

UNIVERSITI PUTRA MALAYSIA

FABRICATION OF CHITOSAN-INTEGRATED SINGLE-MODE TAPERED OPTICAL FIBER DENV II E PROTEIN SENSOR

NADIA BINTI MOHD AMIN @ MOHD NASIR

FK 2021 1



FABRICATION OF CHITOSAN-INTEGRATED SINGLE-MODE TAPERED OPTICAL FIBER DENV II E PROTEIN SENSOR



NADIA BINTI MOHD AMIN @ MOHD NASIR

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

January 2020

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

FABRICATION OF CHITOSAN-INTEGRATED SINGLE-MODE TAPERED OPTICAL FIBER DENV II E PROTEIN SENSOR

By

NADIA MOHD AMIN @ MOHD NASIR

January 2020

Chair Faculty : Muhammad Hafiz bin Abu Bakar, PhD : Engineering

Globally, diseases infected by dengue virus (DENV) prevails among major public health problem, especially in tropical and sub-tropical areas. 500 000 people are estimated infected with severe dengue require hospitalization every year and 2.5% is estimated case fatality. Quantitative assessment by enzyme-linked immunosorbent assay (ELISA) is known to be used by laboratories to produce better clinical monitoring but it needs complex laboratories infrastructure as well as expertise to operate it. For the past decades, tapered single mode fiber has shown versatility and enticing sensitivity towards changes in its surrounding refractive index, making it suitable for sensing applications. In 2018, a research developing tapered optical fiber sensor targeting dengue virus envelope (E) protein originates from DENV II which is among 4 distinct DENV serotypes has been published. DENV II E protein is the interested determinant since it is located at the outermost of dengue virus structure, hence detecting the protein signifies the presence of the virus itself. As a result, high sensitivity and specificity within rapid detection 15 minutes is achieved. This biosensor is enhanced further by utilizing inorganic material which is graphene to facilitate greater surface area for sensing enhancement. However, graphene is known to have mild toxicity and its effect to DENV II E protein is yet to be determined.

This study looks into the use of organic nanomaterial namely chitosan for enhancement of tapered fiber sensing response. A layer of chitosan was introduced to single mode tapered fiber functionalized for the detection of DENV II E protein. Tapered optical fiber was fabricated and functionalized using Sodium Hydroxide (NaOH), 3-(Aminopropyl) triethoxysilane (APTES), and Glutaraldehyde. Chitosan immersion time up to 60 minutes was then tested

i

yielding working immersion time of 20 (CHIT20), 30 (CHIT30), 35 (CHIT35), 40 (CHIT40) and 45 minutes (CHIT45). Subsequently, the experiment proceeded with the immobilization of antibody. The immersion time for antibody was optimized for CHIT20, CHIT30, CHIT35, CHIT40 and CHIT45 at 25, 30, 33, 35 and 38 minutes, respectively.

After that, different concentration of DENV II E protein solution ranging from 0.0nM to 1.0nM with increment of 0.2nM were introduced. Prior to that, optimum incubation time of DENV II E protein for CHIT20, CHIT30, CHIT35, CHIT40 and CHIT45 was observed at 30, 35, 38, 40 and 43 minutes respectively. The spectral shift with the introduction of DENV II E protein was then recorded and analyzed. This set of experiment was conducted in triplicates. Consistent red shift of spectra at increasing concentration is observed for CHIT20. It obeys the linear relationship between concentration and refractive index which altered the effective refractive index and caused the red shift. For CHIT30, CHIT35, CHIT40 and CHIT45, consistent red shift of spectra was also noted. Increment of the sensitivity value is observed as CHIT20, CHIT30 and CHIT35 recorded 6.28 nm/nM, 10.68 nm/nM and 14.19 nm/nM, respectively. However, the sensitivity decreased for CHIT40 and CHIT45 with corresponding value of 12.24 nm/nM and 11.34 nm/nM. From these values, it is noted that the best sensitivity obtained for the sensor is at CHIT35 with 14.19 nm/nM. The work proceeded with the investigation on limit of detection (LOD) and the sensor was tested with different concentration ranging from 0.1pM to 0.1µM. The Langmuir curve plotted from the findings denoted LOD of 1pM. In conclusion, this study highlights the feasibility of using organic nanomaterial which has better biocompatibility and environmental friendly for the enhancement of DENV II E protein detection.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

FABRIKASI PENDERIA PROTEIN E DENV II GENTIAN OPTIK TIRUS SATU MOD BERSEPADUKAN CHITOSAN

Oleh

NADIA BINTI MOHD AMIN @ MOHD NASIR

Januari 2020

Pengerusi : Muhammad Hafiz bin Abu Bakar, PhD Fakulti : Kejuruteraan

Di seluruh dunia, penyakit yang dijangkiti oleh virus denggi (DENV) mengungguli masalah kesihatan awam, terutamanya di kawasan tropika dan sub-tropika. Dianggarkan 500 000 manusia dijangkiti denggi yang kritikal sehingga dimasukkan ke hospital dan 2.5% dianggarkan kes yang membawa kematian. Penilaian kuantitatif dari enzyme-linked immunosorbent assay (ELISA) diketahui untuk digunakan di makmal untuk mendapatkan pengawasan klinikal yang lebih baik tetapi ia memerlukan infrastruktur makmal yang kompleks dan juga kepakaran dalam pengendalian. Beberapa tahun yang lampau, gentian optik tirus telah menunjukkan kebolehan dan kepekaan terhadap perubahan indeks biasan didalam medium menyebabkan kesesuaian untuk diterapkan dalam mekanisma sistem deria. Pada tahun 2018, penyelidikan dalam membangunkan alat penderia gentian optik tirus mensasarkan protein virus denggi envelope (E), berasal daripada DENV II yang mana dalam kalangan 4 serotaip DENV yang berbeza telah diterbitkan. Disebabkan protein DENV II E terletak paling luar pada struktur virus denggi, maka mengesan protein itu menunjukkan kehadiran virus itu sendiri. Kesan daripada itu, sensitiviti yang tinggi dan khusus dalam 15 minit pengesanan pantas telah diperolehi. Penderia bio ini diperbaiki lebih lagi dengan penggunaan bahan bukan organik iaitu grafin untuk menyediakan kawasan permukaan vand lebih besar untuk penambahan penderiaan. Walaubagaimanapun, grafin diketahui untuk mengadungi toksik yang tidak kuat dan kesannya kepada protein DENV II E belum ditentukan lagi.

Kajian ini memperkenalkan penggunaan nanomaterial organik yang bernama Chitosan untuk penambah pada gerak balas penderiaan di gentian tirus. Satu lapisan Chitosan telah dikenalkan kepada satu mod gentian tirus yang telah difungsikan untuk pengesanan protein DENV II E. Gentian optik tirus telah dibuat dan difungsikan menggunakan Natrium Hidroksida (NaOH), 3-(Aminopropyl) triethoxysilane (APTES), dan Glutaraldehyde. Masa rendam

iii

sehingga 60 minit telah diuji menghasilkan masa rendam berfungsi 20 (CHIT20), 30 (CHIT30), 40 (CHIT40) dan 45 minit (CHIT45). Selepas itu, eksperimen diteruskan dengan pelumpuhan antibodi. Masa rendam untuk pelumpuhan antibodi telah dioptimumkan untuk CHIT20, CHIT30, CHIT40 dan CHIT45 dan masa optimum yang dihasilkan untuk setiap keadaan ialah 25, 30, 35 dan 38 minit.

Selepas itu, larutan protein DENV II E dengan kepekatan berbeza antara 0.0nM sehingga 1.0nM dengan kenaikan 0.2nM telah diperkenalkan. Sebelum itu, masa rendam optimum untuk protein DENV II E telah dilihat pada 30, 35, 40 dan 43 minit bagi CHIT20, CHIT30, CHIT40 dan CHIT45. Pengalihan spektrum disebabkan oleh protein DENV II E telah direkod dan dihuraikan. Eksperimen ini dijalalankan sebanyak tiga kali. Pengalihan ke kanan yang berterusan pada kepekatan meningkat telah dilihat untuk CHIT20, CHIT30, CHIT40 dan CHIT45. Kenaikan dalam nilai kepekaan menunjukkan sebagaimana CHIT20, CHIT30 and CHIT35 memperoleh 6.28 nm/nM, 10.68 nm/nM and 14.19 nm/nM. Walaubagaimanapun, kepekaan menurun untuk CHIT40 dan CHIT45 dengan nilai masing-masing 12.24 nm/nM and 11.34 nm/nM. Dari nilai-nilai ini, kepekaan terbaik yang diperoleh ialah 14.19 nm/nM. Kerja ini diteruskan lagi dengan menyiasat had pengesanan dan penderia diuji dengan kepekatan berbeza dalam julat 0.1pM to 0.1µM. Lengkungan Langmuir telah diplot dan LOD yang diperolehi ialah 1pM. Ia mematuhi hubungan lelurus diantara kepekatan dan indeks biasan dan menyebabkan pengalihan ke kanan. Kesimpulannya, kajian ini menonjolkan penggunaan nanomaterial yang boleh dilaksana yang mana mempunyai keserasian bio yang lebih baik dan mesra alam untuk penambah penderiaan protein DENV II E.

ACKNOWLEDGEMENTS

Alhamdulillah, all praise to the Almighty God that I am able to complete this thesis. Throughout this wonderful journey, I have the pleasure and opportunity of working and meeting with many amazing people whom I wish to thank.

I would like to express my deepest appreciation and gratitude to my supervisor, Associate Professor Dr. Muhammad Hafiz Abu Bakar for providing excellent discussion and for sharing his knowledge as well as giving helpful assistance throughout the progress of the research. Through the research, I have learnt in a hard way to be more independent, not only to gain good research ideas but also to succeed in life. My gratitude extends to my co-supervisors, Dr Nadiah Husseini Zainol Abidin and Dr Fatin Hamimi Mustafa for their continuous help and guidance. Last but not least, I would like to deliver special thanks to Dr Yasmin Mustapha Kamil for the guidance and patient throughout the time.

I would like to convey my sincere thanks to Maisarah Mansor for providing me the unfailing support and continuous encouragement throughout my years of study and through the process of researching and writing this thesis. Not to forget, Mr Zamili who has always been helpful whenever I need help with equipment and the fabrication of tapered fiber needed in order to complete this work.

Last but not least, I would like to dedicate my utmost appreciation and gratitude to my beloved husband, Mohd Khuzaipah Mat Yusoff and my lovely son, Noah Abid for the patient, understanding and simply, everything. I thank my beloved parents, Mohd Amin @ Mohd Nasir Hj Ahmad and Rafeah Abdullah for their continuous support and encouragement. This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Muhammad Hafiz bin Abu Bakar, PhD

Associate Professor Faculty of Engineering Universiti Putra Malaysia (Chairman)

Nadiah Husseini binti Zainol Abidin, PhD

Senior Lecturer Centre of Foundations Studies for Agricultural Science Universiti Putra Malaysia (Member)

Fatin Hamimi binti Hamat @ Mustafa, PhD

Senior Lecturer Institute for Research in Molecular Medicine (INFORMM) Universiti Sains Malaysia Health Campus (Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date: 12 August 2021

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

| Signature: Name of Chairman of Supervisory Committee: | Assoc. Prof. Dr. Muhammad Hafiz b. Abu Bakar |
|--|---|
| Signature: Name of Member of Supervisory Committee: | Dr. Nadiah Husseini binti Zainol Abidin |
| Signature: Name of Member of Supervisory Committee: | Dr. Fatin Hamimi binti Hamat @ Mustafa |

TABLE OF CONTENTS

| | Page |
|-----------------------|------|
| ABSTRACT | i |
| ABSTRAK | iii |
| ACKNOWLEDGEMENTS | V |
| APPROVAL | vi |
| DECLARATION | viii |
| LIST OF FIGURES | xii |
| LIST OF ABBREVIATIONS | xv |

| CHAPTER | | | | |
|---------|------|--------------|---|--|
| 1 | INTE | | | |
| • | 11 | Overvie | ew contraction of the second | |
| | 12 | Probler | n Statement | |
| | 1.3 | Aim an | d Objectives | |
| | 14 | Scope | of Work | |
| | 1.5 | Organi | zation of Thesis | |
| 2 | | | | |
| 2 | 2 1 | Overvi | | |
| | 2.1 | Dongu | - Drohlom | |
| | 2.2 | Dengu | abad Dangua Virus Detection | |
| | 2.3 | Establi | | |
| | 2.4 | Ontion | JS Dengue Senser | |
| | 2.4 | Optical | Elber Dengue Sensor | |
| | 2.0 | Optical | Paper Dengue Sensor | |
| | 2.0 | nanom | aterial | |
| | 27 | Chitoo | alenal | |
| | 2.1 | Summ | | |
| | 2.0 | Summa | ary | |
| 3 | TAP | ERED C | PTICAL INTEGRATED WITH | |
| | CHIT | FOSAN | AS BIOSENSOR | |
| | 3.1 | Overvi | ew | |
| | 3.2 | Fabrica | ation and Characterization of | |
| | | single i | mode tapered optical fiber | |
| | 3.3 | Functio | onalization of single mode tapered | |
| | | optical | fiber | |
| | 3.4 | Integra | tion of chitosan on functionalized | |
| | | tapered | d optical fiber | |
| | | 3.4.1 | Immobilization of anti-DENV II E | |
| | | | protein antibodies onto chitosan- | |
| | | | integrated functionalized tapered | |
| | | | optical fiber | |
| | | 210 | Detection of DENIV/ II E protoin | |

3.4.2 Detection of DENV II E protein antibodies with chitosanintegrated bio-functionalized

| | | | tapered optical fiber | |
|---------|------------|--------------------|---|----|
| | | 3.4.3 | Characterization of chitosan integrated tapered optical fiber | 47 |
| | 3.5 | Optimiz tapered | ation of chitosan-integrated l optical fiber performance and rative results | 48 |
| | 3.6 | Summa | ary | 55 |
| 4 | CON CON | NCLUSIC NTRIBUT | DNS, RESEARCH TONS, LIMITATIONS AND | 57 |
| | FUT | URE WC | ORK RECOMMENDATIONS | 57 |
| | 4.1 | Conclus | ion | 57 |
| | 4.2 | Researc Work | ch Contributions and Limitations of | 58 |
| | 4.3 | Future V | Vork | 58 |
| REFEREN | CES | | | 59 |
| BIODATA | UF SI | UDENI | | 67 |
| | | | | |

 \bigcirc

LIST OF FIGURES

| Figure | | Page |
|--------|--|------|
| 1.1 | Scope of Work | 4 |
| 2.1 | The organization of the dengue virus genome and schematic illustration of dengue virus | 7 |
| 2.2 | Sequential infection phase of DENV | 8 |
| 2.3 | PanBio® dengue strip test kit that detects the presence of anti-dengue IgG and IgM antibody | 9 |
| 2.4 | Schematic representation of the development of PSiNs-based biosensor for dengue DNA detection | 11 |
| 2.5 | LSPR-based all-optical fiber sensor setup with an end-face transducer | 12 |
| 2.6 | The experimental set up used for DENV NS1 IgG antibody detection | 13 |
| 2.7 | The crossed section of dengue virus | 14 |
| 2.8 | Light propagation in tapered fiber with a non- adiabatic transition region | 15 |
| 2.9 | Spectra output before and after tapering | 16 |
| 2.10 | Molecular structure of GO with oxygenated groups | 17 |
| 2.11 | The molecular structure of a generation 4 PAMAM dendrimer | 18 |
| 2.12 | The acetylation of chitin to chitosan | 19 |
| 3.1 | Flow chart of the research | 24 |
| 3.2 | (a) Vytran GPX-3000 glass processing workstation. (b) Geometry of tapered optical fiber and (c) microscopic image of the tapered fiber | 26 |
| 3.3 | Experimental setup for characterization of taper sensor | 27 |
| 3.4 | Example of a spectral output example from a single-mode tapered optical fiber | 27 |

| 3.5 | Structural formula of APTES | 28 |
|------|---|----|
| 3.6 | Hydroxylation of tapered optical fiber with NaOH, followed by the deposition of APTES onto the surface of the tapered optical fiber | 29 |
| 3.7 | The structural formula of glutaraldehyde | 29 |
| 3.8 | The activation process with glutaraldehyde on silanized surfaced | 30 |
| 3.9 | Output spectra after immersion of NaOH, APTES and Glutaraldehyde | 31 |
| 3.10 | Raman spectrum of tapered optical fiber after hydroxylation process | 32 |
| 3.11 | Raman spectrum of tapered optical fiber after silanization process with silane agent | 32 |
| 3.12 | Raman spectrum of tapered optical fiber after activation process with glutaraldehyde | 33 |
| 3.13 | FESEM images of tapered optical fiber after the incubation of (a) NaOH, (b) APTES and (c) Glutaraldehyde at 5kV and 50 K magnification | 33 |
| 3.14 | Schematic drawing of chitosan deposition on activated surface of tapered optical fiber | 34 |
| 3.15 | Output spectra after immersion of chitosan at 40 minutes | 35 |
| 3.16 | Raman spectrum of tapered optical fiber after deposition of chitosan at 40 minutes | 36 |
| 3.17 | Schematic diagram of an IgG antibody molecule | 37 |
| 3.18 | Immobilization of anti-Dengue II E protein antibodies onto chitosan-integrated tapered optical fiber surface | 38 |
| 3.19 | Wavelength shift against antibody immersion time for (a) CHIT10, (b) CHIT20, (c) CHIT30, (d) CHIT40, (e) CHIT50 and (f) CHIT60 | 39 |
| 3.20 | AFM and FESEM images of chitosan-deposited tapered optical fiber with different immersion time (a)-(b) 20 minutes, (c)-(d) 30 minutes, and (e)-(f) 40 minutes | 41 |

| 3 | .21 | Wavelength shift against antibody immersion time for CHIT35 and CHIT45 | 42 |
|----|-----|---|----|
| 3. | .22 | AFM and FESEM images of chitosan-deposited tapered optical fiber with immersion time of (a)-(b) 35 minutes and (c)-(d) 45 minutes | 43 |
| 3. | .23 | Schematic drawing of the anti-Dengue II E protein antibody reacting to DENV II E protein | 43 |
| 3. | .24 | Comparison of the output spectra before and after the introduction of DENV II E protein | 44 |
| 3. | .25 | Dynamic response when tapered optical fiber was immersed in Dengue II E protein solution for 60 minutes at different chitosan deposition time (a) CHIT20, (b) CHIT30, (c) CHIT35, (d) CHIT40 and (e) CHIT45 | 47 |
| 3. | .26 | Raman spectrum of tapered optical fiber after antibody immobilization | 48 |
| 3. | .27 | Raman spectrum of tapered optical fiber after the introduction of DENV II E protein | 49 |
| 3. | .28 | FESEM image after (a) anti-Dengue II E protein immobilization (b) the introduction of Dengue II E protein immobilization for CHIT35 at 5kV under magnification of 10 K | 50 |
| 3. | .29 | EDX spectra for before (a) and after (b) the introduction of DENV II E protein | 50 |
| 3 | .30 | Wavelength shift at different concentration of DENV II E protein for (a) non-chitosan integrated bio functionalized tapered optical fiber (b) CHIT20, (c) CHIT30, (d) CHIT35, (e) CHIT40 and (f) CHIT45 | 54 |
| 3. | .31 | Langmuir isotherm adsorption equation fitting of wavelength shift at varied concentration of 0.1 pM to 0.1 μM | 55 |
| | | | |

LIST OF ABBREVIATIONS

| AFM | Atomic force microscope |
|------------------|---|
| APTES | 3- (Aminopropyl) triethoxysilane |
| Au | Gold |
| AuNPs | Gold nanoparticles |
| С | Capsid |
| cDNA | Complementary DNA |
| DENV | Dengue virus |
| DF | Dengue fever |
| DHF | Dengue haemorrhagic fever |
| DNA | Deoxyribonucleic acid |
| DSS | Dengue shock syndrome |
| E | Envelope |
| EDX | Energy-dispersive X-Ray |
| ELISA | Enzyme linked immunosorbent assay |
| FESEM | Field emission scanning electron microscope |
| FSR | Free spectral range |
| GA | Glutaric acid |
| GO | Graphene oxide |
| LOD | Limit of detection |
| LP ₀₁ | Fundamental mode |
| LSPR | Localized surface plasmon resonance |
| Μ | Membrane |
| NaOH | Sodium hydroxide |
| nM | Nanomolar |
| | AFM APTES Au AuNPs C C CDNA DENV DF DHF DHF DHF DNA DSS E EDX ELISA FESEM FSR GA FESEM FSR GA GO LOD LP01 LSPR M NaOH NAOH |

- NS Non-structural
- OH Hydroxyl

C

- OSA Optical spectrum analyzer
- PAMAM Polyamidoamine
- PBS Phosphate buffer solution
- PNA Peptide nucleic acid
- PSiNs Porous silica nanospheres
- SPR Surface plasmon resonance
- WHO World health organization

CHAPTER 1

INTRODUCTION

1.1 Overview

Dengue virus (DENV) infection is the most common viral infection transmitted by mosquito [1]. Four distinct serotypes have been acknowledged which are DENV-1, DENV-2, DENV-3 and DENV-4 [2]. Infection by any of these serotypes can cause classical Dengue fever and the more acute Dengue haemorrhagic fever (DHF) and Dengue shock syndrome (DSS). Up to date, there are no licensed vaccines that have been published for Dengue. Consequently, containment of the infection relies on proper clinical management that should begin during febrile phase which is first three days of infections [3]. Hence, early detection is crucial to ensure the survival of Dengue patients.

Current trend to diagnose this disease for early detection is by using antigenic determinants as a target in sensing systems. DENV is composed of 10 distinct kinds of proteins with its 11 000 nucleotide-based genome [4]. Three of the proteins are structural proteins for the capsid (C), membrane (prM) and envelope (E) glycoprotein, whereas another 7 as non-structural proteins (NS1, NS2a, NS2b, NS3, NS4a, NS4b, NS5) [5]. From the variety types of proteins, invention of detection kits targeting specific protein has been reported [6]. For example, detection kits targeting NS1 proteins are already obtainable in the market [6][7]. Nevertheless, these kits are based on qualitative assessment and only for rapid detection.

Conversely, quantitative assessment produces a better clinical monitoring where clinical laboratories typically employ the enzyme-linked immunosorbent assay (ELISA) method, or the IgM/IgG antibody capture ELISA (MAC-ELISA) [7]. The principle of operation is based on absorbance changes on the interaction of ligand-targeted antibody bound on the sensing solid phase. Although this technique produces quantitative output, the procedure is complex and time consuming which also requires expensive laboratories infrastructure and highly trained personnel [7]. Another quantitative method available is real-time polymerase chain reaction (RT-PCR) [8]. It can specifically detect DENV at very low concentration and possesses high sensitivity. However, it may show false positives when there is a presence of even very small impurities [8]. In light of this, the more common quantitative assessment opted for infection monitoring is platelet count [9]. Nonetheless, monitoring via platelet is not specific to DENV only as other viral infections could also affect platelet count [10].

Hence, the alternative is to pursue a simpler and cheaper quantitative detection method such as the implementation of tapered optical fiber which is known to be small in size, flexible and highly sensitive [11][12]. Recent studies have been published on the use of tapered optical fiber sensors to detect NS1 [13] as well as DENV II E protein [14]. DENV II E protein is of interest since it is located at the outermost of DENV structure, hence it will interact with host cell first compared to other proteins. The detection of the E protein itself allows diagnosis of DENV infection at its onset, thus allowing rapid clinical response.

The transducer in [14] was further enhanced through integration of nanomaterial in the sensing layer [15]. By introducing graphene oxide (GO) onto the single mode tapered optical fiber surface, the effective sensing area was expanded which led to a substantial increase in sensitivity and emphasized the immense potential of nanomaterial integration in the sensing layer of tapered optical fiber transducer. This is proven as the sensitivity obtained was increased from 5.02 nm/nM [14] to 12.77 nm/nM [15].

1.2 Problem Statement

Despite numerous DENV sensing methods available nowadays, there are drawbacks that impede their deployment as comprehensive, accessible and user-friendly diagnostic tools. For example, although ELISA and RT-PCR are sensitive and quantitative, they are highly complex and costly in terms of time and chemical reageants. Whereas, for rapid detection strip test, the results are solely qualitative and it targets NS1 which is not a primary determinant. Hence from the drawbacks mentioned, the emergence of outstanding alternatives such as the tapered fiber sensor are highly desired. Enhancement of the sensor through nanomaterial integration has further solidified the feasibility of taper sensor to address issues pertaining to current DENV detection schemes. Nevertheless, inorganic nanomaterial that has been explored such as graphene oxide could still become an issue in terms of its decomposition as well as toxicity. Additionally, its effect to the protein sensing layer and organic sample remains undetermined.

1.3 Aim and Objectives

This research aims to explore the use of organic nanomaterial, chitosan, for enhancement of functionalized single-mode tapered fiber DENV II E protein sensor. The following objectives of this research are:

- I. To deposit chitosan on functionalized single-mode tapered fiber
- II. To fabricate a chitosan-integrated single-mode tapered optical fiber DENV II E protein sensor
- III. To compare the performance between chitosan-integrated and non chitosan-integrated single mode tapered optical fiber DENV II E protein sensor

1.4 Scope of Work

The scope of work in this research is summarized in Figure 1.1. Generally, this work was focused on dengue diagnostics specifically using optical-based sensor. The optical transducer of choice was tapered single mode fiber due to its fabrication simplicity and high sensitivity accorded by evanescent wave excited along the narrowed region. In addition, the spectral-shift based detection provides accurate and power-independent sensing output. The DENV determinant detected by the single mode fiber transducer was Dengue II E protein. E protein was chosen because it is located at the outermost structure of the virus, hence detecting the E protein signifies the presence of the virus itself while serotype II was selected as it is the most prevalent type in Malaysia. The sensor in this work was integrated with nanomaterial with the aim of enhancing the response by creating more binding sites during antibody immobilization. Chitosan, an organic nanomaterial was chosen for its nontoxicity, biodegradability and biocompatibility. DENV II E protein concentration within nanoMolar (nM) range was chosen for the investigation to make it comparable to other similar prior studies.



1.5 Organization of Thesis

The organization of this thesis is outlined as following:

Chapter 1 consists of the introduction and overview of the research area. It provides a brief explanation on dengue viruses and infection. Current dengue detection method and their challenges are highlighted along with the aim and the objectives that are formed to address those issues. The scope of work and thesis organization are also included in this chapter.

Chapter 2 introduces the classical methods and modern trends to diagnose Dengue. This includes thorough discussion and reviews on established dengue detection methods. Emerging technologies such as tapered optical fiber sensor are also presented in this chapter.

Chapter 3 merges the methodology used in this work as well as results obtained from the whole process. The steps taken starting from the fabrication of single mode tapered optical fiber until the complete detection of DENV II protein are detailed. Analysis of the results as well as comparison to included.

Last but not least, chapter 4 concludes the research work. All the important findings corresponding to the set objectives are highlighted and recommendations for future work are also stated.

REFERENCES

- [1] "Mosquito-Borne Diseases." [Online]. Available: https://www.bcm.edu/departments/molecular-virology-and microbiology/emerging-infections-and-biodefense/mosquitoes. [Accessed: 08-Aug-2019].
- [2] C. Yung, K. Lee, T. Thein, L. Tan, V. Gan, J. Wong, D. Lye, L. Ng and Y. Leo,, "Dengue Serotype-Specific Differences in Clinical Manifestation, Laboratory Parameters and Risk of Severe Disease in Adults, Singapore," The American Journal of Tropical Medicine and Hygiene, vol. 92, no. 5, pp. 999-1005, 2015.
- [3] S. Kalayanarooj, "Clinical Manifestations and Management of Dengue/DHF/DSS," Tropical Medicine and Health, vol. 39, no. 4, pp. S83-S87, 2011.
- [4] Z. Zeng, J. Shi, X. Guo, L. Mo, N. Hu, H. Zhou and Y. Hu, "Complete Genome Sequence Analysis of an Imported Dengue Virus Serotype 1 Strain from Myanmar", Genome Announcements, vol. 6, no. 27, 2018.
- [5] M. Guzman and G. Kouri, "Dengue diagnosis, advances and challenges", International Journal of Infectious Diseases, vol. 8, no. 2, pp. 69-80, 2004.
- [6] S. Chaterji, Y. Leo, A. Chow, E. Ooi and J. Allen, "Evaluation of the NS1 Rapid Test and the WHO Dengue Classification Schemes for Use as Bedside Diagnosis of Acute Dengue Fever in Adults", The American Journal of Tropical Medicine and Hygiene, vol. 84, no. 2, pp. 224-228, 2011.
- [7] S. Pal, A. Dauner, I. Mitra, B. Forshey, P. Garcia, A. Morrison, E. Halsey, T. Kochel and S. Wu, "Evaluation of dengue ns1 antigen rapid tests and elisa kits using clinical samples," PLoS One, vol. 9, no. 11, 2014.
- [8] N. V. Voge, I. Sánchez-Vargas, C. D. Blair, L. Eisen, and B. J. Beaty, "Detection of dengue virus NS1 antigen in infected aedes aegypti using a commercially available kit," The American Journal of Tropical Medicine and Hygiene, vol. 88, no. 2, pp. 260–266, 2013.
- [9] S. Rajapakse, C. Rodrigo and Rajapakse, "Treatment of dengue fever", Infection and Drug Resistance, p. 103, 2012.
- [10] M. Williams, "What are Platelets and Why are They Important?" [Online]. Available: https://www.hopkinsmedicine.org/heart_vascular_institute/clinical_servi ces/centers_excellence/womens_cardiovascular_health_center/patient _information/health_topics/platelets.html. [Accessed: 19-Aug-2019].

- [11] H. Usman, M. Abu Bakar, A. Hamzah and A. Salleh, "A tapered fibre optics biosensor for histamine detection", Sensor Review, vol. 36, no. 1, pp. 40-47, 2016.
- [12] M. Batumalay, Z. Harith, H. A. Rafaie, F. Ahmad, and M. Khasanah, "Sensors and Actuators A: Physical Tapered plastic optical fiber coated with ZnO nanostructures for the measurement of uric acid concentrations and changes in relative humidity," Sensors Actuators A: Physical, vol. 210, pp. 190–196, 2014.
- [13] M. A. Mustapa, M. H. Abu Bakar, Y. Mustapha Kamil, A. Syahir, and M. A. Mahdi, "Bio-Functionalized Tapered Multimode Fiber Coated with Dengue Virus NS1 Glycoprotein for Label Free Detection of Anti-Dengue Virus NS1 IgG Antibody," IEEE Sensor Journal, vol. 18, no. 10, pp. 4066–4072, 2018.
- [14] Y. Mustapha Kamil, M. Abu Bakar, M. Mustapa, M. Yaacob, N. Abidin, A. Syahir, H. Lee and M. Mahdi, "Label-free Dengue E protein detection using a functionalized tapered optical fiber sensor", Sensors and Actuators B: Chemical, vol. 257, pp. 820-828, 2018.
- [15] Y. Mustapha Kamil, M. Abu Bakar, A. Amir Hamzah, M. Yaacob, L. Ngee and M. Mahdi, "Dengue E protein detection using graphene oxide integrated tapered optical fiber sensor", IEEE Journal of Selected Topics in Quantum Electronics, pp. 1-1, 2018.
- [16] K. Versluys, "Review article," English Stud., vol. 60, no. 4, pp. 516– 522, 2008.
- [17] "Dengue Viruses." [Online]. Available: https://www.nature.com/scitable/topicpage/dengue-viruses-22400925/. [Accessed: 29-Aug-2019].
- [18] M. Diamond and T. Pierson, "Molecular Insight into Dengue Virus Pathogenesis and Its Implications for Disease Control", Cell, vol. 162, no. 3, pp. 488-492, 2015.
- [19] R. Perera and R. Kuhn, "Structural proteomics of dengue virus", Current Opinion in Microbiology, vol. 11, no. 4, pp. 369-377, 2008.
- [20] "Molecules of the month: Dengue virus." [Online]. Available: https://pdb101.rcsb.org/motm/103. [Accessed: 31-Aug-2019].
- [21] L. Zonetti, M. Coutinho and A. de Araujo, "Molecular Aspects of the Dengue Virus Infection Process: A Review", Protein & Peptide Letters, vol. 25, no. 8, pp. 712-719, 2018.
- [22] "Dengue and Dengue Hemorrhagic Fever: Information for health care practitioners." [Online] pp.1-4. Available: https://reliefweb.int/report/elsalvador/dengue-and-dengue-hemorrhagic-fever-information-healthcare-practitioners [Accessed 18 Feb. 2020].

- [23] "Dengue and severe dengue." [Online]. Available: https://www.who.int/en/news-room/fact-sheets/detail/dengue-andsevere-dengue. [Accessed: 19-Aug-2019].
- [24] G. Perng, H. Lei, Y. Lin and K. Chokephaibulkit, "Dengue Vaccines: Challenge and Confrontation", World Journal of Vaccines, vol. 01, no. 04, pp. 109-130, 2011.
- [25] A. Jain and U. Chaturvedi, "Dengue in infants: an overview", FEMS Immunology & Medical Microbiology, vol. 59, no. 2, pp. 119-130, 2010.
- [26] D. W. Vaughn, A. Nisalak, S. Kalayanarooj, T. Solomon, N. M. Dung, A. Cuzzubbo, And P. L. Devine, "Evaluation of a Rapid Immunochromatographic Test for Diagnosis of Dengue Virus Infection", Journal of Clinical Microbiology, vol. 36, no. 1, pp. 234-238, 1998.
- [27] S. Chaterji, J. C. Allen, A. Chow, Y. S. Leo, and E. E. Ooi, "Evaluation of the NS1 rapid test and the WHO dengue classification schemes for use as bedside diagnosis of acute dengue fever in adults," The American Journal of Tropical Medicine and Hygiene, vol. 84, no. 2, pp. 224–228, 2011.
- [28] A. J. Cuzzubbo, T. P. Endy, A. Nisalak, S. Kalayanarooj, D. W. Vaughn, S. A. Ogata, D. E. Clements And P. L. Devine, "Use of Recombinant Envelope Proteins for Serological Diagnosis of Dengue Virus Infection in an Immunochromatographic Assay," Clinical and Vaccine Immunology, vol. 8, no. 6, pp. 1150–1155, 2002.
- [29] F. Kassim, M. N. Izati, T. A. R. Tgrogayah, Y. M. Apandi, and Z. Saat, "Use Of Dengue Ns1 Antigen For Early Diagnosis Of Dengue Virus Infection", The Southeast Asian Journal of Tropical Medicine and Public Health, vol. 42, no. 3, pp. 562-569, 2011.
- [30] T. Conceição, A. Da Poian and M. Sorgine, "A real-time PCR procedure for detection of dengue virus serotypes 1, 2, and 3, and their quantitation in clinical and laboratory samples", Journal of Virological Methods, vol. 163,
- [31] K. Yamada, T. Takasaki, M. Nawa and I. Kurane, "Virus isolation as one of the diagnostic methods for dengue virus infection", Journal of Clinical Virology, vol. 24, no. 3, pp. 203-209, 2002.
- [32] S. Alcon, A. Talarmin, M. Debruyne, A. Falconar, V. Deubel, and M. Flamand, "Enzyme-linked immunosorbent assay specific to Dengue virus type 1 nonstructural protein NS1 reveals circulation of the antigen in the blood during the acute phase of disease in patients experiencing primary or secondary infections.," Journal of Clinical Microbiology, vol. 40, no. 2, pp. 376–81, 2002.
- [33] S. Abdul Rahman et al., "Label-Free Dengue Detection Utilizing PNA/DNA Hybridization Based on the Aggregation Process of

Unmodified Gold Nanoparticles," Journal of Nanomaterial, vol. 2014, pp. 1–5, 2014.

- [34] E. Y. Ariffin, L. L. Tan, N. Huda, A. Karim, and L. Y. Heng, "Optical DNA Biosensor Based on Square-Planar Ethyl Piperidine Substituted Nickel(II) Salphen Complex for Dengue Virus Detection", Sensors, vol. 18, no. 4, p. 1173, 2018.
- [35] [13]N. Omar, Y. Fen, J. Abdullah, C. Chik and M. Mahdi, "Development of an optical sensor based on surface plasmon resonance phenomenon for diagnosis of dengue virus E-protein", Sensing and Bio-Sensing Research, vol. 20, pp. 16-21, 2018.
- [36] V. Malachovská, C. Ribaut, V. Voisin, M. Surin, P. Leclère, R. Wattiez, and C. Caucheteu, "Fiber-Optic SPR Immunosensors Tailored to Target Epithelial Cells through Membrane Receptors," Analytical Chemistry, vol. 87, no. 12, pp. 5957–5965, 2015.
- [37] M. Batumalay, Z. Harith, H. Rafaie, F. Ahmad, M. Khasanah, S. Harun, R. Nor, H. Ahmad, "Tapered plastic optical fiber coated with ZnO nanostructures for the measurement of uric acid concentrations and changes in relative humidity," Sensors Actuators A: Physical, vol. 210, pp. 190–196, 2014.
- [38] H. Usman, M. H. A. Bakar, A. S. Hamzah, and A. B. Salleh, "A tapered fibre optics biosensor for histamine detection," Sensor Review, vol. 36, no. 1, pp. 40–47, 2016.
- [39] A. Camara, P. Gouvêa, A. Dias, A. Braga, R. Dutra, R. de Araujo, I. Carvalho, "Dengue immunoassay with an LSPR fiber optic sensor," Optic Express, vol. 21, no. 22, p. 27023, 2013.
- [40] "Evolution of Antibody Response to Dengue Virus Teased Out." [Online]. Available: https://www.technologynetworks.com/biopharma/news/evolution-ofantibody-response-to-dengue-virus-teased-out-317534.
- [41] M. Komanec, T. Nemecek, P. Michal, and T. Martan, "Optical Fiber Technology Structurally-modified tapered optical fiber sensors for longterm detection of liquids," Optic Fiber Technology, vol. 47, no. November 2018, pp. 187–191, 2019.
- [42] S. Korposh, S. James, S. Lee and R. Tatam, "Tapered Optical Fibre Sensors: Current Trends and Future Perspectives", Sensors, vol. 19, no. 10, p. 2294, 2019.
- [43] T. Yadav, M. Mustapa, M. Bakar and M. Mahdi, "Study of single mode tapered fiber-optic interferometer of different waist diameters and its application as a temperature sensor", Journal of the European Optical Society: Rapid Publications, vol. 9, 2014.

- [44] B. Musa, Y. Mustapha Kamil, M. Abu Bakar, A. Noor, A. Ismail and M. Mahdi, "Effects of taper parameters on free spectral range of nonadiabatic tapered optical fibers for sensing applications", Microwave and Optical Technology Letters, vol. 58, no. 4, pp. 798-803, 2016.
- [45] "Tapered fiber." [Online]. Available: https://www.rpphotonics.com/tapered_fibers.html.
- [46] J. Juan Colás, "Dual-Mode Electro-photonic Silicon Biosensors", Ph.D, University of York, UK, 2017.
- [47] T. K. Yadav, R. Narayanaswamy, M. H. Abu Bakar, Y. M. Kamil, and M. A. Mahdi, "Single mode tapered fiber-optic interferometer based refractive index sensor and its application to protein sensing," Optic Express, vol. 22, no. 19, p. 22802, 2014.
- [48] Y. Mustapha Kamil, "Development Of Tapered Optical Fiber Based Sensor For The Detection of II Dengue E Protein", Ph.D, Universiti Putra Malaysia, 2017.
- [49] I. T. Cavalcanti, B. V. M. Silva, N. G. Peres, P. Moura, M. D. P. T. Sotomayor, M. I. F. Guedes, and R. F. Dutra, "A disposable chitosan-modified carbon fiber electrode for dengue virus envelope protein detection," Talanta, vol. 91, pp. 41–46, Mar. 2012.
- [50] X. Wu, F. Mu, Y. Wang and H. Zhao, "Graphene and Graphene-Based Nanomaterials for DNA Detection: A Review", Molecules, vol. 23, no. 8, p. 2050, 2018.
- [51] E. Morales-Narváez and A. Merkoçi, "Graphene oxide as an optical biosensing platform," Advanced Material, vol. 24, no. 25, pp. 3298–3308, 2012.
- [52] C. K. Chua and M. Pumera, "The reduction of graphene oxide with hydrazine: elucidating its reductive capability based on a reaction-model approach," Chemical Communication, vol. 52, no. 1, pp. 72–75, 2016.
- [53] L. Ou, B. Song, H. Liang, J. Liu, X. Feng, B. Deng, T. Sun, L. Shao, "Toxicity of graphene-family nanoparticles: a general review of the origins and mechanisms", Particle and Fibre Toxicology, vol. 13, no. 1, 2016.
- [54] M. Virlan, D. Miricescu, R. Radulescu, C. Sabliov, A. Totan, B. Calenic, M. Greabu, "Organic Nanomaterials and Their Applications in the Treatment of Oral Diseases", Molecules, vol. 21, no. 2, p. 207, 2016.
- [55] N. Siva, K. Gunda, M. Singh, L. Norman, K. Kaur, and S. K. Mitra, "Applied Surface Science Optimization and characterization of biomolecule immobilization on silicon substrates using (3-aminopropy)

) triethoxysilane (APTES) and glutaraldehyde linker," Applied Surface Science, vol. 305, pp. 522–530, 2014.

- [56] M. A. Rahman Bhuiyan, A. Shaid, M. M. Bashar, P. Haque, and M. A. Hannan, "A Novel Approach of Dyeing Jute Fiber with Reactive Dye after Treating with Chitosan," Open Journal of Organic Polymer Materials, vol. 3, no. 04, pp. 87–91, 2013.
- [57] P. K. Dutta, J. Dutta, and V. S. Tripathi, "Chitin and chitosan: Chemistry, properties and applications," Journal of Scientific & Industrial Research, vol. 63, pp. 20-31, 2004.
- [58] M. Mahbubul Bashar and M. A. Khan, "An Overview on Surface Modification of Cotton Fiber for Apparel Use," Journal of Polymers and the Environment, vol. 21, no. 1, pp. 181–190, 2013.
- [59] C. Chou, S. Chen, Y. Li, T. Huang and J. Lee, "Low-molecular-weight chitosan scavenges methylglyoxal and N ε-(carboxyethyl)lysine, the major factors contributing to the pathogenesis of nephropathy", SpringerPlus, vol. 4, no. 1, 2015.
- [60] K. J. Huang, D. J. Niu, W. Z. Xie, and W. Wang, "A disposable electrochemical immunosensor for carcinoembryonic antigen based on nano-Au/multi-walled carbon nanotubes-chitosans nanocomposite film modified glassy carbon electrode," Analytica Chimica Acta, vol. 659, no. 1–2, pp. 102–108, 2010.
- [61] A. M. Yusufu, A. S. M. Noor, H. N. M. Azami, N. Tamchek, and Z. Z. Abidin, "Dinitrobenzene sensing utilizing chitosan-based thin films optical fluorescence sensors via linear and nonlinear excitation," International Conference on Sensing Technology ICST, vol. 2016– March, pp. 629–633, 2016.
- [62] A. H. Ridzwan, K. D. Dambul, S. A. Ibrahim, and A. Mansoor, "Tapered optical fibre coated with chitosan for lead (II) ion sensing," Electronic Letter, vol. 52, no. 12, pp. 1049–1050, 2016.
- [63] Q. Xu, C.-H. Wang, and D. Wayne Pack, "Polymeric Carriers for Gene Delivery: Chitosan and Poly(amidoamine) Dendrimers," Current Pharmaceutical Design, vol. 16, no. 21, pp. 2350–2368, 2010.
- [64] A. R. Camara, P. M. P. Gouvêa, A. C. M. S. Dias, A. M. B. Braga, R. F. Dutra, R. E. de Araujo, and I. C. S. Carvalho, "Dengue immunoassay with an LSPR fiber optic sensor," Optic Express, vol. 21, no. 22, pp. 27023–27031, 2013.
- [65] Y. Sun, M. Yanagisawa, M. Kunimoto, and M. Nakamura, "Applied Surface Science Estimated phase transition and melting temperature of APTES self-assembled monolayer using surface-enhanced antistokes and stokes Raman scattering," Applied Surface Science, vol. 363, pp. 572–577, 2016.

- [66] Y. Liu, Y. Li, X. M. Li, and T. He, "Kinetics of (3aminopropyl)triethoxylsilane (aptes) silanization of superparamagnetic iron oxide nanoparticles," Langmuir, vol. 29, no. 49, pp. 15275–15282, 2013.
- [67] M. P. Byrne and W. E. Stites, "Chemically crosslinked protein dimers: Stability and denaturation effects," Protein Science, vol. 4, no. 12, pp. 2545–2558, 1995.
- [68] P. F. McMillan and R. L. Remmele, "Hydroxyl sites in SiO, glass; a note on infrared and Raman spectra," American Mineralogist, vol. 71, no. 5–6, p. 772 LP-778, Jun. 1986
- [69] G. Walrafen and R. Douglas, "Raman spectra from very concentrated aqueous NaOH and from wet and dry, solid, and anhydrous molten, LiOH, NaOH, and KOH", The Journal of Chemical Physics, vol. 124, no. 11, p. 114504, 2006.
- [70] M. Gnyba, M. Keränen, M. Kozanecki and B. Kosmowski, "Raman investigation of hybrid polymer thin films", Materials Science-Poland, vol. 23, no. 1, pp. 29-39, 2005.
- [71] D. Aureau, Y. Varin, K. Roodenko, O. Seitz, O. Pluchery and Y. Chabal, "Controlled Deposition of Gold Nanoparticles on Well-Defined Organic Monolayer Grafted on Silicon Surfaces", The Journal of Physical Chemistry C, vol. 114, no. 33, pp. 14180-14186, 2010.
- [72] M. Prabhaharan, A. Prabakaran, S. Gunasekaran and S. Srinivasan, "Molecular structure and vibrational spectroscopic investigation of melamine using DFT theory calculations", Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy, vol. 123, pp. 392-401, 2014.
- [73] P. Lagant, G. Vergotten, G. Fleury, and M.-H. Loucheux-Lefbvre, "Raman spectroscopy and normal vibrations of peptides," European Journal of Biochemistry, vol. 139, no. 1, pp. 137–148, Feb. 1984.
- [74] G. Sneddon, A. Ganin and H. Yiu, "Sustainable CO 2 Adsorbents Prepared by Coating Chitosan onto Mesoporous Silicas for Large-Scale Carbon Capture Technology", Energy Technology, vol. 3, no. 3, pp. 249-258, 2015.
- [75] X. D. Ren, Q. S. Liu, H. Feng, and X. Y. Yin, "The characterization of chitosan nanoparticles by raman spectroscopy," Applied Mechanics and Materials, vol. 665, pp. 367–370, 2014.
- [76] Y. Mustapha Kamil, S. Al-Rekabi, M. Yaacob, A. Syahir, H. Chee, M. Mahdi, M. Abu Bakar, "Detection of dengue using PAMAM dendrimer integrated tapered optical fiber sensor," Scientific Reports, vol. 9, no. 1, pp. 1–10, 2019.

- [77] K. Jiang, A. Eitan, L. Schadler, P. Ajayan, R. Siegel, N. Grobert, M. Mayne, M. Reyes-Reyes, H. Terrones, M. Terrones, "Selective Attachment of Gold Nanoparticles to Nitrogen-Doped Carbon Nanotubes", Nano Letters, vol. 3, no. 3, pp. 275-277, 2003.
- [78] R. Kengne-Momo, P. Daniel, F. Lagarde, Y. L. Jeyachandran, J. F. Pilard, and G. Thouand, "Protein Interactions Investigated by the Raman Spectroscopy for Biosensor Applications", International Journal of Spectroscopy, vol. 2012, pp. 1-7, 2012.
- [79] N. Lapin and Y. Chabal, "Infrared Characterization of Biotinylated Silicon Oxide Surfaces, Surface Stability, and Specific Attachment of Streptavidin", The Journal of Physical Chemistry B, vol. 113, no. 25, pp. 8776-8783, 2009.
- [80] D. Aureau, Y. Varin, K. Roodenko, O. Seitz, O. Pluchery, and Y. J. Chabal, "Controlled deposition of gold nanoparticles on well-defined organic monolayer grafted on silicon surfaces," The Journal of Physical Chemistry C, vol. 114, no. 33, pp. 14180–14186, 2010.

BIODATA OF STUDENT

Nadia binti Mohd Amin @ Mohd Nasir was born in Kelantan, Malaysia, in 1992. She received her Bachelor's Degree in Major Physics (Hons) from Universiti Putra Malaysia (UPM) in 2015.

She is now pursuing her Master's Degree in Photonics Engineering at Universiti Putra Malaysia (UPM). She is a member of The Optical Society (OSA).





UNIVERSITI PUTRA MALAYSIA

STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

ACADEMIC SESSION : Second Semester 2020/2021

TITLE OF THESIS / PROJECT REPORT :

FABRICATION OF CHITOSAN-INTEGRATED SINGLE-MODE TAPERED OPTICAL FIBER DENV II E PROTEIN SENSOR

NAME OF STUDENT :

NADIA BINTI MOHD AMIN @ MOHD NASIR

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

- 1. This thesis/project report is the property of Universiti Putra Malaysia.
- 2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
- 3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as:



This thesis is submitted for:

| PATENT | Embargo from | until |
|--|-------------------------------------|--|
| | (date) | (uale) |
| | | Approved by: |
| (Signature of Student) New IC No/ Passport No.: Date : | | (Signature of Chairman of Supervisory Committee) Name: |
| Bato . | | Date : |
| [Note : If the thesis is CONFII the letter from the organizatio confidentially or restricted.] | DENTIAL or RES on/institution wi | STRICTED, please attach with th period and reasons for |
| | | |