



UNIVERSITI PUTRA MALAYSIA

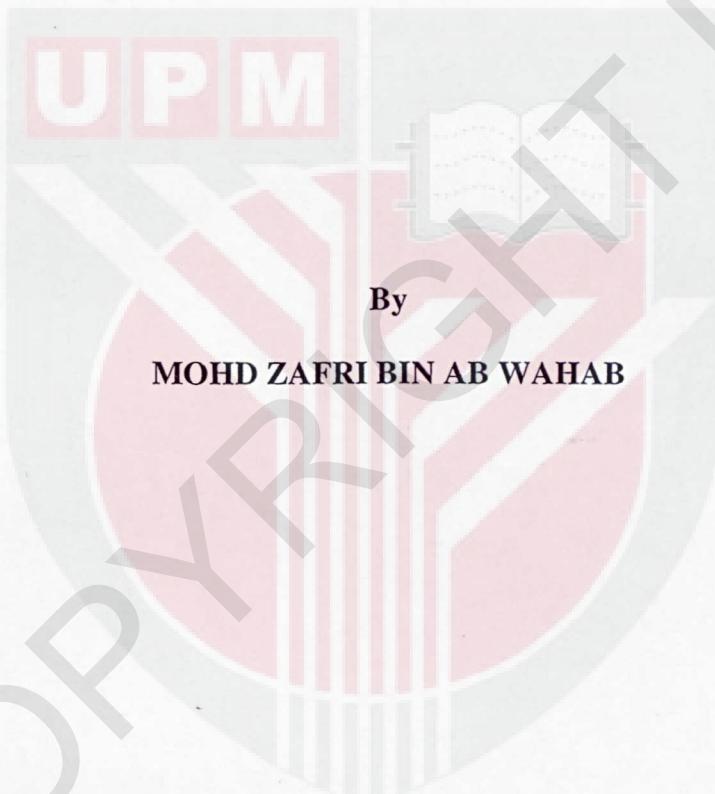
**CHARACTERIZATION OF *Pseudomonas fuscovaginae* CAUSAL  
AGENT SHEATH BROWN ROT OF RICE IN PENINSULAR MALAYSIA**

MOHD ZAFRI AB WAHAB

FP 2014 70



**CHARACTERIZATION OF *Pseudomonas fuscovaginae* CAUSAL AGENT  
SHEATH BROWN ROT OF RICE IN PENINSULAR MALAYSIA**



**Thesis Submitted to the School of Graduate Studies, University Putra Malaysia,  
in Fulfilment of the Requirements for the Degree of Master of Science**

**November 2014**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



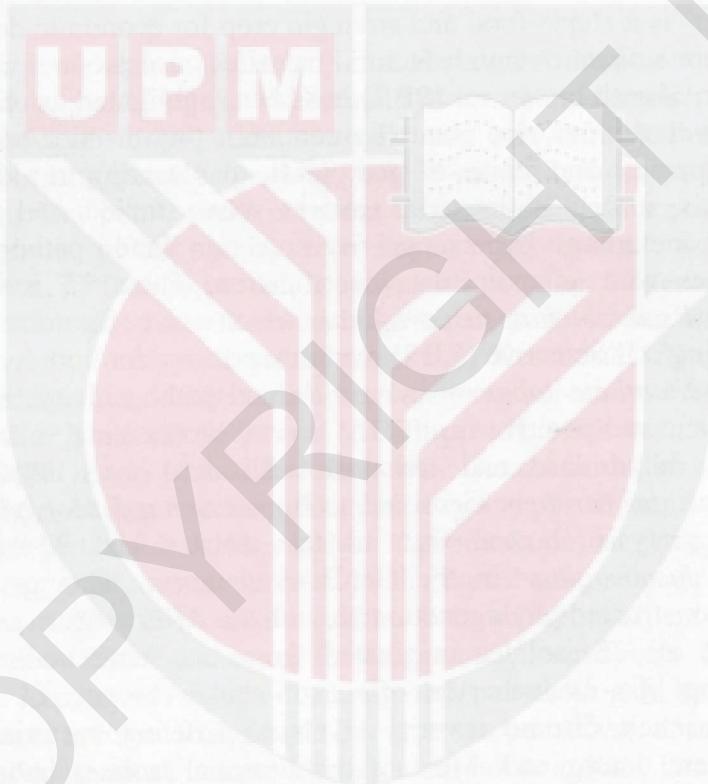
fk

7/7/17

**1000766769**

t  
FP  
2014  
70

*Do whatever you can do, nothing can stop you. To be able to go ahead in life, you have to overcome the struggle and strive. Keep trying till you succeed, such determination is all that you need". – Nor Farah Shamira (Beloved wife and friend)*



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment  
of the requirement for the degree of Master of Science

**CHARACTERIZATION OF *Pseudomonas fuscovaginae* CAUSAL AGENT  
SHEATH BROWN ROT OF RICE IN PENINSULAR MALAYSIA**

By

**MOHD ZAFRI BIN AB WAHAB**

**NOVEMBER 2014**

**Chairman: Associate Professor Dr. Kamaruzaman Bin Sijam**

**Faculty: Agriculture**

Rice (*Oryza sativa*) is a staple food and strategic crop for economic development in Malaysia. There are several downfall factors that affect rice productivity, which are pests and diseases. Sheath brown rot (SBR) has been highlighted as one of the most important diseases that affect rice plants by contaminating the rice seeds. It causes rot to the sheath, panicle and leaves of rice plants, thus leading to a decline in rice production. In this study, 25 bacteria isolates were studied and characterized morphologically, including hypersensitivity reaction and pathogenicity test, biochemical test as well as molecular identification. These 25 isolates had the characteristics of *Pseudomonas fuscovaginae* which were fluoresced under UV illumination on King's B Selective (KBS) medium, positive for both hypersensitivity reaction (HR) on *Nicotiana tabacum* cv. *xanthi* and pathogenicity tests on *Oryza sativa*. They also showed positive results for several biochemical reactions such as oxidase, arginine dihydrolase and trehalose utilization tests. Based on these preliminary tests, all isolates were identified as *Pseudomonas fuscovaginae*. The 16S rDNA sequence analysis showed that all the isolates had 97-99% sequence similarities to *P. fuscovaginae* in the GenBank database. This result was also supported by the constructed phylogenetic tree with the *Acidivorax avenae* outgroup. It was noted that all 25 isolates originated from the same node with the *P. fuscovaginae* group. To evaluate the effect of sheath brown rot pathogen on Malaysian rice varieties, disease severity levels of different bacterial isolates on MR219 variety were determined. Most of the bacterial isolates had virulent and intermediate severity level. Virulent isolate, T1 from Selangor, Malaysia had highest percentage of diseased sheath area. As the effects of sheath brown rot pathogen on Malaysian rice varieties were evaluated, most of the varieties recovered from the disease as they matured. Moreover, results also showed that the panicle's number and grain's weight of infected rice plants were lower than control rice plants but not significant. These findings conclude that under greenhouse conditions, the pathogen of sheath brown rot could not affect Malaysian rice varieties. The information on characteristics of *P. fuscovaginae* in Peninsular Malaysia and its pathogenicity on Malaysian rice varieties could hopefully be used in future studies especially on Malaysian *P. fuscovaginae* biology and their control in Malaysia.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai syarat untuk memenuhi keperluan Ijazah Sarjana Sains

**PENCIRIAN *Pseudomonas fuscovaginae* AGEN PENYEBAB PENYAKIT REPUT SELUDANG PADI DI SEMENANJUNG MALAYSIA**

Oleh

**MOHD ZAFRI BIN AB WAHAB**

**NOVEMBER 2014**

**Pengerusi: Profesor Madya Dr. Kamaruzaman Bin Sijam**

**Fakulti: Pertanian**

Padi (*Oryza sativa*) adalah makanan ruji dan tanaman strategik untuk pembangunan ekonomi di Malaysia. Terdapat beberapa faktor yang memberi kesan kepada kejatuhan produktiviti beras, iaitu perosak dan penyakit. Reput seludang padi merupakan salah satu penyakit penting yang memberi kesan kepada tanaman padi. Selain mencemarkan benih padi, penyakit ini menyebabkan reput pada seludang, tangkai dan daun pokok padi, seterusnya menyebabkan penurunan dalam pengeluaran beras. Dalam kajian ini, sebanyak 25 pencilan bakteria telah dikaji dan diciri secara morfologi, termasuk tindak balas hipersensitiviti dan ujian kepatogenan, ujian biokimia serta kenal pasti secara molekular. Keseluruhananya, 25 pencilan ini mempunyai ciri-ciri *Pseudomonas fuscovaginae* yang bercahaya bawah pencahayaan UV di atas media ‘King’s B Selective’ (KBS), positif untuk kedua-dua tindak balas hipersensitiviti (HR) pada *Nicotiana tabacum* cv. *xanthi* dan ujian kepatogenan pada *Oryza sativa*. Mereka juga menunjukkan hasil yang positif untuk beberapa tindak balas biokimia seperti oxidase, dihydrolase arginina dan ujian penggunaan trehalosa. Berdasarkan ujian awal, semua pencilan telah dikenal pasti sebagai *P. fuscovaginae*. Analisis jujukan 16S rDNA menunjukkan bahawa semua pencilan mempunyai persamaan jujukan 97-99% dengan *Pseudomonas fuscovaginae* dalam pangkalan data GenBank. Keputusan ini juga disokong oleh pohon filogeni yang dijana dengan *Acidivorax avenae* sebagai ‘outgroup’. Berdasarkan pemerhatian, semua 25 pencilan berasal dari nod yang sama dengan kumpulan *P. fuscovaginae*. Untuk menilai kesan patogen reput seludang pada varieti padi Malaysia, tahap kepatogenan penyakit oleh pencilan bakteria yang berbeza pada varieti MR219 telah ditentukan. Kebanyakan pencilan bakteria mempunyai kepatogenan ditahap virulen dan virulen intermidiat. Pencilan yang virulen, T1 dari Selangor, Malaysia mempunyai peratusan tertinggi dalam kawasan seludang berpenyakit. Apabila kesan patogen reput seludang pada varieti padi Malaysia telah dinilai, kebanyakan varieti pulih daripada penyakit ini setelah mereka matang. Selain itu, keputusan juga menunjukkan bilangan ‘panicle’ dan berat bijirin tanaman padi dijangkiti adalah lebih rendah daripada tanaman padi kawalan tetapi tidak ketara. Kesimpulannya, di dalam keadaan rumah hijau, patogen reput perang seludang tidak dapat menjaskan varieti padi Malaysia dengan ketara. Maklumat ciri-ciri *P. fuscovaginae* di Semenanjung Malaysia dan kepatogenannya pada varieti padi Malaysia diharap dapat digunakan di masa hadapan terutamanya dalam kajian tentang biologi *P. fuscovaginae* di Malaysia dan kawalannya.

## **ACKNOWLEDGEMENT**

Alhamdulillah, I want to thanks Almighty Allah for reasons too numerous to mention.

For project financial, I would like to thank to LRGS (Long Research Grant Scheme from Ministry of Education Malaysia and Young Lecturer Fellowship from UiTM scheme for financial assistance.

Several people played an important role in the accomplishing of this thesis, and I would like to acknowledge them here:

I would like to thank Assoc. Prof. Dr Kamaruzaman Sijam (UPM), Prof Razi Ismail (UPM) and Dr Marzukhi Hashim (MARDI) , for assistance and encouragement to pursue to this thesis until completed.

Special thanks also Mrs Erneeza, Mrs. Junaina, Mr. Zawawi, Mr. Yusof, Mr. Johari, all Plant Protection Department staff, and all my friends for the assistance.

To my late wife, Nor Farah Shamira Bt Hamzani, thank you for the kind and infinite love, until we meet again in heaven, InsyaAllah.

To my family, thank you for the generous support in financial, moral aspect as well as for the advice and assistance.

MOHD ZAFRI BIN AB WAHAB

## TABLE OF CONTENTS

	Page
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	ii
<b>ACKNOWLEDGEMENTS</b>	iii
<b>APPROVAL</b>	iv
<b>DECLARATION</b>	vi
<b>LIST OF TABLES</b>	x
<b>LIST OF FIGURES</b>	xi
<b>LIST OF ABBREVIATIONS</b>	xiii
 <b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	<b>1</b>
<b>2 LITERATURE REVIEW</b>	
2.1 Issues of rice production	3
2.2 Rice production in Malaysia	3
2.3 Rice pests and diseases	6
2.4 Fungal and bacterial diseases of rice	6
2.5 Sheath brown rot of rice caused by <i>Pseudomonas fuscovaginae</i>	7
2.5.1 Disease symptoms	9
2.5.2 Taxonomy and morphology of <i>P. fuscovaginae</i>	11
2.5.3 Hypersensitivity reaction, pathogenicity and biochemical characteristics	15
2.5.4 DNA based study on <i>P. fuscovaginae</i>	17
2.6 Sheath brown rot of rice in Malaysia	22
<b>3 MATERIALS AND METHODS</b>	
3.1 Sample collection	23
3.2 Isolation and morphological characterization	26
3.3 Hypersensitivity reaction and pathogenicity tests	26
3.4. Biochemical tests	26
3.5 Molecular identification	27
3.6 Evaluation of disease severity level on different isolates of <i>Pseudomonas fuscovaginae</i>	29
3.6.1 Disease assessment determination	29
3.7 Assessment of effects nine Malaysian rice varieties infected with most virulent <i>Pseudomonas fuscovaginae</i>	31
3.7.1 Host plant preparation	31
3.7.2 Isolate preparation and inoculation	33
3.7.3 Disease severity evaluation	35
3.7.4 Data analysis and experimental design	37

<b>4</b>	<b>RESULTS AND DISCUSSION</b>	
4.1	Results	
4.1.1	Sample collection	39
4.1.2	Isolation and morphological characteristics	39
4.1.3	Hypersensitivity and pathogenicity tests	44
4.1.4	Biochemical tests	47
4.1.5	Molecular Identification	49
4.1.6	Assessment of disease severity level on different isolates of <i>Pseudomonas fuscovaginae</i> isolates	56
4.1.7	Assessment of effects nine Malaysian rice varieties infected with <i>Pseudomonas fuscovaginae</i>	59
4.2	Discussion	
4.2.1	Sample collection	63
4.2.2	Morphological characteristics	63
4.2.3	Hypersensitivity and pathogenicity tests	64
4.2.4	Biochemical tests	64
4.2.5	Molecular Identification	64
4.2.6	Assessment of disease severity level on different <i>Pseudomonas fuscovaginae</i> isolates	65
4.2.7	Assessment of effects on nine Malaysian rice varieties infected with <i>Pseudomonas fuscovaginae</i>	66
<b>5</b>	<b>SUMMARY , CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH</b>	69
<b>REFERENCES/ BIBLIOGRAPHY</b>		71
<b>APPENDICES</b>		83
<b>BIODATA OF STUDENT</b>		102
<b>LIST OF PUBLICATIONS</b>		103

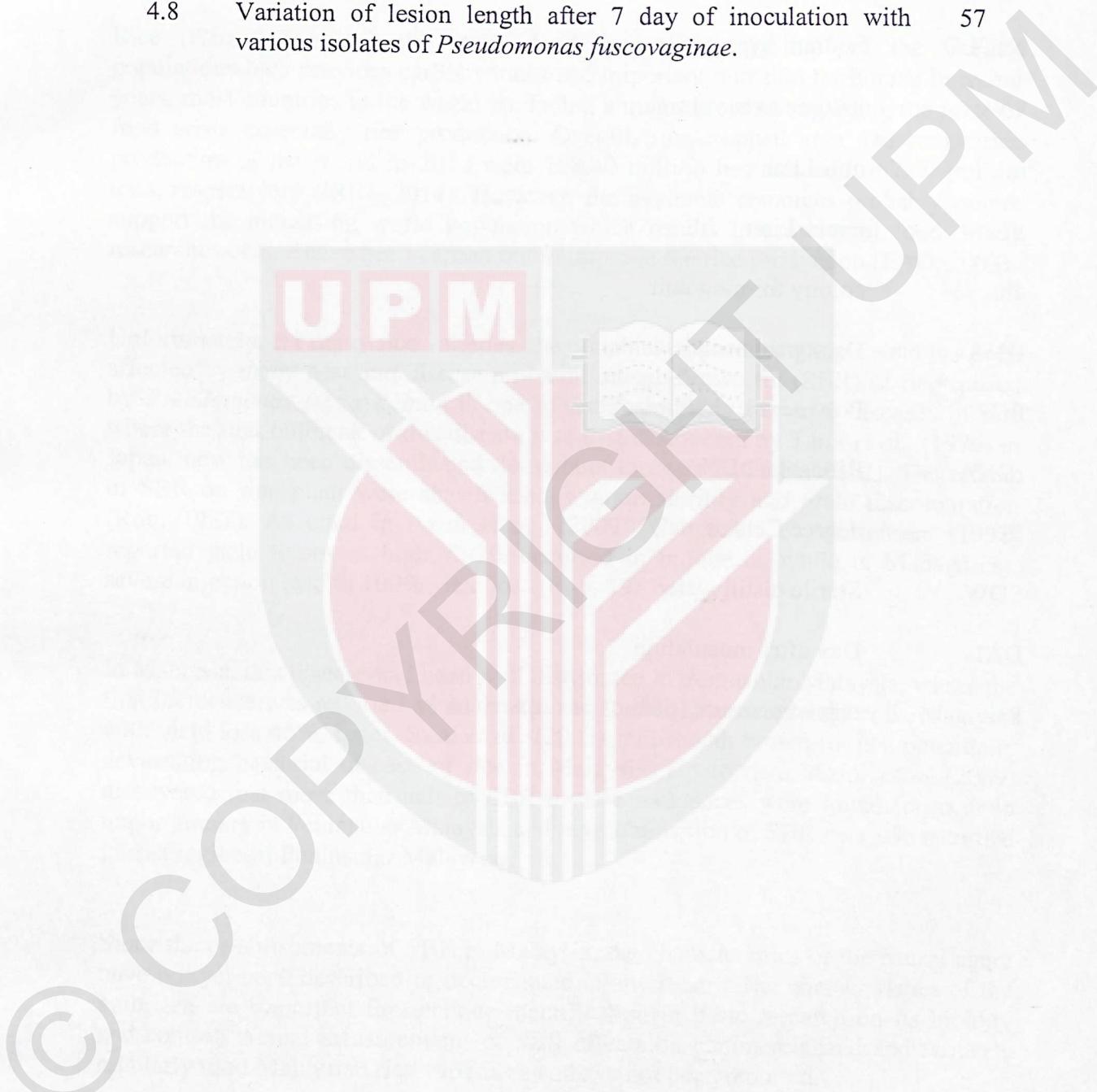
## LIST OF TABLES

<b>Table</b>		<b>Page</b>
2.1	List of rice varieties that have been developed by MARDI	5
2.2	Worldwide reports of sheath brown rot disease of rice	8
2.3	Colony characteristics of <i>Pseudomonas fuscovaginae</i> on Miyajima medium	14
2.4	The characteristics of <i>Pseudomonas fuscovaginae</i>	16
3.1	Source of sheath brown rot disease samples from different location in Peninsular Malaysia	25
3.2	<i>Pseudomonas fuscovaginae</i> from Genbank database used for DNA sequences comparison	28
3.3	Score to determine disease level of <i>Pseudomonas fuscovaginae</i>	30
3.4	Justification selection of varieties in the study	32
3.5	General scale for stress of Standard Evaluation System for Rice	36
3.6	Disease severity scale based on grains discoloration	36
4.1	Bacterial isolates isolated from various rice varieties from different locations in Peninsular Malaysia	41
4.2	Results of Hypersensitivity test and pathogenicity test on 50 bacterial isolates from Peninsular Malaysia	46
4.3	Biochemical tests of bacterial isolates suspected to be <i>Pseudomonas fuscovaginae</i>	48
4.4	Local isolates and accession number.	52
4.5	Results of BLAST search based on 16S rRNA partial sequence gene	54
4.6	Results of disease severity level on different <i>Pseudomonas fuscovaginae</i> isolates	58
4.7	Comparison of disease severity value and reaction of different rice variety from Malaysia infected with <i>Pseudomonas fuscovaginae</i> T1 isolates	60
4.8	Assessment of grains's weight and panicle's number of rice plant infected with <i>Pseudomonas fuscovaginae</i> T1 isolates	62

## LIST OF FIGURES

<b>Figure</b>		<b>Page</b>
2.1	Trend of total Malaysia rice production and import for year 2006-2010 .	4
2.2	Mature rice plant with brown leaf sheath symptoms .	10
2.3	<i>Pseudomonas fuscovaginae</i> showing fluorescent characteristic under UV illumination on selective medium	12
2.4	<i>Pseudomonas fuscovaginae</i> showing the colony morphological characteristics on Miyajima medium.	14
2.5	Genetic diversity among 19 <i>Pseudomonas fuscovaginae</i> isolates was revealed by PCR analysis.	18
2.6	Phylogenetic tree based on neighbor-joining of partial 16S rRNA gene sequences of the rice pseudomonad with related pseudomonad sequences.	19
2.7	Consensus neighbor-joining tree based on sequencing of the 16s RNA gene of <i>Pseudomonas</i> spp.	20
2.8	Phylogenetic tree showing the relationship between the fluorescent pseudomonads isolated from disease rice and the reference strains of the rRNA group 1 pseudomonads.	21
3.1	Map of Peninsular Malaysia showing the locations of rice granary where the isolates were obtained.	24
3.2	Inoculation process of the pathogen in greenhouse experiment.	34
3.3	Plan layout of greenhouse experiment.	38
4.1	Rice plants showing necrosis of sheath and grain discoloration symptoms.	40
4.2	Colony morphology of suspected <i>Pseudomonas fuscovaginae</i> on KBS agar	43
4.3	<i>Nicotiana tabacum</i> cv. <i>xanthi</i> showing the hypersensitivity reaction after 24 hour inoculation	45
4.4	<i>Oryza sativa</i> (MR219 variety) showing SBR brown rot symptom after 3 day inoculation	45
4.5	Genomic DNA fragments of bacterial isolates on 1% (w/v) agarose gel	50

4.6	PCR amplification of total genomic DNA from 25 bacterial isolates of <i>Pseudomonas fuscovaginae</i> based on 16s rRNA region	51
4.7	Phylogenetic tree constructed using a Neighbour joining based on 16s rRNA gene sequence.	55
4.8	Variation of lesion length after 7 day of inoculation with various isolates of <i>Pseudomonas fuscovaginae</i> .	57



## LIST OF ABBREVIATIONS

SBR	Sheath brown rot
KMB	King's B Medium
KBS	King's B selective medium
NaCLO	Sodium hypochlorite
v/v	volume over volume
mL	milli Liter
$\mu$ L	micro Liter
cfu	colony forming unit
DNA	Deoxyribonucleic acid
PCR	Polymerase chain reaction
rDNA	Ribosomal DNA
°C	degree Celsius
SDW	Sterile distill water
DAI	Day after inoculation
Psi	unit of pressure ( <i>pounds per square inch</i> )

## CHAPTER 1

### INTRODUCTION

Rice (*Oriza sativa*) is the major food crop of nearly half of the world's population which provides carbohydrates and important nutrition for human. In recent years, most countries in the world are facing a major problem regarding the issue of food crisis especially rice production. Overall, rice-cropped area and rough rice production of the world in 2012 were 158.40 million hectares and 467.60 million tons, respectively (IRRI, 2014). However, the available resources probably cannot support the increasing world population which results in scarcity of food. Many researches of rice have been carried out to improve the rice production (FAO, 2003).

Unfortunately, for many rice varieties, the potential of producing high yield has been affected by many pest and disease problem. Sheath brown rot (SBR) of rice caused by *Pseudomonas fuscovaginae* is one of the devastating bacterial diseases of rice where the first outbreak of this disease was first discovered by Tanii *et al.*, (1976) in Japan, now has been disseminated throughout the world (CABI, 2012). The effects of SBR on rice plant were sheath lesions, grain sterility and grain discolouration (Rott, 1987). As cited in Azmi *et al.*, (2009), Cahyaniati & Mortensen, (1997) reported yield losses as high 72.2% occurred in Indonesia, while in Madagascar, severe infection lead to 100% yield loss (Rott, 1987).

In Malaysia, this disease had been well distributed in Peninsular Malaysia, where the first incidence was reported by Marzukhi *et al.*, (1991) in Seberang Perak, Malaysia with yield loss 46%. Later, Saad *et al.*, (2003) said sheath brown rot is a potentially devastating bacterial disease of rice in Malaysia. Up to date, Azmi *et al.*, (2009) discovered that more than half of SBR disease incidences were found to occur in major granary in Peninsular Malaysia and severe infection of SBR was also recorded in east region of Peninsular Malaysia.

Since the establishments of SBR in Malaysia, the characteristics of the causal agent have not yet been described or documented in any report. The characteristics of the pathogen are important for accurate identification in basic research on its biology and control. Actual measurements of SBR effects on commercialized and farmer's regularly used Malaysian rice varieties also have not been reported.

This study was encouraged by the report of *P. fuscovaginae* in Malaysia by Azmi *et al.*, (2009), Saad *et al.*, (2003) and Marzukhi *et al.*, (1991) which is based on the fact that the characteristics and pathogenicity of the Malaysian *P. fuscovaginae* had not previously been investigated. Therefore, this study was undertaken with following objectives:

1. To characterize the causal agent of sheath brown rot of rice in Peninsular Malaysia.
2. To evaluate the effect on Malaysian rice varieties after being inoculated with sheath brown rot pathogen.



## REFERENCES

- Adorada, D., L. (2013). Pathogenicity, diversity, biology and sources of resistance to *Pseudomonas fuscovaginae* in rice. PhD Thesis, Charles Sturt University
- Adorada, D. L., Stodart, B.J., Vera Cruz, C. and Ash, G.J. (2013). Analysis of virulence and molecular diversity of Australian strains of *Pseudomonas fuscovaginae* [In: "Pathogenicity, diversity, biology and sources of resistance to *Pseudomonas fuscovaginae* in rice" (Adorada, D., L.) Page 79-99]. PhD Thesis, Charles Sturt University
- Adorada, D., Stodart, B. J., Cruz, C. V., Gregorio, G., Pangga, I., and Ash, G. J. (2012). Standardizing resistance screening to *Pseudomonas fuscovaginae* and evaluation of rice germplasm at seedling and adult plant growth stages. *Euphytica*, 192(1), 1-16.
- Adorada, D., L. (2010). Comparison of inoculation techniques for the sheath brown rot disease in rice cause by *Pseudomonas fuscovaginae*. 3rd International Rice Congress, Hanoi , Vietnam.
- Afolabi, O., Milan, B., Poulin, L., Ongom, J., Szurek, B., Koebnik, R., and Silue, D. (2014). First report of *Xanthomonas oryzaepv. oryzicola* Causing Bacterial Leaf Streak of Rice in Uganda. *Plant Disease*, 98(11), 1579-1579
- Agrios, G. N. (1997) Plant Pathology, 4<sup>th</sup> edn. Academic Press, San Diego. pp 115-142.
- Alexopoulos C J and Mims C W (1979) Introductory mycology. 3d ed. John Wiley and Sons, New York. pp 632
- Altschul, S.F., Madden, T.L., Schäffer, A.A., Zhang, J., Zhang, Z., Miller, W., Lipman, D.J. (1997) Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Research*, 25, 3389–3402.
- Anon. (2002). Standard Evaluation System for Rice (SES). International Rice Research Institute (IRRI). Los Banos, Philippines, 56 pp.
- Arsenijevic, M. (1991). Bacterial sheath brown rot of rice, wheat, maize and sorghum plants. *Savremena Poljoprivreda*, 39, 66-71
- Azegami, K. (2001). *Burkholderia* spp. associated with rice. In 'Seed health and seed-associated microorganisms for rice disease management'. (Eds TW Mew, B Cottyn). Limited Proceedings No. 6. (IRRI: Los Banos, The Philippines)
- Azmi, A. R., Nur Ain Izzati , M. Z., Siti Nordahliawate, M. S., Nor Azliza, I., Nik Mohd Izham, N. M., and Baharuddin, S. (2009). Sheath Brown Rot Disease of Rice Caused by *Pseudomonas Fuscovaginae* in the Peninsular Malaysia. *Journal of Plant Protection Research*, 49(3), 244-249.

- Ballio, A., Bossa, F., Camoni, L., Di Giorgio, D., Flamand, M. C., Maraite, H., . . and Scaloni, A. (1996). Structure of fuscopeptins, phytotoxic metabolites of *Pseudomonas fuscovaginae*. *FEBS Letters*, 381(3), 213-216.
- Batoko, H., Bouharmont, J., Kinet, J.-M., and Maraite, H. (1997a). Inhibition of Rice (*Oryza sativa L.*) Internode Elongation by *Pseudomonas fuscovaginae* Toxins. In K. Rudolph, T. J. Burr, J. W. Mansfield, D. Stead, A. Vivian & J. Kietzell (Eds.), *Pseudomonas Syringae Pathovars and Related Pathogens*, 9, 227-229
- Batoko, H., Flamand, M.-C., Boutry, M., Kinet, J.-M., and Maraite, H. (1997b). Biological Effects of *Pseudomonas fuscovaginae* Toxins on Rice Cells. In K. Rudolph, T. J. Burr, J. W. Mansfield, D. Stead, A. Vivian & J. Kietzell (Eds.), *Pseudomonas Syringae Pathovars and Related Pathogens*, 9, 215-220
- Batoko, H., Bouharmont, J., Kinet, J. M., and Maraite, H. (1997c). Involvement of Toxins Produced by *Pseudomonas fuscovaginae* in Aetiology of Rice Bacterial Sheath Brown Rot. *J. Phytopathology*, 145, 525-531.
- Batoko, H., Bouharmont, J., Maraite, H. (1997d). Biological effects of toxins produced by *Pseudomonas fuscovaginae* on rice and in vitro selection for resistance. In: Poisson C, Rakotoarisoa J (eds) Conference rice for highlands, Centre Cooperation Int Rech Agronomique Development, 75116 Paris, pp 113-118
- Batoko, H., Bouharmont, J., & Maraite, H. (1994). Inhibition of rice (*Oryza sativa L.*) seedling elongation by a *Pseudomonas fuscovaginae* toxin. *Euphytica*, 76(1), 139-143.
- Batoko, H, Kinet JM, Maraite H and Bouharmont. (1991) Susceptibility to brown sheath rot of rice correlated to sensitivity to bacterial toxin. *Parasitica* 47, 151-163
- Benson, DA, Karsch-Mizrachi I, Lipman DJ, Ostell J, and Wheeler DL (2008) GenBank. *Nucleic Acids Research*, 36, 25-30.
- Blair, J.E., Lennete, E. H., and Truant, J. P. (1970). Manual of clinical Microbiology, American Society of Microbiology, Bethesta, MD. 727pp
- CABI (2012). Crop protection compendium. Vol. 2012. CAB International. Retrieved from :  
<http://www.cabi.org/dmpd/FullTextPDF/2005/20056500742.pdf> on 27 November 2014
- Cahyaniati, A. and Mortensen, C.N. (1997). Bacterial Sheath Brown Rot of Rice (*Pseudomonas fuscovaginae*) Grown in Indonesia. Seed Health Testing in the Production of Quality Seed, 195 pp.

- Chun, J., Lee, J.H. and Jung, Y. (2007). EzTaxon: a web-based tool for the identification of prokaryotes based on 16S ribosomal RNA gene sequences. International Jol.of Systematic & Evolutionary Microbiology 57, 2259–61.
- Cortesi, P., Bartoli, F., Pizzatti, C., Bertocchi, D. and Schaad.N.W. (2008). Panicle sterility and grain discolouration: new and emerging bacterial diseases of rice in Italy. In: Fatmi, M., Collmer, A., Iacobellis, N.S., Mansfield, J., Murillo, J., Schaad, N.W., Ullrich, M. (eds) *Pseudomonas syringae* pathovars and related pathogens— identification, epidemiology and genomics. Springer, Dordrecht, pp 391–411
- Cother, E. J., Noble, D. H., van de Ven, R. J., Lanoiselet, V., Ashd, G., Vuthyc, N. and Stodartd, B. (2010). Bacterial pathogens of rice in the Kingdom of Cambodia and description of a new pathogen causing a serious sheath rot disease. *Plant Pathology*, 59, 944–953.
- Cother, E.J., Stodart, B. and Ash, G. (2009a). Improving understanding and management of rice pathogens in Cambodia.Project Final Report. Australian Center for International Agricultural Research, ACIAR, Canberra ACT 2601 Australia, pp 1–46
- Cother, E.J., Stodart, B., Noble, D. H., Reinke, R., and Van de Ven, R. J. (2009b).Polyphasic identification of *Pseudomonas fuscovaginae* causing sheath and glume lesions on rice in Australia.*Australasian Plant Pathology*, 38, 247–261.
- Cother, .EJ, Reinke R, McKenzie C, Lanoiselet VM, and Noble DH (2004). An unusual stem necrosis of rice caused by *Pantoea ananas* and the first record of this pathogen on rice in Australia.*Australasian Plant Pathology* ,33, 495–503
- Cottyn, B.J.G (2003). Bacteria associated with rice seed from Philippine farmers' fields, Universiteit Gent (Belgium) and IRRI, Phd Thesis. Pp 24-25
- Cottyn, B, George T and Vera Cruz CM (2002). Characterization of rice sheath rot from Siniolan, Philippines, International Rice Research Newsletter, IRRI, Los Banos, Laguna, pp 39–40
- Cottyn, B., Regalado, E., Mew, T.W. and Swings, J. (2001). Bacterial population associated with rice seed in the tropical environment. *Phytopathology*, 91, 282–92
- Cottyn, B., Cerez, M. T., Van Outryve, M. F., Barroga, J., Swings, J., and Mew, T. W. (1996a).Bacterial disease of rice. I. Pathogenic bacteria associate with sheath rot complex and grain discoloration of rice in Philippines. *Plant Disease.*, 80, 429-437.
- Cottyn, B., Van-Outryve, M. F., Cerez, M. T., De-Cleene, M., Swings, J., & Mew, T. W. (1996b).Bacterial diseases of rice. II. Characterization of pathogenic bacteria associated with sheath rot complex and grain discoloration of rice in the Philippines. *Plant Disease.*, 80, 438-445.

- Cottyn, B., Cerez, M. T., and Mew, T. W. (1994). Bacterial pathogen. [In: "A Manual of Rice Seed Health Testing" (T.W. Mew, J.K. Misra, eds.)]. (International Rice Research Institute) 42, 91-96.
- Detry, J.F, Chapeaux, J.P. and Tilquin, J.P (1991). Estimation of rice bacterial sheath brown rot (BSR) and rice blast (Bl) severity in five Burundi highland swamps. *Int Rice Res News*, 16, 20–21
- Develey-Riviere and M-P, Galina E (2007). Resistance to pathogens and host developmental stage: a multifaceted relationship within the plant kingdom. *New Phytol* 175, 405–416.
- Duveiller, E., Notteghem, J. L., Rott, P., Snacken, F., and Maraite, H. (1990). Bacterial sheath brown rot of rice caused by *Pseudomonas fuscovaginae* in Malagasy. *Tropical Pest Management*, 36(2), 151-153.
- Duveiller, E., Snacken, F., Maraite, H., and Autrique, A. (1989). First detection of *Pseudomonas fuscovaginae* on Maize and Sorghum in Burundi. *Plant Disease*, 73, 514-517
- Duveiller, E., Miyajima, K., Snacken, F., Autrique, A., and Maraite, H. (1988). Characterization of *Pseudomonas fuscovaginae* and Differentiation from Other Fluorescent Pseudomonads Occurring on Rice in Burundi. *Journal of Phytopathology*, 122(2), 97-107.
- Eden, P.A., Schmidt, T.M., Akemore, R.P., Pace, N.R. (1991) Phylogenetic analysis of *Aquaspirillum magnetotacticum* using polymerase chain reaction-amplified 16S rRNA-specific DNA. *International Journal of Systematic Bacteriology*, 41, 324–325.
- ETP, Economic Transformation Programme (2012) Chapter 15: Transitioning from Agriculture to Agribusiness. Retrieved from <http://etp.pemandu.gov.my/>
- Fahy, P.C., and Hayward, A.C. (1983). Media and methods for isolation and diagnostic tests. 337-370 in: Plant Bacterial Disease. A Diagnostic Guide. P.C. Fahy and G.J. Persley eds. Academic Press, Sydney
- FAO, (2003) Sustainable rice production for food security Proceedings of the 20th Session of the International Rice Commission Bangkok, Thailand  
Retrieved from :<http://www.fao.org/DOCREP/006/Y4751E/y4751e00.htm>
- Freeman, B.C. and G.A. Beattie. (2008). An Overview of Plant Defenses against Pathogens and Herbivores. *The Plant Health Instructor*.
- Flamand, M. C., Pelsser, S., Ewbank, E., and Maraite, H. (1996). Production of syringotoxin and other bioactive peptides by *Pseudomonas fuscovaginae*. *Physiological and Molecular Plant Pathology*, 48(4), 217-231.

- Guanlin, X. (2003).First Report of Sheath Brown Rot of Rice in China and Characterization of the Causal Organism by Phenotypic Tests and Biolog.*International Rice Research Notes*, 8(1), 50-52.
- Gardan, L., Bella, P., Meyer, J. M., Christen, R., Rott, P., Achouak, W., and Samson, G. (2002). *Pseudomonas salomonii* sp nov., pathogenic on garlic, and *Pseudomonas palleroniana* sp. nov., isolated from rice. *International Journal of Systematic and Evolutionary Microbiology*, 52, 2065-2074.
- Ham, J. H., Melanson, R. A., & Rush, M. C. (2011).*Burkholderia glumae*: next major pathogen of rice? [Research Support, Non-U.S. Gov't Review]. *Mol Plant Pathol*, 12(4), 329-339
- Höfte, M., & Vos, P. D. (2006). Plant Pathogenic *Pseudomonas* Species. S.S. Gnanamanickam (ed.), Plant-Associated Bacteria.
- Heong, K. L. (1984). Pest control practices of rice farmers in Tanjong Karang, Malaysia. *International Journal of Tropical Insect Science*, 5(3), 221-226
- Hossain, M.T., Hossain, S.M.M., Bakr, M.A., Matiar Rahman, A.K.M. and Uddin N.S (2010).Survey on major diseases of vegetable and fruit crops in Chittagong region.*Bangladesh J. Agril. Res.* 35(3), 423-429
- Hugh, R., and E. Leifson. (1953). The taxonomic significance of fermentative versus oxidative metabolism of carbohydrates by various gram-negative rods. *J. Bacteriol.*, 66, 24–26.
- IRRI, International Rice Research Institute (2014). World rice statistics. Retrieved from :<http://ricestat.irri.org:8080/wrs2/entrypoint.htm>
- IRRI, International Rice Research Institute (1996). Standard evaluation system for rice. International Rice Research Institute, Manila
- Jaunet, T., Notteghem, J. L., and Rapilly, F. (1996).Pathogenicity Process of *Pseudomonas fuscovaginae*, the Causal Agent of Sheath Brown Rot of Rice.*Journal of Phytopathology*, 144(9-10), 425-430.
- Jaunet, T., Laguerre, G., Lemanceau , P., Frutos, R., and Notteghem, J. L. (1995). Diversity of *Pseudomonas fuscovaginae* and other fluorescent pseudomonads isolated from diseased rice. *Phytopathology*, 85,1534-1541.
- Keen, E. C. (2012). "Paradigms of pathogenesis: Targeting the mobile genetic elements of disease". *Frontiers in Cellular and Infection Microbiology*, 2, 161
- Klement, Z., and Goodman, R. N. (1967).The Hypersensitive Reaction to Infection by Bacterial Plant Pathogens.*Annual Review of Phytopathology*, 5(1), 17-44.

- Klement, Z., Farkas, G. L. and Lovrekovich, L. (1964). Hypersensitive reaction induced by phytopathogenic bacteria in the tobacco leaf. *Phytopathology*, 54,474-477.
- Lai , W. J., Zeng , X. M., Yuan , G.K., and Ju, G. Z. (1983). A note on the identity of the causal bacteria stripe and rice heart rot (*Pseudomonas syringae* pv. *panici*). *J. S. China Agric . Coll.*, 4,79-81
- Lakshmanan, P. (1993). Efficient method of assessing resistance to sheath rot in rice. *Crop Protection*, 12(3), 189-192.
- Leyna, H., and Coyne, D.P. (1985). The effect of inoculation methods, pathogenic variability and inoculum concentrations reactions and genetics of resistance to isolates of *Xanthomonas campestris*pv. *phaseoli* (Smith) dye in leaves and pods of dry beans. Annual Report of the Bean Improvement Cooperative, 28,70-71
- Lelliot, R.A., and Stead, D.E. (1987).Methods for the Diagnosis of Bacterial Diseases of Plants. Blackwell Scientific Publication, Oxford. 216 pp
- Liu, Q., and Wang, Z. (2004).Infection characteristics of *Erwinia chrysanthemi* pv.*zeae* on rice.*Hua nan nong ye da xue xue bao = Journal of South China Agricultural University*, 25(3), 55-57.
- Lozano, J.C., Victoria, J., Volasco, A. C., and Ahn, S. W. (1981). Bacterial brown blotch , a disease of rice in tropical America. Pages 65-73 in : Proc. Int. Conf. Plant Pathog. Bact. 5<sup>th</sup> . J.C. Lozano , ed. Cent. Int. Agric. Trop., Cali , Colombia. 640 pp
- Luo, J., Xie, G., Li, B., and Lihui, X. (2007). First Report of *Burkholderia glumae* Isolated from Symptomless Rice Seeds in China. *Plant Disease*,91, 1363.
- MARDI, Malaysia Agricultural and Development Institute (2014). Information of rice varieties developed by MARDI. Retrieved from <http://agromedia.mardi.gov.my/magritech/>
- Marzukhi, H., Ali A. H., Hassan S. (1991). Kehadiran penyakit baru padi di estet padi Seberang Perak. *Teknologi Padi*, 7,49–52. (in Malay with English abstract).
- Marchetti, M.A., and Petersen, H.D. (1984). The role of *Bipolaris oryzae* in floral abortion and kernel discoloration in rice. *Plant Disease*, 68, 288-291
- Mew, T. W., Leung, H., Savary, S., Vera Cruz, C. M., and Leach, J. E. (2004).Looking Ahead in Rice Disease Research and Management.*Critical Reviews in Plant Sciences*, 23(2), 103-127
- Mew, T. W., and Cottyn, B. (2001).Seed Health and Seed-Associated Microorganisms for Rice Disease Management.*Limited Proceedings No 6. Los Banos (Phillipines) , IRRI*

Mew , T. W., and Misra, J. K. (1994). A Manual of Rice Seed Health Testing. International Rice Research Institute

Meyer, J.M. (2007). Siderotyping and Bacterial Taxonomy: A Siderophore Bank for a Rapid Identification at the Species Level of Fluorescent and Non-Fluorescent *Pseudomonas* Microbial Siderophores. In A. Varma & S. B. Chincholkar (Eds.), 12, 43-65

Misra, J.K., Merca, S.D and Mew, T. W. (1994). Fungal pathogen [In: "A Manual of Rice Seed Health Testing" (T.W. Mew, J.K. Misra, eds.) Page 5-9]. International Rice Research Institute.

Miyajima, K., Tanii, A., and Akita, T. (1983). *Pseudomonas fuscovaginae* sp. nov., nom. rev. *International Journal Of Systematic Bacteriology*, 33( 3), 656-657.

Miyajima, K. (1983). Studies on the sheath brown rot of rice caused by *Pseudomonas fuscovaginae* Tanii, Miyajima and Akita. *Rep. Hokkaido Prefect. Agric. Exp. Stn.* 43, 74

Mondal, K. K., Mani, C., Singh, J., Kim, J. G., and Mudgett, M. B. (2011).A new leaf blight of rice caused by *Pantoea ananatis* in India.*Plant Disease.*, 95, 1582-1583

Nandakumar, R., Shahjahan, A. K. M., Yuan, X. L., Dickstein, E. R., Groth, D. E., Clark, C. A., Cartwright, R. D., and Rush, M. C. (2009). *Burkholderia glumae* and *B. gladioli* cause bacterial panicle blight in rice in the southernUnited States. *Plant Disease*, 93,896-905

NCBI, National center for biotechnology information (2014).Taxonomic classification of *Pseudomonas fuscovaginae*.Retrieved from:  
[http://eol.org/pages/973020/hierarchy\\_entries/53150152/names](http://eol.org/pages/973020/hierarchy_entries/53150152/names)

Norhayati, M., Erneeza, M.H., Kamaruzaman, S., and Radziah O., (2012). Diazotrophic bacteria as biological control agent for *Lasiodiplodia Theobromae* isolated from Kenaf seeds. *Journal of Agricultural and Biological Science*, 7 (12),1076-1082

Onasanya, A., Basso, A., Somado, E., Gasore, E. R., Nwilene, F. E., Ingelbrecht, I., . . . and Onasanya, R. O. (2010). Development of a Combined Molecular Diagnostic and DNA Fingerprinting Technique for Rice Bacteria Pathogens in Africa. *Biotechnology*,9 (2), 89-105.

Ou, S. H. (1985). Rice disease. 2<sup>nd</sup> ed. Commonwealth Mycological Institute, Kew, Surrey, England , 280pp

Parish, R. W., Phan, H. A., Iacuone, S., & Li, S. F. (2012). Tapetal development and abiotic stress: a centre of vulnerability. *Functional Plant Biology*, 39(7), 553-559.

- Patel, H. K., da Silva, D. P., Devescovi, G., Maraite, H., Paszkiewicz, K., Studholme, D. J., and Venturi, V. (2012). Draft genome sequence of *Pseudomonas fuscovaginae*, a broad-host-range pathogen of plants. [Research Support, Non-U.S. Gov't]. *J Bacteriol*, 194(10), 2765-2766.
- Pearson, W.R. and Lipman D.J, (1988). Improved tools for biological sequence analysis. *Proceedings of the National Academy of Sciences, USA* 85, 2444-8.
- Peix, A., Ramirez-Bahena, M. H., and Velazquez, E. (2009). Historical evolution and current status of the taxonomy of genus *Pseudomonas*. [Historical Article Review]. *Infect Genet Evol*, 9(6), 1132-1147.
- Pizzatti, C., and Cortesi, P. (2007). Effect of chemicals, nitrogen, time of sowing and panicle brown spot epidemics on rice grain discolouration in Italy. *Journal of Plant Pathology*, 90(21), 197-209.
- Poulin, L., Raveloson, H., Sester, M., Raboin, L. M., Silué, D., Koebnik, R., and Szurek, B (2014). Confirmation of Bacterial Leaf Streak Caused by *Xanthomonas oryzae* pv. *oryzicola* on Rice in Madagascar. *Plant Dis.*, 98(10), 1423-1423
- Rostami, M., Ghasemi, A. G., Rahimian, H., and Khosravi, V. (2010). Bacterial Sheath Rot of Rice in Mazandaran Province. *Iran. J. Plant Path.*, 45(3), 61-63.
- Rott, P., Honegger, J., Notteghem, J. L., and Ranomenjanahary, S. (1991). Identification of *Pseudomonas fuscovaginae* with biochemical, serological and pathogenicity test. *Plant Dis.*, 75, 843-846
- Rott, P., Notteghem, J. L., & Frossard, P. (1989a). Identification and characterization of *Pseudomonas fuscovaginae*, the causal agent of bacterial sheath brown rot of rice, from Madagascar and other countries *Plant Dis.*, 73, 133-137.
- Rott, P., Honegger, J., and Notteghem, J. L. (1989b). Isolation of *Pseudomonas fuscovaginae* with a semiselective medium (KBS). *Int. Rice Res. Inst. Newslett.*, 14(1), 29.
- Rott, P. (1987). Brown rot (*Pseudomonas fuscovaginae*) of the leaf sheath of rice in Madagascar. Institute de Recherches Agronomiques Tropicales et des Cultures Vivrieres, Montpellier, France, pp 22
- Saad, A., Jatil Aliah T., Azmi A. R. and Normah, I. (2003). Sheath brown rot: A potentially devastating bacterial disease of rice in Malaysia. Proc. of International Rice Conference – Modern Rice Farming. Alor Setar, Kedah, 13–16 October, 2003, Alor Setar, Kedah. MARDI and MAPPS: 352–355.
- Sacchi, C.T., Whitney AM, Mayer LW et al., (2002). Sequencing of 16S rRNA gene: a rapid tool for identification of *Bacillus anthracis*. *Emerging Infectious Diseases*, 8, 1117–23.
- Schadd, N. W., ed. (1980). Laboratory Guide for Identification of Plant Pathogenic Bacteria. American Phytopathological Society , St Paul MN.68pp

Schaad, N. W., Kado, C.I., and Summer D., R. (1975) Synonymy of *Pseudomonas avenae* Mannus 1905 and *Pseudomonas pseudo-alcalagenes* Rosen 1922, *Int. J. Syst. Bacteriol.*, 25,133-137

Samnang, C, (2004). The Rice Industry in Cambodia. Phnom Penh, Cambodia: Economic Institute of Cambodia, Special Report.

Shakya, D., Vinther , F., and Mathur, S. (1985). Worldwide distribution of bacterial stripe pathogen of rice identified as *Pseudomonas avenae*. *Phytopathol. Z*, 114: 256-259

Sharma, S., Sthapit B.R., Pradhanang P.M., Joshi K.D. (1997).Bacterial sheath brown rot of rice caused by *Pseudomonas fuscovaginae* in Nepal. p. 107–112. In: “Rice cultivation in highlandareas” (C. Poisson, J. Rakotoarisoa, eds.). Proc. of the CIRAD Conference, Antananarivo, Madagascar. 29 March–5 April 1996.

Singh, R., and Chand, H. (1985) Rice grain discoloration and its chemical control . *Int Rice Res. Newsl* 10:16

Song, W.Y., Kim H.M., Hwang C.Y. and Schaad N.W. (2004) Detection of *Acidovorax avenae* ssp.*avenae* in rice seeds using BIO-PCR. *Journal of Phytopathology*, 152: 667–676.

Song, W-Y, Kang M-H, Kim H-M (1999).Current status of bacterial brown stripe of rice caused by *Acidovorax avenae* subsp. *avenae*. *Plant Disease and Agriculture*,5, 69–76.

Stead, D.E. (1992). Grouping of plant-pathogenic and some other *Pseudomonas* spp. by using cellular fatty acid profiles.*International Journal of Systematic Bacteriology*,42, 281–295.

Suslow, T. V., Schroth, M.N., and Isaka, M. (1982).Application of a rapid method for Gram differentiation of plant pathogenic and saprophytic bacteria without staining. *Phytopathology*, 72,917-918

Tanii, A., Miyajima, K., and Akita, T. (1976). The Sheath Brown Rot Disease of Rice Plant and Its Causal Bacterium, *Pseudomonas fuscovaginae* A. Tanii, K. Miyajima et T. Akita sp. nov. *Ann. Phytopath. Soc. Japan* ,42, 540-548.

Tanii, A., Baba, T., and Haruki , T. (1974). Bacteria isolated from black rot of rice grains. *Ann, Phytopathol. Soc. Jpn.*, 40, 309-318

Takeuchi, T., Sawada H., Suzuki F. and Matsuda I. (1997) Specific detection of *Bukholderia plantarii* and *B. glumae* by PCR using primers selected from the 16S–23S rDNA spacerregions. *Annals of Phytopathological Society of Japan*, 63, 455–462.

Tamura K and Nei,M (1993). Estimation of the number of nucleotide substitutions in the control region of mitochondrial DNA in humans and chimpanzees.*Molecular Biology and Evolution*, 10, 512–526.

- Tamura, K., Dudley J., Nei M., and Kumar S (2007). MEGA4: molecular evolutionary genetics analysis (MEGA) software version 4.0. *Molecular Biology and Evolution*, 24, 1596–1599.
- Thompson, J.D., Higgins D.G., and Gibson T.J. (1994). CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Research*, 22, 4673–4680.
- Tilquin, J.P. and Detry J.F. (1993). Efficiency of natural selection for bacterial sheath rot (BSR) in bulked families. *Intl Rice ResNotes*, 18, 23–24.
- Tominaga, T., Kimura., and Goh, N, (1981). Bacterial brown stripe of rice in nursery box, caused by *Pseudomonas avenae*. *Ann. Phytopathol. Soc. Jpn .*, 49,92
- Tsushima, S., Mogi S., Saito H. (1985). Effects of inoculum density, incubation temperature and incubation period on the development of rice bacterial grain rot. Proc. of Association of Plant Protection of Kyushu 31: 11–12. (in Japanese with English abstract).
- Turner, S., Pryer, K.M., Miao, V.P.W., and Palmer, J.D. (1999). Investigating deep phylogenetic relationships among cyanobacteria and plastids by small subunit rRNA sequence analysis. *Journal of Eukaryotic Microbiology*, 46, 327–338
- Tvrzová, L., Schumann, P., Spröer, C., Sedlá,\_cek, I., Pá\_cová Z., Šedo O., Zdráhal Z., Steffen, M, and Lang, E. (2006) *Pseudomonas moraviensis* sp. nov. and *Pseudomonas vranovensis* sp. nov., soil bacteria isolated on nitroaromatic compounds, and emended description of *Pseudomonas asplenii*. *International Journal of Systematic and Evolutionary Microbiology* 56, 2657–2663.
- Uematsu, T., Yoshimura, D., Nishiyama, K., Ibaraki, T. and Fujii, H. (1976). Occurrence of bacterial seedling rot in nursery flat, caused by grain rot bacterium *Pseudomonas glumae*. *Annals of Phytopathological Society of Japan*, 42, 310–312.
- Urakami , T., Ito-Yoshida, C., Araki , H., Kijima, T., Suzuki , K.-I., and Komagata, K. (1994). Tranfer of *Pseudomonas plantarii* and *Pseudomonas glumae* to *Burkholderia* as *Burkholderia* spp. and description of *Burkholderia vandii* sp. Nov. *Int. J. Syst. Bacteriol.* , 44, 235-245
- Upadhyay, R. K. (1985). Rice disease status in India. *Int. Rice Res. Newsl.*, 10,17-18
- Valencia-Botín, A. J., Mendoza-Onofre, L. E., Silva-Rojas, H. V., Córdova-Téllez, L., Espinosa-Victoria, D., Valadez-Moctezuma, E., and Villaseñor-Mir, H. E. (2007). Aggressiveness estimates and inoculation methods of pathogenic bacteria on seeds and seedlings of wheat 'Seri M82'.,30(3), 255-259.
- Vematsu, T., Yoshimura, D., Nishiyama, K., Ibaraki, T., and Fujii, H. (1976) Pathogenic bacterium causing seedling rot of rice. *Ann. Phytopathol. Soc. Jpn*, 42,464-471

Vidhyasekaran, P., Ranganathan, K., Rajamanickam, B., and Radhakrishnan, J. (1984), Quality of rice grains from sheath rot-affected plants. *Int. Rice Res. Newslet.*, 9,19

Wakimoto, S., Makoto, A., and Tscchiya, K. (1987). Serological specificity of *Pseudomonas glumae*, the pathogenic bacterium of grain rot disease of rice. *Ann. Phytopathol. Soc. Jpn.*, 53,150-158

Webster, R.K. and Gunnel P.S. (1992) Compendium of Rice Diseases. The American Phytopathological Society, St. Paul, MN, pp. 62.

Willems, A., De Vos, P., Gillis, M., and Kersters, K. (1992a). Towards an improved classification of *Pseudomonas*. In: Identification Methods in Applied and Environmental Microbiology. R. G. Board , D. Jones, and F. A. Skinner , eds. The Society for Applied Bacteriology Technical Series 29. Blackwwall Scientific Publications, Oxford.

Willems, A., Goor, M., Thielemans, S., Gillis, M., Kersters , K., and De Ley, J. (1992b). Tranfer of several phytopathogenic *Pseudomonas* species to *Acidovorax avenae* subsp. *Avenae* subsp. Nov., comb. Nov., *Acidovorax avenae* subsp. *citrulli*, *Acidovorax avenae* subsp. *cattleya*, and *Acidovorax konjaci*. *Inst. J. Syst. Bacteriol.* , 42 , 107-119

Wilson, R. A. & Talbot, N. J. (2009). Under pressure: Investigating the biology of plant infection by *Magnaporthe oryzae*. *Nature Reviews Microbiology* 7 (3): 185–95

Xie, G, Soad A, Swings J, and Mew T.W, (2003). Diversity of gram negative bacteria antagonistic against major pathogens of rice from rice seed in the tropic environment. *Journal of Zhejiang University Science*, 4, 463–8.

Yan, H., Yu, S. H., Xie, G. L., Fang, W., Su, T., and Li, B. (2010). Grain Discoloration of Rice Caused by *Pantoea ananatis* (synonym *Erwinia uredovora*) in China. *Plant Disease.*,94(4), 482-482

Zeier, J (2005).Age-dependent variations of local and systemic defence responses in *Arabidopsis* leaves towards an avirulent strain of *Pseudomonas syringae*. *Physiol Mol Plant Pathol*, 66,30–39

Zeigler, R. S., and Alvarez, E. (1990). Characteristics of *Pseudomonas* spp. causing grain discoloration and sheath rot of rice, and associated pseudomonad epiphytes *Plant Dis.*, 74, 917-922.

Zeigler , R.S., and Alvarez E. (1989). Differential culture medium for *Pseudomonas* species causing sheath rot (ShR) and grain discoloration (GID) of rice. *Int. Rice Res Inst. Newslet.*, 14, 27-28

Zeigler, R. S., & Alvarez, E. (1987).Bacterial sheath brown rot of rice caused by *Pseudomonas fuscovaginae* in Latin America.*Plant Dis.*, 71, 592-597.

Zeigler, R. S., Aricapa, G., and Hoyos, E. (1987). Distribution of fluorescent *Pseudomonas* spp. causing grain and sheath discoloration of rice in Latin America. *Plant Dis.*, 71, 896-900.

Zeigler, R. S., Hoyos , E., and Aricapa, G. (1986) Non-rice host of the causal agent of bacterial sheath brown rot (BSBR) in Latin America. *Int . Rice Res. Inst. Newsl .*, 11,19-20



## BIODATA OF STUDENT

MOHD ZAFRI BIN AB WAHAB

### Personal Background

Date of Birth	05-01-1987
New I/C No.	870105025917
Age	27
Gender	Male
Religion	Islam
Marital status	Married

### Latest info

Address	B82, Taman Derga Jaya, Jalan Datuk Kumbar, 05300, Alor Star, Kedah
Home Tel. No	-
Office Tel. No	-
Hand phone No.	0196338086
Email	zafri_wahab@yahoo.com

### Education

#### Education Level

Program Name	Bachelor with Honest
Field of study	Bac. Hons. Sc. Tech. and Plantation Management
Specification	Management and Biotechnology
Final Result	Management and Biotechnology
Institution	Plant Biotechnology
Graduation date	3.61
Thesis	Universiti Teknologi MARA
	Oktober 2011
	Antagonistic Activity of Effective Microorganism Against Plant Pathogen

#### Education Level

Program Name	Diploma
Field of study	Diploma Microbiology
Specification	Biotechnology
Final Result	Microbiology
Institution	3.40
Graduation date	Universiti Teknologi MARA
Thesis	Oktober 2008
	Isolation of Actinomycetes from soil

### Outside activities and achievement

Asmah, A., Mohd-Zafri, A.W., Shamsiah, Razak, B., Beneficial Microorganism from Fermented Coconut Water, ITEX 2011.

## PUBLICATION

Mohd-Zafri, Kamaruzaman, S., Razi, I., Marzukhi, H., (2013). Characterization the causal agent of sheath brown of rice from rice plant, In proceedings of International Congress of the Malaysian Society for Microbiology, Langkawi, Malaysia.12-15 November 2013.





## UNIVERSITI PUTRA MALAYSIA

### STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

ACADEMIC SESSION : \_\_\_\_\_

#### TITLE OF THESIS / PROJECT REPORT :

---

---

---

#### NAME OF STUDENT : \_\_\_\_\_

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

1. This thesis/project report is the property of Universiti Putra Malaysia.
2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as :

\*Please tick (v )

**CONFIDENTIAL**

(Contain confidential information under Official Secret Act 1972).

**RESTRICTED**

(Contains restricted information as specified by the organization/institution where research was done).

**OPEN ACCESS**

I agree that my thesis/project report to be published as hard copy or online open access.

This thesis is submitted for :

**PATENT**

Embargo from \_\_\_\_\_ until \_\_\_\_\_  
(date) (date)

**Approved by:**

(Signature of Student)  
New IC No/ Passport No.:

(Signature of Chairman of Supervisory Committee)  
Name:

Date :

Date :

[Note : If the thesis is **CONFIDENTIAL** or **RESTRICTED**, please attach with the letter from the organization/institution with period and reasons for confidentiality or restricted. ]