



UNIVERSITI PUTRA MALAYSIA

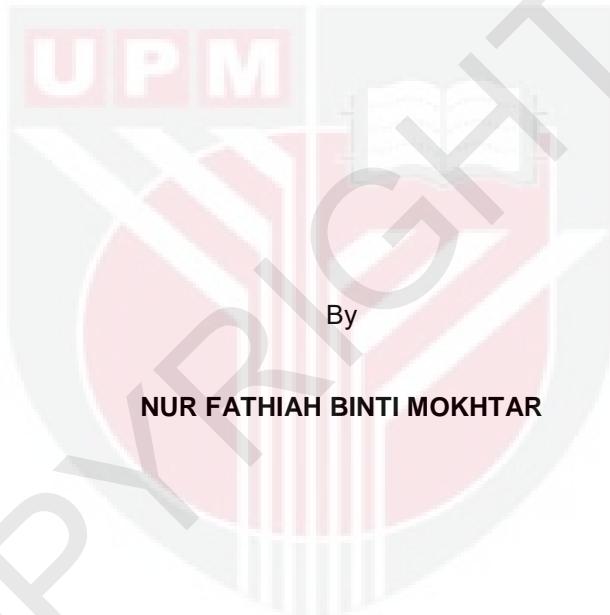
***EXTRACTION, REIMMOBILIZATION AND CHARACTERIZATION OF  
SPENT IMMOBILIZED LIPASE***

NUR FATHIAH BINTI MOKHTAR

FBSB 2020 22



**EXTRACTION, REIMMOBILIZATION AND CHARACTERIZATION OF  
SPENT IMMOBILIZED LIPASE**



Thesis Submitted to the School of Graduate Studies, Universiti Putra  
Malaysia, in Fulfilment of the Requirements for the  
Degree of Master of Science

June 2020

## **COPYRIGHT**

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in  
fulfilment of the requirement for the degree of Master of Science

**EXTRACTION, REIMMOBILIZATION AND CHARACTERIZATION OF  
SPENT IMMOBILIZED LIPASE**

By

**NUR FATHIAH BINTI MOKHTAR**

**June 2020**

**Chairman : Associate Professor Mohd Shukuri Mohamad Ali, PhD**  
**Faculty : Biotechnology and Biomolecular Sciences**

Currently, lipases are considered as one of the important catalysts in substituting the use of chemical reactions in a wide variety of processes. In 2015, the report estimated that the U.S. enzyme's market had a high demand for processed foods and synthesis reactions, which generated more than USD 1 billion in sales. However, the reusability of the industrial immobilized lipase was limited only after several cycles of reactions. After that, the spent immobilized lipase will be replaced with the new immobilized lipase. The inability to reuse the spent immobilized lipase leads to an increased cost required for the new uptake of the enzyme. Practically in industry, the spent immobilized lipase is a waste from industrial users. The spent immobilized lipase still has potential use even though the activity was lower than the new immobilized lipase. There was no study done on the extraction and reimmobilization of the enzyme from the spent immobilized lipase. Therefore, the objective of this research is to study the feasibility of the spent lipase to be extracted, reimmobilized, and characterized.

General methodology involved the recovery of the spent immobilized lipase via chemical and mechanical extraction. The chemical extraction approach via Reverse Micelles Extraction (RME) showed the highest lipase recovery, which was 66% compared to the 34% of lipase yield obtained from the mechanical extraction method. The extracted lipase was reimmobilized via simple adsorption into the ethanol pretreated carrier. The characterization of the reimmobilized lipase at different pH and temperature was conducted. The optimum conditions of immobilization resulted in 96% of the extracted lipase being immobilized. The reimmobilized lipase optimum activity was at 50°C and pH 6. The reimmobilized lipase was incubated for 20 h in pH 6 buffer at 50°C of water bath shaker. The reimmobilized lipase still had 27% residual activity after 18 h of incubation, which indicated higher thermal stability compared to the free lipase.

The Scanning Electron Microscope (SEM) was used to study the morphology of the reimmobilized lipase. The morphological of immobilized lipase was analyzed based on the pore and the particle sizes of the support. SEM also showed oil on the surface of immobilized lipase before and after the solvent treatment. The structural analysis of free lipase and reimmobilized lipase was determined by Fourier-transform infrared spectroscopy (FTIR). The structures of the amide group I (CO stretch) and amide group II (NH bend), which formed the functional group of the free commercial lipase, extracted lipase and reimmobilized lipase, had been identified. In conclusion, the free lipase was successfully extracted from the spent immobilized lipase and reimmobilized into Accurel MP1008 carrier. It exhibited high thermal stability, and the reusability of the spent enzyme will promote continued use of industrial lipase and reduce the cost of the manufacturing process.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk ijazah Master Sains

**PENGEKSTRAKAN, PENYEKATGERAKAN DAN PENCIRIAN LIPASE  
TERSEKATGERAK YANG TELAH DIGUNAKAN**

Oleh

**NUR FATHIAH BINTI MOKHTAR**

**Jun 2020**

**Pengerusi : Profesor Madya Mohd Shukuri Mohamad Ali, PhD**  
**Fakulti : Bioteknologi dan Sains Biomolekul**

Pada masa ini, lipase dianggap sebagai salah satu pemangkin penting dalam mengantikan penggunaan tindak balas kimia di dalam pelbagai proses. Pada tahun 2015, berdasarkan laporan pasaran enzim, A.S dianggarkan mempunyai permintaan enzim yang tinggi untuk pemprosesan makanan dan tindak balas sintesis yang menjana lebih daripada 1 bilion USD jualan. Walau bagaimanapun, kebolehgunaan semula lipase industri hanya terhad selepas beberapa kitaran tindak balas. Kemudian, lipase tersekatgerak lama akan digantikan dengan lipase tersekatgerak yang baru. Ketidakupayaan untuk menggunakan lipase tersekatgerak yang lama akan meningkatkan kos yang diperlukan untuk lipase tersekatgerak yang baru. Secara praktikal, lipase tersekatgerak yang telah digunakan merupakan sisa buangan dari penggunaan industri. Lipase tersekatgerak yang telah digunakan masih mempunyai potensi untuk digunakan semula walaupun aktivitinya lebih rendah dari lipase tersekatgerak yang baru. Tiada kajian dilakukan terhadap pengekstrakan dan penyekatgerakan semula enzim daripada lipase tersekatgerak yang telah digunakan. Oleh itu, objektif kajian ini adalah untuk mengkaji kemungkinan lipase yang telah digunakan untuk diekstrak, disekatgerak semula, dan dicirikan.

Metodologi umum melibatkan pemulihan lipase tersekatgerak yang telah digunakan melalui pengekstrakan kimia dan mekanikal. Pendekatan pengekstrakan kimia melalui ekstraksi misel terbalik (RME) menunjukkan pemulihan lipase tertinggi, iaitu 66% berbanding dengan 34% hasil lipase diperolehi dari kaedah pengekstrakan mekanikal. Lipase yang diekstrak telah disekatgerakan semula melalui penjerapan mudah ke dalam pembawa yang telah dirawat oleh ethanol. Keadaan penyekatgerakan yang optimum menyebabkan 96% lipase yang diekstrak telah disekatgerak. Pencirian lipase tersekatgerak telah dijalankan pada pH dan suhu yang berbeza. Aktiviti optimum untuk lipase tersekatgerak adalah pada 50°C dan pH 6. Lipase tersekatgerak

telah diinkubasi selama 20 jam di dalam penimbal pH 6 dan 50°C penggoncang rendaman air. Lipase tersekatgerak masih mempunyai lebihan aktiviti sebanyak 27% selepas diinkubasi selama 18 jam yang menunjukkan kestabilan haba yang lebih tinggi berbanding dengan lipase bebas.

Mikroskop elektron pengimbas (SEM) telah digunakan untuk mengkaji morfologi lipase tersekatgerak. Morfologi lipase tersekatgerak dianalisis berdasarkan liang pori dan saiz pembawa enzim. SEM juga menunjukkan kehadiran minyak pada permukaan lipase tersekatgerak sebelum dan selepas dirawat oleh pelarut. Analisis struktur lipase bebas dan lipase tersekatgerak ditentukan oleh spektroskopi inframerah transformasi Fourier (FTIR). Struktur amide group I (CO stretch) dan amide group II (NH bend) yang membentuk kumpulan berfungsi dalam lipase komersial bebas, lipase yang diekstrak dan lipase tersekatgerak, telah dikenalpasti.

Kesimpulannya, lipase bebas telah berjaya diekstrak dari lipase tersekatgerak yang telah digunakan dan disekatgerak semula ke dalam pembawa Accurel MP1008. Ia memperbaiki kestabilan haba yang tinggi, dan kebolehgunaan semula enzim buangan akan menggalakkan penggunaan lipase industri secara terus dan mengurangkan kos untuk proses pembuatan.

## **ACKNOWLEDGEMENTS**

Alhamdulillah, all praises to Allah S.W.T for giving me the patience, strength, and opportunity to complete this project with success.

First and foremost, I would like to express my deepest gratitude to my supervisor Prof. Madya Dr. Mohd Shukuri Mohamad Ali for his supports, comments, suggestions, and guidance during the preparation of this thesis.

I would like to express my sincere gratitude to Prof. Dr. Raja Noor Zaliha Bt Raja Abd. Rahman for giving her permission and guidance to use the equipment facilities from The Enzyme and Microbial Technology Research Center (EMTech) at both Biotech 2 and Institute of Bioscience (IBS), UPM.

I would also like to thank all my labmates especially Fatima, Fatin, Farah, Syazwani, Izzati, Fira, and Shida who have been supportive and gave many ideas for my project and sharing the space and equipment throughout these years.

Last but not least, I am grateful to my beloved parents and siblings who always gave their blessing, support, and inspiration for me to finish up this project. Special thanks also to my brother, who always gave his moral and financial supports for my study.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

**Mohd. Shukuri Mohamad Ali, PhD**

Associate Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Chairman)

**Raja Noor Zaliha Raja Abd Rahman, D. Eng**

Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Member)

---

**ZALILAH MOHD SHARIFF, PhD**

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date: 10 September 2020

## **Declaration by graduate student**

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature: \_\_\_\_\_

Date: \_\_\_\_\_

Name and Matric No: Nur Fathiah binti Mokhtar, GS49461

## **Declaration by Members of Supervisory Committee**

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature:

---

Name of Chairman  
of Supervisory  
Committee:

Associate Professor

Dr. Mohd. Shukuri Mohamad Ali

Signature:

---

Name of Member  
of Supervisory  
Committee:

Professor

Dr. Raja Noor Zaliha Raja Abd Rahman, D. Eng

## TABLE OF CONTENTS

	Page
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	iii
<b>ACKNOWLEDGEMENTS</b>	v
<b>APPROVAL</b>	vi
<b>DECLARATION</b>	viii
<b>LIST OF TABLES</b>	xiii
<b>LIST OF FIGURES</b>	xiv
<b>LIST OF ABBREVIATIONS</b>	xvi
 <b>CHAPTER</b>	
 <b>1 INTRODUCTION</b>	 1
1.1 Background of the study	1
1.2 Problem statement	1
1.3 Significance of the study	2
1.4 Research objectives	2
1.4.1 General objective	2
1.4.2 Specific objectives	2
 <b>2 LITERATURE REVIEW</b>	 3
2.1 Lipases	3
2.2 Industrially important lipases	3
2.3 Immobilization of lipases	7
2.3.1 Supports for enzyme immobilization	8
2.3.2 Immobilization techniques	10
2.3.3 The interfacial activation of lipases	11
2.4 Application of immobilized lipase	12
2.5 Recovery of enzymes and extraction of lipases	13
2.5.1 Reverse micellar extraction (RME)	13
2.6 Methods to incorporate enzymes in reverse micelles	14
2.6.1 Phase transfer method	15
2.6.1.1 Forward extraction (FE)	15
2.6.1.2 Backward extraction	15
2.6.2 Injection method	16
2.6.3 Dry addition method	16
2.7 Effects of process parameters on RME	17
2.7.1 Surfactant type and concentrations	17
2.7.2 Aqueous phase pH	18
2.7.3 Salt concentration	18
2.8 Biochemical characterization of immobilized lipase	18
2.8.1 Effect of temperature on the immobilized lipase	18
2.8.2 Storage stability of immobilized lipase	19
2.8.3 Effect of pH on immobilized lipases stability	19

2.9	Morphological and structural analysis of immobilized lipases	20
2.9.1	Scanning Electron Microscope (SEM)	20
2.9.2	Fourier-transform infrared spectroscopy (FTIR)	20
<b>3</b>	<b>MATERIALS AND METHODS</b>	<b>21</b>
3.1	Chemicals, Reagents, and Equipment	21
3.2	Sources of <i>Rhizopus oryzae</i> lipase (ROL) and immobilized lipase	21
3.3	Flow chart of overall experiments	21
3.4	Examination of the spent immobilized lipase	22
3.4.1	Morphological analysis	23
3.4.2	Measurement of enzyme activity	23
3.4.3	Estimation of protein concentration	25
3.5	Extraction of lipase from spent immobilized lipase	25
3.5.1	Mechanical approach using waterbath shaker	25
3.5.1.1	Treatment of spent immobilized lipase	26
3.5.2	Chemical approach via reverse micellar extraction (RME)	26
3.5.2.1	Reverse micellar optimization	27
3.6	Reimmobilization of extracted lipase	27
3.6.1	Optimization of the reimmobilization method	28
3.6.1.1	Amount of support	28
3.6.1.2	Amount of extracted lipase	29
3.6.1.3	Support pretreatment with absolute ethanol	29
3.7	Characterization of the extracted lipase and reimmobilized lipase	29
3.7.1	Effect of temperature on lipase activity	30
3.7.2	Effect of temperature on lipase stability	30
3.7.3	Effect of pH on lipase activity	30
3.7.4	Storage stability	30
3.7.5	Structural analysis via Fourier-transform infrared spectroscopy (FTIR)	30
3.8	Statistical analysis	31
<b>4</b>	<b>RESULTS AND DISCUSSION</b>	<b>32</b>
4.1	Morphology analysis of immobilized lipase using SEM	32
4.2	Immobilized lipase activity	36
4.3	Extraction of lipase from spent immobilized lipase by mechanical and chemical (reverse micelles) methods	39
4.3.1	Mechanical extraction using waterbath shaker	39

4.3.2	Chemical extraction using reverse micellar extraction (RME)	44
4.3.2.1	Effect of surfactant concentration	45
4.3.2.2	Effect of ionic strength in the forward extraction	46
4.3.2.3	Effect of the stripping solution pH	47
4.3.2.4	Effect of ionic strength in the backward extraction	48
4.4	Optimization of lipase reimmobilization	49
4.4.1	Effect of the support amount	50
4.4.2	Effect of extracted lipase amount	51
4.4.3	Effect of support pretreatment with ethanol	52
4.5	Characterization of the extracted lipase and reimmobilized lipase	53
4.5.1	Effect of temperature on lipase activity	53
4.5.2	Thermal stability of the extracted and reimmobilized lipase	55
4.5.3	Effect of pH on lipase activity	57
4.5.4	Storage stability of extracted lipase and reimmobilized lipase	60
4.5.5	Fourier-transform infrared spectroscopy analysis	65
<b>5</b>	<b>CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH</b>	<b>69</b>
5.1	Conclusion	69
5.2	Recommendations for future research	69
<b>REFERENCES</b>		<b>71</b>
<b>APPENDICES</b>		<b>84</b>
<b>BIODATA OF STUDENT</b>		<b>88</b>
<b>PUBLICATION</b>		<b>89</b>

## LIST OF TABLES

<b>Table</b>		<b>Page</b>
1	Effects of immobilization on different sources of lipases	5
2	List of immobilized lipase for morphological analysis	23
3	List of immobilized lipase for activity measurement	24
4	List of immobilized lipase for mechanical separation	26
5	List of sample for the biochemical and structural characterization	29
6	SEM results for fresh support, spent support and denatured support before and after isoctane treatment	36
7	Optimum time for maximum enzyme extracted from support, protein concentration, extracted lipase activity, and lipase recovery after mechanical separation	44
8	The protein concentration, lipase activity, and lipase recovery of extracted lipase after RME	44
9	Optimum conditions for reverse micelles extraction (RME)	49
10	Optimum conditions after biochemical characterization	64

## LIST OF FIGURES

<b>Figure</b>		<b>Page</b>
1	Immobilization of enzyme into a porous carrier	9
2	Scheme of various enzyme immobilization techniques	10
3	The immobilization methods and supports materials	11
4	Scheme of immobilization by interfacial activation on hydrophobic support	12
5	Schematic diagram of Reverse Micellar Extraction (RME) by using the phase transfer method	14
6	Flow chart of overall experiments	22
7	SEM images for supports (a) Empty support	33
8	SEM images of supports before and after treated with isoctane	35
9	Effect of solvents and surfactants treatment on spent immobilized lipase activity=	38
10	Effect of shaking time on lipase activity which extracted from fresh immobilized lipase=	40
11	Effect of shaking time on lipase activity which extracted from spent immobilized lipase=	41
12	Effect of shaking time on lipase activity which extracted from spent immobilized lipase treated with acetone	41
13	Effect of shaking time on lipase activity which extracted from spent immobilized lipase treated with hexane	42
14	Effect of shaking time on lipase activity which extracted from spent immobilized lipase treated with Tween 20	43
15	Effect of surfactant concentration on extracted lipase activity	46
16	Effect of NaCl concentration in initial aqueous phase on extracted lipase activity	47
17	Effect of stripping solution pH on extracted lipase activity	48

18	Effect of KCl concentration in backward phase on extracted lipase activity	49
19	Effect of support amount on immobilization yield (%)	51
20	Effect of enzyme amount on immobilization yield (%)	52
21	Effect of support pretreatment with ethanol on immobilization yield (%)	53
22	Effect of temperature on the extracted lipase (AOT) and reimmobilized lipase (AOT) activity	54
23	Effect of temperature on the extracted lipase (Triton X-100) and reimmobilized lipase (Triton X-100) activity	55
24	Effect of incubation time on extracted lipase (AOT) and reimmobilized lipase (AOT) thermostability from 0 to 20 h of incubation time at 50°C	56
25	Effect of incubation time on extracted lipase (Triton X – 100) and reimmobilized lipase (Triton X – 100) thermostability from 0 to 20 h of incubation time at 50°C	57
26	Effect of pH on the extracted lipase (AOT) and reimmobilized lipase (AOT) activity	58
27	Effect of pH on the extracted lipase (Triton X - 100) and reimmobilized lipase (Triton X - 100) activity	60
28	Storage stability of the extracted lipase (Triton X - 100), reimmobilized lipase (Triton X - 100), extracted lipase (AOT), and reimmobilized lipase (AOT) at 4°C for 150 days	61
29	Storage stability of extracted lipase (Triton X - 100), reimmobilized lipase (Triton X - 100), extracted lipase (AOT) and reimmobilized lipase (AOT) at 25°C for 120 days	63
30	The FTIR spectra of commercial lipase D2, extracted lipase (AOT), reimmobilized lipase (AOT), and the empty support (Accurel MP1008)	66
31	The FTIR spectra of commercial lipase D2, extracted lipase (Triton X - 100), reimmobilized lipase (Triton X - 100), and the empty support (Accurel MP1008)	68

## LIST OF ABBREVIATIONS

Å	Angstrom
µL	Microliter
µm	Micrometer
µmoles	Micromoles
AOT	Bis(2-ethylhexyl) sulfosuccinate sodium salt
BSA	Bovine serum albumin
CaCl <sub>2</sub>	Calcium chloride
g	Gram
h	Hour
HCl	Hydrochloric acid
HEA	Hen egg albumin
KCl	Potassium chloride
K <sub>2</sub> HPO <sub>4</sub>	Dipotassium phosphate
KH <sub>2</sub> PO <sub>4</sub>	Potassium dihydrogen phosphate
L	Liter
M	Molar
mM	Milimolar
mg	Miligram
min	Minute
mL	Mililiter
nm	Nanometer
pH	Negative logarithm of hydrogen ion concentration
RME	Reverse Micelles Extraction

RMs	Reverse Micelles
rpm	Rotation per minute
sec	Second
U/mL	Unit per milliliter



## **CHAPTER 1**

### **INTRODUCTION**

#### **1.1 Background of the study**

Commercial lipase D2 is used in the food processing industry, especially in the production of structured lipids as dietary ingredients derived from fats and oils (Flood & Kondo, 2003; Tecelão et al., 2012). In industrial use, immobilized enzymes had better performance in terms of operational, storage, and thermal stability (Katchalski-Katzir & Kraemer, 2000; Tecelão et al., 2012; Talekar & Chavare, 2012). The study on the reusability of the carrier was conducted for papain enzymes before. The papain enzyme was reimmobilized on the regenerated carrier recovered from spent bound papain by using chelating metal ions to improve enzyme activity (Iqbal & Afaq, 2001). However, the immobilized lipase consists of free lipase and carrier (Accurel MP1008) can only be reused after several cycles (Bosley & Pelow., 1997; Rehm et al., 2016). The simple adsorption technique and extreme condition in the bioreactor system caused the inconsistency of enzyme activity (Rehman et al., 2016). After several usages, the immobilized lipase was considered waste from the industry (Iqbal & Afaq, 2001; Rehman et al., 2016). The waste-immobilized lipase classified into spent and denatured immobilized lipase. The spent immobilized lipase had low activity than the new immobilized lipase but higher activity than the denatured immobilized lipase. Practically, the spent immobilized lipase was treated with boiling water until denatured before being discharged from the bioreactor. The spent immobilized lipase, which potentially had residual activity was retrieved from the bioreactor and used for further analysis. This study is the first attempt to recover the lipase from spent immobilized lipase.

#### **1.2 Problem statement**

Spent immobilized lipase was abundantly present in the reaction after the synthesis process, and once the productivity lost to almost 50% after several cycles of reaction, the spent immobilized lipase will become a waste. The recovery of support or enzyme from the wasted immobilized lipase can significantly reduce the cost required to purchase a new fresh immobilized lipase, especially on both support and enzyme. The price for lipase is very expensive than that carrier. MEMBRANA GMBH Company stated that 1 g of Accurel MP1008 support only cost RM 0.60 per gram. Meanwhile, the price for 1 g of lipase (ROL) is RM 555.00 (Sigma-Aldrich). However, there is a lack of discoveries on the recovery, reimmobilization, and characterization of lipase from spent immobilized lipase.

### **1.3      Significance of the study**

The lipase can be recovered from the spent immobilized lipase, which will significantly reduce the cost needed for the new enzyme. The reimmobilization of extracted lipase was more cost-effective. RME is the most suitable recovery method due the time-saving and easily upscale (Basheer & Thenmozhi, 2010; Gangadharappa et al., 2017; Jalil et al., 2018). No study on the extraction of lipases from spent immobilized lipase was conducted before. After lipases extraction, the optimization of lipase immobilization techniques can be used to enhance the stability of the reimmobilized enzyme using the previous immobilization method (Zhang et al., 2013; Datta et al., 2013). Therefore, the study on lipase extraction from spent immobilized lipase via Reverse Micellar Extraction (RME) can be beneficial. The RME method can be used to upscale the lipase recovery from the spent immobilized lipase.

### **1.4      Research objectives**

#### **1.4.1     General objective:**

To extract, reimmobilize, and characterize the lipase from spent immobilized lipase into industrial use macroporous support (Accurel MP1008). The specifications of the support are listed in Appendix iv.

#### **1.4.2     Specific objectives:**

1. To examine the remaining activity of the spent immobilized lipase
2. To recover the lipase through mechanical and chemical methods
3. To optimize the enzyme reimmobilization
4. To perform biochemical, morphological and structural characterization of the reimmobilized lipase

## REFERENCES

- Adlercreutz, P. (2013). Immobilisation and application of lipases in organic media. *Chemical Society Reviews*, 42(15), 6406.
- Aires-Barros, M. R., & Cabral, J. M. S. (1991). Selective Separation and Purification of two lipases from *chromobacterium viscosum* using AOT reversed micelles. *Biotechnology and Bioengineering*, 38 (11), 1302 - 1307.
- Al-Najada, A. R., Almulaiky, Y. Q., Aldhahri, M., El-Shishtawy, R. M., Mohamed, S. A., Baeshen, M., Al-Harbi, S. A. (2019). Immobilisation of  $\alpha$ -amylase on activated amidrazone acrylic fabric: a new approach for the enhancement of enzyme stability and reusability. *Scientific Reports*, 9(1).
- Ali, Z., Tian, L., Zhang, B., Ali, N., Khan, M., & Zhang, Q. (2017). Synthesis of fibrous and non-fibrous mesoporous silica magnetic yolk-shell microspheres as recyclable supports for immobilization of *Candida rugosa* lipase. *Enzyme and Microbial Technology*, 103, 42 - 52.
- Anand, A., & Weatherley, L.R. (2018). The performance of microbial lipase immobilized onto polyolefin supports for hydrolysis of high oleate sunflower oil. *Process Biochemistry*, 68, 100 - 107.
- Antony, N., Balachandran, S. & Mohanan, P. V. (2016). Immobilization of diastase  $\alpha$ -amylase on nano zinc oxide. *Food Chemistry*, 211, 624 - 630.
- Basheer, S. A., & M.Thenmozhi. (2010). Reverse Micellar Separation of Lipases: A Critical Review. *International Journal of Chemical Sciences*, 8(5), 57 - 67.
- Basri, M., Yunus, W. M. Z. W., Yoong, W. S., Ampon, K., Razak, C. N. A., & Salleh, A. B. (1996). Immobilization of Lipase from *Candida rugosa* on Synthetic Polymer Beads for Use in the Synthesis of Fatty Esters. *Journal of Chemical Technology & Biotechnology*, 66(2), 169 - 173.
- Bassi, J. J., Todero, L. M., Lage, F. A., Khedy, G. I., Ducas, J. D., Custódio, A. P., Mendes, A. A. (2016). Interfacial activation of lipases on hydrophobic support and application in the synthesis of a lubricant ester. *International Journal of Biological Macromolecules*, 92, 900 - 909.
- Bhavya, S. G., Priyanka, B. S., Rastogi, N. K. (2012). Reverse micelles-mediated transport of lipase in liquid emulsion membrane for downstream processing. *Biotechnology Progress*, 28(6), 1542 - 1550.

- Biasutti, M. A., Abuin, E. B., Silber, J. J., Correa, N. M., Lissi, E. A. (2008). Kinetics of reactions catalyzed by enzymes in solutions of surfactants. *Advances in Colloid and Interface Science*, 136, 1 - 24.
- Brady, C., Metcalfe, L., Slaboszewski, D., & Frank, D. (1988). Lipase immobilized on a hydrophobic, microporous support for the hydrolysis of fats. *Journal of the American Oil Chemists Society*, 65(6), 917 - 921.
- Bolivar, J. M., Eisl, I., & Nidetzky, B. (2016). Advanced characterization of immobilized enzymes as heterogeneous biocatalysts. *Catalysis Today*, 259, 66 - 80.
- Bosley, J. A., & Peilow, A. D. (1997). Immobilization of lipases on porous polypropylene: Reduction in esterification efficiency at low loading. *Journal of the American Oil Chemists Society*, 74(2), 107 - 111.
- Bussamara, R., Dallagnol, L., Schrank, A., Fernandes, K. F., & Vainstein, M. H. (2012). Optimal Conditions for Continuous Immobilization of *Pseudozyma hubeiensis* (Strain HB85A) Lipase by Adsorption in a Packed-Bed Reactor by Response Surface Methodology. *Enzyme Research*, 2012, 1 - 12.
- Cabrera, Z., Fernandez-Lorente, G., Fernandez-Lafuente, R., Palomo, J. M., & Guisan, J. M. (2009). Novozym 435 displays very different selectivity compared to lipase from *Candida antarctica B* adsorbed on other hydrophobic supports. *Journal of Molecular Catalysis B: Enzymatic*, 57(1 - 4), 171 - 176.
- Carlson, A., & Nagarajan, R. (1992). Release and Recovery of Porcine Pepsin and Bovine Chymosin from Reverse Micelles: a New Technique Based on Isopropyl Alcohol Addition. *Biotechnology Progress*, 8(1), 85 - 90.
- Cesarini, S., Infanzón, B., Pastor, F., & Diaz, P. (2014). Fast and economic immobilization methods described for non-commercial *Pseudomonas* lipases. *BMC Biotechnology*, 14(1), 27.
- Cesarini, S., Pastor, F. I. J., & Diaz, P. (2014). Improvement of *P.aeruginosa* 42A2 lipase preparations for FAMEs production , both in immobilized and soluble form. *Journal of Molecular Catalysis. B, Enzymatic*, 99, 1 - 7.
- Chang, S. W., Shaw, J. F., Yang, K. H., Chang, S. F., & Shieh, C. J. (2008). Studies of optimum conditions for covalent immobilization of *Candida rugosa* lipase on poly( $\gamma$ -glutamic acid) by RSM. *Bioresource Technology*, 99(8), 2800 - 2805.
- Chaurasiya, R. S., & Hebbar, H. U. (2017). Reverse Micelles for Nanoparticle Synthesis and Biomolecule Separation. *Sustainable Agriculture Reviews Nanoscience in Food and Agriculture*, 4, 181 - 211.

- Chemat, F., & Sawamura, M. (2010). Techniques for Oil Extraction. *Citrus Essential Oils*, 9 - 36.
- Chen, B., Miller, M. E., & Gross, R. A. (2007). Effects of porous polystyrene resin parameters on *Candida antarctica* lipase B adsorption, distribution, and polyester synthesis activity. *Langmuir*, 23(11), 6467 - 6474.
- Çiftçi, O. N., Fadiloğlu, S., & Göögüş, F. (2009). Conversion of olive pomace oil to cocoa butter-like fat in a packed-bed enzyme reactor. *Bioresource Technology*, 100(1), 324 - 329.
- D Voet and J G Voet. (1990). Biochemistry. *John Wiley and Sons, New York*, 1223.
- Datta, S., Christena, L. R., & Rajaram, Y. R. S. (2013). Enzyme immobilization: an overview on techniques and support materials. *Biotech*, 3(1), 1 - 9.
- Deng, H. T., Xu, Z. K., Huang, X. J., Wu, J., & Seta, P. (2004). Adsorption and activity of *Candida rugosa* lipase on polypropylene hollow fiber membrane modified with phospholipid analogous polymers. *Langmuir*, 20(23), 10168 - 10173.
- Dey G., B. Singh and R. Banerjee. (2003). Immobilization of  $\alpha$ - amylase Produced by *Bacillus circulans* GRS 313. *Brazilian Archives of Biology and Technology*, 46(2), 167-176.
- DiCosimo, R., McAuliffe, J., Poulose, A. J., Bohlmann, G., Kumar, H., Satyanarayanan, T., Langan, P. (2013). Industrial use of immobilized enzymes. *Chemical Society Reviews*, 42(15), 6437 - 6474.
- Dizge, N., Keskinler, B., & Tanriseven, A. (2008). Covalent attachment of microbial lipase onto microporous styrene–divinylbenzene copolymer by means of polyglutaraldehyde. *Colloids and Surfaces B: Biointerfaces*, 66(1), 34 - 38.
- Dungan, S. R., Bausch, T., Hatton, T. A., Plucinski, P., & Nitsch, W. (1991). Interfacial transport processes in the reversed micellar extraction of proteins. *Journal of Colloid and Interface Science*, 145(1), 33 - 50.
- Dwevedi, A. (2016). Enzyme immobilization: Advances in industry, agriculture, medicine, and the environment. *Enzyme Immobilization: Advances in Industry, Agriculture, Medicine, and the Environment*, 1 - 132.
- Erdemir, S., Sahin, O., Uyanik, A., & Yilmaz, M. (2009). Effect of the glutaraldehyde derivatives of Calix[n]arene as cross-linker reagents on lipase immobilization. *Journal of Inclusion Phenomena and Macrocyclic Chemistry*, 64(3 - 4), 273 - 282.

- Eslamipour, F. & Hejazi, P. (2016). Evaluating effective factors on the activity and loading of immobilized  $\alpha$ -amylase onto magnetic nanoparticles using a response surface-desirability approach. *RSC Advances*. 6(24), 20187 - 20197.
- Fernandez-Lafuente, R., Armisén, P., Sabuquillo, P., Fernández-Lorente, G., & Guisán, J. M. (1998). Immobilization of lipases by selective adsorption on hydrophobic supports. *Chemistry and Physics of Lipids*, 93(1-2), 185 - 197.
- Fernandez-Lorente, G., Rocha-Martín, J., & Guisan, J. M. (2020). Immobilization of Lipases by Adsorption on Hydrophobic Supports: Modulation of Enzyme Properties in Biotransformations in Anhydrous Media. *Methods in Molecular Biology Immobilization of Enzymes and Cells*, 143 - 158.
- Filho, D. G., Silva, A. G., & Guidini, C. Z. (2019). Lipases: sources, immobilization methods, and industrial applications. *Applied Microbiology and Biotechnology*, 103(18), 7399 - 7423.
- Flood, M. T., & Kondo, M. (2003). Safety evaluation of lipase produced from *Rhizopus oryzae*: summary of toxicological data. *Regulatory Toxicology and Pharmacology*, 37(2), 293 - 304.
- Foresti, M., & Ferreira, M. (2004). Ethanol pretreatment effect and particle diameter issues on the adsorption of *Candida rugosa* lipase onto polypropylene powder. *Applied Surface Science*, 238(1 - 4), 86 - 90.
- Gangadharappa, B. S., Dammalli, M., Rajashekharappa, S., Pandurangappa, K. M. T., & Siddaiah, G. B. (2017). Reverse Micelles As a Bioseparation Tool For Enzymes. *Journal Of Proteins And Proteomics*, 8.
- Gao, S., Wang, Y., Wang, T., Luo, G., & Dai, Y. (2009). Immobilization of lipase on methyl-modified silica aerogels by physical adsorption. *Bioresource Technology*, 100(2), 996 - 999.
- Gashtasbi, F., Ahmadian, G., Noghabi, K. A. (2014). New insights into the effectiveness of alpha-amylase enzyme presentation on the *Bacillus subtilis* spore surface by adsorption and covalent immobilization. *Enzyme Microbial. Technology*, 64, 17 - 23.
- Gerday, C., Aittaleb, M., Bentahir, M., Chessa, J.-P., Claverie, P., Collins, T., Feller, G. (2000). Cold-adapted enzymes: from fundamentals to biotechnology. *Trends in Biotechnology*, 18(3), 103 - 107.
- Guo, Z., Bai, S., & Sun, Y. (2003). Preparation and characterization of immobilized lipase on magnetic hydrophobic microspheres. *Enzyme and Microbial Technology*, 32(7), 776 - 782.

- Haas, M. J., Scott, K., Jun, W., & Janssen, G. (1994). Enzymatic Phosphatidylcholine Hydrolysis in Organic Solvents: An Examination of Selected Commercially Available Lipases. *Journal of the American Oil Chemists Society*, 71, 483 - 490.
- Hasan, F., Shah, A. A., & Hameed, A. (2006). Industrial applications of microbial lipases. *Enzyme and Microbial Technology*, 39(2), 235 - 251.
- Hebbar, H. U., & Raghavarao, K. S. M. S. (2007). Extraction of bovine serum albumin using nanoparticulate reverse micelles. *Process Biochemistry*, 42(12), 1602 - 1608.
- Hemavathi, A., Hebbar, H. U., & Raghavarao, K. (2010). Mixed reverse micellar systems for extraction and purification of  $\beta$ -glucosidase. *Separation and Purification Technology*, 71(2), 263 - 268.
- Hemavathi, A., Hebbar, H., & Raghavarao, K. (2011). Reverse Micellar Extraction of Bioactive Compounds for Food Products. *Enhancing Extraction Processes in the Food Industry*, 399 - 436.
- Hirata, D. B., Albuquerque, T. L., Rueda, N., Sánchez-Montero, J. M., García-Verdugo, E., Porcar, R., & Fernandez-Lafuente, R. (2016). Advantages of Heterofunctional Octyl Supports: Production of 1,2-Dibutyryl by Specific and Selective Hydrolysis of Tributyrin Catalyzed by Immobilized Lipases. *ChemistrySelect*, 1(12), 3259 - 3270.
- Iqbal, J., & Afaq, S. (2001). Immobilization and stabilization of papain on chelating sepharose: a metal chelate regenerable carrier. *Electronic Journal of Biotechnology*, 4(3).
- Jaeger, K.-E., Dijkstra, B. W., & Reetz, M. T. (1999). Bacterial biocatalysts: molecular biology, three-dimensional structures, and biotechnological applications of lipases. *Annual Review of Microbiology*, 53, 315 - 351.
- Jalil, F. A., Rahman, R. R. A., Salleh, A., & Ali, M. M. (2018). Optimization and in Silico Analysis of a Cold-Adapted Lipase from an Antarctic *Pseudomonas* sp. Strain AMS8 Reaction in Triton X-100 Reverse Micelles. *Catalysts*, 8(7), 289.
- Jarudilokkul, S., Poppenborg, L. H., Valetti, F., Gilardi, G., & Stuckey, D. C. (1999). Separation and purification of periplasmic cytochrome c553 using reversed micelles. *Biotechnology Techniques*, 13(3), 159 - 163.
- Jegannathan, K. R., Abang, S., Poncelet, D., Chan, E. S., & Ravindra, P. (2008). Production of Biodiesel Using Immobilized Lipase - A Critical Review. *Critical Reviews in Biotechnology*, 28(4), 253 - 264.
- Jesionowski, T., Zdarta, J., Krajewska, B., Jakub, Z., & Krajewska, B. (2014). Enzyme immobilization by adsorption: a review. *Adsorption*, 20(5–6), 801 - 821.

- Jesudian, C., & Leroy, K. (1975). Lowry assay of dilute protein solutions containing high concentrations of Triton X - 100. *Analytical Biochemistry*, 69(2), 632 - 636.
- Johnson, L. A., & Lusas, E. W. (1983). Comparison of alternative solvents for oils extraction. *Journal of the American Oil Chemists Society*, 60(2Part1), 229 - 242.
- Jun, L. Y., Yon, L. S., Mubarak, N., Bing, C. H., Pan, S., Danquah, M. K., Khalid, M. (2019). An overview of immobilized enzyme technologies for dye and phenolic removal from wastewater. *Journal of Environmental Chemical Engineering*, 7(2), 102961.
- Kalantari, M., Yu, M., Yang, Y., Strounina, E., Gu, Z., Huang, X., Zhang, J., Song, H., Yu, C. (2017). Tailoring mesoporous-silica nanoparticles for robust immobilization of lipase and biocatalysis. *Nano Research*, 10, 605 - 617.
- Karra-Châabouni, M., Bouaziz, I., Boufi, S., Botelho do Rego, A. M., & Gargouri, Y. (2008). Physical immobilization of *Rhizopus oryzae* lipase onto cellulose substrate: Activity and stability studies. *Colloids and Surfaces B: Biointerfaces*, 66(2), 168 - 177.
- Katchalski-Katzir, E., & Kraemer, D. M. (2000). Eupergit® C, a carrier for immobilization of enzymes of industrial potential. *Journal of Molecular Catalysis B: Enzymatic*, 10(1-3), 157 - 176.
- Kharrat, N., Ali, Y. Ben, Marzouk, S., Gargouri, Y.-T. T., & Karra-Châabouni, M. (2011). Immobilization of *Rhizopus oryzae* lipase on silica aerogels by adsorption: Comparison with the free enzyme. *Process Biochemistry*, 46(5), 1083 - 1089.
- Konsoula Z. and M.L. Kyriakides. 2006. Starch hydrolsis by the action of an entrapped in alginate capsules  $\alpha$ -amylase from *Bacillus subtilis*. *Process Biochemistry*, 41, 343 - 349.
- Krei, G. A., & Hustedt, H. (1992). Extraction of enzymes by reverse micelles. *Chemical Engineering Science*, 47(1), 99 - 111.
- Krishna, S. H., Srinivas, N. D., Raghavarao, K. S. M. S., & Karanth, N. G. (2002). Reverse Micellar Extraction for Downstream Processing of Proteins/Enzymes. *History and Trends in Bioprocessing and Biotransformation*, 75, 119 - 183.
- Kumar, A., Dhar, K., Kanwar, S. S., & Arora, P. K. (2016). Lipase catalysis in organic solvents: advantages and applications. *Biological Procedures Online*, 1 - 11.

- Kumar, S., Katiyar, N., Ingle, P., & Negi, S. (2011). Use of evolutionary operation (EVOP) factorial design technique to develop a bioprocess using grease waste as a substrate for lipase production. *Bioresource Technology*, 102(7).
- Kwon, D. Y., & Rhee, J. S. (1986). A Simple and Rapid Colorimetric Method for Determination of Free Fatty Acids for Lipase Assay. *Journal of the American Oil Chemists Society*, 63(1).
- Lage, F.A.P., Bassi, J.J., Corradini, M.C.C., Todero, L.M., Luiz, J.H.H., Mendes, A.A. (2016). Preparation of a biocatalyst via physical adsorption of lipase from *Thermomyces lanuginosus* on hydrophobic support to catalyze biolubricant synthesis by esterification reaction in a solvent-free system. *Enzyme and Microbial Technology*, 84, 56 - 67.
- Leodidis, E. B., & Hatton, T. A. (1989). Specific Ion Effects in Electrical Double Layers: Selective Solubilization of Cations in Aerosol-OT Reversed Micelles. *Langmuir*, 5(3), 741 - 753.
- Li, C., Zhang, G., Liu, N., & Liu, L. (2016). Preparation and Properties of *Rhizopus oryzae* Lipase Immobilized Using an Adsorption-Crosslinking Method. *International Journal of Food Properties*, 19(8), 1776 - 1785.
- Liu, D. M., Chen, J., & Shi, Y. P. (2018). Advances on methods and easy separated support materials for enzymes immobilization. *Trends in Analytical Chemistry*, 102, 332 - 342.
- Liu, T., Liu, Y.; Wang, X., Li, Q., Wang, J., Yan, Y. (2011). Improving catalytic performance of *Burkholderia cepacia* lipase immobilized on macroporous resin NKA. *Journal of Molecular Catalysis B: Enzymatic*, 71, 45 - 50.
- Lowry, O. H., Rosebrough, N. J., Farr, A. L., & Randall, R. J. (1951). Protein measurement with the Folin phenol reagent. *The Journal of Biological Chemistry*, (193), 265 - 275.
- Malcata, F. X., Reyes, H. R., Garcia, H. S., Hill, C. G., & Amundson, C. H. (1990). Immobilized Lipase Reactors for Modification of Fats and Oils-A Review. *Journal of the American Oil Chemists Society*, 67 (12), 890 - 910.
- Manoel, E. A., Dos Santos, J. C. S., Freire, D. M. G., Rueda, N., & Fernandez-Lafuente, R. (2015). Immobilization of lipases on hydrophobic supports involves the open form of the enzyme. *Enzyme and Microbial Technology*, 71, 53 - 57.

- Mateo, C., Palomo, J. M., Fernandez-Lorente, G., Guisan, J. M., & Fernandez-Lafuente, R. (2007). Improvement of enzyme activity, stability and selectivity via immobilization techniques. *Enzyme and Microbial Technology*, 40(6), 1451 - 1463.
- Mathew, D. S., & Juang, R. S. (2005). Improved back extraction of papain from AOT reverse micelles using alcohols and a counter-ionic surfactant. *Biochemical Engineering Journal*, 25(3), 219 - 225.
- Matzke, S. F., Creagh, A. L., Haynes, C. A., Prausnitz, J. M., & Blanch, H. W. (1992). Mechanisms of protein solubilization in reverse micelles. *Biotechnology and Bioengineering*, 40(1), 91 - 102.
- Meireles, M. A. A. (2008). Extraction and Compounds Purification of Bioactive. *Extracting Bioactive Compounds for Food Products Contemporary Food Engineering*, 1 - 8.
- Melo, E. E., & Cabral, J. M. S. (2001). Reverse micelles and protein biotechnology. *Biotechnology Annual Review*, 7, 87 - 129.
- Meng, X., Xu, G., Zhou, Q.L., Wu, J.P., Yang, L.R. (2013). Improvements of lipase performance in high-viscosity system by immobilization onto a novel kind of poly(methylmethacrylate-co-divinylbenzene) encapsulated porous magnetic microspherecarrier. *Journal of Molecular Catalysis B: Enzymatic*, 89, 86 - 92.
- Metin, A. Ü. (2012). Immobilization studies and biochemical properties of free and immobilized *Candida rugosa* lipase onto hydrophobic group carrying polymeric support. *Macromolecular Research*, 21(2), 176 - 183.
- Meullemiestre, A., Breil, C., Abert-Vian, M., & Chemat, F. (2015). Innovative Techniques and Alternative Solvents for Extraction of Microbial Oils. *SpringerBriefs in Molecular Science Modern Techniques and Solvents for the Extraction of Microbial Oils*, 19 - 42.
- Mihailović, M., Stojanović, M., Banjanac, K., Carević, M., Prlainović, N., Milosavić, N., Bezbradica, D. (2014). Immobilization of lipase on epoxy-activated Purolite® A109e® its post-immobilization stabilization. *Process Biochemistry*, 49, 637 -646.
- Mohamad, N., Marzuki, N., Buang, N., Huyop, F., & Wahab, R. (2015). An overview of technologies for immobilization of enzymes and surface analysis techniques for immobilized enzymes. *Biotechnology, Biotechnological Equipment*, 29(2), 205- 220.

- Mohamed SA, Al-Harbi MH, Almulaiky YQ, Ibrahim IH, Salah HA, El-Badry MO, Abdel-Aty AM, Fahmy AS, El-Shishtawy RM. (2018). Immobilization of *Trichoderma harzianum*  $\alpha$ -amylase on PPyAgNp/Fe3O4-nanocomposite: chemical and physical properties. *Artificial Cells, Nanomedicine, Biotechnology*, 1 - 6.
- Mohtar NS, Abdul Rahman MB, Mustafa S, Mohamad Ali MS, Raja Abd Rahman RNZ. (2019). Spray-dried immobilized lipase from *Geobacillus* sp. strain ARM in sago. *PeerJ*.
- Montero, S., Blanco, A., Virto, M. D., Carlos Landeta, L., Agud, I., Solozabal, R., Serra, J. L. (1993). Immobilization of *Candida rugosa* lipase and some properties of the immobilized enzyme. *Enzyme and Microbial Technology*, 15(3), 239 - 247.
- Morhardt, C., Ketterer, B., Heißler, S., Franzreb, M. (2014). Direct quantification of immobilized enzymes by means of FTIR ATR spectroscopy – A process analytics tool for biotransformations applying non-porous magnetic enzyme carriers. *Journal of Molecular Catalysis. B, Enzymatic*, 107, 55 - 63.
- Mosafa, L., Shahedi, M., Moghadam, M. (2013). Magnetite Nanoparticles Immobilized Pectinase: Preparation, Characterization and Application for the Fruit Juices Clarification. *Journal of the Chinese Chemical Society*, 61 (3), 329 - 336.
- Nandini, K. E., & Rastogi, N. K. (2009). Reverse micellar extraction for downstream processing of lipase: Effect of various parameters on extraction. *Process Biochemistry*, 44(10), 1172 - 1178.
- Nandini, K. E., & Rastogi, N. K. (2010). Separation and purification of lipase using reverse micellar extraction: Optimization of conditions by response surface methodology. *Biotechnology and Bioprocess Engineering*, 15(2), 349 - 358.
- Nawani, N., Singh, R., & Kaur, J. (2006). Immobilization and stability studies of a lipase from thermophilic *Bacillus* sp: The effect of process parameters on immobilization of enzyme. *Electronic Journal of Biotechnology*, 9(October 2006), 559 - 565.
- Naya, M., & Imai, M. (2012). Impact of physicochemical character of hydrophobic porous carrier on reactivity of immobilized lipase progressing toward higher reaction rate and high yield in repeated use. *Procedia Engineering*, 42(42), 1004 - 1015.
- Pahujani, S., Kanwar, S. S., Chauhan, G., & Gupta, R. (2008). Glutaraldehyde activation of polymer Nylon-6 for lipase immobilization: Enzyme characteristics and stability. *Bioresource Technology*, 99(7), 2566 - 2570.

- Pawongrat, R., Xu, X., & H-Kittikun, A. (2007). Synthesis of monoacylglycerol rich in polyunsaturated fatty acids from tuna oil with immobilized lipase AK. *Food Chemistry*, 104(1), 251 - 258.
- Peirce, S., Tacias-Pascacio, V., Russo, M., Marzocchella, A., Virgen-Ortíz, J., & Fernandez-Lafuente, R. (2016). Stabilization of *Candida antarctica* Lipase B (CALB) Immobilized on Octyl Agarose by Treatment with Polyethyleneimine (PEI). *Molecules*, 21(6), 751.
- Pencreac'h, G., Leullier, M., & Baratti, J. C. (1997). Properties of Free and Immobilized Lipase From *Pseudomonas cepacia*. *Biotechnology and Bioengineering*, 56(2), 181 - 9.
- Pereira, S. E., Fernandes, K. F., & Ulhoa, C. J. (2016). Immobilization of *Cryptococcus flausa*-amylase on glass tubes and its application in starch hydrolysis. *Starch - Stärke*, 69(3 - 4), 1600189.
- Prakash O. and Jaiswal N. (2011). Immobilization of a Thermostable Amylase on Agarose and Agar Matrices and its Application in Starch Stain Removal. *World Applied Sciences Journal*, 13(3).
- Quintana, P. G., Canet, A., Marciello, M., Valero, F., Palomo, J. M., & Baldessari, A. (2015). Enzyme-catalyzed preparation of chenodeoxycholic esters by an immobilized heterologous *Rhizopus oryzae* lipase. *Journal of Molecular Catalysis B: Enzymatic*, 118, 36 - 42.
- Razib, M. S. M., Raja Noor Zaliha Raja Abd Rahman, Shariff, F. M., & Ali, M. S. M. (2020). Biochemical and Structural Characterization of Cross-Linked Enzyme Aggregates (CLEAs) of Organic Solvent Tolerant Protease. *Catalysts*, 10(1), 55.
- Rehman, S., Bhatti, H. N., Bilal, M., & Asgher, M. (2016). Cross-linked enzyme aggregates (CLEAs) of *Pencillium notatum* lipase enzyme with improved activity, stability and reusability characteristics. *International Journal of Biological Macromolecules*, 91, 1161 - 1169.
- Rehm, F., Chen, S., & Rehm, B. (2016). Enzyme Engineering for In Situ Immobilization. *Molecules*, 21(10), 1370.
- Reyes, H. R., & Hill, C. G. (1994). Kinetic modeling of interesterification reactions catalyzed by immobilized lipase. *Biotechnology and Bioengineering*, 43(2), 171 - 182.
- Robles-Medina, A., González-Moreno, P., Esteban-Cerdán, L., & Molina-Grima, E. (2009). Biocatalysis: Towards ever greener biodiesel production. *Biotechnology Advances*, 27(4), 398 - 408.

- Rodriguez, C. H., Chintanasathien, C., Scamehorn, J. F., Saiwan, C., & Chavadej, S. (1998). Precipitation in solutions containing mixtures of synthetic anionic surfactant and soap. I. effect of sodium octanoate on hardness tolerance of sodium dodecyl sulfate. *Journal of Surfactants and Detergents*, 1(3), 321 - 328.
- Roig, M. G., Estevez, F. B., Velasco, F. G., Ghais, I., & Silverio, J. M. C. (1986). Methods for immobilizing enzymes. *Biotechnology and Applied Biology Section*, 14(4), 180 - 185.
- Sagiroglu, A., & Telefoncu, A. (2004). Immobilization of lipases on different carriers and their use in synthesis of pentyl isovalerates. *Preparative Biochemistry and Biotechnology*, 34(2), 169 - 178.
- Salah, R. B., Mosbah, H., Fendri, A., Gargouri, A., Gargouri, Y., & Mejdoub, H. (2006). Biochemical and molecular characterization of a lipase produced by *Rhizopus oryzae*. *FEMS Microbiology Letters*, 260(2), 241 - 248.
- Sankar, K., & Achary, A. (2019). Bio-ceramic, mesoporous cuttlebone of *Sepia officinalis* is an ideal support for the immobilization of *Bacillus subtilis* AKL13 lipase: Optimization, adsorption, thermodynamic and reaction kinetic studies. *Chemicals Papers*, 74, 459 - 470.
- Séverac, E., Galy, O., Turon, F., Pantel, C.A., Condoret, J.S., Monsan, P., Marty, A. (2011). Selection of *Ca/B* immobilization method to be used in continuous oil transesterification: Analysis of the economical impact. *Enzyme and Microbial Technology*, 48, 61-70.
- Silveira, E.A., Moreno-Perez, S., Basso, A., Serban, S., Pestana-Mamede, R., Tardioli, P.W., Sanchez-Farinias, C., Castejon, N., Fernandez-Lorente, G., Rocha-Martin, J. (2019). Biocatalyst engineering of *Thermomyces lanuginosus* lipase adsorbed on hydrophobic supports: Modulation of enzyme properties for ethanolysis of oil in solvent-free systems. *Journal of Biotechnology*, 289, 126 - 134.
- Singh, K., Bharose, R., Singh, V. K., & Verma, S. K. (2011). Sugar Decolorization through Selective Adsorption onto Functionalized Accurel Hydrophobic Polymeric Support. *Industrial & Engineering Chemistry Research*, 50(17), 10074 - 10082.
- Soumanou, M. M., Edorh, A. P., & Bornscheuer, U. T. (2004). Activity and stability of immobilized lipases in lipase-catalyzed modification of peanut oil. *Oilseeds & fats Crops and Lipids*, 11 (6), 464 - 468.
- Tacias-Pascacio, V. G., Peirce, S., Torrestiana-Sanchez, B., Yates, M., Rosales-Quintero, A., Virgen-Ortíz, J. J., & Fernandez-Lafuente, R. (2016). Evaluation of different commercial hydrophobic supports for the immobilization of lipases: tuning their stability, activity and specificity. *RSC Advances*, 6(102), 100281 - 100294.

- Talekar S., V. Ghodake, A. Kate, N. Samant, C. Kumar and S. Gadagkar. (2010). Preparation and characterization of cross-linked enzyme aggregates of *Saccharomyces cerevisiae* invertase. *Australian Journal of Basic and Applied Sciences*, 4(10), 4760 - 4765.
- Talekar, S., & Chavare, S. (2012). Optimization of immobilization of  $\alpha$ -amylase in alginate gel and its comparative biochemical studies with free  $\alpha$ -amylase, *Recent Research in Science and Technology*, 4(2), 01 - 05.
- Tecelão, C., Guillén, M., Valero, F., & Ferreira-Dias, S. (2012). Immobilized heterologous *Rhizopus oryzae* lipase: A feasible biocatalyst for the production of human milk fat substitutes. *Biochemical Engineering Journal*, 67, 104 - 110.
- Valivety, R. H., Halling, P. J., Peilow, A. D., & Macrae, A. R. (1994). Relationship between water activity and catalytic activity of lipases in organic media: Effects of supports, loading and enzyme preparation. *European Journal of Biochemistry*, 222(2), 461 - 466.
- Vasudevan, M., & Wiencek, J. M. (1996). Mechanism of the Extraction of Proteins into Tween 85 Nonionic Microemulsions. *Industrial and Engineering Chemistry Research*, 35(4), 1085 - 1089.
- Villeneuve, P., Muderhwa, J. M., Graille, J., & Haas, M. J. (2000). Customizing lipases for biocatalysis: a survey of chemical, physical and molecular biological approaches. *Journal of Molecular Catalysis B: Enzymatic*, 9(4 - 6), 113 - 148.
- Virto, M. D., Agud, I., Montero, S., Blanco, A., Solozabal, R., Lascaray, J. de Renobales, M. (1994). Hydrolysis of animal fats by immobilized *Candida rugosa* lipase. *Enzyme and Microbial Technology*, 16(1), 61 - 65.
- Wolbert, R. B. G., Hilhorst, R., Voskuilen, G., Nachtegaal, H., Dekker, M., Riet, K. V., & Bijsterbosch, B. H. (1989). Protein transfer from an aqueous phase into reversed micelles. The effect of protein size and charge distribution. *European Journal of Biochemistry*, 184(3), 627 - 633.
- Wu, J. C., Selvam, V., Teo, H. H., Chow, Y., Talukder, M. M. R., & Choi, W. J. (2006). Immobilization of *Candida rugosa* lipase by cross-linking with glutaraldehyde followed by entrapment in alginate beads. *Biocatalysis and Biotransformation*, 24(5), 352 - 357.
- Xiao, J., Cai, J., & Guo, X. (2013). Reverse micellar extraction of bovine serum albumin – A comparison between the effects of gemini surfactant and its corresponding monomeric surfactant. *Food Chemistry*, 136(2), 1063 - 1069.

- Xu, L., Ke, C., Huang, Y., & Yan, Y. (2016). Immobilized *Aspergillus niger* Lipase with SiO<sub>2</sub> Nanoparticles in Sol-Gel Materials. *Catalysts*, 6(10), 149.
- Yang, D., & Rhee, J. S. (1992). Continuous hydrolysis of olive oil by immobilized lipase in organic solvent. *Biotechnology and Bioengineering*, 40(6), 748 - 752.
- Yu, Y. C., Chu, Y., & Ji, J. Y. (2003). Study of the factors affecting the forward and back extraction of yeast-lipase and its activity by reverse micelles. *Journal of Colloid and Interface Science*, 267(1), 60 - 64.
- Zhang, B., Weng, Y., Xu, H., & Mao, Z. (2012). Enzyme immobilization for biodiesel production. *Applied Microbiology and Biotechnology*, 93(1), 61 - 70.
- Zhang, Yuwen, & Peng. (2013). Parameters affecting the performance of immobilized enzyme, *Journal of Chemistry*, 2013, 1 - 7.
- Zhao, K., Di, Q., Cao, X., Wang, M., Deng, L., & Wang, F. (2016). Production of Biodiesel Using Immobilized Lipase and the Characterization of Different Co-Immobilizing Agents and Immobilization Methods. *Sustainability*, 8(9), 764.
- Zhao, X., Wei, Z., Du, F., & Zhu, J. (2010). Effects of Surfactant and Salt Species in Reverse Micellar Forward Extraction Efficiency of Isoflavones with Enriched Protein from Soy Flour. *Applied Biochemistry and Biotechnology*, 162(7), 2087 - 2097.