

UNIVERSITI PUTRA MALAYSIA

EPIDEMIOLOGY OF Cucumber Mosaic Virus ON GINGER AND TURMERIC, AND ITS SUPPRESSION USING SILVER NANOPARTICLES

MUHAMMAD BUHARI

FP 2021 36



EPIDEMIOLOGY OF Cucumber Mosaic Virus ON GINGER AND TURMERIC, AND ITS SUPPRESSION USING SILVER NANOPARTICLES



By

MUHAMMAD BUHARI

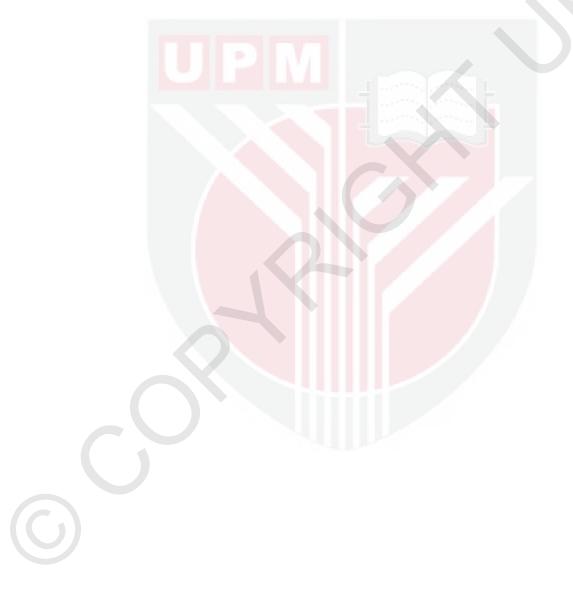
Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of Philosophy

January 2021

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

This thesis is dedicated to my parents and my teachers.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

EPIDEMIOLOGY OF Cucumber Mosaic Virus ON GINGER AND TURMERIC, AND ITS SUPPRESSION USING SILVER NANOPARTICLES

By

MUHAMMAD BUHARI

January 2021

Chairman : Associate Professor Ganesan a/l Vadamalai, PhD Faculty : Agriculture

Plant viruses have hampered vegetable crops production worldwide, causing huge economic losses. Viral diseases have also been reported in ginger with two viruses, Ginger mosaic virus and Ginger chlorotic fleck virus. Virus-like symptoms, such as mosaic, stripping and stunted growth pattern, were observed on the ginger and turmeric crops in the States of Selangor, Pahang and Perak of Malaysia and there was no prior study to unfold the pathogen inciting these symptoms. In principle, for disease management to be successful and feasible, proper and accurate identification of causal organisms must first be achieved. Consequently, a total of 60 ginger leaf samples, 20 from each State and 45 turmeric leaf samples, 15 from each State were collected. Enzyme-linked immunosorbent assay (ELISA) was first employed to index the virus, then nucleic acid extracted using cetyl trimethyl ammonium bromide (CTAB), after which reverse transcription polymerase chain reaction (RT-PCR) was conducted using CMV coat protein (CP) gene-specific primers and primers for GCFV. The expected amplicon of ~500 bp, which encodes 120 amino acids of the coding sequence of CMV CP gene was obtained, which was cloned and sequenced. In ginger plants, 23 % of the total samples were positive for CMV across the three States from the ELISA and RT-PCR assays, with 30 % of the samples in Selangor and 20 % of the samples gotten from each of Pahang and Perak States being detected as CMV-positive. Only one turmeric sample from Selangor was positive for CMV. The ginger and turmeric CMV isolates found in Malaysia, have 100% nucleotide similarity amongst themselves and shared 96% sequence homology with CMV cucumber isolate from Thailand (AJ810264) and tomato CMV isolate from China (KX525736) respectively. The ginger CMV and turmeric CMV isolates obtained from this study, were phylogenetically grouped into CMV subgroup I B. Electron microscopic analysis has confirmed the CMV isolates, when the virus was purified, negatively stained using uranyl acetate and viewed under high resolution electron microscope. GCFV was however not detected from the two crops in this study by



RT-PCR assay. Pathogenicity test was conducted on the ginger and turmeric host plants by mechanical inoculation of the CMV-positive samples; ginger (TM3 isolate) and turmeric (TMR isolate). The inoculated test plants (100 %) showed similar symptoms of mosaic, stunting and stripping as observed in the field and confirmed CMV positive by CP gene through RT-PCR assay. CMV was also detected in threemonth-old plants grown from CMV-infected rhizomes with 90 and 100 % infections in ginger and turmeric plants respectively. Host range studies conducted through a sap transmission technique showed nine plant species from six plant families as potential hosts for CMV isolated from ginger. The efficacy of silver nanoparticles (AgNPs) against CMV in ginger and turmeric plants was investigated by real time PCR. The virus concentration was significantly reduced by AgNPs (p<0.01) in the two plants from one-month post inoculation (MPI) up to 5 MPI. Thus, AgNPs has provided an avenue for the control of CMV infection in ginger and turmeric plants. The virus causing the symptoms of mosaic, stripping and stunting observed on ginger and turmeric plants, was unravelled, and identified as CMV, and its pathogenicity and host range were also conducted and reported in this study. Finally, silver nanoparticles were shown to suppress the CMV multiplication in ginger and turmeric plants.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

EPIDEMIOLOGI Virus Mosaic Cucumber (CMV) PADA HALIA DAN KUNYIT, DAN PENGAWALAN MENGGUNAKAN NANOPARTIKEL PERAK

Oleh

MUHAMMAD BUHARI

Januari 2021

Pengerusi : Profesor Madya Ganesan a/l Vadamalai, PhD Fakulti : Pertanian

Virus tumbuhan menjejaskan pengeluaran tanaman sayuran di seluruh dunia, menyebabkan kerugian ekonomi yang besar. Penyakit virus juga telah dilaporkan pada tanaman halia di mana, dua virus, Ginger mosaic virus dan Ginger chlorotic fleck virus (GCFV) didapati menjangkiti tanaman halia. Simptom penyakit virus, seperti mozek, penjaluran dan pertumbuhan yang terbantut, telah diperhatikan pada tanaman halia dan kunyit di Negeri Selangor, Pahang dan Perak di Malaysia dan tidak ada kajian sebelumnya untuk mengenalpasti patogen yang menyebabkan symptom tersebut. Pada prinsipnya, untuk pengurusan penyakit yang berjaya, pengenalpastian organisma penyebab yang betul dan tepat mesti dicapai terlebih dahulu. Sebanyak 60 daun tumbuhan halia, 20 dari setiap negeri dan 45 daun tumbuhan kunyit, 15 dari setiap negeri telah dikutip berdasarkan keterlihatan symptom penyakit virus. Uji imunosorben berkait enzim (ELISA) digunakan untuk mengindeks virus terlebih dahulu, kemudian asid nukleik diekstrak menggunakan cetly trimetil amonium bromida (CTAB), kemudian reverse transcription polymerase chain reaction (RT-PCR) menggunakan primer spsifik untuk gen kot protin CMV (CP) dan primer khusus untuk GCFV. Amplikon yang dijangakan ~ 500 bp, yang menyandikan 120 asid amino dari urutan kot gen CMV CP telah diperolehi, diklon dan diuraikan. Pada tanaman halia, 23% daripada jumlah sampel dari ketigatiga negeri adalah positif untuk CMV dari ujian ELISA dan RT-PCR, dimana 30% sampel dari Selangor dan 20% dari Negeri Pahang dan Perak masing-masing dikesan positif dengan CMV. Hanya satu sampel kunyit dari Selangor yang positif untuk CMV. Isolat CMV dari halia dan kunyit yang ditemui di Malaysia mempunyai 100% persamaan nukleotida di antara mereka dan berkongsi 96% homologi dengan isolat CMV timun dari Thailand (AJ810264) dan isolat CMV tomato dari China (KX525736) masing-masing. Analisis filogenetik menunjukkan isolat CMV halia dan CMV kunyit yang diperolehi dari kajian ini dikelompokkan ke dalam subkumpulan CMV IB. Analisis mikroskop elektron telah mengesahkan isolat CMV,

dimana partikel virus yang tulen diwarnai secara negatif menggunakan uranyl asetat dapat dilihat di bawah mikroskop elektron resolusi tinggi. Bagaimanapun GCFV tidak dikesan dari dua spesis tanaman dalam kajian ini dengan ujian RT-PCR. Ujian patogenisiti dilakukan pada tumbuhan halia dan kunyit dengan inokulasi mekanikal sampel positif CMV; halia (isolat TM3) dan kunyit (isolate TMR). Pokok yang diinokulasi (100%) menunjukkan simptom yang sama seperti yang dilihat di ladang dan disahkan CMV positif melalui idetifikasi gen CP melalui RT-PCR. CMV juga dikesan pada tanaman berusia tiga bulan yang tumbuh dari rimpang yang dijangkiti CMV dengan 90 dan 100% jangkitan pada tanaman halia dan kunyit masing-masing. Kajian perumah yang dilakukan melalui transmisi sap menunjukkan sembilan spesies tumbuhan dari enam keluarga tumbuhan sebagai perumah yang berpotensi untuk CMV yang diasingkan dari halia. Keberkesanan nanopartikel perak (AgNPs) terhadap CMV pada tanaman halia dan kunyit dikaji mengunakan *Real-Time* PCR. Kepekatan virus dikurangkan dengan signifikan oleh AgNPs (p <0,01) dalam keduadua tanaman dari satu bulan selepas inokulasi (BSI) hingga 5 BSI. Oleh itu, AgNPs boleh menjadi satu kaedah alternatif untuk mengawal jangkitan CMV pada tanaman halia dan kunyit. Virus yang menyebabkan simptom mozek, penjaluran dan pertumbuhan yang terbantut yang dilihat pada tanaman halia dan kunyit, telah dikenalpasti sebagai CMV, kajian patogenisiti dan potensi perumah telah dijalankan dan dilaporkan dalam kajian ini. Akhirnya, nanopartikel perak didapati menindas replikasi CMV dalam tanaman halia dan kunyit.

ACKNOWLEDGEMENTS

All praise is due to Almighty Allah, the Lord of the Universe. May His peace and blessings be upon the noble messenger, Muhammad (SAW), his household, companions and all that follow his footstep until the Last Day.

I would like to express my profound appreciation to my supervisor, Associate Professor Dr. Ganesan Vadamalai for his guidance, mentoring, patience, support and above all, for making my Malaysian life very comfortable. I have no words that can really express my mind in thanking you. My sincere gratitude goes to my cosupervisor, Dr. Kong Lih Ling for her invaluable support and advice during my study. In the same vein, my heartfelt appreciation goes to my co-supervisor, Dr. Lau Wei Hong for her insightful suggestions and comments that made my work a success. I thank my supervisory committee once again.

My special gratitude goes to the Nigerian Government for the scholarship through the Tertiary Education Trust Fund (TETFUND) and the Management of Ahmadu Bello University (ABU), Zaria for nominating me.

I would like to extend my warmest appreciation to Mr Mohamed Nazri Abdul Rahman, Mr. Johari Mohd Sarikat, Mrs Asmalina Abu Bakar, Mrs Mastura Hamit, Mr. Rafiuz Zaman Haroun (IBS), Mr Jamil Muhammad (Vet Medicine) for their technical assistance. I also thank my lab colleagues, Mr. Ahmad Son Turaki, Mr. Khoo, Mrs Najwa, Ms Atiqa and Mrs Nadika for helping one another.

Special prayers go to my parents and my teachers for their continuous motivation, prayers and bringing me up to the level I am today. May Almighty Allah reward them with Jannah. I would like to duly acknowledge the assistance, prayers and support of my brothers and sisters. You are the impetus behind my success. My special appreciation goes to my elder brother, Dr. Nasa'i Muhammad, for taking care of my family during my study abroad and his tireless support for me right from our childhood.

A big thank you to my family friends, the family of Malam Ahmad Ibrahim, through whom I attained many of my life accomplishments. May Allah reward them with Jannah.

The following people contributed in my research work: Dr. Ahmadu Tijjani, Dr. Usman, Dr. Ishaka Ibrahim, Dr. Bashir Tambuwal, Dr. Nafiu, Dr. Jamilu, Dr. Ibrahim Moi, Mr Osamah Bn Zaid, Dr Wael Sultan, Dr. Abdullahi Raji, Dr. Hayatu Aliyu, Dr. Ismai'l Muhammad, Dr Yong Farmatan, Dr. Bello Haliru, Dr Yusuf Oladosun, Dr. Umar (Baba Saraki), Dr. Badmus Kazim, Ms Priya Hamid and Ms

Vinailosni Amirthalingam. I say a big thank you and may Almighty Allah reward them all for making my dream a reality.

I would like to duly acknowledged our educational centres and their teachers, Darul Hadeethis Salafiyyah, Markazussalafiyyah, and Muslim Refresher Course Programme (MRCP) for all my life achievements. Special prayers to the late Malam Abubakar Gumi and Malam Albaniy Zaria for their enthusiastic and methodical teachings that motivated me to continue with my higher education up to this level. May their souls rest in peace.

I cannot finish my acknowledgements without mentioning the contribution of Profs Nafiu Abdu, M.D. Alegbejo, O. O. Banwo and B. D. Kashina for the academic foundation.

Finally, my wholehearted appreciation goes to my dearest wife, Hajiya Hauwa'u M. Sani and my children for their perseverance, motivation, care, patience and understanding during my long absence. May we continue to live together as prosperous family both here and the hereafter.

My list will be endless if I do not stop here. I thank my friends, my students, wellwishers and all those that in one way or the other help me in my life. This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Ganesan a/l Vadamalai, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Kong Lih Ling, PhD Research Officer Institute of Plantation Studies Universiti Putra Malaysia (Member)

Lau Wei Hong, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date: 06 May 2021

TABLE OF CONTENTS

Page

ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vii
DECLARATION	ix
LIST OF TABLES	XV
LIST OF FIGURES	xvii
LIST OF ABBREVIATIONS	xxi
CHAPTER	

 $\overline{()}$

1	INTE	RODUCTION	1
	1.1	Statement of problem	2
	1.2		2 3 3
	1.3	Objectives of the study	3
2	LITF	CRATURE REVIEW	4
	2.1	Vegetable crops	4
		2.1.1 Vegetable industry in Malaysia	4
		2.1.2 Viral diseases of vegetables	4
	2.2	Ginger origin, distribution and production worldwide	5
		2.2.1 Ginger production in Malaysia	5
	2.3	Uses and nutritional value of ginger	6
	2.4	Constraints to ginger production	6
		2.4.1 Pests and diseases of ginger	6
		2.4.1.1 Viral diseases of ginger	
	2.5	Turmeric origin, distribution and production worldwide	8 8
		2.5.1 Turmeric production in Malaysia	9
	2.6	Uses of turmeric	9
	2.7	Constraints to turmeric production	9
		2.7.1 Pests and diseases of turmeric	9
		2.7.1.1 Pests of turmeric	9
		2.7.1.2 Diseases of turmeric	10
	2.8	Cucumber mosaic virus (CMV)	10
		2.8.1 Genomic composition and particle structure	10
		2.8.2 Cucumber mosaic virus Grouping	11
		2.8.3 Symptoms and Pathogenicity of <i>Cucumber mosaic</i>	
		virus Infection	11
		2.8.4 Transmission of Cucumber mosaic virus	12
		2.8.5 Host range of Cucumber mosaic virus	12
		2.8.6 Economic importance of <i>Cucumber mosaic virus</i>	13
		2.8.7 Detection of Cucumber mosaic virus	13
		2.8.8 Management of Cucumber mosaic virus	14
	2.9	Ginger chlorotic mottle virus	14

xi

	2.10		echnologies in controlling plant viral diseases in the	e
		tropics	8	15
		2.10.1	Silver nanoparticles an antiviral agent as the lates means of control	st 16
				10
3	DET	ECTION	N AND CHARACTERIZATION OF VIRUSES	5
	INFE	CTING	GINGER AND TURMERIC USING ELISA	•
	ELE	CTRON	MICROSCOPY, PCR, CLONING ANI)
	SEQ	UENCIN	NG	18
	3.1	Introd	uction	18
	3.2	Materi	als and method	18
		3.2.1	Sample Collection	18
		3.2.2	Double Antibody Sandwich Enzyme-Linker	t
			Immunosorbent Assay (DAS-ELISA)	19
		3.2.3	Nucleic Acid Extraction	19
		3.2.4	Complimentary DNA (cDNA) Synthesis	20
		3.2.5	Polymerase Chain Reaction (PCR)	21
			3.2.5.1 PCR for CMV detection	21
			3.2.5.2 PCR for the detection of Ginge	r
			chlorotic fleck virus (GCFV)	21
		3.2.6	Agarose Gel Electrophoresis and visualization	21
		3.2.7	PCR products cloning	21
			3.2.7.1 Gel purification of PCR products	21
			3.2.7.2 Ligation of PCR product into TOPO-TA	ł
			vector	22
			3.2.7.3 Transformation	22
			3.2.7.4 Selection of recombinant colonies	22
			3.2.7.5 Minipreparation of plasmids	23
			3.2.7.6 Analysis of recombinant inserts	23
			Sequencing analysis	23
			Phylogenetic analysis	24
			Restriction Fragment Length polymorphism	25
		3.2.11	Electron microscopy	25
			3.2.11.1 Purification of virus particles	25
			3.2.11.2 Transmission electron microscopy	
			(TEM)	26
			3.2.11.3 Isolation of RNA from the pure virion	26
			3.2.11.4 Confirmation of the purified viru	
			identity by RT-PCR and sequencing	26
	3.3	Result		26
		3.3.1	Symptoms description and disease incidence	26
			3.3.1.1 Ginger	26
		222	3.3.1.2 Turmeric	29
		3.3.2	6	30
		3.3.3	<i></i>	31
			3.3.3.1 Detection of Cucumber mosaic virus	31
			3.3.3.2 Detection of Ginger chlorotic flech	
			virus	33
		3.3.4	Cloning and analysis of insert in <i>Eco</i> RI	34

		3.3.5 Sequence analysis	34
		3.3.6 Phylogenetic analysis	39
		3.3.7 Restriction fragment length poly (RFLP)	ymorphism 39
		3.3.8 Transmission electron microscopy us CMV ginger isolate	ing TM 3 40
		3.3.9 Confirmation of the pure virion identi PCR	
	3.4	Discussion	42
	3.5	Conclusion	45
4		DGENICITY, RHIZOME TRANSMISSI	
		RANGE STUDIES OF CUCUMBER MOSA	
	4.1	Introduction	46
	4.2	Materials and methods	47
		4.2.1 Sources of planting materials and le study	47
		4.2.2 Pathogenicity test	47
		4.2.3 Experimental design and statistical anal	
		4.2.4 Rhizome transmission of <i>Cucumber m</i>	osaic virus
		in ginger and turmeric	48
		4.2.5 Host range studies	48
		4.2.6 Nucleic acid extraction	49
		4.2.7 cDNA synthesis	49
		4.2.8 Polymerase chain reaction	49
		4.2.9 Agarose gel electrophoresis	49
			from the
		pathogenicity test plants	49
	4.3	Results	49
		4.3.1 Pathogenicity test	49
		4.3.1.1 Effects of CMV infection of	• 1
		development and plant height	
		4.3.1.2 RT-PCR for the confirmatio	
		pathogenicity in ginger and tu	
		4.3.1.3 Sequencing results	55
		4.3.2 Rhizome transmission of CMV in g	
		turmeric	55
		4.3.2.1 RT-PCR for the detection of	
		ginger and turmeric rhizomes	
		4.3.3 Host range	57
		4.3.3.1 Symptomatology	57
		4.3.3.2 RT-PCR for the detection of C	
		hosts range studies	61
	4.4	Discussion	62
	4.5	Conclusion	66

5		CACY OF SILVER NANOPARTICLES IN THE PRESSION OF CUCUMBER MOSAIC VIRUS	
		ECTION IN GINGER AND TURMERIC	67
	5.1	Introduction	67
	5.2	Materials and methods	68
		5.2.1 Experimental design and silver nanoparticles application	68
		5.2.2 RNA extraction	68
		5.2.2 NNA extraction 5.2.3 Measurement of RNA concentration and purity	68
		5.2.4 Primer design	69
		5.2.4 Primer sequence and its properties	69
		5.2.4.2 Temperature gradient for optimum	
		annealing of primers	69 70
		5.2.5 cDNA synthesis 5.2.6 Real-time PCR condition	70
		5.2.6 Real-time PCR condition 5.2.7 Generation of standard curve	70 70
		5.2.7 Generation of standard curve 5.2.8 Real time PCR for quantification of effect of silver	70
		nanoparticles on CMV in ginger and turmeric	70
		5.2.9 Data analysis	70
	5.3	Results	71
	5.5	5.3.1 Analysis of primer properties	71
		5.3.2 The standard curve	71
		5.3.3 Analysis of real time PCR for silver nanoparticles	/1
		against CMV in ginger and turmeric plants	73
	5.4	Discussion	77
	5.5	Conclusion	78
6	SUM	MARY, CONCLUSION AND RECOMMENDATIONS	
		FUTURE RESEARCH	79
	6.1	Summary	79
	6.2	Conclusion	80
	6.3	Recommendations for future studies	80
REF	ERENC	CES	81
	ENDIC		94
		OF STUDENT	144
LIST	T OF PU	JBLICATIONS	145

5

LIST OF TABLES

Table		Page
2.1	The world top twenty producers of ginger	5
3.1	Description of primers used for the detection of CMV and GCFV	20
3.2	Sequences of <i>Cucumber mosaic virus</i> used for phylogenetic analysis from GenBank and those starting with KU in accession and PV in the strain name, were adopted from Bald-Blume et al. (2017)	
3.3	The different symptom characteristics of ginger leaf samples collected from the three States	s 27
3.4	The different symptom characteristics of the turmeric leaf samples collected from the three States	s 29
3.5	ELISA screening across three Malaysian States for Cucumber mosaicvirus in ginger plants	r 31
3.6	ELISA screening across three Malaysian States for <i>Cucumber mosaid</i> <i>virus</i> in turmeric plants	c 31
3.7	RT-PCR assay for the detection of <i>Cucumber mosaic virus</i> in ginge collected from Perak, Selangor and Pahang States	r 32
3.8	RT-PCR assay for the detection of <i>Cucumber mosaic virus</i> in turmeric collected from Perak, Selangor and Pahang	n 33
3.9	Comparison of nucleotide sequences (nt) and amino acids (aa) o CMV CP coding sequence region with isolate TMR and other CMV subgroup members	
4.1	Fertilizer application and watering regime of plants during the hos range studies on weekly intervals	t 48
4.2	Effect of CMV inoculation on the height (cm) of ginger plants from 1 to four months after inoculation (MPI)	n 50
4.3	Effect of CMV inoculation on the height (cm) of turmeric plants from 1 to 4-month post inoculation (MPI)	n 54
4.4	Symptoms on different hosts, number infected plants and days to initial symptom appearance	58
5.1	Sequences used in the primer design	69
5.2	Primer properties for the real time qPCR	69

5.3	Overall performance of silver nanoparticles on CMV in ginger plants	73
5.4	Concentration of CMV in ginger plants treated with silver nanoparticles	75
5.5	Overall performance of silver nanoparticles against CMV in turmeric plants	75
5.6	Concentration of CMV in turmeric plants treated with silver nanoparticles	76
A 1	Symptoms description of ginger samples collected from Selangor State	94
A 2	Symptoms description of ginger samples collected from Pahang State	95
A 3	Symptoms description of ginger samples collected from Perak State	95
A 4	Symptoms description of turmeric samples collected from Selangor State	96
A 5	Symptoms description of turmeric samples collected from Pahang State	96
A 6	Symptoms description of turmeric samples collected from Perak State	97
A 7	ELISA screening of ginger plant samples from Selangor State	97
A 8	ELISA screening of ginger plant samples from Perak State	98
A 9	ELISA screening of ginger plant samples from Pahang State	98
A 10	ELISA screening of turmeric plant samples from Selangor State	99
A 11	ELISA screening of turmeric plant samples from Perak State	99
A 12	ELISA screening of turmeric plant samples from Pahang States	100

LIST OF FIGURES

Figur	°e	Page
2.1	Some pests and diseases of ginger plant A) A larva of shoot borer of ginger Source: cabi.org B) Adult stage of ginger shoot borer Source: plantwise.org C) Bacterial wilt of ginger caused by Ralstonia solanacearum Source: Plantvillage.psu.edu D) Burrowing nematode <i>Radopholus similis</i> on ginger Source: Plantvillage.psu.edu	
2.2	Genome organization of <i>Cucumber mosaic virus</i> showing the different RNAs segments with their corresponding nucleotide number (nt) and protein size (Roossinck, 2002)	
2.3	Cucumber mosaic virus symptoms on various food and cash crops A) Stunting on soyabean B) Necrotic streaks on tobacco C) mosaic symptom on wild pokeweed D) Necrotic symptoms on bell pepper fruit Source: A) and B) cabi.org C) and D) https://extension.psu.edu/ cucumber-mosaic-virus	
2.4	Mechanism of AgNP against pathogenic viruses	17
3.1	Symptoms of sampled ginger plants. A) Mosaic symptom developed on a sample at was 6-month-old in Perak B) Stripping symptom on a sample at 4-month-old in Pahang C) Chlorosis and vein banding on a sample at 2-month-old from Selangor D) Healthy ginger plant at 4- month-old. Arrows indicate the symptom position	
3.2	Minor CMV symptom and virus-like symptom on ginger plants. A) Mild mosaic on the tip of a ginger leaf from Selangor when the plant was 3-month-old. B) Virus-like symptom of leaf curling when the plant was 5-month-old.	
3.3	Turmeric plants collected from Selangor at five-month-old A) Mosaic and chlorotic symptoms, with the symptoms covering the entire foliage in the infected plant B) Healthy turmeric plant with normal leaf colouration	
3.4	RT-PCR product of CMV Coat protein gene of 500 bp separated using 2% (w/v) agarose gel electrophoresis 1; Turmeric CMV isolate from Selangor 2 - 7; Selangor CMV ginger isolates 8 – 11; Pahang CMV ginger isolates 12 - 15; Perak CMV ginger isolates 16; NTC 17; Positive control 18; Negative control M; 100bp DNA ladder	
3.5	PCR products fractionated on 2% (w/v) agarose gel showing no bands of expected size of 340 bp from <i>Sobemovirus</i> conserved replicase gene. Two samples represented each crop from the sampling States, 1-2; Selangor ginger samples, 3 and 4: Selangor turmeric samples, 5 and 6: Perak ginger samples. 7 and 8; Perak turmeric samples, 9 and	

	10; Pahang ginger samples, 11 and 12; Pahang turmeric samples. 13 and 14: No-template control (NTC) and M; 100 bp DNA ladder	33
3.6	Agarose gel 2% (w/v) with pCR 2.1^{TM} TOPO plasmid recombinant digested with <i>Eco</i> RI, having the expected insert ~500 bp and 3.9 kb plasmid. 1; Selangor turmeric isolate2-7; Selangor ginger isolates, 8 – 11; Pahang ginger isolates and 12 – 15; Perak ginger isolates M; 100 bp DNA ladder	34
3.7	A cross section of nucleotide alignment across the Malaysian CMV isolates from fourteen ginger denoted as BTs, TMs and FGs; and a turmeric tagged as TMR samples showing homogeneity in their nucleotide sequence	36
3.8	Comparison of Malaysian ginger CMV isolate, FG5, with other world CMV isolates nucleotide sequences showing a base substitution at 123th nucleotide from "C" to "T" as pointed with arrows	37
3.9	Alignment of amino acid sequence of Malaysian ginger CMV isolate, FG5, with other subgroup representative sequences from the coding sequence region of 3' end of CMV CP gene	38
3.10	Phylogenetic relationship of CMV isolated from ginger and turmeric with other CMV isolates inferred by Neighbour-Joining with 1000 bootstrap replicates	39
3.11	Restriction fragment length polymorphism with <i>Eco</i> RI of the CMV isolates for subgrouping of the virus fractionated using 2% agarose, showing no digestion of 500 bp CP gene. 1; Turmeric isolate, $2 - 7$; Selangor isolates FG 1 – FG 6; 8- 11; Pahang isolates BT 1 – BT 4; $12 - 15$; Perak isolates, TM 1 – TM 4	40
3.12	Sucrose density-gradient using $5 - 25\%$ (w/v) sucrose using isolate TM 3. The virus band was observed between 10 and 15% (w/v) sucrose density along with sucrose particles and other plant materials at the bottom of the tube	40
3.13	Electron micrograph of purified virion showing the icosahedral particles of average diameter of $10 - 12$ nm. The average of 100 particles was measured to confirm the size range	41
3.14	Particles distribution in the purified virions under transmission electron microscope view	41
3.15	RT-PCR of the pure virion after purification replicated three times using 2% (w/v) agarose gel, having the expected amplicon size of ~ 500 bp. M; 100 bp DNA ladder 1 – 3; pure virion RT-PCR products, NTC: No template control and \pm VE: positive control	12
	NTC; No-template control and +VE: positive control	42

xviii

4.1	Symptoms exhibited by ginger plants at pathogenicity test. A) Yellowing and vein banding at 2 MPI B) Leaf curling at 2 MPI C) Stripping at 2 MPI D) Healthy control and 1 MPI E) Mosaic symptoms at 4 MPI	51
4.2	Comparison of height between a sample among the healthy and inoculated ginger plants at the pathogenicity test at 5 MPI A) CMV- inoculated plant showing dwarfism and B) Healthy control at five- month-post-inoculation	52
4.3	Turmeric plants at pathogenicity test at 3-month post inoculation (MPI) A) Healthy plant with no viral symptom B) Stripping symptom parallel to the midrib C) Mosaic symptom intersparsed on leaf surface and D) Yellowing on the leaf along the veins	53
4.4	Comparison between the heights of healthy inoculated and iturmeric plants during pathogenicity test at 4 MPI A) Healthy control and B) Inoculated turmeric plant showing apparent stunted growth and reduced leaf size	54
4.5	RT-PCR assay for the detection of inoculated ginger and turmeric plants at pathogenicity test using 2% (w/v) agarose gel electrophoresis. A) 1–4: Ginger host 5–8: Turmeric host 9: NTC 10: Positive control M; 100 bp DNA ladder B) 1–5: Healthy control ginger 6–10: Healthy control turmeric NTC: No template control +VE: Positive control M: 100bp DNA size marker	55
4.6	Agarose gel 2% (w/v) showing the RT-PCR result from infected ginger and turmeric rhizomes after pathogenicity test when the plants were nine-month-old. $1 - 10$: ginger rhizomes, 11- 20: turmeric rhizomes NTC: No template control +VE: Positive control M: 100 bp DNA ladder	56
4.7	Rhizome transmission of CMV in ginger and turmeric confirmed by RT-PCR separated on 2% (w/v) agarose. $1 - 9$: leaves from grown CMV-infected ginger rhizomes: $10 - 19$: leaves grown from infected turmeric rhizomes	56
4.8	Symptoms of CMV infection on <i>Solanaceae</i> members A) chilli showing mosaic 25 dpi B) Eggplant with distorted leaf and mosaic 30 dpi C) Chlorosis and shoestring on tomato 30 dpi and D) Mild mosaic on tobacco 60 dpi	59
4.9	Some symptoms induced by CMV infection on different hosts tested A) Cowpea showing green mosaic 20 dpi B) Cucumber with mosaic 25 dpi C) Lettuce with mosaic 35 dpi D) Cabbage with mild mosaic 40 dpi and E) Torch ginger with mild mosaic 60 dpi	60
4.10	Plant hosts for host range studies prior to inoculation tested by RT-PCR showing the absence of CMV fractionated using 2% (w/v)	

	7-8: tobacco $9-10$: cowpea $11-12$: cucumber $13-14$: torch ginger $15-16$: lettuce $17 * 18$: cabbage NTC: No template control +VE: Positive control	61
4.11	RT-PCR for the host range studies separated on 2% (w/v) agarose gel. A) Inoculated host plants 1; Chili 2; Tomato 3; Eggplant 4; Tobacco 5; Cowpea 6; Cucumber 7; Torch ginger 8; Lettuce 9; Cabbage 10; No template control (NTC) +VE; Positive control M; 100 bp DNA ladder B) Same plant sequence as in (A) of healthy controls	62
5.1	Standard curve of amplification showing the efficiency, slope and R^2 values from plasmid DNA	72
5.2	Five-point amplification curve generated with 10-fold serially diluted plasmid DNA	72
5.3	Melt curve of amplification for SYBR green I, showing primer specificity with a single peak.	73
5.4	Quantification of CMV in ginger plants for evaluation of silver nanoparticles against the virus at different time interval from pre- treatment to 5-month post inoculation	74
5.5	Quantification of CMV in turmeric plants for evaluation of silver nanoparticles against the virus at different time interval from pre- treatment to 5-month post inoculation	76
J 1	Partial sequence of ginger TM3 isolate pathogenicity host plant blasted against the original host sequence with 100% similarity	143
J 2	Partial sequence of turmeric TMR pathogenicity host plant blasted against the original host sequence with 100% similarity	143

agarose gel electrophoresis. 1 - 2: chilli 3 - 4: tomato 5 - 6: eggplant

XX

LIST OF ABBREVIATIONS

	°C	Degrees Celcius
	μg	Micrcogramme
	μL	Microlitre
	μΜ	Micro molar
	mg	Milligramme
	aa	amino acid
	AMV-RT	Avian myeblastosis virus reverse transcriptase
	bp	base pair
	cDNA	Complimentary deoxyribonucleic acid
	CMV	Cucumber mosaic virus
	CTAB	cetyl trimethylammonium bromide
	dNTP	Deoxynucleotide triphosphate dATP, dCTP. dGTP, dTTP
	g	Gramme
	g	Centrifugal force
	GCFV	Ginger chlorotic fleck virus
	h	Hour
	L	Litre
	Min	Minute
	mL	Millilitre
	PCR	Polymerase chain reaction
	PVP	Polyvinyl pyrrolidone
	RNA	Ribonucleic acid
	RT-PCR	Reverse transcription- Polymerase chain reaction
	ppm	Parts per million
	UV	Ultraviolet
	U	Unit
	rpm	Revolution per minute
	TBE	Tris borate ethylene diamine tetra acetic acid
	SDDW	Sterile double distilled water

SDS	Sodium dodecyl sulphate
v/v	Volume/volume percentage
W/V	Weight/volume percentage
dpi	Day post inoculation
mpi	Month post inoculation
S	Second
WPI	Week post inoculation
MPI	Month post inoculation
EDTA	Ethylene diaminine tetra acetic acid
М	Molar
SOC	Super optimal broth
LB	Luria Bertani
X-gal	5-bromo-4-chloro-3-indolyl-β-D-galactopyranoside
RFLP	Restriction fragment length polymorphism

C

CHAPTER 1

INTRODUCTION

Ginger (*Zingiber officinale* Rosc.) and turmeric (*Curcuma longa* L.) of the *Zingiberaceae* family, are among the economically important vegetable crops in Malaysia. Ginger is of culinary and medicinal importance and it is among the most valued and widely cultivated spice crops in the world (Singletary, 2010; Amadi, 2012; Darshana et al., 2014). Like ginger plant, turmeric has cosmetic, medicinal preservative values besides its culinary function (Akam et al., 2010; Babu et al., 2011; Gul and Bakht, 2015; Abdul Haiyee et al. 2016).

Malaysia is the 13th world producer of ginger (FAOSTAT, 2015), cultivating a highquality Bentong variety, with superior properties compared to many world gingers in terms of mellow taste, richness of gingerols, and long storability. Turmeric production in the country is about 2,640 tonnes and consumed locally (Suhaimi et al., 2014). Both ginger and turmeric have generated revenues to Malaysia at an approximately \$ 24 million and \$ 2 million annually respectively (DOA, 2018).

Fungal, bacterial and viral diseases have constituted a major threat to crop production, which can lead to an enormous yield loss (Koike et al., 2007). Fungi and bacteria constitute a nuisance in ginger and turmeric production, with rhizome rot induced by *Pythium* spp. among the most devastating diseases (American Society for Horticultural Science, 2011; Anusuya and Sathiyabama, 2015).

Many viruses have been associated with important diseases in vegetables but one of the most common and widely distributed viruses is the Cucumber mosaic virus (CMV). CMV has been found in Malaysia on a variety of economic, vegetable, and ornamental crops, as well as weed hosts, indicating the pathogen's long presence in the country's agroecosystem (Mazidah et al. 2012), but it has not been reported in either ginger or turmeric. Among the viruses reported to infect ginger were Ginger mosaic virus, belonging to CMV group (So, 1980). CMV, a member of the genus Cucumovirus with single stranded RNA (ssRNA) and tripartite genome, may reduce crop yield by up to 30% (Zitter and Murphy, 2009; de Breuil et al., 2012). The second virus that has been reported to infect ginger was Ginger chlorotic fleck virus (GCFV), which belongs to the genus Sobemovirus tentatively, with ssRNA and icosahedral particle of 30 nm in diameter (Thomas, 1986). However, there has been no previous report of any attempt to investigate both these viruses in question in Malaysia. Nevertheless, it is important to identify viruses that might have infected ginger and turmeric for proper management strategy to be deployed and in order to avoid the loss that might be incurred.

A rapid and sensitive identification of the causal agent is imperative in management of plant viral diseases. Serological techniques, such as enzyme-linked immunosorbent assay (ELISA) in its various formats have been used in the diagnosis of CMV and GCFV (Thomas, 1986; El-Borollosy and Waziri, 2013; Eni et al., 2013; Bald-Blume et al., 2017). Nucleic acid-based techniques, using RT-PCR have been widely utilized in CMV and GCFV detection and identification; followed by nucleotide sequencing, phylogenetic analysis and restriction digestion of PCR products (Wylie et al., 1993; Sérémé et al., 2008; Kim et al., 2014; Nouri et al., 2014). However, nucleic acid based, RT-PCR, cloning and sequencing had not been applied for the identification of CMV in ginger and turmeric in the past.

Pathogenicity and host range studies remain fundamental in the diagnostic of plant viruses. CMV is known as the plant virus with the widest host range in contrast with GCFV, which only has a single host. CMV infects wild, domestic, vegetables, monocots and dicots, ornamentals as well as tree crops (Carrère et al., 1999; Paradies et al., 2000; Bald-Blume et al., 2017) whereas, GCFV was only found to infect ginger (Thomas, 1986). However, CMV and GCFV infections were not investigated in ginger and turmeric in Malaysia despite the conspicuous viral symptoms observed on the crops.

To control plant viral diseases, various approaches, ranging from cultural, biological and chemical methods have been applied due the fact that there is no unswerving method to control plant viruses (Lecoq and Katis, 2014). Recently, nanotechnology has evolved with remarkable results so far in the field of plant pathology, against plant pathogenic bacteria, fungi as well as viruses (Kim et al., 2008; Ocsoy et al., 2013; Elbeshehy et al., 2015; Pirtarighat et al., 2019). Therefore, the use of silver nanoparticles was conceived in this study, to evaluate its suppression on CMV in ginger and turmeric plants since it has never been conducted before.

1.1 Statement of problem

Many plant viruses were reported on vegetable crops. The viruses have reduced the crop yield or lower the crop quality, thereby decreasing its marketability. Viral symptoms were observed on ginger and turmeric plants in Peninsular Malaysia. The symptoms observed were mosaic, striping, reduced leaf and plant sizes. However, the causal agent has not been identified and characterized. CMV was reported to cause a yield loss ranging from 10 % to 30 % in various crop plants (Zitter and Murphy, 2009; de Breuil et al., 2012). It was then deemed necessary to screen the infected ginger and turmeric for CMV infection, as early and timely identification of the causal organism is the most important aspect of any disease management.

1.2 Justification of study

Since viruses may devastate ginger and turmeric crops, which may lead to a significant loss in the yield either in quantity or quality, and no research was carried out with respect to the identification of viruses reported to be associated with ginger and turmeric in Malaysia, it is of paramount importance to determine the causal organism. This will aid the quarantine services, virus resistance breeding and production of virus-free planting materials and also to meet the demand of Malaysian government in its intended expansion of fruits and vegetables production that are of higher quality, according to the food safety standards and in order to access premium markets in the developed world.

1.3 Objectives of the study

The study aims at generating information needed, which will help in designing ecofriendly and sustainable viral diseases management in ginger and turmeric crops in Peninsular Malaysia as its overall objective.

The specific objectives were:

- 1. To identify and characterize viruses on ginger and turmeric in Peninsular Malaysia using electron microscopy, ELISA, RT-PCR, cloning and sequencing.
- 2. To determine the pathogenicity and host range of the identified virus isolates.
- 3. To evaluate the efficacy of silver nanoparticles to control viral diseases in ginger and turmeric plants.

REFERENCES

- Abdul Haiyee, Z., Mohd Shah, S. H., Ismail, K., Hashim, N., & Wan Ismail, W. I. (2016). Quality parameters of curcuma longa l. extracts by supercritical fluid extraction (sfe) and ultrasonic assisted extraction (UAE). *Malaysian Journal* of Analytical Science, 20(3), 626–632. https://doi.org/10.17576/mjas-2016-2003-23
- Akram, M., Ahmed, A., Usmanghani, K., Hannan, A., Mohiuddin, E., & Asif, M. (2010). Curcuma longa and curcumin: a review article. *Romanian Journal of Biology*, 55(2), 65–70.
- Ali, A., & Kobayashi, M. (2010). Seed transmission of Cucumber mosaic virus in pepper. Journal of Virological Methods, 163(2), 234–237. https://doi.org/10.1016/j.jviromet.2009.09.026
- Amadi, C. O. (2012). Ginger breeding in Nigeria: challenges and prospects. *Journal* of Applied Agricultural Research, 4(2), 155–163.
- American Society for Horticultural Science., V. A., International Society for Horticultural Science., V., Nair, R. R., Zachariah, T. J., Kumar, A., & Prasath, D. (2011). Horticultural reviews. In *Horticultural Reviews* (Vol. 39). AVI Pub. Co.
- Anusuya, S., & Sathiyabama, M. (2015). Protection of turmeric plants from rhizome rot disease under field conditions by β-d-glucan nanoparticle. *International Journal of Biological Macromolecules*, 77, 9–14. https://doi.org/10.1016/j.ijbiomac.2015.02.053
- Arafati, N., Farzadfar, S., & Pourrahim, R. (2013). Characterization of coat protein gene of Cucumber mosaic virus isolates in Iran. *Iranian Journal of Biotechnology*, 11(2), 109–114. https://doi.org/10.5812/ijb.10715
- Babu, K. N., Shiva, K. N., Sabu, M., Divakaran, M., & Ravindran, P. N. (2011). Turmeric. In *Genetic Resources, Chromosome Engineering, and Crop Improvement: Medicinal Plants.* https://doi.org/10.1533/9780857095671.526.
- Bagewadi, B., Naidu, R. A., Shahadath Hossain, & Fayad, A. (2015). First report of cucumber mosaic virus from eggplant (Solanum Melongena) in Bangladesh. *Plant Disease*, 99(2), 14–16. https://doi.org/10.1094/PDIS-09-14-0956-PDN
- Balasuriya, L. B. A. K. M., & Kelaniyangoda, D. B. (2010). Identification and management of pest and diseases of ginger. In *Proceedings of 10th Agricultural Research Symposium Vol* (Vol. 74, p. 78).
- Bald-Blume, N., Bergervoet, J. H. W., & Maiss, E. (2017). Development of a molecular assay for the detection of Cucumber mosaic virus and the discrimination of its subgroups I and II. *Journal of Virological Methods*, 243, 35–43. https://doi.org/10.1016/j.jviromet.2017.01.011

- Betancourt, M., Fraile, A., & García-Arenal, F. (2011). Cucumber mosaic virus satellite RNAs that induce similar symptoms in melon plants show large differences in fitness. *Journal of General Virology*, 92(8), 1930–1938. https://doi.org/10.1099/vir.0.032359-0.
- Bezbuarah, B. J. and Hazarika, M. K. (2014). Generalization of temperature and thickness effects in kinetic studies of turmeric (Curcuma longa) slices drying. *International Food Research Journal*, 21(4), 1529–1532.
- Bhat, A. I., Naveen, K. P., Pamitha, N. S., & Pant, R. P. (2020). Association of two novel viruses with chlorotic fleck disease of ginger. *Annals of Applied Biology*, 177(2), 232-242.
- Carrère, I., Tepfer, M., & Jacquemond, M. (1999). Recombinants of cucumber mosaic virus (CMV): determinants of host range and symptomatology. In *Arch Virol* (Vol. 144).
- Chen, J. C., Huang, L. J., Wu, S. L., Kuo, S. C., Ho, T. Y., & Hsiang, C. Y. (2007). Ginger and its bioactive component inhibit enterotoxigenic Escherichia coli heat-labile enterotoxin-induced diarrhea in mice. *Journal of Agricultural and Food Chemistry*, 55(21), 8390–8397. https://doi.org/10.1021/jf071460f
- Chou, Chun-Nan Chen, Cheng-En Wu, Meng-Ling Su, Hong-Ji and Yeh, H.-H. (2009). Biological and Molecular Characterization of Taiwanese Isolates of Cucumber mosaic virus Associated with Banana Mosaic Disease. 93, 85–93. https://doi.org/10.1111/j.1439-0434.2008.01455.x
- Darshana, C. N., Praveena, R., Ankegowda, S. J., & Biju, C. N. (2014). Morphological variability, mycelial compatibility and fungicidal sensitivity of Colletotrichum gloeosporioides causing leaf spot of ginger (Zingiber officinale Rosc.). 23(2), 211–223.
- de Breuil, S., Giolitti, F.J., Bejerman, N. and Lenardon, S. L. (2012). EFFECTS OF CUCUMBER MOSAIC VIRUS ON THE YIELD AND YIELD COMPONENTS OF PEANUT S. Journal of Plant Pathology, 94(3), 669– 673.
- Deng, T. C., Tsai, C. H., Tsai, H. L., Liao, J. Y., & Huang, W. C. (2010). Disease notes: First report of cucumber mosaic virus on vigna marina in Taiwan. *Plant Disease*, Vol. 94, p. 1267. https://doi.org/10.1094/PDIS-06-10-0459
- Devasahayam, S., & Koya, K. M. A. (2005). Insect pests of ginger. In *Ginger The* genus Zingiber (p. 24).
- Dey, K. K., Li, C., Elliott, M., McVay, J., Whilby, L., Hodges, G., & Smith, T. R. (2019). First Report of Cucumber mosaic virus Infecting Siam Tulip (Curcuma alismatifolia) in Florida. *Plant Health Progress*, 20(3), 132–132. https://doi.org/10.1094/php-03-19-0019-br

DOA. (2018). Malaysia 2018 (1).pdf.

- Doyle, J. J. and Doyle, J. L. (1987). A rapid DNA isolation procedure for small quantities of fresh leaf tissue. *Phytochemical Bulletin*, *19*, 11–15. Retrieved from http://ci.nii.ac.jp/naid/10021087108/en/
- Egbuchua, C. N., & Enujeke, E. C. (2013). Journal of Horticulture and Forestry Growth and yield responses of ginger (Zingiber officinale) to three sources of organic manures in a typical rainforest zone, Nigeria. 5(7), 109–114. https://doi.org/10.5897/JHF2013.0302
- El-Borollosy, A. M., & Waziri, H. M. A. (2013). Molecular characterization of a cucumber mosaic cucumovirus isolated from lettuce in Egypt. Annals of Agricultural Sciences, 58(1), 105–109. https://doi.org/10.1016/j.aoas.2013.01.014
- El-Dougdoug, K. A., Ghaly, M. F., & Taha, M. A. (2012). Biological control of cucumber mosaic virus by certain local Streptomyces isolates: Inhibitory effects of selected five Egyptian isolates. *International Journal of Virology*, 8(2), 151–164. https://doi.org/10.3923/rjv.2012.151.164
- Elbeshehy, E. K. F., Elazzazy, A. M., & Aggelis, G. (2015). Silver nanoparticles synthesis mediated by new isolates of Bacillus spp., nanoparticle characterization and their activity against Bean Yellow Mosaic Virus and human pathogens. *Frontiers in Microbiology*, 6(MAY), 1–13. https://doi.org/10.3389/fmicb.2015.00453
- Eni, A. O., Kumar, P.L., Asiedu, R., Alabi, O. J., Naidu, R. A., Hughes, Jd'A., and Rey, M. E. C. (2013). Characterization of cucumber mosaic virus isolated from yam (Dioscorea spp.) in West Africa. *African Journal of Biotechnology*, 12(22), 3472–3480. https://doi.org/10.5897/AJB2013.12303
- Eni. A.O., Lava Kumar, P., Asiedu, R., Alabi, O.J., Naidu, R.A., Hughes, J., Rey, M. E. C. (2013). Characterization of cucumber mosaic virus isolated from yam (Dioscorea spp.) in West Africa. *African Journal of Biotechnology*, 12(22), 3472–3480. https://doi.org/10.5897/AJB2013.12303
- Eni, A. O., Ogunsanya, P., Oviasuyi, T., & Hughes, J. d. A. (2013). Alarming increase in the incidence of Cucumber mosaic virus in cowpea (Vigna unguiculata (L.) Walp.) in northern Nigeria. Archives of Phytopathology and Plant Protection, 46(16), 1958–1965. https://doi.org/10.1080/03235408.2013.782218
- Ernst, E., & Pittler, M. H. (2000). Efficacy of ginger for nausea and vomiting: a systematic review of randomized clinical trials. *British Journal of Anaesthesia Br J Anaesth*, 84(84), 367–371. https://doi.org/10.1093/oxfordjournals.bja.a013442

FAOSTAT. (2015a). FAOSTAT REPORT.

FAOSTAT. (2015b). Food and Agriculture Organization.

- Feng, J. L., Chen, S. N., Tang, X. S., Ding, X. F., Du, Z. Y., & Chen, J. S. (2006). Quantitative determination of cucumber mosaic virus genome RNAs in virions by real-time reverse transcription-polymerase chain reaction. Acta Biochimica et Biophysica Sinica, 38(10), 669–676. https://doi.org/10.1111/j.1745-7270.2006.00216.x
- Fujisawa, I., Anang, S. H., Shen, Y., & Zhou, A. (1990). Identification of viral diseases affecting some vegetable crops in west Malaysia and the southern part of China. *Tropical Agriculture Research Series*, 23, 218–228.
- Galdiero, S., Falanga, A., Vitiello, M., Cantisani, M., Marra, V., & Galdiero, M. (2011). Silver nanoparticles as potential antiviral agents. *Molecules*. https://doi.org/10.3390/molecules16108894
- Gambino, G., Perrone, I., & Gribaudo, I. (2008). A rapid and effective method for RNA extraction from different tissues of grapevine and other woody plants. *Phytochemical Analysis*, 19(6), 520–525. https://doi.org/10.1002/pca.1078
- Ghasemzadeh, A., Jaafar, H. Z. E., & Rahmat, A. (2010). Elevated Carbon Dioxide Increases Contents of Flavonoids and Phenolic Compounds, and Antioxidant Activities in Malaysian Young Ginger (Zingiber officinale Roscoe.) Varieties. *Molecules*, 15(11), 7907–7922. https://doi.org/10.3390/molecules15117907
- Ghasemzadeh, A., & Ze Jaafar, H. (2011). Anticancer and antioxidant activities of Malaysian young ginger (Zingiber officinale Roscoe) varieties grown under different CO2 concentration Impact of different light intensities and carbon dioxide enrichment on yield and pharmaceutical quality of Malay. *Journal of Medicinal Plants Research*, 5(4), 3247–3255. Retrieved from https://www.researchgate.net/publication/303517188
- Gildow, F. E., Shah, D. A., Sackett, W. M., Butzler, T., Nault, B. A., & Fleischer, S. J. (2008). Transmission efficiency of Cucumber mosaic virus by aphids associated with virus epidemics in snap bean. *Phytopathology*, 98(11), 1233–1241. https://doi.org/10.1094/PHYTO-98-11-1233
- Gillaspie, J. (2001). Resistance to Cucumber mosaic virus in cowpea and implications for control of cowpea stunt disease. *Plant Disease*, 85(9), 1004–1005. https://doi.org/10.1094/pdis.2001.85.9.1004
- Guji, M.J., Yetayew, H.T. & Kidanu, E.D. (2019). Yield loss of ginger (*Zingiber officinale*) due to bacterial wilt (*Ralstonia solanacearum*) in different wilt management systems in Ethiopia. Agric and Food Security 8, 5. https://doi.org/10.1186/s40066-018-0245-6
- Gul, P., & Bakht, J. (2015). Antimicrobial activity of turmeric extract and its potential use in food industry. *Journal of Food Science and Technology*, 52(4), 2272–2279. https://doi.org/10.1007/s13197-013-1195-4
- H. M. Blanck, C. Gillespie, J. E. Kimmons, J.D. Seymour, and M. K. S. (2008).

Trends in fruit and vegetable consumption among U.S. men and women, 1994–2005. *Preventing Chronic Diseases*, 5(2).

- Hadidi, A., Flores, R., Candresse, T., & Barba, M. (2016). Next-generation sequencing and genome editing in plant virology. *Frontiers in Microbiology*, 7(AUG). https://doi.org/10.3389/fmicb.2016.01325
- Hareesh, P. S., Madhubala, R., & Bhat, A. I. (2006). Characterization of Cucumber mosaic virus infecting Indian long pepper (Piper longum L.) and betel vine (Piper betle L.) in India. *Indian Journal of Biotechnology*.
- Hogarth, J., & Joan. (1999). Buderim ginger: An export success story: A history of the ginger industry in Queensland.
- Hull, R. (2009). Mechanical inoculation of plant viruses. Current Protocols in Microbiology, (SUPPL. 13), 6–9. https://doi.org/10.1002/9780471729259.mc16b06s13
- Jacquemond, M. (2012). Cucumber Mosaic Virus. In *Advances in Virus Research* (Vol. 84, pp. 439–504). https://doi.org/10.1016/B978-0-12-394314-9.00013-0
- Jones, R. A. C., Coutts, B. A., Latham, L. J., & McKirdy, S. J. (2008). Cucumber mosaic virus infection of chickpea stands: temporal and spatial patterns of spread and yield-limiting potential. *Plant Pathology*, 57(5), 842-853.
- Kandiannan, K., Sivaraman, K., Thankamani, C. K., & Peter, K. V. (1996). Agronomy of ginger (Zingiber officinule Rose.) a review ! 5(198), 1–27.
- Kareem, K. T., & Taiwo, M. A. (2007). Interactions of viruses in Cowpea: Effects on growth and yield parameters. *Virology Journal*, 4, 1–7. https://doi.org/10.1186/1743-422X-4-15
- Kayode, A. B., Odu, B. O., Ako-Nai, K. A., & Alabi, O. J. (2014). Occurrence of cucumber mosaic virus subgroups IA and IB isolates in tomatoes in Nigeria. *Plant Disease*, Vol. 98, p. 1750. https://doi.org/10.1094/PDIS-08-14-0844-PDN
- Kew, R. B. G. (2016). The state of the world's plants report–2016. *Royal Botanic Gardens*.
- Khandelwal, N., Kaur, G., Kumar, N., & Tiwari, A. (2014). Application of silver nanoparticles in viral inhibition: a new hope for antivirals. *Digest Journal of Nanomaterials & Biostructures (DJNB)*, 9(1).
- Kim, C. H., & Palukaitis, P. (1997). The plant defense response to cucumber mosaic virus in cowpea is elicited by the viral polymerase gene and affects virus accumulation in single cells. *EMBO Journal*, 16(13), 4060–4068. https://doi.org/10.1093/emboj/16.13.4060

- Kim, J. S., Lee, S. I., Park, H. W., Yang, J. H., Shin, T. Y., Kim, Y. C., ... Kim, D. K. (2008). Cytotoxic components from the dried rhizomes of Zingiber officinale Roscoe. Archives of Pharmacal Research, 31(4), 415–418. https://doi.org/10.1007/s12272-001-1172-y
- Kim, K. J., Sung, W. S., Moon, S. K., Choi, J. S., Kim, J. G., & Lee, D. G. (2008). Antifungal effect of silver nanoparticles on dermatophytes. *Journal of Microbiology and Biotechnology*, 18(8), 1482–1484.
- Kim, M. K., Seo, J. K., Kwak, H. R., Kim, J. S., Kim, K. H., Cha, B. J., & Choi, H. S. (2014). Molecular genetic analysis of cucumber mosaic virus populations infecting pepper suggests unique patterns of evolution in Korea. *Phytopathology*, 104(9), 993–1000. https://doi.org/10.1094/PHYTO-10-13-0275-R
- Kim, S. W., Jung, J. H., Lamsal, K., Kim, Y. S., Min, J. S., & Lee, Y. S. (2012). Antifungal effects of silver nanoparticles (AgNPs) against various plant pathogenic fungi. *Mycobiology*, 40(1), 53–58. https://doi.org/10.5941/MYCO.2012.40.1.053
- Kitajima, E. W. (2004). Electron microscopy in plant virology: Past, present and future. *Microscopy and Microanalysis*. https://doi.org/10.1017/S1431927604881467
- Koike, S.T., Glladders, P. and Paulus, A. O. (2007). Vegetable Diseases. In *Vegetable Diseases*. https://doi.org/10.1201/b15147
- Kumar, Sudhir, Stecher, G., & Tamura, K. (2016). MEGA7: Molecular Evolutionary Genetics Analysis Version 7.0 for Bigger Datasets. *Molecular Biology and Evolution*, 33(7), 1870–1874. https://doi.org/10.1093/molbev/msw054
- Kumar, Susheel, Gautam, K. K., & Raj, S. K. (2014). Molecular identification of Cucumber mosaic virus isolates of subgroup IB associated with mosaic disease of eggplant in India. VirusDisease, 25(1), 129–131. https://doi.org/10.1007/s13337-013-0174-8
- Kumari, R., Bhardwaj, P., & Singh, L. (2013). Biological and Molecular Characterization of Cucumber mosaic virus Subgroup II Isolate Causing Severe Mosaic in Cucumber. 24(June), 27–34. https://doi.org/10.1007/s13337-012-0125-9
- Lal, J. (2012). Turmeric , Curcumin and Our Life: A Review. Bulletin of Environment, Pharmacology and Life Sciences, 1(June), 11–17.
- Le, D. P., Smith, M., Hudler, G. W., & Aitken, E. (2014). Pythium soft rot of ginger:Detection and identification of the causal pathogens, and their control. *Crop Protection*, 65, 153-167.
- Lecoq, H., & Katis, N. (2014). Control of Cucurbit Viruses. In Advances in Virus Research. https://doi.org/10.1016/B978-0-12-801246-8.00005-6

- Lefever, S., Pattyn, F., Hellemans, J., & Vandesompele, J. (2013). Single-nucleotide polymorphisms and other mismatches reduce performance of quantitative PCR assays. *Clinical Chemistry*, 59(10), 1470–1480. https://doi.org/10.1373/clinchem.2013.203653
- Libeni, Y., Singh, A. K., & Singh, V. B. (2010). Effect of INM on yield, quality and uptake of N, P and K by ginger. *Agropedology*, 20(1), 74–79. Retrieved from https://www.cabdirect.org/cabdirect/abstract/20103348818
- Liu, S., Cao, X., & Yuan, X. (2017). First report of Cucumber mosaic virus and its associated satellite RNA in celery cabbage in Shandong province of China. *Plant Disease*, 101(10), 1829. https://doi.org/10.1094/PDIS-04-17-0538-PDN
- Loebenstein, G., & Lecoq, H. (2011). Viruses and Viral diseases of Vegetables in the Mediterranean Basin. *Admission Assessment: Exam Review*.
- Lü, J. M., Wang, X., Marin-Muller, C., Wang, H., Lin, P. H., Yao, Q., & Chen, C. (2009). Current advances in research and clinical applications of PLGAbased nanotechnology. *Expert Review of Molecular Diagnostics*, 9(4), 325– 341. https://doi.org/10.1586/erm.09.15
- Lu, L., Sun, R. W. Y., Chen, R., Hui, C. K., Ho, C. M., Luk, J. M., ... Che, C. M. (2008). Silver nanoparticles inhibit hepatitis B virus replication. *Antiviral Therapy*, 13(2), 253.
- M. Yaseer Suhaimi1, Abd. M. Mohamad1, & and M. Nur Farah Hani. (2014). Potential and Viability Analysis for Ginger Cultivation using Fertigation Technology in Malaysia. *International Journal of Innovation and Applied Studies*, 9(1), 421–427.
- Mahdi, H. J., Andayani, R., & Aziz, I. (2013). Determination of phylogenetic and molecular characteristics of three Malaysian ginger cultivars (Zingiber officinale Roscoe) using microsatellite DNA. *Tropical Life Sciences Research*, 24(2), 65–76.
- Maizura, M., Aminah, A., & Aida, W. M. W. (2011). Total phenolic content and antioxidant activity of kesum (Polygonum minus), ginger (Zingiber officinale) and turmeric (Curcuma longa) extract. *International Food Research Journal*, 18(2).
- Maruthi, M. N., Whitfield, E. C., Otti, G., Tumwegamire, S., Kanju, E., Legg, J. P., ... & Mbugua, E. (2019). A method for generating virus-free cassava plants to combat viral disease epidemics in Africa. *Physiological and molecular plant pathology*, 105, 77-87.
- Mascia, T., & Gallitelli, D. (2014). Synergism in plant-virus interactions: A case study of CMV and PVY in mixed infection in tomato. In *Plant Virus-Host Interaction: Molecular Approaches and Viral Evolution. Academic Press* (pp. 195–206). https://doi.org/10.1016/B978-0-12-411584-2.00010-X

- Maule, A. J., & Wang, D. (1996). Seed transmission of plant viruses: A lesson in biological complexity. *Trends in Microbiology*, 4(4), 153–158. https://doi.org/10.1016/0966-842X(96)10016-0
- Mazidah, M., Yusoff, K., Habibuddin, H., Tan, Y. H., & Lau, W. H. (2012). Characterization of cucumber mosaic virus (CMV) causing mosaic symptom on Catharanthus roseus (L.) G. Don in Malaysia. *Pertanika Journal of Tropical Agricultural Science*, 35(1), 41–53.
- Meenu, G., and Kaushal, M. (2017). Diseases infecting ginger (Zingiber officinale Roscoe): A review. *Agricultural Reviews*, 38(1): 15 28.
- Mink, G. I. (1993). Pollen and Seed-Transmitted Viruses and Viroids. Annual Review of Phytopathology. https://doi.org/10.1146/annurev.py.31.090193.002111
- Moury, B. (2004). Differential selection of genes of cucumber mosaic virus subgroups. *Molecular Biology and Evolution*, 21(8), 1602–1611. https://doi.org/10.1093/molbev/msh164
- Moyle, R., Pretorius, L. S., Shuey, L. S., Nowak, E., & Schenka, P. M. (2018). Analysis of the complete genome sequence of Cucumber mosaic virus strain K. Genome Announcements, 6(7). https://doi.org/10.1128/genomeA.00053-18
- Nandhini, M., Pream Sudha, V. and, & Vijaya, M. S. (2016). *Identification and Classification of Leaf Diseases in Turmeric Plants*. 6(2), 48–54.
- Nono-Womdim, R. (2001). An overview of major viral diseases of vegetable crops in Africa and some aspects of their control. *Plant Virology in Sub-Saharan Africa*, 213–232.
- Nouri, S., Arevalo, R., Falk, B. W., & Groves, R. L. (2014). Genetic structure and molecular variability of Cucumber mosaic virus isolates in the United States. *PLoS ONE*, 9(5). https://doi.org/10.1371/journal.pone.0096582
- Ocsoy, I., Paret, M. L., Ocsoy, M. A., Kunwar, S., Chen, T., You, M., & Tan, W. (2013). Nanotechnology in plant disease management: DNA-directed silver nanoparticles on graphene oxide as an antibacterial against Xanthomonas perforans. ACS Nano, 7(10), 8972–8980. https://doi.org/10.1021/nn4034794
- Oreshkovikj, K. B., Rusevski, R., Kuzmanovska, B., Jankulovska, M., & Popovski, Z. T. (2018). Occurrence of plant viruses on pepper cultivated in open fields in R. Macedonia and partial characterization of cucumber mosaic virus isolates. *Journal of Plant Pathology*, 100(3), 485–491. https://doi.org/10.1007/s42161-018-0110-2
- Pallas, V., & García, J. A. (2011). How do plant viruses induce disease? Interactions and interference with host components. *Journal of General Virology*, 92,

2691–2705. https://doi.org/10.1099/vir.0.034603-0

- Papp, I., Sieben, C., Ludwig, K., Roskamp, M., Böttcher, C., Schlecht, S., ... Haag, R. (2010). Inhibition of influenza virus infection by multivalent sialic-acidfunctionalized gold nanoparticles. *Small*, 6(24), 2900–2906. https://doi.org/10.1002/smll.201001349
- Paradies, F., Sialer, M. F., Gallitelli, D., Castellano, M. A., Di Franco, A., Digiaro, M., ... Yilmaz, M. A. (2000). Partial characterization of cucumber mosaic virus isolates from citrus and grapevine. *Journal of Plant Pathology*, 82(2), 133–145. https://doi.org/10.4454/jpp.v82i2.1153
- Pazarlar, S., Gümüs, M., & Öztekin, G. B. (2013). The effects of tobacco mosaic virus infection on growth and physiological parameters in some pepper varieties (Capsicum annuum L.). Notulae Botanicae Horti Agrobotanici Cluj-Napoca, 41(2), 427–433. https://doi.org/10.15835/nbha4129008
- Pirtarighat, S., Ghannadnia, M., & Baghshahi, S. (2019). Green synthesis of silver nanoparticles using the plant extract of Salvia spinosa grown in vitro and their antibacterial activity assessment. *Journal of Nanostructure in Chemistry*, 9(1), 1–9. https://doi.org/10.1007/s40097-018-0291-4
- Pratap, D., Kumar, S., Snehi, S. K., & Raj, S. K. (2012). Biological and molecular characterization of cucumber mosaic virus isolate causing shoestring disease of tomato in India which has closer affinity to European or east Asian isolates of CMV. *Indian Journal of Virology*, 23(1), 57–63. https://doi.org/10.1007/s13337-012-0059-2
- Raji, M. N. A., Ab Karim, S., Ishak, F. A. C., & Arshad, M. M. (2017). Past and present practices of the Malay food heritage and culture in Malaysia. *Journal* of Ethnic Foods, 4(4), 221–231. https://doi.org/10.1016/j.jef.2017.11.001
- Rao, T. N. (1995). Diseases of turmeric (Curcuma longa L.) and their management. Journal of Spices and Aromatic Crops, 4(1), 49–56.
- Ravindran, P. N., Sasikumar, B., George, J. K., Ratnambal, M. J., & Babu, K. N. (1994). Genetic resources of ginger (Zingiber officinale Rosc.) and its conservation in India. *Plant Genetic Resources Newsletter*, 1–4.
- Roossinck, M. J. and White, P. S. (1998). Comovirus isolation and RNA extraction. In *Plant Virology Protocols* (Vol. 81, pp. 189–196). https://doi.org/10.1385/0-89603-385-6:189
- Roossinck, M. J. (2001). Cucumber mosaic virus, a model for RNA virus evolution. *Molecular Plant Pathology*. https://doi.org/10.1046/j.1364-3703.2001.00058.x
- Roossinck, M. J. (2002). Evolutionary History of Cucumber Mosaic Virus Deduced by Phylogenetic Analyses. *Journal of Virology*, 76(7), 3382–3387. https://doi.org/10.1128/jvi.76.7.3382-3387.2002

- Roossinck, Marilyn J. (2001). Cucumber mosaic virus, a model for RNA virus evolution. In *MOLECULAR PLANT PATHOLOGY* (Vol. 2). Retrieved from http://www.ncbi.nlm.nih.gov/ICTVdb/ICTVdB/10040001.htm.
- Rubio, L., Galipienso, L., & Ferriol, I. (2020). Detection of plant viruses and disease management: relevance of genetic diversity and evolution. *Frontiers in plant science*, *11*, 1092.
- Ryder, E. (2011). World vegetable industry: production, breeding, trends. *Horticultural Reviews*, 38, 299.
- Saad, K. A., Mohamad Roff, M. N., Hallett, R. H., & Abd-Ghani, I. B. (2019). Effects of cucumber mosaic virus-infected chilli plants on non-vector Bemisia tabaci (Hemiptera: Aleyrodidae). *Insect Science*, 26(1), 76–85. https://doi.org/10.1111/1744-7917.12488
- Saad, N. B. (2012). INCIDENCE AND DIFFERENTIATION OF Cucumber mosaic virus (CMV) ISOLATES IN PENINSULAR MALAYSIA. University Putra Malaysia.
- Sacristán, S., Fraile, A., & García-Arenal, F. (2004). Population dynamics of Cucumber mosaic virus in melon crops and in weeds in Central Spain. *Phytopathology*, 94, 992–998. https://doi.org/10.1094/PHYTO.2004.94.9.992
- Salleh, A., Naomi, R., Utami, N. D., Mohammad, A. W., Mahmoudi, E., Mustafa, N., & Fauzi, M. B. (2020). The potential of silver nanoparticles for antiviral and antibacterial applications: a mechanism of action. *Nanomaterials*, 10(8), 1566.
- Sastry, K. S. (2013). Seed-borne plant viral diseases. https://doi.org/10.1007/978-81-322-0813-6
- Sastry, S. K. and, & Zitter, T. A. (2014). Management of Virus and Viroid Diseases of Crops in the Tropics. In *Plant Virus and Viroid Diseases in the Tropics*. https://doi.org/10.1007/978-94-007-7820-7_2
- Scholthof, K. B. G., Adkins, S., Czosnek, H., Palukaitis, P., Jacquot, E., Hohn, T., ... Foster, G. D. (2011, December). Top 10 plant viruses in molecular plant pathology. *Molecular Plant Pathology*, Vol. 12, pp. 938–954. https://doi.org/10.1111/j.1364-3703.2011.00752.x
- Sérémé, D., Lacombe, S., Konaté, M., Pinel-Galzi, A., Traoré, V. S. E., Hébrard, E., ... Konaté, G. (2008). Biological and molecular characterization of a putative new sobemovirus infecting Imperata cylindrica and maize in Africa. *Archives* of Virology, 153(10), 1813–1820. https://doi.org/10.1007/s00705-008-0190y
- Shah, H., Yasmin, T., Fahim, M., Hameed, S., & Haque, M. I. (2008). Transmission and host range studies of Pakistani isolate of chilli veinal mottle virus.

Pakistan Journal of Botany, 40(6), 2669–2681.

- Singh, H. B. An assessment of Pre and Post Harvest Factors Affecting Quality of Ginger in the Export Chain : a case Study of Salyan District and Nepalgunj City, Mid-Western Development Region, Nepal., (2013).
- Singletary, K. (2010). Ginger: An Overview of Health Benefits. *Nutrition Today*, 45(4), 171–183. https://doi.org/10.1097/NT.0b013e3181ed3543
- So, I. Y. (1980). Studies on ginger mosaic virus. *Korean Journal of Plant Protection*, 19(2), 67–72.
- Soards, A. J., Murphy, A. M., Palukaitis, P., & Carr, J. P. (2002). Virulence and differential local and systemic spread of Cucumber mosaic virus in tobacco are affected by the CMV 2b protein. *Molecular Plant-Microbe Interactions*, 15(7), 647–653. https://doi.org/10.1094/MPMI.2002.15.7.647
- Suhaimi, M. Y., Mohammad, A. M., Hani, M. N. F., Mohamad, A. M., Hani, M. N. F., Mohammad, A. M., & Hani, M. N. F. (2014). Potential and viability analysis for ginger cultivation using fertigation technology in Malaysia. *International Journal of Innovation and Applied Science*, 9(1), 421–427.
- Sumathi, C. S., Balasubramanian, V., Ramesh, N., & Kannan, V. R. (2008). Influence of Biotic and Abiotic Features on Curcuma longa L. Plantation under Tropical Condition. 3(4), 171–178.
- Syller, J. (2012). Facilitative and antagonistic interactions between plant viruses in mixed infections. *Molecular Plant Pathology*, 13(2), 204–216. https://doi.org/10.1111/j.1364-3703.2011.00734.x
- Tennant, P and Fermin, G. (Ed.). (2015). Viral diseases of tropical and subtropical crops (Vol. 4). CABI.
- Thackray, D. J., Diggle, A. J., Berlandier, F. A., & Jones, R. A. (2004). Forecasting aphid outbreaks and epidemics of Cucumber mosaic virus in lupin crops in a Mediterranean-type environment. *Virus research*, *100*(1), 67-82.
- Thomas, L. and Rajeep, P. (2015). *Turmeric*. ICAR- Indian Institute of Spices Research, Kerala, India.
- THOMAS, J. E. (1986). Purification and properties of ginger chlorotic fleck virus. *Annals of Applied Biology*, *108*(1), 43–50. https://doi.org/10.1111/j.1744-7348.1986.tb01964.x
- Thomas, John E, Services, G., Verification, P., Verification, H., Division, I., Payment, C., ... William, R. (2016). Cucumber Mosaic Virus. Archives of Virology, 1(1), 1–6. https://doi.org/10.1007/s00705-015-2460-9
- Thomson, A. D., & Procter, C. H. (1966). Cucumber mosaic virus in lettuce. *New Zealand Journal of Agricultural Research*, 9(1), 142–144. https://doi.org/10.1080/00288233.1966.10418126

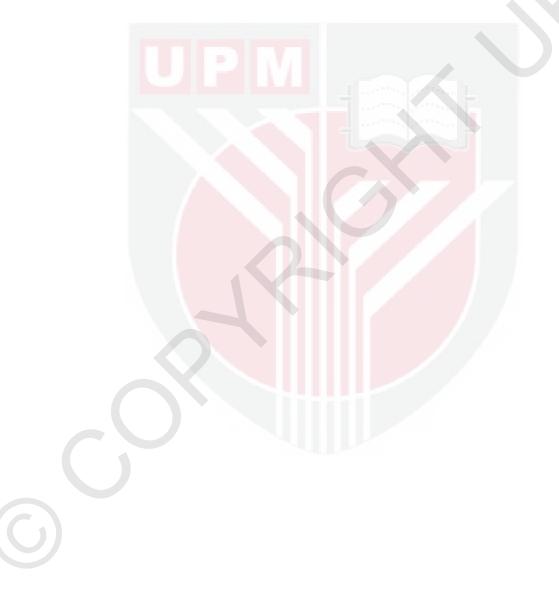
- Thresh, J. M. (2004). Control of plant viral diseases in sub-Saharan Africa: the possibility and feasibility of an integrated approach. *African Crop Science Journal*, *11*(3). https://doi.org/10.4314/acsj.v11i3.27571
- Thresh, J. M. (2006). Control of tropical plant viral diseases. Advances in virus research, 67, 245-295.
- Varadarasan, S., Singh, J., Pradhan, L. N., Gurung, N., & Gupta, S. R. (2000). Bioecology and management of white grub Holotrichia seticollis Mosher (Melolonthinae: Coleoptera), a major pest on ginger in Sikkim. In *Recent Advances in Plantation Crops Research* (pp. 323–326).
- Vargas-Hernandez, M., Macias-Bobadilla, I., Guevara-Gonzalez, R. G., Rico-Garcia, E., Ocampo-Velazquez, R. V., Avila-Juarez, L., & Torres-Pacheco, I. (2020). Nanoparticles as Potential Antivirals in Agriculture. Agriculture, 10(10), 444.
- Vishnoi, R., Kumar, S., & Raj, S. K. (2013). Molecular characterization of a Cucumber mosaic virus isolate associated with mosaic disease of banana in India. *Phytoparasitica*, 41(5), 545–555. https://doi.org/10.1007/s12600-013-0315-z
- Wang, I., Sether, D., Melzer, M., & Borth, W. (2010). First report of banana bract mosaic virus in flowering ginger in Hawaii. *Plant Disease*, 94(7), 921–921. https://doi.org/10.1094/PDIS-94-7-0921A
- Webster, C. G., Wylie, S. J., & Jones, M. G. K. (2004). Diagnosis of plant viral pathogens. *Current Science*.
- Wylie, S., Wilson, C. R., Jones, R. A. C., & Jones, M. G. K. (1993). A polymerase chain reaction assay for cucumber mosaic virus in lupin seeds. *Australian Journal of Agricultural Research*, 44(1), 41–51. https://doi.org/10.1071/AR9930041
- Yadav, A. R., Nawale, R. N., Korake, G. N., & Khandekar, R. G. (2013). Effect of dates of planting and spacing on growth and yield characteristics of ginger (Zingiber officinale Ros.) var. IISR Mahima. 22(2), 209–214.
- Yang, Y., Kim, K. S., & Anderson, E. J. (1997). Seed transmission of cucumber mosaic virus in Spinach. *Phytopathology*, 87(9), 924–931. https://doi.org/10.1094/PHYTO.1997.87.9.924
- Yu, C., Wu, J., & Zhou, X. (2005). Detection and subgrouping of Cucumber mosaic virus isolates by TAS-ELISA and immunocapture RT-PCR. *Journal of Virological Methods*, 123(2), 155–161. https://doi.org/10.1016/j.jviromet.2004.09.014
- Zitter and Murphy. (2009). Cucumber mosaic. *Plant Health Instructor*, 2–3. https://doi.org/10.1094/PHI-I-2009-0518-01

Singh, Z., Jones, R. A. ., & Jones, M.G.K. (1995). Identification of cucumber mosaic virus subgroup I isolates from banana plants affected by infectiuos chlorosis disease using RT-PCR. *Plant Disease*, 79(7), 713–716.



BIODATA OF STUDENT

Buhari Muhammad was born on the 27th August 1983 into the family of Mr. Muhammad Wada Ibrahim and Mrs. Mariyah Abubakar. He attended LEA primary school Tudun Wada, Zaria and Government Secondary School, Zaria, Kaduna State-Nigeria, for his primary and secondary schools' education, respectively. He proceeded to Ahmadu Bello University, Zaria (ABU) where he obtained Bachelor of Agriculture and M.Sc. in Plant Protection certificates in 2009 and 2014, respectively. He is currently a Ph. D. candidate in the Department of Plant Protection, University Putra Malaysia. Buhari is happily married and blessed with children.



LIST OF PUBLICATIONS

International Conference

"Detection and characterization of *Cucumber mosaic virus* (CMV) infecting ginger (*Zingiber officinale* Rosc.) and turmeric (*Curcuma longa* L.) in Peninsular Malaysia". **Buhari Muhammad,** Ganesan Vadamalai, Lau Wei Hong and Kong Lih Ling. Joint Symposium of the 8th International Agriculture Congress and 6th International Symposium for Food and Agriculture 2018 (8TH IAC – 6TH ISFA 2018). 13 – 15 November 2018.

Journal publications

- Buhari Muhammad, Kong Lih Ling, Lau Wei Hong, Ganesan Vadamalai (2021). Detection and Characterization of *Cucumber mosaic virus* Infecting Ginger (*Zingiber officinale* Roscoe) in Malaysia. *International Journal of Sciences: Basic and Applied Research*, 57(1): 9 – 15.
- Buhari Muhammad, Kong Lih Ling, Lau Wei Hong, Ganesan Vadamalai (2021). First Report of *Cucumber mosaic virus* (CMV) infecting Turmeric (*Curcuma longa* L.) in Malaysia. Submitted for publication to the *Journal of Plant Pathology Research*.