesis, and no scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Date:

Signature:			
------------	--	--	--

Name and Matric No.: Khairunnisa' binti Mohd Zin GS51703

TABLE OF CONTENTS

ABSTRACTiABSTRAKiiiACKNOWLEDGEMENTSvAPPROVALviDECLARATIONviiiLIST OF TABLESxivLIST OF FIGURESxviLIST OF ABBREVIATIONSxviii		Page
ABSTRAKiiiACKNOWLEDGEMENTSvAPPROVALviDECLARATIONviiiLIST OF TABLESxivLIST OF FIGURESxviLIST OF ABBREVIATIONSxviii	ABSTRACT	i
ACKNOWLEDGEMENTSvAPPROVALviDECLARATIONviiiLIST OF TABLESxivLIST OF FIGURESxviLIST OF ABBREVIATIONSxviii	ABSTRAK	iii
APPROVALviDECLARATIONviiiLIST OF TABLESxivLIST OF FIGURESxviLIST OF ABBREVIATIONSxviii	ACKNOWLEDGEMENTS	V
DECLARATIONviiiLIST OF TABLESxivLIST OF FIGURESxviLIST OF ABBREVIATIONSxviii	APPROVAL	vi
LIST OF TABLESxivLIST OF FIGURESxviLIST OF ABBREVIATIONSxviii	DECLARATION	viii
LIST OF FIGURES xvi LIST OF ABBREVIATIONS xviii	LIST OF TABLES	xiv
LIST OF ABBREVIATIONS xviii	LIST OF FIGURES	xvi
	LIST OF ABBREVIATIONS	xviii

CHAPTER

4

1	INTRO	DUCTION	1
2	LITER 2.1 2.2	ATURE REVIEW Dye Pollution Direct Dye 2.2.1 Direct Blue 71 Dye Properties And Uses	4 4 5 5 7
	2.3	2.2.2 DB71 Dye Toxicity Dye Effluent Treatments 2.3.1 Biological Treatments 2.3.2 Microbial Technology for Treatment of Azo Dye	7 8 8 8
	2.4	Bacterial Decolorization And Degradation of Azo Dyes 2.4.1 Mixed Bacterial Culture 2.4.2 Mechanisms of Azo Dye Reduction 2.4.2.1 Under anaerobic conditions	11 11 12 14
	2.5 2.6 2.7	Response Surface Methodology (RSM) Artificial Neural Network (ANN) Cell Immobilization 2.7.1 Advantages of Cell Immobilization 2.7.2 Entrapment Method 2.7.3 Alginate as a Gel Entrapment for Dye Removal	15 16 17 18 18 19 19
	2.8 2.9	Metal Ions Co-Contamination Biodegradation Kinetics	20 21
	METH 3.1 3.2	INTRODUCOGY Introduction 3.1.1 Flowchart 3.1.2 Chemicals and Equipment Used Preparation of Culture Media 3.2.1 Minimal Salt Medium (MSM) 3.2.1.1 Minimal Salt Medium containing Glucose 3.2.1.2 Minimal Salt Medium without	25 25 25 25 25 25 25 25

			Glucose and Ammonium	
		3.2.1.3	Minimal Salt Medium	25
		0.2.110	containing Direct Blue 71 dye	20
	3.2.2	Prepara	tion of Nutrient Broth	26
	3.2.3	Prepara	tion of Toxicants	26
3.3	Identifi	cation of	Direct Blue 71 dye-degrading	26
	mix cu	Iture		~~
	3.3.1	Soil san	npling	26
	3.3.Z	Soll Tex		20
	3.3.3	dearadi	no mix culture	21
	334	Screeni	ng of Direct Blue 71 Dve-	27
	0.0.1	degradi	na mix culture	
	3.3.5	Metager	nomics Analysis	28
	3.3.6	Mainten	ance and Growth of Bacterial	29
		Isolates		
3.4	Free C	Cells		29
	3.4.1	Optimiza	ation of decolorization using	29
		Respon	se Surface Methodology (RSM)	20
		3.4.1.1	Optimization of the significant	30
			Design	
	3.4.2	Optimiza	ation of direct blue 71	31
		decolori	zation using Artificial Neural	
		Network	(ANN)	
		3.4.2.1	Residual Analysis (Error	31
			Analysis)	~~
		3.4.2.2	point	32
	3.4.3	Metabol	ites Analysis	32
		3.4.3.1	Extraction and Purification of	32
			the Metabolites from the	
		3132	GCMS Applysis	30
		3433	FTIR Analysis	33
	3.4.4	Effect of	Metal lons on DB71 dve-	33
		decolori	zation activities by Free Cells	
	3.4.5	Kinetics	Modelling of DB71 dye	33
		decolori	zation by free cells	
		3.4.5.1	The effect of different initial	33
			concentration on	
			free colle	
		3452	Kinetics Modelling of DB71	34
		0.1.0.2	dve decolorization by free	01
			cells	
3.5	Immob	ilized Ce	lls	36
	3.5.1	Prepara	tion of Immobilized dye-	36
		degradi	ng mixed culture	~~
		3.5.1.1	Unemicais	36

		3.5.1.2	Immobilization of DB71 dye	36
	3.5.2	Optimiza immobili RSM	ation of dye-degrading zed mixed culture by using	37
		3.5.2.1	Dye decolorization and	38
	3.5.3	Optimiza	ation of dye-degrading	38
	3.5.4	Morphol immobili	ogical Observation of zed dye-degrading mixed	38
	3.5.5	Effect of	metal ions on DB71 dye-	39
	3.5.6	cells Kinetics	modelling of DB71 dye	39
		decoloriz 3.5.6.1	zation by immobilized cells The effect of different initial concentration on	39
			decolorization DB71 dye by immobilized cells	
		3.5.6.2	dye decolorization using immobilized cells	40
RESU 4.1	ILTS AN Isolatio	on and sci	SSION reening of DB71 Dye-degrading	41 41
4.2	Secon	dary Scre	ening of DB71 dye-degrading	43
4.3 4.4	Identif Optim	ication of lization of l	DB71 dye-degrading mix culture Direct Blue 71 dye	45 47
	decolo 4.4.1	Optimization by	y using free cells ation using Response Surface	47
	4.4.2	Optimiza	ation using Artificial Neural	56
	4.4.3	Residua RSM and decoloriz	l Analysis (Error Analysis) of d ANN model on the zation of DB71 dye by using	60
15	Chara	free cells	S of Metabolites by GCMS and	61
4.5	FTIR a	analysis	TOT METADOILLES BY GOMS and	01
	4.5.1 4.5.2	FTIR		61 63
4.6	Effect	of differer	t initial concentration of DB71	64
4.7	Kinetic	cs Modelli	ng of the decolorization of DB71	66
4.8	Optim using	ization of l	, DB71 dye decolorization by ed mixed culture	71

		4.8.1	Optimization of Direct Blue 71 Dye decolorization using RSM	71
		4.8.2	Optimization using Artificial Neural Network	77
		4.8.3	Residual Analysis (Error Analysis) of RSM and ANN model on the decolorization of DB71 dye by using immobilized cells	87
	4.9	Morpho	logical of immobilized cell	87
	4.10	Effect of DB71 d	of different initial dye concentration on lye decolorization by immobilized cells	90
	4.11	Kinetics dye by	Modelling of the decolorization of DB71	92
	4.12	Effect of dye by	of Metal Ions in Decolorization of DB71 Free and Immobilized Cells	97
5	SUMM RECOM	ARY, CO MMEND	ONCLUSION AND ATIONS FOR FUTURE RESEARCH	100
REFER APPEN BIODA PUBLI	RENCES IDICES TA OF S CATION	STUDE	л	102 123 128 129

LIST OF TABLES

Table		Page
2.1	Properties of Direct Blue 71 dye	6
2.2	List of various DB71 dye-degrading microorganisms	10
2.3	Mixed bacterial cultures for azo dye decolorization	13
2.4	Alginate for immobilization of cells and enzyme for dye removal	20
2.5	Various mathematical models developed for growth kinetics involving substrate inhibition	22
2.6	Previous works using different biodegradation kinetics model for various purposes	23
3.1	Lower limit and upper limit of Box Behnken Design	30
3.2	Formula of kinetics modelling	35
3.3	Lower limit and upper limit of Box Behnken Design	37
4.1	The primary screening for samples in static condition	42
4.2	The primary screening for samples in shake condition	43
4.3	Secondary screening on chosen samples from primary screening by varying DB71 dye concentration	44
4.4	Box-Behnken matrix for experimental design and predicted response using RSM and ANN	49
4.5	Analysis of variance (ANOVA) for the fitted quadratic polynomial model for optimization of Direct Blue 71 dye decolorization	50
4.6	Predicted and experimental value for the responses at optimum condition using response surface methodology (RSM)	53
4.7	The effect of different neural network architecture and topologies on R2 and AAD in the estimation of DB71 dye decolorization obtained in the training and testing of neural networks	56
4.8	Predicted and experimental value for the responses at optimum condition using artificial neural network (ANN)	58

- 4.9 Absolute deviation, R2, adjusted R2 and AAD of RSM 61 and ANN models
- 4.10 Summary of kinetics model for degradation of Direct Blue 70 71 dye
- 4.11 Box-Behnken matrix for experimental design and 73 predicted response using RSM

- 4.12 Analysis of variance (ANOVA) for the fitted quadratic polynomial model for optimization of Direct Blue 71 dye decolorization
- 4.13 The effect of different neural network architecture and 77 topologies on coefficient of determination, R2, in the estimation of Direct Blue 71 dye decolorization obtained in the training and testing of neural networks
- 4.14 Predicted and experimental values for the responses at 79 optimum condition using response surface methodology (RSM) and an artificial neural network (ANN) for immobilized cell
- 4.15 Absolute deviation, R2, adjusted R2, RMSE and AAD of 87 RSM and ANN models
- 4.16 Summary of kinetic models for degradation of Direct Blue 97 71 dye by immobilized cells

LIST OF FIGURES

Figure		Page	
2.1	Molecular structure of triazo class DB71 dye	7	
4.1	DB71 dye decolorization by N5 mixed bacterial culture without the presence of carbon and nitrogen source	45	
4.2	Identification of the mixed bacterial culture by Metagenomic Analysis	46	
4.3	Diagnostic plots showing, a) the studentized residuals plotted against the normal probability, b) the predicted versus studentized residuals, c) the run versus studentized residuals, and d) the actual responses plotted against the predicted response values	51	
4.4	Three-dimensional plots showing the effect of (a) pH, yeast extract (b) pH, dye concentration and (c) yeast extract, dye concentration and their mutual effect on the decolorization of Direct Blue 71 dye. Other variables are constant: pH (6.645), yeast extract (3g/L), and dye concentration (150mg/L)	54	
4.5	Neural network topology. The topology of multilayer normal feedforward neural network for the estimation of DB71 dye decolorization	57	
4.6	Nodes no of optimization	57	
4.7	ANN predicted versus actual experimental data values for DB71 dye decolorization	58	
4.8	ANN response surface for a) dye concentration versus yeast extract b) dye concentration versus pH c) yeast extract versus pH with dye decolorization as a response	59	
4.9	FTIR spectrum of control DB71 dye	62	
4.10	FTIR spectrum of DB71 dye after degradation by mixed culture	62	
4.11	Result of GCMS analysis after the degradation of DB71 dye	63	
4.12	Effect of different concentration of DB71 dye on decolorization of DB71 dye by free cells. The error bars represent the mean \pm standard deviation of three	65	

 \mathbf{G}

replicates

- 4.13 Graphs of kinetic modelling for free cells a)Luong model
 b)Monod model c)Webb model d)Teissier model e)Aiba
 model f)Haldane model and g)Yano model
- 4.14 Diagnostic plots showing, a) the studentized residuals plotted against the normal probability, b) the studentized residuals versus predicted, c) the studentized residuals versus run, and d) the predicted response values plotted against the actual responses
- 4.15 Neural network topology. The topology of multilayer normal feedforward neural network for the estimation of DB71 dye decolorization
- 4.16 Three-dimensional plots showing the effect of (a) dye structure conc., alginate conc., b) dye conc., beads size, c) alginate conc., number of beads, d) alginate conc., beads size, e) dye conc., number of beads, and f) number of beads, beads size and and their mutual effect on the decolorization of Direct Blue 71 dye. Other variables are constant: dye conc. (200 mg/L), alginate conc. (1.011%), number of beads (45) and beads size (0.546 cm)
- 4.17 ANN response surface for a) dye conc. versus alginate conc., b) dye conc. versus number of beads, c) dye conc. versus beads size, d) alginate conc. versus number of beads, e) alginate conc. versus beads size, and f) number of beads versus beads size with dye decolorization as a response
- 4.18 Immobilized mixed culture a) before, b) after DB71 dye 88 decolorization and c) surface of immobilized cells
- 4.19 Effect of different initial dye concentration to the 91 degradation of DB71 by immobilized cells
- 4.20 Graphs of kinetic modelling for immobilized cells a)Luong
 b)Haldane c)Aiba d)Teissier e)Webb f)Monod and g) Yano
 model
- 4.21 Effect of metals ions on DB71 dye decolorization by free 99 and immobilized cells. The error bars represent the mean ± standard deviation of three replicates

67

75

LIST OF ABBREVIATIONS

DB71	Direct Blue 71
RSM	Response Surface Methodology
ANN	Artificial Neural Network
AAD	Absolute Average Deviation
PDW	Printing and Dyeing Wastewater
MSM	Minimal Salt Media
DNA	Deoxyribonucleic acid
ANOVA	Analysis of Variance
GPS	Global Positioning System
MNFF	Multilayer Normal Feed Forward
ВВР	Batch Back Propagation
RMSE	Root Mean Square Error
3D	3 Dimensional
NADH	electron carrier
R ²	coefficient of determination
rpm	revolutions per minute
yi, exp	experimental responses
yi, cal	measured responses
p	number of experimental runs
n	number of the experimental data
% w/v	percentage weight per volume
p-value	probability value
Tanh	hyperbolic tangent
GCMS	Gas Chromatography Mass Spectrometry

xviii

 \bigcirc

	FTIR	Fourier Transform Infrared Spectroscopy
	HPLC	High Performance Liquid Chromatography
	K ₂ HPO ₄	Dipotassium phosphate
	KBr	Potassium bromide
	KH ₂ PO ₄	Potassium dihydrogen phosphate
	As	Arsenic
	Pb	Lead
	Zn	Zinc
	Cr	Chromium
	Cd	Cadmium
	Со	Cobalt
	Cu	Copper
	Ni	Nickel
	Ag	Silver
	Hg	Mercury
	DNA	Deoxyribonucleic acid
	CaCl ₂	Calcium chloride
	Mn	Manganese
	MnSO ₄ .H ₂ O	Manganese sulphate
	MSM	Minimal salt media
	Na ₂ SO ₄	Sodium sulphate
	NaCl	Sodium chloride
	NaOH	Sodium hydroxide
	NaM_0O_4	Sodium molybdate
	$(NH_4)_2SO_4$	Ammonium sulphate

mg/L	milligram per liter
SEM	Scanning Electron Microscope
w/v	weight per volume



 \bigcirc

CHAPTER 1

INTRODUCTION

A massive amount of dye effluent was produced by large textile industries during the application of dye for the making of their end product. Approximately two-thirds of the total quantity of dye were constituted to textile effluent. The high concentration of color and a huge variability form of dyes were the issues in handling the textile effluent properly. The coloration process ultimately shed 10% of the dye and was discharged right away into the aqueous effluent for about 2% (K. Singh & Arora, 2011). Moreover, approximately 70% of azo dye that consists of one or more -N=N- double bond was involved in dye formation (Xie et al., 2016).

No exact figure on the production of dyes in the world was found in a year, however, about 1000 000 tons of dyes has been produced as reported from previous study (Gupta, 2009). Batik industry in Malaysia has been the source of economic growth which become the ninth in Asia region for biggest producer of textile fiber in 2008 and fifteenth in worldwide. This industry contribute to 2.3% of total exports of manufactured goods in 2011, therefore, contributed much on the decrease of environmental quality especially in East Coast of Peninsular Malaysia (Tiong, 2015).

Polluted rivers has been increasing through the year of 2006 until 2010 due to the manufacturing industry especially textile industries that contributed about 22% of total industrial wastewater in Malaysia especially in Pulau Pinang, Selangor and Johor states (Pang & Abdullah, 2013). This is due to the huge amount of textile finishing plants located in this states. Malaysia has no centralized treatment system and individual factory needs to treat their own wastewater, however, some of them still failed to meet the discharge quality standards based on the Environmental Quality Act 1974 due to high treatment cost and lack of awareness among small and medium scale factories. Textile waste in Malaysia contributed to four percent of total solid waste in 2013, approximately two million kilogrammes of textile waste per day due to the fast fashion industry. Direct Blue 71 has been extensively used for textile dyeing and 10 to 50 percent of synthetic dye residue end up in the local waterways which remain untreated, became the contributor for water pollution in Malaysia(Tiong, 2015).

The root causes of smelly water, turbidity and awful appearance are the colloidal matter and oily impurities that present along with the dye. Consequently, penetration of sunlight was being obstructed and eventually interrupted the photosynthesis process (Muthu, 2014). The major concern involving the effect of textile waste towards marine life is the deficiency of dissolved oxygen and ultimately, the self-purification process of water was

being halted (Kant, 2012). Besides that, clogged pores of soil was observed as fields were filled with these effluents and causing the dropped in soil efficiency. This is due to the penetration of roots being stagnated as a result of solidified soil. The quality of drinking water that is unfit for human uptake was coming from the corrosion of sewerage tubes due to the wastewater that flows in the drain (Kant, 2012).

Recalcitrant residual leads to environmental pollution that are complex and hazardous due to a lot of water and chemicals used for the procedures in staining for textile industry (Dasgupta et al., 2015). The textile effluent needs specified treatment mainly because dyes, along with chemicals, pigments and high chemical oxygen demand were used in coloration (Lokesh & Sivakiran, 2014). Therefore, the discovery of advanced methods should be done to attenuate the impact of discarding the textile dyes within industrial effluents and causing environmental damage.

Adsorption, flocculation, and photochemical oxidation were some great physical and chemical alternatives for the decolorization of textile wastewater. Previous studies that involved the method of adsorption (Bulut et al., 2007), ultrasound (Tauber et al., 2008), ozonation (Turhan & Turgut, 2009) and Fenton's oxidation(Ertugay & Acar, 2017) have been utilized to deal with Direct Blue 71 dye. However, the primary cons of this solution were the enormous operating expenditure and the introduced of secondary pollutants. These drawbacks of physical and chemical methods can be solved through biological practice that provides easy procedures and economical practices besides that it presently was prevalent in dye treatment method (Schütte et al., 2008). Research on microorganisms were being intensively studied despite the use of physical and chemical methods (Kumaran & Dharani, 2011) such as the utilization of fungi and bacteria for the treatment of dye.

Presently, anaerobic conditions for azo dye degradation were favoured by many of the isolated bacteria (García-Montaño et al., 2008). However, the capability of bacteria was affected by the functional group of the azo dye that created a complex structure. Food chain was badly affected as anoxic-tolerant aromatic amines build up as a result of azo dye decolorization was found to be carcinogenic, toxic and (Dos Santos et al., 2006; Işık & Sponza, 2008). Thereby, discovering the potent azo dye decolorizing bacteria was significant. Complete degradation rather than simply the removal of azo dye colour is the critical element in azo dye removal from the environment (Mohana et al., 2008).

Novel dye decolorizing mixed bacterial culture will be isolated and identified in this study. The significant parameters which will affect the decolorization of DB71 dye will be determined by using Response surface methodology (RSM) and Artificial Neural Network (ANN) along with the optimizing of the decolorization process by free and immobilized mixed culture.

To satisfy the above problem statement, the following objectives are conducted:

- To isolate, screen and identify using Metagenomic Analysis of the best mixed bacterial culture from Malaysian soils which is able to decolorize Direct Blue 71 Dye
- To optimize the decolorization of Direct Blue 71 Dye by the chosen mixed bacterial culture using Response Surface Methodology (RSM) and Artificial Neural Network (ANN).
- 3. To enhance the docolorization of Direct Blue 71 dye through immobilization using sodium alginate and characterize the metabolites from decolorization using Gas Chromatography Mass Spectrometry (GCMS) and Fourier Transform Infrared (FTIR)



REFERENCES

- Abd El–Rahim, W. M., Moawad, H., & Khalafallah, M. (2003). Microflora involved in textile dye waste removal. *Journal of Basic Microbiology: An International Journal on Biochemistry, Physiology, Genetics, Morphology, and Ecology of Microorganisms, 43*(3), 167-174.
- Abdel-El-Haleem, D. (2003). Acinetobacter: environmental and biotechnological applications. *African Journal of Biotechnology*, 2(4), 71-74.
- Aiba, S., Shoda, M., & Nagatani, M. (1968). Kinetics of product inhibition in alcohol fermentation. *Biotechnology and bioengineering*, 10(6), 845-864.
- Aiba, S., Shoda, M., & Nagatani, M. (2000). Kinetics of product inhibition in alcohol fermentation. *Biotechnology and bioengineering*, *67*(6), 671-690.
- Al-Fawwaz, A. T., & Abdullah, M. (2016). decolorization of methylene blue and malachite green by immobilized Desmodesmus sp. isolated from north Jordan. International Journal of Environmental Science and Development, 7(2), 95.
- Alkarkhi, A. F., Ahmad, A., Ismail, N., & Easa, A. M. (2008). Multivariate analysis of heavy metals concentrations in river estuary. *Environmental monitoring and assessment*, 143(1-3), 179-186.
- Amim Jr, J., Petri, D. F., Maia, F. C., & Miranda, P. B. (2010). Ultrathin cellulose ester films: preparation, characterization and protein immobilization. *Química Nova*, 33(10), 2064-2069.
- An, T., Zhou, L., Li, G., Fu, J., & Sheng, G. (2008). Recent patents on immobilized microorganism technology and its engineering application in wastewater treatment. *Recent Patents on Engineering*, 2(1), 28-35.
- Asad, S., Amoozegar, M., Pourbabaee, A. A., Sarbolouki, M., & Dastgheib, S. (2007). Decolorization of textile azo dyes by newly isolated halophilic and halotolerant bacteria. *Bioresource technology*, *98*(11), 2082-2088.
- Asgher, M., Bhatti, H., Shah, S., Asad, M. J., & Legge, R. (2007). Decolorization potential of mixed microbial consortia for reactive and disperse textile dyestuffs. *Biodegradation*, *18*(3), 311-316.
- Ashrafi, S. D., Rezaei, S., Forootanfar, H., Mahvi, A. H., & Faramarzi, M. A. (2013). The enzymatic decolorization and detoxification of synthetic dyes by the laccase from a soil-isolated ascomycete, Paraconiothyrium variabile. *International Biodeterioration & Biodegradation, 85*, 173-181.

- Aslan, N. (2008). Application of response surface methodology and central composite rotatable design for modeling and optimization of a multigravity separator for chromite concentration. *Powder Technology*, *185*(1), 80-86.
- Azizi, N. A. (2018). Senawang Factory Ordered Shut After Pollution Turned Sungai Simin Blue. Retrieved January 26, 2018, from https://www.nst.com.my/news/nation/2018/01/328893/senawangfactory-ordered-shut-after-pollution-turned-sungai-simin-blue
- Banerjee, A., & Ghoshal, A. K. (2011). Phenol degradation performance by isolated Bacillus cereus immobilized in alginate. *International Biodeterioration & Biodegradation*, 65(7), 1052-1060.
- Banerjee, P., Barman, S. R., Sikdar, D., Roy, U., Mukhopadhyay, A., & Das, P. (2017). Enhanced degradation of ternary dye effluent by developed bacterial consortium with RSM optimization, ANN modeling and toxicity evaluation. *Desalin Water Treat*, *7*2, 249-265.
- Bansal, P., & Verma, A. (2017). Synergistic effect of dual process (photocatalysis and photo-Fenton) for the degradation of Cephalexin using TiO2 immobilized novel clay beads with waste fly ash/foundry sand. *Journal of Photochemistry and Photobiology A: Chemistry, 342*, 131-142.
- Bas, D., & Boyaci, I. H. (2007). Modeling and optimization II: comparison of estimation capabilities of response surface methodology with artificial neural networks in a biochemical reaction. *Journal of Food Engineering*, *78*(3), 846-854.
- Baskaran, G., Masdor, N. A., Syed, M. A., & Shukor, M. Y. (2013). An inhibitive enzyme assay to detect mercury and zinc using protease from Coriandrum sativum. *The Scientific World Journal*, 2013.
- Basu, J. K., Bhattacharyya, D., & Kim, T.-h. (2010). Use of artificial neural network in pattern recognition. *International journal of software engineering and its applications, 4*(2).
- Bera, S., Kauser, H., & Mohanty, K. (2019). Optimization of p-cresol biodegradation using novel bacterial strains isolated from petroleum hydrocarbon fallout. *Journal of Water Process Engineering, 31*, 100842.
- Beshay, U., Abd-El-Haleem, D., Moawad, H., & Zaki, S. (2002). Phenol biodegradation by free and immobilized Acinetobacter. *Biotechnology letters*, 24(15), 1295-1297.
- Betiku, E., Omilakin, O. R., Ajala, S. O., Okeleye, A. A., Taiwo, A. E., & Solomon, B. O. (2014). Mathematical modeling and process parameters optimization studies by artificial neural network and

response surface methodology: a case of non-edible neem (Azadirachta indica) seed oil biodiesel synthesis. *Energy*, *7*2, 266-273.

- Bezerra, M. A., Santelli, R. E., Oliveira, E. P., Villar, L. S., & Escaleira, L. A. (2008). Response surface methodology (RSM) as a tool for optimization in analytical chemistry. *Talanta*, *76*(5), 965-977.
- Biglari, H., Javan, N., Khosravi, R., & Zarei, A. (2016). Direct Blue 71 removal from aqueous solutions by adsorption on Pistachio hull waste: Equilibrium, kinetic and thermodynamic studies. *Iranian Journal of Health Sciences*, 4(2), 55-70.
- Bilal, M., & Asgher, M. (2015). Dye decolorization and detoxification potential of Ca-alginate beads immobilized manganese peroxidase. *BMC biotechnology*, *15*(1), 111.
- Blandino, A., Macías, M., & Cantero, D. (2000). Glucose oxidase release from calcium alginate gel capsules. *Enzyme and microbial technology*, 27(3-5), 319-324.
- Boon, B., & Laudelout, H. (1962). Kinetics of nitrite oxidation by Nitrobacter winogradskyi. *Biochemical Journal, 85*(3), 440.
- Boopathy, R. (2000). Factors limiting bioremediation technologies. *Bioresource technology*, 74(1), 63-67.
- Brisson, D., Vohl, M. C., St-Pierre, J., Hudson, T. J., & Gaudet, D. (2001). Glycerol: a neglected variable in metabolic processes? *Bioessays*, 23(6), 534-542.
- Bromley-Challenor, K., Knapp, J., Zhang, Z., Gray, N., Hetheridge, M., & Evans, M. (2000). Decolorization of an azo dye by unacclimated activated sludge under anaerobic conditions. *Water research, 34*(18), 4410-4418.
- Bulut, Y., Gözübenli, N., & Aydın, H. (2007). Equilibrium and kinetics studies for adsorption of direct blue 71 from aqueous solution by wheat shells. *Journal of hazardous materials, 144*(1-2), 300-306.
- Cassidy, M., Lee, H., & Trevors, J. (1996). Environmental applications of immobilized microbial cells: a review. *Journal of Industrial Microbiology*, *16*(2), 79-101.
- Çetin, D., & Dönmez, G. (2006). Decolorization of reactive dyes by mixed cultures isolated from textile effluent under anaerobic conditions. *Enzyme and Microbial Technology*, 38(7), 926-930.
- Chan, D. (2020). Peanut Factory Ordered To Stop Operations After River Turns Green. Retrieved June 9, 2020, from https://www.nst.com.my/news/nation/2020/06/599128/peanut-factoryordered-stop-operations-after-river-turns-green

- Chan, E.-S., Lim, T.-K., Voo, W.-P., Pogaku, R., Tey, B. T., & Zhang, Z. (2011). Effect of formulation of alginate beads on their mechanical behavior and stiffness. *Particuology*, *9*(3), 228-234.
- Chang, J.-S., Chen, B.-Y., & Lin, Y. S. (2004). Stimulation of bacterial decolorization of an azo dye by extracellular metabolites from Escherichia coli strain NO3. *Bioresource technology*, *91*(3), 243-248.
- Chang, J.-S., Chou, C., & Chen, S.-Y. (2001). Decolorization of azo dyes with immobilized Pseudomonas luteola. *Process Biochemistry*, *36*(8-9), 757-763.
- Chang, J. S., & Lin, Y. C. (2000). Fed-Batch Bioreactor Strategies for Microbial Decolorization of Azo Dye Using a Pseudomonasluteola Strain. *Biotechnology progress, 16*(6), 979-985.
- Charumathi, D., & Das, N. (2010). Removal of synthetic dye basic Violet 3 by immobilised Candida tropicalis grown on sugarcane bagasse extract medium. *Int J Eng Sci Technol,* **2**(9), 4325-4335.
- Chaudhari, A. U., Paul, D., Dhotre, D., & Kodam, K. M. (2017). Effective biotransformation and detoxification of anthraquinone dye reactive blue 4 by using aerobic bacterial granules. *Water research, 122*, 603-613.
- Chen, B.-Y. (2002). Understanding decolorization characteristics of reactive azo dyes by Pseudomonas luteola: toxicity and kinetics. *Process Biochemistry*, *38*(3), 437-446.
- Chen, B.-Y., Chen, S.-Y., Lin, M.-Y., & Chang, J.-S. (2006). Exploring bioaugmentation strategies for azo-dye decolorization using a mixed consortium of Pseudomonas luteola and Escherichia coli. *Process Biochemistry*, *41*(7), 1574-1581.
- Chen, C., Shao, Y., Tao, Y., & Wen, H. (2015). Optimization of dynamic microwave-assisted extraction of Armillaria polysaccharides using RSM, and their biological activity. *LWT-Food Science and Technology*, *64*(2), 1263-1269.
- Chen, H., Hopper, S. L., & Cerniglia, C. E. (2005). Biochemical and molecular characterization of an azoreductase from Staphylococcus aureus, a tetrameric NADPH-dependent flavoprotein. *Microbiology (Reading, England), 151*(Pt 5), 1433.
- Chen, K.-C., Wu, J.-Y., Huang, C.-C., Liang, Y.-M., & Hwang, S.-C. J. (2003). Decolorization of azo dye using PVA-immobilized microorganisms. *Journal of Biotechnology*, *101*(3), 241-252.
- Chen, K.-C., Wu, J.-Y., Liou, D.-J., & Hwang, S.-C. J. (2003). Decolorization of the textile dyes by newly isolated bacterial strains. *Journal of Biotechnology*, *101*(1), 57-68.

- Chen, S., Deng, Y., Chang, C., Lee, J., Cheng, Y., Cui, Z., . . . Zhang, L.-H. (2015). Pathway and kinetics of cyhalothrin biodegradation by Bacillus thuringiensis strain ZS-19. *Scientific reports*, *5*, 8784.
- Chen, S., Yin, H., Ye, J., Peng, H., Liu, Z., Dang, Z., & Chang, J. (2014). Influence of co-existed benzo [a] pyrene and copper on the cellular characteristics of Stenotrophomonas maltophilia during biodegradation and transformation. *Bioresource technology*, *158*, 181-187.
- Choudhary, S., & Sar, P. (2009). Characterization of a metal resistant Pseudomonas sp. isolated from uranium mine for its potential in heavy metal (Ni2+, Co2+, Cu2+, and Cd2+) sequestration. *Bioresource technology*, 100(9), 2482-2492.
- Damodar, R. A., & Swaminathan, T. (2008). Performance evaluation of a continuous flow immobilized rotating tube photocatalytic reactor (IRTPR) immobilized with TiO2 catalyst for azo dye degradation. *Chemical Engineering Journal, 144*(1), 59-66.
- Dasgupta, J., Sikder, J., Chakraborty, S., Curcio, S., & Drioli, E. (2015). Remediation of textile effluents by membrane based treatment techniques: a state of the art review. *Journal of environmental management*, 147, 55-72.
- de-Bashan, L. E., Schmid, M., Rothballer, M., Hartmann, A., & Bashan, Y. (2011). CELL-CELL INTERACTION IN THE EUKARYOTE-PROKARYOTE MODEL OF THE MICROALGAE CHLORELLA VULGARIS AND THE BACTERIUM AZOSPIRILLUM BRASILENSE IMMOBILIZED IN POLYMER BEADS 1. Journal of phycology, 47(6), 1350-1359.
- Derringer, G., & Suich, R. (1980). Simultaneous optimization of several response variables. *Journal of quality technology*, *1*2(4), 214-219.
- Desai, K. M., Survase, S. A., Saudagar, P. S., Lele, S., & Singhal, R. S. (2008). Comparison of artificial neural network (ANN) and response surface methodology (RSM) in fermentation media optimization: case study of fermentative production of scleroglucan. *Biochemical Engineering Journal*, *41*(3), 266-273.
- Devi, P., Wahidullah, S., Sheikh, F., Pereira, R., Narkhede, N., Amonkar, D., . . . Meena, R. M. (2017). Biotransformation and detoxification of xylidine orange dye using immobilized cells of marine-derived Lysinibacillus sphaericus D3. *Marine drugs*, *15*(2), 30.
- Dos Santos, A., Bisschops, I., & Cervantes, F. (2006). Closing process water cycles and product recovery in textile industry: perspective for biological treatment. Advanced biological treatment processes for industrial wastewaters, 1, 298-320.

- Dubin, P., & Wright, K. (1975). Reduction of azo food dyes in cultures of Proteus vulgaris. *Xenobiotica*, 5(9), 563-571.
- Dyes, C. A. S. (2016). Screening Assessment Aromatic Azo and Benzidinebased Substance Grouping Certain Azo Solvent Dyes.
- Ebrahimpour, A., Rahman, R. N. Z. R. A., Ch'ng, D. H. E., Basri, M., & Salleh, A. B. (2008). A modeling study by response surface methodology and artificial neural network on culture parameters optimization for thermostable lipase production from a newly isolated thermophilic Geobacillus sp. strain ARM. *BMC biotechnology*, *8*(1), 96.
- Edwards, V. H. (1970). The influence of high substrate concentrations on microbial kinetics. *Biotechnology and bioengineering*, 12(5), 679-712.
- Elisangela, F., Andrea, Z., Fabio, D. G., de Menezes Cristiano, R., Regina, D. L., & Artur, C.-P. (2009). Biodegradation of textile azo dyes by a facultative Staphylococcus arlettae strain VN-11 using a sequential microaerophilic/aerobic process. *International Biodeterioration & Biodegradation*, 63(3), 280-288.
- Ertugay, N., & Acar, F. N. (2017). Removal of COD and color from Direct Blue 71 azo dye wastewater by Fenton's oxidation: Kinetic study. *Arabian Journal of Chemistry*, *10*, S1158-S1163.
- Ertuğrul, S., Bakır, M., & Dönmez, G. (2008). Treatment of dye-rich wastewater by an immobilized thermophilic cyanobacterial strain: Phormidium sp. *Ecological Engineering*, *32*(3), 244-248.
- Ezhumalai, S., & Thangavelu, V. (2010). Kinetic and optimization studies on the bioconversion of lignocellulosic material into ethanol. *Bioresources*, *5*(3), 1879-1894.
- Farid, A. M., Lubna, A., Choo, T. G., Rahim, M. C., & Mazlin, M. (2016). A review on the chemical pollution of Langat River, Malaysia. *Asian Journal of Water, Environment and Pollution, 13*(1), 9-15.
- Fereidouni, M., Daneshi, A., & Younesi, H. (2009). Biosorption equilibria of binary Cd (II) and Ni (II) systems onto Saccharomyces cerevisiae and Ralstonia eutropha cells: Application of response surface methodology. *Journal of hazardous materials, 168*(2-3), 1437-1448.
- Fetyan, N. A., Abdel Azeiz, A. Z., Ismail, I. M., & Shaban, S. A. (2016). Oxidative Decolorization of Direct Blue 71 Azo Dye by Saccharomyces cerevisiae Catalyzed by Nano Zero-valent Iron.
- Franciscon, E., Grossman, M. J., Paschoal, J. A. R., Reyes, F. G. R., & Durrant, L. R. (2012). Decolorization and biodegradation of reactive sulfonated azo dyes by a newly isolated Brevibacterium sp. strain VN-15. *SpringerPlus*, *1*(1), 37.

- Franciscon, E., Zille, A., Fantinatti-Garboggini, F., Silva, I. S., Cavaco-Paulo, A., & Durrant, L. R. (2009). Microaerophilic–aerobic sequential decolourization/biodegradation of textile azo dyes by a facultative Klebsiella sp. strain VN-31. *Process Biochemistry*, 44(4), 446-452.
- García-Montaño, J., Domenech, X., García-Hortal, J. A., Torrades, F., & Peral, J. (2008). The testing of several biological and chemical coupled treatments for Cibacron Red FN-R azo dye removal. *Journal of hazardous materials*, 154(1-3), 484-490.
- Gauthier, P. T., Norwood, W. P., Prepas, E. E., & Pyle, G. G. (2014). Metal– PAH mixtures in the aquatic environment: a review of co-toxic mechanisms leading to more-than-additive outcomes. *Aquatic toxicology*, *154*, 253-269.
- Georgiou, D., Metallinou, C., Aivasidis, A., Voudrias, E., & Gimouhopoulos, K. (2004). Decolorization of azo-reactive dyes and cotton-textile wastewater using anaerobic digestion and acetate-consuming bacteria. *Biochemical Engineering Journal, 19*(1), 75-79.
- Ghodake, G., Jadhav, U., Tamboli, D., Kagalkar, A., & Govindwar, S. (2011). Decolorization of textile dyes and degradation of mono-azo dye amaranth by Acinetobacter calcoaceticus NCIM 2890. Indian journal of microbiology, 51(4), 501-508.
- Ghosh, D. K., Mandal, A., & Chaudhuri, J. (1992). Purification and partial characterization of two azoreductases from Shigella dysenteriae type 1. *FEMS microbiology letters, 98*(1-3), 229-233.
- Gianfreda, L., & Rao, M. A. (2004). Potential of extra cellular enzymes in remediation of polluted soils: a review. *Enzyme and microbial technology*, *35*(4), 339-354.
- Giller, K. E., Witter, E., & McGrath, S. P. (2009). Heavy metals and soil microbes. *Soil Biology and Biochemistry*, *41*(10), 2031-2037.
- Górecka, E., & Jastrzębska, M. (2011). Immobilization techniques and biopolymer carriers. *Biotechnology and Food Science*, *75*(1), 65-86.
- Gupta, V. (2009). Application of low-cost adsorbents for dye removal–a review. *Journal of environmental management, 90*(8), 2313-2342.
- Gusmanizar, N., Halmi, M. I. E., Rusnam, M., Rahman, M. F. A., Shukor, M. S., Azmi, N. S., & Shukor, M. Y. (2016). Isolation and characterization of a molybdenum-reducing and azo-dye decolorizing Serratia marcescens strain neni-1 from Indonesian soil. *Journal of Urban and Environmental Engineering*, 10(1), 113-123.
- Hafshejani, M. K., Ogugbue, C. J., & Morad, N. (2014). Application of response surface methodology for optimization of decolorization and

mineralization of triazo dye Direct Blue 71 by Pseudomonas aeruginosa. *3 Biotech, 4*(6), 605-619.

- Haldane, J. B. S. (1931). The molecular statistics of an enzyme action. *Proceedings of the Royal Society of London. Series B, Containing Papers of a Biological Character, 108*(759), 559-567.
- Hameed, B. B., & Ismail, Z. Z. (2018). Decolorization, biodegradation and detoxification of reactive red azo dye using non-adapted immobilized mixed cells. *Biochemical Engineering Journal*, 137, 71-77.
- Hasan, H. A., Abdullah, S. R. S., Kamarudin, S. K., & Kofli, N. T. (2009). A review on the design criteria of biological aerated filter for COD, ammonia and manganese removal in drinking water treatment. *Inst. Eng*, *70*(4), 25-33.
- Hasan, H. A., Abdullah, S. R. S., Kamarudin, S. K., & Kofli, N. T. (2011). Response surface methodology for optimization of simultaneous COD, NH4+–N and Mn2+ removal from drinking water by biological aerated filter. *Desalination*, 275(1-3), 50-61.
- He, F., Hu, W., & Li, Y. (2004). Biodegradation mechanisms and kinetics of azo dye 4BS by a microbial consortium. *Chemosphere*, *57*(4), 293-301.
- Hill, W. J., & Hunter, W. G. (1966). A review of response surface methodology: a literature survey. *Technometrics*, *8*(4), 571-590.
- Hsueh, C.-C., & Chen, B.-Y. (2007). Comparative study on reaction selectivity of azo dye decolorization by Pseudomonas luteola. *Journal of Hazardous Materials*, 141(3), 842-849.
- Hutchinson, U., Ntwampe, S. K., Ngongang, M. M., Du Plessis, H., Chidi, B., Saulse, C., & Jolly, N. (2018). *Cell immobilization by Gel entrapment in Ca-alginate beads for balsamic-styled vinegar production.* Paper presented at the Proceedings of the 10th International Conference on Advances in Science, Engineering, Technology and Healthcare (ASETH-18) November.
- Isak, S. J., Eyring, E. M., Spikes, J. D., & Meekins, P. A. (2000). Direct blue dye solutions: photo properties. *Journal of Photochemistry and Photobiology A: Chemistry*, 134(1-2), 77-85.
- Işik, M., & Sponza, D. T. (2003). Effect of oxygen on decolorization of azo dyes by Escherichia coli and Pseudomonas sp. and fate of aromatic amines. *Process Biochemistry*, 38(8), 1183-1192.
- Işık, M., & Sponza, D. T. (2008). Anaerobic/aerobic treatment of a simulated textile wastewater. Separation and Purification Technology, 60(1), 64-72.

- Islahuddin, N. K. S., Halmi, M. I. E., Manogaran, M., & Shukor, M. Y. (2017). Isolation and culture medium optimisation using one-factor-at-time and Response Surface Methodology on the biodegradation of the azo-dye amaranth. *Bioremediation Science and Technology Research*, 5(2), 25-31.
- Islam, M. M., Mahmud, K., Faruk, O., & Billah, M. (2011). Textile dyeing industries in Bangladesh for sustainable development. *International Journal of Environmental Science and Development*, 2(6), 428.
- Jadhav, U. U., Dawkar, V. V., Ghodake, G. S., & Govindwar, S. P. (2008). Biodegradation of Direct Red 5B, a textile dye by newly isolated Comamonas sp. UVS. *Journal of hazardous materials*, 158(2-3), 507-516.
- Jadhav, U. U., Dawkar, V. V., Kagalkar, A. N., & Govindwar, S. P. (2011). Effect of metals on decolorization of reactive blue HERD by Comamonas sp. UVS. *Water, Air, & Soil Pollution, 216*(1-4), 621-631.
- Jain, K., Shah, V., Chapla, D., & Madamwar, D. (2012). Decolorization and degradation of azo dye–Reactive Violet 5R by an acclimatized indigenous bacterial mixed cultures-SB4 isolated from anthropogenic dye contaminated soil. *Journal of hazardous materials*, 213, 378-386.
- Jain, N., Shrivastava, S., & Shrivastava, A. (1997). Treatment of pulp mill wastewater by bacterial strain Acinetobacter calcoaceticus. Indian journal of experimental biology, 35(2), 139-143.
- Jin, R., Yang, H., Zhang, A., Wang, J., & Liu, G. (2009). Bioaugmentation on decolorization of CI Direct Blue 71 by using genetically engineered strain Escherichia coli JM109 (pGEX-AZR). *Journal of Hazardous Materials*, 163(2-3), 1123-1128.
- Jobby, R., Jha, P., Kudale, S., Kale, A., & Desai, N. (2019). Biodegradation of textile dye Direct Blue 71 using root nodulating Rhizobium sp.
- Kant, R. (2012). Textile dyeing industry an environmental hazard. *Natural science*, *4*(1), 22-26.
- Kapdan, I. K., Kargi, F., McMullan, G., & Marchant, R. (2000). Decolorization of textile dyestuffs by a mixed bacterial consortium. *Biotechnology Letters*, 22(14), 1179-1181.
- Kapdan, I. K., Tekol, M., & Sengul, F. (2003). Decolorization of simulated textile wastewater in an anaerobic–aerobic sequential treatment system. *Process Biochemistry*, 38(7), 1031-1037.
- Kaur, B., Kumar, B., Garg, N., & Kaur, N. (2015). Statistical optimization of conditions for decolorization of synthetic dyes by Cordyceps militaris MTCC 3936 using RSM. *BioMed research international*, 2015.

- Khataee, A., Gholami, P., & Sheydaei, M. (2016). Heterogeneous Fenton process by natural pyrite for removal of a textile dye from water: effect of parameters and intermediate identification. *Journal of the Taiwan Institute of Chemical Engineers, 58*, 366-373.
- Khehra, M. S., Saini, H. S., Sharma, D. K., Chadha, B. S., & Chimni, S. S. (2005). Decolorization of various azo dyes by bacterial consortium. *Dyes and Pigments*, 67(1), 55-61.
- Khehra, M. S., Saini, H. S., Sharma, D. K., Chadha, B. S., & Chimni, S. S. (2006). Biodegradation of azo dye CI Acid Red 88 by an anoxic– aerobic sequential bioreactor. *Dyes and Pigments*, 70(1), 1-7.
- Khoshdel, A., & Mahmoodzadeh Vaziri, B. (2016). Novel mathematical models for prediction of microbial growth kinetics and contaminant degradation in bioremediation process. *Journal of Environmental Engineering and Landscape Management*, 24(3), 157-164.
- Khuri, A. I., & Mukhopadhyay, S. (2010). Response surface methodology. *Wiley Interdisciplinary Reviews: Computational Statistics, 2*(2), 128-149.
- Kilickap, E. (2010). Modeling and optimization of burr height in drilling of Al-7075 using Taguchi method and response surface methodology. *The International Journal of Advanced Manufacturing Technology, 49*(9-12), 911-923.
- Kim, S., Park, C., Kim, T.-H., Lee, J., & Kim, S.-W. (2003). COD reduction and decolorization of textile effluent using a combined process. *Journal of Bioscience and Bioengineering*, 95(1), 102-105.
- Knezevic, Z., Bobic, S., Milutinovic, A., Obradovic, B., Mojovic, L., & Bugarski, B. (2002). Alginate-immobilized lipase by electrostatic extrusion for the purpose of palm oil hydrolysis in lecithin/isooctane system. *Process Biochemistry*, *38*(3), 313-318.
- Kodam, K., Soojhawon, I., Lokhande, P., & Gawai, K. (2005). Microbial decolorization of reactive azo dyes under aerobic conditions. *World Journal of Microbiology and Biotechnology*, 21(3), 367-370.
- Kramer, M. G. (2016). Determination of plasmid segregational stability in a growing bacterial population *Bacterial Therapy of Cancer* (pp. 125-133): Springer.
- Krishnamoorthy, R., Roy Choudhury, A., Arul Jose, P., Suganya, K., Senthilkumar, M., Prabhakaran, J., . . . Anandham, R. (2021). Long-Term Exposure to Azo Dyes from Textile Wastewater Causes the Abundance of Saccharibacteria Population. *Applied Sciences, 11*(1), 379.

- Krishnan, J., Kishore, A. A., Suresh, A., Murali, A. K., & Vasudevan, J. (2017). Biodegration Kinectics of Azo Dye Mixture: Substrate Inhibition Modeling. RESEARCH JOURNAL OF PHARMACEUTICAL BIOLOGICAL AND CHEMICAL SCIENCES, 8, 365-375.
- Kumar, A., Priyadarshinee, R., Singha, S., Sengupta, B., Roy, A., Dasgupta, D., & Mandal, T. (2019). Biodegradation of alkali lignin by Bacillus flexus RMWW II: analyzing performance for abatement of rice mill wastewater. *Water Science and Technology*, *80*(9), 1623-1632.
- Kumaran, N., & Dharani, G. (2011). Decolorization of textile dyes by white rot fungi Phanerocheate chrysosporium and Pleurotus sajor-caju. *Journal of applied technology in environmental sanitation, 1*(4).
- KUSNIN, N. B. (2015). BIODEGRADATION OF ACRYLAMIDE BY A NEWLY ISOLATED Bacillus sp. strain ZK34.
- Lade, H., Kadam, A., Paul, D., & Govindwar, S. (2015). Biodegradation and detoxification of textile azo dyes by bacterial consortium under sequential microaerophilic/aerobic processes. *EXCLI journal, 14*, 158.
- Li, C., & Fang, H. H. (2007). Inhibition of heavy metals on fermentative hydrogen production by granular sludge. *Chemosphere, 67*(4), 668-673.
- Lin, J., Zhang, X., Li, Z., & Lei, L. (2010). Biodegradation of Reactive blue 13 in a two-stage anaerobic/aerobic fluidized beds system with a Pseudomonas sp. isolate. *Bioresource technology*, *101*(1), 34-40.
- Lin, S. H., & Lai, C. L. (1999). Catalytic oxidation of dye wastewater by metal oxide catalyst and granular activated carbon. *Environment International*, *25*(4), 497-504.
- Liu, H.-L., & Chiou, Y.-R. (2005). Optimal decolorization efficiency of Reactive Red 239 by UV/TiO2 photocatalytic process coupled with response surface methodology. *Chemical Engineering Journal*, *112*(1-3), 173-179.
- Lokesh, K., & Sivakiran, R. (2014). Biological methods of dye removal from textile effluents-A review. *Journal of Biochemical Technology, 3*(5), 177-180.
- Lu, L., Zhao, M., & Wang, Y. (2007). Immobilization of laccase by alginate– chitosan microcapsules and its use in dye decolorization. *World journal* of microbiology and biotechnology, 23(2), 159-166.
- Luong, J. (1985). Kinetics of ethanol inhibition in alcohol fermentation. *Biotechnology and bioengineering*, 27(3), 280-285.

- Luong, J. (1987). Generalization of Monod kinetics for analysis of growth data with substrate inhibition. *Biotechnology and bioengineering*, *29*(2), 242-248.
- Maier, J. r., Kandelbauer, A., Erlacher, A., Cavaco-Paulo, A., & Gübitz, G. M. (2004). A new alkali-thermostable azoreductase from Bacillus sp. strain SF. Applied and environmental microbiology, 70(2), 837-844.
- Mathew, S., & Madamwar, D. (2004). Decolorization of ranocid fast blue dye by bacterial consortium SV5. *Applied biochemistry and biotechnology*, *118*(1-3), 371-381.
- McMullan, G., Meehan, C., Conneely, A., Kirby, N., Robinson, T., Nigam, P., . . . Smyth, W. (2001). Microbial decolourisation and degradation of textile dyes. *Applied microbiology and biotechnology*, *56*(1), 81-87.
- Meng, X., Liu, G., Zhou, J., & Fu, Q. S. (2014). Effects of redox mediators on azo dye decolorization by Shewanella algae under saline conditions. *Bioresource technology*, *151*, 63-68.
- Mirhosseini, H., Tan, C. P., Hamid, N. S. A., & Yusof, S. (2007). Modeling the relationship between the main emulsion components and stability, viscosity, fluid behavior, ζ-potential, and electrophoretic mobility of orange beverage emulsion using response surface methodology. *J. of Agricultural and Food Chemistry*, *55*(19), 7659-7666.
- Mirzaei, N., Mahvi, A. H., & Hossini, H. (2018). Equilibrium and kinetics studies of Direct blue 71 adsorption from aqueous solutions using modified zeolite. *Adsorption Science & Technology*, *36*(1-2), 80-94.
- Mitra, B. (2012). Synthesis and activation of Immobilized beads by natural dye extracts. *Int. J. Drug Dev. & Res, 4*(1), 304-310.
- Mohana, S., Shrivastava, S., Divecha, J., & Madamwar, D. (2008). Response surface methodology for optimization of medium for decolorization of textile dye Direct Black 22 by a novel bacterial consortium. *Bioresource technology*, *99*(3), 562-569.
- Mondol, J. D., Yohanis, Y. G., & Norton, B. (2008). Solar radiation modelling for the simulation of photovoltaic systems. *Renewable Energy*, *33*(5), 1109-1120.
- Monod, J. (1949). The growth of bacterial cultures. *Annual review of microbiology*, *3*(1), 371-394.
- Montgomery, D. (2001). Discovering dispersion effects. *Design and Analysis of Experiments. 5th ed. New york: John Wiley.*
- Montgomery, D. C. (2017). *Design and analysis of experiments*: John wiley & sons.

- Moon, S. H., & Parulekar, S. J. (1991). A parametric study ot protease production in batch and fed-batch cultures of Bacillus firmus. *Biotechnology and bioengineering*, *37*(5), 467-483.
- Moosvi, S., Keharia, H., & Madamwar, D. (2005). Decolourization of textile dye Reactive Violet 5 by a newly isolated bacterial consortium RVM 11.1. *World Journal of Microbiology and Biotechnology, 21*(5), 667-672.
- Moosvi, S., Kher, X., & Madamwar, D. (2007). Isolation, characterization and decolorization of textile dyes by a mixed bacterial consortium JW-2. *Dyes and Pigments, 74*(3), 723-729.
- Muhamad, M. H., Abdullah, S. R. S., Mohamad, A. B., Rahman, R. A., & Kadhum, A. A. H. (2013). Application of response surface methodology (RSM) for optimisation of COD, NH3–N and 2, 4-DCP removal from recycled paper wastewater in a pilot-scale granular activated carbon sequencing batch biofilm reactor (GAC-SBBR). Journal of environmental management, 121, 179-190.
- Munawar, I., Bhatti, I. A., Muhammad, Z.-u.-R., Bhatti, H. N., & Muhammad, S. (2014). Efficiency of advanced oxidation processes for detoxification of industrial effluents. *Asian Journal of Chemistry*, 26(14), 4291-4296.
- Murugesan, K., Kim, Y.-M., Jeon, J.-R., & Chang, Y.-S. (2009). Effect of metal ions on reactive dye decolorization by laccase from Ganoderma lucidum. *Journal of hazardous materials*, *168*(1), 523-529.
- Muthu, S. S. (2014). Assessing the environmental impact of textiles and the clothing supply chain: Elsevier.
- Myers, R. H., Montgomery, D. C., & Anderson-Cook, C. M. (2016). Response surface methodology: process and product optimization using designed experiments: John Wiley & Sons.
- Nachiyar, C. V., & Rajakumar, G. S. (2005). Purification and characterization of an oxygen insensitive azoreductase from Pseudomonas aeruginosa. *Enzyme and Microbial Technology*, *36*(4), 503-509.
- Nakanishi, M., Yatome, C., Ishida, N., & Kitade, Y. (2001). Putative ACP phosphodiesterase gene (acpD) encodes an azoreductase. *Journal of Biological Chemistry*, 276(49), 46394-46399.
- Nasir, N. A. H. A., Pa, N. F. Z. C., Roslani, M. A., Ramli, R., & Zain, N. A. M. (2019). Decolourization of Turqoise Blue (Remazol Blue BB) Dye by Immobilized Penicillium sp. into Sodium-Alginate-Sulfate Beads. *Jurnal Intelek*, 14(2), 153-160.
- Nigam, P., Armour, G., Banat, I., Singh, D., & Marchant, R. (2000). Physical removal of textile dyes from effluents and solid-state fermentation of dye-adsorbed agricultural residues. *Bioresource technology*, *7*2(3), 219-226.

- Nigam, P., Banat, I. M., Singh, D., & Marchant, R. (1996). Microbial process for the decolorization of textile effluent containing azo, diazo and reactive dyes. *Process Biochemistry*, *31*(5), 435-442.
- Noordin, M. Y., Venkatesh, V. C., Sharif, S., Elting, S., & Abdullah, A. (2004). Application of response surface methodology in describing the performance of coated carbide tools when turning AISI 1045 steel. *Journal of Materials Processing Technology*, 145(1), 46-58.
- Okpokwasili, G., & Nweke, C. (2006). Microbial growth and substrate utilization kinetics. *African Journal of Biotechnology*, *5*(4), 305-317.
- Ollis, D. F. (2000). Photocatalytic purification and remediation of contaminated air and water. *Comptes Rendus de l'Académie des Sciences-Series IIC-Chemistry*, 3(6), 405-411.
- Othman, A., Bakar, N., Halmi, M., Johari, W., Ahmad, S., Jirangon, H., . . . Shukor, M. (2013). Kinetics of molybdenum reduction to molybdenum blue by Bacillus sp. strain A. rzi. *BioMed research international*, 2013.
- Özacar, M., & Şengil, I. A. (2003). Adsorption of reactive dyes on calcined alunite from aqueous solutions. *Journal of Hazardous Materials*, *98*(1-3), 211-224.
- Pandey, A., Singh, P., & Iyengar, L. (2007). Bacterial decolorization and degradation of azo dyes. *International Biodeterioration & Biodegradation*, *59*(2), 73-84.
- Pang, Y. L., & Abdullah, A. Z. (2013). Current status of textile industry wastewater management and research progress in Malaysia: a review. *Clean–Soil, Air, Water, 41*(8), 751-764.
- Parales, R. E., Bruce, N., Schmid, A., & Wackett, L. (2002). Biodegradation, biotransformation, and biocatalysis (B3). Appl. Environ. Microbiol., 68(10), 4699-4709.
- Parshetti, G., Kalme, S., Saratale, G., & Govindwar, S. (2006). Biodegradation of Malachite Green by Kocuria rosea MTCC 1532. *Acta Chimica Slovenica*, *53*(4).
- Pearce, C., Lloyd, J., & Guthrie, J. (2003). The removal of colour from textile wastewater using whole bacterial cells: a review. *Dyes and Pigments*, *58*(3), 179-196.
- Pereira, S. I. A., Lima, A. I. G., & Figueira, E. M. d. A. P. (2006). Heavy metal toxicity in Rhizobium leguminosarum biovar viciae isolated from soils subjected to different sources of heavy-metal contamination: effects on protein expression. *Applied Soil Ecology*, 33(3), 286-293.

- Plumb, J. J., Bell, J., & Stuckey, D. C. (2001). Microbial populations associated with treatment of an industrial dye effluent in an anaerobic baffled reactor. *Applied and environmental microbiology*, 67(7), 3226-3235.
- Pointing, S., Bucher, V., & Vrijmoed, L. (2000). Dye decolorization by subtropical basidiomycetous fungi and the effect of metals on decolorizing ability. *World Journal of Microbiology and Biotechnology*, 16(2), 199-205.
- Pourbabaee, A., Malekzadeh, F., Sarbolouki, M., & Najafi, F. (2006). Aerobic decolorization and detoxification of a disperse dye in textile effluent by a new isolate of Bacillus sp. *Biotechnology and bioengineering*, 93(4), 631-635.
- Prabha, S., Gogoi, A., Mazumder, P., Ramanathan, A., & Kumar, M. (2017). Assessment of the impact of textile effluents on microbial diversity in Tirupur district, Tamil Nadu. *Applied water science*, 7(5), 2267-2277.
- Prasad, R. (2017). Mycoremediation and environmental sustainability: Springer.
- Puvaneswari, N., Muthukrishnan, J., & Gunasekaran, P. (2002). Biodegradation of benzidine based azodyes Direct red and Direct blue by the immobilized cells of Pseudomonas fluorescens D41.
- Rafii, F., Franklin, W., & Cerniglia, C. E. (1990). Azoreductase activity of anaerobic bacteria isolated from human intestinal microflora. *Applied* and environmental microbiology, 56(7), 2146-2151.
- Raj, D. S., Prabha, R. J., & Leena, R. (2012). Analysis of bacterial degradation of azo dye congo red using HPLC. *J Ind Pollut Control, 28*(1), 57-62.
- Raja, M. M. M., Raja, A., Salique, S. M., & Gajalakshmi, P. (2016). Studies on effect of marine actinomycetes on amido black (azo dye) decolorization. *J Chem Pharmac Res, 8*(8), 640-644.
- Rehman, K., Shahzad, T., Sahar, A., Hussain, S., Mahmood, F., Siddique, M.
 H., . . . Rashid, M. I. (2018). Effect of Reactive Black 5 azo dye on soil processes related to C and N cycling. *PeerJ, 6*, e4802.
- Reyes, P., Pickard, M. A., & Vazquez-Duhalt, R. (1999). Hydroxybenzotriazole increases the range of textile dyes decolorized by immobilized laccase. *Biotechnology letters, 21*(10), 875-880.
- Rosa, J., Prado, K., Alves, W., Pereira, F., Santana, J., & Tambourgi, E. (2013). Applying of a neural network in effluent treatment simulation as an environmental solution for textile industry. *Chemical Engineering Transactions, 32*.
- Rudakiya, D. M., & Pawar, K. S. (2013). Optimization of culture condition for enhanced decolorization of Reactive Orange 16 by Comamonas

acidovorans MTCC 3364. International Journal of Current Microbiology and Applied Sciences, 2(10), 467-476.

- Russ, R., Rau, J. r., & Stolz, A. (2000). The function of cytoplasmic flavin reductases in the reduction of azo dyes by bacteria. *Applied and environmental microbiology*, *66*(4), 1429-1434.
- Saien, J., & Soleymani, A. (2007). Degradation and mineralization of Direct Blue 71 in a circulating upflow reactor by UV/TiO2 process and employing a new method in kinetic study. *Journal of Hazardous Materials, 144*(1-2), 506-512.
- Salah, Z. (2018). Isolation and Characterization of ISA Degrading Alkaliphilic Bacteria. University of Huddersfield.
- Sandhya, S. (2010). Biodegradation of azo dyes under anaerobic condition: role of azoreductase. *Biodegradation of azo dyes*, 39-57.
- Sandrin, T. R., & Hoffman, D. R. (2007). Bioremediation of organic and metal co-contaminated environments: effects of metal toxicity, speciation, and bioavailability on biodegradation *Environmental Bioremediation Technologies* (pp. 1-34): Springer.
- Saratale, R., Saratale, G., Chang, J.-S., & Govindwar, S. (2009). Ecofriendly degradation of sulfonated diazo dye CI Reactive Green 19A using Micrococcus glutamicus NCIM-2168. *Bioresource technology, 100*(17), 3897-3905.
- Saratale, R., Saratale, G., Chang, J.-S., & Govindwar, S. P. (2010). Decolorization and biodegradation of reactive dyes and dye wastewater by a developed bacterial consortium. *Biodegradation*, 21(6), 999-1015.
- Sarayu, K., & Sandhya, S. (2010). Aerobic biodegradation pathway for Remazol Orange by Pseudomonas aeruginosa. *Applied biochemistry and biotechnology*, *160*(4), 1241-1253.
- Schütte, U. M., Abdo, Z., Bent, S. J., Shyu, C., Williams, C. J., Pierson, J. D., & Forney, L. J. (2008). Advances in the use of terminal restriction fragment length polymorphism (T-RFLP) analysis of 16S rRNA genes to characterize microbial communities. *Applied microbiology and biotechnology*, 80(3), 365-380.
- Shah, T. P., & Shah, P. J. (2013). Connectionist Expert System for Medical Diagnosis using ANN–A case study of skin disease Scabies. *International Journal, 3*(8).
- Shanmugam, B. K., & Mahadevan, S. (2015). Metabolism and biotransformation of azo dye by bacterial consortium studied in a bioreaction calorimeter. *Bioresource technology*, *196*, 500-508.

- Sharma, D., Saini, H., Singh, M., Chimni, S., & Chadha, B. (2004). Biological treatment of textile dye Acid violet-17 by bacterial consortium in an up-flow immobilized cell bioreactor. *Letters in applied microbiology, 38*(5), 345-350.
- Shukor, M. (2014). Statistical Diagnostic Tests of the Luong Model in fitting Molybdenum Reduction from the bacterium Bacillus sp. strain A. rzi. *Journal of Environmental Microbiology and Toxicology*, 2(2).
- Singh, B. K., & Walker, A. (2006). Microbial degradation of organophosphorus compounds. *FEMS microbiology reviews*, *30*(3), 428-471.
- Singh, K., & Arora, S. (2011). Removal of synthetic textile dyes from wastewaters: a critical review on present treatment technologies. *Critical reviews in environmental science and technology, 41*(9), 807-878.
- Singh, R. P., Singh, P. K., & Singh, R. L. (2014). Bacterial decolorization of textile azo dye acid orange by Staphylococcus hominis RMLRT03. *Toxicology international*, 21(2), 160.
- Singh, S., & Pakshirajan, K. (2010). Enzyme activities and decolourization of single and mixed azo dyes by the white-rot fungus Phanerochaete chrysosporium. *International Biodeterioration & Biodegradation, 64*(2), 146-150.
- Sobolev, D., & Begonia, M. (2008). Effects of heavy metal contamination upon soil microbes: lead-induced changes in general and denitrifying microbial communities as evidenced by molecular markers. *International journal of environmental research and public health, 5*(5), 450-456.
- Soil, E. P. o. (2003). Manual on Methods and Criteria for Harmonized Sampling, Assessment, Monitoring and Analysis of the Effects of Air Pollution on Forests, Part IIIa, Annex 1, Methods for Soil Analysis. Sampling and analysis of soil. Int. co-operative programme on assessment and monitoring of air pollution effects on forests.
- Song, S., Ying, H., He, Z., & Chen, J. (2007). Mechanism of decolorization and degradation of CI Direct Red 23 by ozonation combined with sonolysis. *Chemosphere, 66*(9), 1782-1788.
- Soni, R. K., Acharya, P., & Modi, H. (2015). Elucidation of biodegradation mechanism of Reactive Red 35 by Pseudomonas aeruginosa ARSKS20. J. Environ. Sci. Toxicol. Food. Tech, 9, 31-40.
- Soon, A. N., & Hameed, B. (2011). Heterogeneous catalytic treatment of synthetic dyes in aqueous media using Fenton and photo-assisted Fenton process. *Desalination*, 269(1-3), 1-16.

- Sponza, D., & Işik, M. (2002). Decolorization and azo dye degradation by anaerobic/aerobic sequential process. *Enzyme and Microbial Technology*, *31*(1-2), 102-110.
- Sponza, D. T., & Işık, M. (2005). Reactor performances and fate of aromatic amines through decolorization of Direct Black 38 dye under anaerobic/aerobic sequentials. *Process Biochemistry*, 40(1), 35-44.
- Sreedharan, V., Saha, P., & Rao, K. V. B. (2021). Dye degradation potential of Acinetobacter baumannii strain VITVB against commercial azo dyes. *Bioremediation Journal*, 1-27.
- Stoilova, I., Stanchev, V., Dimitrova, G., Angelova, G., & Krastanov, A. (2016). Ggrowth kinetics and biodegradation of high inhibitory concentration of phenol, catechol and 2, 4-dichlorophenol by Trametes versicolor 1. European Journal of Biomedical and Pharmaceutical Sciences, 3(12), 139-144.
- Stolz, A. (2001). Basic and applied aspects in the microbial degradation of azo dyes. *Applied microbiology and biotechnology*, *56*(1), 69-80.
- Su, C.-C., Pukdee-Asa, M., Ratanatamskul, C., & Lu, M.-C. (2011). Effect of operating parameters on decolorization and COD removal of three reactive dyes by Fenton's reagent using fluidized-bed reactor. *Desalination*, *278*(1-3), 211-218.
- Sudha, M., Bakiyaraj, G., Saranya, A., Sivakumar, N., & Selvakumar, G. (2018). Prospective assessment of the Enterobacter aerogenes PP002 in decolorization and degradation of azo dyes DB 71 and DG 28. *Journal of environmental chemical engineering*, 6(1), 95-109.
- Sun, Y., Liu, J., & Kennedy, J. F. (2010). Application of response surface methodology for optimization of polysaccharides production parameters from the roots of Codonopsis pilosula by a central composite design. *Carbohydrate Polymers*, 80(3), 949-953.
- Syed, M., Sim, H., Khalid, A., & Shukor, M. (2009). A simple method to screen for azo-dye-degrading bacteria. *J. Environ. Biol, 30*(1), 89-92.
- Tan, Y., Wang, Z. X., & Marshall, K. C. (1998). Modeling pH effects on microbial growth: a statistical thermodynamic approach. *Biotechnology* and bioengineering, 59(6), 724-731.
- Tauber, M. M., Gübitz, G. M., & Rehorek, A. (2008). Degradation of azo dyes by oxidative processes–laccase and ultrasound treatment. *Bioresource technology*, *99*(10), 4213-4220.
- Tazdait, D., ABDI, N., GRIB, H., LOUNICI, H., PAUSS, A., & MAMERI, N. (2014). Comparison of different models of substrate inhibition in aerobic batch biodegradation of malathion. *Turkish Journal of Engineering and Environmental Sciences*, 37(3), 221-230.

- Tazdaït, D., Salah, R., Grib, H., Abdi, N., & Mameri, N. (2018). Kinetic study on biodegradation of glyphosate with unacclimated activated sludge. International journal of environmental health research, 28(4), 448-459.
- Teissier, G. (1942). Growth of bacterial populations and the available substrate concentration. *Rev Sci Instrum, 3208*(3208), 209-214.
- Tessier, G. (1942). Croissance des populations bactériennes et quantité d'aliment disponible. *Rev. Sci. Paris, 80*, 209.
- Thiruppathi, K., Rangasamy, K., Ramasamy, M., & Muthu, D. (2021). Evaluation of textile dye degrading potential of ligninolytic bacterial consortia. *Environmental Challenges*, 4, 100078.
- Thomas, T., Gilbert, J., & Meyer, F. (2012). Metagenomics-a guide from sampling to data analysis. *Microbial informatics and experimentation*, 2(1), 3.
- Tichonovas, M., Krugly, E., Racys, V., Hippler, R., Kauneliene, V., Stasiulaitiene, I., & Martuzevicius, D. (2013). Degradation of various textile dyes as wastewater pollutants under dielectric barrier discharge plasma treatment. *Chemical Engineering Journal, 229*, 9-19.
- Ting, A. S. Y., Lee, M. V. J., Chow, Y. Y., & Cheong, S. L. (2016). Novel exploration of endophytic Diaporthe sp. for the biosorption and biodegradation of triphenylmethane dyes. *Water, Air, & Soil Pollution,* 227(4), 109.
- Tiong, K. G. (2015). Adsorption studies of methylene blue using selected agrowastes as low cost adsorbents/Tiong Khing Gueok. University of Malaya.
- Tony, B. D., Goyal, D., & Khanna, S. (2009). Decolorization of textile azo dyes by aerobic bacterial consortium. *International Biodeterioration & Biodegradation*, 63(4), 462-469.
- Turhan, K., & Turgut, Z. (2009). Decolorization of direct dye in textile wastewater by ozonization in a semi-batch bubble column reactor. *Desalination*, *242*(1-3), 256-263.
- Ullhyan, A., & Ghosh, U. (2012). Decolorization of irgalite dye by immobilized Pseudomonas putida on activated carbon, prepared from agriculture waste. *African Journal of Environmental Science and Technology, 6*(2), 146-154.
- Verma, M., Brar, S., Blais, J., Tyagi, R., & Surampalli, R. (2006). Aerobic biofiltration processes—Advances in wastewater treatment. *Practice Periodical of Hazardous, Toxic, and Radioactive Waste Management,* 10(4), 264-276.

- Vidali, M. (2001). Bioremediation. an overview. *Pure and Applied Chemistry*, 73(7), 1163-1172.
- Wang, G. G. (2003). Adaptive response surface method using inherited latin hypercube design points. *Journal of Mechanical Design*, 125(2), 210-220.
- Wang, Q., Ma, H., Xu, W., Gong, L., Zhang, W., & Zou, D. (2008). Ethanol production from kitchen garbage using response surface methodology. *Biochemical Engineering Journal*, *39*(3), 604-610.
- Wang, S.-C. (2003). Artificial neural network *Interdisciplinary computing in java programming* (pp. 81-100): Springer.
- Webb, J. L. (1963). Enzyme and metabolic inhibitors.
- Wu, X., Monchy, S., Taghavi, S., Zhu, W., Ramos, J., & van der Lelie, D. (2011). Comparative genomics and functional analysis of niche-specific adaptation in Pseudomonas putida. *FEMS microbiology reviews*, 35(2), 299-323.
- Xie, X., Liu, N., Yang, B., Yu, C., Zhang, Q., Zheng, X., . . . Liu, J. (2016). Comparison of microbial community in hydrolysis acidification reactor depending on different structure dyes by Illumina MiSeq sequencing. *International Biodeterioration & Biodegradation, 111*, 14-21.
- Yano, T., & Koga, S. (1969). Dynamic behavior of the chemostat subject to substrate inhibition. *Biotechnology and bioengineering*, *11*(2), 139-153.
- Yano, T., Nakahara, T., Kamiyama, S., & Yamada, K. (1966). Kinetic studies on microbial activities in concentrated solutions. Part. I. Effect of excess sugars on oxygen uptake rate of a cell free respiratory system. *Agricultural and Biological Chemistry*, 30(1), 42-48.
- Yilmaz, I., & Kaynar, O. (2011). Multiple regression, ANN (RBF, MLP) and ANFIS models for prediction of swell potential of clayey soils. *Expert systems with applications, 38*(5), 5958-5966.
- You, S.-J., & Teng, J.-Y. (2009). Anaerobic decolorization bacteria for the treatment of azo dye in a sequential anaerobic and aerobic membrane bioreactor. *Journal of the Taiwan Institute of Chemical Engineers*, 40(5), 500-504.
- Yu, J., Wang, X., & Yue, P. L. (2001). Optimal decolorization and kinetic modeling of synthetic dyes by Pseudomonas strains. *Water research*, 35(15), 3579-3586.
- Zabłocka-Godlewska, E., Przystaś, W., & Grabińska-Sota, E. (2018). Possibilities of obtaining from highly polluted environments: new bacterial strains with a significant decolorization potential of different synthetic dyes. *Water, Air, & Soil Pollution, 229*(6), 1-13.

- Zanoni, T. B., Lizier, T. M., das Dores Assis, M., Zanoni, M. V. B., & De Oliveira, D. P. (2013). CYP-450 isoenzymes catalyze the generation of hazardous aromatic amines after reaction with the azo dye Sudan III. Food and chemical toxicology, 57, 217-226.
- Zhao, X., & Hardin, I. R. (2007). HPLC and spectrophotometric analysis of biodegradation of azo dyes by Pleurotus ostreatus. *Dyes and Pigments, 73*(3), 322-325.
- Zheng, C., Zhao, L., Zhou, X., Fu, Z., & Li, A. (2013). Treatment technologies for organic wastewater *Water Treatment*: IntechOpen.
- Zheng, Y., & Wang, A. (2010). Removal of heavy metals using polyvinyl alcohol semi-IPN poly (acrylic acid)/tourmaline composite optimized with response surface methodology. *Chemical Engineering Journal, 162*(1), 186-193.
- Zou, Y., Chen, X., Yang, W., & Liu, S. (2011). Response surface methodology for optimization of the ultrasonic extraction of polysaccharides from Codonopsis pilosula Nannf. var. modesta LT Shen. Carbohydrate Polymers, 84(1), 503-508.
- Zouboulis, A. I., Moussas, P. A., & Psaltou, S. G. (2013). Groundwater and soil pollution: bioremediation.

BIODATA OF STUDENT

Khairunnisa' binti Mohd Zin was born in Batu Gajah, Perak in December 1994. In 2015 she completed her Diploma in Science at Universiti Teknologi MARA. In 2018, she finished her first degree of Bachelor of Science (Hons.) Biology from Universiti Teknologi MARA and started her full time Master in research immediately after graduation in 2018 at Universiti Putra Malaysia under supervision of Dr. Mohd Izuan Effendi bin Halmi.



PUBLICATION

Zin, K. M., Effendi Halmi, M. I., Abd Gani, S. S., Zaidan, U. H., Samsuri, A. W., & Abd Shukor, M. Y. (2020). Microbial Decolorization of Triazo Dye, Direct Blue 71: An Optimization Approach Using Response Surface Methodology (RSM) and Artificial Neural Network (ANN). *BioMed research international*, 2020.

