

UNIVERSITI PUTRA MALAYSIA

INVESTIGATION ON INDIGENOUS BACILLUS ISOLATES WITH BIOREMEDIATION PROPERTIES FOR IMPROVING WATER QUALITY AND SHRIMP HEALTH IN MALAYSIAN AQUACULTURE

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By DEVARAJA T.N.

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia in Fulfilment of the Requirement for the Degree of Doctor of Philosophy

U P M

Dedicated to my parents, who supported me to become what I want

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

INVESTIGATION ON INDIGENOUS BACILLUS ISOLATES WITH BIOREMEDIATION PROPERTIES FOR IMPROVING WATER QUALITY AND SHRIMP HEALTH IN MALAYSIAN AQUACULTURE By

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Indigenous marine bacteria of the genus Bacillus were selected to study their properties as potential use for bioremediation owing to their inherent versatility. Bacteria were isolated from water and sediment samples collected along the west coast of Peninsular Malaysia in brackishwater environment. Selected isolates were identified to species level using biochemical and API CH kit and three suitable isolates, Bacillus pumilus AB58, B. subtilis AB65 and B. licheniformis AB69 were selected for the study. Optimum growth requirements of temperature, NaCl and pH were 30°C, 1.5% and 7.5 respectively, determined for the isolates by measuring the optical density and corresponding cell number. The growth curves of the isolates were plotted and all of them reached maximum cell number during a 16-20 h incubation. The cell density in overnight cultures of B. pumilus AB58, B. subtilis AB65 and B. licheniformis AB69 were $5.7x10^9 (\pm 0.8), 3.7x10^8 (\pm 0.6),$ $5.0x10^9$ (± 0.6) cfu/ml respectively. They had the ability to tolerate ammonia levels of up to 20 mg/l without a considerable change in cell numbers for 48 h. However, the growth was suppressed completely at 25 mg/l of ammonia. At 40 ppt salinity, all the isolates survived for 4 days without significant change in initial cell numbers (108 cfu/ml). The selected isolates were found to secrete extracellular enzymes viz., protease, gelatinase, amylase and



lipase as detected by clear zone formation on substrate based agar plates. Bacillus pumilus AB58 and B. subtilis AB65 produced significantly (P < 0.05) bigger protease clear zones $(19.0 \pm 2.0 \text{ and } 23.0 \pm 4.0 \text{ diameter in mm respectively})$ than B. licheniformis AB69. However, B. subtilis AB65 secreted significantly (P < 0.05) more amylase (31.0 \pm 5.0 diameter in mm) than the other two isolates. All the isolates were sensitive to most of the antibiotics tested on MHA plates. These isolates were compatible with each other in mixed culture conditions. They inhibited as well as excluded all the pathogenic vibrios (Vibrio alginolyticus M11, V. alginolyticus M12, V. parahaemolyticus M1, V. parahaemolyticus M3, V. parahaemolyticus M6, V. alginolyticus T, V. parahaemolyticus T, V. harveyi I and V. parahaemolyticus I) tested by diffusion disc, streak plate and common broth methods. Synergistic effect of isolates had significantly higher (P < 0.05) inhibition of all vibrios than the individual isolates. The isolates were confirmed for their non-pathogenicity to shrimp postlarvae (PL 29). All three isolates were tested for their effect on ammonia in simulated pond conditions. All non-aerated treatment tanks had significantly lower ammonia levels (P < 0.05) than the non-aerated control tanks, which were not treated with bacterial isolates both in case of single and combination treatments. Synergistic effect of isolates reduced the ammonia levels at a faster rate than the treatments with single isolate. Sediment properties were not significantly different between treated and control groups except for the total and available phosphorous levels, which were significantly higher in tanks treated with B. licheniformis AB69 (P < 0.05) compared to the others. The selected Bacillus isolates satisfied the criteria to qualify them for bioremediation in aquaculture.



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PENYIASATAN KE ATAS ISOLAT-ISOLAT *BACILLUS* TEMPATAN BERCIRIKAN BIOREMEDIASI BAGI MENINGKATKAN KUALITI AIR DAN KESIHATAN UDANG UNTUK AKUAKULTUR DI MALAYSIA

Oleh

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Dalam kajian ini, spesies *Bacillus* marin tempatan telah dipilih untuk bioremediasi kerana sifat serba boleh yang semulajadi. Bakteria daripada sampel air dan sedimen daripada persekitaran air payau telah di kutip dari sepanjang pantai barat Semenanjung Malaysia. Isolat-isolat yang terpilih telah dikenalpasti ke peringkat spesies dengan menggunakan kaedah biokimia dan kit tersedia API CH dan tiga isolat terbaik, *Bacillus pumilus* AB58, *B. subtilis* AB65 dan *B. licheniformis* AB69 telah dipilih untuk kajian ini. Paras optimum keperluan-keperluan pertumbuhan asas; suhu, garam dan pH masing-masing adalah 30°C, 1.5% dan 7.5, yang mana telah ditentukan untuk isolat-isolat tersebut dengan mengukur densiti optik dan bilangan sel sejajar. Lengkuk pertumbuhan isolat-isolat telah diplot dan didapati bahawa semua isolat mencapai bilangan sel maksimum pada 16-20 jam (j) inkubasi. Densiti sel di dalam kultur semalaman *B. pumilus* AB58, *B. subtilis* AB65 dan *B. licheniformis* AB69 adalah 5.7x10° (± 0.8), 3.7x10⁸ (± 0.6), 5.0x10° (± 0.6) cfu/ml masing-masing. Mereka berupaya menahan paras ammonia sehingga ke 20 mg/l tanpa perubahan pada bilangan sel selama 48 j. Walau bagaimanapun, pertumbuhan terencat sepenuhnya apabila paras ammonia mencapai



tahap 25 mg/l. Pada saliniti 40 ppt, semua isolat berjaya hidup selama 4 hari tanpa perubahan yang signifikan pada bilangan sel permulaan (10⁸cfu/ml). Isolat-isolat terpilih di dapati mengeluarkan enzim-enzim luar sel, iaitu protease, gelatinase, amilase dan lipase berdasarkan zon yang terang terhasil di atas piring agar. Bacillus pumilus AB58 dan B. subtilis AB65 lebih signifikan (P < 0.05) di dalam menghasilkan zon yang lebih terang (19.0 \pm 2.0, dan 23.0 \pm 4.0 diameter dalam mm masing-masing) berbanding dengan B. licheniformis AB69. Walau bagaimanapun, B. subtilis AB65 lebih signifikan di dalam merembes amilase (P < 0.05), iaitu (31.0 \pm 5.0 diameter dalam mm) berbanding dengan kedua-dua isolat yang lain. Kesemua isolat adalah sensitif kepada kebanyakan antibiotik yang diuji di atas piring MHA. Isolat-isolat ini adalah serasi di antara satu sama lain dalam keadaan kultur campuran. Mereka merencat dan menyingkirkan semua vibrios patogenik (Vibrio alginolyticus M11, V. alginolyticus M12, V. parahaemolyticus M1, V. parahaemolyticus M3, V. parahaemolyticus M6, V. alginolyticus T, V. parahaemolyticus T, V. harveyi I and V. parahaemolyticus I) apabila diuji dengan cakera resapan, piring coretan dan medium biasa. Kesan sinergistik oleh isolat-isolat adalah secara signifikannya lebih tinggi (P < 0.05) ke atas perencatan semua vibrios berbanding dengan isolat tunggal. Isolat-isolat juga telah disahkan tidak patogenik terhadap pasca larval udang (PL 29). Ketiga-tiga isolat telah diuji kesannya terhadap ammonia dalam kolam simulasi. Semua tangki rawatan yang tidak mengandungi pengudaraan menunjukkan paras ammonia yang secara signifikannya lebih rendah (P < 0.05) berbanding dengan tangki kawalan yang tidak mengandungi pengudaraan, samada secara berasingan atau campuran. Kesan sinergistik isolat-isolat dapat mengurangkan paras ammonia pada kadar yang lebih cepat berbanding dengan



rawatan menggunakan isolat tunggal. Kandungan sedimen adalah tidak signifikan di antara kumpulan yang dirawat dan kawalan kecuali untuk jumlah dan tahap tersedia fosforus, yang mana secara signifikannya lebih tinggi di dalam tangki yang dirawat dengan *B. licheniformis* AB69 (P < 0.05) berbanding dengan yang lain. Isolat-isolat *Bacillus* yang terpilih memenuhi kriteria untuk melayakkannya sebagai agen bioremediasi di dalam akuakultur.



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LIST OF ABBREVIATIONS

AAHU – aquatic animal health unit

ANOVA – analysis of variance

Bl – Bacillus licheniformis AB69

BOD – biological oxygen demand

Bp — Bacillus pumilus AB58

bp – base pair

Bs – Bacillus subtilis AB65

BWS – bacterial white spot

cfu – colony forming units

COD – chemical oxygen demand

DDW – double distilled water

DNA – deoxyribo nucleic acid

dNTPs – deoxyribonucleoside triphosphates

DO – dissolved oxygen

DW – distilled water

EDTA – ethylene dinitro tetraacetic acid

ERMs – environmentally relevant microorganisms

FAO - Food and Agriculture Organisation

GEMs – genetically engineered microorganisms

h – hour

kDa – kilo dalton

LD₅₀ – lethal dose 50

mmt – million metric tonne

mt – metric tonne

OD – optical density

PCR – polymerase chain reaction

ppt - parts per thousand

RAPD - random amplified polymorphic DNA

16S rRNA - 16 subunit ribosomal ribose nucleic acid

S.E. – standard error

TBE - tris boric acid EDTA

TCBS – thiosulphate citrate bile salt sucrose

TPC – total plate count

TSA – trypticase soy agar

TSB – trypticase soy broth

UPM – Universiti Putra Malaysia

USD – Dollar of United States of America

VaM11 – Vibrio alginolyticus Malaysia 11

VaM12 – Vibrio alginolyticus Malaysia 12

VaT – Vibrio algninolyticus Thailand

VhI – Vibrio harveyi Indonesia

VpI - Vibrio paraheamolyticus Indonesia

VpM1 – Vibrio parahaemolyticus Malaysia 1

VpM3 – Vibrio parahaemolyticus Malaysia 3

VpM6 – Vibrio parahaemolyticus Malaysia 6

VpT - Vibrio parahaemolyticus Thailand

CHAPTER 1

GENERAL INTRODUCTION

1.1 Background and Scope of the Study

In recent decades aquaculture has become a major food production industry helping to meet the increasing demand for food. The world population has crossed six billion during the year 2000 (Census Bureau, 2000) causing an increase in demand for food. Therefore, food production by agriculture and capture fisheries has to be supplemented by other alternatives. Total world fish and shellfish production has reached 126.17 million metric tonnes (mmt) in 1999 (FAO, 2000) of which, 33.31 mmt come from aquaculture.

Dehadrai (1993) predicted that aquaculture has to fill the gap of 19.6 mmt by 2000, 37.5 mmt by 2010 and 62.54 mmt by 2020. Among various aquaculture practices, shrimp culture is gaining increasing importance world-wide due to the short period of culture and high profits. Even though shrimp farming has grown to become a booming export oriented industry, shrimp production and trade have undergone fluctuations during the past four years indicating uncertainty over its sustainability in days to come due to different social and environmental problems.

The shrimp industry has to develop strategies to tackle the viral disease problems, which have disrupted shrimp farming. Generally in shrimp farming, the ultimate goal is to maximise the production with high stocking density, increase feeding and increase water exchange often accompanied with heavy use of chemicals. This has



caused various other problems like disease outbreaks, environmental pollution and other socio-economic fall out (Primavera, 1994). The shrimp industry is now looking for ways to rebuild itself and aims at a long-term sustainability that will not pollute the environment or minimise pollution.

The incidence and severity of several infectious diseases largely depends upon the quality of environment in which the host lives. Outbreak of diseases might be avoided by maintaining a healthy environment through suitable water quality management practices. Healthy environment can also be achieved by stocking at optimum density, providing sufficient water exchange and aeration (Wang et al., 1999a). In the past, water exchange was the only solution widely practised to get rid of the accumulated wastes but recent viral disease outbreak has limited this practice. The recent practice of using closed or semi-closed system with heavy chlorination to reduce introduction of viral infection is not popular due to the extra cost involved. Reducing stocking density and feeding rate to minimise the accumulation of organic matter has also indirectly decreased the rate of production thus reducing the profit margin (Phillips et al., 1993). Frequent water exchange to flush out shrimp pond effluent eutrophicates the coastal environment.

The ultimate strategy is to manipulate the pond environment by enhancing the mineralisation process in culture ponds so as to maintain a healthy environment and minimise the nutrient load to reduce pollution before disposing the effluents (Phillips et al., 1993). Several management practices have been adopted for maintaining optimal water and sediment quality like the use of chemicals, physical methods like bottom

