



UNIVERSITI PUTRA MALAYSIA

***EARLY DETECTION OF *Ganoderma boninense* IN OIL PALM
SEEDLINGS USING HYPERSPECTRAL IMAGES AND MACHINE
LEARNING CLASSIFICATION TECHNIQUES***

AIMAN NABILAH BINTI NOOR AZMI

FK 2021 49



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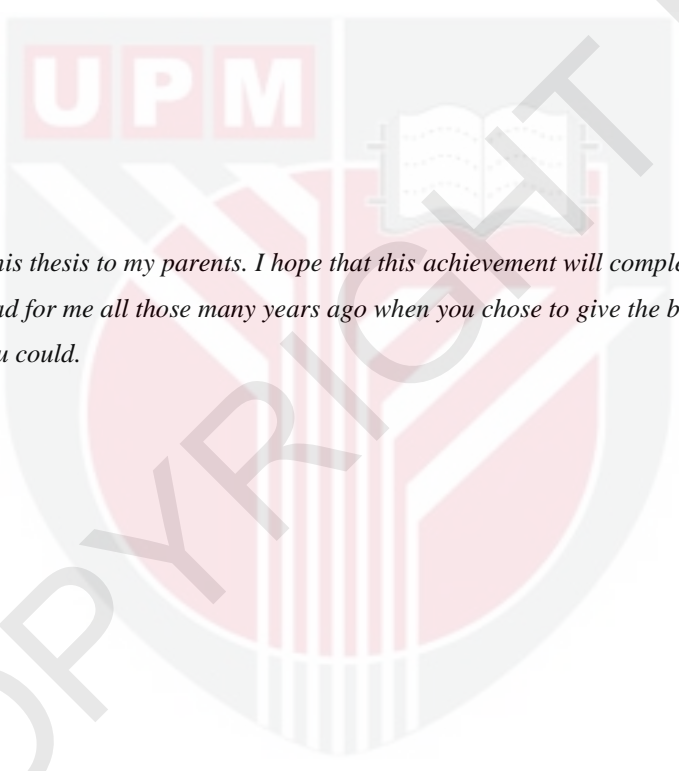
**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Master of Science**

January 2021

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I dedicated this thesis to my parents. I hope that this achievement will complete the dream you had for me all those many years ago when you chose to give the best education you could.

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

**EARLY DETECTION OF *Ganoderma boninense* IN OIL PALM SEEDLINGS
USING HYPERSPECTRAL IMAGES AND MACHINE LEARNING
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AIMAN NABILAH BINTI NOOR AZMI

January 2021

Chair : Siti Khairunniza binti Bejo, PhD
Faculty : Engineering

Basal stem rot (BSR) caused by *Ganoderma boninense* (*G. boninense*) fungus is one of the most destructive diseases of oil palm plantations in Southeast Asia that resulted in losses up to USD500 million annually. Besides mature trees, seedlings are also susceptible to *G. boninense* infection after being transplanted into a plantation. Therefore, early detection, timely prevention and control are crucial because trees with less than 20% infection can still be treated. This study focuses on early detection of *G. boninense* in oil palm seedlings based on the physical growth of seedlings, spectral reflectance of leaves and machine learning classification. Twenty-eight oil palm seedlings aged five months old were used whereby 15 of them were inoculated with the *G. boninense* pathogen. The physical growth of the seedlings was monitored for every two weeks, and parameters recorded were fronds count, chlorophyll content, height, and girth. The physical growth did not provide any significant differences between the uninoculated (H) and inoculated (U) seedlings throughout the study, which indicate no BSR symptoms had appeared; however, the H obtained marginally higher measurements in most weeks. After 20 weeks of inoculation, spectral reflectance oil palm leaflets taken from fronds 1 (F1) and 2 (F2) were obtained using Cubert FirefIEYE S185 hyperspectral camera with wavelength ranging from 450 to 950 nm. The differences between H and U were observed in the NIR and red-edge spectrum for reflectance and first derivative spectra, respectively. Thirty-five bands were found significant for reflectance and 14 bands for first derivative spectra. The bands were later used as input parameters to develop F1, F2, a combination of F1 and F2 (F12), F1 derivative (F1_{dev}), F2 derivative (F2_{dev}), and F12 derivative (F12_{dev}) classification models, i.e., decision trees, discriminant analysis, logistic regression, naïve Bayes, support vector machine (SVM), k-nearest neighbor, and ensemble. These bands were optimised according to the classification accuracy achieved by the models. The result showed that the acceptable

number of bands to develop classification models was 11 bands which obtained accuracies of 100% (F1), 92% (F2), 95% (F12), 97% (F1_{dev}), 90% (F2_{dev}) and 93% (F12_{dev}) which considered the highest of its classes. Overall, 11 bands of F12 provided near good linear SVM model with 95% accuracy and a kappa value of 0.9; it was considered the best model since it did not require complex pre-processing to separate F1 and F2. This information is useful in aerial-view applications when applying an unmanned aerial vehicle (UAV) for image acquisition since both fronds can be clearly seen from the top-view image hence could expedite the detection of the BSR disease.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Sarjana Sains

PENGESANAN AWAL *Ganoderma boninense* PADA ANAK POKOK KELAPA SAWIT MENGGUNAKAN IMEJ HIPERSPEKTRAL DAN TEKNIK PENGELASAN MESIN PEMBELAJARAN

Oleh

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Reput pangkal batang (BSR) yang berpunca daripada kulat *Ganoderma boninense* (G. boninense) adalah salah satu penyakit yang paling bahaya pada ladang kelapa sawit di Asia Tenggara yang mengakibatkan kerugian hingga USD500 juta setiap tahun. Selain pokok matang, anak pokok juga berpotensi untuk terkena jangkitan G. boninense setelah dipindahkan ke ladang komersial. Kajian intensif telah dilakukan untuk mencari teknik yang mudah untuk mengesan tahap jangkitan awal, dan potensi untuk menggabungkan imej dengan maklumat spektral telah dikenalpasti dengan menggunakan penginderaan jauh hiperspektral. Sebelum imej hiperspektral diperoleh, pertumbuhan fizikal anak pokok kelapa sawit dipantau sepanjang 20 minggu inokulasi dengan patogen G. boninense kerana jangkitan G. boninense boleh memberi kesan pada pertumbuhan fizikal kelapa sawit apabila jangkitan semakin parah. Parameter fizikal yang telah direkodkan adalah bilangan pelepah, kandungan klorofil daun, tinggi anak pokok dan lilitan batang. Hasil kajian menunjukkan bahawa secara purata, anak pokok yang tidak diinokulasi (H) lebih tinggi 2.30 cm, lebih lebar 1.35 mm dan mempunyai lebih banyak pelepah daripada anak pokok yang diinokulasi (U). Bagaimanapun, parameter ini tidak memberikan perbezaan yang signifikan secara statistik di antara H dan U, walaupun anak pokok H memperoleh bacaan yang lebih tinggi pada kebanyakan minggu. Pada 20 minggu inokulasi, imej hiperspektral anak pokok diperoleh untuk mengkaji pantulan spektral daun daripada pelepah nombor satu (F1) dan dua (F2) pada keadaan H dan U. Penyingkiran kebisingan dilakukan untuk membersihkan 558 dan 564 pantulan spektra daripada F1 dan F2, masing-masing daripada data yang tidak diingini. Pantulan spektra kemudian dipuratakan dan diplot di mana perbezaan antara H dan U dilihat jelas dalam spektrum inframerah dekat (NIR) untuk spektra pantulan dan rantau tepi merah untuk spektra terbitan pertama. Berdasarkan purata spektra, sejumlah 35 dan 14 jalur dikenalpasti signifikan secara statistik untuk F1, F2 dan gabungan F1 dan F2 (F12) dalam pantulan dan turunan pertama, masing-masing. Jalur signifikan yang telah dikenalpasti digunakan sebagai parameter input untuk membangunkan model klasifikasi mesin pembelajaran. Model klasifikasi yang terlibat adalah pokok keputusan, analisis diskriminan, regresi logistik,

naïve Bayes, mesin vektor sokongan (SVM), k-nearest neighbor, dan ensemble. Jumlah jalur signifikan ini dioptimumkan berdasarkan ketepatan klasifikasi yang dicapai oleh model. Hasilnya, bilangan jalur signifikan untuk pantulan yang dioptimumkan adalah 18, 14, 11 dan 9 jalur, sementara turunan pertama adalah band 11, 10 dan 7 jalur. Model yang dikembangkan menggunakan 11 jalur memperoleh ketepatan 100% (F1), 92% (F2), 95% (F12), 97% (F1_{dev}), 90% (F2_{dev}) dan 93% (F12_{dev}) yang dianggap paling tinggi dalam data tersebut. Selanjutnya, 9 jalur daripada F1 menghasilkan ketepatan yang dapat diterima iaitu 99% dengan nilai Kappa 0.99 ketika menggunakan Pokok halus, Pokok sederhana, Pokok kasar, dan Regresi linear. Walau bagaimanapun, ia memerlukan pra-proses yang kompleks untuk memisahkan F1 dari pelepah lain, yang akan meningkatkan masa pengkomputeran. Oleh itu, 11 jalur daripada F12 dapat memberikan model klasifikasi yang hampir baik dengan ketepatan 95% dan nilai Kappa 0.9 menggunakan mesin vektor sokongan (SVM) linear dan SVM kuadratik. Walaupun begitu, penggunaan F12 dapat mengurangi waktu pra-proses kerana tidak perlu memisahkan antara F1 dan F2. Maklumat ini berguna dalam aplikasi pandangan udara ketika menggunakan kenderaan udara tanpa pemandu (UAV) untuk pemerolehan gambar kerana kedua pelepah dapat dilihat dengan jelas dari gambar pandangan atas sehingga dapat mempercepat pengesanan penyakit BSR.

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Thank you.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS

AISA	Airborne imaging spectrometer for application
ANOVA	Analysis of variance
ASW	Average silhouette width
BSR	Basal stem rot
F1	Frond 1
F2	Frond 2
<i>G. boninense</i>	<i>Ganoderma boninense</i>
H	Uninoculated seedlings
kNN	k-nearest neighbor
LDA	Linear discriminant analysis
NIR	Near-infrared
OIF	Optimum index factor
PCR	Polymerase chain reaction
QDA	Quadratic discriminant analysis
RWB	Rubber wood block
SPAD	Soil plant analysis development
SVM	Support vector machines
U	Inoculated seedlings
UAV	Unmanned aerial vehicle

CHAPTER 1

INTRODUCTION

1.1 Research background

Oil palm (*Elaeis guineensis*) is a palm species that has been extensively planted in Southeast Asia, primarily in Indonesia and Malaysia, to fulfil the global demand for vegetable oil due to increasing population, income, and growing biofuel market. In Malaysia, oil palm is the main commodity crop that has significantly contributed to the economic development and stability of the country. Furthermore, exports of palm-based products such as palm oil, palm kernel oil, palm kernel cake and palm-based oleochemicals rose by 12.1% to 27.88 million tonnes as compared to the previous year (MPOB, 2019), maintaining Malaysia as the second-largest exporter in the world.

Malaysia falls under the tropical rainforest climate, which is typically hot, rainy, and very humid. The average temperature ranges from 24 to 32°C with an annual rainfall of around 2000 mm, which is favourable for oil palm growth. The land is also biophysically suitable for oil palm cultivation (Pirker and Mosnier, 2015). As a result, the area planted with oil palms in Malaysia continues to grow annually, with the latest total of 5.9 million hectares (MPOB, 2019). These plantations accounted for 32% of the world palm oil production, which denoted 38% of world exports (Breslin and Nesadurai, 2020). Besides climate and land suitability, the development of Malaysia's oil palm industry is also driven by government policies, government-private sector network cooperation, and technology advancement (Rasiah and Shahrin, 2006).

Nevertheless, the production of oil palm in Southeast Asia has been affected by a never-ending basal stem rot (BSR) disease caused by the *Ganoderma boninense* (*G. boninense*) pathogen. The BSR disease once infected only mature trees; however, a recent study by Sanderson (2005) has reported that seedlings are also susceptible to the infection where the symptoms appear earlier and more severe. The disease affects a plantation by reducing the number of standing trees and the weight of fresh fruit bunches (FFB) (Flood et al., 2000). According to Subagio and Foster (2003), the FFB yield decreased by an average of 0.16 t ha⁻¹ for every dead palm or 35% when half of the planted trees died. Meanwhile, Malaysia has reported annual losses of up to RM 1.5 billion due to this disease, making BSR the most economically devastating disease. Based on the BSR incident rate, the total area affected in 2020 was estimated to be 443,440 hectares, equivalent to 65.6 million oil palms, which is worrying if preventive measures are not implemented (Roslan and Idris, 2012).

G. boninense is a white-rot fungus that cultivates within oil palm stem through uninjured roots while producing enzymes that could degrade the woody tissues, cellulose, and lignin layers of the stem (Paterson, 2007). It also affects the xylem, causing a major water and nutrient distribution problem to the top part of the tree, making the symptoms appear

similar to water stress and nutrient deficiency (Turner and Gillbanks, 1974; Shu'ud et al., 2007; Paterson, 2007). Nutrient deficiency could cause poor production of new leaves resulting in yellowing and browning of older leaves. In severe cases, there is no development of new leaves and fresh bunches, which leads to growth inhibition and yield reduction. According to Idris et al. (2006), *G. boninense* infection could cause 80% of the affected trees to die and 25% to 40% FFB yield losses. However, the trees with an infection of less than 20% can still be treated (Meor et al., 2009). Therefore, it is crucial to detect the BSR disease at an early stage.

The earliest visual symptom of *G. boninense* infection in oil palm seedlings can be seen in the presence of fruiting bodies at the seedling's bole. Meanwhile, the medium symptoms are partial yellowing of leaves or mottling of basal fronds to form necrosis, indicating that over 50% of the stem base has been internally damaged (Naher et al., 2015). However, the fruiting body at the infected bole may present or may not present before or after developing foliar symptoms (Sariah et al., 1994; Idris et al., 2006), making visual identification complicated and confusing. In short, the disease could change the physical appearance and growth of oil palm seedlings, especially in terms of frond count, height, girth, and chlorophyll content of leaves (Naidu et al., 2018). These are all due to the inability to perform normal photosynthesis due to foliar symptoms and water deficiency (Haniff et al., 2005).

Hyperspectral imaging is a passive sensor that measures the light reflected from each pixel of a scene in narrow and contiguous electromagnetic wavelengths. Every pixel in the hyperspectral image contains a complete spectral reflectance equal to the number of wavebands. Such reflection provides enough information to classify and analyse existing materials within the image that makes it possible to detect plant disease remotely (Susič et al., 2018). The recorded spectral reflectance normally represents chlorophyll content, cell structure and water content of leaves (Khosrokhani et al., 2016) that are important for determining plant health status. This device can cover a large-scale plantation in a single flight to collect information faster and thoroughly than the field workers. Shafri et al. (2011) stated that the rationale of using high dimensionality data is to improve the efficacy of discrimination between more specific features and classes.

Some applications of hyperspectral imaging for early identification, rapid evaluation and visualisation were demonstrated by Bravo et al. (2003), Bauriegel et al. (2011), Hillnhütter et al. (2011), Kumar et al. (2012), Calderón et al. (2013), Lowe et al. (2017), Kuska et al. (2015), Moghadam et al. (2017), Xie et al. (2017), Kong et al. (2018) and Bohnenkamp et al. (2019). These researchers used the acquired hyperspectral images and extracted spectral information to study the disease infection in plants. The results generated a high degree of accuracy, which verified the capability of this technique for the early detection of plant disease.

1.2 Problem statement

To date, there is no convenient method has been developed for the early detection of BSR disease. The common practice in plantations is manual detection through visual

examination of physical symptoms that are considered as low accuracy, inconsistent result, time-consuming and labour-intensive due to human dependency. Another major downside of manual detection is the lack of standard physical symptoms coupled with error-prone methods, which have led to contradictory assessment (Izzati et al., 2008; Kok et al., 2013; Naidu et al., 2018). In contrast, laboratory-based methods are reliable for early disease detection. However, it is only applicable to small scale samples since it comprises stem collection and tedious laboratory procedures (Naher et al., 2013). Meanwhile, a spectroscopy device can detect an early stage of BSR disease in oil palm seedlings using reflectance spectra of leaves (Shafri and Anuar, 2008; Shafri and Hamdan, 2009; Shafri et al., 2011; Izzuddin et al., 2017). However, this device can only take one reading per time for a tiny leaf area, thus increasing data collection duration and impractical for large scale plantations.

Based on previous studies, it can be concluded that hyperspectral imaging is feasible to be used for early detection of BSR; yet no study has been conducted for oil palm seedlings. Oil palm seedlings at the age of 12 – 48 months old are susceptible to *G. boninense* infection after being transferred and planted in a plantation (Sanderson, 2005). Early detection of infection is crucial to ensure sufficient treatment time, which prevents the disease from spreading. The hyperspectral imaging studies available at this time were carried out in oil palm plantations using vegetation indices and conventional classification methods (Shafri and Hamdan, 2009; Shafri and Mohanad, 2009; Shafri et al., 2012; Izzuddin et al., 2015; Izzuddin et al., 2018). The results obtained by Shafri and Mohanad (2009), Shafri and Hamdan (2009), and Shafri et al. (2012) showed a positive correlation of over 80% between reflectance spectra of leaves and BSR status of oil palm trees. These studies supported the potential use of hyperspectral imaging in collecting spectral information to distinguish between healthy and *G. boninense* infected oil palm seedlings.

1.3 Objectives

The general objective of this research is to detect an early stage of *G. boninense* in oil palm seedlings based on the growth of seedlings, spectral reflectance of leaves and machine learning classification.

The specific objectives of this research are:

1. To examine the effects of *G. boninense* infection on the physical growth of oil palm seedlings.
2. To assess the spectral reflectance of leaves taken from different fronds number at the uninoculated (H) and inoculated (U) condition.
3. To identify the best classification models for early detection of *G. boninense* using machine learning techniques.

REFERENCES

- Abdullah, A. H., Adom, A. H., Ahmad, M. N., Saad, M. A., Tan, E. S., Fikri, N. A., & Zakaria, A. (2011). Electronic nose system for *Ganoderma* detection. *Sensor Letters*, 9(1), 353-358.
- Abdullah, J., Hassan, H., Shari, M. R., Mohd, S., Mustapha, M., Mahmood, A. A., & Hamid, N. H. (2013). GammaScorpion: mobile gamma-ray tomography system for early detection of basal stem rot in oil palm plantations. *Optical Engineering*, 52(3), 036502.
- Abdulridha, J., Ampatzidis, Y., Roberts, P., & Kakarla, S. C. (2020). Detecting powdery mildew disease in squash at different stages using UAV-based hyperspectral imaging and artificial intelligence. *Biosystems Engineering*, 197, 135-148.
- Acuña, E., & Rodriguez, C. (2004). On detection of outliers and their effect in supervised classification. *University of Puerto Rico at Mayaguez*.
- Adão, T., Hruška, J., Pádua, L., Bessa, J., Peres, E., Morais, R., & Sousa, J. J. (2017). Hyperspectral imaging: A review on UAV-based sensors, data processing and applications for agriculture and forestry. *Remote Sensing*, 9(11), 1110.
- Ahmadi, P., Muharam, F. M., Ahmad, K., Mansor, S., & Abu Seman, I. (2017). Early detection of *Ganoderma* basal stem rot of oil palms using artificial neural network spectral analysis. *Plant Disease*, 101(6), 1009-1016.
- Alang, Z. C., Moir, G. F. J., & Jones, L. H. (1988). Composition, degradation and utilization of endosperm during germination in the oil palm (*Elaeis guineensis* Jacq.). *Annals of Botany*, 61(2), 261-268.
- Alexander, A., Dayou, J., Sipaut, C. S., ChongKhim, P., & LeePing, C. (2014). Some interpretations on FTIR results for the detection of *Ganoderma boninense* in oil palm tissue. *Advance Environment Biology*, 8, 30-32.
- Ali, H., Lali, M. I., Nawaz, M. Z., Sharif, M., & Saleem, B. A. (2017). Symptom based automated detection of citrus diseases using color histogram and textural descriptors. *Computers and Electronics in agriculture*, 138, 92-104.
- Ariffin, D., Idris A. S., & Marzuki A. (1996). Spread of *Ganoderma boninense* and vegetative compatibility studies of a single field palm isolates. In *Proceedings of the 1996 PORIM International Palm Oil Congress (Agriculture)*.
- Asaari, M. S. M., Mishra, P., Mertens, S., Dhondt, S., Inzé, D., Wuyts, N., & Scheunders, P. (2018). Close-range hyperspectral image analysis for the early detection of stress responses in individual plants in a high-throughput phenotyping platform. *ISPRS Journal of Photogrammetry and Remote Sensing*, 138, 121-138.

- Asraf, H. M., Nooritawati, M. T., & Rizam, M. S. (2012). A comparative study in kernel-based support vector machine of oil palm leaves nutrient disease. *Procedia Engineering*, 41, 1353-1359.
- Ausmus, B. S., & Hilty, J. W. (1971). Reflectance studies of healthy, maize dwarf mosaic virus-infected, and *Helminthosporium maydis*-infected corn leaves. *Remote Sensing of Environment*, 2, 77-81.
- Azuan, N. H., Khairunniza-Bejo, S., Abdullah, A. F., Kassim, M. S. M., & Ahmad, D. (2019). Analysis of changes in oil palm canopy architecture from basal stem rot using terrestrial laser scanner (TLS). *Plant Disease*, 103(12), 3218-3225.
- Baker, J. M., Deng, L., Glass, J., Khudanpur, S., Lee, C. H., Morgan, N., & O'Shaughnessy, D. (2009). Developments and directions in speech recognition and understanding, Part 1 [DSP Education]. *IEEE Signal Processing Magazine*, 26(3), 75-80.
- Bauriegel, E., Giebel, A., & Herppich, W. B. (2011). Hyperspectral and chlorophyll fluorescence imaging to analyse the impact of *Fusarium culmorum* on the photosynthetic integrity of infected wheat ears. *Sensors*, 11(4), 3765-3779.
- Behmann, J., Steinrücken, J., & Plümer, L. (2014). Detection of early plant stress responses in hyperspectral images. *ISPRS Journal of Photogrammetry and Remote Sensing*, 93, 98-111.
- Bevan, J. W. L., & Gray, B. S. (1969). The organization and control of field practice for large-scale oil palm planting in Malaysia. *The Incorporated Society of Planters*, No. L-0101.
- Boatman, S. G., & Crombie, M. (1958). Fat metabolism in the West African oil palm (*Elaeis guineensis*). *Journal of Experimental Botany*, 9(1), 52-74.
- Bohnenkamp, D., Kuska, M. T., Mahlein, A. K., & Behmann, J. (2019). Hyperspectral signal decomposition and symptom detection of wheat rust disease at the leaf scale using pure fungal spore spectra as reference. *Plant Pathology*, 68(6), 1188-1195.
- Bravo, C., Moshou, D., West, J., McCartney, A., & Ramon, H. (2003). Early disease detection in wheat fields using spectral reflectance. *Biosystems Engineering*, 84(2), 137-145.
- Breslin, S., & Nesadurai, H. E. (Eds.). (2020). *Non-State Actors and Transnational Governance in Southeast Asia*. Routledge.
- Breton, F., Hasan, Y., Lubis, Z., & De Franqueville, H. (2005). Characterization of parameters for the development of an early screening test for basal stem rot tolerance in oil palm progenies. *Malaysian Palm Oil Board (MPOB)*.
- Bruce Hapke (2016). Reflectance methods and applications. In *Encyclopedia of spectroscopy and spectrometry* (pp. 931-935). Elsevier.

- Calderón, R., Navas-Cortés, J. A., Lucena, C., & Zarco-Tejada, P. J. (2013). High-resolution airborne hyperspectral and thermal imagery for early detection of Verticillium wilt of olive using fluorescence, temperature and narrow-band spectral indices. *Remote Sensing of Environment*, 139, 231-245.
- Chappelle, E. W., Kim, M. S., & McMurtrey III, J. E. (1992). Ratio analysis of reflectance spectra (RARS): an algorithm for the remote estimation of the concentrations of chlorophyll a, chlorophyll b, and carotenoids in soybean leaves. *Remote Sensing of Environment*, 39(3), 239-247.
- Cha-um, S., Yamada, N., Takabe, T., & Kirdmanee, C. (2013). Physiological features and growth characters of oil palm (*Elaeis guineensis* Jacq.) in response to reduced water-deficit and rewatering. *Australian Journal of Crop Science*, 7(3), 432.
- Chung, C. L., Huang, K. J., Chen, S. Y., Lai, M. H., Chen, Y. C., & Kuo, Y. F. (2016). Detecting Bakanae disease in rice seedlings by machine vision. *Computers and Electronics in Agriculture*, 121, 404-411.
- Collins, W. (1978). Remote sensing of crop type and maturity. *Photogrammetric Engineering and Remote Sensing*, 44(1), 43-55.
- Cooper, R. M., Flood, J., & Rees, R. W. (2011). *Ganoderma boninense* in oil palm plantations: current thinking on epidemiology, resistance and pathology. *Planter*, 87(1024), 515-526.
- Corley, R. H. V. (1979). Palm oil composition and oil palm breeding. *Planter*, 55(642), 467-478.
- Corley, R. H. V., & Tinker, P. B. (2003). The palm oil. *World Agriculture Series*.
- Dale, L. M., Thewis, A., Boudry, C., Rotar, I., Dardenne, P., Baeten, V., & Pierna, J. A. F. (2013). Hyperspectral imaging applications in agriculture and agro-food product quality and safety control: a review. *Applied Spectroscopy Reviews*, 48(2), 142-159.
- Darmono, T. W. (1999). Detection of basal stem rot disease of oil palm using polyclonal antibody. *Menara Perkebunan*, 67(1), 32-39.
- Darus A., Seman A., Khairuddin H. (1993) Confirmation of *Ganoderma* infected palm by drilling technique. In *PORIM International Palm Oil Congress: Update and Vision (Agriculture)*. PORIM, Malaysia, p. 735-738.
- Dayou, J., Alexander, A., Sipaut, C.S., Phin, C.K., & Chin, L.P. (2014). On the possibility of using FTIR for detection of *Ganoderma boninense* in infected oil palm tree. *Journal of Advance Agriculture Environment Engineering*, 1:161–163.
- Dobson, A. J., & Barnett, A. G. (2018). *An introduction to generalized linear models*. CRC press.
- Duckett, J. E. (1999). A Guide to Oil Palm Nursery. *The Incorporated*.

- Ebrahimi, M. A., Khoshtaghaza, M. H., Minaei, S., & Jamshidi, B. (2017). Vision-based pest detection based on SVM classification method. *Computers and Electronics in Agriculture*, 137, 52-58.
- Ezenne, G. I., Jupp, L., Mantel, S. K., & Tanner, J. L. (2019). Current and potential capabilities of UAS for crop water productivity in precision agriculture. *Agricultural Water Management*, 218, 158-164.
- Fairhurst, T. H. (1998). The oil palm-leaf sampling. *Potash and Phosphate Institute, Singapore*.
- Feng, Y., Peng, Y., Cui, N., Gong, D., & Zhang, K. (2017). Modeling reference evapotranspiration using extreme learning machine and generalized regression neural network only with temperature data. *Computers and Electronics in Agriculture*, 136, 71-78.
- Ferentinos, K. P. (2018). Deep learning models for plant disease detection and diagnosis. *Computers and Electronics in Agriculture*, 145, 311-318.
- Flood, J., Hasan, Y., Turner, P. D., & O'Grady, E. B. (2000). The spread of *Ganoderma* from infective sources in the field and its implications for management of the disease in oil palm. *Ganoderma Diseases of Perennial Crops*, 101-112.
- Flood, J., Keenan, L., Wayne, S., & Hasan, Y. (2005). Studies on oil palm trunks as sources of infection in the field. *Mycopathologia*, 159(1), 101-107.
- Friedl, M. A., Brodley, C. E., & Strahler, A. H. (1999). Maximizing land cover classification accuracies produced by Decision trees at continental to global scales. *IEEE Transactions on Geoscience and Remote Sensing*, 37(2), 969-977.
- Frigge, M., Hoaglin, D. C., & Iglewicz, B. (1989). Some implementations of the boxplot. *The American Statistician*, 43(1), 50-54.
- Gausman, H. W. (1977). Reflectance of leaf components. *Remote sensing of Environment*, 6(1), 1-9.
- Goetz, A. F., Vane, G., Solomon, J. E., & Rock, B. N. (1985). Imaging spectrometry for earth remote sensing. *Science*, 228(4704), 1147-1153.
- Goh, K. M., Dickinson, M., Alderson, P., Yap, L. V., & Supramaniam, C. V. (2016). Development of an in planta infection system for the early detection of *Ganoderma* spp. in oil palm. *Journal of Plant Pathology*, 255-264.
- Goh, Y. K., Ng, F. W., Kok, S. M., Goh, Y. K., & Goh, K. J. (2014). Aggressiveness of *Ganoderma boninense* isolates on the vegetative growth of oil palm (*Elaeis guineensis*) seedlings at different ages. *Malaysian Journal of Applied Biology*, 43, 9-16.
- Govender, N. T., Mahmood, M., Seman, I. A., & Wong, M. Y. (2017). The phenylpropanoid pathway and lignin in defense against *Ganoderma boninense*

- colonized root tissues in oil palm (*Elaeis guineensis* Jacq.). *Frontiers in Plant Science*, 8, 1395.
- Grinblat, G. L., Uzal, L. C., Larese, M. G., & Granitto, P. M. (2016). Deep learning for plant identification using vein morphological patterns. *Computers and Electronics in Agriculture*, 127, 418-424.
- Gunn, J. S., Sly, J. M. A., & Chapas, L. C. (1961). The development of improved nursery practices for the oil palm in West Africa. *Journal of the West African Institute for Oil Palm Research*, 3, 198-232.
- Hagen, N. A., & Kudenov, M. W. (2013). Review of snapshot spectral imaging technologies. *Optical Engineering*, 52(9), 090901.
- Halicek, M., Fabelo, H., Ortega, S., Callico, G. M., & Fei, B. (2019). In-vivo and ex-vivo tissue analysis through hyperspectral imaging techniques: Revealing the invisible features of cancer. *Cancers*, 11(6), 756.
- Haniff, M. H., Ismail, S., & Idris, A. S. (2005). Gas exchange responses of oil palm to *Ganoderma boninense* infection. *Asian Journal of Plant Sciences*, 4(4), 438-444.
- Hartley, C. (1988). The oil palm. 3rd Edition, *Editorial Longman*, London, 958.
- Helmi, Z., & Mohanad, S. E. (2009). Quantitative performance of spectral indices in large scale plant health analysis. *American Journal of Agricultural and Biological Sciences*, 4(3), 187-191.
- Heriansyah, C., & Tan, C. (2005). Nursery practices for production of superior oil palm planting materials. *The Planter*, 81(948), 159-171.
- Hillnhütter, C., Mahlein, A. K., Sikora, R. A., & Oerke, E. C. (2011). Remote sensing to detect plant stress induced by *Heterodera schachtii* and *Rhizoctonia solani* in sugar beet fields. *Field Crops Research*, 122(1), 70-77.
- Hu, H., Pan, L., Sun, K., Tu, S., Sun, Y., Wei, Y., & Tu, K. (2017). Differentiation of deciduous-calyx and persistent-calyx pears using hyperspectral reflectance imaging and multivariate analysis. *Computers and Electronics in Agriculture*, 137, 150-156.
- Huang, W., Lamb, D. W., Niu, Z., Zhang, Y., Liu, L., & Wang, J. (2007). Identification of yellow rust in wheat using in-situ spectral reflectance measurements and airborne hyperspectral imaging. *Precision Agriculture*, 8(4-5), 187-197.
- Husin, N. A., Khairunniza-Bejo, S., Abdullah, A. F., Kassim, M. S. M., & Ahmad, D. (2019). Effects of basal stem rot on oil palm inter-frond angles for different severity levels. *Journal of Advanced Agricultural Technologies*, 6(2).
- Husin, N. A., Khairunniza-Bejo, S., Abdullah, A. F., Kassim, M. S., Ahmad, D., & Azmi, A. N. (2020a). Application of ground-based LiDAR for analysing oil palm

canopy properties on the occurrence of basal stem rot (BSR) Disease. *Scientific reports*, 10(1), 1-16.

- Husin, N. A., Khairunniza-Bejo, S., Abdullah, A. F., Kassim, M. S., Ahmad, D., & Aziz, M. H. (2020b). Classification of Basal Stem Rot Disease in Oil Palm Plantations Using Terrestrial Laser Scanning Data and Machine Learning. *Agronomy*, 10(11), 1624.
- Idris, A. S. (2009). Basal Stem Rot in Malaysia-Biology, economic importance, epidemiology, detection and control. In *International Workshop on Awareness, Detection and Control of Oil Palm Devastating Diseases*, 6.
- Idris, A. S., & Rafidah, A. R. (2008). Polyclonal antibody for detection of Ganoderma. *MPOB Information Series, MPOB TT*, 405.
- Idris, A. S., Kushairi, A., Ismail, S., & Ariffin, D. (2004). Selection for partial resistance in oil palm progenies to Ganoderma basal stem rot. *J Oil Palm Res*, 16(2), 12-18.
- Idris, A. S., Kushairi, D., Ariffin, D., & Basri, M. W. (2006). Technique for inoculation of oil palm germinated seeds with Ganoderma. *MPOB Information Series*, 314, 1-4.
- Idris, A. S., Mazliham, M. S., Loonis, P., & Wahid, M. B. (2010). GanoSken for early detection of Ganoderma infection in oil palm. *MPOB Information Series TT*, (442), 4.
- Idris, A.S., Yamaoka, M., Hayakawa, S., Basri, M.W., Noorhasimah, I., Ariffin, D., 2003. PCR technique for detection of Ganoderma. *MPOB Information Series*, No.188,4.
- Ishaq, I., Alias, M. S., Kadir, J., & Kasawani, I. (2014). Detection of basal stem rot disease at oil palm plantations using sonic tomography. *Journal of Sustainability Science and Management*, 9(2), 52-57.
- Izzati, M. Z. N. A., & Abdullah, F. (2008). Disease suppression in Ganoderma-infected oil palm seedlings treated with *Trichoderma harzianum*. *Plant Protection Science*, 44(3), 101-107.
- Izzuddin, M. A., Idris, A. S., Wahid, O., Nishfariza, M. N., & Shafri, H. Z. M. (2013). Field spectroscopy for detection of Ganoderma disease in oil palm. *MPOB Information Series*, 532(4).
- Izzuddin, M. A., Idris, A. S., Nisfariza, N. M., & Ezzati, B. (2015). Spectral based analysis of airborne hyperspectral remote sensing image for detection of ganoderma disease in oil palm. In *International Conference on Biological and Environmental Science (BIOES)* (pp. 13-20).
- Izzuddin, M. A., Seman Idris, A., Nisfariza, M. N., Nordiana, A. A., Shafri, H. Z. M., & Ezzati, B. (2017). The development of spectral indices for early detection of

- Ganoderma disease in oil palm seedlings. *International Journal of Remote Sensing*, 38(23), 6505-6527.
- Izzuddin, M. A., Nisfariza, M. N., Ezzati, B., Idris, A. S., Steven, M. D., & Boyd, D. (2018). Analysis of airborne hyperspectral image using vegetation indices, red edge position and continuum removal for detection of ganoderma disease in oil palm. *Journal of Oil Palm Research*, 30(3), 416-28.
- Jiang, H., Zhang, C., He, Y., Chen, X., Liu, F., & Liu, Y. (2016). Wavelength selection for detection of slight bruises on pears based on hyperspectral imaging. *Applied Sciences*, 6(12), 450.
- Jiménez-Sánchez, G., & Philp, J. (2016). Genomics and the bioeconomy: Opportunities to meet global challenges. In *Genomics and Society* (pp. 207-238). Academic Press.
- Kale, K. V., Solankar, M. M., Nalawade, D. B., Dhumal, R. K., & Gite, H. R. (2017). A research review on hyperspectral data processing and analysis algorithms. *Proceedings of the National Academy of Sciences, India Section A: Physical Sciences*, 87(4), 541-555.
- Kandan, A., Radjacommar, R., Ramanathan, A., Raguchander, T., Balasubramanian, P., & Samiyappan, R. (2009). Molecular biology of Ganoderma pathogenicity and diagnosis in coconut seedlings. *Folia Microbiologica*, 54(2), 147-152.
- Khairudin, H., Lim, T. K., & Abdul Razak, A. R. (1991). Pathogenicity of *Ganoderma boninense* on oil palm seedlings. In *Proceedings of the 1991 PORIM International Congress (Agriculture)*. Kuala Lumpur, Malaysia: Palm Oil Research Institute of Malaysia, 418, pp. 423.
- Khairunniza-Bejo, S., & Vong, C. N. (2014). Detection of basal stem rot (BSR) infected oil palm tree using laser scanning data. *Agriculture and Agricultural Science Procedia*, 2, 156-164.
- Khosrokhani, M., Khairunniza-Bejo, S., & Pradhan, B. (2018). Geospatial technologies for detection and monitoring of Ganoderma basal stem rot infection in oil palm plantations: a review on sensors and techniques. *Geocarto International*, 33(3), 260-276.
- Kitchin, R., & Lauriault, T. P. (2015). Small data in the era of big data. *GeoJournal*, 80(4), 463-475.
- Kok, S., Goh, Y., Tung, H., Goh, K., Wong, W., & Goh, Y. (2013). In vitro growth of *Ganoderma boninense* isolates on novel palm extract medium and virulence on oil palm (*Elaeis guineensis*) seedlings. *Malaysian Journal of Microbiology*, 9(1), 33-42.
- Kong, W., Zhang, C., Huang, W., Liu, F., & He, Y. (2018). Application of hyperspectral imaging to detect *Sclerotinia sclerotiorum* on oilseed rape stems. *Sensors*, 18(1), 123.

- Kotlar, A. M., Iversen, B. V., & de Jong van Lier, Q. (2019). Evaluation of parametric and nonparametric machine - learning techniques for prediction of saturated and near - saturated hydraulic conductivity. *Vadose Zone Journal*, 18(1), 1-13.
- Kumar, A., Lee, W. S., Ehsani, R. J., Albrigo, L. G., Yang, C., & Mangan, R. L. (2012). Citrus greening disease detection using aerial hyperspectral and multispectral imaging techniques. *Journal of Applied Remote Sensing*, 6(1), 063542.
- Kurale, N. G., & Vaidya, M. V. (2018, July). Classification of Leaf Disease Using Texture Feature and Neural Network Classifier. In *2018 International Conference on Inventive Research in Computing Applications (ICIRCA)* (pp. 1-6). IEEE.
- Kushairi, A., Rajanaidu, N., Jalani, B. S., & Zakri, A. H. (1999). Agronomic performance and genetic variability of dura× pisifera progenies. *Journal of Oil Palm Research*, 11(2), 1-24.
- Kuska, M., Wahabzada, M., Leucker, M., Dehne, H. W., Kersting, K., Oerke, E. C., & Mahlein, A. K. (2015). Hyperspectral phenotyping on the microscopic scale: towards automated characterization of plant-pathogen interactions. *Plant Methods*, 11(1), 28.
- Kwak, S. K., & Kim, J. H. (2017). Statistical data preparation: management of missing values and outliers. *Korean journal of anesthesiology*, 70(4), 407.
- Laurikkala, J., Juhola, M., Kentala, E., Lavrac, N., Miksch, S., & Kavsek, B. (2000, August). Informal identification of outliers in medical data. In *Fifth International Workshop on Intelligent Data Analysis in Medicine and Pharmacology, 1*, pp. 20-24.
- Lelong, C. C., Roger, J. M., Brégand, S., Dubertret, F., Lanore, M., Sitorus, N. A., ... & Caliman, J. P. (2010). Evaluation of oil-palm fungal disease infestation with canopy hyperspectral reflectance data. *Sensors*, 10(1), 734-747.
- Li, B., Xu, X., Zhang, L., Han, J., Bian, C., Li, G., & Jin, L. (2020). Above-ground biomass estimation and yield prediction in potato by using UAV-based RGB and hyperspectral imaging. *ISPRS Journal of Photogrammetry and Remote Sensing*, 162, 161-172.
- Liaghat, S., Ehsani, R., Mansor, S., Shafri, H. Z., Meon, S., Sankaran, S., & Azam, S. H. (2014a). Early detection of basal stem rot disease (Ganoderma) in oil palms based on hyperspectral reflectance data using pattern recognition algorithms. *International Journal of Remote Sensing*, 35(10), 3427-3439.
- Liaghat, S., Mansor, S., Ehsani, R., Shafri, H. Z. M., Meon, S., & Sankaran, S. (2014b). Mid-infrared spectroscopy for early detection of basal stem rot disease in oil palm. *Computers and Electronics in Agriculture*, 101, 48-54.
- Liakos, K. G., Busato, P., Moshou, D., Pearson, S., & Bochtis, D. (2018). Machine learning in agriculture: A review. *Sensors*, 18(8), 2674.

- Lim, H. P., & Fong, Y. K. (2005). Research on basal stem rot (BSR) of ornamental palms caused by basidiospores from *Ganoderma boninense*. *Mycopathologia*, 159(1), 171-179.
- Lowe, A., Harrison, N., & French, A. P. (2017). Hyperspectral image analysis techniques for the detection and classification of the early onset of plant disease and stress. *Plant Methods*, 13(1), 80.
- Lv, W., & Wang, X. (2020). Overview of hyperspectral image classification. *Journal of Sensors*, 2020.
- MacFarland, T. W., & Yates, J. M. (2016). Mann–whitney u test. In *Introduction to nonparametric statistics for the biological sciences using R* (pp. 103-132). Springer, Cham.
- Madiah, A. Z., Idris, A. S., & Rafidah, A. R. (2014). Polyclonal antibodies of *Ganoderma boninense* isolated from Malaysian oil palm for detection of basal stem rot disease. *African Journal of Biotechnology*, 13(34), 3455-3463.
- Malenovsky, Z., Ufer, C., Lhotáková, Z., Clevers, J. G., Schaepman, M. E., Albrechtová, J., & Cudlín, P. (2006). A new hyperspectral index for chlorophyll estimation of a forest canopy: Area under curve normalised to maximal band depth between 650-725 nm. *EARSeL eProceedings*, 5(2), 161-172.
- Manikandan, S. (2011). Measures of dispersion. *Journal of Pharmacology and Pharmacotherapeutics*, 2(4), 315.
- Markom, M. A., Shakaff, A. M., Adom, A. H., Ahmad, M. N., Hidayat, W., Abdullah, A. H., & Fikri, N. A. (2009). Intelligent electronic nose system for basal stem rot disease detection. *Computers and Electronics in Agriculture*, 66(2), 140-146.
- Mathews, J., Tan, T. H., Yong, K. K., Chong, K. M., Ng, S. K., & Ip, W. M. (2010). Managing oil palm nursery: IOI's experience. *The Planter*, 86(1016), 771-785.
- Matthews, S. G., Miller, A. L., Plötz, T., & Kyriazakis, I. (2017). Automated tracking to measure behavioural changes in pigs for health and welfare monitoring. *Scientific Reports*, 7(1), 1-12.
- McHugh, M. L. (2012). Interrater reliability: the kappa statistic. *Biochemia medica*, 22(3), 276-282.
- Meor, M. S. Y., Khalid M. A., & Idris, A.S. (2009). Identification of basal stem rot disease in local palm oil by microfocus XRF. *Journal of Nuclear and Related Technologies* 6(1) *Special Edition*, 282-287
- Miller, A. R., Kelley, T. J., & White, B. D. (1995). Nondestructive evaluation of pickling cucumbers using visible-infrared light transmission. *Journal of the American Society for Horticultural Science*, 120(6), 1063-1068.

- Mitchell, T. M. (1997). Machine learning.
- Moghadam, P., Ward, D., Goan, E., Jayawardena, S., Sikka, P., & Hernandez, E. (2017). Plant disease detection using hyperspectral imaging. In *2017 International Conference on Digital Image Computing: Techniques and Applications (DICTA)* (pp. 1-8). IEEE.
- Morales, I. R., Cebrián, D. R., Blanco, E. F., & Sierra, A. P. (2016). Early warning in egg production curves from commercial hens: An SVM approach. *Computers and Electronics in Agriculture*, 121, 169-179.
- Morellos, A., Pantazi, X. E., Moshou, D., Alexandridis, T., Whetton, R., Tziotzios, G., & Mouazen, A. M. (2016). Machine learning based prediction of soil total nitrogen, organic carbon and moisture content by using VIS-NIR spectroscopy. *Biosystems Engineering*, 152, 104-116.
- Naher, L., Siddiquee, S., Yusuf, U. K., & Mondal, M. M. A. (2015). Issues of *Ganoderma* spp. and basal stem rot disease management in oil palm. *American Journal of Agricultural Science*, 2(3), 103-107.
- Naher, L., Yusuf, U. K., Ismail, A., Tan, S. G., & Mondal, M. M. A. (2013). Ecological status of *Ganoderma* and basal stem rot disease of oil palms ('*Elaeis guineensis*' Jacq.). *Australian Journal of Crop Science*, 7(11), 1723.
- Naidu, Y., Siddiqui, Y., Rafii, M. Y., Saud, H. M., & Idris, A. S. (2018). Inoculation of oil palm seedlings in Malaysia with white-rot hymenomycetes: Assessment of pathogenicity and vegetative growth. *Crop Protection*, 110, 146-154.
- Najihah, N. I., Hanafi, M. M., Idris, A. S., & Hakim, M. A. (2015). Silicon treatment in oil palms confers resistance to basal stem rot disease caused by *Ganoderma boninense*. *Crop Protection*, 67, 151-159.
- Nantasenamat, C., Isarankura-Na-Ayudhya, C., & Prachayasittikul, V. (2010). Advances in computational methods to predict the biological activity of compounds. *Expert opinion on drug discovery*, 5(7), 633-654.
- Natarajan, S., Bhaskaran, R., & Shanmugan, N. (1986). Preliminary studies to develop techniques for early detection of Thanjavur wilt in coconut. *Indian Coconut Journal (India)*, 17(3), 3-6.
- Navaratnam, S. J., & Chee, K. (1965). Root inoculation of oil palm seedlings with *Ganoderma* sp. *Plant Disease*, 49, 1011-1012.
- Nisfariza, M. N., Idris, A. S., Shafri, Z. H., Steven, M., Boyd, D. (2009). Hyperspectral derivative band ratios of oil palm stress associated with *Ganoderma* basal stem rot disease. *MRSS 2010, PWTC, Kuala Lumpur, Malaysia*.
- Nisfariza, M. N., Shafri, Z. H., Idris, A., Steven, M., Boyd, D., Mior, M. (2010). Hyperspectral sensing possibilities using continuum removal index in early detection of *Ganoderma* in oil palm plantation. *Conference on World*

- Oettli, P., Behera, S. K., & Yamagata, T. (2018). Climate based predictability of oil palm tree yield in Malaysia. *Scientific reports*, 8(1), 1-13.
- Panchal, G., & Bridge, P. D. (2005). Following basal stem rot in young oil palm plantings. *Mycopathologia*, 159(1), 123-127.
- Pantazi, X. E., Tamouridou, A. A., Alexandridis, T. K., Lagopodi, A. L., Kontouris, G., & Moshou, D. (2017). Detection of *Silybum marianum* infection with *Microbotryum silybum* using VNIR field spectroscopy. *Computers and Electronics in Agriculture*, 137, 130-137.
- Paterson, R. R. M. (2007). Ganoderma disease of oil palm—A white rot perspective necessary for integrated control. *Crop Protection*, 26(9), 1369-1376.
- Pirker, J., & Mosnier, A. (2015). Global oil palm suitability assessment (Interim Report No. IR-13).
- Polder, G., Pekkeriet, E. J., & Snickers, M. (2013). A spectral imaging system for detection of botrytis in greenhouses. *Conference of Sustainable Agriculture through ICT Innovation*.
- Quencez, P., & Dufour, F. (1982). Chemical weed control in palm plantations. II. Usual herbicidal active ingredients for oil palm culture, and treatment techniques. *Oleagineux*, 37(3), 107-113.
- Rakib, M. R. M., Borhan, A. H., & Jawahir, A. N. (2019). The relationship between SPAD chlorophyll and disease severity index in Ganoderma-infected oil palm seedlings. *Journal of the Bangladesh Agricultural University*, 17(3), 355-358.
- Ramos, P. J., Prieto, F. A., Montoya, E. C., & Oliveros, C. E. (2017). Automatic fruit count on coffee branches using computer vision. *Computers and Electronics in Agriculture*, 137, 9-22.
- Rao, V., Lim, C. C., Chia, C. C., & Teo, K. W. (2003). Studies on Ganoderma spread and control. *Planter*, 79(927), 367-383.
- Rapaport, T., Hochberg, U., Rachmilevitch, S., & Karnieli, A. (2014). The effect of differential growth rates across plants on spectral predictions of physiological parameters. *PloS One*, 9(2).
- Rasiah, R., & Shahrin, A. (2006). Development of palm oil and related products in Malaysia and Indonesia. *University of Malaya*.
- Rees, R. W. (2006). *Ganoderma stem rot of oil palm (Elaeis guineensis): Mode of infection, epidemiology and biological control* (Doctoral dissertation, University of Bath (United Kingdom)).

- Rees, R. W., Flood, J., Hasan, Y., & Cooper, R. M. (2007). Effects of inoculum potential, shading and soil temperature on root infection of oil palm seedlings by the basal stem rot pathogen *Ganoderma boninense*. *Plant Pathology*, 56(5), 862-870.
- Rivera, N. V., Gómez-Sanchis, J., Chanona-Pérez, J., Carrasco, J. J., Millán-Giraldo, M., Lorente, D., & Blasco, J. (2014). Early detection of mechanical damage in mango using NIR hyperspectral images and machine learning. *Biosystems Engineering*, 122, 91-98.
- Roslan, A., & Idris, A. S. (2012). Economic impact of *Ganoderma* incidence on Malaysian oil palm plantation—a case study in Johor. *Oil Palm Industry Economic Journal*, 12(1), 24-30.
- Rumpf, T., Mahlein, A. K., Steiner, U., Oerke, E. C., Dehne, H. W., & Plümer, L. (2010). Early detection and classification of plant diseases with support vector machines based on hyperspectral reflectance. *Computers and Electronics in Agriculture*, 74(1), 91-99.
- Sanderson, F. R. (2005). An insight into spore dispersal of *Ganoderma boninense* on oil palm. *Mycopathologia*, 159(1), 139-141.
- Sanderson, F. R., Pilotti, C. A., & Bridge, P. D. (2000). Basidiospores: Their influence on our thinking regarding a control strategy for basal stem rot of oil palm. *Ganoderma diseases of perennial crops*, 113-119.
- Sapak, Z., Meon, S., & Ahmad, Z. A. M (2008). Effect of endophytic bacteria on growth and suppression of *Ganoderma* infection in oil palm. *International Journal of Agriculture and Biology*, 10, 127-132.
- Sariah, M., Hussin, M. Z., Miller, R. N. G., & Holderness, M. (1994). Pathogenicity of *Ganoderma boninense* tested by inoculation of oil palm seedlings. *Plant Pathology*, 43(3), 507-510.
- Schmidt, K. S., & Skidmore, A. K. (2003). Spectral discrimination of vegetation types in a coastal wetland. *Remote Sensing of Environment*, 85(1), 92-108.
- Schwertman, N. C., Owens, M. A., & Adnan, R. (2004). A simple more general boxplot method for identifying outliers. *Computational statistics & data analysis*, 47(1), 165-174.
- Sellar, R. G., & Boreman, G. D. (2005). Classification of imaging spectrometers for remote sensing applications. *Optical Engineering*, 44(1), 013602.
- Shafri, H. Z. M., & Anuar, M. I. (2008). Hyperspectral signal analysis for detecting disease infection in oil palms. In *2008 International Conference on Computer and Electrical Engineering*, pp. 312-316. IEEE.
- Shafri, H. Z., & Hamdan, N. (2009). Hyperspectral imagery for mapping disease infection in oil palm plantation using vegetation indices and red-edge techniques. *American Journal of Applied Sciences*, 6(6), 1031.

- Shafri, H. Z. M., Anuar, M. I., & Saripan, M. I. (2009). Modified vegetation indices for Ganoderma disease detection in oil palm from field spectroradiometer data. *Journal of Applied Remote Sensing*, 3(1), 033556.
- Shafri, H. Z., Anuar, M. I., Seman, I. A., & Noor, N. M. (2011). Spectral discrimination of healthy and Ganoderma-infected oil palms from hyperspectral data. *International Journal of Remote Sensing*, 32(22), 7111-7129.
- Shafri, H. Z. M., Hamdan, N., & Izzuddin Anuar, M. (2012). Detection of stressed oil palms from an airborne sensor using optimized spectral indices. *International Journal of Remote Sensing*, 33(14), 4293-4311.
- Shu'ud, M. M., Loonis, P., & Seman, I. A. (2007). Towards automatic recognition and grading of Ganoderma infection pattern using fuzzy systems. *Journal of Engineering Computer and Technology*, 19.
- Slaton, M. R., Raymond Hunt Jr, E., & Smith, W. K. (2001). Estimating near-infrared leaf reflectance from leaf structural characteristics. *American Journal of Botany*, 88(2), 278-284.
- Su, Y. X., Xu, H., & Yan, L. J. (2017). Support vector machine-based open crop model (SBOCM): Case of rice production in China. *Saudi Journal of Biological Sciences*, 24(3), 537-547.
- Subagio, A., & Foster, H. L. (2003). Implications of Ganoderma disease on loss in stand and yield production of oil palm in North Sumatra. In *Proceedings of the MAPPS Conference*.
- Susič, N., Žibrat, U., Širca, S., Strajnar, P., Razinger, J., Knapič, M., & Stare, B. G. (2018). Discrimination between abiotic and biotic drought stress in tomatoes using hyperspectral imaging. *Sensors and Actuators: Chemical*, 273, 842-852.
- Tan, G. Y., & Hardon, J. J. (1976). *Nursery selection*, 139-141.
- Tawfik O, Shafri HM, Mohammed AA. 2013. Disease detection from field spectrometer data. *IIUM Engineering Journal*, 14, 133-143.
- Torres, I., Sánchez, M. T., Cho, B. K., Garrido-Varo, A., & Pérez-Marín, D. (2019). Setting up a methodology to distinguish between green oranges and leaves using hyperspectral imaging. *Computers and Electronics in Agriculture*, 167, 105070.
- Turner, P. D. (1981). *Oil palm diseases and disorders*. Oxford University Press.
- Turner, P. D., & Gillbanks, R. A. (1974). *Oil palm cultivation and management*.
- Turner, P., & Gillbanks, R. (2003). Oil Palm Management and Cultivation. *The Incorporated Society of Planters*.
- Uchida, R. (2000). Essential nutrients for plant growth: nutrient functions and deficiency symptoms. *Plant Nutrient Management in Hawaii's Soils*, 31-55.

- Uto, K., Seki, H., Saito, G., Kosugi, Y., & Komatsu, T. (2016). Development of a low-cost hyperspectral whiskbroom imager using an optical fiber bundle, a swing mirror, and compact spectrometers. *IEEE Journal of Selected Topics in Applied Earth Observations and Remote Sensing*, 9(9), 3909-3925.
- Utomo, C., & Niepold, F. (2000). Development of diagnostic methods for detecting Ganoderma-infected oil palms. *Journal of Phytopathology*, 148(9-10), 507-514.
- Utomo, C., Werner, S., Niepold, F., & Deising, H. B. (2005). Identification of Ganoderma, the causal agent of basal stem rot disease in oil palm using a molecular method. *Mycopathologia*, 159(1), 159-170.
- Waibel, A., Hanazawa, T., Hinton, G., Shikano, K., & Lang, K. J. (1989). Phoneme recognition using time-delay neural networks. *IEEE Transactions on Acoustics, Speech, and Signal Processing*, 37(3), 328-339.
- Waske, B., Van der Linden, S., Benediktsson, J. A., Rabe, A., & Hostert, P. (2010). Sensitivity of support vector machines to random feature selection in classification of hyperspectral data. *IEEE Transactions on Geoscience and Remote Sensing*, 48(7), 2880-2889.
- Xie, C., Yang, C., & He, Y. (2017). Hyperspectral imaging for classification of healthy and gray mold diseased tomato leaves with different infection severities. *Computers and Electronics in Agriculture*, 135, 154-162.
- Xie, Q., Dash, J., Huang, W., Peng, D., Qin, Q., Mortimer, H., & Dong, Y. (2018). Vegetation indices combining the red and red-edge spectral information for leaf area index retrieval. *IEEE Journal of Selected Topics in Applied Earth Observations and Remote Sensing*, 11(5), 1482-1493.
- Ye, D., Sun, L., Tan, W., Che, W., & Yang, M. (2018). Detecting and classifying minor bruised potato based on hyperspectral imaging. *Chemometrics and Intelligent Laboratory Systems*, 177, 129-139.
- Yin, J., Wang, Y., & Hu, J. (2012). A new dimensionality reduction algorithm for hyperspectral image using evolutionary strategy. *IEEE Transactions on Industrial Informatics*, 8(4), 935-943.
- Zarco-Tejada, P. J., Miller, J. R., Morales, A., Berjón, A., & Agüera, J. (2004). Hyperspectral indices and model simulation for chlorophyll estimation in open-canopy tree crops. *Remote Sensing of Environment*, 90(4), 463-476.
- Zhang, M., Li, C., & Yang, F. (2017). Classification of foreign matter embedded inside cotton lint using short wave infrared (SWIR) hyperspectral transmittance imaging. *Computers and Electronics in Agriculture*, 139, 75-90.
- Zhang, T., Fan, S., Xiang, Y., Zhang, S., Wang, J., & Sun, Q. (2020). Non-destructive analysis of germination percentage, germination energy and simple vigour index on wheat seeds during storage by VIS/NIR and SWIR hyperspectral

imaging. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 118488.

Zwiggelaar, R. (1998). A review of spectral properties of plants and their potential use for crop/weed discrimination in row-crops. *Crop Protection*, 17(3), 189-206.



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LIST OF PUBLICATIONS

- Azmi, A. N., Bejo, S. K., Jahari, M., Muharam, F. M., & Yule, I. (2021). Differences Between Healthy and *Ganoderma boninense* Infected Oil Palm Seedlings Using Spectral Reflectance of Young Leaf Data. *Basrah Journal of Agricultural Sciences*, 34, 171-179.
- Azmi, A. N. N., Bejo, S. K., Jahari, M., Muharam, F. M., Yule, I., & Husin, N. A. (2020). Early Detection of *Ganoderma boninense* in Oil Palm Seedlings Using Support Vector Machines. *Remote Sensing*, 12(23), 3920.





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