



UNIVERSITI PUTRA MALAYSIA

**LEAD TOXICITY AND PHYTOEXTRACTION POTENTIAL OF ACACIA
AURICULIFORMIS A. CUNN. EX BENTH. TO REMEDIATE LEAD-
POLLUTED SOIL**

ZERKOUT ABDERRAHMANE

FS 2021 30



**LEAD TOXICITY AND PHYTOEXTRACTION POTENTIAL OF ACACIA
AURICULIFORMIS A. CUNN. EX BENTH. TO REMEDIATE LEAD-POLLUTED
SOIL**

By

ZERKOUT ABDERAHMANE

Thesis Submitted to the School of Graduate Studies,
Universiti Putra Malaysia, in Fulfilment of the Requirements for the
Degree of Doctor of Philosophy

September 2020

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of
the requirement for the degree of Doctor of Philosophy

**LEAD TOXICITY AND PHYTOEXTRACTION POTENTIAL OF ACACIA
AURICULIFORMIS A. CUNN. EX BENTH TO REMEDIATE LEAD-
POLLUTED SOIL**

By

ZERKOUT ABDERRAHMANE

September 2020

Chairman : Muskhazli Mustafa, PhD

Faculty : Science

Phytoremediation takes a longer time to accomplish than other treatment. Phytoextraction with *Acacia auriculiformis*, also known as Auri, Earleaf acacia, Earpod wattle, Northern black wattle, Papuan wattle and Tan wattle, has not been reported previously even though it is a fast-growing species and able to grow well in very poor soil. This study was conducted to determine the impact of lead (Pb) on *A. auriculiformis* seed germination, the influence of Pb on *A. auriculiformis* growth, the capability of *A. auriculiformis* to remove Pb from the soil and the effect of NPK fertilizer on the speed of growth as well as the absorption of Pb from the soil. Seeds were exposed to Pb by soaked it in different Pb concentrations from 0 to 4 g/L (interval of 0.5 g/L). The evaluation of Pb impact on *A. auriculiformis* seed germination and early radicle development was conducted based on germination percentage (GP), germination index (GI), mean germination time (MGT), seedling vigour index (SVI), relative injury rate (RIR) and histological observation on seed embryo development. Biochemical response of *A. auriculiformis* seeds towards Pb stress was assessed by determining the phenolic compounds, malondialdehyde and starch content. Physical growth parameters such as shoot height, leaves number, basal diameter and root length were measured. Several physiological parameters including net photosynthesis, total chlorophylls, internal CO₂ concentration, transpiration rate, the relative water content (RWC) and water use efficiency (WUE) were also measured. The enzymatic respond to Pb toxicity was assessed by measuring the catalase activity. While the concentration of Pb concentration in the soil and plant part was determined using Inductively coupled plasma atomic emission spectroscopy (ICP AES). The bioconcentration factor (BCF) and the translocation factor (TF) were calculated to assess *A. auriculiformis* phytoremediation potential. The effect of the NPK fertilizer with a ratio of 1:1:1 on *A. auriculiformis* growth was assesed by measuring the same morphological and physiological parameters as ststed previously. All the collected data were analysed using one way analysis of variance (ANOVA). via IBM SPSS Statistic Version 25 software. The triplicates data were presented as mean ± standard deviation based on the Tukey's HSD test. Study reveals that Pb toxicity has does not affect *A. auriculiformis* seeds germination up to 1.5 g/L. Increasing the concentration of Pb up to 3.5 g/L led to a reduction in GP and GI while an

increment in the MGT caused the germination to be delayed by almost 10 days. The SVI and RIR results indicated that Pb has also affected seeds strength and ability to repair the damage. High starch accumulation in *A. auriculiformis* seeds caused a reduction in nutrient availability which adversely reduces the embryo development. The assessment of growth parameters also showed that Pb did not affect *A. auriculiformis* grown in 1 g/kg Pb-treated soil. The plant grown in 1 g/kg Pb-treated soil showed a 12% decrement in WUE and RWC, but no significant difference was recorded between 1 g/kg, 2 g/kg and 3 g/kg Pb-treated soil. This is coherence with the proline and catalase responses. Catalase findings showed that Pb caused a significant overproduction of the CAT enzyme in *A. auriculiformis*. Thus, proving that *A. auriculiformis* has a good antioxidant defence system to overcome the Pb stress. Analysis of BCF and TF values showed that values for *A. auriculiformis* grown in 1 g/kg Pb-treated soil were 3.5 and 1.28, respectively. The BCF and TF results indicated that *A. auriculiformis* could be used as a phytoextractor at Pb concentration of 1 g/kg and as phytostabilizer at 2 g/kg. The NPK fertilizer application enhanced *A. auriculiformis* morphological parameters by 55% and an increment was observed in net photosynthesis, total chlorophylls, the transpiration rate, the internal CO₂ concentration and RWC. Proline accumulation was significantly reduced ranging from 15% to 50% depend on the Pb concentration in the soil. However, no significant difference between the CAT enzyme production in the presence and the absence of the NPK fertilizer. Data on BCF and the TF for *A. auriculiformis* treated with the NPK fertilizer showed lower value compared to without NPK. Thus, indicated that in the presence of NPK, *A. auriculiformis* is unable to absorb and translocate a significant amount of Pb from the soil to the different parts of the plant. Based on the all data and tolerance index, *A. auriculiformis* has the potential to be used as a phytoremediation agent for Pb. Combination of its ability to grow in unfertile soil and resistance to Pb up to 2 g/kg does give it some advantage as phytoremediation agent.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai
memenuhi keperluan untuk ijazah Doktor Falsafah

**KETOKSIKAN PLUMBUM DAN POTENSI FITOEKSTRAK OLEH ACACIA
AURICULIFORMIS A. CUNN. EX BENTH UNTUK MEMULIHAN TANAH
YANG DICEMARI PLUMBUM**

Oleh

ZERKOUT ABDERRAHMANE

September 2020

Pengerusi : Muskhazli Mustafa, PhD

Fakulti : Sains

Fitopemulihan mengambil masa yang lebih lama berbanding dengan rawatan lain. Fitopemulihan dengan menggunakan Acacia auriculiformis yang juga dikenali sebagai auri, ‘Earleaf acacia’, ‘Earpod wattle’, ‘Northern black wattle’, ‘Papuan wattle’ dan ‘Tan wattle’ tidak pernah dilaporkan walaupun ia adalah spesis yang tumbuh dengan pantas dan berupaya tumbuh dengan baik pada tanah yang tidak subur. Kajian ini dijalankan untuk menentukan kesan Pb terhadap percambahan benih *A. auriculiformis*, pengaruh plumbum (Pb) terhadap percambahan biji benih *A. auriculiformis*, pengaruh Pb pada pertumbuhan *A. auriculiformis*, keupayaan *A. auriculiformis* untuk mengeluarkan Pb dari tanah dan kesan baja NPK terhadap kadar tumbesaran serta penyerapan Pb dari tanah. Biji benih didedahkan kepada Pb dengan merendamnya di dalam kepekatan Pb berbeza dari 1 sehingga 4 g/L (selang 0.5 g/L). Penilaian kesan Pb terhadap percambahan biji benih *A. auriculiformis* dan perkembangan awal radikal telah dilaksanakan berdasarkan kepada peratus percambahan (GP), indeks percambahan (GI), purata masa percambahan (MGT), index kekuatan benih (SVI), kadar kecederaan relative (RIR) dan pemerhatian histologi perkembangan embrio biji benih. Tindak balas biokimia biji benih *A. auriculiformis* terhadap tekanan Pb dinilai menerusi penentuan kandungan sebatian prolin, malondaildehid dan kanji. Parameter tumbesaran fizikal seperti tinggi pucuk, bilangan daun, diameter pangkal dan panjang akar telah diukur. Beberapa parameter fisiologi termasuk fotosintesis bersih, jumlah klorofil, kepekatan CO₂ dalaman, kadar transpirasi, kandungan relative air (RWC) dan kecekapan penggunaan air (WUE) juga disukat. Tindak balas enzim terhadap ketoksikan Pb dinilai dengan mengukur aktiviti katalase. Sementara itu, pengukuran kepekatan Pb di dalam tanah dan bahagian tumbuhan ditentukan menggunakan ‘Inductively coupled plasma atomic emission spectroscopy’ (ICP AES). Faktor biokepekatan (BCF) dan faktor translokasi (TF) telah dikira untuk menilai potensi fitopemulihan *A. auriculiformis*. Kesan baja NPK dengan kadar 1:1:1 keatas pertumbuhan *A. auriculiformis* telah dinilai dengan mengukur parameter morfologi dan fisiologi yang sama seperti yang dinyatakan sebelum ini. Semua data yang dikumpul telah dianalisis menggunakan analisis varians satu hala (ANOVA) menggunakan perisian IBM SPSS Statistic Version 25. Data tiga replikat ditunjukkan sebagai purata ± piawai sisihan berdasarkan kepada Tukey’s HSD test. Kajian

mendedahkan bahawa ketoksikan Pb tidak memberikan kesan keatas percambahan biji benih *A. auriculiformis* sehingga 1.5 g/L. Peningkatan kepekatan Pb sehingga 3.5 g/L menyebabkan penurunan dalam GP dan GI manakala peningkatan MGT menyebabkan penagguhan percambahan sehingga hampir 10 hari. Keputusan SVI dan RIR menunjukkan bahawa Pb juga memberi kesan terhadap kekuatan dan keupayaan biji benih membaik pulih kerosakan. Pengumpulan kanji yang tinggi di dalam biji benih *A. auriculiformis* menyebabkan penurunan nutrien tersedia yang mengurangkan perkembangan embrio. Penilaian parameter pertumbuhan juga telah menunjukkan bahawa Pb tidak memberikan kesan terhadap *A. auriculiformis* yang tumbuh di dalam tanah yang rawat dengan 1 g/kg Pb. Pokok yang tumbuh di dalam tanah yang rawat dengan 1 g/kg Pb menunjukkan 12 penurunan dalam WUE dan RWC, tetapi tiada perbezaan bererti dicatatkan diantara tanah yang rawat dengan 1 g/kg, 2g/kg dan 3 g/kg Pb. Ini selaras dengan tindak balas prolin dan katalase. Dapatkan katalase menunjukkan bahawa Pb telah menyebabkan penghasilan yang melampau enzim CAT di dalam *A. auriculiformis*. Oleh itu, ianya membuktikan bahawa *A. auriculiformis* mempunyai sistem pertahanan antioksida yang baik untuk mengatasi tekanan Pb. Analisis terhadap nilai BCF dan TF menunjukkan bahawa nilai bagi *A. auriculiformis* yang ditanam pada tanah yang rawat dengan 1 g/kg Pb ialah masing-masing pada 3.5 dan 1.28. Keputusan BCF dan TF menunjukkan bahawa berupaya *A. auriculiformis* boleh digunakan sebagai fitopengekstrakan pada kepekatan Pb 1 g/kg dan sebagai fitopenstabilan pada 2 g/kg. Penggunaan baja NPK meningkatkan parameter morfologi *A. auriculiformis* sebanyak 55% dan peningkatan dapat dilihat dalam fotosintesis bersih, jumlah klorofil, kadar transpirasi, kepekatan CO₂ dalam dan RWC. Pengumpulan prolin dikurangkan dengan berkesan dari julat 15% sehingga 50%, bergantung kepada kepekatan Pb di dalam tanah. Walau bagaianapun, tiada perbezaan bererti diantara penghasilan enzim CAT semasa kehadiran dan tanpa baja NPK. Data BCF dan TF bagi *A. auriculiformis* yang dirawat dengan baja NPK menunjukkan nilai yang lebih rendah berbanding dengan tanpa NPK. Oleh itu, ianya menunjukkan bahawa dengan kehadiran baja NPK, *A. auriculiformis* tidak berupaya menyerap dan menindahkan secara berkesan sejumlah Pb dari tanah ke bahagian berlainan pada pokok. Berdasarkan kepada semua data dan index toleransi, *A. auriculiformis* mempunyai potensi untuk digunakan sebagai agent fitopemulihan bagi Pb. Gabungan kebolehannya untuk tumbuh pada tanah yang tidak subur dan tahan terhadap Pb sehingga 2 g/kg telah memberikan beberapa kelebihan sebagai agen fitopemulihan.

ACKNOWLEDGEMENTS

First and foremost, praises and thanks to **Allah**, the Almighty, for His showers of blessings throughout my research work to complete the research successfully.

I am extremely grateful to my parents for their love, prayers, caring and sacrifices for educating and preparing me for my future, may **Allah** reward you with Jannatul Firdaus. Special thanks go to my one and only brother Ammar for his support, valuable and courageous advice. I am also very much thankful to my wife Narimen for her love, understanding, prayers and continuing support to complete this research work. I thank her for the moral and emotional support, may **Allah** bless you and keep you always by my side.

I would like to express my deep and sincere gratitude to the chairman of my supervisory committee Associate Professor Dr. Muskhazli Mustafa for giving me the opportunity to do research and providing invaluable guidance throughout this research. He has taught me the methodology to carry out the research and to present the research works as clearly as possible. It was a great privilege and honor to work and study under his guidance. I am extremely grateful for what he has offered me. I would like to thank my supervisor committee Dr. Mohd Hafiz bin Ibrahim and Associate Professor Dr. Hishamuddin bin Omar for helping with this research project.

I would like to say thanks to my friends and research colleagues especially Anna for her dedicated support and for continuously provided encouragement. Finally, many thanks to all participants that took part in the study and enabled this research to be possible directly or indirectly.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Muskhasli Bin Mustafa, PhD

Associate Professor

Faculty of Science

Universiti Putra Malaysia

(Chairman)

Mohd Hafiz Bin Ibrahim, PhD

Senior Lecturer

Faculty of Science

Universiti Putra Malaysia

(Member)

Hishamuddin Bin Omar, PhD

Senior Lecturer

Faculty of Science

Universiti Putra Malaysia

(Member)

Zalilah Mohd Shariff, PhD

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date: 11 March 2021

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	vii
LIST OF TABLES	viii
LIST OF FIGURES	xiii
LIST OF ABBREVIATIONS	xiv
xvi	
 CHAPTER	
1 INTRODUCTION	
1.1 Background of the study	1
1.2 Problem statement	2
1.3 Significant of the study	4
1.3 Objective of the study	5
2 LITERATURE REVIEW	
2.1 Heavy metal	7
2.2 Lead (Pb)	8
2.2.1 History of Pb	8
2.2.2 Physico-chemical properties of Pb	8
2.2.3 Biogeochemical cycle of Pb	9
2.2.4 Origins of Pb pollution	10
2.2.5 Different lead uses	11
2.3 Lead in soil: Origin and mobility	13
2.4 Lead in plant: Phytoavailability and absorption	16
2.4.1 Root absorption mechanism of Pb	17
2.4.2 Pb mobility and sequestration in plant	20
2.4.3 Pb impact on plant	21
2.4.3.1 Oxidative stress generation	22
2.4.3.2 Effects of Pb on plant physiology and development	25
2.5 Lead pollution in Malaysia	29
2.6 Plant defence strategies for Pb stress	30
2.6.1 Antioxidant system	30
2.6.1.1 Enzymatic antioxidants	31
2.6.1.2 Non enzymatic antioxidants	32
2.6.2 Control of Pb toxicity in plants	34
2.6.2.1 Cell activities as a Pb toxicity barrier	35
2.6.2.2 Metal exclusion strategy	36
2.7 Phytoremediation	38
2.7.1 Phytoremediation techniques	38
2.7.2 Phytoremediation benefits and limitations	41
2.8 Overview of the <i>Acacia auriculiformis</i>	42

3 INFLUENCE OF LEAD ON IN VITRO SEED GERMINATION AND EARLY RADICLE DEVELOPMENT OF ACACIA AURICULIFORMIS A. CUNN. EX BENTH.	
3.1 Introduction	46
3.2 Materials and methods	47
3.2.1 Materials preparation	47
3.2.2 Experimental setup	48
3.2.3 Impact of Pb on Seed Germination	49
3.2.4 Biochemical Response of <i>A. auriculiformis</i> seeds towards Pb stress	51
3.2.4.1 Determination of total phenolics contents	51
3.2.4.2 Determination of Malondialdehyde (MDA)	52
3.2.4.3 Determination of starch content	53
3.2.5 Plant embryo development	55
3.2.6 Radicle development	55
3.3 Results and Discussion	56
3.3.1 The effect of Pb on the germination parameters	56
3.3.2 Impact of Pb on selected biochemical responses in <i>A. auriculiformis</i> seeds	59
3.3.3 Effect of Pb on <i>A. auriculiformis</i> seeds embryo	63
3.3.4 Impact of Pb on <i>A. auriculiformis</i> radicle development	65
3.4 Conclusion	68
4 ASSESSMENT OF ACACIA AURICULOFORMIS A. CUNN. EX BENTH. TOLERANCE AND LEAD REMOVAL CAPABILITY VIA PHYTOEXTRACTION	
4.1 Introduction	69
4.2 Materials and methods	70
4.2.1 Growth experiment and parameters measurement	70
4.2.2 Measurement of physiological parameters	71
4.2.3 Determination of Proline and Catalase enzymes in leaves	72
4.2.3.1 Proline determination	72
4.2.3.2 Determination of Catalase	74
4.2.4 Quantification of Pb in plant parts	75
4.2.5 Quantification of macronutrients in <i>Acacia auriculiformis</i> plants	75
4.2.6 Determination of Pb removal factor	76
4.2.7 Statistical analysis	76
4.3 Results and discussions	76
4.3.1 The effect of Pb on <i>A. auriculiformis</i> growth parameters	76
4.3.2 The effect of Pb on <i>A. auriculiformis</i> physiological parameters	80
4.3.3 Proline and Catalase response towards Pb in <i>A. auriculiformis</i>	82
4.3.4 Pb storage capability in different part of <i>A. auriculiformis</i>	85
4.3.5 Impact of Pb on the distribution of macronutrients in <i>A. auriculiformis</i> plant	88
4.3.6 <i>Acacia auriculiformis</i> potential to absorb and accumulate Pb	91
4.4 Conclusion	94

5 EVALUATE THE EFFECT OF THE NPK FERTILIZER ON THE SPEED OF GROWTH AND THE ABSORPTION OF LEAD FROM THE SOIL BY ACACIA AURICULIFORMIS A. CUNN. EX BENTH.	
5.1 Introduction	95
5.2 Materials and methods	96
5.3 Results and discussions	98
5.3.1 The effect of NPK fertilizer on <i>A. auriculiformis</i> growth parameters	98
5.3.2 The effect of NPK fertilizer on <i>A. auriculiformis</i> physiological parameters	103
5.3.3 The effect of NPK fertilizer on Proline and Catalase response towards Pb toxicity in <i>A. auriculiformis</i> plants	108
5.3.4 Impact of NPK fertilizer on Pb storage capability in the soil and different part of <i>A. auriculiformis</i> plants	110
5.3.5 Impact of NPK fertilizer on <i>A. auriculiformis</i> potential to absorb and accumulate Pb	112
5.4 Conclusion	114
6 GENERAL DISCUSSIONS	
7 CONCLUSION AND FUTURE RECOMMENDATIONS	
7.1 Conclusion	118
7.2 Future Recommendations	119
REFERENCES	121
APPENDICES	147
BIODATA OF STUDENT	155
LIST OF PUBLICATIONS	156

LIST OF TABLES

Table		Page
2.1	Major physico-chemical properties of lead	10
2.2	Systematic classification of <i>A. auriculiformis</i> species (Wee, 2003)	44
3.1	Physical proprieties of Lead sulphate	49
3.2	Germination parameter respons of <i>A. auriculiformis</i> seed exposed to selected different concentration of Pb	57
4.1	The effect of Pb on the net photosynthesis, total chlorophylls, transpiration rate, water use efficiency, relative water content and the internal CO ₂ concentration in <i>A. auriculiformis</i> leaves after 90 days planted in soil supplemented with Pb	80
5.1	Comparison between different growth parameters in <i>A. auriculifomris</i> plants supplemented with the NPK and without the NPK fertilizer after 12 weeks of growth	101
5.2	Comparison of the effect of NPK fertilizer and without NPK on the concentration of Pb in soil before and after planting <i>A. auriculiformis</i> and in different plant parts	114

LIST OF FIGURES

Figure		Page
1.1 Summary of research design and attainment of objectives		7
2.1 Biogeochemical cycle of Pb elements		11
2.2 Movement of metal elements from the root surface to the central cylinder		20
2.3 The main ROS production cascade: in orange non-radical ROS; in yellow radical ROS		25
2.4 Phytoremediation techniques (Pilon-Smits, 2005)		40
2.5 <i>Acacia auriculiformis</i> (a) seeds, (b) first leaves, (c) tree and (d) second leaves and flowers (Aggarwal, 1988; Wee, 2003)		45
2.6 Origin and distribution of <i>Acacia auriculiformis</i> (Chakraborty <i>et al.</i> , 2019)		45
3.1 <i>Acacia auriculiformis</i> (a) mature tree; (b) mature seeds.		49
3.2 <i>Acacia auriculiformis</i> seeds placed on filter paper and soaked with selected concentration of Pb		50
3.3 Biochemical response (a) Total phenolics, (b) Maliondialdehyde (MDA) and (c) Starch content in <i>A. auriculiformis</i> seeds when exposed to selected concentration of Pb for 15 days		61
3.4 Comparison in <i>A. auriculiformis</i> embryo development and germination in selected Pb treatment		65
3.5 Effects of different concentrations of lead on (a) the radicle length; (b) the tolerance index and (c) comparison of radicle morphology of <i>A. auriculiformis</i> seedlings		67
4.1 The effect of different Pb concentrations on shoot height, basal diameter, leaves numbers and root length of <i>A. auriculiformis</i> plants after twelve weeks of growth		77
4.2 The effect of Pb on the shoot height; the leaves number; the basal diameter and the root length of <i>A. auriculiformis</i> plants		78
4.3 The effect of Pb on the Proline and Catalase activities in <i>A. auriculiformis</i> leaves		84
4.4 The concentration of Pb in the soil before and after planting <i>A. auriculiformis</i> plant, and in the different part of the plant		86

4.5	The effect of Pb toxicity on the absorption of macronutrients in <i>A. auriculiformis</i> plants	90
4.6	Values of the bioconcentration factor (BCF), and translocation factor (TF) in <i>A. auriculiformis</i> under different Pb concentration	92
5.1	The effect of the NPK fertilizer on <i>A. auriculiformis</i> growth under different Pb concentrations after twelve weeks	100
5.2	Comparison between different growth parameters in <i>A. auriculifomris</i> plants (a) supplemented with the NPK and (b) without the NPK fertilizer in the presence of Pb	101
5.3	Comparision of the effect of NPK fertilizer and without NPK on (a) Net Photosynthesis, (b) Total Chlorophylls, (c) Internal CO ₂ Concentration, (d) Transpiration Rate, (e) Water Use Efficiency, and (f) Relative Water Content in <i>A. auriculiformis</i> leaves after 90 days	106
5.4	The effect of NPK fertilizer on the Proline and Catalase activities in <i>A. auriculiformis</i> leaves under different Pb concentrations	110
5.5	Impact of the NPK fertilizer on the bioconcentration factor (BCF), and translocation factor (TF) Values in <i>A. auriculiformis</i> under different Pb concentration	115

LIST OF ABBREVIATIONS

ANOVA	Analysis of variance
AAS	Atomic Absorption Spectrophotometer
ASC	Ascorbic acid
BCF	Bioconcentration Factor
Ca	Calcium
CAT	Catalases
Cd	Cadmium
CDF	Cation Diffusion Facilitators
Cu	Copper
CNGC	Cyclic nucleotide gated channel
DNA	Deoxyribonucleic acid
DMSO	Dimethyl sulfoxide
DW	Dry weight
EDL	Electron Discharge Lamp
FAA	Formalin-Acetic Acid- Ethanol
FCR	Folin Ciocalteu Reagent
FW	Fresh Weight
g	Gram
GP	Germination Percentage
GI	Germination index
GSH	Glutathione
HMW	High Molecular Weight
H ₂ O ₂	Hydrogen peroxide

h	Hour
ICP-OES	Inductively coupled plasma optical emission spectrophotometer
K	Potassium
kg	Kilogram
LMW	Low Molecular Weight
L	Litre
MTE	Metal Trace Element
mg	Milligram
min	Minute
MGT	Mean Germination Time
MDA	Malondialdehyde
MCW	Methanol:Chloroform:Water
Mg	Magnesium
NSC	Non-Structural Carbohydrate
N	Nitrogen
Ni	Nickel
NRAMP	Natural Resistance-Associated Macrophage Protein
O₂⁻	Superoxide anion
OH	Hydroxyl radical
OH⁻	Hydroxyl anion
Pb	Lead
PCR	Photosynthetic carbon reduction
PCs	Phytochelatins
POD	Peroxidases
Pi	Orthophosphate

P	Phosphorus
ROS	Reactive Oxygen Species
RIR	Relative Pb Injury Rate
RNA	Ribonucleic acid
RO·	Alkoxy radical
RWC	Relative water content
SOD	Superoxide-dismutase
SVI	Seedling Vigor Index
TBA	Thiobarbituric acid
TCA	Trichloroacetic acid
TF	Translocation factor
TPC	Total phenolic contents
TI	Tolerance index
TW	Turgescence weight
WUE	Water use efficiency
Zn	Zinc
ZIP	Zinc-regulated transporter, Iron-regulated transporter, Protein

CHAPTER 1

INTRODUCTION

1.1 Background of the study

Heavy metal pollution has developed as a horrible ecological crisis. It is harmful not only to human beings but also to all other living organisms (Raymond *et al.*, 2011). Rapid industrialization, urbanization and human activities have caused serious environmental hazard in the biosphere (Nagajyoti *et al.*, 2008). Heavy metals are accumulated in soils through precipitation, runoff, spillage, dumping of waste or by-product from industrial areas, mining activities, disposal of industrial, agriculture and domestic wastes (Ali *et al.*, 2013a). Heavy metals do not remain in the soil indefinitely but transferred to the biotic food chain through the primary producer, plant (Sherene, 2010); which threaten their survival, affecting their growth and metabolism (Maryam *et al.*, 2014; Gangaiah *et al.*, 2013)

Heavy metals are classified into two categories: essential heavy metals and non-essential heavy metals (Raymond *et al.*, 2011). The essential heavy metals are required in plant growth such as Zn, Cu and Fe. The non-essential heavy metals such as Pb, Cd and Ni in high concentration can interfere with other nutrient uptake, physiological functions genetic integrity and others (Mihiri *et al.*, 2016). Furthermore, heavy metal pollution may influence the formation of the embryo sac, the vacuolation of embryonic cells, and degeneration of embryo in the late stage of heart-shaped embryo (Mohsenzadeh *et al.*, 2011).

There are many studies on the effect of heavy metals on plants such as germination study, plant development, the effect on DNA, gene expression, respiration, secondary metabolite, heavy metal distribution and accumulation levels (Anișoara *et al.*, 2010; Dane *et al.*, 2010; Chandra & Kang, 2016; Buscaroli, 2017; Rehman *et al.*, 2018). Different plant species responded differently to different heavy metals (Hu *et al.*, 2012). Some plants can accumulate heavy metals to a varying degree without showing physical defect (Amin *et al.*, 2018) and these attributes make plant a very interesting research subject particularly on heavy metals and its primary role in spreading heavy metal in the food chain.

1.2 Problem statement

Among the non-essential heavy metal, lead (Pb) is the most frequently encountered persistent heavy metal in polluted environment (Pourrut *et al.*, 2011a), and it was classed as the second most harmful substance after Arsenic (Patra *et al.*, 2004). Exposure to Pb was identified as a serious public health problem, particularly for children (Nawab *et al.*, 2015). The exploitation of various types of deposits in the ore leads to the accumulation of Pb on the surface (Lutts & Lefevre, 2015), and due to its non-biodegradability, toxicity,

and accumulation, it can enter the human body through intake of food (65%), water (20%) and air (15%), thereby causing hazardous damage to human health, such as kidney damage, disruption of nervous systems, brain damage, high blood pressure, and disruption of the biosynthesis of hemoglobin causing anemia (Zhang *et al.*, 2015; Lei *et al.*, 2016).

The main sources of Pb contamination are anthropogenic releases (Salazar & Pignata, 2014). It comes mostly from paints, cable covering, fertilisers and pesticides, and metal production (Arruti *et al.*, 2010). In soil, paint is the major contributor to Pb contamination; it held tightly on the surfaces of very fine clay and organic matter particles (Srinivasan *et al.*, 2014). In Malaysia, Pb was found to be the second highest metal in contaminated soil after Cu in Perlis state, where the pollution index of Cu and Pb noted the highest by 11 and 6% respectively (Siti *et al.*, 2014).

The issue of Pb in contaminated agricultural soil has been a question of great interest in a wide range of fields (Abbasi *et al.*, 2017). Therefore, several remediation techniques have emphasized on the removal of Pb from soil including soil replacement method, thermal desorption, chemical leaching, chemical fixation, electro-kinetic remediation, and vitrify technology (Yao *et al.*, 2012), but none of them seems to performed satisfactorily. The major issues with these techniques are the increase risk of spreading contaminated soil or dust particles during removal, transport of contaminated soil, and the relatively high cost (Bang *et al.*, 2015).

In recent years, it has been reported that plants can help to reduce or remove heavy metals from the soil by their roots, with the aim of restoring area sites to a condition useable for private or public applications (Couselo *et al.*, 2010; Masvoda *et al.*, 2013; Ochonogor & Atagana, 2014). Phytoremediation has become the way out to extract heavy metal properly without affecting the environment and potentially cost-effective (Illié *et al.*, 2015) especially the phytoextraction technique. Phytoextraction is an important aspect of phytoremediation; it consists especially of the extraction of heavy metals from soil and stored them in roots, shoots and leaves (Souza *et al.*, 2013). Disadvantages for using this technique is that it takes longer time than other treatments, due to plant limitation, where in most cases high contaminant concentration can reduce the speed of plant growth (Laghlimi *et al.*, 2015; Ali *et al.*, 2013b; Moosavi & Mohamad, 2013). However, these drawbacks can be reduced or eliminated through intensive research on identification of fast growing and high heavy metals tolerance plants. The choice of phytoremediation candidates is important. Some plants can be short-lived, too small to be significant as pool to contain heavy metals or serves as food for herbivores. Ideal phytoremediation agents should be a hardy plant, long-lived, fast growing, big and inedible.

1.3 Significance of the study

Acacia plants fit the above description perfectly, they are adapted to a wide range of environments, both tropical and temperate, and this adaptability has made them popular for planting on degraded lands in Asia and elsewhere (Turnbull *et al.*, 1997). Acacias have the potential to rehabilitate the soil through absorption and storage of heavy metals in its leaves, shoots and roots which make them the best phytoremediation candidates (Yang *et al.*, 2005). The phytoremediation potential of acacias has been previously

reported in numerous studies. Majid *et al.*, (2012) showed that *A. mangium* can absorb up to 36.25 ppm of Pb with total plant biomass of 115.39 g in 3 months. Another study by Mohd *et al.*, (2013) showed that the aboveground biomass of a single *A. mangium* tree is projected to accumulate 5.74 g of Zn, 1.15 g of Cu and 0.06 g of Cd in 12 months. It is clearly observed that heavy metals toxicity affects the biomass yield and the speed of growth of *A. mangium* thus reduce its absorption potential and phytoremediation efficiency. Therefore, it is necessary to find other *Acacia* plants that can resist higher concentrations and absorb more toxic metals in their parts.

Acacia auriculiformis could be a perfect candidate in this matter. It has a high biomass yield and adapts well to degraded soil conditions, poor soil quality, even in saline and seasonally waterlogged soils and dry season (Lokman *et al.*, 2009). It is a fast-growing multipurpose tree species in the Leguminosae family that can reach 30 m of height and 30cm of diameter (Gilman & Watson 2013). Its economic impact is predominantly positive, and it is one of the most favoured trees in degraded sites (Ishiguri *et al.*, 2004). However, phytoremediation using *Acacia auriculiformis* species on Pb contaminated soils has never been reported. Based on the attributes mentioned, theoretically, *A. auriculiformis* should be able to tolerate, absorb and accumulate a large amount of Pb from the soil providing a perfect remediation technique to clean up Pb from the soil. If this is the case, the timber from this tree can be used as a building material for the house which can last for 50 years or more (Chakraborty *et al.*, 2019). *Acacia auriculiformis* is an exotic species mostly planted as roadside avenue tree by the side of highways, railway tracks and parks. It is only recently its invasive potential has been noted (Olatunja *et al.*, 2012).

1.4 Objective of the study

Based on the positive attributes of *A. auriculiformis*, the main aim of the study is to assess the potential of using *A. auriculiformis* that grows fast as a phytoremediator agent to absorbs a considerable amount of Pb from the soil. The outline of the research approach is shown in Figure 1.1 and the design of the study was based on the following specific objectives;

- i. To determine the impact of Pb on *A. auriculiformis* seed germination pattern and early seedling development.
- ii. To evaluate the ability of *A. auriculiformis* plant to grow on Pb contaminated soil.
- iii. To determine the distribution of Pb in different parts of *A. auriculiformis* plant.
- iv. To assess the effect of NPK fertiliser on *A. auriculiformis* growth and heavy metal absorption capability.

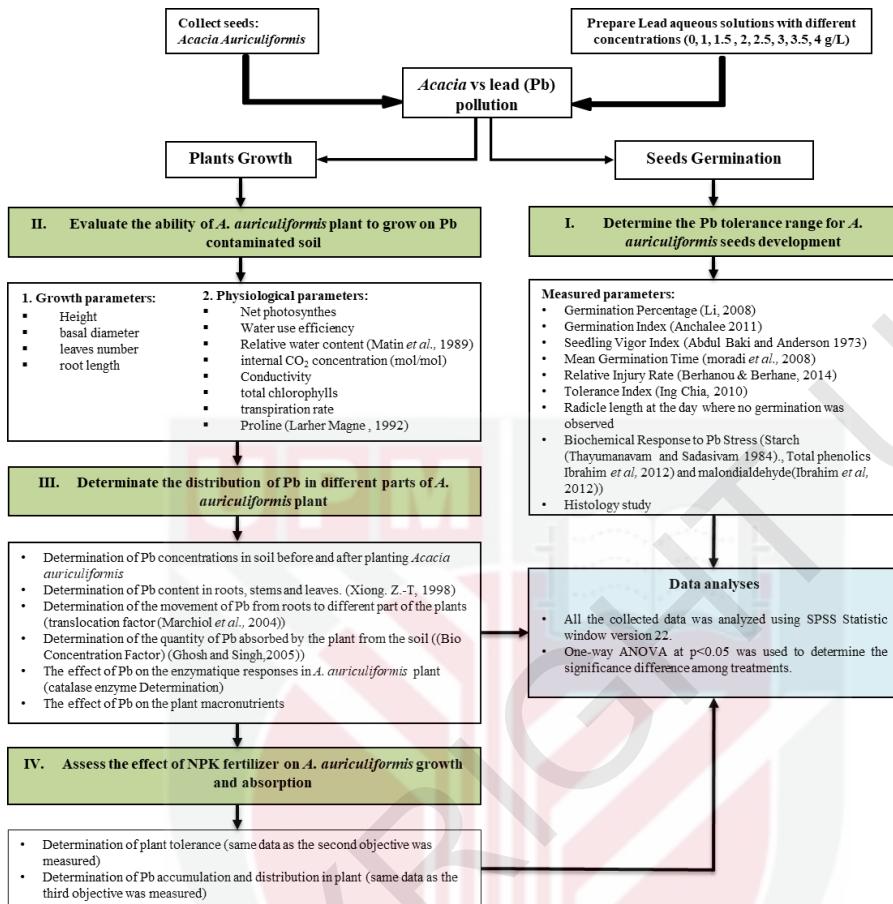


Figure 1.1: Summary of research design and attainment of objectives

REFERENCES

- Abbasi, H., Pourmajidian, M.R., Hodjati, S.M., Fallah, A., & Nath, S. (2017). Effect of soil-applied lead on mineral contents and biomass in *Acer cappadocicum*, *Fraxinus excelsior* and *Platycladus orientalis* seedlings. *iForest - Biogeosciences and Forestry*, 10: 722-728.
- Abuye, C., Urga, K., Knapp, H., Selmar, D., Omwega, A.M., Imungi, J.K., & Winterhalter, P. (2003). A compositional study of *Moringa stenopetala* leaves. *East African Medical Journal*, 80: 247-252.
- Adhikari, T., Kumar, A., Singh, M.V., & Rao, A.S. (2014). Phytoaccumulation of lead by selected wetland plant species. *Communications in Soil Science and Plant Analysis*, 41: 2623-2632.
- Adesmoye, A.O., & Kloepper, J.W. (2009). Plant-microbes interactions in enhanced fertilizer use efficiency. *Applied Microbiology and Biotechnology*, 85: 1-12.
- Afshan, S., Ali, S., Bharwana, S.A., Rizwan, M., Farid, M., Abbas, M., Ibrahim, F., Mehmood, M.A., & Abbasi, G.H. (2015). Citric acid enhances the phytoextraction of chromium, plant growth and photosynthesis by alleviating the oxidative damages in *Brassica napus L.* *Environmental Science and Pollution Research*, 22: 11679-11689.
- Agati, G., Azzarello, E., Pollastri, S., & Tattini, M. (2012). Flavonoids as antioxidants in plants: location and functional significance. *Plant Science*, 196: 67-76.
- Aggarwal, N. (1988). Over half a million Singapore trees go on computer. *The Straits Times Newspaper*, Singapore.
- Ahmad, A., Hadi, F., & Ali, N. (2015). Effective phytoextraction of cadmium (Cd) with increasing concentration of total phenolics and free proline in *Cannabis sativa (L)* plant under various treatments of fertilizers, plant growth regulators and sodium salt. *International Journal of Phytoremediation*, 17: 56-65.
- Ahmad, M.S.A., Ashraf, M., & Hussain, M. (2011). Phytotoxic effects of nickel on yield and concentration of macro- and micro-nutrients in sunflower (*Helianthus annuus L.*) achenes. *Journal of Hazardous Materials*, 185: 1295-1303.
- Ahmad, M.S., & Ashraf, M. (2011). Essential roles and hazardous effects of nickel in plants. *Reviews of Environmental Contamination and Toxicology*, 214: 125-167.
- Ali, A., Iqbal, N., Ali, F., Afzal, B., & Nicholson, G. (2012). *Alternanthera bettzickiana* (Regel) a potential halophytic ornamental plant. *Flora- Morphology, Distribution, Functional Ecology of Plants*, 20: 318-321.
- Ali, H., Ezzat, K., & Muhammad, A. (2013a). Chemosphere phytoremediation of heavy metals Concepts and Applications. *Chemosphere*, 9: 869-81.

- Ali, H., Khan, E., & Sajad, M.A. (2013b). Phytoremediation of Heavy Metals-Concepts and Applications. *Chemosphere*, 91: 869-881.
- Al-Akeel, K. (2016). Lead uptake, accumulation and effects on plant growth of common reed (*Phragmites australis* (Cav.) Trin. Ex Steudel) plants in hydroponic culture. *International Journal of Agricultural and Biological Engineering*, 3: 2349-1531.
- Ali, R., Mahmoud, M.H., Abbas, H., & Fakhr, M. (2017). Physiological studies on the interactive effects of lead and antioxidants on *Carum carvi* plant. *Egyptian Journal of Botany*, 57: 317-333.
- Ali, M., Khadijeh K., Salman A. A., & Reza T. (2014). Lead Accumulation Potential in *Acacia victoria*. *International Journal of Phytoremediation*, 16: 582-592.
- Almansouri, M., Kinet, J.M., & Lutts, S. (2001). Effect of salt and osmotic stresses on germination in durum Wheat (*Triticum durum Desf.*). *Plant and Soil*, 231: 243-254.
- Alscher, R.G., Erturk, N., & Heath, L.S. (2002). Role of superoxide dismutases (SODs) in controlling oxidative stress in plants. *Journal of Experimental Botany*, 53: 1331-1341.
- Amin, H., Arain, B.A., Jahangir, T.M., Abbasi, M.S., & Amin, F. (2018). Accumulation and distribution of lead (Pb) in plant tissues of guar (*Cyamopsis tetragonoloba L.*) and sesame (*Sesamum indicum L.*): profitable phytoremediation with biofuel crops. *Geology, Ecology and Landscapes*, 2: 51-60.
- Ambika, A., Mohnish, P., & Kumar, N. (2016). Effect of Heavy Metals on Plants: An Overview. *International Journal of Application or Innovation in Engineering & Management*. 5: 2319-4847.
- Anwer, S., Khan, S., Ashraf, M.Y., Noman, A., Baloch, S.U., Zafar, S., & Fahad, S. (2017). Impact of chelators induced phytoextraction of cadmium on yield and ionic uptake of maize. *International Journal of Phytoremediation*, 19: 505-513.
- Anchalee, J. (2011). Effects of Different Light Treatments on the Germination of *Nepenthes mirabilis* International Transaction. *Journal of Engineering, Management, Applied Sciences & Technologies*, 2: 1906-9642.
- Anișoara, S., Lupuleac, R., Costică, N., & Alexandrina, M. (2010). The Influence of Certain Heavy Metals on Seeds Germination at *Lens culinaris* medik and *Pisum sativum L.* *Lucrări Științifice*, 20: 25-30.
- Anna, M. R., & Idupulapati, M. R. (2005). 7 Role of Phosphorus in Photosynthetic Carbon Metabolism. *Handbook of Photosynthesis*. Taylor & Francis Group. Poland.
- Anjum, N.A., Umar, S., Ahmad, A., Iqbal, M., & Khan, N.A. (2008). Sulphur protects mustard (*Brassica campestris L.*) from cadmium toxicity by improving leaf ascorbate and glutathione. *Plant Growth Regulation*, 54: 271-279.

- Apel, K., & Hirt, H. (2004). Reactive oxygen species: metabolism, oxidative stress, and signal transduction. *Annual Review of Plant Biology*, 55: 373-399.
- Aravind, P., & Prasad, M.N.V. (2005). Modulation of cadmium-induced oxidative stress in *Ceratophyllum demersum* by zinc involves ascorbate-glutathione cycle and glutathione metabolism. *Plant Physiology and Biochemistry*, 43: 107-116.
- Ariès, S. (2001). Mise en évidence de contaminations métalliques historiques à partir de l'étude d'enregistrements sédimentaires de lacs de haute montagne. Thèse de doctorat, Université de Toulouse 3, Toulouse.
- Ariani, E. (2009). Effect of NPK fertilizers 16:16:16 and various types of mulch for chili plant yield (*Capsicum annum L.*). *Sagu*, 8: 5-9.
- Arora, A., Sairam, R., & Srivastava, G. (2002). Oxidative stress and antioxidative system in plants. *Current Science*, 82: 1227-1238.
- Arruti, A., Fernández-Olmo, I., & Irabien, A. (2010). Evaluation of the contribution of local sources to trace metals levels in urban PM2.5 and PM10 in the Cantabria region (Northern Spain). *Journal of Environmental Monitoring*, 12: 1451-1458.
- Arya, S. K., & Roy, B. K. (2011). Manganese induced changes in growth, chlorophyll content and antioxidants activity in seedlings of broad bean (*Vicia faba L.*). *Journal of Environmental Biology*, 32: 707-711.
- Ashraf, M., & Foolad, M.R. (2007). Roles of glycine betaine and proline in improving plant abiotic stress resistance. *Environmental and Experimental Botany*, 59: 206-216.
- Ashraf, U., Kanul, A.S., Deng1, Q., Mo, Z., Pan1, S., Tian, H., & Tang, X. (2017). Lead (Pb) toxicity: physio-biochemical mechanisms, grain yield, quality, and Pb distribution proportions in scented rice. *Frontiers in Plant Science*, 8: 259-276.
- Asghar, A., Ali, A., Syed, W. H., Asif, M., Khaliq, T., & Abid, A. A. (2010). Growth and yield of maize (*Zea mays L.*) cultivars affected by NPK application in different proportion. *Pakistan Journal of Science*, 62: 211-216.
- Bang, J., Kamala, K., Lee, S., Cho, K.J., Kim, M., Kim, C.H., Bae, Y.J., Kim, J.H., Myung, K.H., & Oh, H. (2015). Phytoremediation of heavy metals in contaminated water and soil using *Miscanthus* sp. Goedae Uksae 1. *International Journal of Phytoremediation*, 17: 515-520.
- Baize, D. (2002). Les éléments traces métalliques dans les sols: Approches fonctionnelles et spatiales. INRA éditions.
- Barrs, H.D., & Weatherley, P.E. (1962). A Re-Examination of the Relative Turgidity Techniques for Estimating Water Deficits in Leaves. *Australian Journal of Biological Sciences*, 15: 413-428.
- Bâtit, M.B., Candan F., Buyuk I., & Aras S. (2015). The determination of physiological and DNA changes in seedlings of maize (*Zea mays L.*) seeds exposed to the waters

- of the Gediz River and copper heavy metal stress. *Environmental Monitoring and Assessment*, 187: 169-178.
- Babatola, L.A. (2006). Effects of NPK on performance and storage life of okra (*Abelmoschus esculentus*). Proceedings of the Horticultural Society of Nigeria Conference Lagos, Nigeria, 125-128.
- Berhanu, A. T., & Berhane G. (2014). The effect of salinity (NaCl) on germination and early seedling growth of *Lathyrus sativus* and *Pisum sativum* var. *abyssinicum*. *African Journal of Plants Science*, 8: 225-231.
- Bernhard A. Z., Che F. I., Mike J. M., & Gill C. (2004). Heavy metals in soils and crops in southeast Asia. *Environmental Geochemistry and Health*, 26: 343-357.
- Beraud, E. (2007). Etude des effets genotoxiques et de l'induction des phytochelatines chez *Vicia faba* (Fabaceae) exposée au cadmium: Application du test Vicia-micronoyaux à des matrices complexes. These de doctorat, Université de Lorraine, Metz.
- Benavides, M.P., Gallego, S.M., & Tomaro, M.L. (2005). Cadmium toxicity in plants. *Brazilian Journal of Plant Physiology*, 17: 21-34.
- Bhargava, A., Carmona, FF., Bhargava, M., & Srivastava, S. (2012). Approaches for enhanced phytoextraction of heavy metals. *Journal of Environmental Management*, 105:103-120.
- Brunet, J., Reppelin, A., Varralult, G., Terryn, N., & Zuily-Fodil, Y. (2008). Lead accumulation in the roots of grass pea (*Lathyrus sativus L.*). *Comptes Rendus Biologies*, 331: 859-864.
- Branvall, M.L., Bindler, R., Emteryd, O., & Renberg, I. (2001). Four thousand years of atmospheric lead pollution in northern Europe: a summary from Swedish lake sediments. *Journal of Paleolimnology*, 25: 421-435.
- Brown, K., & Van den Driessche, R. (2005). Effects of nitrogen and phosphorus fertilization on the growth and nutrition of hybrid poplars on Vancouver Island. *New Forests*, 29: 89-104.
- Buscaroli, A. (2017). An overview of indexes to evaluate terrestrial plants for phytoremediation purposes (Review). *Ecological Indicators*, 82: 367-380.
- Burzynski, M. (1987). The Influence of Lead and Cadmium on the Absorption and Distribution of Potassium, Calcium, Magnesium and Iron in Cucumber Seedlings. *Acta Physiologiae Plantarum*, 9: 229-238.
- Cecchi, M. (2008). Devenir du plomb dans le système Sol-Plante: Cas d'un sol contaminé par une usine de recyclage du plomb et de deux plantes potagères (Fève et Tomate). Thèse de doctorat. Institut National Polytechnique, Toulouse.
- Cenic-Milosevic D., Mileusnic I., Kolak V., Pejanovic D., Ristic T., Jakovljevic A., Popovic M., Pesic D., & Melih, I. (2013). Environmental lead pollution and its

- possible influence on tooth loss and hard dental tissue lesions. *Vojnosanitetski pregled*, 70: 751-756.
- Cenkci S., Yildiz M., Cigerci I.H., Konuk M., & Bozdag A. (2009). Toxic chemicals-induced genotoxicity detected by random amplified polymorphic DNA (RAPD) in bean (*Phaseolus vulgaris L.*) seedlings. *Chemosphere*, 76: 900-906.
- Chandra, R., & Kang, H. (2016). Mixed heavy metal stress on photosynthesis, transpiration rate, and chlorophyll content in poplar hybrids. *Forest Science and Technology*, 12: 55-61.
- Chatterjee, J., & Chatterjee. C. (2003). Management of phytotoxicity of cobalt in tomato by chemical measures. *Plant Science*, 64: 793-801.
- Christine, J.W., & Joseph, J.C. (2009). Measurement of superoxide dismutase, catalase, and glutathione peroxidase in cultured cells and tissue. *Natural Protocols*, 5: 51-66.
- Chaturvedi, R., Favas, P.J.C., Pratas, J., Varun, M., & Paul, M.S. (2018). Effect of *Glomus mosseae* on accumulation efficiency, hazard index and antioxidant defense mechanisms in tomato under metal (loid) stress. *International Journal of Phytoremediation*, 20: 885-894.
- Chakraborty, L., Pandit, P., & Roy Maulik, S. (2019). *Acacia auriculiformis* - A natural dye used for simultaneous coloration and functional finishing on textiles. *Journal of Cleaner Production*, 245: 118921.
- Chenon, P. (2001). Evaluation du pouvoir toxique, génotoxique et tératogène de boues de stations d'épuration valorisées en agriculture. Thèse de doctorat , Université Paul Sabatier, Toulouse.
- Choudhary, M., Jetley, U. K., Khan, M. A., Zutshi, S., & Fatma, T. (2007). Effect of heavy metal stress on proline, malondialdehyde, and superoxide dismutase activity in the cyanobacterium *Spirulina platensis-S5*. *Ecotoxicology and Environmental Safety*, 66: 204-209.
- Choudhury, S., Panda, P., Sahoo, L., & Panda, S. K. (2013). Reactive oxygen species signaling in plants under abiotic stress. *Plant Signaling & Behavior*, 8: 23681.
- Cid, C.V., Rodriguez, J.H., Salazar, M.J., Blanco, A., & Pignata, M.L. (2016) Effects of co-cropping *Bidens pilosa* (L.) and *Tagetes minuta* (L.) on bioaccumulation of Pb in *Lactuca sativa* (L.) growing in polluted agricultural soils. *International Journal of Phytoremediation*, 18: 908-917.
- Clemens, S. (2006). Evolution and function of phytochelatin synthases. *Journal of Plant Physiology*, 163: 319-332.
- Clemens, S. (2001). Molecular mechanisms of plant metal tolerance and homeostasis. *Planta*, 212: 475-786.
- Costa, M.V.J. DA., & Sharma, P.K. (2016). Effect of copper oxide nanoparticles on

- growth, morphology, photosynthesis, and antioxidant response in *Oryza sativa*. *Photosynthetica*, 54: 110-119.
- Conesa, H.M., María-Cervantes, A., Alvarez-Rogel, J., González-Alcaraz, M.N. (2011) Influence of soil properties on trace element availability and plant accumulation in a Mediterranean salt marsh polluted by mining wastes: implications for phytomanagement. *Science of Total Environment*, 409: 4470-4479.
- Couselo, J.L., Navarro-Avino, J., & Ballester, A. (2010). Expression of the phytochelatin synthase TaPCS1 in transgenic aspen, insight into the problems and qualities in phytoremediation of Pb. *International Journal of Phytoremediation*, 12: 358-370.
- Collin, V.C., Eymery, F., Genty, B., Rey, P., & Havaux, M. (2008). Vitamin E is essential for the tolerance of *Arabidopsis thaliana* to metal-induced oxidative stress. *Plant, Cell and Environment*, 31: 244-257.
- Cobbett, C., & Goldsbrough, P. (2002). Phytochelatin and metallothioneins: roles in heavy metal detoxification and homeostasis. *Annual Review of Plant Biology*, 53: 159-182.
- Cuypers, A., Vangronsveld, J., & Clijsters, H. (2002). Peroxidases in roots and primary leaves of *Phaseolus vulgaris* copper and zinc phytotoxicity: a comparison. *Journal of Plant Physiology*, 159: 869-876.
- Dane, T., Lamb, H., Mallavarapu, M., & Ravi, N. (2010). Relative Tolerance of a Range of Australian Native Plant Species and Lettuce to Copper, Zinc, Cadmium, and Lead. *Archives of Environmental Contamination and Toxicology*, 59: 424-432.
- Damm, S., Hofmann, B., Gransee, A. & Christen, O. (2012). The influence of long-standing differentiated potassium fertilization on parameters of the soil water balance. *Archives of Agronomy and Soil Science*. 59: 21-40.
- Department of Environment. (2011). *Malaysia Environmental Quality Report 2011* (MEQR) (Publication Section ISSN 0127-6433). Malaysia: DOE, Malaysia.
- Diáz, J., Bernal, A., pomar, F., & Merino, F. (2001). Induction of shikimate dehydrogenase and peroxidase in pepper (*Capsicum annuum*.) seedlings in response to copper stress and its relation to lignification. *Plant Science*. 161: 179-188.
- Dorogházi, O., Kastori, R., & Maksimović, I. (2010). Nickel translocation from seed during germination and growth of young maize plants. *Proceedings for Natural Sciences*, 119: 17-25.
- Do Huang, C.W.A., & Xing, B. (2006). Phytoextraction: a review on enhanced metal availability and plant accumulation. *Scientia Agricola*, 63: 299-311.
- Dong, J., Wu, F., & Zhang, G. (2006). Influence of cadmium on antioxidant capacity and four microelement concentrations in tomato seedlings (*Lycopersicon esculentum*). *Chemosphere*, 64: 1659-1666.

- Dos Santos, W.D., Ferrarese, M.D.L., Finger, A., Teixeira, A.C.N., & Ferrarese, O. (2004). Lignification and related enzymes in *Glycine max* root growth-inhibition by ferulic acid. *Journal of Chemical Ecology*, 30: 1203-1212.
- Di Salvatore, M., Carafa, A.M., & Carratu, G. (2008). Assessment of heavy metals phytotoxicity using seed germination and root elongation tests: A comparison of two growth substrates. *Chemosphere*, 73: 1461-1464.
- Dugé de Bernonville, T. (2009). Caractérisations histologique, moléculaire et biochimique des interactions compatible et incompatible entre *Erwinia amylovora*, agent du feu bactérien, et le pommier (*Malus x domestica*). Thèse de Doctorat, Université d'Angers, Angers.
- Ducruix, C., Junot, C., Fiévet, J.B., Villiers, F., Ezan, E., & Bourguignon, J. (2006). New insights into the regulation of phytochelatin biosynthesis in *A. thaliana* cells from metabolite profiling analyses. *Biochimie*, 88: 1733-1742.
- Dushenkov, V., Kumar, P.B.A.N., Motto, H. & Raskin, I. (1995). Rhizofiltration: The use of plants to remove heavy metals from aqueous streams. *Environmental Science & Technology*, 29: 1239-45.
- Egilla, J.N., Davies, F.T. & Boutton, T.W. (2005). Drought stress influences leaf water content, photosynthesis, and water-use efficiency of *Hibiscus rosa-sinensis* at three potassium concentrations. *Photosynthetica*, 43: 135-140.
- Erakhrumen, A., & Agbontalor, A. (2007). Review Phytoremediation: an environmentally sound technology for pollution prevention, control and remediation in developing countries. *Educational Research and Review*, 2: 151-156.
- Eranen, JK., & Kozlov, MV. (2007). Competition and facilitation in industrial barrens: Variation in performance of mountain birch seedlings with distance from nurse plants. *Chemosphere*, 67: 1088-1095.
- Farooqi, Z.R., Zafar, Iqbal., M, Kabir, M AND., & Shafiq, M. (2009). Toxic effects of lead and cadmium on germination and seedling growth of *Albizia lebbeck* (L.) benth. *Pakistan Journal of Botany*, 41: 27-33.
- Favier, A. (1997). Le stress oxydant: intérêt de sa mise en évidence en biologie médicale et problèmes posés par le choix d'un marqueur. *Annales de Biologie Clinique*, 55: 9-16.
- Fernando, A., Manuel N. C., Vicente, M., & Francisco, R. (2011). Root K⁺ Acquisition in Plants: The *Arabidopsis Thaliana* Model. *Plant and Cell Physiology*, 52: 1603-12.
- Ferreyra, H.R., Romano, M., & Uhart, M. (2009). Recent and chronic exposure of wild ducks to lead in human-modified wetlands in Santa Fe Province, Argentina. *Journal of Wildlife Diseases*, 45: 823-827.
- Fischer, R.A. (1968). Stomatal opening: role of potassium uptake by guard cells. *Science*,

160: 784-785.

- Foyer, CH., & Noctor, N. (2005). Oxidant and antioxidant signalling in plants: a re-evaluation of the concept of oxidative stress in a physiological context. *Plant, Cell and Environment*, 28: 1056-1071.
- Gangaiah, A., Chandrasekhar, T., Varaprasad, D., Hima Bindu, Y., Keerthi Kumari, M., Chakradhar, T., & Madhava Reddy, C. (2013). Effects of heavy metals on in vitro seed germination and early seedling growth of *Pennisetum glaucum* (L.) R.Br. *International Journal of Food Agriculture and Veterinary Sciences*, 3: 87-93.
- Garg, N., & Manchanda, G. (2009). ROS generation in plants: boon or bane. *Plant Biosystems- An International Journal Dealing with All Aspects of Plant Biology*, 143: 8-96.
- Gilman, E.F., & Dennis, G.W. (2013). *Acacia Auriculiformis*: Earleaf Acacia 1. 1-3.
- Gill, SS., & Tuteja, N. (2010). Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. *Plant Physiology and Biochemistry*, 48: 909-930.
- Girijashankar, V. (2011). Micropropagation of multipurpose medicinal tree *Acacia auriculiformis*. *Journal of Medicinal Plants Research*, 5: 462-6.
- Gomes, M. P., Soaresb, A. M., & Garcia, QS. (2014). Phosphorous and sulfur nutrition modulate antioxidant defenses in *Myracrodroon urundeuva* plants exposed to arsenic. *Journal of Hazardous Materials*, 276: 97-104.
- Gopal, R., & Rizvi, A. H. (2008). Excess lead alters growth, metabolism and translocation of certain nutrients in radish. *Chemosphere*, 70: 1539-1544.
- Grilli, G., Urcelay, C., Galetto, L., Davison, J., Vasar, M., Saks, Ü., Jairus, T., & Öpik, M. (2015). The composition of arbuscular mycorrhizal fungal communities in the roots of a ruderal forb is not related to the forest fragmentation process. *Environmental Microbiology*, 17: 2709-2720.
- Grill, E., Mishra, S., Srivastava, S., & Tripathi, R.D. (2006). Role of phytochelatins in phytoremediation of heavy metals. *Environmental bioremediation technologies*, 101-146.
- Graziani, SN., Salazar, MJ., Pignata, ML., & Rodriguez, JH. (2016). Assessment of the root system of *Brassica juncea* (L.) czern. and *Bidens Pilosa* L. exposed to lead polluted soils using rhizobox systems. *International Journal of Phytoremediation*, 18: 235-244.
- Gupta, D. K., Palma, J. M., & Corpas, F. J. (2015). *Reactive Oxygen Species and Oxidative Damage in Plants Under Stress*. Springer International Publishing. Switzerland.
- Habiba, U., Ali, S., Farid, M., Shakoor, M.B., Rizwan, M., Ibrahim, M., Abbasi, G.H., Hayat, T., & Ali, B. (2015). EDTA enhanced plant growth, antioxidant defense

system and phytoextraction of copper by *Brassica napus* L. *Environmental Science and Pollution Research*, 22: 1534-1544.

Hadi, F., Ali, N., & Fuller, M. P. (2016). Molybdenum (Mo) increases endogenous phenolics, proline and photosynthetic pigments and the phytoremediation potential of the industrially important plant *Ricinus communis* L. for removal of cadmium from contaminated soil. *Environmental Science and Pollution Research*, 23: 20408-20430.

Hall, J.L. (2002). Cellular mechanisms for heavy metal detoxification and tolerance. *Journal of Experimental Botany*, 53: 1-11.

Halliwell, B. (2006). Reactive Species and Antioxidants. Redox Biology is a Fundamental Theme of Aerobic Life. *Plant Physiology*, 141: 312-322.

Hossain, M.A., Piyatida, P., da Silva, J.A.T., & Masayuki, F. (2012). Molecular mechanism of heavy metal toxicity and tolerance in plants: central role of glutathione in detoxification of reactive oxygen species and methylglyoxal and in heavy metal chelation. *Journal of Botany*, 2012: 1-37.

Hu, R., Sunc, K., Suc, X., Pana, Y., & Zhang, Y. (2012). Physiological responses and tolerance mechanisms to Pb in two xerophils: *Salsola passerine* Bunge and *Chenopodium album*. L. *Journal of Hazardous Materials*, 205:131-138.

Hinsinger, P., Schneider, A., & Dufey, J.E. (2005). Le sol: ressource en nutriments et biodisponibilité. In *Sols et Environnement*. Dunod (ed), Paris.

Hovmand, M.F., Nielsen, S.P., & Johansen, I. (2009). Root uptake of lead by norway spruce grown on Pb spiked soils. *Environmental Pollution*, 157: 404-409.

Huang, S., Zhang, W.J., Yu, X.C., & Huang, Q.R. (2010). Effects of long-term fertilization on corn productivity and its sustainability in an ultisol of southern China. *Agriculture, Ecosystems & Environment*. 138: 44-50.

Huang, J., Qin, F., Zang, G., Kang, Z., Zou, H., Hu, F., Yue, C., Li, X., & Wang, G. (2013). Mutation of OsDET1 increases chlorophyll content in rice. *Plant Science*, 210: 241-249.

Huda, S., Sujauddin, M., Shafinat, S., & Uddin, M. (2007). Effects of phosphorus and potassium addition on growth and nodulation of *Dalbergia sissoo* in the nursery. *Journal of Forestry Research*, 18: 279-282.

Ibrahim, M.H., & Jaafar, H.Z.E. (2011). Enhancement of leaf gas exchange and primary metabolites up-regulate the production of secondary metabolites of *Labisia Pumila* Blume seedlings under carbon dioxide enrichment. *Molecules*, 16: 3761-3777.

Ibrahim, M.H., Jaafar, H.Z.E., Asmah, R., & Zaharah, A.R. (2012). Involvement of Nitrogen on Flavonoids, Glutathione, Anthocyanin, Ascorbic Acid and Antioxidant Activities of Malaysian Medicinal Plant *Labisia pumila* Blume. *International Journal of Molecular Sciences*, 13: 393-408.

- Illié Zoran, S., Natasa M., Radmila T., Nikolaos K., Lidija M., & Ljubomir S. (2015). Effect of Pb on germination of different seed and his translocation in bean seed tissues during sprouting. *Fresenius Environmental Bulletin*, 24: 670-75.
- Ishiguri, F., Yokota, S., Yoshizawa, N., & Ona, T. (2004). Radial variation of cell morphology in three *Acacia* species. In *Improvement of Forest Resources For Recyclable Forest Products*. Springer, Kyushu University.
- Islam, E., Yang, X., Li, T., Liu, D., Jin, X., & Meng, F. (2007). Effect of Pb toxicity on root morphology, physiology and ultrastructure in the two ecotypes of *Elsholtzia argyi*. *Journal of Hazardous Materials*, 147: 806-816.
- Islam, E., Liu, D., Li, T., Yang, X., Jin, X., Mahmood, Q., Tian, S., & Li, J. (2008). Effect of Pb toxicity on leaf growth, physiology and ultrastructure in the two ecotypes of *Elsholtzia argyi*. *Journal of Hazardous Material*. 154: 914-926.
- Ismail, A., & Rosniza, R. (1997). Trace metals in sediments and molluscs from an estuary receiving pig farms effluent. *Environmental Technology*. 18: 509-15.
- Ivanov, B.N. (2014). Role of ascorbic acid in photosynthesis. *Biochemistry*, 79: 282-289.
- Jákli, B., Tavakol, E., Tränkner, M., Senbayram, M., & Dittert, K. (2017). Quantitative limitations to photosynthesis in K deficient sunflower and their implications on water-use efficiency. *Journal of Plant Physiology*, 209, 20-30.
- Jayasri, M.A., & Suthindhiran, K. (2017). Effect of zinc and lead on the physiological and biochemical properties of aquatic plant *Lemna minor*: its potential role in phytoremediation. *Applied Water Science*, 7: 1247-1253.
- Jarvis, M.D., & Leung, D.W.M. (2001). Chelated lead transport in *Chamaecytisus proliferus* (L.f.) link ssp. *proliferus* var. *palmensis* (H. Christ): an ultrastructural study. *Plant Science*, 161: 433-441.
- Jaliya, M. M., Falaki, A. M., Mahmud, M., & Sani, Y. A. (2008). Effects of sowing date and NPK fertilizer rate on yield and yield components of quality protein maize (*Zea mays L.*). *ARPN Journal of Agricultural and Biological Science*, 3: 23-29.
- Jenson, W.A. (1962). *Botanical histochemistry-Principles and Practice*. Freeman, San Francisco.
- John, R., Ahmad, P., Gadgil, K., & Sharma, S. (2009). Heavy metal toxicity: Effect on plant growth, biochemical parameters, and metal accumulation by *Brassica juncea L.* *International Journal of Plant Production*, 3: 65-76.
- Kanwal, U., Ali, S., Shakoor, M.B., Farid, M., Hussain, S., Yasmeen, T., Adrees, M., Bharwana, S.A., & Abbas, F. (2014). EDTA ameliorates phytoextraction of lead and plant growth by reducing morphological and biochemical injuries in *Brassica napus L.* under lead stress. *Environmental Science and Pollution Research*, 21: 9899-9910.
- Karimi, L. N., Ahmadi, M. K., & Moradi, B. (2012). Accumulation and phytotoxicity of

lead in *Cynara scolymus*. *Indian Journal of Technology*, 5: 3634-3641.

Kadiriye, U.R.U.Ç., & Dilek, D.Y. (2008). Effect of cadmium lead and nickel on imbibition, water uptake and germination for the seeds of different plants. *Dumlupınar Üniversitesi Fen Bilimleri Enstitüsü*, 17:1-10.

Kaviani, B. (2014). Effect of ascorbic acid concentration on structural characteristics of apical meristems on *in vitro aloe barbadensis mill*. *Acta Scientiarum Polonorum Hortorum Cultus*, 13:49-56.

Kaushik, D., & Aryadeep, R. (2014). Reactive oxygen species (ROS) and response of antioxidants as ROS-scavengers during environmental stress in plants. *Frontiers in Environmental Science*, 2: 53.

Keller, C., Rizwan, M., Davidian, J.C., Pokrovsky, O.S., Bovet, N., Chaurand, P., & Meunier, J.D. (2015). Effect of Silicon on wheat seedlings (*Triticum turgidum L.*) grown in hydroponics under Cu stress. *Planta*, 241: 847-860.

Khalid, A. (2014). Influences of silicate dissolving bacteria and natural potassium on growth and essential oil of rue plant. *Thai Journal of Agricultural Science*, 47: 31-36.

Khan, A., Iqbal, I., Shah, A., Ahmad, A., & Ibrahim, M. (2010). Alleviation of adverse effects of salt stress in brassica (*brassica campestris*) by pre-sowing seed treatment with ascorbic acid. *American-Eurasian Journal of Agricultural and Environmental Science* 2010, 7: 557-560.

Khalid, K.A, & Shedeed, M.R. (2015). Effect of NPK and foliar nutrition on growth, yield, and chemical constituents in *Nigella sativa L.* *Journal of Materials and Environmental Science*, 6: 1709-1714.

Kirpichtchikova, T. (2009). Phytoremédiation par Jardins Filtreurs d'un sol pollué par des métaux lourds: Approche de la phytoremédiation dans des casiers végétalisés par des plantes de milieux humides et étude des mécanismes de remobilisation/immobilisation du zinc et du cuivre. Thèse de doctorat. Université de Joseph Fourier, Grenoble.

Kim, Y.Y., Yang, Y.Y., & Lee, Y. (2003). Pb and Cd uptake in rice roots. *Physiologia Plantarum*, 116: 368-372.

Kováčik, J., Klejdus, B., Hedbavny, J., Štork, F., & Baćkor, M. (2009). Comparison of cadmium and copper effect on phenolic metabolism, mineral nutrients and stress-related parameters in *Matricaria chamomilla* plants. *Plant Soil*. 320: 231-242.

Krupa, Z., Siedlecka, A., Skórzynska-Polit, E., & Maksymiec, W. (2002). Heavy Metal Interactions with Plant Nutrients. *Physiology and Biochemistry of Metal Toxicity and Tolerance in Plants*, 287-301.

Krieger-Liszakay, A., & Trebst, A. (2006). Tocopherol is the scavenger of singlet oxygen produced by the triplet states of chlorophyll in the PSII reaction center. *Journal of Experimental Botany*, 57: 1677-1684.

- Król, A., Amarowicz, R., & Weidner, S. (2014). Changes in the composition of phenolic compounds and antioxidant properties of grapevine roots and leaves (*Vitis vinifera L.*) under continuous of long-term drought stress. *Acta Physiologiae Plantarum*, 36: 1491-1499.
- Küpper, H., & Kochian, L.V. (2010). Transcriptional regulation of metal transport genes and mineral nutrition during acclimatization to cadmium and zinc in the Cd/Zn hyperaccumulator, *Thlaspi caerulescens* (Ganges population). *New Phytologist*, 185: 114-129.
- Kranner, I., & Colville, L. (2011). Metals and seeds: Biochemical and molecular implications and their significance for seed germination. *Environmental and Experimental Botany*. 72: 93-105.
- Lamand, M. (1991). Les oligo-éléments dans la biosphère, dans les oligo-éléments en médecine (Ed), et en biologie. *Médicales Internationales*, Paris.
- Lamhamdi, M., Bakrim, A., Aarab, A., Lafont, R., & Sayah, F. (2011). Lead phytotoxicity on Wheat (*Triticum aestivum L.*) seed germination and seedlings growth. *Comptes Rendus Biologies*, 334: 118-126.
- Laghlimi, M., Bouamar, B., Hassan, E., & Abdelhak, B. (2015). Phytoremediation Mechanisms of Heavy Metal Contaminated Soils: A Review. *Open Journal of Ecology*, 5: 375-388.
- Lakshmi, T., Rajendran, R., & Ezhilarasan, D. (2015). Atomic Absorption Spectrophotometric Analysis of Heavy Metals in *Acacia catechu willd*. *International Journal of Pharmacognosy and Phytochemical Research*, 7: 777-781.
- Lei, K., Giubilato, E., Critto, A., Pan, H., & Lin, C. (2016). Contamination and human health risk of lead in soils around lead/zinc smelting areas in China. *Environmental Science and Pollution Research*, 23: 13128-13136.
- Lee, K.C., Cunningham, B. A., Chung, K.H., Saul Sen, G.M., & Liang, G.H. (1976). Lead effects on several enzymes and nitrogenous compounds in soybean leaf. *Journal of Environment Quality*, 5: 357-359.
- Lemoine, R., La Camera, S., Atanassova, R., Dedald echamp, F., Allario, T., Pourtau, N., Bonnemain, J.L., Laloi, M., Coutos Thevenot, P., Maurousset, L., Faucher, M., Girousse, C., Lemonnier, P., Parrilla, J., & Durand, M. (2013). Source to sink transport and regulation by environmental factors. *Frontiers in Plant Science*. 4: 272.
- Liu, X., Mak, M., Babla, M., Wang, F., Chen, G., Veljanoski, F., Wang, G., Shabala, S., & Zhou, M., & Chen, Z. (2014). Linking stomatal traits and expression of slow anion channel genes HvSLAH1 and HvSLAC- 1with grain yield for increasing salinity tolerance in barley. *Frontiers in Plant Science*, 5: 1-12.
- Liu, HC., Tian, DQ., Liu, JX., Ma, GY., Zou, QC., & Zhu, ZJ. (2013). Cloning and

- functional analysis of a novel ascorbate peroxidase (APX) gene from *Anthurium andraeanum*. *Journal of Zhejiang University Science*, 14: 1110-1120.
- Ling, T., & Jun, R. (2010). Effect of Hg on seed germination, coleoptile growth and root elongation in seven pulses. *Fresenius Environmental Bulletin*, 19: 1144-1150.
- Lin, CC., & Kao., CH. (2000). Effect of NaCl stress on H₂O₂ metabolism in rice leaves. *Plant Growth Regulations*. 30:151-155.
- Lin S., Wang X., Yu I.T., Tang W., Miao J., Li J., Wu S., & Lin X. (2011). Environmental lead pollution and elevated blood lead levels among children in a rural area of China. *American Journal of Public Health*, 101: 834-841.
- Lim J. H., Park K. J., Kim B. K., Jeong J. W. & Kim, H. J. (2012). Effect of salinity stress on phenolic compounds and carotenoids in buckwheat (*Fagopyrum esculentum M.*) sprout. *Food Chemistry*, 135: 1065-1070.
- Li, Y. (2008). Effect of salt stress on seed germination and seedling growth of three salinity plants. *Pakistan Journal of Biological Science*, 11: 1268-1272.
- Liu, J.N., Zhou, Q.X., Sun, T., Ma, L.Q., & Wang, S. (2008). Identification and chemical enhancement of two ornamental plants for phytoremediation. *Bulletin of Environmental Contamination and Toxicology*, 80: 260-265.
- Louise, J.K., Mark, P.T., & Arthur, R.F. (2017). An odyssey of environmental pollution: The rise, fall and remobilization of industrial lead in Australia. *Applied Geochemistry*, 83: 3-13.
- Lo, I.M.C., Tsang, D.C.W., Yip, T.C.M., Wang, F., & Zhang, W. (2011). Influence of injection conditions on EDDS-flushing of metal-contaminated soil. *Journal of Hazardous Materials*, 192 : 667-675.
- Lokman Hossain, Md., Huda, S. M. S., & Hossain, M. K. (2009). Effects of industrial and residential sludge on seed germination and growth parameters of *Acacia auriculiformis* seedlings. *Journal of Forestry Research*, 20: 331-336.
- Lombardi, L., & Sebastiani, L. (2005). Copper toxicity in *Prunus Cerasifera* : growth and antioxidant enzymes responses of in vitro grown plants, *Plant Science*, 3: 797-802.
- Thayumanavan, B., & Sadasivam, S. (1984). Physicochemical basis for the preferential uses of certain rice varieties. *Plant Foods for Human Nutrition*, 34 : 253-259.
- Lutts, S., & Lefèvre, I. (2015). How can we take advantage of halophyte properties to cope with heavy metal toxicity in salt-affected areas? *Annals of Botany*, 115: 509-528.
- Maryam, M.S., Bilal, H.A., Nisar, A., Mohammad, A., & Tariq, M. (2014). Toxic Effects of Heavy Metals (Cd, Cr and Pb) on Seed Germination and Growth and DPPH-Scavenging Activity in Brassica Rapa Var. Turnip. *Toxicology and Industrial Health*, 30: 238-49.

- Mahmood Q., Ahmad R., Kwak SS., Rashid A., & Anjum N.A. (2010). Ascorbate and Glutathione: Protectors of Plants in Oxidative Stress. In: Anjum N., Chan M.T., & Umar, S. (eds) *Ascorbate-Glutathione Pathway and Stress Tolerance in Plants*. Springer, Dordrecht.
- Masarirambi, M.T, Hlawe, M.M., Oseni, O.T. & Sibiya, T.E. (2010). Effects of organic fertilizers on growth, yield, quality and sensory evaluation of red lettuce (*Lactuca sativa L.*) Veneza Roxa. *Agriculture and Biology Journal of North America*, 1: 1319-1324.
- Magwedere, K., Shimwino, J., Hemberger, Y., Hoffman, L.C., Midzi, E.M., & Dziva, F. (2013). Lead and Cadmium Levels in Liver, Kidney and Muscle of Harvested Wild Springbok (*Antidorcus marsupialis*) Under Extensive Management in Southern and Southeastern Namibia. *South African Journal of Wildlife Research*, 43: 52-60.
- Malenčić, D.J., Vasić, D., Popović, M., & Dević, D. (2004). Antioxidant systems in sunflower as affected by oxalic acid. *Biologia Plantarum*, 48: 243-247.
- Magné, C., & Larher, F. (1992). High sugar content of extracts interferes with colorimetric determination of amino acids and free proline. *Analytical Biochemistry*, 200: 115-118.
- Masayuki, S., Aya, W., Sakae, S., Masahiro, I., & Toshikazu, K. (2007). Phytoextraction and phytovolatilization of Arsenic from As-contaminated soils by *Pteris vittata*. *Proceedings of the Annual International Conference on Soils, Sediments, Water and Energy*, 12: 26.
- Mazhoudi, S., Chaoui, A., Ghorbal, M.H., & Ferjani, E.E. (1997). Response of antioxidant enzymes to excess copper in tomato (*Lycopersicon esculentum*). *Plant Science Journal*, 127: 129-137.
- Marschner, P. (2012). *Marschner's Mineral Nutrition of Higher Plants (Third Edition)*. Elsevier Ltd. Adelaide.
- McGrath, S.P., & Cunliffe, C.H. (1995). A simplified method for the extraction of the metals in sewage sludge on soils, microorganisms and plants. *Journal of Industrial Microbiology & Biotechnology*, 14: 94-104.
- Mohsenzadeh, F., Abdolkarim, C., & Nafiseh, Y. (2011). Effect of the Heavy Metals on Developmental Stages of Ovule, Pollen, and Root Proteins in *Reseda Lutea L.* (Resedaceae). *Biological Trace Element Research*, 143: 1777-88.
- Moran, J.F., James, E.K., Rubio, M.C., Sarath, G., Klucas, R.V., & Becana, M. (2003). Functional characterization and expression of a cytosolic iron-superoxide dismutase from cowpea root nodules. *Plant Physiology*, 133: 773-782.
- Mhamdi, A., Queval, G., Chaouch, S., Vanderauwera, S., Van Breusegem, F., & Noctor, G. (2010) Catalase function in plants: a focus on *Arabidopsis* mutants as stress-mimic models. *Journal of Experimental Botany*, 61: 4197-4220.

- Mayank, V., Rohan, D., Devendra, K., & Manoj P.S. (2011). Bioassay as Monitoring System for Lead Phytoremediation through *Crinum Asiaticum L.* *Environmental Monitoring and Assessment*, 178: 373-81.
- Masvodza, D. R., Dzomba, P., Mhandu, F., & Masamha, B. (2013). Heavy Metal Content in *Acacia saligna* and *Acacia polyacantha* on Slime Dams: Implications for Phytoremediation. *American Journal of Experimental Agriculture*, 3: 871-883.
- Malkowski, E., Kita, A., Galas, W., & Michael, K. (2002). Lead distribution in corn seedlings (*Zea mays L.*) and its effect on growth and the concentration of potassium and calcium. *Plant Growth Regulation*, 37: 69-76.
- Mavi, K., Demir, I., & Matthews, S. (2010). Mean germination time estimates the relative emergence of seed lots of three cucurbit crops under stress conditions. *Seed Science. & Technology*, 38: 14-25.
- Małecka, A., Piechalak, A., Morkunas, I., & Tomaszewska, B. (2008). Accumulation of lead in root cells of *Pisum sativum*. *Acta Physiologiae Plantarum*, 30: 629-637.
- Macek, T., Macková, M., & Kás, J., (2000). Exploitation of plants for the removal of organics in environmental remediation. *Biotechnology Advances*, 18: 23-34.
- Majid, N., Islam, M., & Lydia, M. (2012). Heavy metal uptake and translocation by mangium (*Acacia mangium*) from sewage sludge contaminated soil. *Australian Journal of Crop Science*. 6: 1228-1235.
- Manquian-Cerda K., Escudey M., Zuniga G., Arancibia-Miranda, N., Molina, M., & Cruces E. (2016). Effect of cadmium on phenolic compounds, antioxidant enzyme activity and oxidative stress in blueberry (*Vaccinium corymbosum L.*) plantlets grown in vitro. *Ecotoxicology and Environmental Safety*, 133: 316-326.
- Meyers, D.E.R., Auchterlonie, G.J., Webb, R.I., & Wood, B. (2008). Uptake and localization of lead in the root system of *Brassica juncea*. *Environmental Pollution*, 153: 323-332.
- Miguel, P., Mourato, I.N., Moreira, I.L., Filipa, R.P., Joana, R.S., & Luisa, L.M. (2015). Effect of Heavy Metals in Plants of the Genus *Brassica*. *International Journal of Molecular Sciences*, 16: 17975-17998.
- Mihiri, S., Nishanta, R., Muhammad, R., Madawala, Y. S. O., & Meththika, V. (2016). Heavy metal-induced oxidative stress on seed germination and seedling development: a critical review. *Environmental Geochemistry and Health*, 17: 5-8.
- Mihoub, A., Chaoui, A., & El Ferjan, E.I. (2005). Changements biochimiques induit par le cadmium et le cuivre au cours de la germination des grains de petit pois (*Pisum sativum L.*). *Comptes Rendus Biologies*, 328: 33-41.
- Miret, JA., & Munne-Bosch, S. (2015). Redox signaling and stress tolerance in plants: a focus on vitamin E. *Annals of the New York Academy of Sciences*, 1340: 29-38.
- Mohd, SN., Majid, N M., Shazili, N.A.M., & Abdu, A. (2013). Growth performance,

- biomass and phytoextraction efficiency of *Acacia mangium* and *Melaleuca cajuputi* in remediating heavy metal contaminated soil. *American Journal of Environmental Sciences*, 9: 310-316.
- Mombo, S., Foucault, Y., Deola, F., Gaillard, I., Goix, S., Shahid, M., Schreck, E., Pierart, A., & Dumat, C. (2015). Management of human health risk in the context of kitchen gardens polluted by lead and cadmium near a lead recycling company. *Journal of Soils and Sediments*, 16: 1214-1224.
- Moosavi, S.G., & Moham J.S. (2013). Phytoremediation: A Review. *Advance in Agriculture and Biology* 1: 5-11.
- Mostafa, F., El-Banna, MA., Gao, B., Yin, X., Wang, H., & Ahmad, Z. (2019). Scavenging effect of oxidized biochar against the phytotoxicity of lead ions on hydroponically grown chicory: an anatomical and ultrastructural investigation. *Ecotoxicology and Environmental Safety*, 170: 363-374.
- Moradi, D., Sharifzadeh, P. F., & Janmohammadi, M. (2008). Influence of priming techniques on seed germination behavior of maize inbred lines (*Zea mays L.*). *ARPN Journal of Agricultural and Biological Science*, 3: 22-25.
- Moussavou, M. C.F. (2010). Etude des mécanismes d'accumulation du cadmium chez *Arabidopsis thaliana* (écotype Wassilewskija) et chez un mélèzehybride (*Larixx eurolepis*) par des approches moléculaire et développementale. Thèse de doctorat. Université de Limoges.
- Mostafa, H., & Semin, S. (2011). Effects of lead and cadmium on seed germination, seedling growth and antioxidant enzymes activities of mustard (*Sinapis arvensis L.*). *ARPN Journal of Agricultural and Biological Science*, 6: 1990-6145.
- Moise, J.A., Han, S., Gudynaite Savitch, L., Johnson, D.A., & Miki, B.L.A. (2005). Seed coats: structure, development, composition, and biotechnology. *In Vitro Cellular & Developmental Biology. Plant*, 41: 620-644.
- Mozzo, M., Dall'Osto, L., Hienerwadel, R., Bassi, R., & Croce, R. (2008). Photoprotection in the Antenna Complexes of Photosystem II: Role of individual xanthophylls in chlorophyll triplet quenching. *Journal of Biological Chemistry*, 283: 6184-6192.
- Munzuroglu, O., & Geckil, S.W. (2002). Effects of metals on seeds germination, root elongation, and coleoptile and hypocotyl growth in *Triticum aestivum* and *Cucumis sativus*. *Archives of Environmental Contamination and Toxicology*, 43: 203-213.
- Mukherji, S., & Maitra, P. (1976). Toxic effects of lead on growth and metabolism of germination rice (*Oryza sativa L*) root tip cells. *Indian Journal of Experimental Biology*, 14: 519-521.
- Muhammad, S., Iqbal, M. Z., & Mohammad, A. (2008). Effect of lead and cadmium on germination and seedling growth of *Leucaena leucocephala*. *Journal of Applied Sciences and Environmental Management*, 12: 61-66.

- Nawab, J., Sardar K., Mohammad T. S., & Zahir, Q. (2015). Contamination of Soil, Medicinal, and Fodder Plants with Lead and Cadmium Present in Mine-Affected Areas, Northern Pakistan. *Environmental Monitoring and Assessment*, 187: 605.
- Nair, A., Juwarkar, A. A., & Devotta, S. (2008). Study of speciation of metals in an industrial sludge and evaluation of metals chelators for their removal. *Journal of Hazard Materiels*, 152: 545-553.
- Nahed, G., & Aziz, A.E. (2007). Stimulatory effect of NPK fertilizer and Benzyladenine on growth and chemical constituents of *Codiacum variegatum L* plant. *American-Eurasian Journal of Agricultural & Environmental Sciences*, 2: 711-719.
- Nagajyoti, P.C., Dinakar, N., Prasad, T.N.V.K.V., Suresh, C., & Damodharam, T. (2008). Heavy Metal Toxicity: Industrial Effluent Effect on Groundnut (*Arachis Hypogaea L.*) Seedlings. *Journal of Applied Science Research*, 4: 110-21.
- Naz, H., Akram, N.A., Ashraf, M. (2016). Impact of ascorbic acid on growth and some physiological attributes of cucumber (*cucumis sativus*) plants under water-deficit conditions. *Pakistan Journal of Botany*, 48:877-883.
- Needleman, H. (2004). Lead poisoning. *Annual Review of Medicine*, 55: 209-22.
- Nedelkoska, T.V., & Doran, P.M. (2000). Characteristics of heavy metal uptake by plant species with potential for phytoremediation and phytomining. *Minerals Engineering*, 13: 549-561.
- Ochonogor, R.O., & Atagana, H.I. (2014). Phytoremediation of Heavy Metal Contaminated Soil by *Psoralea pinnata*. *International Journal of Environmental Science and Development*, 5: 440-443.
- Oh, K., Cao, T., Li, T., & Cheng, H. (2014). Study on Application of Phytoremediation Technology in Management and Remediation of Contaminated Soils. *Journal of Clean Energy Technologies*, 2: 216-220.
- Olatunja, D, J., Maku, O., & Odumefun, O. P. (2012). Effect of Pre-Treatments on the Germination and Early Seedlings Growth of *Acacia auriculiformis Cunn. Ex Benth.* *African Journal of Plant Science*, 6: 364-69.
- Padmavathiamma, P.K., & Li, L.Y. (2007). Phytoremediation technology: Hyper-accumulation metals in plants. *Water, Air, and Soil Pollution*, 184: 105-126.
- Patrick, L. (2006). Lead toxicity, a review of the literature. *Alternative Medicine Review*, 11: 2-22.
- Pan, X., Zhang, D., Chen, X., Bao, A., & Li, L. (2011). Antimony accumulation, growth performance, antioxidant defense system and photosynthesis of *Zea mays* in response to antimony pollution in soil. *Water, Air, and Soil Pollution*, 21: 517-523.
- Paz-Alberto, A.M., Sigua, G.C., Baui, B.G., Prudente, J.A. (2007) Phytoextraction of lead-contaminated soil using vetivergrass (*Vetiveria zizanioides L.*), cogongrass (*Imperata cylindrica L.*) and carabaograss (*Paspalum conjugatum L.*).

- Pereira, L.B., Tabaldi, L.A., Goncalves, J.F., Jukoski, J.O., & Pauletto, M.M. (2006). Effect of aluminium on inolevulinic acid dehydratase (ALAD) and the development of cucumber (*Cucumis sativus*). *Environmental and Experimental Botany*, 57: 106-115.
- Pérez-de-Luque, A., Lozano-Baena, M.D., Prats, E., Moreno, M.T., & Rubiales, D. (2006). *Medicago truncatula* as a Model for Nonhost Resistance in Legume-Parasitic Plant Interactions. *Plant Physiology*, 145: 2437-449.
- Pilon-Smits, E. (2005). Phytoremediation. *Annual Review of Plant Biology*, 56: 15-39.
- Pourrut, B., Perchet, G., Silvestre, J., Cecchi, M., Guiresse, M., & Pinelli, E. (2008). Potential role of NADPH-oxidase in early steps of lead-induced oxidative burst in *Vicia faba* roots. *Journal of Plant Physiologies*, 165:571-579.
- Pourrut, B., Shahid, M., Dumat, C., Winterton, P., & Pinelli, Eb. (2011a). Lead uptake, toxicity, and detoxification in plants. *Reviews of Environmental Contamination and Toxicology*, 213:113-36.
- Pourrut B., Jean S., Silvestre J., & Pinelli E. (2011b): Lead-induced DNA damage in *Vicia faba* root cells: Potential involvement of oxidative stress. *Mutation Research/Genetic Toxicology and Environmental Mutagenesis*, 726: 123-128.
- Polle, A., & Schützendübel, A. (2003). Heavy metal signaling in plants: linking cellular and organismic responses. *Plant Responses to Abiotic Stress*, 4: 187-215.
- Preiss, J. (1982). Regulation of the biosynthesis and degradation of starch. *Annual Review of Plant Physiology*, 33: 431-454.
- Salazar, MJ., & Pignata, ML. (2014). Lead accumulation in plants grown in polluted soils. Screening of native species for phytoremediation. *Journal of Geochemical Exploration*, 137: 29-36.
- Sanchez, E., Muñoz, E., Anchondo, Á., Ruiz, J.M., & Romero, L. (2009). Nitrogen impact on nutritional status of Phosphorus and its main bioindicator: response in the roots and leaves of green bean plants. *Revista Chapingo. Serie horticultura*, 15: 177-182.
- Sahoo, B., Nedunchezhiyan, M., & Acharyya, P. (2015). Growth and yield of elephant foot yam under integrated nutrient management (INM) in alfisols. *Journal of Root Crops*, 41: 59-64.
- Saouli, N. (2016). Contribution à l'étude de l'effet de quelques engrains sur la disponibilité du phosphore dans les sols calcaires Touggourt. Mémoire de mastère en Agronomie, Université de Ouargla.
- Sardans, J., & Peñuelas, J. (2015). Potassium: a neglected nutrient in global change. *Global Ecology and Biogeography*, 24: 261-275.

- Seregin, IV., & Ivanov, VB. (2001). Physiological Aspects of Cadmium and Lead Toxic Effects on Higher Plants. *Russian Journal of Plant Physiology*, 48: 523-544.
- Sharma, P., & Rama S.D. (2005). Lead toxicity in plants. *Brazilian Journal of Plant Physiology*, 17: 35-52.
- Shoib, A.B., & Shahid, A.M. (2015). Determination of total phenolic and flavonoid content, antimicrobial and antioxidant activity of a root extract of *Arisaema jacquemontii* Blume. *Journal of Taibah University for Science*, 9: 449-454.
- Shaw, B.P., & Rout, N.P. (2002). Hg and Cd induced changes in the level of proline and the activity of proline bio synthesizing enzymes in *Phaseolus aureus* Roxb. and *Triticum aestivum* L. *Biologia. Plantarum*, 45: 267-271.
- Siti, N. M. R., Sharizal, H., Mohd L. K., NorShahrizan, M. H. (2014). Analysis and pollution assessment of heavy metal in soil, perlis. *The Malaysian Journal of Analytical Sciences*, 18: 155-161.
- Soobrattee, M.A., Neergheen, V.S., Luximon-Ramma, A., Aruoma, O.I., & Bahorun, O.T. (2005). Phenolics as potential antioxidant therapeutic agents: mechanism and actions. *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis*, 579: 200-213.
- Sukamto, L.A., Lestari, R., & Putri, W.U. (2014). The effect of biofertilizers on plant growth and growth rate of grafted avocado (*Persea americana* Mill.). *International Journal on Advanced Science Engineering Information Technology*, 4: 205-214.
- Susarla, S., Medina, V.F., & McCutcheon, S.C. (2002). Phytoremediation: An ecological solution to organic chemical contamination. *Ecological. Engineering*, 18: 647-658.
- Sytar, O., Borankulova, A., Hemmerich, I., Rauh, C. & Smetanska, I. (2014). Effect of chlorocholine chloride on phenolic acids accumulation and polyphenols formation of buckwheat plants. *Biological Research*, 47: 1-19.
- Patra, M., Bhowmik, N., Bandopadhyay, B., & Sharma, A. (2004). Comparison of mercury, lead and arsenic with respect to genotoxic effects on plant systems and the development of genetic tolerance. *Environmental and Experimental Botany*, 52: 199-223.
- Pajević, S., Borišev, M., Nikolić, N., Krstić, B., Pilipović, A., & Orlović, S. (2009). Phytoremediation capacity of poplar (*Populus spp.*) and willow (*Salix spp.*) clones in relation to photosynthesis. *Archives of Biological Science*, 61: 239-247.
- Potters, G., De Gara, L., Asard, H., & Horemans, N. (2002). Ascorbate and glutathione: guardians of the cell cycle, partners in crime? *Plant Physiology and Biochemistry*, 40: 537-548.
- Qian, T., Li, X. L., Zhu, X. Y., Huang, Y., & Yang, H. (2008). Effects of plumbum and chromium stress on the growth of tea plants. *Southwest China Journal of Agricultural Sciences*, 21: 156-162.

- Raymond, A.W., & Felix, E.O. (2011). Heavy Metals in Contaminated Soils: A Review of Sources, Chemistry, Risks and Best Available Strategies for Remediation. *International Scholarly Research Network*, 2011: 1-20.
- Remon, E. (2006). Tolérance et accumulation des métaux lourds par la végétation spontanée des friches métallurgiques: vers de nouvelles méthodes de bio-dépollution. Thèse de doctorat. Université Jean Monnet, Saint-Étienne.
- Rehman, A., Bhatti, H.N., & Athar, H.U. R. (2009). Textile effluents affected seed germination and early growth of some winter vegetable crops: a case study. *Water, Air, and Soil Pollution*, 198: 155-163.
- Rehman, K., Fatima, F., Waheed, I., Akash, MSH. (2018). Prevalence of exposure of heavy metals and their impact on health consequences. *Journal of Cellular Biochemistry*, 119: 157-184.
- Ricardo, A.B., Orivaldo, J.S.J., Rosa, M.P., Rosangela, S., Eliane, M.R.S., Silvio, R.L.T., Frederico H.T.G.L., Luiz, F.M., & Antonia, G.T.V. (2012). Selection of leguminous trees associated with symbiont microorganisms for phytoremediation of petroleum – contaminated soil. *Water, Air, and Soil Pollution*, 223: 5659-5671.
- Romualdas, J., Giedrè, V., Milda, R., & Jonė, V. (2012). The impacts of heavy metals on oxidative stress and growth of spring barley. *Central European Journal of Biology*, 7: 299-306.
- Roach, T., Beckett, R.P., Minibayeva, F.V., Colville, L., Whitaker, C., Chen, H., Bailly, C. & Kranner, I. (2010). Extracellular superoxide production, viability, and redox poise in response to desiccation in recalcitrant *Castanea sativa* seeds. *Plant, Cell & Environment*, 33: 59-75.
- Rui, Z., Vincent, L. W., Aixin, H., & Ge, M. (2015). Source of lead pollution, its influence on public health and the countermeasures. *International Journal of Health, Animal science and Food safety*, 2: 18-31.
- Salazar, MJ., Menoyo, E., Fagioli, V., Geml, J., Cabello, M., Rodriguez, JH., Marro, N., Pardo, A., Pignata, ML., & Becerra, AG. (2018). Pb accumulation in spores of arbuscular mycorrhizal fungi. *Science of The Total Environment*, 643: 238-246.
- Scippa, G., Di-Michele, M., Onelli, N.E., Patrignani, G., Chiatante, D., & Bray, E. (2004). The histone-like protein H1-S and the response of tomato leaves to water deficit. *Journal of Experimental Botany*, 55: 99-109.
- Seregin, I.V., Shpigun, L.K., & Ivanov, V.B. (2004). Distribution and Toxic Effects of Cadmium and Lead on Maize Roots. *Russian Journal of Plant Physiology*, 51: 525-533.
- Sethy, S.K., & Ghosh, S. (2013). Effect of heavy metals on germination of seeds. *Journal of Natural Science, Biology, and Medicine*, 4: 272-275.
- Seol, Y., & Javandel, I. (2008). Citric acid-modified Fenton's reaction for the oxidation of chlorinated ethylenes in soil solution systems. *Chemosphere*, 72: 537-542.

- Shenker, M. O., Plessner, E., & Tel-Or, E. (2004). Manganese nutrition effects on tomato growth, chlorophyll concentration, and superoxide dismutase activity. *Journal of Plant Physiology*, 161: 197-202.
- Sherene, T. (2010). Mobility and Transport of Heavy Metals in Polluted Soil Environment. *Biological Forum—An International Journal*, 2: 112-121.
- Shirbhate, N., & Malode, S. N. (2012). Phytoremediation Potential of *Cassia Tora* (L.), Roxb. to remove heavy metals from Waste soil, collect from sukali compost and landfill depot, amravati (M.S.). *Global Journal of Biotechnology & Biochemistry*, 1: 104-109.
- Shamsul, H., Qaiser, H., Mohammed, N.A., Arif, S.W., John, P., & Aqil, A. (2012). Role of proline under changing environments. *Plant Signaling & Behavior*, 7: 1456-1466.
- Shafiq, M., Zafar, I. M., & Athar, M. (2008). Effect of lead and cadmium on germination and seedling growth of *Leucaena leucocephala*. *Journal of Applied Sciences and Environmental Management*, 12: 61-66.
- Sinclair, T.R., Tanner, C.B., & Bennett, J.M. (1984). Water use efficiency in crop production. *BioScience*, 34: 36-40.
- Silvana, R. O., José, A.G.N., Joaquim, A. N., & Bradley, T. J. (2010). Determination of macro- and micronutrients in plant leaves by high-resolution continuum source flame atomic absorption spectrometry combining instrumental and sample preparation strategies. *Spectrochimica Acta Part B*, 65: 316-320.
- Sinha, S., Mishra, R.K., Sinam, G., Mallick, S. & Gupta, A.K. (2013). Comparative Evaluation of Metal Phytoremediation Potential of Trees, Grasses and Flowering Plants from Tannery Wastewater Contaminated Soil in Relation with, Physico-Chemical Properties. *Soil and Sediment Contamination: An International Journal*, 22: 958-983.
- Singh, H.P., Kaur, G., Batish, D.R., & Kohli, R.K. (2011). Lead (Pb)-inhibited radicle emergence in *Brassica campestris* involves alterations in starch-metabolizing enzymes. *Biological Trace Element Research*, 144: 1295-1301.
- Souza, L.A., Fernando, A.P., Roberta, C.N., & Ricardo, A.A. (2013). Use of Non-Hyperaccumulator Plant Species for the Phytoextraction of Heavy Metals Using Chelating Agents. *Scientia Agricola*, 4: 290-95.
- Sosa, M., Salazar, M.J., Zygaldo, J.A., & Wannaz, E.D. (2016). Effects of Pb in *Tagetes minuta* L. (Asteraceae) leaves and its relationship with volatile compounds. *Industrial Crops and Products*, 82: 37-43.
- Srinivas, T.N.R., Kailash, T.B., & Anil, K.P. (2013a). *Silanimonas mangrovi* sp. nov., a member of the family *Xanthomonadaceae* isolated from mangrove sediment, and emended description of the genus *Silanimonas*. *International Journal of Systematic and Evolutionary Microbiology*, 63: 274-279.

- Srinivasan, M., Rajendiran, M.A., Favas, J.C., Shivendra, V.S., & Perumal, V. (2014). Effect of Lead on Phytotoxicity, Growth, Biochemical Alterations and its role on genomic template stability in *Sesbania Grandiflora*: A Potential Plant for Phytoremediation. *Ecotoxicology and Environmental Safety*, 108: 249-57.
- Srinivas, J., Purushotham, A. V., & Murali Krishna, K. V. S. G. (2013b). The Effects of Heavy Metals on Seed Germination and Plant Growth on *Coccinia*, *Mentha* and *Trigonella* Plant Seeds in Timmapuram, E.G. District, Andhra Pradesh, India. *International Research Journal of Environment Sciences*, 2: 20-24.
- Sreekanth, T.V.M., Nagajyothi, P.C., Lee, K.D., & Prasad, T.N.V.K.V. (2013). Occurrence, physiological responses and toxicity of nickel in plants. *International Journal of Environmental Science and Technology*, 10: 1129-1140.
- Sterckeman, T., Douay, F., Proix, N., & Fourrier, H. (2000). Vertical distribution of Cd, Pb, and Zn in soils near smelters in the North of France. *Environmental Pollution*, 107: 377-389.
- Sunil, K.S., & Shyamasree, G. (2013). Effect of heavy metals on germination of seeds. *Biology and Medicine*, 4: 272-275.
- Sultana, T., Arooj, F., Nawaz, M., Alam, S., & Kashif, S.U.R. (2019). Removal of Heavy Metals from Contaminated Soil using Plants: A Mini-Review. *PSM Biological Research*, 4: 113-117.
- Swartjes, F.A. (2011). Dealing with contaminated sites: from theory towards practical application. National Institute of Public Health and the Environment (RIVM), Bilthoven.
- Tanhah, P., Kruatrachue, M., Pokethitiyook, P., & Chaiyarat, R. (2007). Uptake and accumulation of cadmium, lead and zinc by siam weed *Chromolaena odorata* (L.) King & Robinson. *Chemosphere*, 68: 323-329.
- Tong, YP., Kneer, R., & Zhu, YG. (2004). Vacuolar compartmentalization: a second-generation approach to engineering plants for phytoremediation. *Trends in Plant Science*, 9: 7-9.
- Troll, W., & Lindsey, J. (1955). A photometric method for the determination of proline. *Journal of Biological Chemistry*, 215: 655-660.
- Tripathi, D.K., Singh, V.P., Chauhan, D.K., Prasad, S.M., & Dubey, N.K. (2014). Role of macronutrients in plant growth and acclimation: recent advances and future prospective. In *Improvement of crops in the era of climatic changes*, Springer, New York.
- Turnbull, J.W., Midgley, S.J., & Cossalter, C. (1997). Tropical *Acacias* planted in Asia: an overview of recent developments in *Acacias* planting. In: Turnbull, J.W., et al. (Eds.) *Proceedings of Recent Developments in Acacia Planting*. proceedings of an international workshop, Hanoi.

- Valérie, P., & Urs, F. (2015). Heavy Metals in Crop Plants: Transport and Redistribution Processes on the Whole Plant Level. *Agronomy*, 5: 447-463.
- Veeresh, H., Tripathy, S., Chaudhuri, D., & Hart, B.R. (2003). Sorption and distribution of adsorbed metals in three soils of India. *Applied Geochemistry*, 18: 1723-1731.
- Wang, C., Wang, X., Tian, Y., Yu, H., Gu, X., Du, W., & Zhou, H. (2007). Oxidative stress, defence response and early biomarkers for lead-contaminated soil in *Vicia faba* seedlings. *Environmental Toxicology and Chemistry*, 27: 970-977.
- Wang, G., SU, M.Y., Chen, Y.H., Lin, F.F., Luo, D., & Gao, S.F. (2006). Transfer characteristics of cadmium and lead from soil to the edible parts of six vegetable species in southeastern China. *Environmental Pollution*, 144: 127-135.
- Wang, C., Tian, Y., Wang, X., Geng, J., Jiang, J., & Yu, H. (2010) Lead contaminated soil induced oxidative stress, defense response and its indicative biomarkers in roots of *Vicia faba* seedlings. *Ecotoxicology*, 19: 1130-1139.
- Wong, M.H. (2003). Ecological restoration of mine degraded soils, with emphasis on metal contaminated soils. *Chemosphere*, 50: 775-780.
- Weiqiang, Li., Mohammad, A., Khan, S.Y., & Yuji, K. (2005). Effects of Heavy Metals on Seed Germination and Early Seedling Growth of *Arabidopsis Thaliana*. *Plant Growth Regulation*, 46: 45-50.
- Wee, Y.C. (2003). *Tropical Trees and Shrubs: A Selection for Urban Planting*. Sun Tree Publishing, Singapore.
- Wilkins, D. (1957) A technique for the measurement of lead tolerance in plants. *Nature*, 180: 37-38.
- Wierzbicka, M. (1987). Lead translocation and localization in Allium cepa roots. *Canadian Journal of Botany*, 65: 1851-1860.
- Xu, J., Yang, J., Duan, X., Jiang, Y., & Zhang, P. (2014). Increased expression of native cytosolic Cu/Zn superoxide dismutase and ascorbate peroxidase improve tolerance to oxidative and chilling stresses in cassava (*Manihot esculenta* Crantz). *BMC Plant Biology*, 14: 208.
- Yao, Z., Jinhui, L., Henghua, X., & Conghai, Y. (2012). Review on Remediation Technologies of Soil Contaminated by Heavy Metals. *Procedia Environmental Sciences*, 16: 722-29.
- Yahya, Y. A. (2008). Biosorption of selected heavy metals by free and immobilized *pycnoporus sanguineus*: Batch and column studies. Unpublished master's thesis, University of Sciences Malaysia, Penang.
- Yanni, L. (2008). Effect of salt stress on seed germination and seedling growth of three salinity plants. *Journal of Biological Sciences*, 11: 1268-1272.
- Yang, X., Feng, Y., He, Z., & Stoffella, P.J. (2005). Molecular mechanisms of heavy

- metal hyperaccumulation and phytoremediation. *Journal of Trace Elements in Medicine and Biology*, 18: 339-353.
- Yoon, J., Cao, X., Zhou, Q., & Ma, L. Q. (2006). Accumulation of Pb, Cu, and Zn in native plants growing on a contaminated Florida site. *Science of The Total Environment*, 368: 456-464.
- Yusuf, M., Fariduddin, Q., Hayat, S., & Ahmad, A. (2011). Nickel: An Overview of Uptake, Essentiality and Toxicity in Plants. *Bulletin of Environmental Contamination and Toxicology*. 86: 1-17.
- Zhang, X., Zhong, T., Liu, L., & Ouyang, X. (2015). Impact of soil heavy metal pollution on food safety in China. *PLOS One*, 10: 0135182.
- Zhang, X., Xingfeng, Z., Xuehong, Z., Bo, G., Zhian, L., Hanping, X., Haifang, L., & Jian, L. (2014). Effect of cadmium on growth, photosynthesis, mineral nutrition and metal accumulation of an energy crop, king grass (*Pennisetum americanum* × *P. purpureum*). *Biomass and Bioenergy*, 67: 179-187.
- Zhaqiong, C., Xin, L., Yingwei, A., Jiao, C., Xiaoming, L., Jingyao, C., & Shihong, Z. (2018). Effects and mechanisms of revegetation modes on cadmium and lead pollution in artificial soil on railway rock cut slopes. *Science of the Total Environment*, 644: 1602-1611.
- Zheng, L.J., Liu, X.M., Lutz, U., & Peer, T. (2011). Effects of lead and EDTA-assisted lead on biomass, lead uptake and mineral nutrients in *Lespedeza chinensis* and *Lespedeza davidi*. *Water, Air, and Soil Pollution*, 220: 57-68.
- Zheng, N., Wang, Q., & Zheng, D. (2007). Health risk of Hg, Pb, Cd, Zn and Cu to the inhabitants around Huludao zinc plant in China via consumption of vegetables. *Science of The Total Environment*, 383: 81-89.
- Zhang, X., Zhang, X., Gao, B., Li, Z., Xia, H., Li, H., & Li, J. (2014). Effect of cadmium on growth, photosynthesis, mineral nutrition and metal accumulation of an energy crop, king grass (*Pennisetum americanum* × *P. purpureum*). *Biomass and Bioenergy*, 67: 179-187.
- Zhang, Z.C., Chen, B.X., & Qiu, B.S. (2010). Phytochelatin synthesis plays a similar role in shoots of the cadmium hyperaccumulator *Sedum alfredii* as in non-resistant plants. *Plant, Cell & Environment*, 33:1248-1255.

BIODATA OF STUDENT

Abderrahmane Zerkout was born on the 11th of August 1991 in Elkhroub town under Constantine State, Algeria. Abderrahmane attended Bidi Lwiza Primary School, Mouhamed Abdouh Secondary School, and Moustapha Kateb College. He obtained Bachelor of Science in Biology from University Constantine 1, and Master of Science in Conservation of Biodiversity and Sustainable Development from University of Science and Technology Houari Boumediene, all in Algeria. Pursuance to his keen interest in plant ecophysiology drives his intention to study phytoremediation of soil by hyperaccumulator plants at PhD level.

LIST OF PUBLICATIONS

Zerkout, A., Omar, H., Ibrahim, M. H., & Mustafa., M. (2018). Influence of Lead on In vitro Seed Germination and Early Radicle Development of *Acacia auriculiformis Cunn. Ex Benth* Species. *Annual Research & Review in Biology*, 1-12.

Zerkout, A., Omar, H., Ibrahim, M. H., Mustafa., M., & Rusea, G. (2020). *Acacia auriculiformis* as phytoextraction agent: a growth response, physiological tolerance and lead removal capability evaluation. *Jordan Journal of Biological Sciences*, (Accepted)



UNIVERSITI PUTRA MALAYSIA

STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

ACADEMIC SESSION : 2020/2021

TITLE OF THESIS / PROJECT REPORT :

LEAD TOXICITY AND PHYTOEXTRACTION POTENTIAL OF ACACIA AURICULIFORMIS A. CUNN. EX BENTH. TO REMEDIATE LEAD-POLLUTED SOIL

NAME OF STUDENT : ZERKOUT ABDERRAHMANE

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

1. This thesis/project report is the property of Universiti Putra Malaysia.
2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as:

*Please tick (✓)

CONFIDENTIAL

(Contain confidential information under Official Secret Act 1972).

RESTRICTED

(Contains restricted information as specified by the organization/institution where research was done).

✓

OPEN ACCESS

I agree that my thesis/project report to be published as hard copy or online open access.