

UNIVERSITI PUTRA MALAYSIA

THERMOSTABILITY OF THE RECOMBINANT HAEMAGGLUTININ-NEURAMINIDASE GLYCOPROTEINS OF NEWCASTLE DISEASE VIRUS

TANG YIK KIONG

FSAS 2003 41



THERMOSTABILITY OF THE RECOMBINANT HAEMAGGLUTININ-NEURAMINIDASE GLYCOPROTEINS OF NEWCASTLE DISEASE VIRUS

By

TANG YIK KIONG

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

July 2003



Dedicated to,
Yik-Shin, Yik-Jia, Parents, Relatives and
My Friends Graduated from Taiwan...



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in the fulfilment of the requirements for the degree of Master of Science

THERMOSTABILITY OF THE RECOMBINANT HAEMAGGLUTININ-NEURAMINIDASE GLYCOPROTEINS OF NEWCASTLE DISEASE VIRUS

 $\mathbf{B}\mathbf{y}$

TANG YIK KIONG

July 2003

Chairperson: Prof Datin Khatijah Mohd Yusoff, Ph.D.

Faculty

: Science and Environmental Studies

The haemagglutinin-neuraminidase (HN) glycoprotein of Newcastle disease virus (NDV)

is of primary importance in inducing virus-neutralizing antibodies against viral infection in

chicken and has been used in the development of many vaccines. A variant strain of the

vaccine strain V4QUE known as V4UPM(HR) has been developed as a heat stable vaccine

for use in the poultry industry in tropical countries such as Malaysia. This protein may also

be involved in maintaining heat stability of some vaccine strains. In this study, the HN

gene of the heat stable variant NDV strain V4UPM(HR) and its parental strain V4QUE

were cloned and expressed in the Baculovirus Expression Vector System (BEVs) and

characterized for their heat stability.

The 1.9 kb HN genes of these strains were amplified by RT-PCR from their genomic RNA

and unidirectionally cloned into the baculovirus transfer plasmid, pCR Bac4.8. These

iii

DNA, Bac-N-BlueTM DNA into Spodoptera frugiperda (Sf9) insect cell line. The recombinant baculoviruses which were generated as recHNV4UPM(HR) and recHNV4QUE, were purified by plaque assay. The respective recombinant HN glycoproteins (recHNs) which were expressed in Sf9 insect cells showed haemagglutination (HA) and neuraminidase (NA) activities as well as haemagglutination inhibition (HI) and haemadsorption activities in serological assays. The HA and NA activities were also detected on the surface and in the cytoplasm of the infected Sf9 cells. SDS-PAGE and Western blot analysis of the recombinant baculovirus-infected Sf9 cell lysates detected protein bands of approximately ~74 kDa, which corresponded to the glycosylated HN protein of the virion. These results indicated that the recHNs were not only successfully expressed in the Sf9 cells but they also appeared to be biologically active and functional.

Based on HA activity, the thermostabilities of recHNV4QUE and recHNV4UPM(HR) together with recHNAF2240 on HA activity were studied and compared with those of the NDV strains, V4QUE, V4UPM(HR) and AF2240. The latter was earlier showed to be heat stable at 56°C. The results showed that the heat resistance phenotypes of the recombinant baculoviruses were genetically represented identical to NDV individuals in the property of thermostability. NDV heat resistant strains AF2240, V4UPM(HR) and recombinant baculoviruses recHNAF2240, recHNV4UPM(HR) were 50% heat inactivated at ~56°C after 4 hours but the parental NDV strain V4QUE and baculovirus strain recHNV4QUE remained as the temperature sensitive strains.



In addition, the HN genes of the recombinants were sequenced and analyzed by the secondary and three dimensional structure predictions of the computer programs. The roles of individual amino acid residue(s) of the HN protein in thermostability were discussed. The polar/non-polar side chains of the substituted amino acid residues [R32, V413E, N79K] in strain V4UPM(HR); E494K in strain V4UPM, where the polar side chain could generate the dipole to form the hydrogen bonding with the aqueous environment and increased the thermostability of the protein. The hydrophobic values and secondary structural arrangements of these substituted/deleted amino acid residues [F151Y, I175M and G431S in strain V4UPM; A152T, N276K, I280T, V303T, Q372R and Y504H in strain AF2240] and [R32, G221, R305, V413E and S584A in strain V4UPM(HR); H87Q and E494K in strain V4UPM formed the secondary structure of the protein for the protein folding events and the protein packing. The thermostable proteins preferred to form the α -helical/ β -sheet structures rather than the mixture of the \beta-sheet/random coil or random coil structures alone at the putative HN domain (399-GAEGR-403) where was predicted by the secondary structure arrangement. Besides, the 3D structures of the HN protein supported that any amino acid residue substitutions near the putative active sites (HN domain 399-GAEGR-403) could have changed the tertiary structure of the HN protein and these tertiary structure/structural arrangements of the HN protein influenced the heat stability of the HN protein.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia untuk memenuhi keperluan Ijazah Master Sains

KESTABILAN TERMA GLIKOPROTEIN HEMAGLUTININ-NEURAMINIDASE REKOMBINAN DARI VIRUS PENYAKIT NEWCASTLE

Oleh

TANG YIK KIONG

Julai 2003

Pengerusi : Prof D

: Prof Datin Khatijah Mohd Yusoff, Ph.D.

Faculti

: Sains dan Pengajian Alam Sekitar

Glikoprotein hemaglutinin-neuraminidase (HN) dari virus penyakit Newcastle (NDV)

adalah penting dalam merangsangkan antibodi nyah-virus yang berupaya menentang

jangkitan virus dalam ayam, maka ia telah banyak digunakan dalam penghasilan vaksin.

Satu strain asing dari strain vaksin V4QUE yang dikenali sebagai V4UPM(HR) telah di

kembangkan sebagai vaksin stabil haba untuk kegunaan industri penternakan di negara

tropika misalnya Malaysia. Protein in mungkin juga terlibat dalam mengekalkan kestabilan

haba untuk sesetengah strain vaksin. Dalam kajian ini, gen HN dari strain NDV asing

V4UPM(HR) dan strain induknya V4QUE telah diklon dan diekspres dalam Sistem Vektor

Ekspresi Baculovirus (BEVs) dan telah ditentupastikan kestabilan habanya.

Gen HN yang bersaiz 1.9 kb dari strain NDV tersebut telah digandakan dari RNA

genomiknya melalui RT-PCR dan seterusnya diklon secara uni-arah ke dalam plasmid

pemindah baculovirus, pCR Bac4.8TM. Plasmid-plasmid rekombinan baculovirus ini

vi

kemudiannya telah ditranfikasi bersama DNA baculovirus, *Bac-N-Blue*TM yang dilinearkan, ke dalam sel serangga *Spodoptera frugiperda* (*Sf9*). Baculovirus rekombinan yang dinyatakan sebagai *recHNV4UPM(HR)* dan *recHNV4QUE* ini, telah ditulenkan melalui asai plak. Glikoprotein HN rekombinan (*recHN*) yang dihasilkan dalam sel serangga *Sf9* ini menunjukkan aktiviti-aktiviti hemaglutinasi (HA) dan neuraminidase (NA), dan juga aktiviti-aktiviti perencatan hemaglutinasi dan hemaserapan dalam asai-asai serologi. Namum aktiviti-aktiviti HA dan NA ini juga dapat dikesan pada permukaan mahupun dalam sitoplasma sel-sel *Sf9* yang terjangkit. Analisis SDS-PAGE dan Western blot terhadap lisat sel *Sf9* yang dijangkiti baculovirus rekombinan telah menunjukkan jalur protein pada lebih kurang 74 kDa, di mana ia bepadanan dengan protein HN yang berglikolasi dari virus. Keputusan ini menandakan *recHN*s bukan sahaja telah berjaya dihasilkan dalam sel-sel *Sf9*, bahkan ia menampilkan keaktifan dan berfunsi biologinya.

Berdasarkan aktiviti HA, kestabilan terma recHNV4QUE dan recHNV4UPM(HR) bersama dengan recHNAF2240 telah dikaji dan dibandingkan dengan strain-strain NDV lain, misalnya V4QUE, V4UPM(HR) dan AF2240. Namun begitu, strain-strain yang kemudian itu menunjukkan bahawa ia mengekalkan kestabilan haba pada suhu ~56°C terlebih dahulu. Keputusan-keputusan menunjukkan fenotip rintangan haba bagi baculovirus-baculovirus rekombinan adalah bersamaan secara genetik dengan individu-individu NDV dari segi sifat kestabilan terma. Strain-Strain rintang haba seperti AF2240 dan V4UPM(HR) serta baculovirus rekombinan recHNAF2240 dan recHNV4UPM(HR) dinyahaktifkan sebanyak 50% pada ~56°C selepas 4 jam. Namun begitu, strain induk NDV



V4QUE dan baculovirus rekombinan strain recHNV4QUE tetap merupakan strain sensitif haba.

Gen-gen HN rekombinan ini turut dijujuk dan protein-protein telah dianalisis melalui ramalan struktur sekunder dan tiga dimensi. Peranan-peranan jujukan asid amino individu pada protein HN dalam kestabilan termanya juga dibincangkan. Rantai sisi berkutub/tak berkutub pada asid amino yang digantikan residues [R32, V413E, N79K di strain V4UPM(HR); E494K di strain V4UPM, di mana rantai sisi berkutub dapat mewujubkan dwikutub untuk pembentukan ikatan hidrogen dengan persekitaran akueous dan peningkatan kestabilan terma sesuatu protein. Nilai-nilai hidrofobik dan penyusunan struktur sekundernya pada jujukan asid amino terganti [F151Y, I175M, G431S di strain V4UPM; A152T, N276K, I280T, V303T, Q372R, Y504H di strain AF2240] dan [R32, G221, R305, V413E, S584A di strain V4UPM(HR); H87Q, E494K di strain V4UPM membentuk struktur sekunder protein bagi pelipatan dan penyusunannya. Protein-protein yang stabil haba lebih cenderung untuk membentuk struktur heliks-α/kepingan-β daripada campuran kepingan-β/lingkaran rawak atau struktur lingkaran rawak sahaja pada domain HN anggapan (399-GAEGR-403) seperti yang diramalkan menerusi penyusunan struktur sekundernya. Selain itu, struktur 3D protein HN menyokong bahawa sebarang pergantian jujukan asid amino berhampiran dengan tapak domain aktif anggapan (domain HN 399-<u>GAEG</u>R-403) mungkin akan mengubah struktur tertiar protein HN dan struktur tertiar/penyusunan struktur protein HN. Ia seterusnya mempengaruhi kestabilan haba pada protein HN.



ACKNOWLEDGEMENTS

I would like to express my deepest appreciation to Prof Datin Dr. Khatijah Mohd Yusoff, Prof Dr. Abdul Manaf Ali, Dr. Tey Beng Ti and Dr. Alan Ong Han Kiat for their advice, patient guidance and support in my project. My special gratitude also went out to Dr. Tan Wen Siang for his kindness and helpful encouragement which had helped me to complete my project.

It had been a great memory and experience to work with my colleagues in Lab. 143, 134 at Department of Biochemistry and Microbiology and Department of Biotechnology, Animal Tissue Culture Lab. Thank went to Loke, Pria, Subha, Lau, Kho, Ho, Wong, Kok, Majid, Amir, Ong for their friendship, Dr. Azri's support from Jabatan Perkhimatan Haiwan, Petaling Jaya, Miss Chee Siew Yong from Universiti Malaya and Dr. Chan Siew Wee from National University of Singapore, *IMCB*. Mr. Ng from Bio-Diagnostic, Miss Yau from Bio-Syn-Tech and Encik Adali from Linggi Poultry Farm Sdn. Bhd. had also provided technical services and support to my project.

With all the experiments, I would like to thank Prof Datin Khatijah Mohd Yusoff who had provided me the opportunity to attend workshops in *CGAT-UKM*, Universiti Malaya and the support of the Malaysian Ministry of Science, Technology and Environment IRPA grant funding. Finally, a special thank to the Lee, Noe and Tong families, my friends; Lock, Lim Koh, Wee Horng, Foong Yee, Thian Hann, Meng Teck, Bei Looi who graduated from Taiwan and my family, parents' and sisters' supports over the years. Without them, this master thesis would have not been possible.



This study was supported by Malaysian Ministry of Science, Technology and the Environment IRPA Grant No. 01-02-04-0107.



TABLE OF CONTENTS

				Page
DEDI	CATIO	N		ii
ABST	CRACT			iii
ABST	RAK			vi
ACKI	NOWLE	EDGEM	ENTS	ix
APPR	ROVAL			xi
DECI	LARATI	ION		xiii
LIST	OF FIG	URES		xviii
LIST	OF TAI	BLES		xx
LIST	OF ABI	BREVIA	ATIONS	xxi
LIST	OF AM	INO AC	CIDS AND ABBREVIATIONS	xxiv
СНА	PTER			
1	INTR	ODUCI	TION	1
2	LITER	RATUR	E REVIEW	4
	2.1	Newca	astle Disease	4
		2.1.1	Genome Organization and Virion Properties of NDV	4
	2.2	Contro	ol of Newcastle Disease in Malaysia	5
	2.3	Therm	nostability of Newcastle Disease Vaccine	6
	2.4	Therm	nostability of Proteins	9
		2.4.1	The Effects of Temperature on Protein Structure	9
		2.4.2	· · · · · · · · · · · · · · · · · · ·	10
	2.5	The H Virus	Iaemagglutinin-neuraminidase (HN) Gene of Newcastle Disease	12
		2.5.1	The Biological and Molecular Characteristic in Temperature	12
		2.3.1	Sensitive Mutants of the Newcastle Disease Virus	13
		2.5.2	The Crystal Structure of the Multifunctional HN Glycoprotein	14
	2.6		cole of Individual Amino Acid Residues in the Formation of the	14
	2.0		e, Functional HN Glycoprotein of NDV	16
		2.6.1		10
			Arrangement in HN Gene	16
		2.6.2	The Role of the Oligosaccharide Chains in HN Gene	18
		2.6.3		19
	2.7		ovirus Expression Vector System (BEVs)	21
		2.7.1	Biology and Classification of Baculovirus	21
		2.7.2		21
		2.7.3	Principles of BEVs	24
		2.7.4	•	
			Characteristics of Insect Cell Lines	25
		2.7.5	Insect Cell Culture Medium	26
		2.7.6	Allelic Replacement of Homologous Recombinant Techniques	8
			in BEVs	27



	2 8	2 7 7 Advantages of the <i>BEV</i> 's Computational Biology and Bioinformatic	28 31
3	MAT	ERIALS AND METHODS	35
3	3 1	Chemicals, Reagents, Solvents and Tissue Culture Materials	35
	3 2	Viruses, Bacteria and Insect Cell Line	35
	3 3	Vectors, Viral DNA, Plasmids and Primers	37
	3 4	Buffers, Antibiotics Solution and Media	37
	3 5	Nucleic Acid Extraction	41
	3 3	3 5 1 NDV Viral RNA Extraction	41
		3 5 2 Baculoviral DNA Extraction from the Infected <i>Sf9</i> Cell	41
	3 6	RT-PCR Procedures	43
	5 0	3 6 1 RT-PCR Reaction	43
		3 6.2 Purification of RT-PCR Product	44
	3 7	Cloning, Screening and Verification of Putative Recombinant Clones	45
	5 ,	3 7 1 Ligation of RT-PCR Product into Baculovirus Transfer Vector	45
		3 7 2 Preparation of Competent Cells	45
		3 7 3 Introduction of the HN Gene into Competent Cells	45
		3 7 4 Small Scale Plasmid DNA Extraction	46
		3 7 5 Restriction Enzyme (RE) Analysis of the Recombinant	
		Baculovirus Transfer Plasmid	47
		3 7 6 PCR Verification of the HN Gene	47
		3 7 7 DNA Sequencing	48
		3 7 8 RNA Sequencing	48
	3 8	Tissue Culture Techniques of the Insect Cell	49
		3 8 1 General Handling Techniques	49
		3 8 2 Insect Cell Culture Medium	50
		3 8 3 Heat-Inactivation of the Fetal Bovine Serum	50
		3 8 4 Monolayer Cell Culture and Passaging (Subculturing) Cell	
		Culture	51
		3 8 5 Cryopreservation and Storage of Insect Cell Line	51
	3 9	Transfection Procedures	52
		3 9 1 Co-transfection of the Recombinant Transfer Vector and the	
		Linearized Baculoviral DNA	52
		3 9 2 Post-transfection Procedure	53
	3 10	Viral Plaque Assay and Identification of Putative Recombinant	
		Baculovirus	53
		3 10 1 Preparation of Sf9 Insect Cells and Dilution of Recombinant	
		Viruses	53
		3 10 2 Viral Plaque Assay	54
		3 10 3 Visual Screening for Blue Plaques	55
		3 10 4 PCR Verification	55
		3 10 5 Partial DNA Sequencing at End Termini of the Recombinant	
		Baculoviral DNA	56



	3.11	Confo	rmation of the Expressed HN Protein in Baculovirus Expression	
		System	1	57
		3.11.1	Haemagglutinin Activity (HA)	57
		3.11.2	Neuraminidase Activity (NA)	57
			Haemagglutination Inhibition (HI)	58
			Haemadsorption Assay	58
			Sodium Dodecyl Sulphate-Polyacrylamide Gel Electrophoresis	
			(SDS-PAGE) of the Baculovirus Infected Sf9 Cells Lysis	59
		3 11 6	Western Blot Analysis of the Recombinant HN Glycoprotein	60
	3.12		ostability Studies of the Recombinant HN Protein	61
	5.1 2		Thermostability Assay of the Haemagglutinin Activity	61
			Stability Studies of the Haemagglutinin Activity	61
4	RESU	LTS AN	ND DISCUSSION	62
	4.1	Revers	se-Transcriptase Polymerase Chain Reaction (RT-PCR) and	
			erase Chain Reaction (PCR) of NDV Nucleic Acid	62
		4.1.1	· · ·	62
	4.2		ng of the HN Gene into a Cloning and Shuttle Vector	63
		4.2.1	Cloning of the HN Gene into a Cloning Vector, pCR2.1	63
		4.2.2		65
		4.2.3		
		1.2.0	Recombinant Baculovirus Transfer Plasmids	67
		4.2.4	Partial End Termini DNA Sequencing of the Putative	0,
		1.2.1	Recombinant Plasmids	72
	4.3	Mornh	pology of the Sf9 Cells and the Infected Sf9 Cells Culture	72
	1.5	4.3.1	General Morphology of the Sf9 Cells	72
		4.3.2	Morphology of the Baculoviral Infected Sf9 Cells	72
	4.4		nsfection of the Recombinant Baculovirus Plasmid with Linearized	12
	1. 1		oviral DNA into the Sf9 Insect Cells	73
			Confirmation of the Transfection Reaction	73
		4.4.2		13
		4.4.2	· · · · · · · · · · · · · · · · · · ·	77
		1 1 2	Assay Identification of the Putative Recombinant Reculovirus by PCR	//
		4.4.3	Identification of the Putative Recombinant Baculovirus by PCR Procedure	79
		1 1 1		19
		4.4.4	Partial DNA Sequencing at the End Termini of the Recombin Viral DNA	79
	4.5	Evpros	ssion of the HN Glycoprotein in the Sf9 Cells	82
	4.3	4.5.1	Serological Assay and Haemagglutination Inhibition Test	82
		4.5.1	SDS-PAGE and Western Blot Analysis of the Recombinant HN	02
		4.3.2	Glycoprotein	86
	4.6	Thorm	ostability Assay of NDV Strains and the Recombinant	80
	4.0		oviruses on HA Activities	റാ
		4.6.1		92
		4.0.1	Thermostability Assay of NDV Strains and the Recombinant Baculoviruses	റാ
		1.60		92
		4.6.2	Stability Response of the HA Activity at 56°C Heat Stress	93



	4.7	Therm	ostability of the HN Polypeptide	96
		4.7.1	Single Amino Acid Residues Change in the HN Polypeptide	101
		4.7.2	of NDV Strain <i>V4UPM(HR)</i> Single Amino Acid Residues Change in the HN Polypeptide	101
		7.7.2	of NDV Strain V4UPM	107
		4.7.3		
		474	of NDV Strain AF2240	110
		4.7.4	Glycosylation Sites and Cysteine Residues of the HN Polypeptide of NDV Strains	115
		4.7.5		- 10
			Polypeptide	116
5	SUMM	IARY A	AND CONCLUSION	123
REFEI	RENCE	S		127
APPE	NDICES	S		142
VITA				159



LIST OF FIGURES

Figure		Page
2 1	Schematic Diagram of Various Forms of Baculovirus	22
2 2	The Infection Cycle of the Baculovirus BV and OV in Insect Cells	23
2 3	Illustration of Allelic Replacement Reaction in BEVs	29
4 1	Amplified HN Genes from NDV Strains AF2240, V4QUE, V4UPM(HR)	64
4 2(a)	Screening of the Putative Recombinant Clones by Single Digested	
. ,	with <i>Hınd</i> III	66
4 2(b)	Screening of the Putative Recombinant Clones by Double Digested	
` '	with BamHI and HindIII	66
4 3(a)	Screening of the Putative Recombinant Baculovirus Transfer Plasmid	
()	by Double Digested with <i>Hınd</i> III and BamHI	68
4 3(b)	Restriction Enzyme Analysis of the Putative Recombinant Baculovirus	
()	Transfer Plasmids	69
4 4	PCR Verification of the Putative Recombinant Plasmids	70
4 5	The Recombinant Baculovirus Transfer Plasmid	71
4 6	Partial DNA Sequencing at the End Termini of the Recombinant	
	Baculovirus Transfer Plasmid	74
4 7	Sf9 Insect Cell Line	75
4 8	Signs of the Baculoviral Infection	76
4 9	Schematic Diagram of Blue Plaque Formation of the Recombinant	
	Baculovirus	78
4 10	PCR Verification of the Recombinant Baculoviruses	80
4 11	Partial DNA Sequencing at the End Termini of the Recombinant	
	Viral DNA	81
4 12	The Hemagglutinin Activity (HA) of the Recombinant Baculoviruses	
	and Wild Type Baculovirus	83
4 13	The HA Activity of Recombinant Baculoviruses	84
4 14	Haemadsorption Assay of the Recombinant Baculovirus Infected Sf9	
	Cell Culture	85
4 15	The NA Assay was also Done on the Infected Cells Lysates by Using	0.0
	the Quantitative Estimation of <i>N</i> -acetylneuraminic Acid	87
4 16	The Haemagglutination Inhibition Test of Recombinant Baculoviruses	88
4 17	Pattern of Fractionated Protein in 10% SDS-PAGE	90
4 18	Western Blot Analysis of Infected Cell Lysates by Using NDV Antisera	70
, 10	Polyclonal Antibodies	91
4 19	Thermostability Assay on the Haemagglutinin Activity (HA) of NDV	71
	Strains V4QUE, V4UPM(HR), AF2240 and Recombinant Baculovirus	
	Strains recHNV4QUE, recHNV4UPM(HR), recHNAF2240. HA	
	Activity Remaining vs Temperature are Plotted	94
4 20	Stability Response of the Haemagglutinin Activity at 56°C Heat	<i>-</i> 1
. 20	Stress HA Activity Remaining vs Hours of Heat Stress are Plotted	95
4 21	CLUSTAL W (1 81) Multiple Sequence Alignment of the HN Protein	75
1 21	of NDV Strains V4QUE, V4UPM(HR), V4UPM and AF2240	97
	orran a priming a straight of a straight of the straight of th	71



4.22	: Multiple Sequence Alignment of the HN Polypeptide. Secondary		
	Structure of HN Proteins; α -helix, β -sheet and Random Coil		
	Structure were Predicted	104	
4.23	The Tertiary Structure of the HN Protein	121	
4.24	: The Tertiary Structure of the HN Protein Represented in a Ribbon		
	at this Domain	123	



LIST OF TABLES

Table		Page
3 1	List of Baculovirus and Recombinant Virus, NDV, Bacteria and Insect Cell	36
3 2	List of Vectors, Linearized Baculoviral DNA and Plasmids	38
3 3	List of Primers	39
3 4	List of Buffers, Antibiotics Solution and Media	40
4 1	Matrix of Amino Acid Residues between Thermophilic and Mesophilic	
	Protein Sequence	99
4 2	Favorable Amino Acid Replacements from Mesophilic to Thermophilic	
	Proteins	100
4 3	Nucleotide Changes, Deduces and Amino Acid Substitution in Heat	
	Stability NDV Strain V4UPM(HR) were Compared to its Published	
	Parental (Heat Sensitive) Strain V4QUE	102
4 4	Nucleotide Changes, Deduces and Amino Acid Substitution in Heat	
	Stability NDV Strain V4UPM were Compared to its Published Parental	
	(Heat Sensitive) Strain V4QUE	108
4 5	Nucleotide Changes, Deduces and Amino Acid Substitution in Heat	
	Stability NDV Strain AF2240 were Compared to its Published	
	Parental (Heat Sensitive) Strain V4QUE	111
4 6	List of Amino Acid Residues Change in NDV Strains V4UPM(HR),	
	V4UPM and AF2240 that May Contributed towards the Thermostability	
	of HN Polypeptide	114



LIST OF ABBREVIATIONS

A

A Ampere

AcMNPV Autographa Californica Multiple Nuclear Polyhedrosis Virus

Amp Ampicillin α Alpha

B

 β Beta

BCP 1-bromo-3-chloropropane

BEVs Baculovirus Expression Vector system

BmNPV Bombyx mori (Silkworm) Nuclear Polyhedrosis Virus

BV Budded Virus

 \mathbf{C}

cDNA Complementary Deoxyribonucleic Acid

CRBC Chicken Red Blood Cells

CO₂ Carbon Dioxide

D

ddNTP Dideoxynucleotide Triphosphates

DEPC Diethylpyrocarbonate
dH2O Distilled Water
DMF Dimethylformamide

DMSO Dimethyl Sulfoxide
DNA Deoxyribonucleic Acid

dNTP Deoxynucleotide Triphosphate

 \mathbf{E}

ECV Extracellular Virus

EDTA Ethylenediaminetetraacetic Acid Disodium Salt

E. coli Escherichia coli

 \mathbf{F}

F Fusion Protein

Fig. Figure

FBS Fetal Bovine Serum

 \mathbf{G}

Gal Glyco g Gram xg x Gravity

Н

h Hour

UPM

HA Haemagglutinin Activity
HAU Haemagglutinin Activity Unit
HI Haemagglutination Inhibition

HN Haemagglutinin-neuraminidase Protein

HR Heat Resistance

I

ICTV The International Committee in the Taxonomy of Viruses

K

KCl₂ Potassium chloride

Kb Kilobase kDa Kilodalton

L

l liter

ln Base 2 Logarithms L Polymerase Protein

M

Matrix Protein

m Milli ml Milli Liter mM Milli Molar

MCS Multiple Cloning Sites

min Minute M Molar

Mr Molecular Weight

N

NANA N-aceylneuraminic Acid NA Neuraminidase Activity ND Newcastle Disease

NDV Newcastle Disease Virus

n nino

NOVs Non-occluded Virus NP Nucleoprotein

 \mathbf{O}

ooc Absence of Occlusion Bodies ooc Presence of Occlusion Bodies

ORF Open Reading Frame
OV Occlusion Virus

P

PBS Phosphate Buffer Saline



PCR Polymerase Chain Reaction
PIBs Polyhedral Inclusion Bodies
polh Polyhedrin Genes of Baculovirus

P Phosphoprotein

p pico

PVDF Polyvinylidene Difluoride Membranes

R

RBC Red Blood Cells rec Recombinant

recHN Recombinant Haemagglutinin-Neuraminidase Glycoprotein

RE Restriction Enzymes Analysis

RNA Ribonucleic Acid rpm Round per Minute

RT-PCR Reverse Transcriptase-Polymerase Chain Reaction

RT Room Temperature

S

SD Standard Deviation

SDS-PAGE Sodium Dodecyl Sulphate-Polyacrylamide Gel Electrophoresis

sec Second

Sf-900IISFM Sf-900II Serum Free Medium
Sf9 Spodoptera frugiperda
SFM Serum Free Medium

T

TaqThermus AquaticusTAETris-acetate EDTA BufferTBETris-boric EDTA Buffer

TE Tris-base Buffer
Tm Melting Temperature
Tn Trichoplusia ni

U

 $\begin{array}{cc} \mu & \text{micron} \\ \text{UV} & \text{Ultra Violet} \end{array}$

U Unit

UPM Universiti Putra Malaysia

V

v Volume V Volt

W

wt Wild Type Weight



LIST OF AMINO ACIDS AND ABBREVIATIONS

A Alanine (Ala)

C Cysteine (Cys)

D Aspartic Acid (Asp)

E Glutamic Acid (Glu)

F Phenylalanine (Phe)

G Glycine (Gly)

H Histidine (His)

I Isoleucine (Ile)

K Lysine (Lys)

L Leucine (Leu)

M Methionine (Met)

N Asparagine (Asn)

Proline (Pro)

Q Glutamine (Gln)

R Arginine (Arg)

S Serine (Ser)

T Threonine (Thr)

V Valine (Val)

W Tryptophan (Trp)

Y Tyrosine (Tyr)

