



**UNIVERSITI PUTRA MALAYSIA**

**DEVELOPMENT OF AEROSOLIZED NIOSOME FORMULATION  
CONTAINING GEMCITABINE AND CISPLATIN FOR *IN-VITRO*  
CYTOTOXICITY AGAINST MRC5 AND A549 CELL LINES**

**NORFATIN IZZATIE BINTI MOHAMAD SAIMI**

**FS 2021 17**



**DEVELOPMENT OF AEROSOLIZED NIOSOME FORMULATION  
CONTAINING GEMCITABINE AND CISPLATIN FOR *IN-VITRO*  
CYTOTOXICITY AGAINST MRC5 AND A549 CELL LINES**

**By**

**NORFATIN IZZATIE BINTI MOHAMAD SAIMI**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra  
Malaysia, in Fulfilment of the Requirements for the Degree of Master of  
Science**

**November 2020**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

**DEVELOPMENT OF AEROSOLIZED NIOSOME FORMULATION CONTAINING GEMCITABINE AND CISPLATIN FOR *IN-VITRO* CYTOTOXICITY AGAINST MRC5 AND A549 CELL LINES**

By

**NURFATIN IZZATIE BINTI MOHAMAD SAIMI**

**November 2020**

**Chair : Norazlinaliza binti Salim, PhD**  
**Faculty : Science**

Gemcitabine (Gem) and cisplatin (Cis) are currently being used for lung cancer treatment has been proven to improve the life expectancy of cancer patients, but they are highly toxic in high dosages. Aerosolized niosome formulation containing a low-dosage Gem and Cis (NGC), as an alternative formulation for lung cancer chemotherapy treatment was developed. The idea of aerosolization is to able direct delivery of a niosome formulation to the smaller airways of the lung. Niosome is a closed-bilayer vesicle in aqueous media that formed from the self-assembly of nonionic surfactant with cholesterol (Chol). NGC was prepared using a very simple heating method and was further optimized by D-optimal mixture design. The optimized NGC with 2.00 wt.% of surfactant (Tween 65 : Span 60 with ratio of 2:1), 1.01 wt.% of cholesterol and 63.06 wt.% of glycerol solution was obtained, where the other components were kept constant. The actual particle size showed good correlation with the predicted particle size with residual standard error (RSE) value less than 5%. The physicochemical characterization of optimum NGC formulation showed that the particle size, polydispersity index (PDI), and zeta potential were 166.45 nm, 0.16, and -15.28 mV, respectively. Zeta potential obtained indicates good stability against aggregation meanwhile if the zeta potential approaching zero value will lead to aggregation. The optimized NGC remained stable at 27°C with no phase separation for up to 90 days. Optimized NGC surface tension was 35.49 mN/m with an aerosol output of 96.22%, which indicates its suitability as aerosolized formulation. An in vitro drug release study using the dialysis bag diffusion technique showed controlled release for both drugs for up to 24 h penetration. The result showed that percentage of Gem release in pH 7.4 (64.71%) was 2-fold than Gem in pH 6.7 (30.38%). Meanwhile, for Cis, the percentage drug release in pH 7.4 was lower (53.55 %) than in pH 6.7 (99.58%) after 24h. This was due to the nature of drug and pH of the release

medium affect the percentage of drug release. A cytotoxicity study against normal lung (MRC5) and lung cancer (A549) cell lines were performed by diluting and pipetting the formulation onto the cells. The results showed that the optimized NGC had reduced the cytotoxicity effects against both MRC5 and A549 cell lines when compared with the control (Gem + Cis alone) from very toxic ( $IC_{50} < 1.56 \mu\text{g/mL}$ ) to weakly toxic ( $IC_{50} = 280.00 \mu\text{g/mL}$ ) and moderately toxic ( $IC_{50} = 46.00 \mu\text{g/mL}$ ), after 72 h of treatment. These results suggested that optimized NGC has the potential to be used for aerosolized chemotherapy treatment which will be promising less toxicity toward normal cell and able to inhibit cancer cell with low multidrug dosage.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk ijazah Master Sains

**PEMBANGUNAN FORMULASI EROSEL NIOSOM MENGANDUNGI  
GEMCITABINE DAN CISPLATIN UNTUK KESITOTOKSIKAN *IN-VITRO*  
TERHADAP TITISAN SEL MRC5 DAN A549**

Oleh

**NORFATIN IZZATIE BINTI MOHAMAD SAIMI**

**November 2020**

**Pengerusi : Norazlinaliza binti Salim, PhD**  
**Fakulti : Sains**

Gemcitabine (Gem) dan Cisplatin (Cis) yang kini digunakan dalam rawatan kanser paru-paru telah terbukti dapat meningkatkan jangka hayat pesakit kanser, tetapi ianya sangat toksik dalam dos yang tinggi. Formulasi niosom erosol yang mengandungi dos Gem dan Cis yang rendah (NGC), sebagai formulasi alternatif bagi rawatan kemoterapi kanser paru-paru telah dikaji. Idea erosol ini adalah bagi membolehkan penghantaran terus formulasi niosom kepada bahagian saluran udara paru-paru yang paling kecil. Niosom adalah vesikel dwilapisan tertutup dalam medium akues yang terbentuk daripada pengumpulan sendiri surfaktan bukan ionik dengan kolesterol (Chol). NGC telah disediakan dengan menggunakan kaedah pemanasan yang sangat ringkas dan kemudiannya dioptimumkan dengan reka bentuk campuran D-optimum. NGC optimum yang mengandungi 2.00 wt.% surfaktan (Tween 65 : Span 60 dengan nisbah 2:1), 1.01 wt.% kolesterol dan 63.06 wt.% larutan gliserol telah dihasilkan, di mana kandungan komponen lain adalah tetap. Saiz zarah sebenar dan ramalan saiz zarah menunjukkan tiada perbezaan yang ketara dengan baki ralat piawaan (RSE) kurang daripada 5%. Perincian fizikokomia terhadap formulasi NGC optimum menunjukkan saiz zarah, indeks kepoliserakan (PDI), dan potensi zeta masing-masing adalah 166.45 nm, 0.16, dan -15.28 mV. Potensi zeta yang diperoleh menunjukkan tahap kestabilan yang baik melawan pengagregatan manakala sekiranya potensi zeta menghampiri nilai kosong akan menyebabkan pengagregatan berlaku. NGC optimum kekal stabil pada suhu 27°C tanpa pemisahan fasa selama 90 hari. Ketegangan permukaan NGC optimum adalah  $35.49 \pm 0.01$  mN/m dengan keluaran erosol sebanyak 96.22% yang menunjukkan kesesuaiannya sebagai formulasi erosol. Kajian pelepasan ubat secara in-vitro menggunakan teknik resapan beg dialisis menunjukkan pelepasan terkawal bagi kedua-dua ubat tersebut selama 24 jam penembusan. Keputusan menunjukkan kadar pelepasan Gem dalam pH 7.4 (64.71%) adalah 2 kali ganda berbanding Gem

dalam pH 6.7 (30.38%). Manakala bagi Cis, kadar pelepasan dalam pH 7.4 lebih rendah (53.55%) berbanding dalam pH 6.7 (99.58%) selepas 24 jam. Ini adalah bergantung kepada sifat natural ubat dan pH medium pelepasan yang mempengaruhi kadar pelepasan ubat. Kajian kesitotoksikan terhadap titisan sel paru-paru normal (MCR5) dan kanser paru-paru (A549) telah dilakukan. Hasil kajian menunjukkan bahawa NGC optimum telah mengurangkan kesan kesitotoksikan terhadap kedua-dua garis sel MRC5 dan A549 berbanding kawalan (Gem + Cis bersendirian) daripada sangat toksik ( $IC_{50} < 1.56 \mu\text{g/mL}$ ) kepada toksik lemah ( $IC_{50} = 280.00 \mu\text{g/mL}$ ) dan toksik sederhana ( $IC_{50} = 46.00 \mu\text{g/mL}$ ), selepas rawatan selama 72 jam. Keputusan ini mencadangkan bahawa NGC optimum boleh digunakan sebagai rawatan kemoterapi secara erosol yang akan menjanjikan ketoksikan yang lebih rendah terhadap sel normal dan mampu untuk merencat sel kanser dengan dos pelbagai ubat yang rendah.



## ACKNOWLEDGEMENTS

All praises and Gratitude are due to Him for giving me strength, health, knowledge, and passion in the completion of this study and thesis.

Here I would like to express my deepest gratitude and sincere appreciation to my beloved supervisor, Dr. Norazlinaliza Salim for her guidance, encouragement, and support along the study. I am also indebted to my supportive co-supervisors, Prof. Dr. Mohd Basyaruddin Abdul Rahman and Dr. Noraini Ahmad for their help in giving me useful guidance and suggestions to improve my research quality. My appreciation also goes to all my labmates from Universiti Putra Malaysia and Universiti Malaya especially Nadiatul Atiqah, Sharifah Nurfadhlin Afifah, Noor Fazriyana, Azren Aida, Auni Hamimi, Akmarina, Nurul Shahidah and Nurul Faiezin. Thank you for the support, advice, and memorable moments along my master's life.

I also would love to express my greatest love and thanks to my beloved parents, Mohamad Saimi Hamat and Fatimah Mohamed for their help, duas and encouragement throughout my journey. Also, my family, I would like to express my deepest affection of their continues love and support. Last but not least, I would like to express my appreciation to all those unnamed who have contributed directly or indirectly during my research. Alhamdulillah.



I certify that a Thesis Examination Committee has met on (date of viva voce) to conduct the final examination of (student's name) on his (her) thesis entitled ("Title of Thesis") in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the (insert the name of relevant degree).

Members of the Thesis Examination Committee were as follows:

**Name of Chairperson, PhD**

Title (e.g., Professor/Associate Professor/Ir; omit if irrelevant)

Name of Faculty

Universiti Putra Malaysia

(Chairman)

**Name of Examiner 1, PhD**

Title (e.g., Professor/Associate Professor/Ir; omit if irrelevant)

Name of Faculty

Universiti Putra Malaysia

(Internal Examiner)

**Name of Examiner 2, PhD**

Title (e.g., Professor/Associate Professor/Ir; omit if irrelevant)

Name of Faculty

Universiti Putra Malaysia

(Internal Examiner)

**Name of External Examiner, PhD**

Title (e.g., Professor/Associate Professor/Ir; omit if irrelevant)

Name of Department and/or Faculty

Name of Organisation (University/Institute)

Country

(External Examiner)

\_\_\_\_\_  
**(Insert name of current Deputy Dean)**

**(E.g. XXXXX XXXX, PhD)**

Professor and Deputy Dean

School of Graduate Studies

Universiti Putra Malaysia

Date:

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

**Norazlinaliza binti Salim, PhD**

Senior Lecturer  
Centre of Foundation Studies for Agricultural Science  
Universiti Putra Malaysia  
(Chairman)

**Mohd Basyaruddin bin Abdul Rahman, PhD**

Professor  
Faculty of Science  
Universiti Putra Malaysia  
(Member)

**Noraini binti Ahmad, PhD**

Senior Lecturer  
Faculty of Science  
Universiti Malaya  
(Member)

---

**ZALILAH MOHD SHARIFF, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 11 March 2021

## Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No.: Norfatin Izzatie Binti Mohamad Saimi GS46810

## Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: \_\_\_\_\_  
Name of Chairman  
of Supervisory  
Committee: Norazlinaliza binti Salim

Signature: \_\_\_\_\_  
Name of Member of  
Supervisory  
Committee: Mohd Basyaruddin bin Abdul  
Rahman

Signature: \_\_\_\_\_  
Name of Member of  
Supervisory  
Committee: Noraini binti Ahmad

## TABLE OF CONTENTS

	<b>Page</b>
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	iii
<b>ACKNOWLEDGEMENTS</b>	v
<b>APPROVAL</b>	vi
<b>DECLARATION</b>	viii
<b>LIST OF TABLES</b>	xii
<b>LIST OF FIGURES</b>	xiii
<b>LIST OF ABBREVIATIONS</b>	xiv
<b>CHAPTER</b>	
<b>1</b>	
<b>INTRODUCTION</b>	<b>1</b>
1.1 Background of Research	1
1.2 Problem Statement	2
1.3 Scope of Study	3
1.4 Objectives	4
<b>2</b>	
<b>LITERATURE REVIEW</b>	<b>4</b>
2.1 Lung Cancer	4
2.2 Anti-Cancer Drug	4
2.2.1 Gemcitabine	5
2.2.2 Cisplatin	5
2.2.3 Combination of Gemcitabine and Cisplatin	6
2.3 Nanocarrier Usage for Drug Delivery in Cancer Therapy	7
2.3.1 Solid Lipid Nanoparticles	7
2.3.2 Micelle	8
2.3.3 Dendrimer	8
2.3.4 Liposome	8
2.4 Niosome	9
2.4.1 General Composition of Niosome	9
2.4.2 Preparation of Niosome	13
2.4.3 Niosome in Pharmaceutical Application	15
2.5 Aerosol Drug Delivery	17
<b>3</b>	
<b>MATERIALS AND METHODS</b>	<b>20</b>
3.1 Materials	20
3.2 Drug Solubility Study	20
3.3 Preparation of NGC Formulation	20
3.4 Experimental Design	21
3.4.1 Verification of Models	22
3.4.2 Statistical Analysis	22
3.5 Physicochemical Characterization of Optimized Niosome Formulation	23

	3.5.1	Particle Size, Polydispersity Index and Zeta Potential Measurement	23
	3.5.2	pH Measurement	23
	3.5.3	Surface Tension Measurement	23
	3.5.4	Aerosol Performance Analysis	23
	3.6	Drug Entrapment Efficiency	24
	3.7	Morphology	24
	3.8	Stability Study	24
	3.8.1	Centrifugation Test	24
	3.8.2	Different Storage Test	24
	3.9	<i>In-Vitro</i> Drug Release Study	25
	3.9.1	Standard Preparation	25
	3.9.2	Drug Release	25
	3.9.3	Kinetic Release	25
	3.10	Cytotoxicity Analysis	26
	3.10.1	Statistical Analysis	27
<b>4</b>		<b>RESULTS AND DISCUSSION</b>	<b>28</b>
	4.1	Drug Solubility	28
	4.2	Preliminary Analysis of Percentage Range for NGC Composition	28
	4.3	Experimental Design and Model Fitting	29
	4.4	D-Optimal Analysis	31
	4.5	Verification of Model and Optimization of Niosome	32
	4.6	Physicochemical Characterization	34
	4.6.1	Particle Size and Polydispersity Index	34
	4.6.2	Zeta Potential	34
	4.6.3	pH Analysis	35
	4.6.4	Surface Tension Measurement	35
	4.6.5	Aerosol Output	35
	4.7	Drug Entrapment Efficiency	35
	4.8	Morphology	36
	4.9	Stability Analysis	37
	4.10	<i>In-Vitro</i> Drug Release Analysis	38
	4.10.1	Kinetic Release Model	41
	4.11	<i>In-Vitro</i> Cytotoxicity Analysis Against MRC5 and A549 Cell Lines	42
<b>5</b>		<b>CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH</b>	<b>45</b>
	5.1	Conclusion	45
	5.2	Recommendations for Future Research	45
		<b>REFERENCES</b>	<b>47</b>
		<b>APPENDICES</b>	<b>57</b>
		<b>BIODATA OF STUDENT</b>	<b>64</b>
		<b>LIST OF PUBLICATIONS</b>	<b>65</b>

## LIST OF TABLES

Table		Page
3.1	Composition for NGC formulation	21
3.2	Level of independent variables proportions	22
4.1	The solubility of Gem and Cis in NaCl solution (0.90% w/v)	28
4.2	Predicted and actual values of particles size of NGC obtained from D-optimal mixture experimental design	30
4.3	Analysis of variance (ANOVA) for the model derived by D-optimal mixture design	31
4.4	Validation sets of NGC formulation	33
4.5	The optimum composition of NGC formulation	33
4.6	The penetration rate of Gem and Cis release for the first 8 h	41
4.7	The coefficient of determination ( $R^2$ ) for different models of mechanisms of drug release from optimized NGC at different pH of SLF	41
4.8	IC <sub>50</sub> values of the samples after 72 h of treatment	44

## LIST OF FIGURES

Figure		Page
2.1	A molecular structure of gemcitabine	5
2.2	A molecular structure of cisplatin	6
2.3	Structure of niosome	9
2.4	The three types of niosome	9
2.5	A schematic diagram of nonionic surfactant	10
2.6	Molecular structures of (a) Span 60 and (b) Tween 65	11
2.7	A molecular structure of cholesterol	12
2.8	A molecular structure of SDS	13
2.9	(a) A schematic diagram of aerosol drug delivery and (b) Particle size distribution of aerosol particles pass through the lung	18
2.10	(a) Omron Micro-Air vibrating mesh nebulizer and (b) Mechanism operation vibrating-mesh nebulizer	19
4.1	3D surfaces showing the interaction effect of surfactants (T65:S60) ratio 2:1 (A), cholesterol (B), and glycerol solution (C) against particle size	32
4.2	Representative of the particle size distribution of the optimized NGC	34
4.3	TEM image of the optimized NGC at 25,000x magnification	36
4.4	(a) Particle size of the optimized NGC as a function of time at different storage temperatures. (b) Physical appearance of optimized NGC formulation on day 1 and day 90	38
4.5	UV-visible spectra of Gem, Cis, Gem-Cis, and the optimized NGC	39
4.6	Profiles of cumulative percentage release of gemcitabine and cisplatin from the optimized NGC at (a) pH 6.7, and (b) pH 7.4 of SLF for 24 h	40
4.7	Cytotoxicity effect against (a) MRC5 and (b) A549 cell lines	43



## LIST OF ABBREVIATIONS

ANOVA	Analysis of variance
Au	Uranyl acetate
Cis	Cisplatin
CSC	Cancer stem-like cells
°C	Degree Celsius
DLS	Dynamic Light Scattering
DOX	Doxorubicin
DNA	Deoxyribonucleic acid
EE	Entrapment efficiency
g	Gram
Gem	Gemcitabine
h	Hour
HA-Pt	Cisplatin-hyaluronan
HLB	Hydrophile-lipophile balance
IC <sub>50</sub>	Concentration of an inhibitor where the response (or binding) is reduced by half
M°	Initial concentration
MDR	Multidrug resistance
MED	Mixture Experimental Design
mg/L	Milligram/liter
min	Minute
MTT	3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyltetrazolium bromide
mV	Milivolt
NaCl	Sodium chloride
nm	Nanometer
NGC	Niosome formulation containing gemcitabine and cisplatin
NSCLC	Non-small cell lung carcinoma
PAMAM	Polyamidoamine
PBS	Phosphate buffer saline
PDI	Polydispersity index
PEG	Polyethylene glycol
rpm	Revolutions per minute
RSE	Residual standard error
RT	Room temperature
R <sup>2</sup>	Coefficient of determination
SCLC	Small-cell lung carcinoma
SD	Standard deviation
SDS	Sodium dodecyl sulfate
SLF	Simulated lung fluid
SLN	Solid lipid nanoparticles
SPIONS	Superparamagnetic iron oxide nanoparticles
TEM	Transmission electron microscopy
UV/Vis	Ultraviolet-visible

## CHAPTER 1

### INTRODUCTION

#### 1.1 Background of Research

Over the past fifty years, the main public health concern for cancer diseases has increased especially lung cancer which is the most common cancer diagnosed in the world followed by colorectal, liver and breast cancer and contributing to 37% of the leading cause of cancer-related death worldwide from total death in 2018 (Bray *et al.*, 2018). There are two main categories of lung cancer, which are non-small cell lung carcinoma (NSCLC), which contributes 85% of cases while small-cell lung carcinoma (SCLC) contributes 15% cases from all lung cancer cases (Zappa & Mousa, 2016).

A significant factor that leads to lung cancer is cigarette smoking (Hammerschmidt & Wirtz, 2009). However, increasing lung cancer cases among non-smokers are quite alarming nowadays (Planchard & Besse, 2015). Different stages of lung cancer with different treatments are the main reason for the low survival rate of cancer patients. Likewise, in other cancer cases, detection and confirmation usually can only be made at stage four (IV) (Akhter, 2015). The current treatments used for lung cancer are surgery, chemotherapy and radiation therapy, depending on the stage and overall performance of cancer cells. For the advanced stage of lung cancer, chemotherapy is the first-line treatment where most of the treatment is applied as intravenous (IV) formulations into systematic circulation (Quan *et al.*, 2019).

Cancer chemotherapeutic drugs are usually used to destroy cancer cells by preventing the cancer cell from growing, dividing, and producing more cells (Lee *et al.*, 2015). Conventionally, a patient who undergoes chemotherapy will be given a single or multi-drug at a time. For example, it is either cisplatin (Cis) or gemcitabine (Gem) alone or a combination of both drugs (Ma *et al.*, 2017). The combination with other cancer treatments such as radiation therapy, surgery followed by several cycles of chemotherapy has also been applied (Khuda-Bukhsh *et al.*, 2014). The current drugs used for lung cancer chemotherapy treatment are cisplatin, carboplatin, paclitaxel, gemcitabine, docetaxel, and vinorelbine (Zappa & Mousa, 2016).

The demand for nano-carriers for drug administration in medical applications, especially in chemotherapy treatment, has increased as drug development is advancing. Nano-carriers are believed to have great potential in the delivery of single or multiple drugs (at low dosage) to tissue tumours by overcoming biological barriers and can reach the tiniest area in the body due to their small

particle size. Furthermore, most of the nanocarrier systems increase the effectiveness of chemotherapy and minimize side effects by enhancing the deposition of the drug at the tumour based on their stability and permeability (Majumder, Taratula, & Minko, 2019).

Nanocarriers provide a larger surface area, have the potential to increase solubility, improve the controlled release of the drug, and enhance the bioavailability in the delivery of chemotherapy drugs to the targeted tumour area (Majeed *et al.*, 2016). There are a few of nano-carriers used as targeting drugs delivery systems such as micelles, dendrimers, nanoparticles, liposomes and niosomes (Kang *et al.*, 2015; Khodabandehloo *et al.*, 2016). Among those nano-carriers, liposomes and niosomes have great potential in the delivery of multiple drug. Both have efficiencies in encapsulating hydrophilic and hydrophobic drugs into the aqueous layer and the lipid bilayer, respectively (Saraswathi *et al.*, 2019). However, liposomes are less suitable because of their instability due to the oxidation or hydrolysis of phospholipids, where they require special storage conditions in a dark area sealed with nitrogen, and the materials used to produce liposomes are expensive (Wen *et al.*, 2018).

Niosome is a system with bilayer spherical vesicles formed by self-assemblies of nonionic surfactant and cholesterol in an aqueous medium (El-badry *et al.*, 2014; Ertekin *et al.*, 2015). Nonionic surfactants are commonly used in the preparation of niosome due to their high degree of compatibility with other ingredients. As membrane additives to the bilayer composition of niosome, cholesterol enhances the stability and reduces leakage of vesicles which will increase the entrapment efficiency of the drug (Gharbavi *et al.*, 2018). Niosomes are chemically stable, have high compatibility with biological systems, and low toxicity because of their nonionic nature (Ge *et al.*, 2019). Therefore, it is not surprising that niosomes (as nanocarrier) are currently being used to deliver drugs such as (i) 5-Fluorouracil, increasing drug penetration in skin cancer treatment; (ii) tamoxifen citrate, providing higher cytotoxicity against breast cancer cells; and (iii) curcumin, giving more significant apoptotic effect towards ovarian cancer cells.

## 1.2 Problem Statements

Generally, Gem and Cis are administered at a higher dosage of 1.250 and 0.075 g/m<sup>2</sup> per cycle, respectively, which will lead to higher toxicity (Fan *et al.*, 2010). However, it was found that the combination of drugs (Gem + Cis) improved the survival rate among cancer patients compared to the use of a single drug (Pande *et al.*, 2012). This has shown that the optimum dosage of the combination of drugs can be effective in increasing the survival rate of lung cancer patients as well as helping to reduce side effects such as vomiting, sore throat, chest congestion, and nausea (Liu *et al.*, 2015).

Most of the chemotherapeutics drugs are unstable with a short half-life and fast clearance from the body system. Furthermore, cancer chemotherapeutic drugs that are administered orally are always limited due to the first-pass metabolism, where the drug concentration is usually reduced as it passes through the gastrointestinal tract before the drug can access the systematic circulation (Sohail *et al.*, 2018). Multi-drug resistance (MDR) in lung cancer stem-like cells (CSC) becomes a limitation to the combination of cancer therapy even though the dual drug has the potential to improve cancer treatment by reducing the drug dosage which leads to less side effects (Huang *et al.*, 2016). These problems led to less effective lung cancer chemotherapy treatment. Thus, a more efficient carrier for this combination of drugs should be developed to improve the delivery of both drugs to the lung (targeted site) via the alternative route of delivery.

### 1.3 Scope of Study

In this study, aerosolized niosome (NGC) formulation containing Gem and Cis was prepared using the heating method (hydration method of niosome component without using any hazardous chemical). Screening of the compositions of the formulation used has been performed. The optimization of NGC formulation compositions was carried out using a mixture experimental design (MED). The physicochemical characterization and stability of the optimized NGC formulation were investigated. *In-vitro* drug release study in simulated lung fluid was performed at pH 6.7 and 7.4 using dialysis bag method, and a cytotoxicity study against normal healthy MRC5 and lung cancer A549 cells was further investigated. The aerosol output was determined in order to calculate the percentage of the aerosolized formulation.

### 1.4 Objectives

The main objective of this study was to develop aerosolized niosome formulation containing gemcitabine and cisplatin (NGC). In order to successfully achieved the main objective, the following specific objectives have been carried out:

1. To formulate and optimize niosome formulation containing gemcitabine and cisplatin (NGC) using a mixture experimental design (MED).
2. To characterize the physicochemical properties and evaluate the stability of the optimized NGC formulation.
3. To determine the *in vitro* cytotoxicity and drug release of the optimized NGC.

## REFERENCES

- Ahmad, J., Akhter, S., Rizwanullah, M., Amin, S., Rahman, M., Ahmad, M. Z., Rizvi, M. A. Kamal, M. A., Ahmad, F. J. (2015). Nanotechnology-based inhalation treatments for lung cancer: state of the art. *Nanotechnology, Science and Applications*, 8, 55–66.
- Akbarzadeh, I., Tavakkoli, M., & Bourbour, M. (2020). Optimized doxycycline-loaded niosomal formulation for treatment of infection-associated prostate cancer: An in-vitro investigation. *Journal of Drug Delivery Science and Technology*, 57 (February), 101715.
- Aldossary, S. A. (2019). Review on pharmacology of cisplatin: Clinical use, toxicity and mechanism of resistance of cisplatin. *Biomedical and Pharmacology Journal*, 12(1), 7–15.
- Amaani, R., & Dwira, S. (2018). Phytochemical content and in vitro toxicity of Glycine soja ethanol extract on the A549 Lung cancer line cell. *Journal of Physics*, (1073), 1–9.
- Anbarasan, B., Rekha, S., Elango, K., Shriya, B., & Ramaprabhu, S. (2013). Optimization of The Formulation and In-Vitro Evaluation of Capecitabine Niosomes for The Treatment of Colon Cancer. *International Journal of Pharmaceutical Sciences and Research*, 4(4), 1504–1513.
- Antus, B., & Barta, I. (2012). Exhaled breath condensate pH in patients with lung cancer. *Lung Cancer*, 75(2), 178–180.
- Arbain, N. H., Salim, N., Masoumi, H. R. F., Wong, T. W., Basri, M., & Abdul Rahman, M. B. (2018). In vitro evaluation of the inhalable quercetin loaded nanoemulsion for pulmonary delivery. *Drug Delivery and Translational Research*, 1(1), 1–11.
- Arbain, N. H., Salim, N., Wui, W. T., Basri, M., Basyaruddin, M., & Rahman, A. (2018). Optimization of Quercetin loaded Palm Oil Ester Based Nanoemulsion Formulation for Pulmonary Delivery. *Journal of Oleo Science*, 1–8.
- Arul Jothy, M., Shanmuganathan, S., & Nagalakshmi. (2015). An overview on niosome as carrier in dermal drug delivery. *Journal of Pharmaceutical Sciences and Research*, 7(11), 923–927.
- Aryal, S., Jack Hu, C. M., Fu, V., & Zhang, L. (2012). Nanoparticle drug delivery enhances the cytotoxicity of hydrophobic-hydrophilic drug conjugates. *Journal of Materials Chemistry*, 22(3), 994–999.
- Ashutosh, L., Suman, R., Sidhyartha, S., Altaf, S., & Vandana, Y. M. (2012). A Novel Drug Delivery System: Niosomes Review. *Journal of Drug Delivery & Therapeutics*, 2(5), 129–135.

- Asmawi, A. A., Salim, N., Abdulmalek, E., & Rahman, M. B. A. (2020). Development and validation of hplc method for quantification of docetaxel in palm-based nanoemulsion aerosols. *Malaysian Journal of Analytical Sciences*, 24(2), 165–172.
- Barani, M., Mirzaei, M., Torkzadeh-Mahani, M., & Nematollahi, M. H. (2018). Lawsons-loaded Niosome and its antitumor activity in MCF-7 breast Cancer cell line: a Nano-herbal treatment for Cancer. *Journal of Pharmaceutical Sciences*, 26(1), 11–17.
- Barani, M., Nematollahi, M. H., Zaboli, M., Mirzaei, M., Torkzadeh-mahani, M., Pardakhty, A., & Karam, G. A. (2019). In silico and in vitro study of magnetic niosomes for gene delivery: The effect of ergosterol and cholesterol. *Materials Science & Engineering C*, 94, 234–246.
- Basiri, L., Rajabzadeh, G., & Bostan, A. (2017a).  $\alpha$ -Tocopherol-loaded niosome prepared by heating method and its release behavior. *Food Chemistry*, 221, 620–628.
- Bhardwaj, P., Tripathi, P., Gupta, R., & Pandey, S. (2020). Niosomes : A review on niosomal research in the last decade. *Journal of Drug Delivery Science and Technology*, 56(February), 101581.
- Biswal, S., Murthy, P. N., Sahu, J., Sahoo, P., & Amir, F. (2008). Vesicles of non-ionic surfactants (niosomes) and drug delivery potential. *Int J Pharm Sci Nanotechnol*, 1(1), 1–8.
- Bnyan, R., Khan, I., Ehtezazi, T., Saleem, I., Gordon, S., Neill, F. O., & Roberts, M. (2018). Surfactant effects on lipid-based vesicles properties. *Journal of Pharmaceutical Sciences*, 1–25.
- Bray, F., Ferlay, J., Soerjomataram, I., Siegel, R. L., Torre, L. A., & Jemal, A. (2018). Global Cancer Statistics 2018: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA: A Cancer Journal for Clinicians*, 68, 394–424.
- Callion, O. N. M. M., Taylor, K. M. G., Thomas, M., & Taylor, A. J. (1996). The influence of surface tension on aerosols produced by medical nebulisers. *International Journal of Pharmaceutics*, 129, 123–136.
- Chen, S., Hanning, S., Falconer, J., Locke, M., & Wen, J. (2019). Recent advances in non-ionic surfactant vesicles ( niosomes ): Fabrication , characterization , pharmaceutical and cosmetic applications. *European Journal of Pharmaceutics and Biopharmaceutics*, 144(May), 18–39.
- Cipolla, D., Gonda, I., & Chan, H.-K. (2013). Liposomal formulations for inhalation. *Therapeutic Delivery*, 4(8), 1047–1072.
- Crul, M., Van Waardenburg, R. C. A. M., Bocxe, S., Van Eijndhoven, M. A. J., Pluim, D., Beijnen, J. H., & Schellens, J. H. M. (2003). DNA repair mechanisms involved in gemcitabine cytotoxicity and in the interaction

- between gemcitabine and cisplatin. *Biochemical Pharmacology*, 65(2), 275–282.
- Dasari, S., & Bernard Tchounwou, P. (2014). Cisplatin in cancer therapy: Molecular mechanisms of action. *European Journal of Pharmacology*, 740, 364–378.
- De, A., Venkatesh, N., Senthil, M., Sanapalli, B. K. R., Shanmugham, R., & Karri, V. V. S. R. (2018). Smart niosomes of temozolomide for enhancement of brain targeting. *Nanobiomedicine*, 5, 1–11.
- De Sousa Cavalcante, L., & Monteiro, G. (2014). Gemcitabine: Metabolism and molecular mechanisms of action, sensitivity and chemoresistance in pancreatic cancer. *European Journal of Pharmacology*, 741, 8–16.
- Degner, B. M., Chung, C., Schlegel, V., Hutkins, R., & McClements, D. J. (2014). Factors Influencing the Freeze-Thaw Stability of Emulsion-Based Foods. *Comprehensive Reviews in Food Science and Food Safety*, 13, 98–113.
- Du, C., Li, S., Li, Y., Galons, H., Guo, N., Teng, Y., Zhang, Y., Li, M., Yu, P. (2020). F7 and topotecan co-loaded thermosensitive liposome as a nano-drug delivery system for tumor hyperthermia. *Drug Delivery*, 27(1), 836–847.
- Duque, M. D., Kreidel, R. N., Taqueda, M. E. S., Rolim, A., Baby, Kaneko, T. M., Velasco, M., Consiglieri, V. O. (2015). Optimization of primaquine diphosphate tablet formulation for controlled drug release using the mixture experimental design. *Pharmaceutical Development and Technology*, (May), 1–8.
- El-badry, M., Fetih, G., Fathalla, D., & Shakeel, F. (2014). Transdermal delivery of meloxicam using niosomal hydrogels: in vitro and pharmacodynamic evaluation. *Pharmaceutical Development and Technology*, 20(7), 820–826.
- Emamzadeh, M., Emamzadeh, M., & Pasparakis, G. (2019). Dual Controlled Delivery of Gemcitabine and Cisplatin Using Polymer-Modified Thermosensitive Liposomes for Pancreatic Cancer. *ACS Applied Bio Materials*, 2(3), 1298–1309.
- Ertekin, Z., Bayindir, Z., & Yuksel, N. (2015). Stability Studies on Piroxicam Encapsulated Niosomes. *Current Drug Delivery*, 12(2), 192–199.
- Essa, E. (2010). Effect of formulation and processing variables on the particle size of sorbitan monopalmitate niosomes. *Asian Journal of Pharmaceutics*, 4(4), 227–233.
- Fan, Y., Lin, N. M., Ma, S. L., Luo, L. H., Fang, L., Huang, Z. Y., Yu, H. F., Wu, F. Q. (2010). Phase II trial of gemcitabine plus cisplatin in patients with advanced non-small cell lung cancer. *Acta Pharmacologica Sinica*, 31(6),

746–752.

- Fathi-Azarbayjani, A., Jouyban, A., & Chan, S. Y. (2009). Impact of surface tension in pharmaceutical sciences. *Journal of Pharmacy and Pharmaceutical Sciences*, 12(2), 218–228.
- Fraile, R., Geanta, R. M., Escudero, I., Benito, J. M., & Ruiz, M. O. (2014). Formulation of Span 80 niosomes modified with SDS for lactic acid entrapment. *Desalination and Water Treatment*, 56(13), 3463–3475.
- Ge, X., Wei, M., He, S., & Yuan, W. (2019). Advances of Non-Ionic Surfactant Vesicles ( Niosomes ) and Their Application in Drug Delivery. *Pharmaceutics*, 11(55), 1–16.
- Gharbavi, M., Amani, J., Kheiri-Manjili, H., Danafar, H., & Sharafi, A. (2018). Niosome: A Promising Nanocarrier for Natural Drug Delivery through Blood-Brain Barrier. *Advances in Pharmacological Sciences*, 1–15.
- Gong, Z., Chen, M., Ren, Q., Yue, X., & Dai, Z. (2020). Fibronectin-targeted dual-acting micelles for combination therapy of metastatic breast cancer. *Signal Transduction and Targeted Therapy*, 5(12), 1–11.
- Gouda, R., Baishya, H., & Qing, Z. (2017). Application of Mathematical Models in Drug Release Kinetics of Carbidopa and Levodopa ER Tablets. *Journal of Developing Drugs*, 06(02), 1–8.
- Guo, X. Y., Wang, P., Du, Q. G., Han, S., Zhu, S. M., Lv, Y. F., Liu, G. -S., Hao, Z. M. (2015). Paclitaxel and gemcitabine combinational drug-loaded mucoadhesive delivery system in the treatment of colon cancers. *Drug Research*, 65(4), 199–204.
- Gupta, M., Vaidya, B., Mishra, N., & Vyas, S. P. (2011). Effect of Surfactants on the Characteristics of Fluconazole Niosomes for Enhanced Cutaneous Delivery. *Artificial Cells, Blood Substitutes, and Biotechnology*, 39, 376–384.
- Hammerschmidt, S., & Wirtz, H. (2009). Lung cancer: current diagnosis and treatment. *Deutsches Ärzteblatt International*, 106(49), 809–820.
- Hayashi, K., Shimanouchi, T., Kato, K., Miyazaki, T., & Nakamura, A. (2011). Colloids and Surfaces B: Biointerfaces Span 80 vesicles have a more fluid , flexible and “ wet ” surface than phospholipid liposomes. *Colloids and Surfaces B: Biointerfaces*, 87(1), 28–35.
- Hess, D. R. (2008). Aerosol delivery devices in the treatment of asthma. *Respiratory Care*, 53(6), 699–723.
- Honary, S., & Zahir, F. (2013). Effect of Zeta Potential on the Properties of Nano-Drug Delivery Systems - A Review ( Part 1 ). *Tropical Journal of Pharmaceutical Research*, 12(2), 255–264.



- Hong, S.-H., Park, S.-J., Lee, S., Cho, C. S., & Cho, M.-H. (2015). Aerosol gene delivery using viral vectors and cationic carriers for *in vivo* lung cancer therapy. *Expert Opinion on Drug Delivery*, 12(6), 977–991.
- Huang, W. T., Larsson, M., Lee, Y. C., Liu, D. M., & Chiou, G. Y. (2016). Dual drug-loaded biofunctionalized amphiphilic chitosan nanoparticles: Enhanced synergy between cisplatin and demethoxycurcumin against multidrug-resistant stem-like lung cancer cells. *European Journal of Pharmaceutics and Biopharmaceutics*, 109, 165–173.
- Irvin, S. S., & Janet, L. S. (2014). Anticancer Drug. *Encyclopaedia Britannica*, 1–2.
- Jain, C. P., & Vyas, S. P. (1995). Preparation and characterization of niosomes containing rifampicin for lung targeting. *Journal of Microencapsulation*, 12(4), 401–407.
- Jyoti, K., Pandey, R. S., Madan, J., & Jain, U. K. (2016). Inhalable cationic niosomes of curcumin enhanced drug delivery and apoptosis in lung cancer cells. *Indian Journal of Pharmaceutical Education and Research*, 50(2), S21–S31.
- Kanaani, L., Ebrahimifar, M., Ebrahimi, H., Khiyavi, A. A., & Mehrdiba, T. (2017). Effects of Cisplatin-Loaded Niosomal Nanoparticles on BT-20 Human Breast Carcinoma Cells. *Asian Pacific Journal of Cancer Prevention*, 18(2), 365–368.
- Kang, L., Gao, Z., Huang, W., Jin, M., & Wang, Q. (2015). Nanocarrier-mediated co-delivery of chemotherapeutic drugs and gene agents for cancer treatment. *Acta Pharmaceutica Sinica B*, 5(3), 169–175.
- Karim, K., Kuotsu, K., Mandal, A., Chatterjee, S., Guha, A., Biswas, N., & Behera, M. (2011). Niosome: A future of targeted drug delivery systems. *Journal of Advanced Pharmaceutical Technology & Research*, 1(4), 374–380.
- Kaur, T., Kaur, S., & Kaur, P. (2017). Development and validation of UV-spectrophotometric methods for determination of gemcitabine hydrochloride in bulk and polymeric nanoparticles. *International Journal of Applied Pharmaceutics*, 9(5), 60–65.
- Khodabandehloo, H., Zahednasab, H., & Hafez, A. A. (2016). Nanocarriers Usage for Drug Delivery in Cancer Therapy. *Iranian Journal of Cancer Prevention*, 9(2), 1–6.
- Khuda-Bukhsh, A.R., Mondal, J., Panigrahi, A. K. (2014). Conventional chemotherapy : problems and scope for combined therapies with certain herbal products and dietary supplements. *Austin Journal of Molecular and Cellular Biology*, 1(1), 1–10.
- Kroep, J. R., Smit, E. F., Giaccone, G., Born, K. Van der, Beijnen, J. H.,

- Groeningen, C. J. Van, Van der Vijgh, W. J. F., Postmus, P. E., Pinedo, H. M., Peters, G. J. (2006). Pharmacology of the paclitaxel – cisplatin , gemcitabine – cisplatin , and paclitaxel – gemcitabine combinations in patients. *Cancer Chemother Pharmacol*, 58, 509–516.
- Kulkarni, P., & Rawtani, D. (2019). Application of Box-Behnken Design in the Preparation, Optimization, and In Vitro Evaluation of Self-Assembly-Based Tamoxifen- and Doxorubicin-Loaded and Dual Drug-Loaded Niosomes for Combinatorial Breast Cancer Treatment. *Journal of Pharmaceutical Sciences*, 108(8), 2643–2653.
- Kumar, G. P., & Rajeshwarrao, P. (2011). Nonionic surfactant vesicular systems for effective drug delivery—an overview. *Acta Pharmaceutica Sinica B*, 1(4), 208–219.
- Lai, Y.-L., Lin, C.-C., Hsu, S.-R., & Yen, S.-K. (2018). Electrochemical Deposition of Cisplatin on Pure Magnesium. *Journal of The Electrochemical Society*, 165(5), D196–D205.
- Latimer, K. M., & Mott, T. F. (2015). Lung cancer: Diagnosis, treatment principles, and screening. *American Family Physician*, 91(4), 250–256.
- Lee, W., Loo, C., Traini, D., & Young, P. M. (2015). Inhalation of nanoparticle-based drug for lung cancer treatment : Advantages and challenges. *Asian Journal of Pharmaceutical Sciences*, 10(6), 481–489.
- Liu, B., Ezeogu, L., Zellmer, L., Yu, B., Xu, N., & Liao, D. J. (2015). Protecting the normal in order to better kill the cancer. *Cancer Medicine*, 4(9), 1394–1403.
- Ma, D., Wang, J., Hao, X., Wang, Y., Hu, X., Xing, P., & Li, J. (2017). Gemcitabine combined with cisplatin as adjuvant chemotherapy for non-small cell lung cancer : A retrospective analysis. *Thoracic Cancer*, 8, 482–488.
- Mahale, N. B., Thakkar, P. D., Walunj, D. R., & Chaudhari, S. R. (2012). Niosomes : Novel sustained release nonionic stable vesicular systems — An overview. *Advances in Colloid and Interface Science*, 183–184, 46–54.
- Majeed, M., Hussain, A. I., Chatha, S. A. S., Khosa, M. K. K., Kamal, G. M., Kamal, M. A., Xu, Z., Liu, M. (2016). Optimization protocol for the extraction of antioxidant components from *Origanum vulgare* leaves using response surface methodology. *Saudi Journal of Biological Sciences*, 23(3), 389–396.
- Majumder, J., Taratula, O., & Minko, T. (2019). Nanocarrier-based systems for targeted and site specific therapeutic delivery. *Advanced Drug Delivery Reviews*, 144, 57–77.
- Manohar, S., & Leung, N. (2018). Cisplatin nephrotoxicity: a review of the

literature. *Journal of Nephrology*, 31(1), 15–25.

- Manosroi, A., Wongtrakul, P., Manosroi, J., Sakai, H., Sugawara, F., Yuasa, M., & Abe, M. (2003). Characterization of vesicles prepared with various non-ionic surfactants mixed with cholesterol. *Colloids and Surfaces B: Biointerfaces*, 30(1–2), 129–138.
- Marianecci, C., Di, L., Rinaldi, F., Celia, C., Paolino, D., Alhaique, F., Esposito, S., Carafa, M. (2014). Niosomes from 80s to present : The state of the art. *Advances in Colloid and Interface Science*, 205, 187–206.
- Martinho, N., Santos, T. C. B., Florindo, H. F., & Silva, L. C. (2019). Cisplatin-membrane interactions and their influence on platinum complexes activity and toxicity. *Frontiers in Physiology*, 10(JAN), 1–15.
- Moazeni, E., Gilani, K., Sotoudegan, F., Pardakhty, A., Najafabadi, A. R., Ghalandari, R., Fazeli, M. R., Jamalifar, H. (2010). Formulation and in vitro evaluation of ciprofloxacin containing niosomes for pulmonary delivery. *Journal of Microencapsulation*, 27(7), 618–627.
- Moghassemi, S., & Hadjizadeh, A. (2014). Nano-niosomes as nanoscale drug delivery systems: An illustrated review. *Journal of Controlled Release*.
- Mohd Nadzir, M., Tan, W. F., Abdul Rahman, M., & Hisham, S. F. (2017). Size and Stability of Curcumin Niosomes from Combinations of Tween 80 and Span 80. *Sains Malaysiana*, 46(12), 2455–2460.
- Morsy, S. M. I. (2014). Role of Surfactants in Nanotechnology and Their Applications. *International Journal of Current Microbiology Applied Science*, 3(5), 237–260.
- Mozafari, M.Reza. (2005). Liposomes: An overview of manufacturing techniques. *Cellular & Molecular Biology Letters*, 10, 711–719.
- Mozafari, M R. (2005). *A new technique for the preparation of non-toxic liposomes and nanoliposomes: The heating method. Nanoliposomes: From Fundamentals to Recent Developments.*
- Navya, M. N., Parthiban, S., Neasalin, J. A. J., & Vikneswari, A. (2014). Niosome as Novel Vesicular drug Delivery System - A Review. *Asian Journal of Research in Biological and Pharmaceutical Sciences*, 2(2), 62–68.
- Nazari-Vanani, R., Karimian, K., Azarpira, N., & Heli, H. (2019). Capecitabine-loaded nanoniosomes and evaluation of anticancer efficacy. *Artificial Cells, Nanomedicine and Biotechnology*, 47(1), 420–426.
- Niraula, T. P., Bhattarai, A., & Chatterjee, S. K. (2014). Sodium dodecyl sulphate: A very useful surfactant for Scientific Invetigations. *The Journal of Knowledge and Innovation*, 2(1), 111–113.

- Onischuk, A. A., Tolstikova, T. G., Baklanov, A. M., & Khvostov, M. V. (2014). Generation , inhalation delivery and anti-hypertensive effect of nisoldipine nanoaerosol. *Journal of Aerosol Science*, 78, 41–54.
- Osman, N., Kaneko, K., Carini, V., & Saleem, I. (2018). Carriers for the Targeted Delivery of Aerosolized Macromolecules for Pulmonary Pathologies. *Expert Opinion on Drug Delivery*, 0(0), 1.
- Pande, S. B., Doval, D. C., Pavithran, K., Sharma, J. B., Shirali, R., & Jena, A. (2012). Gemcitabine and cisplatin-based combination chemotherapy in advanced hepatocellular carcinoma: An Indian experience. *Indian Journal of Medical and Paediatric Oncology*, 33(1), 42–47.
- Paolino, D., Cosco, D., Muzzalupo, R., Trapasso, E., Picci, N., & Fresta, M. (2008). Innovative bola-surfactant niosomes as topical delivery systems of 5-fluorouracil for the treatment of skin cancer. *International Journal of Pharmaceutics*, 353(1–2), 233–242.
- Pardakhty, A., & Moazeni, E. (2013). Nano-niosomes in drug , vaccine and gene delivery : a rapid overview. *Nanomedicine Journal*, 1(1), 1–12.
- Peate, I. (2018). Lung cancer. *British Journal of Healthcare Assistants*, 12(9), 429–435.
- Planchard, D., & Besse, B. (2015). Lung cancer in never-smokers. *European Respiratory Journal*, 47, 1214–1217.
- Pritchard, J. N., Hatley, R. H. M., Denyer, J., & Von Hollen, D. (2018). Mesh nebulizers have become the first choice for new nebulized pharmaceutical drug developments. *Therapeutic Delivery*, 9(2), 121–136.
- Propst, C. N., Nwabueze, A. O., Kanev, I. L., Pepin, R. E., Gutting, B. W., Morozov, V. N., & Hoek, M. L. Van. (2016). Nanoaerosols reduce required effective dose of liposomal levofloxacin against pulmonary murine Francisella tularensis subsp. novicida infection. *Journal of Nanobiotechnology*, 1–10.
- Quan, A., Chen-xiao, S., Hao, G., Shi-min, X., Ying-ying, Y., Ying-nan, L., Liu, Z., Zhou, C., Feng-ju, N. (2019). Development and characterization of octreotide-modified curcumin plus docetaxel micelles for potential treatment of non-small cell lung cancer. *Pharmaceutical Development and Technology*, 24(9), 1164–1174.
- Ruckmani, K., Jayakar, B., & Ghosal, S. K. (2000). Nonionic surfactant vesicles (niosomes) of cytarabine hydrochloride for effective treatment of leukemias: Encapsulation, storage, and in vitro release. *Drug Development and Industrial Pharmacy*, 26(2), 217–222.
- Sailaja, A. K. (2016). Niosomes- A Novel drug carrier for drug targeting. *Mintage Journal of Pharmaceutical & Medical Sciences*, 5(1), 8–15.

- Sammour, R., Taher, M., Chatterjee, B., Shahiwala, A., & Mahmood, S. (2019). Optimization of Aceclofenac Proniosomes by Using Different Carriers, Part 1: Development and Characterization. *Pharmaceutics*, 11(7), 350.
- Sankhyan, A., & Pawar, P. (2012). Recent Trends in Niosome as Vesicular Drug Delivery System. *Journal of Applied Pharmaceutical Science*, 2(6), 20–32.
- Saraswathi, T. S., Mothilal, M., & Jaganathan, M. K. (2019). Niosomes as an emerging formulation tool for drug delivery-a review. *International Journal of Applied Pharmaceutics*, 11(2), 7–15.
- Seleci, D. A., Seleci, M., Walter, J., Stahl, F., & Scheper, T. (2016). Niosomes as Nanoparticulate Drug Carriers : Fundamentals and Recent Applications. *Journal of Nanomaterials*, 3, 1–13.
- Shen, M., Liu, T., Yu, T., Kv, R., Chiang, W., Tsai, Y., Chen, H., Lin, S., Chiu, H. (2019). Biomaterials Hierarchically targetable polysaccharide-coated solid lipid nanoparticles as an oral chemo / thermotherapy delivery system for local treatment of colon cancer. *Biomaterials*, 197(January), 86–100.
- Skřičková, J., Kadlec, B., Venclíček, O., & Merta, Z. (2018). Lung cancer. *Casopis Lékařů Českých*, 157(5), 226–236.
- Sohail, M. F., Rehman, M., Sarwar, H. S., Naveed, S., Salman, O., Bukhari, N. I., Hussain, I., Webster, T., Shahnaz, G. (2018). Advancements in the oral delivery of docetaxel: Challenges, current state-of-the-art and future trends. *International Journal of Nanomedicine*, 13, 3145–3161.
- Srivastava, A., Amreddy, N., Razaq, M., Towner, R., Zhao, Y. D., Ahmed, R. A., Munshi Ramesh, R. (2018). Exosomes as Theranostics for Lung Cancer. *Advances in Cancer Research*, 139, 1–33.
- Sudheer, P., & Kaushik, K. (2015). Review on Niosomes - A Novel Approach for Drug Targeting. *Journal of Pharmaceutical Research*, 14(1), 20–25.
- Tamam, H., Park, J., Gadalla, H. H., Masters, A. R., Abdel-Aleem, J. A., Abdelrahman, S. I., Lyle, L. T., Yeo, Y. (2019). Development of Liposomal Gemcitabine with High Drug Loading Capacity. *Molecular Pharmaceutics*.
- Torres-pérez, S. A., Ramos-godínez, P., & Ramón-gallegos, E. (2020). Glycosylated one-step PAMAM dendrimers loaded with methotrexate for target therapy in breast cancer cells MDA-MB-231. *Journal of Drug Delivery Science and Technology*, 58(April), 101769.
- Vrignaud, S., Benoit, J. P., & Saulnier, P. (2011). Strategies for the nanoencapsulation of hydrophilic molecules in polymer-based nanoparticles. *Biomaterials*, 32(33), 8593–8604.
- Wagner, M. E., & Rizvi, S. S. H. (2015). Novel method of niosome generation using supercritical carbon dioxide part I : process mechanics. *Journal of*

*Liposome Research*, 25(4), 334–346.

- Wahgiman, N. A., Salim, N., Rahman, M. B. A., & Ashari, S. E. (2019). Optimization of nanoemulsion containing gemcitabine and evaluation of its cytotoxicity towards human fetal lung fibroblast (MRC5) and human lung carcinoma (A549) cells. *International Journal of Nanomedicine*, 14, 7323–7338.
- Wen, A., Choi, M., & Kim, D. (2006). Formulation of Liposome for Topical Delivery of Arbutin. *Arch Pharm Res*, 29(12), 1187–1192.
- Wen, J., Gailani, M. Al, Yin, N., & Rashidinejad, A. (2018). Liposomes and Niosomes. In *Liposomes and Niosomes. Emulsion-Based Systems for Delivery of Food Active Compounds*, 263–292.
- Wu, W., Luo, L., Wang, Y., Wu, Q., Dai, H., Li, J., Durkan, C., Wang, N. (2018). Endogenous pH-responsive nanoparticles with programmable size changes for targeted tumor therapy and imaging applications. *Theranostics*, 8(11), 3038–3058.
- Xie, Y., Aillon, K. L., Cai, S., Christian, J. M., Davies, N. M., Berkland, C. J., & Forrest, M. L. (2010). Pulmonary delivery of cisplatin-hyaluronan conjugates via endotracheal instillation for the treatment of lung cancer. *International Journal of Pharmaceutics*, 392(1–2), 156–163.
- Xu, Y. Q., Chen, W. R., Tsosie, J. K., Xie, X., Li, P., Wan, J. B., He, C., Chen, M. W. (2016). Niosome encapsulation of curcumin: Characterization and cytotoxic effect on ovarian cancer cells. *Journal of Nanomaterials*, 2016, 1–9.
- Yalcin, T. E., Ilbasimis-Tamer, S., Ibisoglu, B., Özdemir, A., Ark, M., & Takka, S. (2017). Gemcitabine hydrochloride-loaded liposomes and nanoparticles: comparison of encapsulation efficiency, drug release, particle size, and cytotoxicity. *Pharmaceutical Development and Technology*, 23(1), 76–86.
- Zappa, C., & Mousa, S. A. (2016). Non-small cell lung cancer: current treatment and future advances. *Translational Lung Cancer Research*, 5(3), 288–300.
- Zhang, Y., Chen, L., Hu, G., Zhang, N., Zhu, X., Yang, K., Jin, F., Shi, M., Chen, Y., Hu, W., Cheng, Z., Wang, S., Tian, Y., Wang, X., Sun, Y., Li, J., Li, W., Tang, Y., Mao, L., Zhou, Y., Sun, G., Liu, R., Guo, R., Long, Liang, G., S., Li, L., Huang, J., Long, J., Zang, J., Liu, Q., L., Su, Q., Zheng, B., Xiao, Y., Guo, Y., Han, F., Mo, H., Lv, J., Du, X., Xu, C., Liu, N., Li, Y., Chua, M., Xie, F., Sun, Y., Ma, J. (2019). Gemcitabine and Cisplatin Induction Chemotherapy in Nasopharyngeal Carcinoma. *The New England Journal of Medicine*, 1124–1135.

## BIODATA OF STUDENT



Norfatin Izzatie binti Mohamad Saimi was born in Pasir Mas, Kelantan on 11<sup>st</sup> May 1992. She received her primary in Sekolah Kebangsaan Temin and secondary education in MRSM Kuala Lipis and MRSM Tun Ghafar Baba. She completed her Matriculation of Science Programme at Matriculation College Pahang in 2011 and was offered to pursue her degree at Universiti Putra Malaysia (UPM) and obtained her Bachelor of Science (Honours) degree in Chemistry in 2016. Then, she pursued her Master's Degree in Nanomedicine under supervision of Dr. Norazlinaliza Salim and co-supervisor of Prof Dr Basyaruddin Abdul Rahman and Dr. Noraini Ahmad at Department of Chemistry, Faculty of Science, Universiti Putra Malaysia. She was offered Graduated Research Foundation under UPM, NanoMITe Research Grant (Vot. No 5526306) and UMRG (RG395-17AFR).

## LIST OF PUBLICATIONS

Mohamad Saimi, N. I., Salim, N., Ahmad, N. Abdul Malek, E., Rahman, A. M. B. (2021). Aerosolized Niosome Formulation Containing Gemcitabine and Cisplatin for Lung Cancer Treatment: Optimization, Characterization and In Vitro Evaluation. *Pharmaceutics*, 13 (59), 1-19.

### List of Conferences/Congress

#### National Conference (2015-2019)

1. Norfatin Izzatie Mohamad Saimi, Norazlinaliza Salim, Mohd Basyaruddin Abd Rahman, and Noraini Ahmad, 2019, Development of Aerosolized Niosome Formulation Containing Gemcitabine and Cisplatin for Lung Cancer Treatment. NanoMITe Annual Symposium & Nanotechnology Malaysia Annual Symposium (NanoSym 2019). (Oral presentation)

#### International Conference (2015-2019)

1. Norfatin Izzatie Mohamad Saimi, Norazlinaliza Salim, Mohd Basyaruddin Abd Rahman, and Noraini Ahmad, 2018, Development of Aerosolized Niosome Formulation Containing Gemcitabine and Cisplatin for Lung Cancer Treatment, 10<sup>th</sup> International Fundamental Science Congress (iFSC2018), p126. (Oral presentation)
2. Norfatin Izzatie Mohamad Saimi, Norazlinaliza Salim, Mohd Basyaruddin Abd Rahman, Noraini Ahmad and Siti Efliza Ashari, 2018, Development of Aerosolized Formulation of Niosome Containing Gemcitabine and Cisplatin for Lung Cancer Treatment, 6th International Symposium on Applied Engineering and Sciences (SAES2018). (Poster presentation)
3. Norfatin Izzatie Mohamad Saimi, Norazlinaliza Salim, Mohd Basyaruddin Abd Rahman, and Noraini Ahmad, 2019. Niosome Encapsulation of Gemcitabine-Cisplatin: Characterization, Stability and Cytotoxicity Effect on Lung Cancer Cell, 7th International Symposium on Applied Engineering and Sciences (SAES2019). (Oral presentation)

### Awards

1. Norfatin Izzatie Mohamad Saimi, Norazlinaliza Salim, Mohd Basyaruddin Abd Rahman, Noraini Ahmad and Siti Efliza Ashari, 2018, Development of Aerosolized Formulation of Niosome Containing Gemcitabine and Cisplatin for Lung Cancer Treatment, 6th International Symposium on Applied Engineering and Sciences (SAES2018). (Best Poster Award)





UNIVERSITI PUTRA MALAYSIA

**STATUS CONFIRMATION FOR THESIS / PROJECT REPORT  
AND COPYRIGHT**

**ACADEMIC SESSION : SECOND SEMESTER 2020/2021**

**TITLE OF THESIS / PROJECT REPORT :**

**Development of Aerosolized Niosome Formulation Containing Gemcitabine and Cisplatin for *In-Vitro* Cytotoxicity against MRC5 and A549 Cell Lines**

**NAME OF STUDENT :**

**Norfatin Izzatie binti Mohamad Suhaimi**

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

1. This thesis/project report is the property of Universiti Putra Malaysia.
2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as:

\*Please tick (√)

**CONFIDENTIAL**

(Contain confidential information under Official Secret Act 1972).

**RESTRICTED**

(Contains restricted information as specified by the organization/institution where research was done).

**OPEN ACCESS**

I agree that my thesis/project report to be published as hard copy or online open access.

This thesis is submitted for:



**PATENT**

Embargo from \_\_\_\_\_ until  
\_\_\_\_\_ (date)  
(date)

**Approved by:**

\_\_\_\_\_  
(Signature of Student)  
New IC No/ Passport No.:

Date :

\_\_\_\_\_  
(Signature of Chairman  
of Supervisory Committee)  
Name:

Date :

**[Note : If the thesis is CONFIDENTIAL or RESTRICTED, please attach with the letter from the organization/institution with period and reasons for confidentiality or restricted.]**

