



**UNIVERSITI PUTRA MALAYSIA**

***ABROGATION OF ONCOGENIC K-RAS FUNCTION BY  
ANDROGRAPHOLIDE DERIVATIVES VIA IN SILICO, IN VITRO, AND IN  
VIVO APPROACHES***

**QUAH SHUN YING**

**FPSK(p) 2021 15**



**ABROGATION OF ONCOGENIC K-RAS FUNCTION BY  
ANDROGRAPHOLIDE DERIVATIVES VIA *IN SILICO*, *IN VITRO*, AND *IN VIVO* APPROACHES**

By

**QUAH SHUN YING**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra  
Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of  
Philosophy**

**April 2021**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

**ABROGATION OF ONCOGENIC K-RAS FUNCTION BY ANDROGRAPHOLIDE DERIVATIVES VIA *IN SILICO*, *IN VITRO*, AND *IN VIVO* APPROACHES**

By

**QUAH SHUN YING**

April 2021

**Chair : Prof. Johnson Stanlas, PhD**  
**Faculty : Medicine and Health Sciences**

The rat sarcoma (Ras) proteins are small guanosine triphosphatases (GTPases) that act as molecular switches in major signalling pathways involved in cell proliferation, differentiation, and survival, such as mitogen-activated protein kinase (MAPK) and phosphatidylinositol 3-kinase (PI3K) cascades. Ras exists in three isoforms – K-Ras, H-Ras, and N-Ras. Approximately 30% of all human cancers harbour Ras mutations, with the most frequently mutated isoform being K-Ras, which exclusively appears in pancreatic ductal adenocarcinoma (PDAC). Mutated K-Ras proteins are constitutively active with the GTPase activity being compromised. Oncogenic K-Ras is currently a valuable oncology target and its inhibition represents an important therapeutic strategy. Recent *in silico* study has revealed a direct binding of andrographolide (AGP) and its benzylidene derivatives, SRJ09 and SRJ23, to K-Ras oncoprotein, which abrogated its function and downstream MAPK signalling. The present study aims to investigate the potential of AGP derivatives as anti-Ras therapeutics through *in silico*, *in vitro*, and *in vivo* approaches. The anticancer potential of SRJ09 and SRJ23 has been well-demonstrated in the human colon (HCT-116) and prostate (PC-3) cancer cells, respectively. These two cell lines have been made resistant to the compounds previously and were used in the present study to examine the altered gene profile in relation to the expression of regulatory genes involved in the compounds' anticancer activity using microarray analysis. Regulatory genes associated with autophagy and apoptotic processes, such as *ATG12* and *HMOX1*, as well as MAPK and PI3K pathways, such as *FGF19* and *SPRY2* that play major roles in promoting cell growth and survival, were found to be altered. New benzylidene derivatives have been previously synthesised using SRJ09 and SRJ23 as parent compounds, yielding SRS compounds. In the present investigation, the most druggable binding pocket on K-Ras mutants namely p2 was revealed through *in silico* simulations. SRJ23 and SRS157 were found to bind *via* intermolecular hydrogen bonding to this pocket. The

anti-PDAC activity of selected AGP derivatives (SRJ23, SRJ09, SRS07, and SRS157) and their mechanisms of action were elucidated *in vitro*. SRJ23 and SRS157 were shown to perform differently particularly in terms of activity on Erk, a crucial signalling protein in the K-Ras-associated MAPK cascade. Its activation was unanticipatedly enhanced by SRJ23 and significantly suppressed by SRS157 upon 24-h treatment of the compounds. SRS07 presented as a superior anti-PDAC agent by promoting oxidative stress, possibly through enhancement of Akt activation in the K-Ras-mediated PI3K pathway. A simple pharmacokinetic study performed in BALB/c mice at a single dose of 100 mg/kg SRJ23 revealed that the compound achieved a maximum plasma concentration of 18.8  $\mu\text{M}$  after 30 min of administration, with long half-life (4.28 h) and mean residence time (6.30 h). Subsequent *in vivo* antitumour study reported that 100 mg/kg SRS157 delayed the doubling of tumour growth in the PDAC-xenograft nude mouse model more effectively than SRJ23 at the same dose. In conclusion, the outcomes of the present study provide a strong indication of the potential of AGP derivatives, which specifically target the oncogenic K-Ras and abrogate its function, as promising clinical anti-pancreatic cancer candidates.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PEMANSUHAN FUNGSI K-RAS ONKOGENIK OLEH DERIVATIF  
ANDROGRAPHOLIDE MELALUI KAEDAH-KAEDAH *IN SILICO*, *IN VITRO*,  
DAN *IN VIVO***

Oleh

**QUAH SHUN YING**

April 2021

**Pengerusi : Prof. Johnson Stanlas, PhD**  
**Fakulti : Perubatan dan Sains Kesihatan**

Protein sarkoma tikus (Ras) adalah trifosfatase guanosin kecil (GTPases) yang bertindak sebagai pertukaran molekul dalam laluan isyarat utama yang terlibat dalam percambahan, perbezaan, dan kelangsungan hidup sel-sel, contohnya laluan isyarat mitogen-activated protein kinase (MAPK) dan phosphatidylinositol 3-kinase (PI3K). Ras wujud dalam tiga isoform – K-Ras, H-Ras dan N-Ras. Anggaran 30% daripada semua kanser manusia mempunyai mutasi Ras dengan isoform yang paling kerap bermutasi adalah K-Ras yang dijumpai khasnya dalam adenokarsinoma saluran pankreas (PDAC). Protein K-Ras yang bermutasi aktif secara konstitutif disertai oleh aktiviti GTPase yang dikompromikan. Kini, K-Ras onkogenik merupakan sasaran onkologi yang berharga dan perencatannya merupakan satu strategi terapi yang penting. Kajian *in silico* terkini telah mendedahkan pengikatan langsung andrographolide (AGP) dan derivatif benzylidene (SRJ09 dan SRJ23) pada protein mutan K-Ras dimana ia dapat memansuhkan fungsi dan isyarat MAPK. Kajian ini bertujuan untuk menyelidik potensi derivatif AGP sebagai terapeutik anti-Ras melalui kaedah-kaedah *in silico*, *in vitro*, dan *in vivo*. Potensi anti-kanser SRJ09 dan SRJ23 terhadap sel-sel kanser usus besar manusia (HCT-116) dan prostat (PC-3) telah didedahkan. Kedua-dua sel ini dicipta untuk menahan rintangan terhadap sebatian tersebut sebelumnya dan ia digunakan dalam kajian ini untuk memeriksa profil gen yang diubah berkait rapat dengan ekspresi gen pengawalseliam melibatkan sebatian aktiviti anti-kanser dengan menggunakan analisis mikroarray. Gen pengawalseliam yang berhubung dengan proses autophagy dan apoptosis, seperti *ATG12* dan *HMOX1*, serta laluan isyarat MAPK dan PI3K, seperti *FGF19* dan *SPRY2* yang memainkan peranan utama dalam menggalakkan pertumbuhan sel dan kelangsungan hidup, telah didapati terjejas. Derivatif benzylidene yang baru disintesis sebelumnya dengan menggunakan SRJ09 dan SRJ23 sebagai sebatian induk telah menghasilkan sebatian SRS. Dalam penyiasatan ini, poket pengikat yang diramalkan paling sesuai digunakan pada protein mutan K-Ras iaitu p2 telah

didedahkan melalui simulasi *in silico*. SRJ23 dan SRS157 didapati mengikat pada poket ini melalui ikatan hidrogen antara molekul. Kegiatan anti-PDAC sesetengah derivatif AGP yang terpilih (SRJ23, SRJ09, SRS07, dan SRS157) dan mekanisme tindakan mereka telah didedahkan secara *in vitro*. SRJ23 dan SRS157 bertindak secara berbeza terutamanya dari segi aktiviti pada Erk (satu protein isyarat yang penting dalam laluan isyarat MAPK berhubungan dengan K-Ras. Pengaktifan ditingkatkan tanpa disangka-sangka oleh SRJ23 tetapi ditindas dengan ketara oleh SRS157 selepas rawatan sebatian selama 24 jam. SRS07 telah ditunjukkan kemampuannya sebagai agen kanser anti-PDAC yang unggul dengan menggalakkan tekanan oksidatif dan meningkatkan pengaktifan Akt dalam laluan isyarat PI3K yang dimediasi oleh K-Ras. Kajian farmakokinetik yang dilakukan pada tikus BALB/c dengan menggunakan dos tunggal 100 mg/kg SRJ23 telah menunjukkan bahawa sebatian tersebut mencapai kepekatan plasma maksimum iaitu 18.8  $\mu\text{M}$  selepas setengah jam pemberian dengan jangka hayat (4.28 jam) dan purata masa ketinggalan (6.30 jam) yang panjang. Kajian anti-kanser *in vivo* selanjutnya melaporkan bahawa 100 mg/kg SRS157 memperlambat penggadaan pertumbuhan tumor pada model tikus PDAC-xenograft dengan lebih berkesan berbanding dengan SRJ23 pada dos yang sama. Kesimpulannya, hasil kajian ini memberi petunjuk yang lebih kuat mengenai potensi derivatif AGP sebagai calon-calon klinikal untuk anti-kanser pankreas yang menjanjikan, di mana secara khususnya mereka boleh menyasarkan dan memansuhkan fungsi K-Ras onkogenik.

## ACKNOWLEDGEMENTS

First and foremost, I would like to express my deep and sincere gratitude to my Ph.D. research supervisor, Prof. Dr. Johnson Stanslas (the head of Pharmacotherapeutics Unit, Department of Medicine, Faculty of Medicine and Health Sciences, University Putra Malaysia), for giving me the opportunity to do research and providing invaluable guidance and advice throughout this research. His enthusiasm, vision, sincerity and motivation have deeply inspired me. He has taught me the methodology to carry out the research and to present the research works as clearly as possible. I am extremely grateful for the funding support provided by him to attend few conferences overseas, giving me the opportunity to learn new skills and enhance knowledge about my research fields as well as meet many passionate researchers and create networks.

My heartfelt thanks and gratitude go to all my co-supervisors, including Dr. Pran Kishore Deb, who has guided me through the work of molecular modelling, Dr. Sreenivasa Rao Sagineedu for the synthesis work of andrographolide derivatives, Dr. Ho Kok Lian, and Dr. Nizar bin Abdul Manan for their constant assistance and contributions throughout my journey of completion of this project.

I am also grateful and honoured to have been awarded with MyBrain15 scholarship (MyPhD) by Ministry of Higher Education, Malaysia, from Mac 2015 to September 2018. This scholarship has lightened my financial burden, which allows me to focus more on my research study.

My sincere appreciation goes to all members of Cancer Research and Drug Discovery (CRDD) group, an informal group formed by Prof. Dr. Johnson Stanslas, for their assistance, troubleshooting ideas, and friendship: Dr. Jonathan Lim Chee Woei, Dr. Sulaiman Ibrahim, Dr. Teh Yuan Han, Dr. Nasir Ibrahim Khatab, Dr. Ibrahim Badamasi, Soo Hon Liong, Tan Chee Yi, and Suzanne Goh. Special thanks and appreciation to Ong Hui Kian, a Ph.D. student from the Laboratory of Chemical Pathology, for his endless help, enthusiastic encouragement and moral active support offered throughout the study. As a best friend of mine since undergraduate studies, he has always inspired me with his genuine enthusiasm and passion in research.

Last but not least, I am extremely thankful to my family members, particularly to my parents whom I dedicate this thesis, for their never-ending supports, understanding and encouragement throughout the years of my study. Their love has been the major spiritual support in my life.



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

**Johnson Stanslas, PhD**

Professor  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Chairman)

**Ho Kok Lian, PhD**

Associate Professor  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Member)

**Nizar bin Abd Manan, PhD**

Senior Lecturer  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Member)

**Pran Kishore Deb, PhD**

Associate Professor  
Faculty of Pharmacy  
Philadelphia University  
(Member)

---

**ZALILAH MOHD SHARIFF, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 9 September 2021

## Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: \_\_\_\_\_  
Name of Chairman  
of Supervisory  
Committee: Prof. Dr. Johnson Stanslas

Signature: \_\_\_\_\_  
Name of Member of  
Supervisory  
Committee: Associate Prof. Dr. Ho Kok Lian

Signature: \_\_\_\_\_  
Name of Member of  
Supervisory  
Committee: Dr. Nizar bin Abd Manan

Signature: \_\_\_\_\_  
Name of Member of  
Supervisory  
Committee: Associate Prof. Dr. Pran Kishore Deb

## TABLE OF CONTENTS

		Page
<b>ABSTRACT</b>		i
<b>ABSTRAK</b>		iii
<b>ACKNOWLEDGEMENTS</b>		v
<b>APPROVAL</b>		vi
<b>DECLARATION</b>		viii
<b>LIST OF TABLES</b>		xv
<b>LIST OF FIGURES</b>		xviii
<b>LIST OF APPENDICES</b>		xxiii
<b>LIST OF ABBREVIATIONS</b>		xxvi
<b>CHAPTER</b>		
<b>1</b>	<b>INTRODUCTION</b>	1
	1.1 Overview of the Study	1
	1.2 Hypothesis	2
	1.3 Objectives	3
	1.3.1 General Objective	3
	1.3.2 Specific Objectives	3
<b>2</b>	<b>LITERATURE REVIEW</b>	4
	2.1 Cancer	4
	2.2 RAS Genes	5
	2.2.1 Ras Proteins and Their Structure Overview	5
	2.2.2 Intracellular Processing and Membrane Localisation	7
	2.2.3 Interactions with Regulators and Effectors of Ras Signalling Pathways	10
	2.3 Ras Isoform Mutations in Human Tumours	16
	2.3.1 K-Ras Mutations and Pancreatic Cancer Tumorigenesis	19
	2.3.2 K-Ras Addiction in Pancreatic Ductal Adenocarcinoma	20
	2.3.3 K-Ras as a Therapeutic Target – Direct Inhibition	22
	2.4 Andrographolide and Its Derivatives	25
	2.4.1 Modulation of Ras-Associated Cancers by Andrographolide and Its Derivatives	28
<b>3</b>	<b>MICROARRAY-BASED IDENTIFICATION OF GENES ASSOCIATED WITH ANDROGRAPHOLIDE DERIVATIVES-INDUCED RESISTANCE IN COLON AND PROSTATE CANCER CELL LINES</b>	29
	3.1 Introduction	29
	3.2 Materials	31
	3.2.1 Compounds Isolation and Synthesis	31
	3.2.2 Cell Lines	31

3.2.3	Reagents and Chemicals	31
3.2.4	Laboratory Wares and Consumables	32
3.2.5	Equipment	32
3.3	Methodology	33
3.3.1	Compounds Preparation and Dilution	33
3.3.2	Cell Culture	33
3.3.3	Cryopreservation and Recovery	34
3.3.4	Cell Plating	35
3.3.5	<i>In Vitro</i> Cell Viability Assay for Resistance Check	36
3.3.6	Total RNA Extraction	37
3.3.7	In-House Assessment of RNA (Quantity, Quality, Integrity)	38
3.3.8	Gene Expression Profiling	38
3.3.8.1	Assessment of Quantity, Quality, and Integrity of RNA	38
3.3.8.2	Preparation and Quality Check of Biotin-Labelled RNA	38
3.3.8.3	Microarray and Data Analysis	39
3.3.9	Conventional Reverse Transcription-Polymerase Chain Reaction (RT-PCR)	40
3.3.10	Statistical Analysis	40
3.4	Results	41
3.4.1	Resistance Developed in Resistant Cell Lines	41
3.4.2	Quantity, Quality, and Integrity of Extracted RNA	42
3.4.3	Microarray Analysis	47
3.4.3.1	Quality Control	47
3.4.3.2	Differential Expression and Clustering	48
3.4.3.3	Prediction of Protein-Protein Interactions, Functional Enrichments, and Pathway Analysis	60
3.4.4	PCR Validation of Altered Genes	65
3.5	Discussion	66
3.6	Conclusion	72
4	<b>IN SILICO MODELLING AND MOLECULAR DYNAMICS SIMULATION STUDIES OF AGP AND ITS BENZYLIDENE DERIVATIVES</b>	73
4.1	Introduction	73
4.2	Materials	75
4.2.1	Hardware	75
4.2.2	Software	75
4.2.3	Protein X-Ray Crystal Structures	76
4.2.4	Ligand Structures	76
4.3	Methodology	79

	4.3.1	Protein and Ligand Preparation	79
	4.3.2	Determination of Binding Sites	79
	4.3.3	Molecular Docking	82
	4.3.4	Binding Energy Calculation	83
	4.3.5	Prediction of Drug-Like Properties	83
	4.3.6	Molecular Dynamics Simulation	85
4.4	Results		87
	4.4.1	Drug-Like Properties of Ligands	87
	4.4.2	Binding Sites on K-Ras Proteins	89
	4.4.3	Molecular Docking and Binding Mode	91
	4.4.4	Molecular Dynamics Simulation	100
4.5	Discussion		106
4.6	Conclusion		112
<b>5</b>	<b>EFFECTS OF BENZYLIDENE DERIVATIVES OF AGP ON RAS-MAPK AND PI3K/AKT SIGNALLING PATHWAYS IN PDAC CELLS</b>		<b>113</b>
	5.1	Introduction	113
	5.2	Materials	115
	5.2.1	Compounds Isolation and Synthesis	115
	5.2.2	Cell Lines	115
	5.2.3	Reagents and Chemicals	115
	5.2.4	Laboratory Wares and Consumables	116
	5.2.5	Equipment	117
	5.3	Methodology	117
	5.3.1	Compounds Preparation and Dilution	117
	5.3.2	Cell Culture and Plating	118
	5.3.3	<i>In Vitro</i> Growth Inhibitory Assay	118
	5.3.4	Western Blot Analysis	118
	5.3.4.1	Total Protein Extraction from Cancer Cells	118
	5.3.4.2	Protein Quantification	119
	5.3.4.3	Protein Sample Preparation	119
	5.3.4.4	SDS Polyacrylamide Gel Electrophoresis (SDS-PAGE) Electrophoresis	119
	5.3.4.5	Semi-Dry Western Blotting and Immunodetection	120
	5.3.5	Intracellular Reactive Oxygen Species (ROS) Analysis	121
	5.3.6	Statistical Analysis	122
5.4	Results		122
	5.4.1	Growth Inhibitory of Compounds on Pancreatic Cancer Cell Lines	122
	5.4.2	Effects of Compounds on Activation and Expression of Ras-Mediated Downstream Signalling Proteins	127
	5.4.3	Effects of Compounds on the Production of ROS	133
5.5	Discussion		134
5.6	Conclusion		141

<b>6</b>	<b>PHARMACOKINETIC PROFILE OF SRJ23 IN MICE</b>	<b>143</b>
6.1	Introduction	143
6.2	Materials	144
6.2.1	Compounds Isolation and Synthesis	144
6.2.2	Animals	144
6.2.3	Chemicals and Reagents	145
6.2.4	Laboratory Wares and Consumables	145
6.2.5	Equipment	145
6.3	Methodology	146
6.3.1	Preparation of Compound	146
6.3.2	Administration of Compound to Mice	146
6.3.3	Blood Sample Collection and Handling	146
6.3.4	High-Performance Liquid Chromatography (HPLC) Analysis	147
6.3.5	HPLC Method Development and Validation	147
6.3.5.1	Preparation of Neat Standards	148
6.3.5.2	Preparation of Post-Spike Standards	148
6.3.5.3	Preparation of Pre-Spike Standards	149
6.3.5.4	Selectivity	149
6.3.5.5	Linearity, Accuracy, and Precision	149
6.3.5.6	Matrix Effect, Recovery, and Process Efficiency	150
6.3.5.7	Limits of Detection and Quantitation	151
6.3.6	Pharmacokinetic Analysis	152
6.4	Results	152
6.4.1	Method Development and Validation	152
6.4.1.1	Selectivity	152
6.4.1.2	Linearity, Accuracy, and Precision	153
6.4.1.3	Matrix Effect, Recovery, and Process Efficiency	157
6.4.1.4	Limits of Detection (LOD) and Quantitation (LOQ)	158
6.4.2	Pharmacokinetic Profile	160
6.5	Discussion	164
6.6	Conclusion	168
<b>7</b>	<b>ANTITUMOUR ACTIVITY OF SRJ23 AND SRS157 AGAINST PANCREATIC CANCER XENOGRAFT IN NUDE MICE</b>	<b>169</b>
7.1	Introduction	169
7.2	Materials	171
7.2.1	Compounds Isolation and Synthesis	171
7.2.2	Animals and Cell Line	171

7.2.3	Chemicals and Reagents	171
7.2.4	Laboratories Wares and Consumables	172
7.2.5	Equipment	172
7.3	Methodology	173
7.3.1	Development of Human Tumour Xenografts in Nude Mice	173
7.3.2	Preparation and Administration of Compounds to Mice	173
7.3.3	Assessment of the Growth of Tumour Xenografts in Nude Mice	174
7.3.4	Histopathological Analysis of Liver, Kidneys, Spleen, and Tumour Tissues	176
7.3.5	Statistical Analysis	178
7.4	Results	178
7.4.1	Body Weight Changes	178
7.4.2	Antitumour Activities	180
7.4.3	Histopathological Changes	184
7.4.3.1	Acute Toxicity	184
7.4.3.2	<i>In Vivo</i> Antitumour Activity	190
7.5	Discussion	192
7.6	Conclusion	197
<b>8</b>	<b>SUMMARY, CONCLUSION, AND RECOMMENDATIONS FOR FUTURE RESEARCH</b>	<b>198</b>
8.1	Research Summary	198
8.2	General Conclusion	201
8.3	Recommendations for Future Research	201
	<b>REFERENCES</b>	<b>205</b>
	<b>APPENDICES</b>	<b>247</b>
	<b>BIODATA OF STUDENT</b>	<b>331</b>
	<b>LIST OF PUBLICATIONS</b>	<b>332</b>

## LIST OF TABLES

Table		Page
2.1	Five most frequent cancer types diagnosed in worldwide	4
3.1	IC <sub>80</sub> values of SRJ09 and SRJ23 against both parental and resistant HCT-116 or PC-3 cancer cells (96 h), and the fold resistance developed in the resistant cell lines	41
3.2	The amount and purity of biological and technical replicates of all RNA samples based on NanoDrop spectrophotometer measurement	45
3.3	Quality assessment of biological and technical replicates of all RNA samples based on NanoDrop spectrophotometer, Agilent Bioanalyzer, and Qubit RNA High Sensitivity Assay	47
3.4	Total number of genes detected for each RNA sample as analysed on microarray platform <i>via</i> GenomeStudio software	48
3.5	Differentially expressed genes with the adjusted p-value ( $p_{adj}$ ) of $< 1 \times 10^{-10}$ (-log base 10) and fold change (FC) of $\geq 2.0$ (log base 2) in expression levels between parental and SRJ09-resistant HCT-116 cells	50
3.6	Differentially expressed genes with the adjusted p-value ( $p_{adj}$ ) of $< 1 \times 10^{-10}$ (-log base 10) and fold change (FC) of $\geq 2.0$ (log base 2) in expression levels between parental and SRJ23-resistant PC-3 cells	55
3.7	Most significant pathways ( $p < 0.05$ ) identified by Reactome, upon dataset input of interconnected 13 significantly modulated genes in SRJ09-resistant HCT-116 in the STRING protein network	62
4.1	X-ray crystal structures of wild-type K-Ras protein and mutant K-Ras oncoproteins selected from Protein Data Bank (PDB)	77
4.2	Binding pockets on K-RasQ61H targeted by AGP and its benzylidene derivatives	81
4.3	Selected ADME-related QikProp properties and descriptors with the suitable ranges or recommended values used to compare studied compounds with those of 95% of known drugs in the market	84
4.4	Drug-like properties of AGP, SRJ09, and SRJ23, as well as eight derivatives of SRJ23 (SRS150 – SRS157), as predicted by QikProp	88



4.5	Possible binding sites on wild-type and mutant K-Ras proteins, generated using SiteMap calculation, with their respective amino acid residues involved and key properties that determine the suitability as docking sites and their druggability	90
4.6	The free energy of binding ( $\Delta G_{\text{bind}}$ ) of each studied compound (the best conformer) to the selected binding sites on both wild-type and mutant K-Ras proteins	94
4.7	Energy contribution of bonded and non-bonded interactions of each studied compound in complex with mutant K-RasG12V (PDB ID: 4EPX) (the most stable complex with the lowest free binding energy score)	95
5.1	Dilutions of primary and secondary antibodies of various proteins	121
5.2	GI <sub>50</sub> values of SRJ09 and SRJ23, as well as their derivatives SRS07 and SRS157, respectively, in comparison to gemcitabine and DCAI, against four pancreatic cancer cells (96 h)	124
5.3	TGI values of SRJ23 and SRJ09, as well as their derivatives SRS157 and SRS07, respectively, in comparison to gemcitabine and DCAI, against four pancreatic cancer cells (96 h)	125
5.4	LC <sub>50</sub> values of SRJ23 and SRJ09, as well as their derivatives SRS157 and SRS07, respectively, in comparison to gemcitabine and DCAI, against four pancreatic cancer cells (96 h)	126
6.1	Pharmacokinetics (PK) parameters and their definitions	153
6.2	Intra- and inter-day accuracy and precision of HPLC analysis of SRJ23 concentrations	156
6.3	Intra-day precision of HPLC analysis of retention times and peak heights for DDAG and SRJ23, as well as peak height ratio determined at each SRJ23 concentration level	157
6.4	Inter-day precision of HPLC analysis of retention times and peak heights for DDAG and SRJ23, as well as peak height ratio determined at each SRJ23 concentration level	157
6.5	Matrix effect (ME), recovery (RE), and process efficiency (PE), and their precision (%CV) for DDAG and SRJ23 in pooled mice plasma	158

6.6	LOD and LOQ of SRJ23 as calculated using calibration data obtained from the intra-day analysis	159
6.7	Pharmacokinetics (PK) parameters obtained after single intraperitoneal bolus administration of 100 mg/kg SRJ23, and comparison to its parent compound, AGP, and SRJ09	163
7.1	Four-point scale scoring system employed for histopathological evaluation of different H&E-stained sections	178
7.2	The period required for both control and treatment groups of PANC-1 tumour-bearing nude mice to reach a tumour volume that was two times greater than the initial volume (RTV2*) and their expected tumour growth delay ( $T_d$ )	180
7.3	Optimal T/C ratio, which was based on relative tumour volumes of nude mice implanted with PANC-1 human pancreatic tumour, treated with gemcitabine (positive control) and two dose levels of SRJ23 or SRS157	182
7.4	Tumour growth inhibition (TuGI), which was based on terminal tumour weights of nude mice implanted with PANC-1 human pancreatic tumour, treated with gemcitabine (positive control) and two dose levels of SRJ23 or SRS157	182
7.5	Scores of liver, kidney, spleen, and tumour tissue H&E sections of mice treated with various treatments as compared to those of the control group to evaluate acute toxicity and antitumour efficacy	186

## LIST OF FIGURES

Figure		Page
2.1	The two conformational states of Ras	6
2.2	Post-translational modifications of the C-terminal membrane-targeting region of Ras proteins	8
2.3	Regulation of Ras function and its activation upon binding of extracellular signal to receptor tyrosine kinases (RTKs)	11
2.4	The Ras-MAPK signalling pathway	14
2.5	The Ras-PI3K signalling cascade	16
2.6	Distribution of oncogenic mutations in H-Ras, N-Ras, and K-Ras	17
2.7	Isoform-specific single-base missense mutations in different tumour types	18
2.8	The progression model for pancreatic cancers	20
2.9	Chemical structures of compounds that interfere with Ras membrane association and prevent Ras-GTP formation	22
2.10	Key interactions involved between residues in GDP-bound K-RasG12C and the G12C-selective compound 9	24
2.11	AGP and its derivatives	26
2.12	Chemical structure of AGP sulfonate	28
3.1	Micrographs of (A) parental HCT-116 and (B) SRJ09-resistant HCT-116 (HCT-116 <sup>rst09</sup> ) cells	43
3.2	Micrographs of (A) parental PC-3 and (B) SRJ23-resistant PC-3 (PC-3 <sup>rst23</sup> ) cells	44
3.3	RNA integrity assessed by formaldehyde (denaturing) agarose gel electrophoresis	46
3.4	Volcano plot illustrating significance versus fold change of gene expression, comparing SRJ09-resistant HCT-116 against parental HCT-116 cells	49
3.5	Volcano plot illustrating significance versus fold change of gene expression, comparing SRJ23-resistant PC-3 against parental PC-3 cells	54

3.6	Hierarchical clustering analysis of differential gene expression in two groups of samples (parental HCT-116 and SRJ09-resistant HCT-116 cell lines)	58
3.7	Hierarchical clustering analysis of differential gene expression in two groups of samples (parental PC-3 and SRJ23-resistant PC-3 cell lines)	59
3.8	Gene interaction analysis on 77 substantially modulated genes in SRJ09-resistant HCT-116 cells using STRING databases on the human whole genome	61
3.9	The number of genes classified under the different biological processes according to Gene Ontology (GO) annotations	64
3.10	Gene interaction analysis on 21 substantially modulated genes in SRJ23-resistant PC-3 cells using STRING databases on the human whole genome	64
3.11	The modulation of <i>ATG12</i> and <i>HMOX1</i> in parental and SRJ23-resistant PC-3 cell lines	65
3.12	A schematic diagram speculating the possible chemoresistance mechanisms of action induced by SRJ09 in the HCT-116 <sup>rst09</sup> cell line	69
3.13	A schematic diagram speculating the possible chemoresistance mechanisms of action induced by SRJ23 in the PC-3 <sup>rst23</sup> cell line	71
4.1	Chemical structures of eight derivatives of SRJ23	78
4.2	The co-crystallised ligand, 0QR, located in the binding site of mutant K-RasG12V (PDB ID: 4EPX)	80
4.3	Structure of the catalytic domain of K-Ras	81
4.4	Superimposition of re-docked (red carbon atoms) and co-crystallised (orange carbon atoms) ligand, 0QR (RMSD: < 2 Å) in mutant K-RasG12V (PDB ID: 4EPX)	92
4.5	Graphical representation of the highest binding affinity of each studied compound in complex with wild-type K-Ras and three K-Ras oncoproteins with mutations G12V, G12C, and G12D	93
4.6	The 2D (right panel) and 3D (left panel) interaction poses of the best conformer of each studied compound in complex with K-RasG12V (PDB ID: 4EPX) in p2 binding site	96

4.7	A close-up of molecular surface illustration showing binding of SRJ23 in the p2 binding site of K-RasG12V (PDB ID: 4EPX)	100
4.8	The water solvent model with SRJ23-K-RasG12V (PDB ID: 4EPX) complex prepared for MD simulation	101
4.9	A snapshot of the MD trajectory of the simulated molecular system of SRJ23-K-RasG12V (PDB ID: 4EPX) complex	102
4.10	The bonded and non-bonded interactions of SRJ23 in complex with K-RasG12V (PDB ID: 4EPX)	102
4.11	Timeline representation of the interactions and contacts (H-bonds, hydrophobic, ionic, water bridges) in the SRJ23-K-RasG12V (PDB ID: 4EPX) complex	103
4.12	RMSD plot obtained for the SRJ23-K-RasG12V (PDB ID: 4EPX) complex during MD simulation	104
4.13	RMSF plot obtained for K-RasG12V (PDB ID: 4EPX) during MD simulation	105
4.14	RMSF plot obtained for the SRJ23-K-RasG12V (PDB ID: 4EPX) complex during MD simulation	106
4.15	A close-up showing protein surfaces of K-RasG12V (PDB ID: 4EPX), with switches and binding pocket of p2	110
5.1	Representative images of immunoblotting of MAPK signalling protein levels in total protein lysate of PANC-1	128
5.2	Semi-quantitative analysis of three independent experiments of phospho- and total (A) C-Raf and (B) Erk1/2 proteins involved in MAPK cascade	129
5.3	Representative images of immunoblotting of PI3K signalling protein levels in total protein lysate of Capan-2	131
5.4	Semi-quantitative analysis of three independent experiments of phospho- and total proteins involved in PI3K cascade	132
5.5	Effect of SRJ09 and SRS07 on the formation of ROS in the DCFDA staining assay, normalised to their effect on percent cell viability of Capan-2 in MTT assay	134
5.6	A schematic diagram illustrating the possible mechanisms involved in the anticancer activity of AGP derivatives (SRJ09, SRJ23, SRS07, and SRS157) on PDAC cells	142

6.1	Representative HPLC chromatograms of (A) mobile phase spiked with DDAG and SRJ23 (50 µM), (B) pooled blank mouse plasma, (C) pooled plasma spiked with DDAG (25 µM) only, and (D) pooled plasma spiked with DDAG and SRJ23 (50 µM)	154
6.2	Calibration curves for SRJ23 standards	155
6.3	Mean plasma concentration-time profile of SRJ23 after administration of a single intraperitoneal bolus dose of 100 mg/kg of SRJ23 to female BALB/c mice	161
6.4	One-compartment model	161
6.5	The (A) predicted and (B) log-transformed plasma concentration-time profiles of SRJ23 after administration of a single intraperitoneal bolus dose of 100 mg/kg of SRJ23 to female BALB/c mice, as computed by PKSolver	162
7.1	Percentage normalised body weight changes of PANC-1 tumour-bearing nude mice relative to day 0	179
7.2	Relative tumour volume (RTV) of PANC-1 tumour-bearing nude mice compared to day 0	181
7.3	Representative images of sacrificed nude mice bearing PANC-1 tumours at the end of the experiment (day 14)	183
7.4	Representative photomicrographs of H&E-stained liver sections (10× magnification) of mice treated intraperitoneally with single dose of (A) vehicle control, (B) 200 mg/kg gemcitabine (C) 100 mg/kg SRJ23, (D) 200 mg/kg SRJ23, (E) 100 mg/kg SRS157, and (F) 200 mg/kg SRS157	187
7.5	Representative photomicrographs of H&E-stained kidney sections (10× magnification) of mice treated intraperitoneally with single dose of (A) vehicle control, (B) 200 mg/kg gemcitabine (C) 100 mg/kg SRJ23, (D) 200 mg/kg SRJ23, (E) 100 mg/kg SRS157, and (F) 200 mg/kg SRS157	188
7.6	Representative photomicrographs of H&E-stained spleen sections (10× magnification) of mice treated intraperitoneally with single dose of (A) vehicle control, (B) 200 mg/kg gemcitabine (C) 100 mg/kg SRJ23, (D) 200 mg/kg SRJ23, (E) 100 mg/kg SRS157, and (F) 200 mg/kg SRS157	189
7.7	Representative photomicrographs of H&E-stained tumour sections (10× magnification) of mice treated intraperitoneally with single dose of (A) vehicle control, (B) 200 mg/kg gemcitabine (C) 100 mg/kg SRJ23, (D) 200 mg/kg SRJ23,	191

(E) 100 mg/kg SRS157, and (F) 200 mg/kg SRS157

- 8.1 A schematic diagram summarising the important findings of the present study 200



## LIST OF APPENDICES

Appendix		Page
3.1	Plate layout and labelling of samples with different treatments	247
3.2	Recipe for formaldehyde agarose gel	249
3.3	An overview of the preparation of biotin-labelled anti-sense RNA (aRNA) and microarray experiment	250
3.4	Primers used in conventional RT-PCR analysis	251
3.5i	Band intensity ratio of 28S:18S rRNA subunits of total RNA samples of HCT-116 and HCT-116 <sup>rst109</sup> assessed by formaldehyde (denaturing) agarose gel electrophoresis	252
3.5ii	Band intensity ratio of 28S:18S rRNA subunits of total RNA samples of PC-3 and PC-3 <sup>rst23</sup> assessed by formaldehyde (denaturing) agarose gel electrophoresis	253
3.6	Functional enrichment analysis <i>via</i> GO annotation on the significantly differentially expressed genes in SRJ09-resistant HCT-116 cells	254
4.1	Superimposition of re-docked (yellow carbon atoms) and co-crystallised (pink carbon atoms) ligand, OQR in mutant K-RasG12V (PDB ID: 4EPX)	255
5.1	Recipe for different types of buffers	256
5.2	Recipe for SDS-PAGE gel	257
5.3	Statistical analysis of growth inhibitory effects of various compounds on four PDAC cell lines based on GI <sub>50</sub> values	258
5.4	Statistical analysis of growth inhibitory effects of various compounds on four PDAC cell lines based on TGI values	263
5.5	Statistical analysis of growth inhibitory effects of various compounds on four PDAC cell lines based on LC <sub>50</sub> values	268
5.6	Immunoblotting images of MAPK signalling protein levels in total protein lysate of PANC-1	274
5.7	Statistical analysis of effects of SRJ23 & SRS157 on	275



	activation and expression of MAPK signalling proteins	
5.8	Immunoblotting images of PI3K signalling protein levels in total protein lysate of Capan-2	283
5.9	Statistical analysis of effects of SRJ09 & SRS07 on activation and expression of PI3K signalling proteins	285
5.10	Statistical analysis of effects of SRJ09 & SRS07 on ROS production	292
6.1	IACUC approval letters	296
6.2	Cursory flow chart detailing experiments needed to prepare for HPLC method validation of SRJ23 determination	298
6.3	Mean peak heights of DDAG and SRJ23	299
6.4	HPLC chromatograms of mice plasma samples collected at different time points after administration with 100 mg/kg SRJ23	300
7.1	Recipe for 10% neutral buffered formalin	302
7.2	Average body weights of nude mice in each group over a fortnight	303
7.3	Statistical analysis of effects of SRJ23 & SRS157 on body weight changes of PANC-1 tumour-bearing nude mice	304
7.4	Average tumour volumes of nude mice in each group over a fortnight	314
7.5	Statistical analysis of effects of SRJ23 & SRS157 on relative tumour volumes of PANC-1 tumour-bearing nude mice	315
7.6	The equations of linear regression in mean RTV-time profile of each animal group used to calculate RTV2	325
7.7	The mean values of $\Delta C$ , $\Delta T$ , and terminal tumour weights used to calculate the parameters of antitumour activity of different treatments	326
7.8i	Photomicrographs of H&E-stained liver sections (10 $\times$ magnification) of mice treated with vehicle control, positive control, and two doses of SRJ23 and SRS157 (100 mg/kg and 200 mg/kg)	327

7.8ii	Photomicrographs of H&E-stained kidney sections (10× magnification) of mice treated with vehicle control, positive control, and two doses of SRJ23 and SRS157 (100 mg/kg and 200 mg/kg)	328
7.8iii	Photomicrographs of H&E-stained spleen sections (10× magnification) of mice treated with vehicle control, positive control, and two doses of SRJ23 and SRS157 (100 mg/kg and 200 mg/kg)	329
7.8iv	Photomicrographs of H&E-stained tumour sections (10× magnification) of mice treated with vehicle control, positive control, and two doses of SRJ23 and SRS157 (100 mg/kg and 200 mg/kg)	330



© COPYRIGHT

## LIST OF ABBREVIATIONS

0QR	N-(6-aminopyridin-2-yl)-4-fluorobenzenesulfonamide
ACN	Acetonitrile
ADME	Absorption, distribution, metabolism, and excretion
AGP	Andrographolide
ANOVA	Analysis of variance
APS	Ammonium persulphate
APT1	Acyl protein thioesterase 1
aRNA	Anti-sense RNA
ATG12	Autophagy-related gene 12
ATCC	American Type Culture Collection
BCA	Bicinchoninic acid
Bcl-2	B-cell lymphoma 2
Bcl-xL	Bcl-2-extra large
BH3	Bcl-2 homology 3
BSA	Bovine serum albumin
BWC	Body weight change
C $\alpha$	Alpha carbon
CADD	Computer-aided drug design
Cdc25	Cell division cycle 25
cDNA	Complementary DNA
CGM	Complete growth medium
CO	Carbon monoxide
CO <sub>2</sub>	Carbon dioxide
COSMIC	The Catalog of Somatic Mutations in Cancer

CPU	Central processing unit
CRD	Cysteine-rich domain
cRNA	Complementary RNA
CSC	Cancer stem cell
CV	Coefficients of variation
CYP	Cytochrome P450s
DCFDA	2',7'-dichlorofluorescein diacetate
DDAG	14-deoxy-11,12-didehydroandrographolide
DEGs	Differentially expressed genes
DEPC	Diethylpyrocarbonate
DMEM	Dulbecco's Modified Eagle Medium
DMSO	Dimethyl sulfoxide
EDTA	Ethylenediaminetetraacetic acid
EGF	Epidermal growth factor
EGFR	EGF receptor
EMT	Epithelial-mesenchymal transition
ERBB4	Erb-B2 receptor tyrosine kinase 4
Erk	Extracellular signal-regulated kinase
EtBr	Ethidium bromide
FBS	Foetal bovine serum
FDR	False discovery rate
FGF19	Fibroblast growth factor 19
FGFR4	Fibroblast growth factor receptor 4
GLOBOCAN	Global Cancer Statistics
GAP	GTPase-activating protein
GAPDH	Glyceraldehyde 3-phosphate dehydrogenase

GDP	Guanosine diphosphate
GEF	Guanine nucleotide exchange factor
GI <sub>50</sub>	Drug concentration that inhibits cell growth by 50%
Glide	Grid-based Ligand Docking with Energetics
GO	Gene Ontology
Grb2	Growth factor receptor-bound protein 2
GST	Glutathione S-transferase
GTP	Guanosine triphosphate
GTPase	Guanosine triphosphatase
H-bond	Hydrogen-bond
H&E	Haematoxylin-eosin
HKP	Housekeeping proteins
HMOX1	Haem oxygenase 1
HPLC	High-performance liquid chromatography
HRP	Horseradish peroxidase
HVR	Hypervariable region
IC <sub>50</sub>	Inhibitory concentration of a drug that reduces cell population to half of the control value
IC <sub>80</sub>	Drug concentration that reduces cell population by 80%, relative to vehicle control
ICH	The International Council for Harmonisation of Technical Requirements for Pharmaceuticals for Human Use
ICMT	Isoprenylcysteine carboxymethyl transferase
IgG	Immunoglobulin
i.p.	Intraperitoneal
IVC	Individually ventilated cage

INHAND	International Harmonization of Nomenclature and Diagnostic Criteria for Lesions in Rats and Mice
KITLG	KIT ligand
LBDD	Ligand-based drug discovery
LC <sub>50</sub>	Drug concentration that kills cells by 50%
LC3	Microtubule-associated protein 1A/1B-light chain 3
LOD	Limit of detection
LOQ	Limit of quantitation
MAPK	Mitogen-activated protein kinase
Mcl-1	Myeloid cell leukemia-1
MD	Molecular dynamics
ME	Matrix effect
Mek	MAPK/ERK kinase
MM-GBSA	Molecular mechanics-generalised Born surface area
MOMP	Mitochondrial outer membrane permeabilisation
MOPS	3-(N-morpholino)propane sulfonic acid
mRNA	Messenger RNA
mTOR	Mechanistic target of rapamycin
mTORC	Mechanistic target of rapamycin complex
MTT	3-[4, 5-dimethylthiazol-2-yl]-2, 5-diphenyltetrazolium bromide
NCI	National Cancer Institute
NCI-60	NCI's panel of 60 cancer cell lines
NF-κB	Nuclear factor kappa B
Nrf-2	Nuclear factor erythroid-2-related factor 2
OECD	The Organisation for Economic Co-operation and Development
OH	Hydroxyl

OPLS	Optimised Potentials for Liquid Simulations
p-loop	Phosphate-binding loop
p70S6K	p70 ribosomal protein S6 kinase
PAGE	Polyacrylamide Gel Electrophoresis
PanIN	Pancreatic intraepithelial neoplasia
PAT	Protein acyltransferase
PBS	Phosphate-buffered saline
PCR	Polymerase chain reaction
PDAC	Pancreatic ductal adenocarcinoma
PDB	Protein Data Bank
PDE6 $\delta$	Phosphodiesterase 6 delta
PDK1	3-phosphoinositide-dependent protein kinase 1
PE	Process efficiency
PH	Pleckstrin homology
PI3K	Phosphoinositide 3-kinase
PIK3CA	Phosphatidylinositol-4,5-bisphosphate 3-kinase catalytic subunit alpha
PIP2	Phosphatidylinositol-(4,5)-bisphosphate
PIP3	Phosphatidylinositol-(3,4,5)-trisphosphate
PK	Pharmacokinetics
PKB	Protein kinase B
PPI	Protein-protein interaction
PTEN	Phosphatase and tensin homolog
R <sup>2</sup>	Correlation coefficient
RAD	Ras-association domain
Ras	Rat sarcoma

RasGRF	Ras guanylnucleotide releasing factors
RBD	Ras binding domain
Rce-1	Ras-converting enzyme type 1
RCSB	Research Collaboratory for Structural Bioinformatics
RE	Recovery
Rheb	Ras homolog enriched in brain
RT	Retention time
RIN	RNA integrity number
RMSD	Root mean square deviation
RMSF	Root mean square fluctuation
RNase	Ribonucleases
ROS	Reactive oxygen species
RPMI	Roswell Park Memorial Institute
rRNA	ribosomal RNA
RTK	Receptor tyrosine kinase
RTV	Relative tumour volume
SAR	Structure-activity relationship
SBDD	Structure-based drug discovery
SD	Standard deviation
SDS	Sodium dodecyl sulfate
SH2	Src homology 2
SHC	Src homology 2 domain-containing
SID	Simulation Interaction Diagram
SOS	Son of Sevenless
SPRY2	Sprouty homolog 2



SPSS	Statistical Package for Social Sciences
SRJ09	3,19-(2-bromobenzylidene)andrographolide
SRJ23	3,19-(3-Chloro-4-fluorobenzylidene)-andrographolide
STD-NMR	Saturation transfer difference nuclear magnetic resonance
STRING	Search Tool for the Retrieval of Interacting Genes/ Proteins
T/C	Optimal treatment-to-control ratio
TAE	Tris-acetate-ethylenediaminetetraacetic acid
T <sub>d</sub>	Tumour growth delay
TGI	Drug concentration that totally inhibits cell growth
TSC	Tuberous sclerosis complex
TuGI	Tumour growth inhibition
vdW	van der Waals

# CHAPTER 1

## INTRODUCTION

### 1.1 Overview of the Study

Cancer is one of the most common chronic diseases, affecting both men and women globally, with increasing prevalence over the past three decades (Tu et al., 2017). In 2018, more than 18 million new cancer cases have been diagnosed and the number has been increasing annually (Bray et al., 2018). The major risk factors of cancer are often associated with aging and growth of the world population (Weir, Thompson, Soman, Møller, & Leadbetter, 2015). Cancer has been recognised as the second leading cause of death worldwide (Bray et al., 2018). Considering population growth and increased urbanisation, the number of cancer patients is expected to rise to 29.5 million by the year 2040, with an estimated 73% increase in the number of cancer-related deaths worldwide (NCI, 2020).

Fundamentally, human cancers involve a transformation of normal cells into tumorous cells at both the genetic and molecular levels of the organism (Bukhtoyarov & Samarin, 2015). One of the initial phases of cancer cell transformation is the activation and/or expression of oncogenes, which drive the tumour initiation and cancer progression (Vicente-Dueñas, Romero-Camarero, Cobaleda, & Sánchez-García, 2013). *RAS* oncogenes, which possess a central role in regulating cell growth and survival, are among the first to be discovered and often found to harbour point mutations in a wide spectrum of human cancers. Three functional *RAS* oncogenes (*HRAS*, *NRAS*, and *KRAS*) have been identified since the 1980s, and they encode highly similar proteins (H-Ras, N-Ras, K-Ras) with inherent guanosine triphosphatase (GTPase) activity (Wennerberg, Rossman, & Der, 2005). It is estimated now that over 30% of all human tumours have activating mutations in one of the *RAS* genes (Kodaz et al., 2017). Mutations on *KRAS* constitute more than 80% of all *RAS* mutations (Arrington et al., 2012; Hobbs, Der, & Rossman, 2016). Analysis of a significant number of human tumours has demonstrated a variation in mutant *KRAS* incidence among different tumour types examined (Bos, 1989). In particular, the highest incidence of mutated *KRAS* genes is found in tumours from the exocrine pancreas (> 90%), typically pancreatic ductal adenocarcinoma (PDAC), followed by colorectal cancers (30% – 50%) and lung cancers (> 30%).

Being a key node in cellular signalling responsible for growth and survival, oncogenic K-Ras activates several downstream effectors, including canonical mitogen-activated protein kinase (MAPK) and phosphoinositide

3-kinases (PI3K) (Eser, Schnieke, Schneider, & Saur, 2014), resulting in constitutive activation of these effector signalling pathways. Tumours with activating *KRAS* mutations were found to be resistant to standard therapies (Lu, Jang, Gu, Zhang, & Nussinov, 2016). To date, the existing cancer drugs available in the market are not catered to cancer patients with mutations in *KRAS*, such as PDAC, which is an aggressive disease characterised by poor prognosis and therapeutic resistance. Therefore, the search for better treatment, using K-Ras oncoprotein as the central anticancer therapeutic target, is especially urgent.

The bioactive compound from the well-known herb *Andrographis paniculata*, andrographolide (AGP), has been studied for its anticancer properties for nearly two decades by the Stanslas' group (Stanslas et al., 2001; Jada et al., 2006; Jada et al., 2007). Structural modifications of AGP led to the generation of benzylidene derivatives, namely 3,19-(2-bromobenzylidene)andrographolide (SRJ09) and 3,19-(3-chloro-4-fluorobenzylidene)-andrographolide (SRJ23), with improved potency and selectivity of anticancer activity (Jada et al., 2008). Further modifications on SRJ09 and SRJ23 yielded SRS07 and eight derivatives (SRS150 – SRS157), respectively. Using *in silico* molecular simulation study of protein-ligand interactions, a previous study revealed that these benzylidene derivatives were capable of binding oncogenic K-Ras directly (Hocker et al., 2013). With this direct binding, these compounds demonstrated remarkable inhibition on the activation of K-Ras and its downstream signalling molecules of the MAPK pathway, thereby jeopardising the viability of the cancer cells harbouring either wild-type or oncogenic K-Ras.

The present investigation is conceptualised based on the possible attenuation of the aberrant mutant K-Ras-mediated signalling pathways by benzylidene derivatives of AGP *via* binding to a druggable allosteric site in the protein. The study further explored the preclinical antitumour potential of these AGP derivatives in PDAC tumour xenograft nude mice model.

## 1.2 Hypothesis

The semisynthetic AGP derivatives bind oncogenic K-Ras proteins, abrogating their functions and exerting anticancer activity through their inhibitory effects on K-Ras-mediated signalling pathways, such as MAPK and PI3K cascades, as well as reduce the tumour growth in preclinical xenograft models.

## 1.3 Objectives

### 1.3.1 General Objective

The main objective of this study was to investigate the potential of AGP derivatives as anti-Ras therapeutics through *in silico*, *in vitro*, and *in vivo* approaches.

### 1.3.2 Specific Objectives

The specific objectives as follows:

- i. To decipher the mechanisms of action and mechanisms of acquired resistance of SRJ09 and SRJ23 in colorectal and prostate cancer cells.
- ii. To identify the binding modes and molecular interactions of the AGP derivatives with K-Ras proteins *via in silico* approach.
- iii. To determine the *in vitro* growth inhibitory effects of AGP derivatives on a panel of PDAC cell lines harbouring either wild-type or oncogenic K-Ras, as well as their effects on K-Ras-mediated MAPK and PI3K pathways.
- iv. To determine the pharmacokinetics of SRJ23 in mice.
- v. To evaluate the preclinical antitumour activity of SRJ23 and SRS157 against PDAC tumour xenografts in nude mice.

## REFERENCES

- Abankwa, D., Gorfe, A. A., Inder, K., & Hancock, J. F. (2010). Ras membrane orientation and nanodomain localization generate isoform diversity. *Proceedings of the National Academy of Sciences of the United States of America*, *107*, 130-1135.
- Adariani, S. R., Buchholzer, M., Akbarzadeh, M., Nakhaei-Rad, S., Dvorsky, R., & Ahmadian, M. R. (2018). Structural snapshots of RAF kinase interactions. *Biochemical Society Transactions*, *46*, 1393-1406.
- Adis Data Information. (2006). Retrieved August 29, 2019, from [https://static.springer.com/sqw/documents/1372030/application/pdf/40262\\_cpk\\_symbols.pdf](https://static.springer.com/sqw/documents/1372030/application/pdf/40262_cpk_symbols.pdf)
- Adjei, A. A. (2001). Blocking oncogenic Ras signaling for cancer therapy. *Journal of the National Cancer Institute*, *93*, 1062-1074.
- Ahearn, I. M., Haigis, K., Bar-Sagi, D., & Phillips, M. R. (2012). Regulating the regulator: post-translational modification of RAS. *Nature Reviews Molecular Cell Biology*, *13*, 39-51.
- Ahmadian, M. R., Stege, P., Scheffzek, K., & Wittinghofer, A. (1997). Confirmation of the arginine-finger hypothesis for the GAP-stimulated GTP-hydrolysis reaction of Ras. *Nature Structural & Molecular Biology*, *4*, 686-689.
- Akhdar, H., Legendre, C., Aninat, C., & Morel, F. (2012). Anticancer drug metabolism: Chemotherapy resistance and new therapeutic approaches. In J. Paxton (Ed.), *Topics on Drug Metabolism* (pp. 137-170). London: IntechOpen.
- Alonso, H., Bliznyuk, A. A., & Gready, J. E. (2006). Combining docking and molecular dynamic simulations in drug design. *Medicinal Research Reviews*, *26*, 531-568.
- AlQahtani, A. D., O'Connor, D., Domling, A., & Goda, S. K. (2019). Strategies for the production of long-acting therapeutics and efficient drug delivery for cancer treatment. *Biomedicine & Pharmacotherapy*, *113*, 108750.
- Aminpour, M., Montemagno, C., & Tuszynski, J. A. (2019). An overview of molecular modeling for drug discovery with specific illustrative examples of applications. *Molecules*, *24*, 1693.
- Amrutkar, M., & Gladhaug, I. P. (2017). Pancreatic cancer chemoresistance to gemcitabine. *Cancers*, *9*, 157.

- Anderson, M. E. (1981). Saturable metabolism and its relationship to toxicity. *Critical Reviews in Toxicology*, 9, 105-150.
- Andreyev, H. J., Norman, A. R., Cunningham, D., Oates, J. R., & Clarke, P. A. (1998). Kirsten ras mutations in patients with colorectal cancer: the multicenter "RASCAL" study. *Journal of the National Cancer Institute*, 90, 675-684.
- Apolloni, A., Prieo, I. A., Lindsay, M., Parton, R. G., & Hancock, J. F. (2000). H-ras but not K-ras traffics to the plasma membrane through the exocytic pathway. *Molecular and Cellular Biology*, 20, 2475-2487.
- Ariffin, A. B., Forde, P. F., Jahangeer, S., Soden, D. M., & Hinchion, J. (2014). Releasing pressure in tumors: What do we know so far and where do we go from here? A review. *Cancer Research*, 74, 8 pages.
- Arrington, A. K., Heinrich, E. L., Lee, W., Duldulao, M., Patel, S., Sanchez, J., . . . Kim, J. (2012). Prognostic and predictive roles of KRAS mutation in colorectal cancer. *International Journal of Molecular Sciences*, 13, 12153-12168.
- Aslantürk, O. S. (2017). *In Vitro* Cytotoxicity and Cell Viability Assays: Principles, Advantages, and Disadvantages. In M. L. Larramendy and S. Soloneski (Ed.), *Genotoxicity - A Predictable Risk to Our Actual World* (pp. 1-17). London: IntechOpen.
- Aston, W. J., Hope, D. E., Nowak, A. K., Robinson, B. W., Lake, R. A., & Lesterhuis, W. J. (2017). A systematic investigation of the maximum tolerated dose of cytotoxic chemotherapy with and without supportive care in mice. *BMC Cancer*, 17, 684.
- Azeloglu, E. U., & Iyengar, R. (2015). Signaling networks: Information flow, computation, and decision making. *Cold Spring Harbor Perspectives in Biology*, 7, a005934.
- Baba, A. I., & Cătoi, C. (2007). Chapter 18 CANCER DIAGNOSIS. In *Comparative Oncology*. Bucharest (RO): The Publishing House of the Romanian Academy. Retrieved from <https://www.ncbi.nlm.nih.gov/books/NBK9550/>
- Baillie, T. A., & Rettie, A. E. (2011). Role of biotransformation in drug-induced toxicity: Influence of intra- and inter-species differences in drug metabolism. *Drug Metabolism and Pharmacokinetics*, 26, 15-29.
- Bao, L., Jaramillo, M. C., Zhang, Z., Zheng, Y., Yao, M., Zhang, D. D., & Yi, X. (2014). Nrf2 induces cisplatin resistance through activation of

autophagy in ovarian carcinoma. *International Journal of Clinical and Experimental Pathology*, 7, 1502-1513.

- Battista, R. A., Resnati, M., Facchi, C., Ruggieri, E., Cremasco, F., Paradiso, F., . . . Milan, E. (2018). Autophagy mediates epithelial cancer chemoresistance by reducing p62/SQSTM1 accumulation. *PLoS ONE*, 13, e0201621.
- Bauer, L. A. (2008). *Applied Clinical Pharmacokinetics* (Second ed.). New York, (NY): The McGraw-Hill Companies, Inc.
- Becker, A. E., Hernandez, Y. G., Frucht, H., & Lucas, A. L. (2014). Pancreatic ductal adenocarcinoma: Risk factors, screening, and early detection. *World Journal of Gastroenterology*, 20, 11182-11198.
- Belounis, A., Nyalendo, C., Gall, R. L., Imbriglio, T. V., Mahma, M., Teira, P., . . . Sartelet, H. (2016). Autophagy is associated with chemoresistance in neuroblastoma. *BMC Cancer*, 16, 891.
- Benerjee, M., Chattopadhyay, S., Choudhuri, T., Bera, R., Kumar, S., Chakraborty, B., & Mukherjee, S. K. (2016). Cytotoxicity and cell cycle arrest induced by andrographolide lead to programmed cell death of MDA-MB-231 breast cancer cell line. *Journal of Biomedical Science*, 23, 16.
- Bennett, B. D., Kimball, E. H., Gao, M., Osterhout, R., Van Dien, S. J., & Rabinowitz, J. D. (2009). Absolute metabolite concentrations and implied enzyme active site occupancy in *Escherichia coli*. *Nature Chemical Biology*, 5, 593-599.
- Berberat, P. O., Dambrauskas, Z., Gulbinas, A., Giese, T., Giese, N., Künzli, B., . . . Friess, H. (2005). Inhibition of heme oxygenase-1 increases responsiveness of pancreatic cancer cells to anticancer treatment. *Clinical Cancer Research*, 11, 3790-3798.
- Berezhkovskiy, L. M. (2015). On the accuracy of calculation of the mean residence time of drug in the body and its volumes of distribution based on the assumption of central elimination. *Xenobiotica*, 1-6.
- Bergo, M. O., Ambroziak, P., Gregory, C., George, A., Otto, J. C., Kim, E., . . . Young, S. G. (2002). Absence of the CAAX endoprotease Rce1: Effects on cell growth and transformation. *Molecular and Cellular Biology*, 22, 171-181.
- Bergo, M. O., Leung, G. K., Ambroziak, P., Otto, J. C., Casey, P. J., & Young, S. G. (2000). Targeted inactivation of the isoprenylcysteine carboxyl methyltransferase gene causes mislocalization of K-Ras

- in mammalian cells. *Journal of Biological Chemistry*, 23, 17605-17610.
- Berman, H. M., Westbrook, J., Feng, Z., Gilliland, G., Bhat, T. N., Weissig, H., . . . Bourne, P. E. (2000). The Protein Data Bank. *Nucleic Acids Research*, 28, 235-242.
- Berndt, N., Hamilton, A. D., & Sebti, S. M. (2011). Targeting protein prenylation for cancer therapy. *Nature Reviews Cancer*, 11, 775-791.
- Bezzera, D. P., de Castro, F. O., Alves, A. P., Pessoa, C., de Moraes, M. O., Silveira, E. R., . . . Costa-Lotufo, L. V. (2007). *In vitro* and *in vivo* antitumor effect of 5-FU combined with piplartine and piperine. *Journal of Applied Toxicology*, 28, 156-163.
- Binenbaum, Y., Na'ara, S., & Gil, Z. (2015). Gemcitabine resistance in pancreatic ductal adenocarcinoma. *Drug Resistance Updates*, 23, 55-68.
- Bister, K. (2015). Discovery of oncogenes: The advent of molecular cancer research. *Proceedings of the National Academy of Sciences of the United States of America*, 112, 15259-15260.
- Boon, V., Glass, B., & Nimmo, A. (2006). High-performance liquid chromatographic assay of indomethacin in porcine plasma with applicability to human levels. *Journal of Chromatographic Science*, 44, 41-44.
- Boriack-Sjodin, P. A., Margarit, S. M., Bar-Sagi, D., & Kuriyan, J. (1998). The structural basis of the activation of Ras by SOS. *Nature*, 394, 337-343.
- Boroujerdi, M. (2001). *Pharmacokinetics: Principles and applications*. New York(NY): McGraw-Hill.
- Bos, J. L. (1989). *ras* oncogenes in human cancer: A review. *Cancer Research*, 49, 4682-4689.
- Bos, J. L., Rehmann, H., & Wittinghofer, A. (2007). GEFs and GAPs: Critical elements in the control of small G proteins. *Cell*, 129, 865-877.
- Bosma, G. C., Custer, R. P., & Bosma, M. J. (1983). A severe combined immunodeficiency mutation in the mouse. *Nature*, 301, 527-530.
- Bosma, M. J., & Carroll, A. M. (1991). The SCID mouse mutant: definition, characterization, and potential uses. *Annual Review of Immunology*, 9, 323-350.



- Bourne, H. R., Sanders, D. A., & McCormick, F. (1991). The GTPase superfamily: conserved structure and molecular mechanism. *Nature*, *349*, 117-127.
- Bournet, B., Muscari, F., Buscail, C., Assenat, E., Barthet, M., Hammel, P., . . . Buscail, L. (2016). KRAS G12D mutation subtype is a prognostic factor for advanced pancreatic adenocarcinoma. *Clinical and Translational Gastroenterology*, *7*, e157.
- Boyartchuk, V. L., Ashby, M. N., & Rine, J. (1997). Modulation of Ras and a-factor function by carboxyl terminal proteolysis. *Science*, *275*, 1796-1800.
- Brake, K., Gumireddy, A., Tiwari, A., Chauhan, H., & Kumari, D. (2017). *In vivo* studies for drug development *via* oral delivery: Challenges, animal models and techniques. *Pharmaceutica Analytica Acta*, *8*, 9.
- Bray, F., Ferlay, J., Soerjomataram, I., Siegel, R. L., Torre, L. A., & Jemal, A. (2018). Global Cancer Statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA: A Cancer Journal for Clinicians*, *68*, 394-424.
- Brooks, E. A., Galarza, S., Gencoglu, M. F., Cornelison, R. C., Munson, J. M. & Peyton, S. R. (2019). Applicability of drug response metrics for cancer studies using biomaterials. *Philosophical Transactions of the Royal Society B*, *374*, 20180226.
- Bruno, V., Grazia, A., Mariella, S. C., Giovanna, C., Leakos, M., Daniele, S., . . . Giuseppe, T. (2016). Drug-induced hepatotoxicity in cancer patients - implication for treatment. *Expert Opinion on Drug Safety*, *15*, 1219-1238.
- Buday, L., & Downward, J. (2008). Many faces of Ras activation. *Biochimica et Biophysica Acta*, *1786*, 178-187.
- Buhrman, G., O'Connor, C., Zerbe, B., Kearney, B. M., Nepoleon, R., Kovrigina, E. A., . . . Mattos, C. (2011). Analysis of binding site hot spots on the surface of Ras GTPase. *Journal of Molecular Biology*, *413*, 773-789.
- Buhrman, G., Wink, G., & Mattos, C. (2007). Transformation efficiency of RasQ61 mutants linked to structural features of the switch regions in the presence of Raf. *Structure*, *15*, 1618-1629.
- Bukhtoyarov, O. V., & Samarin, D. M. (2015). Pathogenesis of cancer: Cancer reparative trap. *Journal of Cancer Therapy*, *6*, 399-412.

- Canon, J., Rex, K., Saiki, A. Y., Mohr, C., Cooke, K., Bagal, D., . . . Lipford, J. R. (2019). The clinical KRAS(G12C) inhibitor AMG 510 drives anti-tumour immunity. *Nature*, *575*, 217-223.
- Capurso, G., & Sette, C. (2019). Drug resistance in pancreatic cancer: New player caught in act. *EBioMedicine*, *40*, 39-40.
- Carlino, M. S., Todd, J. R., & Rizos, H. (2014). Resistance to c-Kit inhibitors in melanoma: insights for future therapies. *Oncoscience*, *1*, 423-426.
- Castellano, E., & Downward, J. (2011). RAS interaction with PI3K: More than just another effector pathway. *Genes & Cancer*, *2*, 261-274.
- Chao, H. P., Kup, C. D., Chiu, J. H., & Fu, S. L. (2010). Andrographolide exhibits anti-invasive activity against colon cancer cells via inhibition of MMP2 activity. *Planta Medica*, *76*, 1827-1833.
- Charan, R. D., Schlingmann, G., Janso, J., Bernan, V., Feng, X., & Carter, G. T. (2004). Diazepinomicin, a new antimicrobial alkaloid from a marine *Micromonospora* sp. *Journal of Natural Products*, *67*, 1431-1433.
- Chen, M., Huang, J., Yang, X., Liu, B., Zhang, W., Huang, L., . . . Ge, J. (2012). Serum starvation induced cell cycle synchronization facilitates human somatic cells reprogramming. *PLoS ONE*, *7*(4), e28203.
- Chen, P., Deng, Y., Bergqvist, S., Falk, M. D., Liu, W., Timofeevski, S., & Brooun, A. (2014). Engineering of an isolated p110 $\alpha$  subunit of PI3K $\alpha$  permits crystallization and provides a platform for structure-based drug design. *Protein Science*, *23*, 1332-1340.
- Chen, S. H., Zhang, Y., Van Horn, R. D., Yin, T., Buchanan, S., Yadav, V., . . . Peng, S. (2016). Oncogenic BRAF deletions that function as homodimers and are sensitive to inhibition by RAF dimer inhibitor LY3009120. *Cancer Discovery*, *6*, 300-315.
- Cherfils, J., & Zeghouf, M. (2013). Regulation of small GTPases by GEFs, GAPs, and GDIs. *Physiological Reviews*, *93*, 269-309.
- Cheymol, G. (1988). Drug pharmacokinetics in the obese. *Fundamental & Clinical Pharmacology*, *2*, 239-256.
- Chial, H. (2008). Proto-oncogenes to oncogenes to cancer. *Nature Education*, *1*, 33.
- Chiou, T. J., Zhang, J., Ferrans, V. J., & Tzeng, W. F. (1997). Cardiac and renal toxicity of menadione in rat. *Toxicology*, *124*, 193-202.

- Chong, H., Lee, J., & Guan, K. (2001). Positive and negative regulation of Raf kinase activity and function by phosphorylation. *EMBO Journal*, *20*, 3716-3727.
- Chow, S. (2014). Bioavailability and bioequivalence in drug development. *Wiley Interdisciplinary Reviews: Computational Statistics*, *6*, 304-312.
- Chung, T. D., Terry, D. B., & Smith, L. H. (2015). *In Vitro* and *In Vivo* Assessment of ADME and PK Properties During Lead Selection and Lead Optimization – Guidelines, Benchmarks and Rules of Thumb. In G. S. Sittampalam, A. Grossman, K. Brimacombe, M. Arkin, D. Auld, C. P. Austin, . . . X. Xu, *The Assay Guidance Manual*. Bethesda (MD): Eli Lilly & Company and the National Center for Advancing Translational Sciences. Retrieved from <https://www.ncbi.nlm.nih.gov/books/NBK326710/>
- Claing, A. (2013).  $\beta$ -Arrestins: The Molecular Biology of Arrestins GTPase Activation and Function. *Progress in Molecular Biology and Translational Science*, *118*, 149-174.
- Cole, S. P., Bhardwaj, G., Gerlach, J. H., Mackie, J. E., Grant, C. E., Almquist, K. C., . . . Deeley, R. G. (1992). Overexpression of a transporter gene in a multidrug-resistant human lung cancer cell line. *Science*, *258*, 1650-1654.
- Collins, M. A., & Pasca di Magliano, M. (2014). Kras as a key oncogene and therapeutic target in pancreatic cancer. *Frontiers in Physiology*, *4*, Article 407, 8 pages.
- Cooper, C. L., O'Toole, S. A., & Kench, J. G. (2013). Classification, morphology and molecular pathology of premalignant lesions of the pancreas. *Pathology*, *45*, 286-304.
- Cooper, G. M. (2000). *The Cell: A Molecular Approach* (2nd edition ed.). Sunderland (MA): Sinauer Associates.
- Cox, A. D., & Der, C. J. (2010). Ras history: The saga continues. *Small GTPases*, *1*, 2-27.
- Cox, A. D., Fesik, S. W., Kimmelman, A. C., Luo, J., & Der, C. J. (2014). Drugging the undruggable Ras: mission possible? *Nature Reviews Drug Discovery*, *13*, 828-851.
- Cree, I. A., & Charlton, P. (2017). Molecular chess? Hallmarks of anti-cancer drug resistance. *BMC Cancer*, *17*, 8 pages.

- Cui, L., Qiu, F., Wang, N. L., & Yao, X. (2005). A new glucuronidated metabolite of andrographolide in human. *Chinese Chemical Letters*, 16, 369-371.
- Cui, L., Chan, W., Qiu, F., Cai, Z., & Yao, X. (2008). Identification of four urea adducts of andrographolide in humans. *Drug Metabolism Letters*, 2, 261-268.
- da Silva, A. R., dos Santos, D. A., Paixão, M. W., & Corrêa, A. G. (2019). Stereoselective multicomponent reactions in the synthesis or transformations of epoxides and aziridines. *Molecules*, 24, 630.
- Dai, Y., Chen, S., Chai, L., Zhao, J., Wang, Y., & Wang, Y. (2019). Overview of pharmacological activities of *Andrographis paniculata* and its major compound andrographolide. *Critical Reviews in Food Science and Nutrition*, 59, S17-S29.
- Davendar, P., Nayak, V. L., Yadav, D. K., Kumae, A. N., Kumar, J. K., Srinivas, K., . . . Ramakrishna, S. (2015). Synthesis and evaluation of anticancer activity of novel andrographolide derivatives. *Medicinal Chemistry Communications*, 6, 898-904.
- Daview, N. M., Takemoto, J. K., Brocks, D. R., & Yáñez, J. A. (2010). Multiple peaking phenomena in pharmacokinetic disposition. *Clinical. Pharmacokinetics*, 49, 351-377.
- Deer, E. L., Gonzalez-Hernandez, J., Coursen, J. D., Shea, J. E., Ngatia, J., Scaife, C. L., . . . Mulvihill, S. J. (2010). Phenotype and genotype of pancreatic cancer cell lines. *Pancreas*, 39, 425-435.
- Degirmenci, U., Wang, M., & Hu, J. (2020). Targeting aberrant RAS/RAF/MEK/ERK signaling for cancer therapy. *Cells*, 9, 198.
- Dekker, F., Rocks, O., Vartak, N., Menninger, S., Hedberg, C., Balamurugan, R., . . . Waldmann, H. (2010). Small-molecule inhibition of APT1 affects Ras localization and signaling. *Nature Chemical Biology*, 6, 449-456.
- Der, C. J., Krontiris, T. G., & Cooper, G. M. (1982). Transforming genes of human bladder and lung carcinoma cell lines are homologous to the ras genes of Harvey and Kirsten sarcoma viruses. *Proceedings of the National Academy of Sciences of the United States of America*, 79, 3637-3640.
- Desnoyers, L. R., Pai, R., Ferrando, R. E., Hötzel, K., Le, T., Ross, J., . . . French, D. M. (2008). Targeting FGF19 inhibits tumor growth in colon cancer xenograft and FGF19 transgenic hepatocellular carcinoma models. *Oncogene*, 27, 85-97.

- Dhillon, A. S., Hagan, S., Rath, O., & Kolch, W. (2007). MAP kinase signalling pathways in cancer. *Oncogene*, *26*, 3279-3290.
- Dhillon, A. S., Meikle, S., Yazici, Z., Eulitz, M., & Kolch, W. (2002). Regulation of Raf-1 activation and signalling by dephosphorylation. *EMBO Journal*, *21*, 64-71.
- Dietel E., Brobeil, A., Gattenlöhner, S., & Wimmer, M. (2018). The importance of the right framework: Mitogen-activated protein kinase pathway and the scaffolding protein PTPIP51. *International Journal of Molecular Sciences*, *19*, 3282.
- Dolado, I., & Nebreda, A. R. (2008) AKT and oxidative stress team up to kill cancer cells. *Cancer Cell*, *14*, 427-429.
- Downward, J. (2003). Targeting RAS signalling pathways in cancer therapy. *Nature Reviews Cancer*, *3*, 11-22.
- Du, F., Wu, X., Liu, Y., Wang, T., Qi, X., Mao, Y., . . . Hua, D. (2013). Acquisition of paclitaxel resistance *via* PI3K-dependent epithelial-mesenchymal transition in A2780 human ovarian cancer cells. *Oncology Reports*, *30*, 1113-1118.
- Dunnington, K., Benrimoh, N., Branquist, C., Cardillo-Marricco, N., Di Spirito, M., & Grenier, J. (2018). Application of Pharmacokinetics in Early Drug Development. In N. Malangu (Ed.), *Pharmacokinetics and Adverse Effects of Drugs - Mechanisms and Risks Factors* (pp. 57-75). London: IntechOpen.
- Dutta, S., Going, J. J., Crumley, A. B., Mohammed, Z., Orange, C., Edwards, J., . . . McMillan, D. C. (2012). The relationship between tumour necrosis, tumour proliferation, local and systemic inflammation, microvessel density and survival in patients undergoing potentially curative resection of oesophageal adenocarcinoma. *British Journal of Cancer*, *106*, 702-710.
- Ebisuy, M., Kondoh, K., & Nishida, E. (2005). The duration, magnitude and compartmentalization of ERK MAP kinase activity: mechanisms for providing signaling specificity. *Journal of Cell Science*, *118*, 2997-3002.
- Ebner, M., Sinkovics, B., Szczygiel, M., Ribeiro, D. W., & Yudushkin, I. (2017). Localization of mTORC2 activity inside cells. *Journal of Cell Biology*, *216*, 343-353.
- El habbash, A. I., Hashim, N. M., Ibrahim, M. Y., Yahayu, M., Elmutaah Omer, F. A., Rahman, M. A., . . . Lian, G. E. (2017). *In vitro* assessment of anti-proliferative effect induced by  $\alpha$ -mangostin from *Cratoxylum arborescens* on HeLa cells. *PeerJ*, *5*, e3460.

- Elghazi, L., Weiss, A. J., Barker, D. J., Callaghan, J., Staloch, L., Sandgren, E. P., . . . Bernal-Mizrachi, E. (2009). Regulation of pancreas plasticity and malignant transformation by Akt signaling. *Gastroenterology*, *136*, 1091-1103.
- Elmore, S. A., Dixon, D., Hailey, J. R., Harada, T., Herbert, R. A., Maronpot, R. R., . . . Creasy, D. M. (2016). Recommendations from the INHAND Apoptosis/Necrosis Working Group. *Toxicologic Pathology*, *44*, 173-188.
- Endrenyi, L., Fritsch, S., & Yan, W. (1991). C<sub>max</sub>/AUC is a clearer measure than C<sub>max</sub> for absorption rates in investigations of bioequivalence. *International Journal of Clinical Pharmacology and Therapeutics*, *29*, 394-399.
- Eser, S., Reiff, N., Messer, M., Seidler, B., Gottschalk, K., Dobler, M., . . . Saur, D. (2013). Selective requirement of PI3K/PDK1 signaling for Kras oncogene-driven pancreatic cell plasticity and cancer. *Cancer Cell*, *23*, 406-420.
- Eser, S., Schnieke, A., Schneider, G., & Saur, D. (2014). Oncogenic KRAS signalling in pancreatic cancer. *British Journal of Cancer*, *111*, 817-822.
- Etti, I., Abdullah, R., Hashim, N. M., Kadir, A., Abdul, A. B., Etti, C., . . . How, C. W. (2016). Artonin E and structural analogs from *Artocarpus* species abrogates estrogen receptor signaling in breast cancer. *Molecules*, *21*, 839.
- European Medicines Agency. (2009). M3(R2) Nonclinical Safety Studies for the Conduct of Human Clinical Trials and Marketing Authorization for Pharmaceuticals (Step 5). Switzerland: International Conference on Harmonisation of Technical Requirements for Registration of Pharmaceuticals for Human Use.
- Fan, Z., Fan, K., Yang, C., Huang, Q., Gong, Y., Cheng, H., . . . Luo, G. (2018). Critical role of KRAS mutation in pancreatic ductal adenocarcinoma. *Translational Cancer Research*, *7*, 1728-1736.
- Feig, C., Gopinathan, A., Neesse, A., Chan, D. S., Cook, N., & Tuveson, D. A. (2012). The pancreas cancer microenvironment. *Clinical Cancer Research*, *18*, 4266-4276.
- Ferreira, L. G., dos Santos, R. N., Oliva, G., & Andricopulo, A. D. (2015). Molecular docking and structure-based drug design strategies. *Molecules*, *20*, 13384-13421.

- Ferro, R., & Falasca, M. (2014). Emerging role of the KRAS-PDK1 axis in pancreatic cancer. *World Journal of Gastroenterology*, *20*, 10752-10757.
- Fischer, A., Hekman, M., Kuhlmann, J., Rubio, I., Wiese, S., & Rapp, U. R. (2007). B- and C-RAF display essential differences in their binding to Ras. *Journal of Biological Chemistry*, *282*, 26503-26516.
- Flanagan, S. P. (1966). 'Nude', a new hairless gene with pleiotropic effects in the mouse. *Genetics Research*, *8*, 295-309.
- Folkman, J., & Kalluri, R. (2003) Beginning of angiogenesis research. In D. W. Kufe, R. E. Pollock, R. R. Weichselbaum, C. B. Robert, T. S. Gansler, J. F. Holland, & E. Frei (Eds.), *Holland-Frei Cancer Medicine* (6th edition ed.). Hamilton (ON): BC Decker.
- Forbes, S. A., Bindal, N., Bamford, S., Cole, C., Kok, C. Y., Beare, D., . . . Futreal, P. A. (2011). COSMIC: mining complete cancer genomes in the Catalogue of Somatic Mutations in Cancer. *Nucleic Acids Research*, *39*, D945-D950.
- Force, T., Bonventre, J. V., Heidecker, G., Rapp, U., Avruch, J., & Kyriakis, J. M. (1994). Enzymatic characteristics of the c-Raf-1 protein kinase. *Proceedings of the National Academy of Sciences of the United States of America*, *91*, 1270-1274.
- Friesner, R. A., Banks, J. L., Murphy, R. B., Halgren, T. A., Klicic, J. J., Mains, D. T., . . . Shenkin, P. S. (2004). Glide: A new approach for rapid, accurate docking and scoring. 1. Method and assessment of docking accuracy. *Journal of Medicinal Chemistry*, *47*, 1739-1749.
- Friesner, R. A., Murphy, R. B., Repasky, M. P., Frye, L. L., Greenwood, J. R., Halgren, T. A., . . . Mainz, D. T. (2006). Extra precision glide: Docking and scoring incorporating a model of hydrophobic enclosure for protein-ligand complexes. *Journal of Medicinal Chemistry*, *49*, 6177-6196.
- Gagliardi, P. A., di Blasio, L., Orso, F., Seano, G., Sessa, R., Taverna, D., . . . Primo, L. (2012). 3-phosphoinositide-dependent kinase 1 controls breast tumor growth in a kinase-dependent but Akt-independent manner. *Neoplasia*, *14*, 719-731.
- Gao, L., Wang, X., Taang, Y., Huang, S., Hu, C.A., Teng, Y., 2017. FGF19/FGFR4 signaling contributes to the resistance of hepatocellular carcinoma to sorafenib. *Journal of Experimental & Clinical Cancer Research*, *36*, 8.

- García-Gómez, R., Bustelo, X. R., & Crespo, P. (2018). Protein-protein interactions: Emerging oncotargets in the RAS-ERK pathway. *Trends in Cancer*, 4, 616-633.
- Genheden, S., & Ryde, U. (2015). The MM/PBSA and MM/GBSA methods to estimate ligand-binding affinities. *Expert Opinion on Drug Discovery*, 10, 449-461.
- Gentile, D. R., Rathinaswamy, M. K., Jenkins, M. L., Moss, S. M., Siempelkamp, B. D., Renslo, A. R., . . . Shokat, K. M. (2017). Ras binder induces a modified switch-II pocket in GTP and GDP states. *Cell Chemical Biology*, 24, 1455-1466.
- Giancotti, F. G. (2014). Deregulation of cell signaling in cancer. *FEBS Letters*, 588, 2558-2570.
- Gleeson, M. P. (2007). Plasma protein binding affinity and its relationship to molecular structure: An *in-silico* analysis. *Journal of Medicinal Chemistry*, 50, 101-112.
- Godfrey, K. R., Arundel, P. A., Zhu, W., Dong, Z., & Bryant, R. (2011). Modelling the double peak phenomenon. *Journal of Bioequivalence & Bioavailability*, 3, 101-107.
- Goldstein, I., Rivlin, N., Shoshana, O., Ezra, O., Madar, S., Goldfinger, N., & Rotter, V. (2013). Chemotherapeutic agents induce the expression and activity of their clearing enzyme CYP3A4 by activating p53. *Carcinogenesis*, 34, 190-198.
- Grant, B. J., Lukman, S., Hocker, H. J., Sayyah, J., Brown, J. H., McCammon, A. J., & Gorfe, A. A. (2011). Novel allosteric sites on Ras for lead generation. *PLoS ONE*, 6, e25711.
- Greenburg, G. & Hay, E. D. (1982) Epithelia suspended in collagen gels can lose polarity and express characteristics of migrating mesenchymal cells. *Journal of Cell Biology*, 95, 333-339.
- Grüner, B. M., Winkelmann, I., Feuchtinger, A., Sun, N., Balluff, B., Teichmann, N., . . . Siveke, J. T. (2016). Modeling therapy response and spatial tissue distribution of erlotinib in pancreatic cancer. *Molecular Cancer Therapeutics*, 15, 1145-1152.
- Gueron, G., De Siervi, A., Ferrando, M., Salierno, M., De Luca, P., Elguero, B., . . . Vazquez, E. S. (2009). Critical role of endogenous heme oxygenase 1 as a tuner of the invasive potential of prostate cancer cells. *Molecular Cancer Research*, 7, 1745-1755.



- Gullo, I., Tomassetti, P., Migliori, M., Casadei, R., & Marrano, D. (2001). Do early symptoms of pancreatic cancer exist that can allow an earlier diagnosis? *Pancreas*, *22*, 210-213.
- Guo, W., Wu, S., Liu, J., & Fang, B. (2008). Identification of a small molecule with synthetic lethality for K-RAS and protein kinase C iota. *Cancer Research*, *68*, 7403-7408.
- Habbe, N., Shi, G., Meguid, R. A., Fendrich, V., Esni, F., Chen, H., . . . Maitra, A. (2008). Spontaneous induction of murine pancreatic intraepithelial neoplasia (mPanIN) by acinar cell targeting of oncogenic Kras in adult mice. *Proceedings of the National Academy of Sciences of the United States of America*, *105*, 18913-18918.
- Haigis, K. M. (2017). KRAS alleles: The devil is in the detail. *Trends in Cancer*, *3*, 686-697.
- Halgren, T. (2007). New method for fast and accurate binding-site identification and analysis. *Chemical Biology & Drug Design*, *69*, 146-148.
- Halgren, T. A. (2009). Identifying and characterizing binding site and assessing druggability. *Journal of Chemical Information and Modeling*, *49*, 377-389.
- Han, L., Jiang, J., Ma, Q., Wu, Z., & Wang, Z. (2018). The inhibition of heme oxygenase-1 enhances the chemosensitivity and suppresses the proliferation of pancreatic cancer cells through the SHH signaling pathway. *International Journal of Oncology*, *52*, 2101-2109.
- Han, Y., & Chen, J. Z. (2013). Oxidative stress induces mitochondrial DNA damage and cytotoxicity through independent mechanisms in human cancer cells. *BioMed Research International*, *2013*, 8 pages.
- Hanada, T., Noda, N. N., Satomi, Y., Ichimura, Y., Fujioka, Y., Takao, T., . . . Ohsumi, Y. (2007). The Atg12-Atg5 conjugate has a novel E3-like activity for protein lipidation in autophagy. *Journal of Biological Chemistry*, *282*, 37298-37302.
- Hanahan, D., & Weinberg, R. A. (2011). Hallmarks of cancer: The next generation. *Cell*, *144*, 646-674.
- Hanahan, D., & Weinberg, R. A. (2016). Biological hallmarks of cancer. In R. C. Bast, Jr., C. M. Croce, W. N. Hait, W. K. Hong, D. W. Kufe, M. Piccart-Gebart, . . . J. F. Holland (Eds.), *Holland-Frei Cancer Medicine* (pp. 1-10). Hoboken (NJ): John Wiley & Sons, Inc.

- Hancock, J. F., & Parton, R. G. (2005). Ras plasma membrane signalling platforms. *Biochemical Journal*, 389, 1-11.
- Hancock, J. F., Magee, A. I., Childs, J. E., & Marshall, C. J. (1989). All ras proteins are polyisoprenylated but only some are palmitoylated. *Cell*, 57, 1167-1177.
- Hancock, J. F., Paterson, H., & Marshall, C. J. (1990). A polybasic domain or palmitoylation is required in addition to the CAAX motif to localize p21ras to the plasma membrane. *Cell*, 63, 133-139.
- Harada, K., Ferdous, T., & Ueyama, Y. (2014). Establishment of 5-fluorouracil-resistant oral squamous cell carcinoma cell lines with epithelial to mesenchymal transition changes. *International Journal of Oncology*, 44, 1302-1308.
- Hata, A. N., Niederst, M. J., Archibald, H. L., Gomez-Caraballo, M., Siddiqui, F. M., Mulvey, H. E., . . . Engelman, J. A. (2016). Tumor cells can follow distinct evolutionary paths to become resistant to epidermal growth factor receptor inhibition. *Nature Medicine*, 22, 262-269.
- Higuchi, M. Honda, T., Proske, R. J., & Yeh, E. T. H. (1998). Regulation of reactive oxygen species-induced apoptosis and necrosis by caspase 3-like proteases. *Oncogene*, 17, 2753-2760.
- Hill, D. P., Smith, B., McAndrews-Hill, M. S., & Blake, J. A. (2008). Gene Ontology annotations: what they mean and where they come from. *BMC Bioinformatics*, 9, S2.
- Hiscox, S., Jiang, W. G., Obermeier, K., Taylor, K., Morgan, L., Burmi, R., . . . Nicholson, R. I. (2006). Tamoxifen resistance in MCF7 cells promotes EMT-like behaviour and involves modulation of beta-catenin phosphorylation. *International Journal of Cancer*, 118, 290-301.
- Hobbs, G. A., Der, C. J., & Rossman, K. L. (2016). RAS isoforms and mutations in cancer at a glance. *Journal of Cell Science*, 129, 1287-1292.
- Hocker, H. J., Cho, K., Chen, C. K., Rambahal, N., Sagineedu, S. R., Shaari, K., . . . Gorfe, A. A. (2013). Andrographolide derivatives inhibit guanine nucleotide exchange and abrogate oncogenic Ras function. *Proceedings of the National Academy of Sciences of the United States of America*, 110, 10201-10206.
- Holbeck, S. L., Camalier, R., Crowell, J. A., Govindharajulu, J. P., Hollingshead, M., Anderson, L. W., . . . Doroshow, J. H. (2017). The National Cancer Institute ALMANAC: A comprehensive screening

- resource for the detection of anticancer drug pairs with enhanced therapeutic activity. *Cancer Research*, 77, 3564-3576.
- Holen, I., Speirs, V., Morrissey, B., & Blyth, K. (2017). *In vivo* models in breast cancer research: progress, challenges and future directions. *Disease Models & Mechanisms*, 10, 359-371.
- Hospital, A., Goñi, J. R., Orozco, M., & Gelpi, J. L. (2015). Molecular dynamics simulations: advances and applications. *Advances and Applications in Bioinformatics and Chemistry*, 8, 37-47.
- Housman, G., Byler, S., Heernoth, S., Lapinska, K., Longacre, M., Snyder, N., & Sarkar, S. (2014). Drug resistance in cancer: An overview. *Cancers*, 6, 1769-1792.
- Hruban, R. H., Goggins, M., Parsons, J., & Kern, S. E. (2000). Progression model for pancreatic cancer. *Clinical Cancer Research*, 6, 2969-2972.
- Hu, L., & Cong, L. (2015). Fibroblast growth factor 19 is correlated with an unfavorable prognosis and promotes progression by activating fibroblast growth factor receptor 4 in advanced-stage serous ovarian cancer. *Oncology Reports*, 34, 2683-2691.
- Huang, E. H., & Wicha, M. S. (2008). Colon cancer stem cells: implications for prevention and therapy. *Trends in Molecular Medicine*, 14, 503-509.
- Huang, J., & Manning, B. D. (2008). The TSC1–TSC2 complex: a molecular switchboard controlling cell growth. *Biochemical Journal*, 412, 179-190.
- Huang, S., & Zou, X. (2010). Advances and challenges in protein-ligand docking. *International Journal of Molecular Sciences*, 11, 3016-3034.
- Huang, W., & Hung, M. (2009). Induction of Akt activity by chemotherapy confers acquired resistance. *Journal of the Formosan Medical Association*, 108, 180-194.
- Hubbard, P. A., Moody, C. L., & Murali, R. (2014). Allosteric modulation of Ras and the PI3K/AKT/mTOR pathway: emerging therapeutic opportunities. *Frontiers in Physiology*, 5, Article 478, 7 pages.
- Hughes, J. P., Rees, S., Kalindjian, S. B., & Philpott, K. L. (2011). Principles of early drug discovery. *British Journal of Pharmacology and Chemotherapy*, 162, 1239-1249.

- Hughes, P., Marshall, D., Reid, Y., Parkes, H., & Gelber, C. (2007). The costs of using unauthenticated, over-passaged cell lines: How much more data do we need? *BioTechniques*, *43*, 575-586.
- Hung, S., Hung, L., Kuo, C., Lee, K., Lee, M., Lin, H., . . . Fu, S. (2010). Andrographolide sensitizes ras-transformed cells to radiation *in vitro* and *in vivo*. *International Journal of Radiation Oncology, Biology, Physics*, *77*, 1232-1239.
- Hunter, J. C., Gurbani, D., Ficarro, S. B., Carraasco, M. A., Lim, S. M., Choi, H. G., . . . Westover, K. D. (2014). *In situ* selectivity profiling and crystal structure of SML-8-73-1, and active site inhibitor of oncogenic K-Ras G12C. *proceedings of the National Academy of Sciences of the United States of America*, *111*, 8895-8900.
- Hunter, J. C., Manandhar, A., Carrasco, M. A., Gurbani, D., Gondi, S., & Westover, K. D. (2015). Biochemical and structural analysis of common cancer-associated KRAS mutations. *Molecular Cancer Research*, *13*, 1325-1335.
- Ibrahim, K. E., Al-Mutary, M. G., Bakhiet, A. O., & Khan, H. A. (2018). Histopathology of the liver, kidney, and spleen of mice exposed to gold nanoparticles. *Molecules*, *23*, 1848.
- ICH. (2005). Validation of Analytical Procedures: Text and Methodology Q2(R1). Switzerland: International Conference on Harmonisation of Technical Requirements for Registration of Pharmaceuticals for Human Use.
- ICH. (2019). ICH Harmonised Guideline: Bioanalytical Method Validation M10 (Draft Version). Switzerland: International Council for Harmonisation of Technical Requirements for Pharmaceuticals for Human Use.
- Jada, S. R., Hamzah, A. S., Lajis, N. H., Saad, M. S., Stevens, M. F., & Stanslas, J. (2006). Semisynthesis and cytotoxic activities of andrographolide analogues. *Journal of Enzyme Inhibition and Medicinal Chemistry*, *21*, 327.
- Jada, S. R., Subur, G. S., Matthews, C., Hamzah, A. S., Lajis, N. H., Saad, M. S., . . . Stanslas, J. (2007). Semisynthesis and *in vitro* anticancer activities of andrographolide analogues. *Phytochemistry*, *68*, 904-912.
- Jada, S. R., Matthews, C., Saad, M. S., Hamzah, A. S., Lajis, N. H., Steven, M. F., & Stanslas, J. (2008). Benzylidene derivatives of andrographolide inhibit growth of breast and colon cancer cells *in vitro* by inducing G1 cell cycle arrest and apoptosis. *British Journal of Pharmacology and Chemotherapy*, *155*, 641-654.

- Jain, R. K. (1997). Delivery of molecular and cellular medicine to solid tumors. *Microcirculation*, 4, 1-23.
- Jakate, A. S., Einhaus, C. M., DeAnglis, A. P., Retzinger, G. S., & Desai, P. B. (2003). Preparation, characterization, and preliminary application of fibrinogen-coated olive oil droplets for the targeted delivery of docetaxel to solid malignancies. *Cancer Research*, 63, 7314-7320.
- Janes, M. R., Zhang, J., Li, L., Hansen, R., Peters, U., Guo, X., . . . Liu, Y. (2018). Targeting KRAS mutant cancers with a covalent G12C-specific inhibitor. *Cell*, 172, 578-589.
- Jansen, P. L. (2017). Fibroblast growth factor 19, a double-edged sword. *Hepatology Oncology*, 4, 1-4.
- Jelinek, T., Dent, P., Sturgill, T. W., & Weber, M. J. (1996). Ras-induced activation of Raf-1 is dependent on tyrosine phosphorylation. *Molecular and Cellular Biology*, 16, 1027-1034.
- Jin, W., Wu, L., Liang, K., Liu, B., Lu, Y., & Fan, Z. (2003). Roles of the PI-3K and MEK pathways in Ras-mediated chemoresistance in breast cancer cells. *British Journal of Cancer*, 89, 185-191.
- Jo, E. J., Park, S. J., & Kim, B. (2016). Propyl gallate sensitizes human lung cancer cells to cisplatin-induced apoptosis by targeting heme oxygenase-1 for TRC8-mediated degradation. *European Journal of Pharmacology*, 788, 321-327.
- John, J., Rensland, H., Schlichting, I., Vetter, L., Borasio, G. D., Goody, R. S., & Wittinghofer, A. (1993). Kinetic and structural analysis of the Mg(2+)-binding site of the guanine nucleotide-binding protein p21H-ras. *Journal of Biological Chemistry*, 268, 923-929.
- Joly, M. M., Hicks, D. J., Jones, B., Sanchez, V., Estrada, M. V., Young, C., . . . Cook, R. S. (2016). Rictor/mTORC2 drives progression and therapeutic resistance of HER2-amplified breast cancers. *Cancer Research*, 76, 4752-4764.
- Jones, G., Willet, P., Glen, R. C., Leach, A. R., & Taylor, R. (1997). Development and validation of a genetic algorithm for flexible docking. *Journal of Molecular Biology*, 267, 727-748.
- Juliano, R. L., & Ling, V. (1976). A surface glycoprotein modulating drug permeability in Chinese hamster ovary cell mutants. *Biochimica et Biophysica Acta*, 455, 152-162.

- Jun, J. E., Rubio, I., & Roose, J. P. (2013). Regulation of Ras exchange factors and cellular localization of Ras activation by lipid messengers in T cells. *Frontiers in Immunology*, 4, 239.
- Jung, J. (2014). Human tumor xenograft models for preclinical assessment of anticancer drug development. *Toxicological Research*, 30, 1-5.
- Kahn, P., & Shin, S. (1979). Cellular tumorigenicity in nude mice. *Journal of Cell Biology*, 82, 1-16.
- Kanda, M., Matthaei, H., Wu, J., Hong, S. M., Yu, J., Borges, M., . . . Goggins, M. (2012). Presence of somatic mutations in most early-stage pancreatic intraepithelial neoplasia. *Gastroenterology*, 142, 730-733.
- Kapoor, A., & Travasset, A. (2015). Differential dynamics of RAS isoforms in GDP- and GTP-bound states. *Proteins*, 83, 1091-1106.
- Kars, M. D., Iseri, O. D., Gündüz, U., Ural, A. U., Arpacı, F., & Molnár, J. (2006). Development of rational *in vitro* models for drug resistance in breast cancer and modulation of MDR by selected compounds. *Anticancer Research*, 26, 4559-4568.
- Kelland, L. R. (2004). "Of mice and men": values and liabilities of the athymic nude mouse model in anticancer drug development. *European Journal of Cancer*, 40, 827-836.
- Keohavong, P., DeMichele, M. A., Melacrinis, A. C., Landreneau, R. J., Weyant, R. J., & Siegfried, J. M. (1996). Detection of K-ras mutations in lung carcinomas: Relationship to prognosis. *Clinical Cancer Research*, 2, 411-418.
- Kerns, E. H., & Di, L. (2008). *Drug-like Properties: Concepts, Structure Design and Methods: from ADME to Toxicity Optimization*. London, United Kingdom: Academic Press, Elsevier.
- Khrenova, M. G., Mironov, V. A., Grigorenko, B. L., & Nemukhin, A. V. (2014). Modeling the role of G12V and G13V Ras mutations in the Ras-GAP-catalyzed hydrolysis reaction of guanosine triphosphate. *Biochemistry*, 53, 7093-7099.
- Khwaja, A., Rodriguez-Viciana, P., Wennstrom, S., Warne, P. H., & Downward, J. (1997). Matrix adhesion and Ras transformation both activate a phosphoinositide 3-OH kinase and protein kinase B/Akt cellular survival pathway. *EMBO Journal*, 16, 2783-2793.
- Knodell, R. G., Farleigh, M., Steele, N. M., & Bond, J. H. (1982). Effects of liver congestion on hepatic drug metabolism in the rat. *Journal of Pharmacology and Experimental Therapeutics*, 221, 52-57.

- Knudsen, E. S., Kumarasamy, V., Ruiz, A., Sivinski, J., Chung S., Grant, A., ... Witkiewicz, A. K. (2019). Cell cycle plasticity driven by MTOR signaling: integral resistance to CDK4/6 inhibition in patient-derived models of pancreatic cancer. *Oncogene*, 38, 3355-3370.
- Kodaz, H., Kostek, O., Hacıoglu, M. B., Erdogan, B., Kodaz, C. E., Hacibekiroglu, I., . . . Cicin, I. (2017). Frequency of RAS mutations (KRAS, NRAS, HRAS) in human solid cancer. *Eurasian Journal of Medicine and Oncology*, 1, 1-7.
- Kolch, W. (2000). Meaningful relationships: the regulation of the Ras/Raf/MEK/ERK pathway by protein interactions. *Biochemical Journal.*, 351, 289-305.
- Kolch, W., Heidecker, G., Lloyd, P., & Rapp, U. R. (1991). Raf-1 protein kinase is required for growth of induced NIH/3T3 cells. *Nature*, 349, 426-428.
- Koong, A. C., Metha, V. K., Le, Q. T., Fisher, G. A., Terris, D. J., Brown, J. M., . . . Vierra, M. (2000). Pancreatic tumors show high levels of hypoxia. *International Journal of Radiation Oncology, Biology, Physics*, 48, 919-922.
- Koundouros, N., & Poulgiannis, G. (2018). Phosphoinositide 3-kinase/Akt signaling and redox metabolism in cancer. *Frontiers in Oncology*, 8, 160.
- Krempley, B. D., & Yu, K. H. (2017). Preclinical models of pancreatic ductal adenocarcinoma. *Chinese Clinical Oncology*, 6, 25.
- Laconi, E., Marongiu, F., & DeGregori, J. (2020). Cancer as a disease of old age: changing mutational and microenvironment landscapes. *British Journal of Cancer*, 122, 943-952.
- Lao, D., Chandramouli, S., Yusoff, P., Fong, C. W., Saw, T. Y., Tai, L. P., . . . Guy, G. R. (2006). A Src homology 3-binding sequence on the C terminus of Sprouty2 is necessary for inhibition of the Ras/ERK pathway downstream of fibroblast growth factor receptor stimulation. *Journal of Biological Chemistry*, 281, 29993-30000.
- Lazarev, V. F., Guzhova, I. V., & Margulis, B. A. (2020). Glyceraldehyde-3-phosphate dehydrogenase is a multifaceted therapeutic target. *Pharmaceutics*, 12, 416.
- Lee, C., & Macgregor, P. F. (2004). Using microarrays to predict resistance to chemotherapy in cancer patients. *Pharmacogenomics*, 5, 611-625.

- Lee, Y. C., Lin, H. H., Ksu, C. H., Wang, C. J., Chiang, T. A., & Chen, J. H. (2010). Inhibitory effects of andrographolide on migration and invasion in human non-small cell lung cancer A549 cells *via* down-regulation of PI3K/Akt signaling pathway. *Journal of Biological Chemistry*, 632, 23-32.
- Lengauer, T., & Rarey, M. (1996). Computational methods for biomolecular docking. *Current Opinion in Structural Biology*, 6, 402-406.
- Lennartsson, J., & Rönstrand, L. (2012). Stem cell factor receptor/c-Kit: from basic science to clinical implications. *Physiological Reviews*, 92, 1619-1649.
- Less, J. R., Posner, M. C., Skalak, T. C., Wolmark, N., & Jain, R. K. (1997). Geometric resistance and microvascular network architecture of human colorectal carcinoma. *Microcirculation*, 4, 25-33.
- Li, J., Zhang, C., Jiang, H., & Cheng, J. (2015). Andrographolide inhibits hypoxia-inducible factor-1 through phosphatidylinositol 3-kinase/AKT pathway and suppresses breast cancer growth. *OncoTargets and Therapy*, 8, 427-435.
- Liao, J., Shima, F., Araki, M., Ye, M., Muraoka, S., Sugimoto, T., . . . Kataoka, T. (2008). Two conformational states of Ras GTPase exhibit differential GTP-binding kinetics. *Biochemical and Biophysical Research Communications*, 369, 327-332.
- Lieber, M., Mazzetta, J., Nelson-Rees, W., Kaplan, M., & Todaro, G. (1975). Establishment of a continuous tumor-cell line (PANC-1) from a human carcinoma of the exocrine pancreas. *International Journal of Cancer*, 15, 741-747.
- Lim, J. C., Jeyaraj, E. J., Sagineedu, S. R., Wong, W. F., & Stanslas, J. (2015). SRS06, a new semisynthetic andrographolide derivative with improved anticancer potency and selectivity, inhibits nuclear factor- $\kappa$ B nuclear binding in the A549 non-small cell lung cancer cell line. *Pharmacology*, 95, 70-77.
- Lim, S. C., Jeon, H. J., Kee, K. H., Lee, M. J., Hong, R., & Han, S. I. (2017). Andrographolide induces apoptotic and non-apoptotic death and enhances tumor necrosis factor-related apoptosis-inducing ligand-mediated apoptosis in gastric cancer cells. *Oncology Letters*, 13, 3837-3844.
- Lim, Z., & Ma, P. C. (2019). Emerging insights of tumor heterogeneity and drug resistance mechanisms in lung cancer targeted therapy. *Journal of Hematology & Oncology*, 12, 134.



- Lin, H. H., Tsai, C. W., Chou, F. P., Wang, C. J., Hsuan, S. W., Wang, C. K., & Chen, J. H. (2011). Andrographolide down-regulates hypoxia-inducible factor-1 $\alpha$  in human non-small cell lung cancer A549 cells. *Toxicology and Applied Pharmacology*, 250, 336-345.
- Lin, J. H., & Lu, A. Y. (1997). Role of pharmacokinetics and metabolism in drug discovery and development. *Pharmacological Reviews*, 49, 403-449.
- Lipinski, C. A., Lombardo, F., Dominy, B. W., & Feeney, P. J. (2001). Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings. *Advanced Drug Delivery Reviews*, 46, 3-26.
- Lu, S., Jang, H., Gu, S., Zhang, J., & Nussinov, R. (2016). Drugging Ras GTPase: a comprehensive mechanistic and signaling structural view. *Chemical Society Reviews*, 45, 929-4952.
- Lu, S., Jang, H., Nussinov, R., & Zhang, J. (2016). The structural basis of oncogenic mutations G12, G13 and Q61 in small GTPase K-Ras4B. *Scientific Reports*, 6, 21949.
- Lukas, G., Brindle, S. D., & Greengard, P. (1971). The route of absorption of intraperitoneally administered compounds. *Journal of Pharmacology and Experimental Therapeutics*, 178, 562-566.
- Luo, Z., Tzivion, G., Belshaw, P. J., Vavvas, D., Marshall, M., & Avruch, J. (1996). Oligomerization activates c-Raf-1 through a Ras-dependent mechanism. *Nature*, 383, 181-185.
- Marchetti, S., & Schellens, J. H. (2007). The impact of FDA and EMEA guidelines on drug development in relation to Phase 0 trials. *British Journal of Cancer*, 97, 577-581.
- Markman, M. (2003). Intraperitoneal antineoplastic drug delivery: rationale and results. *The Lancet Oncology*, 4, 277-283.
- Maenhaut, C., Dumont, J. E., Roger, P. P., & van Staveren, W. C. (2010). Cancer stem cells: a reality, a myth, a fuzzy concept or a misnomer? An analysis. *Carcinogenesis*, 31, 149-158.
- Małyszko, J., Kozłowska, K., Kozłowski, L., & Małyszko, J. (2017). Nephrotoxicity of anticancer treatment. *Nephrology Dialysis Transplantation*, 32, 924-936.
- Manda, G., Isvoranu, G., Comanescu, M. C., Manea, A., Butuner, B. D., & Korkmaz, K. S. (2015). The redox biology network in cancer pathophysiology and therapeutics. *Redox Biology*, 5, 347-357.

- Mann, P. C., Cahle, J., Keenan, C. M., Baker, J. F., Bradley, A. E., Goodman, D. G., . . . Tanaka, T. (2012). International harmonization of toxicologic pathology nomenclature: An overview and review of basic principles. *Toxicologic Pathology*, *40*, 7S-13S.
- Manson, M. M. (1980). Epoxides - Is there a human health problem? *The British Journal of Industrial Medicine*, *37*, 317-336.
- Mansoori, B., Mohammadi, A., Davudian, S., Shirjang, S., & Baradaran, B. (2017). The different mechanisms of cancer drug resistance: A brief review. *Advanced Pharmaceutical Bulletin*, *7*, 339-348.
- Maor, Y., & Malnick, S. (2013). Liver injury induced by anticancer chemotherapy and radiation therapy. *International Journal of Hepatology*, *2013*, 8 pages.
- Marín-Ramos, N. I., Piñar, C., Vázquez-Villa, H., Martín-Fontecha, M., González, Á., Canales, Á., ... López-Rodríguez, M. L. (2017). Development of a nucleotide exchange inhibitor that impairs Ras oncogenic signaling. *Chemistry – A European Journal*, *23*, 1676-1685.
- Marom, M., Haklai, R., Ben-Baruch, G., Marciano, D., Egozi, Y., & Kloog, Y. (1995). Selective inhibition of Ras-dependent cell growth by farnesylthiosalicylic acid. *Journal of Biological Chemistry*, *270*, 22263-22270.
- Maruyama, I. N. (2014). Mechanisms of activation of receptor tyrosine kinases: Monomers or Dimers. *Cells*, *3*, 304-330.
- Massarelli, E., Varella-Garcia, M., Tang, X., Xavier, A. C., Ozburn, N. C., Liu, D. D., . . . Wistuba, I. I. (2007). KRAS mutation is an important predictor of resistance to therapy with epidermal growth factor receptor tyrosine kinase inhibitors in non-small-cell lung cancer. *Clinical Cancer Research*, *13*, 2890-2896.
- Matuszewski, B. K., Constanzer, M. L., & Chavez-Eng, C. M. (2003). Strategies for the assessment of matrix effect in quantitative bioanalytical methods based on HPLC-MS/MS. *Analytical Chemistry*, *75*, 3019-3030.
- Maurer, T., Garrenton, L. S., Oh, A., Pitts, K., Anderson, D. J., Skelton, N. J., . . . Fang, G. (2012). Small-molecule ligands bind to a distinct pocket in Ras and inhibit SOS-mediated nucleotide exchange activity. *Proceedings of the National Academy of Sciences of the United States of America*, *109*, 5299-5304.
- McDermott, M., Eustace, A. J., Busschots, S., Breen, L., Crown, J., Clynes, M., . . . Stordal, B. (2014). *In vitro* development of chemotherapy

and targeted therapy drug-resistant cancer cell lines: a practical guide with case studies. *Frontiers in Oncology*, 4, 16 pages.

- Mecklenburg, L., Nakamura, M., Sundberg, J. P., & Paus, R. (2001). The nude mouse skin phenotype: The role of Foxn1 in hair follicle development and cycling. *Experimental & Molecular Medicine*, 71, 171-178.
- Merika, E. E., Syrigos, K. N., & Saif, M. W. (2012). Desmoplasia in pancreatic cancer. Can we fight it? *Gastroenterology Research and Practice*, 2012, 10 pages.
- Mi, S., Xiang, G., Yuwen, D., Gao, J., Guo, W., Wu, X., . . . Xu, Q. (2016). Inhibition of autophagy by andrographolide resensitizes cisplatin-resistant non-small cell lung carcinoma cells via activation of the Akt/mTOR pathway. *Toxicology and Applied Pharmacology*, 310, 78-86.
- Miao, B., Skidan, I., Yang, J., Lugovskoy, A., Reibarkh, M., Long, K., . . . Degtarev, A. (2010). Small molecule inhibition of phosphatidylinositol-3,4,5-triphosphate (PIP3) binding to pleckstrin homology domains. *Proceedings of the National Academy of Sciences of the United States of America*, 107, 20126-20131.
- Michaelis, M., Wass, M. N., & Cinati jr., J. (2019). Drug-adapted cancer cell lines as preclinical models of acquired resistance. *Cancer Drug Resistance*, 2, 447-456.
- Michel-Reher, M. B., & Michel, M.C. (2015). Regulation of GAPDH expression by treatment with the  $\beta$ -adrenoceptor agonist isoprenaline – is GAPDH a suitable loading control in immunoblot experiments? *Naunyn-Schmiedeberg's Archives of Pharmacology*, 388, 1119-1120.
- Minato, K., Kanzawa, F., Nishio, K., Nakagawa, K., Fujiwara, Y., & Saijo, N. (1990). Characterization of an etoposide-resistant human small-cell lung cancer cell line. *Cancer Chemotherapy and Pharmacology*, 26, 313-317.
- Moodie, S. A., Willumsen, B. M., Weber, M. J., & Wolfman, A. (1993). Complexes of Ras.GTP with Raf-1 and mitogen-activated protein kinase kinase. *Science*, 260, 1658-1661.
- Morton, C. L., & Houghton, P. J. (2007). Establishment of human tumor xenografts in immunodeficient mice. *Nature Protocols*, 2, 247-250.
- Mosmann, T. (1983). Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *Journal of Immunological Methods*, 65, 55-63.

- Mujoo, K., Zhang, L., Klostergaard, J., & Donato, N.J. (2000). Emergence of cisplatin-resistant cells from the OVCAR-3 ovarian carcinoma cell line with p53 mutations, altered tumorigenicity, and increased apoptotic sensitivity to p53 gene replacement. *International Journal of Gynecological Cancer*, 10, 105-114.
- Murillo, M. M., Zelenay, S., Nye, E., Castellano, E., Lassailly, F., Stamp, G., & Downward, J. (2014). RAS interaction with PI3K p110 $\alpha$  is required for tumor-induced angiogenesis. *Journal of Clinical Investigation*, 24, 3601-3611.
- Nagamatsu, H., Teishima, J., Goto, K., Shikuma, H., Kitano, H., Shoji, K., . . . Matsubara, A. (2015). FGF19 promotes progression of prostate cancer. *Prostate*, 75, 1092-1101.
- Nakhaeizadeh, H., Amin, E., Nakhaei-Rad, S., Dvorsky, R., & Ahmadian, M. R. (2016). The RAS-effector interface: Isoform-specific differences in the effector binding regions. *PLoS ONE*, 11, e0167145.
- Nan, X., Tamgüney, T. M., Collisson, E. A., Lin, L., Pitt, C., Galeas, J., . . . Chu, S. (2015). Ras-GTP dimers activate the mitogen-activated protein kinase (MAPK) pathway. *Proceedings of the National Academy of Sciences of the United States of America*, 112, 7996-8001.
- Nancy, V., Callebaut, I., El Marjou, A., & de Gunzburg, J. (2002). The delta subunit of retinal rod cGMP phosphodiesterase regulates the membrane association of Ras and Rap GTPases. *Journal of Biological Chemistry*, 277, 15076-15084.
- Nanduri, S., Nyavanandi, V. K., Sanjeeva Rao Thunuguntla, S., Kasu, S., Pallerla, M. K., Sai Ram, P., . . . Akella, V. (2004). Synthesis and structure–activity relationships of andrographolide analogues as novel cytotoxic agents. *Bioorganic & Medicinal Chemistry Letters*, 14, 4711-4717.
- Naponelli, V., Modernelli, A., Bettuzzi, S., & Rizzi, F. (2015). Roles of autophagy induced by natural compounds in prostate cancer. *BioMed Research International*, 2015, 14 pages.
- Nateewattana, J., Dutta, S., Reabroi, S., Saeeng, R., Kasemsook, S., Chairoungdua, A., . . . Piyachaturawat, P. (2014). Induction of apoptosis in cholangiocarcinoma by an andrographolide analogue is mediated through topoisomerase II alpha inhibition. *European Journal of Pharmacology*, 723, 148-155.
- Nateewattana, J., Saeeng, R., Kasemsook, S., Suksen, K., Dutta, S., Jariyawat, S., . . . Piyachaturawat, P. (2013). Inhibition of topoisomerase II  $\alpha$  activity and induction of apoptosis in

mammalian cells by semi-synthetic andrographolide analogues. *Investigational New Drugs*, 31, 320-332.

- Nayal, M., & Honig, B. (2006). On the nature of cavities on protein surfaces: Application to the identification of drug-binding sites. *Proteins*, 63, 892-906.
- NCI. (2020, September 25). *National Cancer Institute*. Retrieved from Cancer Statistics: <https://www.cancer.gov/about-cancer/understanding/statistics#:~:text=Statistics%20at%20a%20Gance%3A%20The%20Burden%20of%20Cancer%20Worldwide&text=In%202018%2C%20there%20were%2018.1,related%20deaths%20to%2016.4%20million>.
- NCI Staff. (2019, December 5). *National Cancer Institute*. Retrieved from Overcoming a Cancer Nemesis? KRAS Inhibitor Shows Promise in Early Trial: <https://www.cancer.gov/news-events/cancer-currents-blog/2019/kras-inhibitor-amg-510-clinical-trial>
- Neuzillet, C., Hammel, P., Tijeras-Raballand, A., Coulevar, A., & Raymond, E. (2013). Targeting the Ras–ERK pathway in pancreatic adenocarcinoma. *Cancer and Metastasis Reviews*, 32, 147-162.
- Ni, D., Li, X., He, X., Zhang, H., Zhang, J., & Lu, S. (2019). Drugging K-RasG12C through covalent inhibitors: Mission possible? *Pharmacology & Therapeutics*, 202, 1-17.
- Nishant, T., Sathish, K. D., Arun, K., & Phaneendra, M. (2011). Role of Pharmacokinetic Studies in Drug Discovery. *Journal of Bioequivalence & Bioavailability*, 3, 263-267.
- Nogueira, V., Park, Y., Chen, C., Xu, P., Chen, M., Tonic, I., . . . Hay, N. (2008). Akt determines replicative senescence and oxidative or oncogenic premature senescence and sensitizes cells to oxidative apoptosis. *Cancer Cell*, 14, 458-470.
- Noonan, T., Brown, N., Dudycz, L., & Wright, G. (1991). Interaction of GTP derivatives with cellular and oncogenic ras-p21 proteins. *Journal of Medicinal Chemistry*, 34, 1302-1307.
- Nukatsuka, M., Nakagawa, F., Saito, H., Sakata, M., Uchida, J., & Takechi, T. (2015). Efficacy of combination chemotherapy using a novel oral chemotherapeutic agent, TAS-102, with irinotecan hydrochloride on human colorectal and gastric cancer xenografts. *Anticancer Research*, 35, 1437-1446.

- O'Brien, C. A., Pollett, A., Gallinger, S., & Dick, J. E. (2007). A human colon cancer cell capable of initiating tumour growth in immunodeficient mice. *Nature*, *445*, 106-110.
- O'Bryan, J. P. (2019). Pharmacological targeting of RAS: Recent success with direct inhibitors. *Pharmacological Research*, *139*, 503-511.
- O'Neill, E., & Kolch, W. (2004). Conferring specificity on the ubiquitous Raf/MEK signalling pathway. *British Journal of Cancer*, *90*, 283-288.
- OECD. (2009). Chronic Toxicity Studies. Test Guideline No. 452, OECD Guidelines for the Testing of Chemicals. Paris: Organisation for Economic Co-operation and Development.
- OECD. (2009). Draft Guidance Document on the Design and Conduct of Chronic Toxicity and Carcinogenicity Studies. Series on Testing and Assessment No. 116. Paris: Organisation for Economic Co-operation and Development.
- Orhon, I., & Reggiori, F. (2017). Assays to monitor autophagy progression in cell cultures. *Cells*, *6*, 20.
- Ornitz, D. M., & Itoh, N. (2015). The fibroblast growth factor signaling pathway. *Wiley Interdisciplinary Reviews-Developmental Biology*, *4*, 215-266.
- Österberg, F., Morris, G. M., Sanner, M. F., Olson, A. J., & Goodsell, D. S. (2002). Automated docking to multiple target structures: incorporation of protein mobility and structural water heterogeneity in AutoDock. *Proteins*, *46*, 34-40.
- Ostrem, J. M., Peters, U., Sos, M. L., Wells, J. A., & Shokat, K. M. (2013). K-Ras(G12C) inhibitors allosterically control GTP affinity and effector interaction. *Nature*, *503*, 548-551.
- Ottery, F. D. (2000). Bidirectional interplay of nutrition and chemotherapy. In J. B. Mason, & G. Nitenberg (Eds.), *Cancer & Nutrition: Prevention and Treatment* (pp. 183-206). Basel: S. Karger AG.
- Paduch, M., Jeleń, F., & Otlewski, J. (2001). Structure of small G proteins and their regulators. *Acta Biochimica Polonica*, *48*, 829-850.
- Pai, E. F., Krengel, U., Petsko, G. A., Goody, R. S., Kabsch, W., & Wittinghofer, A. (1990). Refined crystal structure of the triphosphate conformation of H-ras p21 at 1.35 Å resolution: implications for the mechanism of GTP hydrolysis. *EMBO Journal*, *9*, 2351-2359.
- Paldino, E., Tesori, V., Casalbore, P., Gasbarrini, A., & Puglisi, M. A. (2014). Tumor initiating cells and chemoresistance: Which is the

best strategy to target colon cancer stem cells? *BioMed Research International*, 2014, 7 pages.

- Pao, W., Wang, T. Y., Riely, G. J., Miller, V. A., Pan, Q., Ladanyi, M., . . . Varmus, H. E. (2005). KRAS mutations and primary resistance of lung adenocarcinomas to gefitinib or erlotinib. *PLoS Medicine*, 2, e17.
- Parker, J. A., & Mattos, C. (2015). The Ras-membrane interface: isoform-specific differences in the catalytic domain. *Molecular Cancer Research*, 13, 595.
- Patel, V., Senderowicz, A. M., Pinto, D. J., Igishi, T., Mark, R., Quitanilla-Martinez, L., . . . Gutkind, J. S. (1998). Flavopiridol, a novel cyclin-dependent kinase inhibitor, suppresses the growth of head and neck squamous cell carcinomas by inducing apoptosis. *Journal of Clinical Investigation*, 102, 1674-1681.
- Patil, R., Venkat, R. P., & Patil, V. (2014). New substituted C(14)-andrographolide derivatives with potent cytotoxic activities. *International Journal of Pharma and Bio Sciences*, 5, 526-532.
- Patricelli, M. P., Janes, M. R., Li, L. S., Hansen, R., Peters, U., Kessler, L. V., . . . Liu, Y. (2016). Selective inhibition of oncogenic KRAS output with small molecules targeting the inactive state. *Cancer Discovery*, 6, 316-329.
- PCAN. (2018). *Pancreatic Cancer Facts*. California: Pancreatic Cancer Action Network.
- Pelkonen, O., Turpeinen, M., & Raunio, H. (2011). *In vivo-in vitro-in silico* pharmacokinetic modelling in drug development: Current status and future directions. *Clinical Pharmacokinetics*, 50, 483-491.
- Pierotti, M. A., Sozzi, G., & Croce, C. M. (2003). Mechanisms of oncogene activation. In D. W. Kufe, R. E. Pollock, R. R. Weichselbaum, C. B. Robert, T. S. Gansler, J. F. Holland, & E. Frei (Eds.), *Holland-Frei Cancer Medicine* (6th edition ed.). Hamilton (ON): BC Decker.
- Pietras, A. (2011). Cancer stem cells in tumor heterogeneity. In D. Gisselsson, *Advances in cancer research* (pp. 255-281). Amsterdam: Elsevier Inc.
- Pijl, H., & Meinders, A. E. (1996). Bodyweight change as an adverse effect of drug treatment. Mechanisms and management. *Drug Safety*, 14, 329-342.
- Plowman, J., Dykes, D. J., Hollingshead, M., Simpson-Herren, L., & Alley, M. C. (1997). Human tumor xenograft models in NCI drug

- development. In B. Teicher, & N. J. Totowa, *Anticancer drug development guide: preclinical screening, clinical trials and approval* (pp. 101-125). Totowa: Humana Press Inc.
- Poerwono, H., Hattori, Y., Kubo, H., & Higashiyama, K. (2007). Structure modification of andrographolide improve its potency as anticancer. *Indonesian Journal of Chemistry*, *7*, 202-207.
- Powis, G. (1983). Dose-dependent metabolism, therapeutic effect, and toxicity of anticancer drugs in man. *Drug Metabolism Reviews*, *14*, 1145-1163.
- Prakash, P., Hancock, J. F., & Gorfe, A. A. (2015). Binding hotspots on K-ras: Consensus ligand binding sites and other reactive regions from probe-based molecular dynamics analysis. *Proteins*, *83*, 898-909.
- Prior, I. A., Lewis, P. D., & Mattos, C. (2012). A comprehensive survey of Ras mutations in cancer. *Cancer Research*, *72*, 2457-2467.
- Qi, L., Sun, B., Liu, Z., Li, H., Gao, J., & Leng, X. (2012). Dickkopf-1 inhibits epithelial-mesenchymal transition of colon cancer cells and contributes to colon cancer suppression. *Cancer Science*, *103*, 828-835.
- Qiu, W., Sahin, F., Iacobuzio-Donahue, C. A., Garcia-Carracedo, D., Wang, W. M., Kuo, C., . . . Su, G. H. (2011). Disruption of p16 and activation of Kras in pancreas increase ductal adenocarcinoma formation and metastasis *in vivo*. *Oncotarget*, *2*, 862-873.
- Qu, L., Pan, C., He, S., Lang, B., Gao, G., Wang, X., & Wang, Y. (2019). The Ras superfamily of small GTPases in non-neoplastic cerebral diseases. *Frontiers in Molecular Neuroscience*, *12*, 121.
- Rai, Y., Pathak, R., Kumari, N., Sah, D. K., Pandey, S., Kalra, N., . . . Bhatt, A. N. (2018). Mitochondrial biogenesis and metabolic hyperactivation limits the application of MTT assay in the estimation of radiation induced growth inhibition. *Scientific Reports*, *8*, 1531.
- Rajagopal, S., Ajaya kumar, R., Deevi, D. S., Satyanarayana, C., & Rajagopalan, R. (2003). Andrographolide, a potential cancer therapeutic agent isolated from *Andrographis paniculata*. *Journal of Experimental Therapeutics and Oncology*, *3*, 147-158.
- Rajakulendran, T., Sahmi, M., Lefrançois, M., Sicheri, F., & Therrien, M. (2009). A dimerization dependent mechanism drives RAF catalytic activation. *Nature*, *461*, 542-545.



- Rajalingam, K., Schreck, R., Rapp, U. R., & Albert, Š. (2007). Ras oncogenes and their downstream targets. *Biochimica et Biophysica Acta - Molecular Cell Research*, 1773, 1177-1195.
- Rajput, A., San Martin, I. D., Rose, R., Beko, A., LeVea, C., Sharratt, E., . . . Wang, J. (2008). Characterization of HCT116 human colon cancer cells in an orthotopic model. *Journal of Surgical Research*, 147, 276-281.
- Rapp, U. R., Goldsborough, M. D., Mark, G. E., Bonner, T. I., Groffen, J., Jr. Reynolds, F. H., & Stephenson, J. R. (1983). Structure and biological activity of v-raf, a unique oncogene transduced by a retrovirus. *Proceedings of the National Academy of Sciences of the United States of America*, 80, 4218-4222.
- Ratain, M. J., & Plunkett, W.K. Jr. (2003). Principles of Pharmacokinetics. In D. W. Kufe, R. E. Pollock, R. R. Weichselbaum, C. B. Robert , T. S. Gansler, J. F. Holland, & E. Frei (Eds.), *Holland-Frei Cancer Medicine (6th edition ed.)*. Hamilton (ON): BC Decker. Retrieved from: <https://www.ncbi.nlm.nih.gov/books/NBK12815/>
- Redza-Dutordoir, M., & Averill-Bates, D. A. (2016). Activation of apoptosis signalling pathways by reactive oxygen species. *Biochimica et Biophysica Acta*, 1863, 2977-2992.
- Reid, T. S., Terry, K. L., Casey, P. J., & Beese, L. S. (2004). Crystallographic analysis of CaaX prenyltransferases complexed with substrates defines rules of protein substrate selectivity. *Journal of Molecular Biology*, 343, 417-433.
- Reiss, Y., Goldstein , J. L., Seabra, M. C., Casey, P. J., & Brown, M. S. (1990). Inhibition of purified p21ras farnesyl:protein transferase by Cys-AAX tetrapeptides. *Cell*, 62, 81-88.
- Ren, K., Zhang, Z., Li, Y., Liu, J., Zhao, D., Zhao, Y., & Gong, T. (2009). Physicochemical characteristics and oral bioavailability of andrographolide complexed with hydroxypropyl- $\beta$ -cyclodextrin. *Pharmazie*, 64, 515-520.
- Resat, H., Straatsma, T. P., Dixon, D. A., & Miller, J. H. (2001). The arginine finger of RasGAP helps Gln-61 align the nucleophilic water in GAP-stimulated hydrolysis of GTP. *Proceedings of the National Academy of Sciences of the United States of America*, 98, 6033-6038.
- Reva, B., Antipin, Y., & Sander, C. (2011). Predicting the functional impact of protein mutations: Application to cancer genomics. *Nucleic Acids Research*, 39, e118.

- Ricci, M. S., & Zong, W. (2006). Chemotherapeutic approaches for targeting cell death pathways. *Oncologist*, *11*, 342-357.
- Ricci-Vitiani, L., Lombardi, D. G., Pilozzi, E., Biffoni, M., Todaro, M., Peschle, C., & De Maria, R. (2007). Identification and expansion of human colon-cancer-initiating-cells. *Nature*, *445*, 111-115.
- Rizk, M. L., Zou, L., Savic, R. M., & Dooley, K. E. (2017). Importance of drug pharmacokinetics at the site of action. *Clinical and Translational Science*, *10*, 133-142.
- Roberts, M. S., Magnusson, B. M., Burczynski, F. J., & Weiss, M. (2002). Enterohepatic Circulation: Physiological, pharmacokinetic and clinical implications. *Clinical Pharmacokinetics*, *41*, 751-790.
- Robey, R. W., Pluchino, K. M., Hall, M. D., Fojo, A. T., Bates, S. E., & Gottesman, M. M. (2018). Revisiting the role of ABC transporters in multidrug-resistant cancer. *Nature Reviews Cancer*, *18*, 452-464.
- Rodriguez-Viciano, P., Sabatier, C., & McCormick, F. (2004). Signaling specificity by Ras family GTPases is determined by the full spectrum of effectors they regulate. *Molecular and Cellular Biology*, *24*, 4943-4954.
- Roidl, A., Berger, H. J., Kumar, S., Bange, J., Knyazev, P., & Ullrich, A. (2009). Resistance to chemotherapy is associated with fibroblast growth factor receptor 4 up-regulation. *Clinical Cancer Research*, *15*, 2058-2066.
- Rojas, J., & Santos, E. (2006). Ras-Gefs and Ras Gaps. In C. Der (Ed.), *RAS Family GTPases* (pp. 15-43). Berlin: Springer.
- Rosenberg, S. A., Tran, E., & Robbins, P. F. (2017). T-cell transfer therapy targeting mutant KRAS. *The New England Journal of Medicine*, *376*, e11.
- Rosnizek, I. C., Graf, T., Spoerner, M., Tränkle, J., Filchtinski, D., Herrmann, C., . . . Kalbitzer, H. R. (2010). Stabilizing a weak binding state for effectors in the human ras protein by cyclen complexes. *Angewandte Chemie International Edition in English*, *49*, 3830-3833.
- Rosnizek, I. C., Spoerner, M., Harsch, T., Kreitner, S., Filchtinski, D., Herrmann, C., . . . Kalbitzer, H. R. (2012). Metal-bis(2-picolyl)-amine complexes as state 1(T) inhibitors of activated ras protein. *Angewandte Chemie International Edition in English*, *51*, 10647-10651.

- Rubinstein, A. D., Eisenstein, M., Ber, Y., Bialik, S., & Kimchi, A. (2011). The autophagy protein Atg12 associates with antiapoptotic Bcl-2 family members to promote mitochondrial apoptosis. *Molecular Cell*, *44*, 698-709.
- Rushworth, L. K., Hindley, A. D., O'Neill, E., & Kolch, W. (2006). Regulation and role of Raf-1/B-Raf heterodimerization. *Molecular and Cellular Biology*, *26*, 2262-2272.
- Russo, G., Zegar, C., Giordano, A. (2003). Advantages and limitations of microarray technology in human cancer. *Oncogene*, *22*, 6497-6507.
- Rygaard, J. (1991). Subcutaneous Xenografts. In E. Boven, & B. Winograd (Eds.), *The Nude Mouse in Oncology Research* (pp. 43-50). Boca Raton: CRC Press, Inc.
- Rygaard, J., & Povlsen, C. O. (1969). Heterotransplantation of a human malignant tumour to "nude" mice. *Acta Path et Microbiol Scand*, *77*, 758-760.
- Ryter, S. W., Kim, H. P., Hoetzel, A., Park, J. W., Nakahira, K., Wang, X., & Choi, A. M. (2007). Mechanisms of cell death in oxidative stress. *Antioxidants & Redox Signaling*, *9*, 49-89.
- Sagineedu, S. R. (2011). *Semisynthesis of selected andrographolide derivatives and in vitro evaluation for cytotoxic and antiangiogenic properties*. (Unpublished PhD's dissertation). Universiti Putra Malaysia, Malaysia.
- Sakai, J. B. (2008). *Practical Pharmacology for the Pharmacy Technician*. Philadelphia (PA): Lippincott Williams & Wilkins.
- Sakamoto, K., Kamada, Y., Sameshima, T., Yaguchi, M., Niida, A., Sasaki, S., . . . Tani, A. (2017). K-Ras(G12D)-selective inhibitory peptides generated by random peptide T7 phage display technology. *Biochemical and Biophysical Research Communications*, *484*, 605-611.
- Salaroglio, I. C., Mungo, E., Gazzano, E., Kopecka, J., & Riganti, C. (2019). ERK is a pivotal player of chemo-immune-resistance in cancer. *International Journal of Molecular Sciences*, *20*, 2505.
- Samid, D., Miller, A. C., Rimoldi, D., Gafner, J., & Clark, E. P. (1991). Increased radiation resistance in transformed and nontransformed cells with elevated ras proto-oncogene expression. *Radiation Research*, *126*, 244-250.
- Sanchez, D., & Cheung, W. L. (2015). Pathology of pancreatic tumors. *Translational Cancer Research*, *4*, 608-615.

- Sartippour, M. R., Pietras, R., Marquez-Garban, M. C., Chen, H., Heber, D., Henning, S. M., ... Brooks, M. N. (2006). The combination of green tea and tamoxifen is effective against breast cancer. *Carcinogenesis*, *27*, 2424-2433.
- Satyanarayanajois, S. D., & Hill, R. A. (2011). Medicinal chemistry for 2020. *Future Medicinal Chemistry*, *3*, 1765-1786.
- Scheffzek, K., Ahmadian, M. R., Kabsch, W., Wiesmüller, L., Lautwein, A., Schmitz, F., & Wittinghofer, A. (1997). The Ras-RasGAP complex: Structural basis for GTPase activation and its loss in oncogenic Ras mutants. *Science*, *277*, 333-338.
- Seabra, M. C., Reiss, Y., Casey, P. J., Brown, M. S., & Goldstein, J. L. (1991). Protein farnesyltransferase and geranylgeranyltransferase share a common alpha subunit. *Cell*, *65*, 429-434.
- Seeburg, P. H., Colby, W. W., Capon, D. J., Goeddel, D. V., & Levinson, A. D. (1984). Biological properties of human c-Ha-ras1 genes mutated at codon 12. *Nature*, *312*, 71-75.
- Shah, A. N., Summy, J. M., Zhang, J., Park, S. I., Parikh, N. U., & Gallick, G. E. (2007). Development and characterization of gemcitabine-resistant pancreatic tumor cells. *Annals of Surgical Oncology*, *14*, 3629-3637.
- Shahinian, S., & Silviu, J. R. (1995). Doubly-lipid-modified protein sequence motifs exhibit long-lived anchorage to lipid bilayer membranes. *Biochemistry*, *34*, 3813-3822.
- Shargel, L., Wu-Pong, S., & Yu, A. B. (2012). Chapter 9. Nonlinear Pharmacokinetics. In *Applied Biopharmaceutics & Pharmacokinetics* (pp. 177-203). New York: The McGraw-Hill Companies, Inc.
- Shen, K., Ji, L., Lu, B., Xu, C., Gong, C., Morahan, G., & Wang, Z. (2014). Andrographolide inhibits tumor angiogenesis via blocking VEGFA/VEGFR2-MAPKs signaling cascade. *Chemico-Biological Interactions*, *218*, 99-106.
- Shima, F., Yoshikawa, Y., Ye, M., Araki, M., Matsumoto, S., Liao, J., . . . Kataoka, T. (2013). *In silico* discovery of small-molecule ras inhibitors that display antitumor activity by blocking the ras-effector interaction. *Proceedings of the National Academy of Sciences of the United States of America*, *110*, 8182-8187.
- Shimizu, K., Goldfarb, M., Perucho, M., & Wigler, M. (1983). Isolation and preliminary characterization of the transforming gene of a human

neuroblastoma cell line. *Proceedings of the National Academy of Sciences of the United States of America*, 80, 383-387.

- Shoemaker, R. H. (2006). The NCI60 human tumour cell line anticancer drug screen. *Nature Reviews Cancer*, 6, 813-823.
- Silvius, J. R., Bhagatji, P., Leventis, R., & Terrone, D. (2006). K-ras4B and prenylated proteins lacking "second signals" associate dynamically with cellular membranes. *Molecular Biology of the Cell*, 17, 192-202.
- Simanshu, D. K., Nissley, D. V., & McCormick, F. (2017). RAS proteins and their regulators in human disease. *Cell*, 170, 17-33.
- Simons, B. W., & Brayton, C. (2017). Challenges and limitations of mouse xenograft models of cancer. In R. Uthamanthil and P. Tinkey (Ed.), *Patient Derived Tumor Xenograft Models: Promise, Potential and Practice* (pp. 25-36). Amsterdam: Elsevier Inc.
- Singh, S. S. (2006). Preclinical pharmacokinetics: An approach towards safer and efficacious drugs. *Current Drug Metabolism*, 7, 165-182.
- Singh, A. K., Arya, R. K., Maheswari, S., Singh, A., Meena, S., Pandey, P., . . . Datta, D. (2015). Tumor heterogeneity and cancer stem cell paradigm: Updates in concept, controversies and clinical relevance. *International Journal of Cancer*, 136, 1991-2000.
- Sirion, U., Kasemsook, S., Suksen, K., Piyachaturawat, P., Suksamrarn, A., & Saeeng, R. (2012). New substituted C-19-andrographolide analogues with potent cytotoxic activities. *Bioorganic & Medicinal Chemistry Letters*, 22, 49-52.
- Siripong, P., Kongkathip, B., Preechanukool, K., Picha, P., Tunsuwan, K., & Taylor, W. C. (1992). Cytotoxic diterpenoid constituents from *Andrographis paniculata* Nees. leaves. *Journal of The Science Society of Thailand*, 18, 187-194.
- Smith, P. K., Krohn, R. I., Hermanson, G. T., Mallia, A. K., Gartner, F. H., Provenzano, M. D., . . . Klenk, D. C. (1985). Measurement of protein using bicinchoninic acid. *Analytical Biochemistry*, 150, 76-85.
- Soltany-Rezaee-Rad, M., Mottaghi-Dastjerdi, N., Setayesh, N., Roshandel, G., Ebrahimifard, F., Sepehrizadeh, Z. (2014). Overexpression of FOXO3, MYD88, and GAPDH identified by suppression subtractive hybridization in esophageal cancer is associated with autophagy. *Gastroenterology Research and Practice*, 2014, 185035.

- Stanslas, J., Liew, P., Lftlkhar, N., Lee, C., Saad, S., Lajis, N., . . . Bibby, M. (2001). Potential of AG in the treatment of breast cancer. *European Journal of Cancer*, *37*, 169.
- Stratton, M. R., Campbell, P. J., & Futreal, P. A. (2009). The cancer genome. *Nature*, *458*, 719-724.
- Sui, X., Chen, R., Wang, Z., Huang, Z., Kong, N., Zhang, M., . . . Pan, H. (2013). Autophagy and chemotherapy resistance: a promising therapeutic target for cancer treatment. *Cell Death & Disease*, *4*, e838.
- Sukardiman, H., Widyawaruyanti, A., Sismindari, & Zaini, N. C. (2007). Apoptosis inducing effect of andrographolide on TD-47 human breast cancer cell line. *African Journal of Traditional, Complementary and Alternative Medicines*, *4*, 345-351.
- Sun, Q., Burke, J. P., Phan, J., Burns, M. C., Olejniczak, E. T., Waterson, A. G., . . . Fesik, S. W. (2012). Discovery of small molecules that bind to K-Ras and inhibit Sos-mediated activation. *Angewandte Chemie International Edition*, *51*, 6140-6143.
- Sun, W. L., Chen, J., Wang, Y. P., & Zheng, H. (2011). Autophagy protects breast cancer cells from epirubicin-induced apoptosis and facilitates epirubicin-resistance development. *Autophagy*, *7*, 1035-1044.
- Suttie, A. W. (2006). Histopathology of the spleen. *Toxicol. Pathol.*, *34*, 466-503.
- Swarthout, J. T., Lobo, S., Farh, L., Croke, M. R., Greentree, W. K., Deschenes, R. J., & Linder, M. E. (2005). DHHC9 and GCP16 constitute a human protein fatty acyltransferase with specificity for H- and N-Ras. *Journal of Biological Chemistry*, *280*, 31141-31148.
- Szadvari, I., Krizanova, O., & Babula, P. (2016). Athymic nude mice as an experimental model for cancer treatment. *Physiological Research*, *65*, S441-S453.
- Szklarczyk, D., Gable, A. L., Lyon, D., Junge, A., Wyder, S., Huerta-Cepas, J., . . . von Mering, C. (2019). STRING v11: protein-protein association networks with increased coverage, supporting functional discovery in genome-wide experimental datasets. *Nucleic Acids Research*, *47*, D607-D613.
- Tai, S., Sun, Y., Squires, J. M., Zhang, H., Oh, W. K., Liang, C., & Huang, J. (2011). PC3 is a cell line characteristic of prostatic small cell carcinoma. *Prostate*, *71*, 1668-1679.

- Takai, Y., Sasaki, T., & Matozaki, T. (2001). Small GTP-binding proteins. *Physiological Reviews*, 81, 153-208.
- Tan, M. S. (2017). *Cytotoxicity of SRJ23 and its derivatives, expression and SRJ23 binding of KRAS G12V oncoprotein*. (Unpublished Master's thesis). Universiti Putra Malaysia, Malaysia.
- Teicher, B. A. (2006). Tumor models for efficacy determination. *Molecular Cancer Therapeutics*, 5, 2435-2443.
- Thorpe, L. M., Yuzugullu, H., & Zhao, J. J. (2015). PI3K in cancer: divergent roles of isoforms, modes of activation, and therapeutic targeting. *Nature Reviews Cancer*, 15, 7-24.
- Tiong, K. H., Tan, B. S., Choo, H. L., Chung, F. F., Hii, L., Tan, S. H., . . . Leong, C. (2016). Fibroblast growth factor receptor 4 (FGFR4) and fibroblast growth factor 19 (FGF19) autocrine enhance breast cancer cells survival. *Oncotarget*, 7, 57633-57650.
- Tong, J., Sun, D., Yang, C., Wang, Y., Sun, S., Li, Q., . . . Liu, Y. (2016). Serum starvation and thymidine double blocking achieved efficient cell cycle synchronization and altered the expression of p27, p53, bcl-2 in canine breast cancer cells. *Research in Veterinary Science*, 105, 10-14.
- Toutain, P. L., & Bousquet-Mélou, A. (2004). Plasma terminal half-life. *Journal of Veterinary Pharmacology and Therapeutics*, 27, 427-439.
- Tu, H., Wen, C. P., Tsai, S. P., Chow, W., Wen, C., Ye, Y., . . . Wu, X. (2017). Cancer risk associated with chronic diseases and disease markers: prospective cohort study. *British Medical Journal*, 360, k134.
- Tuckerman, M., Berne, B. J., & Martyna, G. J. (1992). Reversible multiple time scale molecular dynamics. *The Journal of Chemical Physics*, 97, 1990-2001.
- Turkington, R. C., Longley, D. B., Allen, W. L., Stevenson, L., McLaughlin, K., Dunne, P. D., . . . Johnston, P. G. (2014). Fibroblast growth factor receptor 4 (FGFR4): a targetable regulator of drug resistance in colorectal cancer. *Cell Death & Disease*, 5, e1046.
- Turner, P. V., Brabb, T., Pekow, C., & Vasbinder, M. A. (2011). Administration of substances to laboratory animals: Routes of administration and factors to consider. *Journal of the American Association for Laboratory Animal Science*, 50, 600-613.
- Tzivion, G., Luo, Z., & Avruch, J. (1998). A dimeric 14-3-3 protein is an essential cofactor for Raf kinase activity. *Nature*, 394, 88-92.

- Ubezio, P. (2019). Beyond the T/C ratio: Old and new anticancer activity scores *in vivo*. *Cancer Management and Research*, *11*, 8529-8538.
- Uline, M. J., & Corti, D. S. (2013). Molecular dynamics at constant pressure: Allowing the system to control volume fluctuations *via* a "shell" particle. *Entropy*, *15*, 3941-3969.
- Vasan, N., Baselga, J., & Hyman, D. M. (2019). A view on drug resistance in cancer. *Nature*, *575*, 299-309.
- Veerman, G., Ruiz van Haperen, V. W., Vermorken, J. B., Noordhuis, P., Braakhuis, B. J., Pinedo, H. M., & Peters, G. J. (1996). Antitumor activity of prolonged as compared with bolus administration of 2',2'-difluorodeoxycytidine *in vivo* against murine colon tumors. *Cancer Chemotherapy and Pharmacology*, *38*, 335-342.
- Vetter, I. R., & Wittinghofer, A. (2001). The guanine nucleotide-binding switch in three dimensions. *Science*, *294*, 1299-1304.
- Vicente-Dueñas, C., Romero-Camarero, I., Cobaleda, C., & Sánchez-García, I. (2013). Function of oncogenes in cancer development: a changing paradigm. *EMBO Journal*, *32*, 1502-1513.
- Vigelsø, A., Dybboe, R., Hansen, C. N., Dela, F., Helge, J. W., Guadalupe, G. A. (2015). GAPDH and  $\beta$ -actin protein decreases with aging, making Stain-Free technology a superior loading control in Western blotting of human skeletal muscle. *Journal of Applied Physiology*, *118*, 386.
- Vogt, P. K. (2012). Retroviral oncogenes: A historical primer. *Nature Reviews Cancer*, *12*, 639-648.
- Voskoglou-Nomikos, T., Pater, J. L., & Seymour, L. (2003). Clinical predictive value of the *in vitro* cell line, human xenograft, and mouse allograft preclinical cancer models. *Clinical Cancer Research*, *9*, 4227-4239.
- Wang, D., Zhang, J., Li, Z., Han, J., Gao, Y., Chen, M., & Li, Y. (2019). Upregulation of fibroblast growth factor 19 is associated with the initiation of colorectal adenoma. *Digestive Diseases*, *37*, 214-225.
- Wang, W., Guo, W., Li, L., Fu, Z., Liu, W., Gao, J., . . . Gu, Y. (2016). Andrographolide reversed 5-FU resistance in human colorectal cancer by elevating BAX expression. *Biochemical Pharmacology*, *121*, 8-17.
- Wang, X., Zhang, H., & Chen, X. (2019). Drug resistance and combating drug resistance in cancer. *Cancer Drug Resistance*, *2*, 141-160.



- Wang, Z., Kang, Y., Yang, X., Wang, J., Zhang, Q., Yang, B., . . . Sun, X. (2016). Andrographolide radiosensitizes human esophageal cancer cell line ECA109 to radiation *in vitro*. *Diseases of the Esophagus*, 29, 54-61.
- Was, H., Cichon, T., Smolarczyk, R., Rudnicka, D., Stopa, M., Chevalier, C., . . . Jozkowicz, A. (2006). Overexpression of heme oxygenase-1 in murine melanoma: Increased proliferation and viability of tumor cells, decreased survival of mice. *American Journal of Clinical Pathology*, 169, 2181-2198.
- Waters, A. M., & Der, C. J. (2018). KRAS: The critical driver and therapeutic target for pancreatic cancer. *Cold Spring Harbor Perspectives in Medicine*, 8, a031435.
- Watt, J., & Kocher, H. M. (2013). The desmoplastic stroma of pancreatic cancer is a barrier to immune cell infiltration. *Oncoimmunology*, 2, e26788.
- Weaver, C. H. (2021, March 26). *CancerConnect*. Retrieved from Sotorasib (AMG 510) Precision Cancer Medicine Targets KRAS NSCLC: KRAS a major driver of lung and other cancers for KRAS G12C mutated NSCLC - what patients need to know: <https://news.cancerconnect.com/lung-cancer/sotorasib-amg-510-precision-cancer-medicine-targets-kras-nsclc>
- Wee, P., & Wang, Z. (2017). Epidermal growth factor receptor cell proliferation signaling pathways. *Cancers*, 9, 52.
- Weir, H. K., Thompson, T. D., Soman, A., Møller, B., & Leadbetter, S. (2015). The past, present, and future of cancer incidence in the United States: 1975 through 2020. *Cancer*, 121, 1827-1837.
- Wennerberg, K., Rossman, K. L., & Der, C. J. (2005). The Ras superfamily at a glance. *Journal of Cell Science*, 118, 843-846.
- Willard-Mack, C. L., Elmore, S. A., Hall, W. C., Harleman, J., Kuper, C. F., Losco, P., . . . Keenan, C. M. (2019). Nonproliferative and proliferative lesions of the rat and mouse hematolymphoid system. *Toxicologic Pathology*, 47, 665-783.
- Wilson, J. S., Pirola, R. C., & Apte, M. V. (2014). Stars and stripes in pancreatic cancer: role of stellate cells and stroma in cancer progression. *Frontiers in Physiology*, 5, 11 pages.
- Wlodawer, A., Minor, Q., Dauter, Z., & Jaskolski, M. (2008). Protein crystallography for non-crystallographers, or how to get the best

(but not more) from published macromolecular structures. *The FEBS Journal*, 275, 1-21.

- Wong, C. C. (2015). *Elucidation of the mechanism of anticancer activity of 3,19-(2-bromobenzylidene)andrographolide (SRJ09) in breast and colon cancer cells*. (Unpublished PhD thesis). Universiti Putra Malaysia, Malaysia.
- Wong, C. C., Lim, S. H., Sagineedu, S. R., Lajis, N., & Stanslas, J. (2016). SRJ09, a promising anticancer drug lead: Elucidation of mechanisms of antiproliferative and apoptogenic effects and assessment of *in vivo* antitumor efficacy. *Pharmacological Research*, 107, 66-78.
- Wong, C. C., Sagineedu, S. R., Sumon, S. H., Sidik, S. M., Phillips, R., Lajis, N. H., & Stanslas, J. (2014). NCI *in vitro* and *in silico* anticancer screen, cell cycleperturbation and apoptosis-inducing potential of new acylated, benzylidene and isopropylidenederivatives of andrographolide. *Environmental Toxicology and Pharmacology*, 38, 489-501.
- Wong, H. C. (2013). *In vitro growth inhibition, molecular mechanisms of cell cycle arrest and apoptosis in prostate cancer cells by SRJ23*. (Unpublished Master's thesis). Universiti Putra Malaysia, Malaysia.
- Wong, H. C., Sagineedu, S. R., Lajis, N. H., Loke, S. C., & Stanslas, J. (2011). Andrographolide induces cell cycle arrest and apoptosis in PC-3 prostate cancer cells. *African Journal of Pharmacy and Pharmacology*, 5, 225-233.
- Wong, H. C., Wong, C. C., Sagineedu, S. R., Loke, S. C., Lajis, N. H., & Stanslas, J. (2014). SRJ23, a new semisynthetic andrographolide derivative: *In vitro* growth inhibition and mechanisms of cell cycle arrest and apoptosis in prostate cancer cells. *Cell Biology and Toxicology*, 30, 269-288.
- Wong, H., Choo, E. F., Alicke, B., Ding, X., La, H., McNamara, E., . . . Gould, S. E. (2012). Antitumor activity of targeted and cytotoxic agents in murine subcutaneous tumor models correlates with clinical response. *Clinical Cancer Research*, 18, 10 pages.
- Wu, F., Zhang, Z., & Ding, H. (2006). Simple high-performance liquid chromatographic method for the determination of tetramethylpyrazine phosphate in very small volumes of dog plasma: Application to a pharmacokinetic study. *Journal of Chromatographic Science.*, 44, 13-17.
- Wu, Q., Wang, R., Yang, Q., Hou, X., Chen, S., Hou, Y., . . . Wang, Z. (2013). Chemoresistance to gemcitabine in hepatoma cells induces

epithelial-mesenchymal transition and involves activation of PDGF-D pathway. *Oncotarget*, 4, 1999-2009.

- Xia, Y. F., Ye, B. Q., Li, Y. D., Wang, J. G., He, X. J., Lin, X., . . . Geng, J. G. (2004). Andrographolide attenuates inflammation by inhibition of NF-kappa B activation through covalent modification of reduced cysteine 62 of p50. *The Journal of Immunology*, 173, 4207-4217.
- Xie, M. H., Holcomb, I., Deuel, B., Dowd, P., Huang, A., Vagts, A., . . . Gurney, A. L. (1999). FGF-19, a novel fibroblast growth factor with unique specificity for FGFR4. *Cytokine*, 11, 729-735.
- Xiong, Y., Lu, J., Hunter, J., Li, L., Scott, D., Choi, H. G., . . . Gray, N. S. (2016). Covalent guanosine mimetic inhibitors of G12C KRAS. *ACS Medicinal Chemistry Letters*, 8, 61-66.
- Yamamoto, S., Tomita, Y., Hoshida, Y., Morooka, T., Nagano, H., Dono, K., . . . Aozasa, K. (2004). Prognostic significance of activated Akt expression in pancreatic ductal adenocarcinoma. *Clinical Cancer Research*, 10, 2846-2850.
- Yan, J., Roy, S., Apolloni, A., Lane, A., & Hancock, J. F. (1998). Ras isoforms vary in their ability to activate Raf-1 and phosphoinositide 3-kinase. *Journal of Biological Chemistry*, 273, 24052-24056.
- Yang, A. D., Fan, F., Camp, E. R., van Buren, G., Liu, W., Somcio, R., . . . Ellis, L. M. (2006). Chronic oxaliplatin resistance induces epithelial-to-mesenchymal transition in colorectal cancer cell lines. *Clinical Cancer Research*, 12, 4147-4153.
- Yang, N. J., & Hinner, M. J. (2015). Getting across the cell membrane: An overview for small molecules, peptides, and proteins. *Methods in Molecular Biology*, 1266, 29-53.
- Yang, P. Y., Hsieh, P. L., Wang, T. H., Yu, C. C., Lu, M. Y., Liao, Y. W., . . . Peng, C. Y. (2017). Andrographolide impedes cancer stemness and enhances radio-sensitivity in oral carcinomas *via* miR-218 activation. *Oncotarget*, 8, 4196-4207.
- Yang, S., Evens, A. M., Prachand, S., Singh, A. T., Bhalla, S., David, K., & Gordon, L. I. (2010). Mitochondrial-mediated apoptosis in lymphoma cells by the diterpenoid lactone andrographolide, the active component of *Andrographis paniculata*. *Clinical Cancer Research*, 16, 4755-4768.
- Yang, T., Sheng, H., Feng, N., Wei, H., Wang, Z., & Wang, C. (2013). Preparation of andrographolide-loaded solid lipid nanoparticles and their *in vitro* and *in vivo* evaluations: characteristics, release,

- absorption, transports, pharmacokinetics, and antihyperlipidemic activity. *Journal of Pharmaceutical Sciences*, 102, 4414-4425.
- Yang, Z., & Klionsky, D. J. (2010). Mammalian autophagy: core molecular machinery and signaling regulation. *Current Opinion in Cell Biology*, 22, 124-131.
- Ye, L., Wang, T., Tang, L., Liu, W., Yang, Z., Zhou, J., . . . Liu, Z. (2011). Poor oral bioavailability of a promising anticancer agent andrographolide is due to extensive metabolism and efflux by P-glycoprotein. *Journal of Pharmaceutical Sciences*, 100, 5007-5017.
- Yu, C., Chen, C., Fu, S., Lin, H., Lee, M., Chiou, W., . . . Hung, S. (2018). Andrographolide enhances the anti-metastatic effect of radiation in Ras-transformed cells *via* suppression of ERK-mediated MMP-2 activity. *PLoS ONE*, 13, e0205666.
- Yu, J. S., & Cui, W. (2016). Proliferation, survival and metabolism: the role of PI3K/AKT/mTOR signalling in pluripotency and cell fate determination. *Development*, 143, 3050-3060.
- Yu, J., Zhang, Y., McIlroy, J., Rordorf-Nikolic, T., Orr, G. A., & Backer, J. M. (1998). Regulation of the p85/p110 phosphatidylinositol 3'-kinase: stabilization and inhibition of the p110 $\alpha$  catalytic subunit by the p85 regulatory subunit. *Molecular and Cellular Biology*, 18, 1379-1387.
- Yuan, J., Ng, W. H., Tian, Z., Yap, J., Baccharini, M., Chen, Z., & Hu, J. (2018). Activating mutations in MEK1 enhance homodimerization and promote tumorigenesis. *Science Signaling*, 11, eaar6795.
- Yuan, S., Norgard, R. J., & Stanger, B. Z. (2019). Cellular plasticity in cancer. *Cancer Discovery*, 9, 837-851.
- Yudushkin, I. (2019). Getting the Akt together: Guiding intracellular Akt activity by PI3K. *Biomolecules*, 9, 67.
- Zaidieh, T., Smith, J. R., Ball, K. E., & An, Q. (2019). ROS as a novel indicator to predict anticancer drug efficacy. *BMC Cancer*, 19, 1224.
- Zamek-Gliszczynski, M. J., Hoffmaster, K. A., Nezasa, K., Tallman, M. N., & Brouwer, K. L. (2006). Integration of hepatic drug transporters and phase II metabolizing enzymes: Mechanisms of hepatic excretion of sulfate, glucuronide, and glutathione metabolites. *European Journal of Pharmaceutical Sciences*, 27, 447-486.
- Zang, M., Gong, j., Luo, L., Zhou, J., Xiang, X., Huang, W., . . . Luo, Z. (2008). Characterization of Ser338 phosphorylation for Raf-1 activation. *Journal of Biological Chemistry*, 283, 31429-31437.

- Zeitouni, D., Pylyayeva-Gupta, Y., Der, C. J., & Bryant, K. L. (2016). KRAS mutant pancreatic cancer: No lone path to an effective treatment. *Cancers*, 8, 45.
- Zenonos, K., & Kyprianou, K. (2013). RAS signaling pathways, mutations and their role in colorectal cancer. *World Journal of Gastrointestinal Oncology*, 5, 97-101.
- Zhang, X. F., Settleman, J., Kyriakis, J. M., Takeuchi-Suzuki, E., Elledge, S. J., Marshall, M. S., . . . Avruch, J. (1993). Normal and oncogenic p21ras proteins bind to the amino-terminal regulatory domain of c-Raf-1. *Nature*, 364, 308-313.
- Zhang, X., Ibrahim, O. A., Olsen, S. K., Umemori, H., Mohammadi, M., & Ornitz, D. M. (2006). Receptor specificity of the fibroblast growth factor family. The complete mammalian FGF family. *Journal of Biological Chemistry*, 281, 15694-15700.
- Zhang, X., Kong, M., Zhang, Z., Xu, S., Yan, F., Wei, L., Zhou, J., 2017. FGF19 genetic amplification as a potential therapeutic target in lung squamous cell carcinomas. *Thoracic Cancer*, 8, 655-665.
- Zhao, F., He, E. Q., Wang, L., & Liu, K. (2008). Anti-tumor activities of andrographolide, a diterpene from *Andrographis paniculata*, by inducing apoptosis and inhibiting VEGF level. *Journal of Asian Natural Products Research*, 10, 467-473.
- Zhao, G., Zeng, Q., Zhang, S., Zhong, Y., Wang, C., Chen, Y., . . . Liao, Z. (2019). Effect of carrier lipophilicity and preparation method on the properties of andrographolide-solid dispersion. *Pharmaceutics*, 11, 74.
- Zhe, N., Wang, J., Chen, S., Lin, X., Chai, Q., Zhang, Y., . . . Fang, Q. (2015). Heme oxygenase-1 plays a crucial role in chemoresistance in acute myeloid leukemia. *Hematology*, 20, 384-391.
- Zhou, S. F. (2008). Drugs behave as substrates, inhibitors and inducers of human cytochrome P450 3A4. *Current Drug Metabolism*, 9, 310-322.
- Zhou, Y., Prakash, P., Gorfe, A. A., & Hancock, J. F. (2018). Ras and the plasma membrane: A complicated relationship. *Cold Spring Harbor Perspectives in Medicine*, 8, a031831.
- Zhu, X., Li, W., Ma, J., Zhang, Y., Liu, R., Wu, W., . . . Wang, S. (2015). Knockdown of heme oxygenase-1 promotes apoptosis and autophagy and enhances the cytotoxicity of doxorubicin in breast cancer cells. *Oncology Letters*, 10, 2974-2980.

- Zimmermann, G., Papke, B., Ismail, S., Vartak, N., Chandra, A., Hoffmann, M., . . . Waldmann, H. (2013). Small molecule inhibition of the KRAS-PDEdelta interaction impairs oncogenic KRAS signalling. *Nature*, *497*, 638-642.
- Zong, W., & Thompson, C. B. (2006). Necrotic death as a cell fate. *Genes & Development*, *20*, 1-15.

## BIODATA OF STUDENT

Quah Shun Ying was born in Penang, Malaysia on 12<sup>th</sup> November, 1991 as the eldest daughter in the family. She graduated from Penang Chinese Girls High School, Penang, Malaysia in 2008, and achieved her degree in Bachelor of Science (Chemistry and Biology) with Magna cum Laude in 2013 from Campbell University, Buies Creek, North Carolina, United States of America. During her degree study, she obtained a few awards including Book Prize Award, The Matthew Baillie Prize for Chemistry and The Samuel Martin Brown Prize for Biology.

Same year after graduation, she started work as a research analyst in AcuBiz Consulting Sdn. Bhd., Kuala Lumpur, Malaysia, mainly involved in collecting data needed for consulting projects through extensive secondary and primary research, analysing data collected and preparing project reports under the guidance of project supervisor. She then worked as a research assistant in Pharmacotherapeutics Lab, Department of Medicine, Faculty of Medicine and Health Sciences, Universiti Putra Malaysia (UPM), Malaysia.

In 2015, Shun Ying pursued her study further with a degree of Ph.D. in Pharmacology and Toxicology, under the supervision of Prof. Dr. Johnson Stanslas from the same faculty. Her research studies mainly involved cancer and molecular pharmacology and toxicology. During her course of Ph.D. study, she participated in several scientific conference events, including the 1<sup>st</sup> MACR Scientific Conference organised in MAHSA University, Selangor, Malaysia, of which she was awarded with MACR Student Award (3<sup>rd</sup> Prize). She also became the Assistant Secretary of The International Conference on Drug Discovery and Translational Medicine 2018 (ICDDTM '18) organized by the Malaysian Association for Cancer Research.

## LIST OF PUBLICATIONS

- Quah, S.Y.**, Tan, M.S., Teh, Y.H. & Stanslas, J. (2016). Pharmacological Modulation of oncogenic Ras by natural products and their derivatives: Renewed hope in the discovery of novel anti-Ras drugs. *Pharmacology & Therapeutics*, 162: 35-57.  
doi: 10.1016/j.pharmthera.2016.03.010
- Soo, H.L., **Quah, S.Y.**, Sulaiman, I., Sagineedu, S.R., Lim, J.C.W., Stanslas, J. (2019) Advances and challenges in developing andrographolide and its analogues as cancer therapeutic agents. *Drug Discovery Today*, 24: 1890-1898.  
doi: 10.1016/j.drudis.2019.05.017
- Quah, S.Y.**, Tan, M.S., Ho, K.L., Abdul Manan, N., Gorfe, A.A., Deb, P.K., ... Stanslas, J. (2020). *In silico* and saturation transfer difference NMR approaches to unravel the binding mode of an andrographolide derivative to K-Ras oncoprotein. *Future Medicinal Chemistry*, 12(18): 1611-1631.  
doi: 10.4155/fmc-2020-0104
- Quah, S.Y.**, Wong, C.C., Wong, H.C., Ho, K.L., Abdul Manan, N., Deb, P.K., ... Stanslas, J. (2021). Microarray-based identification of differentially expressed genes associated with andrographolide derivatives-induced resistance in colon and prostate cancer cell lines. *Toxicology and Applied Pharmacology*, 425: 115605.  
doi: 10.1016/j.taap.2021.115605





**UNIVERSITI PUTRA MALAYSIA**

**STATUS CONFIRMATION FOR THESIS / PROJECT REPORT  
AND COPYRIGHT**

**ACADEMIC SESSION :** First semester 2021/2022

**TITLE OF THESIS / PROJECT REPORT :**

ABROGATION OF ONCOGENIC K-RAS FUNCTION BY ANDROGRAPHOLIDE  
DERIVATIVES VIA *IN SILICO*, *IN VITRO*, AND *IN VIVO* APPROACHES

**NAME OF STUDENT :**

QUAH SHUN YING

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

1. This thesis/project report is the property of Universiti Putra Malaysia.
2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as:

\*Please tick (✓)

**CONFIDENTIAL**

(Contain confidential information under Official Secret Act 1972).

**RESTRICTED**

(Contains restricted information as specified by the organization/institution where research was done).

**OPEN ACCESS**

I agree that my thesis/project report to be published as hard copy or online open access.