



UNIVERSITI PUTRA MALAYSIA

***DISPLAY OF HEPATITIS B VIRUS 'a' DETERMINANT ON THE  
SURFACE OF *Macrobrachium rosenbergii* (de Man, 1879)  
NODAVIRUS-LIKE PARTICLE CAPSID PROTEIN***

**NINYIO NATHANIEL NYAKAAT**

**FBSB 2020 29**



**DISPLAY OF HEPATITIS B VIRUS 'a' DETERMINANT ON THE  
SURFACE OF *Macrobrachium rosenbergii* (de Man, 1879) NODAVIRUS-LIKE  
PARTICLE CAPSID PROTEIN**

By

**NINYIO NATHANIEL NYAKAAT**

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,  
in Fulfilment of the Requirements for the Degree of Doctor of Philosophy

October 2020

## **COPYRIGHT**

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment  
of the requirement for the degree of Doctor of Philosophy

**DISPLAY OF HEPATITIS B VIRUS 'a' DETERMINANT ON THE  
SURFACE OF *Macrobrachium rosenbergii* (de Man, 1879) NODAVIRUS-LIKE  
PARTICLE CAPSID PROTEIN**

By

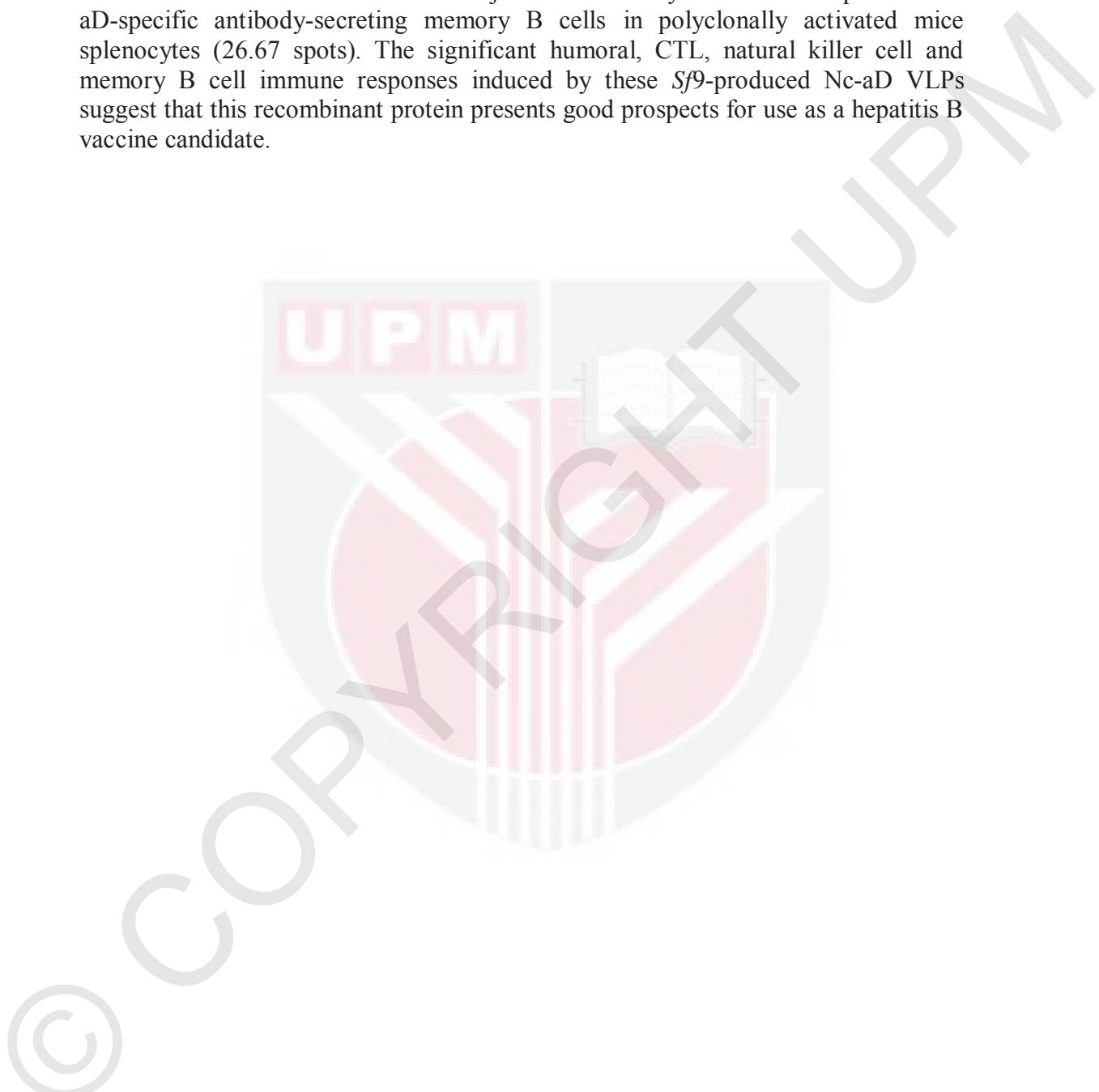
**NINYIO NATHANIEL NYAKAAT**

**October 2020**

**Chairman : Professor Wen Siang Tan, PhD**  
**Faculty : Biotechnology and Biomolecular Sciences**

Complications resulting from hepatitis B account for ~1.45 million deaths each year. To date, none of the available treatments is curative and the prophylactic hepatitis B vaccines can only protect ~90% of vaccinated individuals. Other limitations of current vaccination include poor immunogenicity in people with pre-existing conditions and people with unresponsiveness to yeast-derived vaccines. Overall, this necessitates the continuous development of novel hepatitis B vaccines with improved efficacy. This study is aimed at developing a novel hepatitis B vaccine candidate by producing a chimeric virus-like particle (VLP) displaying the hepatitis B virus (HBV) 'a' determinant (aD). The aD is the immuno-dominant region of HBV that induces the production of HBV-neutralising antibodies during infection. Furthermore, aD is conserved among different strains of HBV making it suitable for use in the development of HBV vaccines. In this study, the aD was fused to the C-terminus of the *Macrobrachium rosenbergii* nodavirus (*MrNV*) capsid protein (Nc) and expressed in *Spodoptera frugiperda* (*Sf9*) cells. SDS-PAGE analysis showed that the expressed protein was ~52 kDa in size. Subsequently, dynamic light scattering (DLS) analysis revealed that the recombinant Nc-aD protein assembled into heterogeneous particles ranging from ~23.4 to ~58.0 nm in diameter. Also, transmission electron microscopy (TEM) confirmed that these particles were spiky spherical virus-like particles (VLPs) with a diameter ranging from ~21 to ~55 nm. Circular dichroism (CD) spectroscopy further revealed that these Nc-aD VLPs consisted of  $\beta$ -sheets (44.8%), random coils (38.7%),  $\alpha$ -helices (16.1%) and  $\beta$ -turns (0.3%) with a melting temperature (Tm) of ~56.2 °C. Furthermore, enzyme-linked immunosorbent assay (ELISA) of these Nc-aD VLPs revealed that the aD was significantly antigenic when probed with the anti-hepatitis B surface antigen (HBsAg) monoclonal antibody. Subcutaneous immunisation of BALB/c mice with three doses of these purified Nc-aD VLPs (100  $\mu$ L; 0.34 mg/mL) elicited a robust humoral immune response that was sustained for 126 days. The elicited humoral immune response was significantly higher ( $p < 0.001$ )

than those elicited by a commercially available hepatitis B vaccine and those of *Escherichia coli*-produced Nc-aD. In addition, immunophenotyping showed that the *Sf9*-produced Nc-aD VLPs induced an increase of cytotoxic T-lymphocytes (CTL) (0.65 CD8+/CD4+ ratio) and NK1.1 natural killer cells (13.8%). Memory B cell enzyme-linked immunospot (ELISPOT) analysis was performed 126 days after the administration of the second booster injection. The analysis showed the presence of aD-specific antibody-secreting memory B cells in polyclonally activated mice splenocytes (26.67 spots). The significant humoral, CTL, natural killer cell and memory B cell immune responses induced by these *Sf9*-produced Nc-aD VLPs suggest that this recombinant protein presents good prospects for use as a hepatitis B vaccine candidate.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PEMAPARAN PENENTU 'a' HEPATITIS B VIRUS PADA PERMUKAAN  
PARTIKEL PROTEIN KAPSID MENYERUPAI NODAVIRUS**  
*Macrobrachium rosenbergii* (de Man, 1879)

Oleh

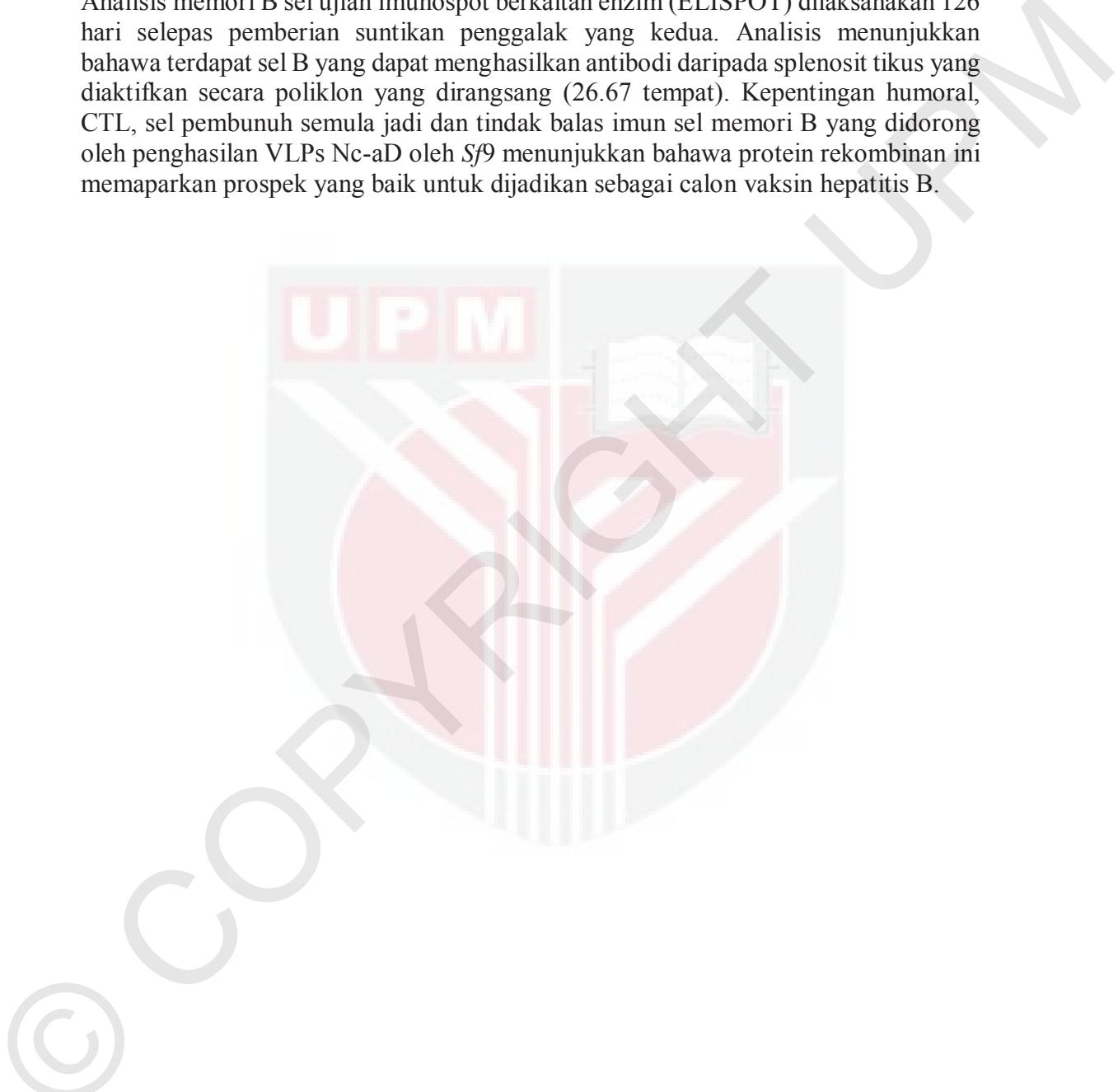
**NINYIO NATHANIEL NYAKAAT**

**Oktober 2020**

Pengerusi : Profesor Wen Siang Tan, PhD  
Fakulti : Bioteknologi dan Sains Biomolekul

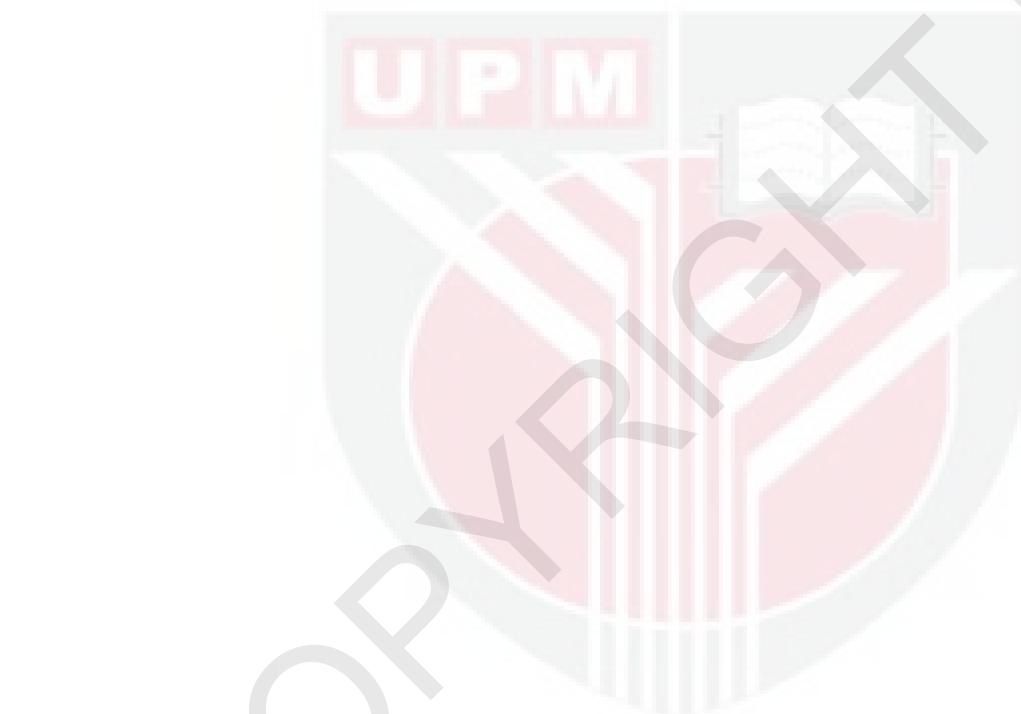
Komplikasi akibat hepatitis B menyumbang kepada ~1.45 juta kematian setiap tahun. Sehingga kini, rawatan yang sedia ada tidak dapat memberikan kesembuhan dan vaksin pencegahan hepatitis B hanya dapat melindungi ~90% individu yang telah divaksinasi. Limitasi lain vaksin semasa termasuklah keimunogenan yang rendah pada individu dengan keadaan sedia ada dan individu yang tidak bertindak balas dengan vaksin yang berasaskan yis. Secara keseluruhannya, vaksin hepatitis B ini perlu dibangunkan secara berterusan dengan keberkesanan yang lebih baik. Kajian ini bertujuan untuk membangunkan vaksin hepatitis B dengan menghasilkan partikel menyerupai virus (VLP) yang terdiri daripada virus hepatitis B (HBV) penentu 'a' (aD). aD adalah kawasan dominan-immuno pada HBV yang akan mendorong penghasilan antibodi peneutralan HBV semasa jangkitan. Tambahan pula, aD ini terpelihara di antara strain HBV yang berbeza yang menjadikannya sesuai digunakan untuk perkembangan vaksin HBV. Di dalam kajian ini aD telah dicantumkan ke terminal-C protein kapsid (Nc) *Macrobrachium rosenbergii* nodavirus (*MrNV*) dan diekspresskan di dalam sel *Spodoptera frugiperda* (*Sf9*). Analisis SDS-PAGE menunjukkan saiz protein yang diekspresskan adalah ~52 kDa. Selain itu, analisis penyebaran cahaya dinamik (DLS) menunjukkan protein rekombinan Nc-aD berkumpul menjadi partikel heterogenus berdiameter di antara ~23.4 hingga ~58 nm. Mikroskopi elektron transmisi (TEM) menunjukkan bahawa zarah-zarah ini adalah zarah menyerupai virus dengan diameter di antara ~21 hingga ~55 nm. Spektroskopi dikromisme pekeliling (CD) seterusnya mendedahkan bahawa VLP Nc-aD terdiri daripada helaian beta (44.8%), gegelung rawak (38.7%), heliks alpha (16.1%) dan selekoh beta (0.3%) dengan suhu lebur (T<sub>m</sub>) ialah ~56.2°C. Seterusnya, pemeriksaan imunosorben berkait enzim (ELISA) dengan VLP Nc-aD ini mendedahkan bahawa aD adalah sangat antigenik apabila diuji dengan antibodi monoklonal terhadap antigen permukaan hepatitis B (HBsAg). Imunisasi subkutan tikus BALB/c dengan tiga dos VLPs Nc-aD yang telah ditulenkhan (100 µL; 0.34 mg/mL) menunjukkan tindak balas

imun humoral yang kuat yang dikekalkan selama 126 hari. Tindak balas imun humoral yang ditunjukkan adalah jauh lebih tinggi ( $p<0.001$ ) daripada yang telah ditunjukkan oleh vaksin hepatitis B yang telah ada secara komersial dan vaksin Nc-aD yang dihasilkan oleh *Escherichia coli*. Selain itu, imunofenotiping menunjukkan bahawa VLPs Nc-aD yang dihasilkan oleh *Sf9* mendorong proliferasi sitotoksik T-limfosit (CTL) (0.65 CD8+/CD4+ ratio) dan sel pembunuhan semula jadi NK1.1(13.8%). Analisis memori B sel ujian imunospot berkaitan enzim (ELISPOT) dilaksanakan 126 hari selepas pemberian suntikan penggalak yang kedua. Analisis menunjukkan bahawa terdapat sel B yang dapat menghasilkan antibodi daripada splenosit tikus yang diaktifkan secara poliklon yang dirangsang (26.67 tempat). Kepentingan humoral, CTL, sel pembunuhan semula jadi dan tindak balas imun sel memori B yang didorong oleh penghasilan VLPs Nc-aD oleh *Sf9* menunjukkan bahawa protein rekombinan ini memaparkan prospek yang baik untuk dijadikan sebagai calon vaksin hepatitis B.



## **ACKNOWLEDGEMENTS**

My sincere appreciation goes to the chairman of my supervisory committee, Prof. Dr. Tan Wen Siang and the supervisory committee members: Associate Professor Dr. Ho Kok Lian, Associate Professor Dr. Chee Hui Yee and Associate Professor Dr. Muhajir Hamid. Thank you for your mentorship and support throughout this project. I am also grateful to my laboratory members for their comradery. To all staff members in the Faculty of Veterinary Medicine, Faculty of Biotechnology and Biomolecular Sciences, Faculty of Medicine and Health Sciences and the Institute of Bioscience, UPM who helped me during the course of my research, thank you for your generous services and technical support. I am also very grateful to my family, siblings and friends for their motivational support.



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

**Wen Siang Tan, PhD**

Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Chairman)

**Muhajir Hamid, PhD**

Associate Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Member)

**Kok Lian Ho, PhD**

Associate Professor

Faculty of Medicine and Health Sciences

Universiti Putra Malaysia

(Member)

**Hui Yee Chee, PhD**

Associate Professor

Faculty of Medicine and Health Sciences

Universiti Putra Malaysia

(Member)

---

**ZALILAH MOHD SHARIFF, PhD**

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date: 10 December 2020

## **Declaration by graduate student**

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No: Ninyio Nathaniel Nyakaat , GS47674

## **Declaration by Members of Supervisory Committee**

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature:

Name of Chairman  
of Supervisory  
Committee:

Professor Dr. Wen Siang Tan

Signature:

Name of Member  
of Supervisory  
Committee:

Associate Professor Dr. Muhajir Hamid

Signature:

Name of Member  
of Supervisory  
Committee:

Associate Professor Dr. Kok Lian Ho

Signature:

Name of Member  
of Supervisory  
Committee:

Associate Professor Dr. Hui Yee Chee

## TABLE OF CONTENTS

	Page
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	iii
<b>ACKNOWLEDGEMENTS</b>	v
<b>APPROVAL</b>	vi
<b>DECLARATION</b>	viii
<b>LIST OF TABLES</b>	xiii
<b>LIST OF FIGURES</b>	xiv
<b>LIST OF ABBREVIATIONS</b>	xvi
 <b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	1
<b>2 LITERATURE REVIEW</b>	4
2.1 Hepatitis B Virus	4
2.1.1 Morphology of HBV	6
2.1.2 Cell tropism and replication of HBV	10
2.1.3 Mechanisms of HBV persistence	11
2.1.4 Hepatitis B virus surface antigen (HBsAg)	12
2.2 Hepatitis B Vaccine	14
2.2.1 Serum-derived hepatitis B vaccines	15
2.2.2 Yeast derived hepatitis B vaccines	15
2.2.3 Hepatitis B vaccine production in prokaryotic cells	16
2.2.4 Plant-based hepatitis B vaccine production	17
2.2.5 Hepatitis B vaccine production in mammalian expression systems	18
2.2.6 Insect cell line-derived hepatitis B vaccines	19
2.2.7 Chimeric HBV vaccines with epitope display	20
2.3 Justification for Continuing Hepatitis B Vaccine Development	21
2.4 <i>Macrobrachium rosenbergii</i> Nodavirus ( <i>MrNV</i> )	22
2.4.1 The <i>Macrobrachium rosenbergii</i> nodavirus genome and the major proteins	23
2.4.2 The <i>MrNV</i> capsid protein	24
2.5 Importance of Virus-like Particles (VLPs)	25
2.6 Animal Models	25
<b>3 MATERIALS AND METHODS</b>	29
3.1 Materials	29
3.2 Plasmid Extraction via Alkaline Lysis	30
3.3 PCR and DNA Purification	31
3.4 Restriction Enzyme Digestion and DNA Ligation	32
3.5 Competent Cell Preparation and Cloning	33
3.6 Preparation of Recombinant Bacmid DNA	34

3.7	Transfection of <i>Sf9</i> Cells and Protein Expression	36
3.8	Production of Virus-like Particles	37
3.9	Protein Purification by Sucrose Density Gradient Ultracentrifugation	37
3.10	Protein Purification by Immobilised Metal Affinity Chromatography (IMAC)	38
3.11	Sodium Dodecyl Sulphate-Polyacrylamide Gel Electrophoresis (SDS-PAGE)	38
3.12	Western Blotting	38
3.13	The Bradford Assay	39
3.14	Dynamic Light Scattering Analysis	39
3.15	Transmission Electron Microscopy (TEM)	39
3.16	Circular Dichroism (CD) Spectroscopy	40
3.16.1	Secondary structure estimation (SSE)	40
3.16.2	Thermal stability analysis	40
3.17	Enzyme-linked Immunosorbent Assay (ELISA)	40
3.18	Immunisation of BALB/c Mice	41
3.19	ELISA to Quantify Immunogenicity of the Nc-aD VLPs	42
3.20	Immunophenotyping of Mice Splenocytes	42
3.21	Enzyme-linked Immunosorbent Spot (ELISPOT) Assay	43
3.22	Statistical Analysis	44
<b>4</b>	<b>RESULTS</b>	<b>45</b>
4.1	Construction of Recombinant Bacmid DNA Harbouiring the <i>Nc-aD</i> gene	45
4.2	Transfection of <i>Sf9</i> Cells	49
4.3	Purification of Nc-aD	53
4.3.1	Sucrose density gradient ultracentrifugation	53
4.3.2	Immobilised+metal+affinity+chromatography (IMAC)	54
4.4	Dynamic Light Scattering (DLS) Analysis	55
4.5	Transmission Electron Microscopy (TEM)	56
4.6	Circular Dichroism (CD) Spectroscopy	57
4.7	Antigenicity of the Nc-aD VLPs	59
4.8	Immunogenicity of the Nc-aD VLPs in BALB/c Mice	60
4.9	Immunophenotyping+of Mice+Splenocytes	61
4.10	Detection of Antibody Secreting Memory B Cells	63
<b>5</b>	<b>DISCUSSION</b>	<b>65</b>
5.1	Protein Expression, Purification and Characterisation	65
5.2	Antigenicity Assays and Immunogenicity	68
5.3	Immunophenotyping	69
5.4	Memory B Cell ELISPOT	70
<b>6</b>	<b>SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH</b>	<b>71</b>
6.1	Research Summary	71
6.2	Conclusion	71
6.3	Recommendations for Future Research	72

<b>REFERENCES</b>	73
<b>APPENDICES</b>	92
<b>BIODATA OF STUDENT</b>	100
<b>LIST OF PUBLICATIONS</b>	101



## LIST OF TABLES

<b>Table</b>	<b>Page</b>
3.1 Liquid and solid media	29
3.2 Buffer solutions	29
3.3 Cell lines and plasmids	30
3.4 List of primers utilised for the amplification and detection of <i>Nc-aD</i> gene insert	32
3.5 Grouping of BALB/c mice into immunisation groups	41
4.1 Temperature interval protein secondary structure estimation for the <i>Sf9</i> -expressed Nc-aD VLPs	59
4.2 Cytotoxic T lymphocyte (CTL) population in mouse splenocytes	62
4.3 ELISPOT analysis of activated mice splenocytes	64

## LIST OF FIGURES

<b>Figure</b>		<b>Page</b>
2.1	Global prevalence of hepatitis B virus infection	5
2.2	Hepatitis B virus morphology	7
2.3	The hepatitis B virus genomic structure	8
2.4	A schematic outline of the entry and replication of hepatitis B virus within hepatocytes	11
2.5	A schematic outline of the expression of hepatitis B surface antigen	13
2.6	Popularity of rodent and non-rodent mammalian models in Pubmed publications from 1970 to 2011	27
4.1	PCR amplification of the chimeric <i>MrNV nodavirus capsid</i> and <i>HBV 'a' determinant (Nc-aD)</i> gene	46
4.2	PCR amplification of the <i>Nc-aD</i> gene in the recombinant pFastBac HT C plasmid	47
4.3	Restriction enzyme double digestion of pFastBac HT C plasmid harbouring the <i>Nc-aD</i> gene	48
4.4	PCR confirmation of recombinant bacmid DNA harbouring the <i>Nc-Ad</i> gene	49
4.5	Schematic representation of the recombinant bacmid DNA constructed in this study	49
4.6	Phase-contrast micrographs of <i>Sf9</i> cells transfected with recombinant bacmid DNA during the 4-day incubation period	50
4.7	Western blotting of culture supernatant of <i>Sf9</i> cells transfected with the recombinant bacmid bearing the <i>Nc-aD</i> gene	51
4.8	The nucleotide and amino acid sequence of the chimeric <i>Nc-aD</i> gene and protein	52
4.9	SDS-PAGE and western blot analyses of Nc-aD purified using sucrose density gradient ultracentrifugation	53
4.10	(A) SDS-PAGE and (B) Western blotting analyses of the concentrated Nc-aD protein with anti-His monoclonal antibody	54
4.11	SDS-PAGE and western blotting of Nc-aD purified via immobilised metal affinity chromatography	55

4.12	Dynamic light scattering (DLS) analysis of the <i>Sf9</i> -expressed Nc-aD protein	56
4.13	Transmission electron microscopy of <i>Sf9</i> -expressed Nc-aD VLPs	57
4.14	Circular+dichroism (CD) spectra of the chimeric Nc-aD VLPs from wavelengths 240 nm to 190 nm	58
4.15	Antigenicity of the chimeric Nc-aD VLPs	60
4.16	Time-course immunogenicity assay of the <i>Sf9</i> -expressed Nc-aD protein in BALB/c mice using ELISA	61
4.17	Frequency of NK1.1 <sup>+</sup> mouse splenocytes	63
4.18	Representative ELISPOT images of the various immunisation groups	64



## LIST OF ABBREVIATIONS

aD	'a' determinant
AcMNPV	<i>Autographa carlifornica</i> multiple nuclear polyhedrosis virus
AIDS	Acquired immunodeficiency syndrome
APC	Allophycocyanin
ASC	Antibody secreting cell
cccDNA	Covalently closed circular DNA
CD	Circular dichroism
CHO	Chinese hamster ovary
Cryo-EM	Cryo-electron microscopy
CTL	Cytotoxic T-lymphocyte
DLS	Dynamic light scattering
ELISA	Enzyme-linked immunosorbent assay
ELISPOT	Enzyme-linked immunospot
ER	Endoplasmic reticulum
FDA	Food and Drug Administration
FITC	Fluorescein isothiocyanate
HBV	Hepatitis B virus
HBcAg	Hepatitis B core antigen
HBeAg	Hepatitis B e-antigen
HBsAg	Hepatitis B surface antigen
HIV	Human immunodeficiency virus
IFN	Interferon
IRF	Interferon regulatory factor
MHC	Major histocompatibility complex
MHR	Major hydrophilic region

<i>MrNV</i>	<i>Macrobrachium rosenbergii</i> nodavirus
Nc-aD	Nodavirus capsid-'a' determinant
Nc	Nodavirus capsid
NK cells	Natural killer cells
NKG2D	Natural killer group 2D
ORF	Open reading frame
PE	Phycoerythrin
pgRNA	Pregenomic RNA
rcDNA	Relaxed circular DNA
RdRp	RNA-dependent RNA polymerase
<i>Sf9</i>	<i>Spodoptera frugiperda</i>
SSE	Secondary structure estimation
TEM	Transmission electron microscopy
Tm	melting temperature
VLP	Virus-like particle
WHO	World Health Organisation

## CHAPTER 1

### INTRODUCTION

Hepatitis B and complications associated with it have since been classified by the World Health Organisation (WHO) to be a major health concern of the 21<sup>st</sup> century. About 250 million people, globally, are living carriers of the hepatitis B virus (HBV) (Tsai et al., 2018) and yearly recorded deaths resulting from HBV infection and related complications have been estimated to be 1.45 million (Anikhindi et al., 2018), higher than yearly deaths that result from malaria and HIV/AIDS with a yearly estimation of 0.44 million and 1.06 million deaths, respectively. Unlike the aforementioned communicable diseases, cases of HBV infections and related deaths have been on the increase, especially in developing nations (Seto et al., 2018; Tan & Ho, 2014). Therefore, more effective curative and preventive measures are being explored.

Since the USA's Food and Drug Administration (FDA) department's approval of the first hepatitis B vaccine in 1981, very significant milestones in the development of prophylactic and therapeutic measures targeted at hepatitis B, have been achieved (Seto et al., 2018). Based on common practice, injection with three separate doses of the HBV vaccine is still the most effective control measure, especially when the first dose is administered within the first 24 hours of birth (Hou et al., 2018).

However, several concerns about the prophylactic efficacies of the currently available hepatitis B vaccines have been raised. These concerns include the capability of these vaccines to elicit sustained protective immunity in some individuals with obesity, advanced age and pre-existing health conditions (Coates et al., 2001; Gerlich, 2017), the ineffectiveness of these vaccines in producing protective immunity in chronically infected subjects (Bengsch & Chang, 2016), induction of a poor immune response in about 10% of adult vaccinated subjects (Lerous-Roels et al., 2001) and their inability to confer protective immunity in subjects that are unresponsive to yeast-derived vaccines (Shouval, 2003). Furthermore, the spread of vaccine HBV escape mutants has made it necessary to develop newer HBV vaccine candidates with improved efficacy against HBV (Carman et al., 1990; Gerlich, 2017; Gerlich, 2015).

In an effort by WHO to eradicate Hepatitis B by 2030, a couple of treatments have been licensed for treatments of HBV infection (Mitra et al., 2018). These treatment regimens include interferons and nucleotide/nucleoside analogues which are capable of inhibiting HBV polymerases involved in viral replication. These treatments are reportedly effective in patients suffering from hepatitis B, however, none of these treatments is curative (Childs et al., 2018; Whitsett et al., 2019). The absence of a cure for hepatitis B further substantiates the need for developing more effective preventive vaccines while a cure is being sought.

Recombinant virus-like particles (VLPs) which are exploited as new hepatitis B vaccine candidates have shown promising prospects in inducing protective immunity in mice (Hyakumura et al., 2015; Kingston et al., 2019; Netter et al., 2001; Netter et al., 2003). Chimeric VLPs, consisting of two or more fused proteins from different viruses, are even more immunogenic and specific than VLPs consisting of a single viral protein (Ryu et al., 1997). Of interest to this study are VLPs of *Macrobrachium rosenbergii*-nodavirus-(*MrNV*) capsid protein (Nc) and their application as nanocarriers to display foreign epitopes. The immunogenicity of *Escherichia coli* (*E. coli*)-expressed chimeric VLPs, which consist of the-Nc displaying foreign viral epitopes, has been studied in mice (Ong et al., 2019; Yong et al., 2015a).

The *MrNV*, from which the recombinant Nc is derived, is implicated in the aetiology of the whitetail disease in *Macrobrachium rosenbergii* (commonly known as the giant freshwater prawns). Outbreaks of *MrNV* infection often result in 100% mortality in the infected prawn population thereby, causing severe financial losses to the aquaculture industry (Murwantoko et al., 2016). The *MrNV* genome is a positive sense single-stranded bipartite RNA molecules, known as RNA 1 (3.1 kb) and RNA 2 (1.2 kb). The former codes for the viral RNA-dependent RNA polymerase (RdRp) while the latter codes for the viral capsid protein (Nc) (Goh et al., 2011; Hanapi et al., 2017).

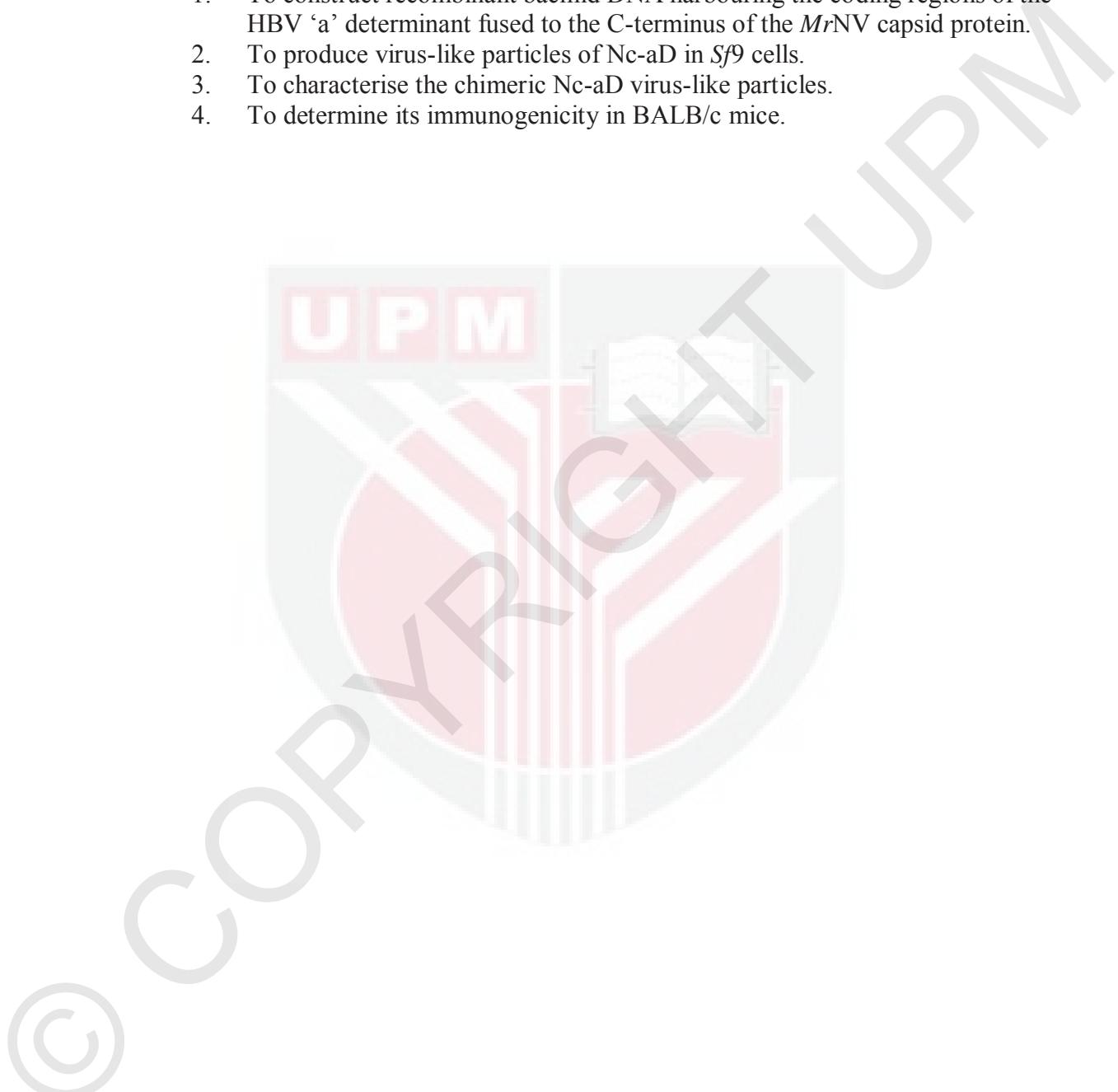
Previously, *E. coli* and *Spodoptera frugiperda* (*Sf9*) were used to express Nc VLPs (Goh et al., 2011; Kueh et al., 2016) and the VLPs were shown to display foreign protein epitopes at the Nc C-terminal region. Chimeric Nc VLPs displaying foreign viral epitopes have been produced in *E. coli* (Yong et al., 2015a; Yong et al., 2015b), suggesting that the Nc is an effective carrier protein to display of foreign viral epitopes in *E. coli* as it has been shown to prevent carrier protein-induced suppression of the displayed epitope (Ong et al., 2019). However, the performance of the Nc as an epitope-displaying carrier protein in *Sf9* expression systems is yet to be explored. Using the baculoviral expression system for insect cell lines, previous studies have shown that *Sf9* cells are better than the *E. coli* expression system with regards to the production of higher yields of stable and distinctly assembled VLPs (Ho et al., 2017; Kueh et al., 2016; López-Vidal et al., 2015; Rendic et al., 2008).

In this study, chimeric virus-like particles consisting of the HBV ‘a’ determinant (aD) fused to the C-terminus of the *MrNV*-capsid-protein-(Nc) were produced in *Sf9* insect cells via the baculovirus expression system. The aD of HBV is located in the S-domain of the HBV surface antigen (HBsAg) and it is conserved among the different strains of HBV (Bengsch & Chang, 2016; Hassemer et al., 2017). Furthermore, during infection, the aD is the component of the virus that induces the production of HBV-neutralising antibodies within the infected hosts and as such, this makes it an important epitope to be exploited in the development of new hepatitis B vaccines (Howard & Allison, 1995).

This study hypothesises that *Sf9* cells are an effective host for the production of chimeric *MrNV* Nc fused with HBV aD, in which Nc functions as a carrier protein. Also, this study hypothesises that *Sf9* cell-expressed Nc-aD will assemble into VLPs,

which will, in turn, elicit an aD-induced humoural, cellular and memory immune responses in BALB/c mice. The general objective of this study was to determine the prospect of the *Sf9*-produced Nc-aD as a new hepatitis B vaccine by measuring its immunogenicity in mice. Specifically, the objectives of this study include:

1. To construct recombinant bacmid DNA harbouring the coding regions of the HBV 'a' determinant fused to the C-terminus of the *MrNV* capsid protein.
2. To produce virus-like particles of Nc-aD in *Sf9* cells.
3. To characterise the chimeric Nc-aD virus-like particles.
4. To determine its immunogenicity in BALB/c mice.



## REFERENCES

- Aghebati-Maleki, L., Bakhshinejad, B., Baradaran, B., Motallebnezhad, M., Aghebati-Maleki, A., Nickho, H., ... Majidi, J. (2016). Phage display as a promising approach for vaccine development. *Journal of Biomedical Science*, 23(1), 1–18. doi:10.1186/s12929-016-0285-9
- Anikhindi, S. A., Kumar, A., Sharma, P., Singla, V., Bansal, N., & Arora, A. (2018). Ideal cure for hepatitis B infection: The target is in sight. *Journal of Clinical and Experimental Hepatology*, 8(2), 188–194. doi:10.1016/j.jceh.2017.10.002
- Arcier, J. M., Herman, F., Lightner, D. V., Redman, R. M., Mari, J., & Bonami, J. R. (1999). A viral disease associated with mortalities in hatchery-reared postlarvae of the giant freshwater prawn *Macrobrachium rosenbergii*. *Diseases of Aquatic Organisms*, 38(3), 177–181. doi:10.3354/dao038177
- Attanasio, R., Lanford, R. E., Dilley, D., Stunz, G. W., Notvall, L., Henderson, A. B., & Kennedy, R. C. (1991). Immunogenicity of hepatitis B surface antigen derived from the baculovirus expression vector system: A mouse potency study. *Biologicals*, 19(4), 347–353. doi:10.1016/S1045-1056(05)80024-7
- Bajaj, S., & Banerjee, M. (2016). In vitro assembly of polymorphic virus-like particles from the capsid protein of a nodavirus. *Virology*, 496, 106–115. doi:10.1016/j.virol.2016.05.025
- Bazan, J., Całkosiński, I., & Gamian, A. (2012). Phage display-a powerful technique for immunotherapy. *Human Vaccines and Immunotherapeutics*, 8(12), 1829–1835. doi:10.4161/hv.21704
- Belisle, E. H., & Strausser, H. R. (1981). Sex-related immunocompetence of BALB/c mice. II. Study of immunologic responsiveness of young, adult and aged mice. *Developmental and Comparative Immunology*, 5(4), 661–670. doi:10.1016/S0145-305X(81)80040-7
- Bengsch, B., & Chang, K. M. (2016). Evolution in our understanding of hepatitis B virus virology and immunology. *Clinics in Liver Disease*, 20(4), 629–644. doi:10.1016/j.cld.2016.06.001
- Bhaskar, R., Shuang, L., Peng, Z., mei xian, W., Fang, Z., Wan, C. C. D., & Yun gen, M. (2011). Probability to produce animal vaccines in insect baculovirus expression system. *African Journal of Biotechnology*, 10(51), 10323–10329. doi:10.5897/ajb09.192
- Biabanikhankahdani, R., Bayat, S., Ho, K. L., Alitheen, N. B. M., & Tan, W. S. (2017). A simple add-and-display method for immobilisation of cancer drug on his-tagged virus-like nanoparticles for controlled drug delivery. *Scientific Reports*, 7(1), 1–12. <https://doi.org/10.1038/s41598-017-05525-4>

- Blumberg, B. S., Sutnick, A. I., London, W. T., & Millman, I. (1970). Australia antigen and hepatitis. *The New England Journal of Medicine*, 283(7), 349–354.
- Bonami, J. R., & Sri Widada, J. (2011). Viral diseases of the giant fresh water prawn *Macrobrachium rosenbergii*: A review. *Journal of Invertebrate Pathology*, 106(1), 131–142. doi:10.1016/j.jip.2010.09.007
- Bornhorst, J. A., & Falke, J. J. (2000). Purification of proteins using polyhistidine affinity tags. *Methods in Enzymology*, 326, 245–254. doi:10.1016/j.immuni.2010.12.017.Two-stage
- Bradford, M. M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry*, 72(1–2), 248–254. doi:10.1016/0003-2697(76)90527-3
- Buchmann, P., Dembek, C., Kuklick, L., Jäger, C., Tedjokusumo, R., von Freyend, M. J., ... Protzer, U. (2013). A novel therapeutic hepatitis B vaccine induces cellular and humoral immune responses and breaks tolerance in hepatitis B virus (HBV) transgenic mice. *Vaccine*, 31(8), 1197–1203. doi:10.1016/j.vaccine.2012.12.074
- Cai, D., Qiu, Y., Qi, N., Yan, R., Lin, M., Nie, D., ... Hu, Y. (2010). Characterization of Wuhan nodavirus subgenomic RNA3 and the RNAi inhibition property of its encoded protein B2. *Virus Research*, 151(2), 153–161. doi:10.1016/j.virusres.2010.04.010
- Cai, X., Zheng, W., Pan, S., Zhang, S., Xie, Y., Guo, H., ... Luo, M. (2018). A virus-like particle of the hepatitis B virus preS antigen elicits robust neutralizing antibodies and T cell responses in mice. *Antiviral Research*, 149, 48–57. doi:10.1016/j.antiviral.2017.11.007
- Carman, W. F., Karayannidis, P., Waters, J., Thomas, H. C., Zanetti, A. R., Manzillo, G., & Zuckerman, A. J. (1990). Vaccine-induced escape mutant of hepatitis B virus. *The Lancet*, 336(8711), 325–329. doi:10.1016/0140-6736(90)91874-A
- Casal, J. I. (2001). Use of the baculovirus expression system for the generation of virus-like particles. *Biotechnology and Genetic Engineering Reviews*, 18(1), 73–87. doi:10.1080/02648725.2001.10648009
- Charnay, P., Gervais, M., Louise, A., Galibert, F., & Tiollais, P. (1980). Biosynthesis of hepatitis B virus surface antigen in *Escherichia coli*. *Nature*, 286(5776), 893–895. doi:10.1038/286893a0
- Chen, N. C., Yoshimura, M., Miyazaki, N., Guan, H. H., Chuankhayan, P., Lin, C. C., ... Chen, C. J. (2019). The atomic structures of shrimp nodaviruses reveal new dimeric spike structures and particle polymorphism. *Communications Biology*, 2(1), 1–14. doi:10.1038/s42003-019-0311-z

- Childs, L., Roesel, S., & Tohme, R. A. (2018). Status and progress of hepatitis B control through vaccination in the. *Vaccine*, 36(1), 6–14. doi:10.1016/j.vaccine.2017.11.027
- Chiou, H. L., Lee, T. S., Kuo, J., Mau, Y. C., & Ho, M. S. (1997). Altered antigenicity of “a” determinant variants of hepatitis B virus. *Journal of General Virology*, 78(10), 2639–2645.
- Chisari, F. V. (1995). Hepatitis B virus. *Annual Review of Immunology*, 13(1), 29–60. doi:10.1201/b14823-10
- Chong, L. C., Ganesan, H., Yong, C. Y., Tan, W. S., & Ho, K. L. (2019). Expression , purification and characterization of the dimeric protruding domain of *Macrobrachium rosenbergii* nodavirus capsid protein expressed in *Escherichia coli*. *PLoS ONE*, 14(2), 1–12. doi:10.1371/journal.pone.0211740
- Chroboczek, J., Szurgot, I., & Szolajska, E. (2014). Virus-like particles as vaccine. *Acta Biochimica Polonica*, 61(3), 531–539. [https://doi.org/10.18388/abp.2014\\_1875](https://doi.org/10.18388/abp.2014_1875)
- Chromy, L. R., Pipas, J. M., & Garcea, R. L. (2003). Chaperone-mediated in vitro assembly of polyomavirus capsids. *Proceedings of the National Academy of Sciences*, 100(18), 10477–10482. doi:10.1073/pnas.1832245100
- Citarasu, T., Lelin, C., Babu, M. M., Anand, S. B., Nathan, A. A., & Vakharia, V. N. (2019). Oral vaccination of *Macrobrachium rosenbergii* with baculovirus-expressed *M. rosenbergii* nodavirus (MrNV) capsid protein induces protective immunity against MrNV challenge. *Fish and Shellfish Immunology*, 86, 1123–1129. doi:10.1016/j.fsi.2018.12.010
- Coates, T., Wilson, R., Patrick, G., André, F., & Watson, V. (2001). Hepatitis B vaccines: Assessment of the seroprotective efficacy of two recombinant DNA vaccines. *Clinical Therapeutics*, 23(3), 392–403. doi:10.1016/S0149-2918(01)80044-8
- Coletti, D., Berardi, E., Aulino, P., Rossi, E., Moresi, V., Li, Z., & Adamo, S. (2013). Substrains of inbred mice differ in their physical activity as a behavior. *The Scientific World Journal*, 2013, 237260. doi:10.1155/2013/237260
- Cox, M. M. J. (2012). Recombinant protein vaccines produced in insect cells. *Vaccine*, 30(10), 1759–1766. doi:10.1016/j.vaccine.2012.01.016
- Crowther, F. A., Kiselev, N. A., Bottcher, B., Berriman, J. A., Borisova, G. P., Ose, V., & Pumpens, P. (1994). Three-dimensional structure of hepatitis B virus core particles determined by electron cryomicroscopy. *Cell*, 77(6), 943–950.
- Dandri, M., & Lütgehetmann, M. (2014). Mouse models of hepatitis B and delta virus infection. *Journal of Immunological Methods*, 410, 39–49. doi:10.1016/j.jim.2014.03.002

- Dane, D. S., Cameron, C. H., & Moya, B. (1970). Virus-like particles in serum of patients with Australia-antigen-associated hepatitis. *The Lancet*, 33(6), 695–698.
- Diminsky, D., Schirmbeck, R., Reimann, J., & Barenholz, Y. (1997). Comparison between hepatitis B surface antigen (HBsAg) particles derived from mammalian cells (CHO) and yeast cells (*Hansenula polymorpha*): Composition, structure and immunogenicity. *Vaccine*, 15(6–7), 637–647. doi:10.1016/S0264-410X(96)00239-3
- Dong, X. F., Natarajan, P., Tihova, M., Johnson, J. E., & Schneemann, A. (1998). Particle polymorphism caused by deletion of a peptide molecular switch in a quasiequivalent icosahedral virus. *Journal of Virology*, 72(7), 6024–6033. doi:10.1128/jvi.72.7.6024-6033.1998
- Dryden, K. A., Wieland, S. F., Whitten-bauer, C., Gerin, J. L., Chisari, F. V., & Yeager, M. (2006). Native hepatitis B virions and capsids visualized by electron cryomicroscopy. *Molecular Cell*, 22(6), 843–850. doi:10.1016/j.molcel.2006.04.025
- Dufour, D. R. (2006). Hepatitis B surface antigen (HBsAg) assays - are they good enough for their current uses? *Clinical Chemistry*, 52(8), 1457–1459. doi:10.1373/clinchem.2006.072504
- Dunn, C., Peppa, D., Khanna, P., Nebbia, G., Jones, M., Brendish, N., ... Maini, M. K. (2009). Temporal analysis of early immune responses in patients with acute hepatitis B virus infection. *Gastroenterology*, 137(4), 1289–1300. doi:10.1053/j.gastro.2009.06.054
- Edman, J. C., Hallewell, R. A., Valenzuela, P., Goodman, H. M., & Rutter, W. J. (1981). Synthesis of hepatitis B surface and core antigens in *E. coli*. *Nature*, 291(5815), 503–506. doi:10.1038/291503a0
- Elghanam, M. S., Attia, A. S., Shoeb, H. A., & Hashem, A. E. M. (2012). Expression and purification of hepatitis B surface antigen S from *Escherichia coli*; a new simple method. *BMC Research Notes*, 5(1), 125. doi:10.1186/1756-0500-5-125
- Ericsson, A. C., Crim, M. J., & Franklin, C. L. (2013). A brief history of animal modeling. *Missouri Medicine*, 110(3), 201–205.
- Escriou, V., Ciolina, C., Helbling-Leclerc, A., Wils, P., & Scherman, D. (1998). Cationic lipid-mediated gene transfer: Analysis of cellular uptake and nuclear import of plasmid DNA. *Cell Biology and Toxicology*, 14(2), 95–104. doi:10.1023/A:1007425803756
- Fan, J., Liang, X., Horton, M. S., Perry, H. C., Citron, M. P., Heidecker, G. J., ... Shiver, J. W. (2004). Preclinical study of influenza virus a M2 peptide conjugate vaccines in mice, ferrets, and rhesus monkeys. *Vaccine*, 22(23-24), 2993–3003. doi:10.1016/j.vaccine.2004.02.021

FAO (2018). The state of world fisheries and aquaculture. *Food and Agriculture Organization of the United Nations* (Vol. 3). doi:10.1093/japr/3.1.101

Fifis, T., Gamvrellis, A., Crimeen-Irwin, B., Pietersz, G. A., Li, J., Mottram, P. L., ... Plebanski, M. (2004). Size-dependent immunogenicity: Therapeutic and protective properties of nano-vaccines against tumors. *The Journal of Immunology*, 173(5), 3148–3154. doi:10.4049/jimmunol.173.5.3148

Gamvrellis, A., Leong, D., Hanley, J. C., Xiang, S. D., Mottram, P., & Plebanski, M. (2004). Vaccines that facilitate antigen entry into dendritic cells. *Immunology and Cell Biology*, 82(5), 506–516. doi:10.1111/j.0818-9641.2004.01271.x

Geier, M. R., Geier, D. A., & Zahalsky, A. C. (2003). A review of hepatitis B vaccination. *Expert Opinion on Drug Safety*, 2(2), 113–122. doi:10.1517/14740338.2.2.113

Gerdts, V., van Drunen Little-van den Hurk, S., Griebel, P. J., & Babiuk, L. A. (2007). Use of animal models in the development of human vaccines. *Future Microbiology*, 2(6), 667–675. doi:10.2217/17460913.2.6.667

Gerdts, V., Wilson, H. L., Meurens, F., van Drunen Littel - van den Hurk, S., Wilson, D., Walker, S., ... Potter, A. A. (2015). Large animal models for vaccine development and testing. *ILAR Journal*, 56(1), 53–62. doi:10.1093/ilar/ilv009

Gerlich, W. H. (2017). Do we need better hepatitis B vaccines? *Indian Journal of Medical Research*, 145(4), 414–419. doi:10.4103/ijmr.IJMR

Gerlich, W. H. (2015). Prophylactic vaccination against hepatitis B: Achievements, challenges and perspectives. *Medical Microbiology and Immunology*, 204(1), 39–55. doi:10.1007/s00430-014-0373-y

Gholson, C. F., Siddiqui, A., & Vierling, J. M. (1990). Cell surface expression of hepatitis B surface and core antigens in transfected rat fibroblast cell lines. *Gastroenterology*, 98(4), 968–975. doi:10.1016/0016-5085(90)90021-R

Goh, Z. H., Tan, S. G., Bhassu, S., & Tan, W. S. (2011). Virus-like particles of *Macrobrachium rosenbergii* nodavirus produced in bacteria. *Journal of Virological Methods*, 175(1), 74–79. doi:10.1016/j.jviromet.2011.04.021

Golding, H., Khurana, S., & Zaitseva, M. (2018). What is the predictive value of animal models for vaccine efficacy in humans? The importance of bridging studies and species-independent correlates of protection. *Cold Spring Harbor Perspectives in Biology*, 10(4), a028902. doi:10.1101/cshperspect.a028902

Gong, D., Zhao, H., Liang, Y., Chao, R., Chen, L., Yang, S., & Yu, P. (2019). Differences in cocaine- and morphine-induced cognitive impairments and serum corticosterone between C57BL/6J and BALB/cJ mice. *Pharmacology Biochemistry and Behavior*, 182, 1–6. doi:10.1016/j.pbb.2019.05.006

- Grgacic, E. V. L., & Anderson, D. A. (2006). Virus-like particles: Passport to immune recognition. *Methods*, 40(1), 60–65. doi:10.1016/jymeth.2006.07.018
- Grompe, M., & Strom, S. (2013). Mice with human livers. *Gastroenterology*, 145(6), 1209–1214. doi:10.1053/j.gastro.2013.09.009
- Guan, Z. jun, Guo, B., Huo, Y. lin, Guan, Z. ping, & Wei, Y. hui. (2010). Overview of expression of hepatitis B surface antigen in transgenic plants. *Vaccine*, 28(46), 7351–7362. doi:10.1016/j.vaccine.2010.08.100
- Guidotti, L. G., Matzke, B., Schaller, H., & Chisari, F. V. (1995). High-level hepatitis B virus replication in transgenic mice. *Journal of Virology*, 69(10), 6158–6169.
- Guidotti, L. G., & Chisari, F. V. (2006). Immunobiology and pathogenesis of viral hepatitis. *Annual Review of Pathology: Mechanisms of Disease*, 1(1), 23–61. doi:10.1146/annurev.pathol.1.110304.100230
- Guidotti, L. G., Rochford, R., Chung, J., Shapiro, M., Purcell, R., & Chisari, F. V. (1999). Viral clearance without destruction of infected cells during acute HBV infection. *Science*, 284(5415), 825–829. doi:10.1126/science.284.5415.825
- Guidotti, L. G., Ishikawa, T., Hobbs, M. V., Matzke, B., Schreiber, R., & Chisari, F. V. (1996). Intracellular inactivation of the hepatitis B virus by inflammatory cytokines. *Immunity*, 4(1), 25–36. doi:10.2957/kanzo.39.supl2\_57
- Hadjji-Abbes, N., Martin, M., Benzina, W., Karray-Hakim, H., Gergely, C., Gargouri, A., & Mokdad-Gargouri, R. (2013). Extraction and purification of hepatitis B virus-like M particles from a recombinant *Saccharomyces cerevisiae* strain using alumina powder. *Journal of Virological Methods*, 187(1), 132–137. doi:10.1016/j.jviromet.2012.09.023
- Hanapi, U. F., Yong, C. Y., Goh, Z. H., Alitheen, N. B., Yeap, S. K., & Tan, W. S. (2017). Tracking the virus-like particles of *Macrobrachium rosenbergii* nodavirus in insect cells. *PeerJ*, 5, e2947. doi:10.7717/peerj.2947
- Hassemer, M., Finkernagel, M., Peiffer, K. H., Glebe, D., Akhras, S., Reuter, A., ... Hildt, E. (2017). Comparative characterization of hepatitis B virus surface antigen derived from different hepatitis B virus genotypes. *Virology*, 502, 1–12. doi:10.1016/j.virol.2016.12.003
- Hayakijkosol, O., & Owens, L. (2012). B2 or not B2: RNA interference reduces *Macrobrachium rosenbergii* nodavirus replication in redclaw crayfish (*Cherax quadricarinatus*). *Aquaculture*, 326–329, 40–45. doi:10.1016/j.aquaculture.2011.11.023
- Herzenberg, L. A., & Tokuhisa, T. (1982). Epitope-specific regulation. I. Carrier-specific induction of suppression for IgG anti-hapten antibody responses. *Journal of Experimental Medicine*, 155(6), 1730–1740. doi:10.1084/jem.155.6.1730

- Hilleman, M. R. (1987). Yeast recombinant hepatitis B vaccine. *Infection*, 15(1), 3–7. doi:10.1007/BF01646107
- Ho, K. L., Gabrielsen, M., Beh, P. L., Kueh, C. L., Thong, Q. X., Streetley, J., ... Bhella, D. (2018). Structure of the *Macrobrachium rosenbergii* nodavirus: A new genus within the Nodaviridae? *PLoS Biology*, 16(10), e3000038. doi:10.1371/journal.pbio.3000038
- Ho, K. L., Kueh, C. L., Beh, P. L., Tan, W. S., & Bhella, D. (2017). Cryo-electron microscopy structure of the *Macrobrachium rosenbergii* nodavirus capsid at 7 angstroms resolution. *Scientific Reports*, 7(2083), 1–8. doi:10.1038/s41598-017-02292-0
- Hoebe, C. J. P. A., Vermeiren, A. P. A., & Dukers-muijrs, N. H. T. M. (2012). Revaccination with Fendrix ® or HBVaxPro ® results in better response rates than does revaccination with three doses of Engerix-B ® in previous non-responders. *Vaccine*, 30(48), 6734–6737. doi:10.1016/j.vaccine.2012.08.074
- Hossain, M. G., & Ueda, K. (2017). Investigation of a novel hepatitis B virus surface antigen (HBsAg) escape mutant affecting immunogenicity. *PLoS ONE*, 12(1), 1–22. doi:10.1371/journal.pone.0167871
- Hou, J., Cui, F., Ding, Y., Dou, X., Duan, Z., Han, G., ... Zhuang, H. (2018). Management Algorithm for Interrupting Mother to Child. *Clinical Gastroenterology and Hepatology*, 17(10), 1–9. doi:10.1016/j.cgh.2018.10.007
- Howard, C. R., & Allison, L. M. C. (1995). Hepatitis B surface antigen variation and protective immunity. *Intervirology*, 38(1), 35–40. doi:10.1159/000150412
- Hyakumura, M., Walsh, R., Thaysen-Andersen, M., Kingston, N. J., La, M., Lu, L., ... Netter, H. J. (2015). Modification of asparagine-linked glycan density for the design of hepatitis B virus virus-like particles with enhanced immunogenicity. *Journal of Virology*, 89(22), 11312–11322. doi:10.1128/JVI.01123-15
- Iannacone, M., & Guidotti, L. G. (2015). Mouse models of hepatitis B virus pathogenesis. *Cold Spring Harbor Perspectives in Medicine*, 5(11), 1–11. doi:10.1101/cshperspect.a021477
- Inada, T., Misumi, Y., Seno, M., Kanezaki, S., Shibata, Y., Oka, Y., & Onda, H. (1989). Synthesis of hepatitis B virus e antigen in *E. coli*. *Virus Research*, 14(1), 27–47. doi:10.1016/0168-1702(89)90067-1
- Iwamoto, T., Mise, K., Takeda, A., Okinaka, Y., Mori, K. I., Arimoto, M., ... Nakai, T. (2005). Characterization of striped jack nervous necrosis virus subgenomic RNA3 and biological activities of its encoded protein B2. *Journal of General Virology*, 86(10), 2807–2816. doi:10.1099/vir.0.80902-0
- Jariyapong, P., Chotwiwatthanakun, C., Somrit, M., Jitrapakdee, S., Xing, L., Cheng, H. R., & Weerachatyanukul, W. (2014). Encapsulation and delivery of plasmid DNA by virus-like nanoparticles engineered from *Macrobrachium rosenbergii*

nodavirus. *Virus Research*, 179(1), 140–146. doi:10.1016/j.virusres.2013.10.021

Jefferies, M., Rauff, B., Rashid, H., Lam, T., & Rafiq, S. (2018). Update on global epidemiology of viral hepatitis and preventive strategies. *World Journal of Clinical Cases*, 6(13), 589–599. doi:10.5500/wjt.v1.i1.4

Jegerlehner, A., Wiesel, M., Dietmeier, K., Zabel, F., Gatto, D., Saudan, P., & Bachmann, M. F. (2010). Carrier induced epitopic suppression of antibody responses induced by virus-like particles is a dynamic phenomenon caused by carrier-specific antibodies. *Vaccine*, 28(33), 5503–5512. doi:10.1016/j.vaccine.2010.02.103

Jenkins, N., & Curling, E. M. A. (1994). Glycosylation of recombinant proteins: Problems and prospects. *Enzyme and Microbial Technology*, 16(5), 354–364. doi:10.1016/0141-0229(94)90149-X

Jing, M., Wang, J., Zhu, S., Ao, F., Wang, L., Han, T., ... Liu, S. (2016). Development of a more efficient hepatitis B virus vaccine by targeting hepatitis B virus preS to dendritic cells. *Vaccine*, 34(4), 516–522. doi:10.1016/j.vaccine.2015.11.069

Jorge, S. A. C., Santos, A. S., Spina, Â., & Pereira, C. A. (2008). Expression of the hepatitis B virus surface antigen in *Drosophila* S2 cells. *Cytotechnology*, 57(1), 51–59. doi:10.1007/s10616-008-9154-z

Kang, C. Y., Bishop, D. H., Seo, J. S., Matsuura, Y., & Choe, M. (1987). Secretion of particles of hepatitis B surface antigen from insect cells using a baculovirus vector. *The Journal of General Virology*, 68(10), 2607–2613. doi: 10.1099/0022-1317-68-10-2607

Kapusta, J., Modelska, A., Figlerowicz, M., Pniewski, T., Letellier, M., Lisowa, O., ... Legocki, A. B. (1999). A plant-derived edible vaccine against hepatitis B virus. *FASEB Journal*, 13(13), 1796–1799. doi: 10.1096/fasebj.13.13.1796

Karayiannis, P. (2003). Hepatitis B virus: Old, new and future approaches to antiviral treatment. *Journal of Antimicrobial Chemotherapy*, 51(4), 761–785. doi:10.1093/jac/dkg163

Keating, G. M., Noble, S., Averhoff, F. M., Belloni, C., Duval, B., Goldwater, P. N., ... Leroux-Roels, G. (2003). Recombinant hepatitis B vaccine (Engerix-B): A review of its immunogenicity and protective efficacy against hepatitis B. *Drugs*, 63(10), 1021–1051. doi: 10.2165/00003495-200363100-00006

Kingston, N. J., Kurtovic, L., Walsh, R., Joe, C., Lovrecz, G., Locarnini, S., ... Netter, H. J. (2019). Hepatitis B virus-like particles expressing *Plasmodium falciparum* epitopes induce complement-fixing antibodies against the circumsporozoite protein. *Vaccine*, 37(12), 1674–1684. doi:10.1016/j.vaccine.2019.01.056

Kirnbauer, R., Booyt, F., Chengt, N., Lowy, D. R., & Schiller, J. T. (1992). Papillomavirus L1 major capsid protein self-assembles into virus-like particles that are highly immunogenic. *Proceedings of the National Academy of Sciences*,

- 89(24), 12180–12184. doi:10.1073/pnas.89.24.12180
- Kondo, Y., Ninomiya, M., Kakazu, E., Kimura, O., & Shimosegawa, T. (2013). Hepatitis B surface antigen could contribute to the immunopathogenesis of hepatitis B virus infection. *ISRN Gastroenterology*, 2013, 1–8. doi:10.1155/2013/935295
- Kong, Q., Richter, L., Yang, Y. F., Arntzen, C. J., Mason, H. S., & Thanavala, Y. (2001). Oral immunization with hepatitis B surface antigen expressed in transgenic plants. *Proceedings of the National Academy of Sciences*, 98(20), 11539–11544. doi:10.1073/pnas.191617598
- Koumbi, L., Pollicino, T., Raimondo, G., Kumar, N., Karayiannis, P., & Khakoo, S. I. (2016). Hepatitis B viral replication influences the expression of natural killer cell ligands. *Annals of Gastroenterology*, 29(3), 348–357. doi:10.20524/aog.2016.0036
- Kramvis, A. (2014). Genotypes and genetic variability of hepatitis B virus. *Intervirology*, 57(3–4), 141–150. doi:10.1159/000360947
- Krugman, S., Giles, J. P., & Hammond, J. (1970). Hepatitis virus : Effect of heat on the infectivity and antigenicity of the MS-1 and MS-2 strains. *The Journal of Infectious Diseases*, 122(5), 432–436. doi: 10.1093/infdis/122.5.432
- Krugman, S., Giles, J. P., & Hammond, J. (1971). Viral hepatitis, type B (MS-2 strain) studies on active immunization. *Journal of the American Medical Association*, 217(1), 41–45. doi:10.1001/jama.1971.03190010023005
- Kueh, C. L., Yong, C. Y., Dezfooli, S. M., Bhassu, S., Tan, S. G., & Tan, W. S. (2016). Virus-like particle of *Macrobrachium rosenbergii* nodavirus produced in *Spodoptera frugiperda* (Sf9) cells is distinctive from that produced in *Escherichia coli*. *Biotechnology Progress*, 33(2), 548–557. doi:10.1002/btpr.2409
- Lamontagne, R. J., Bagga, S., & Bouchard, M. J. (2016). Hepatitis B virus molecular biology and pathogenesis. *Hepatoma Research*, 2(7), 163–186. doi:10.20517/2394-5079.2016.05
- Landford, R. E., Luckow, V., Kennedy, R. C., Dreesman, G. R., Notvall, L., & Summers, M. D. (1989). Expression and characterization of hepatitis B virus surface antigen polypeptides in insect cells with a baculovirus expression system. *Journal of Virology*, 63(4), 1549–1557.
- Lee, T., Inokoshi, J., Namiki, M., Takeshima, H., & Omura, S. (1989). Production of hepatitis B virus surface antigen containing pre-S1 and and pre-S2 domains by Chinese hamster ovary cells. *Archives of Virology*, 106(1-2), 151–158.
- Lerous-Roels, G., Cao, T., De Knibber, A., Meuleman, P., Roobrouck, A., Farhoudi, A., ... Desombere, I. (2001). Prevention of hepatitis B infections: vaccination and its limitations. *Acta Clinica Belgica*, 56(4), 209–219. doi:

10.1179/acb.2001.032

- Li, C., Wang, Y., Liu, T., Niklasch, M., Qiao, K., Durand, S., ... Wang, Y. X. (2019). An *E. coli*-produced single-chain variable fragment (scFv) targeting hepatitis B virus surface protein potently inhibited virion secretion. *Antiviral Research*, 162, 118–129. doi:10.1016/j.antiviral.2018.12.019
- Li, X., Xie, T., Gao, L., Ma, C., Yang, X., & Liang, X. (2018). Prostaglandin E2 facilitates hepatitis B virus replication by impairing CTL function. *Molecular Immunology*, 103, 243–250. doi:10.1016/j.molimm.2018.08.009
- Lihoradova, O. A., Bachurina, E. Y., & Azimova, S. S. (2004). Biosynthesis of the recombinant middle surface antigen of the human hepatitis B virus in silkworm larvae. *Molecular Biology*, 38(4), 603–607. doi:10.1023/B:MBIL.0000037014.85886.fc
- Lin, C. L., & Kao, J. H. (2015). Hepatitis B virus genotypes and variants. *Cold Spring Harbor Perspectives in Medicine*, 5(5), 1–19. doi:10.1101/cshperspect.a021436
- Lindemann, M., Koldehoff, M., Fiedler, M., Schumann, A., Ottlinger, H. D., Heinemann, F. M., ... Beelen, D. W. (2016). Control of hepatitis B virus infection in hematopoietic stem cell recipients after receiving grafts from vaccinated donors. *Bone Marrow Transplantation*, 51(3), 428–431. doi:10.1038/bmt.2015.253
- Locarnini, S. (2004). Molecular virology of hepatitis B virus. *Seminars in Liver Disease*, 24(212), 3–10. doi: 10.1055/s-2004-828672
- López-Vidal, J., Gómez-Sebastián, S., Bárcena, J., Del Carmen Nuñez, M., Martínez-Alonso, D., Dudognon, B., ... Escribano, J. M. (2015). Improved production efficiency of virus-like particles by the baculovirus expression vector system. *PLoS ONE*, 10(10), 1–13. doi:10.1371/journal.pone.0140039
- Lünsdorf, H., Gurramkonda, C., Adnan, A., Khanna, N., & Rinas, U. (2011). Virus-like particle production with yeast: Ultrastructural and immunocytochemical insights into *Pichia pastoris* producing high levels of the hepatitis B surface antigen. *Microbial Cell Factories*, 10(48), 1–10. doi:10.1186/1475-2859-10-48
- Luvisa, B. K., & Hassanein, T. I. (2016). Hepatitis B virus infection and liver decompensation. *Clinics in Liver Disease*, 20(4), 681–692. doi:10.1016/j.cld.2016.07.002
- Ma, X., Zhang, Z., Xie, H., Ma, Y., Liu, C., Liu, S., & Liu, M. (2008). Improved models for animal research. In M. Conn (Ed.), *Sourcebook of models for biomedical research* (pp 17-26). New Jersey, USA: Humana Press. doi:10.1039/c8cc08090a
- Marcondes, J., & Hansen, E. (2008). Transgenic lettuce seedlings carrying hepatitis B virus antigen HBsAg. *Brazilian Journal of Infectious Diseases*, 12(6), 469–471. doi:10.1590/S1413-86702008000600004

- Marshall, H. (2001). Edible vaccine for hepatitis B. *Trends in Immunology*, 22(2), 71. doi:10.1016/s1471-4906(00)01853-6
- Masavuli, M. G., Wijesundara, D. K., Torresi, J., Gowans, E. J., & Grubor-Bauk, B. (2017). Preclinical development and production of virus-like particles as vaccine candidates for hepatitis C. *Frontiers in Microbiology*, 8(2413), 1–11. doi:10.3389/fmicb.2017.02413
- Maurer, K. J., & Quimby, F. W. (2015). Animal models in biomedical research. In J. G. Fox, L. C. Anderson, G. M. Otto, K. R. Pritchett-Corning, M. T. Whary (Eds.), *Laboratory animal medicine: Third Edition* (pp. 1497–1526). Academic Press. doi:10.1016/B978-0-12-409527-4.00034-1
- McAleer, W. J., Bunyak, E. B., Maigetter, R. Z., Wampler, D. E., Miller, W. J., & Hilleman, M. R. (1984). Human hepatitis B vaccine from recombinant yeast. *Nature*, 307(5947), 178–180. doi:10.1038/307178a0
- Meireles, L. C., Marinho, R. T., & Damme, P. Van. (2015). Three decades of hepatitis B control with vaccination, 7(18), 2127–2132. doi:10.4254/wjh.v7.i18.2127
- Michel, M. L., Sobczak, E., Malpièce, Y., Tiollais, P., & Streeck, R. E. (1985). Expression of amplified hepatitis B virus surface antigen genes in chinese hamster ovary cells. *Nature Biotechnology*, 3(6), 561–566. doi:10.1038/nbt0685-561
- Miotto, M., Olimpieri, P. P., Di Renzo, L., Ambrosetti, F., Corsi, P., Lepore, R., ... Milanetti, E. (2019). Insights on protein thermal stability: A graph representation of molecular interactions. *Bioinformatics*, 35(15), 2569–2577. doi:10.1093/bioinformatics/bty1011
- Mirian, M., Taghizadeh, R., Khanahmad, H., Salehi, M., Jahanian-Najafabadi, A., Sadeghi-Aliabadi, H., & Kouhpayeh, S. (2016). Exposition of hepatitis B surface antigen (HBsAg) on the surface of HEK293T cell and evaluation of its expression. *Research in Pharmaceutical Sciences*, 11(5), 366–373. doi:10.4103/1735-5362.192485
- Mitra, B., Thapa, R. J., Guo, H., & Block, T. M. (2018). Host functions used by hepatitis B virus to complete its life cycle : Implications for developing host-targeting agents to treat chronic hepatitis B. *Antiviral Research*, 158, 185–198. doi:10.1016/j.antiviral.2018.08.014
- Mohsen, M. O., Zha, L., Cabral-Miranda, G., & Bachmann, M. F. (2017). Major findings and recent advances in virus-like particle (VLP)-based vaccines. *Seminars in Immunology*, 34(September), 123–132. <https://doi.org/10.1016/j.smim.2017.08.014>
- Murwantoko, M., Bimantara, A., Roosmanto, R., & Kawaichi, M. (2016). *Macrobrachium rosenbergii* nodavirus infection in a giant freshwater prawn hatchery in Indonesia. *SpringerPlus*, 5(1), 1729-1737. doi:10.1186/s40064-016-3127-z

- Naskalska, A., & Pyrc, K. (2015). Virus Like Particles as Immunogens and Universal Nanocarriers. *Polish Journal of Microbiology*, 64(1), 3–13.
- NaveenKumar, S., Karunasagar, I., & Karunasagar, I. (2013a). Protection of *Macrobrachium rosenbergii* against white tail disease by oral administration of bacterial expressed and encapsulated double-stranded RNA. *Fish and Shellfish Immunology*, 35(3), 833–839. doi:10.1016/j.fsi.2013.06.019
- NaveenKumar, S., Shekar, M., Karunasagar, I., & Karunasagar, I. (2013b). Genetic analysis of RNA1 and RNA2 of *Macrobrachium rosenbergii* nodavirus (MrNV) isolated from India. *Virus Research*, 173(2), 377–385. doi:10.1016/j.virusres.2013.01.003
- Netter, H. J., Macnaughton, T. B., Woo, W., & Tindle, R. (2001). Antigenicity and immunogenicity of novel chimeric hepatitis B surface antigen particles with exposed hepatitis C virus epitopes. *Journal of Virological Methods*, 75(5), 2130–2141. doi:10.1128/JVI.75.5.2130
- Netter, H. J., Woo, W., Tindle, R., Macfarlan, R. I., & Gowans, E. J. (2003). Immunogenicity of recombinant HBsAg/HCV particles in mice pre-immunised with hepatitis B virus-specific vaccine. *Vaccine*, 21(21-22), 2692–2697. doi:10.1016/S0264-410X(03)00182-8
- Noad, R., & Roy, P. (2003). Virus-like particles as immunogens. *Trends in Microbiology*, 11(9), 438–444. [https://doi.org/10.1016/S0966-842X\(03\)00208-7](https://doi.org/10.1016/S0966-842X(03)00208-7)
- Ong, H. K., Yong, C. Y., Tan, W. S., Yeap, S. K., Omar, A. R., Razak, M. A., & Ho, K. L. (2019). An influenza A vaccine based on the extracellular domain of matrix 2 protein protects BALB/c mice against H1N1 and H3N2. *Vaccines*, 7(3), 91–106. doi:10.3390/vaccines7030091
- Op Den Brouw, M. L., Binda, R. S., Van Roosmalen, M. H., Protzer, U., Janssen, H. L. A., Van Der Molen, R. G., & Woltman, A. M. (2009). Hepatitis B virus surface antigen impairs myeloid dendritic cell function: A possible immune escape mechanism of hepatitis B virus. *Immunology*, 126(2), 280–289. doi:10.1111/j.1365-2567.2008.02896.x
- Ou, J. H. (1997). Molecular biology of hepatitis B virus e antigen. *Journal of Gastroenterology and Hepatology*, 12(9-10), 178–187. doi:10.1111/j.1440-1746.1997.tb00499.x
- Ou, J. H., Laub, O., & Rutter, W. J. (1986). Hepatitis B virus gene function: the precore region targets the core antigen to cellular membranes and causes the secretion of the e antigen. *Proceedings of the National Academy of Sciences*, 83(6), 1578–1582. doi:10.1073/pnas.83.6.1578
- Pabisch, S., Feichtenschlager, B., Kickelbick, G., & Peterlik, H. (2012). Effect of interparticle interactions on size determination of zirconia and silica based systems - A comparison of SAXS, DLS, BET, XRD and TEM. *Chemical Physics Letters*, 521, 91–97. doi:10.1016/j.cplett.2011.11.049

- Patzer, E. J., Nakamura, G. R., Hershberg, R. D., Gregory, T. J., Crowley, C., Levinson, A. D., & Eichberg, J. W. (1986). Cell culture derived recombinant HBsAg is highly immunogenic and protects chimpanzees from infection with hepatitis B virus. *Nature Biotechnology*, 4, 630–636. doi: 10.1038/nbt0786-630
- Pniewski, T. (2013). The twenty-year story of a plant-based vaccine against hepatitis B: Stagnation or promising prospects? *International Journal of Molecular Sciences*, 14(1), 1978–1998. doi:10.3390/ijms14011978
- Poovorawan, Y., Sanpavat, S., Pongpunlert, W., Chumdermpadetsuk, S., Sentrakul, P., & Safary, A. (1989). Protective efficacy of a recombinant DNA hepatitis B vaccine in neonates of HBe antigen-positive mothers. *The Journal of the American Medical Association*, 261(22), 3278–3281. doi:10.1001/jama.1989.03420220092033
- Potter, M. (1985). History of the BALB/c family. *Current Topics in Microbiology and Immunology*, 122, 1–5. doi:doi:10.1007/978-3-642-70740-7\_1
- Premanand, B., Wee, P. Z., & Prabakaran, M. (2018). Baculovirus surface display of immunogenic proteins for vaccine development. *Viruses*, 10(6). doi:10.3390/v10060298
- Pride, M. W., Shi, H., Anchim, J. M., Scott Linthicum, D., LoVerde, P. T., Thakur, A., & Thanavala, Y. (1992). Molecular mimicry of hepatitis B surface antigen by an anti-idiotype-derived synthetic peptide. *Proceedings of the National Academy of Sciences*, 89(24), 11900–11904. doi:10.1073/pnas.89.24.11900
- Raz, R., Dagantl, R., Gallilz, A., Brill, G., Kassist, I., & Koren, R. (1996). Safety and immunogenicity of a novel mammalian cell-derived recombinant hepatitis B vaccine containing Pre-S1 and Pre-S2 antigens in children. *Vaccine*, 14(3), 207–211. doi:10.1016/0264-410x(95)00185-4
- Reed, J., & Reed, T. A. (1997). A set of constructed type spectra for the practical estimation of peptide secondary structure from circular dichroism. *Analytical Biochemistry*, 254(1), 36–40. doi:10.1006/abio.1997.2355
- Rendic, D., Wilson, I. B. H., & Paschinger, K. (2008). The glycosylation capacity of insect cells. *Croatica Chemica Acta*, 81(1), 7-21. doi:10.1002/chin.200831267
- Richter, L. J., Thanavala, Y., Arntzen, C. J., & Mason, H. S. (2000). Production of hepatitis B surface antigen in transgenic plants for oral immunization. *Nature Biotechnology*, 18(11), 1167-1171. doi:10.1038/81153
- Roldão, A., Mellado, M. C. M., Castilho, L. R., Carrondo, M. J. T., & Alves, P. M. (2010). Virus-like particles in vaccine development. *Expert Review of Vaccines*, 9(10), 1149–1176. <https://doi.org/10.1586/erv.10.115>
- Romani, S., Hosseini, S. M., Mohebbi, S. R., Boonstra, A., Razavi, A. H., & Sharifian, A. (2018). Characterization of the “a” determinant region of the hepatitis B virus genome in Iranian patients at different clinical phases of chronic infection.

*Gastroenterology and Hepatology from Bed to Bench*, 11(2), 131–137.  
doi:10.22037/ghfbb.v0i0.1226

- Ryu, C. J., Gripon, P., Park, H. R., Park, S. S., Kim, Y. K., Guguen-Guilhouzo, C., ... Hong, H. J. (1997). *In vitro* neutralization of hepatitis b virus by monoclonal antibodies against the viral surface antigen. *Journal of Medical Virology*, 52(2), 226–233. doi:10.1002/(SICI)1096-9071(199706)52:2<226::AID-JMV18>3.0.CO;2-I
- Sahul Hameed, A. S., & Bonami, J. R. (2012). White tail disease of freshwater prawn, *Macrobrachium rosenbergii*. *Indian Journal of Virology*, 23(2), 134–140. doi:10.1007/s13337-012-0087-y
- Sato, S., Li, K., Kameyama, T., Hayashi, T., Ishida, Y., Murakami, S., ... Takaoka, A. (2015). The RNA sensor RIG-I dually functions as an innate sensor and direct antiviral factor for hepatitis B virus. *Immunity*, 42(1), 123–132. doi:10.1016/j.immuni.2014.12.016
- Schaefer, S., Melber, K., Jenzelewski, V., Müller, F., Dahlems, U., Bartelsen, O., ... Gellissen, G. (2002). Recombinant Hepatitis B Vaccines: Disease Characterization and Vaccine Production. In G. Gellissen (Ed.), *Production of Recombinant Proteins* (pp 319-359). Weinheim, Germany: Wiley-VCH. doi:10.1002/3527603670.ch15
- Schuch, A., Hoh, A., & Thimme, R. (2014). The role of natural killer cells and CD8+ T cells in hepatitis B virus infection. *Frontiers in Immunology*, 5(258), 1–8. doi:10.3389/fimmu.2014.00258
- Seeger, C., & Mason, W. S. (2003). Hepatitis B virus biology. *Microbiology and Molecular Biology Reviews*, 64(1), 51–68. doi:10.1128/mmbr.64.1.51-68.2000
- Senapin, S., Jaengsanong, C., Phiwsaiya, K., Prasertsri, S., Laisutisan, K., Chuchird, N., ... Flegel, T. W. (2012). Infections of MrNV (*Macrobrachium rosenbergii nodavirus*) in cultivated whiteleg shrimp *Penaeus vannamei* in Asia. *Aquaculture*, 338–341, 41–46. doi:10.1016/j.aquaculture.2012.01.019
- Seto, W., Lo, Y., Pawlotsky, J., & Yuen, M. (2018). Chronic hepatitis B virus infection. *The Lancet*, 392(10161), 2313–2324. doi:10.1016/S0140-6736(18)31865-8
- Shchelkunov, S. N., & Shchelkunova, G. A. (2010). Plant-based vaccines against human hepatitis B virus. *Expert Review of Vaccines*, 9(8), 947–955. doi:10.1586/erv.10.67
- Shouval, D., Ilan, Y., Adler, R., Deepen, R., Panet, A., Even-Chen, Z., ... Gerlich, W. H. (1994). Improved immunogenicity in mice of a mammalian cell-derived recombinant hepatitis B vaccine containing pre-S 1 and pre-S 2 antigens as compared with conventional yeast-derived vaccines. *Vaccine*, 12(15), 1453–1459. doi:10.1016/0264-410X(94)90155-4

- Shouval, D. (2003). Hepatitis B vaccines. *Journal of Hepatology*, 39(1), 70–76. doi:10.1016/S0168-8278(03)00152-1
- Shu, L., Touzjian, N., Nan, D., Kushner, N., Strong, A. J., Zeping, W., ... Lu, Y. (2006). Recombinant hepatitis B large surface antigen, successfully produced in *Escherichia coli*, stimulates T-cell response in mice. *Vaccine*, 24(20), 4409–4416. doi:10.1016/j.vaccine.2006.02.048
- Small, E. J., Fratesi, P., Reese, D. M., Strang, G., Laus, R., Peshwa, M. V., & Valone, F. H. (2000). Immunotherapy of hormone-refractory prostate cancer with antigen-loaded dendritic cells. *Journal of Clinical Oncology*, 18(23), 3894–3903. doi:10.1200/JCO.2000.18.23.3894
- Smith, G. P. (1985). Filamentous fusion phage: Novel expression vectors that display cloned antigens on the virion surface. *Science*, 228(4705), 1315–1317. doi:10.1126/science.4001944
- Sominskaya, I., Skrastina, D., Dislers, A., Vasiljev, D., Mihailova, M., Ose, V., ... Pumpens, P. (2010). Construction and immunological evaluation of multivalent hepatitis B virus (HBV) core virus-like particles carrying HBV and HCV epitopes. *Clinical and Vaccine Immunology*, 17(6), 1027–1033. doi:10.1128/CVI.00468-09
- Somrit, M., Watthammawut, A., Chotwiwatthanakun, C., Ounjai, P., Suntimanawong, W., & Weerachatyanukul, W. (2017). C-terminal domain on the outer surface of the *Macrobrachium rosenbergii* nodavirus capsid is required for Sf9 cell binding and internalization. *Virus Research*, 227(1), 41–48. doi:10.1016/j.virusres.2016.09.017
- Sri Widada, J., Durand, S., Cambournac, I., Qian, D., Shi, Z., Dejonghe, E., ... Bonami, J. R. (2003). Genome-based detection methods of *Macrobrachium rosenbergii* nodavirus, a pathogen of the giant freshwater prawn, *Macrobrachium rosenbergii*: Dot-blot, *in situ* hybridization and RT-PCR. *Journal of Fish Diseases*, 26(10), 583–590. doi:10.1046/j.1365-2761.2003.00493.x
- Stanbridge, L. J., Dussupt, V., & Maitland, N. J. (2003). Baculoviruses as vectors for gene therapy against human prostate cancer. *Journal of Biomedicine and Biotechnology*, 2003(2), 79–91. doi:10.1155/S1110724303209049
- Stephenne, J. (1990). Development and production aspects of a recombinant yeast-derived hepatitis B vaccine. *Vaccine*, 8, S69–S73. doi:10.1016/0264-410X(90)90221-7
- Stibbe, W., & Gerlich, W. H. (1983). Structural relationships between minor and major proteins of hepatitis B surface antigen. *Journal of Virology*, 46(2), 626–628.
- Sun, C., Fu, B., Gao, Y., Liao, X., Sun, R., Tian, Z., & Wei, H. (2012). TGF- $\beta$ 1 down-regulation of NKG2D/DAP10 and 2B4/SAP expression on human NK cells contributes to HBV persistence. *PLoS Pathogens*, 8(3).

doi:10.1371/journal.ppat.1002594

- Sun, Y., Wang, S., Yi, Y., Zhang, J., Duan, Z., Yuan, K., ... Zhu, Y. (2018). The hepatitis B surface antigen binding protein: An immunoglobulin G constant region-like protein that interacts with HBV envelop proteins and mediates HBV entry. *Frontiers in Cellular and Infection Microbiology*, 8, 338. doi:10.3389/fcimb.2018.00338
- Tan, G. H., Yusoff, K., Seow, H. F., & Tan, W. S. (2005). Antigenicity and immunogenicity of the immunodominant region of hepatitis B surface antigen displayed on bacteriophage T7. *Journal of Medical Virology*, 77(4), 475–480. doi:10.1002/jmv.20479
- Tan, W. S., & Ho, K. L. (2014). Phage display creates innovative applications to combat hepatitis B virus. *World Journal of Gastroenterology*, 20(33), 11650–11670. doi:10.3748/wjg.v20.i33.11650
- Thong, Q. X., Biabanikhankahdani, R., Ho, K. L., Alitheen, N. B., & Tan, W. S. (2019). Thermally-responsive virus-like particle for targeted delivery of cancer drug. *Scientific Reports*, 9(1), 1–14. <https://doi.org/10.1038/s41598-019-40388-x>
- Tjia, S. T., zu Altenschildesche, G. M., & Doerfler, W. (1983). *Autographa californica* Nuclear Polyhedrosis Virus (AcNPV) DNA does not persist in mass cultures of mammalian cells. *Virology*, 125, 107–117. doi: 10.1016/0042-6822(83)90067-3
- Tjwa, E. T. T. L., Van Oord, G. W., Hegmans, J. P., Janssen, H. L. A., & Woltman, A. M. (2011). Viral load reduction improves activation and function of natural killer cells in patients with chronic hepatitis B. *Journal of Hepatology*, 54(2), 209–218. doi:10.1016/j.jhep.2010.07.009
- Toprani, V. M., Cheng, Y., Wahome, N., Khasa, H., Kueltzo, L. A., Schwartz, R. M., ... Volkin, D. B. (2018). Structural characterization and formulation development of a trivalent equine encephalitis virus-like particle vaccine candidate. *Journal of Pharmaceutical Sciences*, 107(10), 1–15. doi:10.1016/j.xphs.2018.05.022
- Tsai, K., Kuo, C., & Ou, J. J. (2018). Mechanisms of Hepatitis B Virus Persistence. *Trends in Microbiology*, 26(1), 33–42. doi:10.1016/j.tim.2017.07.006
- Tseng, T. C., & Huang, L. R. (2017). Immunopathogenesis of hepatitis B virus. *Journal of Infectious Diseases*, 216(8), S765–S770. doi:10.1093/infdis/jix356
- Tzeng, H. T., Tsai, H. F., Liao, H. J., Lin, Y. J., Chen, L., Chen, P. J., & Hsu, P. N. (2012). PD-1 blockage reverses immune dysfunction and hepatitis B viral persistence in a mouse animal model. *PLoS ONE*, 7(6), 1–9. doi:10.1371/journal.pone.0039179
- Valaydon, Z. S., & Locarnini, S. A. (2017). The virological aspects of hepatitis B. *Best Practice and Research: Clinical Gastroenterology*, 31(3), 257–264.

doi:10.1016/j.bpg.2017.04.013

- Valenzuela, P., Medina, A., Rutter, W. J., Ammerer, G., & Hall, B. D. (1982). Synthesis and assembly of hepatitis B virus surface antigen particles in yeast. *Nature*, 298(5872), 347–350. doi:10.1038/298347a0
- Van Der Molen, R. G., Sprengers, D., Binda, R. S., De Jong, E. C., Niesters, H. G. M., Kusters, J. G., ... Janssen, H. L. A. (2004). Functional impairment of myeloid and plasmacytoid dendritic cells of patients with chronic hepatitis B. *Hepatology*, 40(3), 738–746. doi:10.1002/hep.20366
- Van Oers, M. M., Pijlman, G. P., & Vlak, J. M. (2015). Thirty years of baculovirus-insect cell protein expression: From dark horse to mainstream technology. *Journal of General Virology*, 96(1), 6–23. doi:10.1099/vir.0.067108-0
- Venkatakrishnan, B., & Zlotnick, A. (2016). The structural biology of hepatitis B virus: Form and function. *Annual Review of Virology*, 3(1), 429–451. doi:10.1146/annurev-virology-110615-042238
- Vogt, G., & Argos, P. (1997). Protein thermal stability: Hydrogen bonds or internal packing? *Folding and Design*, 2(4), 40–46. doi:10.1016/S1359-0278(97)00062-X
- Wang, W., Feng, F., Lv, J., Xie, Z., Chen, J., Zhang, L., & Li, W. (2017). Major immunodominant region of hepatitis B virus core antigen as a delivery vector to improve the immunogenicity of the fusion antigen ROP2-SAG1 multiepitope from *Toxoplasma gondii* in mice. *Viral Immunology*, 30(7), 508–515. doi:10.1089/vim.2016.0135
- Wang, X., Gao, L., Deng, F., Zhang, Y., Li, Y., & Lin, J. (2007). High-level production of a functional recombinant hepatitis B virus polymerase in insect cells with a baculovirus expression system. *Journal of Huazhong University of Science and Technology*, 27(3), 269–273. doi:10.1007/s11596-007-0313-9
- Waters, J. A., Kennedy, M., Voet, P., Hauser, P., Petre, J., Carman, W., & Thomas, H. C. (1992). Loss of the common “a” determinant of hepatitis B surface antigen by a vaccine-induced escape mutant. *Journal of Clinical Investigation*, 90(6), 2543–2547. doi:10.1172/JCI116148
- Wei, S., Lei, Y., Yang, J., Wang, X., Shu, F., Wei, X., ... Wei, S. (2018). Neutralization effects of antibody elicited by chimeric HBV S antigen viral-like particles presenting HCV neutralization epitopes. *Vaccine*, 36(17), 2273–2281. doi:10.1016/j.vaccine.2018.03.036
- Whitsett, M., Feldman, D. M., & Pan, C. Q. (2019). Risk assessment and management of hepatitis B reactivation from direct-acting antivirals for hepatitis C. *Liver Research*, 3(2). doi:10.1016/j.livres.2019.03.002
- WHO (2015). Guidelines for the prevention, care and treatment of persons with chronic hepatitis B infection. *World Health Organization* (pp 1-166). Geneva,

Switzerland: WHO Press.

- Wu, C. C., Chen, Y. S., Cao, L., Chen, X. W., & Lu, M. J. (2018). Hepatitis B virus infection: Defective surface antigen expression and pathogenesis. *World Journal of Gastroenterology*, 24(31), 3488–3499. doi:10.3748/wjg.v24.i31.3488
- Wu, J., Zhang, X. J., Shi, K. Q., Chen, Y. P., Ren, Y. F., Song, Y. J., ... Tang, K. F. (2014). Hepatitis B surface antigen inhibits MICA and MICB expression via induction of cellular miRNAs in hepatocellular carcinoma cells. *Carcinogenesis*, 35(1), 155–163. doi:10.1093/carcin/bgt268
- Xing, J., Singh, S., Zhao, Y., Duan, Y., Guo, H., Hu, C., ... Chen, J. (2017). Increasing vaccine production using pulsed ultrasound waves. *PLoS ONE*, 12(11), 1–17. doi:10.1371/journal.pone.0187048
- Xu, F., Song, H., Xiao, Q., Li, N., Zhang, H., Cheng, G., & Tan, G. (2019). Type III interferon-induced CBF  $\beta$  inhibits HBV replication by hijacking HBx. *Cellular and Molecular Immunology*, 16(4), 357–366. doi:10.1038/s41423-018-0006-2
- Yang, J. T., Wu, C. C., & Martinez, H. M. (1986). Calculation of protein conformation from circular dichroism. *Methods in Enzymology*, 130, 208–269. doi:10.1016/0076-6879(86)30013-2.
- Yang, S., Wang, L., Pan, W., Bayer, W., Thoens, C., Heim, K., ... Liu, J. (2019). MMP2/MMP9-mediated CD100 shedding is crucial for inducing intrahepatic anti-HBV CD8 T cell responses and HBV clearance. *Journal of Hepatology*, 71(4), 685–698. doi:10.1016/j.jhep.2019.05.013
- Ye, Q., Shang, S. Q., & Li, W. (2015). A new vaccine escape mutant of hepatitis B virus causes occult infection. *Human Vaccines and Immunotherapeutics*, 11(2), 407–410. doi:10.4161/21645515.2014.994461
- Yong, C. Y., Yeap, S. K., Goh, Z. H., Ho, K. L., Omar, A. R., & Tan, W. S. (2015a). Induction of humoral and cell-mediated immune responses by hepatitis B virus epitope displayed on the virus-like particles of prawn nodavirus. *Applied and Environmental Microbiology*, 81(3), 882–889. doi:10.1128/AEM.03695-14
- Yong, C. Y., Yeap, S. K., Ho, K. L., Omar, A. R., & Tan, W. S. (2015b). Potential recombinant vaccine against influenza A virus based on M2e displayed on nodaviral capsid nanoparticles. *International Journal of Nanomedicine*, 10, 2751–2763. doi:10.2147/IJN.S77405
- Youm, J. W., Won, Y. S., Jeon, J. H., Ryu, C. J., Choi, Y. K., Kim, H. C., ... Kim, H. S. (2007). Oral immunogenicity of potato-derived HBsAg middle protein in BALB/c mice. *Vaccine*, 25(3), 577–584. doi:10.1016/j.vaccine.2006.05.131
- Zepeda-Cervantes, J., Ramírez-Jarquín, J. O., & Vaca, L. (2020). Interaction Between Virus-Like Particles (VLPs) and Pattern Recognition Receptors (PRRs) From Dendritic Cells (DCs): Toward Better Engineering of VLPs. *Frontiers in Immunology*, 11(June), 1–22. <https://doi.org/10.3389/fimmu.2020.01100>

- Zha, D. (2013). Glycosylation engineering of biopharmaceuticals. In A. Beck (Ed.), *Glycosylation engineering of biopharmaceuticals: Methods and protocols, methods in molecular biology*. Saint Julien-en-Genevois, France: Humana Press. doi:10.1007/978-1-62703-327-5
- Zhai, Y., Zhang, D., Yu, L., Sun, F., & Sun, F. (2019). SmartBac, a new baculovirus system for large protein complex production. *Journal of Structural Biology*, 1, 100003. doi:10.1016/j.jsbx.2019.100003
- Zhang, Q., Zhong, J., & Huan, L. (2011). Expression of hepatitis B virus surface antigen determinants in *Lactococcus lactis* for oral vaccination. *Microbiological Research*, 166(2), 111–120. doi:10.1016/j.micres.2010.02.002
- Zhang, S., Zhao, J., & Zhang, Z. (2018). Humoral immunity, the underestimated player in hepatitis B. *Cellular and Molecular Immunology*, 15(6), 645–648. doi:10.1038/cmi.2017.132
- Zhao, K., Wu, C., Yao, Y., Cao, L., Zhang, Z., Yuan, Y., ... Chen, X. (2017). Ceruloplasmin inhibits the production of extracellular hepatitis B virions by targeting its middle surface protein. *Journal of General Virology*, 98, 1410–1421. doi:10.1099/jgv.0.000794