



**UNIVERSITI PUTRA MALAYSIA**

***HISTOPATHOLOGICAL ASSESSMENT OF ACETAMINOPHEN-INDUCED  
LIVER INJURY AND REGENERATION RESPONSE FOLLOWING EDIBLE  
BIRD'S NEST ADMINISTRATION IN ICR AND BALB/C MICE MODEL***

**MUHAMMAD AZAM BIN FAZIL**

**FPV 2021 17**



**HISTOPATHOLOGICAL ASSESSMENT OF ACETAMINOPHEN-INDUCED  
LIVER INJURY AND REGENERATION RESPONSE FOLLOWING EDIBLE  
BIRD'S NEST ADMINISTRATION IN ICR AND BALB/C MICE MODEL**

**By**

**MUHAMMAD AZAM BIN FAZIL**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfilment of the Requirements for the Degree of Master of Science**

**January 2021**

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of the material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

## **HISTOPATHOLOGICAL ASSESSMENT OF ACETAMINOPHEN-INDUCED LIVER INJURY AND REGENERATION RESPONSE FOLLOWING EDIBLE BIRD'S NEST ADMINISTRATION IN ICR AND BALB/C MICE MODEL**

By

**MUHAMMAD AZAM BIN FAZIL**

**January 2021**

**Chairman: Nur Fazila Binti Saulol Hamid, PhD**  
**Faculty: Veterinary Medicine**

Acetaminophen (APAP) overdose is known to induce liver injury in mice models and humans. Excessive accumulation of reactive metabolite, N-acetyl-p-benzoquinone-imine (NAPQI) increases oxidative stress in tissues, leading to centrilobular necrosis after exceeding the glutathione (GSH) threshold. Although liver recovery takes place after clearance of toxic insults, however, the degree of hepatic regeneration differs between mice strains. Therefore, the first aim of the study was a comparative assessment of APAP-induced liver injury (AILI) and subsequent hepatic regeneration of two mouse models (ICR and BALB/c). Meanwhile, edible bird's nest (EBN) is a well-known natural product with various health-enhancing properties including anti-oxidative and cell proliferative effects. However, the role of EBN from a toxicological perspective against APAP toxicity is lacking. Hence, the second objective aimed to assess the prophylactic and regenerative effect of EBN on AILI response in ICR mice. These studies hypothesized that both models respond differently, and self-regenerating hepatocytes could be prompted by prophylaxis EBN. For the first objective, 25 ICR and 20 BALB/c mice were grouped as controls and treatments of 5, 10, 24, and 48 hours post-APAP dosing (hpd) with 5 animals per each. Secondly, 80 ICR mice were assigned to groups of control, APAP (500 mg/kg) and seven days prophylactic of silymarin (200 mg/kg), and EBN (60, 120, and 250 mg/kg) followed by an APAP induction. Livers were harvested for histopathological assessment by haematoxylin and eosin (H&E) staining and proliferating cell nuclear antigen (PCNA) using immunohistochemistry. Results of the first experiment showed that APAP-treated BALB/c mice had an intense hepatocellular injury at 5 hpd than ICR mice that only exhibited damage at 10 hpd before both underwent almost complete regeneration after 24 hpd. The second study revealed significant differences in histological changes between APAP and prophylactic treatment groups at 10 hpd, with complete recovery of all groups observed at 24 hpd except for EBN 250 that sustained injuries. Hepatocytes proliferation was initiated at 5 hpd in silymarin, EBN 60 and EBN 120, while at 24 hpd, EBN 120 and 250 exhibited higher PCNA expressing hepatocytes. The hepatoprotective role was observed earlier in silymarin, EBN 60 and 120, while cellular proliferation was delayed in EBN 250. In

conclusion, all groups showed liver recovery after clearance of APAP insult at later time points but EBN 60 and 120 enhanced the hepatic proliferation as similar to silymarin. Therefore, this proves EBN 60 and 120 could act as prophylaxis liver supplements to accelerate the hepatic regeneration in AILI.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Master Sains

**PENILAIAN HISTOPATOLOGI TERHADAP PENCETUSAN  
ACETAMINOPHEN KEPADA KECEDERAAN HATI DAN TINDAK BALAS  
PERTUMBUHAN SEMULA SELEPAS PEMAKANAN SARANG BURUNG  
BOLEH DIMAKAN DI DALAM MODEL MENCIT ICR DAN BALB/C**

Oleh

**MUHAMMAD AZAM BIN FAZIL**

**Januari 2021**

**Pengerusi: Nur Fazila Binti Saulul Hamid, PhD**  
**Fakulti: Perubatan Veterinar**

Dos acetaminophen (APAP) yang berlebihan dapat mencetuskan kecederaan pada hati di dalam model mencit dan manusia. Lebihan metabolit reaktif, N-acetyl-p-benzoquinone-imine (NAPQI) menambahkan tekanan oksidatif pada tisu dan menyebabkan nekrosis sentrilobular apabila melepasi tahap ambang glutation (GSH). Walaupun pemulihan hati berlaku selepas toksik dibersihkan, namun, darjah percambahan semula hepatic berbeza mengikut jenis mencit. Oleh itu, sasaran pertama untuk kajian ini adalah menilai perbandingan terhadap pencetusan kecederaan hati oleh APAP (AILI) dan seterusnya penjana semula hepatic pada kedua-dua model mencit (ICR dan BALB/c). Sementara itu, sarang burung boleh dimakan (EBN) yang merupakan produk semula jadi dengan kepelbagaian ciri-ciri peningkatan kesihatan seperti kesan antioksidan dan percambahan sel. Walau bagaimanapun, peranan EBN dalam perspektif toksikologi terhadap ketoksikan APAP masih lagi kurang. Maka, objektif kajian yang kedua adalah untuk menilai kesan profilaksis dan percambahan semula oleh EBN pada tindak balas AILI dalam model mencit ICR. Kajian ini menghipotesis tindak balas kedua-dua model mencit berbeza dan percambahan semula jadi hepatosit boleh ditingkatkan dengan profalaksis EBN. Untuk objektif pertama, 25 mencit ICR dan 20 mencit BALB/c dikelompokkan sebagai kawalan dan rawatan pada 5, 10, 24, dan 48 jam selepas dos APAP (hpd) dengan 5 ekor dalam setiap kumpulan. Keduanya, 80 ekor mencit ICR dibahagikan kepada kumpulan kawalan, APAP (500 mg/kg), dan profilaksis 7 hari oleh silymarin (200 mg/kg) dan EBN (60, 120 dan 250 mg/kg diikuti dengan rawatan APAP. Semua sampel hati dikumpulkan untuk penilaian histopatologi dengan pewarnaan haematoxylin and eosin (H&E) dan proliferasi antigen sel nuklear (PCNA) menggunakan immunohistokimia. Keputusan untuk eksperimen pertama menunjukkan mencit BALB/c dirawat APAP mempunyai kecederaan hepatoselular yang tinggi pada 5 hpd berbanding mencit ICR yang menunjukkan kecederaan yang teruk pada 10 hpd sebelum kedua-duanya melalui proses percambahan semula pada 24 hpd. Kajian kedua menunjukkan perbezaan yang signifikan pada perubahan histologi antara APAP dan kumpulan rawatan profalaksis pada 10 hpd

bersama percambahan semula dengan sempurna dalam semua kumpulan pada 24 hpd kecuali untuk EBN 250 dengan mengekalkan kecederaan. Proliferasi hepatosit dicituskan pada 5 hpd untuk kumpulan silymarin, EBN 60 dan EBN 120, manakala proliferasi untuk EBN 250 telah ditangguhkan. Konklusinya, semua kumpulan menunjukkan pemulihan selepas pengurangan APAP pada titik masa yang akhir walaupun tanpa profilaksis EBN tetapi EBN 60 and 120 meningkatkan percambahan semula hepatic seperti silymarin. Ini membuktikan EBN 60 dan 120 dapat menjadi makanan tambahan secara profilaktik untuk mempercepatkan percambahan semula hepatic pada AILI.



## ACKNOWLEDGEMENTS

First and foremost, I would like to express my gratitude for the opportunity to pursue higher education study in a Master's Science degree in Universiti Putra Malaysia (UPM) that has been listed as one of the top universities in Malaysia. It also had been a good experience for me to continue this Master's study as a research course.

My sincere gratitude to my dedicated research supervisor, Dr Nur Fazila Binti Saulol Hamid, PhD, Senior Lecturer, Faculty of Veterinary Medicine, UPM, for believing in me and allowing me to complete my Master's research study by giving guidance and idea throughout this research. Her optimism, vision, sincerity, and motivation have significantly inspired me to give my best in finishing this study. I also want to express my thanks to my co-supervisors; Prof. Dr Noordin Mohamed Mustapha and Assoc. Prof. Dr Nurhusien Yimer Degu, for their guidance and opinions throughout my study.

I am also highly indebted to all staff who help me during my research works especially for the members of Histopathology Laboratory; Mrs Jamilah Jahari, Mrs Latifah Mohd Hanan, and Miss Zainatulaishah Abdul Manap. My extension of gratitude to the other staff members who assist me either directly or indirectly including; Mr Mohd Jamil Samad (Equipment Room), Mr Ghazali Md Yusoff (Veterinary Post-Mortem Laboratory), Mr Zainuddin Ibrahim, Mr Noraziman Sulaiman (Animal Resource Unit), Mr Rusdam Awang (Veterinary Virology Laboratory), Mr Mohamad Ismail Baharom and Dr Mohd Hafidz Mohd Izhar (COMeT, IBS).

Last but not least, a very special thank you to my parents Fazil Bin Abdullah and Napsiah Binti Abdul Kadir, siblings, Fatin Iylia Binti Fazil, Muhammad Azri Bin Fazil, and Nurul Safina Hazira Binti Baharom for always being supportive.



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

**Nur Fazila Saulol Hamid, PhD.**

Senior Lecturer  
Faculty of Veterinary Medicine  
Universiti Putra Malaysia  
(Chairman)

**Noordin Mohamed Mustapha, PhD.**

Professor  
Faculty of Veterinary Medicine  
Universiti Putra Malaysia  
(Member)

**Nurhusien Yimer Degu, PhD.**

Associate Professor  
Faculty of Veterinary Medicine  
Universiti Putra Malaysia  
(Member)

---

**ZALILAH MOHD SHARIFF, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 10 June 2021

## Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations, and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual-property from the thesis and copyright of the thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from the supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before the thesis is published (in the form of written, printed, or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No.: Muhammad Azam Bin Fazil, GS49763

## Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: \_\_\_\_\_

Name of Chairman of

Supervisory Committee: \_\_\_\_\_

Dr Nur Fazila Saulol Hamid

Signature: \_\_\_\_\_

Name of Member of

Supervisory Committee: \_\_\_\_\_

Professor Dr Noordin Mohamed Mustapha

Signature: \_\_\_\_\_

Name of Member of

Supervisory Committee: \_\_\_\_\_

Assoc. Prof. Dr Nurhusien Yimer Degu

## TABLE OF CONTENTS

	<b>Page</b>
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	iii
<b>ACKNOWLEDGEMENT</b>	v
<b>APPROVAL</b>	vi
<b>DECLARATION</b>	viii
<b>LIST OF FIGURES</b>	xiii
<b>LIST OF ABBREVIATIONS</b>	xv
<b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	<b>1</b>
1.1 Background of study	1
1.2 Problem statement	2
1.3 Objectives of research	3
1.4 Hypothesis	3
<b>2 LITERATURE REVIEWS</b>	<b>4</b>
2.1 Laboratory mouse as an animal model for acetaminophen-induced liver injury (AILI)	4
2.1.1 Laboratory mouse as an animal model for liver diseases	4
2.1.2 Laboratory mouse as a model for AILI	4
2.1.3 Gender and strains differences in AILI model	5
2.2 Liver as a major site for APAP metabolization	6
2.2.1 Mechanism of drugs metabolization in the liver	6
2.2.2 Drug-induced liver injury (DILI)	7
2.2.3 History of APAP usage as antipyretic and analgesic drug	8
2.2.4 APAP-induced liver injury (AILI)	9
2.2.5 Mechanism of AILI	9
2.3 Liver regeneration	10
2.3.1 Previous studies on liver regeneration	10
2.3.2 Mechanism of liver regeneration after AILI	11
2.4 Conventional liver tonic and supplement against DILI	11
2.5 N-acetylcysteine as common drugs used in AILI cases	12
2.6 Edible bird's nest (EBN)	13
2.6.1 EBN as a growing supplementary food	13
2.6.2 Health-promoting benefits of EBN	15
2.6.3 Study of EBN in liver health	15
2.7 Potential protective effects of EBN	16
2.7.1 EBN reduces oxidative stress	16
2.7.2 EBN enhances cellular proliferation	16
2.8 Assessment of hepatocellular changes in AILI	17
2.8.1 Cellular injury based on haematoxylin and eosin (H&E) staining	17

2.8.2	Cellular regeneration based on proliferating cell nuclear antigen (PCNA) immunostaining	17
<b>3</b>	<b>GENERAL MATERIALS AND METHODS</b>	<b>19</b>
3.1	Experimental and sampling protocols	19
3.1.1	Ethical approval	19
3.1.2	Animal housing and care	19
3.1.3	Experimental design, treatment, and sampling protocols	19
3.1.4	Determining the number of animals	21
3.1.5	Preparation of EBN extract	21
3.1.6	Preparation of silymarin solution	23
3.1.7	Preparation of APAP solution	23
3.2	Histological examination by H&E staining	23
3.2.1	Preparation of liver tissue samples	23
3.2.2	Haematoxylin and eosin staining	24
3.2.3	Histological assessment in the liver	25
3.3	Immunohistological examination by PCNA immunostaining	26
3.3.1	Immunohistological examination of PCNA	26
3.3.2	Quantification of PCNA-stained hepatocytes	26
3.4	Serum biochemistry test	27
3.5	Statistical analysis	27
<b>4</b>	<b>ASSESSMENT OF HISTOPATHOLOGICAL CHANGES OF ACETAMINOPHEN-INDUCED LIVER INJURY AND ITS SUBSEQUENT REGENERATION IN TWO MOUSE MODELS</b>	<b>28</b>
4.1	Introduction	28
4.2	Materials and methods	29
4.2.1	Ethical consideration	29
4.2.2	Experimental animals	29
4.2.3	Preparation of liver samples	29
4.2.4	Histological examination by H&E staining	29
4.2.5	Histological assessment based on AILI grading score	30
4.2.6	Statistical analysis	30
4.3	Results	31
4.3.1	Histological features of the liver	31
4.3.2	Histological changes based on AILI grading score	35
4.4	Discussion	36
4.5	Conclusion	38
<b>5</b>	<b>THE PROPHYLACTIC EFFECTS OF EDIBLE BIRD'S NEST EXTRACT ON ACETAMINOPHEN-INDUCED LIVER INJURY RESPONSE</b>	<b>39</b>
5.1	Introduction	39
5.2	Materials and methods	40
5.2.1	Ethical approval	40
5.2.2	Experimental animals	40

5.2.3	Experimental design, treatment, and sampling protocols	40
5.2.4	Histological examination by H&E staining	41
5.2.5	Immunohistological examination by PCNA immunostaining	41
5.2.6	Statistical analysis	41
5.3	Results	42
5.3.1	Assessment of histological examination by H&E staining	42
5.3.1.1	Histological features of the liver	42
5.3.1.2	Histological changes based on AILI grading score	47
5.3.2	Assessment of hepatocellular proliferation based on PCNA immunostaining	52
5.3.2.1	Quantification of proliferative hepatocytes at 5 hpd	52
5.3.2.2	Assessment of PCNA-stained hepatocytes at 10 hpd	54
5.3.2.3	Assessment of PCNA-stained hepatocytes at 24 hpd	56
5.3.2.4	Overall quantification of proliferative hepatocytes	58
5.4	Discussion	60
5.5	Conclusion	62
<b>6</b>	<b>GENERAL DISCUSSION</b>	<b>63</b>
<b>7</b>	<b>CONCLUSION AND RECOMMENDATIONS</b>	<b>67</b>
7.1	Conclusion	67
7.2	Recommendation for future research	67
	<b>REFERENCES</b>	<b>69</b>
	<b>APPENDICES</b>	<b>82</b>
	<b>BIODATA OF STUDENT</b>	<b>86</b>
	<b>LIST OF PUBLICATION</b>	<b>87</b>

## LIST OF FIGURES

Figure		Page
1	Laboratory mice of ICR (outbred) and BALB/c (inbred) used in this study	5
2	Mechanism of drug metabolization in the liver	7
3	Mechanism of the APAP overdose metabolisation process by cytochrome p-450 and detoxification by GSH in which the accumulation of NAPQI caused liver injury	10
4	Types of EBN from raw swiftlet's nest, cleaned swiftlet's nest, and processed EBN by hydrolysis process	13
5	Illustration of <i>Aerodramus</i> sp.; <i>A. fuciphagus</i> and <i>A. maximus</i> , and its nest	14
6	Proliferative hepatocytes stained using PCNA immunostaining methods	18
7	Flowchart of animal study for two distinct mice strains (ICR and BALB/c) in AILI involved 45 mice (25 ICR and 20 BALB/c mice)	20
8	Flowchart of animal study for groups of control, silymarin, and EBN in ICR mice	21
9	Illustration of EBN extract solution preparation as prophylactic prior to APAP dosing	22
10	Histopathological assessment features of the liver sample in ICR and BALB/c mice after 5 and 10 hpd APAP induction	32
11	Assessment of histomorphological features of liver sections in both ICR and BALB/c mice at 24 and 48 hpd after 500 mg/kg APAP induction	34
12	Histopathological AILI grading score for groups of control, and APAP-treated mice of ICR and BALB/c at 5, 10, 24, and 48 hpd	36
13	Photomicrograph of the liver of mice at 5 hpd of AILI (H&E)	43
14	Photomicrograph of the liver of mice at 10 hpd of AILI (H&E)	45
15	Photomicrograph of the liver of mice at 24 hpd of AILI (H&E)	46
16	Assessment of AILI scores between different groups at 5, 10 and 24 hpd	48

17	Assessment of AILI scores in EBN 60, 120 and 250 over time points	50
18	Overall assessment of AILI scores for liver samples in ICR mice	51
19	Photomicrographs of PCNA-stained liver sections of mice at 5 hpd	53
20	Quantification of PCNA expressing hepatocytes at 5 hpd	54
21	Photomicrographs of PCNA-stained liver sections at 10 hpd	55
22	Quantification of PCNA expressing hepatocytes during 10 hpd	56
23	Photomicrographs of PCNA-stained liver sections during 24 hpd	57
24	Quantification of PCNA expressing hepatocytes at 24 hpd	58
25	Overall assessment of PCNA positive cells in ICR mice	59



## LIST OF ABBREVIATIONS

[ <sup>3</sup> H]-TdR	Tritiated thymidine
+ve	Positive
-ve	Negative
1BP	1-bromopropane
A.D	Anno domini
AASLD	American Association for the Study of Liver Diseases
ABTS	Azino-bis[3-ethylbenzothiazoline-6-sulphonic acid]
AILI	APAP-induced liver injury
ALT	Alanine aminotransferase
ANOVA	Analysis of variance
APAP	Acetaminophen
AST	Aspartate aminotransferase
BrDU	Bromodeoxyuridine
BWT	Bodyweight
°C	Degree Celsius
c-met	tyrosine-protein kinase Met
Caco-2	colonic adenocarcinoma cell line
CAT	Catalase
CCl <sub>3</sub> -OO*	Trichlorometane
CCl <sub>4</sub>	carbon tetrachloride
CNS	Central nervous system
Cyp1A2	Cytochrome P-4501A2
Cyp2E1	Cytochrome P-4502E1
CYP-450	Cytochrome P-450

DAB	3,3'-diaminobenzidine tetrahydrochloride
ddH <sub>2</sub> O	Distilled water
DEN	Diethylnitrosamine
DILI	Drug-induced liver injury
DNA	Deoxyribonucleic acid
DPX	Dibutyl phthalate polystyrene xylene
EBN	Edible bird's nest
EGF	Epidermal growth factor
EGFR	Epidermal growth factor receptor
g	Gram
GalN	Galactosamine
galNAc	N-acetylgalactosamine
glcNAc	N-acetylglucosamine
GSH	Glutathione
Gsr	Glutathione reductase
GST	Glutathione S-transferase
Gsx	Glutathione oxidase
H <sub>2</sub> O <sub>2</sub>	Hydrogen peroxide
HAC	Human articular chondrocytes
H&E	Haematoxylin and eosin
HGF	Hepatic growth factor
hpd	Hours post APAP dosing
IACUC	Institutional of Animal Care and Use Committee
i.p	Intraperitoneal
IL-6	Interleukin-6
kg	Kilogram

LPS	Lipopolysaccharide
μL	Millilitre
MDA	Malondialdehyde
mg	Milligram
mL	Millilitre
NAC	N-acetyl-cysteine
NaCl	Sodium chloride
NANA	N-acetylneuraminic acid
NAPQI	N-acetyl-p-benzo-quinine-imine
NO	Nitric oxide
ORAC	Oxygen radical absorbance capacity
OECD	Organizational of Economic Corporation and Development
PCNA	Proliferating cell nuclear antigen
PH	Partial hepatectomy
p.s.i	Pound per square inch
ROS	Reduced reactive oxygen species
SD	Standard deviation
SOD	Superoxide dismutase
SPSS	Statistical Package for the Social Science
TAA	Thioacetamide
TAASO	Thioacetamide S-oxide
TNF-α	Tumour necrosis factor-alpha
TRS	Target retrieval solution
TSST	Toxic shock syndrome toxin
VEGF	Vascular epidermal growth factor

## CHAPTER 1

### INTRODUCTION

#### 1.1 Background of study

Acetaminophen (APAP) has been used as an ideal toxicant to induce liver injury in a xenobiotic study (Ostapowicz et al., 2002). Liver injury is exhibited by hepatocellular necrosis as a result of APAP overdose due to excessive production of a reactive metabolite of APAP that is known as N-acetyl-p-benzo-quinone-imine (NAPQI). Accumulation of NAPQI occurs when exceeding the capacity of the antioxidant threshold due to depletion of glutathione (GSH), a scavenger within hepatocytes, that responsible to detoxify the reactive metabolites (Botta et al., 2006). Then, excessive NAPQI will form a covalent bond with intracellular protein for the formation of protein adducts in mitochondria leading to the occurrence of oxidative stress. As a consequence, the toxic oxygen species generate oxidation to critical cellular protein, deoxyribonucleic acid (DNA), and lipid that later on contribute to hepatocytes necrosis. However, hepatocytes are capable to proliferate by entering the mitotic cell cycle to replace the damaged and dead cells. Hence, the recovery of liver injury has proven to take place after the clearance of toxic insult due to the restitution of the hepatic GSH level.

Many studies have been conducted to obtain a relevant animal model that resembles APAP-induced liver injury (AILI) in humans. Laboratory rodents have been recognised as the most suitable study models in preclinical studies of drug hepatotoxicity (Jaeschke et al., 2014). However, laboratory rat was represented as a poor study model to demonstrate hepatotoxicity as it showed resistance towards APAP intoxication (McGill et al., 2012) due to the absence of mitochondrial dysfunction and reduced protein adducts in mitochondria (McGill et al., 2012). Meanwhile, laboratory mouse has been known to be a relevant study model of AILI as it resembles the responses of APAP toxicity in the human body (Jaeschke, 2015) with the exhibition of mitochondrial damage and DNA fragmentation induced by APAP intoxication in mice (Jollow et al., 1973). The different responses of mice gender on APAP toxicity are also studied where male mice were found to be more susceptible to hepatotoxicity than female counterpart due to induction of glutamate-cysteine ligase that correlate with faster recovery of GSH (Ramachandran & Jaeschke, 2017).

Nowadays, natural products have become favourable to some people. To overcome certain adverse effects of xenobiotics, people may shift to alternative medicinal products. One of the nutritious and natural products that have gained more attention recently is the edible bird's nest (EBN). This glutinous EBN is made from the saliva of swiftlets of the *Apodidae* family mainly from the white nest swiftlets (*Aerodramus fuciphagus*) and black nest swiftlets (*Aerodramus maximus*) (Babji et al., 2015; Sheldon, 2011). The blooming EBN industry has become an excellent opportunity to assure a lucrative return for the producers, as there is a high demand for EBN in the world's market. The value of EBN is worth the labour efforts of the farmer and harvester with the range of RM 4,000

to RM 5,000 per kilogram for raw EBN and RM 6,000 to RM 8,000 per kilogram for cleaned EBN (Mohamad Shukri et al., 2018). Artificial birdhouses built by farmers can attract swiftlets to build their nest as an alternative way for nesting instead of caves. South-East Asian countries are widely known region as EBN consumer and the highest producer of EBN globally in which Malaysia is one of the main EBN producers alongside Indonesia and Thailand.

EBN has been originally introduced as Chinese traditional medicine to be consumed for the benefits of health thousands of years ago. However, the health-promoting effects of EBN are based on anecdotal, historical, and observational reports since the 17<sup>th</sup> century (Vimala et al., 2012). Due to this discovery, EBN becomes one of the interest points in natural medicine products. It was first scientifically studied by Ng et al. (1986) who reported the potential effects of EBN in cell proliferation of the immune cells. Thereafter, many scientific studies were conducted to prove the benefits of EBN with the improvement of life quality by various health promising components such as high protein and carbohydrates, including glycoprotein, sialic acid, and other essential trace elements like zinc, iron, manganese, magnesium, calcium, and sodium (Marcone, 2005; Norhayati et al., 2010). A recent study also believed that it helps to reduce oxidative stress (Yida et al., 2014), increase cognitive function (Careena et al., 2018), enhance cell proliferation (Albishtue et al., 2018) and antiviral agent (Akmal et al., 2017).

## **1.2 Problem statement**

The effect of APAP toxicity in humans and mice are almost undistinguishable. However, the extensively used laboratory mice in AILI have become a crucial reason to understand the different reactions of various mouse strains. Differences in mouse strain contribute to the variation in responses of APAP hepatotoxicity as each strain possess a distinct genetic line-up (Shahid & Subhan, 2014). Outbred stocks were known as genetically undefined mice with a heterogeneous genetic line that resembles humans (Chia et al., 2005). Inbred, however, possibly exhibit a high similarity of APAP toxicity response as inbred possess homogenous genetic line-up (Festing, 2010). Diverse mice strain is a challenge for researchers in choosing the suitable mouse model to ensure data validation for drug hepatotoxicity in humans. Even though the response of different strains of mice is still variable, toxic insult will be metabolised and cleared out throughout the body. Subsequently, recovery of the liver has taken place that could be observed by the regeneration of hepatocytes. Thus, the first part of this study is focusing on the assessment of histological changes of APAP-induced liver injury in different strains of ICR and BALB/c mice based on an AILI grading score system. The degree of hepatic regeneration is also evaluated after the clearance of APAP toxicity.

Positive effects of EBN on health have been noticed in toxicology and pharmacology studies especially as antioxidant and cell proliferative agent (Li et al., 2016). However, studies on the effects of EBN on hepatocytes regeneration after APAP overdose are still scarce and need to be further elucidated to provide a better understanding of the potential of EBN as a supplementary enhancer for liver health. The current study is necessary to prove the proliferation of hepatocytes to determine the degree of the hepatoprotective potential of EBN in AILI. We hypothesise that pre-treatment of mice with EBN extract

enables to enhance the rate of liver cell proliferation following injury. Therefore, in the second part of the study, we aim to explore the hepatoprotective effects of EBN extract in AILI mice.

### **1.3 Objectives of research**

The main objective of this study is to assess the protective role and regenerative enhancement effects of edible bird's nest (EBN) on AILI in mice model

The specific objectives of this study are:

1. to assess and compare the histopathological responses of AILI in ICR and BALB/c mice
2. to assess the degree of protection and hepatic cell regeneration conferred by EBN prophylactic treatment against AILI in ICR mice through histological assessment and proliferation cell nuclear antigen (PCNA) immunostaining

### **1.4 Hypothesis**

The null hypothesis of this study is the histological features of AILI in both outbred and inbred mice are the same with both strains reveal subsequent liver regeneration at later time points after clearance of toxicity whereby the regenerative process is not accelerated by the prophylactic effect of EBN extract.

As the alternative hypothesis, the histological features of AILI in both strains would be significantly different and the regeneration process will occur at a later time point whereby the regeneration process is enhanced by the prophylactic effect of EBN extract.

## REFERENCES

- Abd Rashed, A., & Wan Nazaimoon, W. M. (2010). Effect of Edible Bird's Nest on Caco-2 Cell Proliferation. *Journal of Food Technology*, 8(3), 126–130.
- Akhtar, T., & Sheikh, N. (2013). An overview of thioacetamide-induced hepatotoxicity. *Toxin Reviews*, 32(3), 43–46.
- Akmal, M. N., Intan-Shameha, A. R., Ajat, M., & Ideris, A. (2017). Edible bird's nest (EBN) is a potential natural product against influenza virus infection. *Jurnal Veterinar Malaysia*, 29(2), 14–19.
- Akmal, M. N., Intan-Shameha, A. R., Ajat, M., Mansor, R., Zuki, A. B. Z., & Ideris, A. (2018). Edible Bird's Nest (EBN) Supplementation Ameliorates the Progression of Hepatic changes and Atherosclerosis in Hypercholesterolaemic-Induced Rats. *Malaysian Journal of Microscopy*, 14, 103–114.
- Al-Rasheed, N., Faddah, L., Al-Rasheed, N., Bassiouni, Y. A., Hasan, I. H., Mahmoud, A. M., Mohamad, R. A., & Yacoub, H. I. (2016). Protective effects of silymarin, alone or in combination with chlorogenic acid and/or melatonin, against carbon tetrachloride-induced hepatotoxicity. *Pharmacognosy Magazine*, 12(46), 337–345.
- Albishtue, A. A., Almhanna, H. K., Yimer, N., Zakaria, M. Z. A., Haron, A. W., & Almhanawi, B. H. (2020). Effect of edible bird's nest supplement on hepato-renal histomorphology of rats exposed to lead acetate toxicity. *Jordan Journal of Biological Sciences*, 13(2), 213–218.
- Albishtue, A. A., Yimer, N., Zakaria, M. Z. A., Haron, A. W., Babji, A. S., Abubakar, A. A., Baiee, F. H., Almhanna, H. K., & Almhanawi, B. H. (2019). The role of edible bird's nest and mechanism of averting lead acetate toxicity effect on rat uterus. *Veterinary World*, 12(7), 1013–1021.
- Albishtue, A. A., Yimer, N., Zakaria, M. Z. A., Haron, A. W., Yusoff, R., Assi, M. A., & Almhanawi, B. H. (2018). Edible bird's nest impact on rats' uterine histomorphology, expressions of genes of growth factors and proliferating cell nuclear antigen, and oxidative stress level. *Veterinary World*, 11(13), 71–79.
- Algren, D. A. (2008). *Review of n-acetylcysteine for the treatment of acetaminophen (paracetamol) toxicity in pediatrics*. Paper presented at the meeting of Second Meeting of the Subcommittee of the Expert Committee on the Selection and Use of Essential Medicines, Geneva. October, 2008.
- Aller, R., Izaola, O., Gómez, S., Tafur, C., González, G., Berroa, E., Mora, N., González, J. M., & De Luis, D. A. (2015). Effect of silymarin plus Vitamin E in patients with non-alcoholic fatty liver disease. A randomized clinical pilot study. *European Review for Medical and Pharmacological Sciences*, 19(16), 3118–3124.
- Antoine, D. J., Williams, D. P., Jenkins, A. K., Regan, S. L., Sathish, J. G., Kitteringham, N. R., & Park, B. K. (2009a). High-mobility group box-1 protein and keratin-18,

circulating serum proteins informative of acetaminophen-induced necrosis and apoptosis in vivo. *Toxicological Sciences*, 112(2), 521–

Apfel, C. C., Turan, A., Souza, K., Pergolizzi, J., & Hornuss, C. (2013). Intravenous acetaminophen reduces postoperative nausea and vomiting: A systematic review and meta-analysis. *Pain*, 154(5), 677–689.

Apte, U. (2015). Liver Regeneration: An Introduction. In S. Masucci (Ed.), *Liver Regeneration Basic Mechanisms, Relevant Models and Clinical Applications* (pp. 3–11). Mica Haley.

Apte, U., Singh, S., Zeng, G., Cieply, B., Virji, M. A., Wu, T., & Monga, S. P. S. (2009). Beta-Catenin Activation Promotes Liver Regeneration after Acetaminophen-Induced Injury. *The American Journal of Pathology*, 175(3), 1056–1065.

Australian Rheumatology Association. (2016). Paracetamol. In *Arthritis Australia* (pp. 1–3).

Babji, A. S., Nurfatin, M. H., Ety Syarmila, I. K., & Masitah, M. (2015). Secrets of Edible Bird Nest. *Agricultural Science Journal*, 1(1), 32–37.

Bannwarth, B., & Pehourcq, F. (2003). Pharmacological rationale for the clinical use of paracetamol: pharmacokinetic and pharmacodynamic issues. *Drugs*, 63(2), 1–9.

Baserga, R. (1991). Growth regulation of the PCNA gene. *Journal of Cell Science*, 98, 433–436.

Beckett, G. J., Chapman, B. J., Dyson, E. H., & Hayes, J. D. (1985). Plasma glutathione S-transferase measurements after paracetamol overdose: Evidence for early hepatocellular damage. *Gut*, 26, 26–31.

Bertolini, A., Ferrari, A., Ottani, A., Guerzoni, S., Tacchi, R., & Leone, S. (2006). Paracetamol: New vistas of an old drug. *CNS Drug Reviews*, 12(3–4), 250–275.

Bhushan, B., & Apte, U. (2019). Liver Regeneration after Acetaminophen Hepatotoxicity Mechanisms and Therapeutic Opportunities. *The American Journal of Pathology*, 189(4), 719–729.

Bhushan, B., Walesky, C., Manley, M., Gallagher, T., Borude, P., Edwards, G., Monga, S. P. S., & Apte, U. (2014). Pro-regenerative signaling after acetaminophen-induced acute liver injury in mice identified using a novel incremental dose model. *American Journal of Pathology*, 184(11), 3013–3025.

Boll, M., Weber, L. W. D., Becker, E., & Stampfl, A. (2001). Mechanism of carbon tetrachloride-induced hepatotoxicity. Hepatocellular damage by reactive carbon tetrachloride metabolites. *Zeitschrift Fur Naturforschung - Section C Journal of Biosciences*, 56(7–8), 649–659.

Botta, D., Shi, S., White, C. C., Dabrowski, M. J., Keener, C. L., Srinouanprachanh, S. L., Farin, F. M., Ware, C. B., Ladiges, W. C., Pierce, R. H., Fausto, N., & Kavanagh, T. J. (2006). Acetaminophen-induced liver injury is attenuated in male



- glutamate-cysteine ligase transgenic mice. *Journal of Biological Chemistry*, 281(39), 28865 - 28875.
- Caparrotta, T. M., Antoine, D. J., & Dear, J. W. (2018). Are some people at increased risk of paracetamol-induced liver injury? A critical review of the literature. *European Journal of Clinical Pharmacology*, 74(2), 147–160.
- Carbone, L., Carbone, E. T., Yi, E. M., Bauer, D. B., Lindstrom, K. A., Parker, J. M., Austin, J. A., Seo, Y., Gandhi, A. D., & Wilkerson, J. D. (2012). Assessing cervical dislocation as a humane euthanasia method in mice. *Journal of the American Association for Laboratory Animal Science*, 51(3), 352–356.
- Careena, S., Sani, D., Tan, S. N., Lim, C. W., Hassan, S., Norhafizah, M., Kirby, B. P., Ideris, A., Stanslas, J., Bin Basri, H., & Lim, C. T. S. (2018). Effect of Edible Bird's Nest Extract on Lipopolysaccharide-Induced Impairment of Learning and Memory in Wistar Rats. *Evidence-Based Complementary and Alternative Medicine*, 2018, 1 - 7.
- Charan, J., & Kantharia, N. (2013). How to calculate sample size in animal studies? *Journal of Pharmacology and Pharmacotherapeutics*, 4(4), 303–306.
- Chen, P. J., Pang, V. F., Jeng, Y. M., Chen, T. J., Hu, F. C., Chi, W. T., Chou, H. Y., Chiu, H. C., Lee, Y. C., & Sheen, L. Y. (2012). Establishment of a standardized animal model of chronic hepatotoxicity using acetaminophen-induced hepatotoxicity in the evaluation of hepatoprotective effects of health food. *Journal of Food and Drug Analysis*, 20(1), 41–47.
- Chia, R., Achilli, F., Festing, M. F. W., & Fisher, E. M. C. (2005). The origins and uses of mouse outbred stocks. *Nature Genetics*, 37(11), 1181–1186.
- Chua, K. H., Lee, T. H., Nagandran, K., Md Yahaya, N. H., Lee, C. T., Tjih, E. T. T., & Abdul Aziz, R. (2013). Edible Bird's nest extract as a chondro-protective agent for human chondrocytes isolated from osteoarthritic knee: In vitro study. *BMC Complementary and Alternative Medicine*, 13(19), 1 - 9.
- Comelli, M. C., Mengs, U., Schneider, C., & Prosdociami, M. (2007). Toward the definition of the mechanism of action of silymarin: Activities related to cellular protection from toxic damage induced by chemotherapy. *Integrative Cancer Therapies*, 6(2), 120–129.
- D'Orazio, J. L. (2019). *Isoniazid toxicity Clinical Presentation*. Medscape. Retrieved from <https://emedicine.medscape.com/article/180554-clinical>.
- Dahlin, D. C., Miwa, G. T., Lu, A. Y. H., & Nelson, S. D. (1984). N-acetyl-p-benzoquinone imine: A cytochrome P-450-mediated oxidation product of acetaminophen (reactive metabolite/reduction/conjugation) and tDepartment of Animal Drug Metabolism. *Proceedings of the National Academy of Sciences*, 81, 1327 - 1331.
- Dai, G., He, L., Chou, N., & Wan, J. J. Y. (2006). Acetaminophen metabolism does not contribute to gender difference in its hepatotoxicity in mouse. *Toxicological*

*Sciences*, 92(1), 33–41.

- David, S., & Hamilton, J. P. (2010). Drug-induced liver injury. *US Gastroenterology and Hepatology*, 6(1), 4017–4018.
- Davidson, D. G. D., & Eastham, W. N. (1966). Acute Liver Necrosis Following Overdose of Paracetamol. *British Medical Journal*, 2(5512), 497–499.
- de Jong, M., & Maina, T. (2010). Of mice and humans: Are they the same? - Implications in cancer translational research. *Journal of Nuclear Medicine*, 51(4), 501–504.
- Dias, R. S., Carmo, L. S., Heneine, L. G. D., Rocha, P. H., Barbosa, C. F., Rodrigues, R. J., & Linardi, V. R. (2009). The use of mice as animal model for testing acute toxicity (LD-50) of toxic shock syndrome toxin. *Arquivo Brasileiro de Medicina Veterinária e Zootecnia*, 61(1), 170–173.
- Du, K., Williams, C. D., McGill, M. R., Jaeschke, H., & City, K. (2014). Lower Susceptibility of Female Mice to Acetaminophen Hepatotoxicity: Role of Mitochondrial Glutathione, Oxidant Stress and c-Jun N-Terminal Kinase. *Toxicology and Applied Pharmacology*, 281(1), 58–66.
- Eldridge, S. R., Butterworth, B. E., & Goldsworthy, T. L. (1993a). Proliferating cell nuclear antigen: A marker for hepatocellular proliferation in rodents. *Environmental Health Perspectives*, 101(5), 211–218.
- Faul, F., Erdfelder, E., Lang, A. G., & Buchner, A. (2007). G\*Power 3: A flexible statistical power analysis program for the social, behavioral, and biomedical sciences. *Journal of Materials and Environmental Science*, 39(2), 175–191.
- Feldkamp, M. L., Meyer, R. E., Krikov, S., & Botto, L. D. (2010). Acetaminophen use in pregnancy and risk of birth defects: Findings from the national birth defects prevention study. *Obstetrics and Gynecology*, 115(1), 109–115.
- Fengler, V. H. I., Macheiner, T., Kessler, S. M., Czepukojc, B., Gemperlein, K., Müller, R., Kiemer, A. K., Magnes, C., Haybaeck, J., Lackner, C., & Sargsyan, K. (2016). Susceptibility of different mouse wild type strains to develop diet-induced nafld/aflid-associated liver disease. *PLoS ONE*, 11(5), 1–21.
- Festing, M. F. W. (2010). Inbred strains should replace outbred stocks in toxicology, safety testing, and drug development. *Toxicologic Pathology*, 38(5), 681–690.
- Foley, J., Ton, T., Maronpot, R., Butterworth, B., & Goldsworthy, T. L. (1993). Comparison of proliferating cell nuclear antigen to tritiated thymidine as a marker of proliferating hepatocytes in rats. *Environmental Health Perspectives*, 101, 199–205.
- Freitag, A. F., Cardia, G. F. E., da Rocha, B. A., Aguiar, R. P., Silva-Comar, F. M. de S., Spironello, R. A., Grespan, R., Caparroz-Assef, S. M., Bersani-Amado, C. A., & Cuman, R. K. N. (2015). Hepatoprotective effect of silymarin (silybum marianum) on hepatotoxicity induced by acetaminophen in spontaneously hypertensive rats. *Evidence-Based Complementary and Alternative Medicine*,

2015, 1 - 8.

- Gujral, J. S., Knight, T. R., Farhood, A., Bajt, M. L., & Jaeschke, H. (2002). Mode of cell death after acetaminophen overdose in mice: Apoptosis or oncotic necrosis? *Toxicological Sciences*, 67(2), 322–328.
- Guo, J.-T., Zhou, H., Liu, C., Aldrich, C., Saputelli, J., Whitaker, T., Barrasa, M. I., Mason, W. S., & Seeger, C. (2000). Apoptosis and Regeneration of Hepatocytes during Recovery from Transient Hepadnavirus Infections. *Journal of Virology*, 74(3), 1495–1505.
- Guo, Y., Wang, S., Wang, Y., & Zhu, T. (2016). Silymarin improved diet-induced liver damage and insulin resistance by decreasing inflammation in mice. *Pharmaceutical Biology*, 54(12), 2995–3000.
- Gupta, R. K., Hussain, T., Panigrahi, G., Das, A., & Singh, G. N. (2011). Hepatoprotective effect of Solanum xanthocarpum fruit extract against CCl<sub>4</sub> induced acute liver toxicity in experimental animals. *Asian Pacific Journal of Tropical Medicine*, 4(12), 964–968.
- Haghani, A., Mehrbod, P., Safi, N., Kadir, F. A. in. A., Omar, A. R., & Ideris, A. (2017). Edible bird's nest modulate intracellular molecular pathways of influenza A virus infected cells. *BMC Complementary and Alternative Medicine*, 17(1), 1–13.
- Higgins, G. M. (1931). Experimental pathology of the liver. Restoration of the liver of the white rat following partial surgical removal. *Archives of Pathology*, 12, 186 - 202
- Hinson, J. A., Roberts, D. W., & James, L. P. (2010). Mechanisms of Acetaminophen-Induced Liver Necrosis. *Handbook of Experimental Pharmacology*, 196, 369–405.
- Hou, Z., He, P., Imam, M. U., Qi, J., Tang, S., Song, C., & Ismail, M. (2017). Edible Bird's Nest Prevents Menopause-Related Memory and Cognitive Decline in Rats via Increased Hippocampal Sirtuin-1 Expression. *Oxidative Medicine and Cellular Longevity*, 1- 8.
- Hou, Z., Imam, M. U., Ismail, M., Azmi, N. H., Ismail, N., Ideris, A., & Mahmud, R. (2015). Lactoferrin and ovotransferrin contribute toward antioxidative effects of Edible Bird's Nest against hydrogen peroxide-induced oxidative stress in human SH-SY5Y cells. *Bioscience, Biotechnology and Biochemistry*, 79(10), 1570–1578.
- Ibrahim, S. H., Teo, W. C., & Baharun, A. (2009). A Study on suitable habitat for swiftlet farming. *E-Journal of Civil Engineering*, 1(1), 1–7.
- Iijima, R., Takahashi, H., Namme, R., Ikegami, S., & Yamazaki, M. (2004). Novel biological function of sialic acid (N-acetylneuraminic acid) as a hydrogen peroxide scavenger. *FEBS Letters*, 561(1–3), 163–166.
- Ioannides, C., Steele, C. M., & Parke, D. V. (1983). Species variation in the metabolic activation of paracetamol to toxic intermediates: role of cytochromes p-450 and p-448. *Toxicology Letters*, 16, 55–61.

- Jadeja, R. N., Upadhyay, K. K., Devkar, R. V., & Khurana, S. (2016). Naturally occurring Nrf2 activators: Potential in treatment of liver injury. *Oxidative Medicine and Cellular Longevity*, 1- 13.
- Jaeschke, H. (2015). Acetaminophen: Dose-Dependent Drug Hepatotoxicity and Acute Liver Failure in Patients. *Digestive Diseases*, 33(4), 464–471.
- Jaeschke, H., & Bajt, M. L. (2006). Intracellular signaling mechanisms of acetaminophen-induced liver cell death. *Toxicological Sciences*, 89(1), 31–41.
- Jaeschke, H., Williams, C. D., Ramachandran, A., & Bajt, M. L. (2011). Acetaminophen hepatotoxicity and repair: The role of sterile inflammation and innate immunity. *Liver International*, 32(1), 8 - 20
- Jaeschke, H., Xie, Y., & McGill, M. R. (2014). Acetaminophen-induced Liver Injury: from Animal Models to Humans. *Journal of Clinical and Translational Hepatology*, 2, 153–161.
- Jahan, S., Khan, M., Imran, S., & Sair, M. (2015). The hepatoprotective role of Silymarin in isoniazid induced liver damage of rabbits. *Journal of the Pakistan Medical Association*, 65(6), 620–622.
- Jefferies, S., Saxena, M., & Young, P. (2012). Paracetamol in critical illness: A review. *Critical Care and Resuscitation*, 14(1), 74–80.
- Jiang, Y., Fan, X., Wang, Y., Chen, P., Zeng, H., Tan, H., Gonzalez, F. J., Huang, M., & Bi, H. (2015). Schisandrol B protects against acetaminophen-induced hepatotoxicity by inhibition of CYP-mediated bioactivation and regulation of liver regeneration. *Toxicological Sciences: An Official Journal of the Society of Toxicology*, 143(1), 107–115.
- Johnson, M. (2012). Laboratory Mice and Rats. *Materials and Methods*, 2(113), 1 -13.
- Jollow, D. J., Mitchell, J. R., Potter, W. Z., Davis, D. C., Gillette, J. R., & Brodie, B. B. (1973). Acetaminophen-Induced Hepatic Necrosis. II. Role of Covalent Binding In Vivo. *The Journal of Pharmacology and Experimental Therapeutics*, 187(1), 195–202.
- Kabiri, N., Ahangar-Darabi, M., Setorki, M., & Rafieian-Kopaei, M. (2013). The effect of silymarin on liver injury induced by Thioacetamide in rats Hepatoprotection Thioacetamide Silymarin Rat. *Journal of HerbMed Pharmacology*, 2(2), 29–33.
- Kar, G., Chan, L., Chun, Z., Wong, F., Yin, K., Lam, C., Kwan, L., Cheng, W., Zhang, L. M., Lin, H., Dong, T. T., Wah, K., & Tsim, K. (2015). Edible Bird's Nest, an Asian Health Food Supplement, Possesses Skin Lightening Activities: Identification of N-Acetylneuraminic Acid as Active Ingredient. *Journal of Cosmetics, Dermatological Sciences and Applications*, 5, 262–274.
- Kong, D. Y., Park, J. H., Lee, K. W., Park, H., & Cho, J. A. (2016). Comparative Analysis of 3 Experimental Mouse Model for Blood Hematology and Chemistry.

- Kong, Y. C., Keung, W. M., Yip, T. T., Ko, K. M., Tsao, S. W., & Ng, M. H. (1987). Evidence that epidermal growth factor is present in swiftlet's (*Collocalia*) nest. *Comparative Biochemistry and Physiology Part B: Comparative Biochemistry*, 87(2), 221–226.
- Kubben, F. J. G. M., Peeters-Haesevoets, A., Engels, L. G. J. B., Baeten, C. G. M. I., Schutte, B., Arends, J. W., Stockbrugger, R. W., & Blijham, G. H. (1994). Proliferating cell nuclear antigen (PCNA): A new marker to study human colonic cell proliferation. *Journal of Clinical Pathology*, 35, 530–535.
- Lee, D. Y. Y., Kim, M. H. H., Suh, H. R. R., Jung, Y. S. S., Hwang, D. Y. Y., & Kim, K. S. S. (2017). Comparison of the response using ICR mice derived from three different sources to multiple low-dose streptozotocin-induced diabetes mellitus. *Laboratory Animal Research*, 33(2), 150–156.
- Lee, W. M. (2004). Acetaminophen and the U.S. acute liver failure study group: Lowering the risks of hepatic failure. *Hepatology*, 40(1), 6–9.
- Li, N., & Hua, J. (2017). Immune cells in liver regeneration. *Oncotarget*, 8(2), 3628–3639.
- Li, Q., Yuan, Y., Zhang, R., Yang, L., Han, L., Cai, T., and Cai, Y. Edible bird's nest: extraction and pharmacological. *SCIREA Journal of Preclinical Medicine*, 1(1), 1–10.
- Liang, C. M., Tai, M. C., Chang, Y. H., Chen, Y. H., Chen, C. L., Chien, M. W., & Chen, J. T. (2010). Glucosamine inhibits epidermal growth factor-induced proliferation and cell-cycle progression in retinal pigment epithelial cells. *Molecular Vision*, 2010(16), 2559–2571.
- Liu, B. Q., Meng, X., Li, C., Gao, Y. Y., Li, N., Niu, X. F., Guan, Y., & Wang, H. Q. (2011). Glucosamine induces cell death via proteasome inhibition in human ALVA41 prostate cancer cell. *Experimental and Molecular Medicine*, 43(9), 487–493.
- Liu, F., Ichihara, S., Mohideen, S. S., Sai, U., Kitoh, J., & Ichihara, G. (2009). Comparative study on susceptibility to 1-bromopropane in three mice strains. *Toxicological Sciences*, 112(1), 100–110.
- Looi, Q. H., & Omar, A. R. (2016). Swiftlets and Edible Bird's Nest Industry in Asia. *Pertanika Journal of Scholarly Research Reviews*, 2(1), 32–48.
- Lubek, B. M., Avaria, M., Basu, P. K., & Wells, P. G. (1988). Pharmacological studies on the in vivo cataractogenicity of acetaminophen in mice and rabbits. *Toxicological Sciences*, 10(4), 596–606.
- Macherey, A.-C., & Dansette, P. M. (2008). Biotransformations leading to toxic metabolites: Chemical aspect. In *Wermuth's The Practice of Medical Chemistry* (33), 1–9.

- Maga, G., & Hübscher, U. (2003). Proliferating cell nuclear antigen (PCNA): a dancer with many partners. *Diabetes, Stoffwechsel Und Herz*, 116, 3051–3060.
- Mahmoudi, G. A., Astaraki, P., Mohtashami, A. Z., & Ahadi, M. (2015). N-acetylcysteine overdose after acetaminophen poisoning. *International Medical Case Reports Journal*, 8, 65–69.
- Mangipudy, R. S., Chanda, S., & Mehendale, H. M. (1995). Tissue repair response as a function of dose in thioacetamide hepatotoxicity. *Environmental Health Perspectives*, 103(3), 260–267.
- Marcone, M. F. (2005). Characterization of the edible bird's nest the "Caviar of the East." *Food Research International*, 38, 1125–1134.
- Mastellos, D., Papadimitriou, J. C., Franchini, S., Tsonis, P. A., & Lambris, J. D. (2001). A Novel Role of Complement: Mice Deficient in the Fifth Component of Complement (C5) Exhibit Impaired Liver Regeneration. *The Journal of Immunology*, 166(4), 2479–2486.
- Matias, M., Canário, C., Silvestre, S., Falcão, A., & Alves, G. (2014). Cytochrome P450-mediated toxicity of therapeutic drugs. *Cytochrome P450 Enzymes: Biochemistry, Pharmacology and Health Implications*, 13–50.
- McGill, M. R., Sharpe, M. R., Williams, C. D., Taha, M., Curry, S. C., & Jaeschke, H. (2012). The mechanism underlying acetaminophen-induced hepatotoxicity in humans and mice involves mitoch. *The Journal of Clinical Investigation*, 122(4), 1574–1583.
- McGill, M. R., Williams, C. D., Xie, Y., Ramachandran, A., & Jaeschke, H. (2012). Acetaminophen-induced liver injury in rats and mice: comparison of protein adducts, mitochondrial dysfunction, and oxidative stress in the mechanism of toxicity. *Toxicology and Applied Pharmacology*, 264(3), 387–394.
- Mehendale, H. M. (2005). Tissue Repair: AN Important Determinant of Final Outcome of Toxicant-Induced Injury. *Toxicologic Pathology*, 33(1), 41–51.
- Metushi, I. G., & Uetrecht, J. (2014). Isoniazid-induced liver injury and immune response in mice. *Journal of Immunotoxicology*, 11(4), 383–392.
- Michalopoulos, G. K. (2007). Liver regeneration. *Journal of Cell Physiology*, 213(2), 286–300.
- Michalopoulos, G. K. (2010). Liver regeneration after partial hepatectomy: Critical analysis of mechanistic dilemmas. *American Journal of Pathology*, 176(1), 2–13.
- Mirochnitchenko, O., Weisbrot-Lefkowitz, M., Reuhl, K., Chen, L., Yang, C., & Inouye, M. (1999). Acetaminophen toxicity: Opposite effects of two forms of glutathione peroxidase. *Journal of Emergency Nursing: JEN: Official Publication of the Emergency Department Nurses Association*, 274(15), 10349–10355.
- Mohamad Shukri, N. N. H., Mohd Nawi, N., Abdullah, A. M., & Man, N. (2018). Consumer's perception on the quality of controversial contents in edible bird's nest products. *Pertanika Journal of Scholarly Research Reviews*, 4(1), 1–9.

- Moreno-Marín, N., Barrasa, E., Morales-Hernández, A., Paniagua, B., Blanco-Fernández, G., Merino, J. M., & Fernández-Salguero, P. M. (2017). Dioxin Receptor Adjusts Liver Regeneration After Acute Toxic Injury and Protects Against Liver Carcinogenesis. *Scientific Reports*, 7(1), 1–12.
- Muhammad-Azam, F., Nur-Fazila, S. H., Ain-Fatin, R., Noordin, M. M., & Yimer, N. (2019). Histopathological changes of acetaminophen-induced liver injury and subsequent liver regeneration in BALB/C and ICR mice. *Veterinary World*, 12(11), 1682–1688.
- Nassar, I., Pasupati, T., Judson, J. P., & Segarra, I. (2010). Histopathological study of the hepatic and renal toxicity associated with the co-administration of imatinib and acetaminophen in a preclinical mouse model. *Malaysian Journal of Pathology*, 32(1), 1–11.
- Newsome, P. N., Plevris, J. N., Nelson, L. J., & Hayes, P. C. (2000). Animal models of fulminant hepatic failure: a critical evaluation. *Liver Transplantation*, 6(1), 21–31.
- Ng, M., Chan, K.-H., & Kong, Y. C. (1986). Potentiation of mitogenic response by extracts of the swiftlet's (Collocalia) nest. *Biochemistry International*, 13, 521–531.
- Norhayati, M., Azman, O., & Wan Nazaimoon, W. M. (2010). Preliminary Study of the Nutritional Content of Malaysian Edible Bird's Nest. In *Malaysian Journal of Nutrition*, 16(3), 389 - 396.
- Ogata, K., Kurki, P., Celis, J. E., Nakamura, R. M., & Tan, E. M. (1987). Monoclonal antibodies to a nuclear protein (PCNA/cyclin) associated with DNA replication. *Experimental Cell Research*, 168(2), 475–486.
- Oghenesuvwe, E. E., Nwoke, E. E., & Lotanna, D. A. (2014). Guidelines on dosage calculation and stock solution preparation in experimental animals' studies. *Journal of Natural Sciences Research*, 37(8), 831–838.
- Ostapowicz, G., Fontana, R. J., Schioødt, F. V., Larson, A., Davern, T. J., Han, S. H. B., McCashland, T. M., Shakil, A. O., Hay, J. E., Hynan, L., Crippin, J. S., Blei, A. T., Samuel, G., Reisch, J., Lee, W. M. (2002). Results of a prospective study of acute liver failure at 17 tertiary care centers in the United States. *Annals of Internal Medicine*, 2002(137), 947 - 954
- Otto, G. P., Rathkolb, B., Oestereicher, M. A., Lengger, C. J., Moerth, C., Micklich, K., Fuchs, H., Gailus-Durner, V., Wolf, E., & de Angelis, M. H. (2016). Clinical Chemistry Reference Intervals for C57BL/6J, C57BL/6N, and C3HeB/FeJ Mice (*Mus musculus*). *Journal of the American Association for Laboratory Animal Science*, 55(4), 375–386.
- Papackova, Z., Heczko, M., Dankova, H., Sticova, E., Lodererova, A., Bartonova, L., Poruba, M., & Cahova, M. (2018). Silymarin prevents acetaminophen-induced hepatotoxicity in mice. *PLoS ONE*, 13(1), 1–20.

- Peng, L., & Zhong, X. (2015). Epigenetic regulation of drug metabolism and transport. *Acta Pharmaceutica Sinica B*, 5(2), 106–112.
- Phillipps, Q., & Phillipps, K. (2011). *Phillipps' Field Guide To The Birds Of Borneo Sabah, Sarawak, Brunei & Kalimantan*. England: John Beaufoy Publishing.
- Rahman, T. M., & Hodgson, H. J. F. (2000). Animal models of acute hepatic failure. *International Journal of Experimental Pathology*, 81(2), 145–157.
- Ramachandran, A., & Jaeschke, H. (2017). Mechanisms of acetaminophen hepatotoxicity and their translation to the human pathophysiology. *Journal of Clinical and Translational Research*, 3(1), 157–169.
- Rivera, P., Pastor, A., Arrabal, S., Decara, J., Vargas, A., Sánchez-Marín, L., Pavón, F. J., Serrano, A., Bautista, D., Boronat, A., de la Torre, R., Baixeras, E., Lucena, M. I., de Fonseca, F. R., & Suárez, J. (2017). Acetaminophen-induced liver injury alters the Acyl ethanolamine-based anti-inflammatory signaling system in liver. *Frontiers in Pharmacology*, 8(705), 1–16.
- Roh, K. B., Lee, J., Kim, Y. S., Park, J., Kim, J. H., Lee, J., & Park, D. (2012). Mechanisms of edible bird's nest extract-induced proliferation of human adipose-derived stem cells. *Evidence-Based Complementary and Alternative Medicine*, 1 - 11.
- Ruegg, M. A., & Meinen, S. (2012). Histopathology in Hematoxylin & Eosin stained muscle sections. *TREAT-NMD Neuromuscular Network*, Id, 1–9.
- Ruepp, S. U., Tonge, R. P., Shaw, J., Wallis, N., & Pognan, F. (2002). Genomics and proteomics analysis of acetaminophen toxicity in mouse liver. *Toxicological Sciences*, 65, 135–150.
- Sabaté, M., Ibáñez, L., Pérez, E., Vidal, X., Buti, M., Xiol, X., Mas, A., Guarner, C., Forné, M., Solà, R., Castellote, J., Rigau, J., & Laporte, J. R. (2011). Paracetamol in therapeutic dosages and acute liver injury: Causality assessment in a prospective case series. *BMC Gastroenterology*, 11(8), 1 - 7.
- Saito, C., Zwingmann, C., & Jaeschke, H. (2010). Novel mechanisms of protection against acetaminophen hepatotoxicity in mice by glutathione and N-acetylcysteine. *Hepatology*, 51(1), 246–254.
- Saleem, T. H., Abo El-Maali, N., Hassan, M. H., Mohamed, N. A., Mostafa, N. A. M., Abdel-Kahaar, E., & Tammam, A. S. (2018). Comparative Protective Effects of N-Acetylcysteine, N-Acetyl Methionine, and N-Acetyl Glucosamine against Paracetamol and Phenacetin Therapeutic Doses-Induced Hepatotoxicity in Rats. *International Journal of Hepatology*, 1 - 8.
- Saljoughian, M. (2016). Acetaminophen intoxication: A critical-care emergency. *U.S. Pharmacist*, 41(12), 38–41.
- Salomone, F., Godos, J., & Zelber-Sagi, S. (2016). Natural antioxidants for non-alcoholic



- fatty liver disease: Molecular targets and clinical perspectives. *Liver International*, 36(1), 5–20.
- Scheller, F. W., Yarmana, A., Wollenberger, U., Matias, M., Canário, C., Silvestre, S., Falcão, A., Alves, G., & Wang, X. (2014). Cytochrome P450 Enzymes: Biochemistry, Pharmacology and Health Implications. In J. Wu (Ed.) *Biochemistry Research Trends* (pp 16 - 24). New York: Nova Publisher
- Schilling, A., Corey, R., Leonard, M., & Eghtesad, B. (2010). Acetaminophen: Old drug, new warnings. *Cleveland Clinic Journal of Medicine*, 77(1), 19–27.
- Shahid, M., & Subhan, F. (2014). Comparative histopathology of acetaminophen induced hepatotoxicity in animal models of mice and rats. *Pharmacologyonline*, 3, 32–43.
- Sheen, C. L., Dillon, J. F., Bateman, D. N., Simpson, K. J., & Macdonald, T. M. (2002). Paracetamol Toxicity: Epidemiology, Prevention and Costs to the Health-Care System. *Quarterly Journal of Medicine*, 95, 609–619.
- Siddiqui, E. A., Jagdale, P., Ahire, K., Jadhav, S., Khan, S. A., Bhosle, S., Pal, A., Jamdagni, P., & Chaudhari, B. (2016). Relevance of small laboratory animals as models in translational research: Challenges and road ahead. *Journal of Applied Pharmaceutical Science*, 6(5), 198–209.
- Sreejith, G. S., Ittop, J. A., Anitha, M. G., & Ravishankar, B. (2015). Hepatoprotective Activity of Dushivishari Agada in Paracetamol Induced Hepatotoxicity of Wistar Rats. *International Journal of Research in Ayurveda & Pharmacy*, 6(5), 602–608.
- Stoner, H. B. (1955). The mechanism of toxic hepatic necrosis. *British Journal of Experimental Pathology*, 1953, 37(2), 176–198.
- Suckow, M. A., Danneman, P., & Brayton, C. (2001). The Laboratory Mouse. *Introduction* (pp. 1). Florida: CRC Press LLC.
- Tan, C. J. Y., & Sklar, G. E. (2017). Characterisation and outcomes of adult patients with paracetamol overdose presenting to a tertiary hospital in Singapore. *Singapore Medical Journal*, 58(12), 695–702.
- Tanaka, M., & Miyajima, A. (2016). Liver regeneration and fibrosis after inflammation. *Inflammation and Regeneration*, 36(19), 1–6.
- Tittarelli, R., Pellegrini, M., Scarpellini, M. G., Marinellix, E., Bruti, V., Di Luca, N. M., Busardò, F. P., & Zaami, S. (2017). Hepatotoxicity of paracetamol and related fatalities. *European Review for Medical and Pharmacological Sciences*, 21(1), 95–101.
- Tuer, A., Tokarz, D., Prent, N., Cisek, R., Alami, J., Dumont, D. J., Bakueva, L., Rowlands, J., & Barzda, V. (2010). Nonlinear multicontrast microscopy of hematoxylin-and-eosin-stained histological sections. *Journal of Biomedical Optics*, 15(2), 1 - 9.
- Tung, B. T., Hai, N. T., & Son, P. K. (2017). Hepatoprotective effect of Phytosome









- Curcumin against paracetamol-induced liver toxicity in mice. *Brazilian Journal of Pharmaceutical Sciences*, 53(1), 1–13.
- Twycross, R., Pace, V., Mihalyo, M., & Wilcock, A. (2013). Acetaminophen (Paracetamol). *Journal of Pain and Symptom Management*, 46(5), 747–755.
- Uchida, N. S., Silva-Filho, S. E., Cardia, G. F. E., Cremer, E., De Souza Silva-Comar, F. M., Silva, E. L., Bersani-Amado, C. A., & Cuman, R. K. N. (2017). Hepatoprotective Effect of Citral on Acetaminophen-Induced Liver Toxicity in Mice. *Evidence-Based Complementary and Alternative Medicine*, 1 - 9.
- Uhl, E. W., & Warner, N. J. (2015). Mouse Models as Predictors of Human Responses: Evolutionary Medicine. *Current Pathobiology Reports*, 3, 219–223.
- Vaickus, L. J., Bouchard, J., Kim, J., Natarajan, S., & Remick, D. G. (2010). Inbred and outbred mice have equivalent variability in a cockroach allergen-induced model of asthma. *Comparative Medicine*, 60(6), 420–426.
- Vandamme, T. (2014). Use of rodents as models of human diseases. *Journal of Pharmacy & Bioallied Sciences*, 6, 2–9.
- Vimala, B., Hussain, H., & Wan Nazaimoon, W. M. (2012). Effects of edible bird's nest on tumour necrosis factor-alpha secretion, nitric oxide production and cell viability of lipopolysaccharide-stimulated RAW 264.7 macrophages. *Food and Agricultural Immunology*, 23(4), 303 - 314.
- Wan Syahidah, H., Normah, M., & Albert, T. (2014). Protein content and amino acid composition of farmed edible bird's nests in Malaysia. *Malaysian Journal of Veterinary Research*, 5(1), 342–346.
- Wang, B., & Brand-Miller, J. (2003). The role and potential of sialic acid in human nutrition. *European Journal of Clinical Nutrition*, 57(11), 1351–1369.
- Wang, P., Pradhan, K., Zhong, X. bo, & Ma, X. (2016). Isoniazid metabolism and hepatotoxicity. *Acta Pharmaceutica Sinica B*, 6(5), 384–392.
- Williams, C. D., Koerner, M. R., Lampe, J. N., Farhood, A., & Jaeschke, H. (2011). Mouse strain-dependent caspase activation during acetaminophen hepatotoxicity does not result in apoptosis or modulation of inflammation. *Toxicology and Applied Pharmacology*, 257(3), 449–458.
- Woolbright, B. L., & Jaeschke, H. (2017). Review Role of the inflammasome in acetaminophen-induced liver injury and acute liver failure. *Journal of Hepatology*, 66(4), 836–848.
- Xie, Y., Zeng, H., Huang, Z., Xu, H., Fan, Q., Zhang, Y., & Zheng, B. (2018). Effect of maternal administration of edible bird's nest on the Learning and memory abilities of suckling offspring in mice. *Neural Plasticity*, 1 - 13.
- Yan, M., Huo, Y., Yin, S., & Hu, H. (2018a). Mechanisms of acetaminophen-induced liver injury and its implications for therapeutic interventions. *Redox Biology*, 17,

- Yew, M. Y., Koh, R. Y., Chye, S. M., Othman, I., & Ng, K. Y. (2014). Edible bird's nest ameliorates oxidative stress-induced apoptosis in SH-SY5Y human neuroblastoma cells. *BMC Complementary and Alternative Medicine*, *14*(391), 1 - 12.
- Yida, Z., Imam, M. U., Ismail, M., Hou, Z., Abdullah, M. A., Ideris, A., & Ismail, N. (2015). Edible Bird's Nest attenuates high fat diet-induced oxidative stress and inflammation via regulation of hepatic antioxidant and inflammatory genes. *BMC Complementary and Alternative Medicine*, *15*(310), 1 - 7.
- Yida, Z., Imam, M. U., Ismail, M., Ooi, D. J., Sarega, N., Azmi, N. H., Ismail, N., Chan, K. W., Hou, Z., & Yusuf, N. B. (2015). Edible bird's nest prevents high fat diet-induced insulin resistance in rats. *Journal of Diabetes Research*, *1- 11*.
- Yida, Z., Imam, U. U., & Ismail, M. (2014). In vitro bioaccessibility and antioxidant properties of edible bird's nest following simulated human gastro-intestinal digestion. *BMC Complementary and Alternative Medicine*, *14*(486), 1–7.
- Yoon, E., Babar, A., Choudhary, M., Kutner, M., & Pysopoulos, N. (2016). Acetaminophen-Induced Hepatotoxicity: a Comprehensive Update. *Journal of Clinical and Translational Hepatology*, *4*(2), 131–142.
- Zainal Abidin, F., Hui, C. K., Luan, N. S., Mohd Ramli, E. S., Hun, L. T., & Abd Ghafar, N. (2011). Effects of edible bird's nest (EBN) on cultured rabbit corneal keratocytes. *BMC Complementary and Alternative Medicine*, *11*(94), 1 - 10.
- Zanger, U. M., & Schwab, M. (2013). Cytochrome P450 enzymes in drug metabolism: Regulation of gene expression, enzyme activities, and impact of genetic variation. *Pharmacology and Therapeutics* *138*, 103 - 141.
- Zhiping, H., Imam, M. U., Ismail, M., Ismail, N., Yida, Z., Ideris, A., Sarega, N., & Mahmud, R. (2015). Effects of edible bird's nest on hippocampal and cortical neurodegeneration in ovariectomized rats. *Food and Function*, *2015*(6), 1701–1711.
- Zoubair, B., Fatima Azzahra, L., Fouzia, H., Mohammed, L., Brahim, B., & Nouredine, B. (2013). Evaluation of acetaminophen effect on oxidative stressed mice by peroxide hydrogen. *American Journal of Biomedical Research*, *1*(4), 75–79.

## APPENDICES

### Appendix A

#### Approval letter of ethical committee (IACUC), UPM



PEJABAT TIMBALAN NAIB CANCELOR (PENYELIDIKAN DAN INOVASI)  
OFFICE OF THE DEPUTY VICE CHANCELLOR (RESEARCH AND INNOVATION)

**INSTITUTIONAL ANIMAL CARE AND USE COMMITTEE**

Date: 10<sup>th</sup> November 2017

AUP No.: UPM/IACUC/AUP-R078/2017

Project Title: The Hepatoprotective Effect of Edible Bird Nest (EBN) After Acetaminophen-Induced Hepatotoxicity In ICR Mice

Principal Investigator: Dr. Nur Fazila Saulol Hamid

Members: Prof. Dr. Noordin Mohamed Mustapha; Muhammad Azam Bin Fazli

Attending Veterinarian: Dr. Nur Fazila Saulol Hamid

Committee Decision: The committee has reviewed and approved the proposed animal utilisation protocol, subject to relevant permit and/ or owner's consent.

Project Classification: Acute

Category of Invasiveness: D


Source of Animals: UPM Animal Research Unit, Faculty of Veterinary Medicine, Universiti Putra Malaysia

Number of Animals Approved: 78 Mice

Housing: Animal Research Facility, Faculty of Veterinary Medicine, Universiti Putra Malaysia

Duration: 10 November 2017 – 10 November 2018

Ethical approval is required in the case of amendments to the approved animal utilisation protocol (AUP). Please apply using Form 105. Kindly submit a final/annual report (Form 106) upon study completion, or before expiry of approval.

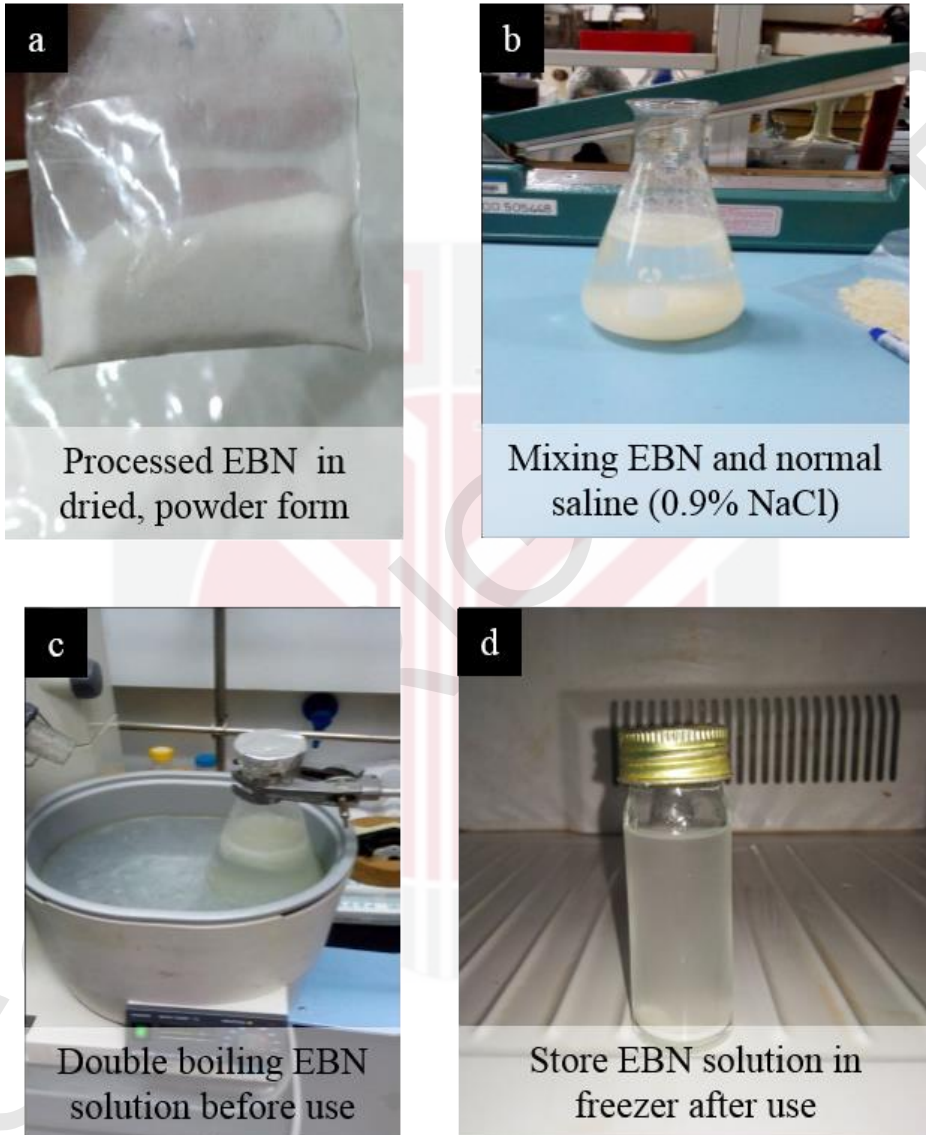
  
**PROF. DR. MOHD HAIR BEJO**  
Chairman  
Institutional Animal Care and Use Committee  
Universiti Putra Malaysia

✉ Pejabat Timbalan Naib Canselor (Penyelidikan dan Inovasi), Universiti Putra Malaysia, 43400 UPM Serdang, Selangor Darul Ehsan, Malaysia  
Pejabat Timbalan Naib Canselor (P&I) ☎ 603-8947 1002 📠 603-8945 1646, Pejabat Pentadbiran TNCPi ☎ 603-8947 1608 📠 603-8945 1673,  
Pejabat Pengarah, Pusat Pengurusan Penyelidikan (RMC) ☎ 603-8947 1601 📠 603-8945 1536, Pejabat Pengarah, Putra Science Park (PSP)  
☎ 603-8947 1291 📠 603-8946 4121 🌐 <http://www.tncpi.upm.edu.my>

Figure 24. Institutional of Animal Care and Use Committee (IACUC) approval for the animal utilisation protocol in this study.

## Appendix B

### EBN Preparation Process



**Figure 25.** EBN preparation process for the prophylactic study of EBN consumption followed by AILI in ICR mice. (a) EBN was bought in powder form after being processed. (b) EBN powder was diluted with normal saline (0.9% NaCl) to obtain doses of 60, 120, and 250 mg/kg. (c) The mixed solution was double-boiled and let to cool at room temperature. (d) EBN solution was stored in a chiller of 4 °C and will be heated to 40 °C before it was instilled in the mouse using oral gavage.

## Appendix C

### Summary of histopathological description of the liver section

**Table 2. Histopathological description of the liver section in each group after 5 hours of APAP induction. The histological features were summarised as a group according to the AILI mean scores calculated for each group.**

Group	Score range (Mean $\pm$ SD)	Description
Control	0 (0)	Normal hepatocellular features with no histological abnormalities
APAP	0-0.5 (0.15 $\pm$ 0.22)	Early hydropic degeneration with variably sized cytoplasmic vacuoles and showed minimal necrosis
Silymarin	0-0.25 (0.15 $\pm$ 0.14)	Minimal to none centrilobular necrosis with variably size vacuoles in the centrilobular area
EBN 60	0.5-1 (0.75 $\pm$ 0.25)	Minimal hepatocellular necrosis and early hydropic degeneration in the centrilobular area with few mitotic figures were seen
EBN 120	0-0.25 (0.05 $\pm$ 0.11)	Scattered individual necrotic cells in the centrilobular area with variable size vacuolation and infiltration of inflammatory cells
EBN 250	1-3.5 (2.67 $\pm$ 1.44)	Severe centrilobular necrosis and intense haemorrhage. Macrovascular steatosis was evident by large fat droplets

**Table 3. Description of the histological finding of each group after 10 hours post APAP dosing (hpd). The histological descriptions were done based on the AILI mean scores in each group represented by a total of 5 animals per group.**

Group	Range score (Mean $\pm$ S.D)	Description
APAP	3.5-4 (3.75 $\pm$ 0.29)	Extensive centrilobular necrosis and haemorrhage that spread out to almost $\frac{3}{4}$ of liver sections
Silymarin	0 (0)	Completely intact liver cells with no histological abnormalities
EBN 60	0-0.5 (0.25 $\pm$ 0.25)	An extensive area of normal hepatocytes compared to individual necrotic cells with few presences of mitotic figures
EBN 120	0.25-1.5 (0.6 $\pm$ 0.52)	Normal hepatocytes cover a broad area of liver sections and some scattered individual necrosis particularly the centrilobular area
EBN 250	0-2.5 (1.33 $\pm$ 1.26)	Moderate to severe necrosis and haemorrhage at the centrilobular area

**Table 4. The histological findings of APAP-induced ICR mice were described in each treatment group based on the AILI mean scores by assessing the hepatocytes changes for individual animals in the groups at 24 hpd.**

Group	Range score (Mean $\pm$ S.D)	Description
APAP	0.5 (0.5 $\pm$ 0)	Minimal cell damage and hydropic degeneration at the centrilobular area of the liver in most of the individual animals in the group, also some degree of mitotic figures
Silymarin	0-0.1 (0.07 $\pm$ 0.06)	Normal hepatocytes present with no histological changes and a very minimal degree of hydropic
EBN 60	0-0.25 (0.06 $\pm$ 0.13)	The liver section covered mostly with the normal architecture of liver cells and a very minimal degree of hydropic degeneration
EBN 120	0 (0)	Normal hepatocellular features with no histological abnormalities are observed
EBN 250	0-3.5 (1.17 $\pm$ 1.75)	Continuous variable hepatocytes necrosis at the centrilobular area with extensive haemorrhage and mild macrovascular steatosis

## BIODATA OF STUDENT

Muhammad Azam Bin Fazil is a post-graduate student in the Faculty of Veterinary Medicine, Universiti Putra Malaysia, Serdang campus under the Department of Veterinary Pathology and Microbiology. His Master's study focuses on the research project of the histopathological assessment on acetaminophen-induced liver injury with subsequent regeneration and its response to the edible bird's nest extract in a mouse model. During the post-graduate candidature, an article research paper was published at Veterinary World Journal (Vet World) and another had been submitted which still in the process to be published. This student had participated in a poster presentation at the 10<sup>th</sup> Malaysian Association of Veterinary Pathology (MAVP) Scientific Conference at Hotel Casuarina @ Meru, Ipoh, Perak in 2018.

Before the post-graduate study, Muhammad Azam had been graduated from Universiti Malaysia Sarawak (UNIMAS) in Kota Samarahan, Sarawak. He enrolled in the Animal Science and Resources Management course from 2013 to 2016. During the final year project, he worked on a project entitled the effect of preservation and fixation process in bone and cartilage staining of juvenile tilapia specimens. After completing the study period, Muhammad Azam attended an internship at World Wide Fund for Nature (WWF) in the Turtle Conservation project at Kerteh, Terengganu for 3 months period and due in September 2016.



## LIST OF PUBLICATIONS

### Publications

Muhammad-Azam, F., Nur-Fazila, S. H., Ain-Fatin, R., Noordin, M.M., & Yimer, N. (2019). Histopathological changes of acetaminophen-induced liver injury and subsequent liver regeneration in BALB/C and ICR mice. *Veterinary World*, 12(11), 1682–1688.

Ain-Fatin R., Nur-Fazila S.H., Nur-Mahiza M.I., Yasmin A.R., Muhammad-Azam F. & Nur-Kuain H. (2020). Incidental findings of *Heterakis spumosa* and *Chirodiscoides caviae* with pinworms in Sprague Dawley rats. *Sains Malaysiana*, 49(5), 1097-1106.

Muhammad-Azam F., Nur-Fazila S.H., Ain-Fatin R., Noordin M.M., Yasmin A.R. & Yimer N. Prophylactic effect of edible bird's nest on acetaminophen-induced liver injury response in mice model. *Accepted in Pakistan Journal of Pharmaceutical Sciences*.

### Proceeding

Poster presentation on “The effects of acetaminophen-induced liver injury in ICR mice” at 10<sup>th</sup> Malaysian Association of Veterinary Pathology (MAVP) Scientific Conference 2018.



**UNIVERSITI PUTRA MALAYSIA**

**STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT**

**ACADEMIC SESSION : Second Semester 2020/2021**

**TITLE OF THESIS / PROJECT REPORT :**

HISTOPATHOLOGICAL ASSESSMENT OF ACETAMINOPHEN-INDUCED LIVER INJURY  
AND REGENERATION RESPONSE FOLLOWING EDIBLE BIRD'S NEST ADMINISTRATION  
IN ICR AND BALB/C MICE MODEL

**NAME OF STUDENT: MUHAMMAD AZAM BIN FAZIL**

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

1. This thesis/project report is the property of Universiti Putra Malaysia.
2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as :

\*Please tick (v )

**CONFIDENTIAL**

(Contain confidential information under Official Secret Act 1972).

**RESTRICTED**

(Contains restricted information as specified by the organization/institution where research was done).

**OPEN ACCESS**

I agree that my thesis/project report to be published as hard copy or online open access.

This thesis is submitted for :

**PATENT**

Embargo from \_\_\_\_\_ until \_\_\_\_\_  
(date) (date)

**Approved by:**

\_\_\_\_\_  
(Signature of Student)  
New IC No/ Passport No.:

Date :

\_\_\_\_\_  
(Signature of Chairman of Supervisory Committee)  
Name:

Date :

**[Note : If the thesis is CONFIDENTIAL or RESTRICTED, please attach with the letter from the organization/institution with period and reasons for confidentiality or restricted. ]**