



**UNIVERSITI PUTRA MALAYSIA**

***IN VITRO ANTIPROLIFERATIVE EFFECTS AND UNDERLYING  
MECHANISMS OF BISMUTH DITHIOCARBAMATE DERIVATIVES  
AGAINST BREAST CANCER CELL LINE***

**CHAN PIT FOONG**

**FPSK(p) 2021 5**



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AGAINST BREAST CANCER CELL LINE**

By

**CHAN PIT FOONG**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra  
Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of  
Philosophy**

**July 2019**

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in  
fulfilment of the requirement of the degree of Doctor of Philosophy

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July 2019

**Chair : Roslida Abdul Hamid @ Abdul Razak, PhD**  
**Faculty : Medicine and Health Sciences**

Drastic increment of cancer incidence in recent years has driven metal complexes including bismuth complexes as a promising approach in anticancer drug development due to its well-known low toxicity, environmental friendliness and medical therapeutic benefits. The complexes with R = (CH<sub>2</sub>CH<sub>2</sub>OH)(iPr), Et<sub>2</sub>, (CH<sub>2</sub>)<sub>4</sub> and (CH<sub>2</sub>CH<sub>2</sub>OH)(CH<sub>3</sub>) in bismuth-dithiocarbamate, Bi[S<sub>2</sub>CN-R]<sub>3</sub> labelled as C3, C4, C5 and C9, have been studied, respectively. This study aimed at determining the antiproliferative effect of the bismuth complexes and their underlying mechanism(s) in breast cancer cell (MCF-7) through various *in vitro* assays. The antiproliferative effect of the bismuth complexes was studied using Methylthiazolyldiphenyl-tetrazolium bromide (MTT) assay. Apoptotic cell death activities of C3, C4, C5 and C9 were performed via DNA fragmentation, acridine orange/propidium iodide (AO/PI), Annexin V and caspase activity assays. Whilst, this study further employed Human Cancer Drug Targets RT<sup>2</sup> Profiler PCR array to study the expression of 84 actively sought targets for anticancer therapeutics and drug development to delineate the underlying mechanism(s). Results showed all tested compounds (C3, C4, C5 and C9) exhibited antiproliferative effects against MCF-7 cell line with IC<sub>50</sub> of 10.33 ± 0.06 μM, 1.26 ± 0.02 μM, 1.07 ± 0.01 μM and 25.37 ± 0.12 μM, respectively. The morphology of apoptosis including formation of DNA fragments, phosphatidylserine translocation, chromatin condensation, and membrane blebbing was shown. The compounds were found to variably increase reactive oxygen species (ROS) generation thus increased mitochondrial membrane potential (MMP). Consequently, this led to the release of cytochrome c from mitochondria, demonstrated by the data obtained by flow cytometric analysis. All four tested compounds were revealed to induce both intrinsic and extrinsic apoptotic pathways, conferred by the data obtained from Human Cancer Drug Targets RT<sup>2</sup> Profiler PCR array, along with caspases activities assay. The compounds were also reported to significantly reduce several key gene

expressions such as *AKT1*, *BIRC5*, *CDK1* and *NFκB1*. The NF-κB signalling pathway was inhibited with the activation of Lys48-linked polyubiquitination thus led to NF-κB degradation. Conclusively, this study evidenced the anticancer property of C3, C4, C5 and C9 against breast cancer by initiating intrinsic and extrinsic apoptosis pathway and lays the foundation in the development of new bismuth based chemotherapeutic drugs.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

**KESAN *IN VITRO* ANTIPROLIFERATIF DAN MEKANISME- MEKANISME  
TERBITAN BISMUTH DITHIOKARBAMAT KE ATAS SEL KANSER  
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Peningkatan kejadian kanser secara drastik kebelakangan ini telah mendorong perubahan kanser yang berasaskan kompleks logam termasuk kompleks bismuth sebagai langkah yang menjanjikan dalam perkembangan drug antikanser, disebabkan ketoksikannya yang rendah, mesra alam sekitar dan bermanfaat sebagai terapeutik perubahan. Terbitan kompleks dengan R= (CH<sub>2</sub>CH<sub>2</sub>OH)(iPr), Et<sub>2</sub>, (CH<sub>2</sub>)<sub>4</sub> dan (CH<sub>2</sub>CH<sub>2</sub>OH)(CH<sub>3</sub>) dalam bismuth dithiokarbamat, Bi[S<sub>2</sub>CN-R]<sub>3</sub> dilabelkan sebagai C3, C4, C5 serta C9, masing-masing telah diuji untuk menentukan kesan anti-proliferatif dan mekanisme-mekanisme sebatian-sebatian tersebut ke atas sel kanser payudara (MCF-7) melalui pelbagai asai *in vitro*. Kesan anti-proliferatif kompleks bismuth dikaji melalui asai Metilthiazolidifenil-tetrazolium bromide (MTT). Kematian sel apoptotik sebatian C3, C4, C5 dan C9 masing-masing dilakukan melalui pemecahan DNA, akridin oren/propidium iodida (AO/PI), asai Annexin V dan asai aktiviti caspase. Sementara itu, kajian ini turut menggunakan pemprofilan RT<sup>2</sup> PCR Sasaran Kanser Manusia untuk mengkaji ekspresi 84 sasaran yang aktif dalam terapeutik antikanser dan perkembangan drug, di samping menjelaskan mekanismenya. Kajian menunjukkan semua sebatian yang diuji (C3, C4, C5 dan C9) menunjukkan kesan anti-proliferatif terhadap sel MCF-7 masing-masing dengan IC<sub>50</sub> 10.33 ± 0.06 μM, 1.26 ± 0.02 μM, 1.07 ± 0.01 μM dan 25.37 ± 0.12 μM. Morfologi apoptosis menunjukkan perubahan-perubahan termasuk pembentukan pecahan DNA, translokasi fosfatidilserin, kondensasi kromatin, dan pembengkakkan membran. Sebatian-sebatian tersebut juga didapati menyebabkan peningkatan yang berbeza terhadap penjanaan spesies oksigen reaktif (ROS) seterusnya meningkatkan potensi membran mitokondria (MMP). Di samping itu, data yang diperolehi daripada analisis aliran sitometrik juga menunjukkan pelepasan sitokrom c dari mitokondria. Data yang diperolehi daripada pemprofilan RT<sup>2</sup> PCR Sasaran Kanser Manusia bersama-sama

dengan asai aktiviti caspase menunjukkan keempat-empat sebatian yang diuji menyebabkan kematian sel kanser MCF-7 secara apoptosis berperantaraan aruhan laluan intrinsik dan ekstrinsik. Sebatian-sebatian ini juga dilaporkan dapat merendahkan ekspresi beberapa gen utama seperti *AKT1*, *BIRC5*, *CDK1* dan *NFKB1*. Laluan isyarat NF- $\kappa$ B telah direncat dengan pengaktifan poliubiquitinasi yang dikaitkan dengan Lys48 seterusnya menyebabkan degradasi NF- $\kappa$ B. Kesimpulannya, kajian ini membuktikan sifat anti-kanser C3, C4, C5 dan C9 terhadap kanser payudara melalui laluan apoptosis intrinsik dan ekstrinsik seterusnya menyediakan pengetahuan asas dalam membantu pembangunan drug kemoterapeutik baru yang berasaskan penggunaan logam bismuth.



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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

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## LIST OF ABBREVIATIONS

AO/PI	Acridine orange/propidium iodide
APAF-1	Apoptosis activating factor-1
Apo2L	Apo2 ligand
Apo3L	Apo3 ligand
BAX	BCL2-Associated X Protein
Bcl-2	B-cell lymphoma 2
Bcl-XL	B-cell lymphoma-extra large
BIRC5	Baculoviral inhibitor of apoptosis repeat-containing 5
BRK	Breast tumour kinase
BSS	Bismuth subsalicylate
Calu-6	Human lung adenocarcinoma cells
CBS	Colloidal bismuth subcitrate
CDKs	Cyclin dependent kinases
CDKIs	Cyclin dependent kinase inhibitors
cFLIP	Cellular FLICE (FADD-like IL-1 $\beta$ -converting enzyme)-inhibitory protein
DCFDA	5(6)-carboxy-2',7'-dichlorofluorescein diacetate
DCIS	Ductal carcinoma in situ
DISC	Death-inducing signaling complex
DMEM	Dulbecco's Modified Eagle Medium
DMSO	Dimethyl sulfoxide
DNA	Deoxyribonucleic acid
DOTA	1,4,7,10- tetra-azacyclododecane 1,4,7,10-tetracetate
DR3	Death receptor 3

DR4	Death receptor 4
DR5	Death receptor 5
DTPA	Diethylenetriaminepenta-acetate
ECM	Extracellular matrix
EGF	Epidermal growth factor
EGFR	Epidermal growth factor receptor
ELAM-1	Endothelial leucocyte adhesion molecule-1
ER	Estrogen receptor
ETC	Electron transport chain
ErbB	Epidermal growth factor receptor
EtBr	Ethidium bromide
FADD	Fas-associated death domain protein
FasL	Fas ligand
FasR	Fas receptor
FBS	Fetal bovine serum
<i>H. pylori</i>	<i>Helicobacter pylori</i>
HER2	Human epidermal growth receptor 2
HRT	Hormone replacement therapy
HSAB	Hard-soft acid-base
IAP	Inhibitor of apoptosis protein
ICAM-1	Intercellular adhesion molecule-1
IDC	Infiltrating ductal carcinoma
IKK	I $\kappa$ B kinase
IL	Interleukin
LCIS	Lobular carcinoma in situ
LPS	Lipopolysaccharides

MAP	Mitogen-activated protein
MAPK	Mitogen-activated protein kinase
MCF-7	Human breast carcinoma cells
MPT	Mitochondrial permeability transition
MTT	3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide
NF- $\kappa$ B	Nuclear factor-kappa B
PARP	Poly (ADP-ribose) polymerase
PCD	Programmed cell death
PI3K	Phosphatidylinositol 3-kinase
PPI-based	Proton pump inhibitor-based
PS	Phosphatidylserine
PTPs	Permeability transition pores
RBC	Ranitidine bismuth citrate
ROS	Reactive oxygen species
SHH	Sonic hedgehog
TAE	Tris-acetate-EDTA
TDLU	Terminal ductal lobular units
TNF	Tumor necrosis factor
TNF- $\alpha$	Tumor necrosis factor alpha
TNFR1	Tumor necrosis factor receptor 1
TNFR2	Tumor necrosis factor receptor 2
TRADD	TNF receptor-associated death domain
Trx	Thioredoxin
TrxR1	Thioredoxin reductase 1
VCAM-1	Vascular cell adhesion molecule-1
VEGF-1	Vascular endothelial growth factor-1

XIAP

X-linked inhibitor of apoptosis



## CHAPTER 1

### INTRODUCTION

The development of cancer is a multistep process in which normal and healthy cells in the body go through stages that eventually develop into continual unregulated growth of abnormal cells. Normal cells in the body proliferate to replace worn-out cells. When cancer occurs, cells proliferate in an uncontrolled manner rather than responding appropriately to the signals that control normal cell behavior, invading normal tissues and organs and ultimately spreading throughout the body (Cooper, 2000; National Institutes of Health, 2007).

Cancer is now one of the world's largest killer, ranked top three with number of cases set to explode in coming years as reported in The World Cancer Report 2014 (Stewart and Wild, 2014). Based on GLOBOCAN 2018, cancer has become one of the biggest cause of morbidity and mortality globally with estimated 18.1 million new cases and approximately 9.6 million deaths in 2018 which responsible for nearly 1 in 6 deaths due to cancer (Bray *et al.*, 2018). Statistics released shows nearly 65% of the rise in the number of all cancer deaths will occur in less developed regions and simultaneously developing countries will bear the burden of the predicted 23.6 million new cases per year by 2030 (National Cancer Institute, 2018).

Breast cancer is the most common malignancy in women worldwide after lung cancer (Bray *et al.*, 2018). It was reported with nearly 1.7 million new cases diagnosed in 2012. This represents around 12% of all new cancer cases and 25% of all cancer cases in women (Ferlay *et al.*, 2012). Hence, the change in the global distribution of female breast cancer cases is emerging as a major health issue for women in Asia, Africa and South America.

In Asia, breast cancer incidence has been increasing rapidly in recent years. According to Moore *et al.* (2003), the disease may happen at a relatively young age. Breast cancer is a major cause of morbidity and cancer related mortality among women. Meanwhile, the breast cancer incidence was reported to be increasing in most of the Asian countries. Malaysia is also sharing the same movement.

Health Facts (2013) released by Ministry of Health (MoH) Malaysia stated that cancer is one of the top ten causes of hospitalization and one of the top five

causes of death with an estimation of 30000 cases annually. Furthermore, cancer has overtaken heart disease as the number one killer by 2014 in Malaysia. Based on the report released by Malaysian National Cancer Registry Report (2016), a total of 103507 new cancer cases were diagnosed for the period of 2007-2011 in Malaysia. The top three most common cancers among Malaysians are breast followed by colorectal and lung cancer, with one in 19 Malaysians developing breast cancer, one in 33 developing colorectal cancer and one in 40 developing lung cancer, respectively. Breast cancer appeared to be the most common cancer among female residents in Malaysia which accounted for 32.1%.

Rapid societal and economic transitions have occurred in many countries, any reduction in infection-related cancers has been offset by an increasing number of new cases related to reproductive, dietary and hormonal factors (Bray *et al.*, 2012). Numerous factors that affect the risk of breast cancer have been suggested such as decreased childbearing and breast-feeding, exposure to oestrogen, late menopause, and detrimental dietary and lifestyle changes, including high fat diet and less physical activity (Sun *et al.*, 2017).

There are several types of cancer treatments suggested by medical institutions comprise of surgery, radiotherapy, chemotherapy, endocrine therapy and targeted therapies. The most common method is chemotherapy which uses anti-neoplastic drugs to interact with cancer cells by either eradicating or controlling the growth of cancer. The first developed major metal-based antineoplastic drug is derived from platinum variants such as carboplatin, oxaliplatin, and picoplatin (Donzelli *et al.*, 2004). However, most of the therapeutic agents are associated with severe adverse effects such as significant nausea and vomiting, which indirectly leading to poor compliance with scheduled chemotherapy, eventually leading to poor outcomes.

Previous studies on the bismuth-based compounds do not only reveal the promising outcomes of these compounds in combating several cancers (Köpf-Maier and Klapötke, 1988; Bandyopadhyay *et al.*, 2012; Ishak *et al.*, 2014) but it was also found that the use of bismuth compounds in chemotherapy is able to alleviate the adverse effects of cisplatin by increasing the metallothionein production, without significantly altering the anticancer drug activity (Kondo *et al.*, 1992). *Helicobacter pylori* is believed in causing gastric lymphoma and treatment with bismuth not only resulted in regression but even a cure (Steinbach *et al.*, 1999). Hamer (1986) discovered bismuth subnitrate enhanced the bismuth distribution in kidneys and significantly protected against the toxicity of heavy metals, alkylating agents and free radicals. However, from the foregoing, it is clear that the exploration of the anti-tumor activity of bismuth-based compounds is relatively undeveloped. Hence, elucidation of bismuth dithiocarbamate compounds' cytotoxicity and possible underlying pathway(s) against breast cancer cell line were focused in the current study.



## 1.1 Problem statements

Breast cancer appears to be one of the main concern and major cause of death in women globally. Cisplatin, was the first platinum based chemotherapy drug discovered by scientists and continues its application in a wide variety of cancer treatments such as breast cancer for about forty years. Notwithstanding, there is drawback in the use of cisplatin to the cancer patients especially its toxicity profile and drug resistance by cancer cells (Kelland and Farrell, 2000; Boyiadzis, 2007; Dasari and Tchounwou, 2014). Other than that, administration of other chemotherapeutic drugs, for instance vinblastine and vincristine may cause immune system and bone marrow suppression, led to decrease amount of white blood cells, red blood cells and platelets, eventually results in anemia, neutropenia and thrombocytopenia (Krzyzanowska *et al.*, 2016). Doxorubicin has been used against cancer for more than 30 years. However, it produces a range of adverse effects such as toxicity in heart, kidney and liver (Tacar *et al.*, 2013). Lacking of safe and effective chemotherapy regimens against breast cancer, on top of limitations from current treatments including surgery, chemotherapy, radiation or targeted therapies, has led to the new development of chemotherapy regimen. Bismuth-based compound which has been found to be effective in treating a variety of diseases such as peptic ulcer and infection due to *Helicobacter pylori*, which may also lead to gastric lymphoma. Bismuth has also been found to have unusually unexpected low toxicity and is believed to act as protective agent for cancer patients from some of the toxic adverse effects caused by cisplatin, without affecting its anti-cancer activity (Yang *et al.*, 2014; Kondo *et al.*, 1992).

## 1.2 Objectives

### 1.2.1 General objectives

This study aimed at investigating the anticancer potential of four bismuth dithiocarbamate complexes with four different functional groups substitution (isopropyl ethanol, **C3**; diethyl, **C4**; pyrrolidine, **C5** and methyl ethanol, **C9**) against human breast adenocarcinoma cell lines (MCF-7) and its possible underlying molecular mechanisms.

### 1.2.2 Specific objectives

- To determine the IC<sub>50</sub> value of bismuth dithiocarbamate complexes with four different functional groups substitution (isopropyl ethanol, C3; diethyl, C4; pyrrolidine, C5 and methyl ethanol, C9) against MCF-7 cell lines.

- To assess the cell death mode induced by bismuth dithiocarbamate complexes and their morphological features.
- To delineate the underlying pathway of breast cancer cell inhibition by bismuth dithiocarbamate complexes through caspase-3, caspase-8, caspase-9, caspase- 10, cytochrome c, cell cycle and reactive oxygen species (ROS)
- To relate the involvement of Nuclear Factor-kappa B (NF-κB) in breast cancer cell inhibition by bismuth dithiocarbamate complexes via ubiquitination activity.
- To construe the cross talk signaling pathway via the gene expression analysis by Human Cancer Drug Targets RT<sup>2</sup> Profile PCR Array.

### 1.3 Hypothesis

Based on the previous preliminary studies on other cancer cell lines, it is anticipated that bismuth dithiocarbamate coordinated compounds may possess antiproliferative properties on human breast adenocarcinoma cell lines. Apoptosis pathway can be induced through intrinsic and extrinsic apoptosis pathway by mediation of multiple genes expression.

## REFERENCES

- Abbas, R. S. M., Li, y., Song, E. Y. J., Qu, C. F., Raja, C., Morgenstern, A., Apostolidis, C., Allen, B. J. (2006). Preclinical Studies of Bismuth-213 Labeled Plasminogen Activator Inhibitor Type 2 (PAI2) in a Prostate Cancer Nude Mouse Xenograft Model. *Cancer Biology & Therapy* 5(4): 386-393.
- Acehan, D., Jiang, X., Morgan, D. G., Heuser, J. E., Wang, X., Akey, C. W. (2002). Three-dimensional structure of the apoptosome: Implications for assembly, procaspase-9 binding, and activation. *Molecular Cell* 9: 423-432.
- Adhikary, A., Mohanty, S., Lahiry, L., Hossain, D. M., Chakraborty, S., Das, S. T. (2010). Theaflavins retard human breast cancer cell migration by inhibiting NF-kappaB via p53-ROS cross-talk. *FEBS Letters* 584(1): 7-14.
- Aggarwal, S., Ichikawa, H., Takada, Y., Sandur, S. K., Shishodia, S., Aggarwal, B. B. (2006). Curcumin (diferuloylmethane) down-regulates expression of cell proliferation and antiapoptotic and metastatic gene products through suppression of I kappa B alpha kinase and Akt activation. *Molecular Pharmacology* 69: 195-206.
- Al-Hussaini, H., Subramanyam, D., Reedijk, M., Sridhar, S. S. (2011). Notch Signaling Pathway as a Therapeutic Target in Breast Cancer. *Molecular Cancer Therapeutics* 10(1): 9-15.
- Alberts B, Johnson A, Lewis J, et al. (2002). *Molecular Biology of the Cell*. 4th edition. New York: Garland Science. Cancer as a Microevolutionary Process.
- Alenzi, F. Q. B. (2005). Links between Apoptosis, Proliferation and the Cell Cycle. *Journal of Continuing Education Topics & Issues* 292: 86-90.
- Alnemri, E. S., Livingston, D. J., Nicholson, D. W., Salvesen, G., Thornberry, N. A., Wong, W. W., Yuan, J. (1996). Human ICE/CED-3 protease nomenclature. *Cell* 87: 171.
- Altieri, D. C. (2001). The molecular basis and potential role of survivin in cancer diagnosis and therapy. *Trends in Molecular Medicine* 7: 542-547.
- Altieri, D. C. (2003). Survivin, versatile modulation of cell division and apoptosis in cancer. *Oncogene* 22: 8581-8589.
- Altieri, D. C. (2008). Survivin, cancer networks and pathway-directed drug discovery. *Nature Reviews Cancer* 8; 61-70.
- Altieri, D. C. (2010). Survivin and IAP proteins in cell-death mechanisms. *Biochemical Journal* 430: 199-205.

- American Cancer Society. Breast Cancer 2015. <http://www.cancer.org/cancer/breastcancer/detailedguide/breast-cancer-breast-cancer-types>. Accessed on 22 December 2015.
- Andersen, J. and Poulsen, H.S. (1989). Immunohistochemical estrogen receptor determination in paraffin-embedded tissue. Prediction of response to hormonal treatment in advanced breast cancer. *Cancer* 64: 1901-1908.
- Arendt, L. M., Keller, P. J., Skibinski, A., Goncalves, K., Naber, S. P., Buchsbaum, R. J., Gilmore, H., Come, S. E., and Kuperwasser, C. (2014). Anatomical localization of progenitor cells in human breast tissue reveals enrichment of uncommitted cells within immature lobules. *Breast Cancer Research* 16:453.
- Arnér, E. S. and Holmgren, A. (2000). Physiological Functions of Thioredoxin and Thioredoxin Reductase. *European Journal of Biochemistry* 267: 6012-6019.
- Ashkenazi, A. (2015). Targeting the extrinsic apoptotic pathway in cancer: lessons learned and future directions. *The Journal of clinical investigation* 125(2): 487-9.
- Aubele, M., Vidojkovic, S., Braselmann, H., Ritterswürden, D., Auer, G., Atkinson, M. J., Bartlett, J. M. S. (2009). Overexpression of PTK6 (breast tumor kinase) protein—a prognostic factor for long-term breast cancer survival—is not due to gene amplification. *Virchows Archiv* 455(2): 117–123.
- Badve, S., Dabbs, D. J., Schnitt, S. J., Baehner, F. L., Decker, T., Eusebi, V., Fox, S. B., Ichihara, S., Jacquemier, J., Lakhani, S. R., Palacios, J., Rakha, E. A., Richardson, A. L., Schmitt, F. C., Tan, P., Tse, G. M., Weigelt, B., Ellis, I. O., Reis-Filho, J. S. (2011). Basal-like and Triple-negative Breast Cancers. *Modern Pathology* 24(2): 157-167.
- Bandyopadhyay, D., Maldonado, S., Banik, B. K. (2012). A Microwave-Assisted Bismuth Nitrate-Catalyzed Unique Route Toward 1,4-Dihydropyridines. *Molecules* 17(3): 2643-2662.
- Bandyopadhyay, D., Mukherjee, S., Granados, J. C., Short, J. D., Banik, B. K. (2012). Ultrasound-assisted bismuth nitrate-induced green synthesis of novel pyrrole derivatives and their biological evaluation as anticancer agents. *European Journal of Medicinal Chemistry* 50: 209-215.
- Bao W, Zhu F, Duan Y, et al. HtrA1 resensitizes multidrug-resistant hepatocellular carcinoma cells by targeting XIAP. *Biomedicine & Pharmacotherapy* 2015; 70, 97-102.
- Bartok, B. and Firestein, G. S. (2010). Fibroblast-like synoviocytes: Key effector cells in rheumatoid arthritis. *Immunological Reviews* 233: 233–255.

- Baust, J. G., Gao, D., Baust, J. M. (2009). Cryopreservation. An emerging paradigm change. *Organogenesis* 5(3): 90-96.
- Baxter, G. F. (1992). Settling the stomach. *Chemistry in Britain* 28: 445-448.
- Beales, I. L. (2001). Efficacy of Helicobacter pylori eradication therapies: a single centre observational study. *BMC Gastroenterology* 1: 7.
- Bohgaki, M., Tsukiyama, T., Nakajima, A., Maruyama, S., Watanabe, M., Koike, T., Hatakeyama, S. (2008). Involvement of Ymer in suppression of NF- $\kappa$ B activation by regulated interaction with lysine-63-linked polyubiquitin chain. *Biochimica et Biophysica Acta* 1783: 826-837.
- Boidot, R., Vegran, F., Jacob, D., Chevrier, S., Gangneux, N., Taboureau, J., Oudin, C., Rainville, V., Mercier, L., Lizard-Nacol, S. (2008). The expression of BIRC5 is correlated with loss of specific chromosomal regions in breast carcinomas. *Genes Chromosomes Cancer* 47(4): 299-308.
- Born, M., Quintanilla-Fend, L., Braselmann, H., Reich, U., Richter, M., Hutzler, P., Aubele, M. (2005) Simultaneous over-expression of the Her2/neu and PTK6 tyrosine kinases in archival invasive ductal breast carcinomas. *The Journal of Pathology* 205: 592–596.
- Boyiadzis, M. M. (2007). Hematology-Oncology Therapy. 2007, New York, McGraw Hill, Medical Publishing Division.
- Bray, F., Jemal, A., Grey, N., Ferlay, J., Forman, D. (2012). Global cancer transitions according to the human development index (2008-2030): a population based study. *The Lancet Oncology* 13: 790–801.
- Bray, F., Ferlay, J., Soerjomataram, I., Siegel, R.L., Torre, L.A., Jemal, A. (2018). Global Cancer Statistics 2018: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA: A Cancer Journal for Clinicians* 68:394–424.
- Breastcancer.org. (2017). [https://www.breastcancer.org/symptoms/diagnosis/lymph\\_nodes](https://www.breastcancer.org/symptoms/diagnosis/lymph_nodes). Accessed on 29 June 2017.
- Briand, G. G. and Burford, N. (1999). Bismuth Compounds and Preparations with Biological or Medicinal Relevance. *Chemical Reviews* 99(9): 2601-2658.
- Canadian Cancer Society. (2018). The breasts. <http://www.cancer.ca/en/cancer-information/cancer-type/breast/breast-cancer/the-breasts/?region=on>. Accessed on 12 December 2018.
- Cancer Research UK (2014). What is cancer? <http://www.cancerresearchuk.org/about-cancer/what-is-cancer/how-cancer-starts/cancer-cells>. Accessed on 8 October 2015.

- Cancer Research UK (2014). Types of Cancer. <http://www.cancerresearchuk.org/about-cancer/what-is-cancer/how-cancer-starts/types-of-cancer>. Assessed on 16 October 2015.
- Cancer Research UK. (2017). <https://www.cancerresearchuk.org/about-cancer/what-is-cancer/how-cancer-can-spread>. Assessed on 4 January 2018.
- Cardone, M. H., Roy, N., Stennicke, H. R., Salvesen, G. S., Franke, T. F., Stanbridge, E., Frisch, S., Reed, J. C. (1998). Regulation of cell death protease caspase-9 by phosphorylation. *Science* 282(5392): 1318-21.
- Castro, N. E. and Lange, C. A. (2010). Breast tumor kinase and extracellular signal-regulated kinase 5 mediate Met receptor signaling to cell migration in breast cancer cells. *Breast Cancer Research* 12(4): R60.
- Chen, G., Goeddel, D. V. (2002). TNF-R1 signaling: a beautiful pathway. *Science* 296(5573): 1634-1635.
- Chen, H. Y., Shen, C. H., Tsai, Y. T., Lin, F. C., Huang, Y. P. and Chen, R. H. (2004). Brk activates rac1 and promotes cell migration and invasion by phosphorylating paxillin. *Molecular and Cellular Biology* 24(24): 10558-10572.
- Cheng, Y. and Qi, Y. (2017). Current Progresses in Metal-based Anticancer Complexes as Mammalian TrxR Inhibitors. *Anti-Cancer Agents in Medicinal Chemistry* 17(8): 1046-1069.
- Chiodino, C., Ottani, D., Fantini, F., Giannetti, A., Pincelli, C., Cesinaro, A. M., Trentini, G. P. (1999). Expression of the Novel Inhibitor of Apoptosis Survivin in Normal and Neoplastic Skin. *Journal of Investigative Dermatology* 113(3): 415-418.
- Circu, M. L. and Aw, T. Y. (2010). Reactive Oxygen Species, Cellular Redox Systems, and Apoptosis. *Free Radical Biology & Medicine* 48: 749-762.
- Cohen, J. J., Duke, R. C., Fadok, V. A., Sellins, K. S. (1992). Apoptosis and programmed cell death in immunity. *Annual Review of Immunology* 10: 267-93.
- Colditz, G. A. and Bohlke, K. (2014). Priorities for the primary prevention of breast cancer. *CA: A Cancer Journal for Clinicians* 64:186-194.
- Connolly, J., Kempson, R., LiVolsi, V., Page, D., Patchefsky, A., Silverberg, S. (2004). Recommendations for the reporting of breast carcinoma. Association of Directors of Anatomic and Surgical Pathology.
- Cooper GM. The Cell: A Molecular Approach. 2nd edition. Sunderland (MA): Sinauer Associates; 2000. The Development and Causes of Cancer. <https://www.ncbi.nlm.nih.gov/books/NBK9963/>. Accessed on 15 October 2017.

- Cross, C. E., Halliwell, B., Borish, E. T., Pryor, W. A., Ames, B. N., Saul, R. L., McCord, J. M., Harman, D. (1987). Oxygen Radicals and Human Disease. *Annals of Internal Medicine* 4: 526-545.
- Cryns, V., Yuan, J. (1998). Proteases to die for. *Genes & Development* 12(11): 1551-70.
- Dasari, S., & Tchounwou, P.B. (2014). Cisplatin in cancer therapy: molecular mechanisms of action. *European Journal of Pharmacology* 740: 364-78.
- Daugaard, G. (1990). Cisplatin nephrotoxicity: experimental and clinical studies. *Danish Medical Bulletin* 37(1): 1-12.
- David, L. S., Rapheal, R., and Rubin, E. (2008). Rubin's pathology: clinicopathologic foundations of medicine. Philadelphia: Wolters Kluwer/Lippincott Williams & Wilkins. pp. 138-139.
- Dayang Hazwani, A. I., Ooi, K. K., Ang, K. P., Akim, A. M., Cheah, Y., Nordin, N., Halim, S. N. B. A., Seng, H., Tiekink, E. R. T. (2014). A bismuth diethyldithiocarbamate compound promotes apoptosis in HepG2 carcinoma, cell cycle arrest and inhibits cell invasion through modulation of the NF- $\kappa$ B activation pathway. *Journal of Inorganic Biochemistry* 130: 38–51.
- Deng, Y. T., Huang, H. C., Lin, J. K. (2010). Rotenone Induces Apoptosis in MCF-7 Human Breast Cancer Cell-Mediated ROS through JNK and p38 Signaling. *Molecular Carcinogenesis* 49: 141-151.
- Derry, J. J., Prins, G. S., Ray, V., and Tyner, A. L. (2003). Altered localization and activity of the intracellular tyrosine kinase BRK/Sik in prostate tumor cells. *Oncogene* 22: 4212-4220.
- Donzelli, E., Carfi, M., Miloso, M., Strada, A., Galbiati, A., Bayssas, M., Griffon-Etienne, G., Caveletti, G. (2004). Neurotoxicity of platinum compounds: comparison of the effects of cisplatin and oxaliplatin on the human neuroblastoma cell line SH-SY5Y. *Journal of Neuro-oncology* 67: 65-73.
- Easty, D. J., P. J. Mitchell, K. Patel, V. A. Florenes, R. A. Spritz, and D. C. Bennett. (1997). Loss of expression of receptor tyrosine kinase family genes PTK7 and SEK in metastatic melanoma. *International Journal of Cancer* 71: 1061–1065.
- Eckhardt, B. L., Francis, P. A., Parker, B. S., Anderson, R. L. (2012). Strategies for the discovery and development of therapies for metastatic breast cancer. *Nature Reviews Drug Discovery* 11: 479–497.
- Elmore S. (2007). Apoptosis: a review of programmed cell death. *Toxicologic Pathology* 35(4): 495-516.

- Elmore, S. (2007). Apoptosis: A Review of Programmed Cell Death. *Toxicologic Pathology* 35:495–516.
- Enari, M., Sakahira, H., Yokohama, H., Okawa, K., Iwamatsu, A., and Nagata, S. (1998). A caspase-activated DNase that degrades DNA during apoptosis, and its inhibitor ICAD. *Nature* 391: 43–50.
- Fadok, V. A., Voelker, D. R., Campbell, P. A., Cohen, J. J., Bratton, D. L., Henson, P. M. (1992). Exposure of phosphatidylserine on the surface of apoptotic lymphocytes triggers specific recognition and removal by macrophages. *The Journal of Immunology* 148(7): 2207-2216.
- Ferlay, J., Soerjomataram, I., Ervik, M., Dikshit, R., Eser, S., Mathers, C., Rebelo, M., Parkin, D.M., Forman, D., Bray, F. GLOBOCAN 2012 v1.1, Cancer Incidence and Mortality Worldwide: IARC CancerBase No. 11 [Internet]. Lyon, France: International Agency for Research on Cancer; 2014. Available from: <http://globocan.iarc.fr>. Accessed on 16/01/2015.
- Ferri, K. F., Kroemer, G. (2001). Organelle-specific initiation of cell death pathways. *Nature Cell Biology* 3(11):E255-63.
- Finn, R. S., Aleshin, A., Slamon, D. J. (2016). Targeting the cyclin-dependent kinases (CDK) 4/6 in estrogen receptor-positive breast cancers. *Breast Cancer Research* 18: 17.
- Friebolin, W., Schilling, G., Zoller, M., Amtmann, E. (2005). Antitumoral activity of non-platinum xanthate complexes. *Journal of Medicinal Chemistry* 48: 7925-7931.
- Gabriel, A. (2016). Breast Anatomy. <https://reference.medscape.com/article/1273133-overview#a3>. Accessed on 12 December 2017.
- Gefen, A. and Dilmoney, B. (2007). Mechanics of the normal woman's breast. *Technology and Health Care* 15: 259–271.
- Geyer, F. C., Marchiò, C., Reis-Filho, J. S. (2009). The role of molecular analysis in breast cancer. *Pathology* 41(1): 77–88.
- Ghobrial, I. M., Witzig, T. E., Adjei, A. A. (2005). Targeting Apoptosis Pathways in Cancer Therapy. *CA: A Cancer Journal for Clinicians* 55:178–194.
- Ghosh, G., Wang, V. Y., Huang, D. B. and Fusco, A. (2012). NF-kappaB regulation: lessons from structures. *Immunological Reviews* 246: 36–58.
- Globocan. (2012). Estimate cancer incidence, mortality & prevalence worldwide in 2012. [http://globocan.iarc.fr/Pages/fact\\_sheets\\_cancer.aspx](http://globocan.iarc.fr/Pages/fact_sheets_cancer.aspx). Accessed on 3 November 2016.



- Going, J. J., and Moffat, D. F. (2004). Escaping from Flatland: clinical and biological aspects of human mammary duct anatomy in three dimensions. *The Journal of Pathology* 203: 538-544.
- Gomes, A., Fernandes, E., Lima, J. L. F. C. (2005). Fluorescence probes used for detection of reactive oxygen species. *Journal of Biochemical and Biophysical Methods* 65: 45–80.
- Graham, D. Y., and Lee, S. Y. (2015). How to Effectively Use Bismuth Quadruple Therapy: The Good, the Bad, and the Ugly. *Gastroenterology Clinics of North America* 44(3): 537-63.
- Groner, B., and Weiss, A. (2013). Targeting survivin in cancer: novel drug development approaches. *BioDrugs : clinical immunotherapeutics, biopharmaceuticals and gene therapy* 28(1): 27-39.
- Guo, Z. and Sadler, P. J. (1999). Metals in medicine. *Angewandte Chemie International Edition* 38: 1512-31.
- Gupta, S. C.; Kim, J. H.; Prasad, S., Aggarwal, B. B. (2010). Regulation of survival, proliferation, invasion, angiogenesis, and metastasis of tumor cells through modulation of inflammatory pathways by nutraceuticals. *Cancer Metastasis Review* 29: 405–434.
- Häcker, G. (2000). The morphology of apoptosis. *Cell Tissue Research* 301(1): 5-17.
- Halime, Z., Michaudet, L., Lachkar, M., Brossier, P., Boitrel, B. (2004). Influence of Pendant Arms Bearing Ligating Groups on the Structure of Bismuth Porphyrins: Implications for Labeling Immunoglobulins Used in Medical Applications. *Bioconjugate Chemistry* 15(6): 1193–1200.
- Hamer, D. H. (1986). Metallothionein. *Annual Review of Biochemistry* 55: 913-51.
- Hanahan, D., and Weinberg, R. A. (2011). Hallmarks of Cancer: The Next Generation. *Cell* 144: 646-674.
- Harvey, A. J., Pennington, C. J., Porter, S., Burmi, R. S., Edwards, D. R., Court, W., Eccles, S. A., Crompton MR. (2009). Brk protects breast cancer cells from autophagic cell death induced by loss of anchorage. *The American Journal of Pathology* 175(3): 1226-34.
- Hassfjell, S. and Brechbiel, M. W. (2001). The Development of the  $\alpha$ -Particle Emitting Radionuclides  $^{212}\text{Bi}$  and  $^{213}\text{Bi}$ , and Their Decay Chain Related Radionuclides, for Therapeutic Applications. *Chemical Reviews* 101(7): 2019–2036.

Health Facts 2013, Ministry of Health Malaysia, Health Informatics Centre Planning Division, July 2013.

Hecht, F., Pessoa, C. F., Gentile, L. B., Rosenthal, D., Carvalho, D. P., Fortunato, R. S. (2016). The Role of Oxidative Stress on Breast Cancer Development and Therapy. *Tumor Biology* 37: 4281-4291.

Helgason, C.D. and Miller, C.L. Basic Cell Culture Protocols, 3rd ed., Vol. 290 in Methods in Molecular Biology Series, series ed. J.M. Walker (Totowa, NJ: Humana Press, 2005).

Hengartner, M. O. (2001). Apoptosis. *Cell* 104(3): 325-328.

Herschkowitz, J. I., Simin, K., Weigman, V. J., Mikaelian, I., Usary, J., Hu, Z., Rasmussen, K. E., Jones, L. P., Assefnia, S., Chandrasekharan, S., Backlund, M. G., Yin, Y., Khramtsov, A. I., Bastein, R., Quackenbush, J., Glazer, R. I., Brown, P. H., Green, J. E., Kopelovich, L., Furth, P. A., Palazzo, J. P., Olopade, O. I., Bernard, P. S., Churchill, G. A., Van Dyke, T., Perou, C. M. (2007). Identification of conserved gene expression features between murine mammary carcinoma models and human breast tumors. *Genome Biology* 8(5), R76.

Hitoshi, Y., Lorens, J., Kitada, S. I., Fisher, J., LaBarge, M., Ring, H. Z., Francke, U., Reed, J. C., Kinoshita, S., Nolan, G. P. (1998). Toso, a cell surface, specific regulator of Fas-induced apoptosis in T cells. *Immunity* 8(4): 461-71.

Hsieh, C. J., Kuo, P. L., Hsu, Y. C., Huang, Y. F., Tsai, E. M., Hsu, Y. L. (2014). Arctigenin, a Dietary Phytoestrogen, Induces Apoptosis of Estrogen Receptor-Negative Breast Cancer Cells Through the ROS/p38 MAPK Pathway and Epigenetic Regulation. *Free Radical Biology & Medicine* 67: 159-170.

Hu, D., Liu, S., Shi, L., Li, C., Wu, L., Fan, Z. (2010). Cleavage of survivin by Granzyme M triggers degradation of the survivin-X-linked inhibitor of apoptosis protein (XIAP) complex to free caspase activity leading to cytolysis of target tumor cells. *The Journal of Biological Chemistry* 285(24): 18326–18335.

Ikeda, O., Miyasaka, Y., Sekine, Y., Mizushima, A., Muromoto, R., Nanbo, A., Yoshimura, A. and Matsuda, T. (2009). STAP-2 is phosphorylated at tyrosine-250 by Brk and modulates Brk-mediated STAT3 activation. *Biochemical and Biophysical Research Communications* 384(1): 71-75.

Imam, S. K. (2001). Advancements in cancer therapy with alpha-emitters: a review. *International Journal of Radiation Oncology, Biology, Physics* 51(1): 271–278.

Jaiswal, P. K., Goel, A., and Mittal, R. D. (2015). Survivin: A molecular biomarker in cancer. *The Indian Journal of Medical Research* 141(4): 389–397.

- Jiang, B. P., Le, L., Xu, L. J., Xiao, P. G. (2014). Minocycline inhibits ICAD degradation and the NF- $\kappa$ B activation induced by 6-OHDA in PC12 cells. *Brain Research* 1586: 1-11.
- John, R. (2010). General Guide for Cryogenically Storing Animal Cell Cultures. Corning Technical Bulletin.
- Julian, L., and Olson, M. F. (2015). Apoptotic membrane dynamics in health and disease. *Cell Health and Cytoskeleton* 7: 133–142.
- Jurcic, J. G., Larson, S. M., Sgouros, G., McDevitt, M. R., Finn, R. D., Divgi, C. R. (2002). Targeted  $\alpha$  particle immunotherapy for myeloid leukemia. *Blood* 100: 1233-39.
- Kamalati, T., Jolin, H. E., Mitchell, P. J., Barker, K. T., Jackson, L. E., Dean, C. J., Page, M. J., Gusterson, B. A., Crompton, M. R. (1996). Brk, a breast tumor-derived non-receptor protein-tyrosine kinase, sensitizes mammary epithelial cells to epidermal growth factor. *The Journal of Biological Chemistry* 271(48): 30956-63.
- Kamdje, A. H. N., Etet, P. F. S., Vecchio, L., Tagne, R. S., Amvene, J. M., Muller, J. M., Krampera, M., Lukong, K. E. (2014). New targeted therapies for breast cancer: A focus on tumor microenvironmental signals and chemoresistant breast cancers. *World Journal of Clinical Cases* 2(12): 769-86.
- Kapuscinski, J., Darzynkiewicz, Z., Melamed, M. R. (1983). Interactions of acridine orange with nucleic acids. Properties of complexes of acridine orange with single stranded ribonucleic acid. *Biochemical Pharmacology* 32(24): 3679-94.
- Karin, M., Cao, Y., Greten, F. R., Li, Z. W. (2002). NF- $\kappa$ B in cancer: From innocent bystander to major culprit. *Nature Reviews Cancer* 2: 301–310.
- Karin, M. and Delhase, M. (2000). The I kappa B kinase (IKK) and NF-kappa B: key elements of proinflammatory signalling. *Seminars in Immunology* 12(1): 85-98.
- Karin, M. and Lin, A. (2002). NF-kappaB at the crossroads of life and death. *Nature Immunology* 3(3): 221-7.
- Karp, G. (2008). Cell and molecular biology: Concepts and experiments 5th edition with study. *John Wiley and Sons* 653–7.
- Kasibhatla, S. and Tseng, B. (2003). Why target apoptosis in cancer treatment? *Molecular Cancer Therapeutics* 2(6):573-80.
- Kataoka, T., Schroter, M., Hahne, M., Schneider, P., Irmeler, M., Thome, M., Froelich, C. J., Tschopp, J. (1998). FLIP Prevents Apoptosis Induced by

- Death Receptors But Notby Perforin/Granzyme B, Chemotherapeutic Drugs, and Gamma Irradiation. *The Journal of Immunology* 161: 3936–3942.
- Kaufmann, S. H. and Hengartner, M. O. (2001). Programmed cell death: alive and well in the new millennium. *Trends in Cell Biology* 11(12): 526-34.
- Kelland, L. R., Farrell, N.P. (2000). Platinum-based Drugs in Cancer Therapy. Humana Press, Totawa, New Jersey.
- Kelly, S. L., Basu, A., Teicher, B. A., Hacker, M. P., Hamer, D. H. and Lazo, J. S. (1988). Overexpression of metallothionein confers resistance to anticancer drugs. *Science* 241: 1813-1815.
- Kerr, J. F., Wyllie, A. H., Currie, A. R. (1972). Apoptosis: a basic biological phenomenon with wide-ranging implications in tissue kinetics. *British Journal of Cancer* 26(4): 239-57.
- Kim, S. J., Miyoshi, Y., Taguchi, T., Tamaki, Y., Nakamura, H., Yodoi, J., Kato, K., Noguchi, S. (2005). High thioredoxin expression is associated with resistance to docetaxel in primary breast cancer. *Clinical Cancer Research* 11: 8425–8430.
- King, C. R., Kraus, M. H., Aaronson, S. A. (1985). Amplification of a novel v-erbB-related gene in a human mammary carcinoma. *Science* 229: 974–976.
- Kondo, Y., Himeno, S., Satoh, M., Naganum, A., Nishimura, T., Imura, N. (2004). Citrate enhances the protective effect of orally administered bismuth subnitrate against the nephrotoxicity of cis-diamminedichloroplatinum. *Cancer Chemotherapy and Pharmacology* 53(1): 33-38.
- Kondo, Y., Satoh, M., Imura, N., Akimoto, M. (1991). Effect of bismuth nitrate given in combination with cis-diamminedi-chloroplatinum(II) on the antitumor activity and renal toxicity of the latter in nude mice inoculated with human bladder tumor. *Cancer Chemotherapy and Pharmacology* 29:19.
- Kondo, Y., Satoh, M., Imura, N., Akimoto, M. (1992). Tissue-specific induction of metallothionein by bismuth as a promising protocol for chemotherapy with repeated administration of cisdiamminedichloro-platinum (II) against bladder tumor. *Anticancer Research* 12: 2303-2307.
- Konturek, S. J., Radecki, T., Piastucki, I., Brzozowski, T., Drozdowicz, D. (1987). Gastroprotection by colloidal bismuth subcitrate (DE-NOL) and sucralfate. Role of endogenous prostaglandins. *Gut* 28: 201-5.
- Köpf-Maier, P., Klapötke, T. (1988). Antitumor activity of some organometallic bismuth(III)thiolates. *Inorganica Chimica Acta* 152(1): 49-52.
- Krammer, P. H. (1999). CD95(APO-1/Fas)-Mediated Apoptosis: Live and Let Die. *Advances in Immunology* 71: 163–210.

- Kroemer, G., El-Deiry, W. S., Golstein, P., Peter, M. E., Vaux, D., Vandenabeele, P., Zhivotovsky, B., Blagosklonny, M. V., Malorni, W., Knight, R. A., Piacentini, M., Nagata, S., Melino, G. (2005). Classification of cell death: recommendations of the Nomenclature Committee on Cell Death. *Cell Death & Differentiation* 12(2): 1463-7.
- Kroemer, G., Galluzzi, L., Brenner, C. (2007). Mitochondrial membrane permeabilization in cell death. *Physiological Reviews* 87(1): 99-163.
- Krzyzonowska, M. K., Walker-Diks, C., Morris, A. M., Gupta, R., Halligan, R., Kouroukis, C. T., McCann, K., and Atzema, C. L. (2016). Approach to evaluation of fever in ambulatory cancer patients receiving chemotherapy: A systematic review. *Cancer Treatment Reviews*. 51: 35-45.
- Kuschinsky, G. and Lullmann, H. (1974). 'Lehrbuch der Pharmakologie', Georg Thieme Verlag, Stuttgart.
- Lambert, J.R. and Midolo, P. (1997). The actions of bismuth in the treatment of Helicobacter pylori infection. *Alimentary Pharmacology & Therapeutics* 11(1): 27-33.
- Landis, M. W., Pawlyk, B. S., Li, T., Sicinski, P., Hinds, P. W. (2006). Cyclin D1-dependent kinase activity in murine development and mammary tumorigenesis. *Cancer Cell* 9: 13-22.
- Laura, M., Maria, A. C., Elena, M., Gloriano, M., Stefania, N., Ida, L., Enrico, M., Annalisa, G., Chiara, G., and Luigi, M. (2011). Structural and solution chemistry, protein binding and antiproliferative profiles of gold (I)/(III) complexes bearing the saccharinato ligand. *Journal of Inorganic Biochemistry* 105: 348-355.
- LeBel, C. P., Ischiropoulos, H., Bondy, S. C. (1992). Evaluation of the probe 2',7'-dichlorofluorescein as an indicator of reactive oxygen species formation and oxidative stress. *Chemical Research in Toxicology* 5: 227-231.
- Le Bras, M., Clément, M. V., Pervaiz, S., Brenner, C. (2005). Reactive oxygen species and the mitochondrial signalling pathway of cell death. *Histology and Histopathology* 20: 205-219.
- Lester, S. C., Bose, S., Chen, Y. Y., Connolly, J. L., de Baca, M. E., Fitzgibbons, P. L. (2009). Protocol for the examination of specimens from patients with invasive carcinoma of the breast. *Archives of Pathology & Laboratory Medicine* 133: 1515-38.
- Liang, H., Zhang, Y., Shi, X., Wei, T., & Lou, J. (2014). Role of Notch-1 signaling pathway in PC12 cell apoptosis induced by amyloid beta-peptide (25-35). *Neural Regeneration Research* 9(13): 1297-302.

- Li, F., He, Z., Shen, J., Huang, Q., Li, W., Liu, X., He, Y., Wolf, F., Li, C. Y. (2010). Apoptotic caspases regulate induction of iPSCs from human fibroblasts. *Cell Stem Cell* 7: 508-520.
- Linke, S. P., Clarkin, K. C., Di Leonardo, A., Tsou, A., Wahl, G. M. (1996). A reversible, p53-dependent G0/G1 cell cycle arrest induced by ribonucleotide depletion in the absence of detectable DNA damage. *Genes & Development* 10(8): 934-47.
- Liu, Y., Li, Y., Yu S., and Zhao, G. (2012). Recent Advances in the Development of Thioredoxin Reductase Inhibitors as Anticancer Agents. *Current Drug Targets* 13: 1432-1444.
- Llor, X., Serfas, M. S., Bie, W., Vasioukhin, V., Polonskaia, M., Derry, J., Abbott, C. M., and Tyner, A. L. (1999). BRK/Sik expression in the gastrointestinal tract and in colon tumors. *Clinical cancer research: an official journal of the American Association for Cancer Research* 5: 1767-1777.
- Logue, S. E., and Martin, S. J. (2008). Caspase activation cascades in apoptosis. *Biochemical Society Transactions* 36(1): 1-9.
- Ludyga, N., Anastasov, N., Gonzalez-Vasconcellos, I., Ram, M., Hofler, H., Aubele, M. (2011). Impact of protein tyrosine kinase 6 (PTK6) on human epidermal growth factor receptor (HER) signalling in breast cancer. *Molecular BioSystems* 7: 1603–1612.
- Majno, G. and Joris, I. (1995). Apoptosis, oncosis, and necrosis. An overview of cell death. *The American Journal of Pathology* 146(1): 3-15.
- Malaysia National Cancer Registry Report 2007-2011 Malaysia Cancer Statistics, Data and Figure. (2016). Azizah, A.M., Saleha, I.T., Hashimah, A., Asmah, Z.A., Mastulu, W. Ministry of Health.
- Malhotra, G. K., Zhao, X., Band, H. and Band, V. (2010). Histological, molecular and functional subtypes of breast cancers. *Cancer Biology & Therapy* 10(10): 955-960.
- Margulis, V., Lotan, Y. and Shariat, S. F. (2008). Survivin: a promising biomarker for detection and prognosis of bladder cancer. *World Journal of Urology* 26: 59–65.
- Marino, G. and Kroemer, G. (2013). Mechanisms of apoptotic phosphatidylserine exposure. *Cell Research* 23: 1247-1248.
- Martin, T. A., Ye, L., Sanders, A. J., Lane, J., Jiang, W. G. (2013). Cancer Invasion and Metastasis: Molecular and Cellular Perspective. In: Madame Curie Bioscience Database [Internet]. Austin (TX): Landes Bioscience; 2000-2013. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK164700/>

- Massagué, J. and Obenauf, A. C. 2016. Metastatic colonization. *Nature* 529(7586): 298-306.
- Maxwell, G. P. and Gabriel, A. (2009). The evolution of breast implants. *Clinics in Plastic Surgery* 36(1):1-13.
- McCubrey, J. A., Lertpiriyapong, K., Fitzgerald, T. L. (2017). Roles of TP53 in determining therapeutic sensitivity, growth, cellular senescence, invasion and metastasis. *Advances in Biological Regulation* 63: 32-48.
- McDevitt, M. R., Finn, R. D., Sgouros, G., Ma, D., Scheinberg, D. A. (1999). An  $^{225}\text{Ac}/^{213}\text{Bi}$  generator system for therapeutic clinical applications: construction and operation. *Applied Radiation and Isotopes* 50(5): 895-904.
- McDevitt, M. R., Nikula, T. N., Finn, R. D., Curcio, M. J., Gansow O. A., Geerlings, M. W. Sr., Larson, S. M., Scheinberg, D. A. (1996). Bismuth labelled antibodies for therapy of leukemias, lymphomas, and carcinomas: preclinical studies. *Tumor Targeting* 2: 182.
- McGuire, S. (2016). World Cancer Report 2014. Geneva, Switzerland: World Health Organization, International Agency for Research on Cancer, WHO Press, 2015. *Advances in nutrition* (Bethesda, Md.), 7(2), 418-9.
- Miah, S., Martin, A., & Lukong, K. E. (2012). Constitutive activation of breast tumor kinase accelerates cell migration and tumor growth in vivo. *Oncogenesis* 1(5): e11.
- Mincey, B. A. (2003). Genetics and the Management of Women at High Risk for Breast Cancer. *The Oncologist* 8(5): 466-473.
- Mitchell, P. J., Barker, K. T., Martindale, J. E., Kamalati, T., Lowe, P. N., Page, M. J., Gusterson, B. A., Crompton, M. R. (1994). Cloning and characterisation of cDNAs encoding a novel non-receptor tyrosine kinase, brk, expressed in human breast tumours. *Oncogene* 9(8):2383-90.
- Mohan, S., Abdul, A. B., Abdelwahab, S. I., Al-Zubairi, A. S., Sukari, M. A., Abdullah, R., Elhassan Taha M. M., Ibrahim, M. Y., Syam, S. (2010). Typhonium flagelliforme induces apoptosis in CEMss cells via activation of caspase-9, PARP cleavage and cytochrome C release: Its activation coupled with G0/G1 phase cell cycle arrest. *Journal of Ethnopharmacology* 131: 592–0.
- Moore, M. A., Kazuo, T., Anh, P. H., Aydemir, G., Basu, P. S., Bhurgri, Y., Chen, K., Gajalakshmi, V., Hirose, K., Jarrahi, A. M., Ngoan le T., Qiao, Y. L., Shin, H. R., Sriamporn, S., Srivatanakul, P., Tokudome, S., Yoo, K.Y., Tsuda, H. (2003). Grand challenges in global health and the practical prevention program? Asian focus on cancer prevention in females of the developing world. *Asian Pacific Journal of Cancer Prevention* 4: 153-65.

- Morgan, M. J., and Liu, Z. G. (2010). Crosstalk of reactive oxygen species and NF- $\kappa$ B signaling. *Cell Research* 21(1): 103-15.
- Mulford, D. A., Scheinberg, D. A., Jurcic, J. G. (2005). The promise of targeted  $\alpha$ -particle therapy. *Journal of Nuclear Medicine* 46(1): 199S-204S.
- Nadji, M., Gomez-Fernandez, C., Ganjei-Azar, P., Morales, A.R. (2005). Immunohistochemistry of estrogen and progesterone receptors reconsidered: experience with 5993 breast cancers. *American Journal of Clinical Pathology* 123: 21-27.
- Natalija, D.P., Ana, K., Damir, V. (2012). Role of HER2 signaling pathway in breast cancer: biology, detection and therapeutical implications. *Periodicum Biologorum* 114(4): 505-510.
- National Cancer Institute (2018). Cancer Statistics. <https://www.cancer.gov/about-cancer/understanding/statistics>. Accessed on 30 November 2018.
- National Institutes of Health (US); Biological Sciences Curriculum Study. NIH Curriculum Supplement Series [Internet]. Bethesda (MD): National Institutes of Health (US); 2007. Understanding Cancer. <https://www.ncbi.nlm.nih.gov/books/NBK20362/>. Accessed on 15 October 2017.
- Nielsen, D. L., Kümler, I., Palshof, J. A., Andersson, M. (2013). Efficacy of HER2-targeted therapy in metastatic breast cancer. Monoclonal antibodies and tyrosine kinase inhibitors. *Breast* 22(1): 1-12.
- Nie, Y., Wu, H., Sha, W., Du, H., Dai, S., Wang, H., Li, Q. (1999). Colloidal Bismuth Pectin: An Alternative to Bismuth Subcitrate for the Treatment of Helicobacter pylori- Positive Duodenal Ulcer. *Helicobacter* 4(2): 128-134.
- Norbury, C. J. and Zhivotovsky, B. (2004). DNA damage-induced apoptosis. *Oncogene* 23(16): 2797-808.
- Nowell, P.C. (1976). The clonal evolution of tumor cell populations. *Science* 194(4260): 23-8.
- Nurse, P. (2002). Cyclin Dependent Kinases and Cell Cycle Control. *ChemBioChem* 3(7): 596-603.
- O'Brien, M. A. and Kirby, R. (2008). Apoptosis: A review of pro-apoptotic and anti-apoptotic pathways and dysregulation in disease. *Journal of Veterinary Emergency and Critical Care* 18(6): 572-585.
- O'Connor, C. M. and Adams, J. U. (2010). Essentials of Cell Biology. Cambridge, MA: NPG Education.



- O'Neill, C. F., Urs, S., Cinelli, C., Lincoln, A., Nadeau, R. J., León, R., Toher, J., Mouta-Bellum, C., Friesel, R. E., ... Liaw, L. (2007). Notch2 signaling induces apoptosis and inhibits human MDA-MB-231 xenograft growth. *The American Journal of Pathology* 171(3): 1023-36.
- Ocker, M. and Höpfner, M. (2012). Apoptosis-Modulating Drugs for Improved Cancer Therapy. *European Surgical Research* 48: 111–120.
- Odier, L. (1786). Observations on the effects of the bismuth magisterium. *Journal of Medicine, Surgery and Pharmacy* 68: 49-56.
- Orphanos, G., and Kountourakis, P. (2012). Targeting the HER2 receptor in metastatic breast cancer. *Hematology/Oncology and Stem Cell Therapy* 5(3): 127-37.
- Ostrander, J. H., Daniel, A. R., & Lange, C. A. (2010). Brk/PTK6 signaling in normal and cancer cell models. *Current Opinion in Pharmacology* 10(6): 662-9.
- Overall, C. M. and Lopez-Otin, C. (2002). Strategies for MMP inhibition in cancer: Innovations for the post-trial era. *Nature Reviews Cancer* 2: 657–672.
- Pang, Y., Qin, G., Wu, L., Wang, X., Chen, T. (2016). Artesunate induces ROS-dependent apoptosis via a Bax-mediated intrinsic pathway in Huh-7 and Hep3B cells. *Experimental Cell Research* 347: 251-260.
- Parajuli, B., Lee, H. G., Kwon, S. H., Cha, S. D., Shin, S. J., Lee, G. H., Bae, I., Cho, C. H. (2013). Salinomycin inhibits Akt/NF-κB and induces apoptosis in cisplatin resistant ovarian cancer cells. *Cancer Epidemiology* 37(4): 512-7.
- Park M. H. and Hong J. T. (2016) Roles of NF-κB in Cancer and Inflammatory Diseases and Their Therapeutic Approaches. *Cells* 5(2): 15.
- Parr, C., Watkins, G., Jiang, W. G. (2004). The possible correlation of Notch-1 and Notch-2 with clinical outcome and tumour clinicopathological parameters in human breast cancer. *International Journal of Molecular Medicine* 14(5): 779-86.
- Perou, C. M. (2011). "Molecular Stratification of Triple-Negative Breast Cancers". *The Oncologist* 16: 61–70.
- Perou, C. M., Sorlie, T., Eisen, M. B., van de Rijn, M., Jeffrey, S. S., Rees, C.A. (2000). Molecular portraits of human breast tumours. *Nature* 406:747-52.
- Pickup, M. W., Mouw, J. K., Weaver, V. M. (2014). The extracellular matrix modulates the hallmarks of cancer. *EMBO Reports* 15: 1243–1253.

- Place, A. E., Jin Huh, S., & Polyak, K. (2011). The microenvironment in breast cancer progression: biology and implications for treatment. *Breast Cancer Research* 13(6): 227.
- Plavetić, N. D., Kulić, A., Vrbanec, D. (2012). Role of HER2 signaling pathway in breast cancer: Biology, detection and therapeutical implications. *Periodicum Biologorum* 114(4): 505–510.
- Polyak, K., Kato, J. Y., Solomon, M. J., Sherr, C. J., Massague, J., Roberts, J. M., Koff, A. (1994). p27Kip1, a cyclin-Cdk inhibitor, links transforming growth factor-beta and contact inhibition to cell cycle arrest. *Genes & Development* 8(1): 9-22.
- Prasad, S., Ravindran, J., Aggarwal, B. B. NF- $\kappa$ B and cancer: How intimate is this relationship. *Molecular and Cellular Biochemistry* 336: 25–37.
- Prat, A., Parker, J.S., Karginova, O., Fan, C., Livasy, C., Herschkowitz, J.I., He, X. and Perou, C. M. (2010). Phenotypic and molecular characterization of the claudin-low intrinsic subtype of breast cancer. *Breast Cancer Research* 12: R68.
- Pusztai, L., Mazouni, C., Anderson, K., Wu, Y. and Symmans, W. F. (2006). Molecular Classification of Breast Cancer: Limitations and Potential. *The Oncologist* 11(8): 868-877.
- Qin, L. F. and Ng, I. O. (2002). Induction of apoptosis by cisplatin and its effect on cell cycle-related proteins and cell cycle changes in hepatoma cells. *Cancer Letters* 175(1): 27-38.
- Rahman, M., and Mohammed, S. (2015). Breast cancer metastasis and the lymphatic system. *Oncology letters* 10(3): 1233-1239.
- Reed, J. C. (1997). Bcl-2 family proteins: regulators of apoptosis and chemoresistance in hematologic malignancies. *Semin Hematol* 34(4 Suppl 5): 9-19.
- Reedijk, M. (2012). Notch signaling and breast cancer. *Advances in Experimental Medicine and Biology* 727: 241-57.
- Ricci, J. E., Gottlieb, R. A., and Green, D. R. (2003). Caspase-mediated loss of mitochondrial function and generation of reactive oxygen species during apoptosis. *The Journal of Cell Biology* 160: 65–75.
- Ricci, J. E., Munoz-Pinedo, C., Fitzgerald, P., Bailly-Maitre, B., Perkins, G. A., Yadava, N., Scheffler, I. E., Ellisman, M. H., and Green, D. R. (2004). Disruption of mitochondrial function during apoptosis is mediated by caspase cleavage of the p75 subunit of complex I of the electron transport chain. *Cell* 117: 773–786.

- Rizzo, P., Osipo, C., Foreman, K., Golde, T., Osborne, B., Miele, L. (2008). Rational targeting of Notch signaling in cancer. *Oncogene* 27(38): 5124-31.
- Rouzier, R., Perou, C.M., Symmans, W.F., Ibrahim, N., Cristofanilli, M., Anderson, K., Hess, K.R., Stec, J., Ayers, M., Wagner, P., Morandi, P., Fan, C., Rabiul, I., Ross, J. S., Hortobagyi, G. N., Pusztai, L. (2005). Breast cancer molecular subtypes respond differently to preoperative chemotherapy. *Clinical Cancer Research* 11: 5678–5685.
- Roy, S., & Nicholson, D. W. (2000). Cross-talk in cell death signaling. *The Journal of experimental medicine* 192(8): F21-5.
- Russo, J., Rivera, R., Russo, I. H. (1992). Influence of age and parity on the development of the human breast. *Breast Cancer Research and Treatment* 23: 211-218.
- Russo, J. and Russo, I. H. (2004). Development of the human breast. *Maturitas* 49: 2-15.
- Saelens, X., Festjens, N., Vande Walle, L., van Gorp, M., van Loo G, Vandenabeele, P. (2004). Toxic proteins released from mitochondria in cell death. *Oncogene* 23(16): 2861-74.
- Saleh, A., Srinivasula, S. M., Acharya, S., Fishel, R., Alnemri, E. S. (1999). Cytochrome c and dATP-mediated oligomerization of Apaf-1 Is a prerequisite for procaspase-9 activation. *The Journal of Biological Chemistry* 274: 17941–17945.
- Sarvothaman, S., Undi, R. B., Pasupuleti, S. R., Gutti, U., & Gutti, R. K. (2015). Apoptosis: role in myeloid cell development. *Blood Research* 50(2): 73-9.
- Sato, S., Sanjo, H., Takeda, K., Ninomiya-Tsuji, J., Yamamoto, M., Kawai, T., Matsumoto, K., Takeuchi, O., Akira, S. (2005). *Nature Immunology* 6(11):1087-1095.
- Scaffidi, C., Schmitz, I., Zha, J., Korsmeyer, S. J., Krammer, P. H., Peter, M. (1999). Differential modulation of apoptosis sensitivity in CD95 type I and type II cells. *The Journal of Biological Chemistry* 274: 22532–22538.
- Schmandt, R. E., Bennett, M., Clifford, S., Thornton, A., Jiang, F., Broaddus, R. R., Sun, C. C., Lu, K. H., Sood, A. K., and Gershenson, D. M. (2006). The BRK tyrosine kinase is expressed in high-grade serous carcinoma of the ovary. *Cancer Biology & Therapy* 5: 1136-1141.
- Schweitzer, V. G. (1993). Cisplatin-induced ototoxicity: the effect of pigmentation and inhibitory agents. *Laryngoscope* 103(4 Pt 2): 1-52.
- Sethi, G., Sung, B., Aggarwal, B. B. (2008). Nuclear factor-kappaB activation: from bench to bedside. *Experimental Biology and Medicine* 233(1): 21-31.

- Shariat, S. F., Ashfaq, R., Karakiewicz, P. I., Saeedi, O., Sagalowsky, A. I., Lotan, Y. (2007). Survivin expression is associated with bladder cancer presence, stage, progression, and mortality. *Cancer* 109: 1106–1113.
- Shibuya, Y., Tanimoto, H., Umeda, M., Yokoo, S., Komori, T. (2004). Induction Chemotherapy with Docetaxel, Cisplatin and 5-fluorouracil for Tongue Cancer. *Kobe Journal of Medical Sciences* 50(1): 1-7.
- Shim, J. H., Xiao, C., Paschal, A. E., Bailey, S. T., Rao, P., Hayden, M. S., Lee, K. Y., Bussey, C., Steckel, M., Tanaka, N., Yamada, G., Akira, S., Matsumoto, K., ... Ghosh, S. (2005). TAK1, but not TAB1 or TAB2, plays an essential role in multiple signaling pathways in vivo. *Genes & development* 19(22): 2668-2681.
- Skinner, S. M., Lewis, R. W. (1977). Anti leukemia activity (L1210) of 6-mercaptopurine and its metallo complexes in mice. *Research Communications in Chemical Pathology and Pharmacology* 16(1): 183-6.
- Skulachev, V. P. (1996). Why are mitochondria involved in apoptosis? Permeability transition pores and apoptosis as selective mechanisms to eliminate superoxide-producing mitochondria and cell. *FEBS Letters* 397: 7–10.
- Slamon, D. J., Clark, G. M., Wong, S. G., Levin, W. J., Ullrich, A., McGuire, W. L. (1987). Human breast cancer: correlation of relapse and survival with amplification of the HER-2/neu oncogene. *Science* 235(4785): 177-82.
- Slee, E. A., Adrain, C., Martin, S. J. (2000). Executioner caspase-3, -6, and -7 perform distinct, non-redundant roles during the demolition phase of apoptosis. *The Journal of Biological Chemistry* 276(10): 7320-6.
- Slamon, D.J., Godolphin, W., Jones, L.A., Holt, J.A., Wong, S. G., Keith, D. E., Levin, W. J., Stuart, S. G., Udove, J., Ullrich, A. et al. (1989). Studies of the HER-2/neu proto-oncogene in human breast and ovarian cancer. *Science* 244: 707-712.
- Smith, S. M., Lyu, Y. L., Cai, L. (2014). NF-κB Affects Proliferation and Invasiveness of Breast Cancer Cells by Regulating CD44 Expression. *PLoS ONE* 9(9): e106966.
- Sorlie, T., Perou, C. M., Tibshirani, R., Aas, T., Geisler, S., Johnsen, H. (2001). Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications. *Proceedings of the National Academy of Sciences of the United States of America* 98:10869-74.
- Sorlie, T., Tibshirani, R., Parker, J., Hastie, T., Marron, J. S., Nobel, A. (2003). Repeated observation of breast tumor subtypes in independent gene expression data sets. *Proceedings of the National Academy of Sciences of the United States of America* 100: 8418-23.

- Sovak, M. A., Bellas, R. E., Kim, D. W., Zanieski, G. J., Rogers, A. E., Traish, A. M., Sonenshein, G. E. (1997). Aberrant nuclear factor- $\kappa$ B/rel expression and the pathogenesis of breast cancer. *Journal of Clinical Investigation* 100: 2952–2960.
- Steinbach, G., Ford, R., Guber, G., Sample, D., Hagemester, F. B., Lynch, P. M., McLaughlin, P. W., Rodriguez, M. A., Romaguera, J. E., Sarris, A. H., Younes, A., Luthra, R., Manning, J. T., Johnson, C. M., Lahoti, S., Shen, Y., Lee, J. E., Winn, R. J., Genta, R. M., Graham, D. Y., Cabanillas, F. F. (1999). Antibiotic treatment of gastric lymphoma of mucosa-associated lymphoid tissue. An uncontrolled trial. *Annals of Internal Medicine* 131(2): 88-95.
- Stewart, B. W. and Wild, C.W. (2014). World Cancer Report 2014. International Agency for Research on Cancer.
- Stierer, M., Rosen, H., Weber, R., Hanak, H., Spona, J., Tuchler, H. (1993). Immunohistochemical and biochemical measurement of estrogen and progesterone receptors in primary breast cancer. Correlation of histopathology and prognostic factors. *Annals of Surgery* 218: 13-21.
- Stingl, J., and Caldas, C. (2007). Molecular heterogeneity of breast carcinomas and the cancer stem cell hypothesis. *Nature Reviews Cancer* 7: 791-9.
- Strasser, A., O'Connor, L., and Dixit, V. M. (2000). Apoptosis signalling. *Annual Review of Biochemistry* 69: 217-45.
- Suliman, A., Lam, A., Datta, R., Srivastava, R. K. (2001). Intracellular mechanisms of TRAIL: apoptosis through mitochondrial-dependent and -independent pathways. *Oncogene* 20: 2122–33.
- Sun, H., Li, H., Harvey, I., Sadler, P. J. (1999). Interactions of Bismuth Complexes with Metallothionein(II). *The Journal of Biological Chemistry* 274(41): 29094-29101.
- Sun, Y. S., Zhao, Z., Yang, Z. N., Xu, F., Lu, H. J., Zhu, Z. Y., Shi, W., Jiang, J., Yao, P. P., Zhu, H. P. (2017). Risk Factors and Preventions of Breast Cancer. *International Journal of Biological Sciences* 13(11): 1387-1397.
- Sylvester, P. W. (2011). Optimization of the tetrazolium dye (MTT) colorimetric assay for cellular growth and viability. *Methods in Molecular Biology* 716: 157-68.
- Tacar, O., Sriamornsak, P., Dass, C.R. (2013). Doxorubicin: an update on anticancer molecular action, toxicity and novel drug delivery systems. *Journal of Pharmacy and Pharmacology* 65(2): 157-70.
- Taherian-Fard, A., Srihari, S., Ragan, M. A. (2015). Breast cancer classification: linking molecular mechanisms to disease prognosis. *Briefings in Bioinformatics* 16(3): 461-474.

- Tamaian, R., Mot, A., Silaghi-Dumitrescu, R., Ionut, I., Stana, A., Oniga, O., Nastasa, C., Benedec, D., Tiperciuc, B. (2015). Study of the Relationships between the Structure, Lipophilicity and Biological Activity of Some Thiazolyl-carbonyl-thiosemicarbazides and Thiazolyl-azoles. *Molecules* 20: 22188–22201.
- Tiekink, E. R. (2002). Antimony and bismuth compounds in oncology. *Critical Reviews in Oncology/Hematology* 42: 217–24.
- Tonissen, K. F. and Di Trapani, G. (2009). Thioredoxin system inhibitors as mediators of apoptosis for cancer therapy. *Molecular Nutrition & Food Research* 53: 87-103.
- Toyoshima, H. and Hunter, T. (1994). p27, a novel inhibitor of G1 cyclin-Cdk protein kinase activity, is related to p21. *Cell* 78(1): 67-74.
- Tsujimoto, Y. and Shimizu, S. (2007). Role of the mitochondrial membrane permeability transition in cell death. *Apoptosis* 12(5): 835-840.
- Tu, D., Zhu, Z., Zhou, A. Y., Yun, C., Lee, K. Y., Yoms, A. V., Li, Y., Dunn, G. P., Chan, E., Thai, T., Yang, S., Ficarro, S. B., Marto, J. A., Jeon, H., Hahn, W. C., Barbic, D. A., Eck, M. J. (2013). Structure and ubiquitination-dependent activation of TANK-binding kinase 1. *Cell Reports* 3: 747-758.
- Uemura, N., Okamoto, S., Yamamoto, S., Matsumura, N., Yamaguchi, S., Yamakido, M., Taniyama, K., Sasaki, N., Schlemper, R. J. (2001). *The New England Journal of Medicine* 345(11): 784-9.
- Ukaji, T., Umezawa, K. (2014). Novel approaches to target NF- $\kappa$ B and other signaling pathways in cancer stem cells. *Advances in Biological Regulation* 56: 108-115.
- Verweij, J., de Wit, R. and de Mulder, P. H. (1996). Optimal control of acute cisplatin-induced emesis. *Oncology* 53 (1): 56-64.
- Vogelstein, B., Kinzler, K. W. (2004). Cancer genes and the pathways they control. *Nature Medicine* 10: 789-99.
- Wahab, S. I. A., Abdul, A. B., Alzubairi, A. S., Elhassan, M. M., Mohan, S. (2009). In Vitro Ultramorphological Assessment of Apoptosis Induced by Zerumbone on (HeLa). *Journal of Biomedicine and Biotechnology* 10 pages.
- Wajant, H. (2002). The Fas signaling pathway: more than a paradigm. *Science* 296(5573): 1635-6.
- Walczak, H., Bouchon, A., Stahl, H., Krammer, P. H. (2000). Tumor necrosis factor-related apoptosis-inducing ligand retains its apoptosis-inducing capacity on Bcl-2- or Bcl-xL-overexpressing chemotherapy-resistant tumor cells. *Cancer Research* 60: 3051-3057.

- Wang, C., Deng, L., Hong, M., Akkaraju, G. R., Inoue, J., Chen, Z. J. (2001). TAK1 is a ubiquitin-dependent kinase of MKK and IKK. *Nature* 412(6844): 346-351.
- Yang, Y., Ouyang, R., Xu, L., Guo, N., Li, W., Feng, K., Ouyang, L., Yang, Z., Zhou, S., Miao, Y. (2014). Review: Bismuth complexes: synthesis and applications in biomedicine. *Journal of Coordination Chemistry* 68(3): 379-397.
- Yu, Q., Sicinska, E., Geng, Y., Ahnstrom, M., Zagozdzon, A., Kong, Y., Gardner, H., Kiyokawa, H., Harris, L. N., Stål, O., Sicinski, P. (2006). Requirement for CDK4 kinase function in breast cancer. *Cancer Cell* 9: 23–32.
- Zhang, H., Berezov, A., Wang, Q., Zhang, G., Drebin, J., Murali, R., Greene, M. I. (2007). ErbB receptors: from oncogenes to targeted cancer therapies. *Journal of Clinical Investigation* 117(8): 2051-2058.
- Zhang, X., Zhang, Y., Liu, X., Fang, A., Li, P., Li, Z., Liu, T., Yang, Y., Du, L., Wang, C. (2015). MicroRNA-203 Is a Prognostic Indicator in Bladder Cancer and Enhances Chemosensitivity to Cisplatin via Apoptosis by Targeting Bcl-w and Survivin. *PLoS ONE* 10(11): e0143441.
- Zhang, Y., Chen, X., Gueydan, C., and Han, J. (2018). Plasma membrane changes during programmed cell deaths. *Cell Research* 28: 9–21.
- Zhou, A. Y., Shen, R. R., Kim, E., Lock, Y. J., Xu, M., Chen, Z. J., Hahn, W. C. (2013). IKK $\epsilon$ -mediated tumorigenesis required k63-linked polyubiquitination by a ciAP1/ciAP2/TRAF2 E3 ubiquitin ligase complex. *Cell Reports* 3: 724-733.