



**UNIVERSITI PUTRA MALAYSIA**

***HABITAT, MORPHOLOGY, POPULATION GENETICS AND  
REPRODUCTIVE BIOLOGY OF HARD CLAM (BIVALVIA: VENERIDAE)  
FROM TWO LOCATIONS IN SARAWAK***

**HADI BIN HAMLİ**

**FSPM 2015 9**



**UPM**  
UNIVERSITI PUTRA MALAYSIA  
BERILMU BERBAKTI

**HABITAT, MORPHOLOGY, POPULATION GENETICS AND  
REPRODUCTIVE BIOLOGY OF HARD CLAM (BIVALVIA: VENERIDAE)  
FROM TWO LOCATIONS IN SARAWAK**

By

**HADI BIN HAMLI**

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfillment of the Requirement for the Degree of Philosophy

December 2015

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

**HABITAT, MORPHOLOGY, POPULATION GENETICS AND  
REPRODUCTIVE BIOLOGY OF HARD CLAM (BIVALVIA: VENERIDAE)  
FROM TWO LOCATIONS IN SARAWAK**

By

**HADI BIN HAMLİ**

**December 2015**

**Chairman: Associate Professor Mohd Hanafi Idris, PhD**

**Faculty: Agriculture and Food Sciences (Bintulu)**

Biology and habitat characteristic of most common and abundance hard clam *Meretrix* spp. was investigated at two selected division of Sarawak namely Kuching and Betong from May 2013 to April 2014. Both areas comprised intertidal areas with sandy type sediment which able to support the growth of *Meretrix* spp. Environmental factor play important role to influence biology and physiology of *Meretrix* spp. for the present study. Habitat area for *Meretrix* spp. in the present study recorded the concentration of hydrogen ion (pH) range from 7.73–8.31, total dissolved solid 35.358–50.467 mg/L, salinity 22.5–31.8 psu, temperature 25.3–27.7 °C, turbidity 82.9–999 ntu, conductivity 3.473–4.683 S/cm, dissolved oxygen 3.74–5.58 mg/L, total rainfall 162.6–729.4 mm, ammonia 0.023–0.223 mg/L, nitrite 0.004–0.017 mg/L, nitrate 0.167–1.233 mg/L, phosphate 0.097–0.43 mg/L, total suspended solid 0.041–0.147 mg/L and chlorophyll *a* 0.419–0.147 µg/L.

A total of 3 hard clam species identified based on morphological characteristic were *Meretrix lyrata*, *M. meretrix* and *M. lusoria*. Pallial sinus scar profile was the main morphological characteristic used for the differentiation of these 3 *Meretrix* spp. While, a total of 13 morphometric characteristics were used to differentiate the three species of *Meretrix* recorded from Sarawak. Significant differences (ANOVA,  $p < 0.05$ ) on 7 morphometric characteristics (SW; Shell Width, AL; Anterior Length, LCT; Length of Cardinal Tooth, AW; Anterior Adductor Scar Width, PW; Posterior Adductor Scar Width, PS; Pallial Sinus Open Scar and LL; Ligament Length) based on proportion ratio with SL (Shell Length) were found among three *Meretrix* species recorded. Variation among *Meretrix* shell characteristic also was strengthened by results from clustering analysis, Principal Component Analysis (PCA) and genetic characteristic.

Genetic characteristic between *M. lyrata*, *M. meretrix* and *M. lusoria* were investigated based on universal marker cytochrome C oxidase subunit I (COI) with sequence LCO1490: 5'-ggtaacaaatcataaagatattgg-3' and HCO2198: 5'-taaacttcagggtgacccaaaaatca-3'. Genetic characteristic between this *Meretrix* spp. was able to be distinguished through phylogenetic analysis. The result was crucial to support the morphology and morphometric characterization. However, the present study using molecular approach was unable to verify the species of *Meretrix* sp. Therefore, additional genetic approaches are needed for further verification.

Present investigation on reproductive biology was able to determine the reproductive stages and spawning period of *Meretrix lyrata* within one year. Determination was made based on quantitative (Gonad Index and Condition Index) and qualitative approach (histological procedure). This study showed that the spawning period of *M. lyrata* is from May to September. Most of the *M. lyrata* in the studied samples undergoes continuous gonad development for 7 months from October 2013 to April 2014. This indicated *M. lyrata* only has one cycle of reproductive development in a year. Reproductive development of *M. lyrata* positively correlated to the abundance of phytoplankton in coastal area.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan ijazah Doktor Falsafah

**HABITAT, MORFOLOGI, POPULASI GENETIK DAN BIOLOGI  
PEMBIAKAN KERANG KERAS (BIVALVIA: VENERIDAE) DARI DUA  
LOKASI DI SARAWAK**

Oleh

**HADI HAMLİ**

**Disember 2015**

**Pengerusi: Professor Madya Mohd Hanafi Idris, PhD**

**Fakulti: Sains Pertanian dan Makanan (Bintulu)**

Biologi dan ciri habitat kerang keras *Meretrix* spp. yang paling biasa dan banyak telah dikaji di dua bahagian terpilih di Sarawak iaitu Kuching dan Betong bermula dari Mei 2013 hingga April 2014. Kedua-dua kawasan terdiri daripada kawasan pasang surut dengan mendapan jenis berpasir yang dapat menyokong pertumbuhan *Meretrix* spp. Faktor persekitaran memainkan peranan yang penting dalam mempengaruhi biologi dan fisiologi *M. lyrata* dalam kajian ini. Kajian ini merekodkan julat kepekatan ion hydrogen (pH) antara 7.73–8.31, jumlah pepejal terlarut 35.358–50.467 mg/L, kemasinan 22.5–31.8 psu, suhu 25.3–27.7 °C, kekeruhan 82.9–999 ntu, kekonduksian 3.473–4.683 S/cm, oksigen terlarut 3.74–5.58 mg/L, jumlah hujan 162.6–729.4 mm, nitrogen ammonia 0.023–0.223 mg/L, nitrit 0.004–0.017 mg/L, nitrat 0.167–1.233 mg/L, fosfat 0.097–0.43 mg/L, jumlah pepejal terampai 0.041–0.147 mg/L dan klorofil *a* 0.419–0.147 µg/L.

Tiga spesis kerang keras telah dikenalpasti dengan jelas berdasarkan ciri morfologi iaitu *Meretrix lyrata*, *M. meretrix* and *M. lusoria*. Bentuk parut sinus pallial adalah ciri utama morfologi yang boleh membezakan tiga spesis *Meretrix* yang direkod di Sarawak. Perbezaan ketara dikenalpasti antara tiga rekod spesis *Meretrix* (ANOVA,  $p < 0.05$ ) terhadap tujuh ciri morfometrik (SW; Lebar Cengkerang, AL; Panjang Anterior, LCT; Panjang Gigi Utama, AW; Lebar Parut Adductor Anterior, PW; Lebar Parut Adductor Posterior, PS; Bukaan Parut Sinus Pallial and LL; Panjang Ligament) berdasarkan nisbah perkadaran dengan SL (Panjang Cengkerang). Variasi antara ciri cengkerang *Meretrix* juga telah disokong oleh keputusan analisis kelompok, Analisis Komponen Utama (PCA) dan ciri genetik.

Ciri genetik di antara *M. lyrata*, *M. meretrix* dan *M. lusoria* telah dikaji berdasarkan penanda umum “cytochrome C oxidase” subunit I (COI) dengan jujukan LCO1490: 5'-ggcacaacaatcataaagatattgg-3' and HCO2198: 5'-taaactcagggtgaccaaataatca-3'. Ciri genetik di antara *Meretrix* spp. telah dapat dibezakan melalui analisis phylogenetik. Hasil keputusan adalah penting untuk menyokong pencirian morfologi dan morfometrik. Walau bagaimanapun, kajian ini menggunakan pendekatan molekular tidak dapat mengesahkan spesies *Meretrix* sp. Oleh itu, pendekatan genetik tambahan diperlukan untuk mengesahkan spesies tersebut.

Kajian semasa keatas biologi pembiakan telah menentukan peringkat gonad dan tempoh bertelur untuk *Meretrix lyrata* dalam setahun. Penentuan adalah berdasarkan pendekatan kuantitatif (Index Gonad dan Index Keadaan) dan pendekatan kualitatif (tatacara histologi). Kajian menunjukkan *M. lyrata* dalam sample yang dikaji mempunyai tempoh bertelur dari Mei hingga September. Kebanyakan individu menjalani perkembangan gonad yang berterusan selama 7 bulan bermula dari Oktober hingga April 2014. Hasil kajian menunjukkan *M. lyrata* mempunyai satu kitaran pembiakan dalam setahun. Perkembangan pembiakan *M. lyrata* menunjukkan kaitan positif terhadap kelimpahan fitoplankton di perairan tersebut.

## ACKNOWLEDGEMENTS

All praises be unto the Allah Almighty whose blessings and gave me strength to finish up my PhD project and completing this thesis. I would like to thanks to my beloved family that gave me support and inspired me to be more spirited. They taught me to never give up in anything I do. Special thanks to supervisor committee members Associate Professor Dr. Mohd Hanafi Idris, Dr. Abu Hena Mustafa Kamal and Dr. Amy Halimah Rajae for their valuable advice and guidance during the completing this project. They gave high commitment to ensure I can finish my project smoothly. Also thanks to lab assistants Mr. Awangku Ahmad Nizam and Mr. Mohd Ezwandi Bin Mahrit for their help in water quality analysis and plankton observation. Special thanks to my friends Mohd. Hafizbillah Bin Zawawi for his support during sanpling and Azimah Binti Abdul Rahim for her support in completing the journal. Thanks also to Faculty of Agriculture and Food Sciences, Universiti Putra Malaysia Bintulu Sarawak Campus for technical, logistic supports and laboratory facilities provided. Lastly special thanks Ministry of Education Malaysia, for the research grant (5524237 FRGS) which make this study possible.



I certify that a Thesis Examination Committee has met 10 December 2015 to conduct the final examination of Hadi Bin Hamli on his thesis entitled “Habitat, Morphology, Population Genetics and Reproductive Biology of Hard Clam (*Bivalvia: Veneridae*) from Two Locations in Sarawak” in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as Follows:

**Osumanu Haruna Ahmed, PhD**

Associate Professor  
Faculty of Agriculture and Food Sciences  
Universiti Putra Malaysia  
(Chairman)

**Hishamuddin Bin Omar, PhD**

Senior Lecturer  
Faculty of Science  
Universiti Putra Malaysia  
(Internal Examiner)

**Annie Christianus, PhD**

Senior Lecturer  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Internal Examiner)

**Sukree Hajisamae, PhD**

Associate Professor  
Faculty of Science and Technology  
Prince of Songkla University  
Thailand  
(External Examiner)

---

**ZULKARNAIN ZAINAL, PhD**

Professor and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 10 March 2016

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

**Mohd Hanafi Idris, PhD**

Associate Professor  
Faculty of Agriculture and Food Sciences  
Universiti Putra Malaysia  
(Chairman)

**Abu Hena Mustafa Kamal, PhD**

Senior lecturer  
Faculty of Agriculture and Food Sciences  
Universiti Putra Malaysia  
(Member)

**Amy Halimah Rajae, PhD**

Senior lecturer  
Faculty of Agriculture and Food Sciences  
Universiti Putra Malaysia  
(Member)

---

**BUJANG KIM HUAT, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 10 March 2016

## TABLE OF CONTENTS

	<b>Page</b>
ABSTARCT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
LIST OF TABLES	xiv
LIST OF FIGURES	xvi
LIST OF ABBREVIATIONS	xix
<b>CHAPTER</b>	
<b>1 GENERAL INTRODUCTION</b>	<b>1</b>
1.1 Background	1
1.2 Problem Statement	2
1.3 Objectives	3
<b>2 LITERATURE REVIEW</b>	<b>4</b>
2.1 Veneridae Morphology	4
2.1.1 Classification	5
2.2 Habitat	5
2.2.1 Sediment Characteristic	5
2.2.2 Temperature	6
2.2.3 Dissolved Oxygen	7
2.2.4 Salinity	8
2.3 Feeding and Nutrient	8
2.4 Culture	9
2.4.1 Shellfish Production	10
2.5 Gonad Development	13
2.6 Embryonic and Larva Development	16
2.7 Genetic Characterization	18
2.7.1 Allozyme	18
2.7.2 Mitochondrial DNA Marker (mtDNA)	18
2.7.3 Nuclear DNA Marker	19
<b>3 GENERAL METHODOLOGY</b>	<b>21</b>
3.1 Description of Study Area	21
3.2 <i>Meretrix</i> spp. Habitat Study	22
3.2.1 Nutrient and Chlorophyll <i>a</i> Analysis	22
3.2.2 Sediment Structure Analysis	22
3.3 Collection and Identification of <i>Meretrix</i> spp.	22

<b>4</b>	<b>NATURAL HABITAT CHARACTERISTIC OF <i>Meretrix</i></b>	<b>24</b>
4.1	Introduction	24
4.1.1	Objectives	25
4.2	Materials and Methods	25
4.2.1	Study Site Description	25
4.2.2	Seawater and Sediment Collection	27
4.2.3	Determination of Ammonia Nitrogen ( $\text{NH}_4^+$ )	27
4.2.4	Determination of Nitrite ( $\text{NO}_2^-$ )	27
4.2.5	Determination of Nitrate ( $\text{NO}_3^-$ )	28
4.2.6	Determination of Ortho-phosphate ( $\text{PO}_4^{3-}$ )	29
4.2.7	Analysis of Total Suspended Solids (TSS)	29
4.2.8	Determination of Chlorophyll <i>a</i>	30
4.2.9	Analysis of Sediment Characteristic	30
4.2.10	Statistical Analysis	32
4.3	Results	32
4.3.1	Tidal Pattern	32
4.3.2	Physico-Chemical Parameters	33
4.3.3	Sediment Texture and Grain Size	37
4.4	Discussion	37
4.4.1	Tidal Pattern and Sediment Characteristic	39
4.5	Conclusion	40
<b>5</b>	<b>MORPHOLOGY AND MORPHOMETRIC CHARACTERISTICS OF HARD CLAM <i>Meretrix</i></b>	<b>41</b>
5.1	Introduction	41
5.1.1	Objectives	42
5.2	Materials and Methods	42
5.2.1	Sample Preservation	42
5.2.2	Morphological Identification	43
5.2.3	Morphometric Study	43
5.2.4	Morphometric Data Analysis	44
5.3	Results	45
5.3.1	Morphological Characters of <i>Meretrix</i> Species	45
5.3.2	Exterior Shell Morphology Variation of <i>Meretrix</i> Species	52
5.3.3	Interior Shell Morphology Variation	53
5.3.4	Morphometric Study of <i>Meretrix</i> Species	53
5.3.5	Cluster Analysis	56
5.3.6	Principal Component Analysis (PCA)	57

	5.3.7 Dichotomus Key for <i>Meretrix</i> spp.	59
5.4	Discussion	59
5.5	Conclusion	62
<b>6</b>	<b>GENETIC CHARACTERISTIC OF HARD CLAM <i>Meretrix</i></b>	<b>63</b>
6.1	Introduction	63
	6.1.1 Objectives	63
6.2	Materials and Methods	64
	6.2.1 Sample Collection	64
	6.2.2 DNA Extraction	64
	6.2.3 DNA Quantitation and Electrophoresis	64
	6.2.4 Polymerase Reaction Chain (PCR) Amplification	65
	6.2.5 Purification and Sequencing	66
	6.2.6 Statistical Analysis	66
6.3	Results	67
	6.3.1 Sequences Characteristics	68
	6.3.2 Phylogenetic Analysis	69
6.4	Discussion	71
6.5	Conclusion	73
<b>7</b>	<b>SOME ASPECT ON REPRODUCTIVE BIOLOGY OF <i>Meretrix lyrata</i></b>	<b>74</b>
7.1	Introduction	74
	7.1.1 Objectives	75
7.2	Materials and Methods	75
	7.2.1 Length-Weight relationship	76
	7.2.2. Histological Procedure	76
	7.2.3 Condition Index (CI) Analysis	79
	7.2.4 Wet and Dry Weight	79
	7.2.5 Statistical Analysis	79
7.3	Results	79
	7.3.1 Length-Weight Relationship	81
	7.3.2 Male Gonad Maturation Stages	81
	7.3.3 Female Gonad Maturation Stages	83
	7.3.4 Sex Ratio	85
	7.3.5 Monthly Variation in <i>Meretrix lyrata</i> Gamete Phases	86
	7.3.6 Monthly Gonad Index (GI) of <i>Meretrix lyrata</i>	87
	7.3.7 Monthly Condition Index (CI) of <i>Meretrix lyrata</i>	88
	7.3.8 Correlation of <i>Meretrix lyrata</i> Condition Index (CI) with Gonad Index (GI), Wet and Dry Weight	88

7.3.9	Correlation of <i>Meretrix lyrata</i> Gonad Index (GI) with Environmental Factors	91
7.3.10	Correlation of <i>Meretrix lyrata</i> Condition Index (CI) with Environmental Factors	92
7.4	Discussion	93
7.4.1	Size Distribution	93
7.4.2	Length-Weight Relationships	92
7.4.3	Monthly Gonad Maturation Stages of <i>Meretrix lyrata</i>	92
7.4.4	Sex Ratio	95
7.4.5	Monthly Gonad Index (GI) of <i>Meretrix lyrata</i>	96
7.4.6	Monthly Condition Index (CI) of <i>Meretrix lyrata</i>	97
7.4.7	Correlation between Condition Index (CI) with Gonad Index (GI) and Tissue Weight of <i>Meretrix lyrata</i>	97
7.4.8	Correlation of <i>Meretrix lyrata</i> Gonad Development with Environmental Factors	98
7.5	Conclusion	99
<b>8</b>	<b>SUMMARY, CONCLUSION AND RECOMMENDATIONS</b>	<b>100</b>
	<b>REFERENCES</b>	<b>103</b>
	<b>APPENDICES</b>	<b>121</b>
	<b>BIODATA OF STUDENT</b>	<b>126</b>
	<b>LIST OF PUBLICATIONS</b>	<b>127</b>

## LIST OF TABLES

<b>Table</b>	<b>Page</b>
1.1 Shellfish landing in Malaysia from 2009 to 2013 in tone	2
2.1 Major marine cultured bivalve in the world (Garibaldi, 1996)	10
2.2 Classification stages of bivalve gonad	14
2.3 Clams species that have been propagate through artificial breeding	16
4.1 Grain size classification scheme	32
4.2 Monthly variation of different water parameters in the Buntal estuary from May 2013 to April 2014	34
4.3 Correlations among physico-chemical parameters by Pearson ( <i>r</i> ) correlation	35
4.4 Sediment texture and grain size	37
5.1 Morphometric characteristic abbreviation used for <i>Meretrix</i> spp identification	43
5.2 Analysis of one way ANOVA with Tukey's HSD test for 12 morphometric characteristic proportion with shell length of three <i>Meretrix</i> species	54
5.3 Loading of variables on the first four principal components <i>Meretrix</i> species	58
6.1 List of <i>Meretrix</i> accession sequences from the GeneBank (NCBI)	67
6.2 Similarity percentage of present study on <i>Meretrix</i> spp. with 16 selected <i>Meretrix</i> spp. mtDNA accession sequences from GenBank (NCBI)	68
6.3 Nucleotide base composition for accession sequences	69
7.1 Stages in dehydration and paraffin wax infiltration	77
7.2 Staining procedure	78
7.3 Monthly length-weight relationship of collected <i>Meretrix lyrata</i> for gonad development study	81
7.4 Monthly distribution of male and female of <i>Meretrix lyrata</i>	86

7.5	Reproductive stages of <i>Meretrix lyrata</i> from Buntal estuary, Sarawak	86
7.6	Monthly male and female Gonad Index (GI) for <i>Meretrix lyrata</i> from Buntal estuary	87
7.7	Pearson Correlation between Condition Index (CI) with other variables	89
7.8	Pearson correlation analysis of Gonad Index (GI) of <i>Meretrix lyrata</i> with different environment parameters at the Buntal estuary	91
7.9	Pearson correlation analysis of Condition Index (CI) of <i>Meretrix lyrata</i> with different environment parameters at the Buntal estuary	92
7.10	Previously published values of the coefficients $a$ and $b$ for <i>Meretrix</i>	94
7.11	Comparison of Condition Index (CI) of <i>M. lyrata</i> with those of other shellfish	98
1A	Maximum Likelihood fits of 24 different nucleotide substitution models	123



## LIST OF FIGURES

Figure		Page
2.1	Comparison of cultured and wild captured bivalve for 1991, 1995, 2000 and 2005. Adapted from Globefish-FAO (2007) and Helm <i>et al.</i> , (2004)	10
2.2	World production of cultured bivalve in year 1999 (FAO, 2001a)	11
2.3	Malaysian bivalve production in year 2013 (Department of Fisheries Malaysia, 2013)	12
2.4	Stages of ovary development. Stage 0: rest, Stage 1: early development, Stage 2: late development, Stage 3: Mature (Duinker <i>et al.</i> , 2008)	15
2.5	Stage of testis development. Stage 0: rest, Stage 1: early development, Stage 2: late development, Stage 3: Mature (Duinker <i>et al.</i> , 2008)	15
2.6	Bivalve larval development stages after Reverol <i>et al.</i> (2004) and Costa <i>et al.</i> (2008)	17
3.1	Study area for natural habitat of <i>Meretrix</i> spp.	21
4.1	Map of <i>Meretrix</i> spp. distribution at two division of Sarawak	25
4.2	Habitat of <i>Meretrix</i> spp. at the Sungai Buntal estuary	26
4.3	<i>Meretrix lyrata</i> burrowed itself in sediment during the high tide	26
4.4	Triangular classification of soil texture, adapted from Hawver and Bassuk (2007)	31
4.5	Monthly high tide at the study area (Sarawak Marine Department Malaysia, 2013; Sarawak Marine Department, 2014)	33
5.1	Map of <i>Meretrix</i> spp. distribution at two division of Sarawak	42
5.2	Measurements of shell characters for <i>Meretrix</i> spp. Note: 1(SL), 2(SH), 3(SW), 4(LL), 5(PL), 6(AL), 7(UL), 8(LCT), 9(LPAS), 10(PW), 11(AW), 12(PVM), 13(PS)*	44

5.3	Morphological structure of <i>Meretrix lyrata</i> (Sowerby, 1851) A; Outer view, B; Inner view, C; Sketch of inner view	47
5.4	Morphological structure of <i>Meretrix meretrix</i> (Linnaeus, 1958), A; Outer view, B; Inner view, C; Sketch of inner view	49
5.5	Morphological structure of <i>Meretrix lusoria</i> (Roding, 1798), A; Outer view, B; Inner view, C; Sketch of inner view	51
5.6	Outer shell characteristic between <i>Meretrix</i> species. (A) <i>M. lyrata</i> ; (B) <i>M. Meretrix</i> ; (C) <i>M. lusoria</i>	52
5.7	Pallial sinus scar pattern for (A) <i>Meretrix lyrata</i> ; (B) <i>M. meretrix</i> ; (C) <i>M. lusoria</i>	53
5.8	Differences of pallial sinus open scar (PS) between three local hard clams. (A) <i>M. lyrata</i> , (B) <i>M. meretrix</i> (C) <i>M. lusoria</i>	55
5.9	Box plot of morphometric characteristic for <i>Meretrix</i> spp.	56
5.10	Hierarchical cluster between three local hard clams from Sarawak	57
5.11	Principal Component Analysis (PCA) for three local hard clams from Sarawak	58
6.1	Study area of <i>Meretrix</i> spp. for genetic study	64
6.2	Fragment size produce by <i>Meretrix</i> spp. using COI marker. 1: <i>M. lyrata</i> , 2: <i>M. lusoria</i> , 3: <i>M. meretrix</i>	67
6.3	Phylogenetic tree of <i>Meretrix</i> accession sequence inferred from the Maximum Likelihood analysis using Tamura three-parameter model. Only bootstrap score greater than 50% are shown	70
7.1	Study site for monthly <i>Meretrix lyrata</i> collection	75
7.2	Total length class of <i>Meretrix lyrata</i> used for gonad development study	80
7.3	Total body weight class of <i>Meretrix lyrata</i> used for gonad development study	80
7.4	Male gonad stages for <i>Meretrix lyrata</i> , A: Rest, B: Early development, C: Late development, D: Matured, E: Spawning, F: Spent, Scale: 50 $\mu$ m. Note: ct = connective tissue; dgs = degenerate spermatozoa/ spermatid/ spermatocyte; f = follicle; fw = follicle wall; lu = lumen; mt = muscular tissue; sz = spermatozoa; sd = spermatid; s = spermatocyte	83

7.5	Female gonad stages for <i>Meretrix lyrata</i> , A: Rest, B: Early development, C: Late development, D: Matured, E: Spawning, F: Spent, Scale: 50 $\mu$ m. Note: dgs = degenerate oocyte; do= developing oocyte; gv=germinal vesicle; mo= matured oocyte; mt= muscular tissue; n=nucleus; ct=connective tissue; lu= lumen; f=follicle; fw=follicle wall	85
7.6	Gonad Index (GI) values for <i>Meretrix lyrata</i> from Buntal estuary	87
7.7	Temporal Condition Index (CI) for <i>Meretrix lyrata</i> from Buntal estuary (Mean $\pm$ standard error)	88
7.8	Monthly correlation graph between Condition Index (CI) and wet weight of <i>Meretrix lyrata</i>	89
7.9	Monthly correlation graph between Condition Index (CI) and dry weight of <i>Meretrix lyrata</i>	90
7.10	Monthly correlation graph between Condition Index (CI) and Gonad Index (GI) of <i>Meretrix lyrata</i>	90
7.11	Monthly correlation graph between <i>Meretrix lyrata</i> Gonad Index (GI) and chlorophyll <i>a</i>	92
7.12	Monthly graph pattern between <i>Meretrix lyrata</i> Condition Index (CI) and chlorophyll <i>a</i> concentration in the seawater	93

## LIST OF ABBREVIATIONS

ANOVA	Analysis of Variance
AL	Anterior length
AW	Anterior adductor scar width
Bp	Base pair
DNA	Deoxyribonucleic Acid
FAO	Food and Agriculture Organization
LCT	Length of cardinal tooth
LL	Ligament length
LPAS	Length of posterior adductor scar to anterior adductor scar
NA	Not available
Ns	No significant difference
PL	Posterior length
PCA	Principal Component Analysis
PCR	Polymerase Reaction Chain
PRIMER	Plymouth Routines In Multivariate Ecological Research
PS	Distance of the pallial sinus opening
PVM	Pallial line to ventral margin
PW	Posterior adductor scar width
SAS	Statistical Analysis of Software
SH	Shell height
SL	Shell length
SW	Shell Width
UL	Umbone length
$\mu\text{L}$	Microliters
mL	Mililiters
Ng	Nanogram
M	Meter
mm	Millimeter
$\mu\text{g/L}$	Microgram per liter
mg/L	Miligram per liter
g/L	Gram per liter
$\text{mg/m}^3$	Miligram per cubic mater
psu	Particle salinity unit
NTU	Nephelometric turbidity units
S/cm	Siemens per centimeter
$^{\circ}\text{C}$	Degree Celsius

# CHAPTER 1

## INTRODUCTION

### 1.1 Background

Veneridae (bivalve) is listed under mollusca phylum and as one of the important invertebrate that generates the source of animal protein for human consumption in the modern world. Veneridae is among of 82 families of bivalve and approximately 10,000 species, included of oyster, clams, scallop and mussels (Okutani, 2000; Wye, 2007). Veneridae generally found inhabit at marine area particularly intertidal area such as coastal and estuary. Favourable habitat condition can increase number and diversity of bivalve for instant, Southeast Asia turn out to be the utmost diversity bivalve faunas compared with other 29 regions around the world (Crame, 2000). Furthermore, they can be found distributed around the Western Central Pacific area (Poutiers, 1998). Slight changes to the environment condition will definitely affect distribution and diversity pattern of mollusc that occupied the habitat.

Mollusc diversity and behaviour highly correlated to nutrients cycle (Thakur *et al.*, 2012), physico-chemical variable (Khade and Mane 2012), and sediment properties (Suresh *et al.*, 2012). This environmental property discussed in Chapter 4 for Buntal estuary of Kuching Division and Kabong of Betong division which are suitable habitat area for *Meretrix* spp. Abundance and distribution of *Meretrix* spp. is corresponding to the habitat condition and different between *Meretrix* spp. East Asia region such as China, Japan and Korea are preference habitat for native *M. petechialis* and *M. lusoria* (Yamakawa and Imai, 2012). Species differences can be identified through the external feature such as shell morphology (Rosewater, 1961). However, two similar species that inhabit at the similar habitat area probably can cause confusion through morphology identification particularly when involve hybrid species. Therefore, extra approach such as molecular work will facilitate a lot for species identification. This molecular method has been widely practiced for many metazoan species included *Meretrix* spp. Torii *et al.* (2010) has used molecular work to study the phylogentic among *Meretrix* spp. from the Japan, Korea and China locality. Moreover, *Meretrix* of similar species from different locality or habitat areas will have different genetic sequences. Hence, both morphology and molecular work has significant influence in determination of *Meretrix* spp. and both methods have been discussed detail in Chapter 5 and 6 of this dissertation.

Habitat area such as coastal area and mangrove comprised of flora and fauna to sustain each other to form substantial ecosystem. Sarawak itself sustains large area of wetland approximately 1.24 million ha or 13% of the total land area (Page, 2011). This large area of wetland can support large number of fauna included *Meretrix* spp. with proper and adequate supplies of nutrient to help prolong heredity. Suitable of habitat condition such as salinity, temperature, nutrient and food are significantly important for the reproductive development of bivalve (Saxby, 2002; Chu and Kumar, 2008; Enriquez-Diaz *et al.*, 2009). Reproductive development of bivalve only can be determined based on histological procedure, Gonad Index (GI) and Condition Index (CI). Based on these

approaches, *M. lyrata* has been selected for reproductive development study due to its abundance found at the Buntal estuary. Detail on reproductive development study and environmental factor that influence the gonad development has been elaborated in Chapter 7.

## 1.2 Problem Statement

Mariculture on bivalve are widely applied at Western coast of Peninsular Malaysia. Most of bivalve cultures are *Anadara granosa*, *Paphia undulata*, *Perna viridis*, and *Crassostrea* spp. (Vakily, 1989; Poutiers, 1998). In Sarawak *A. granosa* and *Crassostrea* spp. are only culture in small scale. While *Meretrix* sp., *Placuna* spp., *Polymesoda* spp., *Pinna* spp. and *Modolus* spp. are collected from its natural environment to meet local market demand (Lovatelli, 1988).

Shellfish contribute to the source of Malaysian fish landing and on year 2009 shellfish landing was 23,746 tonne (Department of Fisheries Malaysia, 2013) (Table 1.1). Surf clam became the largest contributor during this year with 22,039 tonne. The lowest shellfish landing was recorded on year 2010 with 2,458 tonne. Total of shellfish landing was started to increase from year of 2011 to 2012 with 2,694 and 5,038 tonne respectively. However, the total shellfish landing decrease on year of 2013 with 4,910 tonne. High shellfish landing on year of 2009 may due to the high demand on surf clam. Therefore, this cause over exploitation on surf clam which was lead to significant decrease on total shellfish landing for the consequent year.

**Table 1.1. Shellfish landing in Malaysia from 2009 to 2013 in tone**

Species	2009	2010	2011	2012	2013
Oyster	11	14	13	10	9
Mussels	179	129	100	79	30
Sea-green mussel	11	0	1	1	2
Surf clam	22,039	623	686	1,501	1,548
Hard clam	176	132	76	100	52
Miscellaneous	1,330	1,560	1,818	3,347	3,269
<b>Total</b>	<b>23,746</b>	<b>2,458</b>	<b>2,694</b>	<b>5,038</b>	<b>4,910</b>

(Source: Department of Fisheries Malaysia, 2013)

In Sarawak, *Meretrix* spp. (Veneridae) are widely exploited as alternative meat source especially community that live close to coastal area. This bivalve was only sold at two divisions in Sarawak (Hamli *et al.*, 2012). However, there is no record for *Meretrix* spp. farming practice in Malaysia particularly Sarawak. Thus, fishermen need to collect it from its natural habitat in the mud flat area during the low tide.

## REFERENCES

- Abu Hena, M.K., Kohinoor, S.M.S., Siddique, M.A.M., Ismail, J., Idris, M.H. and S.M.N. Amin. 2012. Composition of macrobenthos in the Bakkhali Channel system. Cox's Bazar with note on soil parameter. *Journal of Fisheries and Aquatic Science*, 15(13): 641–646.
- Adjei-Boateng, D. and Wilson, J.G. 2013. Body condition and gametogenic cycle of *Galatea paradoxa* (Mollusca: Bivalvia) in the Volta River estuary, Ghana. *Estuarine Coastal and Shelf Science*, 132: 94–98.
- Akester, R.J. and Martel, A.L. 2000. Shell shape, dysodont tooth morphology, and hinge ligament thickness in the bay mussel *Mytilus trossulus* correlate with wave exposure. *Canadian Journal of Zoological*, 78: 240–253.
- Al-Barwani, S.M., Arshad, A., Nurul Amin. S.M.. 2011. *Reproductive Biology of Green Mussel Perna viridis (Linnaeus, 1758)*. Lambert Academic Publishing, 90p.
- Alongi, D.M. 1996. The dynamic of benthic nutrients pools and fluxes in tropical mangrove forests. *Journal of Marine Research*, 54: 123–148.
- Alunno-Bruscia M., Bourget, E. and Frechette, M. 2001. Shell allometry and length mass density relationship for *Mytilus edulis* in an experimental food regulated situation. *Marine Ecology Progress Series*, 219:177–188.
- APHA. 2005. Standard methods for the examination of water and wastewater. Second edition. American Public Health Association, Washington, DC.
- Araujo, R., Moreno, D. and Ramos, M.A. 1993. The Asiatic clam *Corbicula fluminea* (Muller 1774) (Bivalvia: Corbiculidae) in Europe. *American Malacological Bulletin*, 10(1): 39–49.
- Azimah, A.R., Idris, M.H., Abu Hena, M.K., Wong, S.K. and Arshad, A. 2013. Analysis of condition index in *Polymesoda expansa* (Mousson 1849). *Pakistan Journal of Biological Sciences*. 15(13): 629–634.
- Baba, K., Tada, M., Kawajiri, T. and Kuwahara, Y. 1999. Effects of temperature and salinity on spawning of brackish water bivalve *Corbicula japonica* in Lake Abashiri, Hokkaido, Japan. *Marine Ecology Progress Series*, 180: 213–221.
- Babaei, M.M., Sahafi, H.H., Ardalan, A.A., Ghaffari, H. and Abdollahi, R. 2010. Morphometric relationship and of weight and size of clam *Amiantis umbonella* L., 1818 (Bivalvia: Veneridae) In the Eastern Coasts of Bandar Abbas, Persian Gulf. *Advances in Environmental Biology*, 4(3): 376-382.
- Ballard, J.W.O. and Rand, D.M. 2005. The population biology of mitochondrial DNA and its phylogenetic implications. *Annual Review of Ecology, Evolution and Systematics*, 36: 621–642.



- Bantoto, V. and Ilano, A. 2012. The reproductive biology of *Lutraria philippinarum* (Veneroidea: Mactridae) and its fishery in the Philippines. *International Journal of Tropical Biology*, 60(4): 1807–1818.
- Baron, J. and Clavier, J. 1992. Effects of environmental factors on the distribution of the edibles bivalves *Atactodea striata*, *Gafrarium tumidum* and *Anadara scapha* on the coast of New Caledonia (SW Pacific). *Aquatic Living Resource*, 5:107–114.
- Bayne, B.L. 1975. Reproductive in bivalve mollusc under environmental tress. In: *Physiological ecology of estuarine organisms* ed. F.J. Vernberg, pp. 259–77. University of South Carolina Press, Columbia, USA.
- Bayne, B.L. and Newell, R.C. 1983. Physiological energetic of marine mollusc. In: the mollusca, Vol, 4, Part 1, ed. A.S.M. Saleuddin and K.M. Wilbur, pp 407–515. Physiology.
- Belda, C.A. and Del Norte, A.G.C. 1988. Notes on the induced spawning and larval rearing of the Asian moon scallop, *Amusium pleuronectes* (Linne'), in the laboratory. *Aquaculture*, 72: 173–179.
- Beukema, J.J. and Meehan, B.W. 1985. Latitudinal variation in linear growth and other shell characteristic of *Macoma balthic*. *Marine Biology*, 90:27–33.
- Bouillon, S., Dahdouh-Guebas, F., Rao, A.V.V.S., Koedam, N. and Dehairs, F. 2003. Source of organic carbon in mangrove sediment: variability and possible ecological implications. *Hydrobiologia*, 495: 33–39.
- Bouyoucos, G.J. 1962. Hydrometer method improved for making particle size analysis of soils. *Agronomy Journal*, 54:464–465.
- Bromham, L., Eyre-Walker, A., Smith, N.H. and Smith, J.M. 2003. Mitochondrial Steve: Paternal inheritance of mitochondria in humans. *TRENDS in Ecology and Evolution*, 18(1): 2–4.
- Broom, M. J. 1985. The biology and culture of marine bivalves molluscs of genus *Anadara*. ICLARM Studies and Reviews, 12:37.
- Brown, W.M., Prager, E.M., Wang, A. and Wilson, A.C. 1982. Mitochondrial DNA sequences of primates: tempo and mode evolution. *Journal of Molecular Evolution*, 18: 225–239.
- Camacho-Mondragon, M., Arellano-Martinez, M. and Ceballos-Vazquez, B.P. 2012. Particular features of gonadal maturation and size at maturity in *Atrina maura* (Bivalvia: Pinnidae). *Scintia Marina*, 76(3): 539–548.
- Carmichael, R.H., Shriver, A.C. and Valiela, V. 2004. Changes in shell and soft tissue growth, tissue composition and survival of quahogs, *Mercenaria mercenaria*, and softshell clam, *Mya arenaria*, in response to eutropic-driven changes in food supply and habitat. *Journal of Experimental Marine Biology and Ecology*, 313: 75–104.



- Carlucci, A.F. and MacNally, P.M. 1960. Nitrification by marine bacteria in low in low concentration of substrate and oxygen. *Limnology and Oceanography*, 19: 136–139.
- Carpenter, K.E. 1998. An Introduction to the Oceanography, Geology, Biogeography, and Fisheries of the Tropical and Subtropical Western and Central Pacific. In *The living marine resources of the Western Central Pacific: FAO species identification guide for fishery purpose Volume 1*. ed. K.E. Carpenter and V.H. Niem, pp.1–17. Rome, Italy: Food and Agriculture Organization of the United Nation.
- Caterino, M.S., Cho, S. and Sperling, F.A.H. 2000. The current state of insect molecular systematics: A thriving tower of Babel. *Annual Review of Entomology*, 45: 1–54.
- Ceballos-Vazquez, B.P., M. Arellani-Martinez, F. Garcia-Dominguez and M. Villalejo-Fuerte, M. 2000. Reproductive cycle of *rugosa* pen shell *Pinna rugosa*, sowebey, 1835 (Mollusca: Bivalvia) from Bahia Concepcion, Gulf of California and its relation to temperature and photoperiod. *Journal of shellfish Research*. 19(1): 95-99.
- Celekli, A., Ozturk, B. and Kapi, M. 2014. Relationship between phytoplankton composition and environmental variables in an artificial pond. *Algal Research*, 5: 37–41.
- Chanley, P. and Andrews, J.D. 1971. Aids for identification of bivalve larvae of Virginia. *Malacologia*, 11(1): 45–119.
- Charnov, E.L. and Bull, J.J. 1989. Non-fisherian sex-ratios with sex change and environmental sex determination. *Letters to Nature*, 338: 148–149.
- Chen, J., Li, Q., Kong, L. and Zheng, X. 2011a. Molecular phylogeny of venus clams (Mollusca, Bivalvia, Veneridae) with emphasis on the systematic position of taxa along the coast of mainland China. *Zoologica Scripta*, 40(3), 260–271.
- Chen, J., Li, Q., Kong, L. and Yu, H. 2011b. How DNA barcodes complement taxonomy and explore species diversity: The case study of a poorly understood marine fauna. *Public Library of Science*, 6(6): E21326
- Chen, A., Li, Z. and Feng, G. 2009. Phylogenetic relationships of genus *Meretrix* (Mollusca: Veneridae) base on mitochondrial COI gene sequences. *Zoological Research*, 30(3): 233–239.
- Cheng, H.L., Zhou, M.C., Peng, Y.X., Meng, X.P., Dong, Z.G. and Shen, X. 2011. Direct Submission. Jiangsu Key Laboratory of Marine Biotechnology, HuaiHai Institute of Technology China.
- Chu, C.T. and M.S. Kumar. 2008. *Clam (Meretrix lyrata) hatchery manual*. Ministry of Agriculture and Rural Development, Australian Government.

- Chung, E. 2007. Oogenesis and sexual maturation in *Meretrix lusoria* (Roding 1798) (Bivalvia: Veneridae) in Western Korea. *Journal of Shellfish Research*, 26(1): 71–80.
- Clarke, K.R., Gorley, R.N., 2001. *Primer Version 5. Primer-E, Plymouth, UK.*
- Claxton, W.T., Wilson, A.B., Mackie, G.L. and Boulding, E.G. 1998. A genetic and morphological comparison of shallow and deep water populations of the introduced dreissenid bivalve *Dreissena bugensis*. *Canadian Journal of Zoology*, 76:1269–1276.
- Claudi, R. and Mackie, G.L. 1994. *Practical manual for zebra mussel monitoring and control*. Lewis Publishers, Boca Raton.
- Costa, F.D., Darriba, S. and Martinez-Patino. 2008. Embryonic and larval development of *Ensis Arcuatus* (Jeffreys, 1865) (Bivalvia: Pharidae). *Journal of Molluscan Studies*, 74: 103–109.
- Crame, J.A. 2000. Evolution of taxonomic diversity gradients in the marine realm: Evidence from the composition of recent bivalves faunas. *Paleobiology*, 26(2): 188–214.
- Davenport, J. and Chen, X. 1987. A Comparison of methods for the assessment of condition in mussel (*Mytilus edlis* L.). *Journal of Molluscan Studies*, 53: 293–297.
- Davenport, J., Smith R.J.J.W. and Packer, M. 2000. Mussels *Mytilus edulis*, significant consumers and destroyers of mezooplankton. *Marine Ecology Program Series*, 198: 131–137.
- Davy, F B. and Graham, M. ed. 1982. Proceedings of Workshop held in Singapore. Ottawa: Bivalve culture in Asia and the Pacific.
- Department of Fisheries Malaysia. 2013. Landings of Marine fish by Month and state 2011. Retrieved 20 January from [http:// www.dof.gov.my](http://www.dof.gov.my).
- Dharmaraj, S., Shanmugasundaram, K. and Suja, C.P. 2004. Larval rearing and spat production of the windowpane shell *Placuna placenta*. *Aquaculture Asia*, 9(2): 20–28.
- Dobson, M. and Frid, C. 2009. *Ecology of aquatic systems*. Oxford University Press.
- Dridi, S. Romdhane, M.S. and Ecafsi, M. 2007. Seasonal variation in weight and biochemical composition of the Pacific oyster, *Crassostrea gigas* in relation to the gametogenic cycle and environmental conditions of the Bizert lagoon, Tunisia. *Aquaculture*, 263: 238–248.
- Duinker, A., Haland, L. and Mortensen, S. 2008. Gonad development and spawning in one and two year old mussels (*Mytilus edulis*) from Western Norway. *Journal of Marine Biological Association of the United Kingdom*, 1–9.

- Durve, V.S. 1964. Preliminary observations on the seasonal gonadal changes and spawning in the clam *Meretrix casta* (Chemnitz) from the Fish Farm. *Marine Journal of the Marine Biological of India Association*, 6(2): 241–248.
- Edgar, R.C. 2004. MUSCLE: A multiple sequence alignment method with reduced time and space complexity. *BMC Bioinformatics*, 5(113): 1–19.
- Edwards, P. 2000. Aquaculture, poverty impacts and livelihoods, *Overseas Development Institute*, 56: 1–4.
- Endler, J.A. 1986. *Natural selection in the wild*. Princeton University Press, New Jersey.
- Enriquez-Diaz, M., Pouvreau, S., Chavez-Villalba, J. and Le Penec, M. 2009. Gametogenesis, reproductive investment, and spawning behavior of the Pacific giant oyster *Crassostrea gigas*: evidence of an environment-dependent strategy. *Aquaculture International*, 17(5): 491–506.
- FAO. 2001. 1999. *Fisheries Statistics: Aquaculture Production*. Vol. 88/2.FAO, Rome.
- Feinberg, H.S. 2003. *Shells*. Simon and Schuster Inc.
- Felsenstein J. 1985. Confidence limits on phylogenies: An approach using the bootstrap. *Evolution*, 39:783–791.
- Folmer, O., Black, M., Hoeh, W., Lutz, R. and Vrijenhoek, R. 1994. DNA primers for amplification of mitochondrial cytochrome C oxidase subunit I from diverse metazoan invertebrates. *Molecular Marine Biology and Biotechnology*, 3(5): 294–299.
- Franz, D.R. 1993. Allometry of shell and body weight in relation to shore level in the intertidal bivalves *Geukensia demissa* (Bivalvia: Mytilidae). *Journal of Experimental Marine Biology and Ecology*, 174:193–207.
- Fuiman, L.A., Gage, J.D. and Lamont, P.A. 1999. Shell morphometry of deep sea protobranch bivalve *Ledella pustulosa* in the Rockall Trough, North- East Atlantic. *Journal of the Marine Biological Association of the United Kingdom*, 79:661–671.
- Gan, H.H., Perlow, R.A., Roy, S., Ko, J., Wu, M., Huang, J., Yan, S., Nicoletta, A., Vafai, J., Sun, D., Wang, L., Noah, J.E. Pasquali, S. and Schlick, T. 2002. Analysis of protein sequence/structure similarity relationships. *Biophysical Journal*, 83: 2781–2791.
- Garibaldi, L. 1996. *List of Animal Species used in Aquaculture*. FAO Fisheries Circular No. 914 FIRI/C914.
- Garrison, L.P. and Morgan, J.A. 1999. Abundance and vertical distribution of drifting post-larval *Macoma* spp. (Bivalvia: Telinidea) in the York River, Virginia, USA. *Marine Ecology Progress Series*, 183: 175–185.

- Gribben, P.E. 2005. Gametogenic development and spawning of the razor clam *Zenatia acinaces* in northeastern New Zealand. *New Zealand Journal of Marine and Freshwater Research*, 39(6): 1287–1296.
- Gibbs, P.E. 1978. Macrofauna of the intertidal sand flats on Low Wooded Islands Northern Great Barrier Reef. *Philosophical Transaction of the Royal Society*, 283:81-97.
- Gieskes, J.M. 1969. Effect of temperature on the pH of the seawater. *Limnology and Oceanography*, 14: 679–685.
- Globefish-FAO. 2007. *Bivalves-Commodity Update*. Globefish (FAO), Rome, Italy.
- Gonneea, M.E., Paytan, A. and Herrera-Silveira, J.A. 2004. Tracing organic matter sources and carbon burial in mangrove sediments over past 160 years. *Estuarine Coastal and Shelf Science*, 61: 211–227.
- Gosling, E. 2003. *Bivalve Molluscs: biology, ecology and culture*. Fishing News Books.
- Guo, X., Ford, S.E. and Zhang, F. 1999. Molluscan aquaculture in China. *Journal Shellfish Research*, 18: 19–31.
- Guo, X., Liu, S. and Liu, Y. 2006. Evidence for recombination of mitochondrial DNA in triploid Crucian Carp. *Genetic*, 172: 1745–1749.
- Haley, L.E. 1979. Genetic of sex determination in the American oyster. *Proceeding of the National Shellfisheries Association*, 69: 54–57.
- Hamli, H. and Idris M.H. 2014. *Biology of edible bivalve and gastropod from Sarawak Malaysia*. Lap Lambert Academic Publishing.
- Hamli, H., Idris, M.H., Abu Hena, M.K. and Wong, S.K. 2012. Taxonomic study of edible bivalve from selected division of Sarawak. *International Journal of Zoological Research*, 8(1): 52–58.
- Hamaguchi, M., Sasaki, M. and Higano, J. 2001. Direct Submission. Fisheries Research Institute of Seto Inland Sea, Japan.
- Harding, J.M. 2007. Northern quahog (=hard clam) *Mercenaria mercenaria* age at length relationships and growth pattern in the York River, Virginia 1954 to 1970. *Journal of Shellfish Research*, 26(1): 101–107.
- Hare, M.P., Palumbi, S.R. and Butman, C.A. 2000. Single-step species identification of bivalve larvae using multiplex polymerase chain reaction. Springer-Verlag, *Marine Biology*, 137: 953–961.
- Hawkins, A.J.S. and Bayne, B.L. 1992. Physiological interrelations and the regulation of production. In: *The mussel Mytilus: ecology, physiology, genetic and culture*. Ed. Gosling E. pp. 171–212. Elsevier, Amsterdam.

- Hawver, G.A. and Bassuk, N.L. 2007. Soil: The key to successful established of urban vegetation. Springer, Netherlands.
- Hayes, T.B., Collins, A., Lee, M., Mendoza, M., Norlega, N., Stuart, A.A. and Vonk, A. 2002. Hermaphroditic, demasculinized frogs after exposure to the herbicide atrazine at low ecologically relevant doses. *PNAS*, 99(8): 5476–5480.
- He, C.B., Cong, L.L., Ge, L.L., Liu, W.D., Zhou, Z.C. and Gao, X.G. 2008. ALFP analysis of cultured and wild hard clam (*Meretrix meretrix*) population. *Journal of Fisheries Science of China*, 15(2): 204–214.
- He, C., Wang, J., Gao, X., Liu, W., Fu, L., Li, Y. and Bao, X. 2009. Direct Submission. Liaoning key laboratory of marine fishery molecular biology, Liaoning Ocean and Fisheries Science Research Institute, China.
- Helm, M.M., N. Bourne and A. Lovatelli, A. 2004. *Hatchery culture of bivalves: A practical manual*. FAO Fisheries Technical Paper, Rome.
- Hinch, S.G. and Bailey, R.C. 1988. Within- and among-lake variation in shell morphology of the freshwater clam *Elliptio complanata* (Bivalvia: Unionidae) from south-central Ontario lakes. *Hydrobiologia* 157: 27–32.
- Humason, G.L. 1972. Animal tissue techniques. 4<sup>th</sup> edition. W.H. Freeman Co., San Francisco, CA.
- Hur, Y.B., Bae, J.H. and Hur, S.B. 2005. Comparison of development and larval growth of four venerid clam. *Journal of the World Aquaculture Society*, 36(2): 179–187.
- Idris, M.H., Arshad, A., Japar Sidik, B., Mazlan, A.G. and Daud, S.K. 2009. Morphological characteristics of *Pinna bicolor* Gmelin and *Pinna deltodes* Menke from seagrass bed of Sungai Pulai, Johor, Peninsular Malaysia. *Sains Malaysiana*, 38(3): 333–339.
- Japar Sidik, B. Lim, L.H., Muta Harah, Z., Arshad, A. and Hisao, O. 2008. Laboratory culture of the seagrass, *Halophila ovalis* (R.Br.) Hooker f. *Marine Research in Indonesia* 33(1): 1–6.
- Japar Sidik, B., Muta Harah Z. and Arshad, A. 2006. Distribution and significance of seagrass ecosystem in Malaysia. *Aquatic Ecosystem Health and Management*, 9(2): 203–214.
- Jarne, P. and Charlesworth, D. 1993. The evolution of the selfing rate in functionally hermaphrodite plants and animals. *Annual Review of and Ecology Systematics*. 24: 441–466.
- Jayabal, R. and Kalyani, M. 1987. Reproductive cycle of the estuarine bivalve, *Meretrix meretrix* (Linn) of the Vellar Estuary. *Indian Journal of Fisheries*, 34(2): 229–232.

- Jorgensen, C.B. 1990. *Bivalve filter feeding. Hydrodynamics, bioenergetics, physiology and ecology*. Olsen and Olsen, Fredensborg, Denmark.
- Kang, J.H., Kim, B.H., Park, J.Y., Lee, J.M., Jeong, J.E., Lee, J.S., Ko, H.S. and Lee, Y.S. 2012. Novel microsatellite markers of *Meretrix petechialis* and cross-species amplification in related taxa (Bivalvia: Veneroida). *International Journal of Molecular Sciences*, 13: 15942 – 15954.
- Khade, S.N. and Mane, U.H. 2012. Diversity of bivalve and gastropod, molluscs of some localities from Raigad district, Maharashtra, west coast of India. *Recent Research in Science and Technology*, 4(10): 43–48.
- Kitamura H, Ishitani, H., Kuge, Y. and Nakamoto, M. 1982 Determination of nitrate in freshwater and sea Water by a hidrazine reduction method. *Suishitu Odaku Kenkyu* 5: 35–42.
- Kimura, T., Nakano, T., Yamaguchi, T., Sato, M., Ogawa, T., Muramoto, K., Yokoyama, T. Kan-no, N., Nagahisa, E., Frank, J. and Manfred, K.G. 2005. Complementary DNA cloning and molecular evolution of opine dehydrogenases in some marine invertebrates. *Marine Biotechnology*, 6: 493–502.
- Kovitvadhi, S., Kovitvadhi, U., Sawangwong, P., Thongpan, A. and Machado, J. 2006. Optimization of diet and culture environment for larvae and juvenile freshwater Pearl Mussels, *Hyriopsis (Limnoscapha) myersiana* Lea, 1856. *Invertebrate Reproduction and Development*, 49: 61–70.
- Kovitvadhi, S., Kovitvadhi, U., Swangwong, P., Trisaranuwatana, P. and Machado, J. 2009. Morphometric relationship of weight and size of cultured freshwater pearl mussel, *Hyriopsis (Limnoscapha) myersiana*, under laboratory conditions and Earthen Pond Phases. *Aquaculture International*, 17: 57–67.
- Laing, I. and B.E. Spencer. 2006. Bivalve cultivation: Criteria for selecting a site. Science Series Technical Report no. 136.
- Lajtner, J., Marusic, Z., Klobucar, G.I.V., Maguire, I. and Erben, R. 2004. Comparative shell morphology of the zebra mussel, *Dreissena polymorpha* in the Drava River (Croatia). *Biologia*, 59:595–600.
- Laxmilatha, P. 2013. Population dynamic of the edible clam *Meretrix casta* (Chemnitz) (International Union for Conservation of Nature Status: Vulnerable) from two estuaries of North Karala south west coast of India. *Global Journal of Fisheries and Aquaculture*, 1(1): 031–039.
- Laxmilatha, P., Rao, G.S., Patnaik, P., Rao, T.N., Rao, M.P. and Dash, B. 2011. Potential for the hatchery production of spat of the green mussel *Perna viridis* Linnaeus (1758). *Aquaculture* ,312: 88–94.
- Layton, K.K.S., Hebert, P.D.N. and Martel, A.L. 2010. Direct Submission. Biodiversity Institute of Ontario, University of Guelph, Canada.



- Lee, A-C., Lee, Y-C. and Chin, T-S. 2012. Effects of low dissolved oxygen on the digging behaviour and metabolism of the hard clam (*Meretrix lusoria*). *Aquaculture Research*, 43: 1–13.
- Lewis, D.E. and Cerrato, R.M. 1997. Growth uncoupling and the relationship between shell growth and metabolism in the soft shell clam *Mya arenaria*. *Marine Ecology Progress Series*, 158: 177–189.
- Leal, J.H. 2002. Bivalves. In *The Living Marine Resources of the Western Central Atlantic: FAO Species Identification Guide for Fishery Purpose Volume 1*, ed. K.E. Carpenter, pp. 25–98. Rome, Italy: Food and Agriculture Organization of the United Nations.
- Lee, J.S., Ku, K., Kim, H., Park, J.S., Park, J.J., Shin, Y.K. and Jeon, M.A. 2012. Indirect evidence on sex reversal with sex ratio of *Tegillarca granosa* (Bivalvia: Arcidae) and *Ruditapes philippinarum* (Bivalvia: Veneridae). *Developmental and Reproductive*, 16(3): 177–183.
- Lin, Z-H., Dong, Y-H., Li, N., Lu, R-M., Xiao, G-Q., Chai, X-L., Liu, B-Z., Sun, C-S., Bao, Z-M. and Hu, J-J. 2007. The genetic structure and diversity analysis of different geographic populations of *Meretrix meretrix* using different morphological parameters and AFLP markers. *Oceanologia et Limnologia Sinica*, 39: 245–251.
- Liu, Z.J. and Cordes, J.F. 2004. DNA markers technologies and their application in aquaculture genetics. *Aquaculture*, 238: 1–37.
- Li Xuan, S., Duc, T.T. and Kim, C.D. 2011. Study on growth's role of hard clam (*Meretrix lyrata*) in Bach Dang Estuary, Viet Nam. *Environment and Natural Research*, 1(1): 139–151.
- Lovatelli, A. 1988. Status of mollusc culture in selected Asian countries. Food and Agriculture Organization. Retrieved 13 July 2014 from <http://www.fao.org/docrep/field/003/AB718E/AB718E00.htm>
- Low, K.C. *Application of nowcasting techniques towards strengthening national warning capabilities on hydrometeorological and landslides hazards*. Paper presented at Public Weather Services Workshop on Warning of Real-Time Hazards by Using Nowcasting Technology. Sydney, Australia, 9 to 13 October 2006.
- Lu, X. Wang, H., Liu, B. and Xiang, J. 2011a. An effective method for parentage determination of the clam (*Meretrix meretrix*) base on SSR and COI markers. *Aquaculture*, 318: 223–228.
- Lu X., Wang, H.X., Dai, P. and Liu, B.Z. 2011b. Characterization of EST-SSR and genomic-SSR markers in the clam, *Meretrix meretrix*. *Conservation Genetics Resources*. 3:655–658.
- Lucas, A. and Beninger, P.G. 1985. The use of physiological condition indices in marine bivalve aquaculture. *Aquaculture*, 44: 187–200.

- MacLachlan, A. and Erasmus, T. 1974. Temperature tolerance and osmoregulation in some estuarine bivalves. *African Zoology*, 9(1): 1–13.
- Manahan, D.T. Wright, S.H., Stephens, G.C. and Rice, M.A. 1982. Transport of dissolved amino acid by the mussel, *Mytilus edulis*: demonstration of net uptake from natural seawater. *Science N.Y.*, 215: 1253–1255.
- Marsden, I.D. and Pilkington, R.M. 1995 Spatial and temporal variations in the condition of *Austrovenus stutchburyi* Finlay, 1927 (Bivalvia: Veneridae) from the Avon-Heathcote Estuary, Christchurch. *New Zealand Natural Sciences*, 22: 57–67.
- Mass, P.A.Y., O'Mullan, G.D., Lutz, R.A. and Vrijenhoek, R.C. 1999. Genetic and Morphometric Characterization of Mussels (Bivalvia: Mytilidae) from Mid-Atlantic Hydrothermal Vents. *Biological Bulletin*, 196: 265–272.
- Massey, R.C. and Buckling A. 2002. Environmental regulation of mutation rates at specific sites. *TRENDS in Microbiology*, DOI: 10.1016/S0966-842X(02)02475-7.
- Marcado-Silva, N. 2005. Condition index of the eastern oyster, *Crassostrea virginica*, (Gmelin, 1791) in Sapelo Island Georgia—Effect of site, position on bed and pea crab parasitism. *Journal of Shellfish Research*, 24(1): 121–126.
- Maretto, F, Reffo, E., Baracaccia, G. and Mantovani, R. *Genetic traceability of clam species using mitochondrial markers*. Proceeding of the XIIX Italian Society of Agricultural Genetics Annual Congress Potenza, Italy 12/15, September 2005.
- McLusky, D. 1973. The effect of temperature on the oxygen consumption and filtration rate of Chlamys (*Aequipecten*) *opercularis* (L.) (Bivalvia). *Ophelia*, 10: 114–54.
- Mikulich, L.V. and Tsikhon-Lukanina, E.A. 1981. Food composition of the yesso scallop. *Oceanology*, 21: 633–635.
- Mishmar, D., Ruiz-Pesini, E., Golik, P., Macaulay, V., Clark, A.G., Hosseini, S., Brandon, M., Easley, K., Chen, E., Brown, M.D., Sukernik, R.I. and Olckers, A. 2003. Natural selection shaped regional mtDNA variation in human. *PNAS*, 100(1): 171–176.
- Mohamat-Yusuff, F., Zulkifli, Z.S., Ismail, A., Yusof, M.K. and Tsuuguo, O. 2012. Field survey and spatial distribution of tropical Neogastropod, *Thais* spp., along Malaysian coastal area. *Acta Biologica Malaysiana*, 1(1): 9–17.
- Mohan, M.V., Damodaran, R., and Salih, K.Y.M. 1984. Allometric studies in the clam *Meretrix casta* Chemnitz. *Mahasagar Bulletin of the National Institute of Oceanography*, 17(2): 119–1923.
- Moneva, C.S., Baquiano, P.M., Blasco, J., Borlaza, K.M., Burias, D.M., Flores, K., Fuentes, G.R., Pancho, A.G. and Sanchez, R.R. *Comparative morphological descriptions of interior shell patterns of the venerid. Bivalves: Meretrix lyrata, Mercenaria mercenaria, and Venerupis philippinarum using landmark-based*



*geometric morphometric analysis*. Paper presented at The Fourth International Congress on Interdisciplinary Research and Development, Thailand. May 2014.

- Muhammad, T.T.S. and Nor, M.S.A. 2004. Elektroforesis Gel .In *Penanda DNA untuk kajian kepelbagaian genetik*, ed A.S. Othman, pp. 36–43. Universiti Sains Malaysia, Pulau Pinang.
- Muta Harah, Z. and Japar Sidik, B. 2013. Occurrence and distribution of seagrass in waters of Perhentian Island Archipelago, Malaysia. *Journal of Fisheries and Aquatic Science*, 8(3): 441–451.
- Mutshinda, C.M., Finkel, Z.V. and Irwin, A.J. 2013. Which environmental factors control phytoplankton populations? A Bayesian variable selection approach. *Ecological Modeling*, 269: 1–8.
- Najafpour, G.D. 2007. Dissolved oxygen measurement and mixing. *Biochemical Engineering and Biotechnology*, 14–21.
- Nakamura, Y., Nakano, T., Yurimoto, T., Maeno, Y., Koizumi, T. and Tamali, A. 2010. Reproductive cycle of the venerid clam *Meretrix lusoria* in Ariake Sound and Tokyo Bay, Japan. *Fisheries Science*, 76: 931–941.
- Narasimham, K.A., Muthiah, P., Sundararajan, D. and Vaithinathan, N. 1988. Biology of the great clam, *Meretrix meretrix* in the Korampallam Creek, Tuticorin. *Indian Journal of Fisheries*, 35(4): 288–293.
- Narayanan, B. and P. Sivadas. 1986. Studies on the intertidal macrofauna of the sandy beach at Kavaratti Atoll (Lakshadweep). *Mahasagar Bulletin National Institute Oceanography*, 19:84–98.
- Neo, M.L., Todd, P.A., Chou, L.M. and Teo, S.L-M. 2011. Spawning induction and larval development in the fluted Giant Clam, *Tridacna Squamosa* (Bivalvia: Tridacnidae). *Nature in Singapore*, 4: 157–161.
- Newell, C.R. and Hidu, H. 1982. The effects of sediment type on growth rate and shell allometry in the soft shelled clam *Mya arenaria* (L). *Journal of Experimental Marine Biology and Ecology*, 65: 285–295.
- Nicholas, F.H. 1985. Increased benthic grazing: an alternative explanation for low phytoplankton biomass in northern San Francisco Bay during the 1976–1977 drought. *Estuarine, Coastal and Shelf Science*, 21: 379–388.
- Nicolini, M.H. and Penry, D.L. 2000. Spawning, fertilization and larval development of *Potamocorbula amurensis* (Mollusca: Bivalvia) from San Francisco Bay, California. *Pacific Science*, 54(4): 377–388.
- Okumus, I. and Ciftci, Y. 2003. Fish population genetic and molecular markers: II- Molecular markers and their applications in fisheries and aquaculture. *Turkish Journal of Fisheries and Aquatic Sciences*, 3: 51–79.
- Okutani, T. 2000. *Marine mollusks in Japan*. Tokai University Press.

- Othman, A.S. 2004. Teknik-teknik asas penanda DNA. In *Penanda DNA untuk kajian kepelbagaian genetik*, ed A.S. Othman, pp. 44–60. Universiti Sains Malaysia, Pulau Pinang.
- Overton, L.C. and Rhoads, D. 2004. Molecular phylogenetic relationships based on mitochondrial and nuclear gene sequences for the *Todies* (Todus, Todidae) of the Caribbean. *Molecular Phylogenetic and Evolution*, 32: 524–538.
- Page, S., 2011. Biodiversity information on peat swamp forest in S.E. Asia. Retrieved 9 December from <http://www.strapeat.alterra.nl/download/6%20biodiversity%20information.pdf>
- Parsons, T.R., Yoshiaki, M. and Lalli, C.M. 1984. *A manual of chemical and biological methods for seawater analysis*. Pergamon Press.
- Passamonti, M., Boore, J.L. and Scali, V. 2003. Molecular evolution and recombination in gender-associated mitochondrial DNAs of the Manila clam *Tapes philippinarum*. *Genetics*, 164: 603–611.
- Pathansali, D. 1966. Notes on the biology cockle *Anadara granosa* L. *Proceeding. Indo-Pacific Fish Council*, 11:84-98.
- Peharda, M., Mladineo, I., Bolotin, J. Kekez, L. and Skaramuca, B. 2006. The reproductive cycle and potential protandric development of the Noah's Ark shell, *Arca noae* L.: Implications for aquaculture. *Aquaculture*, 252: 317–327.
- Perez Camacho, A. Delgado, M., Fernandez-Reiriz, M.J. and Labarta, U. 2003. Energy balance, gonad development and biochemical composition in the clam *Ruditapes decussatus*. *Marine Ecology Progress Series*, 258: 133–145.
- Philippart, C.J.M., Van Aken, H.M., Beukema, J.J., Bos, O.G., Cadee, G.C. and Dekker, R. 2003. Climate-related changes in recruitment of the bivalve *Macoma balthica*. *Limnology Oceanography*, 48(6): 2171–2185.
- Poutiers, M. 1998. Bivalves. In *The living marine resources of the Western Central Pacific: FAO species identification guide for fishery purpose Volume 1*. ed. K.E. Carpenter and V.H. Niem, pp.123–362. Rome, Italy: Food and Agriculture Organization of the United Nations,
- Pronker, A.E., Nevejan, N.M., Peene, F., Geijssen, P. and Sorgeloos, P. 2008. Hatchery broodstock conditioning of the blue mussel *Mytilus edulis* (Linnaeus 1758). Part 1. Impact of different micro-algae mixtures on broodstock performance. *Aquaculture International*, 16: 297–307.
- Purchon, R.D. and Purchon, D.E.A. 1981. The marine shelled mollusca of West Malaysia and Singapore. I. General introduction and an account of collections. *Journal of Molluscan Studies*, 47:290–312.
- Quayle, D.B. and Newkirk, G.F. 1989. *Farming bivalve mollusc: Methods for study and development*. The World Aquaculture Society.

- Ranjith, K.R., Vijayan, K.K., Reynold, P., Thomas, P.C., Gopalakrishnan, A. and Mohamed, K.S. 2012. Direct Submission. Marine Biotechnology Division, Central Marine Fisheries Research Institute. India.
- Rao, G.S. 1988. Biology of *Meretrix casta* (Chemnitz) and *Paphia malabarica* (Chemnitz) from Mulky Estuary, Dakshina Kannada. *National Seminar on Shellfish Resources and Farming*, 148–153.
- Reise, K. 1983. Biotic enrichment of intertidal sediments aggregates of deposit-feeding bivalve *Macoma balthica*. *Marine Ecology Progress Series*, 12: 229–236.
- Ren, J., Shen, X., Sun, M., Jiang, F., Yu, Y., Chi, Z. and Liu, B. 2009. The complete mitochondrial genome of the clam *Meretrix petechialis* (Mollusca: Bivalvia: Veneridae). *Mitochondrial DNA*, 20(4): 78–87.
- Rengarajan, K., Cristol, S.M., Mehta, M. and Nickerson, J.M. 2002. Quantifying DNA concentrations using fluometry: A comparison of fluorophores. *Molecular vision*, 8: 416–421.
- Reverol, Y.M., J.G. Severeyn, Y.G. and Severeyn, H.J. 2004. Embryonary and larval development of the marine clam *Tivela mactroides* (Bivalvia: Veneridae) in Zulia State, Vanuezela. *Revista de Biologia Tropical*, 52(4): 1–6.
- Rizzo, J. and Rouchka, E.C. 2007. *Review of hylogenetic tree construction*. University of Louisville Bioinformatics Laboratory Technical Report Series, 1: 1–7.
- Rosewater, J. 1961. The family Pinnidae in the Indo-Pacific. *Indo-Pacific Mollusca*, 1(4): 175-226.
- Saucedo, P.E., Ocampo, L., Monteforte, M. and Bervera, H. 2004. Effect of temperature on oxygen consumption and ammonia excretion in the Calafia mother-of-pearl oyster, *Pinctada mazatlanica* (Hanley, 1856). *Aquaculture*, 229: 377–387.
- Sarawak Marine Department. 2013. *2013 Sarawak hourly and high and low tide tables (Including standard ports of Sabah)*. The Director of Marine Sarawak, Malaysia.
- Sarawak Marine Department. 2014. *2014 Sarawak hourly and high and low tide tables (Including standard ports of Sabah)*. The Director of Marine Sarawak, Malaysia.
- SAS® Institute Inc. 2004. *SAS ODBC Driver 9.1: User's Guide and Programmer's Reference*. Cary, NC: SAS Institute Inc.
- Sastry, A.N. and Blake, N.J. 1971. Regulation of gonad development in the bay scallop, *Aequipecten irradians* Lamarck. *The Biological Bulletin*, 140: 274–282.
- Sawant, P.P. and Mohite, S.A. 2013. Study of proximate composition of *Meretrix meretrix* (Linnaeus, 1758) of the Ratnagiri coast, Maharashtra, India. *Biosciences Biotechnology Research Asia*, 10(1): 311–317.

- Saxby, S.A. 2002. A review of food availability, sea water characteristics and bivalve growth performance at coastal culture site in temperate and warm temperate regions of the world. *Fisheries Research Report* No. 132, pp 44.
- Scheltema, A. 1983. *Pinna deltodes* Menke newly described and differentiated from *P. Bicolor* Gmelin (Bivalvia, Ptreioida). *Journal Malacological Society of Australia*, 6(1-2): 37-52.
- Scherrer, B. 1984. Biostatistique. Morin, Montreal Canada. pp.850.
- Seed, R. 1975. Reproduction in *Mytilus* (Mollusca: Bivalvia) in European waters. *Pubblicazioni Stazione Zoologica Napoli*. 39: 227-316.
- Seed, R. 1976. Ecology. In *Marine Mussels: Their Ecology and Physiology*. Ed. B.L. Bayne. pp. 13-65. Cambridge University Press: London, New York, Melbourne.
- Seike, Y., Oka, H., Mitamura, O., Okumura, M. Fujinaga, K. and Singa, Y. 2000. A pretreatment method for the determination of nitrate in brackish water and seawater based on the hydrazinium reduction technique. *Limnology*, 1: 129-132.
- Serdar, S. and Lok, A. 2009. Gametogenic cycle and biochemical composition of the transplanted carpet shell clam *Tapes decussatus* Linnaeus 1758 in Sufa (Homa) Lagoon, Izmir, Turkey. *Aquaculture*, 293: 81-88.
- Serdar, S., Lok, A., Kirtik, A., Acarh, S., Kucukdermenci, A., Guller, M. and Yigitkurt, S. 2010. Comparison of Gonadal Development of Carpet Shell Clam (*Tapes decussatus*, Linnaeus 1758) in Inside and Outside of Çakalburnu Lagoon, Izmir Bay. *Turkish Journal of Fisheries and Aquatic Sciences*, 10: 395-401.
- Siebers, D. and Winkler, A. 1984. Amino acid uptake by mussels, *Mytilus edulis*, from natural sea water in a flow-through system. *Helgolander Meeresunters*, 38: 189-199.
- Sophia, A.J.A. and Balasubramanian, T. 1992. Changes in the physical condition of *Meretrix casta* exposed to water-soluble fraction of refined and crude oil. *Achieves of Environmental Contamination and Toxicology*, 22: 471-474.
- Spencer, B.E. 2002. Molluscan shellfish farming. Fishing News Books. pp. 274.
- Sreenivasan, P.V. and Rao, K.S. 1991. Spawning and larval development and spat settlement of the clam *Meretrix casta* (Chemnitz) in the laboratory. *Indian Journal of Fisheries* 38(1): 1-8.
- Stanley, J.G. 1985. *Hard clam. Species profiles: Life histories and environmental requirement of coastal fisheries and invertebrates (mid-Atlantic)*. Research and development branch of the U.S.fish and wildlife service.
- Stevens, F.J., Kuemmel, C., Babnigg, G. and Collart, F.R. 2007. Efficient recognition of protein fold at low sequence identity by conservative application of Psi-BLAST: Application. *Journal of Molecular Recognition*, 18: 150-157.

- Stewart, M.G. and Bamford, D.R., 1975. Kinetics of alanine uptake by the gills of the soft-shelled clam *Mya arenaria*. *Comparative Biochemistry and Physiology*, 52A: 67–74.
- Subba Rao, N.V. 1993. Freshwater mollusc of India. In *Recent Advances in freshwater biology*, ed. K.S. Roa, Volume 2. pp. 187-202. New Delhi. Anmol Publication.
- Suhaila, J., Mohd Deni, S., Wan Zain, W.Z. and Jemain, A.A. 2010. Trends in Peninsular Malaysia rainfall data during the Southwest Monsoon and Northeast Monsoon Seasons: 1975–2004. *Sains Malaysiana*, 39(4): 533–542.
- Suja, N. and Muthiah, P. 2007. The reproductive biology of the baby clam, *Marcia opima*, from two geographically separated areas of India. *Aquaculture*, 273: 700–710.
- Sullivan, D.J. 2000. *Nutrients and suspended solids in surface waters of the upper Illinois River Basin in Illinois, Indiana, and Wisconsin, 1978–97*. U.S. Department of the Interior, U.S. Geological Survey: Middleton, Wisconsin.
- Sundaram K.S. and M. Syed Shafee. 1989. Salinity tolerance of some bivalves of Ennore Estuary. *Journal of the Marine Biological Association of India*. 31(1&2): 299–302.
- Suquet, M., Rimond, F., Cosson, J., Wilson-Leedy, J., Leburn, L., Queau, I., Mingant, C. and Fauvel, C. 2013. Effect of age and environment conditions on the movement characteristics of Pacific Oyster (*Crassostrea gigas*) trochophores. *Journal of Applied Ichthyology*, 29(5), 1145–1148.
- Suresh, M., Arularasan, S. and Ponnusamy, K. 2012. Distribution of molluscan fauna in the artificial mangroves of Pazhayar back water canal, Southeast Coast of India. *Advances in Applied Science Research*, 3(3): 1795–1798.
- Suwanjarat, J., Pituksalee, C. and Thongchai, S. 2009. Reproductive cycle of *Anadara granosa* at Pattani Bay and its relationship with metal concentrations in the sediments. *Songklanakarin Journal of Science and Technology*, 31(5): 471–479.
- Tamura K. 1992. Estimation of the number of nucleotide substitutions when there are strong transition-transversion and G + C-content biases. *Molecular Biology and Evolution*, 9:678–687.
- Tamura, K., Stecher, G., Peterson, D., Filipski, A. and Kumar S. 2013. MEGA6: Molecular evolutionary genetics analysis version 6.0. *Molecular Biology and Evolution*, 30: 2725–2729.
- Tang, B., Liu, B., Wang, G., Zhang, T. and Xiang, J. 2006. Effects of various algal diets and starvation on larval growth and survival of *Meretrix meretrix*. *Aquaculture*, 254: 526–533.
- Tedengren, M. and Kautsky, N. 1986. Comparative study of physiology and its probable effect on size in blue mussels (*Mytilus edulus* L.) from the North Sea and the Northern Baltic Proper. *Ophelia*, 25(3): 147–155.

- Thakur, S., Veragi, S.G., and Yeragi, S.S. 2012. *Population density and biomass of organisms in the mangrove region of Akshi Creek, Alibag Taluka, Raigad District, Maharashtra*. Paper presented at International Day for Biological Diversity, Marine Biodiversity. May 2012.
- Thanh, N.X. and Thung, D.C. 2014. The reproductive biology of Lyrate Asiatic hard clam (*Meretrix lyrata*) in the intertidal zone of Nam Dinh provinve. *Vietnam Journal of Marine Science and Technology*, 14(2) DOI: 10.15625/1859-3097/14/2/4483.
- Tirado, C., Salas, C. and Marquez, I. 2003. Reproduction of *Venus verrucosa* L., 1758 (Bivalvia: Veneridae) in the littoral of Málaga (southern Spain). *Fisheries Research*, 63: 437–445.
- Torii, H., Sato, S., Hamaguchi, M. and Henmi, Y. 2010. The comparison of shell morphology and genetic relationship between *Meretrix lusoria* and *Meretrix petechialis* in Japan and Korea. *Plankton and Benthos Research*, 5: 231–241.
- Touchette, B.W. and Burkholder, J.M. 2000. Review of nitrogen and phosphorus metabolism in seagrasses. *Journal of Marine Experimental Biology and Ecology*, 250: 133–167.
- Wallace, D.C. 1999. Mitochondrial disease in man and mouse. *Science*, 283: 1482–1488.
- Wang, G. and Dunbrack Jr. R.L. 2003. PISCES: A protein sequence culling server. *Bioinformatics Applications Note*, 19(12): 1589–1591.
- Wang, H., Huan, P., Lu, X. and Liu, B. 2011. Mining of EST-SSR markers in clam *Meretrix meretrix* larvae from 454 shotgun transcriptome. *Gene and Genetic Systems*, 86: 197–205.
- Wang, H., Zhang, S. and Liu, B. 2011a. Direct Submission. Key laboratory of the experimental marine biology, Institute of Oceanology, Chinese Academy of Sciences, China.
- Wang, H., Zhang, S., Xiao, G. and Liu, B. 2011b. Complete mtDNA of the *Meretrix lamarckii* (Bivalvia: Veneridae) and molecular identification of suspected *M. lamarckii* based on the whole mitochondrial genome. *Marine Genomics*, 4(4): 263–271.
- Wang, H., Zhang, S., Li, Y. and Liu, B. 2010. Complete mtDNA of *Meretrix lusoria* (Bivalvia: Veneridae) reveals the presence of an atp8 gene, length variation and heteroplasmy in the control region. *Comparative Biochemistry and Physiology, Part D Genomics Proteomics*, 5(4): 256–264.
- Wang, S.-L., Niu, D.-H. and Li, J.-L. 2009. Isolation and characterization of 10 polymorphic microsatellites in *Meretrix meretrix*. *Conservation Genetic Resource*, 1: 111–113.
- Webb, J.E., Wallwork, J.A. and Elgood, J.H. 1978. Guide to invertebrate animals, Second Edition, Macmillan Press Ltd. pp 310.



- Weber, K., Sturmer, L., Hoover, E. and Baker, S. 2010. The role of water temperature in hard clam aquaculture. University of Florida IFAS Extension, Retrieved 12 January 2013 from <https://edis.ifas.ufl.edu/fa151>.
- Widdows, J. and Johnson, D. 1988. Physiological energetics of *Mytilus edulis*: Scope for growth. *Marine Ecology Progress Series*, 46: 113–12.
- Wilson, J.H. and Seed, R. 1974. Reproduction in *Mytilus edulis* L. (Mollusca: Bivalve) in Carlingford Lough, Northern Ireland. *Irish Fisheries Investigation, Series B (Marine)*, 15:30pp
- Wilson, K-J. 2002. A survey of coastal and marine birds at Bako National Park and Samunsam Wildlife Sanctuary, Sarawak. Wildlife Management Report No. 28. 19pp.
- Wu, X., Xiao, S., Li, X., Li, L., Shi, W. and Yu, Z. 2014. Evolution of the tRNA gene family in mitochondrial genomes of five *Meretrix clams* (Bivalvia, Veneridae). *Gene*, 533(1): 439–446.
- Wye, K. 2007. *Pocket guide to shell*. Silverdale Books.
- Vakily, J.M. 1989. *The biology and culture of mussels of the Genus Perna*. International Center for Living Aquatic Resources Management, Manila, Philippines.
- Van der Schalie, H. 1970. Hermaphroditism among North American freshwater mussels. *Malacologia*, 10: 93–112.
- Verween, A., Vincx, M. and Degraer, S. 2007. The effect of temperature and salinity on the survival of *Mytilopsis leucophaeata* larvae (Mollusca, Bivalvia): The search for environmental limits. *Journal of Experimental Marine Biology and Ecology*, 348: 111–120.
- Ward, B.B. 2011. Measurement and Distribution of Nitrification Rates in the Oceans. In *Methods in Enzymology*, Vol. 486, ed. M.G., Klotz, pp. 308–320. Elsevier.
- Wentworth, C.K. 1922. A scale of grade and class terms for clastic sediments. *The Journal of Geology*, 30(5): 377–392.
- Xia, X.H., Yang, Z.F., Huang, G.H., Zhang, X.Q., Yu, H. and Rong, X. 2004. Nitrification in natural waters with high suspended-solid content—A study for the Yellow River. *Chemosphere*, 57: 1017–1029.
- Xu, X., Wu, X. and Yu, Z. 2012. Comparative studies of the complete mitochondrial genomes of four *Paphia* clams and reconsideration of subgenus *Neotapes* (Bivalvia: Veneridae). *Gene*, 494: 17–23.
- Yamakawa, A.Y. and Imai, H. 2012. PCR-RFLP typing reveals a new invasion of Taiwanese *Meretrix* (Bivalvia: Veneridae) to Japan. *Aquatic Invasions*, 8(4): 407–415.

- Yamakawa, A.Y. and Imai, H. 2013. Direct Submission. Okinawa International University, Department of Regional Economics and Environmental Policy, Japan.
- Yamakawa, A.Y., Yamaguchi, Y. and Imai, H. 2008. Genetic relationships among species of *Meretrix* (Mollusca: Veneridae) in the Western Pacific Ocean. *Pacific Science*, 62(3): 385–394.
- Yoloye, V. 1975. The habitat and functional anatomy of the West African bloody cockle *Anadara senilis* (L). *Proceeding of the Malacological Society of London*, 41: 277–299.
- Yoon, J-M., Park, K-I. and Choi, S-H. 2012. Variation of shell color in three geographic white clam (*Meretrix lusoria*) population of the Yellow Sea. *Developmental Reproduction*, 16(1): 47–51.
- Yu, D-s., Peng, Y-z. and Zhang, K. 2004. Effects of seawater salinity on nitrite accumulation in short range nitrification to nitrite as end product. *Journal of Environmental Sciences*, 16(2): 247–252.
- Yusa, Y. 2007. Causes of variation in sex ratio and modes of sex determination in the Mollusca—an overview. *American Malacological Society*, 23(1): 89–98.
- Zhang, J-Z. And Huang, X-L. 2011. Effect of temperature and salinity on phosphate sorption on marine sediments. *Environmental Science and Technology*, 45: 6831–6837.
- Zwamm, A. de, and Mathieu, M. 1992. Cellular biochemistry and endocrinology. In: *The mussel Mytilus: ecology, physiology, genetic and culture*, ed. E.M. Gosling, pp. 223–307. Elsevier Science Publisher B.V., Amsterdam.