

# **UNIVERSITI PUTRA MALAYSIA**

TRANSFORMATION OF OIL PALM MESOCARP FIBER TO CONVERTIBLE MATERIALS IN WATER AT SUBCRITICAL STATE

SANGGITHAPRIYA A/P MAHANDRAN

FK 2021 32



## TRANSFORMATION OF OIL PALM MESOCARP FIBER TO CONVERTIBLE MATERIALS IN WATER AT SUBCRITICAL STATE



# SANGGITHAPRIYA A/P MAHANDRAN

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirement for the Degree of Master of Sciences

November 2020

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia

Copyright © Universiti Putra Malaysia

 $\mathbf{G}$ 



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Sciences

#### TRANSFORMATION OF OIL PALM MESOCARP FIBER TO CONVERTIBLE MATERIALS IN WATER AT SUBCRITICAL STATE

By

#### SANGGITHAPRIYA A/P MAHANDRAN

November 2020

Chair Faculty : Shamsul Izhar bin Siajam, PhD, Ir : Engineering

In this research, water at subcritical state was studied as an alternative way to breakdown oil palm mesocarp fibre (OPMF) by analysing their structural and compositional changes at different reaction temperature. High ion products of water in subcritical conditions and the dielectric constant of water at temperature above 200°C makes water to behave as acid catalyst and environmentally friendly solvent. The decomposition of OPMF using subcritical water (Sub CW) method has been carried out at reaction temperature ranging from 200°C to 300°C and with constant reaction time of 5 min using a batch stainlesssteel tube as reactor. The highest oil yield was obtained at 240°C with result values 219.6 mg g<sup>-1</sup> dry OPMF due to the behaviour of water to act like ethanol solvent at 240°C ( $\varepsilon$ = 27.0). High-performance liquid chromatography (HPLC) analysis identified that the highest number of monosaccharides such as glucose and fructose presented at 240°C was due to optimum condition for hydrolysis process with sugar yield, 22.17mg g<sup>-1</sup> dry OPMF. Moreover, morphological and particle size analyses proven that water at Sub critical state ease hydrolysis process occur as the Sub CW temperature increase to 240°C. When the reaction temperature gets higher than 240°C, cellulose of OPMF degrades through pyrolysis process. Increase in Sub CW reaction temperature decreases yield of solid from 0.286 g g<sup>-1</sup> dry OPMF at 200°C to 0.037 g g<sup>-1</sup> dry OPMF at 300°C. Thus, Sub CW at 240°C was able to extract optimum oil and sugar at high yield and reduces solid residue from mesocarp fibre with the absence of any chemical solvent.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

#### TRANSFORMASI SERAT MESOCARP OIL PALM (OPMF) KEPADA BAHAN BOLEH UBAH MENGGUNAKAN AIR DALAM KEADAAN SUB-KRITIKAL

Oleh

#### SANGGITHAPRIYA A/P MAHANDRAN

November 2020

Pengerusi Fakulti : Shamsul Izhar bin Siajam, PhD, Ir : Kejuruteraan

Dalam penyelidikan ini, air pada keadaan sub-kritical dikaji sebagai kaedah alternatif untuk memecahkan serat mesocarp kelapa sawit (OPMF) dengan menganalisis perubahan struktur dan komposisi OPMF pada suhu tindak balas yang berbeza. Produk ion tinggi air dalam keadaan sub-kritical dan pemalar dielektrik air pada suhu melebihi 200°C adalah serupa dengan pemalar dielektrik metanol pada suhu bilik, yang menjadikan air berperilaku sebagai pemangkin asid dan pelarut mesra alam. Penguraian OPMF di bawah menggunakan kaedah air sub-kritical telah dilakukan pada suhu tindak balas antara 200°C hingga 300°C dan dengan masa tindak balas berterusan 5 min menggunakan tiub keluli tahan karat kelompok sebagai reaktor. Hasil minyak tertinggi diperoleh pada suhu tindak balas 240°C (selama 5 minit) dengan nilai hasil 219.6 mg g-1 OPMF kering kerana tingkah laku air bertindak seperti pelarut etanol pada  $240^{\circ}$ C ( $\varepsilon =$ 27.0). Analisis HPLC mengenal pasti bahawa jumlah monosakarida tertinggi seperti glukosa dan fruktosa ditunjukkan pada suhu tindak balas 240°C kerana keadaan optimum untuk proses hidrolisis dengan hasil gula 22.17 mg g-1 kering OPMF. Lebih-lebih lagi, analisis morfologi dan ukuran zarah membuktikan bahawa air pada keadaan sub-kritical memudahkan proses hidrolisis berlaku ketika suhu reaksi meningkat hingga 240°C dan suhu reaksi lebih tinggi daripada 240°C, hasil selulosa OPMF berkurang melalui proses pirolisis. Peningkatan suhu tindak balas sub-kritical menurunkan hasil pepejal dari 0.286 g g-1 OPMF kering pada 200°C hingga 0.037 g g-1 OPMF kering pada 300°C. Oleh itu, air sub-kritical pada suhu reaksi 240°C dapat mengekstrak hasil minyak dan gula yang optimum dan mengurangkan sisa pepejal dari serat mesocarp dengan tidak menggunakan pelarut kimia.



#### ACKNOWLEDGEMENTS

I would like to thank all the people who have been concern in accomplishment of this research work successfully. First and foremost, I would like to express my gratitude to my supervisor Ir. Dr. Shamsul Izhar bin Siajam from Department of Chemical and Environmental Engineering, Universiti Putra Malaysia for guiding me from the beginning until the end of research work completion. It was great honour to complete this master thesis under his supervision. Greatly appreciate his skilful guidance with innovative ideas and provide me needed information during research work.

Besides that, I would like to thank my co-supervisor, Dr. Nordin Bin Hj. Sabli, my supportive research lab mate, Raja Munawarrah binti Raja Zazalli who had always helped me with their suggestions and guidance in completion of this research work successfully.

This acknowledgement would not complete without mentioning my parents Mr. Mahandran Subrayan and Mrs. Puvaneswary Narayanan for their guidance, support and motivation during my research work. Thank you for your support, encouragement and determination in helping me deal with the difficult moments of depressions.

Finally, I would like to express my gratitude to those who directly or indirectly helps me by providing their valuable assistance and support through this research work.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Sciences. The members of the Supervisory Committee were as follows:

#### Shamsul Izhar bin Siajam, PhD

Associate Professor, Ir Faculty of Engineering Universiti Putra Malaysia (Chairman)

Nordin Bin Hj. Sabli, PhD Senior Lecturer Faculty of Engineering Universiti Putra Malaysia (Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date: 11 March 2021

#### **Declaration by graduate student**

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_

Date:

Name and Matric No.: Sanggithapriva A/P Mahandran, GS52357

#### **Declaration by Members of Supervisory Committee**

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: Name of chairman supervisory comm	of ttee: Shamsul	Izhar bin Siaiam
Signature: Name of member of supervisory comm	f ttee: Nordin H	3in Hi, Sabli
supervisory comm		

# TABLE OF CONTENTS

			Page
ABSTRACT ABSTRAK ACKNOWL APPROVAL DECLARAT LIST OF FIG LIST OF TA LIST OF AB	EDGE TION GURE BLES BREV	EMENTS S TATIONS	i iii iv vi x xiiii xv
CHAPTER			
1	INTI	RODUCTION	
	1.1	Research Background	1
	1.2	Problem Statement	2
	1.3	Research Objective	3
	1.4	Scopes of study	3
2	LITI	CRATURE REVIEW	
	2.1	Oil Palm in Malaysia	5
		2.1.1 Palm oil production	6
		2.1.2 Type, Structure and Use of Palm Oil	8
	2.2	Oil Palm Mesocarp Fibre (OPMF)	9
	2.2	2.2.1 Composition of Lignocellulosic Biomass	9
	23	Oil Extraction Method	11
	2.3	Hydrolysis	12
	2.1	Pyrolysis	12
	2.5	Degradation of Lignocellulosic Biomass	13
	2.0	2.6.1 Cellulose Degradation	15
		2.6.2 Alternative Cell Disruption Method	16
	27	Sub Critical Water (Sub CW)	16
		2.7.1 High Ion Production at Elevated Temperature	18
		2.7.2 Low Relative Dielectric Constant	18
3	MET	HODOLOGY	
U	31	Overview of Experiment	20
	3.2	Material and Sample Preparation	20
	33	Moisture content	21
	3.4	Subcritical Fluid Extraction System	22
	3.5	Phases recovery process	23
	5.5	3.5.1 Recovery of Oil Layer Using Hexane	23
		3.5.2 Recovery of Aqueous Phase Using Illtrapure	2 <del>4</del> 24
		water	27
		3.5.3 Recover Solid Phase Content Using Acetone	25
	3.6	Oil Phase Analysis	25
	3.7	Aqueous Phase Analysis	25

C

		3.7.1 Measurement of Total Sugar	26
	3.8	Solid Phase Analysis	26
	3.9	Morphological Analysis	27
		3.9.1 Microscope View	27
		3.9.2 Scanning Electron Microscopy (SEM)	27
	3.10	Chemical compositional Analysis	28
		3.10.1 High-performance liquid chromatography (HPLC)	28
		3.10.2 Fourier transform infra-red (FTIR)	28
	3.11	Particle size analysis (PSA)	28
4	RESU	ULTS AND DISCUSSION	
	4.1	Moisture Content	29
	4.2	Sub CW extraction	30
	4.3	Oil Phase Analysis	30
	4.4	Aqueous Phase Analysis	32
	4.5	Solid Phase Analysis	33
		4.5.1 Yield of Solid Residue	33
		4.5.2 Yield of Tar	35
	4.6	Morphological Analysis	36
		4.6.1 Light Microscopic Analysis	36
		4.6.2 Scanning Electron Microscopic (SEM)	38
		Analysis	•
	4.7	Chemical Compositional Analysis	39
		4.7.1 High-performance liquid chromatography (HPLC, Sugar) analysis	39
		4.7.2 Fourier Transform Infra-Red (FTIR) Analysis	41
		of treated OPMF	
	4. <mark>8</mark>	Particle size of Treated OPMF	43
	4.9	Effect of Sub CW Reaction Temperature Towards Its	44
		Product	
5	CON	CLUSION AND RECOMMENDATION	
	5.1	Conclusion	50
	5.2	Recommendation	51
REFERF	INCES		52
APPEND	ICES		61
BIODAT	A OF STI	JDENT	71
LIST OF	PUBLICA	ATION	72

 $\bigcirc$ 

# LIST OF FIGURES

Figure		Page
2.1	Oil Palm Biomass Waste	5
2.2	Statistical Analysis of Production of Oil Palm Biomass such as Oil Palm Kernel Shell (OPKS), Oil Palm Empty Fruit Bunch (OPEFB), Oil Palm Mesocarp Fiber (OPMF), Oil Palm Trunk (OPT) and Oil Palm Frond (OPF)	6
2.3	Palm Oil Milling Process Flow Diagram	7
2.4	Structure of Different Type of Oil Palm Fruit	8
2.5	Types of Oil Palm	8
2.6	Schematic Diagram of Lignocellulosic Biomass Material consists of Cellulose, Hemicellulose and Lignin	10
2.7	Molecular Chain Structure of Cellulose (i), Hemicellulose (ii) and Lignin (iii)	11
2.8	The arrangement of cellulose molecules and hydrogen bonds	12
2.9	Acid Catalyzed Cellulose Hydrolysis Reaction	13
2.10	The Process of Cellulose Dissolution and Hydrolysis	15
2.11	Phase diagram of water in P-T plane	17
2.12	The influence of temperature on water's ionizing constant	18
2.13	The influence of temperature on water's dielectric constants, $\boldsymbol{\epsilon}$	19
3.1	Overview of Research Flow Diagram	20
3.2	Raw Material, OPMF Collected from Gomali Mill	21
3.3	Sub CW extraction system, Molten Salt Bath Celsius 600H, Tomasu, Japan	22
3.4	Schematic diagram of Sub CW Reactor	23
3.5	Flow diagram of Recovery of Sub CW Product Phase for Yield Analysis	24

 $\overline{C}$ 

4.1	Moisture Content of untreated OPMF Sample	29
4.2	Effect of Sub CW Reaction Temperature towards Yield of Oil Obtained	32
4.3	Yield of Total Sugar Obtained from Different Sub CW Reaction Temperature	33
4.4	Yield of Solid Residue at Different Sub CW Temperature Reaction	34
4.5	Yield of Tar from Solid Phase Obtained at Different Sub CW reaction Temperature	35
4.6	Microscopic View of Untreated OPMF (a) and Treated OPMF After Sub CW Reaction at 200°C (b), 240°C (c) and 300°C (d)	37
4.7	SEM image of Untreated OPMF (a) and Treated OPMF After Sub CW Reaction at 200°C (b), 240°C (c) and 300°C (d)	38
4.8	HPLC Analysis of OPMF Treated at Different Sub CW Reaction Temperature	40
4.9	FTIR analysis of OPMF Treated at Different Sub CW Reaction Temperature	41
4.10	Effect of Sub CW temperature on Particle Size of Solid Residue of OPMF Treated at Different Sub CW Reaction Temperature	43
4.11	Relationship between Yield of Sugar and Yield of Oil obtained under Sub-CW treatment	44
4.12	Structure of Cellulose and Its Hydrolysis Products Cellobiose and Glucose	45
4.13	Relationship between Yield of Sugar and Ion Production of Water	46
4.14	Relation between Sugar Concentration and Yield of Tar at Different Sub-CW Temperature	47
4.15	Degradation of Cellulose by Dehydration	47
4.16	Relationship between Yield of Tar and Dielectric Constant of Water	48
4.17	Relationship Between Changes in Particle Size and Production of Total Sugar and Tar	49

# xi

64



# LIST OF TABLES

Table		Page
4.1	Centrifuged Sub CW Reaction Product Obtained at Different Reaction Temperature	30
4.2	Oil Extracted at Different Sub CW Reaction Temperature	31
4.3	Solid Sediment at Bottom of Centrifuge Tube After Separation of Aqueous Phase and Solid Residue Obtained by Filtration	33
4.4	Production of tar at Different Sub CW reaction Temperature	36
4.5	FTIR Wavelength Characterization	42
A.1	Weight and MC of wet sample over series of drying hours	61
A.2	Dry weight of sample based on dry basis MC	62
A.3	Total Oil Collected at Different Sub CW Temperature Reaction	63
A.4	Yield of Oil at Different Sub CW Temperature Reaction	63
A.5	Standard Curve of Total Sugar	64
A.6	Total Sugar obtained at Different Sub CW Temperature Reaction	65
A.7	Yield of Total Sugar at Different Sub CW Temperature Reaction	65
A.8	Total Solid Collected at Different Sub CW Temperature Reaction	66
A.9	Yield of Solid Collected at Different Sub CW Temperature Reaction	66
A.10	Yield of Tar Collected at Different Sub CW Temperature Reaction	67
A.11	Sugar Component Standard Equations	68
A.12	Summary of HPLC Analysis and Sugar Content (mg) Obtained at Different Sub CW Temperature Reaction	69
A.13	Ion Production and Yield of Total Sugar at Different Sub CW Temperature Reaction	70

G

 A.14 Dielectric Constant (ε) and Yield of Tar at Different Sub CW 70 Temperature Reaction



6

# LIST OF ABBREVIATIONS

С	Carbon
FFA	Free Fatty Acid
FTIR	Fourier Transform Infra-Red
HPLC	High Performance Liquid Chromatography
kg	Kilogram
K <sub>w</sub>	Ion Product of Water
L	Litre
МС	Moisture Content
mg	Milli Gram
min	Minute
ml	Milli Litre
mol	Molecule
Mpa	MegaPascals
МРОВ	Malaysian Palm Oil Board
OPEFB	Oil Palm Empty Fruit Brunch
OPF	Oil Palm Frond
OPMF	Oil Palm Mesocarp Fibre
OPT	Oil Palm Truck
Р	Pressure
PKS	Palm Kernel Shell
POME	Palm Oil Mill Effluent
ppm	Parts Per Million

6

PSA	Particle Size Analysis
rpm	Rotations Per Minute
S	Second
SEM	Scanning Electron Microscopy
Sub CW	Sub Critical Water
Т	Temperature
UV μm	Ultraviolet Micro Meter

#### **CHAPTER 1**

#### **INTRODUCTION**

#### 1.1 Research Background

Oil palm mesocarp fibre (OPMF) is known as oil palm fruit lignocellulose fibre that can be obtained after extraction process of palm oil (Iberahim *et al*, 2013). Cellulose, hemicelluloses and lignin are the common components found in mesocarp fibres of oil palms.

Hemicelluloses, according to Yang *et al.*, (2007), loses its weight at temperature between of  $220^{\circ}$ C to  $315^{\circ}$ C while cellulose loses its weight at temperature between of  $315^{\circ}$ C to  $400^{\circ}$ C. Nevertheless, at temperature between of  $160^{\circ}$ C to  $900^{\circ}$ C, lignin loses its weight which makes it difficult to decompose. Hemicellulose has high tendency to degrade compared to lignin and cellulose due to its lesser thermal stability. The complex structure of the oil palm cell makes it difficult to break down the cellulose and lignin using a method of physical industrial scale (Ariffin *et al.*, 2008).

Hydrothermal degrade of cellulose involves chemical reaction such as hydrolysis, pyrolysis, rearrangement reaction and splitting of aldol (Kruse, 2008). Because of its ability to extract high quality product in less time without harming the environment, subcritical water technology is much desirable to be used for the palm oil extraction process instead of solvent as extraction tool. Organic solvents like hexane and ether are usually used in commercial extraction processes. The properties of water in a subcritical state are identical to organic solvent. Such water properties make subcritical water (Sub-CW) as an alternative method of extraction of palm oil, thus reducing the consumption of toxic solvent in industries. Cellulose hydrolysis to fermentable sugars is effective in subcritical water than water at supercritical state (Olanrewaju, 2012).

The water pressure increases with the increasing water temperature to maintain the liquid state of water in the process. At high temperature, water behaves like acid and base catalyst. As a consequence, the order magnitude of  $H_3O^+$  and OH<sup>-</sup> ion concentrations in Sub-CW is higher at a temperature level greater than 200°C and the production of ions goes up to  $1 \times 10^{-11} \text{ mol}^2/\text{L}^2$  at 250°C. Higher ion production facilitates a strong hydrolysis reaction (Omid, 2010). However, as the temperature increases above 250°C, pyrolysis reaction takes place and reduces the ion production drastically (Omid, 2010). The dielectric constant of methanol, ethanol and water are 32.5, 27.0 and 79.9 respectively at room temperature to 17 at 312°C (Pourali *et al.*, 2009). This shows that water at subcritical state can behave as organic solvent like methanol and ethanol. These properties of water

are beneficial for use in treatment of OPMF and transform OPMF into value added products.

#### 1.2 Problem Statement

In 2017, the crude palm oil production rose by 15% to 19.92 million tonnes with respect to 17.32 million tonnes in 2016, due to an increase in the yield of fresh fruit bunches by 5.3 percent (Kushairi *et al.*, 2018). Only 10% of palm oil can be extracted from the total biomass harvested creates the problem of disposal and degradation of environment (Kurnia *et al.*, 2016). The type of biomass waste produced from oil palm industry includes oil palm empty fruit bunches (OPEFB), oil palm trunks (OPT), oil palm mesocarp fibre (OPMF), palm kernel shell (PKS), oil palm fronds (OPF) and palm oil mill effluent (POME) (Abdullah & Sulaiman, 2013). According to Noorshamsiana *et. al* (2017), 6.95 million tons of OPMF is being produced in 2016 after palm oil extraction from 453 palm oil mills in Malaysia. The amount of OPMF produced is more than needed by the oil palm industry to utilize as a source of useful gases production, thus OPMF is used for mulching which can harm the environment (Iberahim *et al.*, 2013).

Cellulose, hemicelluloses and lignin are the common components that can be found in OPMF in the percentage of 42%, 32% and 22% respectively (Nordin *et al.*, 2013). The complex structure of palm oil cell causes difficulty in breaking down the cellulose using the physical method used at current industrial such as press digester, screw press, bead mill and etc. (Ariffin *et al.*, 2008). Cellulose is classified as recalcitrant substance due to its molecules arrangement and hydrogen bonds that make the degradation process difficult (Sasaki et. al, 2004; Olanrewaju, 2012). Arrangement of molecules in cylindrical conformation structure with crystallites form and hydrogen bonds between microfibre's layers makes the cellulose formation stable (Yunos *et al.*, 2012). Cellulose decomposes into water-soluble substance in the absence of catalyst between 200°C to 400°C which proves that hydrothermal reaction is able to hydrolyse cellulose (Nordin *et al.*, 2013). Pyrolysis reaction occurs at a higher temperature in the absence of oxygen to release carbon dioxide, carbon monoxide and a significant amount of char. According to Tolonen, (2016), slow pyrolysis and fast pyrolysis reactions take place below 300°C and above 300°C respectively.

The commercial method which is the Soxhlet extraction currently uses organic solvents such as hexane and ether in the extraction process of the oil (Azmir *et al.*, 2013; Mohammad and Fereshteh, 2007). Subcritical water technology is much preferable to be used for palm oil extraction process instead of organic solvent as extraction medium due to its capability to extract high-quality product in a lesser amount of time without harming the environment (Abdelmoez *et al.*, 2014). Value of dielectric constant of water above 200°C is similar as the dielectric constant of methanol at room temperature, which makes Sub-CW to be a suitable solvent for hydrophobic organic (Amashukeli *et al.*, 2007 and Singh and Saldana, 2011). At subcritical state, water behaves as acid and base catalyst (Carr *et al.*, 2011). This technology gets attention due to its behaviour of producing high ion and low dielectric constant at an increased temperature (Adachi, 2009).

Subcritical water reaction is capable of facilitating structural conversion of OPMF to sugar as 240°C and alternative oil at higher than 240°C without any chemical solvent being used. Hydrolysis process occurs at temperature below 240°C by breakdown hydrogen bond present in cell wall of OPMF and release lignocellulose material. As the temperature gets higher, dehydration occur in process of pyrolysis which alter sugar content into alternative oil. According to Agensi Inovasi Malaysia (2013), the highest biomass from palm oil plantation is from Oil Palm Frond (OPF). However, OPMF was used to identify alteration after treat with Sub CW due to it is known as sustainable resources that able to utilise for various of valuable processes and products (Iberahim *et al.*, 2013).

Thus, OPMF treated in Sub-CW at a 240°C reaction temperature provides optimum sugar yield with less alternative oil output by ease cell disruption of OPMF. Structural transformation and conversion of lignocellulosic biomass such as OPMF after treat with Sub CW was not studied. The relationship between the structural and compositional alteration of OPMF and the substance formed at water in sub-critical state needed to be identifying to maximize Sub CW reaction effect towards conversion of OPMF.

#### 1.3 Research Objective

The main objective of this research is to study on the transformation of mesocarp fibre composition in water at different temperature in subcritical state. The specific objectives of the studies are as following:

- 1. To determine optimum Sub-CW reaction temperature in breaking down of fibre structure for optimum biomass conversion into desired product through hydrolysis and pyrolysis reactions.
- 2. To examine the structural changes of oil palm mesocarp fibre after Sub-CW reaction at different reaction temperature
- 3. To identify chemical composition of oil palm mesocarp fibre after undergoes Sub-CW reaction.

#### 1.4 Scopes of Study

This study was carried out mainly to analyse the structural and compositional transformation of mesocarp fibre treated in water at different temperature of subcritical state. The OPMF samples used in this research was collected from Gomali Mill, Gemas before and after press digester. Moisture content of sample was determined by dry the samples in oven at 80°C. The samples were viewed under a light microscope and a Scanning Electronic Microscope (SEM) before and after the samples were treated by Sub-CW to study the breakdown and structural changes of OPMF's cellulose at different temperatures. Reaction temperatures were between 200°C and 300°C to obtain the

optimum temperature to produce desirable product from OPMF. Compositional changes of OPMF were identified using HPLC, UV Spectroscopy and IR spectroscopy.



#### REFERENCES

- Abdelmoez, W., Abdelfatah, R., Tayeb, A., & Yoshida, H. (2011). Extraction of cottonseed oil using subcritical water technology. AIChE journal, 57(9), 2353-2359.
- Abdelmoez, W., Nage, S. M., Bastawess, A., Ihab, A., & Yoshida, H. (2014). Subcritical water technology for wheat straw hydrolysis to produce value added products. *Journal of Cleaner Production*, 70, 68-77.
- Abdullah, N., & Sulaiman, F. (2013). The oil palm wastes in Malaysia. Biomass nowsustainable growth and use, 1(3), 75-93.
- Abdullah, R., & Wahid, M. B. (2010). World palm oil supply, demand, price and prospects: focus on Malaysian and Indonesian palm oil industry. Malaysian Palm Oil Board Press, Malaysia.
- Abdulrazik, A., Elsholkami, M., Elkamel, A., & Simon, L. (2017). Multi-products productions from Malaysian oil palm empty fruit bunch (EFB): Analyzing economic potentials from the optimal biomass supply chain. Journal of cleaner production, 168, 131-148.
- Adachi, S. (2009). Properties of subcritical water and its utilization. Division of Food Science and Biotechnology, Graduate School of Agriculture, Kyoto University.
- Agirre, I., Griessacher, T., Rösler, G., & Antrekowitsch, J. (2013). Production of charcoal as an alternative reducing agent from agricultural residues using a semi-continuous semi-pilot scale pyrolysis screw reactor. Fuel Processing Technology, 106, 114-121.
- Akiya, N., & Savage, P. E. (2002). Roles of water for chemical reactions in high-temperature water. Chemical reviews, 102(8), 2725-2750.
- Alhattab, M., Kermanshahi-Pour, A., & Brooks, M. S. L. (2019). Microalgae disruption techniques for product recovery: influence of cell wall composition. Journal of Applied Phycology, 31(1), 61-88.
- Alvira, P., Tomás-Pejó, E., Ballesteros, M., & Negro, M. J. (2010). Pretreatment technologies for an efficient bioethanol production process based on enzymatic hydrolysis: a review. Bioresource technology, 101(13), 4851-4861.
- Amashukeli, X; Pelletier, C C; Kirby, J P and Grunthaner, F J (2007). Subcritical water extraction of amino acids from Atacama Desert soils, J. Geophysical Res., (112): 1–10
- APV, An SPX Brand (2009); Cell disruption by Homogenization; http://www.apvhemisan.com/uploads/images/Cell\_Disruption\_by\_Homogeniz ation\_3006\_01\_06\_2008\_US.pdf

- Ariffin, H; Hassan, M A; Umi Kalsom, M S; Abdullah, N and Shirai, Y (2008). Effect of physical, chemical and thermal pretreatments on the enzymatic hydrolysis of oil palm empty fruit bunch (OPEFB). J. Trop. Agric. Food Sci. 2008, 36: 259– 268
- Asadieraghi, M., & Daud, W. M. A. W. (2014). Characterization of lignocellulosic biomass thermal degradation and physiochemical structure: effects of demineralization by diverse acid solutions. Energy Conversion and Management, 82, 71-82.
- Azmir, J., Zaidul, I. S. M., Rahman, M. M., Sharif, K. M., Mohamed, A., Sahena, F., ... & Omar, A. K. M. (2013). Techniques for extraction of bioactive compounds from plant materials: A review. Journal of food engineering, 117(4), 426-436.
- Balat, M (2011). Production of bioethanol from lignocellulosic materials via the biochemical pathway: a review. Energy Conversion and Management, 52(2): 858-875
- Barcelos, E., Rios, S. D. A., Cunha, R. N., Lopes, R., Motoike, S. Y., Babiychuk, E., & Kushnir, S. (2015). Oil palm natural diversity and the potential for yield improvement. Frontiers in plant science, 6, 190.
- Baryeh, E. A. (2001). Effects of palm oil processing parameters on yield. Journal of Food Engineering, 48(1), 1-6.
- Brenes, M. D. (2006). Biomass and bioenergy: new research. Nova Publishers.
- Bridgwater, A. V. (2012). Review of fast pyrolysis of biomass and product upgrading. Biomass and bioenergy, 38, 68-94.
- Calvini P, Gorassini A, Merlani AL (2008) On the kinetics of cellulose degradation: looking beyond the pseudo zero order rate equation. Cellulose 15:193–203. doi:10.1007/s10570-007-9162-8
- Carr, A. G., Mammucari, R., & Foster, N. R. (2011). A review of subcritical water as a solvent and its utilisation for the processing of hydrophobic organic compounds. Chemical Engineering Journal, 172(1), 1-17
- Carrier, M., Loppinet-Serani, A., Absalon, C., Aymonier, C., & Mench, M. (2012). Degradation pathways of holocellulose, lignin and α-cellulose from Pteris vittata fronds in sub-and super critical conditions. Biomass and bioenergy, 43, 65-71
- Chaturvedi, V., & Verma, P. (2013). An overview of key pretreatment processes employed for bioconversion of lignocellulosic biomass into biofuels and value added products. 3 Biotech, 3(5), 415-431.
- Chen, B., McClements, D. J., Gray, D. A., & Decker, E. A. (2012). Physical and oxidative stability of pre-emulsified oil bodies extracted from soybeans. Food chemistry, 132(3), 1514-1520

- Chen, H. (2014). Chemical composition and structure of natural lignocellulose. In Biotechnology of lignocellulose (pp. 25-71). Springer, Dordrecht
- Cyr, M., & Tagnit-Hamou, A. (2001). Particle size distribution of fine powders by LASER diffraction spectrometry. Case of cementitious materials. Materials and Structures, 34(6), 342-350.
- Demirbaş, A. (2003). Biodiesel fuels from vegetable oils via catalytic and non-catalytic supercritical alcohol transesterifications and other methods: a survey. Energy conversion and Management, 44(13), 2093-2109.
- Demirbas, A. (2007). Products from lignocellulosic materials via degradation processes. Energy Sources, Part A: Recovery, Utilization, and Environmental Effects, 30(1), 27-37.
- Eikani, M. H., Golmohammad, F., & Rowshanzamir, S. (2007). Subcritical water extraction of essential oils from coriander seeds (Coriandrum sativum L.). Journal of Food Engineering, 80(2), 735-740.
- Farrokhi, N., Burton, R. A., Brownfield, L., Hrmova, M., Wilson, S. M., Bacic, A., & Fincher, G. B. (2006). Plant cell wall biosynthesis: genetic, biochemical and functional genomics approaches to the identification of key genes. Plant biotechnology journal, 4(2), 145-167
- Franssen, M. C., Steunenberg, P., Scott, E. L., Zuilhof, H., & Sanders, J. P. (2013). Immobilised enzymes in biorenewables production. Chemical Society Reviews, 42(15), 6491-6533
- Garrote, G. D. H. P., Dominguez, H., & Parajo, J. C. (1999). Hydrothermal processing of lignocellulosic materials. Holz als Roh und Werkstoff, 57, 191-202
- Giannoccaro, E., Wang, Y. J., & Chen, P. (2008). Comparison of two HPLC systems and an enzymatic method for quantification of soybean sugars. Food Chemistry, 106(1), 324-330.
- Gomez del Pulgar, E. M., & Saadeddin, A. (2014). The cellulolytic system of Thermobifida fusca. Critical reviews in microbiology, 40(3), 236-247
- Hadi, S., Ahmad, D., & Akande, F. B. (2009). Determination of the bruise indexes of oil palm fruits. Journal of food engineering, 95(2), 322-326.
- Hendriks, A. T. W. M., & Zeeman, G. (2009). Pretreatments to enhance the digestibility of lignocellulosic biomass. Bioresource technology, 100(1), 10-18.
- Hodgman, C D and Lange N A (1924). Handbook of Chemistry and Physics. Cleveland: Chemical Rubber Co.: 312-313
- Hosseinaei, O; Wang, S; Enayati, A A and Rials, T G (2012). Effects of hemicellulose extraction on properties of wood flour and wood-plastic composites. Compos. Part A Appl. Sci. Manuf. 43(4): 686-694

- Iberahim, N. I., Jahim, J. M., Harun, S., Nor, M. T. M., & Hassan, O. (2013). Sodium hydroxide pretreatment and enzymatic hydrolysis of oil palm mesocarp fiber. International Journal of Chemical Engineering and Applications, 4(3), 101.
- Jirasatid, S., Chaikham, P., & Nopharatana, M. (2018). Thermal degradation kinetics of total carotenoids and antioxidant activity in banana-pumpkin puree using Arrhenius, Eyring-Polanyi and Ball models. International Food Research Journal, 25(5).
- Kang, X., Kirui, A., Widanage, M. C. D., Mentink-Vigier, F., Cosgrove, D. J., & Wang, T. (2019). Lignin-polysaccharide interactions in plant secondary cell walls revealed by solid-state NMR. Nature communications, 10(1), 1-9.
- Koc, B., Eren, I., & Ertekin, F. K. (2008). Modelling bulk density, porosity and shrinkage of quince during drying: The effect of drying method. Journal of food engineering, 85(3), 340-349.
- Kruse, A. (2008). Supercritical water gasification. Biofuels, Bioproducts and Biorefining: Innovation for a sustainable economy, 2(5), 415-437
- Kurnia, J. C., Jangam, S. V., Akhtar, S., Sasmito, A. P., & Mujumdar, A. S. (2016). Advances in biofuel production from oil palm and palm oil processing wastes: a review. Biofuel Research Journal, 3(1), 332-346
- Kurnin, N A A; Ismail, M H S; Yoshida, H and Izhar, S (2016). Recovery of Palm Oil and Valuable Material from Oil Palm Empty Fruit Bunch by Sub-critical Water. J. Oleo Sci., 65 (4): 283-289
- Kushairi, A., Loh, S. K., Azman, I., Hishamuddin, E., Ong-Abdullah, M., Izuddin, Z. B. M. N., & Parveez, G. K. A. (2018). Oil palm economic performance in Malaysia and R&D progress in 2017. Journal of Oil Palm Research, 30(2), 163-195.
- Lemmon, E. W., McLinden, M. O., & Friend, D. G. (2017). Thermophysical Properties of Fluid Systems in NIST Chemistry WebBook, NIST Standard Reference Database Number 69; Linstrom, PJ, Mallard, WG, Eds.; National Institute of Standards and Technology: Gaithersburg MD. J., Mallard, WG, Eds
- Liu, D., Yu, Y., Hayashi, J. I., Moghtaderi, B., & Wu, H. (2014). Contribution of dehydration and depolymerization reactions during the fast pyrolysis of various salt-loaded celluloses at low temperatures. Fuel, 136, 62-68
- Loh, S. K. (2017). The potential of the Malaysian oil palm biomass as a renewable energy source. Energy conversion and management, 141, 285-298.
- Mazaheri, H., Lee, K. T., Bhatia, S., & Mohamed, A. R. (2010). Sub/supercritical liquefaction of oil palm fruit press fiber for the production of bio-oil: effect of solvents. Bioresource technology, 101(19), 7641-7647.

- Mezzomo, N., & Ferreira, S. R. (2016). Carotenoids functionality, sources, and processing by supercritical technology: a review. Journal of Chemistry, 2016
- Möller, M., & Schröder, U. (2013). Hydrothermal production of furfural from xylose and xylan as model compounds for hemicelluloses. Rsc Advances, 3(44), 22253-22260.
- MPOB. Malaysian oil palm statistics 2017. 37th ed. Bangi, Malaysia: Malaysian Palm Oil Board; 2018. p. 205
- Muda, N. A., Yoshida, H., Ishak, H., Ismail, M. H. S., & Izhar, S. (2019). Conversion of oil palm trunk into bio-oil via treatment with subcritical water. Journal of wood chemistry and technology, 39(4), 255-269.
- Neoh, B. K., Thang, Y. M., Zain, M. Z. M., & Junaidi, A. (2011). Palm pressed fibre oil: A new opportunity for premium hardstock?. Int. Food Res. J, 18(2), 769-773
- Nevell, T. P., & Zeronian, S. H. (1985). Cellulose chemistry and its applications.
- Nikiforidis, C. V., Matsakidou, A., & Kiosseoglou, V. (2014). Composition, properties and potential food applications of natural emulsions and cream materials based on oil bodies. RSC Advances, 4(48), 25067-25078
- Njoku, P. C., Egbukole, M. O., & Enenebeaku, C. K. (2010). Physio-chemical Characteristics and Dietary Metal Levels of Oil from Elaesis guineensis species.
- Nomanbhay, S. M., Hussain, R., & Palanisamy, K. (2013). Microwave-assisted alkaline pretreatment and microwave assisted enzymatic saccharification of oil palm empty fruit bunch fiber for enhanced fermentable sugar yield.
- Noorshamsiana, A. W., Nur, E., Fatiha, I., & Astimar, A. A. (2017). A review on extraction processes of lignocellulosic chemicals from oil palm biomass. Journal of Oil Palm Research, 29(4), 512-527
- Nordin, N.I.; Ariffin, H.; Andou, Y.; Hassan, M.A.; Shirai, Y.; Nishida, H.; Yunus, W.Z.; Karuppuchamy, S. and Ibrahim, N.A. (2013). Modification of oil palm mesocarp fibre characteristics using superheated steam treatment. Molecules, 18(8): 9132-46
- Ohno, E., & Miyafuji, H. (2014). Decomposition of cellulose in an ionic liquid, 1-ethyl-3-methylimidazolium chloride. Journal of wood science, 60(6), 428-437
- Olanrewaju, K. B. (2012). Reaction kinetics of cellulose hydrolysis in subcritical and supercritical water. PhD Thesis, University of Iowa
- Omid, P. (2010) Production of Valuable Materials from Rice Bran Biomass Using Subcritical Water. Doctoral Thesis at Osaka Prefecture University

- Oriez, V., Peydecastaing, J., & Pontalier, P. Y. (2019). Lignocellulosic biomass fractionation by mineral acids and resulting extract purification processes: Conditions, yields, and purities. Molecules, 24(23), 4273.
- Ouellette, R. J., & Rawn, J. D. (2014). 1-Structure and Bonding in Organic Compounds. Organic Chemistry. Boston: Elsevier, 1-39
- Pohl, H. (2010). A scanning electron microscopy specimen holder for viewing different angles of a single specimen. Microscopy Research and Technique, 73(12), 1073-1076.
- Pourali, O. (2010). Production of valuable materials from rice bran biomass using subcritical water. Osaka prefecture University.
- Pourali, O; Salak, F and Yoshida, H (2009a). Simultaneous rice bran oil stabilization and extraction using sub-critical water medium. J. Food Eng., 95(3): 510–516.
- Provesi, J. G., Dias, C. O., & Amante, E. R. (2011). Changes in carotenoids during processing and storage of pumpkin puree. Food Chemistry, 128(1), 195-202.
- Ravber, M., Knez, Ž., & Škerget, M. (2015). Simultaneous extraction of oil-and watersoluble phase from sunflower seeds with subcritical water. Food chemistry, 166, 316-323
- Rizal, N. F. A. A., Ibrahim, M. F., Zakaria, M. R., Abd-Aziz, S., Yee, P. L., & Hassan, M. A. (2018). Pre-treatment of oil palm biomass for fermentable sugars production. Molecules, 23(6), 1381.
- Rytioja, J., Hildén, K., Yuzon, J., Hatakka, A., de Vries, R. P., & Mäkelä, M. R. (2014). Plant-polysaccharide-degrading enzymes from basidiomycetes. Microbiology and Molecular Biology Reviews, 78(4), 614-649
- Sabil, K. M., Aziz, M. A., Lal, B., & Uemura, Y. (2013). Effects of torrefaction on the physiochemical properties of oil palm empty fruit bunches, mesocarp fiber and kernel shell. Biomass and Bioenergy, 56, 351-360.
- Sadasivam, S. (1996). Biochemical methods. New age international.
- Saha, P., Manna, S., Chowdhury, S. R., Sen, R., Roy, D., & Adhikari, B. (2010). Enhancement of tensile strength of lignocellulosic jute fibers by alkali-steam treatment. Bioresource technology, 101(9), 3182-3187.
- Saka, S., & Ueno, T. (1999). Chemical conversion of various celluloses to glucose and its derivatives in supercritical water. Cellulose, 6(3), 177-191
- Sampaio, K A; Ayala, J V; Silva, S M; Ceriani, R; Verhé, R and Meirelles, A J (2013). Thermal degradation kinetics of carotenoids in palm oil. J. Am. Oil Chem. Soc., 90(2): 191-198

- Sasaki, M., & Goto, M. (2008). Recovery of phenolic compounds through the decomposition of lignin in near and supercritical water. Chemical Engineering and Processing: Process Intensification, 47(9-10), 1609-1619
- Sasaki, M., Adschiri, T., & Arai, K. (2004). Kinetics of cellulose conversion at 25 MPa in sub-and supercritical water. AIChE Journal, 50(1), 192-202
- Siew, W. L., & MPOB, J. (2007). Palm Oil and Fractions. OFI Middle East Proceeding.
- Singh, P P and Saldaña, M D A (2011). Subcritical water extraction of phenolic compounds from potato peel, Food Res. Int., 44(8): 2452–2458
- Singh, R. K., Pandey, D., Patil, T., & Sawarkar, A. N. (2020). Pyrolysis of banana leaves biomass: physico-chemical characterization, thermal decomposition behavior, kinetic and thermodynamic analyses. Bioresource technology, 310, 123464.
- Sinha, E., & Rout, S. K. (2009). Influence of fibre-surface treatment on structural, thermal and mechanical properties of jute fibre and its composite. Bulletin of materials science, 32(1), 65.
- Sreekala, M. S., Kumaran, M. G., Joseph, S., Jacob, M., & Thomas, S. (2000). Oil palm fibre reinforced phenol formaldehyde composites: influence of fibre surface modifications on the mechanical performance. Applied Composite Materials, 7(5-6), 295-329.
- Stanbury, P. F., Whitaker, A., & Hall, S. J. (2016). Principles of fermentation technology. Elsevier. (third edition)
- Sterling, J. D., Atmodjo, M. A., Inwood, S. E., Kolli, V. K., Quigley, H. F., Hahn, M. G., & Mohnen, D. (2006). Functional identification of an Arabidopsis pectin biosynthetic homogalacturonan galacturonosyltransferase. Proceedings of the National Academy of Sciences, 103(13), 5236-5241
- Subramaniam, Vijaya., Menon, N. R., Sin, H., & May, C. Y. (2013). The development of a residual oil recovery system to increase the revenue of a palm oil mill. J Oil Palm Res, 25(1), 116-122.
- Sue, T. T., & Pantzaris, T. (2009). Pocketbook of palm oil uses. Kuala Lumpur, Malaysia: Malaysian palm oil board.
- Teh, C. C., Ibrahim, N. A., & Yunus, W. M. Z. W. (2013). Response surface methodology for the optimization and characterization of oil palm mesocarp fiber-graft-poly (butyl acrylate). BioResources, 8(4), 5244-5260.
- Teoh, C. H. (2002). The palm oil industry in Malaysia: from seed to frying pan. Report of WWF, Malaysia.
- Teoh, Y. P., & Don, M. M. (2011). Kinetic model for the hydrolysis of sterilized palm press fibre. Chemical engineering science, 66(15), 3523-3530

- Then, Y. Y., Ibrahim, N. A., Zainuddin, N., Ariffin, H., Yunus, W. M. Z. W., & Chieng, B. W. (2014). Surface modifications of oil palm mesocarp fiber by superheated steam, alkali, and superheated steam-alkali for biocomposite applications. BioResources, 9(4), 7467-7483
- Ting, S. V. (1956). Fruit juice assay, rapid colormetric methods for simultaneous determination of total reducing sugars and fructose in citrus juices. Journal of Agricultural and Food Chemistry, 4(3), 263-266.
- Tunchaiyaphum, S., Eshtiaghi, M. N., & Yoswathana, N. (2013). Extraction of bioactive compounds from mango peels using green technology. International Journal of Chemical Engineering and Applications, 4(4), 194
- Uematsu, M., & Frank, E. U. (1980). Static dielectric constant of water and steam. Journal of Physical and Chemical Reference Data, 9(4), 1291-1306.
- Vamvuka, D. (2011). Bio-oil, solid and gaseous biofuels from biomass pyrolysis processes—an overview. International journal of energy research, 35(10), 835-862.
- Verheye, W. (2010). Growth and production of oil palm. In Land use, land cover and soil sciences. UNESCO-EOLSS Publishers
- Wagner, W., & Prub, A. (2002). The IAPWS formulation 1995 for the thermodynamic properties of ordinary water substance for general and scientific use. Journal of physical and chemical reference data, 31(2), 387-535.
- Wang, K., Jiang, J. X., Xu, F., & Sun, R. C. (2009). Influence of steaming pressure on steam explosion pretreatment of Lespedeza stalks (Lespedeza crytobotrya): Part 1. Characteristics of degraded cellulose. Polymer degradation and stability, 94(9), 1379-1388.
- Weemaes, C. A., Ooms, V., Van Loey, A. M., & Hendrickx, M. E. (1999). Kinetics of chlorophyll degradation and color loss in heated broccoli juice. Journal of Agricultural and Food Chemistry, 47(6), 2404-2409.
- Xiao, B., Sun, X., & Sun, R. (2001). Chemical, structural, and thermal characterizations of alkali-soluble lignins and hemicelluloses, and cellulose from maize stems, rye straw, and rice straw. Polymer degradation and stability, 74(2), 307-319.
- Yahya, M. B., Lee, H. V., & Abd Hamid, S. B. (2015). Preparation of nanocellulose via transition metal salt-catalyzed hydrolysis pathway. BioResources, 10(4), 7627-7639.
- Yang, H., Yan, R., Chen, H., Lee, D. H., & Zheng, C. (2007). Characteristics of hemicellulose, cellulose and lignin pyrolysis. Fuel, 86(12-13), 1781-1788.
- York, W. S., Darvill, A. G., McNeil, M., Stevenson, T. T., & Albersheim, P. (1986). Isolation and characterization of plant cell walls and cell wall components. In Methods in enzymology(Vol. 118, pp. 3-40). Academic Press

- Yoshida, H; Izhar, S.; Nishio, E., Utsumi, Y.; Kakimori, N. and Feridoun, S. A. (2018). Application of Sub-Critical Water for Recovery of Tin and Glass Substrates from LCD Panel E-Waste. Detritus, 4: 98-103
- Young, H. A., & Extraordinary, A. (1977). Food and agriculture organization of the united nation Rome
- Yunos, N. S. H. M., Baharuddin, A. S., Yunos, K. F. M., Naim, M. N., & Nishida, H. (2012). Physicochemical property changes of oil palm mesocarp fibers treated with high-pressure steam. BioResources, 7(4), 5983-5994
- Zhou, X., Broadbelt, L. J., & Vinu, R. (2016). Mechanistic understanding of thermochemical conversion of polymers and lignocellulosic biomass. Advances in chemical engineering, 49, 95-198.

### APPENDICES

#### Appendix 1: Raw data for moisture content based on dry basis and wet basis

Equation (1) used to obtain moisture content of average sample for 1 hour duration based on dry basis:



Table A.1: Weight and MC of wet sample over series of drying hours

		Sample 1 (g)	Sample 2 (g)	Sample 3 (g)	Average (g)	Dry Basis (%)
D	ish Weight	0.5495	0.602	0.5055	0.5523	
Sa	mple Weight	0.4994	0.4991	0.5168	0.5051	
Т	otal Weight	1.0489	1.1011	1.0223	1.0574	
	1	0.7971	0.8489	<mark>0.7737</mark>	0.8066	23.7%
Ĺ.	2	0.7925	0.8400	0.7699	0.8008	24.3%
1) na	3	0.7876	0.8335	0.7693	0.7968	24.6%
ratic	4	0.7853	0.8330	0.7688	0.7957	24.8%
Ĩ	5	0.7845	0.8327	0.7683	0.7952	24.8%
	24	0.7843	0.8327	0.7680	0.7950	24.8%

# Appendix 2: Raw Data of dry sample weight used in different reaction temperature in Sub-CW reaction (wet basis and dry basis)

Equation (6) was used to obtain the dry weight of sample used in temperature 200°C in 5 minutes' reaction time:

Weight of dry sample =1 - 
$$\frac{MC}{100}$$
 X weight of wet sample (6)  
=  $(1 - \frac{24.8}{100})$  X 0.5072 g dry sample  
= 0.3814 g dry sample

Temperature (°C)	Sample Weight (g)	Dry Basis (g)
200	0.5072	0.3814
210	0.5066	0.3810
220	0.5016	0.3772
230	0.5041	0.3791
240	0.5122	0.3852
250	0.5001	0.3761
260	0.5140	0.3865
300	0.5089	0.3827

# Table A.2: Dry weight of sample based on dry basis MC

# Appendix 3: Raw data of yield of oil obtained from different reaction temperature in Sub-CW reaction

Equation (2) was used to obtain yield of oil at reaction temperature 200°C in 5 minutes' reaction time:

Oil phase yield (g oil/g dry OPMF) =  $\frac{dry oil phase weight}{charged dry sample weight based on wet basis}$  (2)

 $= \frac{0.0080 \, (g)}{0.3814 \, (g \, dry \, sample)}$ 

= 0.0210 g/g dry sample

#### Table A.3: Total Oil Collected at Different Sub-CW Temperature Reaction

Temperature	Vial Weight	Vial + Oil Weight	Total Oil Collected
(°C)	(g)	(g)	(g)
200	15.1350	15.1430	0.0080
210	19.2132	19.2276	0.0144
220	15.0456	15.0644	0.0188
230	19.2731	19.2972	0.0241
240	19.0460	19.0814	0.0354
250	14.9743	14.9903	0.0160
260	19.4392	19.4480	0.0088
300	19.0963	19.0971	0.0008

#### Table A.4: Yield of Oil at Different Sub-CW Temperature Reaction

Temperature (°C)	Yield of Oil (g/g-dry sample)	Yield of Oil (mg/g-dry sample)
200	0.0210	20.97
210	0.0378	37.80
220	0.0498	49.84
230	0.0636	63.57
240	0.0919	91.91
250	0.0425	42.54
260	0.0228	22.77
300	0.0021	2.09

Appendix 4: Raw data of yield of sugar obtained from different reaction temperature in Sub-CW reaction

	Absor	Absorbance		
Glucose (ppm)	A1	A2	Average	
0	0.2210	0.2500	0.2355	
10	0.3110	0.3530	0.3320	
25	0.4730	0.5520	0.5125	
50	0.7226	0.7310	0.7268	
100	1.2776	1.3200	1.2988	

Table A.5: Standard Curve of Total Sugar



Figure A.1: Graph of Sugar Substance Standard Curve

Temperature (°C)	Absorbance Reading	Concentration of Sugar (g/ml)	Total Sugar (g)
200	0.324	8.868	0.0022
210	0.358	12.075	0.0030
220	0.422	18.113	0.0045
230	0.474	23.019	0.0058
240	0.592	34.151	0.0085
250	0.516	26.981	0.0067
260	0.432	19.057	0.0048
300	0.271	3.868	0.0010

Table A.6: Total Sugar obtained at Different Sub-CW Temperature Reaction

Absorbance value and concentration of sugar of reacted sample at different reaction temperature identified using standard curve of total sugar.

Equation (3) was used to obtain yield of total sugar at reaction temperature 200°C in 5 minutes' reaction time:

Total sugar yield (g/g-dry sample) = weight of total sugar (g) (3)

> = 0.002 g 0.3814 (g-dry sample)

= 0.0065 g/g-dry sample

Table A.7:	Yield	of Total	Sugar	at Different	Sub-CW	Temperature	Reaction

Temperature (°C)	Yield of Suga <mark>r</mark> (g/g-dry sample)	Yield of Total Sugar (mg/g-dry sample)
200	0.0058	5.831
210	0.0079	7.924
220	0.0120	12.005
230	0.0152	15.181
240	0.0222	22.166
250	0.0179	17.936
260	0.0123	12.326
300	0.0025	2.527

Appendix 5: Raw data of solid residue obtained from different reaction temperature in Sub-CW reaction

Temperature (°C)	Solid Collected from Centrifuge Tube (g)	Solid Collected from Filter Paper (g)	Total Solid Collected (g)
200	0.0371	0.2494	0.2865
210	0.0322	0.2150	0.2472
220	0.0321	0.2124	0.2445
230	0.0310	0.1512	0.1822
240	0.0244	0.1368	0.1612
250	0.0226	0.1106	0.1332
260	0.0212	0.1096	0.1308
300	0.0120	0.0246	0.0366

Table 11.0. Total Dona Concella at Different Dab-Cit Temperature Reaction	Table A.8: 7	<b>Fotal Solid</b>	Collected at	t Different Sub-CV	W Tem	perature Reaction
---	--------------	--------------------	--------------	--------------------	-------	-------------------

Equation (4) was used to obtain yield of oil at reaction temperature 200°C in 5 minutes' reaction time:

Solid phase residue yield

weight of solid phase residue(g) weight of charged sample (g-dry sample)

(4)

0.2865 g 0.3814 (g-dry sample)

= 0.7512 g/g-dry sample

Table A.9: Yield of Solid Collected at Different Sub-CW Temperature Reaction

=

Temperature (°C)	Yield of Solid (g/g-dry sample)	Yield of Solid (mg/g-dry sample)
200	0.7512	751.1515
210	0.6489	648.8816
220	0.6482	648.1917
230	0.4806	480.6333
240	0.4185	418.5117
250	0.3542	354.1845
260	0.3384	338.3972
300	0.0956	95.6381

## Appendix 6: Raw data of tar obtained from different reaction temperature in Sub-CW reaction

Equation (5) was used to obtain yield of tar at reaction temperature 200°C in 5 minutes' reaction time:

Yield of Tar

= weight of tar(g) weight of charged sample (g-dry sample)

(5)

 $= \frac{0.0021 \text{ g}}{0.3814 (\text{g-dry sample})}$ 

# = 0.0055 g/g-dry sample

#### Table A.10: Yield of Tar Collected at Different Sub-CW Temperature Reaction

Temperature	Weight of	Yield of Tar (g/g-dry	Yield of Tar (mg/g-dry
(°C)	tar (g)	sample)	sample)
200	0.0021	0.0055	5.5058
210	0.0057	0.0150	14.9621
220	0.0069	0.0183	18.2925
230	0.0101	0.0266	26.6432
240	0.0231	0.0600	59.9728
250	0.0443	0.1178	117.7956
260	0.0632	0.1635	163.5069
300	0.0718	0.1876	187.6178

Appendix 7: Raw data of HPLC analysis on aqueous phase obtained from different reaction temperature in Sub-CW reaction



**Table A.11: Sugar Component Standard Equations** 



Table A.12: Summary	of HPLC Analysis	and Sugar	Content (mg	) Obtained at
Different Sub-CW Tem	perature Reaction			

Temperature (°C)	tR	Type of Sugar	Area	ppm	Sugar Content (mg/ml)	Sugar Content (mg)
200	17.267	Cellobiose	13511	30.733	76.834	1.536
200	19.258	Glucose	33343	81.886	204.714	4.094
220	17.942	Cellobiose	21191	48.099	120.247	2.405
220	19.075	Glucose	44461	104.415	261.038	5.220
	17.442	Cellobiose	4151	9.569	23.924	0.478
240	18.492	Glucose	34836	84.911	212.278	4.245
240	22.017	Galactose	3100	9.621	24.053	0.481
	22.875	Mannose	30700	69.080	172.701	3.454
	17.033	Cellobiose	326	0.920	2.302	0.046
200	18.392	Glucose	28190	71.444	178.61	3.572
260	23.183	Mannose	25410	57.319	143.297	2.866
	22.100	Galactose	1006	5.013	12.532	0.250

Appendix 8: Raw Data of Sub-CW Reaction Product and Ion Production

Temperature (°C)	Ion Product Constant in base 10 log (mol/kg)2	Yield of Total Sugar (mg/g-dry sample)
200	-11.16	5.81
250	-11.01	22.17
300	-11.14	2.53

 Table A.13: Ion Production and Yield of Total Sugar at Different Sub-CW

 Temperature Reaction

# Appendix 9: Raw Data of Sub-CW Reaction Product and Ion Production

Table A.14: Dielectric Constant (ε) and Yield of Tar at Different Sub-CW Temperature Reaction

Temperature (°C)	Dielectric Const. (ɛ)	Yield of Tar (mg/g-dry sample)
200	34.742	5.506
210	33.091	14.962
220	31.496	18.293
230	29.952	26.643
240	28.455	59.973
250	26.999	117.796
260	25.579	163.507
300	20.135	187.618

#### **BIODATA OF STUDENT**

Sanggithapriya Mahandran was born on 19th October 1994 in Negeri Sembilan. Her primary school at Sekolah Jenis Kebangsaan (tamil) Ladang Senama. She proceeds her secondary school at Sekolah Menengah Kebangsaan Batu Kikir (Model Khas). Then, she continued her study in matriculation level at Perak Matriculation College in 2012. She furthered her degree with Bachelor of Chemical Engineering at Universiti Putra Malaysia in 2013. She graduated her degree in November 2017 and enrolled for the master study in September 2018. She struggles to finish her master study in December 2020 and looking forward any opportunity to further for PhD study.



## LIST OF PUBLICATIONS

Mahandran, S., Sabli, N., Ismail, M. H. S., Yoshida, H., & Izhar, S. (2020). Structural transformation of oil palm mesocarp fibre (OPMF) in water at subcritical state. Journal of Oil Palm Research, 32(4), 590-598.





## **UNIVERSITI PUTRA MALAYSIA**

## STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

# ACADEMIC SESSION : Second Semester 2020/2021

# TITLE OF THESIS / PROJECT REPORT :

# TRANSFORMATION OF OIL PALM MESOCARP FIBER TO CONVERTIBLE

# MATERIALS IN WATER AT SUBCRITICAL STATE

## NAME OF STUDENT :

## Sanggithapriya A/P Mahandran

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

- 1. This thesis/project report is the property of Universiti Putra Malaysia.
- 2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
- 3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as:



This thesis is submitted for:

PATENT	Embargo from	until
	(date)	(uale)
		Approved by:
(Signature of Student) New IC No/ Passport No.: Date :		(Signature of Chairman of Supervisory Committee) Name: Date :
[Note : If the thesis is CONFI the letter from the organizati confidentially or restricted.]	DENTIAL or RES on/institution wi	STRICTED, please attach with th period and reasons for