



**UNIVERSITI PUTRA MALAYSIA**

***ASSESSMENT OF MORPHOMETRIC VARIATION, GROWTH  
PERFORMANCE, DISEASE RESISTANCE AND NUTRITIONAL  
VALUE OF CLARIID CATFISH HYBRID (*Clarias macrocephalus*  
GUNTHER, 1864 x *Clarias gariepinus* BURCHELL, 1822)***

**NORA FATEN AFIFAH BINTI MOHAMAD**

**FPV 2021 18**



**ASSESSMENT OF MORPHOMETRIC VARIATION, GROWTH  
PERFORMANCE, DISEASE RESISTANCE AND NUTRITIONAL VALUE OF  
CLARIID CATFISH HYBRID (*Clarias macrocephalus* GUNTHER, 1864 x  
*Clarias gariepinus* BURCHELL, 1822)**

By

**NORA FATEN AFIFAH BINTI MOHAMAD**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra  
Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of  
Philosophy**

**November 2020**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in  
fulfilment of the requirement for the degree of Doctor of Philosophy

**ASSESSMENT OF MORPHOMETRIC VARIATION, GROWTH  
PERFORMANCE, DISEASE RESISTANCE AND NUTRITIONAL VALUE OF  
CLARIID CATFISH HYBRID (*Clarias macrocephalus* GUNTHER, 1864 x  
*Clarias gariepinus* BURCHELL, 1822)**

By

**NORA FATEN AFIFAH BINTI MOHAMAD**

**November 2020**

**Chair : Assoc. Prof. Hassan Bin Haji Mohd Daud, PhD**  
**Faculty : Veterinary Medicine**

The present study was carried out to access the morphometric variation and performance of crossbreed of *Clarias macrocephalus* (♀) and *Clarias gariepinus* (♂), (hybrid CMxCG) and its parental species; female, CM (*C. macrocephalus*♀ x *C. macrocephalus* ♂); male, CG (*C. gariepinus*♀ x *C. gariepinus*♂). The first objective was conducted to evaluate the effect of hybridization on the breeding performance, growth and survival of hybrid CMxCG and its parental species as a control for a period of 60 days, with fortnightly sampling. For the purposes, broodstock fishes were induced to spawn by intramuscular injection of Ovaprim hormone at the dose of 0.5 mL kg<sup>-1</sup> body weight (female) and 0.25 mL kg<sup>-1</sup> body weight (male). The result revealed that hybrid CMxCG achieved better in breeding performance than *C. macrocephalus* in regard to fertilization and hatching rate as well as growth and survival. Percentage of fertilization and hatching rate of hybrid CMxCG were 72.62±2.51% and 64.01±9.17%, respectively, which were higher than in *C. macrocephalus* which recorded 60.76±0.64 % and 28.14±6.51 %, respectively. As for growth performance, the body weight and total length of hybrid CMxCG showed significantly larger and longer ( $p<0.05$ ) than *C. macrocephalus* at the end of the study period which resulted in 1.54±0.48 g and 5.81±1.09 cm, respectively for hybrid CMxCG and 0.98±0.28 g and 4.77±0.47 cm for *C. macrocephalus*. Higher survival rate of 58% which significantly different ( $p<0.05$ ) was noticed in hybrid CMxCG followed by *C. macrocephalus* (49%) and *C. gariepinus* (41%).

The second objective of this study employed the morphological variation of hybrid CMxCG and its parental species. A total of 30 morphometric measurements and 5 meristic counts were carried out on each specimen using method from Teugels (1986) and Agnese et al. (1997) with slight modification. For the morphometric characters in this present study, eight new measurements were added with one new measurement in the head and seven new measurement in toothplate. The results emphasized that the shape of the occipital process and the shape of the premaxillary as well as vomerine toothplate appears to be highly reliable index of morphological discrimination because it's showed high phenotypic differentiation among all three fish species. Most of the hybrids also showed intermediate morphological characteristics and exhibits more similarities of phenotypic featured to their paternal species after juvenile stage which might be due to paternal dominating characteristics.

The third objective of this study was to evaluate the nutritional composition of hybrid CMxCG, *C. gariepinus* and *C. macrocephalus*. Analyses on proximate composition revealed that hybrid CMxCG had the highest protein content ( $40.07 \pm 0.09\%$ ) than both parental species (CM:  $39.84 \pm 0.07\%$ ; CG:  $38.54 \pm 0.05\%$ ). Fatty acid analysis was also done in triplicate consisted of two consecutive steps which were preparations of fatty acid methyl ester (FAME) and gas liquid chromatography. In this study, fatty acids varied between species, demonstrating high polyunsaturated fatty acids (PUFA), EPA, DHA in hybrid CMxCG with the value of  $31.41 \pm 0.94\%$ ,  $2.02 \pm 0.09$  and  $2.42 \pm 0.40\%$ , respectively. However, no significant difference ( $p > 0.05$ ) was observed in total amino acids (EAA) content of hybrid CMxCG.

Apart from that, this study also was conducted to assess the resistance of hybrid CMxCG and its parental species towards bacteria, *Aeromonas hydrophila*. The PCR primers used for specific detection of *A. hydrophila* was tested successfully and a desired PCR product of 685 bp was obtained. For aerolysin gene detection, bacteria isolates produced a 309 bp amplicon as expected. In the next phase of the study, fishes were intraperitoneally injected (0.1 mL) with different *A. hydrophila* concentrations;  $T_0$ = control (0.85% saline solution); and five serial dilutions ( $10^{-9}$ ,  $10^{-8}$ ,  $10^{-7}$ ,  $10^{-6}$ , and  $10^{-5}$ ) of *A. hydrophila* into juveniles of hybrid CMxCG, *C. gariepinus* and *C. macrocephalus*. The last objective of this present study revealed that 96 h-LD<sub>50</sub> calculated using the Reed and Muench (1938) method was higher in hybrid CMxCG ( $\times 10^{6.09}$  cfu mL<sup>-1</sup>) than *C. macrocephalus* ( $\times 10^{5.36}$  cfu mL<sup>-1</sup>) but lower than *C. gariepinus* ( $\times 10^{6.33}$  cfu mL<sup>-1</sup>). Intraperitoneal injection resulted in 100% mortality at the highest concentration of bacteria ( $10^9$  cfu mL<sup>-1</sup>). Lesions scores were observed to be more severe in *C. macrocephalus* particularly in the kidney tubules with significantly higher ( $p < 0.05$ ) mean lesion score as compared to *C. gariepinus* and hybrid CMxCG.

In conclusion, the present study has provided important information on morphometric variation which can be used as quick and cheap method in identifying and distinguishing the parental species and the hybrids especially for field application. Hybrid CMxCG also have showed good performance of important traits which is beneficial for future aquaculture industry in regard to growth and disease resistance. Moreover, hybrid CMxCG revealed better performance of flesh quality content such as protein, PUFA, EPA and DHA, hence, aquaculturing of hybrid can contribute to a better source of protein diet for human being.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PENILAIAN VARIASI MORFOMETRIK, PRESTASI PERTUMBUHAN,  
RINTANGAN PENYAKIT DAN NILAI PEMAKANAN IKAN KELI CLARIID  
HIBRID (*Clarias macrocephalus* GUNTHER, 1864 x *Clarias gariepinus*  
BURCHELL, 1822)**

Oleh

**NORA FATEN AFIFAH BINTI MOHAMAD**

**November 2020**

**Pengerusi : Prof. Madya Hassan Bin Haji Mohd Daud, PhD**  
**Fakulti : Perubatan Veterinar**

Kajian ini dijalankan untuk menilai variasi morfometrik dan prestasi kacukan *Clarias macrocephalus* (♀) dan *Clarias gariepinus* (♂), (hibrid CMxCG) dan spesies induk; betina, CM (*C. macrocephalus*♀ x *C. macrocephalus* ♂); jantan, CG (*C. gariepinus*♀ x *C. gariepinus*♂). Objektif pertama dijalankan untuk menilai kesan hibridisasi terhadap prestasi pembiakan, pertumbuhan dan kelangsungan hidup hibrid CMxCG dan spesies induk sebagai kawalan untuk tempoh 60 hari, dengan persampelan dua minggu. Untuk tujuan itu, induk ikan telah dirangsang melalui suntikan intramuskular dengan menggunakan hormon Ovaprim pada dos 0.5 mL kg<sup>-1</sup> berat badan (betina) dan 0.25 mL kg<sup>-1</sup> berat badan (jantan). Hasil menunjukkan bahawa hibrid CMxCG mencapai prestasi yang lebih baik dalam pembiakan berbanding *C. macrocephalus* berkaitan dengan persenyawaan dan kadar penetasan serta pertumbuhan dan kelangsungan hidup. Peratus persenyawaan dan penetasan kadar hibrid CMXCG adalah masing-masing 72.62 ± 2.51% dan 64.01 ± 9.17%, yang mana lebih tinggi daripada *C. macrocephalus* yang mencatatkan 60.76 ± 0.64% dan 28.14 ± 6.51%. Bagi prestasi pertumbuhan, berat badan dan jumlah keseluruhan hibrid CMXCG menunjukkan secara signifikan lebih besar dan lebih lama ( $p < 0.05$ ) daripada *C. macrocephalus* pada akhir tempoh kajian yang menghasilkan 1.54 ± 0.48 g dan 5.81 ± 1.09 cm, masing-masing untuk hibrid CMXCG dan 0.98 ± 0.28 g dan 4.77 ± 0.47 cm untuk *C. macrocephalus*. Kadar kelangsungan hidup yang lebih tinggi sebanyak 58% yang mana kadar perbezaan adalah ketara ( $p < 0.05$ ) telah dilihat di hibrid CMxCG diikuti oleh *C. macrocephalus* (49%) dan *C. gariepinus* (41%).

Objektif kedua kajian ini menggunakan variasi morfologi hibrid CMxCG dan spesies induk. Sebanyak 29 pengukuran morfometrik dan 5 kiraan meristik dilakukan pada setiap spesimen menggunakan kaedah dari Teugels (1986) dan Agnese et al. (1997) dengan sedikit pengubahsuaian. Untuk watak morfometrik dalam kajian ini, lapan pengukuran baru ditambah dengan satu pengukuran baru di kepala dan tujuh pengukuran baru dalam plat gigi. Hasilnya menekankan bahawa bentuk proses occipital dan bentuk premaxillary serta vomerine gigi-palatal ternyata menjadi indeks diskriminasi morfologi yang sangat boleh dipercayai kerana menunjukkan perbezaan fenotipik yang tinggi di antara ketiga spesies ikan ini. Kebanyakan hibrid juga menunjukkan ciri-ciri morfologi perantaraan dan mempamerkan lebih banyak persamaan fenotip yang diketengahkan kepada spesies induk jantan mereka selepas peringkat juvenil yang mungkin disebabkan oleh ciri-ciri keturunan induk jantan.

Objektif ketiga kajian ini adalah untuk menilai komposisi nutrisi hibrid CMxCG, *C. gariepinus* dan *C. macrocephalus*. Analisis komposisi menunjukkan bahawa CMxCG hibrid mempunyai kandungan protein tertinggi ( $40.07 \pm 0.09\%$ ) daripada kedua-dua spesies induk (CM:  $39.84 \pm 0.07\%$ ; CG:  $38.54 \pm 0.05\%$ ). Analisis asid lemak bervariasi antara spesies juga dilakukan sebanyak tiga kali ulangan terdiri daripada dua langkah berturut-turut iaitu penyediaan metil ester asid lemak (FAME) dan kromatografi cecair gas. Dalam kajian ini, asid lemak berbeza antara spesies, menunjukkan asid lemak tidak tepu (PUFA), EPA, DHA dalam hibrid CMxCG dengan nilai masing-masing  $31.41 \pm 0.94\%$ ,  $2.02 \pm 0.09$  dan  $2.42 \pm 0.40\%$ . Walau bagaimanapun, tidak perbezaan yang signifikan ( $p > 0.05$ ) diperhatikan dalam jumlah kandungan asid amino (EAA) CMxCG hibrid.

Selain itu, kajian ini juga dilakukan terhadap ketahanan hibrid CMxCG dan spesies induknya terhadap bakteria, *Aeromonas hydrophila*. Primer PCR yang digunakan untuk pengesanan spesifik *A. hydrophila* diuji dengan jayanya dan produk PCR yang diinginkan sebanyak 685 bp diperoleh. Untuk pengesanan gen aerolysin, isolat bakteria menghasilkan amplicon 309 bp seperti yang diharapkan. Dalam fasa kajian seterusnya, ikan disuntik secara intraperitoneal (0.1 mL) dengan penggunaan kepekatan *A. hydrophila* yang berbeza;  $T_0$  = kawalan (larutan garam 0.85%); dan lima pencairan bersiri ( $10^{-9}$ ,  $10^{-8}$ ,  $10^{-7}$ ,  $10^{-6}$ , dan  $10^{-5}$ ) *A. hydrophila* terhadap hibrid CMxCG, *C. gariepinus* dan *C. macrocephalus*. Objektif terakhir dari kajian ini menunjukkan 96j LD<sub>50</sub> yang dikira dengan kaedah Reed dan Muench (1938) lebih tinggi pada hibrid CMxCG ( $\times 10^{6.09}$  cfu mL<sup>-1</sup>) daripada *C. macrocephalus* ( $\times 10^{5.36}$  cfu mL<sup>-1</sup>) tetapi lebih rendah daripada *C. gariepinus* ( $\times 10^{6.33}$  cfu mL<sup>-1</sup>). Kaedah suntikan intraperitoneally mengakibatkan kematian 100% pada kepekatan bakteria tinggi ( $10^9$  cfu mL<sup>-1</sup>). Luka juga dilihat lebih teruk pada *C. macrocephalus* dengan skor



luka yang lebih tinggi ( $p < 0.05$ ) berbanding dengan *C. gariepinus* dan CMxCG hibrid.

Kesimpulannya, kajian ini telah memberikan maklumat penting tentang variasi morfometrik yang boleh digunakan sebagai kaedah yang cepat dan murah dalam mengenalpasti dan membezakan spesies induk dan hibrid mereka terutama untuk aplikasi di ladang. Hibrid CMxCG juga telah menunjukkan prestasi yang baik terhadap ciri-ciri penting yang bermanfaat untuk industri akuakultur di masa depan mengenai pertumbuhan dan rintangan penyakit. Selain itu, hibrid CMxCG menunjukkan prestasi yang lebih baik terhadap kandungan kualiti daging seperti protein, PUFA, EPA dan DHA, oleh itu, akuakultur hibrid boleh menyumbang kepada sumber makanan protein yang lebih baik untuk manusia.



## ACKNOWLEDGEMENTS

In the name of ALLAH, the Most Gracious, the Most Merciful.

First, and most of all, Alhamdulillah, praises and thanks to Allah S.W.T for the wisdom He bestowed upon me, the strength, peace of my mind and good health in order to finish this research.

Immeasurable appreciation and deepest gratitude for the help and kind support are also extended to the following persons who made significant impact on my doctoral journey and accomplishment:

- To my dearest supervisor, Assc. Prof. Dr. Hassan Bin Haji Mohd Daud, for his endless support, advice, guidance, valuable comments and suggestions during my research. It was a great privilege and honor to work and study under his guidance.
- To my co-supervisors, Dr. Annie Christianus and Asst. Prof. Dr. Maheran Yaman for their consultations and concerns on my research work and thesis preparation.
- To the Faculty of Veterinary Medicine, Universiti Putra Malaysia for giving me the opportunity to do research and letting me fulfill my dream of being a student here.
- To my fellow labmates especially Ruhil, Raina, Fairus, Nadia, Sofia, Daya, Kumari and Shida for your friendship, empathy and never ending motivations I've been getting all this while. They were always beside me during the happy and hard moments and have made my time at the Universiti Putra Malaysia more positive and enjoyable.
- To my beloved parents who have always been there for me no matter where I am. It was their unconditional love, care, tolerance, and valuable prayers which made the hardship of writing the thesis worthwhile. Without their supports, I do not think that I could overcome the difficulties during these years. I salute all the pain and sacrifice they did to shape my life. I would never be able to pay back the love and affection showered upon by my parents.
- To my brother, sisters, brother in laws, sister in law for their love, understanding and continuing supports, either morally, financially and physically to complete this research work.
- To a very special person, my husband, for his unfailing love, constant support and understanding during my pursuit of Ph.D degree that made the completion of thesis possible. You were always around at times I thought that it is impossible to continue, you helped me to keep things in perspective. I deeply appreciate his belief in me.
- To my little girl, for abiding my ignorance and the patience she showed during my thesis writing.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

**Hassan Bin Haji Mohd Daud, PhD**

Associate Professor  
Faculty of Veterinary Medicine  
Universiti Putra Malaysia  
(Chairman)

**Annie Christianus, PhD**

Senior Lecturer  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Member)

**Maheran Yaman, PhD**

Associate Professor  
Kullliyyah of Architecture and Environmental Design  
International Islamic University Malaysia  
(Member)

**ZALILAH MOHD SHARIFF, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 11 November 2021

## Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- There is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No.: Nora Faten Afifah Binti Mohamad, GS39396

## TABLE OF CONTENTS

<b>ABSTRACT</b>	<b>Page</b>
<b>ABSTRAK</b>	i
<b>ACKNOWLEDGEMENTS</b>	iv
<b>APPROVAL</b>	vii
<b>DECLARATION</b>	viii
<b>LIST OF TABLES</b>	x
<b>LIST OF FIGURES</b>	xvii
<b>LIST OF ABBREVIATIONS</b>	xix
	xxii

### CHAPTER

<b>1</b>	<b>INTRODUCTION</b>	<b>1</b>
1.1	World Fisheries and Aquaculture	1
1.2	Statement of Problems and Significance of Study	3
1.3	Research Hypothesis	5
1.4	Objectives of Study	5
<b>2</b>	<b>LITERATURE REVIEW</b>	<b>6</b>
2.1	Freshwater aquaculture in Malaysia	6
2.2	Family of Clariidae	7
2.3	Catfish of <i>Clarias</i> species	8
2.3.1	<i>Clarias gariepinus</i> (Burchell, 1822)	9
2.3.2	Taxonomy	9
2.3.3	Morphological identification, Biology and Geographical Distributions	9
2.4	<i>Clarias macrocephalus</i> (Gunther, 1864)	10
2.4.1	Taxonomy	11
2.4.2	Morphological identification, Biology and Geographical Distributions	11
2.5	Hybridization	12
2.5.1	Intra-specific Hybridization	12
2.5.2	Inter-specific Hybridization	13
2.5.3	Hybridization in Catfishes	13
2.6	Morphology Differentiation among Fish Species	16
2.7	Biochemical Compositions	18
2.7.1	Proximate Compositions	19
2.7.2	Fatty Acids	20
2.7.3	Amino Acids	21
		22

2.8	Bacterial Disease Associated with Fish Culture	23
2.8.1	<i>Aeromonas hydrophila</i>	23
2.8.2	<i>A. hydrophila</i> infections	
<b>3</b>	<b>EVALUATION OF BREEDING PERFORMANCE, GROWTH AND SURVIVAL OF HYBRID CMXCG AND ITS PARENTAL SPECIES</b>	<b>25</b>
3.1	Introduction	25
3.2	Materials and Methods	26
3.2.1	Breeding Program	26
3.2.2	Broodstock Procurement and Management	26
3.2.3	Selection of Broodstock	27
3.2.4	Hormone Treatment and Artificial Fertilization	27
3.2.5	Embryonic Chronology	28
3.2.6	Larvae Rearing Conditions	29
3.2.7	Breeding Performance, Growth and Survival	29
3.2.8	Data Analysis	30
3.3	Results	30
3.3.1	Fertilization and Hatchability	30
3.3.2	Embryogenesis	31
3.3.3	Growth and Survival	33
3.3.4	Heterosis	35
3.4	Discussion	35
3.5	Conclusion	39
<b>4</b>	<b>MORPHOLOGICAL VARIATION OF HYBRID CMxCG AND ITS PARENTAL SPECIES</b>	<b>40</b>
4.1	Introduction	40
4.2	Materials and Methods	41
4.2.1	Study Site and Sample Collection	41
4.2.2	Data Collections and Fish Identifications	41
4.2.3	Morphological Measurements	42
4.2.3.1	Morphometric Characteristic	42
4.2.3.2	Meristic Characteristic	43
4.2.4	Morphometric and Meristic Analysis	43
4.2.5	Statistical Analysis	43
4.3	Results	45
4.3.1	Morphometric Analysis	45
4.3.2	Meristic Analysis	49
4.3.3	Correlation Between Morphometric Variables	49
4.3.4	Principal Component Analysis	50

	(PCA)	
4.4	Discussion	60
4.5	Conclusion	64



<b>5</b>	<b>COMPARISON OF PROXIMATE COMPOSITION, FATTY ACIDS AND AMINO ACIDS PROFILE OF HYBRID CMxCG AND ITS PARENTAL SPECIES</b>	<b>65</b>
5.1	Introduction	65
5.2	Materials and Methods	66
5.2.1	Sample collection	66
5.2.2	Analysis of Proximate Compositions, Fatty Acids and Amino Acids Profiles	66
5.2.2.1	Fish Sample Collection, Storage and Preparation	66
5.2.2.2	Proximate Composition of Moisture Content, Ash, Crude Protein and Crude Lipid	66
5.2.3	Moisture Content Determination	66
5.2.4	Ash Content Determination	67
5.2.5	Crude Protein Determination	67
5.2.6	Crude Fat Determination	68
5.2.7	Dry Matter and Ash Determination	69
5.2.8	Fatty Acid Profile Determination	70
5.2.9	Extraction of Total Lipids	70
5.2.10	Preparation of Fatty Acid Methyl Ester (FAME)	71
5.2.10.1	Gas Liquid Chromatography	71
5.2.10.2	Amino Acid Analysis	71
5.2.11	Statistical Analysis	72
5.3	Results	72
5.3.1	Proximate Compositions	72
5.3.2	Fatty Acid Profiles	73
5.3.3	Amino Acid Profiles	76
5.4	Discussion	78
5.5	Conclusion	84
<b>6</b>	<b>RESISTANT OF THE HYBRID BETWEEN <i>Clarias macrocephalus</i> AND <i>Clarias gariepinus</i> TO <i>Aeromonas hydrophila</i></b>	<b>85</b>
6.1	Introduction	85
6.2	Materials and Methods	86
6.2.1	Experimental Design	86
6.2.2	Bacteria	86
6.2.2.1	Bacteria Isolation	86
6.2.2.2	Bacteria DNA Extraction and Purification	87
6.2.2.3	Quantification of DNA Samples	88
6.2.2.4	16S rDNA PCR Amplification	88



6.2.2.5	Standard Concentration Curve of Bacteria	90
6.2.3	Lethal Median Pathogenicity Test, (LD <sub>50</sub> at 96h) of the isolated <i>A. hydrophila</i>	90
6.2.4	Histopathological Examination	91
6.2.5	Statistical Analysis	91
6.3	Result	92
6.3.1	Bacteriological Observation and Morphological Characteristics of Isolates	92
6.3.2	Biochemical Characterization	93
6.3.3	Analysis of 16S rDNA PCR	96
6.3.4	Fish Mortality and Pathogenicity Test, 96 h-LD <sub>50</sub>	97
6.3.5	Clinical Signs and Gross Pathology of Experimentally Infected Fish	98
6.3.6	Histopathology	101
6.4	Discussion	109
6.5	Conclusion	112
7	<b>SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH</b>	113
7.1	Summary	113
7.2	General Conclusion	114
7.3	Recommendation	115
	<b>REFERENCES</b>	116
	<b>APPENDICES</b>	144
	<b>BIODATA OF STUDENT</b>	148
	<b>LIST OF PUBLICATIONS</b>	149

## LIST OF TABLES

Table		Page
2.1	The most cultured freshwater fish species in Malaysia	7
2.2	Summarization of hybrid fishes that commonly used in global aquaculture	15
3.1	Brood fish used in the mating process	28
3.2	Percentage of fertilization and hatching in pure parental lines of <i>C. gariepinus</i> (♀CG x ♂CG) and <i>C. macrocephalus</i> (♀CM x ♂CM) and their hybrids (♀CM x ♂CG).	31
4.1	Body morphometric measurements (mean ± SD) of <i>C. macrocephalus</i> (CM), <i>C. gariepinus</i> (CG) and CMxCG	47
4.2	Head morphometric measurements (mean ± SD) of <i>C. macrocephalus</i> , <i>C. gariepinus</i> and hybrid CMxCG	48
4.3	Range of meristic parameters and (mean ± SD) of <i>C. macrocephalus</i> (CM), <i>C. gariepinus</i> (CG) and hybrid (CMxCG)	49
4.4	PCA loadings of transformed meristic variables of two parental species and their hybrids ( <i>C. macrocephalus</i> , <i>C. gariepinus</i> and hybrid	51
4.5	PCA loadings of transformed morphometric variables of two parental species and their hybrids ( <i>C. macrocephalus</i> , <i>C. gariepinus</i> and hybrid CMxCG)	52
4.6	Pearson's correlation coefficients between the twenty-nine morphometric and parameters of cultured <i>C. gariepinus</i> .	53
4.7	Pearson's correlation coefficients between the twenty nine morphometric parameters of cultured <i>C. gariepinus</i> .	54
5.1	Proximate composition (Mean±SD) of <i>Clarias macrocephalus</i> , <i>Clarias gariepinus</i> , and hybrids CMxCG	73

5.2	Fatty acid profiles of <i>Clarias macrocephalus</i> , <i>Clarias gariepinus</i> , and hybrids CMxCG	75
5.3	Amino acid profiles of <i>Clarias macrocephalus</i> , <i>Clarias gariepinus</i> , and hybrids CMxCG	77
6.1	Oligonucleotide primers used for detection specific 16S rDNA of <i>Aeromonas hydrophila</i> and Aerolysin gene detection	89
6.2	List of the freshly collected bacterial isolates from <i>C. gariepinus</i>	94
6.3	Morphological, physiological and biochemical characteristics of <i>Aeromonas hydrophila</i>	95
6.4	Lesion scores (Mean±SD) in the kidneys of hybrid CMxCG, <i>Clarias macrocephalus</i> and <i>Clarias gariepinus</i>	102
6.5	Lesion scores (Mean±SD) in the livers of hybrid CMxCG, <i>Clarias macrocephalus</i> and <i>Clarias gariepinus</i>	103
6.6	Lesion scores (Mean±SD) in the spleens of hybrid CMxCG, <i>Clarias macrocephalus</i> and <i>Clarias gariepinus</i>	104

## LIST OF FIGURES

Figure		Page
2.1	Pie chart showed the production of major aquaculture species in Malaysia	8
2.2	<i>Clarias gariepinus</i>	9
2.3	<i>Clarias macrocephalus</i>	10
3.1	Typical embryological development of hybrid crosses of <i>Clarias macrocephalus</i> and <i>Clarias gariepinus</i> : (A) fertilized egg; (B) two-cell stage at first cleavage; (C) four-cell stage; (D) eight-cell stage; (E) 16-cell stage; (F) 32-cell stage; (G); (H) 64-cell stage; (I) Early morula; (J) Blastula stage; (K) Early gastrula stage; (L) Late gastrula; (M) Blastophore; (N) Early embryo formation; (O) 5-myomere; (P) 9-somite cell; (Q) 15-somite cell; (R) 19-somite cell; (S) 21-somite cell; (T) 26-somite cell; (U) Prime (start moving); (V) Hatchling. Scale bar: 1mm. Magnification: x40. Scale bar=0.5 mm.	32
3.2	Post-embryonic developmental stages in hybrid ♀CMx♂CG. (A) heart formed (H); optic primordium (OP); otoliths (OT). (B) yolk sac (YS). (C) barbells as diminutive knots at 1 DAH; (B) myotomal muscle (MM). (D) developed caudal fins at 3 DAH (CF); developed operculum (O). Scale bar: 1 mm. Magnification: x40. Scale bar=0.5 mm.	33
3.3	Growth curve showing fortnightly length increase of fry of <i>Clarias gariepinus</i> , <i>Clarias macrocephalus</i> and hybrid CMxCG for a rearing period of 8 weeks	34
3.4	Growth curve showing fortnightly weight increase of fry of <i>Clarias gariepinus</i> , <i>Clarias macrocephalus</i> and hybrid CMxCG for a rearing period of 8 weeks	34
3.5	Survival of larvae of <i>Clarias gariepinus</i> , <i>Clarias macrocephalus</i> and hybrid CMxCG for a rearing period of 8 weeks	35
4.1	Schematic illustration of morphometric characters showing location of measurements on <i>Clarias</i> specimens according to Teugels (1986) and Agnese et al. (1997).	44

4.2	Schematic representation of meristic counts among <i>Clarias</i> specimens: DFR, Dorsal fin rays; AFR, Anal fin rays; PFR, Pectoral fin Rays; VR, Ventral Rays, Caudal Rays and NOB, Number of Barbels.	45
4.3	Plot of morphometric variables of Clariids. (a) Percentage in parenthesis represents the variation of each component. (b) The biplot shows the positions of PC score of <i>Clarias</i> species according to PC1 and PC2	55
4.4	Plot of meristic variables of Clariids. (a) Percentage in parenthesis represents the variation of each component. b) The biplot shows the positions of PC score of <i>Clarias</i> species according to PC1 and PC2	56
4.5	Morphological comparison between (A) <i>C. macrocephalus</i> (B) <i>C. gariepinus</i> (C) hybrid CMxCG in Malaysia used in the present study. Note the <i>C. macrocephalus</i> have large numbers of small white spot along its body sides.	57
4.6	The shape of the head, occipital process, and frontal fontanelle of (A) <i>C. gariepinus</i> (CG), (B) <i>C. macrocephalus</i> (CM) and (C) hybrid CMxCG.	58
4.7	White double arrow (left picture) showing the distance between occipital process and dorsal fin (DODF) of (A) <i>C. gariepinus</i> , (B) <i>C. macrocephalus</i> , and (C) hybrid CMxCG while black double arrow and white doble arrow (right picture) showing distance between dorsal and caudal fin (DDCF) and Caudal peduncle depth (CPD) of (A) hybrid CMxCG, (B) <i>C. macrocephalus</i> , and (C) <i>C. gariepinus</i> , respectively.	58
4.8	The premaxillary and vomerine toothplates of (A) <i>C. macrocephalus</i> , (B) <i>C. gariepinus</i> , (C) hybrid CMxCG	59
4.9	The premaxillary and vomerine toothplates of (A) <i>C. macrocephalus</i> , (B) <i>C. gariepinus</i> , (C) hybrid CMxCG	59
6.1	Morphological characteristics of <i>A. hydrophila</i> . (A-B) Single colonies of <i>A. hydrophila</i> on TSA. (C) <i>A.</i>	92

*hydrophila* on horse blood agar (D) *A. hydrophila* colonies on RSA

6.2	Gram staining of <i>A. hydrophila</i> under light microscope showed Gram negative short rod (magnification 1000x)	93
6.3	PCR amplicon (16S rDNA) amplified from bacteria samples	96
6.4	Aerolysin gene detected in <i>Aeromonas hydrophila</i> isolated from <i>C. gariepinus</i>	97
6.5	Cumulative mortality of <i>Clarias gariepinus</i> , <i>Clarias macrocephalus</i> and hybrid CMxCG at $2.1 \times 10^6$ cfu mL <sup>-1</sup> .	98
6.6	Clinical signs observed on moribund of hybrid CMxCG (A1, A2, A3), <i>Clarias macrocephalus</i> (B1, B2, B3), <i>Clarias gariepinus</i> (C1, C2, C3) challenged with $2.1 \times 10^6$ cfu mL <sup>-1</sup> of <i>Aeromonas hydrophila</i> .	99
6.7	Internal organs of (A) hybrid CMxCG, (B) <i>Clarias gariepinus</i> , and (C) <i>Clarias macrocephalus</i> challenged with $2.1 \times 10^6$ cfu mL <sup>-1</sup> of <i>Aeromonas hydrophila</i> shows kidney (K) congestion and enlargement of spleen (S) with pale liver (L).	100
6.8	Histopathological changes in the kidney (A), liver (B) and spleen (C) of <i>Clarias gariepinus</i> control group. H & E 400x	105
6.9	Histopathological changes in the kidney (A), liver (B) and spleen (C) of infected <i>Clarias gariepinus</i> at 96 h post challenged.	106
6.10	Histopathological changes in the kidney (A), liver (B) and spleen (C) of infected <i>Clarias macrocephalus</i> at 96 h post challenged.	107
6.11	Histopathological changes in the kidney (A), liver (B) and spleen (C) of infected hybrid CMxCG at 96 h post challenged.	108

## LIST OF ABBREVIATIONS

°C	Degree celcius
m	Meter
cm	Centimeter
mg	Miligram
mM	Milimolar
mL	Mililiter
μl	Microliter
bp	Base pair
kb	Kilobase
rpm	Revolution per minute
V	Volt
MgCl <sub>2</sub>	Magnesium chloride
NCBI	National Center for Biotechnology Information
Sp. Or spp.	Species or species (plural)
Cfu	Colony-forming unit
dH <sub>2</sub> O	Distilled water
DNA	Deoxyribonucleic acid
dpi	dpi
EDTA	Ethylenediaminetetraacetic acid
H&E	Hematoxylin and eosin
HCL	Hydrochloric acid
LD <sub>50</sub>	Median lethal dose
NaOH	Sodium hydroxide

NaCl	Sodium chloride
PCR	Polymerase chain reaction
TBE	Tris-borate-EDTA
TSA	Trypticase soy agar
RSA	Rimler-Shott Agar
UV	Ultra violet
h	Hour
EFA	Essential fatty acid
EAA	Essential amino acid
MUFA	Monounsaturated fatty acid
PUFA	Polyunsaturated fatty acid



## CHAPTER 1

### INTRODUCTION

#### 1.1 World Fisheries and Aquaculture

Human population is increasing rapidly where it expected to reach 9.6 billion people by 2050 (FAO 2014). Reports of large numbers of people that are facing social problems such as poverty and chronic malnourishment has made the need for food production one of the worldwide issues of concern. Fisheries is one of the activities that contribute to major food production since people have depended so greatly on fish and other aquatic food as the primary sources of dietary animal protein for their well-being. Apart from being a economical source of animal protein of the household compare to poultry and red meat, fish also contains high amounts of vitamins and good quality of amino acids such as lysine, methionine and tryptophan as well as high polyunsaturated fatty acids (PUFA) omega three  $\omega$ -3 (n-3) and omega six  $\omega$ -6 (n-6) which are good for human health (Simopoulos, 2004; Sidhu, 2003; Lovell, 1989; Benitez, 1999).

According to FAO (2014), world per capita apparent fish consumption increased each year and it is in fact highly concentrated in Asia. Total food fish consumption demand particularly in South-East Asia is expected to increase by 36.3% from 14.2 million tonnes in 2010 to 19.3 million tonnes in 2030 (Worldbank, 2014). Apparently however, recent reports obviously shows that fish supplies from global captured fisheries are declining rapidly due to several factors including over-fishing, habitat destruction, and pollution (Eyo 2001; Dunham et al 2001; Olufeagba et al 2007; Adewumi & Olaleye 2011). Due to this current issues, it is undisputable that fish supplies from capture fisheries sector solely will not able to meet the heavy demand fish and fish based products in the near future (FAO, 2006). Hence, aquaculture or fish farming became one of the recommended activities that can fits exactly into the role as an alternative fish production system and satisfy the high domestic demand for fish and export demand of fish products as well.

Generally, there are several species of catfish belong to the family *Clariidae* are present in Malaysia. They are *Clarias anfractus*, *Clarias leiocanthus*, *Clarias planiceps*, *Clarias sulcatus*, *Clarias teijsmanni*, *Clarias batrachus* and *Clarias macrocephalus*. The English “walking catfish”, or “Ikan Keli” in English and Malay respectively, is a generic name for a group of catfish species belongs to the family *Clariidae*. Among them, *Clarias batrachus*, or also known as “keli kayu” and *Clarias macrocephalus*, which is locally known as “keli bunga” are the most common and important native species in Malaysia.

*Clarias macrocephalus* is found to be generally restricted to the northern region of Peninsular Malaysia, particularly in Kedah and Perlis which are famous with

paddy fields as their natural environment and habitat (Mohsin and Ambak, 1983; Lee et al., 1993). On the other hand, *C. macrocephalus* also can be found in irrigation canals, stagnant pools or streams. As an air-breathing fish, this species of catfish is tolerant to harsh environmental conditions such as extreme dry season and consequently have a capability to move to another secure habitats using their pectoral fins especially during spawning season, feeding and seeking for shelter (Pouyaud et al., 2009).

On the other hand, *Clarias gariepinus* (Burchell, 1822), which is another common species of *Clarias* catfish, was introduced to Malaysia from North Africa. This African catfish was introduced to Malaysia from Thailand in the early 1980's which is between 1986-1989 (Csavas I, 1995) and is currently represents as dominant finfish culture not only in Malaysia but also in various regions in the world (Akeem et al., 2018). The first report of *C. gariepinus* culture in Malaysia can be traced back to 1987 with an annual output of 6.46 metric tonnes, and this has developed and grown quickly since then (Akeem et al. 2018). Available official statistical data was obtained from Department of Fisheries (DOF), Ministry of Agriculture and Food Industries, Malaysia, showed that freshwater catfish, *Clarias* sp. had the highest total production of 555, 822 MT between 2007 and 2018 (DOF, 2018; Saba et al., 2020). However, according to FAO (2017), the statistics of total production of African catfish especially in Malaysia is currently reported inaccurate production amount. This inconsistency was said to be due to farming of African catfish hybrids and it was difficult to separate the data obtained from farmers for pure African catfish and that of hybrid. Other than that, misclassifications also were found between *C. gariepinus* and hybrids. Therefore, FAO did not capture the output under the name African catfish but were usually reported as *Clarias* sp. for both hybrid and nonhybrid (Xiaowei Zhou, 2017).

Furthermore, the success in gonadal maturation in captivity followed by induced spawning and mass seed production resulted in the rapid expansion of the hybrid catfish culture by local hatcheries. In fact, the catfish that is broadly cultivated now is the hybrid between local female clariid species, *Clarias macrocephalus* and exotic male, *Clarias gariepinus*. The *Clarias* hybrid with fast growth, high disease resistance and environmental conditions (from paternal genes), high meat quality and good taste (from maternal genes), are attractive to farmers and has become one of the most popular freshwater fishes by consumers (Senanan et al., 2004). As a result, hybrid Clariid catfishes almost completely replaced the native clariid catfish aquaculture in our countries and other countries such as Thailand (Poompuang and Na-Nakorn, 2004). In addition, it has given a great potential to aquaculture of clariid catfishes in many Asian countries and positively impacted the livelihoods of many catfish farmers. Since Clariid catfishes has become a great economic importance in Malaysia, especially as a food fish and vital in the local sustainability of the aquaculture activity, it has been widely cultures uncontrollably both in ponds and artificial tanks without considering the proper husbandry and management practices of culture systems (Akeem et al., 2018; FAO, 2010).

Likewise, the culture methods also have become more intensive in order to produce higher yields which simultaneously caused disease problems in the aquaculture industry. Disease issues has become a great concern in aquaculture production because it can impacts negatively on market value and lead to great monetary losses which is related to increase production costs such as high mortality of cultured animals, increase cost of treatment, decreased quality and quantity of yield and loss of the opportunity to sell the fish due to undesirable appearance which affects customer's choice (Idowu et al., 2017; Akoll and Mwanja, 2012; Subasinghe et al., 2002). Studies showed that almost fifty percent of production loss is because of diseases-related problems and become more severe in developing countries (Ayalew and Fufa, 2018). In addition to disease problems, abiotic stresses such as water quality factors including high water temperature and low levels of dissolved oxygen also can reduce the effectiveness of the fishes immune systems and then lead to significant economic losses (Welker et al., 2007). Stressors can be defined as the sum of the physiological responses the fish makes both physiologically and behaviourally to adapt with deterioration of environmental conditions and then regain its normal balance (Chris et al., 2003). However, if a fish is constantly exposed to any stressor which is exceeds the fish's ability to adapt, it may be lethal or will facilitate the infection by opportunistic pathogens or parasite which may present in the water.

## **1.2 Statement of Problems and Significance of Study**

Genetic improvement of economically important traits of farm-raised catfish using interspecific hybridization has become one of a short-cut approach to increase the production of fish, lower the production costs and improve profitability for catfish farmers. Attributes that have been associated with hybrid catfish include faster growth rates, higher survival rate, improved yield and meat quality and improve disease resistance. These attributes would motivates the community and catfish farmers to breed this type of fish and will contribute to the increased of inland fisheries production through aquaculture (Bartley et al., 2001; Aminur Rahman, 2013; Omeji et al., 2013).

However, to date, as the production of *Clarias* hybrids draw attention among catfish farmers. In Thailand, hybrid catfish production was introduced since 1988 by artificially crossing males of African catfish (*Clarias gariepinus*), with females of the Thai walking catfish (*Clarias macrocephalus*). These hybrid catfish is already adopted for commercial aquaculture in Thailand since 1996 with more than 80% of catfish farmers raise hybrid catfish (Na-Nakorn, 1999). Although hybrid catfish is currently became the dominant fish culture in Thailand, this present study must be done particularly in Malaysia to develop better understanding on the gene interactions and degrees of genetic differences (Falconer, 1989) contributed by two distinct populations, species or inbred lines produced by the hybrid catfish in the future. Additionally, differences of strains used could effects the superior performance of the interspecific catfish hybrids despite the large genetic differentiation between the parental species (Koolboon et al., 2014).

On the other hand, since culture of the hybrids catfish are widely used in aquaculture, the possibilities to increase production through inappropriate hybridization will also increase. Some hybrids are accidentally produced through mixed spawning of different species in a hatchery, misidentification of species by hatchery personnel, or by contamination of the aquaculture facility with wild fish (Senanan et al., 2004; Akeem et al., 2018). Such uncontrolled and unintentional hybridization could decrease the performance of cultured stocks and produce unrecognized introgressed individuals, thus, make future use of the contaminated stocks as broodstock questionable (Senanan et al., 2004). Other problem that hybrids presently cause with regard to their inclusion in FAO database due to the incorporation of production information on hybrids by catfish farmers or aquaculturist (Bartley et al., 2001). Consequently, most of the hybrid catfish production is simply list and reported to the taxonomic level of genera only which is *Clarias* spp. The report with incompletely identified species reflected official FAO statistics and reduces the effectiveness of the data for monitoring the utilization of aquatic biodiversity for aquaculture. On the other hand, many fish geneticists and molecular biologists realized that the use of inter-specific hybrid is not well-reported (Aminur Rahman et al., 2013). The lack of constant reporting hybrid species may be due to the difficulty of assessing parental origins of inter-specific hybrids.

Therefore, since catfish hybrids (*C. macrocephalus* x *C. gariepinus*) are produced from the Clariid species that belong to the same genus and the external features of hybrids show very similar in appearance with their parental species, identifying these individuals become critical not only for development of management strategies in aquaculture and fisheries industry but also to allow for a better understanding of biodiversity among aquaculturist. It can reduce the potential of ecological problems by preventing the risk of fish from hybridizing in nature which is considered as one of good efforts to conserve native catfish as well. A morphological analysis and molecular genetics markers such as microsatellite DNA are important methods to differentiate inter-specific hybrids species (Chaiparinya, 1996; Na-Nakorn, 1999; Senanan et al., 2004; Nazia and Siti Azizah, 2014).

Other than that, since interest in hybridization of Clariid catfish in aquaculture has been purely for genetic and economic importance, this current study aimed to evaluate the successful of hybridizations between *Clarias gariepinus* and *Clarias macrocephalus*, and demonstrated the possible reproductive performance such as growth and survival, disease resistance and proximate composition of hybrids of this species compared to their parental lines crosses. Assessment of morphometric variation of the parental species and its hybrids also is considered as critical for identification purposes. Hence, the hypotheses and research objectives were as follows:

### 1.3 Research Hypothesis

Hybridization between African catfish, *C. gariepinus* and Asian catfish, *C. macrocephalus* holds high potential for global aquaculture production in the coming years due to general achievement of better growth rate, survival rate, high disease resistance, flesh quality and good nutritional value. The offspring of this *Clarias* sp. is considered exhibit positive heterosis for culture traits as they can perform better than both or at least either one of the parental lines. Other than that, there is a variation in the morphometric characteristics of hybrid CMxCG and its parental species.

### 1.4 Objectives of the study

1. To determine the growth rates and survival rates of the hybrid of CMxCG in comparison with its parental lines as a control
2. To examine the differentiation of morphological characteristic of hybrid CMxCG and its parental species
3. To compare the nutritional value of hybrid CMxCG and its parental species
4. To determine the 96 h-LD<sub>50</sub> values of hybrid CMxCG and its parental species towards *A. hydrophila*
5. To examine the clinical signs and histopathological changes in kidney, liver and spleen of hybrid CMxCG and its parental species



## REFERENCES

- Abbott, J.C., & Dill, L.M. (1985). Patterns of aggressive attack in juvenile steelhead trout (*Salmo gairdneri*). *Can J Fish Aquat Science*, 42, 1702-6.
- Abdelhamed, H., Ibrahim, I., Baumgartner, W., Lawrence, M.L., & Karsi, A. (2017). Characterization of histopathological and ultrastructural changes in channel catfish experimentally infected with virulent *Aeromonas hydrophila*. *Frontiers in Microbiology*, 8: 1519.
- Abimbola, A. O., Kolade, O. Y., Ibrahim, A. O., Oramadike, C. E., & Ozor, P. A. (2010). Proximate and anatomical weight composition of wild brackish *Tilapia guineensis* and *Tilapia melanotheron*. *Journal of Food Safety*, Volume 2, Nigerian Institute for Oceanography and Marine Research, Victoria Island, Lagos, pp. 100-103.
- Abdullahi, S. A. (2001). Investigation of nutritional status of *Chrysichthys nigrodigitatus*, *Barus filamentous* and *Auchenoglanis occidentals*: Family Barigidae. *Journal Arid Zone Fisheries*, 1, 39-50.
- Abdullwahab, R., Auta, J., & Jatau, A. (2016). Comparative proximate composition of *Clarias gariepinus* (Burchell, 1822), *Oreochromis niloticus* (Linnaeus, 1754) and *Brycinus nurse* (J. P Muller and Troichel, 1846) caught from Bakori dam in Katsina state. *Journal of Arts and Science*, 9, 1-6.
- Aboyadak, I. M., Nadia, GMA1., Ashraf, A. G., Walaa, H., A., & Asmaa, A. (2015). Molecular detection of *Aeromonas hydrophila* as the main cause of outbreak in Tilapia farms in Egypt. *Journal of Aquaculture and Marine Science*, 2(5), 00045.
- Abol-Munafi, A. B., Masduki, M. M., Liem, P.T., & Ambak, M. A. (2003). Study on the production of hybrid catfish. Poster presented at "Research and Development Exposition 2003", PWTC, Kuala Lumpur, 9-12 October 2003.
- Ackman, R. G. (1989). Nutritional composition of fats in seafoods. *Progress in Food and Nutrition Science*, 13: 161-241.
- Ackman, R. G. (2007) Fatty Acids in Fish and Shellfish. Fatty acids in foods and their health implications. 19:155.
- Adah, P. M., Onyia, L.U., & Obande, R. A. (2014). Fish Hybridization in Some Catfishes: A Review. *Biotechnology*, 13:248-251.
- Adebambo, A., Adeleke, M., Whetto, M., Peters, S., Ikeobi, C., Ozoje, M., Oduguwa, O., & Adebambo, O.A. (2010). Combining abilities of carcass traits among pure and crossbred meat type chickens. *International Journal of Poultry Science*, 9, 777-783.

- Adebayo, O.T. (2006). Reproductive performance of African Clariid catfish *Clarias gariepinus* broodstock on varying maternal stress. *Journal Fish International*, 1(1-2), 17-20.
- Adefemi, O.S. (2011). Chemical composition of Tilapia mossambis fish from major dams in Ekiti-State. *Nigeria Africa Journal of Food Science*, 5(10), 550-554.
- Adeyeye, E. L., & Adamu, A. S. (2005). Chemical composition and food properties of *Gymnarchus niloticus* (Trunk fish). *Journal of Bioscience and Biotechnology Research Asia*, 3(2), 265-272.
- Adewolu, M.A., Ogunsanmi, A.O., & Yunusa, A. (2008). Studies on Growth Performance and Feed utilization of two clariid catfish and their hybrid reared under different culture systems. *Europe Journal Science Research*, 23 (2): 252-260.
- Adewumi, A. A., Olaleye, V.F. (2011). Catfish culture in Nigeria: Progress, prospects and problems. *African Journal of Agricultural Research*, 6, 1281-1285.
- Adwoye, S. O., & Omotosho, J. S. (1997). Nutrient composition of some freshwater fishes. *Nigeria Bioscience research and Communication*, 11(4), 333-3336.
- Afifi , S. H., Al-Thobiati, S., & Hazaa, M. S. (2000). Bacteriological and histopathological studies on *Aeromonas hydrophila* infection on Nile tilapia (*Oreochromis niloticus*) from fish farms in Saudi Arabia. *Assiut Veterinary Medical Journal*. 42: 195-205.
- Agius, C., & Roberts, R. J. (2003). Melano-macrophage centers and their role in fish pathology. *Journal of Fish Disease*, 26: 499-509.
- Agnese, J.F., Teugels, G.G., Galbusera, P., Guyomard, R., &Volckaert, F. (1997). Morphometric and genetic characterization of sympatric populations of *Clarias gariepinus* and *C. anguillaris* from Senegal. *Journal of Fish Biology*, 50, 1143–1157.
- Ahmed, E. O., Ali, M. E., Khalid, R.A., Taha, H. M., Mahamad, A.A. (2010). Investigating the quality changes of raw and hot smoked *Oreochromis niloticus* and *Clarias lazera*. *Pakistan Journal of Nutrition*, 9, 481-484.
- Airina, N. M., & Jamaludin, M. (2012). Fatty acids composition of selected Malaysian fishes. *Sains Malaysiana*, 41(1), 81-94.
- Akeem, B. D., Natrah, I., Karim, M., Kamarudin, M. S., & Bichi, A. H. (2018). African catfish aquaculture in Malaysia and Nigeria: Status, Trends and Prospects. *Fisheries and Aquaculture Journal*, 9, 1.
- Akinwande, A. A., Fagbenro, O. A., & Adebayo, O. T. (2013). Phenotypic characterization in interspecific and intergeneric hybrids of the clariid

- catfishes *Heterobranchus longifilis*, *Clarias gariepinus* and *Clarias anguillaris* in Nigeria. *Afr. J. Aquat. Sci.* 38, 109–113.
- Akoll, P., & Mwanjaa, W. W. (2012). Fish health status, research and management in East Africa: past and present. *African Journal of Aquatic Science*, 37(2), 117-129.
- Aleev, Y.G. (1969). Function and gross morphology in fish. Israel Program for Scientific Translations, Jerusalem, TT 67-51391 (Transl. from Russian).
- Ali Farag, A., Hassan, M. D., Saleha, A. A., Siti Khairani, B., & Alsaied, M. (2011). Pathogenicity of *Streptococcus agalactiae* isolated from a fish farm in Selangor to juvenile red tilapia (*Oreochromis* sp.). *Journal of Animal and Veterinary Advances*, 10(7): 914-919.
- Ali, M., Igbal, F., Salam, A., Iram, S., & Athar, M. (2005). Comparative study of body composition of different fish species from brackish water pond. *International Journal of Environmental Sciences and Technology*, 2(3), 229-232.
- Allendorf, F.W., Leary, R.F., Spruell, P., & Wenburg, J.K. (2001). The problems with hybrids: Setting conservation guidelines. *Trends Ecol. Evol*, 16(11), 613-622.
- Ama-Abasi, O., & Ogar, A. (2013). Proximate analysis of snakehead fish-*Parachanna obscura* (Gunther, 1861) of the Cross River Nigeria. *Journal of Fisheries and Aquatic Science*, 8, 295-298.
- Amer, H.A., Sedik, M. F., Khalafalla, F. A., & Awad, H.A. (2005). Results of chemical analysis of prawn muscle as influenced by sex variations. *Food/ Nahrung*, 35, 133-138.
- Aminur Rahman, M., Arshad, A. B., Marimuthu, K., Ara, R., & Amin, S.M.N. (2013). Inter-specific hybridization and its potential for aquaculture of fin fishes. *Asian Journal of Animal and Veterinary Advances*, 8(2), 139-153.
- Anetekhai, M. A. (2013). Catfish Aquaculture Industry Assessment in Nigeria. African Union – Inter African Bureau for Animal Resources.
- Anyanwu, M. U., Kennedy, F. C., & Vincent, S. S. (2015). Evaluation of pathogenicity of motile *Aeromonas* species in African catfish. *International Journal of Fisheries and Aquatic Studies*, 2(3), 93-98.
- Angka, S. L. (1990). The pathology of the walking catfish, *Clarias batrachus* (L.), infected intraperitoneally with *Aeromonas hydrophila*, *Asian Fish Science*, 3: 343-351.
- Angka, S.L., Lam, T.J., & Sin, Y.M. (1995). Some virulence characteristics of *Aeromonas hydrophila* in walking catfish (*Clarias gariepinus*). *Aquaculture*, 130: 103-112.



- Association of Official Analytical Chemist (2012): Official Methods of Analysis of (AOAC) 19th Edition by George Latimer (Author) Maryland, USA.
- Argue, B.J., Lui, Z., & Dunham, R.A. (2003). Dress-out and fillet yields of Channel Catfish, *Ictalurus punctatus*, Blue Catfish, *Ictalurus furcatus*, and their F1, F2 and backcross hybrids. *Aquaculture* 288:81-90
- Arias, C. R., Cai, W., Peatman, E., & Bullard, S. A. (2012). Catfish hybrid (*Ictalurus punctatus* x *I. furcatus*) exhibits higher resistance to columnaris disease than the parental species. *Disease of Aquatic Organisms*, 100(1), 77-81.
- Ataguba, G. A., Annune, P.A., & Ogbe, F. G. (2010). Growth performance of two African catfishes *Clarias gariepinus* and *Heterobranchus longifilis* and their hybrids in plastic aquaria. *Livestock Research for Rural Development* [online serial] 22:30
- Austin, B., & Austin, D. A. (2012). Bacterial fish pathogens.: Disease of farmed and wild fish, 3rd edition, pp: 112-115. Al-Yahya, S. A., Fuad, A., Khalidah, S. A. N., Bachar, A. S., Sarfaraz, H., & Ashraf, A. M. (2018). Histopathological studies of experimental *Aeromonas hydrophila* infection in blue tilapia, *Oreochromis aureus*. *Saudi Journal of Biological Sciences*, 25:182-185.
- Ayelew, A., & Fufa, A. (2018). Maintenance of Fish Health in Aquaculture: Review of Epidemiological Approaches for Prevention and Control of Infectious Disease of Fish. *Veterinary Medicine International*, 4-12.
- Azim, M. A., Islam, M. R., Hossain, M. B., & Minar, M. H. (2012). Seasonal variations in the proximate composition of gangetic sillago (*Sillaginopsis panijus*, Perciformes): Sillaginidae. *Middle East Journal Science and Research*, 11, 559-562.
- Bartley, D. M., Rana, K., & Immink, A.J. (2000). The use of inter-specific hybrids in aquaculture and fisheries. *Reviews in Fish Biology and Fisheries*, 10(3): 325 - 337.
- Bartley, D.M., Rana, K., & Immink A.J. (2001). The use of interspecific hybrid in aquaculture and fisheries. *Review Fish Biology Fish*, 10:325-37.
- Begum, M., Akter, T., & Minar, M. H. (2012). Analysis of the proximate composition of domesticated stock Pangas (*Pangasianodon hypophthalmus*) in Laboratory condition. *Journal of Environmental science and Natural Resources*, 5(1), 69-74.
- Bhowmick, R. M., Jana, R. K., Gupta, S. D., Kowtal, G. V., & Rout, M. (1981). Studies on some aspects of biology and morphometry of the intergeneric hybrid *Catla catla* (Hamilton) x *Labeo rohita* (Hamilton) produced by hypophysation. *Aquaculture* 23, 367-371.
- Bleeker, 1846 and *Pangasionodon hypophthalmus* Sauvage, 1878. *Indones. J. Agric. Sci.* 5, 70-74.

- Boyde, C. (2012). Water quality. In: Aquaculture: farming aquatic animals and plants (2nd edition). John L. S., Paul S. C. (eds), Blackwell Publishing Ltd., pp. 52-83.
- Boulanger, Y., Lallier, R. & Cousineau, G. (1977). Isolation of enterotoxigenic *Aeromonas* from fish. *Canadian Journal of Microbiology*, 23: 1161-1164.
- Bosakowski, T. and Wagner, E.J. 1994. Assessment of fin erosion by comparison of relative fin length in hatchery and wild trout in Utah Can J Fish Aquat Sci. 51(3):636-41.
- Buckley, J. T. (1991). Secretion and mechanism of action of the hole-forming toxin aerolysin. *Review Experientia*, 47(5), 418-419.
- Bullock, G. L. (1961). The identification and separation of *Aeromonas liquefaciens* from *Pseudomonas fluorescens* and related organisms occurring in diseased fish. *Journal of applied Microbiology*, 9: 587-590.
- Burgess, W.E.(1989). An atlas of freshwater and marine catfishes. A preliminary survey of the Siluriformes. T.F.H. Publications, Inc., Neptune City, New Jersey (USA), p:784.
- Cai, W. Q., Li, S. F., & Ma, J. Y. (2004). Disease resistance of Nile tilapia (*Oreochromis niloticus*), blue tilapia (*Oreochromis aureus*) and their hybrid female Nile tilapia x male blue tilapia to *Aeromonas sobria*. *Aquaculture*, 229(1-4), 79-87.
- Chaijan, M. A., Jongjareonrak, S., Phatcharat, S., Benjakul, & Rawdkuen, S. (2010). Chemical compositions and characteristics of farm raised giant catfish (*Pangasianodon gigas*) muscle. *LWT-Food Science and Technology*, 43(3), 452-457.
- Chakwa, O., & Shaba, I. M. (2009). Effect of drying method on proximate composition of catfish (*Clarias gariepinus*). *Journal of World Agricultural Sciences*, 5(4), 114-116.
- Chatruvedi, C.S., & Pandey A.K. (2012). Successful induced breeding and hatchery development of Asian catfish, *Clarias batrachus* in port Blair. *Biochem. Cell. Arch.*, 12: 321-325.
- Cheah, M.S.H. & Lee, C.L. (2000). *Asian Fisheries Science*, 13:87-96.
- Chedoloh, R., Karrila, T. T., & Pakdeechanuan, P. (2011). Fatty acid composition of important aquatic animals in Southern Thailand. *International Food Research Journal*, 18, 783-790.
- Chen, C., Sander, J. E., & Dale, N. M. (2003). The effect of dietary lysine deficiency on the immune response to Newcastle disease vaccination in chickens. *Avian diseases*, 47(4), 1346-1351.
- Chevassus, B. (1979). Hybridization in salmonids: results and perspectives. *Aquaculture*, 17(2), 113-128.

- Chevassus, B. (1983). Hybridization in fish. *Aquaculture*, 33, 245-262.
- Chu, W. H., & Lu, C. P. (2005). Multiplex PCR assay for the detection of pathogenic *Aeromonas hydrophila*. *Journal of Fish Disease*, 28, 437-441.
- Cipriano, R. C. (2001). *Aeromonas hydrophila* and motile aeromonad septicemias of fish. Disease Leaflet 68. [https://articles.extension.org/sites/default/files/w/1/1e/Aeromonas\\_hydrophila.pdf](https://articles.extension.org/sites/default/files/w/1/1e/Aeromonas_hydrophila.pdf).
- Cruz, C.N.B, Cruz, C.P.E., & Mahecha, H.S. (2012). Characterization of the Nutritional quality of the meat in some species of catfish. A review. *Rev. Fac. Nal. Agr. Medellin* 65(2):6799-6709.
- Csavas, I. (1995). Status and perspectives of culturing catfishes in East and South-East Asia. Presented at the International Workshop on the Biological Bases for Aquaculture of Siluriformes, May, Montpellier, France.
- Dhara, K., & Saha, N. C. (2013). Controlled Breeding of Asian Catfish *Clarias batrachus* using Pituitary Gland Extracts and Ovaprim at different Temperatures, Latency Periods and their Early Development. *Journal of Aquaculture*, 4,4.
- Dempson, I. B., Schwarz, C. J., Shears, M., & Furey, G. (2004). Comparative proximate body composition of Atlantic salmon with emphasis on parr from fluvial and lacustrine habitats. *Journal of Fish Biology*, 64, 1257-1271.
- De-Silva, S. S., Gunasekera, R. M., Ingram, B.A. (2004). Performance of intensively farmed murray cod *Maccullochella peelii peelii* (Mitchell) fed newly formulated vs. Currently used commercial diets, and a comparison of filled composition of farmed and wild fish. *Aquaculture Research*, 35, 1039-1052.
- De-Silva M. P. K. S. K. (2015). Genetic diversity of genetically improved farmed Tilapia (GIFT) brood stocks in Sri Lanka. *International Journal of Scientific Research and Innovative Technology*, 2(3):66-76.
- Dezhabad, A., Dalirie, M. S., & Toudar, S. (2012). Amino acid profile of Kutum (*Rutilus frisii*), silver carp (*Hypophthalmichthys molitrix*) and rainbow trout (*Onchorhynchus mykiss*). *African Journal of Agricultural Research*, 7(34), 4845-4847.
- Diyaware, M. Y., & Onyila, L. U. (2014). Growth and survival of intergeneric hybrids of *Clarias anguillaris* and *Heterobranchus bidorsalis* in semi arid zone of Nigeria. *Journal of Fisheries and Aquatic Science*, 9:398–406.
- Doan, H. Van. (2013). The LD50 of Asian catfish (*Pangasius bocourti*) (Sauvage 1870) challenge to pathogen *Aeromonas hydrophila* FW52 strain. *Pensee Journal*, 57(10), 287-293.

- DOF. (2016). Annual Fisheries Statistics 2015. Department of Fisheries, Malaysia, Ministry of Agriculture and Agro-based industries, Putrajaya.
- Dorsch, M., Ashbolt, N. J., Cox, P. T., & Goodman, A. E. (1994). Rapid identification of *Aeromonas* species using 16S rDNA targeted oligonucleotide primers: a molecular approach based on screening of environmental isolates. *Journal of Applied Bacteriology*, 77: 722-726.
- Dorson, M., Chevassus, B., & Torhy, C. (1991). Comparative susceptibility of three species of char and rainbow trout x char triploid hybrids to several pathogenic salmonid viruses. *Disease of Aquatic Organisms*, 11(3), 217-224.
- Duong, T.Y., Nguyen, T.T., & Pham, T.L. (2017). Morphological differentiation among cultured and wild *Clarias macrocephalus*, *C. macrocephalus* x *C. gariepinus* hybrids, and their parental species in the Mekong delta, Viet Nam. *International Journal of Fisheries and Aquatic Studies*, 5(1): 233-240.
- Dunham, R. A., Majumdar, K., Hallerman, E., & Main, G. (2001). Review of the status of aquaculture genetics. In K.R.P. Subasinghe, P. Buemo, M.J. Philipa; C. Haugh; S.E. and J.R. Arhur (eds). *Aquaculture in the third millennium. Proceedings of the conference on Aquaculture in the third millennium*, Bangkok, Thailand, 20-25 February, NACA, Bangkok and FAO, Rome, 137-186.
- Eccles, D.H. (1992). FAO Species Identification Sheets for Fishery Purposes. Field Guide to the Freshwater Fishes of Tanzania. UNDP Project URT/87/016, Rome.
- Effiong BN, Mohammed I. (2008). Effect of seasonal variation on the nutrient composition in selected fish species in Lake Kainji Nigeria. *Nature and Science* 6(2): 1545 – 1590.
- Effiong, B. N., & Fakunle, J. O. (2013). Proximate composition and fatty acid profile in some commercially important fish species from Lake Kainji, Nigeria. *International Journal of Biology Pharmacy Science*, 2, 849-856.
- El-Hawarry, W. N., Soliman, H. A. R., Ramy, M. S. (2016). Breeding response and larval quality of African catfish (*Clarias gariepinus*, Burchell 1822) using different hormones/hormonal analogues with dopamine antagonist. *Egyptian Journal of Aquatic Research*, 42, 231-239.
- Elliott, N.G., Haskard, K., & Koslow, J.A. (1995). Morphometric analysis of orange roughy (*Hoplostethus atlanticus*) off the continental slope of southern Australia. *Journal of Fish Biology*, 46:202-20.
- El-Zaeem, S.Y., & Salam, G.M. (2013). Production of genetically male tilapia through interspecific hybridization between *Oreochromis niloticus* and *O. Aureus*. 12 (4): 802- 812

- Erkihun, M., Hintsu, M., Paul, L., & Ashagrie, Z. (2017). Proximate composition and fatty acid content of commercially important fish species from Ethiopian Lakes: A review. *World Journal of Food Science and Technology*, 1(3), 105-114.
- Ersoy, B., & Özeren, A. (2009). The effect of cooking methods on mineral and vitamin contents of African catfish. *Food Chemistry*, 115(2): 419-422.
- Etzel, M. R. (2004). Manufacture and use of dairy protein fractions. *Journal of Nutrition*, 134(4), 996-1002.
- Eyo, & Joseph, E. (2004). Differences in meristic counts of the genus *Clarias* (Pisces: Clariidae) in Anambra River, Nigeria. *Animal Research International*, 1(1), 31-35.
- Eyo, A.A. (2001). Fish Processing Technology in the tropics. National Institute for Freshwater Fisheries Research, University of Ilorin, Nigeria 430pp.
- Ezeafulukwe, C.F., Njoku, D.C., Ekeledo, C.B., & Adaka, G.S. (2015). Morphomeristic characteristics of selected cichlid fishes from two aquatic environments in Imo state, Nigeria. *Int J. Vet Science*, 4(3):131-5.
- Fagbenro, O. A., Adeparusi, E. O., & Jimoh, W.A. (2007). Evaluation and nutrients quality of detoxified jackbean seeds cooked in distilled water or trona solution, as a substitute for soybean meal in Nile tilapia, *Oreochromis niloticus* diets. *Journal of Applied Aquaculture*, 19(2), 83-100.
- Fagbuaro, O., Oso, J.A, Olurotimi, M.B., & Akinyemi, O. (2015). Morphometric and meristic characteristics of *Clarias gariepinus* from controlled and uncontrolled population from South Western Nigeria. *Journal of agriculture and ecology research international*. Article no: JAERI.2015.005.
- FAO.(1996). Artificial Reproduction and Pond Rearing of the African Catfish *Clarias gariepinus* in Sub-Saharan Africa-A Handbook. Food and Agriculture Organization of the United Nations, Rome, Italy, p:179.
- FAO. (1997). FAO Database on Introduced Aquatic Species. FAO, Rome, Italy: Food and Agricultural Organization of the United Nations.
- FAO. (2006). The State of World Fisheries and Aquaculture 2006. FAO Fisheries and Aquaculture Department. Food and Agriculture Organization of the United Nations Rome, 2007. 5 – 22.
- FAO. (2010). Fishstat Plus Version 2.30. (Available source: <http://www.fao.org/fi/ststist/FISOFT/FISHPLUS>)
- FAO. (2014). Fisheries and Aquaculture Statistics. Rome, Italy. 105p.
- FAO. (2017). Fishery and Aquaculture Statistics. Global aquaculture production 1950-2015 (Fishstat). In: FAO Fisheries and Aquaculture Department. Rome.



- FAO/WHO/UNU (1985). Energy and protein requirement. WHO Technical Report Series: No 724. Geneva, Switzerland.
- Fang, H. M., Ling, K. C., & Sin, G. Y. M. (2000). Enhancement of protective immunity in blue gourami, *Trichogaster trichopterus* (Pallas), against *Aeromonas hydrophila* and *Vibrio anguillarum* by *A. hydrophila* major adhesin. *Journal of Fish Disease*, 23: 137-145.
- Falconer D. S. (1989). Introduction to quantitative genetics. (3rd Edn) Logman House. *Burnt Mill, Harlow, Essex, England: Longman Scientific and Technical*, 438.
- Falconer, D. S., & Mackay, T. F. C. (1996). Introduction to Quantitative Genetics, Fourth edition. *Pearson Education, Ltd.*, Essex, UK (464 pp).
- Farahvash, T., Shodja, J., Rafat, A., & Keshtkaran, A. (2011). The effect of bilateral crossbreeding between Arkhamerino and Ghezel Sheep on the quality of wool of their F1 crosses. *Asian Journal of Animal and Veterinary Advances*, 6(4), 397-400.
- Faskin, K. (2006). Fish farming (aquaculture) made easy. *LUSI J Publication Lagos*, 33.
- Favaloro, E., Lopiano, L., & Mazzola, A. (2002). Rearing of sharpsnout seabream (*Diplodus puntazzo*, Cetti 1777) in a Mediterranean fish farm: monoculture versus polyculture. *Aquaculture Research*, 33:137-40.
- Flos, R., Reig, L., Oca, J., & Ginovart, M. (2002). Influence of marketing and different land-based systems on gilthead sea bream (*Sparus aurata*) quality. *Aquaculture International*, 10:189-206.
- Frimodt, C. (1995). Multilingual illustrated guide to the world's commercial warmwater fish. Fishing News Books, Osney Mead, Oxford, England. 215 p.
- Froese, R., & Pauly, D. (2011). "*Clarias macrocephalus*" in FishBase.
- Folch, J., M., Lees, G.H.S., & Stanley. (1956). *Journal of Biology and Chemistry*. 195".497
- Fuentes, A., Fernandez-Segovia, I., Serra, J.A., & Barat, J.M. (2010). Comparison of wild and cultured sea bass (*Dicentrarchus labrax*) quality. *Food chemistry*, 119, 1514-1518.
- Funmilayo, S. M. (2016). Proximate composition and amino acid profiles of snakehead (*Parachanna obscura*) mudfish (*Clarias gariepinus*) and African pike (*Hepsetus odoe*) in Igboho dam, South-West Nigeria. *Journal of Global Science Research*, 4(4), 317-324.
- Garcia, F., Moraes, F. R., & Martins, M. L. (2010). Challenge of pacu (*Piaractus mesopotamicus*) fed diets supplemented with vitamins C and E by

- Aeromonas hydrophila* under different temperatures. *Arquivo Brasileiro de Medicina Veterinariae Zootecnia*, 61(2), 378-385.
- Garibaldi, L. (1996). List of animal species used in aquaculture. FAO Fisheries Circular 914.
- Gatz, A.J. (1979). Ecological morphology of freshwater stream fishes. *Tulane Studies in Zoology and Botanical*, 21, 91–124.
- George, R. (1995). Fat composition of free living and farmed sea species: implications for human diet and sea-farming techniques. *Britannica Food Journal*, 97, 19022.
- Gopakumar, K. (2002). Biochemical composition of fish. Textbook of fish processing technology. *Indian Council of Agricultural Research, New Delhi*, 18-30.
- Gopakumar, K. Indian food fishes: Biochemical composition. Central Institute of Fisheries Technology. Indian Council of Agricultural Research, Cochin, India, 22-28.
- Gopinath, B., Buyken, A. E., & Flood, V. M. (2011). Consumption of polyunsaturated fatty acids, fish, and nuts and risk of inflammatory disease mortality. *The American Journal of Clinical Nutrition*, 93(5):1073–9.
- Gupta, S. (2006). General and Applied Ichthyology: *Fish and Fisheries*. S. Chand. p. 993.
- Gustiano, R. (2004). Biomeric analysis of the artificial hybridization between *Pangasius djambal*.
- Haddon, M., & Willis, T. J. (1995). Morphometric and Meristic Comparison of Orange Roughy (*Hoplostethus atlanticus*, Trachichthyidae) From the Puysegur Bank and Lord-Howe- Rise, New-Zealand, and Its Implications For Stock Structure. *Marine Biology*, 123, 19–27.
- Haliloglu, H. I., Bayu, A., Surkeciglu, A. N., Aras, N. M., & Atamanalp, M. (2004). Comparison of fatty acid composition in some tissues of rainbow trout (*Onchorhynchus mykiss*) living in seawater and freshwater. *Food Chemistry*, 86: 55-59.
- Hamid, N. H., Hassan, M.D., Sabri, M.Y.M., Hasliza, A. H., Hamdan, R.H., Afifah, M.N.F., Raina, M. S., Nadia, A. B. S., & Fuad, M. M. (2017). Studies on Pathogenicity effect of *Aeromonas hydrophila* infection in juvenile red hybrid tilapia *Oreochromis* sp. *Proceedings of International Seminar on Livestock Production and Veterinary Technology*, 532-539.
- Hamid, N. H., Hassan, M. D., Md Sabri, M. Y., Hasliza, A. H., Hamdan, R. H., Afifah, M. N. F., Raina, M. S., Nadia, A. B. S., & Fuad, M. M. (2016). Studies on Pathogenicity Effect of *Aeromonas hydrophila* Infection in Juvenile Red Hybrid Tilapia *Oreochromis* sp. *Proceedings of*

- Haniffa, M. A. K., & Sridhar, S. (2002). Induced spawning of spotted murrel (*Channa punctatus*) and catfish (*Heteropneustes fossilis*) using human chorionic gonadotrophin and synthetic hormone (ovaprim). *Veterinary Archiv*, 72(1), 51-56.
- Hao, F., Zhuang, P., Song, C., Shu, Z. H., Zhang, L. Z. (2010). Amino acid and fatty acid composition and nutritional quality of muscle in the pomfret, *Pampus punctatissimus*. *Food chemistry*, 118(2), 224-227.
- Harikrisnan, R., & Balasundaram, C. (2005). Modern Trends in *Aeromonas hydrophila* disease management with fish. *Reviews in Fisheries Science*, 13(4): 281-320.
- Harris, W. S. (1997). N-3 fatty acids and serum lipoproteins, human studies. *American Journal of Clinical Nutrition*, 65, 16456-16545.
- Hassan, A., Azmi Ambak, M., & Samad, A. P. A. (2011). Crossbreeding of *Pangasianodon hypophthalmus* (Sauvage 1878) and *Pangasius nasutus* (Bleeker, 1863) and their larval development. *Journal of sustainability Science and Management*, 6(1), 28-35.
- Hecht, T., and Lublinkhof. (1985). *Clarias gariepinus* x *Heterobranchus longifilis* (Clariidae) Pieces: A new hybrid for aquaculture. *South Afr. J. Sci.*, 81:620-621.
- Hecht, T. (2013). A review of on-farm feed management practices for North African catfish (*Clarias gariepinus*) in sub-Saharan Africa – In: On-farm feeding and feed management in aquaculture (Eds) M.R. Hasan, M.B. New, FAO Fisheries and Aquaculture Technical Paper No. 583. Rome: 463-479.
- Hecht, T., Oellermann, L., & Verheust, L. (1996). Perspectives on clariid catfish culture in Africa – *Aquat. Living Resources* 9:197-206.
- Hibiya, T. (1982). An atlas of fish histology, normal and pathological features. Stuttgart (Germany): Fischer. P. 64-65.
- HMSO (Her Majesty's Stationery Office). (1994). Nutritional aspects of cardiovascular disease. Report on health and social subjects N 46. London, HMSO Department of Health, UK.
- Ho, B. T., & Paul, D.R. (2009). Fatty acid profile of Tra Catfish (*Pangasius hypophthalmus*) compared to Atlantic Salmon (*Salmo solar*) and Asian Seabass (*Lates calcarifer*). *International Food Research Journal*, 16: 501-506.
- Hogendoorn, H., and Vismans, M. M. (1980). Controlled propagation of the African catfish *Clarias lazera* (C and V). II. Artificial reproduction. *Aquaculture*, 21:39-53.



- Hosomi, R., Yoshida, M., & Fukunaga, K. (2012). Seafood consumption and components for health. *Global Journal of Health Science*, 4(3), 72-86.
- Ibhadon, S., Abdulsalami, M. S., Emere, M. C., & Yilwa, V. (2015). Comparative study of proximate, fatty and amino acids composition of wild and farm-raised African Catfish, *Clarias gariepinus* in Kaduna, Nigeria. *Pakistan Journal of Nutrition*, 14(1), 56-61.
- Idowu, T. A., Adedeji, H. A., & Sogbesan, O. A. (2017). Fish disease and health management in aquaculture production. *International Journal of Environment and Agriculture Science*, 1,1.
- Iwasaki, M., & Harada, R. (1985). Proximate and amino acid composition of the roe and muscle of selected marine species. *Journal of Food Science*, 50, 1586-1587.
- Jabeen, F., & Chaudhry, A.S. (2011). Chemical compositions and fatty acid profiles of three freshwater fish species. *Food Chemistry*, 125(3), 991-996.
- Jag Pal, B.N., Shukla, Ashish, K. M., Hari, O. V., Gayatri, P., & Amitha. (2018). A review on role of fish in human nutrition with special emphasis to essential fatty acid. *International Journal of Fisheries and Aquatic Studies*, 6(2), 427-430.
- Jakhar, J. K., Pal, A. K., Reddy, A. D., Sahu, N. P., Venkateshwarlu, G., & Vardia, H. K. (2012). Fatty acid composition of some selected Indian fishes. *African Journal of Basic & Applied Science*, 4(5), 155-160.
- Jana, R.K. 1993. Studies on some aspects of biology and cytogenetics of rohu-catla and catla-rohu hybrids. PhD Thesis, Department of Fisheries, University of Calcutta, 53 pp.
- Janda, J. M., & Abbott, S. L. (2010). Evolving concepts regarding the genus *Aeromonas*: an expanding panorama of species, disease presentations, and unanswered questions. *Clinical Infectious Disease*, 27, 332-334.
- Janda, J.M., & Abbott, S.L. (2010). The Genus *Aeromonas*: Taxonomy, Pathogenicity, and Infection. *Clinical Microbiology Review*, 23: 35-73.
- Jayavignesh, V., Sandesh, K. K., & Abhijith, D. B. (2011). Biochemical characterization and cytotoxicity of *Aeromonas hydrophila* isolated from catfish. *Scholar Research Library*, 3(3): 85-93.
- Job, B. E., Antai, E. E., Inyang-Etoh, A. P., Otogo, G.A., Ezekiel, H. S. (2015). Proximate composition and fatty acid profile in some commercially important fish species from lake Kainji, Nigeria. *International Journal Biology Pharmacology Applied Science*, 2, 849-856.

- Jothilakshmanan, N., & Karal Marx, K. (2013). Hybridization between Indian catfish, female *Heteropneustes fossilis* (Bloch) and Asian catfish, male *Clarias batrachus* (Linn). 12 (9), 976-981.
- Jiang, J. F., Han, X. Q., Fu, Z. R., & Xie, G. (2012). Comparative analysis of the main nutritional components in muscle and skin of male and female *Silurus asotus*. *Journal of Jimei University. Natural Science*, 17, 6-12.
- Juliano, R.O., Guerrero, R. III., Ronquillo, I.(1989). The introduction of exotic aquatic species in the Philippines. In: De Silva SS, ed. Proceedings of the Workshop on Introduction of Exotic Aquatic Organisms in Asia: The Asian Fisheries Society, 83-90.
- Kaya, Y., Turan, H., & Erdem, M. E. (2008). Fatty acid and amino acid composition of raw and hot smoked sturgeon (*Huso huso*, L. 1758). *International Journal of Food Science and Nutrition*, 59(7-8), 635-642.
- Kenari, A. A., Regenstein, J. M., Rezai, M., Tahergorabi, R., Nazari, R. M., Mogaddasi, M., & Kaboli, S. A. (2009). Amino acid and fatty acid composition of cultured Beluga (*Huso huso*) of different ages. *Journal of Aquatic Food Product and Technology*, 18, 245-265.
- Kessler, R.K., Casper, A.F., & Weddle, G.K. (1995). Temporal variation in microhabitat use and spatial relations in the benthic fish community of a stream. *Am. Midl. Nat.* 134:361 – 70.
- Khalil, A. H., & Mansour, E. H. (1997). Toxicity of crude extracellular products of *Aeromonas hydrophila* in tilapia, *Tilapia nilotica*. *Letters in Applied Microbiology*, 25: 269-272.
- Kiem, N. V. (2003). African catfish (*Clarias gariepinus*) and its role in fish composition in the Mekong Delta, Vietnam. Paper presented at the workshop "Role and Impacts of Exotic aquatic species in Vietnam". Cantho University, 20-21 May 2003.
- Kim, S. K., Yoon, S.C., Youn, S.H., Park, S.U., Corpus, L.S., & Jang, I.K. (2013). Morphometric changes in the cultured starry flounder, *Platichthys stellatus*, in openmarine ranching areas. *J Environ Biol.* 34(2):197-204.
- Klug, W.S., Cummings, M.R, Spencer, C.A. and Palladino, M.A. 2011. Concept of genetics. 10th ed. Pearson Benjamin Cummings, California.
- Koolboon, U., Koonawootrittriron, S., Kamolrat, W., & Na-nakorn, U. (2014). Effects of parental strains and heterosis of the hybrid between *Clarias macrocephalus* and *Clarias gariepinus*. *Aquaculture*, 424-425, 131-139.
- Kris-Etherton, P. M., Harris, W. S., & Appel, L. J. (2002). Fish consumption, fish oil, omega-3 fatty acids and cardiovascular disease. *Arterioscler Thromb. Vasc. Biology*, 23, 20-31.

- Kreuzer, Rudolf (1974). *Fishery Products*. Fishing News [for] the Food and Agriculture Organization. p. 271.
- Kumar, R. (2005). Fish as a food commodity: Biochemical composition of fish, National Association of Biology Teachers (NABT) Bulletin Board.
- Kumar, R., Pande, V., Singh, L., Sharma, L., & Saxena, N. (2016). Pathological findings of experimental *Aeromonas hydrophila* infection in golden mahseer (*Tor putitora*). *Journal of Fish Aquaculture*, 7: 160
- Kumaran, R., Ravi, V., Gunalan, B., Murugan, S., & Sundramanickam, A. (2012). Estimation of proximate, amino acids, fatty acids and mineral composition of mullet (*Mugil cephalus*) of Parangipettai, Southeast Coast of India. *Advances in Applied Science Research*, 3(4), 2015-2019.
- Laith, A. R., & Najiah, M. (2013). *Aeromonas hydrophila*: Antimicrobial susceptibility and histopathology of isolates from diseased catfish, *Clarias gariepinus* (Burchell). *Journal of Aquaculture Research and Development*, 5:215.
- Laleh, Y. G., Mohammad, E. J. Z., & Milad, A. (2015). The study on Effect of Temperature Stress on Occurrence of Clinical Signs Caused by *Aeromonas hydrophila* in *Capoeta damascina* in *In Vitro* Condition. *Advances in Animal and Veterinary Sciences*, 3(7): 406.
- Laoka, N., Juhel, F., Fazilleau, & Loonis, P. V. (2004). A novel colorimetry analysis used to compare different drying fish processes. *Food Control*, 15, 327-334.
- Lawonyawut, K. (1995). *Hybridization and genetic manipulation in Clarias catfish*, PhD thesis, University of Sterling, Scotland.
- Liem, P.T. 2008. Breeding performance and traits inheritance of hybrid catfish, *Clarias Macrocephalus* and *Clarias gariepinus* Ph.D dissertation, Universiti Malaysia Terengganu, Malaysia.
- Lee, S., Najiah, M., Wendy, W., & Nadirah, M. (2010). Antibigram and heavy metal resistance of pathogenic bacteria isolated from moribund cage cultured silver catfish (*Pangasius sutchi*) and red hybrid tilapia (*Tilapia* sp.). *Frontiers of Agriculture in China* 4(1), 116-120.
- Legendre, M, & Antoine, P. (1998). The biological diversity and aquaculture of clariid and pangasiid catfishes in South-East Asia: Proceedings of the mid-term workshop of the "Catfish Asia project". Jakarta (IDN); Can Tho: IRD; Can Tho.
- Legendre, M., Teugels, G.G., Cauty, C., & Jalabert, B. (1992). A comparative study on morphology, growth rate and reproduction of *Clarias gariepinus* (Burchell, 1822), *Heteropneustes longifilis* (Valenciennes, 1840) and their reciprocal hybrids (Pisces, Clariidae). *Journal of Fish Biology* 40:59-79.

- Legendre, M., & Antoine, P. (1998). The biological diversity and aquaculture of clariid and pangasiid catfishes in South-East Asia: Proceedings of the mid-term workshop of the "Catfish Asia project". Jakarta (IDN); Can Tho: IRD; Can Tho.
- Liao, I. C., & Chao, N. H. (2009). Aquaculture and food crisis: opportunities and constraints. *Asia Pacific Journal of Clinical Nutrition*, 18(4):564–9.
- Limin, L, Feng, X, Jing, H. (2006). Amino acids composition differences and nutritive evaluation of the muscle of five species of marine fish (common sea perch); (*Pagrosomus major* red seabream) *Seriola dumerili* (*Dumerili samber*) Jack and *Hepalogenys nitens* (blackgrunt) from Xiamen Bay of China. *Aquat. Nutr.* 12: 55:59.
- Lovell, R.T. (1989). *Nutrition and Feeding of Fish*. Van Nostrand Reinhold, New York. <http://dx.doi.org/10.1007/978-1-4757-1174-5>
- Martinez-Murcia, A., Beaz-Hidalgo, R., Svec, P. S., Figueras, M. J., & Sedlacek, I. (2013). *Aeromonas cavernicola* sp. nov., isolated from freshwater of a brook in a cavern. *Current Microbiology*, 66: 197-204.
- Martino, R.C., Cyrino, J.E., Portz, L., & Trugo, L.C. (2002). Performance and fatty acid composition of surubim (*Pseudoplatystoma coruscans*) fed diets with animal and plant lipids. *Aquaculture*, 209(1-4): 233- 246.
- Mateos, H. T., Lewandowski, P. A., Su, X. Q. (2011). Dietary fish oil supplements increase tissue n-3 fatty acid composition and expression of delta-6 desaturase and elongase-2 in Jade Tiger hybrid abalone. *Lipids*, 46, 741-751.
- Mathiesen, A.M. (2012). The State of the World Fisheries and Aquaculture 2012. Food and Agriculture Organization. Stearns, S.C. 1983. A natural experiment in life-history evolution: field data on the introduction of mosquitofish (*Gambusia affinis*) to Hawaii. *Evolution*, 37, 601-617.
- Matthews, W.J. (1988). *Patterns in Freshwater Fish Ecology*. Chapman & Hall.
- Mazumder, M. S. A., Rahman, M. M., Ahmed, A. T. A., Begum, M., & Hossain, M. A. (2008). Proximate composition of some small indigenous fish species (SIS) in Bangladesh. *Journal of International Sustainable Crop Production*, 3(4), 18-23.
- Megbowon, I., Fashina-Bombata, H. A., Akinwale, M. M. A., Hammed, A. M., Okunade, O. A., & Mojekwu, T.O. (2014). Breeding performance of v obtained from Nigerian waters. *Journal of Agriculture and Veterinary Science*, 6(3), 06-09.
- Mengumphan, K. and Panase, P. 2015. Morphometric and Meristic Divergence of Two Hybrid Catfish: Backcross (F1 hybrid female x *Pangasianodon gigas* Chevey 1931 male) and Reciprocal Backcross (*P. gigas*, female x F1 hybrid male). *Asian Fisheries Science* 28:37-46

- Mekawwy, I. A. A. and A. S. Mohammad. 2011. Morphometrics and meristics of the three Epinepheline species: *Cephalopholis argus* (Bloch and Schneider, 1801), *Cephalopholis miniata* (Forsskal, 1775) and *Variola louti* (Forsskal, 1775) from the red sea, Egypt. *Journal of Biological Science* 11:10-21.
- Men, L. T., Thanh, V. C., Hirata, Y., & Yamasaki, S. (2005). Evaluation of the genetic diversities and the nutritional values of the Tra (*Pangasius hypophthalmus*) and the Basa (*Pangasius bacourti*) catfish cultivated in the Mekong River Delta of Vietnam. *Journal of Asian-Australian Animal Science*, 18(5), 671-676.
- Miyazaki, T., & Kaige, N. (1985). A histopathological study on motile aeromonad disease in Crucian carp. *Fish pathology*, 21: 181-185.
- Monalisa, K., Islam, M. Z., Khan, T. A., Abdulla, A. T. M., & Hoque, M. M. (2013). Comparative study on nutrient contents of native and hybrid koi (*Anabas testudineus*) and Pangas (*Pangasius pangasius*, *Pangasius hypophthalmus*) fish in Bangladesh. *International Food Research Journal*, 20(2), 791-797.
- Mohammed, A., Asmar, Z., Abdul, R., Shahid, M., & Naureen, Q. (2011). Nutritional values of wild and cultivated silver carp (*Hypophthalmichthys molitrix*) and Crass (*Ctenopharygodon idella*). *International Journal of Agriculture and Biology*, 210-214.
- Mohanty, B. P., Ganguly, S., Mahanty, A., Sankar, T. V., Anandan, R., Chackraborty, K., Paul, B. N., Sarma, D., Dayal, J. S., Venkateshwarlu, G., Mathew, S., Asha, K.K., Karunakaran, D., Mitra, T., Banerjee, S., Chanda, S., Shahi, N., Das, P., Akhtar, M. S., Vijayagopal, P., & Sridhar, N. (2016). DHA, EPA content and fatty acid profile of 39 food fishes from India. *Journal of International Biomedical Research*, 1, 1-7.
- Mohanty, B. P., Ganguly, S., Mahanty, A., Sankar, T. V., Anandan, R., Chackraborty, K., Paul, B. N., Sarma, D., Dayal, J. S., Venkateshwarlu, G., Mathew, S., Asha, K.K., Karunakaran, D., Mitra, T., Banerjee, S., Chanda, S., Shahi, N., Das, P., Akhtar, M. S., Vijayagopal, P., & Sridhar, N. (2014). Amino acid composition of 27 food fishes and their importance in clinical nutrition. *Journal of Amino Acid*, 1, 1-7.
- Mohsin, A. K. M., & Ambak, M. A. (1983). Freshwater fishes of peninsular Malaysia. Universiti Pertanian Publishing House, Serdang, p.282.
- Mollah, M. F. and Khan, M. R. 1997. Comparative studies on growth and survival of *Clarias gariepinus*, *Clarias batrachus* and F1 hybrid fry under laboratory conditions. *Bangladesh J. Agricult. Sci.* 24, 17–20.
- Moreira, A. B., Visentainer, J. V., de Souza, N. E., & Matsushita, M. (2001). Fatty acids profile and cholesterol contents of three Brazilian Brycon freshwater fishes. *Journal of Food Composition and Analysis*, 14, 565-574.



- Morni, M.M. (2003). Study on cross breeding between Asian and African catfish (*Clarias macrocephalus* and *Clarias gariepinus*) and some aspects of the hybrid larvae development and rearing. Kolej University of Science and Technology Malaysia, Master of Science and Science Thesis, 137 pp.
- Morris, M. C., Evans, D. A., & Bienias, J. L. (2003). Consumption of fish and n-3 fatty acids and risk of incident Alzheimer disease. *Archives of Neurology*, 60(7):940–6.
- Mozaffarian, D, & Rimm, E. B. Fish intake, contaminants, and human health: evaluating the risks and the benefits. *The Journal of the American Medical Association*, 296(15):1885–99.
- Murta, A.G. (2000). Morphological variation of horse mackerel (*Trachurus trachurus*) in the Iberian and North Africa Atlantic: implications for stock identification. *ICES J. Mar. Science*, 57(4):1240-1248.
- Najiah, M., Lee, S. W., Wendy, W., & Nadirah, M. (2008). Bacterial diseases outbreak of African catfish (*Clarias gariepinus*) from Manir River, Terengganu, Malaysia. *Journal of Life Science*, 3(5), 10-13.
- Najiah, M., Lee, S. W., Wendy, W., & Nadirah, M. (2009). Bacterial disease outbreak of African catfish (*Clarias gariepinus*) from Manir River, Terengganu, Malaysia. *Journal of Life Science*, 3(5), 10-13.
- Na-Nakorn, U. (1999). Genetic factors in fish production: a case study of the catfish *Clarias*. In: *Mustafa, S. (Ed.), Genetics in Sustainable Fisheries Management*. Fishing News Books, London, pp. 175-187.
- Na-Nakorn, U., Kamonrat, W., & Ngamsiri, T. (2004). Genetic diversity of walking catfish, *Clarias macrocephalus*, in Thailand and evidence of genetic introgression from introduced farmed *C. gariepinus*. *Aquaculture*, 240(1-4):145-63.
- Nargis, A. (2006). Seasonal variation in the chemical composition of body flesh of koi fish *Anabas testudineus* (Bloch) (Anabantidae: Perciformes). *Bangladesh Journal Science Ind Research*. 41:219-26.
- Nazia, A. K., & Siti Azizah, M. N. (2014). Isolation of microsatellites in the bighead catfish, *Clarias macrocephalus* and cross-amplification in selected *Clarias* species. *Molecular Biology Rep.*, 41, 1207-1213.
- Ndimele, P. E., & Owodeinde, F. G. (2012). Comparative reproductive and growth performance of *Clarias gariepinus* (Burchell, 1822) and its hybrid induced with synthetic hormone and pituitary gland of *Clarias gariepinus*. Lagos, Nigeria. *Turkish Journal Fisheries and Aquaculture Science*, 12, 619-626.
- Nestel, P. J. N. (2000). Fish oil and cardiovascular disease lipids and arterial function. *Journal of Clinical Nutrition*, 228.

- Nelson, K. L., Raja, S. M., & Buckley, J. T. (1997). The glycol sylphosphatidy inositol anchored surface glycoprotein Thy-1 is a receptor for the channel-forming toxin Aerolysin. *Journal of Biological Chemistry*, 272: 12170-12174.
- Nielsen, M. E., Hoei, L., Schmidt, A. S., Qian, D., Shimada, T., Shen, J. Y., & Larsen, J. L. (2001). Is *Aeromonas hydrophila* the dominant motile *Aeromonas* species that cause disease outbreaks in aquaculture production in the Zhejiang Province of China? *Disease of aquatic Organisms*, 46: 23-29.
- Nlewadim A.A, Onuoh, G.C., & Aluko, P.O. (2004). Studies on the growth and survival of fry and fingerlings of clariid catfish species: *Clarias gariepinus* (Burchell, 1822), *Heterobranchus bidorsalis* (Geoffroy, 1809) and *H. longifilis* (Valenciennes, 1840). *Journal of Aquaculture Tropical*, 19 (1): 1-14
- Noga, E. J. (2000). Skin ulcers in fish: Pfiesteria and other etiologies. *Toxicologic Pathology*, 28:807-823.
- Noor El Deen, A. E., Sohad, Dorgham, M., Azza, H. M. H., & Hakim, A.S. (2014). Studies on *Aeromonas hydrophila* in Cultured *Oreochromis niloticus* at Kafr El Sheikh Governorate, Egypt with Reference to Histopathological Alterations in Some Vital Organs. *World Journal of Fish and Marine Sciences*, 6(3), 233-240.
- Nukwan, S., Tangtrongpiros, M., Lawanyawut, K., Veerasidith, P. (1990). Crossbreeding between *Clarias macrocephalus* and *Clarias gariepinus*. Proceeding of the 28th Kasetsart University Conference, Fisheries Section, Bangkok. 29-31 January 1990. Pp. 553-267 (In Thailand).
- Nurnadia, A. A., Azrina, A., & Amin. (2011). Proximate composition and energetic value of selected marine fish and shellfish from the West coast of Peninsular Malaysia. *International Food Research*, 1(18), 137-148.
- Nwadukwe, F.O. (1995). Hatchery propagation of five hybrid groups by artificial hybridization of *Clarias gariepinus* and *Heterobranchus longifilis* (Clariidae) using dry powdered carp pituitary hormone. *Journal Aquaculture Tropical*, 10: 1-11.
- Ochokwu, I. J., 1Bichi, A. H., & Onyia, L.U. (2016). Intra-specific Hybridization between Two Strains of *Clarias gariepinus* from SouthWest and North Western Nigeria. Nigerian. *Journal of Fisheries and Aquaculture*, 4(1): 34 – 41.
- Okeyo, D.O.(2003). On the biodiversity and the distribution of freshwater fish of Namibia: an annotated update. p.156-194. In M.L.D. Palomares, B. Samb, T. Diouf, J.M. Vakily and D. Pauly (eds.) Fish biodiversity: local studies as basis for global inferences. ACP-EU Fish. Res. Rep. 14:281 p.

- Okomoda, V. T., Koh, I. C. C., & Shahreza, M. S. (2018). First report on the successful hybridization of *Pangasianodon hypophthalmus* (Sauvage, 1878) and *Clarias gariepinus* (Burchell, 1822). *Zygote* 25: 443–452.
- Olagunju, A., Mohammad, A., Mada, S. B., Mohammed, A., Mohammed, H. A., & Mahmoud, K. T. (2012). Nutrient composition of *Tilapia zilli*, *Hemisyndontis membranacea*, *Clupea harengus* and *Scomberscombrus* consumed in Zaria. *World Journal of Life Science and Medical Research*, 2, 16.
- Olaniyi, W. A., & Omitogun, O. G. (2012). Induction of diploid gynogeneic larvae of African catfish, *Clarias gariepinus* Burchell (1822). *Life Journal Agriculture*, 25, 73-82.
- Oliveira, S. R., Souza, T. Y. B., Brasil, E. M., Andrade, J.I.A., Nunes, E.S.S. Nono, E. A., & Affonso, E. G. (2011). LD50 of the bacteria *Aeromonas hydrophila* to matrinxa (*Brycon amazonicus*), *Acta Amazonica*, 41(2).
- Olufeagba, O., & Okomoda, V.T. (2015). Preliminary Report on Genetic Improvement of *Heterobranchus longifilis* through Intraspecific Hybridization of Different strains from Nigeria. *Journal of Aquaculture engineering and fisheries research*, 1(1), 45-48
- Olufeagba, S. O., Okomoda, V. T., & Shaibu, G. (2016). Embryogenesis and early growth of pure strains and hybrids between *Clarias gariepinus* (Burchell, 1822) and *Heterobranchus longifilis* Valenciennes, 1840. *North Am. J. Aquaculture*, 78, 346-355.
- Olufeagba, S. O., & Okomoda, V. T. (2016). Cannibalism and performance evaluation of hybrids between *Clarias batrachus* and *Clarias gariepinus*. *Croatian J. Fish*, 74, 124-129.
- Omeji, S., Obande, R. A., & Oyaje, J. (2013). Intra-specific hybridization of local and exotic *Clarias gariepinus*, *IJMBR*, 1, 35-41.
- Onyia, L. U., Milam, C, Manu, J. M., & Allison, D. S. (2010). Proximate and mineral composition in some freshwater fishes in upper river Benue, Yola, Nigeria. *Journal of Food Science and Technology*, 4, 1-6.
- O'Reilly, K.M., & Horn, M.H., (2004). Phenotypic variation among populations of *Atherinops affinis* (Atherinopsidae) with insights from a geometric morphometric analysis. *J. Fish Biol.* 64, 1117–1135.
- Osibona, A. O. (2011). Comparative study of proximate composition, amino and fatty acids of some economically important fish species in Lagos, Nigeria. *Journal of African Food Science*, 5, 581.
- Ozogul, Y., & Ozogul, F. (2007). Fatty acid profiles of commercially important fish species from the Mediterranean, Aegean and Black Seas. *Food Chemistry*, 100: 1634-1638.



- Olakunle, O.M. (2012). Chemical analysis of flesh and some body parts of different fresh fish south west Nigeria. *Pakistan Journal of Nutrition*, 11(1), 14-15.
- Oluwaniyi, O., & Dosumu, O.O. (2009). Preliminary studies on the effect of processing methods on the quality of three commonly consumed marine fishes in Nigeria. *Biochemistry*, 21(1), 1-7.
- Osibona, A. O., Kusemiju, K., & Akande, G. R., (2006). Proximate composition and fatty acids profile of the African catfish *Clarias gariepinus*. *Acta SATECH*, 3(1): In Press.
- Padmakumar, K.G., Bindu, L., Sreerekha, P.S., Gopalakrishnan, A., Basheer, V.S., Nitta Joseph, P. S. Manu & Anuradha Krishnan (2011). Current Science, 100(25), 1232-1236
- Paiboon, B., & Kittichon, U. (2016). Growth performance and disease resistance against *Vibrio vulnificus* infection of novel hybrid grouper (*Epinephelus lanceolatus* x *Epinephelus fuscoguttatus*). *Journal of Aquaculture Research*, 1-13.
- Palumbo, S. A., Bencivengo, M. M., Corral, F., Williams, A. C., Buchanan, R. L., & Del-Corral, F. (1989). Characterization of *Aeromonas hydrophila* group isolated from retail foods of animal origin. *Journal of Clinical Microbiology*, 27: 854-859.
- Pandey, A.K., Koteeswaran, R. & Singh, B.N. (2002). *Aquaculture*, 3 :137–142.
- Paul, B. N., Chanda, S., Shridhar, N., Saha, G. S., & Giri, S. S. (2016). Proximate, mineral and vitamin contents of Indian Major Carp. *Indian Journal of Animal Nutrition*, 32(2), 221-226.
- Park, I. S., Nam, Y. K., Douglas, S. E., Johnson, S., & Kim, D. S. (2003). Genetic characterisation, morphometrics and gonad development of induced interspecific hybrids between yellow tail flounder, *Pleuronectes ferrugineus*, and winter flounder. *Pleuronectes americanus*. *Aquacult. Res.* 34, 389–396
- Parker, M. W., Buckley, J. T., Postma, J. P., Tucker, A. D., Leonard, K., Pattus, F., & Tsernoglou, D. (1994). Structure of *Aeromonas* toxin proaerolysin in its water-soluble and membrane-channel states. *Nature*, 367(6460), 292-295.
- Payne, S. A., Johnson, B. A., Otto, R. S. (1999). Proximate composition of some north-eastern Pacific forage fish species. *Fish Oceanography*, 8(3), 159-177.
- Perea, Gómez, A., E., Mayorga, Y., & Triana, C.Y. (2008). Caracterización nutricional de pescados de producción y consumo regional en Bucaramanga, Colombia. *Archivos Latinoamericanos de Nutrición*, 58: 91-97.

- Pervin, T., Yeasmin, R., Islam, R., Kamruzzaman, A., Rahman, A., & Sattar, A. (2012). Studies on nutritional composition and characterization of lipids of *Lates calcarifer* (Bhetki). *Bangladesh Journal of Science India Research*, 47(4), 393-400.
- Poompuang, S., & Na-Nakorn, U. (2004). A preliminary genetic map of walking catfish (*Clarias macrocephalus*). *Aquaculture*, 232:195-203
- Popoff, M., & Vernon, M. (1976). *Journal of General Microbiology*, 94, 11-22.
- Pouyaud, L., & Sudarto Paradis, E. (2009). The phylogenetic structure of habitat shift and morphological convergence in Asian *Clarias* (Teleostei, Siluriformes: Clariidae). *Journal of Zoology Systematic Evolution Research* 47:344-356.
- Ravichandran, S., Kumaravel, K., & Pamela Florence, E. (2011). Nutritive composition of some edible fin fishes. *International Journal of Zoological Research*, 7(3), 241-251.
- Rahayu, K., Kismiyati, Sudarno, Hendi, K., & Yudha, T. P. (2017). Isolation and Identification of *Aeromonas hydrophila* and *Saprolegnia* sp. on Catfish (*Clarias gariepinus*) in Floating cages in Bozem Moro Krembangan Surabaya. *IOP Conference Series: Earth and Environmental Science*, 55.
- Rahman, M. A., Arshad, A., Marimuthu, K., Ara, R., & Amin, S. M.N. (2012). Inter-specific hybridization and its potential for aquaculture of fin fishes, Selangor, Malaysia. *Asian Journal Animal Veterinary Advances*, 8(2), 139-153.
- Rahman, M. H., Suzuki, S., & Kawai, K. (2001). The effect of temperature on *Aeromonas hydrophila* infection in goldfish, *Carassius auratus*. *Journal of Applied Ichthyology*, 17: 282-285.
- Rahman, M.A., Bhadra, A., Begum, N., Islam, M.S., & Hussain M.G. (1995). Production of hybrid vigor through cross breeding between *Clarias batrachus* Lin. and *Clarias gariepinus* Burchel, 125-130.
- Rao, G.R.M., Tripathi, S.D., & Sahu, A.K., (1994). Breeding and seed production of the Asian catfish *Clarias batrachus* (Lin.). In: Manual Series 3. A publication of CIFA, Kausalyaganga, India. pp. 5
- Reddy, P. V. G. K. (2000). Genetics resources of Indian major carps. Optimization of feminisation protocol. *Aquaculture*, 89, 329-339.
- Rey, A., Verjan, N., Ferguson, H. W., & Iregui, C. (2009). Pathogenesis of *Aeromonas hydrophila* strain KJ99 infection and its extracellular products in two species of fish. *Veterinary Record*, 164: 493-499.
- Reyes-Becerril, M., Lopez-Medina, T., Ascencio-Valle, F., & Esteban, M.A. (2011). Immune response of gilthead seabream (*Sparus aurata*)

- following experimental infection with *Aeromonas hydrophila*. *Fish Shellfish Immun*, 31: 564-570.
- Rimler, R., & Shoots, E.B. (1973). Medium for the isolation of *Aeromonas hydrophila*. *Applied Microbiology*, 26: 550-553.
- Roberts, R. J. (2001). *Aeromonas hydrophila* in fish pathology. 3<sup>rd</sup> Edition, pp: 3.
- Ruiz-Capillas, C., & Moral, A. (2004). Free amino acids in muscle of Norway lobster (*Nephrops norvegicus* L.) in controlled and modified atmospheres during chilled storage. *Food Chemistry*, 86, 85-91.
- Sahu, S., Das, B.K., Pradhan, J., Mohapatra, B.C., Mishra, B.K. (2007). Effect of *Mangifera indica* kernel as a feed additive on immunity and resistance to *Aeromonas hydrophila* in Labeo rohita fingerlings. *Fish Shellfish Immun*, 23, 109-118.
- Sahoo, S. K., Giri, S. S., Sahu, A. K., & Ayyappan, S. (2003). Experimental hybridization between catfish *Clarias batrachus* (Linn.) x *Clarias gariepinus* (Bur.) and performance of the offspring in rearing operations. *Asian Fisheries Science*, 16, 157-166.
- Sahoo, P. K., Meher, P. K., Mahapatra, K. D., Saha, J. N., Jana, R. K., & Reddy, P. V. (2004). Immune responses in different fullsib families of Indian major carp (*Labeo rohita*), exhibiting differential resistance to *Aeromonas hydrophila* infection. *Aquaculture*, 238(1-4), 115-125.
- Sahoo, S.K., Giri, S.S., Chandra, S., & Sahu, A. K. (2007). Spawning performance and egg quality of Asian catfish *Clarias batrachus* at various doses of HCG injection and latency periods during spawning induction. *Aquaculture*, 266: 289-292.
- Sahin, S. A., Nadin, B., Mehmet, K., Bekir, T., Sevim, K., & Ibrahim, O. (2011). Evaluation of meat yield, proximate composition and fatty acid profile of cultured brook trout (*Salvelinus fontinalis* Mitchell, 1814) and Black Sea Trout (*Salmo trutta labrax* Pallas, 1811) in comparison with their hybrid. *Turkish Journal of fisheries and Aquatic Sciences*, 11: 261-271.
- Sankar, T.V., & Ramachandran, A. (2001). Changes in biochemical composition in Indian major carps in relation to size. *Fishery Technology*, 38:22-27.
- Santour, M., Mary, P., Chihib, N. E., & Hornez, J. P. (2003). The effects of temperature, water activity and pH on the growth of *Aeromonas hydrophila* and its subsequent survival in microcosm water. *Journal of Applied Microbiology*, 95: 807-813.
- Satio, H., Yamashiro, R., Alasalvar, C., & Konno, T. (1999). Influence of diet on fatty acids of three subtropical fish, subfamily caesioninae (*Casio digrumna* and *C. tile*). And family siganidae (*Siganus canaliculatus*). *Lipids*, 34(10), 1073-1082.

- Scribner, K.T., Page, K.S., & Bartron, M.L. (2000). Hybridization in freshwater species: a review of case studies and cytonuclear methods of biological inference *Rev Fish Biology Fish* 10:293-323.
- Seegers, L.(2008). The catfishes of Africa: A handbook for identification and maintenance. Aqualog Verlag A.C.S. GmbH, Germany. 604 p.
- Šegvic-Bubic, T., Talijancic, I., Grubisic, L., Izquierdo- Gomez, D., & Katavic, I. (2014). Morphological and Molecular differentiation of wild and farmed gilthead sea bream *Sparus aurata*: implications for management. *Aquac Environ Interact*, 6(1):43-54.
- Shady, M. E.S., Ali, A. G. A., & Hamed, M. A., M. (2016). Amino acids pattern and fatty acids composition of the most important fish species of Saudi Arabia. *International Journal of Food Science and Nutrition Engineering*, 6(2), 32-41.
- Shahi, N., Mallik, S. K., Sahoo, M., & Das, P. (2013). Biological characteristics and pathogenicity of a virulent *Aeromonas hydrophila* associated with ulcerative syndrome in farmed rainbow trout, *Onchorhynchus mykiss* (Walbaum), in India. *Israeli Journal of Aquaculture*, 39:11.
- Shankar Murthy, K., Kiran, B. R., & Venkateshwarlu, M. (2013). A review on induced breeding of catfishes, murrels and climbing perches in India. *Advances in Applied Science Research*, 4(4), 310-323.
- Sharma, K., Yadava, N.K., & Jindal, M. (2010). Livestock Research for Rural Development, 22 (4).
- Sidhu, K. S. (2003). Health benefits and potential risks related to consumption of fish or fish oil. *Regul Toxicology Pharmacology*, 38, 336-344.
- Senanan, W., Kapuscinski, A.R., Na-Nakorn, U., & Miller, L.M. (2004) Genetic impacts of hybrid catfish farming (*Clarias macrocephalus* x *C. gariepinus*) on native catfish populations in central Thailand. *Aquaculture*, 235(1-4):167-84.
- Simon, K.D., Bakar, Y. Temple, S.E. and Mazlan, A.G. 2010. Morphometric and meristic variation in two congeneric archer fishes *Toxotes chatareus* (Hamilton, 1822) and *Toxotes jaculatrix* (Pallas 1767) inhabiting Malaysian coastal waters. *Journal of Zhejiang Univ-Sci B (Biomed & Biotechnol)* 11(11):871-879.
- Simopoulos, A. P. (2002). Omega-3 fatty acids in inflammation and autoimmune diseases. *Journal of the American College of Nutrition*, 21(6), 495-505.
- Simopoulos, M. D. (2004). Omega-6/Omega-3 essential fatty acid ratio and chronic diseases. *Food Reviews International*, 20(1), 77-90.
- Silva, B. C., Mourino, J. L. P., Vieira, F. N., Jatoba, A., Seiffert, W. Q., & Martins, M. L. (2012). Haemorrhagic septicaemia in the hybrid surubim

- (*Pseudoplatystoma corruscans* x *Pseudoplatystoma fasciatum*) caused by *Aeromonas hydrophila*. *Aquaculture Research*, 43(6), 908-916.
- Silva, A. (2003). Morphometric variation among sardine (*Sardina pilchardus*) populations from the northeastern Atlantic and the western Mediterranean. *ICES J. Mar. Sci.* 60: 1352 -1360.
- Skelton, P. (1993). A Complete Guide to the Freshwater Fishes of Southern Africa. Southern Book Publishers (Pty) Ltd., Halfway House.
- Skelton, P. 2001. A complete guide to the freshwater fishes of Southern Africa. Struik Publishers, Cape Town, 1–262.
- Snieszko, S. F. (1978). Control of fish diseases. *Marine Fisheries Review*, 40: 65-68.
- Steffens, W. (1997). Effects of variation in essential fatty acids in fish feeds on nutritive value of freshwater fish for humans. *Aquaculture*, 151,97-119.
- Srivastava, P.P, Raizada, S., Dayal, R., Chowdhary, S., &Lakra, W.S. (2012). Breeding and larval rearing of Asian Catfish, *Clarias batrachus* (Linn) on live and artificial feed. *Journal Aquaculture Research Development*, 3: 134.
- Sunita, G., Ram, C. S., & Rajender, K. G. (2015). Effect of pond fertilization with vermicompost and Some other manures on the pathogenic bacterial populations of treated waters. *Research journal of Microbiology*, 10(6), 230-245.
- Sun, S., Zhu, J., Jiang, X., Li, B., & Ge, X. (2014). Molecular cloning, tissue distribution and expression analysis of a manganese superoxide dismutase in blunt snout bream *Megalobrama amblycephala*. *Journal of Fish and Shellfish Immunology*, 38: 340-347.
- Solomon, S. G., Okomoda, V. T., &Ogbenyikwu, A. I. (2015). Intraspecific morphological variation between cultured and wild *Clarias gariepinus* (Burchell) (Clariidae, Siluriformes). *Arch. Polish Fish.* 23, 53–61.
- Somnuek, C., Boonphakdee, C., Cheevaporn, V., &Tanaka, K. (2009). Gene expression of acetylcholinesterase in hybrid catfish (*Clarias gariepinus* x *Clarias macrocephalus*) exposed to chlorpyrifos and carbaryl. *Journal of Environmental and Biology*, 30(1):83–88.
- Smolin, L.A., & Grosvenor, M. B. (2003). Nutrition: Science and applications, 4th Edition, USA: John Wiley and Sons Inc.
- Stansby, M. E. (1985). Fish or fish oil in the diet and heart attack. *Fish Review*, 46(2), 60-63.
- Strauss R.E. (1985). Evolutionary allometry and variation in body form in the South American catfish genus *Corydoras* Callichthyidae. *Syst. Biol.* 34, 381-396.



- Swain, D.P., Ridell, B.E. and Murray, C.B. 1991: Morphological differences between hatchery and wild populations of coho salmon (*Oncorhynchus kisutch*): environmental versus genetic origin. *Can. Journal of Fish. Aquaculture Science* 48(9): 1783–1791.
- Swapna, H. C., Kumar, R. A., Bhaskar, N., & Sachindra, N. M. (2010). Lipid classes and fatty acid profile of selected Indian freshwater fishes. *Journal of Food Science and Technology*, 47(4), 394-400.
- Szymanski, K. M., Wheeler, D. C., & Mucci, L. A. (2010). Fish consumption and prostate cancer risk: a review and meta-analysis. *The American Journal of Clinical Nutrition*, 92(5):1223–33.
- Tarnchalanukit, W. (1995). Experimental hybridization between catfish of the families Clariidae and Pangasiidae in Thailand., Kasetsart University, Fishery Research Buletin Number 16. P.8.
- Tao, N. P., Wang, L. Y., Gong, X., & Liu, Y. (2012). Comparison of nutritional composition of farmed pufferfish muscles among *Fugu obscurus*, *Fugu flavidus*, and *Fugu rubripes*. *Journal of Food Composition and Analysis*, 28, 40-45.
- Taufik, P. (1984). Faktor Kualitas Air Dapat Mempengaruhi Timbulnya Suatu Penyakit Pada Ikan. Majalah Pertanian No. 3. Departemen Pertanian. Jakarta.
- Tawfik, M.S. (2009). Proximate Composition and fatty acids profiles in most common available fish species in Saudi market. *Asian. J. Clin. Nutr.* 1(1): 50-57
- Tave, D. 1993. Genetics for Fish Hatchery Managers. 2nd ed. Van Nostrand Reinhold New York 415
- Teugels, G.G.(1986). A systematic revision of the African species of the genus *Clarias* (Pisces; Clariidae). *Ann. Mus. R. Afr. Centr., Sci. Zool.*, 247:199 p.
- Teugels, G.G., Legendre, M., & Le, T.H. (1998). Preliminary results on the morphological characterisation of natural populations and cultured strains of *Clarias* species (Siluriformes, Clariidae) from Vietnam. Proceedings of the mid-term workshop of the “Catfish Asia project (1):27-30.
- Teugels, G. G. (1986). A systematic revision of the African species of the genus *Clarias* (Pisces: Clariidae). *Annales du Musée Royal de l'Afrique Centrale* 247, 199 p.
- Thammapat, P., Raviyan, P., & Siriamornpun, S. (2010). Proximate and fatty acids composition of the muscles and viscera of Asian catfish (*Pangasius bocourti*). *Food chemistry*, 122:223-227.

- Thomas, J. Madan, N., Nambi, K. S. S., Majeed, S. A., Basha, A. N., & Hameed, S.S. (2013). Studies on ulcerative disease caused by *Aeromonas caviae*-like bacterium in Indian catfish, *Clarias batrachus* (Linn). *Aquaculture*, 376-379: 146-150.
- Tilahun, G., Kiran, D., Chtruvedi, C. S., & Bindhi, K. (2016). Assessment of Reproductive Performance, Growth and Survival of Hybrids of African Catfish (*Clarias gariepinus*) and Indian Catfish (*Clarias batrachus*) Compared to Their Parental Lines Crosses. *Turkish Journal of Fisheries and aquatic Science*, 16, 123-133.
- Topic Popovic, N., Coz-Racovac, R., & Strunjak-Perovic, I. (2007). Commercial phenotypic tests (API 20E) in diagnosis of fish bacteria: a review. *Veterinarni Medicina*, 52(2): 49-53.
- Tran, N.T., Gao, Z.X., Zhao, H.H., Yi, S.K., & Chen, B.X. (2015) Transcriptome analysis and microsatellite discovery in the blunt snout bream (*Megalobrama amblycephala*) after challenge with *Aeromonas hydrophila*. *Fish Shellfish Immun*, 45: 72-82.
- Turan, C., Yalcin, S., Turan, F., Okur, E., Akyurt, I.I., & Yalcin, S. (2005). Morphometric comparisons of African catfish, *Clarias gariepinus*, populations in Turkey Africa (Lond) 54:165 72.
- Turan, C. (2004). Stock identification of Mediterranean horse mackerel (*Trachurus mediterraneus*) using morphometric and meristic characters. *Journal Marine Sciences*, 61(5):774-81.
- Koolboon, U., Koonawootrittriron, S., Kamolrat, W., Na-Nakorn, U. (2014). Effects of parental strains and heterosis of the hybrid between *Clarias macrocephalus* and *Clarias gariepinus*. *Journal of Aquaculture*, 131-139.
- Vaccaro, A.M., Buffa, G., Messina, C. M., Santulli, A., & Mazzola, A. (2008). Fatty acid composition of a cultured sturgeon hybrid (*Acipenser naccarii* x *A. baeri*). *Food Chemistry*, 93, 627-631.
- Vidthayanon, C., & Allen, D. (2013). *Clarias macrocephalus*. The IUCN Red List of Threatened Species.
- Vieira, V.A., Hilsdorf, A.W., Moreira, R.G. (2012). The fatty acid profiles and energetic substrates of two Nile tilapia (*Oreochromis niloticus*, Linnaeus) strains, Red-Stirling and Chitralada, and their hybrid. *Aquaculture Research*, 43:565-76.
- Viveen, W.J., Richter, C.J., Janssen, J.A., Van Oordt, P.G., and Huisman, E.A. 1986. Practical manual for the culture of the African catfish (*Clarias gariepinus*). Department of Fish Culture and Fisheries of the Agricultural University of Wageningen, the Netherlands, 121pp.
- Wang, S., Su, Y., Ding, S., Cai, Y., & Wang, J. (2011). Cytogenetic analysis of orange-spotted grouper, *Epinephelus coioides*, using chromosom banding and fluorescence in situ hybridization. *Hydrobiologia*, 638(1), 1.



- Wang, Y., Shiliang, Y., Gunjum, M., Songho, C., Ye, S., & Yuhong, Y. (2014). Comparative study of proximate composition and amino acid in farmed and wild *Pseudobagrus ussuriensis* muscles. *International Journal of Food Science and Technology*, 49, 983-989.
- Wang, L., Wu, J., Wang, C., Li, J., Zhao, Z., Luo, L., Du, X., & Xu, Q. (2016). Dietary arginine requirement of juvenile hybrid sturgeon (*Acipenser schrenckii* x *Acipenser baerii*). *Aquaculture Research*, 1-9.
- Wangcharoen, W., Kriangsak, M., & Doungporn, A. (2015). Fatty acid composition, physical properties, acute oral toxicity and antioxidant activity of crude lipids from adipose tissue of some commercialized freshwater catfish. *Chiang Mai Journal of Science*, 42(3), 626-636.
- Weber, J., Bochi, V.C., Ribeiro, C.P., Victorio, A., & Emanuelli, T. (2008). Effect of different cooking methods on the oxidation, proximate and fatty acid composition of silver catfish (*Rhamdia quelen*) fillets. *Food Chemistry*, 106(1): 140-146.
- Welcomme, R.L. (1988). International introductions of inland aquatic species. FAO Fisheries Technical Paper, No. 294, pp: 318
- Whitman, K. A., & MacNair, N. G. (2004). Finfish and shellfish bacteriology: Manual Techniques and Procedures. USA: Iowa State Press.
- Wimberger, P.H. (1992). Plasticity of fish body shape – the effects of diet, development, family and age in two species of *Geophagus* (Pisces: Cichlidae). *Biol. J. Linn. Soc.* 45: 197–218.
- Witte, M.B., Thorton, F.J., Tantry, U., & Barbul, A. (2002). L Arginine supplementation enhances diabetic wound healing, involvement of the nitric oxide synthesis and arginase pathways. *Metabolism*, 51 (10): 1269-1273.
- Witono, Y., Taruna, I., & Windrat, W. (2014). Amino acids profiles and chemical properties of four inferior sea fishes in Madura Indonesia. *International Journal of Chemistry and Technology Research*, 6(1), 311-315.
- Wolker, R. E. G., George, C. J., & Blazer, V. S. (1985). Pigmented macrophage accumulations (MMC, PMB): Possible monitors of fish health. In: Hargis WJ, editor. NOAA Technical Report NMFS 25. Washington DC (USA): NOAA. P, 93-97.
- Wolters, W. R., Wise, D. J., & Klesius, P. H. (1996). Survival and antibody response of channel catfish, blue catfish, and channel catfish female x blue catfish male hybrids after exposure to *Edwardsiella ictaluri*. *J. Aquat. Anim. Health*, 8, 249–254.
- Wringe, B.F., & Fleming, I.A. (2015). Purchase CF. Rapid morphological divergence of cultured cod of the northwest Atlantic from their source population. *Aquac Environ Interact*, 9(167-177).

- Xu, D., & Klesius, P. H. (2013). Comparison of serum antibody responses and host protection against parasite *Ichthyophthirius multifiliis* between channel catfish and channel × blue hybrid catfish. *Fish Shellfish Immunol.* 34, 1356–1359.
- Yardimci, B., & Aydin, Y. (2011). Pathological findings of experimental *Aeromonas hydrophila* infection in Nile tilapia (*Oreochromis niloticus*). *Ankara University Veterinary Faculty Derg.*, 58: 47-64.
- Yu, H. B., Zhang, Y. L., Lau, F., Yao, S., Vilches, S., Merino, S., J. M., Tomas, S.P., Howard, S. P., & Leung, K.Y. (2005). Identification and characterization of putative virulence genes and gene clusters in *Aeromonas hydrophila*. *Journal of Applied Environmental Microbiology*, 71(8), 4469-4477.
- Zenebe, T. (2010). Diet composition impacts the fatty acid contents of Nile Tilapia, *Oreochromis niloticus* L, in Ethiopian highland lakes. *Verh Internat Verein Limnol.* 30: 1363-1368.
- Zonneveld, N., Rustidja, W.J.A., Viven, & Maduna, W. (1988). Induced spawning and egg incubation of the Asian Catfish, *Clarias batrachus*. *Aquaculture*, 74:41-47.