

UNIVERSITI PUTRA MALAYSIA

CLONAL PROPAGATION AMENABILITY OF SELECTED ELITE (DxP) AND CLONAL TENERA ORTETS OF OIL PALM (Elaeis guineensis Jacq.)

SITI RAHMAH BINTI ABDUL RAHMAN

FBSB 2021 16



CLONAL PROPAGATION AMENABILITY OF SELECTED ELITE (DxP) AND CLONAL TENERA ORTETS OF OIL PALM (*Elaeis guineensis* Jacq.)



SITI RAHMAH BINTI ABDUL RAHMAN

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the Requirement for the Degree of Master of Science

November 2019

COPYRIGHT

All material contained within the thesis, including without limitatin text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

CLONAL PROPAGATION AMENABILITY OF SELECTED ELITE (DxP) AND CLONAL TENERA ORTETS OF OIL PALM (*Elaeis guineensis* Jacq.)

By

SITI RAHMAH ABDUL RAHMAN

November 2019

Chairman Faculty

: Associate Professor Mohd. Puad Abdullah, PhD : Biotechnology and Biomolecular Sciences

The scale up of high quality oil palm ramets production is a main priority for all commercial tissue culture laboratories. To date, oil palm cloning technology is generally inefficient with low embryogenesis rate, genotype dependency and insufficient elite ortets. As the trends in culture development for mass propagation remains unpredictable, a thorough investigation of the relevant process is immensely important. This study aims to evaluate and expound the performance of selected elite tenera ortets for large scale production specifically during the stages of callogenesis, embryogenesis and embryoids maintenance en route for enhanced oil palm yield. Leaf explants were cultured on Murashige and Skoog (1962) medium with the current standard production protocol. Clonal ortets yielded callus of 1,340 ± 149 lines significantly (F=7.46, P=0.014) which refers to the frequency of callus initiated from the edge of cultured immature leaflet explants at 0.52-fold higher compared to elite tenera (DxP) ortets at 698 \pm 182 lines. Besides, clonal ortets demonstrated shorter callus induction time than elite tenera ortets. The earliest callus formation for clonal and elite *tenera* ortets were observed after 93.90 ± 4.27 days and 107.00 ± 7.11 days respectively (F=2.73, df=1, 18; P>0.05). Embryoid lines achieved were significantly higher (F=38.00, P=0.00) in clonal ortets (794 \pm 108 lines) by 8.6-fold against elite tenera ortets (91.6 \pm 35.0 lines). In addition to that, differences of the embryoid line production rapidity between ortet types were found to be statistically significant (F=5.71, df=1, 18: P<0.05) in which clonal ortets developed embryoids faster by an average of 68 days earlier than the elite tenera. Statistical analysis of the data showed that there was positive correlation (r=0.69449, P<0.05) between the total of embryoid lines formed with the total production of callus derived from elite tenera and clonal ortets throughout the production process. The embryoid lines are represented as the frequency of embryoid developed from the callus which were observed as whitish in colour and appear with smooth surface. This indicates that for both types of ortet, the initiation of embryoid is proportionate to the production of callus lines (r=0.69449, P<0.05). The growth models for describing polyembryoids (PE) development for clonal and elite tenera ortets versus time of subculture or pattern were established. Meanwhile, the growth rate pattern of elite tenera ortets appeared to be non-uniformed,

or uncertainty in the culture growth development compared to the series of clonal ortets tested. The potential of ortets development therefore could be estimated from a logistic growth curve fitted through the PE culture growth (in weight, g) against the duration of week after the initial culture. The growth patterns of clonal ortets are observed to be more predictive which facilitate further growth prediction. Therefore, the clonal ortets offers the best source for oil palm propagation optimization towards upscaling the ramets production in commercial scales.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

KEBOLEH-TERIMAAN PEMBIAKAN KLON PADA ORTET ELIT (DxP) DAN KLON TENERA SAWIT (*Elaeis guineensis* Jacq.) TERPILIH

Oleh

SITI RAHMAH ABDUL RAHMAN

November 2019

Pengerusi Fakulti : Profesor Madya Mohd. Puad Abdullah, PhD : Bioteknologi dan Sains Biomolekul

Penghasilan ramet sawit berskala besar dan berkualiti menjadi keutamaan bagi setiap makmal kultur tisu komersial. Sehingga kini, proses pengklonan sawit secara amnya adalah tidak cekap, dengan kadar embriogenesis yang rendah, kebergantungan kepada genotip dan kekurangan sumber ortet terpilih. Memandangkan corak perkembangan bahan kultur ke arah penghasilan ramet sawit berskala besar masih tidak dapat dijangka, adalah amat penting untuk kajian terperinci dilakukan. Kajian ini bertujuan untuk menilai dan mendapatkan pemahaman terhadap prestasi ortet untuk pengeluaran berskala besar dengan penumpuan kajian pada peringkat pembentukan kalus, embrioid dan multiplikasi embrioid ke arah meningkatkan hasil minyak sawit. Eksplan daun telah dikultur ke dalam medium Murashige and Skoog (1962) menggunakan protokol pengeluaran piawai sedia ada. Ortet klonal secara signifikannya (F=7.46, P=0.014) menghasilkan bilangan kalus lebih tinggi sebanyak 0.52-ganda iaitu 1.340 \pm 149 baris berbanding ortet tenera elit (DxP) iaitu 698 ± 182 baris. Baris merujuk kepada kekerapan kalus terbentuk daripada pinggir eksplan daun muda yang dikultur. Selain itu, ortet klonal juga menunjukkan tempoh penghasilan kalus yang lebih pendek dengan penghasilan kalus terawal bagi ortet klonal adalah selepas 93.90 ± 4.27 hari dan ortet tenera elit pula hanya selepas 107.00 ± 7.11 hari (F=2.73, df=1, 18; P>0.05). Pencapaian pembentukan embrioid juga lebih tinggi secara signifikan (F=38.00, P=0.00) bagi ortet klonal (794 \pm 108 baris), iaitu 8.6-ganda berbanding ortet *tenera* elit (91.6 ± 35.0). Malah, perbezaan tempoh penghasilan embrioid di antara kedua-dua jenis ortet juga adalah signifikan (F=5.71, df=1, 18: P<0.05), iaitu ortet klonal dapat menghasilkan embrioid seawal 68 hari berbanding ortet tenera elit. Analisis statistik menunjukkan terdapat korelasi secara positif (r=0.69449, P<0.05) di antara bilangan embrioid yang dihasilkan, yang diwakili oleh struktur berwarna putih dan mempunyai permukaan yang licin serta terbentuk daripada kalus, berbanding dengan jumlah kalus dihasilkan daripada ortet elit tenera dan ortet klonal ketika proses pengeluaran. Ini menunjukkan bahawa pembentukan embrioid adalah berkadar langsung dengan pengeluaran kalus bagi kedua-dua jenis ortet (r=0.69449, P<0.05). Model pertumbuhan perkembangan poliembrioid untuk ortet tenera elit dan klonal terhadap masa subkultur atau corak pertumbuhan telah dibangunkan. Sementara itu, corak pertubuhan bagi ortet

tenera elit didapati tidak seragam berbanding ortet klonal yang dibuktikan lebih sekata. Justeru, potensi pertumbuhan ortet dapat dijangka berdasarkan lengkung pertumbuhan logistik dari pertumbuhan kultur PE (berat, g) dan tempoh kultur PE daripada permulaan kultur. Corak pertumbuhan ortet klonal didapati lebih dapat diramal yang memudahkan jangkaan pertumbuhan pada masa seterusnya. Justeru, ortet klonal adalah sumber terbaik bagi mengoptimumkan pembiakan ramet sawit ke arah meningkatkan pengeluaran ramet berskala komersial.

ACKNOWLEDGEMENT

'My Lord, increase me in knowledge..!'

Surah Taha (20: 114)

First and foremost, all the praises is due to Allah for giving me the strength and courage to complete this thesis and for granting me with beneficial knowledge and His abundant sustenance.

I would like to express my deepest and sincerest gratitude to my research supervisor, Associate Prof. Dr. Mohd. Puad Abdullah for his kindness and continuous guidance, assistance, advice and encouragement throughout this research. Similar thanks go to my co-supervisors, Associate Prof. Dr. Noor Azmi Shaharuddin and Dr. Ahmad Tarmizi Hashim for their constructive ideas and expertise.

My special appreciation extends to my beloved parents, Abdul Rahman Mat Zain and Siti Hajar Sulaiman, for their constant support, inspiration and prayers.

I wish to express my warm and sincere thanks to my mentor Tn. Hj. Zamzuri Ishak, a former tissue culturist from MPOB for his motivation and guidance. I am also indebted to my former boss, Tn. Hj. Mad Jais Che Yem for his unwavering faith in scientific research in general, and in my ability as a scientist.

Tributes are also due to Noor Idayu Tahir, Marina Roseli, Dalilah Abu Bakar, Asmahanim Amir, colleagues from both the Malaysian Palm Oil Board (MPOB) and the RISDA Tissue Culture Laboratories, and to friends and acquaintance whose names do not appear here due to space constraints who was involved in any way to the successful realization of this thesis.

This thesis submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Mohd. Puad bin Abdullah, PhD

Associate Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Chairman)

Noor Azmi bin Shaharuddin, PhD

Associate Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Member)

Ahmad Tarmizi bin Hashim, PhD

Principal Research Officer Advanced Biotechnology & Breeding Centre Malaysian Palm Oil Board (Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date: 14 January 2021

Declaration by graduate student

I hereby confirm that:

- This thesis is my original work;
- Quotations, illustration and citation have been dully referenced;
- This thesis has not been submitted previously or concurrently for other degree at any other institutions;
- Intellectual property from thesis and copyright of thesis are fully owned by • Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- Written permission must be obtain from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or electronic form) including books, journals, modules, proceeding, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- There is no plagiarism or data falsification / fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date:

Name and Matric No: Siti Rahmah binti Abdul Rahman, GS42360

Declaration by Members of Supervisory Committee

This is to confirm that:

- The research conducted and the writing of this thesis was under our supervision;
- Supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: Name of Chairman of Supervisory Committee:	Associate Professor Dr. Mohd. Puad Abdullah
Committee.	Associate Trofessor D1. World. Tudu Abdurian
Signature:	
Name of Member of	
Supervisory	
Committee:	Associate Professor Dr. Noor Azmi Shaharuddin
Signature:	
Name of Member of	
Supervisory	
Committee:	Dr. Ahmad Tarmizi Hashim

TABLE OF CONTENTS

AB AC AP DE LIS	PROV CLAF ST OF ST OF	K WLEDGEMENT	Page i iii v vi vii viii xii xiv xvi	
СН	APTE	CR		
1	INTI	RODUCTION	1	
-			-	
2	LITE	CRATURE REVIEW	3	
	2.1	Oil Palm	3	
		2.1.1 Origin	3	
		2.1.2 Biology of <i>Elaeis guineensis</i>	3	
	2.2	Research and Development (R&D)	4	
	2.3	Socio Economic Impact	5	
	2.4	Oil Palm Industry in Malaysia	7	
	2.5	Tissue Culture of Oil Palm	9	
		2.5.1 Oil Palm Clonal Material	9	
		2.5.2 Oil Palm Tissue Culture Protocols	11	
		2.5.2.1 Process of <i>In Vitro</i> Propagation	11	
		2.5.2.2 Ortet Selection	12	
		2.5.2.3 Tissue Culture Medium	12	
		2.5.2.4 Explant Culture	13	
		2.5.2.5 Callogenesis, Embryogenesis and Embryoids Multiplication	14	
		2.5.2.6 Shoot Development, Rooting and Hardening	15	
	2.6	Tissue Culture Database System (TCDS)	15	
	2.7	Field Performance of Clonal Materials	17	
	2.8	Polyembryoids Maintenance	18	
3		FE TENERA (SELECTED D x P) AND CLONAL ORTET	20	
		ENTIAL FOR CALLOGENESIS AND EMBRYOGENESIS		
		Introduction	20	
	3.2	Materials and Methods	21	

UI	ENIL	AL FOR CALLOGENESIS AND EMIBRYOGENESIS			
1	Introduction				
2	Materials and Methods				
	3.2.1	Source of Ortets	21		
	3.2.2	Ortet Sampling	23		
	3.2.3	Cabbage and Explant Cutting	24		
	3.2.4	Medium Preparation	25		
		3.2.4.1 Explant Culture and Callus Induction	25		
		3.2.4.2 Callus Maintenance	26		
	3.2.5	Callogenesis and Embryogenesis Stage	26		
	3.2.6	Tissue Culture Database System (TCDS)	28		
	3.2.7	Statistical Analysis	29		

(C)

	3.3	Result	s and Discussion	29
		3.3.1	Explant Culture	29
		3.3.2	Initiation of Callus-like Masses	31
		3.3.3	Embryoids Induction	39
	3.4	Conclu	ision	43
4	OIL	PALN	1 POLYEMBRYOID (PE) GROWTH PATTERN	44
			FURE MAINTENANCE	
	4.1	Introd	uction	44
	4.2	Mater	ials and Methods	45
		4.2.1	Stock of PE Cultures	45
		4.2.2	Data Collection and Processing	45
		4.2.3	Mathematical Modelling in Computing Growth Models and Growth Rate	46
			4.2.3.1 Growth Models	46
			4.2.3.2 Growth Rates	47
		4.2.4	Allometric Basis	47
			4.2.4.1 Relative Growth Rate (RGR)	47
		4.2.5	Statistical Analysis	48
	4.3	Result	s and Discussion	48
		4.3.1	Growth Models	48
		4.3.2	Polyembryoids Growth Increment	56
	4.4	Conclu	usion	61
5			Y, CONCLUSION AND RECOMMENDATIONS	63
	FOI	R FUTI	URE STUDIES	
	5.1			63
			al Conclusion	64
	5.3	Recon	nmendations for Future Studies	65
		INCES		67
		ICES		78
BIO	DAT	A OF S	STUDENT	80

 \bigcirc

LIST OF TABLES

Table		Page
2.1	Phylogenetic lineage of <i>Elaeis guineensis</i>	4
2.2	Interest traits in oil palm breeding and selection	5
2.3	Oil palm traits for genetic improvement program in Malaysia	5
2.4	Word major producers of palm oil ('000 tonnes)	б
2.5	Oil palm planted area in Malaysia (million hectares)	7
2.6	Yield of fresh fruit bunches, crude palm oil and palm kernel (tonnes/hectare)	8
2.7	World production of oil palm tissue culture plantlets	10
2.8	Oil palm tissue culture ramets production in Malaysia	10
2.9	Oil palm explants and growth regulators towards callogenesis and somatic embryogenesis	13
2.10	Performance of MPOB outstanding clones	18
3.1	Details on elite <i>tenera</i> ortet samples from MPOB Research Station	22
3.2	Details on reclone ortet samples from MPOB Research Station	23
3.3	Mean of callus production of ten clonal ortets and ten elite <i>tenera</i> ortets using One-Way ANOVA	33
3.4	Days of the fastest callus formation of ten clonal ortets and ten elite <i>tenera</i> ortets using One-Way ANOVA	33
3.5	Callus production of ten clonal ortets at different callogenesis duration	34
3.6	Callus production of ten elite <i>tenera</i> ortets at different callogenesis duration (callus production vs. time)	34
3.7	Performance of callus production for ten clonal ortets during four observation points	35
3.8	Performance of callus production for ten elite <i>tenera</i> ortets during four observation points	36
3.9	Callus production of ten elite <i>tenera</i> and ten clonal ortets at different callogenesis stages / time using One-Way ANOVA	36

 \bigcirc

3.10	Callus production of ten clonal and ten elite <i>tenera</i> ortets vs. ortet type x callogenesis stages (time) using One-Way ANOVA		
3.11	Response of callus production of ten elite <i>tenera</i> and ten clonal ortets cultured on fresh and non-fresh induction media using One-way ANOVA	38	
3.12	Response of callus production of ten elite <i>tenera</i> and ten clonal ortets cultured on fresh and non-fresh induction media using One-Way ANOVA	38	
3.13	Mean of embryoids production of ten clonal ortets and ten elite <i>tenera</i> ortets using One-Way ANOVA	39	
3.14	Day of the fastest embryoids formation of ten clonal ortets and ten elite <i>tenera</i> ortets using One-Way ANOVA	39	
3.15	Correlation of total polyembryoid lines produced versus total callus and first day embryogenic calli form for elite <i>tenera</i> and clonal ortets (Pearson Correlation Coefficients, N=20) Prob > $ \mathbf{r} $ under H0: Rho=0	42	
3.16	Correlation of total polyembryoids lines produced versus total callus and first day embryogenic calli form for clonal ortets (Pearson Correlation Coefficients)	43	
3.17	Correlation of total polyembryoids lines produced versus total callus and first day embryogenic calli form for elite <i>tenera</i> ortets (Pearson Correlation Coefficients)	43	
4.1	Details of elite <i>tenera</i> ortets sampled from MPOB-UKM Research Station, Bangi, Selangor and clonal ortets from MPOB Keratong Research Station, Pahang	45	

xiii

(G)

LIST OF FIGURES

I	Figure		Page
	2.1	Design view of inoculation or leaf explant stage-0 table	16
	2.2	Linking a common field from two tables	17
	3.1	Process of ortet sampling in MPOB research station (a) Removing thorns and preparing a working platform. (b) Cutting of the shoot cabbage (c) The sample is tied with a rope before it is lowered to the ground and (d) The cut cabbage is about 50 to 70 cm long dependening on the ortet age	24
	3.2	Cabbage and explant cutting for cloning process (a) A longitudinal is made to open the outermost petiole until all the leaf spears comprising stacks of young leaflets are exposed (b) Strips of six-layer stack were inoculated vertically on the callus induction media (c) Leaf explants with callus and (d) Smooth surfaced structure of primary embryoids	25
	3.3	Tissue culture schemes for callogenesis and embryogenesis at RISDA Tissue Culture laboratory	27
	3.4	Screenshot image of data generated from the TCDS 'Query' function	29
	3.5	Figure 3.5 Initial leaf explants culture for clonal and elite <i>tenera</i> ortets (a): Quantity of young leaf explants culture at the Explant Stage 0 (Ep0) for ten clonal ortets and (b) for ten elite <i>tenera</i> ortets	30
	3.6	Initiation of callus-like masses incubated in the continous darkness (a) Leaf explants cultured in MS medium supplemented with 10 mg/L NAA, (b) yellowish and hardened callus of age 12 weeks was initiated at the edge of explant cutting and (c) CLTM structure aged 32 weeks	32
	3.7	Correlation of the total embryoid lines (PE) formed versus callus production from elite <i>tenera</i> and clonal ortets	40
	3.8	Correlation of the first embryoid lines formed versus callus production from elite <i>tenera</i> and clonal ortets	41
	3.9	Correlation of the embryoid lines formed versus the first embryoid days formed in elite <i>tenera</i> and clonal ortets	41
	3.10	Scatter plot of correlation of elite <i>tenera</i> and clonal ortets here variables Pearson Correlation Coefficients, $N = 20$, Prob > r under H0: Rho=0	42

2	4.1	The PE culture of clonal ortets RC1-HPR15 derived from the fourth subculture (a) Initial weight of 1.25 g and (b) 2.5 g	46
2	4.2	The growth curve of ortet RC1-HPR15 PE culture maintained at the duration of 12 weeks	49
2	4.3	The growth curve of ortet RC44-KRR5 polyembryoids culture maintained at the duration of 12 weeks	50
2	4.4	The growth curve of ortet RC45-KRR9 polyembryoids culture maintained at the duration of 12 weeks	51
2	4.5	Growth pattern and stationary points of clonal ortets during PE maintenance	52
2	4.6	The growth curve of ortet RC37-UKR33 PE culture maintained at the duration of 12 weeks	53
2	4.7	The growth curve of ortet RC38-UKR35 PE culture maintained at the duration of 12 weeks	54
2	4.8	The growth curve of ortet RC40-UKR37 PE culture maintained at the duration of 12 weeks	55
2	4.9	Growth pattern and stationary points of elite <i>tenera</i> ortets during PE maintenance	56
4	.10	PE culture selected for PE maintenance in oil palm ramet production	57
4	.11	Growth rate of clonal ortets during PE maintenance	58
4	.12	Growth rate of elite tenera ortets during PE maintenance	59
4	.13	The acceleration of clonal PE cultures during 12 weeks in PE maintenance media	60
4	.14	The acceleration of elite <i>tenera</i> PE cultures during 12 weeks in PE maintenance media	61

LIST OF ABBREVIATIONS

ANOVA	Analysis of Variance
cm	centimetre
g	gram
ha	hectare
m	meter
mm	milimeter
kg	kilogram
t	tonne
r	correlation coefficient
R ²	coefficient of determination
%	percentage
0°C	degree celcius
2,4-D	2,4-dichlorophenoxyacetic acid
MS	Murashige and Skoog media
FFB	Fresh fruit bunches
СРО	Crude palm oil
РК	Palm kernel
МРОВ	Malaysian Palm Oil Board
PGRs	Plant growth regulator
TCDS	Tissue culture database system
SpID	Sampling identification
ОТВ	oil to bunch
G	

CHAPTER 1

INTRODUCTION

The golden crop of Malaysia, the oil palm was originally introduced to Malaysia in 1870 from Ghana. The cultivation began in the last century by planting the *dura* fruit type in the plantation. However, the yield was marginal (Kushairi and Rajanaidu, 2000). The first wave of yield improvement of up to 30% was by switching from dura to the tenera (DxP) planting materials (Beirnaert and Vanderweyen, 1941). Since then, oil palm breeders have continuously devoted great efforts in developing traits for genetic improvement program (Soh et al., 2009). The genetic potential of the palm has been reported to be 18.2 tonnes of oil/hectare/year (t/ha/yr) (Corley, 1998; Chan et al., 2000) and cloning such palm would provide high-yielding planting materials for the industry, which would be the second wave in yield improvement. In 2018, the national average annual oil yield has been stagnating at 3.42 tonnes of oil/ha/yr with the fresh fruit bunches (FFB) achievement of 17.9 t/ha/yr. Most of the current oil palm planting materials are from the high yielding materials which are propagated through crosspollination of selected dura x pisifera (DxP) seed materials with improved genetic potential. Generally, conventional breeding methodology for superior traits is time consuming and often produces a plenty of variation.

Reports on success in oil palm cloning was first announced in the mid-seventies (Jones, 1974; Rabechault and Martin, 1976) and a little later in the early eighties in Malaysia (Wooi et al., 1981; Paranjothy and Othman, 1982).

Oil palm possesses a single growing point and due to its feature that does not have suckers or bud grafting, oil palm ramets cannot be vegetatively multiplied through common technique as applied to other plants e.g. roses, bamboo and rattans (Corley and Tinker, 2016). Thus, cloning via plant tissue culture techniques is the only method to produce and propagate uniform planting materials.

Soh (2018) described the selection of outstanding ortets based on oil to bunch is preferrable due to higher heritability against fresh fruit bunches (FFB). Inefficient oil palm tissue culture process, incidence of mantling and high cost of ramets are also the current prevailing drawbacks. The ortet source limitation has led the tissue culturist to investigate and assess the potential and performance of clonal and elite *tenera* (DxP) ortets in the cloning process for mass propagation. Clonal ortet refers to the selected planted palms derived from tissue culture process that meets the requirement of Malaysian Standard MS 2099: 2008 - Oil palm clones for commercial planting – Specification for ortet selection. The inclination of utilizing clonal ortets in commercial tissue culture laboratory for mass propagation has been recently documented (Roowi et al., 2010; Tarmizi et al., 2017; Febrianti, 2018; Choo et al., 2018; Ong-Abdullah et al., 2018; Soh, 2018). Since information on the oil palm tissue culture protocols are scarce, thorough investigation on the culture development from the initial stages of callogenesis, embryogenesis and polyembryoids maintenance are needed to optimize the ortets potential.

Therefore, this study was undertaken with a prime goal to improve the efficiency of the oil palm cloning process with the following specific objectives. The objectives of this study are:

- 1. to differentiate between the callogenesis and embryogenesis potentials of elite *tenera* and clonal ortets.
- 2. to establish the growth patterns of polyembryoids derived from elite *tenera* and clonal ortets

This work will advance the current knowledge on oil palm cloning, specifically on ortets performances at the stage of callogenesis, embryogenesis and polyembryoid maintenance that eventually will contribute further to the development of a growth model which will eventually provide efficient information for oil palm ramets projection, particularly for commercial scale propagation.



REFERENCES

- Al-Khayri, J. M. (2012). Determination of the date palm suspension growth curve, optimum plating efficiency and influence of liquid medium on somatic embryogenesis. *Emirates Journal of Food and Agriculture*, 24(5): 444-455.
- Barcelos, E., Rios, S. A., Cunha, R. N. V., Lopes, R., Motoike, S. Y., Babiychuk, E., Skirycz, A. and Kushnir, S. (2015). Oil palm natural diversity and the potential for yield improvement. *Frontiers in Plant Science*, 6(190): 1-16.
- Basiron, Y. (2000). Techno-economic aspects of research and development in the Malaysian oil palm industry. In Y. Basiron, B. S. Jalani, and K. W. Chan (Eds.), *Advances in Oil Palm Research*, Vol. 1. Chapter 1, pp. 1-18. Malaysian Palm Oil Board.
- Basiron, Y. (2007). Palm oil production through sustainable plantations. European Journal of Lipid Science and Technology. 109: 289-295.
- Basiron, Y. and Fong, K. Y. (2015). Land use impacts of the lifestock and palm oil industries. *Journal of Oil Palm, Environment and Health*, 6: 1-9.
- Baudouin, L., Meunier, J. and Noiret, J. M. (1995). Methods used for ortet choice. In Rao, V., Henson, I. E. and Rajanaidu, N. (Eds). *Recent developments in oil palm tissue culture and biotechnology*, pp. 217-224. Palm Oil Research Institute.
- Beirnaert, A. and Vanderweyen, R. (1941). Contribution a l'etude genetique et biometrique des varieties d'*Elaeis guineensis* Jacquin. *Pub. Inst. Nat. Etude Agron, Congo Belge Ser. Sci.*, 27: 1–101.
- Buttery, B. R. (1969). Analysis of the growth of soybeans as affected by plant population and fertilizer. *Canadian Journal of Plant Science*, 49: 675-684.
- Chan, K. W., Tarmizi, A. and Wahid, O. (2000). Advances in fertilizer management in the oil palm industry. *Oil Palm Bulletin*, 41: 39-46.
- Choo, C. N., Wong, C. K., Nur Akilla, M. R., Ee, C. C., Ilham, A. A. and Tan, C. C. (2014). Genotype effect on oil palm tissue culture callogenesis and embryogenesis. *Proceedings of the International Oil Palm Conference (IOPC)*, International Oil Palm Research Institute, Bali, Indonesia.
- Choo, C. N., Wong, C. K. and Melody, M. P. (2018). Experience and challenges in commercial production of elite oil palm clones in Applied Agricultural Resources Sdn. Bhd. *International Seminar on Status of Oil Palm Tissue Culture Technology*, July 16, 2018, pp. 31-41. International Society for Oil Palm Breeders, Medan, Indonesia.
- Corley, R. H. V. and Gray, B. S. (1976). Yield and yield components. In R. H. V. Corley, J. J. Hardon and B. J. Wood (Eds.), *Oil Palm Research*, pp. 77-86. Amsterdam: Elsevier.

- Corley, R. H. V., Wooi, K. C., and Wong, C. Y. (1979). Progress with vegetative propagation of oil palm. *Planter*, 55: 377-380.
- Corley, R. H. V., Lee, C. H., Law, I. H. and Wong, C. Y. (1986). Abnormal flower development in oil palm clones. *The Planter*, 62: 233-240.
- Corley, R. H. V. (1998). What is the upper limit to oil extraction ratio? Proceedings of the International Conference on Oil and Kernel Production in Oil Palm -A Global Perspective, pp. 256-269. 27-28 September 1996. Palm Oil Research Institute of Malaysia, Kuala Lumpur, Malaysia.
- Corley, R. H. V. and Tinker, P. B. (2003). Vegetative propagation and biotechnology. In R. H. V. Corley and P. B. Tinker (Eds.), *The Oil Palm*, Fourth Edition, pp. 201-215. United Kingdom: Blackwell Science.
- Corley, R. H. V. and Palat, T. (2015). Maximising lifetime yield for greater economic sustainability. Paper presented at *International Palm Oil Congress, Green Opportunities from the Golden Crop*, Kuala Lumpur, 6-8 October 2015.
- Corley, R. H. V. and Tinker, P. B. (2016a). The climate of the oil palm-growing regions. In R. H. V. Corley and P. B. Tinker (Eds.), *The Oil Palm*, Fifth Edition, pp. 53-67. United Kingdom: Blackwell Science.
- Corley, R. H. V. and Tinker, P. B. (2016b). Oil palm and sustainability. In R. H. V. Corley and P. B. Tinker (Eds.), *The Oil Palm*, Fifth Edition, pp. 519-534. United Kingdom: Blackwell Science.
- Corley, R. H. V. and Tinker, P. B. (2016c). Vegetative propagation and biotechnology. In R. H. V. Corley and P. B. Tinker (Eds.), *The Oil Palm*, Fifth Edition, pp. 208-224. United Kingdom: Blackwell Science.
- Cross, D (2017). Genomic selection for oil palm. In A. C. Soh, S. Mayes and R. Jeremy (Eds.), Oil Palm Breeding Genetics and Genomics, pp. 263-269. Boca Raton: CRC Press.
- Cynthia, L. M. M., Nurita, T-M., Reflini, C. U. and Tony, L. (2015). Micropropagation of embryogenic callus of oil palm (*Elaeis guineensis* Jacq.). *Procedia Chemistry* 14: 122-129.
- Dantu, P. K. and Tomar, U. K. (2010). Somatic embryogenesis. In G. Thipathi (Ed.), *Celular and Biochemical Sciences*, pp. 892-902. New Delhi: IL International Publishing House Pt. Ltd.
- Denis, M. and Bouvet, J. M. (2013). Efficiency of genomic selection with models including dominance effect in the context of *Eucalyptus* breeding. *Tree Genetics* & *Genomes*, 9: 37-51.

- Durand-Gasselin, T., Duval, Y., Baudouin, L., Maheran, A. B., Konan, K. & Noiret, J. M. (1995). Description and degree of the mantled flowering abnormality in oil palm (*Elaeis guineensis* Jacq.) clones produced using the Orstrom-CIRAD procedure. In *Recent Developments In Oil Palm Tissue Culture and Biotechnology*, pp. 48-63.
- Durand-Gasselin, T. (2010). Strategies to develop oil palm clones for Latin American countries and Africa and its potential development. Paper presented at the *International Seminar on Advances in Oil Palm Tissue Culture*, International Society for Oil Palm Breeder (ISOPB), May 29, Yogyakarta, Indonesia.
- Duval, Y., Aberlenc, F. and de Touchet, B.(1995). Use of embryogenic suspensions for oil palm micropropagation. In. (eds. Rao, V., Henson, I. E. and Rajanaidu, N.). Proceedings of The 1993 ISOPB International Symposium on Recent Developments in Oil Palm Tissue Culture and Biotechnology. Palm Oil Research Institute of Malaysia, Bangi. Pp. 38-47.
- Eeuwens, C. J. (1976) Mineral requirements for growth and callus induction of tissue explants excised from mature coconut palms (*Cocos nucifera*) and cultured *in vitro*. *Physiology Plant*, 34: 23-28.
- Efretuei, A. O. (2018). Suspension culture, callus culture and bud culture. In *Plant Propagation by Tissue Culture*, pp. 105-109. Canada: Delve Publishing.
- El-Bellaj, M. (2000). Study on biochemical parameters in relation to embryogenesis potential and somatic embryo maturation of date palm (*Phoenix dactylifera* L.).
 PhD. Thesis. Faculty of Sciences Semlalia, University of Cadi Ayyad, Morocco.
- Escobar, R., Alvarado, A., Guzmin, N. and Chinchilla, C. (2005). An overview of the ASD approach for using its broad genetic pool and reducing the risk of abnormalities in oil palm clones. *Proceedings of the MPOB International Palm Oil Congress. Technological Breakthrough and commercialization the way forward*, pp. 144-166, 25-29 September, Kuala Lumpur, Malaysia.
- Febrianti, I., Boon Beng, A., Izarul, I., Manjit, S., Samosir, Y. and Sharma, M. (2018). Status of oil palm cloning program at Asian Agri. *International Seminar on Status of Oil Palm Tissue Culture Technology*, pp. 42-54, July 16, International Society for Oil Palm Breeders, Medan, Indonesia.
- Gardener, F. P., Pearce, R. B., and Mitchell, R. L. (1985). Physiology of crop plants. First edition. Iowa, USA: The Iowa State University Press.
- Gaspar, T., Kevers, C., Penel, C., Greppin, H., Reid, D. M. and Thrope, T. A. (1996). Plants hormones and plant growth regulators in plant tissue culture. *In Vitro Cellular and Development Biology-Plant*, 32: 272-289.
- Goddijn, O. J. M., De Kam, R. J., Zanetti, A., Schilperoort, R. A. and Hoge, J. H. C. (1992). Auxin rapidly down-regulates transcription of the tryptophan decarboxylase gene from *Catharanthus roseus*. *Plant Molecular Biology*, 18: 1113-1120.

- Haliza Dahlia, A. H., Norrizah, J. S. and Tarmizi, A. H (2011) *In vitro* studies of biomass yield and its phytochemical on different clones of oil palm cell suspension culture. Universiti Malaysia Terengganu 10th International Annual Symposium (UMTAS) 2011 -Empowering Science, Technology and Innovation towards a Better Tomorrow, pp. 778-783, Terengganu, Malaysia.
- Hassan, H. H. E. (2015). Reproductive allometry and plasticity in relation to plant population density (*Glycine max* L. Merrill). PhD. Thesis, Universiti Putra Malaysia.
- Ho, Y. W., Tan, C. C., Soh, A. C., Wong, G., Chong, S. P., Choo, C. N. and Norazura,
 A. (2009). Biotechnological approaches in producing oil palm planting materials a success story. Oil Palm for farmers' prosperity and edible oil security. In Rethinam, P., Reddy, V. M., Mandal, P. K., Suresh, K. and Prasad,
 M. V. (Eds). *Proceedings of The National Conference on Oil Palm*, pp. 86-93 SOPOPRAD, Pedavegi, India.
- Hunt, R. (1982). Plant growth curves. The functional approach to plant growth analysis. London, UK: Edward Arnold Ltd.
- Jaafar, M. and Jalani, B.S. (1999). The oil palm in Malaysia in the next millennium. *Tropical Agriculture Association Newsletter*, 19(3): 9-13.
- Jalani, B. S., Chan, K. W, Rajanaidu, N. and Ariffin, D. (2001). Review of the stagnating yield in oil palm plantations and its possible remedies. Proceeding of the 2001 International Palm Oil Congress -Agriculture, Malaysian Palm Oil Board, 20-22 September, Kuala Lumpur, Malaysia.
- Jones, L. H. (1974). Propagation of clonal palms by tissue culture. *Oil Palm News*, 17: 1-8.
- Jones, L. H. (1995). Clonal propagation of oil palm, past, present and future: a personal view. In V. Rao, I. E. Henson and N. Rajanaidu, N. (Eds.), *Recent developments* in oil palm tissue culture and biotechnology, pp. 1-20, Palm Oil Research Institute of Malaysia, Kuala Lumpur, Malaysia.
- Kushairi, A. and Rajanaidu, N. (2000) Breeding populations, seed production and nursery management. In Y. Basiron, B. S. Jalani and K. W. Chan (Eds.), *Advances in oil palm research*, Vol. I, pp. 39-98. Malaysian Palm Oil Board, Kuala Lumpur, Malaysia.
- Kushairi, A., Tarmizi, A. H., Zamzuri, I., Ong-Abdullah, M., Rohani, O., Samsul Kamal, R., Ooi, S. E., Ravigadevi, S. and Mohd Basri, W. (2006). Current status of oil palm tissue culture in Malaysia. Paper presented at the *Clonal and Quality Replanting Material One-Day Workshop*, pp. 1-11, Kuala Lumpur, Malaysia.

- Kushairi, A., Tarmizi, A. H., Zamzuri, I., Ong-Abdullah, M., Samsul Kamal, R., Ooi S E., Ravigadevi, S., Mohd. Basri, W. and Rajanaidu, N. (2010). Production, performance and advances in oil palm tissue culture. Paper presented at the *International Seminar on Advances in Oil Palm Tissue Culture*, pp. 1-22, International Society for Oil Palm Breeders (ISOPB), May 29, Yogyakarta, Indonesia.
- Kushairi, A., Singh, R. and Ong-Abdullah, M. (2017). The oil palm industry in Malaysia: thriving with transformative technologies. *Journal of Oil Palm Research*, 29(4): 431-439.
- Latiff, A. (2000). The biology of the genus *Elaeis*. In Y. Basiron, B. S. Jalani, and K. W. Chan (Eds.), *Advances in Oil Palm Research*, Vol. 1. Chapter 2, pp. 19-38. Malaysian Palm Oil Board.
- Machakova, I., Zazimalove, E. and George, E. F. (2008). Plant growth regulators I: Introduction: auxins, their analogues and inhibitors. In E. F. George, M. A. Hall, and G-J De Klerk (Eds.), Plant Propagation by Tissue Culture, Vol. 1, pp. 175-204. Netherlands: Springer Publ.
- Maheran, A. B., Abu Zarin, O., Aw, K. T. and Chin, C. W. (1995). FELDA's early experiences with vegetative propagation of the oil palm (*Elaeis guineensis* Jacq.) In *Proceedings of the 1993 PORIM International Palm Oil Congress -Agriculture*, pp. 99-113. Palm Oil Research Institute of Malaysia, Kuala Lumpur, Malaysia.
- Majerus, F. and Pareilleux, A. (1986). Alkaloid accumulation in Ca-alginate entrapped cells *Catharanthus roseus*, using a limiting growth medium. *Plant Cell Reports*, 5: 302-305.
- Mandak, B. and Pysek, P. (1999). How does density and nutrient stress affect allometry and fruit production in the heterocarpic species *Altriplex sagittata* (Chenopodiaceae)? *Canadian Journal of Botany*, 77: 1106-1119.
- MPOB (2018). Malaysian Oil Palm Statistics 2017, 37th Edition. Selangor, Malaysia: Malaysian Palm Oil Board.
- MPOB (2019a. Malaysian Oil Palm Statistics 2018, 38th Edition. Selangor, Malaysia: Malaysian Palm Oil Board.
- MPOB (2019b. Review of the Malaysian Oil Palm Industry 2018. Selangor, Malaysia: Malaysian Palm Oil Board.
- MPOB (2019c. MPOB Industry Oil Palm Breeders and Tissue Culturist (BTC) Committee Meeting (1/2019), Personal Communication.
- Mondjeli, C., Walter, A. N., Ntsomboh, N. G., Ni Made, A. W., Ade, W. and Ngando, E. G. F. (2015). Induction of oil palm (*Elaeis guineensis* Jacq. Var *Tenera*) callogenesis and somatic embryogenesis from young leaf explants. *Journal of Applied Biology and Biotechnology*, 3(4): 4-10.

- Murashige, T. and Skoog, F. (1962). A revised medium for rapid growth and bio-assay with tobacco tissue cultures. *Physiology Plant*, 15: 473-497.
- Murphy, D. J. (2014). The future of oil palm as a major global crop: opportunities and challenges. *Journal of Oil Palm Research*, 26(1): 1-24.
- Nurul, I. O., Asmah, A., Norrizah, J. S. and Shamsiah, A. (2013). Callus induction and somatic embryogenesis from leaf and nodal explants of *Lycium barbarum* L. *Biotechnology*, 12: 36-45.
- Nwanko, B. A. and Krikorian, A. D. (1983). Morphogenic potential of embryo and seedling-derived callus of *Elaeis guineensis* Jacq. Var. *Pisifera. Annual of Botany*, 51: 65-76.
- Ong-Abdullah, M., Tarmizi, A. H., Zamzuri, I., Samsul Kamal, R., Ooi, S. E., Naqiuddin, M. H., Dalilah, A. B., Norashikin, S., Nuraziyan, A., Pang, J. T. Y., Siti Rahmah A. R., Mohd Isa, Z. A., Sing, R., Low, E. T. L., Nookiah, R., Ordway, J. M., Jiang, N., Smith, S. W., Lakey, N., Mohamad Arif, A. M., Ahmad Parveez, G. K., Martienssen, R. A. and Sambanthamurthi, R. (2018). A holistic perspective of current progress in oil palm tissue culture. *International Seminar on Status of Oil Palm Tissue Culture Technology*, pp. 15-16. International Society for Oil Palm Breeders, July 16, Medan, Indonesia.
- Ooi, S. E., Zamzuri, I., Ang, C. L., Mohd Azmir, M., and Ong-Abdullah, M. (2009). Response of hormone -responsive genes to 2,4-D vs. NAA treatment of explant cultures via gene expression analysis. *Proceedings of the Agriculture, Biotechnology and Sustainability Conference*, pp. 1339, Vol. III. International Palm Oil Congress PIPOC 2009, KLCC, Kuala Lumpur, Malaysia.
- Paranjothy, K. and Othman, R. (1982). In vitro propagation of oil palm. Proceedings of the Fifth Congress Plant Tissue and Cell Culture, pp. 747-748, Maruzen, Tokyo, Japan.
- Paranjothy, K., Othman, R., Tarmizi, A. H., Tan, C. S. and Tan, C. C. (1990). Current status and strategies of oil palm tissue culture research. *Proceedings of the 1989 International Palm Oil Development Conference Agriculture*, pp. 109-121, Palm Oil Research Institute of Malaysia, Kuala Lumpur, Malaysia.
- Paranjothy, K., Othman, O., Tan, C. C., Wong, G. and Soh, A. C. (1995). Incidence abnormalities in relation to in vitro protocols. In. (eds. Rao, V., Henson, I. E. and Rajanaidu, N.). Proceedings of The 1993 ISOPB International Symposium on Recent Developments in Oil Palm Tissue Culture and Biotechnology. Palm Oil Research Institute of Malaysia, Bangi. Pp. 77-85.
- Pasquali, G., Goddijn, O. J. M., De Waal, A., Verpoorte, R., Schilperoort, R. A., Hoge, J. H. C. and Memelink, J. (1992). Coordinated regulation of two indole alkaloid biosynthetic genes from *Catharanthus roseus* by auxin and elicitors. *Plant Molecular Biology*, 18: 1121-1131.
- Rabechault, H. and Martin, J-P. (1976). Multiplication vegetative du palmier a huile (*Elaeis guineensis* Jacq.) al'aide de cultures de tissue de tissusfoliares. *C.R.Acad. Sci. Paris*, 283: 1735-1737.

- Rajanaidu, N., Rohani, O. and Jalani, B. S. (1997). Oil palm clones: current status and prospects for commercial production. *The Planter*, 73(853): 163-184.
- Rajanaidu, N., Kushari, A., Rafii, M., Mohd. Din, A., Maizura, I and Jalani, B. S. (2000). Oil palm breeding and genetic resources. In Y. Basiron, B. S. Jalani and K. W. Chan (Eds.), *Advances in oil palm research*, Vol. 1, pp. 171-237, Malaysian Palm Oil Board, Kuala Lumpur, Malaysia.
- Rajesh, M. K., Radha, E., Karun, A., and Parthasarathy, V. A. (2003). Plant regeneration from embryo-derived callus of oil palm - The effect of exogenous polyamines. *Plant Cell. Tissue Organ Cult.* 75, 41–47.
- Rival, A., Bernard, F. and Mathieu, Y. (1997). Changes in peroxidase activity during in vitro rooting of oil palm (*Elaeis guineensis* Jacq.). Science Horticulture, 71: 103-112.
- Rival, A. (2000). Somatic embryogenesis in oil palm. In S. M. Jain, P. K. Gupta and R.
 J. Newton (Eds.), *Somatic embryogenesis in woody plants*, pp. 249-289, Dordrecht, Netherlands: Kluwer Academic Publishers.
- Rohani, O., Sharifah, S. R. S. A., Mohd Rafii, Y., Ong-Abdullah, M., Tarmizi, A. H and Zamzuri, I. (2000). Tissue culture of oil palm. In Basiron, Y., Jalani, B. S and Chan, K. W (Eds.), *Advances In Oil Palm Research*, pp. 238-283. Malaysian Palm Oil Board.
- Rohani, O., Zamzuri, I and Tarmizi, A. H. (2003). Oil Palm Cloning: MPOB Protocol. In: *MPOB Technology*, No. 26. Malaysian Palm Oil Board.
- Roowi, S. H., Ho, C. L., Syed Alwee, S. S. R., Ong-Abdullah, M. and Napis, S. (2010). Isolation and characterization of differentially expressed transcripts from the suspension cells of oil palm (*Elaeis guineensis* Jacq.) in response to different concentration of auxin. *Molecular Biotechnology*, 46: 1-19.
- Roowi, S. H., Rafidah, M. K., Bahiyah, R. N. R. H., Fatah, A. D., Nazmi, M. B., Noorsusilawati, M., Farid, M. A. R., Latif, M. K., Mahfuz, M. R., Naderman, S., Adibah, N. I., Nasruddin, M., Tan, J. H. and Syed Alwee, S. S. R. (2018). Is there a future in tissue culture for the oil palm industry? *International Seminar on Status of Oil Palm Tissue Culture Technology*, pp. 59-71, International Society for Oil Palm Breeders. July 16, Medan, Indonesia.
- Rosenquist, E. A. (1990). An overview of breeding technology and selection in *Elaeis* guineensis, Proceedings of the 1989 International Palm Oil Development Conference Agriculture, pp. 5-25, Palm Oil Research Institute of Malaysia, Kuala Lumpur, Malaysia.
- Selamat, A., Wahab, Z., Awang, Y., Mohamed, M. M. T. and Osman, M. (2008). Use of absolute function and its formation and 'redevelopment' of mathematical models in some plant-related quantitative physiology: Salinity effects on leaf development of *Schefflera arboricola* and harvest index in rice. Proc. 976, pp. 165-167, American Institute of Physics, Meville, USA.

- Selamat, A., Atiman, S. A., Puteh, A., Abdullah, N. A. P., Mohamed, M. T. M., Zulkeefli, A. A. and Othman, S. (2012)."Allometry" deterministic approaches in cell size, cell number and crude fiber content related to the physical quality of kangkong (*Ipomoea reptans*) grown under different plant density pressures, Vol. 9, pp. 30-34, *International Journal of Modern Physics*: Conference Series. Singapore: World Scientific Publishing Co. Pte. Ltd.
- Sharifah, S. R. S. A., Roowi, S. H. Aw, K. T. and Abu Zarin, O. (2010). Progress of oil palm tissue culture in FELDA and its challenges. *International Seminar on Advances In Oil Palm Tissue Culture*. pp. 1-8. International Society for Oil Palm Breeders, May 29, Yogyakarta, Indonesia.
- Sharma, M. and Tan, Y. P. (1999). Oil palm breeding programmes and the performance of DxP planting materials at United Plantations Berhad. *Proceedings of the Seminar on Sourcing of Oil Palm Plating Materials for Local and Overseas Joint Venture*. Rajanaidu, N. & Jalani, B S. (Ed.), pp. 118-135. Palm Oil Research Institute of Malaysia, Kuala Lumpur, Malaysia.
- Simon, S., Hendry, T., Chang, S. W. and Kiaw, C. W. (1998). Early year performance of clonal oil palm (*Elaeis guineensis* Jacq.) plantings in PPB Oil Palm Bhd., Sabah – a case study. *The Planter*, 74(866): 257-269.
- Simpson, A. and Robinson, C. (1999). Mastering Access 2000 Premium Edition. San Francisco, USA: Sybex Inc. and John Wiley & Sons Inc.
- Smith, D. R., Yimbang, R., Lee, C. and Lord, S. (2010). High efficiency vegetative amplification -a new oil palm improvement system. Paper presented at the Seminar of Advances In Oil Palm Tissue Culture, International Society for Oil Palm Breeders, 29 May 2010, Yogyakarta, Indonesia.
- Sogeke, A. K. (1996). Rapid callus proliferation, somatic embryogenesis and organogenesis of oil palm (*Elaeis guineensis* Jacq.). *Elaeis*, 8: 92-103.
- Sogeke, A. K. (1998). Stages in the vegetative propagation of oil palm (*Elaeis guineensis* Jacq.) through tissue culture. *Journal of Oil Palm Research*, 10(2): 1-9.
- Sogeke, A. K., Odewale, J. O., Exe, C. R. and Omamor, I. B. (1999). Vegetative propagation of oil palm (*Elaeis guineensis* Jacq.). Initiation of callus from leaf explants. *Proceedings of the 1999 PORIM International Palm Oil Congress Agriculture Conference*, pp. 536-541, Kuala Lumpur, Malaysia.
- Soh, A. C., Wong, G., Tan, C. C., Chew, P. S., Hor, T.Y., Chong, S. P. and Gopal, K (2001). Recent advanced towards commercial production of elite oil palm clones. *Proceedings of the 2001 International Palm Oil Congress Agriculture*, pp. 3, 20-11 August 2001, Kuala Lumpur, Malaysia.
- Soh, A. C., Wong, G., Tan, C. C. (2006). Advances and issues in commercial propagation of oil palm clones. Paper presented at Ministry of Plantation Industry and Commodities Workshop, Kuala Lumpur, Malaysia.

- Soh, A. C., Wong, C. K., Ho, Y. W. and Choong, C. W. (2009). Oil palm. In J. Vollmann and I. Rajcan (Eds.), *Oil Crops-Handbook for Plant Breeding*, Chapter 11, pp. 333-367. Berlin, Germany: Springer Science+Business Media, LLC.
- Soh, A. C., Wong, G., Tan, C. C., Chew, P S., Chong, S. P. Ho, Y. W., Wong, C. K., Choo, C. N., Nor Azura, H. and Kumar, K. (2011). Commercial scale propagation and planting of elite oil palm clones: research and development towards realization. *Journal of Oil Palm Research*, 23: 935-952.
- Soh, A. C., Mayes, S., Roberts, J., Ong-Abdullah, M; Ooi, S. E., Tarmizi, A. H., Zamzuri, I., Samsul Kamal, R., Wong, W. C., Chin, N. C., Kok, S. Y., Nuraziyan, A. and Norashikin, S. (2017). Clonal Propagation. In A. C. Soh, S. Mayes and R. Jeremy (Eds.), *Oil Palm Breeding Genetics and Genomics*, pp. 191-224. Boca Raton, USA: CRC Press.
- Soh, A. C. (2018). Quo vadis (wither) oil palm tissue culture clonal propagation? International Seminar on Status of Oil Palm Tissue Culture Technology, pp. 7-77. International Society for Oil Palm Breeders, July 16, Medan, Indonesia.
- Tahardi, J. S., Riyadi, I. and Dodd, W. A. (2003). Enhancement of somatic embryo development and plantlet recovery in *Camellia sinensis* by temporary liquid immersion. *Jurnal Bioteknologi Pertanian*, 8(1): 1-7.
- Tarmizi, A. H. (2002). Oil Palm Liquid Culture -MPOB Protocol. *MPOB Information* Series, TT No. 138. Malaysian Palm Oil Board.
- Tarmizi, A. H., Zamzuri, I., Samsul Kamal, R., Ong-Abdullah, M., Ooi, S. E., Naqiuddin, M. H., Dalilah, A. B., Pang, J. T. Y., Siti Rahmah, A. R. and Mohd Isa, A. A. (2017). Realizing clonal potential: current status. *Proceedings of The* 2017 International Palm Oil Congress – Agriculture Conference, pp. 106-126, Kuala Lumpur, Malaysia.
- Te-chato, S. (1998a). Callus induction from cultured zygotic embryo of oil palm subsequent to plantlet regeneration. Songklanakarin Journal of Science and Technology, 20: 1-6.
- Te-chato, S. (1998b). Fertile plant from young leaves-derived somatic embryos of oil palm. *Songklanakarin Journal of Science and Technology*, 20: 7-13.
- Teh, S. H. (2008). Use of growth analysis and model for predicting the 'Quality harvest' of kangkong (*Ipomoea reptans*) at three nitrogen fertilizer rates. Master Thesis. Universiti Putra Malaysia, Malaysia.
- Texeira, J. B., Sondahl, M. R. and Kirby, E. G. (1993). Somatic embryogenesis from immature zygotic embryos of oil palm. *Plant Cell, Tissue and Organ Culture*, 40: 105-11
- Texeira, J. B., Sondahl, M. R. and Kirby, E. G. (1994). Somatic embryogenesis from immature inflorescences of oil palm. *Plant Cell Reports*, 13: 247-250.

- Thuzar, M., Vanavichit, A., Tragoonrung, S. and Jantasuriyarat, C. (2011). Efficient and rapid plant regeneration of oil palm zygotic embryo cv. '*Tenera*' through somatic embryogenesis. *Acta Physiologiae Plantarum*, 33: 123-128.
- Tinker, P. B. (2000). The future research requirements for the oil palm plantation. Proceeding of the International Planters Conference -Plantation tree crops in the new millennium: the way ahead (Ed. E. Pushparajah), pp. 3-40, Incorporated Society of Planters, Kuala Lumpur, Malaysia.
- Wahid, M. B. and Simeh, M.A. (2009). Issues related to production cost of palm oil in Malaysia. Oil Palm Industry Economic Journal, 9(2): 1-12.
- Weckx, S., Inzé, D. and Maene, L. (2019). Tissue culture of oil palm: finding the balance between mass propagation and somaclonal variation. *Front Plant Sci.*, 10(722): 1-12.
- Woittiez, L. S., Wijk, M. T., Slingerland, M., Noordwijk, M. and Giller, K. E. (2017). Yield gaps in oil palm: A quantitative review of contributing factors. *European Journal of Agronomy*, 83: 55-57.
- Wong, G., Tan, C. C. and Soh, A. C. (1996). Large scale propagation of oil palm clones – experience to date. Paper presented at *International Symposium on in vitro Culture and Horticultural Breeding*. Jerusalem, Israel, pp. 9.
- Wong, G., Chong, S. P., Tan, C. C. and Soh, A. C. (1999). Liquid suspension culture- a potential technique for mass production of oil palm clones. *Proceedings of the* 1999 PORIM Palm Oil Congress, pp. 3-11. Palm Oil Research Institute of Malaysia, Kuala Lumpur, Malaysia.
- Wooi, K. C., Wong, C. Y. and Corley, R. H. V. (1981). Tissue culture of palms a review. Proceedings COSTED Symposium 'Tissue culture of economically important plants' (Eds. A. N. Rao), pp. 138-144, Singapore.
- Wooi, K. C. (1995). Oil palm tissue culture practice and constraints. Proceedings of the 1993 ISOPB International Symposium on Recent Development in Oil Palm Tissue Culture and Biotechnology (Eds. V. Rao, I. E. Hensen and N. Rajanidu), pp. 21-32. International Society for Oil Palm Breeders, Kuala Lumpur, Malaysia.
- Yamamoto, O. and Yamada. Y. (1986). Production of reserpine and its optimization in cultured *Rauwolfia serpentina* Benth cells. *Plant Cell Rep.* 5: 50-53.
- Yusnita, Y. and Dwi, H. (2011). In vitro callus induction and embryogenesis of oil palm (*Elaeis guineensis* Jacq.) from leaf explants. *Hayati Journal of Biosciences*, 18(2): 61-65.
- Zamzuri, I. (1998). Efficient rooting of oil palm *in vitro* plantlets using double- layer technique. *PORIM Bulletin*, 36: 23-36.
- Zamzuri, I. (2001). Tissue culture database system. MPOB Viva No. 184/2001 (51). *Internal Report.* Malaysian Palm Oil Board.

- Zamzuri, I. (2011). MPOB Clonal Propagation Programme. International Seminar on Breeding for Sustainability in Oil Palm, pp. 110-124. November 18, 2011. Kuala Lumpur, Malaysia. Jointly organized by the International Society for Oil Palm Breeders (ISOPB) and Malaysian Palm Oil Board (MPOB).
- Zati Hanani, T., Norrizah, J. S., Tarmizi, A. H. and Norizan, A. (2014). The effects of different concentrations of NAA on oil palm (*Elaeis guineensis*) embryoid cultures and phytosterols production. *Australian Journal of Crop Science*, (6): 840-847.