



**UNIVERSITI PUTRA MALAYSIA**

***DEVELOPMENT OF FEED-BASED WHOLE-CELLS INACTIVATED  
BIVALENT VACCINE AND ITS IMMUNOPROTECTIVE ABILITY AGAINST  
STREPTOCOCCOSIS AND MOTILE AEROMONAD SEPTICEMIA IN RED  
HYBRID TILAPIA (*Oreochromis spp.*)***

**MD SHIRAJUM MONIR**

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By

**MD SHIRAJUM MONIR**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,  
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**June 2021**

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

**DEVELOPMENT OF FEED-BASED WHOLE-CELLS INACTIVATED BIVALENT VACCINE AND ITS IMMUNOPROTECTIVE ABILITY AGAINST STREPTOCOCCOSIS AND MOTILE AEROMONAD SEPTICEMIA IN RED HYBRID TILAPIA (*Oreochromis spp.*)**

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**June 2021**

**Chairman : Associate Professor Ina Salwany Md Yasin, PhD**  
**Faculty : Agriculture**

Streptococcosis and motile aeromonad septicemia (MAS) are well-known bacterial diseases in tilapia culture, which cause mass mortality with significant economic losses to aquaculture globally. As therapy resistance is an increasing problem, development of efficient fish vaccines seems to be an alternative to minimize the streptococcosis and MAS diseases. The development of feed-based monovalent vaccines in controlling these diseases has been attempted; however, the mechanism of immunity of feed-based bivalent vaccine against streptococcosis and MAS infections, and the cross-protective ability of these two diseases are still understudied. To explore the immunological role of the feed-based bivalent vaccine, we compared the immune responses of red hybrid tilapia after immunization with both feed-based bivalent and monovalent vaccines, and compared the relative percentage survival (RPS) and cross-immunization protections of red hybrid tilapia following challenged with *Streptococcus iniae*, *Aeromonas hydrophila*, *S. agalactiae* and *A. veronii*. A total of five groups of fish were vaccinated orally through two different techniques; bivalent vaccine (inactivated *S. iniae* and *A. hydrophila*) sprayed on feed pellets (BS group); bivalent vaccine (inactivated *S. iniae* and *A. hydrophila*) incorporated in fish feed (BI group); monovalent inactivated *S. iniae* and *A. hydrophila* vaccine separately incorporated into feed as monovalent *S. iniae* (MS group) and monovalent *A. hydrophila* (MA group); and control group (without vaccine). The feed based vaccines were delivered orally at 5% of body weight for five consecutive days and also the double booster doses were administered in the same manner on weeks 2 and 6. The haematological results revealed that BI vaccinated group exhibited significantly the highest ( $P < 0.05$ ) number of leucocytes ( $45.39 \pm 1.34 \times 10^3/\mu\text{L}$ ) and granulocytes ( $7.68 \pm 0.29 \times 10^3/\mu\text{L}$ ) on weeks 3 post-vaccination. The lysozyme activity demonstrated a significant ( $P < 0.05$ ) increase in BI vaccinated group particularly on 8 (313.77 units/mL) and 12 (303.62 units/mL) weeks post-vaccination. The significantly ( $P < 0.05$ ) highest phagocytic activity was also observed in BI group (53.83%), while the lowest was obtained in BS group (37.33%) on weeks 12 post-vaccination. The

enzyme-linked immunosorbent assay (ELISA) analysis showed that BI group developed a strong and significantly ( $P < 0.05$ ) higher systemic and mucosal IgM responses against both *S. iniae* and *A. hydrophila*, and also cross-protective antigen *S. agalactiae* and *A. veronii* as compared to the BS vaccine and unvaccinated groups. On weeks 10 post-vaccination, all fish were challenged through the intraperitoneally (i.p.) route, where relative percentage survival (RPS) in the BI vaccinated group were observed  $82.22 \pm 3.85\%$  when challenged with *S. iniae*,  $77.78 \pm 3.85\%$  when challenged with *A. hydrophila* and  $77.78 \pm 3.85\%$  when co-challenged with both *S. iniae* and *A. hydrophila*, which were significantly higher ( $P < 0.05$ ) compared to the other groups. Simultaneously, the BI vaccinated group also showed significantly ( $P < 0.05$ ) higher partial cross-protections following challenges with *S. agalactiae* (RPS at  $60.00 \pm 6.67\%$ ) and *A. veronii* (RPS at  $57.78 \pm 7.70\%$ ). Quantitative real-time PCR results also showed that the relative expressions of IL-1 $\beta$ , C-type lysozyme, TNF- $\alpha$ , TGF- $\beta$ , CD4, MHC-I, MHC-II and IgT genes in the BI vaccinated fish spleen, head kidney and hindgut exhibited various significant ( $P < 0.05$ ) rising trends following both the early-phase vaccination and post-infections. Notably, the highest relative expression of IL-1 $\beta$ , MHC-II and IgT genes in BI vaccinated group were observed in the co-infected (*S. iniae* and *A. hydrophila*) fish spleen (9.8 - fold), head kidney (9.6 - fold) and hindgut (24.5 - fold), respectively.

Combining our results demonstrate that the BI vaccine could elicit significant immunological responses and this vaccine is highly effective to control *S. iniae* and *A. hydrophila* virulence in red hybrid tilapia, but have moderate efficacy when challenged with *S. agalactiae* and *A. veronii*. Nevertheless, this newly developed feed-based bivalent incorporated (BI) vaccine can effectively protect tilapia against streptococcosis and MAS infections, and also could offer a promising strategy for mass fish vaccination in aquaculture industry.

Keywords: Feed-based; Bivalent vaccine; IgM responses; Cross-protection; Gene expression; Red hybrid tilapia

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PENGHASILAN VAKSIN SEL UTUK TIDAK AFTIF SECARA BIVALEN  
BERASASKAN MAKANAN SERTA KEUPAYAAN IMUNOPROTEKTIFNYA  
DALAM MENENTANG JANGKITAN STREPTOKOKUSIS DAN  
AEROMONAD SEPTISEMIA MOTIL PADA TILAPIA MERAH HIBRID  
(*Oreochromis spp.*)**

Oleh

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Penyakit bakteria, streptococcus and aeromonad septicemia motile (MAS) cukup sinonim dalam kultur ikan tilapia. Penyakit bakteria ini menyebabkan kematian ikan yang sangat besar dengan kerugian yang signifikan dari sudut ekonomi bagi industri akuakultur di peringkat global. Oleh kerana rintangan terhadap terapi semakin meningkat, penghasilan vaksin ikan yang efisien dilihat sebagai alternatif bagi meminimumkan jangkitan penyakit seperti streptokokus dan MAS. Penghasilan vaksin monovalen berasaskan makanan dalam mengawal penyakit-penyakit ini telah dijalankan; namun, mekanisme imuniti disebalik vaksin bivalen berasaskan makanan terhadap streptokokus dan jangkitan MAS, dan kemampuan perlindungan silang kedua penyakit ini masih kurang difahami. Bagi meneroka peranan imunologi vaksin bivalen berasaskan makanan, kami membandingkan tindak balas imun dari tilapia merah hibrid selepas imunisasi dengan vaksin bivalen dan monovalen berasaskan makanan, dan membandingkan kelangsungan peratusan relatif (RPS) dan perlindungan silang-imunisasi tilapia merah hibrid selepas dicabar dengan bakteria *Streptococcus iniae*, *Aeromonas hydrophila*, *S. agalactiae* dan *A. veronii*. Sejumlah lima kumpulan ikan divaksinasi berasaskan makanan melalui dua teknik yang berbeza; vaksin bivalen (*S. iniae* dan *A. hydrophila* yang tidak aktif) disembur pada pelet makanan (kumpulan BS); vaksin bivalen (*S. iniae* dan *A. hydrophila* yang tidak aktif) yang dimasukkan dalam makanan (kumpulan BI); vaksin monovalen *S. iniae* dan *A. hydrophila* yang tidak aktif secara berasingan dimasukkan ke dalam makanan sebagai monovalen *S. iniae* (kumpulan MS) dan monovalen *A. hydrophila* (kumpulan MA); dan kumpulan kawalan (tanpa vaksin). Vaksin berasaskan makanan tersebut diberikan secara oral pada kadar 5% dari berat badan selama lima hari berturut-turut dan juga dos penganda diberikan dengan cara yang sama pada minggu ke-2 dan 6. Hasil hematologi pula menunjukkan kumpulan vaksin BI memberikan jumlah leukosit tertinggi ( $P < 0.05$ ) ( $45.39 \pm 1.34 \times 10^3/\mu\text{L}$ ) dan granulosit ( $7.68 \pm 0.29 \times 10^3/\mu\text{L}$ ) pada minggu ke-3 selepas vaksinasi. Aktiviti lisozim

menunjukkan peningkatan yang signifikan ( $P < 0.05$ ) dalam kumpulan vaksin BI terutamanya pada minggu ke-8 (313.77 units/mL) dan minggu ke-12 (303.62 units/mL) selepas vaksinasi. Aktiviti fagositik tertinggi ( $P < 0.05$ ) yang tinggi juga diperhatikan pada kumpulan BI (53.83%), sementara yang terendah diperoleh pada kumpulan BS (37.33%) pada minggu ke-12 selepas vaksinasi. Analisis imunoserben berkait enzim (ELISA) menunjukkan bahawa pemberian dos pertama vaksin berasaskan makanan telah merangsang tahap antibodi IgM yang berlanjutan sehingga minggu ke-4, sementara penggalak kedua memastikan tahap IgM tetap tinggi selama 16 minggu pada Kumpulan vaksin BI dan MS. Di samping itu, kumpulan BI telah membentuk tindak balas IgM sistemik (serum) dan mukosa (lendir badan dan cecair lavaj usus) yang kuat dan lebih tinggi ( $P < 0.05$ ) secara signifikan terhadap *S. iniae* dan *A. hydrophila* dan juga memberikan perlindungan silang terhadap antigen *S. agalactiae* dan *A. veronii* berbanding dengan kumpulan BS dan kawalan. Pada minggu ke-10 selepas vaksinasi, semua ikan dicabar melalui intraperitoneum (i.p.), di mana peratusan survival relatif (RPS) dalam kumpulan vaksin BI dilihat pada kadar  $82.22 \pm 3.85\%$  ketika dicabar dengan *S. iniae*,  $77.78 \pm 3.85\%$  ketika dicabar dengan *A. hydrophila* dan  $77.78 \pm 3.85\%$  ketika dicabar bersama dengan *S. iniae* dan *A. hydrophila*, yang jauh lebih tinggi ( $P < 0.05$ ) berbanding kumpulan lain. Pada masa yang sama, kumpulan yang diberi vaksin BI juga menunjukkan perlindungan silang separa yang lebih tinggi ( $P < 0.05$ ) berikutan cabaran dengan *S. agalactiae* (RPS pada  $60.00 \pm 6.67\%$ ) dan *A. veronii* (RPS pada  $57.78 \pm 7.70\%$ ). Hasil PCR kuantitatif masa nyata juga menunjukkan bahawa gen IL-1 $\beta$ , lisozim jenis C, TNF- $\alpha$ , TGF- $\beta$ , CD4, MHC-I, MHC-II dan IgT dalam limpa, ginjal bahagian atas dan usus bahagian belakang ikan yang divaksinasi BI menunjukkan corak peningkatan ekspresi yang pelbagai secara signifikan ( $P < 0.05$ ) selepas vaksinasi fasa awal dan selepas jangkitan. Ekspresi relatif tertinggi gen IL-1 $\beta$ , MHC-II dan IgT dalam kumpulan vaksin BI yang dilihat tertinggi masing-masing pada limpa ikan (9.8 kali ganda), ginjal bahagian atas (9.6 kali ganda) dan usus bahagian belakang (24.5 kali ganda) yang dicabar secara rentas dengan *S. iniae* and *A. hydrophila*.

Gabungan hasil kajian kami menunjukkan bahawa vaksin BI dapat menimbulkan tindak balas imunologi yang signifikan dan vaksin ini sangat berkesan untuk mengawal jangkitan *S. iniae* dan *A. hydrophila* pada ikan tilapia merah hibrid, namun mempunyai keberkesanan sederhana ketika dicabar dengan *S. agalactiae* dan *A. veronii*. Walaupun begitu, vaksin bivalen (BI) berasaskan makanan yang baru dihasilkan ini dapat melindungi ikan tilapia merah hibrid dengan berkesan terhadap streptokokus dan jangkitan MAS, dan juga dapat menawarkan strategi vaksinasi ikan yang berkesan dan efektif dalam industri akuakultur.

Kata kunci: Vaksin berasaskan makanan; Vaksin bivalen; Tindak balas IgM; Perlindungan rentas; Ekspresi gen; Tilapia merah hibrid

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## LIST OF ABBREVIATIONS

s	percentage
μL	microlitre
μM	micromolar
®	registeredp
™	trademark
°C	degree celcius
<i>A. hydrophila</i>	<i>Aeromonas hydrophila</i>
APC	antigen-presenting cell
<i>A. veronii</i>	<i>Aeromonas veronii</i>
BSA	bovine serum albumin
BHIA	brain heart infusion agar
BHIB	brain heart infusion broth
BLAST	basic local alignment search tool
bp	base pair
CaCl <sub>2</sub>	calcium chloride
CFU	colony forming units
CD4	cluster of differentiation 4
dATP	deoxyadenosine triphosphate
dNTP	deoxynucleotide triphosphate
DNA	deoxyribonucleic acid
ECPs	extracellular products
g	gram
GALT	gut-associated lymphoid tissue

H & E	haematoxylin and eosin
hpi	hour post infection
Ig	immunoglobulin
IL-1 $\beta$	Interleukins -1 beta
IP	intraperitoneal
kb	kilobase pair
LD <sub>50</sub>	median lethal dosage
LPS	lipopolysaccharide
M	molar
MAS	motile aeromonad septicemia
MHC	major histo-compatibility complex
mg	milligram
MgCl <sub>2</sub>	magnesium chloride
mM	millimolar
MS-222	tricaine methanesulfonate solution
NaCl	sodium chloride
OMPs	outer membrane proteins
PAMPs	pathogen-associated molecular patterns
PBS	phosphate buffered saline
PCR	polymerase chain reaction
RNA	ribonucleic acid
RPM	revolutions per minute
RPS	relative percentage survival
rRNA	ribosomal ribonucleic acid

<i>S. agalactiae</i>	<i>Streptococcus agalactiae</i>
<i>S. iniae</i>	<i>Streptococcus iniae</i>
sp	species
TGF- $\beta$	transforming growth factor-beta
TNF- $\alpha$	tumor necrosis factor-alpha
TSA	tryptic soy agar
TSB	tryptic soy broth
V	voltage
v/v	volume per volume
wpv	week post vaccination
w/v	weight per volume

# CHAPTER 1

## INTRODUCTION

### 1.1 Background of the study

Tilapia (*Oreochromis* spp.) is one of the most important freshwater fish species that is cultured intensively all over the world. The global production of cultured tilapia represent 6.93 million tons in 2020, around 40 percent of the combined production of the most important commercial whitefish species (FAO, 2021). Tilapia is very popular due to rapid growth, suitability for aquaculture, low production cost, high acceptability in the market (Gu et al., 2017; Dawood et al., 2019), and it is easy to adapt in tropical and sub-tropical regions of the world (Shelton, 2002). Subsequently, tilapia is increasingly demanded in global markets ranging from the poorest segments in developing countries to highly developed western markets (Aminudin, 2017). However, the production of tilapia is increasing day by day from the particular ecosystem, but this sector in globally has been facing several health risks for affecting with emerging and re-emerging bacterial diseases, which causes the severe economic losses in this industry (Adikesavalu et al., 2017; Vásquez-Machado et al., 2019).

Amongst infectious bacterial diseases, the occurrence of streptococcosis and motile aeromonad septicemia (MAS) have been characterized as one of the major obstacle to the sustainable establishment of tilapia sector particularly in Asian countries (Sumiati et al., 2015; Verner-Jeffreys et al., 2017; Pasaribu et al., 2018; Abdelkhalek et al., 2020; Han et al., 2020). Streptococcosis is a septicemic infection mainly by Gram-positive bacteria *Streptococcus agalactiae* and *S. iniae*, and major bacterial pathogens of tilapia that can cause severe symptoms, and result in high morbidities and mortalities rates of up to 60 - 80% (Noraini et al., 2013; Abdelkhalek et al., 2020; Wang et al., 2020a). In Malaysia, the outbreak of streptococcosis is frequently reported in red hybrid tilapia (*Oreochromis* spp.), particularly in dry and hot (April - June) seasons and causes economic losses every year in tilapia industry (Zamri-Saad et al., 2014; Ali et al., 2020; Syuhada et al., 2020). The annual global economic loss due to streptococcosis has been estimated at around USD 10 billion (Xu et al., 2007; Cui et al., 2019). In another bacterial disease of motile aeromonad septicemia (MAS) mainly by Gram-negative bacteria of *Aeromonas hydrophila*, recognized to have a devastating effect on tilapia production (Aboyadak et al., 2015; Pasaribu et al., 2018; Mahrous et al., 2020), and is also responsible for huge economic losses in the aquaculture sector globally (AlYahya et al., 2018; Han et al., 2020). The motile aeromonads of *A. veronii* and *A. sobria* also observed to cause adverse effects in tilapia farming (Dong et al., 2017; Amal et al., 2018; Chirapongsatunkul et al., 2019).

The mortality of cultured fish mostly in intensive farming system is not only caused by single bacteria, but most of the cases due to multiple co-infection with different bacteria (Dong et al., 2015). Tilapia mortalities due to bacterial co-infections has also been observed, such as co-infections of *Streptococcus* sp. and *Aeromonas* sp. (Sugiani et al.,

2012; Sumiati et al., 2015), *S. agalactiae* and *Francisella noatunensis* (Assis et al., 2017), *A. veronii* and *Flavobacterium columnare* (Dong et al., 2015), and *F. noatunensis* and *Shewanella putrefaciens* (Pradeep et al., 2016). In Malaysia, a recent report revealed that mass mortality due to co-infection of tilapia lake virus (TiLV) and *A. veronii* was observed in cultured red hybrid tilapia (Amal et al., 2018).

The most common practice in treating fish bacterial diseases of streptococcosis and MAS involves the application of different antibiotics (Darwish et al., 2002; Zamri-Saad et al., 2014; Assane et al., 2019). Unfortunately, serious complications raised through the spread of antibiotics resistance bacteria and the accumulation of antibiotics residues in the environment and food product, creating serious problem to human and environmental health (Defoirdt et al., 2011; Cabello et al., 2016; Stratev & Odeyemi, 2016a; Watts et al., 2017). Moreover, antibiotics are presently ineffective for controlling streptococcosis and MAS infections (Klesius et al., 2000; Musa et al., 2009; Aisyhah et al., 2014; Mahrous et al., 2020). Owing to the issues, the application of antibiotic is no longer encouraged. Therefore, vaccination is now considered as the best approach and feasible way to prevent bacterial diseases outbreaks.

To date, three widely vaccine delivery systems have been applied in aquaculture with variable success, depending on the nature of each vaccine and production stages of fish; injection, bath immersion and oral (Adams, 2019; Abu-Elala et al. 2019). In general, Southeast Asian countries particularly Indonesia, Malaysia, Thailand and Vietnam, most fish farmers operate on small-scale basis with little technical supports (Aminudin, 2017; Kayansamruaj et al., 2020). Thus, the costs of manpower and technical supports, and facilities required to carry out vaccination using injection and immersion routes are unaffordable (Aminudin, 2017; Ismail et al., 2017). However, the oral route is considered to be the ideal approach as it is less stressful, more practical, applicable to all sized fish, mass immunization of large scale fish, reduces the cost of labour and provide a good technique for boosting several times during culture period in ponds and cages (Firdaus-Nawi et al., 2014; Laith et al., 2019; Jun et al., 2020; Sotomayor-Gerding et al., 2020). Moreover, vaccines administered via oral route have the ability to elicit both mucosal and systemic immunities; thereby protecting the fish at the portal of entry of the pathogen and preventing the spread of infections systemically (Munang'andu et al., 2015; Somamoto & Nakanishi, 2020). As for now, Chile, Scotland and Norway are the only few countries that apply feed-based vaccines for fish (Laith et al., 2019). The limited application of this feed-based vaccine could be due to little information and research documentation on the assessment of immunogenicity and protective efficacy of feed-based vaccination in different cultured fish other than injection and immersion vaccination methods.

Inactivated vaccines prepared in formalin solution have been widely applied in aquaculture in recent decades and reached a satisfactory level of protections against pathogens (Nguyen et al., 2017; Wei et al., 2020). Meanwhile, some previous studies have observed that inactivated vaccine combined with adjuvant could provide more effective protections to prevent bacterial diseases compared to the use of alone vaccine (Nguyen et al., 2017; Xu et al., 2019a). Thus, commercially available different fish vaccines are currently developed from different mineral oil as an adjuvant, whereas these

oils can cause damage to fish, with consequent retardation of growth and negative effects on the welfare of the fish (Mutoloki et al., 2006; Heegaard et al., 2011; Xu et al., 2019a; Li et al., 2020). On the other hand, Freund's complete adjuvant (FCA) and Freund's incomplete adjuvant (FIA) are very effective in fish vaccination with low toxicity levels (Tafalla et al., 2013; Jiang et al., 2015; Wang et al., 2016), however, it extremely expensive, particularly for the commercial fish vaccine development (Aminudin, 2017). Some studies showed palm oil as an adjuvant for newcastle disease virus (NDV) vaccine in chicken and caseous lymphadenitis vaccine in rat model could effectively enhance immune protection without any side effects (Wanasawaeng et al., 2009; Roslindawani et al., 2016). Therefore, palm oil can be a potential adjuvant for fish vaccine that could stimulate good immunities and subsequent protections at a cheaper rate (Roslindawani et al., 2016; Aminudin et al., 2018).

## 1.2 Problem statement

A number of monovalent vaccines have been developed to combat *S. iniae*, *S. agalactiae* (Klesius et al., 2000; Chen et al., 2012; Diab et al., 2019) and *A. hydrophila* (Aly et al., 2015; Sukenda et al., 2017a). Moreover, the effective commercial streptococcosis vaccines such as AQUAVAC® Strep Sa (Merck Animal Health Company, USA) and NORVAX® STREP Si (Merck Animal Health Company, USA), are administered via intraperitoneal (i.p.) and immersion routes, respectively. These developed monovalent vaccines have high protective properties against single target bacteria through injection or immersion immunization, but they are unable to protect multiple bacteria through a single vaccination. However, considering the cost and timing of fish vaccination, developing feed-based bivalent or cross-protective vaccines to prevent these diseases in a single immunization is desirable (Bastardo et al., 2012; Shoemaker et al., 2012; Abu-Elala et al., 2019; Guo et al., 2020).

The commercial adjuvants of FCA, FIA and montanide have been currently used for developing fish vaccines against *S. iniae* (Wang et al., 2016), *S. agalactiae* and (Abu-Elala et al. 2019) and *A. hydrophila* (Heloisa et al., 2020), respectively and showed 70 - 75% relative percent survival (RPS) with long-term protection, but the cost of using these commercial adjuvants in fish vaccines is too expensive. Thus, we require to find a lower ratio of adjuvant to minimize the cost of vaccine production and give comparable protection to the fish. Due to the increasing awareness of animal welfare and environmental issues, a safer and natural ingredient of palm oil was selected as the best candidate to substitute the present expensive commercial adjuvants. However, this research provides a better understanding of the use of feed-based palm oil combined bivalent vaccine that is capable of providing adequate immuno-protective against streptococcosis and MAS infections.

## 1.3 Justification of the research

To date from our knowledge, there is no report that has been claimed to develop feed-based bivalent vaccine, which can confer protection against *S. iniae*, *A. hydrophila* and cross-protection ability of *S. agalactiae* and *A. veronii* in red hybrid tilapia. However,

preliminary investigations performed by Wang et al. (2020b) suggest sufficient antigenic similarity exists between *S. iniae* and *S. agalactiae*, and cross-protection was observed, as the *S. iniae* monovalent vaccine protected fish from *S. agalactiae* infection (Diab et al. 2019; Rahmatullah, 2018). On the other hand, cross-protection of *A. hydrophila* vaccine against *A. veronii* in tilapia has not been reported. Given the increased incidence and prevalence of *S. agalactiae* and *A. veronii* also associated with red hybrid tilapia production in Malaysia, it would be fortuitous if the newly developed feed-based bivalent vaccine based on whole-cells inactivated *S. iniae* and *A. hydrophila* would provide adequate cross-protection against *S. agalactiae* and *A. veronii*.

Moreover, as described by Behera & Swain (2014), Firdaus-Nawi et al. (2014), Dubey et al. (2016) and Wang et al. (2018a), developed feed-based adjuvant added monovalent vaccine or antigen encapsulated oral vaccines those stimulated immune responses with high protection against target bacterial strains in fish. In general however, the manufacture of these non-cross-protective encapsulated or commercial adjuvant added oral vaccine formulations is elaborate and expensive, and could protect only one pathogen with a single vaccination. In order to fill in the research gap, the main purpose of this study is to develop a feed-based bivalent or cross-protective vaccine formulated from two prominent pathogenic bacteria, *S. iniae* and *A. hydrophila*, which can be a novel vaccine strategy for the prevention of both streptococcosis and MAS infections in the aquaculture industry.

#### 1.4 Objectives

The general objective of this study is to develop a feed-based bivalent vaccine combined palm oil as an adjuvant, which can protect against two major bacterial diseases of streptococcosis and motile aeromonad septicemia (MAS) in red hybrid tilapia (*Oreochromis* spp.).

There are four specific objectives of this study, which are stated as below:

- 1) to develop a feed-based palm oil combined bivalent vaccine containing whole-cells inactivated *Streptococcus iniae* and *Aeromonas hydrophila*.
- 2) to assess the systemic and mucosal immune responses by red hybrid tilapia (*Oreochromis* spp.) following oral vaccination using the newly developed feed-based bivalent vaccine.
- 3) to evaluate the effectiveness of the newly developed feed-based bivalent vaccine in protecting red hybrid tilapia (*Oreochromis* spp.) against *S. iniae*, *A. hydrophila*, *S. agalactiae* and *A. veronii* infections.
- 4) to analyze the relative expression of immune-related genes at early phase-immunization and post-infections by quantitative real-time PCR in feed-based bivalent vaccinated red hybrid tilapia (*Oreochromis* spp.).

## 1.5 Research hypotheses

### Hypothesis 1:

**H<sub>0</sub>:** The newly developed feed-based bivalent vaccines against streptococcosis and motile aeromonad septicemia (MAS) are not safe and negative impact on growth performances in red hybrid tilapia (*Oreochromis* spp.).

**H<sub>a</sub>:** The newly developed feed-based bivalent vaccines against streptococcosis and motile aeromonad septicemia (MAS) are safe and no negative impact on growth performances in red hybrid tilapia (*Oreochromis* spp.).

### Hypothesis 2:

**H<sub>0</sub>:** Vaccination of red hybrid tilapia (*Oreochromis* spp.) with newly developed feed-based bivalent vaccine could not induce certain level of systemic and mucosal immune responses against *Streptococcus* sp. and *Aeromonas* sp.

**H<sub>a</sub>:** Vaccination of red hybrid tilapia (*Oreochromis* spp.) with newly developed feed-based bivalent vaccine could induce certain level of systemic and mucosal immune responses against *Streptococcus* sp. and *Aeromonas* sp.

### Hypothesis 3:

**H<sub>0</sub>:** Vaccination with the newly developed feed-based bivalent vaccine could not confer better protection against *S. iniae* and *A. hydrophila* as well as cross-protection against *S. agalactiae* and *A. veronii* in red hybrid tilapia (*Oreochromis* spp.).

**H<sub>a</sub>:** Vaccination with the newly developed feed-based bivalent vaccine could confer better protection against *S. iniae* and *A. hydrophila* as well as cross-protection against *S. agalactiae* and *A. veronii* in red hybrid tilapia (*Oreochromis* spp.).

### Hypothesis 4:

**H<sub>0</sub>:** The newly developed feed-based bivalent vaccine could not induce the expression of immune-related genes at early phase-immunization and post-infections in vaccinated red hybrid tilapia (*Oreochromis* spp.).

**H<sub>a</sub>:** The newly developed feed-based bivalent vaccine could induce the expression of immune-related genes at early phase-immunization and post-infections in vaccinated red hybrid tilapia (*Oreochromis* spp.).



## REFERENCES

- Abdel-Aziz, E. S. H., Abdu, S. B. S., Ali, T. E. S., & Fouad, H. F. (2010). Haemopoiesis in the head kidney of tilapia, *Oreochromis niloticus* (Teleostei: Cichlidae): a morphological (optical and ultrastructural) study. *Fish Physiology & Biochemistry*, 36(3), 323-336.
- Abdelhamed, H., Ibrahim, I., Baumgartner, W., Lawrence, M. L., & Karsi, A. (2017). Characterization of histopathological and ultrastructural changes in channel catfish experimentally infected with virulent *Aeromonas hydrophila*. *Frontiers in Microbiology*, 1-15. DOI: 10.3389/fmicb.2017.01519
- Abdelkhalek, N. K., Risha, E., El-Adl, M. A., Salama, M. F., & Dawood, M. A. O. (2020). Antibacterial and antioxidant activity of clove oil against *Streptococcus iniae* infection in Nile tilapia (*Oreochromis niloticus*) and its effect on hepatic hepcidin expression. *Fish & Shellfish Immunology*, 104, 478-488. <https://doi.org/10.1016/j.fsi.2020.05.064>
- Abdel-Latif, H. M. R., & Khafaga, A. F. (2020). Natural co-infection of cultured Nile tilapia *Oreochromis niloticus* with *Aeromonas hydrophila* and *Gyrodactylus cichlidarum* experiencing high mortality during summer. *Aquaculture Research*, 51(5), 1880–1892. <https://doi.org/10.1111/are.14538>
- Aboyadak, I. M., Ali, N. G. M., Goda, A. M. A. S., Aboelgalagel, W. H., & Salam, A. M. E. (2015). Molecular detection of *Aeromonas hydrophila* as the main cause of outbreak in tilapia farms in Egypt. *Journal of Aquaculture & Marine Biology*, 2, 2-5. DOI: 10.15406/jamb.2015.02.00045
- Abu-Elala, N. M., Samir, A., Wasfy, M., & Elsayed, M. (2019). Efficacy of injectable and immersion polyvalent vaccine against streptococcal infections in broodstock and offspring of Nile tilapia (*Oreochromis niloticus*). *Fish & Shellfish Immunology*, 88, 293-300. DOI: 10.1016/j.fsi.2019.02.042
- Adams, A. (2019). Progress, challenges and opportunities in fish vaccine development. *Fish & Shellfish Immunology*, 90, 210–214. <https://doi.org/10.1016/j.fsi.2019.04.066>
- Adikesavalu, H., Banerjee, S., Patra, A., & Abraham, T. J. (2017). Meningoencephalitis in farmed monosex Nile tilapia (*Oreochromis niloticus* L.) caused by *Streptococcus agalactiae*. *Archives of Polish Fisheries*, 25(3), 187-200. DOI: 10.1515/aopf-2017-0018
- Agnew, W., & Barnes, A. C. (2007). *Streptococcus iniae*: an aquatic pathogen of global veterinary significance and a challenging candidate for reliable vaccination. *Veterinary Microbiology*, 122, 1-15. DOI: 10.1016/j.vetmic.2007.03.002
- Ahmadivand, S., Soltani, M., Behdani, M., & Evensen, Ø. (2017). Oral DNA vaccines based on CS-TPP nanoparticles and alginate microparticles confer high protection against infectious pancreatic necrosis virus (IPNV) infection in trout. *Developmental & Comparative Immunology*, 74, 178–189. <https://doi.org/>

- Ahmed, O. M. (2019). Bacteria Associated with Fresh-water Aquaculture Tilapia Fish (*Oreochromis niloticus*) in Suez, Egypt. *Food Science & Nutrition*, 2(1), 1-7.
- Aisyhah, M A S, Amal, M N A Zamri-Saad, M, A Siti-Zahrah, A, & Shaqinah, N. N. (2014). *Streptococcus agalactiae* isolates from cultured fishes in Malaysia manifesting low resistance pattern towards selected antibiotics. *Journal of Fish Diseases*. <https://doi.org/10.1111/jfd.12351>
- Alcamo, I. E. (1997). In: Fundamentals of Microbiology. 5th edn., Addison Wesley Longman, Inc., California. pp. 766.
- Alcorn, S., Ostland, V., LaPatra, S., Harbell, S., Friedman, C. & Winton, J. (2008). Measurement of the innate cellular immune responses of hybrid striped bass and rainbow trout. WRAC Publications.
- Alejo, A., & Tafalla, C. (2011). Chemokines in teleost fish species. *Developmental & Comparative Immunology*, 35(12), 1215-1222. DOI: 10.1016/j.dci.2011.03.011
- Ali, S. H. M., Ridzuan, M. S. M., Abdullah, S. Z., Mansor, N. N., Nawi, M. F., Zamri-Saad, M., & Amal, M. N. A. (2020). Retrospective identification of bacterial depository revealed that *Streptococcus iniae* was responsible for some of the streptococcosis cases in cultured red tilapia in Malaysia since 2006. *Pertanika Journal of Tropical Agricultural Science*, 43(2), 231-238.
- Alsaid, M., Daud, H. H. M., Mustapha, N. M., Bejo, S. K., Abdelhadi, Y. M., Abuseliane, A. F., & Hamdan, R. H. (2013). Pathological findings of experimental *Streptococcus agalactiae* infection in red hybrid tilapia (*Oreochromis* sp.). International Conference on Chemical, Agricultural and Medical Sciences (CAMS-2013), Kuala Lumpur, Malaysia: 70-73.
- Aly, S. M., Albutti, A. S., Rahmani, A. H., & Abdel Atti, N. M. (2015). The response of new-season Nile tilapia to *Aeromonas hydrophila* vaccine. *International Journal of Clinical and Experimental Medicine*, 8(3), 4508-4514.
- Aly, S. M., Zohairy, A., Rahmani, M. A., Fathi, A. H. M., & Atti, N. M. A. (2016). Trials to improve the response of *Oreochromis niloticus* to *Aeromonas hydrophila* vaccine using immunostimulants (garlic, Echinacea) and probiotics (Organic Green and Vet-Yeast). *African Journal of Biotechnology*, 15, 989-994.
- Aly, Salah M., Albutti, A. S., Rahmani, A. H., & Abdel Atti, N. M. (2015). The response of new-season Nile tilapia to *Aeromonas hydrophila* vaccine. *International Journal of Clinical & Experimental Medicine*, 8(3), 4508-4514.
- Aly, Salah Mesalhy, Zohairy, M. A. Al, Rahmani, A. H., Fathi, M., & Nashwa M Abdel Atti. (2016). Trials to improve the response of *Oreochromis niloticus* to *Aeromonas hydrophila* vaccine using immunostimulants (garlic, Echinacea) and probiotics (Organic Green™ and Vet-Yeast™). *African Journal of Biotechnology*, 15(21), 989-994. <https://doi.org/10.5897/ajb2015.15155>

- Aly, T. M. PhD Thesis. Studies on the effect of different adjuvant on the efficiency of FMD vaccine in farm animal. Zagazig University, Egypt, 1981.
- AlYahya, S. A., Ameen, F., Al-Niaeem, K. S., Al-Sa'adi, B. A., Hadi, S., & Mostafa, A. A. (2018). Histopathological studies of experimental *Aeromonas hydrophila* infection in blue tilapia, *Oreochromis aureus*. *Saudi Journal of Biological Sciences*, 25(1), 182-185. DOI: 10.1016/j.sjbs.2017.10.019
- Amal, M. N. A., & Zamri-Saad, M. (2011). Streptococcosis in Tilapia (*Oreochromis niloticus*): A review. *Pertanika Journal of Tropical Agricultural Science*, 34(2), 195-206.
- Amal, M. N. A., Koh, C. B., Nurliyana, M., Suhaiba, M., Nor-Amalina, Z., Santha, S., Diyana-Nadhirah K., P., Yusof, M. T., Ina-Salwany, M., Y., & M. Zamri-Saad, M. (2018). A case of natural co-infection of Tilapia Lake Virus and *Aeromonas veronii* in a Malaysian red hybrid tilapia (*Oreochromis niloticus* × *O. mossambicus*) farm experiencing high mortality. *Aquaculture*, 485, 12-16. DOI: 10.1016/j.aquaculture.2017.11.019
- Amal, M. N. A., Zamri-Saad, M., Siti-Zahrah, A., Zulkafli, A.R., Nur-Nazifah, M. (2013). Molecular characterization of *Streptococcus agalactiae* strains isolated from fishes in Malaysia. *Journal of Applied Microbiology*, 115(1), 20-29.
- Amal, M. N.A., Koh, C. B., Nurliyana, M., Suhaiba, M., Nor-Amalina, Z., Santha, S., & Zamri-Saad, M. (2018). A case of natural co-infection of Tilapia Lake Virus and *Aeromonas veronii* in a Malaysian red hybrid tilapia (*Oreochromis niloticus* × *O. mossambicus*) farm experiencing high mortality. *Aquaculture*, 485, 12-16. <https://doi.org/10.1016/j.aquaculture.2017.11.019>
- Amend, D. F. (1981). Potency testing of fish vaccines. *Developments in Biological Standardization*, 49, 447-454.
- Aminudin, S. A, Kamal, F. M., Zamri-Saad, M., Abdullah, S., Ridzuan, M. S., Yusoff, H. M, Hashim, S., Sudirwan, F., Salihin I. M., & Sulaiman, S. A. (2018). Effect of incorporating different concentrations of palm oil as adjuvant in fish vaccine. *International Journal of Biosciences*, 12, 35-41. DOI: 10.12692/ijb/12.1.35-41
- Aminudin, S. A. PhD Thesis. Development of optimal ratio of Freund's Incomplete and Palm Oil Adjuvants Incorporated into Feed-based Oral Vaccine for Fish. Universiti Putra Malaysia, Malaysia, 2017.
- Anderson, D. P. (1977). Fish Immunology. (S. F. Snieszko & H. R. Axelrod, Eds), Hong Kong: TFH Publications, Inc. Ltd.
- Anderson, D. P., Capstiek, P. B. & Mowat, G. N. (1970). In vitro method for safety of FMD. *Journal of Hygiene*, 68, 159-172.
- Andrian, U. H., & Mempel, T. R. (2003). Homing and cellular traffic in lymph nodes, *Nature Reviews Immunology*, 3, 867-878.

- Anshary, H., Kurniawan, R. A., Sriwulan, S., Ramli, R., & Baxa, D. V. (2014). Isolation and molecular identification of the etiological agents of streptococcosis in Nile tilapia (*Oreochromis niloticus*) cultured in net cages in Lake Sentani, Papua, Indonesia. *SpringerPlus*, 3(1), 1–11. <https://doi.org/10.1186/2193-1801-3-627>
- Anuradha, K., Foo, H. L., Mariana, N. S., Loh, T. C., Yusoff, K., Hassan, M. D. Sasan, H., & Raha, A. R. (2010). Live recombinant *Lactococcus lactis* vaccine expressing aerolysin genes D1 and D4 for protection against *Aeromonas hydrophila* in tilapia (*Oreochromis niloticus*). *Journal of Applied Microbiology*, 109(5), 1632-1642. DOI: 10.1111/j.1365-2672.2010.04789.x
- AOAC. (2002). Association of official Analytical Chemist. Official Method of Analysis (17<sup>th</sup> Edition), Maryland USA
- Argüello-Guevara, W. & C. Molina-Poveda. (2013). Effect of binder type and concentration on prepared feed stability, feed ingestion and digestibility of *Litopenaeus vannamei* broodstock diets. *Aquaculture Nutrition*, 19(4), 515-22.
- Arijo, S., Rico, R., Chabrillon, M., Diaz-Rosales, P., Martínez-Manzanares, E., Balebona, M. C., Magariños, B., Toranzo, A. E., & Moriñigo, M. A. (2005). Effectiveness of a divalent vaccine for sole, *Solea senegalensis* (Kaup), against *Vibrio harveyi* and *Photobacterium damsela* subsp. piscicida. *Journal of Fish Diseases*, 28(1), 33-38. DOI: 10.1111/j.1365-2761.2004.00597.x
- Assane, I. M., Gozi, KS, Valladão, G. M. R., & Pilarski, F., (2019). Combination of antimicrobials as an approach to reduce their application in aquaculture: Emphasis on the use of thiamphenico or florfenicol against *Aeromonas hydrophila*. *Aquaculture*, 507, 238-245. DOI: 10.1016/j.aquaculture.2019.04.021
- Assis, G. B. N., Tavares, G. C., Pereira, F. L., Figueiredo, H. C. P., & Leal, C. A. G. (2017). Natural coinfection by *Streptococcus agalactiae* and *Francisella noatunensis* subsp. orientalis in farmed Nile tilapia (*Oreochromis niloticus* L.). *Journal of Fish Diseases*, 40(1), 51-63. DOI: 10.1111/jfd.12493
- Attaya, A., Secombes, C. J., & Wang, T. (2020). Effective isolation of GALT cells: Insights into the intestine immune response of rainbow trout (*Oncorhynchus mykiss*) to different bacterin vaccine preparations. *Fish & Shellfish Immunology*. <https://doi.org/10.1016/j.fsi.2020.06.051>
- Austin, B., & Austin, D. (2007) Bacterial fish pathogens disease of farmed and wild fish (7th revised ed). Chichester: Springer-Praxis Publishing
- Austin, B., & Zhang, X. H. (2006). *Vibrio harveyi*: a significant pathogen of marine vertebrates and invertebrates. *Letters in applied microbiology*, 43(2), 119-124. DOI: 10.1111/j.1472-765X.2006.01989.x
- Ayoub, H. F. (2018). The use of coated and uncoated oral inactivated *Aeromonas hydrophila* vaccine on the immune response of Nile Tilapia (*Oreochromis niloticus*). *Egyptian Journal for Aquaculture*, 8(2), 57–69.

- Azad, I. S., Shankar, K. M., Mohan, C.V., & Kilata, B. (2000). Protective response of common carp vaccinated with biofilm and free cells of *Aeromonas hydrophila*. *Journal of Aquaculture in the Tropics*, 15(1), 65-70.
- Azad, I., Shankar, K., Mohan, C. V., & Kalita, B. (1999). Biofilm vaccine of *Aeromonas hydrophila* - standardization of dose and duration for oral vaccination of carps. *Fish & Shellfish Immunology*, 9, 519-528. DOI: 10.1006/fsim.1998.0206
- Bactol, I. D. C., Padilla, L. V., & Hilario, A. L. (2018). Immune response of tilapia (*Oreochromis niloticus*) after vaccination with autoclavekilled, heat-killed, and formalin-killed whole cell *Aeromonas hydrophila* vaccines as possible serotype-independent vaccines. *International Journal of Agriculture and Biology*, 20(4), 846–850. <https://doi.org/10.17957/IJAB/15.0575>
- Baiano, J. C. F., & Barnes, A. C. (2009). Towards control of *Streptococcus iniae*. *Emerging Infectious Diseases*, 15(12), 1891–1896. <https://doi.org/10.3201/eid1512.090232>
- Bailone, R. L., Martins, M. L., Mourino, J. L. P., Vieira, F. N., Pedrotti, F. S., Nunes, G. C., & Silva, B. C. (2010). Hematology and agglutination titer after polyvalent immunization and subsequent challenge with *Aeromonas hydrophila* in Nile tilapia (*Oreochromis niloticus*). *Archivos de Medicina Veterinaria*, 42 (3), 221-227.
- Bao, P., Sun, X., Liu, Q., Zhang, Y., & Liu, X. (2019). Synergistic effect of a combined live *Vibrio anguillarum* and *Edwardsiella piscicida* vaccine in turbot. *Fish & Shellfish Immunology*, 88, 84–90. <https://doi.org/10.1016/j.fsi.2019.02.014>
- Barnes, A. C., Young, F. M., Horne, M. T., & Allis, A. E. (2003). *Streptococcus iniae*: Serological differences, presence of capsule and resistance to immune serum killing. *Diseases of Aquatic Organisms*, 53, 241- 247.
- Bartie, K. L., Taslima, K., Bekaert, M., Wehner, S., Syaifudin, M., Taggart, J. B., & Penman, D. J. (2020). Species composition in the Molobicus hybrid tilapia strain. *Aquaculture*, 526, 735433. <https://doi.org/10.1016/j.aquaculture.2020.735433>
- Bastardo, A., Ravelo, C., Castro, N., Calheiros, J., & Romalde, J. L. (2012). Effectiveness of bivalent vaccines against *Aeromonas hydrophila* and *Lactococcus garvieae* infections in rainbow trout *Oncorhynchus mykiss* (Walbaum). *Fish & Shellfish Immunology*, 32(5), 756-761. DOI: 10.1016/j.fsi.2012.01.028
- Baums, C. G., Hermeyer, K., Leimbach, S., Adamek, M., Czerny, C. P., Horstgen-Schwark, G., Valentin-Weigand, P., Baumgärtner, W., & Steinhagen, D. (2013). Establishment of a model of *Streptococcus iniae* meningoencephalitis in Nile tilapia (*Oreochromis niloticus*). *Journal of Comparative Pathology*, 149:94-102.
- Bear R. & Rintoul D. (2013) Adaptive Immunity, Connexions. [online] Available at: <http://cnx.org/content/m47434/1.1/> [Accessed 12 November 2020].
- Beckman, M. J., Saraiva, M., McLaggan, D., Pottinger, T. G., & West, P. (2020). Saprolegnia infection after vaccination in Atlantic salmon is associated with differential expression of stress and immune genes in the host. *Fish & Shellfish*

- Behera, T., & Swain, P., (2014). Antigen encapsulated alginate-coated chitosan microspheres stimulate both innate and adaptive immune responses in fish through oral immunization. *Aquaculture International*, 22, 673-688. DOI: 10.1007/s10499-013-9696-8.
- Belmonte, R., Wang, T., Duncan, G. J., Skaar, I., Mérida, H., Bulone, V., & Secombes, C. J. (2014). Role of Pathogen-Derived Cell Wall Carbohydrates and Prostaglandin E 2 in Immune Response and Suppression of Fish Immunity by the Oomycete *Saprolegnia parasitica*. *Infection and Immunity*. 83, 454. doi:10.1128/IAI.02821-14
- Bene, C., Arthur, R., Norbury, H., Allison, E. A., Beveridge, M. C. M., Bush, S., Campling, L., Leschen, W., Little, D. C., Squires, D., Thilsted, S. H., Troell, M., & Williams, M. (2016). Contribution of fisheries and aquaculture to food security and poverty reduction: assessing the current evidence. *World Development*, 77, 179-196.
- Benli, A. C. K. & Yildiz, H. Y. (2004). Blood parameters in Nile tilapia (*Oreochromis niloticus* L.) spontaneously infected with *Edwardsiella tarda*. *Aquaculture Research*, 35, 1388-1390.
- Biller-Takahashi, J. D., & Urbinati, E. C. (2014). Fish immunology. The modification and manipulation of the innate immune System: Brazilian studies. *Anais Da Academia Brasileira de Ciencias*, 86(3), 1483–1495. <https://doi.org/10.1590/0001-3765201420130159>
- Boerlage, A. S., Dung, T. T., Hoa, T. T. T., Davidson, J., Stryhn, H. & Hammell, K. L. (2017). Production of red tilapia (*Oreochromis* spp.) in floating cages in the Mekong Delta, Vietnam: mortality and health management. *Diseases of Aquatic Organisms*, 124, 131-144.
- Burakova, Y., Madera, R., McVey, S., Schlup, J. R., & Shi, J. (2018). Adjuvants for Animal Vaccines. *Viral Immunology*, 31(1), 11–22. <https://doi.org/10.1089/vim.2017.0049>
- Bruijn, I. De, Belmonte, R., Anderson, V. L., Saraiva, M., Wang, T., West, P. Van, & Secombes, C. J. (2012). Immune gene expression in trout cell lines infected with the fish pathogenic oomycete *Saprolegnia parasitica*. *Developmental and Comparative Immunology*, 38(1), 44–54. <https://doi.org/10.1016/j.dci.2012.03.018>
- Brun, A. (2016). Vaccines and vaccination for veterinary viral diseases: A general overview. *Methods in Molecular Biology*, 1349, 1-24.
- Cabello, F. C., Godfrey, H. P., Buschmann, A. H., & Dölz, H. J. (2016). Aquaculture as yet another environmental gateway to the development and globalisation of antimicrobial resistance. *Lancet Infectious Diseases*, 16, 127-133. DOI: 10.1016/S1473-3099(16)00100-6

- Cabillon, N. A. R., & Lazado, C. C. (2019). Mucosal barrier functions of fish under changing environmental conditions. *Fishes*, 4(1), 1–10. <https://doi.org/10.3390/fishes4010002>
- Cai, W. Q., Li, S. F. & Ma, J. Y. (2004). Diseases resistance of Nile tilapia (*Oreochromis niloticus*), blue tilapia (*Oreochromis aureus*) and their hybrid (female Nile tilapia × male blue tilapia) to *Aeromonas sobria*. *Aquaculture*, 229, 79-87.
- Cai, X., Wang, B., Peng, Y., Li, Y., Lu, Y., Huang, Y., & Wu, Z. (2017). Construction of a *Streptococcus agalactiae* phoB mutant and evaluation of its potential as an attenuated modified live vaccine in golden pompano, *Trachinotus ovatus*. *Fish & Shellfish Immunology*, 63, 405–416. <https://doi.org/10.1016/j.fsi.2016.11.050>
- Cain, K. D., LaPatra, S. E., Baldwin, T. J., Shewmaker, B., Jones, J., & Ristow, S. S. (1996). Characterization of mucosal immunity in rainbow trout *Oncorhynchus mykiss* challenged with infectious hematopoietic necrosis virus: Identification of antiviral activity. *Disease of Aquatic Organisms*, 27(3), 161-172. DOI: 10.3354/dao027161
- Camus, A. C, Durborow, R. M, Hemstreet, W. G, Thune, R. L, & Hawke, J. P (1998). *Aeromonas* bacterial infections: motile aeromonad septicemia (No. 478). Southern Regional Aquaculture Center.
- Chalmers, L., Thompson, K. D., Taylor, J. F., Black, S., Migaud, H., North, B., & Adams, A. (2016). A comparison of the response of diploid and triploid Atlantic salmon (*Salmo salar*) siblings to a commercial furunculosis vaccine and subsequent experimental infection with *Aeromonas salmonicida*. *Fish & shellfish immunology*, 57, 301-308
- Chen, C., Chao, C., & Bowser, P. R. (2007). Comparative histopathology of *Streptococcus iniae* and *Streptococcus agalactiae* - infected tilapia. *Bulletin-European Association of Fish Pathologists*, 27, 2.
- Chen, L., Klaric, G., Wadsworth, S., Jayasinghe, S., Kuo, T. Y., Evensen, Ø., & Mutoloki, S. (2014). Augmentation of the antibody response of Atlantic salmon by oral administration of alginate-encapsulated IPNV antigens. *PLoS ONE*, 9(10). DOI: 10.1371/journal.pone.0109337
- Chen, M., Wang, R., Li, L. P., Liang, W. W., Li, J., Huang, Y., Lei, A., Y., Huang, W. Y., & Gan, X. (2012). Screening vaccine candidate strains against *Streptococcus agalactiae* of tilapia based on PFGE genotype. *Vaccine*, 30, 6088-6092
- Chen, P. L. Lamy, B. & Ko, W. C. (2016). *Aeromonas dhakensis*, an increasingly recognized human pathogen. *Front Microbiol*, 7, 793. DOI: 10.3389/fmicb.2016.00793
- Chen, Y., Cai, S., & Jian, J. (2019). Protection against *Vibrio alginolyticus* in pearl gentian grouper (♀*Epinephelus fuscoguttatus* × ♂*Epinephelus lanceolatu*) immunized with an acfA-deletion live attenuated vaccine. *Fish & Shellfish Immunology*, 86, 875–881. <https://doi.org/10.1016/j.fsi.2018.12.030>

- Chettri, J. K., Deshmukh, S., Holten-Andersen, L., Jafaar, R. M., Dalsgaard, I., & Buchmann, K., 2013. Comparative evaluation of administration methods for a vaccine protecting rainbow trout against *Yersinia ruckeri* O1 biotype 2 infections. *Veterinary Immunology and Immunopathology*, 154, 42-47.
- Chettri, J. K., Jaafar, R. M., Skov, J., Kania, P. W., Dalsgaard, I., & Buchmann, K. (2015). Booster immersion vaccination using diluted *Yersinia ruckeri* bacterin confers protection against ERM in rainbow trout. *Aquaculture*, 440, 1-5. DOI: 10.1016/j.aquaculture.2015.01.027
- Chiew, I. K. M., Salter, A. M., & Lim, Y. S. (2019). The significance of major viral and bacterial diseases in Malaysian aquaculture industry. *Pertanika Journal of Tropical Agricultural Science*, 42(3), 1023–1047.
- Chilmonczyk, S. (1992). The thymus in fish: Development and possible function in the immune response. *Annual Review of Fish Diseases*, 2, 181-200.
- Chin, A., & Woo, P. T. (2005). Innate cell-mediated immune response and peripheral leukocyte populations in Atlantic salmon, *Salmo salar* L., to a live *Cryptobia salmositica* vaccine. *Parasitology Research*, 95, 299-304.
- Chin, Y. K., Al-saari, N., Zulperi, Z., Mohd-Aris, A., Salleh, A., Silvaraj, S., Silvaraj S., Mohamad, A., Lee, J. Y., Zamri-Saad, M., & Ina-Salwany, M. Y. (2019). Efficacy of bath vaccination with a live attenuated *Vibrio harveyi* against vibriosis in Asian seabass fingerling, *Lates calcarifer*. *Aquaculture Research*, 1-11. DOI: 10.1111/are.14386
- Chirapongsatonkul, N., Mueangkan, N., Wattitum, S., & U-taynapun, K. (2019). Comparative evaluation of the immune responses and disease resistance of Nile tilapia (*Oreochromis niloticus*) induced by yeast  $\beta$ -glucan and crude glucan derived from mycelium in the spent mushroom substrate of Schizophyllum commune. *Aquaculture Reports*, 15, 100205. DOI: 10.1016/j.aqrep.2019.100205
- Chun, D., Gen, H., Kangsen, M., Huihui, Z., & Wei, X. (2016.) Palatability of water-soluble extracts of protein sources and replacement of fishmeal by a selected mixture of protein sources for juvenile turbot (*Scophthalmus maximus*). *Journal of Ocean University of China*, 15(3), 561-567. DOI: 10.1007/s11802-016-2898-8
- Clark, T.G., & Dickerson, H.W. (1997). Antibody-mediated effects on parasite behaviour: evidence of a novel mechanism of immunity against a parasitic protist. *Parasitology Today*, 13, 477-480.
- Correia, D., Escarcega-Miranda, B., Barreto-Curiel, F., Mata-Sotres, J., del Rio-Zaragoza, O., Viana, M. T., & Rombenso, A. N. (2019). Growth performance and body composition of hybrid red tilapia (*Oreochromis mossambicus*  $\times$  *O. aureus*) fed with different protein levels raised in saltwater. *Latin American Journal of Aquatic Research*, 47(5), 853-859. <https://doi.org/10.3856/vol47-issue5-fulltext-15>
- Cui, M., Zhang, H., Li, J., Liu, R., Wu, M., Xu, D., & Zhang, Q. (2019). Differential PCR detection of *Streptococcus agalactiae* and *Streptococcus iniae* in tilapia



- (*Oreochromis niloticus*) by a single primer pair. *Journal of Fish Diseases*, 42(8), 1211–1216. <https://doi.org/10.1111/jfd.13014>
- Crumlish, M., Thanh, P. C., Koesling, J., Tung, V. T., & Gravningen, K. (2010). Experimental challenge studies in Vietnamese catfish, *Pangasianodon hypophthalmus* (Sauvage), exposed to *Edwardsiella ictaluri* and *Aeromonas hydrophila*. *Journal of Fish Diseases*, 33(9), 717-722. DOI: 10.1111/j.1365-2761.2010.01173.x
- Darwish, A. M., Rawles, S. D., & Griffin, B. R. (2002). Laboratory efficacy of oxytetracycline for the control of *Streptococcus iniae* infection in blue tilapia. *Journal of Aquatic Animal Health*, 14(3), 184-190. DOI: 10.1577/1548-8667(2002)014<0184:LEOFT>2.0.CO;2
- Dash, P., Sahoo, P. K., Gupta, P. K., Garg, L. C., & Dixit, A. (2014). Immune responses and protective efficacy of recombinant outer membrane protein R (rOmpR)-based vaccine of *Aeromonas hydrophila* with a modified adjuvant formulation in rohu (*Labeo rohita*). *Fish & Shellfish Immunology*, 39(2), 512–523. <https://doi.org/10.1016/j.fsi.2014.06.007>
- Dash, P., Yadav, S. K., Garg, L. C., Dixit, A., & Sahoo, P. K. (2017). Post-challenge immune gene expression profiling in rohu, *Labeo rohita* vaccinated with modified adjuvant-based *Aeromonas hydrophila* outer membrane protein R formulation. *Veterinarski Arhiv*, 87 (5), 607-622.
- Davina, J. H. M., Parmentier, H. K., & Timmermans, L. P. M. (1982). Effect of oral administration of *Vibrio* bacteria on the intestine of cyprinid fish. *Developmental & Comparative Immunology Supplement*, 2, 157-166.
- Dawood, M. A. O., Eweedah, N. M., Moustafa, E. M., & Shahin, M. G. (2019). Synbiotic effects of *Aspergillus oryzae* and  $\beta$ -Glucan on growth and oxidative and immune responses of Nile Tilapia, *Oreochromis niloticus*. probiotics and antimicrobial proteins. *Probiotics and Antimicrobial Proteins*, 12(1), 172-183. DOI: 10.1007/s12602-018-9513-9
- Day, M. J., Horzinek, M. C., Schultz, R. D., & Squires, R. A. (2016). WSAVA Guidelines for the vaccination of dogs and cats. *Journal of Small Animal Practice*, 57(1), 1-45.
- Day, M. J., Karkare, U., Schultz, R. D., Squires, R., & Tsujimoto, H. (2015). Recommendations on vaccination for Asian small animal practitioners: a report of the WSAVA Vaccination Guidelines Group. *Journal of Small Animal Practice*, 56(2), 77-95.
- Defoirdt, T., Sorgeloos, P., & Bossier, P. (2011). Alternatives to antibiotics for the control of bacterial disease in aquaculture. *Current Opinion in Microbiology*, 14, 251-258. DOI: 10.1016/j.mib.2011.03.004
- Delannoy, C. M. J., Crumlish, M., Fontaine, M. C., Pollock, J., Foster, G., Dagleish, M. P., Turnbull, J. F., & Zadoks, R. N. (2013). Human *Streptococcus agalactiae* strains in aquatic mammals and fish. *BMC microbiology*, 13, 41.

- Department of Fisheries Malaysia (2020). Annual Fisheries Statistics, 2020. Assessed on 14<sup>th</sup> January 2021. Retrieved from:[Online Resource] dwi- language version. [https://www.dof.gov.my/dof2/resources/user\\_29/Documents/Buku%20Perangkaan%20Tahunan%20Perikanan/Perangkaan\\_Perikanan\\_2019\\_Jilid\\_1.pdf](https://www.dof.gov.my/dof2/resources/user_29/Documents/Buku%20Perangkaan%20Tahunan%20Perikanan/Perangkaan_Perikanan_2019_Jilid_1.pdf)
- Department of Statistics Malaysia (2017). Report of Household Income and Basic Amenities Survey 2016. Assessed on 14 April 2020. Retrieved from:[Online Resource].[https://www.dosm.gov.my/v1/index.php?r=column/cthemByCat&cat=120&bul\\_id=RUZ5REwveU1ra1hGL21JWV1PRmU2Zz09&menu\\_id=amVoWU54UT10a21NWmdhMjFMMWcyZz09](https://www.dosm.gov.my/v1/index.php?r=column/cthemByCat&cat=120&bul_id=RUZ5REwveU1ra1hGL21JWV1PRmU2Zz09&menu_id=amVoWU54UT10a21NWmdhMjFMMWcyZz09).
- Deshmukh, S., Kania, P. W., Chetri, J. K., Skov, J., Bojesen, A. M., & Dalsgaard, I. (2013). Insight from molecular, pathological, and immunohistochemical studies on cellular and humoral mechanisms responsible for vaccine-induced protection of rainbow trout against *Yersinia ruckeri*. *Clinical & Vaccine Immunology*, 20(10), 1623–1641. <https://doi.org/10.1128/CVI.00404-13>
- Devadason, C. (2018). Lysozyme level during acute infection of bacterium *Aeromonas salmonicida* SUBSP *Salmonicida* in halibut and atlantic salmon. *Journal of Experimental Biology and Agricultural Sciences*, 6(1), 236–242. [https://doi.org/10.18006/2018.6\(1\).236.242](https://doi.org/10.18006/2018.6(1).236.242)
- Dhar, A. K., & Allnut, F. C. T. (2011). Challenges and Opportunities in Developing Oral Vaccines against Viral Diseases of Fish. *Journal of Marine Science Research & Development*, 3, 1-6. DIO: 10.4172/2155-9910.S1-003
- Diab, A. M., Khalil, R. H., & Khallaf, M. (2019). Autogenous bacterins cross-protection as a trial for Streptococcosis control in *Oreochromis niloticus*. *Aquaculture International*, 27(6), 1787-1800. DIO: 10.1007/s10499-019-00432-z
- Dias, M. K. R., Sampaio, L. S., Proietti, A. A., Yoshioka, E. T. O., Rodrigues, D. P., Rodriguez, A. F. R., Fernando S. E. D. V., & Tavares-Dias, M. (2016). Lethal dose and clinical signs of *Aeromonas hydrophila* in *Arapaima gigas* (Arapaimidae), the giant fish from Amazon. *Veterinary Microbiology*, 188, 12-15. DOI: 10.1016/j.vetmic.2016.04.001
- Dinarello, C. A. (1997). Interleukin-1. *Cytokine & Growth Factor Reviews*, 8(4), 253-65.
- Dissel, V. J. T, van Langevelde. P., Westendorp, R. G. J., Kwappenberg, K., & Frölich, M. (1998). Anti-inflammatory cytokine profile and mortality in febrile patients. *Lancet*, 351, 950-953.
- Dockree, T., Holland, C. J., Clement, M., Ladell, K., McLaren, J. E., Van Den Berg, H. A., Wooldridge, L. (2017). CD8 + T-cell specificity is compromised at a defined MHCI/CD8 affinity threshold. *Immunology & Cell Biology*, 95(1), 68–76. <https://doi.org/10.1038/icb.2016.85>
- Dong, H. T., Nguyen, V. V., Le, H. D., Sangsuriya, P., Jitrakorn, S., Saksmerprom, V., Senapin, S., & Rodkhum, C. (2015). Naturally concurrent infections of bacterial and viral pathogens in disease outbreaks in cultured Nile tilapia (*Oreochromis*

- niloticus*) farms. *Aquaculture*, 448, 427-435. DOI: 10.1016/j.aquaculture.2015.06.027
- Dong, H. T., Techatanakitarnan, C., Jindakittikul, P., Thaiprayoon, A., Taengphu, S., Charoensapsri, W., Khunrae, P., Rattanarojpong, T., & Senapin, S. (2017). *Aeromonas jandaei* and *Aeromonas veronii* caused disease and mortality in Nile tilapia, *Oreochromis niloticus* (L.). *Journal of Fish Diseases*, 40(10), 1395-1403. DOI: 10.1111/jfd.12617
- Dryden, G. McL. (2008). *Animal Nutrition Science*. Cambridge University Press, UK.
- Du, L., & Liu, W. (2012). Occurrence, fate, and ecotoxicity of antibiotics in agroecosystems. A review. *Agronomy for Sustainable Development*, 32, 309-327. DOI: 10.1007/s13593-011-0062-9
- Dubey, S., Avadhani, K., Mutalik, S., Sivadasan, S. M., Maiti, B., Girisha, S. K., & Munang'andu, H. M. (2016). *Edwardsiella tarda* OmpA encapsulated in chitosan nanoparticles shows superior protection over inactivated whole cell vaccine in orally vaccinated fringed-lipped peninsula carp (*Labeo fimbriatus*). *Vaccines*, 4(4). DOI: 10.3390/vaccines4040040
- Eissa, I. A. M., El-Lamei, M., Sherif, M., Desuky, E., & Zaki, M., (2015). *Aeromonas veronii* biovar *sobria* a Causative Agent of Mass Mortalities in Cultured Nile Tilapia in El-Sharkia governorate, Egypt. *11*(7), 160–166
- El-Barbary, M. I. (2017). Serum biochemical and histopathological changes associated with *Aeromonas hydrophila* isolated from *Oreochromis niloticus* and *Sparus aurata* with multiple antibiotic resistance index. *Journal of Biological Sciences*, 17(5), 222-234. DOI: 10.3923/jbs.2017.222.234
- Eldar, A., Horovitz, A., & Bercovier, H. (1997). Development and efficacy of a vaccine against *Streptococcus iniae* infection in farmed rainbow trout. *Veterinary Immunology & Immunopathology*, 56, 175-183.
- Eldar, A., Shapiro, O., Bejerano, Y., & Bercovier, H. (1995). Vaccination with whole-cell vaccine and bacterial protein extract protects tilapia against *Streptococcus difficile* meningoencephalitis. *Vaccine*, 13, 867-870.
- Eldridge, J. H., Hammond, C. J., Meulbroek, J. A., Staas, J. K., Gilley, R. M., & Tice, T. R. (1990). Controlled vaccine release in the gut-associated lymphoid tissues. I. Orally administered biodegradable microspheres target the peyer's patches. *Journal of Controlled Release*, 11(1-3), 205-214. DOI: 10.1016/0168-3659(90)90133-E
- El-Sherif, M. S., & El-Feky, A. M. I. (2009). Performance of Nile tilapia (*Oreochromis niloticus*) fingerlings. II. Influence of different water temperatures. *International Journal of Agriculture & Biology*, 11(3), 301-305.
- Embregts, C. W. E., & Forlenza, M. (2016). Oral vaccination of fish: Lessons from humans and veterinary species. *Developmental & Comparative Immunology*, 64, 118-137. DOI: 10.1016/j.dci.2016.03.024

- Embregts, C. W.E., Rigaudeau, D., Tacchi, L., Pijlman, G. P., Kampers, L., Veselý, T., & Forlenza, M. (2019). Vaccination of carp against SVCV with an oral DNA vaccine or an insect cells-based subunit vaccine. *Fish & Shellfish Immunology*, *85*, 66–77. <https://doi.org/10.1016/j.fsi.2018.03.028>
- Esfandiari, K. M., Ina-Salwany, M. Y., Mohamad, A., Soltani, M., Sabili, A., Karimi, & M., Muthukrishnan, S. (2019). Growth performance, palatability and water stability of oral feed- based vaccines against *Streptococcus agalactiae* in Red Tilapia (*Oreochromis* sp.). *Journal of Biochemical Technology*, *2*, 106-115.
- Evans, J. J., Klesius, P. H., & Shoemaker, C. A. (2004). Efficacy of *Streptococcus agalactiae* (group B) vaccine in tilapia (*Oreochromis niloticus*) by intraperitoneal and bath immersion administration. *Vaccine*, *22*(27-28), 3769-3773. DOI: 10.1016/j.vaccine.2004.03.012
- Evensen, Ø. (2016). Immunization strategies against *Piscirickettsia salmonis* infections: review of vaccination approaches and modalities and their associated immune response profiles. *Frontiers in Immunology*, *7*, 482. DOI: 10.3389/fimmu.2016.00482
- FAO (2015). Fisheries and Aquaculture Statistics. FAO: Rome, Italy.
- FAO (2016). National aquaculture sector overview. Malaysia. National aquaculture sector overview fact sheets. Fisheries and aquaculture information and statistics branch.
- FAO (2017). Cultured Aquatic Species Information Programme *Oreochromis niloticus* (Linnaeus, 1758). [Cited 20 Dec 2017.]. Retrieved at [http://www.fao.org/fishery/cultured-species/Oreochromis\\_niloticus/en](http://www.fao.org/fishery/cultured-species/Oreochromis_niloticus/en).
- FAO (2018a). Food and Agriculture Organization. Fish Stat J. Rome: FAO. [Accessed 2020 July 5].
- FAO (2018b). Food and Agriculture Organization. FAOSTAT. Rome: FAO. [Accessed 2019 July 7]. <http://www.fao.org/faostat/en/#home>.
- FAO (2020a). The State of World Fisheries and Aquaculture 2020. Sustainability in action. Rome. <https://doi.org/10.4060/ca9229en>
- FAO (2020b). Fishery and Aquaculture Country Profiles. Malaysia. Food and Agriculture Organization of the United Nations. Assessed on 14 April 2020. Retrieved from: [Online Resource]. <http://www.fao.org/fishery/facp/MYS/en>.
- FAO (2021). GLOBEFISH Highlights - A quarterly update on world seafood markets 1st issue 2021 January–September 2020 *Statistics*. Globefish Highlights No. 1–2021. Rome. <https://doi.org/10.4060/cb4129en>
- Ferguson, H. W., Morales, J. A., & Ostland, V. E. (1994) Streptococcosis in aquarium fish. *Diseases of Aquatic Organisms*, *19*(1), 1-6.

- Fernandez, J. B., Yambot, A. V., & Almeria, O. (2014). Vaccination of Nile tilapia (*Oreochromis niloticus*) using Lipopolysaccharide (LPS) prepared from *Aeromonas hydrophila*. *International Journal of Fauna & Biological Studies*, 1, 1-3.
- Figueiredo, H. C. P., Klesius, P. H., Arias, C. R., Evans, J., Shoemaker, C. A., & Jr, D. J. P. (2005). Isolation and characterization of strains of *Flavobacterium columnare* from Brazil. *Fish Diseases*, 28, 199-204.
- Figueiredo, H. C. P., Netto, L. N., Leal, C. A. G., Pereira, U. P., & Mian, G. F. (2012). *Streptococcus iniae* outbreaks in Brazilian Nile tilapia (*Oreochromis niloticus* L.) farms. *Brazilian Journal of Microbiology*, 43, 576-580.
- Fink, I. R., Pietretti, D., Voogdt, C. G. P., Westphal, A. H., Savelkoul, H. F. J., Forlenza, M., & Wiegertjes, G. F. (2016). Molecular and functional characterization of Toll-like receptor (Tlr)1 and Tlr2 in common carp (*Cyprinus carpio*). *Fish & Shellfish Immunology*, 56, 70-83.
- Firdaus-Nawi, M., Yusoff, S. M., Yusof, H., Abdullah, S. Z., & Zamri-Saad, M. (2014). Efficacy of feed-based adjuvant vaccine against *Streptococcus agalactiae* in *Oreochromis* spp. in Malaysia. *Aquaculture Research*, 45(1), 87-96. DOI: 10.1111/j.1365-2109.2012.03207.x
- Fisheries Development Authority of Malaysia (2020a). Official Portal. Fisheries Development Authority of Malaysia. Ministry of Agriculture & Agro-based Industry Malaysia. Assessed on 14 April 2020. Retrieved from:[Online Resource]. <https://www.lkim.gov.my/en/>.
- Fisheries Development Authority of Malaysia (2020b). Laporan Risikoan Pasaran (Annual Market Intelligence Report) 2018 (Edisi Ke-6). Assessed on 2 August 2020. Retrieved from: <https://www.lkim.gov.my/en/annual-market-intelligence-report/>.
- Frederick, W. G., Planas, J. V., & MacKenzie, S. (2004). Tumor necrosis factor. *Developmental & Comparative Immunology*, 28, 487-497.
- Fredriksen, B. N., Olsen, R. H., Furevik, A., Souhoka, R. A., Gauthier, D., & Brudeseth, B. (2013). Efficacy of a divalent and a multivalent water-in-oil formulated vaccine against a highly virulent strain of *Flavobacterium psychrophilum* after intramuscular challenge of rainbow trout (*Oncorhynchus mykiss*). *Vaccine*, 31(15), 1994-1998. DOI: 10.1016/j.vaccine.2013.01.016
- Fu, X., Lin, Q., Liu, L., Liang, H., Huang, Z., & Li, N. (2016). Display of ISKNV orf086 protein on the surface of *Aeromonas hydrophila* and its immunogenicity in Chinese perch (*Siniperca chuatsi*). *Fish & Shellfish Immunology*, 56, 286-293. DOI: 10.1016/j.fsi.2016.07.023
- Fuglem, B., Jirillo, E., Bjerkas, I., Kiyono, H., Nochi, T., Yuki, Y, Raida, M., Fischer, U. (2010). Antigen-sampling cells in the salmonid intestinal epithelium. *Developmental & Comparative Immunology*, 34(7), 768-774. DIO: 10.1016/j.dci.2010.02.007

- Gao, Yan, Pei, C., Sun, X., Zhang, C., Li, L., & Kong, X. (2018). Novel subunit vaccine based on grass carp reovirus VP35 protein provides protective immunity against grass carp hemorrhagic disease. *Fish & Shellfish Immunology*, 75(46), 91–98. <https://doi.org/10.1016/j.fsi.2018.01.050>
- Gao, Yingli, Tang, X., Sheng, X., Xing, J., & Zhan, W. (2016). Antigen uptake and expression of antigen presentation-related immune genes in flounder (*Paralichthys olivaceus*) after vaccination with an inactivated *Edwardsiella tarda* immersion vaccine, following hyperosmotic treatment. *Fish & Shellfish Immunology*. <https://doi.org/10.1016/j.fsi.2016.05.042>
- Garcia, F., Pilarski, F., Onaka, E. M., Moraes, F. R., & Martins, M. L. (2007). Hematology of *Piaractus mesopotamicus* fed diets supplemented with vitamins C and E, challenged by *Aeromonas hydrophila*. *Aquaculture*, 271, 39–46.
- Garlock, T., Asche, F., Anderson, J., Bjørndal, T., Kumar, G., Lorenzen, K., & Tveteras, R. (2019). A Global Blue Revolution: Aquaculture Growth Across Regions, Species, and Countries. *Reviews in Fisheries Science and Aquaculture*, 28(1), 107–116. <https://doi.org/10.1080/23308249.2019.1678111>
- Geven, E. J. W., & Klaren, P. H. M. (2017). The teleost head kidney: Integrating thyroid and immune signalling. *Developmental and Comparative Immunology*, 66, 73–83. <https://doi.org/10.1016/j.dci.2016.06.025>
- Giri, S. S., Sen, S. S., Chi, C., Kim, H. J., & Yun, S. (2015). Effect of guava leaves on the growth performance and cytokine gene expression of *Labeo rohita* and its susceptibility to *Aeromonas hydrophila* infection. *Fish & Shellfish Immunology*. DOI: 10.1016/j.fsi.2015.05.051
- Gomes, S., Afonso, A., & Gartner, F. (2006). Fish vaccination against infections by Streptococcal species and the particular case of Lactococcus. *Returned Peace Corps Volunteers*, 101, 25–35.
- Gomez, D., Sunyer, J. O., & Salinas, I. (2013). The mucosal immune system of fish: the evolution of tolerating commensals while fighting pathogens. *Fish & Shellfish Immunology*, 35(6): 1729–1739.
- Gong, Y., Feng, S., Li, S., Zhang, Y., Zhao, Z., Hu, M., Xu, P., & Jiang, Y. (2017). Genome-wide characterization of Toll-like receptor gene family in common carp (*Cyprinus carpio*) and their involvement in host immune response to *Aeromonas hydrophila* infection. *Comparative Biochemistry & Physiology*, 24, 89–98. <https://doi.org/10.1016/j.cbd.2017.08.003>
- Gu, D., Hu Y., Wei, H., Zhu, Y., Mu, X., Luo, D., Xu, M., & Yang, Y. (2017). Nile Tilapia *Oreochromis niloticus* (L.). In: F. Wan, M. Jiang, & A. Zhan (Eds.), *Biological Invasions and Its Management in China: Vol. 2*. Springer Singapore, Singapore (pp. 77–89).
- Gudmundsdottir, B. K., & Bjornsdottir, B. (2017). *Aeromonas* spp. (*A. salmonicida* and *A. hydrophila*). In P. T. K. Woo, & R.C. Cipriano (Eds.), *Fish Viruses and*

*Bacteria: Pathobiology and Protection* (ed.). England: CABI, Nosworthy Way, Wallingford, Oxfordshire.

- Gulati K, Guhathakurta S, Joshi J, & Rai N, (2016). Cytokines and their Role in Health and Disease: A Brief Overview. *MOJ Immunology*, 4(2), 1–9. <https://doi.org/10.15406/moji.2016.04.00121>
- Guo, Q., Zheng, H., Liu, X., Chi, S., Xu, Z., & Wang, Q. (2019). Nutrient sensing signaling functions as the sensor and regulator of immunometabolic changes in grass carp during *Flavobacterium columnare* infection. *Fish and Shellfish Immunology*, 93, 278–287. <https://doi.org/10.1016/j.fsi.2019.07.041>
- Guo, S., He, L., Wu, L., Xiao, Y., Zhai, S., & Yan, Q. (2020). Immunization of a novel bivalent outer membrane protein simultaneously resisting *Aeromonas hydrophila*, *Edwardsiella anguillarum* and *Vibrio vulnificus* infection in European eels (*Anguilla anguilla*). *Fish & Shellfish Immunology*, 97(43), 46-57. DOI: 10.1016/j.fsi.2019.12.044
- Haddad, G., Hanington, P. C., Wilson, E. C., Grayfer, L., & Belosevic, M. (2008). Molecular and functional characterization of goldfish (*Carassius auratus* L.) transforming growth factor beta. *Developmental & Comparative Immunology*, 32(6), 654-63. DIO: 10.1016/j.dci.2007.10.003
- Halimi, M., Alishahi, M., Abbaspour, M. R., Ghorbanpoor, M., & Tabandeh, M. R. (2019). Valuable method for production of oral vaccine by using alginate and chitosan against *Lactococcus garvieae*/*Streptococcus iniae* in rainbow trout (*Oncorhynchus mykiss*). *Fish & Shellfish Immunology*, 90, 431-439. DOI: 10.1016/j.fsi.2019.05.020
- Hamid, N. H., Hassan, M. D., Sabri, M. M., Hasliza, A. H., Hamdan, R. H., Afifah, M. N., & Fuad, M. M. (2016). Studies on pathogenicity effect of *Aeromonas hydrophila* infection in juvenile red hybrid tilapia *Oreochromis* sp. In D. Yulistiani, A. H. Wardhana, I. Inounu, S. Bahri, S. Iskandar, E. Wina & D. L. Aquino (Eds.), *Proceedings of International Seminar on Livestock Production and Veterinary Technology* (pp. 532-539). Jakarta, Indonesia: IAARD Press.
- Han, B., Xu, K., Liu, Z., Ge, W., Shao, S., Li, P., & Zhang, Z. (2019). Oral yeast-based DNA vaccine confers effective protection from *Aeromonas hydrophila* infection on *Carassius auratus*. *Fish & Shellfish Immunology*. <https://doi.org/10.1016/j.fsi.2018.10.065>
- Han, Z., Zhou, Y., Zhang, X., Yan, J., Xiao, J., Luo, Y., & Zhong, H. (2020). Ghrelin modulates the immune response and increases resistance to *Aeromonas hydrophila* infection in hybrid tilapia. *Fish & Shellfish Immunology*, 98(100), 100-108. <https://doi.org/10.1016/j.fsi.2020.01.006>
- Handfield, M., Simard, P., Couillard, M., & Letarte, R. (1996). *Aeromonas hydrophila* isolated from food and drinking water: hemagglutination, hemolysis, and cytotoxicity for a human intestinal cell line (HT-29). *Applied & Environmental Microbiology*, 62, 3459-3461.

- Hansen, J. D., Landis, E. D., & Phillips, R. B. (2005). Discovery of a unique Ig heavy-chain (IgT) in rainbow trout: Implications for a distinctive B cell developmental pathway in teleost fish. *Proceedings of the National Academy of Sciences of the United States of America*, 102(19), 6919-6924. DOI: 10.1073/pnas.0500027102
- Harikrishnan, R., & Balasundaram, C. (2005). Modern trends in *Aeromonas hydrophila* disease management with fish. *Reviews in Fisheries Science*, 13(4), 281-320. DOI: 10.1080/10641260500320845
- Harikrishnan, Ramasamy, Devi, G., Paray, B. A., Al-Sadoon, M. K., Hoseinifar, S. H., & Gokul, E. (2019). Study the immunomodulation of anthracenedione in striped dwarf catfish, *Mystus vittatus* against pathogenic bacteria, *Aeromonas hydrophila*. *Fish & Shellfish Immunology*, 95, 117-127. <https://doi.org/10.1016/j.fsi.2019.10.033>
- Harikrishnan, R., Jawahar, S., Thamizharasan, S., Paray, B. A., Al-Sadoon, M. K., & Balasundaram, C. (2018). Immune defense of emodin enriched diet in *Clarias batrachus* against *Aeromonas hydrophila*. *Fish & Shellfish Immunology*, 76, 13-20. <https://doi.org/10.1016/j.fsi.2018.02.035>
- Harun, N. O., Wang, T., & Secombes, C. J. (2011). Gene expression profiling in naïve and vaccinated rainbow trout after *Yersinia ruckeri* infection: Insights into the mechanisms of protection seen in vaccinated fish. *Vaccine*, 29(26), 4388-4399. DOI: 10.1016/j.vaccine.2011.04.003
- Hashim, R., Chong, A. S. C., Fatan, N. A., Layman, N., & Ali, A. (2002). Production of hybrid red tilapia, *Oreochromis mossambicus* x *O. niloticus*, at varying stocking densities in portable canvas tanks. *Journal of Applied Aquaculture*, 12(3), 1-12. DOI: 10.1300/J028v12n03\_01
- Hayat, M., Sabri, M. Y., Intan-Shameha, A. R., Ina-Salwany, M. Y., & Thompson, K. D. (2020). Localisation of antigens in the gut post-challenge with *Streptococcus iniae* in vaccinated and non-vaccinated red hybrid tilapia (*Oreochromis* sp.). *Aquaculture International*, 28(4), 1739-1752. <https://doi.org/10.1007/s10499-020-00554-9>
- He, L., Wu, L. Q., Tang, Y. J., Lin, P., Zhai, S. W., Xiao, Y. Q., & Guo, S. L. (2020). Immunization of a novel outer membrane protein from *Aeromonas hydrophila* simultaneously resisting *A. hydrophila* and *Edwardsiella anguillarum* infection in European eels (*Anguilla anguilla*). *Fish & Shellfish Immunology*, 97(43), 300-312. <https://doi.org/10.1016/j.fsi.2019.12.060>
- He, S., Ran, C., Qin, C., Li, S., Zhang, H., & Vos, W. M. De. (2017). *Anti-Infective Effect of Adhesive Probiotic Lactobacillus in Fish is Correlated With Their Spatial Distribution in the Intestinal Tissue*. (9037), 1-12. <https://doi.org/10.1038/s41598-017-13466-1>
- He, Y. F., Wang, L. M., Zhu, W. B., Dong, Z. J. & Liu, N. (2017a). Effects of salinity on cold tolerance of Malaysian red tilapia. *Aquaculture International*, 25(2), 777-792.



- Heath, C., Gillen, C. M., Chrysanthopoulos, P., Walker, M. J., & Barnes, A. C. (2016). *Streptococcus iniae* cpsG alters capsular carbohydrate composition and is a cause of serotype switching in vaccinated fish. *Veterinary Microbiology*, *193*, 116-124. DOI: 10.1016/j.vetmic.2016.08.011
- Heegaard, P. M. H., Dedieu, L., Johnson, N., Le Potier, M. F., Mockey, M., Mutinelli, F., & Sørensen, N. S. (2011). Adjuvants and delivery systems in veterinary vaccinology: Current state and future developments. *Archives of Virology*, *156*(2), 183–202. <https://doi.org/10.1007/s00705-010-0863-1>
- Heloisa, T., Farias, V., Arijo, S., Medina, A., Pala, G., Jhonny, E., & Belo, D. A. (2020). Fish and Shellfish Immunology Immune responses induced by inactivated vaccine against *Aeromonas hydrophila* in pacu, *Piaractus mesopotamicus*. *Fish & Shellfish Immunology*, *101*, 186-191. <https://doi.org/10.1016/j.fsi.2020.03.059>
- Hirono, I., Nam, B., Kurobe, T., & Alerts, E. (2000). Molecular Cloning, Characterization, and Expression of TNF cDNA and Gene from Japanese Flounder *Paralichthys olivaceus*. *Immunology*, *2000*, 165, 4423-4427. DIO: 10.4049/jimmunol.165.8.4423
- Hirschmann, R. (2019). Number of People Employed in the Fishing Industry in Malaysia 2010-2017. Statista. Assessed on 14 April 2020. Retrieved from:[Online Resource]. <https://www.statista.com/statistics/809640/annual-employment-in-the-fishingindustry-malaysia/>.
- Hoare, R., Ngo, T. P. H., Bartie, K. L., & Adams, A. (2017). Efficacy of a polyvalent immersion vaccine against *Flavobacterium psychrophilum* and evaluation of immune response to vaccination in rainbow trout fry (*Onchorynchus mykiss* L.). *Veterinary Research*, *48*(1), 1-13. DOI: 10.1186/s13567-017-0448-z
- Hodgkinson, J. W., Grayfer, L., & Belosevic, M. (2015). Biology of bony fish macrophages. *Biology*, *4*(4), 881–906. <https://doi.org/10.3390/biology4040881>
- Holland, M. C., & Lambris, J. D. (2002):. The complement system of teleosts. *Fish & Shellfish Immunology*, *12*, 399-420.
- Hoshina, T., Sano, T., & Morimoto, Y. (1958). A Streptococcus pathogenic for fish. *Tokyo University of Fisheries*, *44*, 57-68.
- Hossain, S., De Silva, B. C. J., Wimalasena, S. H. M. P., Pathirana, H. N. K. S., Dahanayake, P. S., & Heo, G. J. (2018). Distribution of antimicrobial resistance genes and class 1 integron gene cassette arrays in motile aeromonas spp. Isolated from goldfish (*Carassius auratus*). *Microbial Drug Resistance*, *24*(8), 1217–1225. <https://doi.org/10.1089/mdr.2017.0388>
- Huang, L. Y., Wang, K. Y., Xiao, D., Chen, D. F., Geng, Y., Wang, J., He, Y., Wang, E. L., Huang, J. L. & Xiao, G. Y. (2014). Safety and immunogenicity of an oral DNA vaccine encoding Sip of *Streptococcus agalactiae* from Nile tilapia *Oreochromis niloticus* delivered by live attenuated *Salmonella typhimurium*. *Fish & Shellfish Immunology*, *38*, 34-41.

- Hwang, J. Y., Kwon, M. G., Seo, J. S., Hwang, S. D., Jeong, J. M., Lee, J. H., Jeong, A. R., & Jee, B. Y. (2020). Fish and Shellfish Immunology Current use and management of commercial fish vaccines in Korea. *Fish and Shellfish Immunology*, 102, 20–27. <https://doi.org/10.1016/j.fsi.2020.04.004>
- Ibrahim, A. Z., Hassan, K. H., Kamarudin, R., Anuar, & A. R. (2018). Measuring sustainable livelihood for Malaysia's poor: the Sustainable Livelihoods Index and the B40 group of coastal fishermen in northern Peninsular Malaysia. *LIFEWAYS-International Journal of Society, Development and Environment in the Developing World* 2, 39-47.
- Ighwela, K. A., Ahmad, A. B., & Abol-Munaf A. B. (2012). Haematological Changes in Nile Tilapia (*Oreochromis Niloticus*) Fed with Varying Dietary Maltose Levels. *World Journal of Fish & Marine Sciences*, 4(4), 376-81.
- Iliev, D. B., Thim, Lagos, H. L., Olsen, R., & Jørgensen, J. B. (2013). Homing of antigen-presenting cells in head kidney and spleen salmon head kidney hosts diverse APC types. *Frontiers in Immunology*, 4.
- Ingunn, S., Bjorn, Eirik, B., & Petter, F. (2005). Vaccines for fish in aquaculture. *Expert Review of Vaccines*, 4(1), 89-101.
- Intervet (2006). Streptococcosis in Tilapia. Available from [http://aqua.intervet.com/news/2006-06-20\\_streptococcosis\\_in\\_tilapia.asp](http://aqua.intervet.com/news/2006-06-20_streptococcosis_in_tilapia.asp).
- Irungu, F. G., Mutungi, C. M., Faraj, A. K., Affognon, H., Kibet, N., Tanga, C., & Fiaboe, K. K. M. (2018). Physico-chemical properties of extruded aquafeed pellets containing black soldier fly (*Hermetia illucens*) larvae and adult cricket (*Acheta domesticus*) meals. *Journal of Insects as Food and Feed*, 4(1), 19–30. <https://doi.org/10.3920/JIFF2017.0008>
- Ismail, M. S., Siti-Zahrah, A., Syafiq, M. R. M., Amal, M. N. A., Firdaus-Nawi, M., & Zamri-Saad, M. (2016). Feed-based vaccination regime against streptococcosis in red tilapia, *Oreochromis niloticus* × *Oreochromis mossambicus*. *BMC Veterinary Research*, 12(1), 1-6. DOI: 10.1186/s12917-016-0834-1
- Ismail, M. S., Syafiq, M. R., Siti-Zahrah, A., Fahmi, S., Shahidan, H., Hanan, Y., Amal, M. N. A., & Zamri Saad, M., (2017). The effect of feed-based vaccination on tilapia farm endemic for streptococcosis. *Fish & Shellfish Immunology*, 60, 21-24. DOI: 10.1016/j.fsi.2016.11.040
- Ito, T., & Maeno, Y. (2015). Effect of booster shot and investigation of vaccination efficacy period against herpesviral haematopoietic necrosis (HVHN) in goldfish, *Carassius auratus*. *Veterinary microbiology*, 175(1), 139-144.
- Iwasaki, A., & Medzhitov, R. (2015). Control of adaptive immunity by the innate immune system. *Cancer Research*, 16(4), 343-353. DOI: 10.1038/ni.3123
- Jaafar, R. M., Chettri, J. K., Dalsgaard, I., Al-jubury, A., Kania, P. W., Skov, J., & Buchmann, K. (2015). Effects of adjuvant Montanide™ ISA 763 A VG in rainbow trout injection vaccinated against *Yersinia ruckeri*. *Fish and Shellfish Immunology*,

47(2), 797–806. <https://doi.org/10.1016/j.fsi.2015.10.023>

- Jahari, M. A., Mustafa, S., Akhmal, M., Roslan, H., Manap, Y. A., Lamasudin, U., & Jamaludin, F. I. (2018). The effects of synbiotics and probiotics supplementation on growth performance of red hybrid tilapia, *Oreochromis mossambicus* x *Oreochromis niloticus*. *Journal of Biochemistry, Microbiology and Biotechnology*, 6(1), 5–9.
- Janda, J. M., & Abbott, S. L. (2010). The Genus *Aeromonas*: Taxonomy, Pathogenicity, and Infection. *Clinical Microbiology Reviews*, 23(1), 35-73. DOI: 10.1128/cmr.00039-09
- Jansen, M. D., Dong, H. T., & Mohan, C. V. (2019). Tilapia lake virus: a threat to the global tilapia industry? *Reviews in Aquaculture*, 11(3), 725-739. DOI: 10.1111/raq.12254
- Jantrakajorn, S., Maisak, H., & Wongtavatchai, J. (2014). Comprehensive investigation of streptococcosis outbreaks in cultured Nile tilapia, *Oreochromis niloticus*, and Red Tilapia, *Oreochromis* sp., of Thailand. *Journal of the World Aquaculture Society*, 45, 392-402.
- Jeffery, K. R., Stone, D., Feist, S. W., & Verner-jeffreys, D. W. (2010). An outbreak of disease caused by *Francisella* sp. in Nile tilapia *Oreochromis niloticus* at a recirculation fish farm in the UK. *Diseases of Aquatic Organisms*, 91, 161-165.
- Jiang, J., Zheng, Z., Wang, K., Wang, J., He, Y., Wang, E., Wang, E., Chen, D., Ouyang, P., Geng, Y., & Huang, X. (2015). Adjuvant immune enhancement of subunit vaccine encoding pSCPI of *Streptococcus iniae* in channel catfish (*Ictalurus punctatus*). *International Journal of Molecular Sciences*, 16(12), 28001-28013. DOI: 10.3390/ijms161226082
- Jinendiran, S., Nathan, A. A., Ramesh, D., Vaseeharan, B., & Sivakumar, N. (2019). Modulation of innate immunity, expression of cytokine genes and disease resistance against *Aeromonas hydrophila* infection in goldfish (*Carassius auratus*) by dietary supplementation with *Exiguobacterium acetylicum* S01. *Fish & Shellfish Immunology*, 84, 458–469. <https://doi.org/10.1016/j.fsi.2018.10.026>
- Jørgensen, L. V. G., Nemli, E., Heinecke, R. D., Raida, M. K., & Buchmann, K. (2008). Immune relevant genes expressed in rainbow trout following immunization with a live vaccine against *Ichthyophthirius multifiliis*. *Diseases of Aquatic Organisms*, 80, 189-97.
- Jun, J. W., Kang, J. W., Giri, S. S., Yun, S., Kim, H. J., Guen Kim, S., & Park, S. C. (2020). Immunostimulation by starch hydrogel-based oral vaccine using formalin-killed cells against edwardsiellosis in Japanese eel, *Anguilla japonica*. *Vaccine*, 38(22), 3847–3853. <https://doi.org/10.1016/j.vaccine.2020.03.046>
- Kadowaki, T., Yasui, Y., Takahashi, Y., Kohchi, C., Soma, G., & Inagawa, H. (2009). Comparative immunological analysis of innate immunity activation after oral administration of wheat fermented extract to teleost fish. *Anticancer Research*, 29(11), 4871-7.

- Kahieshesfandiari, M. (2018). *Application of bio-film chitin oral vaccine on streptococcosis infcted red hybrid tilapia (Oreochromis sp.)*. (Unpublished doctoral dissertation) Universiti Putra Malaysia, Malaysia.
- Kahieshesfandiari, M., Sabri, M. Y., Ina-salwany, M. Y., Hassan, M. D., Noraini, O., Ajadi, A. A., & Isiaku, A. I. (2019). Streptococcosis in *Oreochromis* sp.: is feed-based biofilm vaccine of *Streptococcus agalactiae* effective? *Aquaculture International*, 27, 817-832. DOI: 10.1007/s10499-019-00372-8
- Kany, S., Vollrath, J. T., & Relja, B. (2019). Cytokines in inflammatory disease. *International Journal of Molecular Sciences*, 20(23), 1–31.
- Karsi, A., Lawrence, M. L. & Abdelhamed, H. (2017). Live attenuated *Edwardsiella ictaluri* vaccine and method of using the same. US Patent US2017/0065695 A1.
- Kato, G., Kondo, H., Aoki, T., & Hirono, I. (2012). Mycobacterium bovis BCG vaccine induces non-specific immune responses in Japanese flounder against *Nocardia seriolae*. *Fish & Shellfish Immunology*, 33(2), 243–250. <https://doi.org/10.1016/j.fsi.2012.05.002>
- Katzenback, B. A. (2015). Antimicrobial peptides as mediators of innate immunity in teleosts. *Biology*, 4(4), 607-639.
- Kayansamruaj, P., Areechon, N., & Unajak, S. (2020). Development of fish vaccine in Southeast Asia: A challenge for the sustainability of SE Asia aquaculture. *Fish and Shellfish Immunology*, 103, 73-87. <https://doi.org/10.1016/j.fsi.2020.04.031>
- Kayansamruaj, P., Pirarat, N., Katagiri, T., Hirono, I. & Rodkhum, C. (2014). Molecular characterization and virulence gene profiling of pathogenic *Streptococcus agalactiae* populations from tilapia (*Oreochromis* sp.) farms in Thailand. *Veterinary Diagnostic Investigation*, 26, 488-495.
- Kim, J. H., Sohn, S., Kim, S. K., & Hur, Y. B. (2020). Effects on hematological parameters, antioxidant and immune responses, AChE, and stress indicators of olive flounders, *Paralichthys olivaceus*, raised in bio-floc and seawater challenged by *Edwardsiella tarda*. *Fish & Shellfish Immunology*, 97, 194–203. <https://doi.org/10.1016/j.fsi.2019.12.011>
- Kim, S. H., & Jang, Y. S. (2014). Antigen targeting to M cells for enhancing the efficacy of mucosal vaccines. *Experimental & Molecular Medicine*, 46(3), e85-8. DOI: 10.1038/emm.2013.165
- Kirov, S. M. (1997) *Aeromonas* and *Plesiomonas* species. In Food Microbiology Fundamentals and Frontiers ed. Doyle, M, Beuchat, L and Montville, T pp. 265-287. Washington: American Society for Microbiology.
- Kirubakaran, C. J. W., Subramani, P. A., & Michael, R. D. (2016). Methanol extract of *Nyctanthes arbortristis* seeds enhances non-specific immune responses and protects *Oreochromis mossambicus* (Peters) against *Aeromonas hydrophila* infection. *Research in Veterinary Science*, 105, 243–248. <https://doi.org/10.1016/j.rvsc.2016.02.013>

- Kitao, T., Aoki, T., & Sakoh, R. (1981). Epizootic caused by beta-haemolytic *Streptococcus* species in cultured freshwater fish. *Fish Pathology*, *15*, 301-307.
- Klesius, P. H., Evans, J. J., Shoemaker, C. A., & Pasnik, D. J. (2006). A vaccination and challenge model using calcein marked fish. *Fish & Shellfish Immunology*, *20*(1), 20-28. DOI: 10.1016/j.fsi.2005.03.003
- Klesius, P. H., Shoemaker, C. A., & Evans, J. J. (2000). Efficacy of single and combined *Streptococcus iniae* isolate vaccine administered by intraperitoneal and intramuscular routes in tilapia (*Oreochromis niloticus*). *Aquaculture*, *188*(3-4), 237-246. DOI: 10.1016/S0044-8486(00)00345-8
- Kole, S., Qadiri, S. S. N., Shin, S. M., Kim, W. S., Lee, J., & Jung, S. J. (2019). Nanoencapsulation of inactivated-viral vaccine using chitosan nanoparticles: Evaluation of its protective efficacy and immune modulatory effects in olive flounder (*Paralichthys olivaceus*) against viral haemorrhagic septicaemia virus (VHSV) infection. *Fish and Shellfish Immunology*, *91*, 136-147. <https://doi.org/10.1016/j.fsi.2019.05.017>
- Komai, T., Inoue, M., Okamura, T., Morita, K., Iwasaki, Y., Sumitomo, S., & Fujio, K. (2018). Transforming growth factor- $\beta$  and interleukin-10 synergistically regulate humoral immunity via modulating metabolic signals. *Frontiers in Immunology*, *9*, 1-15. <https://doi.org/10.3389/fimmu.2018.01364>
- Kondera, E. (2014). Cell composition of the head kidney of European chub (*Squalius cephalus* L.). *Archives of Polish Fisheries*, *22*(4), 271-280.
- Kordon, A. O., Abdelhamed, H., Ahmed, H., Baumgartner, W., Karsi, A., & Pinchuk, L. M. (2019). Assessment of the live attenuated and wild-type edwardsiella ictaluri-induced immune gene expression and langerhans-like cell profiles in the immune-related organs of catfish. *Frontiers in Immunology*, *10*, 1-12. DOI: 10.3389/fimmu.2019.00392
- Kubista, M., Andrade, J. M., Bengtsson, M., Forootan, A., Jonák, J., Lind, K., Sindelka, R., Sjöback, R., Sjögreen, B., & Strömbom, L. (2006). The real-time polymerase chain reaction. *Molecular Aspects of Medicine*, *27*, 95-125. DOI: 10.1016/j.mam.2005.12.007
- Kumar, B. V., (2019). Macrophages: The Potent Immunoregulatory Innate Immune Cells Submitted: November 22nd 2018 Reviewed: June 12th 2019 Published: July 18th 2019 DOI: 10.5772/intechopen.88013
- Kumar, M., Gupta G, Vikas, & Sharma, S. (2018). Feed Based Vaccine in Aquaculture. Pro Aqua Farm Marine Biol. *Progress in Aqua Farming and Marine Biology Review Article*, *1*(2), 180013. Retrieved from <https://chembiopublishers.com/submit-manuscript.php>
- Kumar, R., Pande, V., Singh, L., Sharma, L., & Saxena, N. (2016b). Pathological findings of experimental *Aeromonas hydrophila* infection in golden mahseer (*Tor putitora*). *Fish Aquaculture*, *7*, 160. DOI: 10.4172/2150-3508.1000160

- Kumar, V., Roy, S., Meena, D. K., & Sarkar, U. K. (2016a). Application of probiotics in shrimp aquaculture: importance, mechanisms of action, and methods of administration. *Reviews in Fisheries Science & Aquaculture*, 24 (4), 342-368
- Kusuda, R., Hamaguchi, M. (1988). The efficacy of attenuated live bacterin of *Pasteurella piscicida* against pseudotuberculosis in yellowtail. *Bulletin- European Association of Fish Pathologists*, 8(3), 50.
- Lacerda, I. P. S., Goncalves, Y. S., Oliveira, S. T. L., Demarqui, F. N., Krewer, C., Gouveia, G. V., & Costa, M. M. (2015). Efficacy of *Aeromonas hydrophila* S-layer bacterins with different protein profiles as a vaccine in Nile tilapia (*Oreochromis niloticus*). *African Journal of Microbiology Research*, 9(29), 1770–1777. <https://doi.org/10.5897/ajmr2015.7586>
- Laith, A. A., Abdullah, M. A., Nurhafizah, W. W. I., Hussein, H. A., Aya, J., Effendy, A. W. M., & Najiah, M. (2019). Efficacy of live attenuated vaccine derived from the *Streptococcus agalactiae* on the immune responses of *Oreochromis niloticus*. *Fish & Shellfish Immunology*, 90, 235-243.
- Laith, A. A., Ambak, M. A., Hassan, M., Sheriff, S., & Nadirah, M. (2017). Molecular identification and histopathological study of natural *Streptococcus agalactiae* infection in hybrid tilapia (*Oreochromis niloticus*). *Veterinary World*, 10(1), 101-111. DOI: 10.14202/vetworld.2017.101-111
- Laith, A. R., & Najiah, M. (2013). *Aeromonas hydrophila*: Antimicrobial susceptibility and histopathology of isolates from diseased catfish, *Clarias gariepinus* (Burchell). *Journal of Aquaculture Research & Development*, 5(2), 1-7. DOI: 10.4172/2155-9546.1000215
- Lan, N. G. T., Salin, K. R., Longyant, S., Senapin, S., & Dong, H. T. (2020). Systemic and mucosal antibody response of freshwater cultured Asian seabass (*Lates calcarifer*) to monovalent and bivalent vaccines against *Streptococcus agalactiae* and *Streptococcus iniae*. *Fish & Shellfish Immunology*, 108, 7-13. <https://doi.org/10.1016/j.fsi.2020.11.014>
- Le Breton, A. D. (2009). Vaccines in Mediterranean aquaculture : Practice and needs. *Options Méditerranéennes. Séries A. Mediterranean Seminars*, 154(86), 147-154. Retrieved from <http://om.ciheam.org/om/pdf/a86/00801068.pdf>
- Lee, N. H., Lee, J. A., Park, S. Y., Song, C. S., Choi, I. S., & Lee, J. B. (2012). A review of vaccine development and research for industry animals in Korea. *Clinical & Experimental Vaccine Research*, 1(1), 18-34.
- Leong, J. A. (1993). Molecular and biotechnological approaches to fish vaccines. *Current Opinion in Biotechnology*, 4(3), 286-293.
- Li, J., Tang, L., Li, S., Li, G., & Mo, Z. (2020). The efficacy and side-effects of oil-based adjuvants emulsified *Vibrio anguillarum* bivalent inactivated vaccine in turbot (*Scophthalmus maximus*) under production mode. *Aquaculture*, 524, 735259. <https://doi.org/10.1016/j.aquaculture.2020.735259>

- Li, L. P., Wang, R., Liang, W. W., Huang, T., Huang, Y., Luo, F. G., & Gan, X. (2015). Development of live attenuated *Streptococcus agalactiae* vaccine for tilapia via continuous passage invitro. *Fish & Shellfish Immunology*, 45(2), 955–963. <https://doi.org/10.1016/j.fsi.2015.06.014>
- Li, M. O., & Flavell, R. A. (2008). TGF-beta: a master of all T cell trades. *Cell*, 134(3), 392-404.
- Li, W., Li, Y., Hu, Y. Z., Mo, X. B., Xu, G. H., Li-Wei Xie, L. W., & Li, A. X. (2019). GroEL, a novel vaccine candidate of piscine *Streptococcus agalactiae* identified by immunoproteome. *Fish and Shellfish Immunology*, 84, 377–383. <https://doi.org/10.1016/j.fsi.2018.10.020>
- Lin, S., Mao, S., Guan, Y., Luo, L., Luo, L., & Pan, Y. (2012). Effects of dietary chitosan oligosaccharides and Bacillus coagulans on the growth, innate immunity and resistance of koi (*Cyprinus carpio* koi). *Aquaculture*, 342–343, 36–41. <https://doi.org/10.1016/j.aquaculture.2012.02.009>
- Ling, X. dong, Dong, W. tao, Zhang, Y., Qian, X., Zhang, W. dong, He, W. hong & Liu, J. X. (2019). Comparative transcriptomics and histopathological analysis of crucian carp infection by atypical *Aeromonas salmonicida*. *Fish and Shellfish Immunology*, 94, 294–307. <https://doi.org/10.1016/j.fsi.2019.09.006>
- Lipiäinen, T., Peltoniemi, M., Sarkhel, S., Yrjönen, T., Vuorela, H., Urtti, A., & Juppo, A. (2015). Formulation and stability of cytokine therapeutics. *Journal of Pharmaceutical Sciences*, 104(2), 307–326. <https://doi.org/10.1002/jps.24243>
- Liu, C., Hu, X., Cao, Z., Sun, Y., Chen, X., & Zhang, Z. (2019). Construction and characterization of a DNA vaccine encoding the SagH against *Streptococcus iniae*. *Fish & Shellfish Immunology*, 89, 71–75. <https://doi.org/10.1016/j.fsi.2019.03.045>
- Liu, F., Tang, X., Sheng, X., Xing, J., & Zhan, W. (2016b). *Edwardsiella tarda* outer membrane protein C: An immunogenic protein induces highly protective effects in flounder (*Paralichthys olivaceus*) against edwardsiellosis. *International Journal of Molecular Sciences*, 17, 1117. DOI:10.3390/ijms17071117.
- Liu, G., Zhang, W., Liu, Y., Yao, H., Lu, C., & Xu, P. (2014). Identification of a virulence-related surface protein XF in piscine *Streptococcus agalactiae* by pre-absorbed immunoproteomics. *BMC Veterinary Research*, 10(1), 1-11. DOI: 10.1186/s12917-014-0259-7.
- Liu, G., Zhu, J., Chen, K., Gao, T., Yao, H., Liu, Y., Zhang, W., & Lu, C. (2016a). Development of *Streptococcus agalactiae* vaccines for tilapia. *Diseases of Aquatic Organisms*, 122(2), 163-170. DOI: 10.3354/dao03084
- Liu, L., Gao, S., Luan, W., Zhou, J., & Wang, H. (2018). Generation and functional evaluation of a DNA vaccine co-expressing Cyprinid herpesvirus-3 envelope protein and carp interleukin-1 beta. *Fish & Shellfish Immunology*, 80, 223–231. <https://doi.org/10.1016/j.fsi.2018.05.046>

- Liu, Z., Liu, W., Ran, C., Hu, J., & Zhou, Z. (2016c). Abrupt suspension of probiotics administration may increase host pathogen susceptibility by inducing gut dysbiosis. *Scientific Reports*, 6(12), 1–12. <https://doi.org/10.1038/srep23214>
- Locati, M., Mantovani, A., & Sica, A. (2013). Macrophage activation and polarization as an adaptive component of innate immunity. *Advances in Immunology*, 120, 163–184.
- Locke, J. B., Aziz, R. K., Vicknair, M. R., Nizet, V., & Buchanan, J. T. (2008). *Streptococcus iniae* M-like protein contributes to virulence in fish and is a target for live attenuated vaccine development. *PLoS ONE*, 3(7), 1–13. <https://doi.org/10.1371/journal.pone.0002824>
- Lu, D., Bei, J., Feng, L., Zhang, Y., Liu, X., Wang, L., Chen, J. L., & Lin, H. (2008). Interleukin-1 beta gene in orange-spotted grouper, *Epinephelus coioides*: Molecular cloning, expression, biological activities and signal transduction. *Molecular Immunology*, 45, 857–867. DOI: 10.1016/j.molimm.2007.08.009
- Ma, J., Bruce, T. J., Jones, E. M., & Cain, K. D. (2019). *A Review of Fish Vaccine Development Strategies: Conventional Methods and Modern Biotechnological Approaches*. *Microorganisms* 2019, 7, 569; doi:10.3390/microorganisms7110569
- Ma, Y., Hao, L., Liang, Z., Ma, J., Ke, H., Yang, H. & Liu, Z. (2020). Characterization of novel antigenic vaccine candidates for Nile tilapia (*Oreochromis niloticus*) against *Streptococcus agalactiae* infection. *Fish & Shellfish Immunology*. <https://doi.org/10.1016/j.fsi.2020.07.024>
- Mahrous, K. F., Mabrouk, D. M., Aboelenin, M. M., Abd El-Kader, H. A. M., Gaafar, A. Y., Younes, A. M., & Hassanane, M. S. (2020). Molecular characterization and immunohistochemical localization of tilapia piscidin 3 in response to *Aeromonas hydrophila* infection in Nile tilapia. *Journal of Peptide Science*, 1-11. <https://doi.org/10.1002/psc.3280>
- Mamun, M. A. A., Nasren, S., Abhiman, P. B., Rathore, S. S., Sowndarya, N. S., Ramesh, K. S., & Shankar, K. M. (2019). Effect of biofilm of *Aeromonas hydrophila* oral vaccine on growth performance and histopathological changes in various tissues of striped catfish, *Pangasianodon hypophthalmus* (Sauvage 1878). *Indian Journal of Animal Research*, DIO: 10.18805/ijar.b-3814
- Mangunwudoyo, W., Ismayasari, & R., Riani, E. (2009). The activity of chitinase, lechitinase, and hemolycine of isolated bacteria from *Oreochromis niloticus* Lin. cultured in floating net cage at Jatiluhur, Purwakarta. *Jurnal Riset Akuakultur*, 4(2), 257-265.
- Mankar, S. A. (2016). Evaluation of immunostimulant and vaccine on *Labeo rohita* (Rohu). *International Journal of Current Microbiology & Applied Sciences*, 5(4), 626-634. DOI: 10.20546/ijcmas.2016.504.071
- Martinez, G., Harel, J., & Gottschalk, M. (2001). Specific detection by PCR of *Streptococcus agalactiae* in milk. *Canadian Journal of Veterinary Research*, 65, 68-72.



- Martins, M. L., Xu, D. H., Shoemaker, C. A., & Klesius, P. H. (2011). Temperature effects on immune response and hematological parameters of channel catfish *Ictalurus punctatus* vaccinated with live theronts of *Ichthyophthirius multifiliis*. *Fish & Shellfish Immunology*, 31(6), 774-780. DOI: 10.1016/j.fsi.2011.07.015
- Masso-Silva, J. A., & Diamond, G. (2014). Antimicrobial peptides from fish. *Pharmaceuticals*, 7(3), 265-310.
- Matusin, S. B. (2015). *Molecular characterization of Aeromonas hydrophila and development of recombinant cells vaccine expressing outer membrane proteins against its in African catfish (Clarias gariepinus)*. (Unpublished master's dissertation). Universiti Putra Malaysia, UPM, Malaysia.
- McClusky, L. M., & Sulikowski, J. (2014). The epigonal organ and mature pole of the testis in the recreationally fished blue shark (*Prionace glauca*): Histochemico-functional correlates. *Journal of Anatomy*, 225(6), 614-624.
- McDonald, P., Edwards, R. A., Greenhalgh, J. F. D., Morgan, C. A. L., Sinclair, A., & Wilkinson, R. G. (2011). *Animal Nutrition (7<sup>th</sup> ed) Harlow: Pearson*.
- Mishra, A., Nam, G., Gim, J., Lee, H., Jo, A., & Kim, H. (2018). Molecules and cells current challenges of streptococcus infection and effective molecular, cellular, and environmental control methods in aquaculture. *Molecules and Cells*, 41(6), 495-505.
- Mjoun, K., Rosentrater, K. A., & Brown, M. L. (2010) Tilapia: profile and economic importance. Fact Sheets. Paper 163. Available at [http://openprairie.sdstate.edu/extension\\_fact/163](http://openprairie.sdstate.edu/extension_fact/163).
- Mmanda, F. P., Zhou, S., Zhang, J., Zheng, X., An, S., & Wang, G. (2014). Massive mortality associated with *Streptococcus iniae* infection in cage-cultured red drum (*Sciaenops ocellatus*) in Eastern China. *African Journal of Microbiology Research*, 8(16), 1722-1729. DOI: 10.5897/AJMR2014.6659
- Mohammadian, T., Alishahi, M., Tabandeh, M. R., Ghorbanpoor, M., & Gharibi, D. (2017). Changes in immunity, expression of some immune-related genes of shabot fish, tor grypus, following experimental infection with *Aeromonas hydrophila*: Effects of autochthonous probiotics. *Probiotics and Antimicrobial Proteins*, 10(4), 616-628. <https://doi.org/10.1007/s12602-017-9373-8>
- Mohd-Aris, A., Saad, M. Z., Daud, H. M., Yusof, M. T., & Ina-Salwany, M. Y. (2018). *Vibrio harveyi* protease deletion mutant as live attenuated vaccine candidate against vibriosis and transcriptome profiling following vaccination for *Epinephelus fuscoguttatus*. *Aquaculture International*, 1-16. DOI: 10.1007/s10499-018-0311-x
- Monir, M. S., Yusoff, S. B. M., Zulperi, Z. B. M., Hassim, H. B. A., Mohamad, A., Ngoo, M. S. B. M. H., & Ina-Salwany, M. Y. (2020b). Haemato-immunological responses and effectiveness of feed-based bivalent vaccine against *Streptococcus iniae* and *Aeromonas hydrophila* infections in hybrid red tilapia (*Oreochromis*

- mossambicus* × *O. niloticus*). *BMC Veterinary Research*, 16(1), 226. <https://doi.org/10.1186/s12917-020-02443-y>
- Monir, M. S., Yusoff, S. M., Mohamad, A., & Ina-Salwany, M. Y. (2020a). Vaccination of tilapia against motile aeromonas septicemia: A Review. *Journal of Aquatic Animal Health*, 1–8. <https://doi.org/10.1002/aah.10099>
- Muktar, Y., & Tesfaye, S. (2016). Present Status and Future Prospects of Fish Vaccination: A Review. *Journal of Veterinary Science & Technology*, 7(2). <https://doi.org/10.4172/2157-7579.1000299>
- Munang'andu, H. M., & Evensen, Ø. (2015). A review of intra- and extracellular antigen delivery systems for virus vaccines of finfish. *Journal of Immunology Research*, 1-19. DOI: 10.1155/2015/960859
- Munang'andu, H. M., Paul, J., & Evensen, Ø. (2016). An overview of vaccination strategies and antigen delivery systems for *Streptococcus agalactiae* vaccines in Nile tilapia (*Oreochromis niloticus*). *Vaccines*, 4, 1-13. DOI: 10.3390/vaccines4040048
- Munang'andu, H. M., Fredriksen, B. N., Mutoloki, S., Brudeseth B., Kuo, T. Y., Marjara, I. S., Dalmo, R. A., & Evensen, Ø. (2012). Comparison of vaccine efficacy for different antigen delivery systems for infectious pancreatic necrosis virus vaccines in Atlantic salmon (*Salmo salar* L.) in a cohabitation challenge model. *Vaccine*, 30(27), 4007-16.
- Munang'andu, H. M., Fredriksen, B. N., Mutoloki, S., Dalmo, R. A., & Evensen, Ø. (2013). Antigen dose and humoral immune response correspond with protection for inactivated infectious pancreatic necrosis virus vaccines in Atlantic salmon (*Salmo salar* L.). *Veterinary Research*, 44(1), 7.
- Musa, N., Wei, L. S., Musa, N., Hamdan, R. H., Leong, L. K., Wee, W., Amal M., N., Kutty, B. M., & Abdullah S., Z. (2009). Streptococcosis in red hybrid tilapia (*Oreochromis niloticus*) commercial farms in Malaysia. *Aquaculture Research*, 40(5), 630-632. DOI: 10.1111/j.1365-2109.2008.02142.x
- Mutoloki, S., Munang'andu, H. M., & Evensen, Ø. (2015). Oral vaccination of fish - antigen preparations, uptake, and immune induction. *Frontiers in Immunology*, 6, 519. DOI: 10.3389/fimmu.2015.00519
- Mutoloki, S., Reite, O. B., Brudeseth, B., Tverdal, A., & Evensen, Ø. (2006). A comparative immunopathological study of injection site reactions in salmonids following intraperitoneal injection with oil-adjuvanted vaccines. *Vaccine*, 24(5), 578-588. <https://doi.org/10.1016/j.vaccine.2005.08.070>
- Nahar, S., Mizanur Rahman, M., Uddin Ahmed, G., Ali Reza Faruk, M., & Shamsun Nahar, C. (2016). Isolation, identification, and characterization of *Aeromonas hydrophila* from juvenile farmed pangasius (*Pangasianodon hypophthalmus*). *International Journal of Fisheries and Aquatic Studies*, 4(4), 52–60. [www.fisheriesjournal.com](http://www.fisheriesjournal.com)

- Najiah, M., Aqilah, N. I., Lee, K. L., Khairulbarryyah, Z., Mithun, S., Jalal, K. C. A., & Nadirah, M. (2012). Massive mortality associated with *Streptococcus agalactiae* infection in cage- cultured red hybrid tilapia *Oreochromis niloticus* in Como River, Kenyir Lake, Malaysia. *Journal of Biological Sciences*, 12(8), 438-442. doi: 10.3923/jbs.2012.438.442
- Nakatsugawa, T. (1983). A streptococcal disease of cultured flounder. *Fish pathology*, 17, 281-285.
- Nasrullah, H., Nababan, Y. I., Yanti, D. H., Hardiantho, D., Nuryati, S., Junior, M. Z. & Alimuddin, A. (2019). Identification and expression analysis of c-type and g-type lysozymes genes after *Aeromonas hydrophila* infection in African catfish. *Jurnal Akuakultur Indonesia*, 18(2), 110–119. <https://doi.org/10.19027/jai.18.2.1-10>
- Netea, M. G., Simon, A., van de Veerdonk, F., Kullberg, B. J., Van der Meer, J. W., & Joosten, L. A. (2010). IL-1beta processing in host defense: beyond the inflammasomes. *PLoS Pathog*, 6, 1000661.
- Newman, S. G. (1993). Bacterial vaccines for fish. *Annual Review of Fish Diseases*, 3, 145-185.
- Nguyen, H. T., Thu Nguyen, T. T., Tsai, M. A., Ya-Zhen, E., Wang, P. C., & Chen, S. C. (2017). A formalin-inactivated vaccine provides good protection against *Vibrio harveyi* infection in orange-spotted grouper (*Epinephelus coioides*). *Fish & Shellfish Immunology*, 65, 118-126. DOI: 10.1016/j.fsi.2017.04.008
- Nguyen, T. T. T., Nguyen, H. T., Chen, Y. C., Hoang, H. H., Wang, P. C., & Chen, S. C. (2018). Effectiveness of a divalent *Streptococcus dysgalactiae* inactivated vaccine in cobia (*Rachycentron canadum* L.). *Aquaculture*, 495, 130-135. DIO: 10.1016/j.aquaculture.2018.05.034
- Nicholson, P., Monon, N., Jaemwimol, P., Tattiyapong, P., & Surachetpong, W. (2019). Coinfection of tilapia lake virus and *Aeromonas hydrophila* synergistically increased mortality and worsened the disease severity in tilapia (*Oreochromis* spp.). *Aquaculture*, 734746. <https://doi.org/10.1016/j.aquaculture.2019.734746>
- Nielsen, M. E., Høi, L., Schmidt, A. S., Qian, D., Shimada, T., Shen, J. Y., & Larsen, J. L. (2001). Is *Aeromonas hydrophila* the dominant motile *Aeromonas* species that causes disease outbreaks in aquaculture production in the Zhejiang Province of China? *Diseases of Aquatic Organisms*, 46, 23-29.
- Nikoskelainen, S., Verho, S., Jarvinen, S., Madetoja, J., Wiklund, T., & Lilius, E. M. (2007). Multiple whole bacterial antigens in polyvalent vaccine may result in inhibition of specific responses in rainbow trout (*Oncorhynchus mykiss*). *Fish & Shellfish Immunology*, 22(3), 206-217. DIO: 10.1016/j.fsi.2006.04.010
- Niu, G., Khattiya, R., Zhang, T., Boonyayatra, S., & Wongsathein, D. (2020). Phenotypic and genotypic characterization of *Streptococcus* spp. isolated from tilapia (*Oreochromis* spp.) cultured in river-based cage and earthen ponds in Northern Thailand. *Journal of Fish Diseases*, 43(3), 391–398. <https://doi.org/10.1111/jfd.13137>

- NOAA (2015). Fisheries, National Marine Fisheries Service, Fisheries Statistics Division. <http://www.nmfs.noaa.gov/aquaculture.html>
- Noraini, O., Sabri, M. Y., & Siti-Zahrah, A. (2013). Efficacy of spray administration of formalin-killed *Streptococcus agalactiae* in hybrid red tilapia. *Journal of Aquatic Animal Health*, 25(2), 142-148. DOI: 10.1080/08997659.2013.781553
- Nur-Nazifah, M., Sabri, M. Y., & Siti-Zahrah, A. (2014). Development and efficacy of feed-based recombinant vaccine encoding the cell wall surface anchor family protein of *Streptococcus agalactiae* against streptococcosis in *Oreochromis* sp. *Fish & Shellfish Immunology*, 37(1), 193-200. DOI: 10.1016/j.fsi.2014.01.011
- Odeyemi, O. A., Asmat, A., & Usup, G. (2012). Antibiotics resistance and putative virulence factors of *Aeromonas hydrophila* isolated from estuary. *Journal of Microbiology, Biotechnology & Food Sciences*, 1, 1339-1357.
- Oliveira, T. F., Queiroz, G. A., Teixeira, J. P., Figueiredo, H. C. P., & Leal, C. A. G. (2018). Recurrent *Streptococcus agalactiae* infection in Nile tilapia (*Oreochromis niloticus*) treated with florfenicol. *Aquaculture*, 493, 51-60. <https://doi.org/10.1016/j.aquaculture.2018.04.037>
- Olsen, M. M., Kania, P. W., Heinecke, R. D., Skjoedt, K., Rasmussen, K. J., & Buchmann, K. (2011). Cellular and humoral factors involved in the response of rainbow trout gills to *Ichthyophthirius multifiliis* infections: molecular and immunohistochemical studies. *Fish & Shellfish Immunology*, 30, 859-69.
- Ortega, C., García, I., Irgang, R., Fajardo, R., Tapia-Cammas, D., Acosta, J., & Avendaño-Herrera, R. (2018). First identification and characterization of *Streptococcus iniae* obtained from tilapia (*Oreochromis aureus*) farmed in Mexico. *Journal of Fish Diseases*, 41(5), 773-782. DOI: 10.1111/jfd.12775
- Osman, K. M., Mohamed, L. A., Eman, H., Rahman, E. H. A., Soliman, W. S. (2009). Trials for vaccination of Tilapia fish against *Aeromonas* and *Pseudomonas* infections using monovalent, bivalent and polyvalent vaccines. *World Journal of Fish and Marine Sciences*, 1(4), 297-304. Retrieved from [http://www.idosi.org/wjfms/wjfms1\(4\)09/8.pdf](http://www.idosi.org/wjfms/wjfms1(4)09/8.pdf)
- Osman, K. M., Al-Maary, K. S., Mubarak, A. S., Dawoud, T. M., Moussa, I. M. I., Ibrahim, M. D. S., & Fawzy, N. M. (2017). Characterization and susceptibility of streptococci and enterococci isolated from Nile tilapia (*Oreochromis niloticus*) showing septicemia in aquaculture and wild sites in Egypt. *BMC Veterinary Research*, 13(1), 1-10. <https://doi.org/10.1186/s12917-017-1289-8>
- Pablos, M., Rodríguez-Calleja, J. M., Santos, J. A., Otero, A., & García-López, M. L. (2009). Occurrence of motile *Aeromonas* in municipal drinking water and distribution of genes encoding virulence factors. *International Journal of Food Microbiology*, 135, 158-164.
- Palikova, M., Papezikova, I., Markova, Z., Navratil, S., Mares, J., Mares, L., & Schmidt-Posthaus, H. (2017). Proliferative kidney disease in rainbow trout (*Oncorhynchus mykiss*) under intensive breeding conditions: Pathogenesis and haematological and

immune parameters. *Veterinary Parasitology*, 238, 5–16. <https://doi.org/10.1016/j.vetpar.2017.03.003>

Parra, D., Takizawa, F., & Sunyer, J. O. (2013). Evolution of B cell immunity. *Annual Review of Animal Biosciences*, 1(1):65-97.

Pasaribu, W., Sukenda, S., & Nuryati, S., (2018). The efficacy of Nile tilapia (*Oreochromis niloticus*) broodstock and larval immunization against *Streptococcus agalactiae* and *Aeromonas hydrophila*. *Fishes*, 3, 1-14. DOI: 10.3390/fishes3010016.

Pasnik, D. J., Evans, J. J., & Klesius, P. H. (2005). Duration of protective antibodies and correlation with survival in Nile tilapia *Oreochromis niloticus* following *Streptococcus agalactiae* vaccination. *Diseases of Aquatic Organisms*, 66, 129-134.

Pasnik, D. J., Evans, J. J., & Klesius, P. H. (2014). A microwave-irradiated *Streptococcus agalactiae* vaccine provides partial protection against experimental challenge in Nile tilapia, *Oreochromis niloticus*. *World Journal of Vaccines*, 4, 184-189.

Peatman, E., Lange, M., Zhao, H., & Beck, B. H. (2015). Physiology and immunology of mucosal barriers in catfish (*Ictalurus spp.*). *Tissue Barriers*, 3(4), 1–14. <https://doi.org/10.1080/21688370.2015.1068907>

Pennacchi, Y., Adams, M. B., Nowak, B. F., & Bridle, A. R. (2016). Immune gene expression in the gills of Atlantic salmon (*Salmo salar* L.) following experimental reinfection with *Neoparamoeba perurans*. *Aquaculture*, 464, 410–419. <https://doi.org/10.1016/j.aquaculture.2016.07.025>

Pereira, G.V., Silva, B. C., Vieira, F. N., Seiffert, W. Q., Ushizima, T. T., Mourino, J. L. P., & Martins, M. L. (2015). Vaccination strategies with oral booster for surubim hybrid (*Pseudoplatystoma corruscans* x *P. reticulatum*) against haemorrhagic septicemia. *Aquaculture Research*, 46(8), 1831-1841. DOI: 10.1111/are.12339

Pérez-Sánchez, T., Balcázar, J. L., Merrifield, D. L., Carnevali, O., Gioacchini, G., de Blas, I., & Ruiz-Zarzuola, I. (2011). Expression of immune-related genes in rainbow trout (*Oncorhynchus mykiss*) induced by probiotic bacteria during *Lactococcus garvieae* infection. *Fish & Shellfish Immunology*, 31(2), 196–201. <https://doi.org/10.1016/j.fsi.2011.05.005>

Piazzon, M. C., Galindo-Villegas, J., Pereiro, P., Estensoro, I., Caldach-Giner, J. A., Gómez-Casado, E., & Pérez-Sánchez, J. (2016). Differential modulation of IgT and IgM upon parasitic, bacterial, viral, and dietary challenges in a perciform fish. *Frontiers in Immunology*, 7, 1–16. <https://doi.org/10.3389/fimmu.2016.00637>

Pinsky, M. R., Vincent, J. L., Devière, J., Alegre, Kahn, R. J. M., & Dupont, E. (1993). Serum cytokine levels in human septic shock: relation to multiple-system organ failure and mortality. *Chest*, 103, 565-75. DOI: 10.1378/chest.103.2.565

Polinski, M., Bridle, A., & Nowak, B. (2013). Temperature-induced transcription of

- inflammatory mediators and the influence of Hsp70 following LPS stimulation of southern blue fin tuna peripheral blood leukocytes and kidney homogenates. *Fish & Shellfish Immunology*, 34(5), 1147-1157. DOI: 10.1016/j.fsi.2013.01.026
- Pongthana, N., Hong, N., & Ponzoni, R. W. (2010). Comparative performance of four red tilapia strains and their crosses in fresh- and saline water environments. *Aquaculture*, 308, 109-114. DOI: 10.1016/j.aquaculture.2010.07.033
- Ponzoni, R. W., Nguyen, N. H., Khaw, H. L., Hamzah, A., Bakar, K. R. A., & Yee, H. Y. (2011). Genetic improvement of Nile tilapia (*Oreochromis niloticus*) with special reference to the work conducted by the World Fish Center with the GIFT strain. *Reviews in Aquaculture*, 3, 27-41.
- Poynter, S., Lisser, G., Monjo, A., & DeWitte-Orr, S. (2015). Sensors of infection: Viral nucleic acid PRRs in fish. *Biology*, 4, 460-493.
- Prabu, E., Rajagopalsamy, C. B. T., Ahilan, B., Jeevagan, I. J. M. A., & Renuhadevi, M. (2019). Tilapia. An Excellent Candidate Species for World Aquaculture: A Review. *Annual Research & Review in Biology*, 1-14. <https://doi.org/10.9734/arrb/2019/v31i3330052>
- Pradeep, P. J., Suebsing, R., Sirithammajak, S., Kampeera, J., Turner, W., Jeffs, A., Kiatpathomchai, W., & Withyachumanarnkul, B. (2016). Vertical transmission and concurrent infection of multiple bacterial pathogens in naturally infected red tilapia (*Oreochromis* spp.). *Aquaculture Research*, 48(6), 2706-2717. DOI: 10.1111/are.13102
- Prasad, S., & Areechon, N. (2011). Efficacy of Formalin-killed *Aeromonas hydrophila* and *Streptococcus* sp. Vaccine in Red Tilapia. *Our Nature*, 8, 231-240.
- Press, C. M., & Evensen, Ø. (1999). The morphology of the immune system in teleost fishes. *Fish & Shellfish Immunology*, 9, 309-318.
- Pretto-Giordano, L. G., Müller, E. E., Klesius, P., & da Silva, V. G. (2010). Efficacy of an experimentally inactivated *Streptococcus agalactiae* vaccine in Nile tilapia (*Oreochromis niloticus*) reared in Brazil. *Aquaculture Research*, 41(10), 1539-1544.
- Pridgeon, J. W., & Klesius, K. (2012). Major bacterial diseases in aquaculture and their vaccine development. *CAB Reviews*, 7, 1-16.
- Pridgeon, J. W., Klesius, P. H., & Yildirim-Aksoy, M. (2013a). Attempt to develop live attenuated bacterial vaccines by selecting resistance to gossypol, proflavine hemisulfate, novobiocin, or ciprofloxacin. *Vaccine*, 31(18), 2222-2230. DOI: 10.1016/j.vaccine.2013.03.004
- Pridgeon, J. W., Klesius, P. H., Dominowski, P. J., Yancey, R. J., & Kievit, M. S. (2013b). Chicken-type lysozyme in channel catfish: Expression analysis, lysozyme activity, and efficacy as immunostimulant against *Aeromonas hydrophila* infection. *Fish & Shellfish Immunology*, 35(3), 680-688. DOI: 10.1016/j.fsi.2013.05.018

- Pulpipat, T., Maekawa, S., Wang, P., & Chen, S. (2020). Immune Responses and Protective Efficacy of a Formalin-Killed *Francisella koatunensis* Subsp. *Orientalis* Vaccine Evaluated through Intraperitoneal and Immersion Challenge Methods in *Oreochromis niloticus*. *Vaccines*, 8(163), 1–14.
- Pulsford, A. L., Lemaire-Gony, S., Tomlinson, M., Collingwood, N., & Glynn, P. J. (1994). Effects of acute stress on the immune system of the dab, *Limanda limanda*. *Comparative Biochemistry and Physiology*, 109(2), 129–139. [https://doi.org/10.1016/0742-8413\(94\)00053-D](https://doi.org/10.1016/0742-8413(94)00053-D)
- Purwanti, C. S., Suminto, & Sudaryono, A. (2014). The description of blood profile catfish *Clarias gariepinus* that is fed with a combination of artificial feed and earthworm *Lumbricus rubellus*. *Journal of Aquaculture Management & Technology*, 3, 53-60.
- Qiang, J., He, J., Yang, H., Xu, P., Habte-Tsion, H. M., Ma, X. Y., & Zhu, Z. X. (2016). The changes in cortisol and expression of immune genes of GIFT tilapia *Oreochromis niloticus* (L.) at different rearing densities under *Streptococcus iniae* infection. *Aquaculture International*, 24(5), 1365-1378. DOI: 10.1007/s10499-016-9995-y
- Rahman, A. N. A., Khalil, A. A., Abdallah, A. A. & ElHady, H. M. (2018). The effects of the dietary supplementation of Echinacea purpurea extract and/or vitamin C on the intestinal histomorphology, phagocytic activity, and gene expression of the Nile tilapia. *Fish & Shellfish Immunology*, 82, 312–318. <https://doi.org/10.1016/j.fsi.2018.08.024>
- Rahman, M. M., & McFadden, G. (2006). Modulation of tumor necrosis factor by microbial pathogens, 2(2).
- Rahmatullah, M. (2018). *Development of monovalent recombinant vaccine against streptococcus infection in hybrid red tilapia (Oreochromis spp.)*. (Unpublished doctoral dissertation). Universiti Putra Malaysia, Malaysia,
- Rahmatullah, M., Ariff, M., Kahieshesfandiari, M., Daud, H. M., Zamri-Saad, M., Sabri, M. Y., Amal, N. A., & Ina-Salwany, M. Y. (2017). Isolation and pathogenicity of *Streptococcus iniae* in cultured red hybrid tilapia in Malaysia. *Aquatic Animal Health*, 29(4), 208-213. DOI: 10.1080/08997659.2017.1360411
- Ram, M. K., Kumar, B. T. N., Poojary, S. R., Abhiman, P. B., Patil, P., Ramesh, K. S., & Shankar, K. M. (2019). Evaluation of biofilm of *Vibrio anguillarum* for oral vaccination of Asian seabass, *Lates calcarifer* (BLOCH, 1790). *Fish & Shellfish Immunology*, 94, 746-751. DOI: 10.1016/j.fsi.2019.09.053
- Ran, C. (2013). *Isolation and Characterization of Bacillus spp. As Potential Probiotics for Channel Catfish, Ictalurus punctatus*. (Unpublished doctoral dissertation). Auburn University, United States.
- Rao, A., Strauss, O., Kokkinou, E., Bruchard, M., Tripathi, K. P., Schlums, H., & Mjösberg, J. (2020). Cytokines regulate the antigen-presenting characteristics of human circulating and tissue-resident intestinal ILCs. *Nature Communications*,

11(1). <https://doi.org/10.1038/s41467-020-15695-x>

- Rauta, P. R., Nayak, B., Monteiro, G. A., & Mateus, M. (2017). Design and characterization of plasmids encoding antigenic peptides of Aha1 from *Aeromonas hydrophila* as prospective fish vaccines. *Journal of Biotechnology*, 241, 116-126.
- Ravindra, Pradhan, P. K., Paria, A., Pande, V., Verma, D. K., Arya, P. & Sood, N. (2019). Expression of immune genes in Indian major carp, *Catla catla* challenged with *Flavobacterium columnare*. *Fish & Shellfish Immunology*, 94, 599–606. <https://doi.org/10.1016/j.fsi.2019.09.052>
- Reite, O. B., & Evensen, Ø. (2006). Inflammatory cells of teleostean fish: A review focusing on mast cells/eosinophilic granule cells and rodlet cells. *Fish & Shellfish Immunology*, 20(2), 192-208.
- Reyes, M., Ramírez, C., Ñancuqueo, I., Villegas, R., Schaffeld, G., Krizan, L., Gonzalez, J., & Oyarzun, P. (2017). A novel “in-feed” delivery platform applied for oral DNA vaccination against IPNV enables high protection in Atlantic salmon (*Salmon salar*). *Vaccine*, 35(4), 626-632. DOI: 10.1016/j.vaccine.2016.12.013
- Reyes-Cerpa, S., Maisey, K., Reyes-López, F. & Toro-Ascuy, D. (2012). *Fish Cytokines and Immune*. Response Ana María Sandino and Mónica Imarai. New Advances and Contributions to Fish Biology. DOI: 10.5772/53504
- Reygaert, C W. (2018). An overview of the antimicrobial resistance mechanisms of bacteria. *AIMS Microbiology*, 4(3), 482–501. <https://doi.org/10.3934/microbiol.2018.3.482>
- Rodkhum, C., Kayansamruaj, P., Pirarat, N., & Wongtawatchai, J. (2012). Duplex PCR for simultaneous and unambiguous detection of *Streptococcus iniae* and *Streptococcus agalactiae* associated with Streptococcosis of cultured tilapia in Thailand. *Thai Veterinary Medicine*, 42(2), 153-158.
- Rodrigues, A. P., Hirsch, D., Figueiredo, H. C. P., Logato, P. V. R., & Moraes, Â. M. (2006). Production and characterisation of alginate microparticles incorporating *Aeromonas hydrophila* designed for fish oral vaccination. *Process Biochemistry*, 41(3), 638–643. <https://doi.org/10.1016/j.procbio.2005.08.010>
- Rombout, J. H. W. M., Abelli, L., Picchiatti, S., Scapigliati, G., & Kiron, V. (2013). Teleost intestinal immunology. *Fish & Shellfish Immunology*, 31, 616-26.
- Rombout, J. H. W. M., Lamers, C. H. J., Helfrich, M. H., Dekker, A., & Taverne-Thiele J. J. (1985). Uptake and transport of ferritin in the epithelium of carp (*Cyprinus Carpio* L.) and the possible immunological implications. *Cell & Tissue Research*, 239, 519-30. DIO: 10.1016/0309-1651(85)90003-7
- Rombout, J. H. W. M., Taverne N., van de Kamp, M., & Taverne-Thiele, A. J. (1993). Differences in mucus and serum immunoglobulin of carp (*Cyprinus carpio* L.). *Developmental & Comparative Immunology*, 17, 309- 317.



- Roslindawani, M N R, Syafiqah, A S, Jesse, F F A, Effendy, A W, & Zamri-Saad, M. (2016). Recombinant caseous lymphadenitis vaccine with palm oil as adjuvant enhances the humoral and cell-mediated immune responses in rat model. *Journal of Animal Health & Production*, 4(1), 25. <https://doi.org/10.1080/00102207908946886>
- Roy, B., Singh, R., Kumar, S., & Rai, U. (2008). Diurnal variation in phagocytic activity of splenic phagocytes in freshwater teleost *Channa punctatus*: Melatonin and its signaling mechanism. *Journal of Endocrinology*, 199(3), 471–480. <https://doi.org/10.1677/JOE-08-0270>
- Rozas-Serri, M., Ildefonso, R., Peña, A., Enríquez, R., Barrientos, S., & Maldonado, L. (2017). Comparative pathogenesis of piscirickettsiosis in Atlantic salmon (*Salmo salar* L.) post-smolt experimentally challenged with LF-89-like and EM-90-like *Piscirickettsia salmonis* isolates. *Journal of Fish Diseases*, 1-22. DOI: 10.1111/jfd.12671
- Rozas-Serri, M., Peña, A., & Maldonado, L. (2019). Gene expression associated with immune response in Atlantic salmon head-kidney vaccinated with inactivated whole-cell bacterin of *Piscirickettsia salmonis* and pathogenic isolates. *Fish & Shellfish Immunology*, 93, 789-795. DOI: 10.1016/j.fsi.2019.08.031
- Ruangpan, L., Kitao, T., & Yoshida, T. (1986). Protective efficacy of *Aeromonas hydrophila* vaccines in Nile tilapia. *Veterinary Immunology & Immunopathology*, 12, 345-350.
- Sáez, M. I., Vizcaíno, A. J., Alarcón, F. J., & Martínez, T. F. (2018). Feed pellets containing chitosan nanoparticles as plasmid DNA oral delivery system for fish: In vivo assessment in gilthead sea bream (*Sparus aurata*) juveniles. *Fish & Shellfish Immunology*, 80, 458–466. <https://doi.org/10.1016/j.fsi.2018.05.055>
- Salinas, I. (2015). The mucosal immune system of teleost fish. *Biology*, 4(3), 525-539. DOI: 10.3390/biology4030525
- Salinas, I., Zhang, Y., & Sunyer, J. O. (2011). Mucosal immunoglobulins and B cells of teleost fish. *Developmental & Comparative Immunology*, 35(12), 1346-1365.
- Salonius, K., Siderakis, C., MacKinnon, A. M., & Griths, S. G. (2005). Use of *Arthrobacter davidanieli* as a live vaccine against *Renibacterium salmoninarum* and *Piscirickettsia salmonis* in salmonids. *Developmental Biology*, 121, 189-197.
- Sandeep, M., Sun, F. E. I., Liu, F., Li, J., David, P. B. A. L., & Yue, G. E. N. H. U. A. (2012). Novel polymorphic microsatellites from Florida red tilapia and cross-species amplification in Mozambique and Nile tilapia. *Journal of Genetics*, 91, 97-99.
- Sato, A., & Okamoto, N. (2010). Induction of virus-specific cell-mediated cytotoxic responses of isogenic ginbuna crucian carp, after oral immunization with inactivated virus. *Fish & Shellfish Immunology*, 29(3), 414-21.

- Savan, R., Aman, A., Sato, K., Yamaguchi, R., & Sakai, M. (2005). Discovery of a new class of immunoglobulin heavy chain from fugu. *European Journal of Immunology*, 35(11), 3320-3331. DOI: 10.1002/eji.200535248
- Saxena, V., Lienesch, D. W., Zhou, M., Bommireddy, R., Azhar, M., Doetschman, T., & Singh, R. R. (2008). Dual roles of immunoregulatory cytokine TGF-beta in the pathogenesis of autoimmunity-mediated organ damage. *Journal of Immunology*, 180(3), 1903-12.
- Scapigliati, G. (2013). Functional aspects of fish lymphocytes. *Developmental & Comparative Immunology*, 41(2), 200-208.
- Scapigliati, G., Fausto, A. M., & Picchiatti, S. (2018). Fish lymphocytes: An evolutionary equivalent of mammalian innate-like lymphocytes? *Frontiers in Immunology*, 9, 1-8. <https://doi.org/10.3389/fimmu.2018.00971>
- Secombes, C. J., & Ellis, A. E. (2012). The immunology of teleosts. In R. Roberts (Ed.), *Fish Pathology (4th ed., pp. 144-166)*. Oxford, UK: Wiley-Blackwell.
- Sepúlveda, D. & Lorenzen, N. (2016). Can VHS virus bypass the protective immunity induced by DNA vaccination in rainbow trout. *PLoS ONE*, 11 (4), 153306.
- Sfanos, K., Harmody, D., Dang, P., Ledger, A., Pomponi, S., McCarthy, P., & Lopez, J. (2005). A molecular systematic survey of cultured microbial associates of deep-water marine invertebrates. *Systematic & Applied Microbiology*, 28, 242-264. DOI: 10.1016/j.syapm.2004.12.002
- Shahi, N., Mallik, S. K., Sahoo, M., & Das, P. (2013). Characteristics and Pathogenicity of a Virulent *Aeromonas hydrophila* Associated with Ulcerative Syndrome in Farmed Rainbow Trout, *Oncorhynchus mykiss* (Walbaum). *The Israeli Journal of Aquaculture*, 65(1), 926-936.
- Shahin, K., Shinn, A. P., Metselaar, M., Ramirez-Paredes, J. G., Monaghan, S. J., Thompson, K. D., Hoarea, R., & Adams, A. (2019). Efficacy of an inactivated whole-cell injection vaccine for Nile tilapia, *Oreochromis niloticus* (L), against multiple isolates of *Francisella noatunensis* subsp. *orientalis* from diverse geographical regions. *Fish & Shellfish Immunology*, 89, 217-227. DOI: 10.1016/j.fsi.2019.03.071.
- Shamsuddin, A. F. (2012). Palm oil adjuvant for use in veterinary vaccine. Universiti Kebangsaan Malaysia. Retrieved August 14, 2017 from [www.ukm.my/pik/11\\_175\\_palm-oil-adjuvant-for-use-in-veterinary-vaccines/](http://www.ukm.my/pik/11_175_palm-oil-adjuvant-for-use-in-veterinary-vaccines/)
- Sheehan, B., Lee, Y. S., Wong, E. S., Chan, J., Labrie, L., Komar, C., Wendover, N., & Grisez, L. (2009). Aquavac Strep Sa: Una novedosa vacuna para el control de las infecciones causadas por *Streptococcus agalactiae* Biotipo 2 en tilapia de granja. *En memorias: Manejo de Streptococcus en peces de aguas cálidas*, 21-26
- Shelby, R. A., Klesius, P. H., Shoemaker, C. A., & Evans, J. J. (2002). Passive immunization of tilapia, *Oreochromis niloticus* (L.), with anti-*Streptococcus iniae* whole sera. *Journal of Fish Diseases*, 25, 1-6. DOI: 10.1046/j.1365-

- Shelton, W. L. (2002). Tilapia culture in the 21st century. In: Proceedings of the International Forum on Tilapia Farming in the 21st Century (Tilapia Forum 2002). In R. D. I. Guerrero & M.R. Guerrero-Del Castillo. (Eds) (pp. 1-20).
- Sheng, X., Chai, B., Wang, Z., Tang, X., Xing, J., & Zhan, W. (2019). Polymeric immunoglobulin receptor and mucosal IgM responses elicited by immersion and injection vaccination with inactivated *Vibrio anguillarum* in flounder (*Paralichthys olivaceus*). *Aquaculture*, 505, 1-11. DOI: 10.1016/j.aquaculture.2019.02.045
- Sheng, X., Liu, M., Liu, H., Tang, X., Xing, J., & Zhan, W. (2018). Identification of immunogenic proteins and evaluation of recombinant PDHA1 and GAPDH as potential vaccine candidates against *Streptococcus iniae* infection in flounder (*Paralichthys olivaceus*). *PloS One*, 1–14.
- Shishido, S. N., Varahan, S., Yuan, K., Li, X., & Fleming, S. D. (2012). Humoral innate immune response and disease. *Clinical Immunology*, 144(2), 142–158. <https://doi.org/10.1016/j.clim.2012.06.002>
- Shoemaker, C. A., Klesius, P. H., Drennan, J. D., & Evans, J. J. (2011). Efficacy of a modified live *Flavobacterium columnare* vaccine in fish. *Fish & Shellfish Immunology*, 30(1), 304-308. DOI: 10.1016/j.fsi.2010.11.001
- Shoemaker, C. A., LaFrentz, B. R., & Klesius, P. H. (2012). Bivalent vaccination of sex reversed hybrid tilapia against *Streptococcus iniae* and *Vibrio vulnificus*. *Aquaculture*, 354-355, 45-49. DOI: 10.1016/j.aquaculture.2012.04.033
- Shoemaker, C. A., Lim, C., Yildirim-Aksoy, M., Welker, T. L., & Klesius, P. (2006). Growth response and acquired resistance of Nile tilapia, *Oreochromis niloticus* (L.) that survived *Streptococcus iniae* infection. *Aquaculture Research*, 37, 1238-1245.
- Shoemaker, C. A., Mohammed, H. H., Bader, T. J., Peatman, E., & Beck, B. H. (2018). Immersion vaccination with an inactivated virulent *Aeromonas hydrophila* bacterin protects hybrid catfish (*Ictalurus punctatus* X *Ictalurus furcatus*) from motile *Aeromonas* septicemia. *Fish & Shellfish Immunology*, 82, 239-242. DOI: 10.1016/j.fsi.2018.08.040
- Shoemaker, C. A., Vandenberg, G. W., Désormeaux, A., Klesius, P. H., & Evans, J. J. (2006). Efficacy of a *Streptococcus iniae* modified bacterin delivered using Oralject TM technology in Nile tilapia (*Oreochromis niloticus*), *Aquaculture*, 255, 151-156. DOI: 10.1016/j.aquaculture.2005.12.016
- Silva, B. C., Martins, M. L., Jatobá, A., Buglione Neto, C. C., Vieira, F. N., Pereira, G. V., Jerônimo, G. T., Seiffert W. Q., & Mourião J. L. P. (2009). Hematological and immunological responses of Nile tilapia after polyvalent vaccine administration by different routes. *Pesquisa Veterinária Brasileira*, 29(11), 874-880. DOI: 10.1590/s0100-736x2009001100002

- Sirimanapong, W., Thompson, K. D., Kledmanee, K., Thaijongrak, P., Collet, B., Ooi, E. L., & Adams, A. (2014). Optimisation and standardisation of functional immune assays for striped catfish (*Pangasianodon hypophthalmus*) to compare their immune response to live and heat killed *Aeromonas hydrophila* as models of infection and vaccination. *Fish & Shellfish Immunology*, *40*(2), 374–383. <https://doi.org/10.1016/j.fsi.2014.07.021>
- Siriypagouder, P., Shankar, K. M., K. B. T., & Patil, R. & Byadgi, O. V. (2014). Evaluation of biofilm of *Aeromonas hydrophila* for oral vaccination of *Channa striatus*. *Fish & Shellfish Immunology*, *41*(2), 581–585. <https://doi.org/10.1016/j.fsi.2014.09.021>
- Siti-Zahrah, A., Padilah, B., Azila, A., Rimatulhana, R., & Shahidan., H. (2008). Multiple streptococcal species infection in cage-cultured Red Tilapia but showing similar clinical signs. Pages 313–320 in M. G. Bondad-Reantaso, C. V. Mohan, M. Crumlish, and R. P. Subasinghe, editors. Diseases in Asian aquaculture VI. Asian Fisheries Society, Fish Health Section, Manila.
- Slotved, H. C., Kong, F., Lambertsen, L., Sauer, S., Gilbert, G. L. (2007). Serotype IX, a proposed new *Streptococcus agalactiae* serotype. *Journal of clinical microbiology*, *45*, 2929–2936.
- Smith, N. C., Rise, M. L., & Christian, S. L. (2019). A Comparison of the Innate and Adaptive Immune Systems in Cartilaginous Fish, Ray-Finned Fish, and Lobe-Finned Fish. *Frontiers in Immunology*, *10*. <https://doi.org/10.3389/fimmu.2019.02292>
- Soller, F., Roy, L. A., & Davis, D. A. (2019). Replacement of fish oil in plant-based diets for Pacific white shrimp, *Litopenaeus vannamei*, by stearine fish oil and palm oil. *Journal of the World Aquaculture Society*, *50*(1), 186–203. <https://doi.org/10.1111/jwas.12571>
- Somamoto, T., & Nakanishi, T. (2020). Mucosal delivery of fish vaccines: Local and systemic immunity following mucosal immunisations. *Fish and Shellfish Immunology*, *99*, 199–207. <https://doi.org/10.1016/j.fsi.2020.01.005>
- Sommerset, I., Skern, R., Biering, E., Bleie H., Fiksdal I. U., Grove S., & Nerland A. H. (2005). Protection against Atlantic halibut nodavirus in turbot is induced by recombinant capsid protein vaccination but not following DNA vaccination. *Fish & Shellfish Immunology*, *18*, 13–29.
- Song, M., Kang, Y., Zhang, D., Chen, L., Bi, J., Zhang, H., & Shan, X. (2018). Fish and Shell fish Immunology Immunogenicity of extracellular products from an inactivated vaccine against *Aeromonas veronii* TH0426 in koi, *Cyprinus carpio*. *Fish & Shellfish Immunology*, *81*, 176–181. <https://doi.org/10.1016/j.fsi.2018.07.004>
- SongLin, G., PanPan, L., JianJun, F., JinPing, Z., Peng, L., & LiHua, D. (2015). A novel recombinant bivalent outer membrane protein of *Vibrio vulnificus* and *Aeromonas hydrophila* as a vaccine antigen of American eel (*Anguilla rostrata*). *Fish & Shellfish Immunology*, *43*, 477–84.

- Sotomayor-Gerding, D., Troncoso, J. M., Pino, A., Almendras, F., & Diaz, M. R. (2020). Assessing the immune response of atlantic salmon (*Salmo salar*) after the oral intake of alginate-encapsulated piscirickettsia salmonis antigens. *Vaccines*, 8(3), 1–15. <https://doi.org/10.3390/vaccines8030450>
- Srinivasan, R., Karaoz, U., Volegova, M., MacKichan, J., Kato-Maeda, M., Miller, S., Nadarajan, R., Brodie, E. L., & Lynch, S. V. (2015). Use of 16S rRNA gene for identification of a broad range of clinically relevant bacterial pathogens. *PLoS One*, 10, 1-22. DOI: 10.1371/journal.pone.0117617
- Stone, H. D. (1997). Newcastle disease oil emulsion vaccines prepared with animal, vegetable and synthetic oils. *Avian Diseases*, 41, 591-597.
- Stosik, M., Tokarz-Deptuła, B., & Deptuła, W. (2018). Specific humoral immunity in Osteichthyes. *Central European Journal of Immunology*, 43(3), 335–340. <https://doi.org/10.5114/ceji.2018.80054>
- Stosik, M., Tokarz-Deptuła, B., & Deptuła, W. (2020). Major histocompatibility complex in Osteichthyes. *Journal of Veterinary Research (Poland)*, 64(1), 127–136. <https://doi.org/10.2478/jvetres-2020-0025>
- Stratev, D., & Odeyemi, O. A. (2016a). Antimicrobial resistance of *Aeromonas hydrophila* isolated from different food sources: a mini-review. *Journal of Infection & Public Health*, 9, 535-544. DOI: 10.1016/j.jiph.2015.10.006
- Stratev, D., & Odeyemi, O. A. (2016b). An overview of motile *Aeromonas septicaemia* management. *Aquaculture International*, 25(3), 1095-1105. DOI: 10.1007/s10499-016-0100-3
- Suanyuk, N., & Itsaro, A. (2011). Efficacy of inactivated *Streptococcus iniae* vaccine and protective effect of  $\beta$ -(1,3/1,6)-glucan on the effectiveness of vaccine in red tilapia *Oreochromis niloticus* x *O. mossambicus*. *Songklanakarin Journal of Science & Technology*, 33(2), 143-149.
- Sudhagar, A., Khan, F. & Prabu, L. (2016). Fish vaccination: A health management tool for aquaculture (<http://aquafind.com/articles/Vaccination.php>).
- Sugiani, D., Sukenda, S., Harris, E., & Lusiastuti, A. M. (2012). Haemato responses and histopathology of tilapia (*Oreochromis niloticus*) to co-infection *Streptococcus agalactiae* and *Aeromonas hydrophila*. *Jurnal riset akuakultur*, 7, 85-91. DOI: 10.15578/jra.7.1.2012.85-91
- Sukenda, S., Carman, O., Rahman, R., Hidayatullah, D., & Yumaidawati, N. S. (2017b). Vaccination in Nile tilapia broodstock with whole cell vaccine and disease resistance in its fry against *Aeromonas hydrophila*. *Jurnal Akuakultur Indonesia*, 16, 268-276. DOI: 10.19027/jai.16.2.268-276
- Sukenda, S., Sumiati, T., Nuryati, S., Lusiastuti, A. M., & Hidayatullah, D. (2017a). Specific immune response kinetics and mortality patterns of Tilapia *Oreochromis niloticus* on post-cocktail vaccination period against the infection of *Aeromonas hydrophila* and *Streptococcus agalactiae*. *Omni-Akuatika*, 13, 6-15.

- Sumiati, T., Sukenda, S., Nuryati, S., & Lusiastuti, A. M. (2015). Development of ELISA method to detect specific immune response in Nile tilapia (*O. niloticus*) vaccinated against *A. hydrophila* and *S. agalactiae*. *Jurnal Riset Akuakultur*, 10, 243-250. DOI: 10.15578/jra.10.2.2015.243-250
- Sun, X., Jin, P., Liu, Q., Wang, Q., Zhang, Y., & Liu, X. (2020). Fish and Shell fish Immunology A CpG-riched plasmid as vaccine adjuvant reduce antigen dose of an inactivated *Vibrio anguillarum* vaccine in turbot (*Scophthalmus maximus* L.). *Fish and Shellfish Immunology*, 98, 312–317. <https://doi.org/10.1016/j.fsi.2020.01.03>
- Suwannasang, A., Suanyuk, N., Issaro, A., Phromkunthong, W., Tantikitti, C., Itami, T., & Yoshida, T. (2017). Growth, immune responses and protection of Nile tilapia *Oreochromis niloticus* immunized with formalin-killed *Streptococcus agalactiae* serotype Ia and III vaccines. *Songklanakarin Journal of Science & Technology*, 39(4).
- Swain, P., Behura, A., Dash, S., & Nayak, S. K. (2007). Serum antibody response of Indian major carp, *Labeo rohita* to three species of pathogenic bacteria; *Aeromonas hydrophila*, *Edwardsiella tarda* and *Pseudomonas fluorescens*. *Veterinary Immunology & Immunopathology*, 117(1-2), 137-141. DOI: 10.1016/j.vetimm.2007.02.010
- Swann, L., & White, M. R. (1991). Diagnosis and Treatment of *Aeromonas hydrophila* in Fishes. Aquaculture Extension. Available at: <http://nsgl.gso.uri.edu/ilin/iling91003.pdf> (Accessed: 01 June 2016).
- Syahputra, K., Kania, P. W., Al-Jubury, A., Marnis, H., Cahyo Setyawan, A., & Buchmann, K. (2019). Differential immune gene response in gills, skin, and spleen of rainbow trout *Oncorhynchus mykiss* infected by *Ichthyophthirius multifiliis*. *PLoS ONE*, 14(6), 1–14. <https://doi.org/10.1371/journal.pone.0218630>
- Syuhada R., Zamri-Saad M., Ina-Salwany M. Y., Mustafa M., Nasruddin N. N., Desa M. N. M., Nordin S. A., Barkham T., & Amal, M. N. A. (2020). Molecular characterization and pathogenicity of *Streptococcus agalactiae* serotypes Ia ST7 and III ST283 isolated from cultured red hybrid tilapia in Malaysia. *Aquaculture*, 515, 734543. <https://doi.org/10.1016/j.aquaculture.2019.734543>
- Tacchi, L., Bron, J. E., Taggart, J. B., Secombes, C. J., Adler, M. A., Takle, H., & Martin, S. A. M. (2011). Multiple tissue transcriptomic responses to *Piscirickettsia salmonis* in Atlantic salmon (*Salmo salar*) Multiple tissue transcriptomic responses to *Piscirickettsia salmonis* in Atlantic. *Physiological Genomics*, 43, 1241-1254. DOI: 10.1152/physiolgenomics.00086.2011
- Tafalla, C., Bøgwald, J., & Dalmo, R. A. (2013). Adjuvants and immunostimulants in fish vaccines: Current knowledge and future perspectives. *Fish & Shellfish Immunology*, 35, 1740-1750. DIO: 10.1016/j.fsi.2013.02.029
- Tafalla, C., Bøgwald, J., Dalmo, R. A., Munang'andu, H. M., & Evensen, Ø. (2014). Adjuvants in Fish Vaccines. *Fish Vaccination*, 68–84. <https://doi.org/10.1002/9781118806913.ch7>

- Tantawy, M. M. E., & Ayoub, H. F. (2016). Efficiency of oral *Aeromonas hydrophila* vaccine and tumeric powder mixture on immune response of Nile tilapia (*Oreochromis niloticus*). *Egyptian Journal of Aquatic Research*, 6(1), 47-66.
- Thompson, J. D., Higgins, D. G., & Gibson, T. J. (1994). CLUSTAL W: Improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Research*, 22, 4673-80.
- Thompson, K. (2017). Immunology: Improvement of innate and adaptive immunity. In G. Jeney (Ed.), *Fish diseases: prevention & control strategies* (1st ed., pp. 3-14). London, UK: Elsevier.
- Tinsley, J. W., Lyndon, A. R., & Austin, B. (2011). Antigenic and cross-protection studies of biotype 1 and biotype 2 isolates of *Yersinia ruckeri* in rainbow trout, *Oncorhynchus mykiss* (Walbaum). *Journal of Applied Microbiology*, 111(1), 8–16. <https://doi.org/10.1111/j.1365-2672.2011.05020.x>
- Tizard, I. (2012). *Veterinary Immunology* (9th ed.). London, UK: WB Saunders Company.
- Uribe, C., Folch, H., Enriquez, R., & Moran, G. (2011). Innate and adaptive immunity in teleost fish: A review. *Veterinari Medicina*, 56(10), 486-503. DOI: 10.17221/3294-VETMED
- Tobar, J. A., Jerez, S., Caruffo, M., Bravo, C., Contreras, F., Bucarey, S. A., & Harel, M. (2011). Oral vaccination of Atlantic salmon (*Salmo salar*) against salmonid rickettsial septicaemia. *Vaccine*, 29, 2336-2340.
- Toranzo, A. E., Romalde, J. L., Magariños, B., & Barja, J. L. (2009). Present and future of aquaculture vaccines against fish bacterial diseases. *Options Méditerranéennes: Série A*, 86(86), 155-176.
- Tort, L., Balasch, J., & Mackenzie, S. (2003). Fish immune system. A crossroads between innate and adaptive responses. *Immunología*, 22, 277-86.
- Trichet, V. V., Santigosa, E., Cochin, E., & Gabaudan, J. (2015). The Effect of Vitamin C on Fish Health. *Dietary Nutrients, Additives & Fish Health*, 151-171.
- Tu, F. P., Chu, W. H., Zhuang, X. Y., & Lu, C. P. (2010). Effect of oral immunization with *Aeromonas hydrophila* ghosts on protection against experimental fish infection. *Letters in Applied Microbiology*, 50(1), 13-17. DOI: 10.1111/j.1472-765X.2009.02746.x
- Tu, X., Qi, X., Huang, A., Ling, F., & Wang, G. (2019). Cytokine gene expression profiles in goldfish (*Carassius auratus*) during *Gyrodactylus kobayashii* infection. *Fish & Shellfish Immunology*, 86, 116–124. <https://doi.org/10.1016/j.fsi.2018.11.035>
- Tukmechi, A., Rahmati Andani, H. R., Manaffar, R., & Sheikhzadeh, N. (2011). Dietary administration of beta-mercapto-ethanol treated *Saccharomyces cerevisiae* enhanced the growth, innate immune response and disease resistance of the rainbow trout, *Oncorhynchus mykiss*. *Fish & Shellfish Immunology*, 30(3), 923–

928. <https://doi.org/10.1016/j.fsi.2011.01.016>

- Uribe, C., Folch, H., Enriquez, R., & Moran, G. (2011). Innate and adaptive immunity in teleost fish: a review. *Veterinarni Medicina*, 56(10), 486-503.
- Vásquez-Machado, G., Barato-Gómez, P., & Iregui-Castro, C. (2019). Morphological characterization of the adherence and invasion of *Streptococcus agalactiae* to the intestinal mucosa of tilapia *Oreochromis* sp.: An in vitro model. *Journal of Fish Diseases*, 42(9), 1223-1231. DOI: 10.1111/jfd.13042
- Velázquez, J., Acosta, J., Lugo, J. M., Reyes, E., Herrera, F., González, O., Morales, A., Carpio, Y., & Estrada, M. P. (2018). Discovery of immunoglobulin T in Nile tilapia (*Oreochromis niloticus*): A potential molecular marker to understand mucosal immunity in this species. *Developmental & Comparative Immunology*, 88, 124-136. <https://doi.org/10.1016/j.dci.2018.07.013>
- Verner-Jeffreys, D. W., Wallis, T. J., Cejas C., Ryder, D., Haydon, D. J., Domazoro, J. F., Field, T. R., Adjei-Boteng, D., Wood, G., Bean, T., & Feist, S. W. (2017). *Streptococcus agalactiae* Multilocus sequence type 261 is associated with mortalities in the emerging Ghanaian tilapia industry. *Journal of Fish Diseases*, 41(1), 175-179. DOI: 10.1111/jfd.12681
- Vigneulle, M., & Baudin-Laurencin, F. (1991). Uptake of *Vibrio anguillarum* bacterin in the posterior intestine of rainbow trout *Oncorhynchus mykiss*, sea bass *Dicentrarchus labrax* and turbot *Scophthalmus maximus* after oral administration or anal intubation. *Diseases of Aquatic Organisms*, 11, 85-92.
- Villumsen, K. R., Neumann, L., Ohtani, M., Strøm, H. K., & Raida, M. K., (2014). Oral and anal vaccination confers full protection against enteric redmouth disease (ERM) in rainbow trout. *PLoS One*, 9, 93845. DOI: 10.1371/journal.pone.0093845.
- Waiho, K., Fazhan, H., Ishak, S. D., Kasan, N. A., Liew, H. J., Norainy, M. H., & Ikhwanuddin, M. (2020). Potential impacts of COVID-19 on the aquaculture sector of Malaysia and its coping strategies. *Aquaculture Reports*, 18. <https://doi.org/10.1016/j.aqrep.2020.100450>
- Wakabayashi, H., & Egusa, S. (1972). Preliminary experiments on environmental factors influencing the prevalence of columnaris disease. *Fish Pathology*, 7, 58-63.
- Walz, P. H., Edmondson, M. A., Riddell, K. P., Braden, T. D., Gard, J. A., Bayne, J., Joiner, K. S., Galik, P. K., Zuidhof, S., & Givens, M. D. (2015). Effect of vaccination with a multivalent modified-live viral vaccine on reproductive performance in synchronized beef heifers. *Theriogenology*, 83(5), 822-831.
- Walz, P. H., Givens, M. D., Rodning, S. P., Riddell, K. P., Brodersen, B. W., Scruggs, D., Short, T., & Grotelueschen, D. (2017). Evaluation of reproductive protection against bovine viral diarrhoea virus and bovine herpesvirus-1 afforded by annual revaccination with modified-live viral or combination modified-live/killed viral vaccines after primary vaccination with modified-live viral vaccine. *Vaccine*, 35(7), 1046-1054.



- Wanasawaeng, W., Tawatsin, A., Sasipreeyajan, J., Poomvises, P., & Chansiripornchai, N. (2009). Development of inactivated Newcastle disease vaccine using palm oil as an adjuvant. *Thai Journal of Veterinary Medicine*, 39(1), 9-16.
- Wang, A., Ran, C., Wang, Y., Zhang, Z., Ding, Q., Yang, Y., & Zhou, Z. (2019a). Use of probiotics in aquaculture of China- a review of the past decade. *Fish and Shellfish Immunology*, 86, 734–755. <https://doi.org/10.1016/j.fsi.2018.12.026>
- Wang, E., Liu, T., Wu, J., Wang, K., Chen, D., Geng, Y., Huang, X., Ouyang, P., Lai, W., & Ai, X. (2019b). Molecular characterization, phylogenetic analysis and adjuvant effect of channel catfish interleukin-1 $\beta$ s against *Streptococcus iniae*. *Fish & Shellfish Immunology*, 87, 155-165. DOI: 10.1016/j.fsi.2019.01.007
- Wang, E., Wang, J., Long, B., Wang, K., He, Y., Yang, Q., & Lai, W. (2016). Molecular cloning, expression and the adjuvant effects of interleukin-8 of channel catfish (*Ictalurus Punctatus*) against *Streptococcus iniae*. *Scientific Reports*, 6, 1-12. <https://doi.org/10.1038/srep29310>
- Wang, E., Wang, X., Wang, K., He, J., Zhu, L., He, Y., & Lai, W. (2018a). Preparation, characterization and evaluation of the immune effect of alginate/chitosan composite microspheres encapsulating recombinant protein of *Streptococcus iniae* designed for fish oral vaccination. *Fish & Shellfish Immunology*, 73, 262–271. <https://doi.org/10.1016/j.fsi.2017.12.034>
- Wang, J., Zou, L. L., & Li, A. X. (2014b). Construction of a *Streptococcus iniae* sortase A mutant and evaluation of its potential as an attenuated modified live vaccine in Nile tilapia (*Oreochromis niloticus*). *Fish & Shellfish Immunology*, 40(2), 392-398. DOI: 10.1016/j.fsi.2014.07.028
- Wang, Jing, Lu, D. Q., Jiang, B., Mo, X. B., Du, J. J., & Li, A. X. (2020a). Influence of temperature on the vaccine efficacy against *Streptococcus agalactiae* in Nile tilapia (*Oreochromis niloticus*). *Aquaculture*, 521, 734943. <https://doi.org/10.1016/j.aquaculture.2020.734943>
- Wang, Q., Fu, T., Li, X., Luo, Q., Huang, J., Sun, Y., & Wang, X. (2020b). Cross-immunity in Nile tilapia vaccinated with *Streptococcus agalactiae* and *Streptococcus iniae* vaccines. *Fish & Shellfish Immunology*, 97, 382–389. DOI: 10.1016/j.fsi.2019.12.021
- Wang, Q., Wang, X., Wang, X., Feng, R., Luo, Q., & Huang, J. (2018b). Generation of a novel *Streptococcus agalactiae* ghost vaccine and examination of its immunogenicity against virulent challenge in tilapia. *Fish & Shellfish Immunology*, 81, 49-56. DOI: 10.1016/j.fsi.2018.06.055
- Wang, R., Li, L., Huang, Y., Luo, F., Liang, W., Gan, X., Huang, T., Lei, A., Chen, M., & Chen, L. (2015). Comparative genome analysis identifies two large deletions in the genome of highly-passaged attenuated *Streptococcus agalactiae* strain YM001 compared to the parental pathogenic strain HN016. *BMC Genomics*, 16, 897. DOI: 10.1186/s12864-015-2026-y

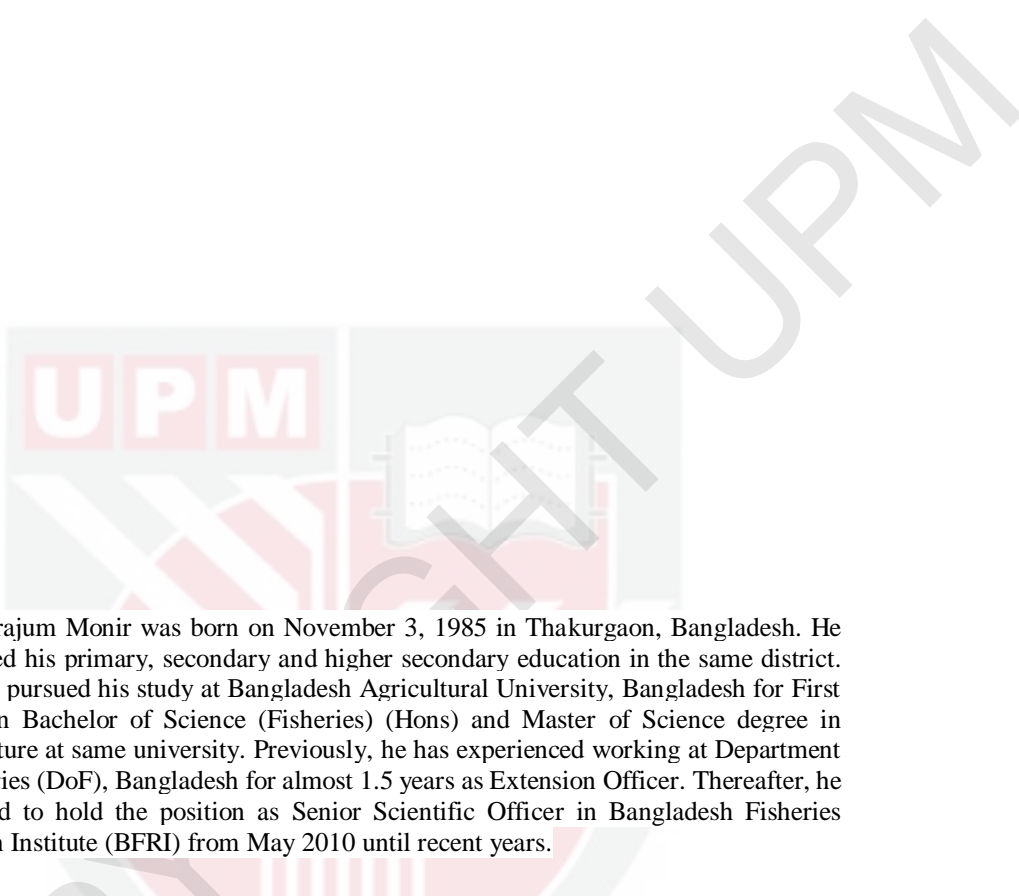
- Wang, S., Cheng, Z., Ling, X., Chu, X., Peng, X., & Li, H. (2018). Construction, immune protection and innate immune response of shuffled polyvalent ompAs vaccines. *Fish & Shellfish Immunology*, *74*, 325-331. DOI: 10.1016/j.fsi.2017.12.048
- Wang, T., & Secombes, C. J. (2013). The cytokine networks of adaptive immunity in fish. *Fish & Shellfish Immunology*, *35*(6), 1703-1718.
- Wang, Y. T., Huang, H. Y., Tsai, M. A., Wang, P. C., Jiang, B. H., & Chen, S. C. (2014a). Phosphoglycerate kinase enhanced immunity of the whole cell of *Streptococcus agalactiae* in tilapia, *Oreochromis niloticus*. *Fish & Shellfish Immunology*, *41*, 250-259.
- Watts, J. E. M., Schreier, H. J., Lanska, L., & Hale, M. S. (2017). The rising tide of antimicrobial resistance in aquaculture: sources, sinks and solutions. *Marine Drugs*, *15*, 1-16. DOI: 10.3390/md15060158
- Wei, G., Cai, S., Wu, Y., Ma, S., & Huang, Y. (2020). Immune effect of *Vibrio harveyi* formalin-killed cells vaccine combined with chitosan oligosaccharide and astragalus polysaccharides in ♀ *Epinephelus fuscoguttatus* × ♂ *Epinephelus lanceolatus*. *Fish & Shellfish Immunology*, *98*(1), 186-192. <https://doi.org/10.1016/j.fsi.2020.01.015>
- Wei, S., Huang, Y., Cai, J., Huang, X., Fu, J., & Qin, Q. (2012). Molecular cloning and characterization of c-type lysozyme gene in orange-spotted grouper, *Epinephelus coioides*. *Fish & Shellfish Immunology*, *33*, 186-196.
- World Health Organization (WHO). (2019). *Vaccines*. Retrieved <https://www.who.int/topics/vaccines/en/>
- Xu, C., Guo, T., Mutoloki, S., Haugland, Ø., & Evensen, Ø. (2012). Gene expression studies of host response to Salmonid alphavirus subtype 3 experimental infections in Atlantic salmon. *Veterinary Research*, *43*, 78.
- Xu, D. H., Shoemaker, C. A., & Klesius, P. H. (2007). Evaluation of the link between grodactylosis and *Streptococcus* of Nile tilapia. L. *Fish Disease*, *30*, 230-238.
- Xu, D. H., Zhang, D., Shoemaker, C., & Beck, B. (2019b). Immune response of channel catfish (*Ictalurus punctatus*) against *Ichthyophthirius multifiliis* post vaccination using DNA vaccines encoding immobilization antigens. *Fish & Shellfish Immunology*, *94*, 308-317.
- Xu, D., Moreira, G. S. A., Shoemaker, C. A., Zhang, D., & Benjamin, H. (2017). Expression of immune genes in systemic and mucosal immune tissues of channel catfish vaccinated with live theronts of *Ichthyophthirius multifiliis*. *Fish & Shellfish Immunology*. <https://doi.org/10.1016/j.fsi.2017.05.051>
- Xu, W., Jiao, C., Bao, P., Liu, Q., Wang, P., Zhang, R., & Zhang, Y. (2019a). Efficacy of Montanide™ ISA 763 A VG as aquatic adjuvant administered with an inactivated *Vibrio harveyi* vaccine in turbot (*Scophthalmus maximus* L.). *Fish & Shellfish Immunology*, *84*, 56-61. <https://doi.org/10.1016/j.fsi.2018.09.024>

- Yadav, S. K., Dash, P., Sahoo, P. K., Garg, L. C., & Dixit, A. (2018). Modulation of immune response and protective efficacy of recombinant outer-membrane protein F (rOmpF) of *Aeromonas hydrophila* in *Labeo rohita*. *Fish and Shellfish Immunology*. <https://doi.org/10.1016/j.fsi.2018.06.041>
- Yang, C. G., Wang, X. L., Tian, J., Liu, W., Wu, F., Jiang, M., & Wen, H. (2013b). Evaluation of reference genes for quantitative real-time RT-PCR analysis of gene expression in Nile tilapia (*Oreochromis niloticus*). *Gene*, 527(1), 183-192. DOI: 10.1016/j.gene.2013.06.013
- Yang, D., Liu, Q., Ni, C., Li, S., Wu, H., Wang, Q., & Zhang, Y. (2013a). Gene expression profiling in live attenuated edwardsiella tarda vaccine immunized and challenged zebrafish: Insights into the basic mechanisms of protection seen in immunized fish. *Developmental & Comparative Immunology*, 40(2), 132-141. <https://doi.org/10.1016/j.dci.2013.01.014>
- Yang, W., & Li, A. (2009). Isolation and characterization of *Streptococcus dysgalactiae* from diseased *Acipenser schrenckii*. *Aquaculture*, 294(1), 14-17.
- Yanong, R. P. E., & Floyd, F. R. (2002). Streptococcal infections of fish. Report from University of Florida. Series from the Department of Fisheries and Aquatic Sciences, Florida Cooperative Extension Service, Institute of Food and Agricultural Sciences, University of Florida.
- Yao, Y. Y., Chen, D. D., Cui, Z. W., Zhang, X. Y., Zhou, Y. Y., Guo, X., Li, A. H., & Zhang, Y. A. (2019). Oral vaccination of tilapia against *Streptococcus agalactiae* using *Bacillus subtilis* spores expressing Sip. *Fish & Shellfish Immunology*, 86, 999-1008. DOI: 10.1016/j.fsi.2018.12.060
- Ye, X., Li, J., Lu, M., Deng, G., Jiang, X., Tian, Y., & Jian, Q. (2011). Identification and molecular typing of *Streptococcus agalactiae* isolated from pond-cultured tilapia in China. *Fisheries Science*, 77(4), 623-632. <https://doi.org/10.1007/s12562-011-0365-4>
- Yonar, M. E., Mişer Yonar, S., İspir, Ü., & Ural, M. Ş. (2019). Effects of curcumin on haematological values, immunity, antioxidant status and resistance of rainbow trout (*Oncorhynchus mykiss*) against *Aeromonas salmonicida* subsp. achromogenes. *Fish & Shellfish Immunology*, 89. <https://doi.org/10.1016/j.fsi.2019.03.038>
- Yu, Y., Wang, Q., Huang, Z., Ding, L., & Xu, Z. (2020). Immunoglobulins, Mucosal Immunity and Vaccination in Teleost Fish. *Frontiers in Immunology*, 11, 1-14. <https://doi.org/10.3389/fimmu.2020.567941>
- Yus, E., Maier, T., Michalodimitrakis, K., Noort, V., Yamada, T., Chen, W. H., Wodke, J. A., Güell, M., Martínez, S., Bourgeois, R., Kühner, S., Raineri, E., Letunic, I., Kalinina, O. V., Rode, M., Herrmann, R., Gutiérrez-Gallego, R., Russell, R.B., Gavin, A.C., Bork, P., & Serrano, L. (2009). Impact of Genome Reduction on Bacterial Metabolism and Its Regulation. *Science*, 326, 1263-368.

- Zamri-Saad, M., Amal, M. N. A., Siti-Zahrah, A., & Zulkafli, A. R. (2014). Control and prevention of streptococcosis in cultured tilapia in malaysia: A review. *Pertanika Journal of Tropical Agricultural Science*, 37(4), 389-410.
- Zamri-Saad, M., Amal, M. N., & Siti-Zahrah, A. (2010). Pathological changes in red tilapias (*Oreochromis* spp.) naturally infected by *Streptococcus agalactiae*. *Journal of Comparative Pathology*, 2-3, 227-229.
- Zapata, A. G., Chiba, A., & Vara, A. (1996). Cells and tissues of the immune system of fish. In: *The Fish Immune System: Organism, Pathogen and Environment*. Fish Immunology Series. (eds. G. Iwama & T. Nakanishi,), New York, Academic Press, pp. 1-55.
- Zhang, C., Li, L. H., Wang, J., Zhao, Z., Li, J., Tu, X., & Zhu, B. (2018). Enhanced protective immunity against spring viremia of carp virus infection can be induced by recombinant subunit vaccine conjugated to single-walled carbon nanotubes. *Vaccine*, 36(42), 6334–6344. <https://doi.org/10.1016/j.vaccine.2018.08.003>
- Zhang, D. X., Kang, Y. H., Chen, L., Siddiqui, S. A., Wang, C. F., Qian, A. D., & Shan, X. F. (2017). Oral immunization with recombinant *Lactobacillus casei* expressing OmpAI confers protection against *Aeromonas veronii* challenge in common carp, *Cyprinus carpio*. *Fish & Shellfish Immunology*. <https://doi.org/10.1016/j.fsi.2017.10.043>
- Zhang, D., Li, A., Guo, Y., Zhang, Q., Chen, X., & Gong, X. (2013). Molecular characterization of *Streptococcus agalactiae* in diseased farmed tilapia in China. *Aquaculture*, 412-413, 64-69.
- Zhang, J., Hu, Y., Sun, Q., Li, X., & Sun, L. (2021). An inactivated bivalent vaccine effectively protects turbot (*Scophthalmus maximus*) against *Vibrio anguillarum* and *Vibrio harveyi* infection. *Aquaculture*, 544(June).
- Zhang, L., Ma, J., Fan, Y., Zhou, Y., Xu, J., Liu, W., & Zeng, L. (2016). Immune response and protection in gibel carp, *Carassius gibelio*, after vaccination with  $\beta$ -propiolactone inactivated cyprinid herpesvirus 2. *Fish & Shellfish Immunology*, 49, 344–350. <https://doi.org/10.1016/j.fsi.2016.01.003>
- Zhang, X. Y., Fan, H. P., Zhong, Q. F., Zhuo, Y. C., Lin, Y., & Zeng, Z. Z. (2008). Isolation, identification and pathogenicity of *Streptococcus agalactiae* from tilapia. *Journal of Fisheries of China*, 5, 772-779.
- Zhang, Y. A., Salinas, I., Li, J., Parra, D., Bjork, S., Xu, Z., LaPatra, S. E., Bartholomew, J., & Sunyer, J. O. (2010). IgT, a primitive immunoglobulin class specialized in mucosal immunity. *Nature Immunology*, 11(9), 827-835. DOI: 10.1038/ni.1913
- Zhang, Y.A., Salinas, I., & OriolSunyer, J. (2011). Recent findings on the structure and function of teleost IgT. *Fish & Shellfish Immunology*, 31, 627-634
- Zhang, Z., Chi, H., & Dalmo, R. A. (2019). Trained innate immunity of fish is a viable approach in larval aquaculture. *Frontiers in Immunology*, 10, 1–5. <https://doi.org/10.3389/fimmu.2019.00042>

- Zhu, B., Zhang, C., Yang, B., Guo, Z. R., Zheng, Y. Y., Gong, Y. M., & Wang, G. X. (2019). Preliminary screening and immunogenicity analysis of antigenic epitopes of spring viremia of carp virus. *Fish & Shellfish Immunology*, *94*, 833–841. <https://doi.org/10.1016/j.fsi.2019.09.063>
- Zhu, B., Zhang, C., Zhao, Z., & Wang, G. X. (2020). Targeted delivery of mannosylated nanoparticles improve prophylactic efficacy of immersion vaccine against fish viral disease. *Vaccines*, *8*(1). <https://doi.org/10.3390/vaccines8010087>
- Zhu, J., Fu, Q., Ao, Q., Tan, Y., Luo, Y., Jiang, H., Li, C., & Gan, X. (2017). Transcriptomic profiling analysis of tilapia (*Oreochromis niloticus*) following *Streptococcus agalactiae* challenge. *Fish & Shellfish Immunology*, *62*, 202-212. DOI: 10.1016/j.fsi.2017.01.023
- Zhu, L. Y., Nie, L., Zhu, G., Xiang, L. X., & Shao, J. Z. (2013). Advances in research of fish immune-relevant genes: a comparative overview of innate and adaptive immunity in teleosts. *Developmental & Comparative Immunology*, *39*, 39-62. DOI: 10.1016/j.dci.2012.04.001
- Zlotkin, A., Hershko, H., & Eldar, A. (1998). Possible Transmission of *Streptococcus iniae* from Wild Fish to Cultured Marine Fish. *Applied & Environmental Microbiology*, *64*, 4065-4067.
- Zou, J., & Secombes, C. J. (2016). The function of fish cytokines. *Biology*, *5* (2), 23. DOI: 10.3390/biology5020023.

## BIODATA OF STUDENT



Md. Shirajum Monir was born on November 3, 1985 in Thakurgaon, Bangladesh. He completed his primary, secondary and higher secondary education in the same district. Then, he pursued his study at Bangladesh Agricultural University, Bangladesh for First degree in Bachelor of Science (Fisheries) (Hons) and Master of Science degree in Aquaculture at same university. Previously, he has experienced working at Department of Fisheries (DoF), Bangladesh for almost 1.5 years as Extension Officer. Thereafter, he appointed to hold the position as Senior Scientific Officer in Bangladesh Fisheries Research Institute (BFRI) from May 2010 until recent years.

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## LIST OF PUBLICATIONS

- Monir, M. S., Yusoff, S. M., Zulperi, Z. M. Z., Hassim, H. A., Mohamad, A., Ngoo, M. S. M. H. & Ina-Salwany, M. Y. (2020). Haemato-immunological responses and effectiveness of feed-based bivalent vaccine against *Streptococcus iniae* and *Aeromonas hydrophila* infections in hybrid red tilapia (*Oreochromis mossambicus* × *O. niloticus*). *BMC Veterinary Research*, 16 (226), 1-14. DOI: 10.1186/s12917-020-02443-y
- Monir, M. S., Yusoff, S. M., Mohamad, A., & Ina-Salwany, M. Y. (2020). Vaccination of tilapia against motile aeromonas septicemia: a review. *Journal of Aquatic Animal Health*, 32(2), 65-76. DOI: 10.1002/aah.10099
- Monir, M. S., Yusoff, S. M., Zulperi, Z. M. Z., Hassim, H. A., Zamri-Saad, M., Amal, M.N.A., Salleh, A., Aslah Mohamad, A., Yie, L. J. & Ina-Salwany, M. Y. (2021). Immuno-protective efficiency of feed-based whole-cell inactivated bivalent vaccine against *Streptococcus* and *Aeromonas* infections in red hybrid tilapia (*Oreochromis niloticus* × *Oreochromis mossambicus*). *Fish & Shellfish Immunology*, 113, 162-175. <https://doi.org/10.1016/j.fsi.2021.04.006>
- Mohamad, A., Zamri-Saad, M., Amal, M. N. A., Al-Saari, N., Monir, M. S., Chin, Y. K., & Ina-Salwany, M. Y. (2021). Vaccine efficacy of a newly developed feed-based whole-cell polyvalent vaccine against vibriosis, streptococcosis and motile aeromonad septicemia in asian seabass, *Lates calcarifer*. *Vaccines*, 9(4). <https://doi.org/10.3390/vaccines9040368>
- Azzam-Sayuti, M., Ina-Salwany, M. Y., Zamri-Saad, M., Yusof, M. T., Annas, S., Najihah, M. Y., Liles, M. R., Monir, M. S., Zaidi, Z., & Amal, M. N. A. (2021). The prevalence, putative virulence genes and antibiotic resistance profiles of *Aeromonas* spp. isolated from cultured freshwater fishes in peninsular Malaysia. *Aquaculture*, 540, 736719. <https://doi.org/10.1016/j.aquaculture.2021.736719>
- Rahman M., Rahman, M., Monir, M. S., Haque, M., Siddique, M., Khasruzzaman, A., Rahman, M., & Islam, M. (2021). Isolation and molecular detection of *Streptococcus agalactiae* from popped eye disease of cultured Tilapia and Vietnamese koi fishes in Bangladesh. *Journal of Advanced Veterinary and Animal Research*, 8(1), 1. <https://doi.org/10.5455/javar.2021.h480>
- Azzam-Sayuti M., Ina-Salwany M.Y., Zamri-Saad M., Yusof M.T., Annas S., Najihah M.Y., Liles M.R., Monir M.S., Zaidi, Z., & Amal, M. N. A. (2021). Comparative pathogenicity of *Aeromonas* spp. in cultured red hybrid tilapia (*Oreochromis niloticus* × *O. mossambicus*). *Journal of Biology*.
- Monir, M. S., Yusoff, S. M., Zulperi, Z. M. Z., Hassim, H. A., Zamri-Saad, M., Amal, M.N.A., Salleh, A., Aslah Mohamad, A., Yie, L. J. & Ina-Salwany, M. Y. (2021). Feed-based bivalent vaccine induced expressions of immune-related genes in systemic and mucosal tissues of red hybrid tilapia (*Oreochromis niloticus* × *O. mossambicus*) against *Streptococcus iniae* and *Aeromonas hydrophila*. *PLoS ONE* (Submitted).

Monir, M. S., Yusoff, S. M., Zulperi, Z. M. Z., Hassim, H. A., Zamri-Saad, M., Amal, M.N.A., Salleh, A., Aslah Mohamad, A., Yie, L. J. & Ina-Salwany, M. Y. (2021). Effect of oral bivalent vaccine on immune response and immune gene profiling in vaccinated red tilapia (*Oreochromis* spp.) during infections with *Streptococcus iniae* and *Aeromonas hydrophila*. *Aquaculture Research* (Submitted).

Mohamad, A., Mursidi, F. A., Zamri-Saad, M., Amal, M. N. A., Annas, S., Al-Saari, N., Monir, M. S., Loqman, M., Hairudin, F., & Ina-Salwany, M. Y. (2021). Laboratory and field assessments of oral *Vibrio* vaccine provides good protection against vibriosis in cultured marine fishes. *Animals*.

Monir, M. S., Ahammed, T., Borty, S. C., Bagum, N., Islam, M. A. & Mahmud, Y. (2015). Pathogenesis of *Aeromonas* species in stinging catfish shing (*Heteropneustes fossilis*) of Bangladesh. *International Journal of Trends in Fisheries Research*. 4 (1): 7-13.

Monir, M. S., Bagum, N., Rahman, S., Doulah, M.A.U., Bhadra, A. & Borty S. C. (2015). Parasitic diseases and estimation of loss due to infestation of parasites in Indian major carp culture ponds in Bangladesh. *International Journal of Fisheries and Aquatic Studies*. 2(5): 118-122.

#### **Papers Presented in Seminars/Conferences**

Monir, M. S., Yusoff, S. M., Zulperi, Z. M. Z., Hassim, H. A. & Ina-salwany, M. Y. (2019). Efficacy of feed-based bivalent vaccine against streptococcosis and motile aeromonad septicemia in red tilapia, *Oreochromis* spp. The 9<sup>th</sup> International Fisheries Symposium, 18-21 November 2019, Kuala Lumpur, Malaysia. p. 471.

Monir, M. S., Ina-salwany, M. Y., Yusoff, S. M., Zulperi, Z. M. Z., & Hassim, H. A. (2020). Immunity assessment of feed based bivalent vaccine against streptococcosis and motile aeromonad septicemia in hybrid red tilapia, *Oreochromis niloticus* × *Oreochromis mossambicus*. International Conference on Marine Science & Aquaculture (ICOMSA), 9-10 December, 2020, Malaysia. OADM-6.

#### **Patent**

Composition and method for preparing and using oral vaccine for vibriosis in marine fish. Inventors: Ina-Salwany, M.Y., Zamri-Saad, M., Amal, M. N. A. Al-sarri, N. B., Mohamd, M. A. & Monir, M. S. Patent No:PA-2010/024, University Putra Malaysia, Malaysia and Date of Patent: Jan. 08, 2021.





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