



UNIVERSITI PUTRA MALAYSIA

***SYNTHESIS AND CHARACTERIZATION OF ION-IMPRINTED POLYMER
FOR DETERMINATION OF IONIZED CALCIUM IN HUMAN BLOOD
SERUM***

RAFAH JASIM MOHAMMED AL-MAIBD

FS 2021 55



**SYNTHESIS AND CHARACTERIZATION OF ION-IMPRINTED
POLYMER FOR DETERMINATION OF IONIZED CALCIUM IN HUMAN
BLOOD SERUM**

By

RAFAH JASIM MOHAMMED AL-MAIBD

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirements for the Degree of Master of Science**

January 2021

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DEDICATION

To my lovely husband, Wissam, who has supported and encouraged me all the way to finish my Master study at Universiti Putra Malaysia

...I dedicate this research.

Thank you,
My love for you can never be quantified. Allah bless you.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

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RAFAH JASIM MOHAMMED AL-MAIBD

January 2021

Chairman : Sazlinda Kamaruzaman, PhD
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Calcium (Ca) is a vital element in the human body as it maintains the integrity of the bone system and acts as regulatory ion. Total calcium (tCa) circulates the human body in different forms with only ionized calcium (iCa) is the physiology active fraction. Direct measurement of tCa in human serum remains the most common way of assessing calcium status in patients. The majority of clinical laboratories do not have iCa analysers but instead predict iCa from the tCa value measured in the lab. However, one of the pressing issues is the potential of underdiagnoses of Ca deficiency due to the false prediction of iCa.

The adsorption properties of ion-imprinted polymers (IIPs) make them ideal for the selective removal of Calcium ions [Ca(II)] from human blood serum and the subsequent determination of iCa level. Ca(II)-IIP was developed from two naturally formed biopolymers, cellulose and sodium alginate. The polymers were dissolved in co-solvent and casted into films, Ca(II) ions were added to the polymer matrix using CaCl_2 bath, the polymer films were cross-linked using Epichlorohydrin (ECH) and then the template Ca(II) ions were extracted using EDTA. The final Ca(II)-IIP was in the form of white, porous film with high selectivity to Ca(II) ion.

Taguchi method was utilized to determine the optimum adsorption conditions of the Ca(II)-IIP, the resulted optimum conditions were pH 5.9, initial concentration (50 mg/l), dosage (300 mg) and 90 min contact time. The effects of pH, dosage, initial concentration on the Ca(II) adsorption process were investigated. The kinetic study of Ca(II) adsorption fitted well with the pseudo-first-order while, the adsorption isotherm is well fitted with Langmuir isotherm model.

The Ca(II)-IIP has a good selectivity towards Ca(II) in the presence of one or more competing ions. The removal percentage of Ca(II) remains high in the presence of one competing ion but showed a lower removal percentage of Ca(II) ion in the presence of multiple competing ions. The developed Ca(II) exhibits good reusability which can be recycled for 5 times before its efficiency starts to degrade significantly.

The Ca(II)-IIP was successfully applied for the determination of iCa in human blood serum. The optimum adsorption conditions (determined by Taguchi analysis) to achieve the best prediction of iCa level in human blood serum are IIP to serum ratio (60 mg/mL), dilution times (10) and dilution mixing time (30 min).



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SINTESIS DAN KARAKTERISASI POLIMER TERKINI ION UNTUK PENGHASILAN KALSIUM IONISASI DALAM SERUM DARAH MANUSIA

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Kalsium (Ca) adalah elemen penting dalam tubuh manusia kerana ia mengekalkan keutuhan sistem tulang dan bertindak sebagai ion pengawal selia. Kalsium total (tCa) beredar dalam tubuh manusia dalam pelbagai bentuk dan hanya kalsium terion (iCa) wujud sebagai pecahan aktif fisiologi. Pengukuran secara langsung tCa dalam serum manusia merupakan kaedah paling umum bagi menilai status kalsium pesakit. Sebilangan besar makmal klinikal tidak memiliki penganalisis iCa, sebaliknya akan meramal iCa berdasarkan nilai tCa yang diukur di dalam makmal. Namun begitu, antara masalah mendesak yang mungkin timbul ialah diagnosis tersasar tentang kekurangan Ca akibat ramalan iCa yang salah.

Sifat penjerapan polimer tercetak ion (ion-imprinted polymers, IIP) menjadikannya sesuai untuk penyingkiran selektif ion Kalsium [Ca(II)] daripada serum darah manusia dan menentukan tahap iCa yang seterusnya. Ca(II)-IIP telah dihasilkan daripada dua biopolimer yang terbentuk secara semula jadi, iaitu selulosa dan natrium alginat. Polimer-polimer ini dilarutkan dalam pelarut bersama dan dituang menjadi filem nipis, dan ion Ca(II) ditambah ke dalam matriks polimer menggunakan rendaman CaCl₂. Filem polimer yang terhasil diikat silang menggunakan epiklorohidrin (epichlorohydrin, ECH) dan kemudian, templat ion Ca(II) diekstrak menggunakan EDTA. Ca(II)-IIP yang terhasil adalah dalam bentuk filem putih yang berliang dengan kepilahan ion yang tinggi terhadap ion Ca(II).

Kaedah Taguchi digunakan bagi menentukan syarat penjerapan Ca(II)-IIP yang optimum. Syarat optimum yang diperolehi adalah pH 5.9, kepekatan awal (50 mg/L), dos (300 mg) dan tempoh sentuhan selama 90 minit. Kesan pH, dos, dan kepekatan awal terhadap proses penjerapan Ca(II) dikaji. Kajian kinetik penjerapan Ca(II)

padan dengan tertib pseudo pertama, sementara isoterma penjerapan padan dengan model isoterma Langmuir.

Ca(II)-IIP memiliki kepilahan ion yang baik terhadap Ca(II) dengan kewujudan satu atau lebih ion bersaing. Peratusan penyingkiran Ca(II) kekal tinggi dengan kewujudan satu ion bersaing, namun menunjukkan peratusan penyingkiran ion Ca(II) yang lebih rendah dengan kewujudan pelbagai ion bersaing. Ca(II) yang dihasilkan menunjukkan sifat kebolehgunaan semula yang baik dan ia boleh diguna semula sebanyak 5 kali sebelum kecekapannya mula menurun dengan ketara.

Ca(II)-IIP berjaya digunakan bagi menentukan kandungan iCa dalam serum darah manusia. Syarat penjerapan optimum (ditentukan melalui analisis Taguchi) untuk memperoleh ramalan terbaik tahap iCa dalam serum darah manusia adalah nisbah IIP kepada serum (60 mg/mL), pengulangan pencairan (10) dan tempoh pengadunan pencairan (30 minit).

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LIST OF ABBREVIATIONS

4-VP	4-Vinylpyridine
AAS	Atomic absorption spectrophotometry
AIBN	2,2'-azobisisobutyronitrile
ANOM	Analysis of mean
ANOVA	Analysis of variance
APS	3-amino propyl tri-methoxy silane
b	Langmuir adsorption constant
BET	Brunauer Emmett Teller
BPO	Benzoyl peroxide
C_0	Initial concentration
Ca	Calcium
Ca(II)-IIP	Calcium ion imprinted polymer
CE	Cellulose
C_e	Final concentration
C_f	Final concentration
C_F	Final Ca(II) concentration in diluted serum after IIP treatment
C_i	Initial concentration
COVID-19	Coronavirus disease
C_R	Ca(II) concentration of the reference sample (serum without IIP treatment)
DEM	2-(diethylamino) ethyl methacrylate
DMF	Dimethylformamide
DMSO	Dimethyl sulfoxide
DoE	Design of Experiment
DVB	Bi-vinylated monomer divinylbenzene
ECH	Epichlorohydrin

EDMA	3,4-Ethylenedioxy-N-methylamphetamine
EDTA	Ethylenediaminetetraacetic acid
FESEM	Field emission scanning electron microscopy
FMHS	Faculty of medicine and health sciences
FTIR	Fourier transform infrared spectroscopy
HEMA	2-hydroxyethyl methacrylate
iCa	Ionized (free) calcium
ICP-MS	Inductively coupled plasma mass spectrometry
ICP-OES	Inductively coupled plasma - optical emission spectrometry
IIP	Ion imprinted polymer
IPT	Imprinted polymer technology
ISE	Ion-selective electrodes
K'	Effectiveness of imprinting factor
k_1	Pseudo-first-order rate constant
k_2	Pseudo-second-order rate constant
k_3	Intraparticle transport rate constant
K_F	Freundlich constant
MAA	Methacrylic acid
MBA	N,N0-methylene bis (acrylamide)
MCO	Movement control order
MI	Molecular Imprinting
MIP	Molecular imprinting polymer
MIT	Molecular imprinting technology
n	Freundlich exponent
NIIP	Non ion imprinted polymer
PETRA	Pentaerythritol triacrylate
PPE	Percentage prediction error
Q_e	Total adsorption capacity

q_e	Amount of Ca(II) ions removed at equilibrium
Q_{max}	Maximum adsorption capacity
q_t	Amount of Ca(II) ions removed at specific contact time (t)
R^2	Slope
R_L	Separation factor
S	Selectivity coefficient
SA	Sodium alginate
SNR	Signal to Noise Ratio
SPE	Solid-phase extraction
SR	Swelling ratio
Sr	Selectivity ratio
t	Contact time
tCa	Total calcium
TGA	Thermogravimetric analysis
THF	Tetrahydrofuran
TRIM	Trimethylolpropane trimethacrylate
UPM	Universiti Putra Malaysia
V	Volume of the solution
V_f	Final volume
V_i	Initial volume
WIIP	IIP dosage
Wt.%	Weight percentage
XRD	X-ray diffraction

CHAPTER 1

INTRODUCTION

1.1 Background of Research

Calcium is the fifth most abundant element in Earth's crust and critical prevalent cation present in the human body. Calcium has a vital role in the mineralization and maintenance of the skeletal system of the human body. It also plays an essential role in blood coagulation, nerve signal transmission, the excitability of cardiac muscle and the preservation of cell membrane integrity and permeability [1]. Calcium exists in the human extracellular fluid (including serum or plasma) in soluble form as 0.1 wt.% of the body's total calcium mass [2]. Serum or plasma calcium exists in the human body in three fractions: ionized calcium (iCa), complexed or reacted calcium, and protein-bound calcium. Around 40 % of the total circulating calcium in human blood is bound to protein (mainly albumin), 10 % is complexed with different ions with the residual 50 % circulates as free or ionized calcium [2].

Different type of illness causes the concentration of iCa to be disrupted, and the body will be no longer able to supply and maintain the desired levels of calcium for the intracellular functions. The disruption of calcium levels in the body is the leading cause of hypercalcemia or hypocalcaemia to develop [3]. Hence, in most large hospitals, calcium is monitored daily for inpatients with calcium disorder related illness. iCa is commonly tested for patients with low total calcium (tCa) [4].

It is increasingly evidenced that iCa represents the physiologically active fraction of calcium that significantly affects the body's health. The direct measurement of iCa provides a more accurate picture of the calcium level in the human body. Nowadays, tCa can be measured using different modern methods such as spectrophotometry, atomic absorption spectrophotometry [5], and inductively coupled plasma mass spectrometry [6]. In contrast, iCa is mainly measured by potentiometry with ion-selective electrodes (ISEs) in some clinical laboratory [7]. Other methods that depend on mathematical formulas based on statistical data to adjust tCa from albumin is commonly used in clinical laboratories [8].

The development of portable ISE clinical analysers marks the next milestone in the measurement of iCa in clinical laboratories. These analysers utilize disposable cartridges that contain impregnated iCa biosensors for use with whole heparinized blood [9]. Despite further advancement in the ISEs, cost remains a disadvantage derived from frequent equipment maintenance, electrode replacement, and associated downtime [10]; thus, the clinical application of this technique is still limited due to the relatively higher maintenance cost of analysis, high probability of errors from the likely CO₂ losses during sample handling and the subsequent impact on pH and iCa measurement accuracy. Meanwhile, albumin adjusted formulas

remain the commonly used methods in the majority of the clinical laboratory to estimate iCa quickly but less accurately.

1.2 Problem Statement

Measurement of tCa in human blood serum is a routine test for assessing calcium status in patients. Many laboratories do not have iCa analysers but instead utilize published formulas to predict iCa from the tCa value measured in the lab. One of the pressing issues is the potential of underdiagnoses the calcium deficiency due to the false prediction of iCa.

In a recent study, ion imprinting polymer (IIP) was reported as a suitable candidate for the removal of Ca(II) ions from aqueous water samples [11]. It showed adequate adsorption capacity, high selectivity to Ca(II) ion. Unfortunately, the study has not provided full adsorption study to determine the effect of some critical factors such as pH, initial concentration and dosage on the removal of the Ca(II) ions.

Given above, IIP was proposed as an alternative approach for the removal of calcium ions (iCa fraction) from human blood serum for the subsequent determination of iCa level. The present work proposes the synthesis and characterization of IIP in the form of porous film for the initial removal of Ca(II) from aqueous solution and then apply it to human blood serum for the determination of iCa level.

Finally, Taguchi design of experiments (DoE) was utilized in the current study for designing and optimizing all adsorption and application experiments.

1.3 Objectives of the Current Research

The general objective of the current research is to prepare calcium ion-imprinted polymer [Ca(II)IIP] for the determination of iCa level in human blood serum. The following specific objectives are derived to achieve the general aim of the study:

- i. To prepare and characterize Ca(II)-IIP for the selective removal of Ca(II) ion from Ca(II) aqueous solutions.
- ii. To utilize Taguchi DoE for the optimization of the IIP synthesis, adsorption and application processes.
- iii. To carry out adsorption, kinetic, isotherm, reusability and selectivity studies on the Ca(II)-IIP.
- iv. To apply Ca(II)-IIP in the removal of Ca(II) ions from an actual sample (human blood serum) and study the predictivity of iCa level using Ca(II)-IIP.

REFERENCES

- [1.] Buege, M.J., B. Do, H.C. Lee, D.M. Weber, S.B. Horowitz, L. Feng, Y. Qing, and B.R. Shank, *Corrected calcium versus ionized calcium measurements for identifying hypercalcemia in patients with multiple myeloma*. Cancer Treatment and Research Communications, 2019. 21: p. 100159
- [2.] Danner, J.A., *Predictive Model to Estimate Ionized calcium from Routine Serum Biochemical Profile in Dogs*, in *Veterinary Clinical Medicine* 2017, University of Illinois at Urbana: Urbana, Illinois. p. 81.
- [3.] Catalano, A., D. Chilà, F. Bellone, G. Nicocia, G. Martino, I. Loddo, N. Morabito, S. Benvenga, and S. Loddo, *Incidence of hypocalcemia and hypercalcemia in hospitalized patients: Is it changing?* Journal of Clinical & Translational Endocrinology, 2018. 13: p. 9-13.
- [4.] Szymanski, J.J., Z.K. Otroock, K.K. Patel, and M.G. Scott, *Incidence of humoral hypercalcemia of malignancy among hypercalcemic patients with cancer*. Clinica Chimica Acta, 2016. 453: p. 190-193.
- [5.] Baird, G.S., *Ionized calcium*. Clinica Chimica Acta, 2011. 412(9): p. 696-701.
- [6.] Yan, Y., M. Ge, R. Ma, H. Zhao, D. Wang, C. Hu, J. Wang, W. Chen, and C. Zhang, *A candidate reference method for serum calcium measurement by inductively coupled plasma mass spectrometry*. Clinica Chimica Acta, 2016. 461: p. 141-145.
- [7.] D'Orazio, P., H. Visnick, and S. Balasubramanian, *Accuracy of commercial blood gas analyzers for monitoring ionized calcium at low concentrations*. Clinica Chimica Acta, 2016. 461: p. 34-40.
- [8.] Rivara, M.B., V. Ravel, K. Kalantar-Zadeh, E. Streja, W.L. Lau, A.R. Nissenson, B. Kestenbaum, I.H. de Boer, J. Himmelfarb, and R. Mehrotra, *Uncorrected and Albumin-Corrected Calcium, Phosphorus, and Mortality in Patients Undergoing Maintenance Dialysis*. J Am Soc Nephrol, 2015. 26(7): p. 1671-81.
- [9.] Tappin, S., F. Rizzo, S. Dodkin, K. Papasouliotis, S. Tasker, and K. Murphy, *Measurement of ionized calcium in canine blood samples collected in prefilled and self-filled heparinized syringes using the i-STAT point-of-care analyzer*. Vet Clin Pathol, 2008. 37(1): p. 66-72.
- [10.] Björkman, M.P., A.J. Sorva, and R.S. Tilvis, *Calculated serum calcium is an insufficient surrogate for measured ionized calcium*. Archives of Gerontology and Geriatrics, 2009. 49(3): p. 348-350.
- [11.] Wang, Y., S. Zhu, Y. Liao, and X. Xiong, *A calcium ion-imprinted porous film prepared from a cellulose-alginate composite*. Journal of Polymer Research, 2014. 21(12): p. 612.

- [12.] Rosol, T.J. and C.C. Capen, *Pathophysiology of Calcium, Phosphorus, and Magnesium Metabolism in Animals*. Veterinary Clinics of North America: Small Animal Practice, 1996. 26(5): p. 1155-1184.
- [13.] de Brito Galvão, J.F., P.A. Schenck, and D.J. Chew, *A Quick Reference on Hypocalcemia*. Veterinary Clinics of North America: Small Animal Practice, 2017. 47(2): p. 249-256.
- [14.] Pfitzenmeyer, P., I. Martin, P. d'Athis, Y. Grumbach, M.-C. Delmestre, F. Blondé-Cynober, B. Derycke, and L. Brondel, *A new formula for correction of total calcium level into ionized serum calcium values in very elderly hospitalized patients*. Archives of Gerontology and Geriatrics, 2007. 45(2): p. 151-157.
- [15.] Li, M., X. Meng, X. Liang, J. Yuan, X. Hu, Z. Wu, and X. Yuan, *A novel In(III) ion-imprinted polymer (IIP) for selective extraction of In(III) ions from aqueous solutions*. Hydrometallurgy, 2018. 176: p. 243-252.
- [16.] Shifrin, A., *Chapter 6 - Brief Overview of Calcium, Vitamin D, Parathyroid Hormone Metabolism, and Calcium-Sensing Receptor Function*, in *Advances in Treatment and Management in Surgical Endocrinology*, A.L. Shifrin, Editor. 2020, Elsevier. p. 63-70.
- [17.] Titchenal, C.A. and J. Dobbs, *A system to assess the quality of food sources of calcium*. Journal of Food Composition and Analysis, 2007. 20(8): p. 717-724.
- [18.] Ross, A.C., *The 2011 report on dietary reference intakes for calcium and vitamin D*. Public Health Nutr, 2011. 14(5): p. 938-9.
- [19.] Brown, E.M., *Extracellular Ca²⁺ sensing, regulation of parathyroid cell function, and role of Ca²⁺ and other ions as extracellular (first) messengers*. Physiol Rev, 1991. 71(2): p. 371-411.
- [20.] Gelli, R., F. Ridi, and P. Baglioni, *The importance of being amorphous: calcium and magnesium phosphates in the human body*. Advances in Colloid and Interface Science, 2019. 269: p. 219-235.
- [21.] Inzucchi, S.E., *Understanding hypercalcemia. Its metabolic basis, signs, and symptoms*. Postgrad Med, 2004. 115(4): p. 69-70, 73-6.
- [22.] Higgins, C. *Ionized calcium*. July 2007.
- [23.] Schreckenber, R. and K.-D. Schlüter, *Calcium sensing receptor expression and signalling in cardiovascular physiology and disease*. Vascular Pharmacology, 2018. 107: p. 35-42.
- [24.] Schenck, P., D. Chew, L. Nagode, and T. Rosol, *Disorders of Calcium: Hypercalcemia and Hypocalcemia*. Fluid Therapy in Small Animal Practice, 2006: p. 122-194.

- [25.] Hastbacka, J. and V. Pettila, *Prevalence and predictive value of ionized hypocalcemia among critically ill patients*. Acta Anaesthesiol Scand, 2003. 47(10): p. 1264-9.
- [26.] Suh, S., J.C. Bae, S.-M. Jin, J.H. Jee, M.K. Park, D.K. Kim, and J.H. Kim, *Serum calcium changes and risk of type 2 diabetes mellitus in Asian population*. Diabetes Research and Clinical Practice, 2017. 133: p. 109-114.
- [27.] Kimura, S., T. Iwasaki, K. Oe, K. Shimizu, T. Suemori, T. Kanazawa, N. Shioji, Y. Kuroe, Y. Matsuoka, and H. Morimatsu, *High Ionized Calcium Concentration Is Associated With Prolonged Length of Stay in the Intensive Care Unit for Postoperative Pediatric Cardiac Patients*. Journal of Cardiothoracic and Vascular Anesthesia, 2018. 32(4): p. 1667-1675.
- [28.] Rudnicki, M., A. Frolich, A. Haaber, and J. Thode, *Actual ionized calcium (at actual pH) vs adjusted ionized calcium (at pH 7.4) in hemodialyzed patients*. Clin Chem, 1992. 38(7): p. 1384.
- [29.] Thode, J., S.N. Holmegaard, I. Transbol, N. Fogh-Andersen, and O. Siggaard-Andersen, *Adjusted ionized calcium (at pH 7.4) and actual ionized calcium (at actual pH) in capillary blood compared for clinical evaluation of patients with disorders of calcium metabolism*. Clin Chem, 1990. 36(3): p. 541-4.
- [30.] Pfitzenmeyer, P., I. Martin, P. d'Athis, Y. Grumbach, M.C. Delmestre, F. Blonde-Cynober, B. Derycke, L. Brondel, and N. Club Francophone de Geriatrie et, *A new formula for correction of total calcium level into ionized serum calcium values in very elderly hospitalized patients*. Arch Gerontol Geriatr, 2007. 45(2): p. 151-7.
- [31.] Zheng, Y., F. Zhuang, Q. Zhu, S. Ma, Y. Xu, J. Lu, G. Hao, Y. Gu, C. Hao, M. Zhu, and F. Ding, *Albumin-corrected total/ionized calcium ratio is not superior to total/ionized calcium ratio as an indicator of citrate accumulation*. Int J Artif Organs, 2017. 40(11): p. 602-606.
- [32.] Payne, R.B., A.J. Little, R.B. Williams, and J.R. Milner, *Interpretation of serum calcium in patients with abnormal serum proteins*. Br Med J, 1973. 4(5893): p. 643-6.
- [33.] Costa, M., C. Lanna, M. Braga, and S. Magalhães, *Assessment of asymptomatic hypercalcemia in outpatients*. Jornal Brasileiro de Patologia e Medicina Laboratorial, 2008. 44: p. 329-335.
- [34.] Bushinsky, D.A. and R.D. Monk, *Electrolyte quintet: Calcium*. Lancet, 1998. 352(9124): p. 306-11.
- [35.] Hastings, F.C.M.a.A.B., *The classic: the state of calcium in the fluids of the body. I. The conditions affecting the ionization of calcium*. Clin Orthop Relat Res, 1970. 69: p. 4-27.

- [36.] Byrnes, M.C., K. Huynh, S.D. Helmer, C. Stevens, J.M. Dort, and R.S. Smith, *A comparison of corrected serum calcium levels to ionized calcium levels among critically ill surgical patients*. Am J Surg, 2005. 189(3): p. 310-4.
- [37.] Basseto, T.P., L.A. Azzalis, E.C. Pereira, V.B.C. Junqueira, L.Z. Caputo, A.L.A. Fonseca, D. Feder, and F.L.A. Fonseca, *Comparison between two methods of ionized calcium measurement in newborns*. Jornal Brasileiro de Patologia e Medicina Laboratorial, 2013. 49: p. 317-319.
- [38.] Ijaz, A., T. Mehmood, A. Qureshi, M. Anwar, M. Dilawar, I. Hussain, F. Khan, D. Khan, S. Hussain, and I. Khan, *Estimation of ionized calcium, total calcium and albumin corrected calcium for the diagnosis of hypercalcaemia of malignancy*. Journal of the College of Physicians and Surgeons--Pakistan : JCPSP, 2006. 16: p. 49-52.
- [39.] Evenepoel, P., B. Bammens, K. Claes, D. Kuypers, B.K. Meijers, and Y. Vanrenterghem, *Measuring total blood calcium displays a low sensitivity for the diagnosis of hypercalcemia in incident renal transplant recipients*. Clin J Am Soc Nephrol, 2010. 5(11): p. 2085-92.
- [40.] Jain, A., S. Bhayana, M. Vlasschaert, and A. House, *A formula to predict corrected calcium in haemodialysis patients*. Nephrol Dial Transplant, 2008. 23(9): p. 2884-8.
- [41.] Bienzle, D., R.M. Jacobs, and J.H. Lumsden, *Relationship of serum total calcium to serum albumin in dogs, cats, horses and cattle*. Can Vet J, 1993. 34(6): p. 360-4.
- [42.] Meuten, D.J., D.J. Chew, C.C. Capen, and G.J. Kociba, *Relationship of serum total calcium to albumin and total protein in dogs*. J Am Vet Med Assoc, 1982. 180(1): p. 63-7.
- [43.] Carreira, L.M., D. Dias, and P. Azevedo, *Relationship Between Gender, Age, and Weight and the Serum Ionized Calcium Variations in Dog Periodontal Disease Evolution*. Topics in Companion Animal Medicine, 2015. 30(2): p. 51-56.
- [44.] Weller, R.E., R.L. Buschbom, H.A. Ragan, J.F. Baer, and C.A. Malaga, *Relationship of serum total calcium and albumin and total protein in owl monkeys (Aotus nancymai)*. J Med Primatol, 1990. 19(5): p. 439-46.
- [45.] Berlin, D. and I. Aroch, *Concentrations of ionized and total magnesium and calcium in healthy horses: Effects of age, pregnancy, lactation, pH and sample type*. The Veterinary Journal, 2009. 181(3): p. 305-311.
- [46.] Szenci, O., B.P. Chew, A.C. Bajcsy, P. Szabo, and E. Brydl, *Total and ionized calcium in parturient dairy cows and their calves*. J Dairy Sci, 1994. 77(4): p. 1100-5.

- [47.] Jafri, L., A.H. Khan, and S. Azeem, *Ionized calcium measurement in serum and plasma by ion selective electrodes: comparison of measured and calculated parameters*. Indian journal of clinical biochemistry : IJCB, 2014. 29(3): p. 327-332.
- [48.] Wang, B., D. Li, Y. Gong, B. Ying, and B. Cheng, *Association of serum total and ionized calcium with all-cause mortality in critically ill patients with acute kidney injury*. Clinica Chimica Acta, 2019. 494: p. 94-99.
- [49.] Moore, E.W., *Ionized calcium in normal serum, ultrafiltrates, and whole blood determined by ion-exchange electrodes*. The Journal of clinical investigation, 1970. 49(2): p. 318-334.
- [50.] Looney, A.L., J. Ludders, H.N. Erb, R. Gleed, and P. Moon, *Use of a handheld device for analysis of blood electrolyte concentrations and blood gas partial pressures in dogs and horses*. J Am Vet Med Assoc, 1998. 213(4): p. 526-30.
- [51.] Lindemans, J., P. Hoefkens, A.L. van Kessel, M. Bonnay, W.R. Kulpmann, and J.D. van Suijlen, *Portable blood gas and electrolyte analyzer evaluated in a multiinstitutional study*. Clin Chem, 1999. 45(1): p. 111-7.
- [52.] Zamani, H., J. Abedini-Torghabeh, and M. Ganjali, *A Highly Selective and Sensitive Calcium(II)-Selective PVC Membrane Based on Dimethyl 1-(4-Nitrobenzoyl)-8-oxo-2,8-dihydro-1H-pyrazolo[5,1-a]isoindole-2,3-dicarboxylate as a Novel Ionophore*. Bulletin of the Korean Chemical Society, 2006. 27.
- [53.] Schenck, P.A., D.J. Chew, and C.L. Brooks, *Effects of storage on serum ionized calcium and pH values in clinically normal dogs*. Am J Vet Res, 1995. 56(3): p. 304-7.
- [54.] Unterer, S., H. Lutz, B. Gerber, T.M. Glaus, M. Hassig, and C.E. Reusch, *Evaluation of an electrolyte analyzer for measurement of ionized calcium and magnesium concentrations in blood, plasma, and serum of dogs*. Am J Vet Res, 2004. 65(2): p. 183-7.
- [55.] Tung, J.K. and R.A.R. Bowen, *Impact of underfilling heparinized collection tubes on ionized calcium measurements*. Clin Chim Acta, 2019.
- [56.] Dewitte, K., D. Stöckl, and L.M. Thienpont, *pH dependency of serum ionised calcium*. The Lancet, 1999. 354(9192): p. 1793-1794.
- [57.] Burnett, R.W., T.F. Christiansen, A.K. Covington, N. Fogh-Andersen, W.R. Kulpmann, A. Lewenstam, A.H.J. Maas, O. Muller-Plathe, C. Sachs, O.S. Andersen, A.L. VanKessel, W.G. Zijlstra, C. International Federation of Clinical, and W.G.o.S.E. Laboratory Medicine. Ifcc Scientific Division, *IFCC recommended reference method for the determination of the substance concentration of ionized calcium in undiluted serum, plasma or whole blood*. Clin Chem Lab Med, 2000. 38(12): p. 1301-14.

- [58.] Paul D'Orazio, J.G. Toffaletti, and Jesper Wandrup, *Ionized Calcium Determinations: Precollection Variables, Specimen Choice, Collection, and Handling; Approved Guideline*. Vol. 21. 2001, USA: NCCLS.
- [59.] Iacob, B.-C., E. Bodoki, C. Farcau, L. Barbu-Tudoran, and R. Oprean, *Study of the Molecular Recognition Mechanism of an Ultrathin MIP Film-Based Chiral Electrochemical Sensor*. *Electrochimica Acta*, 2016. 217: p. 195-202.
- [60.] Zhu, G.-j., H.-y. Tang, H.-l. Zhang, L.-l. Pei, P. Zhou, Y.-l. Shi, Z.-h. Cai, H.-b. Xu, and Y. Zhang, *A novel ion-imprinted polymer for selective removal of trace Fe(III) from Cr(III)-containing solutions*. *Hydrometallurgy*, 2019. 186: p. 105-114.
- [61.] Polyakov, M.V., *Adsorption properties and structure of silica gel*. *Zhurnal fizicheskoi khimii*, 1931: p. 799-804.
- [62.] Wulff, G. and A. Sarhan, *The use of polymers with enzyme-analogous structures for the resolution of racemates*. *Angew. Chem. Int. Ed.*, 1972. 11: p. 341-344.
- [63.] Fu, J., L. Chen, J. Li, and Z. Zhang, *Current status and challenges of ion imprinting*. *Journal of Materials Chemistry A*, 2015. 3(26): p. 13598-13627.
- [64.] Hiroyuki, N., D. Jichio, and T. Eishun, *SELECTIVE ADSORPTION OF METAL IONS ON CROSSLINKED POLY(VINYLPYRIDINE) RESIN PREPARED WITH A METAL ION AS A TEMPLATE*. *Chemistry Letters*, 1976. 5(2): p. 169-174.
- [65.] Sagiv, J., *Organized Monolayers by Adsorption. III. Irreversible Adsorption and Memory Effects in Skeletonized Silane Monolayers*. *Israel Journal of Chemistry*, 1979. 18(3-4): p. 346-353.
- [66.] Vlatakis, G., L.I. Andersson, R. Müller, and K. Mosbach, *Drug assay using antibody mimics made by molecular imprinting*. *Nature*, 1993. 361(6413): p. 645-647.
- [67.] Whitcombe, M.J., M.E. Rodriguez, P. Villar, and E.N. Vulfson, *A New Method for the Introduction of Recognition Site Functionality into Polymers Prepared by Molecular Imprinting: Synthesis and Characterization of Polymeric Receptors for Cholesterol*. *Journal of the American Chemical Society*, 1995. 117(27): p. 7105-7111.
- [68.] Sreenivasan, K. and R. Sivakumar, *Imparting recognition sites in poly(HEMA) for two compounds through molecular imprinting*. *Journal of Applied Polymer Science*, 1999. 71(11): p. 1823-1826.
- [69.] Mafu, L.D., T.A. Msagati, and B.B. Mamba, *Ion-imprinted polymers for environmental monitoring of inorganic pollutants: synthesis, characterization, and applications*. *Environ Sci Pollut Res Int*, 2013. 20(2): p. 790-802.

- [70.] Saraji, M. and H. Yousefi, *Selective solid-phase extraction of Ni(II) by an ion-imprinted polymer from water samples*. Journal of Hazardous Materials, 2009. 167(1): p. 1152-1157.
- [71.] Rao, T.P., R. Kala, and S. Daniel, *Metal ion-imprinted polymers--novel materials for selective recognition of inorganics*. Anal Chim Acta, 2006. 578(2): p. 105-16.
- [72.] Andaç, M., E. Özyapı, S. Şenel, R. Say, and A. Denizli, *Ion-Selective Imprinted Beads for Aluminum Removal from Aqueous Solutions*. Industrial & Engineering Chemistry Research, 2006. 45(5): p. 1780-1786.
- [73.] Saatçılar, Ö., N. Şatiroğlu, R. Say, S. Bektaş, and A. Denizli, *Binding behavior of Fe³⁺ ions on ion-imprinted polymeric beads for analytical applications*. Journal of Applied Polymer Science, 2006. 101(5): p. 3520-3528.
- [74.] Özkara, S., R. Say, C. Andaç, and A. Denizli, *An Ion-Imprinted Monolith for in Vitro Removal of Iron out of Human Plasma with Beta Thalassemia*. Industrial & Engineering Chemistry Research, 2008. 47(20): p. 7849-7856.
- [75.] Germiniano, T.O., M.Z. Corazza, M.G. Segatelli, E.S. Ribeiro, M.J.S. Yabe, E. Galunin, and C.R.T. Tarley, *Synthesis of novel copper ion-selective material based on hierarchically imprinted cross-linked poly(acrylamide-co-ethylene glycol dimethacrylate)*. Reactive and Functional Polymers, 2014. 82: p. 72-80.
- [76.] Walas, S., A. Tobiasz, M. Gawin, B. Trzewik, M. Strojny, and H. Mrowiec, *Application of a metal ion-imprinted polymer based on salen-Cu complex to flow injection preconcentration and FAAS determination of copper*. Talanta, 2008. 76(1): p. 96-101.
- [77.] Yasinzai, M., G. Mustafa, N. Asghar, I. Ullah, M. Zahid, P.A. Lieberzeit, D. Han, and U. Latif, *Ion-Imprinted Polymer-Based Receptors for Sensitive and Selective Detection of Mercury Ions in Aqueous Environment*. Journal of Sensors, 2018. 2018: p. 6.
- [78.] Godlewska-Żyłkiewicz, B., E. Zambrzycka, B. Leśniewska, and A.Z. Wilczewska, *Separation of ruthenium from environmental samples on polymeric sorbent based on imprinted Ru(III)-allyl acetoacetate complex*. Talanta, 2012. 89: p. 352-359.
- [79.] Fayazi, M., M. Ghanei-Motlagh, M.A. Taher, R. Ghanei-Motlagh, and M.R. Salavati, *Synthesis and application of a novel nanostructured ion-imprinted polymer for the preconcentration and determination of thallium(I) ions in water samples*. Journal of Hazardous Materials, 2016. 309: p. 27-36.

- [80.] Chauvin, A.-S., J.-C.G. Bünzli, F. Bochud, R. Scopelliti, and P. Froidevaux, *Use of Dipicolinate-Based Complexes for Producing Ion-Imprinted Polystyrene Resins for the Extraction of Yttrium-90 and Heavy Lanthanide Cations*. Chemistry – A European Journal, 2006. 12(26): p. 6852-6864.
- [81.] Sebastian, M. and B. Mathew, *Carbon nanotube-based ion imprinted polymer as electrochemical sensor and sorbent for Zn(II) ion from paint industry wastewater*. International Journal of Polymer Analysis and Characterization, 2018. 23(1): p. 18-28.
- [82.] Otero-Romaní, J., A. Moreda-Piñeiro, P. Bermejo-Barrera, and A. Martín-Esteban, *Ionic imprinted polymer for nickel recognition by using the bi-functionalized 5-vinyl-8-hydroxyquinoline as a monomer: Application as a new solid phase extraction support*. Microchemical Journal, 2009. 93(2): p. 225-231.
- [83.] Wang, J., J. Wei, and J. Li, *Straw-supported ion imprinted polymer sorbent prepared by surface imprinting technique combined with AGET ATRP for selective adsorption of La³⁺ ions*. Chemical Engineering Journal, 2016. 293: p. 24-33.
- [84.] James, D., J.M. Gladis, A.K. Pandey, G.R.K. Naidu, and T. Prasada Rao, *Design of two-dimensional biomimetic uranyl optrode and its application to the analysis of natural waters*. Talanta, 2008. 74(5): p. 1420-1427.
- [85.] Okay, O., *Macroporous copolymer networks*. Progress in Polymer Science, 2000. 25(6): p. 711-779.
- [86.] Baghel, A., M. Boopathi, B. Singh, P. Pandey, T.H. Mahato, P.K. Gutch, and K. Sekhar, *Synthesis and characterization of metal ion imprinted nanoporous polymer for the selective recognition of copper*. Biosensors and Bioelectronics, 2007. 22(12): p. 3326-3334.
- [87.] Tobiasz, A., S. Walas, B. Trzewik, P. Grzybek, M.M. Zaitz, M. Gawin, and H. Mrowiec, *Cu(II)-imprinted styrene–divinylbenzene beads as a new sorbent for flow injection-flame atomic absorption determination of copper*. Microchemical Journal, 2009. 93(1): p. 87-92.
- [88.] Gomes, A.C.S.A., L.C. Costa, D.C. Brito, R.J. França, and M.R.C. Marques, *Development of a new ion-imprinted polymer (IIP) with Cd²⁺ ions based on divinylbenzene copolymers containing amidoxime groups*. Polymer Bulletin, 2019.
- [89.] Lopes Pinheiro, S.C., A.B. Descalzo, I.M. Raimundo, G. Orellana, and M.C. Moreno-Bondí, *Fluorescent ion-imprinted polymers for selective Cu(II) optosensing*. Analytical and Bioanalytical Chemistry, 2012. 402(10): p. 3253-3260.

- [90.] Lin, C.X., M.H. Liu, and H.Y. Zhan, *Pb(II)-Imprinted Polymer Prepared by Graft Copolymerization of Acrylic Acid onto Cellulose*. Advanced Materials Research, 2011. 295-297: p. 2045-2048.
- [91.] Zhai, Y., Y. Liu, X. Chang, S. Chen, and X. Huang, *Selective solid-phase extraction of trace cadmium(II) with an ionic imprinted polymer prepared from a dual-ligand monomer*. Analytica Chimica Acta, 2007. 593(1): p. 123-128.
- [92.] Zhu, L., Z. Zhu, R. Zhang, J. Hong, and Y. Qiu, *Synthesis and adsorption performance of lead ion-imprinted micro-beads with combination of two functional monomers*. Journal of Environmental Sciences, 2011. 23(12): p. 1955-1961.
- [93.] Dakova, I., I. Karadjova, V. Georgieva, and G. Georgiev, *Ion-imprinted polymethacrylic microbeads as new sorbent for preconcentration and speciation of mercury*. Talanta, 2009. 78(2): p. 523-529.
- [94.] Guo, J., J. Cai, and Q. Su, *Ion imprinted polymer particles of neodymium: synthesis, characterization and selective recognition*. Journal of Rare Earths, 2009. 27(1): p. 22-27.
- [95.] Alizadeh, T., *An imprinted polymer for removal of Cd²⁺ from water samples: Optimization of adsorption and recovery steps by experimental design*. Chinese Journal of Polymer Science, 2011. 29(6): p. 658.
- [96.] Branger, C., W. Meouche, and A. Margaillan, *Recent advances on ion-imprinted polymers*. Reactive and Functional Polymers, 2013. 73(6): p. 859-875.
- [97.] Rao, T.P., R. Kala, and S. Daniel, *Metal ion-imprinted polymers—Novel materials for selective recognition of inorganics*. Analytica Chimica Acta, 2006. 578(2): p. 105-116.
- [98.] Rahman, M.L., P.Y. Puah, M.S. Sarjadi, S.E. Arshad, B. Musta, and S.M. Sarkar, *Ion-Imprinted Polymer for Selective Separation of Cerium(III) Ions from Rare Earth Mixture*. Journal of Nanoscience and Nanotechnology, 2019. 19(9): p. 5796-5802.
- [99.] Maciejewska, M. and J. Gawdzik, *Preparation and Characterization of Sorption Properties of Porous Microspheres of 1-Vinyl-2-Pyrrolidone-Divinylbenzene*. Journal of Liquid Chromatography & Related Technologies, 2008. 31(7): p. 950-961.
- [100.] Ahmadi, E., J. Gatabi, and Z. Mohamadnia, *Preparation and characterization of Zn(II) ion-imprinted polymer based on salicylic acrylate for recovery of Zn(II) ions*. Polímeros, 2016. 26: p. 242-248.

- [101.] Ara, B., M. Muhammad, M. Salman, R. Ahmad, N. Islam, and T.u.H. Zia, *Preparation of microspheric Fe(III)-ion imprinted polymer for selective solid phase extraction*. Applied Water Science, 2018. 8(1): p. 41.
- [102.] Hande, P.E., A.B. Samui, and P.S. Kulkarni, *Selective nanomolar detection of mercury using coumarin based fluorescent Hg(II)—Ion imprinted polymer*. Sensors and Actuators B: Chemical, 2017. 246: p. 597-605.
- [103.] Meouche, W., K. Laatikainen, A. Margaillan, T. Silvonen, H. Siren, T. Sainio, I. Beurroies, R. Denoyel, and C. Branger, *Effect of porogen solvent on the properties of nickel ion imprinted polymer materials prepared by inverse suspension polymerization*. European Polymer Journal, 2017. 87: p. 124-135.
- [104.] Yusoff, M.M., N.R.N. Mostapa, M.S. Sarkar, T.K. Biswas, M.L. Rahman, S.E. Arshad, M.S. Sarjadi, and A.D. Kulkarni, *Synthesis of ion imprinted polymers for selective recognition and separation of rare earth metals*. Journal of Rare Earths, 2017. 35(2): p. 177-186.
- [105.] Singh, D.K. and S. Mishra, *Synthesis and characterization of Hg(II)-ion-imprinted polymer: Kinetic and isotherm studies*. Desalination, 2010. 257(1): p. 177-183.
- [106.] Liu, Y., Z. Liu, J. Dai, J. Gao, J. Xie, and Y. Yan, *Selective Adsorption of Co(II) by Mesoporous Silica SBA-15-Supported Surface Ion Imprinted Polymer: Kinetics, Isotherms, and Thermodynamics Studies*. Chinese Journal of Chemistry, 2011. 29(3): p. 387-398.
- [107.] Ebrahimzadeh, H., M. Behbahani, Y. Yamini, L. Adlnasab, and A.A. Asgharinezhad, *Optimization of Cu(II)-ion imprinted nanoparticles for trace monitoring of copper in water and fish samples using a Box–Behnken design*. Reactive and Functional Polymers, 2013. 73(1): p. 23-29.
- [108.] Monier, M., I.M. Kenawy, and M.A. Hashem, *Synthesis and characterization of selective thiourea modified Hg(II) ion-imprinted cellulosic cotton fibers*. Carbohydr Polym, 2014. 106: p. 49-59.
- [109.] Zhang, T., X. Yue, K. Zhang, F. Zhao, Y. Wang, and K. Zhang, *Synthesis of Cu(II) ion-imprinted polymers as solid phase adsorbents for deep removal of copper from concentrated zinc sulfate solution*. Hydrometallurgy, 2017. 169: p. 599-606.
- [110.] Ahamed, M.E.H., X.Y. Mbianda, A.F. Mulaba-Bafubiandi, and L. Marjanovic, *Ion imprinted polymers for the selective extraction of silver(I) ions in aqueous media: Kinetic modeling and isotherm studies*. Reactive and Functional Polymers, 2013. 73(3): p. 474-483.
- [111.] Jiang, Y. and D. Kim, *Effect of solvent/monomer feed ratio on the structure and adsorption properties of Cu²⁺-imprinted microporous polymer particles*. Chemical Engineering Journal, 2011. 166(1): p. 435-444.

- [112.] Alizadeh, T. and S. Amjadi, *Preparation of nano-sized Pb²⁺ imprinted polymer and its application as the chemical interface of an electrochemical sensor for toxic lead determination in different real samples*. Journal of Hazardous Materials, 2011. 190(1): p. 451-459.
- [113.] Abdullah, A. Balouch, F.N. Talpur, A. Kumar, M.T. Shah, A.M. Mahar, and Amina, *Synthesis of ultrasonic-assisted lead ion imprinted polymer as a selective sorbent for the removal of Pb²⁺ in a real water sample*. Microchemical Journal, 2019. 146: p. 1160-1168.
- [114.] Hu, S., X. Xiong, S. Huang, and X. Lai, *Preparation of Pb(II) Ion Imprinted Polymer and Its Application as the Interface of an Electrochemical Sensor for Trace Lead Determination*. Analytical Sciences, 2016. 32(9): p. 975-980.
- [115.] Dam, A.H. and D. Kim, *Metal ion-imprinted polymer microspheres derived from copper methacrylate for selective separation of heavy metal ions*. Journal of Applied Polymer Science, 2008. 108(1): p. 14-24.
- [116.] Hoai, N.T. and D. Kim, *Synthesis, structure, and selective separation behavior of copper-imprinted microporous polymethacrylate beads*. AIChE Journal, 2009. 55(12): p. 3248-3254.
- [117.] Singh, D.K. and S. Mishra, *Synthesis, characterization and analytical applications of Ni(II)-ion imprinted polymer*. Applied Surface Science, 2010. 256(24): p. 7632-7637.
- [118.] Tamahkar, E., M. Bakhshpour, M. Andaç, and A. Denizli, *Ion imprinted cryogels for selective removal of Ni(II) ions from aqueous solutions*. Separation and Purification Technology, 2017. 179: p. 36-44.
- [119.] Jalilzadeh, M. and S. Şenel, *Removal of Cu(II) ions from water by ion-imprinted magnetic and non-magnetic cryogels: A comparison of their selective Cu(II) removal performances*. Journal of Water Process Engineering, 2016. 13: p. 143-152.
- [120.] Candan, N., N. Tüzmen, M. Andac, C.A. Andac, R. Say, and A. Denizli, *Cadmium removal out of human plasma using ion-imprinted beads in a magnetic column*. Materials Science and Engineering: C, 2009. 29(1): p. 144-152.
- [121.] Birlik, E., A. Ersöz, E. Açıkkalp, A. Denizli, and R. Say, *Cr(III)-imprinted polymeric beads: Sorption and preconcentration studies*. Journal of Hazardous Materials, 2007. 140(1): p. 110-116.
- [122.] Andaç, M., S. Mirel, S. Şenel, R. Say, A. Ersöz, and A. Denizli, *Ion-imprinted beads for molecular recognition based mercury removal from human serum*. International Journal of Biological Macromolecules, 2007. 40(2): p. 159-166.

- [123.] Bhaskarapillai, A., N.V. Sevilimedu, and B. Sellergren, *Synthesis and Characterization of Imprinted Polymers for Radioactive Waste Reduction*. Industrial & Engineering Chemistry Research, 2009. 48(8): p. 3730-3737.
- [124.] Biju, V.M., J.M. Gladis, and T.P. Rao, *Ion imprinted polymer particles: synthesis, characterization and dysprosium ion uptake properties suitable for analytical applications*. Analytica Chimica Acta, 2003. 478(1): p. 43-51.
- [125.] Otero-Romaní, J., A. Moreda-Piñeiro, P. Bermejo-Barrera, and A. Martín-Esteban, *Synthesis, characterization and evaluation of ionic-imprinted polymers for solid-phase extraction of nickel from seawater*. Analytica Chimica Acta, 2008. 630(1): p. 1-9.
- [126.] Zambrzycka-Szelewa, E., B. Leśniewska, and B. Godlewska-Żyłkiewicz, *Chapter Ten - Preparation and application of ion-imprinted polymer sorbents in separation process of trace metals*, in *Comprehensive Analytical Chemistry*, M. Maré, Editor. 2019, Elsevier. p. 261-293.
- [127.] Metilda, P., J.M. Gladis, G. Venkateswaran, and T. Prasada Rao, *Investigation of the role of chelating ligand in the synthesis of ion-imprinted polymeric resins on the selective enrichment of uranium(VI)*. Analytica Chimica Acta, 2007. 587(2): p. 263-271.
- [128.] Yuan, Y., Y. Yang, X. Ma, Q. Meng, L. Wang, S. Zhao, and G. Zhu, *Molecularly Imprinted Porous Aromatic Frameworks and Their Composite Components for Selective Extraction of Uranium Ions*. Advanced Materials, 2018. 30(12): p. 1706507.
- [129.] Gawin, M., J. Konefał, B. Trzewik, S. Walas, A. Tobiasz, H. Mrowiec, and E. Witek, *Preparation of a new Cd(II)-imprinted polymer and its application to determination of cadmium(II) via flow-injection-flame atomic absorption spectrometry*. Talanta, 2010. 80(3): p. 1305-1310.
- [130.] Moussa, M., V. Pichon, C. Mariet, T. Vercouter, and N. Delaunay, *Potential of ion imprinted polymers synthesized by trapping approach for selective solid phase extraction of lanthanides*. Talanta, 2016. 161: p. 459-468.
- [131.] Luo, X., S. Luo, Y. Zhan, H. Shu, Y. Huang, and X. Tu, *Novel Cu (II) magnetic ion imprinted materials prepared by surface imprinted technique combined with a sol-gel process*. Journal of Hazardous Materials, 2011. 192(3): p. 949-955.
- [132.] Fu, X.-C., J. Wu, L. Nie, C.-G. Xie, J.-H. Liu, and X.-J. Huang, *Electropolymerized surface ion imprinting films on a gold nanoparticles/single-wall carbon nanotube nanohybrids modified glassy carbon electrode for electrochemical detection of trace mercury(II) in water*. Analytica Chimica Acta, 2012. 720: p. 29-37.

- [133.] Yu, K.Y., K. Tsukagoshi, M. Maeda, and M. Takagi, *Metal Ion-Imprinted Microspheres Prepared by Reorganization of the Coordinating Groups on the Surface*. Analytical Sciences, 1992. 8(5): p. 701-703.
- [134.] Dam, H.A. and D. Kim, *Selective Copper(II) Sorption Behavior of Surface-Imprinted Core–Shell-Type Polymethacrylate Microspheres*. Industrial & Engineering Chemistry Research, 2009. 48(12): p. 5679-5685.
- [135.] Li, T., S. Chen, H. Li, Q. Li, and L. Wu, *Preparation of an Ion-Imprinted Fiber for the Selective Removal of Cu²⁺*. Langmuir, 2011. 27(11): p. 6753-6758.
- [136.] Zhou, Z., X. Liu, M. Zhang, J. Jiao, H. Zhang, J. Du, B. Zhang, and Z. Ren, *Preparation of highly efficient ion-imprinted polymers with Fe₃O₄ nanoparticles as carrier for removal of Cr(VI) from aqueous solution*. Science of The Total Environment, 2020. 699: p. 134334.
- [137.] Jiang, N., X. Chang, H. Zheng, Q. He, and Z. Hu, *Selective solid-phase extraction of nickel(II) using a surface-imprinted silica gel sorbent*. Analytica Chimica Acta, 2006. 577(2): p. 225-231.
- [138.] Chang, X., N. Jiang, H. Zheng, Q. He, Z. Hu, Y. Zhai, and Y. Cui, *Solid-phase extraction of iron(III) with an ion-imprinted functionalized silica gel sorbent prepared by a surface imprinting technique*. Talanta, 2007. 71(1): p. 38-43.
- [139.] Chang, P.Y., R. Doppalapudi, J. Bakke, A. Puey, and S. Lin, *Evaluation of the impact of shielding materials in radiation protection in transgenic animals*. Radiation and Environmental Biophysics, 2007. 46(2): p. 113-118.
- [140.] Milja, T.E., K.P. Prathish, and T. Prasada Rao, *Synthesis of surface imprinted nanospheres for selective removal of uranium from simulants of Sambhar salt lake and ground water*. Journal of Hazardous Materials, 2011. 188(1): p. 384-390.
- [141.] Li, F., H. Jiang, and S. Zhang, *An ion-imprinted silica-supported organic–inorganic hybrid sorbent prepared by a surface imprinting technique combined with a polysaccharide incorporated sol–gel process for selective separation of cadmium(II) from aqueous solution*. Talanta, 2007. 71(4): p. 1487-1493.
- [142.] Pan, J., W. Guan, Z. Zhang, X. Wang, C. Li, and Y. Yan, *Selective Adsorption of Co(II) Ions by Whisker Surface Ion-Imprinted Polymer: Equilibrium and Kinetics Modeling*. Chinese Journal of Chemistry, 2010. 28(12): p. 2483-2488.
- [143.] Zhang, Z., X. Xu, and Y. Yan, *Kinetic and thermodynamic analysis of selective adsorption of Cs(I) by a novel surface whisker-supported ion-imprinted polymer*. Desalination, 2010. 263(1): p. 97-106.

- [144.] Felix, C.S.A., D.G. Silva, H.M.C. Andrade, V.B. Riatto, M.M. Victor, and S.L.C. Ferreira, *An on-line system using ion-imprinted polymer for preconcentration and determination of bismuth in seawater employing atomic fluorescence spectrometry*. Talanta, 2018. 184: p. 87-92.
- [145.] Zhu, C., T. Hu, L. Tang, G. Zeng, Y. Deng, Y. Lu, S. Fang, J. Wang, Y. Liu, and J. Yu, *Highly efficient extraction of lead ions from smelting wastewater, slag and contaminated soil by two-dimensional montmorillonite-based surface ion imprinted polymer absorbent*. Chemosphere, 2018. 209: p. 246-257.
- [146.] Behbahani, M., M. Salarian, A. Bagheri, H. Tabani, F. Omid, and A. Fakhari, *Synthesis, characterization and analytical application of Zn(II)-imprinted polymer as an efficient solid-phase extraction technique for trace determination of zinc ions in food samples*. Journal of Food Composition and Analysis, 2014. 34(1): p. 81-89.
- [147.] Arbab-Zavar, M.H., M. Chamsaz, G. Zohuri, and A. Darroudi, *Synthesis and characterization of nano-pore thallium (III) ion-imprinted polymer as a new sorbent for separation and preconcentration of thallium*. Journal of Hazardous Materials, 2011. 185(1): p. 38-43.
- [148.] Liu, Y., Y. Zai, X. Chang, Y. Guo, S. Meng, and F. Feng, *Highly selective determination of methylmercury with methylmercury-imprinted polymers*. Analytica Chimica Acta, 2006. 575(2): p. 159-165.
- [149.] Jo, S.-H., S.-Y. Lee, K.-M. Park, S.C. Yi, D. Kim, and S. Mun, *Continuous separation of copper ions from a mixture of heavy metal ions using a three-zone carousel process packed with metal ion-imprinted polymer*. Journal of Chromatography A, 2010. 1217(45): p. 7100-7108.
- [150.] Alizadeh, T., M.R. Ganjali, P. Nourozi, M. Zare, and M. Hoseini, *A carbon paste electrode impregnated with Cd²⁺ imprinted polymer as a new and high selective electrochemical sensor for determination of ultra-trace Cd²⁺ in water samples*. Journal of Electroanalytical Chemistry, 2011. 657(1): p. 98-106.
- [151.] Prasad, K., R. Kala, T. Prasada Rao, and G.R.K. Naidu, *Ion imprinted polymer based ion-selective electrode for the trace determination of dysprosium(III) ions*. Analytica Chimica Acta, 2006. 566(1): p. 69-74.
- [152.] Qin, X., A. Lu, and L. Zhang, *Gelation behavior of cellulose in NaOH/urea aqueous system via cross-linking*. Cellulose, 2013. 20.
- [153.] Sikorski, P., F. Mo, G. Skjåk-Bræk, and B.T. Stokke, *Evidence for Egg-Box-Compatible Interactions in Calcium-Alginate Gels from Fiber X-ray Diffraction*. Biomacromolecules, 2007. 8(7): p. 2098-2103.

- [154.] Braccini, I. and S. Pérez, *Molecular Basis of Ca²⁺-Induced Gelation in Alginates and Pectins: The Egg-Box Model Revisited*. Biomacromolecules, 2001. 2(4): p. 1089-1096.
- [155.] Zhu, H., R. Srivastava, and M.J. McShane, *Spontaneous Loading of Positively Charged Macromolecules into Alginate-Templated Polyelectrolyte Multilayer Microcapsules*. Biomacromolecules, 2005. 6(4): p. 2221-2228.
- [156.] Alizadeh, T., A.N. Shamkhali, Y. Hanifehpour, and S. Joo, *A Ca²⁺ selective membrane electrode based on calcium-imprinted polymeric nanoparticles*. New J. Chem., 2016. 40.
- [157.] Rosatzin, T., L. Andersson, W. Simon, and K. Mosbach, *Preparation of Ca²⁺ selective sorbents by molecular imprinting using polymerisable ionophores*. Journal of The Chemical Society-perkin Transactions 2 - J CHEM SOC PERKIN TRANS 2, 1991. 28.
- [158.] Yoshida, M., K. Uezu, M. Goto, and S. Furusaki, *Required Properties for Functional Monomers To Produce a Metal Template Effect by a Surface Molecular Imprinting Technique*. Macromolecules, 1999. 32(4): p. 1237-1243.
- [159.] Kala, R., V.M. Biju, and T.P. Rao, *Synthesis, characterization, and analytical applications of erbium(III) ion imprinted polymer particles prepared via γ -irradiation with different functional and crosslinking monomers*. Analytica Chimica Acta, 2005. 549(1): p. 51-58.
- [160.] Kariş, D., D. Tarhan, K. Boyacıoğlu, C. Köksal, and A.M. Ercan, *The comparison of zinc, copper and iron levels in serum, aorta and left internal mammarian artery tissues in coronary by-pass graft surgery patients*. Journal of Trace Elements in Medicine and Biology, 2019. 51: p. 86-90.
- [161.] Bursalıoğlu, E.O., F.A. Alkan, Ü.B. Barutçu, M. Demir, Y. Karabul, B. Balkan, E. Öz, and O. İçelli, *Prediction of electron density and trace element concentrations in human blood serum following radioiodine therapy in differentiated thyroid cancer patients*. Measurement, 2017. 100: p. 19-25.
- [162.] Mohammed Nawi, A., S.-F. Chin, and R. Jamal, *Simultaneous analysis of 25 trace elements in micro volume of human serum by inductively coupled plasma mass spectrometry (ICP-MS)*. Practical Laboratory Medicine, 2020. 18: p. e00142.
- [163.] Lorenzo, R.A., A.M. Carro, C. Alvarez-Lorenzo, and A. Concheiro, *To remove or not to remove? The challenge of extracting the template to make the cavities available in Molecularly Imprinted Polymers (MIPs)*. Int J Mol Sci, 2011. 12(7): p. 4327-47.

- [164.] Chang, C., B. Duan, and L. Zhang, *Fabrication and characterization of novel macroporous cellulose–alginate hydrogels*. Polymer, 2009. 50(23): p. 5467-5473.
- [165.] Sefrou, Z. and N.-E. Belkhouche, *Cloud point extraction of La(III) by C13E10 non-ionic surfactant: Statistical refinement of experimental optimization by L9 Taguchi's design*. Chemical Engineering Research and Design, 2020. 153: p. 819-828.
- [166.] Zhou, J. and L. Zhang, *Structure and properties of blend membranes prepared from cellulose and alginate in NaOH/urea aqueous solution*. Journal of Polymer Science Part B: Polymer Physics, 2001. 39(4): p. 451-458.
- [167.] Huang, R.Y.M., R. Pal, and G.Y. Moon, *Characteristics of sodium alginate membranes for the pervaporation dehydration of ethanol–water and isopropanol–water mixtures*. Journal of Membrane Science, 1999. 160(1): p. 101-113.
- [168.] Daemi, H. and M. Barikani, *Synthesis and characterization of calcium alginate nanoparticles, sodium homopolymannuronate salt and its calcium nanoparticles*. Scientia Iranica, 2012. 19(6): p. 2023-2028.
- [169.] Fan, L., Y. Du, R. Huang, Q. Wang, X. Wang, and L. Zhang, *Preparation and characterization of alginate/gelatin blend fibers*. Journal of Applied Polymer Science, 2005. 96(5): p. 1625-1629.
- [170.] Ben Dekhil, A., Y. Hannachi, A. Ghorbel, and B. Taoufik, *Comparative Study of the Removal of Cadmium from Aqueous Solution by using Low-cost Adsorbents*. Journal of Environmental Science and Technology, 2011. 4: p. 520-533.
- [171.] Kuchen, W. and J. Schram. *Metal-Ion-Selective Exchange Resins by Matrix Imprint with Methacrylates*. 1988.
- [172.] Nacano, L.R., M.G. Segatelli, and C.R.T. Tarley, *Selective sorbent enrichment of nickel ions from aqueous solutions using a hierarchically hybrid organic-inorganic polymer based on double imprinting concept*. Journal of the Brazilian Chemical Society, 2010. 21: p. 419-430.
- [173.] Svenson, J. and I.A. Nicholls, *On the thermal and chemical stability of molecularly imprinted polymers*. Analytica Chimica Acta, 2001. 435(1): p. 19-24.
- [174.] Yusof, N.F., F. Mehamod, M. Abdul Kadir, and F. Mohd Suah, *Characteristics of adsorption isotherm and kinetic study for newly prepared Co 2+ -imprinted polymer linkage with dipicolinic acid*. IOP Conference Series: Materials Science and Engineering, 2018. 440: p. 012005.

- [175.] Mittal, A., L. Kurup, and J. Mittal, *Freundlich and Langmuir Adsorption Isotherms and Kinetics for the Removal of Tartrazine from Aqueous Solutions Using Hen Feathers*. Journal of hazardous materials, 2007. 146: p. 243-8.
- [176.] Rais, S., A. Islam, I. Ahmad, S. Kumar, A. Chauhan, and H. Javed, *Preparation of a new magnetic ion-imprinted polymer and optimization using Box-Behnken design for selective removal and determination of Cu(II) in food and wastewater samples*. Food Chemistry, 2021. 334: p. 127563.
- [177.] Neolaka, Y.A.B., G. Supriyanto, H. Darmokoesoemo, and H.S. Kusuma, *Characterization, isotherm, and thermodynamic data for selective adsorption of Cr(VI) from aqueous solution by Indonesia (Ende-Flores) natural zeolite Cr(VI)-imprinted-poly(4-VP-co-EGDMA)-ANZ (IIP-ANZ)*. Data in Brief, 2018. 17: p. 1020-1029.
- [178.] Sathish, T., N.V. Vinithkumar, G. Dharani, and R. Kirubakaran, *Efficacy of mangrove leaf powder for bioremediation of chromium (VI) from aqueous solutions: kinetic and thermodynamic evaluation*. Applied Water Science, 2015. 5(2): p. 153-160.
- [179.] Malkoc, E. and Y. Nuhoglu, *Potential of Tea Factory Waste for Chromium (VI) Removal From Aqueous Solutions: Thermodynamic and Kinetic Studies*. Separation and Purification Technology - SEP PURIF TECHNOL, 2007. 54: p. 291-298.
- [180.] Li, Y.-H., S. Wang, Z. Luan, J. Ding, C. Xu, and D. Wu, *Adsorption of cadmium(II) from aqueous solution by surface oxidized carbon nanotubes*. Carbon, 2003. 41(5): p. 1057-1062.
- [181.] Robati, D., *Pseudo-second-order kinetic equations for modeling adsorption systems for removal of lead ions using multi-walled carbon nanotube*. Journal of Nanostructure in Chemistry, 2013. 3(1): p. 55.
- [182.] George, A.M. and A.R. Tembhurkar, *Taguchi experimental design for adsorptive removal of fluoride from water using novel Ficus Glomerata Bark-developed biosorbent*. International Journal of Environmental Science and Technology, 2020.
- [183.] Guang, W., M. Baraldo, and M. Furlanut, *Calculating percentage prediction error: A user's note*. Pharmacological Research, 1995. 32(4): p. 241-248.

BIODATA OF STUDENT

Rafah Al-maibd was born in Basra (Iraq) on the 16th May 1972. She started her primary education at Al-Fayhaa primary school in Basra. Then, she completed her secondary education at Al- Basra school secondary. She obtained her Baccalaureate level in 1990. Later, she completed her Bachelor of Science (Hons) degree in Chemistry in 1995 from Basra University.

In 1997, she worked as chemist in the quality control department at the Advanced pharmaceutical industry in Amman, Jordan. In 2012, Rafah was offered a government job in Al-Fayhaa general hospital in Basra as Chemist in AL-FAIHA SPECIALIZED DIABETES ENDOCRINE AND METABOLISM CENTER in which she was in charge of the daily analysis of daily patient's samples using automated clinical biochemistry analyzers. She was responsible of trouble shooting and maintaining the analyzers at the laboratory.

Rafah has enrolled as a full time Master student in September 2018 at Universiti Putra Malaysia under the supervision of Dr. Sazlinda Kamurzamman.

LIST OF PUBLICATIONS

Peer review papers:

Rafah AL-Maibd, Wissam Al-Ashaq, Norhazlin Zainuddin, Nor Azowa Ibrahim, Intan Nureslyna Samsudin, Sazlinda Kamaruzaman, *Synthesis and optimization selective ion-imprinted polymer for the elimination of Ca II ions using Taguchi design*", Journal of Polymer Research, Vol 28, Issue 3, Pages 1-16, Springer, (March 202)

Conferences

Rafah AL-Maibd and Sazlinda Kamurzamman, *Optimization of the synthesis and application of Novel Ca(II) Ion-Imprinted Polymer (IIP) using Taguchi Method*, The International Virtual Conference on Fundemnatl and Applied Scinces (IFSC 2020), Kasetsart University, Thailand.

Rafah AL-Maibd, 11th International Fundamental Science Conference (iFSC 2019), Palm Garden Hotel, Putrajaya, Malaysia (October 2019)



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