



UNIVERSITI PUTRA MALAYSIA

***IDENTIFICATION OF LONG NON-CODING RNAs IN SERUM
EXOSOMES FROM COLORECTAL CANCER PATIENTS AND
EFFECTS OF EXOSOMES ON TUBE FORMATION BY ENDOTHELIAL
CELLS***

NG CHIN TAT

FPSK(p) 2021 39



**IDENTIFICATION OF LONG NON-CODING RNAS IN SERUM
EXOSOMES FROM COLORECTAL CANCER PATIENTS AND EFFECTS
OF EXOSOMES ON TUBE FORMATION BY ENDOTHELIAL CELLS**

By

NG CHIN TAT

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

September 2019

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

**IDENTIFICATION OF LONG NON-CODING RNAS IN SERUM
EXOSOMES FROM COLORECTAL CANCER PATIENTS AND EFFECTS
OF EXOSOMES ON TUBE FORMATION BY ENDOTHELIAL CELLS**

By

NG CHIN TAT

September 2019

Chairman : Prof.essor Seow Heng Fong, PhD
Faculty : Medicine and Health Sciences

Extracellular vesicles (exosome-like vesicles) are small membrane vesicles ranging from 20-150nm in size that are released by various cells into the extracellular space. This extracellular vesicles play a major role in cell-to-cell communication and contain materials, such as proteins, mRNAs, microRNAs (miRNAs) and long non-coding RNAs (lncRNAs). Recent studies have reported that the components in exosomes vary based on the type of cell. Long non-coding RNAs (lncRNAs) are non-protein-coding RNAs consisting of more than 200 nucleotides in length. It has been shown that lncRNAs play an important role in various cellular processes including angiogenesis by interacting with RNA, DNA or proteins through diverse mechanisms to regulate gene expression. However, the effect of exosomes derived from an invasive colorectal cancer cell line on angiogenesis is unclear and the potential of lncRNAs in circulating exosomes as biomarkers have not been completely studied. Hence, the aims of this study is to investigate the effect of exosomes derived from an invasive colorectal cancer cell line on angiogenesis of endothelial cells and to identify the potential of lncRNAs in circulating exosomes as biomarkers for detection of colorectal cancer. In the present study, the exosomes from the cell culture supernatants of an invasive colorectal cancer cell line SW480-7 were characterized and lncRNAs in these exosomes were profiled. The effect on tube formation and expression of angiogenic genes in a microvascular endothelial cell, telomerase-immortalized microvascular endothelial cell (TIME) cocultured with exosomes were also determined. In addition, RT² lncRNA PCR array was used for exosomal lncRNA profiling to determine the relative expression level of lncRNAs in the exosomes of sera from 18 CRC (colorectal carcinoma) and 21 non-cancer patients. The expression level of lncRNAs was compared between 8 early-stage (stages I and II) and 10 advanced-stage (stages III and IV) of CRC patients. Results showed that exosomes derived from SW480-7 increased tube formation and up-regulated expression FGFR3 mRNA in TIME. Zetasizer result showed average diameter of exosomes derived from SW480-7 was

274.6 nm and morphology analysis showed the majority of exosomes size less than 200 nm. Western blot analysis demonstrated that exosomes from SW480-7 contained exosomal protein, Alix. A total number of 32 lncRNAs were detected in SW480-7-derived exosomes and some have been reported to be associated with angiogenesis. In clinical studies, six lncRNAs, namely GAS5, H19, LINC00152, SNHG16, RMRP, and ZFAS1 were detected in the sera of 18 CRC patients. These lncRNAs may serve as potential biomarkers for detection of CRC. Among these six lncRNAs, expression level of LINC00152 was found to be significantly lower in CRC patients as compared to non-cancer patients ($p=0.04$). The expression level of lncRNAs was compared for early versus advanced-stage of CRC patients. Expression level of exosomal lncRNA H19 was significantly up-regulated in advanced-stage of CRC (p value= 0.04). In conclusion, exosomal lncRNAs derived from SW480-7 increased tube formation and up-regulated expression of FGFR3 mRNA in TIME. Among the six exosomal lncRNAs detected in the sera, LINC00152 and H19 may be useful as biomarkers in liquid biopsies for diagnosis and treatment monitoring in colorectal cancer.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**IDENTIFIKASI LONG NON-CODING RNA DI DALAM SERUM EKSOSOM
DARIPADA PESAKIT KANSER KOLOREKTAL DAN KESAN EKSOSOM
TERHADAP FORMASI TIUB SEL ENDOTELIAL**

Oleh

NG CHIN TAT

September 2019

Pengerusi : Profesor Seow Heng Fong, PhD
Fakulti : Perubatan dan Sains Kesihatan

Vesikel ekstraselular (vesikel menyerupai eksosom) adalah vesikel membran kecil, saiz di antara 20-150 nm dan dirembeskan oleh pelbagai jenis sel ke dalam ruang ekstraselular. Vesikel ekstraselular ini memainkan peranan penting dalam komunikasi di antara sel-sel dan mengandungi komponen seperti protein, mRNAs, microRNAs (miRNAs) dan long non-coding RNAs (lncRNAs). Kebelakangan ini, banyak kajian melaporkan bahawa komponen dalam eksosom adalah berbeza dan bergantung kepada jenis sel yang merembesnya. LncRNAs adalah non-protein-coding RNAs (RNA yang tidak terlibat dalam pengekodan protein) yang melebihi 200 nukleotida. Kajian telah membuktikan lncRNAs memainkan peranan penting dalam pelbagai proses sel termasuk angiogenesis (pembentukan pembuluh darah baru) yang melibatkan interaksi di antara RNA, DNA atau protein melalui pelbagai mekanisme yang mengawal ekspresi gen. Walau bagaimanapun, kesan eksosom dari sel kanser kolorektal yang invasif terhadap angiogenesis dan potensi lncRNAs di dalam eksosom sebagai biopenanda kanser kolorektal (CRC) masih belum ditentukan. Oleh itu, tujuan penyelidikan ini adalah untuk mengkaji kesan eksosom dari sel kanser kolorektal yang invasif terhadap angiogenesis sel endotelial dan potensi lncRNAs sebagai biopenanda kanser kolorektal. Dalam kajian ini, ciri-ciri eksosom ditentukan dan lncRNAs daripada eksosom sel kanser kolorektum SW480-7 diprofilkan. Kesan eksosom terhadap formasi tiub dan ekspresi angiogenik gen dalam sel endotelial mikrovaskular, *telomerase-immortalized microvascular endothelial cell* (TIME) juga dikaji. Selain itu, RT² lncRNA PCR array digunakan untuk pemprofilan lncRNA di dalam eksosom untuk menentukan tahap ekspresi lncRNAs dari 18 pesakit kanser kolorektal dan 21 pesakit bukan kanser kolorektal. Tahap ekspresi lncRNA dibandingkan antara 8 pesakit CRC yang berperingkat awal (peringkat I dan II) dengan 10 pesakit CRC yang berperingkat lanjut (peringkat III dan IV). Hasil kajian menunjukkan bahawa eksosom yang berasal dari SW480-7 meningkatkan formasi tiub dan meningkatkan ekspresi *FGFR3* mRNA di dalam sel TIME. Keputusan *Zetasizer* menunjukkan purata

diameter eksosom yang berasal dari SW480-7 ialah 274.6 nm dan analisis morfologi menunjukkan majoriti saiz eksosom kurang daripada 200 nm. Analisis *Western Blot* menunjukkan bahawa eksosom yang berasal dari SW480-7 mengandungi protein eksosom, *Alix*. Sebanyak 32 lncRNAs dikesan di dalam eksosom yang berasal dari SW480-7 termasuk yang telah dilaporkan yang terlibat dengan angiogenesis. Dalam kajian klinikal, enam lncRNAs iaitu GAS5, H19, LINC00152, SNHG16, RMRP dan ZFAS1 dikesan di dalam serum eksosom daripada 18 pesakit CRC. LncRNA ini berpotensi sebagai biopenanda dalam pengesanan CRC. Di antara enam lncRNAs ini, tahap ekspresi LINC00152 adalah lebih rendah di dalam pesakit CRC berbanding dengan pesakit bukan CRC ($p = 0.04$). Tahap ekspresi lncRNA dibandingkan antara pesakit CRC peringkat awal dengan pesakit CRC peringkat lanjut. Tahap ekspresi lncRNA H19 di dalam eksosom meningkat dengan ketara bagi pesakit CRC peringkat lanjut (nilai $p = 0.04$). Kesimpulannya, lncRNA di dalam eksosom yang berasal dari SW480-7 meningkatkan formasi tiub dan meningkatkan ekspresi mRNA FGFR3 dalam TIME. Antara 6 lncRNAs di dalam serum eksosom yang dikesan, LINC00152 dan H19 mungkin berguna sebagai biopenanda dalam biopsi cecair untuk diagnosis dan rawatan pemantauan bagi kanser kolorektal.

ACKNOWLEDGEMENTS

I would like to express my profoundest appreciation to my supervisor, Prof. Dr. Seow Heng Fong for her guidance, motivation, knowledge and endless support throughout this study. Her words can always inspire me and lead me to a higher level of thinking. Without her guidance and persistent help, it is impossible for me to complete this thesis. Also, special thanks to my co-supervisors, Assoc. Prof. Dr. Mohd Faisal Jabar, Assoc. Prof. Dr. Norhafizah Mohtarrudin and Dr. Norren Haneezah Binti Sahak for their valuable advice, assistance and review for my work. They offer their unreserved help and suggestion to guide me step by step throughout my work.

My deepest appreciation goes to my fellow lab mates for their helpfulness and encouragement. I would like to express my gratitude to the staff of Immunology Laboratory for their kind assistance and support.

Finally, and most importantly, I would like to thank my wife, Cheong Fui Ying for her endless support, encouragement, understanding, patience and love. I am also deeply grateful to my parents for supporting me throughout all my studies at University Putra Malaysia.

This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Seow Heng Fong, PhD

Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Chairman)

Norhafizah binti Mohtarrudin, PhD

Associate Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Member)

Mohd Faisal bin Jabar, PhD

Associate Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 09 December 2021

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature: _____

Date: _____

Name and Matric No.: Ng Chin Tat, GS42735

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: _____
Name of Chairman
of Supervisory Committee: Professor
Dr. Seow Heng Fong

Signature: _____
Name of Member
of Supervisory Committee: Associate Professor
Dr. Norhafizah binti Mohtarrudin

Signature: _____
Name of Member
of Supervisory Committee: Associate Professor
Dr. Mohd Faisal bin Jabar

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiv
LIST OF FIGURES	xv
LIST OF APPENDICES	xvii
LIST OF ABBREVIATIONS	xviii
 CHAPTER	
1 INTRODUCTION	1
1.1 Background of Study	1
1.2 Objectives	2
2 LITERATURE REVIEW	3
2.1 Colorectal cancer (CRC)	3
2.1.1 Diagnostic and therapeutic challenges in CRC	6
2.1.2 Migration, invasion and metastasis	6
2.2 Long non-coding RNA	8
2.2.1 Biogenesis of lncRNAs	8
2.2.2 Classification and characterization of lncRNAs in colorectal cancer	9
2.3 The mechanism action of lncRNAs in colorectal cancer	10
2.3.1 Epigenetic modifications	11
2.3.2 LncRNA-miRNA interaction	11
2.3.3 LncRNA-protein interaction	11
2.3.4 LncRNAs act as pseudogenes or miRNA precursors	11
2.4 Pathological function of lncRNAs in tumourigenesis of CRC	11
2.5 LncRNAs as diagnostic and prognostic disease biomarkers	13
2.5.1 Oncogenic lncRNAs in colorectal cancer	13
2.5.1.1 Colon cancer-associated transcript - CCAT	13
2.5.1.2 Colorectal neoplasia differentially expressed –CRNDE	14
2.5.1.3 Colorectal liver metastasis-associated transcript 3 –CLMAT3	14
2.5.1.4 Differentiation antagonizing non-protein coding RNA-DANCR	14
2.5.1.5 Five prime to Xist -FTX	14

2.5.1.6	HOXA distal transcript antisense RNA - HOTTIP	15
2.5.1.7	HOX transcript Antisense intergenic RNA -HOTAIR	15
2.5.1.8	Human universal load carrier-HULC	15
2.5.1.9	Long non-coding RNA activated by transforming growth factor- β - lncRNA-ATB	15
2.5.1.10	Metastasis-associated lung adenocarcinoma transcript 1-MALAT1	16
2.5.1.11	Prostate cancer-associated ncRNA transcript 1-PCAT-1	16
2.5.1.12	Plasmacytoma variant translocation 1-PVT-1	16
2.5.1.13	Urothelial cancer associated 1 -UCA-1	17
2.5.1.14	SBDSP1	17
2.5.2	Tumour suppressive lncRNAs in colorectal cancer	17
2.5.2.1	Growth arrest specific 5 -GAS5	17
2.5.2.2	Maternally expressed gene 3 -MEG3	18
2.5.2.3	Non-coding RNA expressed in aggressive neuroblastoma -NcRAN	18
2.5.2.4	Non-protein coding RNA, upstream of F2R/PAR1- ncRuPAR	18
2.5.2.5	lncRNA-RP11-462.C24.1	18
2.5.2.6	Tumor Suppressor Candidate 7- TUSC7	18
2.5.3	Dual function of lncRNAs in colorectal cancer	19
2.6	Exosomes	20
2.6.1	Biogenesis of exosomes	20
2.6.2	Regulation of angiogenesis by exosomal lncRNAs	21
2.6.3	Roles of exosomes as potential diagnosis and prognosis marker	22
2.7	Potential applications of circulating lncRNA for diagnostic and therapeutic of CRC	23
3	MATERIALS AND METHODS	25
3.1	Flow chart of project activities	25
3.2	Reagent Preparation	26
3.2.1	RPMI Medium	26
3.2.2	Endothelial Cell Basal Medium	26
3.3	Cell Line and cell cultures	26
3.4	Isolation of exosomes	27
3.4.1	Isolation of exosomes derived from supernatant of SW480-7	27
3.4.2	Reconstitution of standard exosomes	28
3.5	Exosomal protein determination by BCA protein assay kit	28
3.6	Characterization of exosomes derived from SW480-7	28
3.6.1	Size distribution analysis of exosomes by Zetasizer	28
3.6.2	Determination of exosome morphology by transmission electron microscope (TEM)	28

3.6.3	Detection of Axil in exosomes by Western blotting	29
3.7	Selection of exosome-depleted FBS percentage for tube formation assay	30
3.7.1	Cell seeding and Pre-treatment	30
3.7.2	Preparation of Growth Factor-Reduced BD Matrigel Matrix	30
3.7.3	Coating of μ -Slide Angiogenesis with Growth Factor-Reduced BD Matrigel	30
3.7.4	Cell Harvesting and Seeding in μ -Slide Angiogenesis	30
3.8	Endothelial Tube Formation Assay	31
3.8.1	Cell Seeding and Pre-Treatment	31
3.8.2	Preparation of Growth Factor-Reduced BD Matrigel Matrix	31
3.8.3	Coating of μ -Slide Angiogenesis with Growth Factor-Reduced BD Matrigel	31
3.8.4	Cell Harvesting and Seeding in μ -Slide Angiogenesis	31
3.8.5	Quantification of Capillary-Like Tube Formation	32
3.9	Effect of SW480-7 exosomes on expression of angiogenic genes in TIME	32
3.9.1	Cells seeding and pre-treatment	32
3.9.2	RNA extraction	32
3.9.3	Reverse transcription	33
3.9.4	RT ² Angiogenesis PCR Array	34
3.10	LncRNA RT ² Profiler PCR Array in clinical samples	35
3.10.1	Flow chart of exosomal lncRNAs work	35
3.10.2	Serum, reagent and equipment	35
3.10.3	Serum exosomal RNA preparation	36
3.10.4	RNA isolation from serum exosomes	36
3.10.4.1	Vesicle isolation	36
3.10.4.2	RNA isolation	37
3.10.5	Reverse transcription of lncRNA	38
3.10.6	Preamplification of cDNA Target Templates	39
3.10.7	Detection and profiling of lncRNA in exosomes by RT-PCR	39
3.11	Statistical Analysis	40
4	RESULTS	41
4.1	Particle size distribution and protein detection with SW480-7 derived exosomes	41
4.2	Morphology Characterization of Exosomes by TEM	43
4.2.1	Morphological Characterization of Exosomes derived from SW480	43
4.2.2	Morphological Characterization of Exosomes derived from SW480-7	45
4.2.3	Morphological Characterization of Exosomes derived from serum sample	46
4.3	Detection of exosomal protein derived from SW480-7	49

4.4	Selection of a suitable concentration of exosome-depleted FBS for low (minimal) tube formation by TIME	49
4.5	Effect of exosomes derived from SW480-7 on tube formation by TIME	52
4.6	Effect of exosome derived from cell culture supernatants of SW480-7 on FGFR3 mRNA expression in TIME.	55
4.7	Detection of lncRNAs derived from SW480-7 cell line by RT ² lncRNAs PCR array	57
4.8	Detection of lncRNAs derived from serum sample by RT ² lncRNAs PCR array	60
4.9	Selection of lncRNAs and reference gene from serum samples	62
4.10	Detection of serum exosomal lncRNAs expression in CRC patients and non-cancer patients	63
4.11	Detection of exosomal lncRNAs expression in early and advanced stage of CRC patients	66
5	DISCUSSION	68
5.1	Characterization of exosomes derived from SW480-7	68
5.2	Effect of exosomes derived from SW480-7 in telomerase-immortalized microvascular endothelial cell	69
5.3	Expression of exosomal lncRNAs from serum of CRC patients and non-cancer patients	71
6	CONCLUSION	76
6.1	Conclusion	76
6.2	Future perspectives	76
	REFERENCES	78
	APPENDICES	93
	BIODATA OF STUDENT	109
	PUBLICATION	110

LIST OF TABLES

Table	Page
2.1 Tumour node metastasis clinical classification	5
2.2 Stage grouping of CRC	6
2.3 Biological functions of lncRNAs in colorectal cancer	12
2.4 Role of tumour-derived lncRNAs in endothelial cells	22
3.1 Genomic DNA elimination mix	33
3.2 Reverse transcription mix	33
3.3 PCR array components mix	34
3.4 Thermal cycle programming	38
3.5 Pre-amplification mix	39
3.6 Real-time cycler program	39
3.7 PCR array component mix	40
4.1 Expression of exosomal long non-coding RNAs derived from SW480-7 by RT2 PCR array	58
4.2 Clinical and pathological characteristics of CRC patients included in this study	60
4.3 Detection of serum exosomal lncRNAs by RT 2 lncRNA PCR Array	61
4.4 Average Ct values (< 30) of long non-coding RNAs expressed in serum exosomes from colorectal cancer patients and non-cancer patients	62
4.5 The fold change of long non-coding RNAs, GAS5, H19, LINC00152, RMRP, SNHG16 and ZFAS1 expression level in non-cancer patients (non-cancer), early stage (stage I &II) and advanced stage (stage III & IV) of colorectal cancer patients	64
4.6 The fold change of lncRNAs, GAS5, H19, LINC00152, RMRP, SNHG16 and ZFAS1 expression level in early stage (stage I &II) and advanced stage (stage III & IV) of CRC patients	66

LIST OF FIGURES

Figure	Page
2.1 Staging of colorectal cancer	4
2.2 Metastatic cascade. Metastasis is a multistep process	7
2.3 Biogenesis and Classification of lncRNAs	8
2.4 Categories of lncRNAs	9
2.5 Mechanisms of lncRNAs function in colorectal cancer	10
2.6 Potential prognostic marker of lncRNAs in human colorectal cancer	13
2.7 Biogenesis and classification of lncRNA in human and other animals	20
2.8 Exosomal lncRNAs in angiogenesis of endothelial cell	21
3.1 Flow chart of project activities	25
3.2 Workflow of exosomal lncRNAs profiling from serum samples and cell culture supernatant	35
3.3 Workflow for isolating RNA from extracellular vesicles using membrane affinity columns	36
4.1 Size distribution of exosomes	42
4.2 Exosomes derived from SW480 (commercially available exosome) were stained using 1 % (w/v) uranyl acetate and viewed by transmission electron microscopy	44
4.3 Exosomes derived from SW480-7 (cell culture supernatant) were stained using 1 % (w/v) uranyl acetate and viewed by transmission electron microscopy	45
4.4 Exosomes derived from serum samples from Hospital Serdang were stained using 1 % (w/v) uranyl acetate and viewed by transmission electron microscopy	47
4.5 Exosomes derived from serum samples from Hospital Serdang were stained using 1 % (w/v) uranyl acetate and viewed by transmission electron microscopy	48
4.6 Western blot analysis of exosomal protein Alix. Exosomal protein derived from SW480-7 culture medium	49

4.7	Effect of various concentrations of exosomes-depleted FBS on Capillary Tube Formation by Telomerase Immortalized Microvascular Endothelial Cells (TIME)	51
4.8	Effect of Exosomes Derived from SW480-7 on Capillary Tube Formation in Cultured Telomerase Immortalized Microvascular Endothelial Cells (TIME)	53
4.9	Quantification of Capillary-like Tube Formation by TIME	54
4.10	FGFR3 mRNA expression in TIME cell line was analyzed by qRT-PCR	56
4.11	Cycle threshold values of long non-coding RNAs expressed in exosomes derived from cell culture supernatants of SW480-7	59
4.12	Average Ct value of target genes from 84-genes in the array	63
4.13	Comparison of lncRNAs expression level ($2^{(-\Delta CT)}$) in serum exosomes from CRC and non-cancer patients	65
4.14	Comparison of lncRNAs expression level ($2^{(-\Delta CT)}$) in serum exosomes from early (I and II) and advanced (III and IV) stages of CRC patients	67

LIST OF APPENDICES

Appendix		Page
A	Laemmli resolving gel, 29:1 ratio	93
B	Details of Clinical Pathology Data for Cancer Patients and Non-Cancer Patients	95
C	Details of Clinical Pathology Data and CEA Level for Cancer Patients	98
D	Data of Sera Volume, RNA Yield in Concentration and Total Volume of Purified RNA Obtained for Cancer Patients and Non-Cancer Patients	100
E	Normalized gene expression level of lncRNAs for early versus advanced-stage of CRC patients	101
F	Normalized gene expression level of lncRNAs for non-cancer individuals versus CRC patients	102
G	Normalized gene expression level of lncRNAs for early versus advanced-stage of CRC patients	104
H	Normalized gene expression level of lncRNAs for non-cancer individuals versus CRC patients	107

LIST OF ABBREVIATIONS

%	Percent
μ	Micro
AEG-1	Astrocyte elevated gene-1
AKAP-9	A-kinase anchor protein 9
ANGPT2	Angiopoietin 2
BCAR4	Breast cancer anti-estrogen resistance 4
bFGF	Basic fibroblast growth factor
bFGFR	Basic fibroblast growth factor receptor
BLACAT1	Bladder cancer associated transcript 1
BSA	Bovine serum albumin
CA19-9	Carbohydrate antigen 19-9
CA242	Carbohydrate antigen 242
CBR3-AS1	Cbr3 antisense RNA 1
CCAT1	Colon cancer associated transcript 1
CCAT1-L	Colorectal cancer-associated transcript 1-long
CCAT2	Colon cancer-associated transcript-2
CEA	Carcinoembryonic antigen
ceRNAs	Competing endogenous RNAs
CLMAT3	Colorectal liver metastasis-associated transcript 3
CRNDE	Colorectal neoplasia differentially expressed
CRNDE-h	Colorectal neoplasia differentially expressed – h
DANCR	Differentiation antagonizing non-protein coding RNA
DLEU2	Human leukemia-associated protein 2
DNMT1	Dna methyltransferase 1

DNMT3A	Dna methyltransferase 3a
DNMT3B	Dna methyltransferase 3b
ECM	Extracellular matrix
EMT	Epithelial–mesenchymal transition
ESCRT	Endosomal sorting complexes required for transport
EZH2	Enhancer of zeste homolog 2
FGF2	Fibroblast growth factor 2
FGFR3	Fibroblast growth factor receptor 3
FTX	Five prime to xist
GAS5	Growth arrest-specific 5
GDF15	Growth differentiation factor 15
HCC	Hepatocellular carcinoma
hEGF	Human recombinant epidermal growth factor
HEIH	Hepatocellular carcinoma up-regulated ezh2-associated long non-coding RNA
hFGF-B	Human fibroblast growth factor basic with heparin
HOTAIR	Hox transcript antisense RNA
HOTAIRM1	Hoxa transcript antisense RNA, myeloid-specific 1
HOTTIP	Hoxa transcript at the distal tip
HULC	Highly upregulated in liver cancer
JAK-STAT	Janus kinase/signal transducer and activator of transcription
LINC00152	Long intergenic non-coding RNA 00152
LINC00261	Long intergenic non-protein coding RNA 261
LINC00312	Long intergenic non-protein coding RNA 321
LINC00963	Long intergenic non-protein coding RNA 963
LINC01234	Long intergenic non-protein coding RNA 1234

LINC01296	Long intergenic non-coding RNA 1296
lincRNAs	Large intervening non-coding RNAs
lncRNA-ATB	LncRNA activated by $\text{tgf-}\beta$
lncRNAs	Long non-coding RNAs
LSD1	Lysine-specific demethylase 1
MALAT1	Metastasis-associated lung adenocarcinoma transcript 1
MDM2	Murine double minute 2
MEG3	Maternally expressed gene 3
miR	MicroRNA
MIR17HG	Mir-17-92 cluster host gene
MIR31HG	Mir31 host gene
miRNAs	MicroRNAs
MMP	Matrix metalloproteinase
mRNAs	Messenger RNAs
MTX	Methotrexate
MVB	Multivesicular body
MYC	Myelocytoma
NaHCO ₃	Sodium bicarbonate
NBR2	Neighbor of brca1 gene 2
ncRNA	Non-protein ribonucleic RNA
NEAT1	Nuclear paraspeckle assembly transcript 1
OXA	Oxaliplatin
OS	Overall survival
PAR-1	Protease-activated receptor-1
PBS	Phosphate buffered saline
PCA3	Prostate cancer antigen 3

PCAT1	Prostate cancer-associated transcript 1
piRNAs	Piwi-interacting RNAs
POU3F3	Pou class 3 homeobox 3
PRNCR1	Prostate cancer associated non-coding RNA 1
PTBP2	Polypyrimidine tract binding protein 2
PTENP1	Phosphatase and tensin homolog pseudogene 1
PVDF	Polyvinylidene fluoride
PVT1	Plasmacytoma variant translocation 1
R3-IGF-1	Human recombinant insulin-like growth factor
REST	Re1 silencing transcription factor
RIPA	Radioimmune precipitation assays
RMRP	RNA component of mitochondrial RNA-processing
RMRP	RNA component of mitochondrial RNA processing endoribonuclease
RN7SK	RNA, 7sk small nuclear
RPLP0	Ribosomal proteins ribosomal protein, large, p0
SBDSP1	Shwachman-bodian-diamond syndrome pseudogene 1
SCID	Severe combined immunodeficiency disease
SFPQ	Splicing factor proline and glutamine rich
shRNA	Small hairpin RNA
siRNAs	Small interfering RNAs
SNHG 16	Small nucleolar RNA host genes 16
SNORA73A	Small nucleolar RNA, h/aca box 73a
SNP	Single nucleotide polymorphism
SPIA	Single primer isothermal amplification
SRPK1	Serine-arginine protein kinase 1

TCF7L2	Transcription factor 7-like 2
TEM	Transmission electron microscopy
TERC	Telomerase RNA component
TGF- β	Transforming growth factor beta
TNM	Tumour node metastasis
TUG1	Taurine up-regulated gene 1
TUSC7	Tumor suppressor candidate 7
UCA1	Urothelial carcinoma-associated 1
VDR	Vitamin d receptor
VEGF	Vascular endothelial growth factor
VEGFA	Vascular endothelial growth factor a
VEGFR	Vascular endothelial growth factor receptor
ZEB	Zinc finger e-box-binding protein
ZFAS1	Zinc finger antisense 1

CHAPTER 1

INTRODUCTION

1.1 Background of Study

Colorectal cancer (CRC) is the third most common type of cancer (Moriarty et al., 2016) that causes high mortality worldwide (Arnold et al., 2017). Despite decades of research, the mortality rate of CRC patients is still high. In 2018, CRC was estimated to cause 551,269 deaths (5.8% of total cancer death) and about 1.09 million (6.1% of total new cases) new cases were reported at the same year from 185 countries (Bray et al., 2018). Most previous genomic studies in the pathogenesis of CRC are mainly about protein-coding genes (Xie et al., 2016). Surprisingly, current genomic studies via latest next generation sequencing methods and transcriptome analysis have discovered that larger part of genome is transcribe into RNA including non-protein ribonucleic RNA (ncRNA)(Saus et al., 2016). These ncRNAs encompass a variety of subclasses. One of the subclasses known as regulatory ncRNA consists of two main groups namely short RNAs (miRNAs siRNAs and piRNAs) and long non-coding RNA (lncRNAs).

lncRNAs are transcripts more than 200 nucleotides (Han et al., 2015) in length with no or limited protein-coding potential and are found in significant amounts in exosomes. In the past, lncRNAs were described as transcriptional ‘noise’ (Raut and Khullar, 2018; Ward et al., 2015) but recent studies have demonstrated that lncRNAs play an important role in central cellular process such as epigenetic modulation, transcription and translation. When compared with protein-coding genes, lncRNAs are tissue-specific although expressed at low levels (Balas and Johnson, 2018; Ward et al., 2015). In CRC, lncRNAs have been shown to play significant roles in carcinogenesis via various mechanisms such as cell proliferation (Smolle et al., 2014; Ye et al., 2016; Yin et al., 2015), apoptosis (Takahashi et al., 2014), migration (Qi et al., 2015; Smolle et al., 2014), invasion and metastasis (Smolle et al., 2014). Exosomes are extracellular vesicles ranging between 20 nm – 120 nm in size and contain significant amount of nucleic acids including lncRNAs. These vesicles are secreted into the extracellular space by various cells including cancer cells. Cancer derived exosomes are able to trigger the angiogenic process and enhance metastasis of tumour. Exosomes can be used as liquid biomarker for various cancer diagnosis as exosomal contents are originated from parental cells (Li et al., 2017d; Roma-Rodrigues et al., 2014). Currently, the screening tests are inadequate and factors such as false positive or negative results, cumbersome preparatory procedures and expensive molecular testing are major obstacles to early detection of CRC. Hence, there is a dire need for development of a diagnostic test for cancer detection. Exosomes carried biomarkers specific to the origin of cancer cells (Roma-Rodrigues et al., 2014) that are present in serum and their profiles may potentially be useful as a novel biomarkers for CRC patients and are emerging as a new diagnostic approach.

To date, no circulating exosomal marker in serum has been developed for use in CRC as of yet. Recently circulating miRNAs or exosomal miRNAs have been recognized as promising biomarkers for ovarian cancer (Yokoi et al., 2017), lung cancer (Farran et al., 2018; Yu et al., 2018), colon cancer (Fang et al., 2015; Zhang et al., 2018; Zheng et al., 2014a), prostate cancer (Sapre and Selth, 2013) and breast cancer (Hamam et al., 2017). It is likely that circulating exosomes in body fluids containing lncRNAs may present new, relatively non-invasive cancer biomarkers. The results will enable the development of diagnostic and prognostic tests (Saus et al., 2016) for non-invasive and early detection of CRC. Hence, the aims of this study was to isolate the exosomal lncRNAs in sera from apparently non-cancer patients and patients with colorectal adenoma and carcinoma, examine the lncRNA expression profile and identify the differentially expressed serum lncRNAs related to the colorectal carcinogenesis. Recently, many studies revealed the effect of exosomes derived from various cancer cells on angiogenesis. However, the effect of exosomes from invasive colorectal cancer cell line on angiogenesis is unknown. Hence, this study also determine the effects of invasive colorectal cancer cell line (SW480-7) derived exosomes on angiogenesis and determine angiogenic gene expression in telomerase-immortalized microvascular endothelial cells (TIME). SW480-7 is an invasive subpopulation of SW480 that was established *via* 7 sequential passages through matrigel-coated transwells. The hypothesis of the present study is that, there are differences in lncRNA expression between non-cancer patients and patients with colorectal adenoma and carcinoma. For the *in vitro* study, it is hypothesized that exosomes derived from SW480-7 will increase tube formation of TIME, and up-regulate angiogenic gene expression.

1.2 Objectives

- a. To purify and characterize the exosomes secreted by SW480-7 colon cancer cell line.
- b. To identify lncRNAs in exosomes secreted by SW480-7.
- c. To determine the effects of SW480-7 derived-exosomes on angiogenesis.
- d. To isolate and identify the exosomal lncRNAs in sera from non-cancer patients and patients with colorectal adenoma and carcinoma.
- e. To examine the profile of lncRNA expression and identify the differentially expressed serum exosomal lncRNAs related to stages of colorectal carcinoma.

REFERENCES

- Alaiyan, B., N. Ilyayev, A. Stojadinovic, M. Izadjoo, M. Roistacher, V. Pavlov, V. Tzivin, D. Halle, H. Pan, B. Trink, A.O. Gure, and A. Nissan. 2013. Differential expression of colon cancer associated transcript1 (CCAT1) along the colonic adenoma-carcinoma sequence. *BMC Cancer*. 13:196.
- Arnold, M., M.S. Sierra, M. Laversanne, I. Soerjomataram, A. Jemal, and F. Bray. 2017. Global patterns and trends in colorectal cancer incidence and mortality. *Gut*. 66:683-691.
- Balas, M.M., and A.M. Johnson. 2018. Exploring the mechanisms behind long noncoding RNAs and cancer. *Non-coding RNA Research*. 3:108-117.
- Barbagallo, C., D. Brex, A. Caponnetto, M. Cirnigliaro, M. Scalia, A. Magnano, R. Caltabiano, D. Barbagallo, A. Biondi, A. Cappellani, F. Basile, C. Di Pietro, M. Purrello, and M. Ragusa. 2018. LncRNA UCA1, Upregulated in CRC Biopsies and Downregulated in Serum Exosomes, Controls mRNA Expression by RNA-RNA Interactions. *Molecular Therapy. Nucleic Acids*. 12:229-241.
- Benetatos, L., G. Vartholomatos, and E. Hatzimichael. 2011. MEG3 imprinted gene contribution in tumorigenesis. *International Journal of Cancer*. 129:773-779.
- Bouckenheimer, J., S. Assou, S. Riquier, C. Hou, N. Philippe, C. Sansac, T. Lavabre-Bertrand, T. Commes, J.-M. Lemaître, A. Boureux, and J. De Vos. 2016. Long non-coding RNAs in human early embryonic development and their potential in ART. *Human Reproduction Update*. 23:19-40.
- Brooks, S.A., H.J. Lomax-Browne, T.M. Carter, C.E. Kinch, and D.M. Hall. 2010. Molecular interactions in cancer cell metastasis. *Acta Histochemica*. 112:3-25.
- Cao, H.L., Z.J. Liu, P.L. Huang, Y.L. Yue, and J.N. Xi. 2019. lncRNA-RMRP promotes proliferation, migration and invasion of bladder cancer via miR-206. *European Review for Medical and Pharmacological Sciences*. 23:1012-1021.
- Cao, X., S. Zhuang, Y. Hu, L. Xi, L. Deng, H. Sheng, and W. Shen. 2016. Associations between polymorphisms of long non-coding RNA MEG3 and risk of colorectal cancer in Chinese. *Oncotarget*. 7:19054-19059.
- Carramusa, L., F. Contino, A. Ferro, L. Minafra, G. Perconti, A. Giallongo, and S. Feo. 2007. The PVT-1 oncogene is a Myc protein target that is overexpressed in transformed cells. *Journal of Cellular Physiology*. 213:511-518.

- Charoenviriyakul, C., Y. Takahashi, M. Morishita, A. Matsumoto, M. Nishikawa, and Y. Takakura. 2017. Cell type-specific and common characteristics of exosomes derived from mouse cell lines: Yield, physicochemical properties, and pharmacokinetics. *European Journal of Pharmaceutical Sciences : Official Journal of the European Federation for Pharmaceutical Sciences*. 96:316-322.
- Chen, L., W. Yang, Y. Guo, W. Chen, P. Zheng, J. Zeng, and W. Tong. 2017a. Exosomal lncRNA GAS5 regulates the apoptosis of macrophages and vascular endothelial cells in atherosclerosis. *PLoS one*. 12:e0185406.
- Chen, S., D. Bu, Y. Ma, J. Zhu, G. Chen, L. Sun, S. Zuo, T. Li, Y. Pan, X. Wang, Y. Liu, and P. Wang. 2017b. H19 Overexpression Induces Resistance to 1,25(OH)2D3 by Targeting VDR Through miR-675-5p in Colon Cancer Cells. *Neoplasia (New York, N.Y.)*. 19:226-236.
- Chen, X., K. Zeng, M. Xu, X. Hu, X. Liu, T. Xu, B. He, Y. Pan, H. Sun, and S. Wang. 2018. SP1-induced lncRNA-ZFAS1 contributes to colorectal cancer progression via the miR-150-5p/VEGFA axis. *Cell Death & Disease*. 9:982.
- Christensen, L.L., K. True, M.P. Hamilton, M.M. Nielsen, N.D. Damas, C.K. Damgaard, H. Ongen, E. Dermitzakis, J.B. Bramsen, J.S. Pedersen, A.H. Lund, S. Vang, K. Stribolt, M.R. Madsen, S. Laurberg, S.E. McGuire, T.F. Orntoft, and C.L. Andersen. 2016. SNHG16 is regulated by the Wnt pathway in colorectal cancer and affects genes involved in lipid metabolism. *Molecular Oncology*. 10:1266-1282.
- Cui, M., L. You, X. Ren, W. Zhao, Q. Liao, and Y. Zhao. 2016. Long non-coding RNA PVT1 and cancer. *Biochemical and Biophysical Research Communications*. 471:10-14.
- Delilhas, N. 2013. Editorial on the Special Issue: Regulation by non-coding RNAs. *International Journal of Molecular Sciences*. 14:21960-21964.
- Deng, H., J.M. Wang, M. Li, R. Tang, K. Tang, Y. Su, Y. Hou, and J. Zhang. 2017. Long non-coding RNAs: New biomarkers for prognosis and diagnosis of colon cancer. *Tumour biology : the journal of the International Society for Oncodevelopmental Biology and Medicine*. 39:1010428317706332.
- Deng, Q., B. He, T. Gao, Y. Pan, H. Sun, Y. Xu, R. Li, H. Ying, F. Wang, X. Liu, J. Chen, and S. Wang. 2014. Up-regulation of 91H promotes tumor metastasis and predicts poor prognosis for patients with colorectal cancer. *PLoS one*. 9:e103022.
- Dhanoa, J.K., R.S. Sethi, R. Verma, J.S. Arora, and C.S. Mukhopadhyay. 2018. Long non-coding RNA: its evolutionary relics and biological implications in mammals: a review. *Journal of Animal Science and Technology*. 60:25.

- Dragomir, M., B. Chen, and G.A. Calin. 2018. Exosomal lncRNAs as new players in cell-to-cell communication. *Translational Cancer Research*. 7:S243-s252.
- Enderle, D., A. Spiel, C.M. Coticchia, E. Berghoff, R. Mueller, M. Schlumpberger, M. Sprenger-Haussels, J.M. Shaffer, E. Lader, J. Skog, and M. Noerholm. 2015. Characterization of RNA from Exosomes and Other Extracellular Vesicles Isolated by a Novel Spin Column-Based Method. *PLoS one*. 10:e0136133.
- Fan, S., C. Fan, N. Liu, K. Huang, X. Fang, and K. Wang. 2018. Downregulation of the long non-coding RNA ZFAS1 is associated with cell proliferation, migration and invasion in breast cancer. *Molecular Medicine Reports*. 17:6405-6412.
- Fan, Y.H., C.X. Ji, B. Xu, H.Y. Fan, Z.J. Cheng, and X.G. Zhu. 2017. Long noncoding RNA activated by TGF-beta in human cancers: A meta-analysis. *Clinica Chimica Acta; International Journal of Clinical Chemistry*. 468:10-16.
- Fang, Z., J. Tang, Y. Bai, H. Lin, H. You, H. Jin, L. Lin, P. You, J. Li, Z. Dai, X. Liang, Y. Su, Q. Hu, F. Wang, and Z.Y. Zhang. 2015. Plasma levels of microRNA-24, microRNA-320a, and microRNA-423-5p are potential biomarkers for colorectal carcinoma. *Journal of Experimental & Clinical Cancer Research : CR*. 34:86.
- Farran, B., G. Dyson, D. Craig, A. Dombkowski, J.L. Beebe-Dimmer, I.J. Powell, I. Podgorski, L. Heilbrun, S. Bolton, and C.H. Bock. 2018. A study of circulating microRNAs identifies a new potential biomarker panel to distinguish aggressive prostate cancer. *Carcinogenesis*. 39:556-561.
- Ferlay, J., I. Soerjomataram, R. Dikshit, S. Eser, C. Mathers, M. Rebelo, D.M. Parkin, D. Forman, and F. Bray. 2015. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *International Journal of Cancer*. 136:E359-386.
- Folkman, J. 1995. Angiogenesis in cancer, vascular, rheumatoid and other disease. *Nature medicine*. 1:27-31.
- Ganguly, K.K., S. Pal, S. Moulik, and A. Chatterjee. 2013. Integrins and metastasis. *Cell Adhesion & Migration*. 7:251-261.
- Ge, X., Y. Chen, X. Liao, D. Liu, F. Li, H. Ruan, and W. Jia. 2013. Overexpression of long noncoding RNA PCAT-1 is a novel biomarker of poor prognosis in patients with colorectal cancer. *Medical Oncology (Northwood, London, England)*. 30:588.
- Gómez-Cuadrado, L., N. Tracey, R. Ma, B. Qian, and V.G. Brunton. 2017. Mouse models of metastasis: progress and prospects. *Disease Models & Mechanisms*. 10:1061-1074.

- Grange, C., M. Tapparo, F. Collino, L. Vitillo, C. Damasco, M.C. Deregibus, C. Tetta, B. Bussolati, and G. Camussi. 2011. Microvesicles released from human renal cancer stem cells stimulate angiogenesis and formation of lung premetastatic niche. *Cancer Research*. 71:5346-5356.
- Guan, X. 2015. Cancer metastases: challenges and opportunities. *Acta Pharmaceutica Sinica B*. 5:402-418.
- Gudbergsson, J.M., K.B. Johnsen, M.N. Skov, and M. Duroux. 2016. Systematic review of factors influencing extracellular vesicle yield from cell cultures. *Cytotechnology*. 68:579-592.
- Guo, X.B., Z. Hua, C. Li, L.P. Peng, J.S. Wang, B. Wang, and Q.M. Zhi. 2015. Biological significance of long non-coding RNA FTX expression in human colorectal cancer. *International Journal of Clinical and Experimental Medicine*. 8:15591-15600.
- Gupta, R.A., N. Shah, K.C. Wang, J. Kim, H.M. Horlings, D.J. Wong, M.C. Tsai, T. Hung, P. Argani, J.L. Rinn, Y. Wang, P. Brzoska, B. Kong, R. Li, R.B. West, M.J. van de Vijver, S. Sukumar, and H.Y. Chang. 2010. Long non-coding RNA HOTAIR reprograms chromatin state to promote cancer metastasis. *Nature*. 464:1071-1076.
- Hajjari, M., and A. Salavaty. 2015. HOTAIR: an oncogenic long non-coding RNA in different cancers. *Cancer Biology & Medicine*. 12:1-9.
- Hamam, R., D. Hamam, K.A. Alsaleh, M. Kassem, W. Zaher, M. Alfayez, A. Aldahmash, and N.M. Alajez. 2017. Circulating microRNAs in breast cancer: novel diagnostic and prognostic biomarkers. *Cell Death & Disease*. 8:e3045.
- Han, D., X. Gao, M. Wang, Y. Qiao, Y. Xu, J. Yang, N. Dong, J. He, Q. Sun, G. Lv, C. Xu, J. Tao, and N. Ma. 2016. Long noncoding RNA H19 indicates a poor prognosis of colorectal cancer and promotes tumor growth by recruiting and binding to eIF4A3. *Oncotarget*. 7:22159-22173.
- Han, D., M. Wang, N. Ma, Y. Xu, Y. Jiang, and X. Gao. 2015. Long noncoding RNAs: novel players in colorectal cancer. *Cancer Letters*. 361:13-21.
- Han, P., J.W. Li, B.M. Zhang, J.C. Lv, Y.M. Li, X.Y. Gu, Z.W. Yu, Y.H. Jia, X.F. Bai, L. Li, Y.L. Liu, and B.B. Cui. 2017. The lncRNA CRNDE promotes colorectal cancer cell proliferation and chemoresistance via miR-181a-5p-mediated regulation of Wnt/beta-catenin signaling. *Molecular Cancer*. 16:9.
- Han, Y., Y.N. Yang, H.H. Yuan, T.T. Zhang, H. Sui, X.L. Wei, L. Liu, P. Huang, W.J. Zhang, and Y.X. Bai. 2014. UCA1, a long non-coding RNA up-regulated in colorectal cancer influences cell proliferation, apoptosis and cell cycle distribution. *Pathology*. 46:396-401.

- Hansji, H., E.Y. Leung, B.C. Baguley, G.J. Finlay, D. Cameron-Smith, V.C. Figueiredo, and M.E. Askarian-Amiri. 2016. ZFAS1: a long noncoding RNA associated with ribosomes in breast cancer cells. *Biology Direct*. 11:62.
- Hao, C., G. Zhang, and L. Zhang. 2019. Serum CEA levels in 49 different types of cancer and noncancer diseases. *Progress in Molecular Biology and Translational Science*. 162:213-227.
- Harding, C., J. Heuser, and P. Stahl. 1983. Receptor-mediated endocytosis of transferrin and recycling of the transferrin receptor in rat reticulocytes. *The Journal of Cell Biology*. 97:329-339.
- He, P.Y., W.K. Yip, B.L. Chai, B.Y. Chai, M.F. Jabar, N. Dusa, N. Mohtarrudin, and H.F. Seow. 2017. Inhibition of cell migration and invasion by miR29a3p in a colorectal cancer cell line through suppression of CDC42BPA mRNA expression. *Oncology Reports*. 38:3554-3566.
- He, X., X. Tan, X. Wang, H. Jin, L. Liu, L. Ma, H. Yu, and Z. Fan. 2014. C-Myc-activated long noncoding RNA CCAT1 promotes colon cancer cell proliferation and invasion. *Tumour biology : the journal of the International Society for Oncodevelopmental Biology and Medicine*. 35:12181-12188.
- Helwa, I., J. Cai, M.D. Drewry, A. Zimmerman, M.B. Dinkins, M.L. Khaled, M. Seremwe, W.M. Dismuke, E. Bieberich, W.D. Stamer, M.W. Hamrick, and Y. Liu. 2017. A Comparative Study of Serum Exosome Isolation Using Differential Ultracentrifugation and Three Commercial Reagents. *PloS one*. 12:e0170628.
- Hu, D., Y. Zhan, K. Zhu, M. Bai, J. Han, Y. Si, H. Zhang, and D. Kong. 2018. Plasma Exosomal Long Non-Coding RNAs Serve as Biomarkers for Early Detection of Colorectal Cancer. *Cellular Physiology and Biochemistry : International Journal of Experimental Cellular Physiology, Biochemistry, and Pharmacology*. 51:2704-2715.
- Hu, Z.Y., X.Y. Wang, W.B. Guo, L.Y. Xie, Y.Q. Huang, Y.P. Liu, L.W. Xiao, S.N. Li, H.F. Zhu, Z.G. Li, and H. Kan. 2016. Long non-coding RNA MALAT1 increases AKAP-9 expression by promoting SRPK1-catalyzed SRSF1 phosphorylation in colorectal cancer cells. *Oncotarget*. 7:11733-11743.
- Iguchi, T., R. Uchi, S. Nambara, T. Saito, H. Komatsu, H. Hirata, M. Ueda, S. Sakimura, Y. Takano, J. Kurashige, Y. Shinden, H. Eguchi, K. Sugimachi, Y. Maehara, and K. Mimori. 2015. A long noncoding RNA, lncRNA-ATB, is involved in the progression and prognosis of colorectal cancer. *Anticancer Research*. 35:1385-1388.
- Jahroudi, N., and J.S. Greenberger. 1995. The role of endothelial cells in tumor invasion and metastasis. *Journal of Neuro-oncology*. 23:99-108.

- Kahlert, C., and R. Kalluri. 2013. Exosomes in tumor microenvironment influence cancer progression and metastasis. *Journal of Molecular Medicine (Berlin, Germany)*. 91:431-437.
- Kasagi, Y., E. Oki, K. Ando, S. Ito, T. Iguchi, M. Sugiyama, Y. Nakashima, K. Ohgaki, H. Saeki, K. Mimori, and Y. Maehara. 2017. The Expression of CCAT2, a Novel Long Noncoding RNA Transcript, and rs6983267 Single-Nucleotide Polymorphism Genotypes in Colorectal Cancers. *Oncology*. 92:48-54.
- Katoh, M. 2013. Therapeutics targeting angiogenesis: genetics and epigenetics, extracellular miRNAs and signaling networks (Review). *International Journal of Molecular Medicine*. 32:763-767.
- Kino, T., D.E. Hurt, T. Ichijo, N. Nader, and G.P. Chrousos. 2010. Noncoding RNA gas5 is a growth arrest- and starvation-associated repressor of the glucocorticoid receptor. *Science Signaling*. 3:ra8.
- Kita, Y., K. Yonemori, Y. Osako, K. Baba, S. Mori, K. Maemura, and S. Natsugoe. 2017. Noncoding RNA and colorectal cancer: its epigenetic role. *Journal of Human Genetics*. 62:41-47.
- Kung, J.T., D. Colognori, and J.T. Lee. 2013. Long noncoding RNAs: past, present, and future. *Genetics*. 193:651-669.
- Lang, H.L., G.W. Hu, Y. Chen, Y. Liu, W. Tu, Y.M. Lu, L. Wu, and G.H. Xu. 2017a. Glioma cells promote angiogenesis through the release of exosomes containing long non-coding RNA POU3F3. *European Review for Medical and Pharmacological Sciences*. 21:959-972.
- Lang, H.L., G.W. Hu, B. Zhang, W. Kuang, Y. Chen, L. Wu, and G.H. Xu. 2017b. Glioma cells enhance angiogenesis and inhibit endothelial cell apoptosis through the release of exosomes that contain long non-coding RNA CCAT2. *Oncology Reports*. 38:785-798.
- Levin, T.R., and D.A. Corley. 2013. Colorectal-cancer screening--coming of age. *The New England Journal of Medicine*. 369:1164-1166.
- Li, H., S.Q. Ma, J. Huang, X.P. Chen, and H.H. Zhou. 2017a. Roles of long noncoding RNAs in colorectal cancer metastasis. *Oncotarget*. 8:39859-39876.
- Li, J., Y. Wang, C.-G. Zhang, H.-J. Xiao, J.-M. Hou, and J.-D. He. 2018. Effect of long non-coding RNA Gas5 on proliferation, migration, invasion and apoptosis of colorectal cancer HT-29 cell line. *Cancer Cell International*. 18:4.
- Li, J., W. Xue, J. Lv, P. Han, Y. Liu, and B. Cui. 2017b. Identification of potential long non-coding RNA biomarkers associated with the progression of colon cancer. *Oncotarget*. 8:75834-75843.

- Li, N., X.B. Feng, Q. Tan, P. Luo, W. Jing, M. Zhu, C. Liang, J. Tu, and Y. Ning. 2017c. Identification of Circulating Long Noncoding RNA Linc00152 as a Novel Biomarker for Diagnosis and Monitoring of Non-Small-Cell Lung Cancer. *Disease Markers*. 2017:7439698.
- Li, Q., Y. Dai, F. Wang, and S. Hou. 2016. Differentially expressed long non-coding RNAs and the prognostic potential in colorectal cancer. *Neoplasma*. 63:977-983.
- Li, W., C. Li, T. Zhou, X. Liu, X. Liu, X. Li, and D. Chen. 2017d. Role of exosomal proteins in cancer diagnosis. *Molecular Cancer*. 16:145.
- Li, Y., Y. Li, S. Huang, K. He, M. Zhao, H. Lin, D. Li, J. Qian, C. Zhou, Y. Chen, and C. Huang. 2017e. Long non-coding RNA growth arrest specific transcript 5 acts as a tumour suppressor in colorectal cancer by inhibiting interleukin-10 and vascular endothelial growth factor expression. *Oncotarget*. 8:13690-13702.
- Lian, Y., Z. Cai, H. Gong, S. Xue, D. Wu, and K. Wang. 2016a. HOTTIP: a critical oncogenic long non-coding RNA in human cancers. *Molecular BioSystems*. 12:3247-3253.
- Lian, Y., J. Ding, Z. Zhang, Y. Shi, Y. Zhu, J. Li, P. Peng, J. Wang, Y. Fan, W. De, and K. Wang. 2016b. The long noncoding RNA HOXA transcript at the distal tip promotes colorectal cancer growth partially via silencing of p21 expression. *Tumour biology : The Journal of the International Society for Oncodevelopmental Biology and Medicine*. 37:7431-7440.
- Liang, W.C., W.M. Fu, C.W. Wong, Y. Wang, W.M. Wang, G.X. Hu, L. Zhang, L.J. Xiao, D.C. Wan, J.F. Zhang, and M.M. Waye. 2015. The lncRNA H19 promotes epithelial to mesenchymal transition by functioning as miRNA sponges in colorectal cancer. *Oncotarget*. 6:22513-22525.
- Liu, L., T. Meng, X.H. Yang, P. Sayim, C. Lei, B. Jin, L. Ge, and H.J. Wang. 2018. Prognostic and predictive value of long non-coding RNA GAS5 and mircoRNA-221 in colorectal cancer and their effects on colorectal cancer cell proliferation, migration and invasion. *Cancer biomarkers : section A of Disease Markers*. 22:283-299.
- Liu, Q., J. Huang, N. Zhou, Z. Zhang, A. Zhang, Z. Lu, F. Wu, and Y.Y. Mo. 2013. LncRNA loc285194 is a p53-regulated tumor suppressor. *Nucleic Acids Research*. 41:4976-4987.
- Liu, T., X. Zhang, S. Gao, F. Jing, Y. Yang, L. Du, G. Zheng, P. Li, C. Li, and C. Wang. 2016. Exosomal long noncoding RNA CRNDE-h as a novel serum-based biomarker for diagnosis and prognosis of colorectal cancer. *Oncotarget*. 7:85551-85563.

- Liu, Y., M. Zhang, L. Liang, J. Li, and Y.X. Chen. 2015. Over-expression of lncRNA DANCR is associated with advanced tumor progression and poor prognosis in patients with colorectal cancer. *International Journal of Clinical and Experimental Pathology*. 8:11480-11484.
- Luo, P., C. Liang, X. Zhang, X. Liu, Y. Wang, M. Wu, X. Feng, and J. Tu. 2018. Identification of long non-coding RNA ZFAS1 as a novel biomarker for diagnosis of HCC. *Bioscience Reports*. 38.
- Ma, C., X. Shi, Q. Zhu, Q. Li, Y. Liu, Y. Yao, and Y. Song. 2016. The growth arrest-specific transcript 5 (GAS5): a pivotal tumor suppressor long noncoding RNA in human cancers. *Tumour Biology : The Journal of the International Society for Oncodevelopmental Biology and Medicine*. 37:1437-1444.
- Ma, L., V.B. Bajic, and Z. Zhang. 2013. On the classification of long non-coding RNAs. *RNA biology*. 10:925-933.
- Meng, Q., M. Ren, Y. Li, and X. Song. 2016. LncRNA-RMRP Acts as an Oncogene in Lung Cancer. *PloS one*. 11:e0164845.
- Misawa, A., K.I. Takayama, and S. Inoue. 2017. Long non-coding RNAs and prostate cancer. *Cancer Science*. 108:2107-2114.
- Molloy, T., and L.J. van 't Veer. 2008. Recent advances in metastasis research. *Current Opinion in Genetics & Development*. 18:35-41.
- Moriarty, A., J. O'Sullivan, J. Kennedy, B. Mehigan, and P. McCormick. 2016. Current targeted therapies in the treatment of advanced colorectal cancer: a review. *Therapeutic Advances in Medical Oncology*. 8:276-293.
- Mourtada-Maarabouni, M., V.L. Hedge, L. Kirkham, F. Farzaneh, and G.T. Williams. 2008. Growth arrest in human T-cells is controlled by the non-coding RNA growth-arrest-specific transcript 5 (GAS5). *Journal of Cell Science*. 121:939-946
- Mourtada-Maarabouni, M., M.R. Pickard, V.L. Hedge, F. Farzaneh, and G.T. Williams. 2009. GAS5, a non-protein-coding RNA, controls apoptosis and is downregulated in breast cancer. *Oncogene*. 28:195-208.
- Ni, B., X. Yu, X. Guo, X. Fan, Z. Yang, P. Wu, Z. Yuan, Y. Deng, J. Wang, D. Chen, and L. Wang. 2015. Increased urothelial cancer associated 1 is associated with tumor proliferation and metastasis and predicts poor prognosis in colorectal cancer. *International Journal of Oncology*. 47:1329-1338.
- Nissan, A., A. Stojadinovic, S. Mitrani-Rosenbaum, D. Halle, R. Grinbaum, M. Roistacher, A. Bochem, B.E. Dayanc, G. Ritter, I. Gomceli, E.B. Bostanci, M. Akoglu, Y.T. Chen, L.J. Old, and A.O. Gure. 2012. Colon cancer associated transcript-1: a novel RNA expressed in malignant and pre-malignant human tissues. *International Journal of Cancer*. 130:1598-1606.

- Ohno, S., A. Ishikawa, and M. Kuroda. 2013. Roles of exosomes and microvesicles in disease pathogenesis. *Advanced Drug Delivery Reviews*. 65:398-401.
- Pan, L., W. Liang, M. Fu, Z.H. Huang, X. Li, W. Zhang, P. Zhang, H. Qian, P.C. Jiang, W.R. Xu, and X. Zhang. 2017. Exosomes-mediated transfer of long noncoding RNA ZFAS1 promotes gastric cancer progression. *Journal of Cancer Research and Clinical Oncology*. 143:991-1004.
- Panzitt, K., M.M. Tschernatsch, C. Guelly, T. Moustafa, M. Stradner, H.M. Strohmaier, C.R. Buck, H. Denk, R. Schroeder, M. Trauner, and K. Zatloukal. 2007. Characterization of HULC, a novel gene with striking up-regulation in hepatocellular carcinoma, as noncoding RNA. *Gastroenterology*. 132:330-342.
- Pickard, M.R., M. Mourtada-Maarabouni, and G.T. Williams. 2013. Long non-coding RNA GAS5 regulates apoptosis in prostate cancer cell lines. *Biochimica et Biophysica Acta*. 1832:1613-1623.
- Prensner, J.R., M.K. Iyer, O.A. Balbin, S.M. Dhanasekaran, Q. Cao, J.C. Brenner, B. Laxman, I.A. Asangani, C.S. Grasso, H.D. Kominsky, X. Cao, X. Jing, X. Wang, J. Siddiqui, J.T. Wei, D. Robinson, H.K. Iyer, N. Palanisamy, C.A. Maher, and A.M. Chinnaiyan. 2011. Transcriptome sequencing across a prostate cancer cohort identifies PCAT-1, an unannotated lincRNA implicated in disease progression. *Nature Biotechnology*. 29:742-749.
- Qi, P., M.D. Xu, S.J. Ni, X.H. Shen, P. Wei, D. Huang, C. Tan, W.Q. Sheng, X.Y. Zhou, and X. Du. 2015. Down-regulation of ncRAN, a long non-coding RNA, contributes to colorectal cancer cell migration and invasion and predicts poor overall survival for colorectal cancer patients. *Molecular Carcinogenesis*. 54:742-750.
- Qi, P., X.Y. Zhou, and X. Du. 2016. Circulating long non-coding RNAs in cancer: current status and future perspectives. *Molecular Cancer*. 15:39.
- Rashed, M., E. Bayraktar, K.H. G, M.F. Abd-Ellah, P. Amero, A. Chavez-Reyes, and C. Rodriguez-Aguayo. 2017. Exosomes: From Garbage Bins to Promising Therapeutic Targets. *International Journal of Molecular Sciences*. 18.
- Raut, S.K., and M. Khullar. 2018. The Big Entity of New RNA World: Long Non-Coding RNAs in Microvascular Complications of Diabetes. *Front Endocrinol (Lausanne)*. 9:300.
- Ren, Y.K., Y. Xiao, X.B. Wan, Y.Z. Zhao, J. Li, Y. Li, G.S. Han, X.B. Chen, Q.Y. Zou, G.C. Wang, C.M. Lu, Y.C. Xu, and Y.C. Wang. 2015. Association of long non-coding RNA HOTTIP with progression and prognosis in colorectal cancer. *International Journal of Clinical and Experimental Pathology*. 8:11458-11463.

- Roma-Rodrigues, C., A.R. Fernandes, and P.V. Baptista. 2014. Exosome in tumour microenvironment: overview of the crosstalk between normal and cancer cells. *BioMed Research International*. 2014:179486.
- Sapre, N., and L.A. Selth. 2013. Circulating MicroRNAs as Biomarkers of Prostate Cancer: The State of Play. *Prostate Cancer*. 2013:539680.
- Saus, E., A. Brunet-Vega, S. Iraola-Guzman, C. Pegueroles, T. Gabaldon, and C. Pericay. 2016. Long Non-Coding RNAs As Potential Novel Prognostic Biomarkers in Colorectal Cancer. *Frontiers in Genetics*. 7:54.
- Shao, Y., M. Ye, Q. Li, W. Sun, G. Ye, X. Zhang, Y. Yang, B. Xiao, and J. Guo. 2016. LncRNA-RMRP promotes carcinogenesis by acting as a miR-206 sponge and is used as a novel biomarker for gastric cancer. *Oncotarget*. 7:37812-37824.
- Shi, D., L. Liang, H. Zheng, G. Cai, X. Li, Y. Xu, and S. Cai. 2017. Silencing of long non-coding RNA SBDSP1 suppresses tumor growth and invasion in colorectal cancer. *Biomedicine & pharmacotherapy = Biomedecine & Pharmacotherapie*. 85:355-361.
- Shi, D., H. Zheng, C. Zhuo, J. Peng, D. Li, Y. Xu, X. Li, G. Cai, and S. Cai. 2014. Low expression of novel lncRNA RP11-462C24.1 suggests a biomarker of poor prognosis in colorectal cancer. *Medical Oncology (Northwood, London, England)*. 31:31.
- Shi, J., X. Li, F. Zhang, C. Zhang, Q. Guan, X. Cao, W. Zhu, X. Zhang, Y. Cheng, K. Ou, Q. Chen, and S. Hu. 2015. Circulating lncRNAs associated with occurrence of colorectal cancer progression. *American Journal of Cancer Research*. 5:2258-2265.
- Smolle, M., S. Uranitsch, A. Gerger, M. Pichler, and J. Haybaeck. 2014. Current status of long non-coding RNAs in human cancer with specific focus on colorectal cancer. *International Journal of Molecular Sciences*. 15:13993-14013.
- Song, W., D. Yan, T. Wei, Q. Liu, X. Zhou, and J. Liu. 2018. Tumor-derived extracellular vesicles in angiogenesis. *Biomedicine & pharmacotherapy = Biomedecine & Pharmacotherapie*. 102:1203-1208.
- Suwakulsiri, W., A. Rai, R. Xu, M. Chen, D.W. Greening, and R.J. Simpson. 2018. Proteomic profiling reveals key cancer progression modulators in shed microvesicles released from isogenic human primary and metastatic colorectal cancer cell lines. *Biochimica et biophysica acta. Proteins and proteomics*.
- Takahashi, Y., G. Sawada, J. Kurashige, R. Uchi, T. Matsumura, H. Ueo, Y. Takano, H. Eguchi, T. Sudo, K. Sugimachi, H. Yamamoto, Y. Doki, M. Mori, and K. Mimori. 2014. Amplification of PVT-1 is involved in poor prognosis via apoptosis inhibition in colorectal cancers. *British Journal of Cancer*. 110:164-171.

- Thorenoor, N., P. Faltejskova-Vychytilova, S. Hombach, J. Mlcochova, M. Kretz, M. Svoboda, and O. Slaby. 2016. Long non-coding RNA ZFAS1 interacts with CDK1 and is involved in p53-dependent cell cycle control and apoptosis in colorectal cancer. *Oncotarget*. 7:622-637.
- Tsai, M.C., O. Manor, Y. Wan, N. Mosammaparast, J.K. Wang, F. Lan, Y. Shi, E. Segal, and H.Y. Chang. 2010. Long noncoding RNA as modular scaffold of histone modification complexes. *Science (New York, N.Y.)*. 329:689-693.
- Tseng, Y.Y., B.S. Moriarity, W. Gong, R. Akiyama, A. Tiwari, H. Kawakami, P. Ronning, B. Reuland, K. Guenther, T.C. Beadnell, J. Essig, G.M. Otto, M.G. O'Sullivan, D.A. Largaespada, K.L. Schwertfeger, Y. Marahrens, Y. Kawakami, and A. Bagchi. 2014. PVT1 dependence in cancer with MYC copy-number increase. *Nature*. 512:82-86.
- Valastyan, S., and R.A. Weinberg. 2011. Tumor metastasis: molecular insights and evolving paradigms. *Cell*. 147:275-292.
- Veettil, S.K., K.G. Lim, N. Chaiyakunapruk, S.M. Ching, and M.R. Abu Hassan. 2017. Colorectal cancer in Malaysia: Its burden and implications for a multiethnic country. *Asian Journal of Surgery*. 40:481-489.
- Verma, M., T.K. Lam, E. Hebert, and R.L. Divi. 2015. Extracellular vesicles: potential applications in cancer diagnosis, prognosis, and epidemiology. *BMC Clinical Pathology*. 15:6.
- Waldenstrom, A., and G. Ronquist. 2014. Role of exosomes in myocardial remodeling. *Circulation Research*. 114:315-324.
- Wang, C., J. Yu, Y. Han, L. Li, J. Li, T. Li, and P. Qi. 2016. Long non-coding RNAs LOC285194, RP11-462C24.1 and Nbla12061 in serum provide a new approach for distinguishing patients with colorectal cancer from healthy controls. *Oncotarget*. 7:70769-70778.
- Wang, J., Y.X. Song, B. Ma, J.J. Wang, J.X. Sun, X.W. Chen, J.H. Zhao, Y.C. Yang, and Z.N. Wang. 2015. Regulatory Roles of Non-Coding RNAs in Colorectal Cancer. *International Journal of Molecular Sciences*. 16:19886-19919.
- Wang, J.S., Q.H. Liu, X.H. Cheng, W.Y. Zhang, and Y.C. Jin. 2018. The long noncoding RNA ZFAS1 facilitates bladder cancer tumorigenesis by sponging miR-329. *Biomedicine & pharmacotherapy = Biomedecine & Pharmacotherapie*. 103:174-181.
- Wang, T., X. Qu, J. Jiang, P. Gao, D. Zhao, X. Lian, and X. Li. 2017. Diagnostic significance of urinary long non-coding PCA3 RNA in prostate cancer. *Oncotarget*. 8:58577-58586.

- Wang, W., and C. Xing. 2016. Upregulation of long noncoding RNA ZFAS1 predicts poor prognosis and prompts invasion and metastasis in colorectal cancer. *Pathology, Research and Practice*. 212:690-695.
- Ward, M., C. McEwan, J.D. Mills, and M. Janitz. 2015. Conservation and tissue-specific transcription patterns of long noncoding RNAs. *Journal of Human Transcriptome*. 1:2-9.
- Wu, K.F., W.C. Liang, L. Feng, J.X. Pang, M.M. Waye, J.F. Zhang, and W.M. Fu. 2017. H19 mediates methotrexate resistance in colorectal cancer through activating Wnt/beta-catenin pathway. *Experimental Cell Research*. 350:312-317.
- Wu, Y., W. Deng, and D.J. Klinke, 2nd. 2015. Exosomes: improved methods to characterize their morphology, RNA content, and surface protein biomarkers. *The Analyst*. 140:6631-6642.
- Wu, Y., L. Zhang, Y. Wang, H. Li, X. Ren, F. Wei, W. Yu, X. Wang, L. Zhang, J. Yu, and X. Hao. 2014. Long noncoding RNA HOTAIR involvement in cancer. *Tumour Biology : The Journal of the International Society for Oncodevelopmental Biology and Medicine*. 35:9531-9538.
- Xiao, H., F. Zhang, Y. Zou, J. Li, Y. Liu, and W. Huang. 2018. The Function and Mechanism of Long Non-coding RNA-ATB in Cancers. *Frontiers in Physiology*. 9:321.
- Xie, X., B. Tang, Y.F. Xiao, R. Xie, B.S. Li, H. Dong, J.Y. Zhou, and S.M. Yang. 2016. Long non-coding RNAs in colorectal cancer. *Oncotarget*. 7:5226-5239.
- Xie, Y., W. Dang, S. Zhang, W. Yue, L. Yang, X. Zhai, Q. Yan, and J. Lu. 2019. The role of exosomal noncoding RNAs in cancer. *Molecular Cancer*. 18:37.
- Xin, Y., Z. Li, J. Shen, M.T. Chan, and W.K. Wu. 2016. CCAT1: a pivotal oncogenic long non-coding RNA in human cancers. *Cell Proliferation*. 49:255-260.
- Xu, M.D., P. Qi, and X. Du. 2014. Long non-coding RNAs in colorectal cancer: implications for pathogenesis and clinical application. *Modern pathology : an official journal of the United States and Canadian Academy of Pathology, Inc*. 27:1310-1320.
- Xue, M., W. Chen, and X. Li. 2016. Urothelial cancer associated 1: a long noncoding RNA with a crucial role in cancer. *Journal of Cancer Research and Clinical Oncology*. 142:1407-1419.
- Yan, B., W. Gu, Z. Yang, Z. Gu, X. Yue, Q. Gu, and L. Liu. 2014. Downregulation of a long noncoding RNA-ncRuPAR contributes to tumor inhibition in colorectal cancer. *Tumour Biology : The Journal of the International Society for Oncodevelopmental Biology and Medicine*. 35:11329-11335.

- Yang, M.H., Z.Y. Hu, C. Xu, L.Y. Xie, X.Y. Wang, S.Y. Chen, and Z.G. Li. 2015. MALAT1 promotes colorectal cancer cell proliferation/migration/invasion via PRKA kinase anchor protein 9. *Biochimica et Biophysica Acta*. 1852:166-174.
- Yang, W., N. Ning, and X. Jin. 2017a. The lncRNA H19 Promotes Cell Proliferation by Competitively Binding to miR-200a and Derepressing beta-Catenin Expression in Colorectal Cancer. *BioMed Research International*. 2017:2767484.
- Yang, X.J., C.Q. Huang, C.W. Peng, J.X. Hou, and J.Y. Liu. 2016. Long noncoding RNA HULC promotes colorectal carcinoma progression through epigenetically repressing NKD2 expression. *Gene*. 592:172-178.
- Yang, Y., P. Junjie, C. Sanjun, and Y. Ma. 2017b. Long non-coding RNAs in Colorectal Cancer: Progression and Future Directions. *Journal of Cancer*. 8:3212-3225.
- Yang, Y., Z. Shen, Y. Yan, B. Wang, J. Zhang, C. Shen, T. Li, C. Ye, Z. Gao, G. Peng, Y. Ye, K. Jiang, and S. Wang. 2017c. Long non-coding RNA GAS5 inhibits cell proliferation, induces G0/G1 arrest and apoptosis, and functions as a prognostic marker in colorectal cancer. *Oncology Letters*. 13:3151-3158.
- Ye, L.C., T. Chen, D.X. Zhu, S.X. Lv, J.J. Qiu, J. Xu, F.L. Yuan, and Y. Wei. 2016. Downregulated long non-coding RNA CLMAT3 promotes the proliferation of colorectal cancer cells by targeting regulators of the cell cycle pathway. *Oncotarget*. 7:58931-58938.
- Ye, L.C., L. Ren, J.J. Qiu, D.X. Zhu, T. Chen, W.J. Chang, S.X. Lv, and J. Xu. 2015. Aberrant expression of long noncoding RNAs in colorectal cancer with liver metastasis. *Tumour biology : the journal of the International Society for Oncodevelopmental Biology and Medicine*. 36:8747-8754.
- Yin, D., X. He, E. Zhang, R. Kong, W. De, and Z. Zhang. 2014. Long noncoding RNA GAS5 affects cell proliferation and predicts a poor prognosis in patients with colorectal cancer. *Medical Oncology (Northwood, London, England)*. 31:253.
- Yin, D.D., Z.J. Liu, E. Zhang, R. Kong, Z.H. Zhang, and R.H. Guo. 2015. Decreased expression of long noncoding RNA MEG3 affects cell proliferation and predicts a poor prognosis in patients with colorectal cancer. *Tumour biology : the journal of the International Society for Oncodevelopmental Biology and Medicine*. 36:4851-4859.
- Yokoi, A., Y. Yoshioka, A. Hirakawa, Y. Yamamoto, M. Ishikawa, S.I. Ikeda, T. Kato, K. Niimi, H. Kajiyama, F. Kikkawa, and T. Ochiya. 2017. A combination of circulating miRNAs for the early detection of ovarian cancer. *Oncotarget*. 8:89811-89823.

- Yoshimura, H., Y. Matsuda, M. Yamamoto, S. Kamiya, and T. Ishiwata. 2018. Expression and role of long non-coding RNA H19 in carcinogenesis. *Frontiers in bioscience (Landmark edition)*. 23:614-625.
- Yu, B., and S. Wang. 2018. Angio-LncRs: LncRNAs that regulate angiogenesis and vascular disease. *Theranostics*. 8:3654-3675.
- Yu, H., Z. Guan, K. Cuk, H. Brenner, and Y. Zhang. 2018. Circulating microRNA biomarkers for lung cancer detection in Western populations. *Cancer medicine*. 7:4849-4862.
- Yu, Y., J. Yang, Q. Li, B. Xu, Y. Lian, and L. Miao. 2017. LINC00152: A pivotal oncogenic long non-coding RNA in human cancers. *Cell proliferation*. 50.
- Yue, B., S. Qiu, S. Zhao, C. Liu, D. Zhang, F. Yu, Z. Peng, and D. Yan. 2016. LncRNA-ATB mediated E-cadherin repression promotes the progression of colon cancer and predicts poor prognosis. *Journal of Gastroenterology and Hepatology*. 31:595-603.
- Zhang, J., G.S. Raju, D.W. Chang, S.H. Lin, Z. Chen, and X. Wu. 2018. Global and targeted circulating microRNA profiling of colorectal adenoma and colorectal cancer. *Cancer*. 124:785-796.
- Zhang, K., Z. Luo, Y. Zhang, L. Zhang, L. Wu, L. Liu, J. Yang, X. Song, and J. Liu. 2016a. Circulating lncRNA H19 in plasma as a novel biomarker for breast cancer. *Cancer Biomarkers : Section A of Disease Markers*. 17:187-194.
- Zhang, S., S. Chen, G. Yang, F. Gu, M. Li, B. Zhong, J. Hu, A. Hoffman, and M. Chen. 2014a. Long noncoding RNA HOTAIR as an independent prognostic marker in cancer: a meta-analysis. *PloS one*. 9:e105538.
- Zhang, W., Y. Bi, J. Li, F. Peng, H. Li, C. Li, L. Wang, F. Ren, C. Xie, P. Wang, W. Liang, Z. Wang, and D. Zhu. 2017. Long noncoding RNA FTX is upregulated in gliomas and promotes proliferation and invasion of glioma cells by negatively regulating miR-342-3p. *Laboratory Investigation; A Journal of Technical Methods and Pathology*. 97:447-457.
- Zhang, Y.H., J. Fu, Z.J. Zhang, C.C. Ge, and Y. Yi. 2016b. LncRNA-LINC00152 down-regulated by miR-376c-3p restricts viability and promotes apoptosis of colorectal cancer cells. *American Journal of Translational Research*. 8:5286-5297.
- Zhang, Z., C. Wang, T. Li, Z. Liu, and L. Li. 2014b. Comparison of ultracentrifugation and density gradient separation methods for isolating Tca8113 human tongue cancer cell line-derived exosomes. *Oncology Letters*. 8:1701-1706.

- Zhao, J., J. Xu, A.Q. Shang, and R. Zhang. 2018. A Six-LncRNA Expression Signature Associated with Prognosis of Colorectal Cancer Patients. *Cellular Physiology and Biochemistry : International Journal of Experimental Cellular Physiology, Biochemistry, and Pharmacology*. 50:1882-1890.
- Zheng, G., L. Du, X. Yang, X. Zhang, L. Wang, Y. Yang, J. Li, and C. Wang. 2014a. Serum microRNA panel as biomarkers for early diagnosis of colorectal adenocarcinoma. *British Journal of Cancer*. 111:1985-1992.
- Zheng, H.T., D.B. Shi, Y.W. Wang, X.X. Li, Y. Xu, P. Tripathi, W.L. Gu, G.X. Cai, and S.J. Cai. 2014b. High expression of lncRNA MALAT1 suggests a biomarker of poor prognosis in colorectal cancer. *International Journal of Clinical and Experimental Pathology*. 7:3174-3181.
- Zheng, J., L. Hu, J. Cheng, J. Xu, Z. Zhong, Y. Yang, and Z. Yuan. 2018. lncRNA PVT1 promotes the angiogenesis of vascular endothelial cell by targeting miR26b to activate CTGF/ANGPT2. *International Journal of Molecular Medicine*. 42:489-496.
- Zheng, Y., D. Song, K. Xiao, C. Yang, Y. Ding, W. Deng, and S. Tong. 2016. LncRNA GAS5 contributes to lymphatic metastasis in colorectal cancer. *Oncotarget*. 7:83727-83734.
- Zhou, M., L. Zhong, W. Xu, Y. Sun, Z. Zhang, H. Zhao, L. Yang, and J. Sun. 2016. Discovery of potential prognostic long non-coding RNA biomarkers for predicting the risk of tumor recurrence of breast cancer patients. *Scientific Reports*. 6:31038.
- Zhou, Y., X. Zhang, and A. Klibanski. 2012. MEG3 noncoding RNA: a tumor suppressor. *Journal of Molecular Endocrinology*. 48:R45-53.
- Zhuang, G., X. Wu, Z. Jiang, I. Kasman, J. Yao, Y. Guan, J. Oeh, Z. Modrusan, C. Bais, D. Sampath, and N. Ferrara. 2012. Tumour-secreted miR-9 promotes endothelial cell migration and angiogenesis by activating the JAK-STAT pathway. *The EMBO Journal*. 31:3513-3523.