

UNIVERSITI PUTRA MALAYSIA

BIODEGRADATION OF SODIUM DODECYL SULPHATE BY A LOCALLY ISOLATED BACTERIUM, *Klebsiella oxytoca* DR.Y14

> WAN SURINI BINTI WAN HUSIN FBSB 2006 42



BIODEGRADATION OF SODIUM DODECYL SULPHATE BY A LOCALLY ISOLATED BACTERIUM, *Klebsiella oxytoca* DR.Y14

By

WAN SURINI BINTI WAN HUSIN

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

June 2006



Dedicated to my father Wan Husin Sapien,, my mother Khasiah, Isa, sisters and brother



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirements for the degree of Master of Science

BIODEGRADATION OF SODIUM DODECYL SULPHATE BY A LOCALLY ISOLATED BACTERIUM, *Klebsiella oxytoca* DR.Y14

By

WAN SURINI BINTI WAN HUSIN

June 2006

Chairman: Professor Mohd Arif Syed, PhD

Faculty : Biotechnology and Biomolecular Sciences

This study was conducted on sodium dodecyl sulphate (SDS) biodegradation. Bacteria capable of utilizing SDS as a sole source of carbon were isolated from water samples exposed with surfactants. Enrichment culture yielded several isolates capable of metabolizing SDS. Of these, Isolate S11 was selected for further studies based on its biodegradative capability as determined using methylene blue active substance (MBAS) assay. The isolate was identified as Klebsiella sp. using BiologTM identification system and was confirmed using 16S rRNA molecular phylogenetic analysis. Isolate S11 exhibited optimum growth at 37 °C in media containing high SDS concentrations (up to 1.0 g/L SDS), and is able to degrade 99% of 1.0 g/L SDS in 3 days. It requires minimal nitrogen source as low as 0.5 mg/L ammonium sulphate for optimum growth consistent with the *Klebsiella* genus ability to fix atmospheric nitrogen. Partially purified alkylsulphatase from S11 showed optimum enzyme activity at 80°C and at pH 8 using Tris-HCl buffer when tested using MBAS assay. The apparent K_m and apparent V_{max} of the SDS-degrading enzyme were determined to be 0.232 mM and 1.391 µmol per minute per mg protein



respectively. The enzyme was found to be stable at room temperature for 50 days in Tris-HCl buffer at pH 7.5.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

PENGURAIAN SODIUM DODECIL SULFAT DARIPADA BAKTERIA TEMPATAN, *Klebsiella oxytoca* DR.Y14

Oleh

WAN SURINI BINTI WAN HUSIN

June 2006

Pengerusi: Profesor Mohd Arif Syed, PhD

Fakulti : Bioteknologi dan Sains Biomolekul

Kajian mengenai bio-penguraian natrium dodesil sulfat (SDS) telah dijalankan. Bakteria yang berupaya menggunakan SDS sebagai sumber tunggal karbon telah dipencilkan daripada sampel air yang terdedah dengan surfaktan. Beberapa isolat pengurai SDS telah diperolehi daripada teknik kultur pengkayaan. Isolat S11 telah dipilih untuk kajian seterusnya berdasarkan kebolehannya mengurai SDS setelah diuji melalui asai MBAS (Methylene Blue Active Substance). Isolat ini telah dikenalpasti sebagai *Klebsiella* sp. menggunakan sistem pengenalpastian Biolog[™] dan telah dipastikan dengan lebih lanjut lagi menggunakan analisis filogenetik molekul 16S rRNA. Isolat S11 telah mempamerkan pertumbuhan yang optimum pada suhu 37 °C dalam media yang mengandungi kepekatan SDS yang tinggi sehingga 1.0 g/L dan berupaya mengurai 99% daripada amaun ini dalam masa 3 hari. Untuk pertumbuhan optima, ia memerlukan sumber nitrogen terbaik seiring dengan kebolehan genus *Klebsiella* untuk mengikat nitrogen daripada udara. Penulenan separa alkilsulfatase daripada S11 menunjukkan aktiviti optima pada 80 °C



menggunakan penimbal Tris-HCl pada pH 8.0 apabila diuji menggunakan asai MBAS. Nilai K_m dan V_{max} yang diperolehi daripada enzim pengurai SDS ini ialah masing-masing 0.232 mM dan 1.391 µmol per minit per mg protein. Enzim ini didapati stabil pada suhu bilik selama lebih 50 hari di dalam penimbal Tris-HCl pada pH 7.5.



ACKNOWLEDGEMENTS

Thankfulness and glories to Allah the Almighty for His blessings.

First and foremost, I would like to extend my greatest and deepest gratitude to my supervisor, Prof. Dr Mohd Arif Syed for his invaluable guidance throughout the completion of this project. My warmest gratitude also goes to. Dr. Mohd Yunus Abdul Shukor for his supervision, advice, constructive suggestions and review of my work during the period of this study.

I would also like to express my sincere appreciation to Ms. Taznim Begam Mohd Mohidin, Mr. Ariff Khalid, and Mr. Shahrul Izham Rojali for their assistance, supervision, knowledge and encouragement in making this project a successful one for me.

I would also like to take this opportunity to thank all members, postgraduates as well as the undergraduates of Enzymology and Bioremediation Lab (115 and 204) especially Fazuriana, Sim Han Koh, Natarajan, Farrah Aini and Aqlima for their kind assistance and for sharing their experiences and knowledge, directly or indirectly. Not forgetting my fellow coursemates, Nor Azlina, Nor Adeela, Tang Kah Fai, Siti Salwa and Mokrish for their encouragement and companionship through thick and thin.

Last but not least, I would like to recognize my parents Wan Husin Sapien and Khasiah Isa, sisters, Wan Nor Adianti and Wan Sofia and brother, Wan Mohd Amir



for their unconditional sacrifices, love and undying support. Thank you for believing in me and for making me the person I am today.



I certify that an Examination Committee has met on 21st June 2006 to conduct the final examination of Wan Surini binti Wan Husin on her Master of Science thesis entitled "Biodegradation of Sodium Dodecyl Sulphate by a locally isolated bacterium, *Klebsiella oxytoca* Dr.Y14" in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The committee recommends that the candidate be awarded the relevant degree. Members of the Examination Committee are as follows:

Chairman, PhD

Professor Dr. Mohd Arif Syed Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Chairman)

Examiner 1, PhD

Associate Professor Dr. Juzu Hayati Arshad Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Internal examiner)

Examiner 2, PhD

Associate Professor Dr. Johari Ramli Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Internal examiner)

Independent Examiner, PhD

Professor, Name of faculty/institute Universiti Putra Malaysia (External Examiner)

HASANAH MOHD GHAZALI

Professor/Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date :



This thesis submitted to the Senate of Universiti Putra Malaysia has been accepted as fulfilment of the requirements for the degree of Master of Science. The members of the Supervisory Committee are as follows:

Mohd Arif Syed, PhD

Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Chairman)

Mohd Yunus Abd Shukor, PhD

Lecturer Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Member)

AINI IDERIS, PhD

Professor/Dean School of Graduate Studies Universiti Putra Malaysia

Date :



DECLARATION

I hereby declare that the thesis is based on my original work except for quotation and citations, which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at UPM or other institutions.

WAN SURINI BINTI WAN HUSIN

Date:



TABLE OF CONTENTS

Page

DEDICATION	ii
ABSTRACT	iii
ABSTRAK	V
ACKNOWLEDGEMENTS	vii
APPROVAL	ix
DECLARATION	xi
LIST OF TABLES	XV
LIST OF FIGURES	xvi
LIST OF ABBREVIATIONS	xviii

CHAPTER

1	INTE	RODUCTION	1
2	LITE	ERATURE REVIEW	3
	2.1	Surfactants	3
	2.2	Classification of Surfactants	4
	2.3	Surfactants Pollutions in Malaysia	7
	2.4	Problems Caused by Anionic Surfactants	9
		2.4.1 Surfactants in Wastewater	10
		2.4.2 Surfactants in Sewage Sludge	11
		2.4.3 Surfactants in Land	12
	2.5	Sodium Dodecyl Sulphate (SDS) as a Surfactant	13
		2.5.1 Characterization of SDS	13
		2.5.2 Human Exposure of SDS	14
		2.5.3 Toxicity of SDS	16
		2.5.4 Analysis of Anionic Surfactant in Environmental Samples	19
	2.6	Bioremediation	20
		2.6.1 Introduction	20
		2.6.2 Principles of Bioremediation	20
		2.6.3 Advantages of Bioremediation	21
	2.7	Biodegradation	22
		2.7.1 Introduction	22
		2.7.2 Anionic Surfactant-degrading Bacteria	23
		2.7.3 Factors Affecting Bacterial Growth and Activity	26
	2.8	Mechanisms and Pathways of SDS Biodegradation	27
	2.9	Alkylsulphatase	30
3	МАТ	TERIALS AND METHODS	33
	3.1	Chemicals and Equipments	34
	3.2	Isolation of SDS-degrading Bacteria	34
		3.2.1 Bacterial Sampling	34
		3.2.2 Isolation of Bacteria	35
		3.2.3 Maintenance and Growth of Bacterial Isolates	36



	3.2.4 Measurement of Bacterial Growth	36
3.3	Screening of SDS-degrading Bacteria	38
	3.3.1 Preliminary Screening	38
	3.3.2 Secondary Screening	39
	3.3.3 Methylene Blue Active Substance (MBAS) Assay	40
3.4	Growth Optimization of Isolate S11	41
	3.4.1 Optimization of pH	41
	3.4.2 Optimization of Temperature	42
	3.4.3 Effects of SDS Concentrations	42
	3.4.4 Optimization of Nitrogen Sources	43
	3.4.5 Effects of Nitrogen Source Concentrations	44
3.5	Identification of SDS-degrading Bacteria	44
	3.5.1 Biochemical Test	44
	3.5.2 16S rRNA Analysis	46
	3.5.2.1 Genomic Extraction	47
	3.5.2.2 Polymerase Chain Reaction (PCR)	47
	3.5.2.3 Purification of Amplified PCR Products	48
	3.5.2.4 Quantification of DNA Concentration	48
	3.5.2.5 Sequence Analysis	49
	3.5.2.6 Phylogenetic Analysis	50
3.6	Partial Purification of Alkylsulphatase	51
	3.6.1 Scale-up Culture of Isolate S11	51
	3.6.2 Preparation of Enzyme Extracts	52
	3.6.3 Determination of Protein Concentration	52
	3.6.4 Partial Purification Using Macro-Prep High-Q TM	53
	Anion Exchanger	
	3.6.5 Partial Purification Using Agilent Zorbax TM (GF-250)	54
	Gel Filtration	
	3.6.6 SDS Polyacrylamide Gel Electrophoresis	55
3.7	Enzymatic Studies on Degradation of SDS	57
	3.7.1 Alkylsulphatase Assay	57
	3.7.2 Determination of Alkylsufatase K_m and V_{max}	58
	3.7.3 Effect of Different Temperatures on Alkylsulphatase	59
	Activity	
	3.7.4 Effect of Different pH on Alkylsulphatase Activity	59
	3.7.5 Alkylsulphatase Stability Study	60
	3.7.5.1 Determination of Alkylsulphatase pH Stability	60
	3.7.5.2 Determination of Alkylsulphatase Temperature	61
	Stability	
RES	ULTS AND DISCUSSION	62
4.1	Isolation of SDS-degrading Bacteria	62
4.2	Screening of SDS-degrading Bacteria	64
	4.2.1 Preliminary Screening of SDS-degrading Bacteria	64
	4.2.2 Secondary Screening of SDS-degrading Bacteria	68
4.3	Growth Optimization of Isolate S11	70
	4.3.1 Optimization of pH	70
	4.3.2 Optimization of Temperature	72
	4.3.3 Effects of SDS Concentrations	74
	4.3.4 Optimization of Nitrogen Sources	76



4

	4.3.5	Effects of Ammonium Sulphate Concentrations	79
4.4	SDS I	Degradation Study	81
4.5	Identi	fication of SDS-degrading Bacteria	86
	4.5.1	Morphological Observation	86
	4.5.2	Gram Identification	86
	4.5.3	Biochemical Test	89
	4.5.4	16S rRNA Analysis	92
		4.5.4.1 Extraction of Genomic DNA	92
		4.5.4.2 Polymerase Chain Reaction (PCR)	92
		4.5.4.3 16S rRNA Gene Sequencing	94
		4.5.4.4 Phylogenetic Analysis	96
4.6	Partia	l Purification of Alkylsulphatase	100
	4.6.1	SDS Polyacrylamide Gel Electrophoresis	105
4.7	Enzyn	natic Study on SDS Degradation	107
	4.7.1	Alkylsulphatase Assay	107
	4.7.2	Effect of Different Temperatures on Alkylsulphatase Activity	108
	4.7.3	Effect of Different pH on Alkylsulphatase Activity	110
		Determination of Alkylsulphatase $K_{m(app)}$ and $V_{max(app)}$	112
		Determination of Alkylsulphatase pH Stability	115
	4.7.6	Determination of Alkylsulphatase Temperature Stability	117
CON		ON	101

5	CONCLUSION	121
RE	FERENCES	123
AP	ENDICES	131
BIC	ODATA OF THE AUTHOR	149



LIST OF TABLES

Table		Page
1	Acute toxicity of SDS in selected animals	17
2	Toxicity of SDS in various organisms	18
3	Examples of anionic surfactant-degrading bacteria	25
4	List of successfully isolated SDS-degrading bacteria	63
5	Microscopic and macroscopic observations of Isolate S11	87
6	Growth of Isolate S11 on various carbon sources in Biolog's ID microplates	90
7	Alkylsulphatase purification table	104



LIST OF FIGURES

Figure

Page

1	Illustrative examples of the structural classification of surfactants	6
2	Distribution of water pollution sources in Malaysia, 2003	8
3	Pathways of incorporated of SDS into bacterial cellular products	29
4	Enzymatic hydrolysis of sulfate esters	30
5	The overview of methods used in this study	33
6	The absorbance readings of sixteen bacterial isolates from the MTT assay during the preliminary screening	66
7	SDS degradation of six isolates in SDS basal salt media with 1.0 g/L SDS as the initial concentration	69
8	The effect of pH on the growth of Isolate S11 using three overlapping 50 mM buffers	71
9	The effect of temperatures for the growth of Isolate S11	73
10	The effects of SDS concentrations on the growth of Isolate S11	75
11	The effect of different nitrogen sources (0.1%) on the growth of Isolate S11	78
12	The effect of ammonium sulphate concentration on the growth of Isolate S11	80
13	SDS degradation and growth profile of Isolate S11 before growth optimizations	82
14	SDS degradation and growth profile of Isolate S11 after growth optimizations	84
15	SDS degradation culture of Isolate S11	85
16	Photomicrograph of Isolate S11 which is Gram-negative (pink-red) rod, by observation under light microscope (Olympus BX40.F4, Japan) with 1000 X magnification	88
17	Agarose gel electrophoresis	93
18	The region of homology between the forward and reverse	95



complement of Isolate S11

19	Phylogenetic tree of newly isolated SDS-degrading bacteria	97
20	The 16S rRNA sequence of Isolate S11 and its accession number as deposited in GenBank	99
21	Alkylsulphatase elution profile using Macro-Prep High- Q^{TM} anion-exchanger	100
22	Alkylsulphatase elution profile using Zorbax TM (GF250) gel filtration column	103
23	SDS-10% polyacrylamide gel analysis and Coomasie blue staining of partially purified alkylsulphatase	106
24	Effect of temperatures on alkylsulphatase activity with 100 mM SDS as the substrate	109
25	Effect of different pH on alkylsulphatase activity with 100 mM SDS as the substrate	111
26	Michaelis-Menten curve for alkylsulphatase with SDS as the substrate	113
27	Lineweaver-Burke graph for alkylsulphatase activity with SDS as the substrate	114
28	Effects of pre-incubation at different pHs and buffers on alkylsulphatase stability	116
29	Effects of different pre-incubation temperatures on alkylsulphatase	118
30	Effects of prolonged pre-incubation temperatures on alkylsulphatase	119



LIST OF ABBREVIATIONS

°C	degree Celsius
%	percent
bp	base pair
CFU	colony forming unit
dH ₂ O	distilled water
DNA	deoxyribonucleic acid
EDTA	ethylene diamine tetraacetic acid
GPS	global positioning system
HPLC	High Performance Liquid Chromatography
IUPAC	International Union of Pure and Applied Chemistry
kb	kilobase
kDa	kilodalton
K_m	Michaelis-Menten Constant
K _{m(app)}	Apparent Michaelis-Menten Constant
μl	microlitre
μΜ	micromolar
М	molar
mA	milliampere
mAu	milli absorbance unit
MBAS	methylene blue active substance
mg	milligram
mM	millimolar
nm	Nanometer



OD	optical density
PBS	phosphate-buffered saline
pH	-log concentration of H^+ ion (<i>Puissance hydrogene</i>)
PCR	polymerase chain reaction
PMSF	phenylmethylsulfonylfluoride
RNA	ribonucleic acid
rpm	revolutions per minute
SDS	sodium dodecyl sulphate
TCA cycle	Tricarboxylic acid cycle
TCA cycle PAGE	Tricarboxylic acid cycle polyacrylamide gel electrophoresis
·	
PAGE	polyacrylamide gel electrophoresis
PAGE UV	polyacrylamide gel electrophoresis ultraviolet
PAGE UV w/v	polyacrylamide gel electrophoresis ultraviolet weight/ volume
PAGE UV w/v v/v	polyacrylamide gel electrophoresis ultraviolet weight/ volume volume/ volume



CHAPTER 1

INTRODUCTION

Study on surfactant biodegradation has assumed importance as a consequence of the chemical revolution which occurred in the detergent industry during the decade centering on 1950 (Swisher, 1987). Historically, potential surfactant contamination of the environment followed the shift from the use of soap-based detergent to synthetic surfactants (Scott and Jones, 2000).

Based on their favorable physicochemical properties, synthetic surfactants are extensively used in many fields of technology and research for example in pharmacy, cosmetics, textile industry, agriculture and biotechnology (Bizukojc and Bizukojc, 2005). Worldwide production of surfactant increased from 3500 tons in 1950 to approximately 4.3 million tons in 1990 (Jerabkova *et al.*, 1999).

Due to extensive application, an appreciable amount of anionic surfactant is released in aquatic and terrestrial environment causing serious water pollution (Cserhati *et al.*, 2002). Even though surfactants are essentially nontoxic to humans, there is general agreement that their presence in drinking water is undesirable, (Swisher, 1987; Jerabkova *et al.*, 1999). Surfactants caused foaming in aerated bioreactor and decreased the settling ability of the sludge. Besides that, surfactant was reported to be toxic to microorganisms (Jorge and Moreira, 2005).



Taking into account the potential environmental impact of surfactant, many studies concerning biodegradability and toxicity of surfactants have been performed (Petterson *et al.*, 2000; Bizukojc *et al.*, 2005). Due to current laws on the banning of importation of microbes as well as the highly cautious use of genetically-modified organism (GMO) to be used for the bioremediation of xenobiotics (Walter *et al.*, 2003) in Malaysia, it is important to isolate local bacteria for bioremediation of anionic surfactants in Malaysia. Research into bioremediation or the use of microbes or their enzymes to biodegrade the contaminated environment to their original state are currently still in the early ages (Thassitou and Arvanitoyannis, 2001). To date, no such publications exist for isolation of sodium dodecyl sulphate (SDS)-degrading microbes from Malaysia and this work is thus of high importance for fundamental knowledge as well as application. As SDS is the most common surfactant found in detergent, shampoos and cleaning formulations, its biodegradation is vital to be studied compared to other anionic surfactants.

In this study, four major objectives will be accomplished. The objectives of this study are:

- 1. to isolate and screen local SDS-degrading bacteria.
- to determine the optimum growth characteristics of the isolated SDSdegrading bacterium.
- 3. to identify the SDS-degrading bacterium to species level.
- 4. to partially purify and characterize the SDS-degrading enzyme.



CHAPTER 2

LITERATURE REVIEW

2.1 Surfactants

Anionic surfactants, as major components of synthetic detergents used for both domestic and industrial applications, contribute significantly to the pollution profile of sewage and wastewaters. In 1995, world production of 9.3 million tons anionic surfactants reflects the high demand for this type of surfactant (Douib *et al.*, 2003). Anionic surfactant is the major surfactant used as it represents 59% of surfactants used worldwide (Dhouib *et al.*, 2003). In terms of environmental issues, the focus of concern has largely been on the effects of the surfactant in detergent formulation. The most widely used surfactants in detergent formulations are those containing anionic group such as alkyl sulphate (Jerabkova *et al.*, 1999).

Surfactant is a large group of structurally-diverse molecules and possesses surfaceactive properties. Surfactant molecules are amphiphiles, contain both strongly hydrophobic and hydrophilic group (Cserhati *et al.*, 2002). Thus, they tend to concentrate at the surfaces and interfaces of the aqueous systems including air, oily material and particles (White and Russell, 1994).

Detergent is distinct from surfactant which refers to a commercial formulation or product that is designed with particular cleansing properties (White and Russell, 1994). These formulations contain one-third surfactant, larger amounts of a



"builder" which acts as chelating agents and smaller amounts of perfumes, colouring agents, whiteners, enzymes and other components (White and Russell, 1994).

2.2 Classification of Surfactants

Surfactants are classified broadly based on the chemical nature of the polar group as being anionic, non-ionic, cationic or zwitterionic (Figure 1) (White and Russell, 1994). The structural characteristics and surface activity of surfactants are based on simple principles. The molecule of a surfactant is formally constructed by bonding one or more lipophilic groups to one more hydrophilic group (Hummel, 2000). The surface activity of a surfactant is determined by the nature of and relationship between the lipophilic and hydrophilic groups and by their spatial arrangement. The hydrophilic groups can consist of electrically charged (ionic) and also uncharged polar structures (Hummel, 2000).

Anionic surfactants produce negatively charged ions in aqueous solution, originated from sulphate or sulphonate groups. Cationic surfactants produce positively charged ions in solution, frequently quaternary ammonium ions, which constitute less than 10% of the ionics and are used for fabric softening, disinfection and other specialized applications. Zwiterionic surfactants contain both cationic and anionic moieties in the same molecule while non-ionic surfactants contain hydrophobic and hydrophilic that is organic and do not ionize (White and Russell, 1994).

Bioavailability of surfactants changes under anaerobic conditions and may affect the outcome of toxicity and inhibition studies, and to some extent biodegradation or



removal rates. The specific chemical structure of some surfactants contributes to a rapid precipitation with water hardness ions (Ca, Mg) into insoluble forms, as well as adsorption to the surrounding solid matrix. This highlights the need to use the real environmental form of a surfactant in inhibition and biodegradation tests, in order to obtain a realistic test result (George and White, 1999).

