

# **UNIVERSITI PUTRA MALAYSIA**

SUBCRITICAL WATER EXTRACTION PROCESS FOR MICROALGAL BIODIESEL PRODUCTION

SELVAKUMAR THIRUVENKADAM

FK 2021 31



# SUBCRITICAL WATER EXTRACTION PROCESS FOR MICROALGAL BIODIESEL PRODUCTION

By

# SELVAKUMAR THIRUVENKADAM

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of Philosophy

November 2020

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

### SUBCRITICAL WATER EXTRACTION PROCESS FOR MICROALGAL BIODIESEL PRODUCTION

By

#### SELVAKUMAR THIRUVENKADAM

November 2020

Chair Faculty : Assoc. Prof. Mohd Razif Harun, PhD : Engineering

Microalgae have been used as a substrate for biodiesel production due to their numerous advantages, including fast growth rate, non-edibility, and the ability to accumulate substantial amounts of carbohydrates, lipids, and proteins. However, current extraction technologies used to produce oil from microalgal biomass are expensive, unsustainable, involve lengthy processing steps and result in lower product yields. One of the newer extraction techniques is subcritical water extraction which offers lower production costs, milder operating conditions, and a shorter production period compared to other conventional methods, such as chemical and biological extraction. The current subcritical water extraction (SCW) for biodiesel production involves two steps: extraction and transesterification. These two steps can be combined in one-step by the proposed subcritical methanol extraction (SCM) process for microalgal biodiesel production.

In this study, SCW and SCM were used to treat *Chlorella pyrenoidosa*. The operational factors such as reaction temperature, reaction time and biomass loading influence the oil yield during the extraction process. In this study, response surface methodology was employed to identify the desired extraction conditions for maximum extraction yield. SCW experiments were carried out in batch reactors as per the central composite design with three independent factors: reaction temperature (170 to 370 °C), reaction time (1 to 20 min) and biomass loading (1 to 15%). The maximum oil yield of 12.89 wt.% was obtained at 320 °C and 15 min, with 3% biomass loading. Sequential model tests showed the good fit of experimental data to the second-order quadratic model. The extracted oil from SCW is converted to biodiesel via second-step, transesterification. Recent developments in subcritical studies utilize subcritical alcohol solvents as a single step process to produce biodiesel from algae. Here, the algal biomass is subjected to SCM in the second phase of this study. The

effects of three operational factors: reaction temperature (140 to 220 °C), reaction time (1 to 15 min) and methanol to algae ratio (1 to 9 wt.%) were investigated. A maximum yield of crude biodiesel of 7.1 wt.% was obtained at 160 °C, 3 min reaction time and 7 wt.% methanol to algae ratio. The analysis of variance revealed that methanol to algae ratio is the most significant factor for maximizing biodiesel yield. Regression analysis showed a good fit of the experimental data to the second-order polynomial model. Higher cetane number (74.92) and low iodine value (58.81 g l<sub>2</sub>/100 g) crude biodiesel produced from SCM were found in compliance with the European standard (EN 14214). The SCW and SCM experimental data were fitted with three models, namely firstorder kinetic model, second-order kinetic model and Fick's law kinetic model. A comparison between SCW and SCM in terms of mass flow and energy consumption is provided through the LCA study. In terms of energy requirements, SCM has a lower energy demand than SCW. The use of subcritical technology for high-grade algal biodiesel production is expected to be promising and will result in a positive outlook for commercially viable production of high-guality biodiesel.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

### PROSES PENGEKSTRAKAN AIR SUBKRITIKAL UNTUK PENGELUARAN BIODIESEL MICROALGAL

Oleh

#### SELVAKUMAR THIRUVENKADAM

November 2020

Pengerusi Fakulti : Prof. Madya Mohd Razif Harun, PhD : Kejuruteraan

Microalga telah digunakan sebagai substrat untuk pengeluaran biodiesel kerana banyak kelebihannya, termasuk kadar pertumbuhan yang cepat, tidak dapat dipertahankan, dan keupayaan untuk mengumpul sejumlah besar karbohidrat, lipid, dan protein. Walau bagaimanapun, teknologi pengekstrakan semasa yang digunakan untuk menghasilkan minyak daripada biojisim mikro adalah mahal, tidak lestari, melibatkan langkah pemprosesan yang panjang dan menghasilkan hasil produk yang lebih rendah. Salah satu teknik pengekstrakan yang lebih baru adalah teknik pengekstrakan subkritikal yang menawarkan kos pengeluaran yang lebih rendah, keadaan operasi yang lebih ringan, dan tempoh pengeluaran yang lebih pendek berbanding dengan kaedah konvensional yang lain, seperti pengekstrakan kimia dan biologi. Teknik pengekstrakan subkritikal air (SCW) untuk penghasilan biodiesel melibatkan dua langkah iaitu: pengekstrakan dan transesterifikasi. Dua langkah ini boleh digabungkan menjadi satu langkah seperti yang dicadangkan daripada proses pengekstrakan subkritikal methanol (SCM) untuk penghasilan biodiesel daripada mikroalga.

Dalam kajian ini, SCW dan SCM digunakan untuk merawat *Chlorella pyrenoidosa*. Faktor-faktor operasi seperti suhu tindak balas, masa tindak balas dan pemuatan biomas mempengaruhi hasil minyak semasa proses pengekstrakan. Dalam kajian ini, metodologi permukaan tindak balas digunakan untuk mengenal pasti keadaan pengekstrakan yang dikehendaki bagi hasil pengekstrakan maksimum. Eksperimen SCW dilakukan dalam reaktor batch sebagai satu reka bentuk komposit pusat dengan tiga faktor bebas termasuk suhu tindak balas (170 hingga 370 °C), masa tindak balas (1 hingga 20 min) dan pemuatan biomas (1 hingga 15%). Hasil minyak maksimum sebanyak 12.89 wt% diperoleh pada 320 °C dan 15 min, dengan 3% biomass loading. Ujian model urutan menunjukkan kesesuaian data eksperimen dengan model kuadrat urutan kedua. Minyak yang diekstrak dari SCW ditukarkan kepada biodiesel

melalui langkah kedua, transesterifikasi. Perkembangan terbaru dalam kajian subkritikal menggunakan pelarut alkohol subkritikal sebagai satu proses langkah untuk menghasilkan biodiesel dari alga. Di sini, biomas alga tertakluk kepada SCM dalam fasa kedua kajian ini. Kesan tiga faktor operasi: suhu tindak balas (140 hingga 220 °C), masa tindak balas (1 hingga 15 min) dan metanol kepada nisbah alga (1 hingga 9%) telah disiasat. Hasil maksimum biodiesel mentah sebanyak 7.1% diperolehi pada 160 °C, masa reaksi 3 min dan 7% methanol kepada nisbah alga. Analisis varians mendedahkan bahawa nisbah metanol kepada alga adalah faktor paling penting untuk memaksimumkan hasil biodiesel. Analisis regresi menunjukkan kesesuaian data eksperimen untuk model polinomial pesanan kedua. Bilangan cetane yang lebih tinggi (74.92) dan nilai iodin yang rendah (58.81 g l2/100 g) oleh biodiesel mentah yang dihasilkan dari SCM didapati mematuhi piawaian Eropah (EN 14214). Data eksperimen SCW dan SCM diuji dengan menggunakan tiga model, iaitu model kinetik pertama, model kinetik kedua dan model kinetik berdasarkan undang-undang Fick. Perbandingan antara SCW dan SCM dari segi aliran masa dan penggunaan tenaga disediakan melalui kajian LCA. Dari segi keperluan tenaga, tenaga yang diperlukan oleh proses SCM adalah lebih rendah daripada SCW. Model matematik telah dicadangkan berdasarkan baki pemindahan massa semasa proses pengekstrakan subkritikal. Penggunaan teknologi subkritikal bagi pengeluaran biodiesel alga gred tinggi dijangka menjanjikan dan akan menghasilkan tinjauan positif untuk menghasilkan biodiesel berkualiti tinggi secara komersial.

#### ACKNOWLEDGEMENTS

First and foremost, I would like to extend my gratitude to my supervisor, Assoc. Prof. Mohd Razif Harun, for the guidance, encouragement, and advice he has provided throughout my time as his student. He has helped me through extremely difficult times over the course of the analysis and the writing of the thesis and for that, I sincerely thank him.

I would additionally like to thank Prof. Hiroyuki Yoshida, Assoc. Prof. Shamsul Izhar Siajam, and Assoc. Prof. Rabitah Zakaria for their support by cosupervising my PhD studies. My friends in Malaysia deserve special thanks for providing a much-needed form of escape from my studies and helping me keep things in perspective. Completing this work would have been more difficult were it not for the help provided by fellow graduate students of the Process Control Laboratory in the Faculty of Engineering.

Finally, I would like to extend my deepest gratitude to my dad, my mom, and my brother, without whose love, support, and understanding, I could never have completed this PhD degree.

This research was supported by the Ministry of Science, Technology, and Innovation, Malaysia (grant number 5450818).

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

#### Mohd Razif Harun, PhD

Associate Professor Faculty of Engineering Universiti Putra Malaysia (Chairman)

#### Shamsul Izhar Siajam, PhD

Associate Professor, Ir. Faculty of Engineering Universiti Putra Malaysia (Member)

#### Rabitah Zakaria, PhD

Associate Professor Faculty of Engineering Universiti Putra Malaysia (Member)

### ZALILAH MOHD SHARIFF, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date: 08 April 2021

# Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: Name of Chairman of Supervisory Committee:	Assoc. Prof. Mohd Razif Harun
Signature:	
Name of Member of Supervisory	
Committee:	Assoc. Prof. Ir. Shamsul Izhar Siajam
Signature	
Name of Member of	
Supervisory	
Committee:	Assoc. Prof. Rabitah Zakaria

# TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	V
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xii
LIST OF FIGURES	xiv
LIST OF ABBREVIATIONS	xvi

# CHAPTER

1	INTF 1.1 1.2 1.3	Research Background Problem Statement Objectives 1.3.1 Overall Objective 1.3.2 Specific Objectives Scope of the Study	1 1 2 4 4 4 5
2	LITE	RATURE REVIEW	6
	2.1	Theory of subcritical water extraction	6
	2.2	Biocrude	10
		2.2.1 Effect of residence time	17
		2.2.2 Effect of water density and biomass loading	14
	23	Theory of subcritical methanol extraction	15
	2.4	Extraction kinetics modelling	16
		2.4.1 Kinetic models derived from rate law	17
		2.4.2 Kinetic model derived from Fick's law	20
		2.4.3 Previous kinetics literature	23
	2.5	Life cycle assessment (LCA)	23
	2.6	Summary	26
3	MET	HODOLOGY	27
	3.1	Materials	29
	3.2	Biomass characterization	29
	3.3	Subcritical water extraction	29
		3.3.1 Experimental procedure	29
		3.3.2 Product separation and recovery	30
	3 1	Subcritical methanol extraction	30
	5.4	3.4.1 Experimental procedure	30
		3.4.2 Product separation and recovery	32
		3.4.3 Characterization of oil	32

		3.5 3.6	Charao Statisti	cterization of algae crude biodiesel cal analysis	32 33
		3.7	Extract	tion kinetics modelling	33
			3.7.1	First-order rate law	34
			3.7.2	Second-order rate law	34
			3.7.3	Fick's law	35
		3.8	Life-cv	cle assessment	35
			3.8.1	Goal and scope	35
			3.8.2	System boundary	36
			3.8.3	Life cycle inventory	36
	4	RES	ULTS A	ND DISCUSSION	39
		4.1	Chara	cterization of C. pyrenoidosa	39
		4.2	Subcri	tical water extraction (SCW)	40
			4.2.1	Response surface analysis	40
			4.2.2	Effect of temperature on oil yield	45
			4.2.3	Effect of reaction time on oil yield	46
			4.2.4	Effect of biomass loading on oil yield	46
			4.2.5	Process optimization	46
		4.3	Subcri	tical methanol extraction (SCM)	48
			4.3.1	Response surface analysis	48
			4.3.2	Effect of temperature on oil yield	53
			4.3.3	Effect of reaction time on oil yield	54
			4.3.4	Effect of biomass loading on oil yield	54
			4.3.5	Process optimization	58
		4.4	Chara	cteristics of crude biodiesel via subcritica	al 58
		45	Extrac	tion kinetics modelling of SCW	59
		1.0	4.5.1	First-order rate law	59
			4.5.2	Second-order rate law	60
			453	Fick's law	62
			454	Analysis of kinetic modelling of SCW	63
		46	Extrac	tion kinetics modelling of SCM	64
			461	First-order rate law	64
			462	Second-order rate law	65
			463	Fick's law kinetic model	66
			464	Analysis of kinetic modelling of SCM	67
		47		f subcritical extraction	68
			471	Production system overview	68
			4.7.2	Production chain analysis	71
			7.7.2		11
	5	CON	ICLUSI	ON AND RECOMMENDATIONS	76
		5.1	Conclu	usion	76
		5.2	Recon	nmendations	77
RE	FERENC	CES			78
			91		
BIU		7L 91			104
LIS	I OF PL	JRFIC	ATIONS	>	105

6

# LIST OF TABLES

Table		Page
2.1	Biocrude yield from SCW of various algae species	11
3.1	Levels of independent factors used for SCW optimization	33
3.2	Levels of independent factors used for SCM optimization	33
4.1	Proximate, ultimate, and biochemical analysis of freshwater algae species	40
4.2	Response values of the oil yield via SCW	41
4.3	Selection of a suitable model for SCW system (Fit Summary)	42
4.4	ANOVA for the regression model for the prediction of oil yield via SCW	44
4.5	Major compounds in biocrude oil from SCW of <i>C. pyrenoidosa</i> at 320° C, 15 min and 3 wt.% biomass loading detected by GC analysis.	45
4.6	Respo <mark>nse values of the crude biodiesel yield via S</mark> CM	49
4.7	Selection of a suitable model for SCM system (Fit Summary)	50
4.8	ANOVA of the regression model for the prediction of crude biodiesel yield via SCM.	52
4.9	Major compounds in biocrude oil from SCM of <i>C. pyrenoidosa</i> at 160 °C, 3 min and 7 wt.% of methanol to algae ratio.	53
4.10	Oil yield from SCW extraction	59
4.11	First-order kinetic parameters for SCW of <i>C. pyrenoidosa</i>	60
4.12	Second-order kinetic parameters for SCW of C. pyrenoidosa	61
4.13	Fick's law coefficients for SCW of C. pyrenoidosa	62
4.14	Goodness of fit of the various kinetic models from SCW data	63

4.15	Oil yield from SCM extraction	64
4.16	First-order kinetic parameters for SCM of C. pyrenoidosa	64
4.17	Second-order kinetic parameters for SCM of C. pyrenoidosa	65
4.18	Fick's law coefficients for SCM of C. pyrenoidosa	66
4.19	Goodness of fit of the various kinetic models from SCM data	67
4.20	Mass and Energy flows generated from operating 1000 t of algae	73
4.21	Lifecycle emissions by phase inventories	75

C

# LIST OF FIGURES

Figure		Page
2.1	The phase diagram of water representing the subcritical region	7
2.2	Algae biorefinery model featuring SCW	9
2.3	A schematic drawing of a typical batch scale SCW process	10
2.4	LCA framework	24
3.1	Overall research flowchart	28
3.2	Schematic diagram of experimental apparatus	31
3.3	Process block flowcharts of SCW and SCM	36
3.4	System boundary for conventional SCM process	37
3.5	System boundary of the SCM process	38
4.1	Interactions between the independent variables in 3D response surface plots of SCW	47
4.2	The effect of (a) reaction temperature, (b) reaction time, and (c) biomass loading on the oil yield via SCW	48
4.3	Interactions between the independent variables in 3D response surface plots of SCM	55
4.4	The effect of (a) reaction temperature, (b) reaction time, and (c) biomass loading on the biodiesel yield via SCM	56
4.5	The effects of process parameters interactions on biodiesel yield via SCM	57
4.6	Desirability ramp for numerical optimization of three selected goals	58
4.7	Plot of In (Yt) vs. time of SCW extraction of 170–320 °C	60
4.8	Plot of $t/Y_t$ vs. time of SCW extraction of 170–320 °C	61
4.9	Plot of In (( $M_{\infty}$ - $M_t$ )/ $M_{\infty}$ ) vs. time of SCW extraction at 170–320°C	62
4.10	Plot of ln $D_e$ vs. 1/T of SCW extraction of 170–320 °C	63

- 4.11 Plot of ln (Yt) vs. time of SCM extraction of 140–220 °C 65
- 4.12 Plot of  $t/Y_t$  vs. time of SCM extraction of 140–220 °C 66
- 4.13 Plot of ln ( $(M_{\infty} M_t)/M_{\infty}$ ) vs. time of SCM extraction at 67 140–220°C
- 4.14 Plot of ln *k* vs. time of SCM extraction of 140–220 °C 68

74

- 4.15 Material balances for SCW and SCM
- 4.16 Energy requirements of SCW and SCM processes 75 operating 1000 t algae feedstock



# LIST OF ABBREVIATIONS

ANOVA	Analysis of variance
CHF	Catalytic hydrothermal gasification
EROI	Energy return on investment
FAME	Fatty acid methyl ester
FCCCD	Face-centered central composite design
FFA	Free fatty acids
HHV	Higher heating value
LCA	Life cycle assessment
LC-GHG	Life-cycle greenhouse gas
LE	Lipid extraction
LEA	Lipid-extracted algae
LHV	Lower heating value
MUFA	Monounsaturated fatty acid
PUFA	Polyunsaturated fatty acid
RSM	Response surface methodology
SCM	Subcritical methanol extraction
SCW	Subcritical water extraction
SEM	Scanning electron microscopy
SEQSCW	Sequential subcritical water
TGA	Thermo-gravimetric analyzer
UAE	Ultrasound-assisted extraction
UCAS	Utility-connected algae system
WWTP	Wastewater treatment plant

G

#### CHAPTER 1

#### INTRODUCTION

This chapter briefly introduces the background and context of this thesis work. The motivation for this research has been established in the problem statement. Also, the objectives and scope of the study are further outlined in this chapter.

### 1.1 Research Background

Subcritical water has been utilized for solid waste resource recovery and is gaining interest as a potential solvent and catalyst for organic reactions. Among the different media used for the reaction, water is attractive because of its safety and low cost. Subcritical states of water are described at temperatures between its boiling point (100 °C) and its critical point (374 °C) and at pressures high enough to maintain the liquid state of water. At such conditions, the dielectric constant of water decreases, thereby lowering its polarity. Secondly, the magnitude of the ionic product of water increases three orders higher around 250 °C compared to room temperature. These properties are advantageous for the hydrolysis and decomposition of organic compounds including polymeric materials (Tavakoli & Yoshida, 2006a). Extensive studies led by Yoshida and coworkers have concluded that valuable and useful substances, such as organic acids, amino acids, proteins, fatty acids, oils, and nutrition were made recoverable by utilizing the subcritical extraction technique for waste treatment. For instance, fish waste was easily liquefied by hydrolysis with subcritical extraction, which enabled the recovery of organic acids, amino acids and the extraction of fatty acids (Yoshida, Terashima, & Takahashi, 1999). Similar results were also obtained with squid waste where free fatty acids (FFA) containing eicosapentaenoic acid and docosahexaenoic acid were produced during subcritical extraction (Tavakoli & Yoshida, 2006b).

The rapid depletion of fossil fuels, together with the uncertain global climate in the past decade, has inevitably led to an increased commercial interest in renewable fuels. Biodiesel and bioethanol are viewed as attractive potential solutions to alleviate the existing dependency on petroleum-based fuels (Lang, Dalai, Bakhshi, Reaney, & Hertz, 2001). The current production of biodiesel involves methanolic transesterification of extracted plant lipids, while bioethanol is presently synthesized via anaerobic yeast fermentation of sugar molecules found in the biomass of different food crops (Harun, Danquah, & Forde, 2010). Due to its high biomass productivity, perceived rapid lipid accumulation, and the suitability of its carbohydrate biochemistry for the fermentation process, microalgae are identified as a promising alternative feedstock for both biofuels (Chisti, 2007; Halim, Gladman, Danquah, & Webley, 2011; Sheehan, Dunahay, Benemann, & Roessler, 1998). Additionally, unlike other fuel-producing crops, most marine microalgae can be grown with saline water in non-agricultural lands,

thereby exempting their large-scale cultivation from placing additional demands on precious freshwater and arable lands required for food production (Sheehan et al., 1998). Since the biochemical products used in the synthesis of either biofuel (neutral lipids for biodiesel and simple sugars for bioethanol) are encapsulated within the microalgal cellular structures, disintegrating the cells to liberate these intracellular products will render them more readily accessible and subsequently enhance production yield.

A body of work has been reported on the application of the subcritical technique for the extraction of essential oils from coriander seeds (Eikani, Golmohammad, & Rowshanzamir, 2007), citrus fruit Yuzu (Ueno, Tanaka, Hosino, Sasaki, & Goto, 2008), and herb zaatar (Ozel, Gogus, & Lewis, 2003); proteins and amino acids from de-oiled rice bran (Sereewatthanawut et al., 2008); antioxidant compounds from canola meal (Hassas-Roudsari, Chang, Pegg, & Tyler, 2009), and winery waste (Aliakbarian, Fathi, Perego, & Dehghani, 2012); phenolic compounds from bitter melon (Budrat & Shotipruk, 2009), and pomegranate (He et al., 2012). The high moisture content of algal feedstock has made this technique a promising technology to produce biocrude with minimal dewatering requirements. Numerous methods for the extraction of biochemical products from microalgae have been applied; but most common methods are expeller/oil press, solvent extraction, supercritical fluid extraction, and ultrasound techniques (Popoola & Yangomodou, 2006). The cost of these extraction methods for commercial applications could be high. One of the techniques to overcome the existing problem is employing a subcritical extraction technique.

Life-cycle assessment (LCA) is a standardized tool for evaluating the environmental impact of a product. The international standard for life-cycle assessment, ISO 14040:2006, which was reviewed and confirmed in 2016, states that "LCA addresses the environmental aspects and potential environmental impacts (e.g., use of resources and the environmental consequences of releases) throughout a product's life-cycle from raw material acquisition through production, use, end-of-life treatment, recycling, and final disposal (i.e., cradle-to-grave)". All life-cycle stages of the algae-to-biodiesel process via subcritical extraction are thoroughly studied during LCA analysis. The cradle-to-gate assessment provides complete information from the start to the end of the product's life, comprising all processes from the supply of raw materials to the production of the final product.

# 1.2 Problem Statement

Algae are a unique biomass feedstock for the sustainable production of biofuels. Algae, one of the fastest-growing photosynthetic organisms on earth, have biomass productivity rates higher than terrestrial plants (Alba, Torri, Fabbri, Kersten, & Brilman, 2013). Benefits of algae over food crops include fast growth rates, less water intake, adaptation to various water sources (fresh, seawater, saline/brackish and wastewater), high photosynthetic efficiency, carbon dioxide (CO<sub>2</sub>) bio-sequestration, phytoremediation, inexpensive cultivation techniques using non-arable land and short harvesting periods. Notwithstanding these benefits, algal biofuel development faces a few drawbacks which include low biomass densities and high operating costs for biomass generation and conversion (Harun et al., 2014). Although algal-based biofuels generate approximately 13% CO<sub>2</sub> lower emissions from combustion relative to CO<sub>2</sub> emissions from petroleum diesel, in terms of absolute emission levels, algal biofuels can be significantly high for full-scale applications (Nair & Paulose, 2014). The development of biofuels from algal biomass has been significantly successful under lab-scale conditions (Hannon, Gimpel, Tran, Rasala, & Mayfield, 2010). However, opportunities for commercial-scale applications should focus on addressing related environmental, technological, and economic drawbacks (Khan, Shin, & Kim, 2018).

A body of work has been reported on algae extraction using alkaline treatment (Harun, Jason, Cherrington, & Danquah, 2011), enzymatic treatment (Harun & Danquah, 2011a), acidic treatment (Harun & Danquah, 2011b). Though these methods were found to be effective in the extraction process, the use of toxic organic solvents, expensive enzymes and treatment conditions makes the process non-commercially feasible. In view of the economic and environmental needs, it is desirable to explore the use of subcritical water extraction (SCW) as fundamental research in recovering valuable materials from microalgal biomass.

The SCW technique is simpler, more environmentally friendly and can reach high extraction in a very short time (Kumar, Yadav, Kumar, Vyas, & Dhaliwal, 2017). In general, this technique is the most promising technology that can be the first step to the fractionation and obtaining high-value products according to the biorefinery concept (Alba et al., 2011). Depending on the operational conditions (temperature, residence time, particle size, moisture, and reactor configuration), subcritical extraction can cause several effects over the product yield and its quality. The conventional method for biodiesel production from algae via SCW is a two-step process, i.e., extraction of oil from algae via SCW and catalyst-mediated transesterification of extracted oil to biodiesel. Hence, the conventional process has many disadvantages, including expensive and complicated downstream processing steps (Bi, He, & McDonald, 2015). Therefore, to circumvent these disadvantages, this research study proposes a one-step, *insitu* algal biodiesel production using subcritical methanol extraction (SCM).

This research work will unite oil extraction and transesterification as a single process for producing high-quality biodiesel from algae, a potential and promising feedstock. High capital costs due to low lipid productivity of microalgae is a major bottleneck, hindering the commercial production of microalgal oil-derived biodiesel (Dahmani, Zerrouki, Ramanna, Rawat, & Bux, 2016). *Chlorella pyrenoidosa* was selected for conducting the feasibility study of high biomass productivity low-lipid algal strain for maximum oil extraction. Reaction temperature, reaction time and biomass loading were identified as major influencing factors from previous literature on subcritical studies (Edeh, Overton, & Bowra, 2019; Ravber, Knez, & Škerget, 2015). The batch experimental setup in this research project limits the consideration of these three variables as independent variables as these variables can be systematically manipulated

during the experimental run. Therefore, this study aims to evaluate the crude biodiesel production from *C. pyrenoidosa* under two-step SCW and one-step SCM along with three independent variables.

Response surface methodology (RSM) is used to assess the importance of independent variables and their interactions; hence, it is applied in the optimization of independent variables (Bai, Saren, & Huo, 2015). In this work, RSM was applied to assess the oil yield from SCW and SCW of *C. pyrenoidosa* from the optimization of three independent variables – reaction temperature, reaction time and biomass loading. Thus, RSM allows understanding the interactions among independent variables over oil yield. Kinetic studies on SCM of algae remains unexplored. The extraction rate of oil is an important parameter for constructing large-scale extraction units (Saxena, Sharma, & Sambi, 2011). Three kinetic models were evaluated to fit the experimental data of SCW and SCM. LCA is an important tool for evaluating new technologies and it helps to identify the technical bottlenecks and therefore encourages the eco-design of an effective and sustainable production chain (Lardon, Hélias, Sialve, Steyer, & Bernard, 2009). Hence, the economic and environmental impacts of the subcritical extraction systems were investigated through the LCA analysis.

### 1.3 Objectives

### 1.3.1 Overall Objective

To develop a one-step algal crude biodiesel production process to eliminate expensive and complicated downstream processing steps associated with conventional two-step approaches.

# 1.3.2 Specific Objectives

The following are the specific objectives:

- a. To evaluate the effects of process parameters via RSM for the two-step SCW and one-step SCM of algal crude biodiesel production processes.
- b. To evaluate three kinetic models for SCW and SCM of *C. pyrenoidosa* for the algal biodiesel production process.
- c. To analyze the LCA of the algae-to-biodiesel process via subcritical extraction.

# 1.4 Scope of the Study

The scope of the study is summarized based on the thesis objectives as below:

#### a. Evaluation of the effects of process parameters

First, standard methods were implemented to characterize the algae, *C. pyrenoidosa*, including proximate and ultimate analyses. Secondly, SCW experiments were carried out as per RSM with three independent factors: reaction temperature (170

to 370 °C), reaction time (1 to 20 min) and biomass loading (1 to 15%). Thirdly, SCM experiments were carried out as per RSM with three independent factors: reaction temperature (140 to 220 °C), reaction time (1 to 15 min) and methanol to algae ratio (1 to 9 wt.%). The effects of process parameters are evaluated and the optimum conditions for maximum oil yield from each subcritical extraction process were achieved.

#### b. Investigation of the kinetic and thermodynamic parameters

The experimental data of SCW and SCM were fitted with three kinetic models, namely, first-order rate law kinetic model, second-order rate law kinetic model and Fick's law kinetic model— to presume the extraction mechanism parameters.

# c. Evaluation of LCA analysis of the algae-to-biodiesel process via subcritical extraction

Mass and energy balances were evaluated to perform LCA analysis of the algae-to-biodiesel process. This study also compared the energy requirements of subcritical water and subcritical methanol processes.

# 1.5 Significance of the Study

The purpose of this thesis was to compare SCW and SCM techniques on the production of biodiesel from *C. pyrenoidosa*. The emphasis is put on the extraction of maximum oil yield which is a basis for such extraction techniques. To compare the effects of process parameters, RSM is utilized to optimize the reaction conditions. The commercial application of the subcritical technique is limited due to high operating and investment costs. Kinetics data are crucial in understanding the extraction process in detail. LCA studies provide a better insight into operating costs between these two extraction techniques. The goal of this study is to generate a major interest in utilizing subcritical extraction in a wide-scale commercial biofuel production facility exploiting algae as a renewable source.

#### REFERENCES

- Abdelmoez, W., & Abdelfatah, R. (2017). Therapeutic Compounds From Plants Using Subcritical Water Technology *Water Extraction of Bioactive Compounds* (pp. 51-68): Elsevier.
- Acién, F., Fernández, J., Magán, J., & Molina, E. (2012). Production cost of a real microalgae production plant and strategies to reduce it. *Biotechnology advances*, 30(6), 1344-1353.
- Aguilera, J. M. (2005). Why food microstructure? *Journal of food engineering*, 67(1-2), 3-11.
- Ahmad, A., Yasin, N. M., Derek, C., & Lim, J. (2014). Kinetic studies and thermodynamics of oil extraction and transesterification of Chlorella sp. for biodiesel production. *Environmental technology*, 35(7), 891-897.
- Akhtar, J., & Amin, N. A. S. (2011). A review on process conditions for optimum bio-oil yield in hydrothermal liquefaction of biomass. *Renewable and Sustainable Energy Reviews*, 15(3), 1615-1624.
- Alba, L. G., Torri, C., Fabbri, D., Kersten, S. R., & Brilman, D. W. W. (2013). Microalgae growth on the aqueous phase from hydrothermal liquefaction of the same microalgae. *Chemical engineering journal*, 228, 214-223.
- Alba, L. G., Torri, C., Samorì, C., van der Spek, J., Fabbri, D., Kersten, S. R., & Brilman, D. W. (2011). Hydrothermal treatment (HTT) of microalgae: evaluation of the process as conversion method in an algae biorefinery concept. *Energy & Fuels*, 26(1), 642-657.
- Aliakbarian, B., Fathi, A., Perego, P., & Dehghani, F. (2012). Extraction of antioxidants from winery wastes using subcritical water. *The Journal of Supercritical Fluids*, 65, 18-24.
- Anastasakis, K., & Ross, A. (2011). Hydrothermal liquefaction of the brown macro-alga *Laminaria Saccharina*: Effect of reaction conditions on product distribution and composition. *Bioresource technology, 102*(7), 4876-4883.
- Aslam, A., Thomas-Hall, S. R., Manzoor, M., Jabeen, F., Iqbal, M., Uz Zaman, Q., Tahir, M. A. (2018). Mixed microalgae consortia growth under higher concentration of CO2 from unfiltered coal fired flue gas: Fatty acid profiling and biodiesel production. *Journal of Photochemistry and Photobiology B: Biology, 179*, 126-133.
- Asri, N. P., Machmudah, S., Wahyudiono, W., Suprapto, S., Budikarjono, K., Roesyadi, A., & Goto, M. (2013). Non catalytic transesterification of vegetables oil to biodiesel in sub-and supercritical methanol: A kinetic's study. *Bulletin of Chemical Reaction Engineering & Catalysis, 7*(3), 215-223.
- Bach, Q.-V., Sillero, M. V., Tran, K.-Q., & Skjermo, J. (2014). Fast hydrothermal liquefaction of a Norwegian macro-alga: Screening tests. *Algal Research*, 6, 271-276.

- Bai, Y., Saren, G., & Huo, W. (2015). Response surface methodology (RSM) in evaluation of the vitamin C concentrations in microwave treated milk. *Journal of food science and technology*, 52(7), 4647-4651.
- Baümler, E. R., Crapiste, G. H., & Carelli, A. A. (2010). Solvent extraction: kinetic study of major and minor compounds. *Journal of the American Oil Chemists' Society*, 87(12), 1489-1495.
- Bhuana, D., Qadariyah, L., Panjaitan, R., & Mahfud, M. (2020). Optimization of biodiesel production from Chlorella sp through in-situ microwaveassisted acid-catalyzed trans-esterification. Paper presented at the IOP Conference Series: Materials Science and Engineering.
- Bi, Z., He, B. B., & McDonald, A. G. (2015). Biodiesel production from green microalgae Schizochytrium limacinum via in situ transesterification. *Energy & Fuels*, 29(8), 5018-5027.
- Biller, P., Riley, R., & Ross, A. (2011). Catalytic hydrothermal processing of microalgae: decomposition and upgrading of lipids. *Bioresource technology*, 102(7), 4841-4848.
- Biller, P., & Ross, A. (2011). Potential yields and properties of oil from the hydrothermal liquefaction of microalgae with different biochemical content. *Bioresource technology*, *102*(1), 215-225.
- Bird, R. B. (2002). Transport phenomena. Appl. Mech. Rev., 55(1), R1-R4.
- Bucić-Kojić, A., Sovová, H., Planinić, M., & Tomas, S. (2013). Temperaturedependent kinetics of grape seed phenolic compounds extraction: experiment and model. *Food Chemistry*, *136*(3-4), 1136-1140.
- Budrat, P., & Shotipruk, A. (2009). Enhanced recovery of phenolic compounds from bitter melon (*Momordica charantia*) by subcritical water extraction. *Separation and Purification Technology*, 66(1), 125-129.
- Cacace, J., & Mazza, G. (2003). Mass transfer process during extraction of phenolic compounds from milled berries. *Journal of food engineering*, *59*(4), 379-389.
- Caporgno, M. P., Pruvost, J., Legrand, J., Lepine, O., Tazerout, M., & Bengoa,
  C. (2016). Hydrothermal liquefaction of Nannochloropsis oceanica in different solvents. *Bioresource technology*, *214*, 404-410.
- Chan, C.-H., Yusoff, R., & Ngoh, G.-C. (2014). Modeling and kinetics study of conventional and assisted batch solvent extraction. *Chemical engineering research and design, 92*(6), 1169-1186.
- Channiwala, S., & Parikh, P. (2002). A unified correlation for estimating HHV of solid, liquid and gaseous fuels. *Fuel*, *81*(8), 1051-1063.
- Chen, M., Liu, T., Chen, X., Chen, L., Zhang, W., Wang, J., Peng, X. (2012). Subcritical co-solvents extraction of lipid from wet microalgae pastes of *Nannochloropsis* sp. *European Journal of Lipid Science and Technology*, *114*(2), 205-212.
- Chen, W.-T., Zhang, Y., Zhang, J., Schideman, L., Yu, G., Zhang, P., & Minarick, M. (2014). Co-liquefaction of swine manure and mixed-culture algal

biomass from a wastewater treatment system to produce bio-crude oil. *Applied Energy*, *128*, 209-216.

- Chen, W.-T., Zhang, Y., Zhang, J., Yu, G., Schideman, L. C., Zhang, P., & Minarick, M. (2014). Hydrothermal liquefaction of mixed-culture algal biomass from wastewater treatment system into bio-crude oil. *Bioresource technology*, *152*, 130-139.
- Cheng, J., Huang, R., Yu, T., Li, T., Zhou, J., & Cen, K. (2014). Biodiesel production from lipids in wet microalgae with microwave irradiation and bio-crude production from algal residue through hydrothermal liquefaction. *Bioresource technology*, *151*, 415-418.
- Chisti, Y. (2007). Biodiesel from microalgae. *Biotechnology advances, 25*(3), 294-306.
- Chisti, Y. (2016). Large-scale production of algal biomass: raceway ponds *Algae Biotechnology* (pp. 21-40): Springer.
- Cissé, M., Bohuon, P., Sambe, F., Kane, C., Sakho, M., & Dornier, M. (2012). Aqueous extraction of anthocyanins from Hibiscus sabdariffa: Experimental kinetics and modeling. *Journal of food engineering, 109*(1), 16-21.
- Collet, P., Hélias, A., Lardon, L., Ras, M., Goy, R.-A., & Steyer, J.-P. (2011). Lifecycle assessment of microalgae culture coupled to biogas production. *Bioresource technology*, *10*2(1), 207-214.
- Cooney, M., Young, G., & Nagle, N. (2009). Extraction of Bio-oils from Microalgae. Separation & Purification Reviews, 38(4), 291-325.
- Crank, J. (1975). The mathematics of diffusion, 414 p: Oxford University Press, New York.
- Crossley, J., & Aguilera, J. (2001). Modeling the effect of microstructure on food extraction. *Journal of Food Process Engineering*, 24(3), 161-177.
- Curran, M. A. (2012). Life cycle assessment handbook: a guide for environmentally sustainable products: John Wiley & Sons.
- D'Oca, M. G. M., Viêgas, C. V., Lemoes, J. S., Miyasaki, E. K., Morón-Villarreyes, J. A., Primel, E. G., & Abreu, P. C. (2011). Production of FAMEs from several microalgal lipidic extracts and direct transesterification of the Chlorella pyrenoidosa. *biomass and bioenergy*, *35*(4), 1533-1538.
- Dahmani, S., Zerrouki, D., Ramanna, L., Rawat, I., & Bux, F. (2016). Cultivation of Chlorella pyrenoidosa in outdoor open raceway pond using domestic wastewater as medium in arid desert region. *Bioresource technology*, 219, 749-752.
- Davis, M. E., & Davis, R. (2003). Effects of Transport Limitations on Rates of Solid-Catalyzed Reactions. Fundamentals of Chemical Reaction Engineering, 1.
- Delrue, F., Li-Beisson, Y., Setier, P.-A., Sahut, C., Roubaud, A., Froment, A.-K., & Peltier, G. (2013). Comparison of various microalgae liquid biofuel

production pathways based on energetic, economic and environmental criteria. *Bioresource technology*, *136*, 205-212.

- Durling, N. E., Catchpole, O. J., Grey, J. B., Webby, R. F., Mitchell, K. A., Foo, L. Y., & Perry, N. B. (2007). Extraction of phenolics and essential oil from dried sage (Salvia officinalis) using ethanol–water mixtures. *Food Chemistry*, 101(4), 1417-1424.
- Eboibi, B., Lewis, D., Ashman, P., & Chinnasamy, S. (2014). Effect of operating conditions on yield and quality of biocrude during hydrothermal liquefaction of halophytic microalga *Tetraselmis* sp. *Bioresource technology*, *170*, 20-29.
- Edeh, I., Overton, T., & Bowra, S. (2019). Optimization of subcritical watermediated lipid extraction from activated sludge for biodiesel production. *Biofuels*, 1-7.
- Eikani, M. H., Golmohammad, F., & Rowshanzamir, S. (2007). Subcritical water extraction of essential oils from coriander seeds (*Coriandrum sativum* L.). *Journal of food engineering*, *80*(2), 735-740.
- El-Hefny, N. E. (2017). Chemical Kinetics and Reaction Mechanisms in Solvent Extraction: New Trends and Applications. *Journal of Physical Science*, *28*(1), 129-156.
- Elliott, D. C. (2011). Hydrothermal processing. *Thermochemical Processing of Biomass*, 200-231.
- Elliott, D. C., Hart, T. R., Schmidt, A. J., Neuenschwander, G. G., Rotness, L. J., Olarte, M. V., . . . Holladay, J. E. (2013). Process development for hydrothermal liquefaction of algae feedstocks in a continuous-flow reactor. *Algal Research*, 2(4), 445-454.
- Fasaei, F., Bitter, J., Slegers, P., & Van Boxtel, A. (2018). Techno-economic evaluation of microalgae harvesting and dewatering systems. *Algal Research*, *31*, 347-362.
- Felix, C., Ubando, A., Madrazo, C., Culaba, A., Go, A. W., Sutanto, S., Chang, J.-S. (2017). Uncatalyzed direct biodiesel production from wet microalgae under subcritical conditions. Paper presented at the 2017IEEE 9th International Conference on Humanoid, Nanotechnology, Information Technology, Communication and Control, Environment and Management (HNICEM).
- Felix, C., Ubando, A., Madrazo, C., Gue, I. H., Sutanto, S., Tran-Nguyen, P. L., Chang, J.-S. (2019). Non-catalytic in-situ (trans) esterification of lipids in wet microalgae Chlorella vulgaris under subcritical conditions for the synthesis of fatty acid methyl esters. *Applied Energy*, 248, 526-537.
- Fernández, F. A., Sevilla, J. F., Pérez, J. S., Grima, E. M., & Chisti, Y. (2001). Airlift-driven external-loop tubular photobioreactors for outdoor production of microalgae: assessment of design and performance. *Chemical Engineering Science*, *56*(8), 2721-2732.
- Fillières, R., Benjelloun-Mlayah, B., & Delmas, M. (1995). Ethanolysis of rapeseed oil: Quantitation of ethyl esters, mono-, di-, and triglycerides

and glycerol by high-performance size-exclusion chromatography. *Journal of the American Oil Chemists' Society*, 72(4), 427-432.

- Fortier, M.-O. P., Roberts, G. W., Stagg-Williams, S. M., & Sturm, B. S. (2014). Life cycle assessment of bio-jet fuel from hydrothermal liquefaction of microalgae. *Applied Energy*, 122, 73-82.
- Franco, D., Pinelo, M., Sineiro, J., & Núñez, M. J. (2007). Processing of Rosa rubiginosa: Extraction of oil and antioxidant substances. *Bioresource technology*, 98(18), 3506-3512.
- Fuad, F. M., & Karim, K. A. (2017). Kinetics study of oil extraction from Calophyllum inophyllum seeds using ultrasonic-assisted extraction technique. *Journal of Physical Science*, 28(2), 57-69.
- Gai, C., Li, Y., Peng, N., Fan, A., & Liu, Z. (2015). Co-liquefaction of microalgae and lignocellulosic biomass in subcritical water. *Bioresource technology*, *185*, 240-245.
- Gai, C., Zhang, Y., Chen, W.-T., Zhang, P., & Dong, Y. (2014). Energy and nutrient recovery efficiencies in biocrude oil produced via hydrothermal liquefaction of *Chlorella pyrenoidosa*. *RSC Advances*, 4(33), 16958-16967.
- García-Pérez, J. V., Ozuna, C., Ortuño, C., Cárcel, J. A., & Mulet, A. (2011). Modeling ultrasonically assisted convective drying of eggplant. *Drying Technology*, 29(13), 1499-1509.
- Go, A. W., Sutanto, S., Zullaikah, S., Ismadji, S., & Ju, Y.-H. (2016). A new approach in maximizing and direct utilization of whole Jatropha curcas L. kernels in biodiesel production-technological improvement. *Renewable Energy*, *85*, 759-765.
- Halim, R., Danquah, M. K., & Webley, P. A. (2012). Extraction of oil from microalgae for biodiesel production: A review. *Biotechnology advances*, 30(3), 709-732.
- Halim, R., Gladman, B., Danquah, M. K., & Webley, P. A. (2011). Oil extraction from microalgae for biodiesel production. *Bioresource technology*, 102(1), 178-185.
- Hannon, M., Gimpel, J., Tran, M., Rasala, B., & Mayfield, S. (2010). Biofuels from algae: challenges and potential. *Biofuels*, *1*(5), 763-784.
- Harun, R., & Danquah, M. K. (2011a). Enzymatic hydrolysis of microalgal biomass for bioethanol production. *Chemical engineering journal*, 168(3), 1079-1084.
- Harun, R., & Danquah, M. K. (2011b). Influence of acid pre-treatment on microalgal biomass for bioethanol production. *Process Biochemistry*, 46(1), 304-309.
- Harun, R., Danquah, M. K., & Forde, G. M. (2010). Microalgal biomass as a fermentation feedstock for bioethanol production. *Journal of Chemical Technology and Biotechnology*, 85(2), 199-203.

- Harun, R., Jason, W., Cherrington, T., & Danquah, M. K. (2011). Exploring alkaline pre-treatment of microalgal biomass for bioethanol production. *Applied Energy*, *88*(10), 3464-3467.
- Harun, R., Yip, J. W., Thiruvenkadam, S., Ghani, W. A., Cherrington, T., & Danquah, M. K. (2014). Algal biomass conversion to bioethanol–a stepby-step assessment. *Biotechnology journal*, 9(1), 73-86.
- Hassas-Roudsari, M., Chang, P. R., Pegg, R. B., & Tyler, R. T. (2009). Antioxidant capacity of bioactives extracted from canola meal by subcritical water, ethanolic and hot water extraction. *Food Chemistry*, *114*(2), 717-726.
- He, L., Zhang, X., Xu, H., Xu, C., Yuan, F., Knez, Ž., Gao, Y. (2012). Subcritical water extraction of phenolic compounds from pomegranate (*Punica granatum* L.) seed residues and investigation into their antioxidant activities with HPLC-ABTS<sup>+</sup> assay. *Food and bioproducts processing*, 90(2), 215-223.
- Herrero, M., Martín-Álvarez, P. J., Señoráns, F. J., Cifuentes, A., & Ibáñez, E. (2005). Optimization of accelerated solvent extraction of antioxidants from Spirulina platensis microalga. Food Chemistry, 93(3), 417-423.
- Hirschfelder, J., Curtiss, C. F., & Bird, R. (1954). Molecular theory of gases and liquids, New York. *J. Wiley*.
- Hojnik, M., Škerget, M., & Knez, Ž. (2008). Extraction of lutein from Marigold flower petals–Experimental kinetics and modelling. *LWT-Food Science* and Technology, 41(10).
- Imahara, H., Minami, E., Hari, S., & Saka, S. (2008). Thermal stability of biodiesel in supercritical methanol. *Fuel*, *87*(1), 1-6.
- Islam, M. A., Ayoko, G. A., Brown, R., Stuart, D., & Heimann, K. (2013). Influence of fatty acid structure on fuel properties of algae derived biodiesel. procedia engineering, 56, 591-596.
- Janulis, P. (2004). Reduction of energy consumption in biodiesel fuel life cycle. *Renewable Energy*, *29*(6), 861-871.
- Jazrawi, C., Biller, P., Ross, A. B., Montoya, A., Maschmeyer, T., & Haynes, B. S. (2013). Pilot plant testing of continuous hydrothermal liquefaction of microalgae. *Algal Research*, 2(3), 268-277.
- Jesikha, M. (2012). Fatty acid methyl esters characteristic and esterification of some vegetable oils for production of biodiesel. *Res Inventy*, *1*(12), 50-53.
- Jin, B., Duan, P., Xu, Y., Wang, F., & Fan, Y. (2013). Co-liquefaction of microand macroalgae in subcritical water. *Bioresource technology*, 149, 103-110.
- Jin, B., Duan, P., Zhang, C., Xu, Y., Zhang, L., & Wang, F. (2014). Non-catalytic liquefaction of microalgae in sub-and supercritical acetone. *Chemical engineering journal, 254*, 384-392.

- Johnson, M. C., & Tester, J. W. (2013). Lipid Transformation in Hydrothermal Processing of Whole Algal Cells. *Industrial & engineering chemistry research, 52*(32), 10988-10995.
- Kabuba, J., & Huberts, R. (2009). Steam extraction of essential oils: investigation of process parameters. *The Canadian Journal of chemical Engineering*, 87(6), 915-920.
- Käferböck, A., Smetana, S., de Vos, R., Schwarz, C., Toepfl, S., & Parniakov, O. (2020). Sustainable extraction of valuable components from Spirulina assisted by pulsed electric fields technology. *Algal Research, 48*, 101914.
- Kanmaz, E. Ö., & Ova, G. (2013). The effective parameters for subcritical water extraction of SDG lignan from flaxseed (Linum usitatissimum L.) using accelerated solvent extractor. *European Food Research and Technology*, 237(2), 159-166.
- Kaymak-Ertekin, F., & Sultanoğlu, M. (2000). Modelling of mass transfer during osmotic dehydration of apples. *Journal of food engineering, 46*(4), 243-250.
- Khan, M. I., Shin, J. H., & Kim, J. D. (2018). The promising future of microalgae: current status, challenges, and optimization of a sustainable and renewable industry for biofuels, feed, and other products. *Microbial cell factories, 17*(1), 36.
- Krishna, I. M., Manickam, V., Shah, A., & Davergave, N. (2017). Environmental management: science and engineering for industry: Butterworth-Heinemann.
- Kumar, K., Yadav, A. N., Kumar, V., Vyas, P., & Dhaliwal, H. S. (2017). Food waste: a potential bioresource for extraction of nutraceuticals and bioactive compounds. *Bioresources and Bioprocessing*, 4(1), 18.
- Lang, X., Dalai, A. K., Bakhshi, N. N., Reaney, M. J., & Hertz, P. (2001). Preparation and characterization of bio-diesels from various bio-oils. *Bioresource technology*, *80*(1), 53-62.
- Lardon, L., Hélias, A., Sialve, B., Steyer, J.-P., & Bernard, O. (2009). Life-cycle assessment of biodiesel production from microalgae (pp. 6475-6481): ACS Publications.
- Lewis, T., Nichols, P. D., & McMeekin, T. A. (2000). Evaluation of extraction methods for recovery of fatty acids from lipid-producing microheterotrophs. *Journal of Microbiological Methods*, 43(2), 107-116.
- Li, H., Hu, J., Zhang, Z., Wang, H., Ping, F., Zheng, C., He, Q. (2014). Insight into the effect of hydrogenation on efficiency of hydrothermal liquefaction and physico-chemical properties of biocrude oil. *Bioresource technology*, *163*, 143-151.
- Li, H., Liu, Z., Zhang, Y., Li, B., Lu, H., Duan, N., Si, B. (2014). Conversion efficiency and oil quality of low-lipid high-protein and high-lipid lowprotein microalgae via hydrothermal liquefaction. *Bioresource technology*, *154*, 322-329.

- Li, J., Wang, G., Chen, M., Li, J., Yang, Y., Zhu, Q., Liu, H. (2014). Deoxyliquefaction of three different species of macroalgae to high-quality liquid oil. *Bioresource technology, 169*, 110-118.
- Liotta, C., Hallett, J., Pollet, P., & Eckert, C. (2007). *Reactions in nearcritical water*: Blackwell Publishing Ltd: Oxford, UK.
- López Barreiro, D., Zamalloa, C., Boon, N., Vyverman, W., Ronsse, F., Brilman, W., & Prins, W. (2013). Influence of strain-specific parameters on hydrothermal liquefaction of microalgae. *Bioresource technology*, 146, 463-471.
- Lou, D.-W., Lee, X., & Pawliszyn, J. (2008). Extraction of formic and acetic acids from aqueous solution by dynamic headspace-needle trap extraction: temperature and pH optimization. *Journal of Chromatography A*, 1201(2), 228-234.
- Meher, L. C., Sagar, D. V., & Naik, S. (2006). Technical aspects of biodiesel production by transesterification—a review. *Renewable and Sustainable Energy Reviews*, 10(3), 248-268.
- Meziane, S., & Kadi, H. (2008). Kinetics and thermodynamics of oil extraction from olive cake. *Journal of the American Oil Chemists' Society, 85*(4), 391-396.
- Molina, E., Fernández, J., Acién, F., & Chisti, Y. (2001). Tubular photobioreactor design for algal cultures. *Journal of biotechnology*, *9*2(2), 113-131.
- Nair, S., & Paulose, H. (2014). Emergence of green business models: the case of algae biofuel for aviation. *Energy Policy*, *65*, 175-184.
- Narbutt, J. (2020). Fundamentals of Solvent Extraction of Metal Ions Liquid-Phase Extraction (pp. 121-155): Elsevier.
- Neveux, N., Yuen, A., Jazrawi, C., Magnusson, M., Haynes, B., Masters, A., de Nys, R. (2014). Biocrude yield and productivity from the hydrothermal liquefaction of marine and freshwater green macroalgae. *Bioresource technology*, *155*, 334-341.
- Ozel, M. Z., Gogus, F., & Lewis, A. C. (2003). Subcritical water extraction of essential oils from *Thymbra spicata*. *Food Chemistry*, *8*2(3), 381-386.
- Panpraneecharoen, S., & Punsuvon, V. (2016). Biodiesel from crude Pongamia pinnata oil under subcritical methanol conditions with calcium methoxide catalyst. *Energy Sources, Part A: Recovery, Utilization, and Environmental Effects, 38*(15), 2225-2230.
- Patel, A., Matsakas, L., Sartaj, K., & Chandra, R. (2020). Extraction of lipids from algae using supercritical carbon dioxide *Green Sustainable Process for Chemical and Environmental Engineering and Science* (pp. 17-39): Elsevier.
- Peng, X., Ma, X., Lin, Y., Wang, X., Zhang, X., & Yang, C. (2016). Effect of process parameters on solvolysis liquefaction of Chlorella pyrenoidosa in ethanol–water system and energy evaluation. *Energy Conversion and Management*, 117, 43-53.

- Petkov, G., & Garcia, G. (2007). Which are fatty acids of the green alga Chlorella? *Biochemical Systematics and Ecology*, *35*(5), 281-285.
- Pham, M., Schideman, L., Sharma, B. K., Zhang, Y., & Chen, W.-T. (2013). Effects of hydrothermal liquefaction on the fate of bioactive contaminants in manure and algal feedstocks. *Bioresource technology*, 149, 126-135.
- Phong, H. Q., Au, T. D., Huong, H. L., & Van Dat, N. (2016). Biodiesel synthesis from algae (Chlorella sp.) in condition of subcritical methanol. *Tap chí Khoa học Trường Đại học Cần Thơ, 4*, 1-5.
- Plaza, M., Santoyo, S., Jaime, L., Avalo, B., Cifuentes, A., Reglero, G., Ibáñez, E. (2012). Comprehensive characterization of the functional activities of pressurized liquid and ultrasound-assisted extracts from *Chlorella vulgaris*. *LWT-Food Science and Technology*, *46*(1), 245-253.
- Popoola, T., & Yangomodou, O. (2006). Extraction, properties and utilization potentials of cassava seed oil. *Biotechnology, 5*(1), 38-41.
- Posten, C. (2012). Design and performance parameters of photobioreactors. *TATuP-Zeitschrift für Technikfolgenabschätzung in Theorie und Praxis*, 21(1), 38-45.
- Provo, J., Fava, J., & Baer, S. (2013). Life Cycle Assessment and the chemical engineer: a marriage of convenience. *Current Opinion in Chemical Engineering*, 2(3), 278-281.
- Qu, W., Pan, Z., & Ma, H. (2010). Extraction modeling and activities of antioxidants from pomegranate marc. *Journal of food engineering, 99*(1), 16-23.
- Qunju, H., Wenzhou, X., Fangfang, Y., Tao, L., & Guanghua, W. (2016). Evaluation of Five Nannochloropsis Sp. Strains for Biodiesel and Poly-Unsaturated Fatty Acids (PUFAs) Production. Curr Synthetic Sys Biol, 4(128), 2332-0737.1000128.
- Rakotondramasy-Rabesiaka, L., Havet, J.-L., Porte, C., & Fauduet, H. (2009). Solid–liquid extraction of protopine from Fumaria officinalis L.—Kinetic modelling of influential parameters. *Industrial crops and products, 29*(2-3), 516-523.
- Ravber, M., Knez, Ž., & Škerget, M. (2015). Optimization of hydrolysis of rutin in subcritical water using response surface methodology. *The Journal of Supercritical Fluids*, 104, 145-152.
- Recchia, L., Boncinelli, P., Cini, E., Vieri, M., Pegna, F. G., & Sarri, D. (2011). *Multicriteria analysis and LCA techniques: With applications to agroengineering problems*: Springer Science & Business Media.
- Reddy, H. K., Muppaneni, T., Rastegary, J., Shirazi, S. A., Ghassemi, A., & Deng, S. (2013). ASI: Hydrothermal extraction and characterization of bio-crude oils from wet *Chlorella sorokiniana* and *Dunaliella tertiolecta*. *Environmental Progress & Sustainable Energy*, 32(4), 910-915.
- Rickman, M., Pellegrino, J., Hock, J., Shaw, S., & Freeman, B. (2013). Life-cycle and techno-economic analysis of utility-connected algae systems. *Algal Research*, 2(1), 59-65.

- Ruiz, H. A., Rodriguez-Jasso, R. M., Fernandes, B. D., Vicente, A. A., & Teixeira, J. A. (2013). Hydrothermal processing, as an alternative for upgrading agriculture residues and marine biomass according to the biorefinery concept: a review. *Renewable and Sustainable Energy Reviews*, 21, 35-51.
- Saxena, D. K., Sharma, S., & Sambi, S. (2011). Kinetics and thermodynamics of cottonseed oil extraction. grasas y aceites, 62(2), 198-205.
- Sayyar, S., Abidin, Z. Z., Yunus, R., & Muhammad, A. (2009). Extraction of oil from Jatropha seeds-optimization and kinetics. *American Journal of Applied Sciences, 6*(7), 1390.
- Sazdanoff, N. (2006). *Modeling and simulation of the algae to biodiesel fuel cycle*. The Ohio State University.
- Schott, H. (1992). Swelling kinetics of polymers. Journal of Macromolecular Science, Part B: Physics, 31(1), 1-9.
- Sereewatthanawut, I., Prapintip, S., Watchiraruji, K., Goto, M., Sasaki, M., & Shotipruk, A. (2008). Extraction of protein and amino acids from deoiled rice bran by subcritical water hydrolysis. *Bioresource technology, 99*(3), 555-561.
- Sheehan, J., Dunahay, T., Benemann, J., & Roessler, P. (1998). A look back at the US Department of Energy's aquatic species program: biodiesel from algae. *National Renewable Energy Laboratory, 3*28.
- Shuping, Z., Yulong, W., Mingde, Y., Kaleem, I., Chun, L., & Tong, J. (2010). Production and characterization of bio-oil from hydrothermal liquefaction of microalgae *Dunaliella tertiolecta* cake. *Energy*, 35(12), 5406-5411.
- Singh, R., Bhaskar, T., & Balagurumurthy, B. (2013). *Biofuels from Algae:* Chapter 11. Hydrothermal Upgradation of Algae into Value-added Hydrocarbons: Elsevier
- Slade, R., & Bauen, A. (2013). Micro-algae cultivation for biofuels: cost, energy balance, environmental impacts and future prospects. *biomass and bioenergy*, 53, 29-38.
- Sokoto, M., Hassan, L., Dangoggo, S., Ahmad, H., & Uba, A. (2011). Influence of fatty acid methyl esters on fuel properties of biodiesel produced from the seeds oil of Curcubita pepo. *Nigerian Journal of Basic and Applied Sciences, 19*(1), 81-86.
- Soumya, C., Avadhani, H. P., & Vidhya, R. (2015). Production of Biofuel from Micro Algae (Chlorella pyrenoidosa) Using Vertical Reactor System and Effect of Nitrogen on Growth and Lipid Content. *Journal of Academia* and Industrial Research (JAIR), 4(7), 179.
- Spigno, G., & De Faveri, D. (2009). Microwave-assisted extraction of tea phenols: a phenomenological study. *Journal of food engineering*, *93*(2), 210-217.
- Sudhakar, K., Suresh, S., & Premalatha, M. (2011). An overview of CO2 mitigation using algae cultivation technology. *International Journal of Chemical Research*, *3*(3), 110-117.

- Suganya, T., & Renganathan, S. (2012). Optimization and kinetic studies on algal oil extraction from marine macroalgae Ulva lactuca. *Bioresource technology*, *107*, 319-326.
- Tan, X., Chu, H., Zhang, Y., Yang, L., Zhao, F., & Zhou, X. (2014). Chlorella pyrenoidosa cultivation using anaerobic digested starch processing wastewater in an airlift circulation photobioreactor. *Bioresource technology*, 170, 538-548.
- Tao, Y., Zhang, Z., & Sun, D.-W. (2014). Experimental and modeling studies of ultrasound-assisted release of phenolics from oak chips into model wine. *Ultrasonics sonochemistry*, 21(5), 1839-1848.
- Tavakoli, O., & Yoshida, H. (2006a). Conversion of scallop viscera wastes to valuable compounds using sub-critical water. *Green Chemistry, 8*(1), 100-106.
- Tavakoli, O., & Yoshida, H. (2006b). Squid oil and fat production from squid wastes using subcritical water hydrolysis: free fatty acids and transesterification. *Industrial & engineering chemistry research, 45*(16), 5675-5680.
- Teo, C. C., Tan, S. N., Yong, J. W. H., Hew, C. S., & Ong, E. S. (2008). Evaluation of the extraction efficiency of thermally labile bioactive compounds in Gastrodia elata Blume by pressurized hot water extraction and microwave-assisted extraction. *Journal of Chromatography A, 1182*(1), 34-40.
- Thiruvenkadam, S., Izhar, S., Yoshida, H., Danquah, M. K., & Harun, R. (2015). Process application of Subcritical Water Extraction (SWE) for algal bioproducts and biofuels production. *Applied Energy*, 154, 815-828.
- Toda, T. A., Sawada, M. M., & Rodrigues, C. E. (2016). Kinetics of soybean oil extraction using ethanol as solvent: Experimental data and modeling. *Food and bioproducts processing, 98*, 1-10.
- Toor, S. S., Reddy, H., Deng, S., Hoffmann, J., Spangsmark, D., Madsen, L. B., Rosendahl, L. A. (2013). Hydrothermal liquefaction of *Spirulina* and *Nannochloropsis salina* under subcritical and supercritical water conditions. *Bioresource technology*, 131, 413-419.
- Toor, S. S., Rosendahl, L., & Rudolf, A. (2011). Hydrothermal liquefaction of biomass: a review of subcritical water technologies. *Energy*, *36*(5), 2328-2342.
- Tsigie, Y. A., Huynh, L. H., Ismadji, S., Engida, A. M., & Ju, Y.-H. (2012). *In situ* biodiesel production from wet *Chlorella vulgaris* under subcritical condition. *Chemical engineering journal, 213*, 104-108.
- Ueno, H., Tanaka, M., Hosino, M., Sasaki, M., & Goto, M. (2008). Extraction of valuable compounds from the flavedo of *Citrus junos* using subcritical water. Separation and Purification Technology, 62(3), 513-516.
- Valdez, P. J., Nelson, M. C., Wang, H. Y., Lin, X. N., & Savage, P. E. (2012). Hydrothermal liquefaction of *Nannochloropsis* sp.: Systematic study of

process variables and analysis of the product fractions. *biomass and bioenergy*, *46*, 317-331.

- Vallance, C. (2017). Reaction Kinetics.
- Van den Berg, N., Dutilh, C. E., & Huppes, G. (1995). *Beginning LCA: a guide into environmental life cycle assessment*: Centrum voor Milieukunde.
- Vardon, D. R., Sharma, B., Scott, J., Yu, G., Wang, Z., Schideman, L., Strathmann, T. J. (2011). Chemical properties of biocrude oil from the hydrothermal liquefaction of *Spirulina* algae, swine manure, and digested anaerobic sludge. *Bioresource technology*, 102(17), 8295-8303.
- Venteris, E. R., Skaggs, R. L., Wigmosta, M. S., & Coleman, A. M. (2014). A national-scale comparison of resource and nutrient demands for algaebased biofuel production by lipid extraction and hydrothermal liquefaction. *biomass and bioenergy*, *64*, 276-290.
- Welty, J. R., Wicks, C. E., Rorrer, G., & Wilson, R. E. (2009). Fundamentals of momentum, heat, and mass transfer: John Wiley & Sons.
- Winterbone, D., & Turan, A. (2015). Advanced thermodynamics for engineers: Butterworth-Heinemann.
- Wong, Y., & Shahirah, R. (2019). Effect of Different Solvent and Ratio Towards Microalgae Oil Production by Ultrasonic Assisted Soxhlet Extraction Techniques. Oriental Journal of Chemistry, 35(4), 1377.
- Wongkittipong, R., Prat, L., Damronglerd, S., & Gourdon, C. (2004). Solid–liquid extraction of andrographolide from plants—experimental study, kinetic reaction and model. *Separation and Purification Technology, 40*(2), 147-154.
- Xiao, X., Song, W., Wang, J., & Li, G. (2012). Microwave-assisted extraction performed in low temperature and in vacuo for the extraction of labile compounds in food samples. *Analytica chimica acta*, 712, 85-93.
- Xu, C. C., Shao, Y., Yuan, Z., Cheng, S., Feng, S., Nazari, L., & Tymchyshyn, M. (2014). Hydrothermal Liquefaction of Biomass in Hot-Compressed Water, Alcohols, and Alcohol-Water Co-solvents for Biocrude Production *Application of Hydrothermal Reactions to Biomass Conversion* (pp. 171-187): Springer.
- Xu, H.-N., & He, C.-H. (2007). Extraction of isoflavones from stem of Pueraria lobata (Willd.) Ohwi using n-butanol/water two-phase solvent system and separation of daidzein. Separation and Purification Technology, 56(1), 85-89.
- Xu, H., Miao, X., & Wu, Q. (2006). High quality biodiesel production from a microalga Chlorella protothecoides by heterotrophic growth in fermenters. *Journal of biotechnology*, 126(4), 499-507.
- Xu, Y.-Q., Ji, W.-B., Yu, P., Chen, J.-X., Wang, F., & Yin, J.-F. (2018). Effect of extraction methods on the chemical components and taste quality of green tea extract. *Food Chemistry*, 248, 146-154.

- Xu, Y., Yu, H., Hu, X., Wei, X., & Cui, Z. (2014). Bio-oil production from algae via thermochemical catalytic liquefaction. *Energy Sources, Part A: Recovery, Utilization, and Environmental Effects, 36*(1), 38-44.
- Yang, L., Li, Y., & Savage, P. E. (2014). Catalytic Hydrothermal Liquefaction of a Microalga in a Two-Chamber Reactor. *Industrial & engineering chemistry research*, 53(30), 11939-11944.
- Yoshida, H., Terashima, M., & Takahashi, Y. (1999). Production of Organic Acids and Amino Acids from Fish Meat by Sub-Critical Water Hydrolysis. *Biotechnology progress, 15*(6), 1090-1094.
- Zanoni, B., Pagliarini, E., & Peri, C. (1992). Modelling the aqueous extraction of soluble substances from ground roasted coffee. *Journal of the Science* of Food and Agriculture, 58(2), 275-279.
- Zhang, Y. (2010). Hydrothermal liquefaction to convert biomass into crude oil. Biofuels from agricultural wastes and byproducts, 201-232.
- Zhang, Z.-S., Li, D., Wang, L.-J., Ozkan, N., Chen, X. D., Mao, Z.-H., & Yang, H.-Z. (2007). Optimization of ethanol–water extraction of lignans from flaxseed. Separation and Purification Technology, 57(1), 17-24.
- Zhu, Y., Albrecht, K. O., Elliott, D. C., Hallen, R. T., & Jones, S. B. (2013). Development of hydrothermal liquefaction and upgrading technologies for lipid-extracted algae conversion to liquid fuels. *Algal Research*, 2(4), 455-464.
- Zighmi, S., Ladjel, S., Goudjil, M. B., & Bencheikh, S. E. (2017). Renewable energy from the seaweed chlorella pyrenoidosa cultivated in developed systems. *International Journal of Renewable Energy Research*, *7*, 49-57.
- Zullaikah, S., Rahkadima, Y. T., & Ju, Y.-H. (2017). A non-catalytic in situ process to produce biodiesel from a rice milling by-product using a subcritical water-methanol mixture. *Renewable Energy*, *111*, 764-770.

# **BIODATA OF STUDENT**

Selvakumar Thiruvenkadam completed his bachelor's degree in Biotechnology from Anna University, India, with the thesis "Utilization of Oil Cakes for Biogenesis of Methane". He then received his master's degree in Biotechnology from KTH Royal Institute of Technology, Sweden. His master's thesis, entitled "Evaluation of Cellruptor pre-treatment on biogas yield from various substrates" was carried out at Scandinavian Biogas Fuels International AB, Sweden. With research interests in biofuels, his PhD research study focuses on biodiesel production from algal biomass.



# LIST OF PUBLICATIONS

- Thiruvenkadam, S., Izhar, S., Hiroyuki, Y., & Harun, R. (2019). One-step microalgal biodiesel production from *Chlorella pyrenoidosa* using subcritical methanol extraction (SCM) technology. *Biomass and Bioenergy*, 120, 265-272.
- Thiruvenkadam, S., Izhar, S., Hiroyuki, Y., & Harun, R. (2018). Subcritical Water Extraction of *Chlorella pyrenoidosa*: Optimization through Response Surface Methodology. *BioMed research international*, 2018.
- Thiruvenkadam, S., Izhar, S., Hiroyuki, Y., Danquah, M. K., & Harun, R. (2015). Process application of Subcritical Water Extraction (SWE) for algal bioproducts and biofuels production. *Applied energy*, *154*, 815-828.
- Thiruvenkadam, S., & Harun, R. (2016). Subcritical Water Extraction of algal oil from *Chlorella pyrenoidosa*. World Research & Innovation Convention on Engineering & Technology 2016, Langkawi



# **UNIVERSITI PUTRA MALAYSIA**

# STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

# ACADEMIC SESSION: Second Semester 2020/2021

# TITLE OF THESIS / PROJECT REPORT:

# SUBCRITICAL WATER EXTRACTION FOR MICROALGAL BIODIESEL

PRODUCTION

### NAME OF STUDENT:

### SELVAKUMAR THIRUVENKADAM

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

- 1. This thesis/project report is the property of Universiti Putra Malaysia.
- 2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
- 3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as:

