



**UNIVERSITI PUTRA MALAYSIA**

***MOLECULAR CHARACTERIZATION OF NADPH OXIDASE GENES  
AND EXPRESSION OF MarbohB1 GENE IN RESPONSE TO FUNGAL  
PATHOGEN *Fusarium oxysporum* f. sp. *ubense* IN BANANA***

**CHAI SIEW YING**

**FBSB 2020 24**



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By

CHAI SIEW YING

**Thesis Submitted to the School of Graduate Studies, Universiti Putra  
Malaysia, in Fulfilment of the Requirements for the Master of Science**

**July 2019**

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master Science

**MOLECULAR CHARACTERIZATION OF NADPH OXIDASE GENES AND EXPRESSION OF *MarbohB1* GENE IN RESPONSE TO FUNGAL PATHOGEN *Fusarium oxysporum* f. sp. *ubense* IN BANANA**

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July 2019

**Chair : Noor Baity binti Saidi, PhD**  
**Faculty : Biotechnology and Biomolecular Sciences**

Plant NADPH oxidases or also known as respiratory burst oxidase homolog (*rboh*) catalyzes the production of reactive oxygen species (ROS) which play crucial roles in plant development, hormone signalling and defense reactions. ROS production via plasma membrane-localized RBOHs is one of the earliest responses during pathogen infection in plants. *Fusarium* wilt is recognized as one of the most destructive banana diseases in the world that is caused by *Fusarium oxysporum* f. sp. *ubense* (*Foc*), a soil-borne fungus. Despite numerous reports on the involvement of ROS in *Foc*-banana interaction, *rboh* genes have not yet been identified in banana thus hindering the profiling of *Marbohs* expression in response to *Fusarium* wilt infection. The aim of this study is to characterize *rbohs* in banana (*Marbohs*) and identify defense-related *Marbohs* through genome-wide and gene expression analysis in relation with hydrogen peroxide and electrolyte leakage level. In this study, we have identified nineteen *Marbohs* distributed on nine chromosomes of DH Pahang through database search. The functional domain organization of *Marbohs* include respiratory burst NADPH oxidase, EF-hand calcium binding domain (EFh), EF-hand domain pair (EFh-7), Nox/Duox transmembrane protein and NAD binding domain (NAD binding 6) are important for ROS production. Phylogenetic analysis clustered *Marbohs* into four subgroups and had a closer relationship with those from *Manihot esculenta*, *Arabidopsis thaliana* and *Brassica rapa*. Among all *Marbohs*, only *MarbohB1* is located in subgroup III where most of the *rbohs* are related with plant defense response against pathogen. Based on the digital gene expression analysis of *Marboh* transcripts, *MarbohB1* was significantly downregulated at 48 hpi and 96 hpi following *Fusarium oxysporum* f. sp. *ubense* Tropical Race 4 (*Foc*TR4) infection. Based on the phylogenetic and digital gene expression analysis, *MarbohB1* was chosen for further analysis in this study. The full length of *MarbohB1* (*MabrbohB1*) was isolated via primer walking. The amino

acid alignment of DH Pahang *MarbohB1* and Berangan *MabrbohB1* revealed 13 single nucleotide polymorphisms (SNPs) which caused 6 nonsynonymous and 6 synonymous amino acid changes in which 4 nonsynonymous occurred in conserve domains. The organ-specific and temporal expression analysis of *MabrbohB1* upon hemibiotrophic fungal pathogen *FocTR4* infection showed that *MabrbohB1* was specifically expressed in root and exhibited a transient up-regulation after 2 hpi of *FocTR4* inoculation, followed by down regulation at 48 hpi and 96 hpi. The increased expression was accompanied by steady increment of H<sub>2</sub>O<sub>2</sub> level and electrolyte leakage in the infected root, suggesting that *MabrbohB1* may be involved in defense responses in banana. Characterization of *Marbohs* and identification of *MarbohB1* as defense-related gene add new knowledge about *FocTR4*-banana interaction and might be useful to counter fusarium wilt infection in the future. Overall, this research supports the suggested role of *rboh* genes in oxidative burst during plant response to pathogen.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk ijazah Master Sains

**PENCIRIAN MOLEKUL GEN-GEN *NADPH* OKSIDASE DAN EKSPRESI  
*MarbohB1* DALAM TINDAK BALAS TERHADAP PATOGEN KULAT  
*Fusarium oxysporum* f. sp. *cubense* DALAM PISANG**

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*NADPH* oksidase tumbuhan atau dikenali sebagai *homolog ledakan pernafasan oksidase (rboh)* merupakan pemangkin pengeluaran spesies oksigen reaktif (ROS) yang memainkan peranan yang penting dalam perkembangan tumbuhan, isyarat hormon dan mekanisme pertahanan. Pengeluaran ROS oleh RBOHs yang terletak di membran plasma merupakan salah satu tindakbalas yang paling awal semasa serangan patogen dalam tumbuhan. *Fusarium wilt* dikenali sebagai salah satu penyakit pisang yang paling merosakkan di seluruh dunia yang disebabkan oleh *Fusarium oxysporum* f. sp. *cubense* (*Foc*), sejenis kulat di dalam tanah. Walaupun terdapat banyak laporan tentang penglibatan ROS dalam interaksi *Foc*-pisang, walaubagaimanapun, gen *rboh* dalam pisang masih belum dikenal pasti, maka menghadkan pemprofilan ekspresi *Marbohs* terhadap serangan *Fusarium* wilt. Matlamat utama kajian ini ialah untuk mencirikan *rbohs* dalam pisang (*Marbohs*) dan mengenalpasti *Marbohs* yang berkaitan dengan pertahanan melalui analisis ekspresi gen dan genomyang dikaitkan dengan tahap hidrogen peroksida dan kebocoran elektrolit. Dalam kajian ini, sembilan belas gen *Marbohs* telah dikenal pasti dalam sembilan kromosom DH Pahang melalui pencarian pangkalan data. Organisasi domain fungsian *Marbohs* termasuk ledakan pernafasan *NADPH* oksidase, domain pengikat kalsium *EF-hand* (EFh), pasangan domain *EF-hand* (EFh-7), Nox/ Duox protein transmembran dan domain pengikat NAD (*NAD binding 6*) yang penting untuk pengeluaran ROS. Analisis filogenetik telah mengelompokkan *Marbohs* ke dalam empat subkelompok dan mempunyai hubungan yang lebih rapat dengan *Manihot esculenta*, *Arabidopsis thaliana* dan *Brassica rapa*. Di antara semua *Marbohs*, hanya *MarbohB1* terletak di kumpulan III yang mana kebanyakan *rbohs* terlibat dalam tindakbalas pertahanan menentang patogen. Berdasarkan analisis ekspresi gen digital transkrip *Marbohs*, *MarbohB1* menunjukkan penurunan

yang ketara pada 48 dan 96 jam selepas inokulasi *Foc* Tropical Race 4 (*Foc*TR4). Berdasarkan analisis filogenetik dan ekspresi gen digital, *MarbohB1* telah dipilih untuk analisis yang seterusnya. Jujukan penuh *MarbohB1* dari Berangan (*MabrbohB1*) telah dipencilkan menggunakan kaedah *primer walking*. Penjajaran asid amino *MarbohB1* dari DH Pahang dan Berangan mendedahkan 13 polimorfisme nukleotida tunggal (SNPs) yang menyebabkan 6 perubahan asid amino tanpa sinonim dan 6 perubahan asid amino sinonim di mana 4 perubahan asid amino tanpa sinonim berlaku di domain abadi. Analisa ekspresi organ dan temporal *MabrbohB1* semasa infeksi kulat hemibiotrof *Foc*TR4 menunjukkan bahawa *MabrbohB1* diekspreskan secara khusus di dalam akar dan menunjukkan peningkatan secara sementara 2 jam selepas inokulasi oleh *Foc*TR4, diikuti penurunan pada 48 dan 96 jam selepas inokulasi. Peningkatan ekspresi tersebut disertai dengan peningkatan mantap tahap  $H_2O_2$  dan kebocoran elektrolit dalam akar yang diinokulasi, menunjukkan bahawa *MabrbohB1* mungkin terlibat dalam tindak balas pertahanan dalam pisang. Pencirian *Marbohs* dan pengenalpastian *MarbohB1* sebagai gen dalam pertahan menyumbang pengetahuan baru tentang interaksi *Foc*-pisang dan mungkin berguna untuk menangani penyakit *Fusarium wilt* dalam pisang pada masa akan datang. Secara keseluruhan, kajian ini menyokong peranan gen *rboh* yang dicadangkan dalam ledakan oksidatif semasa tindak balas tumbuhan dengan patogen.

## ACKNOWLEDGEMENTS

This dissertation is written to fulfil part of the requirement for Master Science in Plant Biotechnology. I would like to express my deepest appreciation to all those who provided me the possibility to complete my master research.

Firstly, I would like to express my gratitude to my respected supervisor, Dr. Noor Baity binti Saidi from Cell and Molecular Biology Department for continuously support of my master research. Her patience, immense knowledge, motivation and guidance helped me throughout my research and thesis writing.

Besides my supervisor, I would like to thank the rest of my thesis committees: Dr. Teo Chee How and Dr. Nur Fatihah Mohd Yusoff, for their insightful comments and suggestions, and further widen my research from various perspectives. Apart from that, I would like to thank Dr. Nadiya Akmal Baharum for always sharing her knowledge in plant molecular field especially related to banana. My sincere thanks also go to Dr. Lai Kok Song, who gave me access to his laboratory and facilities.

Furthermore, I would also like to acknowledge with much appreciation to the staff of Plant Molecular Biology Lab, Encik Faizzul, who gave the permission to use all required equipment and the materials to complete the research. Not to forget my lab members Nurul Najihah, Farah Wahida, Haira and Nisa for their help and support.

Lastly, I must express my very profound gratitude to my family members for providing me with unfailing support psychologically and financially throughout my master study. This accomplishment would not have been possible without them. Thank you.



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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## LIST OF ABBREVIATIONS

•O <sub>2</sub> <sup>-</sup>	Superoxide
•OH	Hydroxyl radical
1O <sub>2</sub>	Singlet oxygen
BIK1	Botrytis-induced kinase 1
bp	Base pair
Ca <sup>2+</sup>	Calcium ions
cDNA	Complementary deoxyribonucleic acid
CTAB	Cetyl trimethylammonium bromide
cv.	Cultivar
DEPC	Diethyl pyrocarbonate
DNA	Deoxyribonucleic acid
DUOX	Dual oxidase
EDTA	Ethylenediaminetetraacetic acid
EF	Elongation factor
EL	Electrolyte leakage
ETI	Effector-triggered immunity
FAD	Flavin adenine dinucleotide
FAOSTAT	Food and Agriculture Organization of the United Nations statistic
Fe	Iron
Foc	Fusarium oxysporium f. sp. cubense
FW	Fresh weight
GCTCV	Giant Cavendish Tissue Culture Variants
H <sup>+</sup>	Hydrogen ion
H <sub>2</sub> O <sub>2</sub>	Hydrogen peroxide
hpi	Hour post infection
HR	Hypersensitive cell death response
HWC	Hyphal wall components
K <sup>+</sup>	Potassium ion
kb	Kilo base pair
LiCl	Lithium chloride
MAMPs	Microbe-associated molecular patterns
NaCl	Sodium chloride
NAD	Nicotinamide-adenine dinucleotide
NADPH	Nicotinamide adenine dinucleotide phosphate
NCBI	National Centre for Biotechnology Information
NLR	Nucleotide-binding leucine-rich repeat
NO	Nitric oxide
NOX	NADPH oxidase
PA	Phosphatidic acid
PAMPs	Pathogen-associated molecular patterns
PCR	Polymerase chain reaction
PDA	Potato dextrose agar
PRRs	Pathogen recognition receptors
PTI	PAMP-triggered immunity
PVP	Polyvinylpyrrolidone
RBOH	Respiratory burst oxidase homologs

RNA	Ribonucleic acid
ROI	Reactive oxygen intermediates
ROS	Reactive oxygen species
S	Sulphur
SA	Salicylic acid
SAR	Systemic acquired resistance
SNP	Single nucleotide polymorphism
SOD	Superoxide dismutase
spp	Species
SR4	Subtropical Race 4
subsp.	Subspecies
TR4	Tropical Race 4
TTSS	Type III secretion system
VCG	Vegetative compatibility group
UPM	Universiti Putra Malaysia





## CHAPTER 1

### INTRODUCTION

In plants, respiratory burst oxidase homologs (RBOHs) or also known as NADPH oxidase play an important role in reactive oxygen species (ROS) production. ROS are the key components for plants to carry out basic biological processes such as response to biotic and abiotic stresses as well as modulation of protein and gene expression. RBOHs are membrane-localized proteins where C-terminal consists of FAD- and NADPH-binding sites, N-terminal consists of Ca<sup>2+</sup> binding EF hand motifs and six conserved transmembrane helices in between (Liu et al., 2019). Since the discovery of the first *rboh* gene in rice (Groom et al., 1996), many *rboh* genes have been identified in other plants including *Arabidopsis thaliana* (*Arabidopsis*), tobacco, tomato and potato (Hyodo et al., 2017, Kadota, Shirasu & Zipfel, 2015; Li et al., 2015; Wang et al., 2013; Torres and Dangl, 2005). Studies on different members of *rbohs* showed that they control different biological processes in plant such as plant development, hormone signalling and defense reactions (Wang et al., 2018).

ROS production via oxidative burst is one of the earliest responses during pathogen infection. Several plasma membrane-localized RBOHs for example AtRBOHD and its orthologues are responsible for ROS production during both pathogen-associated molecular pattern-triggered and effector-triggered immunities in plants (Kadota, Shirasu & Zipfel, 2015). In addition, *rbohB* from tomato, potato, rice and tobacco also positively regulates resistance response to pathogen (Li et al., 2015; Hajianfar et al., 2016; Kosami et al., 2014; Yoshioka et al., 2003).

Fusarium wilt is recognized as one of the most destructive banana diseases in the world (Köberl et al., 2017). The disease is caused by *Fusarium oxysporium* f. sp. *cubense* (*Foc*), a soil-borne fungus. Among four *Foc* races, *Foc* Tropical Race 4 (*Foc*TR4) is the most virulent, infecting almost 80% of banana cultivars worldwide (Luo et al., 2016). In Malaysia, the losses caused by *Foc*TR4 infection was reported to be about \$253 million in 2017 (FAO, 2017). The disease mostly affected banana that belongs to the AAA genome group including the famous dessert banana in Malaysia, *Musa acuminata* cv Berangan (Berangan). ROS have been reported to benefit hemibiotrophic pathogens such as *Foc*, which change from biotroph to necrotroph, stimulating the host to produce ROS or induce their own ROS production and feeding dead plant tissues to survive and reproduce. To our knowledge, *rboh* genes have not yet been characterized in banana. Thus, this study aims to identify *rboh* homologs in *Musa acuminata* subsp. *Malaccensis* (DH Pahang) and characterize selected *rboh* homolog in the local Berangan during a compatible interaction with *Foc*TR4. It is hypothesized that *Musa acuminata rboh* (*Marboh*)

genes share a common structure with RBOH from other plants and selected *Marboh(s)* with predicted role in defence response is induced by *FocTR4*.

The work described in this thesis was designed to achieve the following objectives:

1. To characterize *Marboh* genes in the resistant DH Pahang by in silico analysis
2. To isolate defense-related *Marboh* from the susceptible Berangan and analyse its expression in response to *FocTR4* infection
3. To determine the effect of *FocTR4* infection on the level of reactive oxygen species (ROS) and the extent of cell death in Berangan banana



## REFERENCES

- Abbas, H. M. K., Xiang, J., Ahmad, Z., Wang, L., & Dong, W. (2018). Enhanced *Nicotiana benthamiana* immune responses caused by heterologous plant genes from *Pinellia ternata*. *BMC plant biology*, 18(1), 357.
- Adachi, H., Ishihama, N., Nakano, T., Yoshioka, M., & Yoshioka, H. (2016). *Nicotiana benthamiana* MAPK-WRKY pathway confers resistance to a necrotrophic pathogen *Botrytis cinerea*. *Plant signaling & behavior*, 11(6), e1183085.
- Agunbiade, S. O., Olanlokun, J. O., & Olaofe, O. A. (2006). Quality of chips produced from rehydrated dehydrated plantain and banana. *Pakistan Journal of Nutrition*, 5(5), 471-473.
- Ajilogba, C. F., & Babalola, O. O. (2013). Integrated management strategies for tomato Fusarium wilt. *Biocontrol science*, 18(3), 117-127.
- Akhter, A., Hage-Ahmed, K., Soja, G., & Steinkellner, S. (2016). Potential of Fusarium wilt-inducing chlamydospores, in vitro behaviour in root exudates and physiology of tomato in biochar and compost amended soil. *Plant and soil*, 406(1-2), 425-440.
- Alloosh, M., Hamwieh, A., Ahmed, S., & Alkai, B. (2019). Genetic diversity of *Fusarium oxysporum* f. sp. *ciceris* isolates affecting chickpea in Syria. *Crop protection*, 124, 104863.
- Al-Obaidi, J. R., Mohd-Yusuf, Y., Chin-Chong, T., Mhd-Noh, N., & Othman, R. Y. (2010). Identification of a partial oil palm polygalacturonase-inhibiting protein (EgPGIP) gene and its expression during basal stem rot infection caused by *Ganoderma boninense*. *African journal of biotechnology*, 9(46), 788-7797.
- Avila, J., & Serrano, L. (2018). Structure and Function of Tubulin Regions. In *Microtubule Proteins* (pp. 67-88). CRC Press.
- Bai, T. T., Xie, W. B., Zhou, P. P., Wu, Z. L., Xiao, W. C., Zhou, L., Sun, J., Ruan, X., & Li, H. P. (2013). Transcriptome and expression profile analysis of highly resistant and susceptible banana roots challenged with *Fusarium oxysporum* f. sp. *ubense* tropical race 4. *Plos one*, 8(9), e73945.
- Bánfi, B., Tirone, F., Durussel, I., Knisz, J., Moskwa, P., Molnár, G. Z., Krause, K., & Cox, J. A. (2004). Mechanism of Ca<sup>2+</sup> activation of the NADPH oxidase 5 (NOX5). *Journal of biological chemistry*, 279(18), 18583-18591.

- Baştaş, K. K. (2015). Importance of reactive oxygen species in plants-pathogens interactions. *Selcuk Journal of Agriculture and Food Sciences*, 28(1), 11-21.
- Beautiful Banana Tree*. Retrieved 1 December 2018 from <http://www.123rf.com>.
- Berrocal-Lobo, M., & Molina, A. (2008). Arabidopsis defense response against *Fusarium oxysporum*. *Trends in plant science*, 13(3), 145-150.
- Bhatia, S., & Dahiya, R. (2015). Concepts and techniques of plant tissue culture science. *Modern Applications of Plant Biotechnology in Pharmaceutical Sciences*.
- Blanc, C., Coluccia, F., L'Haridon, F., Torres, M., Ortiz-Berrocal, M., Stahl, E., Reymond, P., Schreiber, L., Nawrath, C., Metraux, J., & Serrano, M. (2018). The cuticle mutant *eca2* modifies plant defense responses to biotrophic and necrotrophic pathogens and herbivory insects. *Molecular plant-microbe interactions*, 31(3), 344-355.
- Booth, C. (1971). The genus *Fusarium*. *The genus Fusarium*.
- Bos, J. I., Prince, D., Pitino, M., Maffei, M. E., Win, J., & Hogenhout, S. A. (2010). A functional genomics approach identifies candidate effectors from the aphid species *Myzus persicae* (green peach aphid). *PLoS genetics*, 6(11), e1001216.
- Buddenhagen, I. (2007). Understanding strain diversity in *Fusarium oxysporum* f. sp. *cubense* and history of introduction of Tropical Race 4 to better manage banana production. In *III International Symposium on Banana: ISHS-ProMusa Symposium on Recent Advances in Banana Crop Protection for Sustainable 828* (pp. 193-204).
- Campos, P. S., nia Quartin, V., chicho Ramalho, J., & Nunes, M. A. (2003). Electrolyte leakage and lipid degradation account for cold sensitivity in leaves of *Coffea* sp. plants. *Journal of plant physiology*, 160(3), 283-292.
- Carrasco-Castilla, J., Ortega-Ortega, Y., Jáuregui-Zúñiga, D., Juárez-Verdayes, M. A., Arthikala, M. K., Monroy-Morales, E., Nava, N., Santana, O., Sánchez-López, R., & Quinto, C. (2018). Down-regulation of a *Phaseolus vulgaris* annexin impairs rhizobial infection and nodulation. *Environmental and experimental botany*, 153, 108-119.
- Chaouch, S., Queval, G., & Noctor, G. (2012). *AtRbohF* is a crucial modulator of defence-associated metabolism and a key factor in the interplay between intracellular oxidative stress and pathogenesis responses in Arabidopsis. *The plant journal*, 69(4), 613-627.

- Chapman, J. M., Muhlemann, J. K., Gayomba, S. R., & Muday, G. K. (2019). RBOH-dependent ROS synthesis and ROS scavenging by plant specialized metabolites to modulate plant development and stress responses. *Chemical research in toxicology*, 32(3):370-396.
- Chen, L., Zhong, H. Y., Kuang, J. F., Li, J. G., Lu, W. J., & Chen, J. Y. (2011). Validation of reference genes for RT-qPCR studies of gene expression in banana fruit under different experimental conditions. *Planta*, 234(2), 377.
- Christelová, P., De Langhe, E., Hřibová, E., Čížková, J., Sardos, J., Hušáková, M., Houwe, I. V. D., Sutanto, A., Kepler, A. K., Swennen, R., Roux, N., & Doležel, J. (2017). Molecular and cytological characterization of the global *Musa* germplasm collection provides insights into the treasure of banana diversity. *Biodiversity and conservation*, 26(4), 801-824.
- Cove, D. J. (1979). Genetic studies of nitrate assimilation in *Aspergillus nidulans*. *Biological Reviews*, 54(3), 291-327.
- Dale, J., James, A., Paul, J. Y., Khanna, H., Smith, M., Peraza-Echeverria, S., Garcia-Bastidas, F., Kema, G., Waterhouse, P., Mengersen, K., & Harding, R. (2017). Transgenic Cavendish bananas with resistance to *Fusarium* wilt tropical race 4. *Nature communications*, 8(1), 1496.
- Damodaran, T., Mishra, V. K., Jha, S. K., Gopal, R., Rajan, S., & Ahmed, I. (2019). First report of *Fusarium* wilt in banana caused by *Fusarium oxysporum* f. sp. *cubense* tropical race 4 in India. *Plant disease*, 103(5), 1022-1022.
- Dangl, J. L., Horvath, D. M., & Staskawicz, B. J. (2013). Pivoting the plant immune system from dissection to deployment. *Science*, 341(6147), 746-751.
- Das, K., & Roychoudhury, A. (2014). Reactive oxygen species (ROS) and response of antioxidants as ROS-scavengers during environmental stress in plants. *Frontiers in environmental science*, 2, 53.
- De Langhe, C., Vrydaghs, L., De Maret, P., Perrier, X., & Denham, T. (2009). Why bananas matter: an introduction to the history of banana domestication. *Ethnobotany research and applications*, 7, 165-177.
- Demidchik, V. (2014). Mechanisms and physiological roles of K<sup>+</sup> efflux from root cells. *Journal of plant physiology*, 171(9), 696-707.
- Demidchik, V., Straltsova, D., Medvedev, S. S., Pozhvanov, G. A., Sokolik, A., & Yurin, V. (2014). Stress-induced electrolyte leakage: the role of K<sup>+</sup>-permeable channels and involvement in programmed cell death and metabolic adjustment. *Journal of experimental botany*, 65(5), 1259-1270.

- Dexter, S. T., Tottingham, W. E., & Graber, L. F. (1932). Investigations of the hardiness of plants by measurement of electrical conductivity. *Plant physiology*, 7(1), 63.
- Dita, M. A., Waalwijk, C., Buddenhagen, I. W., Souza Jr, M. T., & Kema, G. H. J. (2010). A molecular diagnostic for tropical race 4 of the banana fusarium wilt pathogen. *Plant pathology*, 59(2), 348-357.
- Dita, M., Barquero, M., Heck, D., Mizubuti, E. S., & Staver, C. P. (2018). Fusarium wilt of banana: current knowledge on epidemiology and research needs toward sustainable disease management. *Frontiers in plant science*, 9, 1468.
- Dodds, P. N., & Rathjen, J. P. (2010). Plant immunity: towards an integrated view of plant–pathogen interactions. *Nature reviews genetics*, 11(8), 539.
- Drerup, M. M., Schlücking, K., Hashimoto, K., Manishankar, P., Steinhorst, L., Kuchitsu, K., & Kudla, J. (2013). The Calcineurin B-like calcium sensors CBL1 and CBL9 together with their interacting protein kinase CIPK26 regulate the Arabidopsis NADPH oxidase RBOHF. *Molecular plant*, 6(2), 559-569.
- El-Khishin, D. A., Belatus, E. L., El-Hamid, A. A., & Radwan, K. H. (2009). Molecular characterization of banana cultivars (*Musa* spp.) from Egypt using AFLP. *Research journal of agriculture and biological sciences*, 5(3), 271-279.
- FAOSTAT (2017). Retrieved 1 December 2018 from <http://www.fao.org/faostat/en/#data/QC>.
- Foreman, J., Demidchik, V., Bothwell, J. H., Mylona, P., Miedema, H., Torres, M. A. & Davies, J. M. (2003). Reactive oxygen species produced by NADPH oxidase regulate plant cell growth. *Nature*, 422(6930), 442-446.
- Franco Ortega, S., Tomlinson, J., Gilardi, G., Spadaro, D., Gullino, M. L., Garibaldi, A., & Boonham, N. (2018). Rapid detection of *Fusarium oxysporum* f. sp. *lactucae* on soil, lettuce seeds and plants using loop-mediated isothermal amplification. *Plant pathology*, 67(7), 1462-1473.
- Fu, L., Penton, C. R., Ruan, Y., Shen, Z., Xue, C., Li, R., & Shen, Q. (2017). Inducing the rhizosphere microbiome by biofertilizer application to suppress banana Fusarium wilt disease. *Soil biology and biochemistry*, 104, 39-48.
- Galletti, R., Denoux, C., Gambetta, S., Dewdney, J., Ausubel, F. M., De Lorenzo, G., & Ferrari, S. (2008). The *AtrbohD*-mediated oxidative burst elicited by oligogalacturonides in *Arabidopsis* is dispensable for the activation of defense responses effective against *Botrytis cinerea*. *Plant physiology*, 148(3), 1695-1706.

- Garcia-Bastidas, F. A., Van der Veen, A., Nakasato-Tagami, G., Meijer, H. J., Arango Isaza, R. E., & Kema, G. H. (2019). An improved phenotyping protocol for Panama disease in banana. *Frontiers in plant science*, *10*, 1006.
- Grabherr, M. G., Haas, B. J., Yassour, M., Levin, J. Z., Thompson, D. A., Amit, I., Adiconis, X., Fan, L., Raychowdhury, R., Zheng, Q., Chen, Z., Mauceli, E., Hacohen, N., Gnirke, A., Rhind, N., Palma, F. D., Birren, B. W., Nusbaum, C., Lindblad-Toh, K., Friedman, N., & Regev, A. (2011). Full-length transcriptome assembly from RNA-Seq data without a reference genome. *Nature biotechnology*, *29*(7), 644.
- Groenewald, S., Van den Berg, N., Marasas, W. F. O., & Viljoen, A. (2006). Biological, physiological and pathogenic variation in a genetically homogenous population of *Fusarium oxysporum* f. sp. *cubense*. *Australasian Plant pathology*, *35*(4), 401-409.
- Groom, Q. J., Torres, M. A., Fordham-Skelton, A. P., Hammond-Kosack, K. E., Robinson, N. J., & Jones, J. D. (1996). *rbohA*, a rice homologue of the mammalian gp91phox respiratory burst oxidase gene. *The Plant journal*, *10*(3), 515-522.
- Hajianfar, R., Kolics, B., Cernák, I., Wolf, I., Polgár, Z., & Taller, J. (2016). Expression of biotic stress response genes to *Phytophthora infestans* inoculation in White Lady, a potato cultivar with race-specific resistance to late blight. *Physiological and molecular plant pathology*, *93*, 22-28.
- Hasanuzzaman, M., Nahar, K., Gill, S. S., Alharby, H. F., Razafindrabe, B. H., & Fujita, M. (2017). Hydrogen peroxide pretreatment mitigates cadmium-induced oxidative stress in *Brassica napus* L.: an intrinsic study on antioxidant defense and glyoxalase systems. *Frontiers in plant science*, *8*, 115.
- Hernández, J. A., Gullner, G., Clemente-Moreno, M. J., Künstler, A., Juhász, C., Díaz-Vivancos, P., & Király, L. (2016). Oxidative stress and antioxidative responses in plant-virus interactions. *Physiological and molecular plant pathology*, *94*, 134-148.
- Hirt, H. (2016). Aquaporins link ROS signaling to plant immunity. *Plant physiology*, *171*(3), 1540-1540.
- Huang, Y. H., Wang, R. C., Li, C. H., Zuo, C. W., Wei, Y. R., Zhang, L., & Yi, G. J. (2012). Control of Fusarium wilt in banana with Chinese leek. *European journal of plant pathology*, *134*(1), 87-95.
- Huq, M. A., Akter, S., Nou, I. S., Kim, H. T., Jung, Y. J., & Kang, K. K. (2016). Identification of functional SNPs in genes and their effects on plant phenotypes. *Journal of plant biotechnology*, *43*(1), 1-11.

- Hwang, S. C., & Ko, W. H. (2004). Cavendish banana cultivars resistant to Fusarium wilt acquired through somaclonal variation in Taiwan. *Plant disease*, 88(6), 580-588.
- Hyodo, K., Suzuki, N., Mise, K., & Okuno, T. (2017). Roles of superoxide anion and hydrogen peroxide during replication of two unrelated plant RNA viruses in *Nicotiana benthamiana*. *Plant signaling & behavior*, 12(6), e1338223.
- Jacob, F., Kracher, B., Mine, A., Seyfferth, C., Blanvillain-Baufumé, S., Parker, J. E., Tsuda, K., Schulze-Lefert, P., & Maekawa, T. (2018). A dominant-interfering camta3 mutation compromises primary transcriptional outputs mediated by both cell surface and intracellular immune receptors in *Arabidopsis thaliana*. *New phytologist*, 217(4), 1667-1680.
- Jakubowska, D., & Janicka, M. (2017). The role of brassinosteroids in the regulation of the plasma membrane H<sup>+</sup>-ATPase and NADPH oxidase under cadmium stress. *Plant science*, 264, 37-47.
- Jang, Y. J., Seo, M., Hersh, C. P., Rhee, S. J., Kim, Y., & Lee, G. P. (2019). An evolutionarily conserved non-synonymous SNP in a leucine-rich repeat domain determines anthracnose resistance in watermelon. *Theoretical and applied genetics*, 132(2), 473-488.
- Jeong, B. R., van Dijk, K., & Alfano, J. R. (2018). Pseudomonas Syringae Type III-Secreted Proteins and their Activities and Effects on Plant Innate Immunity. *Annual plant reviews online*, 48-76.
- Jones, J. D., & Dangl, J. L. (2006). The plant immune system. *Nature*, 444(7117), 323
- Kadota, Y., Shirasu, K., & Zipfel, C. (2015). Regulation of the NADPH oxidase RBOHD during plant immunity. *Plant and cell physiology*, 56(8), 1472-1480.
- Kadota, Y., Sklenar, J., Derbyshire, P., Stransfeld, L., Asai, S., Ntoukakis, V., Jones, J. DG., Shirasu, K., Menke, F., Jones, A., & Zipfel, C. (2014). Direct regulation of the NADPH oxidase RBOHD by the PRR-associated kinase BIK1 during plant immunity. *Molecular cell*, 54(1), 43-55.
- Karanjalkar, G. R., Ravishankar, K. V., Shivashankara, K. S., Dinesh, M. R., Roy, T. K., & Rao, D. S. (2018). A study on the expression of genes involved in carotenoids and anthocyanins during ripening in fruit peel of green, yellow, and red colored mango cultivars. *Applied biochemistry and biotechnology*, 184(1), 140-154.



- Kaur, G., & Pati, P. K. (2018). In silico physicochemical characterization and topology analysis of respiratory burst oxidase homolog (Rboh) proteins from *Arabidopsis* and rice. *Bioinformation*, *14*(3), 93.
- Kaur, G., Guruprasad, K., Temple, B. R., Shirvanyants, D. G., Dokholyan, N. V., & Pati, P. K. (2018). Structural complexity and functional diversity of plant NADPH oxidases. *Amino acids*, *50*(1), 79-94.
- Kobayashi, M., Yoshioka, M., Asai, S., Nomura, H., Kuchimura, K., Mori, H., Doke, N., & Yoshioka, H. (2012). StCDPK5 confers resistance to late blight pathogen but increases susceptibility to early blight pathogen in potato via reactive oxygen species burst. *New phytologist*, *196*(1), 223-237.
- Köberl, M., Dita, M., Martinuz, A., Staver, C., & Berg, G. (2017). Members of Gammaproteobacteria as indicator species of healthy banana plants on *Fusarium* wilt-infested fields in Central America. *Scientific reports*, *7*, 45318.
- Kosami, K. I., Ohki, I., Nagano, M., Furuita, K., Sugiki, T., Kawano, Y., Kawasaki, T., Fujiwara, T., Nakagawa, A., Shimamoto, K., & Kojima, C. (2014). The crystal structure of the plant small GTPase OsRac1 reveals its mode of binding to NADPH oxidase. *Journal of biological chemistry*, *289*(41), 28569-28578.
- Kuang, J., Yan, X., Genders, A. J., Granata, C., & Bishop, D. J. (2018). An overview of technical considerations when using quantitative real-time PCR analysis of gene expression in human exercise research. *PLoS one*, *13*(5), e0196438.
- Kühlbrandt, W. (2015). Structure and function of mitochondrial membrane protein complexes. *BMC biology*, *13*(1), 89.
- Kumar, A., Rajendran, V., Sethumadhavan, R., Shukla, P., Tiwari, S., & Purohit, R. (2014). Computational SNP analysis: current approaches and future prospects. *Cell biochemistry and biophysics*, *68*(2), 233-239.
- Kumar, S., Stecher, G., & Tamura, K. (2016). MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. *Molecular biology and evolution*, *33*(7), 1870-1874.
- Lal, N. K., Nagalakshmi, U., Hurlburt, N. K., Flores, R., Bak, A., Sone, P., Ma, X., Song, G., Walley, J., Shan, L., He, P., Casteel, C., Fisher, A. J., & Dinesh-Kumar, S. P. (2018). The receptor-like cytoplasmic kinase BIK1 localizes to the nucleus and regulates defense hormone expression during plant innate immunity. *Cell host & microbe*, *23*(4), 485-497.
- Laluk, K., Luo, H., Chai, M., Dhawan, R., Lai, Z., & Mengiste, T. (2011). Biochemical and genetic requirements for function of the immune response regulator Botrytis-induced kinase1 in plant growth, ethylene

- signaling, and PAMP-triggered immunity in Arabidopsis. *The plant cell*, 23(8), 2831-2849.
- Letunic, I., & Bork, P. (2017). 20 years of the SMART protein domain annotation resource. *Nucleic acids research*, 46(D1), D493-D496.
- Li, C. Y., Deng, G. M., Yang, J., Viljoen, A., Jin, Y., Kuang, R. B., Zuo, C. W., Lv, Z. C., Sheng, Ou., Hu. C. H., Dong. Tao, Yi. G. J. & Wei, Y. R. (2012). Transcriptome profiling of resistant and susceptible Cavendish banana roots following inoculation with *Fusarium oxysporum* f. sp. *cubense* tropical race 4. *BMC genomics*, 13(1), 374.
- Li, D., Wu, D., Li, S., Dai, Y., & Cao, Y. (2019). Evolutionary and functional analysis of the plant-specific NADPH oxidase gene family in *Brassica rapa* L. *Royal Society open science*, 6(2), 181727.
- Li, L., Li, M., Yu, L., Zhou, Z., Liang, X., Liu, Z., Cai, G., Zhang, X., Wang, Y., Zhou, J., & Chen, S. (2014). The FLS2-associated kinase BIK1 directly phosphorylates the NADPH oxidase *RbohD* to control plant immunity. *Cell host & microbe*, 15(3), 329-338.
- Li, L., Tan, J., Miao, Y., Lei, P., & Zhang, Q. (2015). ROS and autophagy: interactions and molecular regulatory mechanisms. *Cellular and molecular neurobiology*, 35(5), 615-621.
- Li, W. M., Qian, C. M., Mo, Y. W., Hu, Y. L., & Xie, J. H. (2011). Tolerance of banana for fusarium wilt is associated with early H<sub>2</sub>O<sub>2</sub> accumulation in the roots. *African journal of biotechnology*, 10(55), 11378-11387.
- Li, X., Zhang, H., Tian, L., Huang, L., Liu, S., Li, D., & Song, F. (2015). Tomato *SIRbohB*, a member of the NADPH oxidase family, is required for disease resistance against *Botrytis cinerea* and tolerance to drought stress. *Frontiers in plant science*, 6, 463.
- Lim, S. D., Kim, S. H., Gilroy, S., Cushman, J. C., & Choi, W. G. (2019). Quantitative ROS bioreporters: A robust toolkit for studying biological roles of ROS in response to abiotic and biotic stresses. *Physiologia plantarum*, 165(2), 356-368.
- Liu, J., Lu, H., Wan, Q., Qi, W., & Shao, H. (2019). Genome-wide analysis and expression profiling of respiratory burst oxidase homologue gene family in *Glycine max*. *Environmental and experimental botany*, 161, 344-356.
- Liu, Y., & He, C. (2016). Regulation of plant reactive oxygen species (ROS) in stress responses: learning from AtRBOHD. *Plant cell reports*, 35(5), 995-1007.
- Luo, J. Y., Pan, X. L., Peng, T. C., Chen, Y. Y., Hui, Z. H. A. O., Lei, M. U., ... & Hua, T. A. N. G. (2016). DNA methylation patterns of banana leaves in

response to *Fusarium oxysporum* f. sp. *cubense* tropical race 4. *Journal of integrative agriculture*, 15(12), 2736-2744.

- Lyons, R., Stiller, J., Powell, J., Rusu, A., Manners, J. M., & Kazan, K. (2015). *Fusarium oxysporum* triggers tissue-specific transcriptional reprogramming in *Arabidopsis thaliana*. *PLoS one*, 10(4), e0121902.
- Mak, C., Ho, Y. W., Liew, K. W., Asif, J. M., Jain, S. M., & Swennen, R. (2004). Biotechnology and in vitro mutagenesis for banana improvement. In *Banana improvement: cellular, molecular biology, and induced mutations. Proceedings of a meeting held in Leuven, Belgium, 24-28 September 2001.* (pp. 59-77). *Science publishers*.
- Marchler-Bauer, A., Derbyshire, M. K., Gonzales, N. R., Lu, S., Chitsaz, F., Geer, L. Y., Geer, R. C., He, J., Gwadz, M., Hurwitz DI, Lanczycki, C. J., Lu F, Marchler GH, Song JS, Thanki S, Wang Z., Yamashita, R. A., Zhang, D., Zheng, C., Bryant, S. H. (2014). CDD: NCBI's conserved domain database. *Nucleic acids research*, 43(D1), D222-D226.
- Marino, D., Andrio, E., Danchin, E. G., Oger, E., Gucciardo, S., Lambert, A., Puppo, A., & Pauly, N. (2011). A *Medicago truncatula* NADPH oxidase is involved in symbiotic nodule functioning. *New phytologist*, 189(2), 580-592.
- Marino, D., Dunand, C., Puppo, A., & Pauly, N. (2012). A burst of plant NADPH oxidases. *Trends in plant science*, 17(1), 9-15.
- Maryani, N., Lombard, L., Poerba, Y. S., Subandiyah, S., Crous, P. W., & Kema, G. H. J. (2019). Phylogeny and genetic diversity of the banana *Fusarium oxysporum* f. sp. *cubense* in the Indonesian centre of origin. *Studies in mycology*, 92, 155-194.
- Masdek bin Nik Kassan (2002). Banana industry and R&D in Malaysia. *Advancinbanana and plantain R&D in Asia and the Pacific. INIBAPAP, Los Banos*, 99-106.
- Meldrum, R. A., Daly, A. M., Tran-Nguyen, L. T. T., & Aitken, E. A. B. (2013). Are banana weevil borers a vector in spreading *Fusarium oxysporum* f. sp. *cubense* tropical race 4 in banana plantations? *Australasian plant pathology*, 42(5), 543-549.
- Melillo, M. T., Leonetti, P., Leone, A., Veronico, P., & Bleve-Zacheo, T. (2011). ROS and NO production in compatible and incompatible tomato-*Meloidogyne incognita* interactions. *European journal of plant pathology*, 130(4), 489-502.
- Miller, M., Tunc-Ozdemir, G., Song, L., Kim, J., Sodek, A., Koussevitzky, S., & Shintani, D. (2009). Thiamin confers enhanced tolerance to oxidative stress in *Arabidopsis*. *Plant physiology*, 151(1), 421-432.

- Mokhtarud-din, H., & William, R. (2011). Status of banana cultivation and disease incidences in Malaysia. In *Abstract of the Workshop on Integrated Approaches in Banana Disease Management, 22th March* (p. 5).
- Montiel, J., Arthikala, M. K., & Quinto, C. (2013). Phaseolus vulgaris *RbohB* functions in lateral root development. *Plant signaling & behavior*, 8(1), e22694.
- Morales, J., Kadota, Y., Zipfel, C., Molina, A., & Torres, M. A. (2016). The *Arabidopsis* NADPH oxidases *RbohD* and *RbohF* display differential expression patterns and contributions during plant immunity. *Journal of experimental botany*, 67(6), 1663-1676.
- Morales, J., Kadota, Y., Zipfel, C., Molina, A., & Torres, M. A. (2016). The *Arabidopsis* NADPH oxidases *RbohD* and *RbohF* display differential expression patterns and contributions during plant immunity. *Journal of experimental botany*, 67(6), 1663-1676.
- Mostert, D., Molina, A. B., Daniells, J., Fourie, G., Hermanto, C., Chao, C. P., Fabregar, E., Sinohin, V. G., Masdek, N., Yi, G., Mostert, L., Viljoen, A., & Li, C. (2017). The distribution and host range of the banana Fusarium wilt fungus, *Fusarium oxysporum* f. sp. *cubense*, in Asia. *PLoS one*, 12(7), e0181630.
- Mostofa, M. G., Fujita, M., & Tran, L. S. P. (2015). Nitric oxide mediates hydrogen peroxide-and salicylic acid-induced salt tolerance in rice (*Oryza sativa* L.) seedlings. *Plant growth regulation*, 77(3), 265-277.
- Nagano, M., Ishikawa, T., Fujiwara, M., Fukao, Y., Kawano, Y., Kawai-Yamada, M., & Shimamoto, K. (2016). Plasma membrane microdomains are essential for Rac1-RbohB/H-mediated immunity in rice. *The plant cell*, 28(8), 1966-1983.
- Naing, K. W., Nguyen, X. H., Anees, M., Lee, Y. S., Kim, Y. C., Kim, S. J., ... & Kim, K. Y. (2015). Biocontrol of Fusarium wilt disease in tomato by *Paenibacillus ehimensis* KWN38. *World journal of microbiology and biotechnology*, 31(1), 165-174.
- Nicaise, V., & Candresse, T. (2017). Plum pox virus capsid protein suppresses plant pathogen-associated molecular pattern (PAMP)-triggered immunity. *Molecular plant pathology*, 18(6), 878-886.
- Niu, L., & Liao, W. (2016). Hydrogen peroxide signaling in plant development and abiotic responses: crosstalk with nitric oxide and calcium. *Frontiers in plant science*, 7, 230.
- Noor, M. R. M., Malik, T. A., Maamun T., & Sharif H. (2012). Challenges to banana production in Malaysia: A threat to food security. *The planter*, Kuala Lumpur; 88 (1030):13-21.

- O'Brien, J. A., Daudi, A., Finch, P., Butt, V. S., Whitelegge, J. P., Souda, P., Ausubel, F. M., & Bolwell, G. P. (2012). A peroxidase-dependent apoptotic oxidative burst in cultured *Arabidopsis* cells functions in MAMP-elicited defense. *Plant physiology*, 158(4), 2013-2027.
- Pereira, A., & Maraschin, M. (2015). Banana (*Musa* spp) from peel to pulp: ethnopharmacology, source of bioactive compounds and its relevance for human health. *Journal of ethnopharmacology*, 160, 149-163.
- Pérez Vicente, L., Dita, M., & Martínez De La Parte, E. (2014). Technical Manual: Prevention and diagnostic of Fusarium Wilt (Panama disease) of banana caused by *Fusarium oxysporum* f. sp. *cubense* Tropical Race 4 (TR4). Conference proceedings of International Conference on Innovative Approaches in Applied Sciences and Technologies.
- Ploetz, R. C. (1990). Population biology of *Fusarium oxysporum* f. sp. *cubense*. *Fusarium wilt of banana*, 63-76.
- Ploetz, R. C. (2006). Fusarium wilt of banana is caused by several pathogens referred to as *Fusarium oxysporum* f. sp. *cubense*. *Phytopathology*, 96(6), 653-656.
- Ploetz, R. C. (2015). Fusarium wilt of banana. *Phytopathology*, 105(12), 1512-1521.
- Ploetz, R. C., & Pegg, K. G. (2000). Fusarium wilt. *Diseases of banana, abaca and enset*, 143-159.
- Potocký, M., Jones, M. A., Bezdová, R., Smirnov, N., & Žárský, V. (2007). Reactive oxygen species produced by NADPH oxidase are involved in pollen tube growth. *New phytologist*, 174(4), 742-751.
- Qi, S., Feng, Z., Li, Q., Qi, Z., & Zhang, Y. (2017). Myricitrin modulates NADPH oxidase-dependent ROS production to inhibit endotoxin-mediated inflammation by blocking the JAK/STAT1 and NOX2/p47phox pathways. *Oxidative medicine and cellular longevity*, 2017: 9738745.
- Ranjan, A., Jayaraman, D., Grau, C., Hill, J. H., Whitham, S. A., Ané, J. M., Smith, D. L., & Kabbage, M. (2018). The pathogenic development of *Sclerotinia sclerotiorum* in soybean requires specific host NADPH oxidases. *Molecular plant pathology*, 19(3), 700-714.
- Ren, L., Zhu, B. Q., Zhang, Y. B., Wang, H. Y., Li, C. Y., Su, Y. H., & Ba, C. F. (2004). The research of applying primer premier 5.0 to design PCR primer. *Journal of Jinzhou medical college*, 25(6), 43-46.
- Resmi, L., Kumari, R., Bhat, K. V., & Nair, A. S. (2011). Molecular characterization of genetic diversity and structure in South Indian *Musa* cultivars. *International journal of botany*, 7(4), 274-282.

- Roy, S., & Nandi, A. K. (2017). *Arabidopsis thaliana* methionine sulfoxide reductase B8 influences stress-induced cell death and effector-triggered immunity. *Plant molecular biology*, 93(1-2), 109-120.
- Sabiiti, G., Ininda, J. M., Ogallo, L. A., Ouma, J., Artan, G., Basalirwa, C., Opijah, F., Nimusiima, A., Ddumba, S. D., Mwesigwa, J. B., Nanteza, J., & Otieno, G. (2018). Adapting agriculture to climate change: suitability of banana crop production to future climate change over Uganda. In *Limits to Climate Change Adaptation* (pp. 175-190).
- Sagi, M., & Fluhr, R. (2006). Production of reactive oxygen species by plant NADPH oxidases. *Plant physiology*, 141(2), 336-340.
- Sagi, M., Davydov, O., Orazova, S., Yesbergenova, Z., Ophir, R., Stratmann, J. W., & Fluhr, R. (2004). Plant respiratory burst oxidase homologs impinge on wound responsiveness and development in *Lycopersicon esculentum*. *The plant cell*, 16(3), 616-628.
- Saijo, Y., Loo, E. P. I., & Yasuda, S. (2018). Pattern recognition receptors and signaling in plant-microbe interactions. *The plant journal*, 93(4), 592-613.
- Samajdar, S., Mukherjee, S., & Das, P. P. (2018). Seed Germination Inhibitors: Molecular and Phytochemical Aspects. *International journal of applied pharmaceutical sciences and research*, 3(2), 12-23.
- Segura, R. A. (2018). Nutrition and soil management in banana cultivation Jetse J. Stoorvogel, Wageningen University, The Netherlands; and. In *Achieving sustainable cultivation of bananas* (pp. 243-254).
- Shamsinah, N., & Suhaimi, M. (2017). *Diversity of bacterial communities in bacterial wilt-diseased banana plants* (Doctoral dissertation, University of Malaya).
- Shang, Y., Dai, C., Lee, M. M., Kwak, J. M., & Nam, K. H. (2016). BRI1-associated receptor kinase 1 regulates guard cell ABA signaling mediated by open stomata 1 in *Arabidopsis*. *Molecular plant*, 9(3), 447-460.
- Simmonds, N. W., & Weatherup, S. T. C. (1990). Numerical taxonomy of the wild bananas (*Musa*). *New phytologist*, 115(3), 567-571.
- Singh, H. P., Batish, D. R., Kohli, R. K., & Arora, K. (2007). Arsenic-induced root growth inhibition in mung bean (*Phaseolus aureus* Roxb.) is due to oxidative stress resulting from enhanced lipid peroxidation. *Plant growth regulation*, 53(1), 65-73.
- Stover, R. H. (1959). Studies of *Fusarium* wilt of bananas: IV. Clonal differentiation among wild types isolates of *F. oxysporum* f. *cubenses*. *Canadian journal of botany*, 37(2), 245-255.

- Stover, R. H. (1962). Fusarial wilt (Panama Disease) of bananas and other *Musa* species. *Fusarial wilt (Panama disease) of bananas and other Musa species* (pp. 117).
- Stover, R. H., & Simmonds, N. W. (1987). Bananas. Tropical agricultural series. *John Wiley and Sons*, 10, 158-468.
- Su, N. Y., & Scheffrahn, R. H. (1986). A method to access, trap, and monitor field populations of the *Formosan subterranean* termite (Isoptera: Rhinotermitidae). *Sociobiology*, 12(2).
- Sun, E. J., & Su, H. J. (1985). Fluoride injury to rice plants caused by air pollution emitted from ceramic and brick factories. *Environmental pollution series A, Ecological and Biological*, 37(4), 335-342.
- Suzuki, N., & Katano, K. (2018). Coordination between ROS regulatory systems and other pathways under heat stress and pathogen attack. *Frontiers in plant science*, 9, 490.
- Swennen, R., Vuylsteke, D., & Ortiz, R. (1995). Phenotypic diversity and patterns of variation in West and Central African plantains (*Musa* spp., AAB group Musaceae). *Economic botany*, 49(3), 320-327.
- Sytykiewicz, H. (2016). Deciphering the role of NADPH oxidase in complex interactions between maize (*Zea mays* L.) genotypes and cereal aphids. *Biochemical and biophysical research communications*, 476(2), 90-95.
- Teufel, R., Miyanaga, A., Michaudel, Q., Stull, F., Louie, G., Noel, J. P., Baran, P. S., Palfey, B., & Moore, B. S. (2013). Flavin-mediated dual oxidation controls an enzymatic Favorskii-type rearrangement. *Nature*, 503(7477), 552.
- Thatcher, L. F., Kidd, B. N., & Kazan, K. (2016). Belowground defence strategies against *Fusarium oxysporum*. In *Belowground defence strategies in plants* (pp. 71-98).
- Thaysen, A. C., & Butlin, K. R. (1945). Inhibition of the development of *Fusarium oxysporum cubense* by a growth substance produced by Meredith's actinomycetes. *Nature*, 156(3974), 781.
- Tinzaara, W., Stoian, D., Ocimati, W., Kikulwe, E., Otieno, G., & Blomme, G. (2018). Challenges and opportunities for smallholders in banana value chains. In *Achieving sustainable cultivation of bananas*. Cambridge (UK).
- Torres, M. A., & Dangl, J. L. (2005). Functions of the respiratory burst oxidase in biotic interactions, abiotic stress and development. *Current opinion in plant biology*, 8(4), 397-403.
- Torres, M. A., Jones, J. D., & Dangl, J. L. (2006). Reactive oxygen species signaling in response to pathogens. *Plant physiology*, 141(2), 373-378.

- Waite, B. H., & Stover, R. H. (1960). Studies on Fusarium wilt of bananas: VI Variability and the cultivar concept in *Fusarium oxysporum* f. cubenses. *Canadian journal of botany*, 38(6), 985-994.
- Wang, G. F., Li, W. Q., Li, W. Y., Wu, G. L., Zhou, C. Y., & Chen, K. M. (2013). Characterization of rice NADPH oxidase genes and their expression under various environmental conditions. *International journal of molecular sciences*, 14(5), 9440-9458.
- Wang, W., Chen, D., Zhang, X., Liu, D., Cheng, Y., & Shen, F. (2018). Role of plant respiratory burst oxidase homologs in stress responses. *Free radical research*, 52(8), 826-839.
- Wang, W., Zhang, H., Wei, X., Yang, L., Yang, B., Zhang, L., Li, J., & Jiang, Y. Q. (2018). Functional characterization of calcium-dependent protein kinase (CPK) 2 gene from oilseed rape (*Brassica napus* L.) in regulating reactive oxygen species signaling and cell death control. *Gene*, 651, 49-56.
- Wang, X., Zhang, M. M., Wang, Y. J., Gao, Y. T., Li, R., Wang, G. F., Li, W. Q., Liu, W. T., & Chen, K. M. (2016). The plasma membrane NADPH oxidase *OsRbohA* plays a crucial role in developmental regulation and drought-stress response in rice. *Physiologia plantarum*, 156(4), 421-443.
- Wi, S. J., Ji, N. R., & Park, K. Y. (2012). Synergistic biosynthesis of biphasic ethylene and reactive oxygen species in response to hemibiotrophic *Phytophthora parasitica* in tobacco plants. *Plant physiology*, 159(1), 251-265.
- Wong, H. L., Pinontoan, R., Hayashi, K., Tabata, R., Yaeno, T., Hasegawa, K., Kojima, C., Yoshioka, H., Iba, K., Kawasaki, T., & Shimamoto, K. (2007). Regulation of rice NADPH oxidase by binding of Rac GTPase to its N-terminal extension. *The plant cell*, 19(12), 4022-4034.
- Xiong, J., Yang, Y., Fu, G., & Tao, L. (2015). Novel roles of hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) in regulating pectin synthesis and demethylesterification in the cell wall of rice (*Oryza sativa*) root tips. *New Phytologist*, 206(1), 118-126.
- Xu, G., Guo, C., Shan, H., & Kong, H. (2012). Divergence of duplicate genes in exon-intron structure. *Proceedings of the national academy of sciences*, 109(4), 1187-1192.
- Xue, C., Penton, C. R., Shen, Z., Zhang, R., Huang, Q., Li, R., Ruan, Y., & Shen, Q. (2015). Manipulating the banana rhizosphere microbiome for biological control of Panama disease. *Scientific reports*, 5, 11124.
- Yan, M., Jing, W., Xu, N., Shen, L., Zhang, Q., & Zhang, W. (2016). *Arabidopsis thaliana* constitutively active ROP11 interacts with the NADPH oxidase respiratory burst oxidase homologue F to regulate



- reactive oxygen species production in root hairs. *Functional plant biology*, 43(3), 221-231.
- Yang, C., Li, W., Cao, J., Meng, F., Yu, Y., Huang, J., Jiang, L., Liu, M., Zhang, Z., Chen, X., Yamane, H., Zhang, J., Chen, S., Liu, J., & Miyamoto, K. (2017). Activation of ethylene signaling pathways enhances disease resistance by regulating ROS and phytoalexin production in rice. *The Plant journal*, 89(2), 338-353.
- Yoshie, Y., Goto, K., Takai, R., Iwano, M., Takayama, S., Isogai, A., & Che, F. S. (2005). Function of the rice gp91phox homologs *OsrbohA* and *OsrbohE* genes in ROS-dependent plant immune responses. *Plant biotechnology*, 22(2), 127-135.
- Yoshioka, H., Numata, N., Nakajima, K., Katou, S., Kawakita, K., Rowland, O., Jones, J. D. G., & Doke, N. (2003). *Nicotiana benthamiana* gp91phox homologs *NbrbohA* and *NbrbohB* participate in H<sub>2</sub>O<sub>2</sub> accumulation and resistance to *Phytophthora infestans*. *The plant cell*, 15(3), 706-718.
- Yoshioka, H., Sugie, K., Park, H. J., Maeda, H., Tsuda, N., Kawakita, K., & Doke, N. (2001). Induction of plant gp91 phox homolog by fungal cell wall, arachidonic acid, and salicylic acid in potato. *Molecular Plant-Microbe Interactions*, 14(6), 725-736.
- You, J., & Chan, Z. (2015). ROS regulation during abiotic stress responses in crop plants. *Frontiers in plant science*, 6, 1092.
- Zakpaa, H. D., Mak-Mensah, E. E., & Adubofour, J. (2010). Production and characterization of flour produced from ripe apem plantain (*Musa sapientum* L. var. paradisiacal; French horn) grown in Ghana. *Journal of agricultural biotechnology and sustainable development*, 2(6), 92-99.
- Zepeda-Jazo, I., Velarde-Buendía, A. M., Enríquez-Figueroa, R., Bose, J., Shabala, S., Muñiz-Murguía, J., & Pottosin, I. I. (2011). Polyamines interact with hydroxyl radicals in activating Ca<sup>2+</sup> and K<sup>+</sup> transport across the root epidermal plasma membranes. *c*, 157(4), 2167-2180.
- Zhang, X., Köster, P., Schlücking, K., Balcerowicz, D., Hashimoto, K., Kuchitsu, K., Vissenberg, K., & Kudla, J. (2018). CBL1-CIPK26-mediated phosphorylation enhances activity of the NADPH oxidase RBOHC, but is dispensable for root hair growth. *FEBS letters*, 592(15), 2582-2593.
- Zhang, Y., Li, Y., He, Y., Hu, W., Zhang, Y., Wang, X., & Tang, H. (2018). Identification of NADPH oxidase family members associated with cold stress in strawberry. *FEBS open bio*, 8(4), 593-605.
- Zhang, Y., Zhu, H., Zhang, Q., Li, M., Yan, M., Wang, R., Welti, R., Zhang, W., & Wang, X. (2009). Phospholipase Dα1 and phosphatidic acid regulate NADPH oxidase activity and production of reactive oxygen species in ABA-mediated stomatal closure in *Arabidopsis*. *The plant cell*, 21(8), 2357-2377.

Zhou, J., Xia, X. J., Zhou, Y. H., Shi, K., Chen, Z., & Yu, J. Q. (2013). RBOH1-dependent H<sub>2</sub>O<sub>2</sub> production and subsequent activation of MPK1/2 play an important role in acclimation-induced cross-tolerance in tomato. *Journal of experimental botany*, 65(2), 595-607.

Zuo, C., Deng, G., Li, B., Huo, H., Li, C., Hu, C., Kuang, R., Yang, Q., Dong, T., Sheng, O., & Yi, G. (2018). Germplasm screening of *Musa* spp. for resistance to *Fusarium oxysporum* f. sp. cubense tropical race 4 (*Foc* TR4). *European journal of plant pathology*, 151(3), 723-734.

