

UNIVERSITI PUTRA MALAYSIA

PHYLOGEOGRAPHY OF GENUS *Tenualosa* AND POPULATION STRUCTURE OF *Tenualosa toli* (Valenciennes, 1847) INFERRED FROM CYTOCHROME B MITOCHONDRIAL DNA

PUVANESWARI PUVANASUNDRAM

FP 2018 16



PHYLOGEOGRAPHY OF GENUS Tenualosa AND POPULATION STRUCTURE OF Tenualosa toli (Valenciennes, 1847) INFERRED FROM CYTOCHROME B MITOCHONDRIAL DNA



By

PUVANESWARI A/P PUVANASUNDRAM

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the Requirements for the Degree of Master of Science

March 2017

COPYRIGHT

All material within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Used may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright© Universiti Putra Malaysia



DEDICATION

To my beloved father and mother

Mr. Puvanasundram a/l VM Rajagopal and Mrs. Subethra a/p Perumal

Family members:

Jeevaperagasi a/p Puvanasundram

Sumathi a/p Puvanasundram



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

PHYLOGEOGRAPHY OF GENUS *Tenualosa* AND POPULATION STRUCTURE OF *Tenualosa toli* (Valenciennes, 1847) INFERRED FROM CYTOCHROME B MITOCHONDRIAL DNA

By

PUVANESWARI A/P PUVANASUNDRAM



This study was conducted to examine the systematic and evolutionary relationship among members of genus Tenualosa as well as genetic diversity of Tenualosa toli from selected populations in Sarawak using cytochrome b (cyt b), mitochondrial Deoxyribonucleic acid (mtDNA). A total of 111 T. toli, 24 T. macrura and four T. ilisha samples were obtained. All T. toli and T. macrura samples were collected from various localities in Sarawak and imported samples were collected from Satok market, Sarawak. Samples of T. toli were collected from Sebuyau (N= 25), Sadong Jaya (N=20), Satok market (N=9), Batang Lupar (N=20), Daro (N=12) and Mukah (N=25). Samples of T. macrura on the other hand, were collected from Sadong Java (N=7), Kota Samarahan (N=11) and Daro (N=6). Tenualosa ilisha samples were obtained from Bangladesh. These samples in the form of muscle tissue and fin clips were stored in 95% ethanol before being stored in -20°C freezer for long term storage. Polymerase Chain Reaction (PCR) was conducted on samples using cyt b primer and products were then sent for sequencing. All the sequences obtained were first validated through Basic Local Alignment Search Tool (BLAST) analysis. Phylogenetic analysis supported the monophyletic status between the three species of shad. Pairwise genetic distance (13.9%-15.3%) between species in genus *Tenualosa* supports the taxonomic status of T. toli and T. macrura as distinct species.

Nucleotide diversity in all populations were low (0.001) and the haplotype diversity ranged from the lowest value of 0.417 (Imported Toli shad) to the highest value of 0.656 (Sebuyau samples). Gene flow (Nm) was equal to infinite among all the populations of *T. toli* in Sarawak showing very high gene flow between them due to homogeneity of terubok population from different parts of Sarawak as *T. toli* move from different rivers for spawning. Pairwise fixation index (Fst) values for genetic differentiation among population showed significant levels of genetic differentiation in all comparisons between imported samples from India of *T. toli* population and samples obtained from Sarawak. However, there were no significant differences recorded in

pairwise Fst values in most comparisons among populations of Toli shad collected from Sarawak. Analysis of molecular variance (AMOVA) results revealed that the majority of variance including percentage of variation was among population variance. Negative Tajima's D value was obtained for samples collected from Sebuyau, Batang Lupar, and Daro as well as imported samples which could be due to recent bottleneck events leading to population expansion. Fu's FS showed negative value for samples obtained from Sebuyau, Batang Lupar and imported samples where this value is expected due to recent population expansion. The utilization of mtDNA cyt b in this study has managed to provide an insight on the genetic makeup of T. toli collected from five different locations in Sarawak. This study supports the high genetic differences between the imported and locally collected samples where it could mean that the imported samples belong to a different gene pool or breeding groups. This study showed that cyt b is one of the suitable genes which could be employed for the purpose of species identification, phylogeography and population structure studies. Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

FILOGEOGRAFI GENUS Tenualosa DAN STRUKTUR POPULASI Tenualosa toli (Valenciennes, 1847) MENGGUNAKAN MITOKONDRIA GEN SITOKROM B

Oleh

PUVANESWARI A/P PUVANASUNDRAM



Kajian ini dijalankan untuk mengetahui hubungan evolusi dan sistematik antara ikan daripada genus Tenualosa dan juga kepelbagaian genetik antara Tenualosa toli daripada populasi yang dipilih di Sarawak menggunakan sitokrom b, mitokondria Deoxyribonucleic acid (DNA). Sebanyak 111 sampel T. toli, 24 sampel T. macura dan empat sampel T. ilisha telah diperoleh. Kesemua sampel T. toli dan T. macrura diperoleh daripada beberapa lokasi di Sarawak manakala sampel yang diimport diperoleh dari Pasar Satok, Sarawak. Sampel T. toli yang digunakan dalam kajian ini diperoleh dari Sebuyau (N= 25), Sadong Jaya (N=20), Pasar Satok (N=9), Batang Lupar (N=20), Daro (N=12) dan Mukah (N=25). Sampel T. macrura pula diperoleh dari Sadong Jaya (N= 7), Kota Samarahan (N=11) and Daro (N=6). Sampel T. ilisha pula diperoleh dari Bangladesh. Kesemua sampel dalam bentuk tisu otot dan sirip ini disimpan di 95% ethanol sebelum disimpan di dalam peti sejuk bersuhu -20°C untuk jangka masa panjang. Polymerase Chain Reaction (PCR) telah dijalankan menggunakan primer sitokrom b dan sampel dihantar untuk proses penjujukan DNA. Kesemua jujukan yang diperoleh disahkan menerusi Basic Local Alignment Search Tool (BLAST). Analisa filogenetik mengesahkan status monofiletik di antara tiga spesies di bawah Genus Tenualosa. Jarak genetik di antara spesies-spesies dalam genus Tenualosa (13.9%-15.3%) mengesahkan status taksonomi T. toli dan T. macrura sebagai spesies yang berbeza.

Kepelbagaian nukleotida di antara semua populasi adalah rendah (0.001) dan kepelbagaian haplotaip pula adalah di antara 0.417 (ikan *T. toli* yang diimport) dan 0.656 (sampel *T. toli* dari Sebuyau). Nilai *gene flow* (Nm) pula di antara kesemua populasi *T. toli* di Sarawak adalah infiniti menunjukkan aliran gen yang tinggi di antara kesemua populasi ini kerana ikan ini bergerak dari sungai untuk bertelur dan ini menyebabkan kehomogenan populasi terubok. Nilai *Fixation index* (Fst) di antara sampel *T. toli* yang diimport dan sampel *T. toli* dari Sarawak menunjukkan perbezaan

genetik di tahap yang signifikan. Tiada perbezaan yang signifikan dijumpai dalam kalangan *T. toli* di Sarawak. Nilai analisis varian molekular (AMOVA) yang tinggi menunjukkan bahawa majoriti daripada varian dan peratusan tinggi variasi genetik adalah berpunca daripada perbezaan di antara populasi. Nilai Tajima's D yang negatif yang diperoleh dari sampel di Sebuyau, Batang Lupar, Daro dan sampel yang diimport mungkin berlaku akibat proses 'bottleneck' yang seterusnya menyebabkan pengembangan populasi. Nilai negatif Fu's FS yang diperoleh dari sampel di Sebuyau, Batang Lupar dan sampel diimport juga menunjukkan berlakunya pengembangan populasi. Penggunaan sitokrom *b*, DNA mitokondria dalam kajian ini mampu memberikan informasi mengenai genetik *T. toli* yang diperoleh dari lima populasi berbeza di Sarawak. Kajian ini yang membuktikan kepelbagaian genetik yang tinggi di antara sampel *T. toli* dari Sarawak dan sampel yang diimport menunjukkan sampel yang diimport adalah daripada kumpulan gen yang berbeza. Kajian ini juga membuktikan keberkesanan penggunaan sitokrom *b* dalam mengetahui identiti species, filogeografi and kajian struktur populasi.

ACKNOWLEDGEMENT

First and foremost, I would like to thank Universiti Putra Malaysia (UPM) for giving me the opportunity to further my studies in Masters Degree, and also for providing the required space and equipments to run my research throughout my studies. I would also like to thank my supervisor Assoc. Prof Dr Yuzine Esa for guiding me throughout my research journey by giving constructive comments. A special thanks to my cosupervisors Dr Ina Salwany Binti Md Yasin and Dr S. M. Nurul Amin for assisting me throughout my studies. I am also grateful for the assistance provided by Prof Madya Dr. Khairul Adha A. Rahim and staff as well as postgraduate students from Faculty of Resource Science and Technology (FRST) Universiti Malaysia Sarawak. My gratitude to all the staff in Aquaculture Department, Universiti Putra Malaysia for their assistance. I would also like to acknowledge my laboratory mates namely Nurnadia Marshita Binti Abdul Aziz, Nor Syakina binti Nor Rahim for your assistance and idea whenever I face problem regarding my research. I have learnt a lot from them. Thank you to Diyana Nadhirah Khairul Parman for assisting me in some laboratory works which were new to me. I am also grateful to Nur Jasmin Mohd Yaminudin, Nur Ain Sofea Mohd Taher and Zalikha Mohad Hoszaini for being such wonderful friends and assisting me whenever I needed help.

A very special thanks to my parents Mr Puvanasundram s/o Rajagopal and Madam Subethra d/o Perumal who have never stopped believing in me and supported me through thicks and thin throughout my masters journey, my sisters, Madam Jeevaperagasi and Madam Sumathi for being a role model and back bone of my life. I am who I am now because of my family. Thank you for all the tolerance you gave me. I love you all and thank you so much for everything.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Yuzine B. Esa, PhD Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Ina Salwany Binti Md. Yasin, PhD Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Member)

S. M. Nurul Amin, PhD Associate Professor

Faculty of Agriculture Universiti Putra Malaysia (Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- This thesis is my original work;
- Quotations, illustrations, and citations have been duly referenced;
- This thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- Intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Malaysia (Research) Rules 2012;
- Written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture motes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- There is no plagiarism or data falsification/ fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:	Date:

Name and Matric No.: Puvaneswari a/p Puvanasundram/ GS41372

Declaration by Members of Supervisory Committee

This is to confirm that:

- The research conducted and the writing of this thesis was under our supervisions;
- Supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (revision 2012-2013) are adhered to.

Signature: _____

Name of Chairman of supervisory Committee: Yuzine B. Esa



Signature: ____

Name of Member of Supervisory Committee: Ina Salwany Binti Md. Yasin

Signature: _____

Name of Member of Supervisory Committee: S. M. Nurul Amin

TABLE OF CONTENTS

		Page
	STRACT	i
	STRAK	iii
AC	KNOWLEDGEMENT	v
	PROVAL	vi
	CLARATION	vii
	T OF TABLES	xii
	T OF FIGURES	xiii
LIS	T OF ABBREVIATIONS	XV
CII	ADTEDC	
	APTERS INTRODUCTION	1
1	INTRODUCTION	1
2	LITERATURE REVIEW	
2	2.1 Genus <i>Tenualosa</i>	
	2.1.1 Taxonomy of genus <i>Tenualosa</i>	5
	2.1.2 The biology of genus <i>Tenualosa</i>	5
	2.2 Economic importance of <i>Tenualosa toli</i>	8
	2.3 Molecular genetics methods in fisheries and aquaculture	8
	management	0
	2.4 Mitochondrial DNA	10
	2.4.1 Cytochrome b	10
	2.4.2 Advantages and limitations of cytochrome <i>b</i>	11
	2.5 Nuclear marker: Microsatellites	12
	2.6 Methods in molecular phylogenetics analysis	13
	2.7 Molecular works on <i>Tenualosa toli</i> and other clupeids	14
3	METHODOLOGY	
5	3.1 Study areas	15
	3.2 Laboratory works	10
	3.2.1 DNA Extraction	17
	3.2.2 Gel Electrophoresis	19
	3.2.3 Polymerase Chain Reaction (PCR)	20
	3.2.4 DNA Purification	21
	3.2.5 DNA Sequencing	22
	3.3 Data analysis	
	3.3.1 Phylogeography of Genus Tenualosa	22
	3.3.2 Population structure of Tenualosa toli	23
	3.4 Microsatellite marker analysis	
	3.4.1 DNA Extraction	24
	3.4.2 Polymerase chain reaction (PCR): Optimization	24
	3.4.3 Gel electrophoresis	24
4	RESULTS	
	4.1 PCR amplification	26
	4.2 Characteristics of cytochrome b dataset	27
	4.3 Species validation through BLAST	28

6

	4.4 Phylogeography of genus <i>Tenualosa</i>	30
	4.5 Genetic distance and population genetic structure of <i>Tenualosa toli</i>	34
	4.6 Analysis of microsatellite marker	41
5	DISCUSSIONS	
	5.1 Phylogeography of genus <i>Tenualosa</i>	43
	5.2 Genetic distance and population genetic structure	45
	5.3 Population structure of <i>Tenualosa toli</i> inferred from microsatellite loci	47
6	CONCLUSION AND RECOMMENDATIONS FOR	
	FUTURE RESEARCH	
	6.1 Conclusion	49
	6.2 Recommendations for future research	49
REFE	CRENCES	51
	INDICES	62
	ATA OF STUDENT	72
	OF PUBLICATIONS	73

 \bigcirc

LIST OF TABLES

Table		Page	
3.1	Sequence of forward and reverse cytochrome b primers utilized in this study	20	
3.2	The list of the used microsatellite markers obtained from various references with their respective primer sequences		
4.1	List of samples, haplotype code, number of samples, location and Genbank Accession numbers	29	
4.2	Pairwise genetic distance between species from Genus <i>Tenualosa</i> in percentage value (%)	34	
4.3	Distribution of 15 observed mitochondrial DNA cytochrome b haplotypes, nucleotide diversity, number of haplotypes, haplotype diversity and number of polymorphic sites among populations of <i>Tenualosa toli</i>	35	
4.4	Below the diagonal: pairwise Tamura-Nei genetic distances among 6 populations of <i>Tenualosa toli</i> . Above the diagonal: population subdivision (Fst) values Upper diagonal population subdivision (Fst) values and probability test (Chi-square) for population differentiation based on 1000 permutations of the sequence data, significance levels (P<0.05)	38	
4.5	Hieracrhical analysis of molecular variance (AMOVA) among populations and within populations of <i>Tenualosa toli</i>	38	
4.6	Neutrality test (Tajima's D and Fu' FS) and mismatch distribution parameter estimates (signifance level: $P < 0.01$) for <i>Tenualosa toli</i> from different population	40	
4.7	Population genetic indices between different populations of <i>Tenualosa toli</i> calculated from nucleotide sequences of mitochondrial cytochrome <i>b</i> gene	41	

LIST OF FIGURES

Figure

- 2.1 Images of *Tenualosa toli* (a), *Tenualosa macrura* (b) and *Tenualosa ilisha* (c)
- 2.2 Illustrative diagram showing morphological differences between *Tenualosa toli* (a) and *Tenualosa macrura* (b). Differences could be seen at their head, tail and upper jaw. Photo source: Food and Agriculture Organization of the United Nations (FAO) Species catalogue, Volume 7. Clupeoid fishes of the world
- 3.1 Map of Malaysia where *Tenualosa toli* and *Tenualosa macrura* 15 samples utilized in this study were collected from selected localities in Sarawak
- 3.2 Map indicating sampling location of *Tenualosa ilisha* (N=4) 16 which were obtained from Meghna River, Chandpur, Bangladesh
- 3.3 Map indicating sampling locations and sample size (N) of all 17 *Tenualosa toli* (TT) and *Tenualosa macrura* (TM) samples utilized in this study. *Tenualosa toli* samples were collected from Sebuyau (N=25), Sadong Jaya (N=20), Batang Lupar (N=20), Daro (N=12) and Mukah (N=25) whereas *Tenualosa macrura* samples were collected from Sadong Jaya (N=7), Kota Samarahan (N=11) and Daro (N=6).

3.4 Methodology flowchart for the laboratory works 18

- 3.5 The primer positions of L14724 and H15149 on mitochondrial 21 DNA. Numbering is based on human mitochondrial DNA (Hsieh *et al.*, 2001)
- 4.1 The products of Polymerase Chain Reaction (PCR) using 26 mitochondrial DNA cytochrome *b* gene on Genus *Tenualosa* utilized in this study which were viewed under 1% agarose gel produces band in between 1000bp and 1500 bp
- 4.2 Plot of transition (x) and transversion (Δ) against divergence 28 using Tamura and Nei (1993) distance method onto the third codon position shows little saturation in the cytochrome *b* gene of genus *Tenualosa*

6

re

- 4.3 Phylogenetic relationships among *Tenualosa toli*, *Tenualosa macrura*, *Tenualosa ilisha* and outgroup (*Sardinella maderensis*) haplotypes utilized in the present study. The bootstrap percentage (%) values presented at each nodes corresponds to the Neighbour-Joining/Maximum Likelihood (NJ/ML) analyses (consensus tree) based on 1000 pseudoreplications for NJ and ML analysis
- 4.4 Phylogenetic relationships among *Tenualosa toli*, *Tenualosa macrura*, *Tenualosa ilisha* and outgroup (*Sardinella maderensis*) haplotypes utilized in the present study. The bootstrap percentage values presented at each nodes corresponds to the Bayesian analyses
- 4.5 Phylogenetic relationships among *Tenualosa toli*, *Tenualosa* 33 *macrura*, *Tenualosa ilisha* and outgroup (*Sardinella maderensis*) haplotypes utilized in the present study. The bootstrap percentage (%) values presented at each nodes corresponds to the Maximum Parsimony (MP) analyses based on 1000 pseudoreplications
- 4.6 Minimum spanning network showing relationships among 15 36 mitochondrial DNA cytochrome *b* haplotypes of *Tenualosa toli* with their respective population. The hatch marks among the haplotypes shows single mutational steps
- 4.7 Observed and expected mismatch distribution for expanding 37 population of *Tenualosa toli* from all selected populations and whole population utilized in this study
- 4.8 Gel electrophoresis image of amplification product using 42 AsaC010 microsatellite primer. The size of fragment ranges from 200 bp to 350 bp. This primer managed to amplify all 111 *Tenualosa toli* samples collected in this study

31

32

LIST OF ABBREVIATIONS

°C	Degree celcius
Δ	Transversion
%	Percentage
R	Registered trademark
TM	Trademark symbol
©	Copyright
-ve	Negative
μΙ	Microlitre
μΜ	Micromolar
А	Adenosine
AFLP	Amplified Fragment Length Polymorphism
AMOVA	Hierarchical analysis of molecular variance
BBA	Binding buffer
BI	Bayesian inference
BLAST	Basic Local Alignment Search Tool
Вр	Base pair
С	Cytosine
СЕ	Capillary electrophoresis
CEPA	Community Education and Public Awareness
CLB	Cell Lysis Buffer
CNI	Close neighbour interchange
CWA	Column Wash Solution
Cyt b	Cyrochrome <i>b</i>
DAMBE	Data Analysis in Molecular Biology and Evolution

DNADeoxyribonucleic acidDnaSpSoftware package for comprehensive analysis of DNA polymorphism datadNTPDeoxyribonucleotidesDxyNucleotide substitution per sitedr alAnd othersFAOFood and Agriculture Organization of the United NationsFstFixation indexGGuanineHPLCHigh Performance Liquid ChromatographyHTLTenualosa illsha haplotypesHTMTenualosa illsha haplotypesHTMSolectric fusingIUCNInternational Union for Conservation of NaturekgKilogrammekxyPairwise nucleotide differenceLKMLenst ConcernLIFDCSLow-income food deficit countriesLKMLensta ConcernMicMCMarkov Chain Monte CarloMEGAMolecular Evolutionary Genetics Analysesminminutes		ddH20	Double distilled water
polymorphism data dNTP Deoxyribonucleotides Dxy Nucleotide substitution per site et al And others FAO Food and Agriculture Organization of the United Nations Fst Fixation index G Guanine HPLC High Performance Liquid Chromatography HTL Tenualosa ilisha haplotypes HTM Tenualosa dilisha haplotypes HTT Tenualosa dilisha baplotypes IEF Isoelectric fusing IUCN International Union for Conservation of Nature kg Kilogramme Kxy Pairwise nucleotide difference LEFM Low-income food deficit countries LKIM Lembaga Kemajuan Ikan Malaysia M Molar MCMC Markov Chain Monte Carlo MEGA Molecular Evolutionary Genetics Analyses		DNA	Deoxyribonucleic acid
dNTPDeoxyribonucleotidesDxyNucleotide substitution per siteet alAnd othersFAOFood and Agriculture Organization of the United NationsFstFixation indexGGuanineHPLCHigh Performance Liquid ChromatographyHTLTenualosa illsha haplotypesHTMTenualosa macrura haplotypesHTTSoelectric fusingIUCNInternational Union for Conservation of NaturekgKilogrammeKxyPairwise nucleotide differenceLCNLeast ConcernLIFDCSLow-income food deficit countriesLKIMLembaga Kemajuan Ikan MalaysiaMMolarMCMCMarkov Chain Monte CarloMEGAMolecular Evolutionary Genetics Analyses		DnaSp	Software package for comprehensive analysis of DNA
DxyNucleotide substitution per siteet alAnd othersFAOFood and Agriculture Organization of the United NationsFsiFixation indexGGuanineHPLCHigh Performance Liquid ChromatographyHTLTenualosa ilisha haplotypesHTMTenualosa ilisha haplotypesHTMTenualosa toli haplotypesHTGIsoelectric fusingIUCNInternational Union for Conservation of NaturekgSilogrammeKxyPairwise nucleotide differenceLFDCsLow-income food deficit countriesLKIMLembaga Kemajuan Ikan MalaysiaMMolarMCMCMarkov Chain Monte CarloMEGAMolecular Evolutionary Genetics Analyses			polymorphism data
et alAnd othersFAOFood and Agriculture Organization of the United NationsFstFixation indexFstGitation indexGGuanineHPLCHigh Performance Liquid ChromatographyHTLTenualosa ilisha haplotypesHTMTenualosa ilisha haplotypesHTMTenualosa toli haplotypesHTTTenualosa toli haplotypesHTTSoelectric fusingIUCNInternational Union for Conservation of NaturekgKilogrammeLCLeast ConcernLIFDCsLow-income food deficit countriesILKIMLembaga Kemajuan Ikan MalaysiaMMolarMCMCMarkov Chain Monte CarloMEGAMolecular Evolutionary Genetics Analyses		dNTP	Deoxyribonucleotides
FAOFood and Agriculture Organization of the United NationsFstFixation indexGGuanineHPLCHigh Performance Liquid ChromatographyHTLCenualosa ilisha haplotypesHTMCenualosa macrura haplotypesHTTSoelectric fusingHCNInternational Union for Conservation of NaturekgKilogrammekgAirwise nucleotide differenceLTPCsLeast ConcernLTPCsLow-income food deficit countriesLKMLembaga Kemajuan Ikan MalaysiaMCNCMakov Chain Monte CarloMEGAMicoult Evolutionary Genetics Analyses		Dxy	Nucleotide substitution per site
FstFixation indexGGuanineHPLCHigh Performance Liquid ChromatographyHTLTenualosa ilisha haplotypesHTMTenualosa macrura haplotypesHTMTenualosa toli haplotypesHTTSoelectric fusingIUCNInternational Union for Conservation of NaturekgKilogrammeKxyPairwise nucleotide differenceLFDCsLow-income food deficit countriesLKIMLeast ConcernLIFDCsLow-income food deficit countriesMMolarMCMCMakov Chain Monte CarloMEGAMolecular Evolutionary Genetics Analyses		et al	And others
GGuanineHPLCHigh Performance Liquid ChromatographyHTLTenualosa ilisha haplotypesHTMTenualosa macrura haplotypesHTMTenualosa toli haplotypesHTTSoelectric fusingIUCNInternational Union for Conservation of NaturekgKilogrammeKxyPairwise nucleotide differenceLFDCsLow-income food deficit countriesLKIMLow-income food deficit countriesMMolarMCMCMarkov Chain Monte CarloMEGAMolecular Evolutionary Genetics Analyses		FAO	Food and Agriculture Organization of the United Nations
HPLCHigh Performance Liquid ChromatographyHTL <i>Emualosa ilisha</i> haplotypesHTM <i>Emualosa macrura</i> haplotypesHTT <i>Temualosa toli</i> haplotypesHTTSoelectric fusingIUCNInternational Union for Conservation of NaturekgKilogrammeLCNLeast ConcernLFDCSLow-income food deficit countriesLFDCSLow-income food deficit countriesMAMolarMCNCMakov Chain Monte CarloMEGAMiceular Evolutionary Genetics Analyses		Fst	Fixation index
HTL <i>Tenualosa ilisha</i> haplotypesHTM <i>Tenualosa macrura</i> haplotypesHTM <i>Tenualosa macrura</i> haplotypesHTT <i>Tenualosa toli</i> haplotypesIEFIsoelectric fusingIUCNInternational Union for Conservation of NaturekgKilogrammeKxyPairwise nucleotide differenceLCLeast ConcernLIFDCsLow-income food deficit countriesLKMLembaga Kemajuan Ikan MalaysiaMMolarMCMCMarkov Chain Monte CarloMEGAMolecular Evolutionary Genetics Analyses		G	Guanine
HTMTenualosa macrura haplotypesHTTTenualosa toli haplotypesHTTTenualosa toli haplotypesIEFIsoelectric fusingIUCNInternational Union for Conservation of NaturekgKilogrammeKxyPairwise nucleotide differenceLCLeast ConcernLIFDCsLow-income food deficit countriesLKIMLembaga Kemajuan Ikan MalaysiaMMolarMCMCMarkov Chain Monte CarloMEGAMolecular Evolutionary Genetics Analyses		HPLC	High Performance Liquid Chromatography
HTTTenualosa toli haplotypesIEFIsoelectric fusingIUCNInternational Union for Conservation of NaturekgKilogrammeKxyPairwise nucleotide differenceLCLeast ConcernLIFDCsLow-income food deficit countriesLKIMLembaga Kemajuan Ikan MalaysiaMMolarMCMCMarkov Chain Monte CarloMEGAMolecular Evolutionary Genetics Analyses		HTL	Tenualosa ilisha haplotypes
IEFIsoelectric fusingIUCNInternational Union for Conservation of NaturekgKilogrammeKxyPairwise nucleotide differenceLCLeast ConcernLIFDCsLow-income food deficit countriesLKIMLembaga Kemajuan Ikan MalaysiaMMolarMCMCMarkov Chain Monte CarloMEGAMolecular Evolutionary Genetics Analyses		HTM	Tenualosa macrura haplotypes
IUCNInternational Union for Conservation of NaturekgKilogrammeKxyPairwise nucleotide differenceLCLeast ConcernLIFDCsLow-income food deficit countriesLKIMLembaga Kemajuan Ikan MalaysiaMMolarMCMCMarkov Chain Monte CarloMEGAMolecular Evolutionary Genetics Analyses		HTT	Tenualosa toli haplotypes
kgKilogrammeKxyPairwise nucleotide differenceLCLeast ConcernLIFDCsLow-income food deficit countriesLKIMLembaga Kemajuan Ikan MalaysiaMMolarMCMCMarkov Chain Monte CarloMEGAMolecular Evolutionary Genetics Analyses		IEF	Isoelectric fusing
KxyPairwise nucleotide differenceLCLeast ConcernLIFDCsLow-income food deficit countriesLKIMLembaga Kemajuan Ikan MalaysiaMMolarMCMCMarkov Chain Monte CarloMEGAMolecular Evolutionary Genetics Analyses		IUCN	International Union for Conservation of Nature
LCLeast ConcernLIFDCsLow-income food deficit countriesLKIMLembaga Kemajuan Ikan MalaysiaMMolarMCMCMarkov Chain Monte CarloMEGAMolecular Evolutionary Genetics Analyses		kg	Kilogramme
LIFDCsLow-income food deficit countriesLKIMLembaga Kemajuan Ikan MalaysiaMMolarMCMCMarkov Chain Monte CarloMEGAMolecular Evolutionary Genetics Analyses		Кху	Pairwise nucleotide difference
LKIMLembaga Kemajuan Ikan MalaysiaMMolarMCMCMarkov Chain Monte CarloMEGAMolecular Evolutionary Genetics Analyses		LC	Least Concern
M Molar MCMC Markov Chain Monte Carlo MEGA Molecular Evolutionary Genetics Analyses		LIFDCs	Low-income food deficit countries
MCMC Markov Chain Monte Carlo MEGA Molecular Evolutionary Genetics Analyses		LKIM	Lembaga Kemajuan Ikan Malaysia
MEGA Molecular Evolutionary Genetics Analyses		М	Molar
	(\mathbf{G})	MCMC	Markov Chain Monte Carlo
min minutes		MEGA	Molecular Evolutionary Genetics Analyses
		min	minutes

	mg	Miligramme
	Mg Cl ₂	Magnesium chloride
	ml	Millilitre
	ML	Maximum likelihood
	mM	milimolar
	MP	Maximum Parsimony
	MSN	Minimum spanning network
	mtDNA N	Mitochondrial DNA Number of samples
	NCBI	National Centre for Biotechnology Information
	ng	Nanogramme
	NJ	Neighbour Joining
	Nm	Gene flow
	nuDNA	Nuclear DNA
	Р	Probability
	PCR	Polymerase Chain Reaction
	PBS	Phosphate-buffered saline
	РК	Proteinase K
	PopArt	Population Analysis with Reticulate Trees
	PUFAs	Polyunsaturated fatty acids
	RAPD	Randomly Amplified Polymorphic DNA
	R	Transition to transversion ratio
(\mathbf{C})	Rpm	Revolutions per minute
	S	Seconds
	SSR	Simple Sequence Repeat

Standard length

T Thymine

TBE Tris base, boric acid and EDTA

Transition

Times

TRIP Terubok Rehabilitation Integrated Programma

- TL Total length
- UV Ultraviolet
- V Volt
- х
- Х

CHAPTER 1

INTRODUCTION

The global total capture of fishery production in 2014 was 93.4 million tonnes, where 81.5 million tonnes were from marine waters and 11.9 million tonnes were from inland waters according to Food and Agriculture Organization of the United Nations (FAO, 2016). On the other hand, per capita consumption of fish in 2013 for developing regions were 18.8 kilogramme (kg) whereas for low-income food deficit countries (LIFDCs) were 7.6 kg. The total marine capture production in Malaysia specifically, for the year 2014 was 1.46 million tonnes which was 1.7% lower compared to 2013. Besides that, Malaysia seems to be one of the top 25 producers and main group of farmed species in 2015, where the total aquaculture production which includes both inland and marine aquaculture was 521 thousand tonnes (FAO, 2016) thus portraying the significance of this sector in Malaysia.

Clupeiformes is globally known as one of the most important commercial fisheries. This order is classified into five different families namely Denticipidae, Pristigasteridae, Engraulidae, Clupeidae and Chirocentridae (Froese and Pauly, 2004). Genus *Tenualosa* which is one of the member of family Clupeidae consists of five main species worldwide but only two species (*Tenualosa toli and Tenualosa macrura*) could be mainly found in Malaysia especially in East Malaysia also known as Borneo. *Tenualosa toli* commonly known as 'terubok sungai' is one of the most important species in Sarawak fisheries as it has a lot of economical and cultural value. This fish has various other local names such as 'bekawal', 'bengkalis', 'temparik', 'terubuk padi', 'terubuk mulut besar' and Chinese herring (Atan *et al.*, 2010).

The order Clupeiformes which includes anchovies, herrings, sardines, menhadens, shads and other members plays a major role in world fisheries as the global capture production of Clupeiformes was 19,000,000 tonnes in 2003, which constitutes 25% of total annual catch (FAO, 2003). Likewise, in 2014, there were many Clupeid species listed under the major species and genera in marine capture production (FAO, 2016). One of the twenty highest imported fish in Malaysia for the year 2015 was terubok fish where it is imported in frozen form [Fisheries Development Authority of Malaysia (LKIM, 2015)]. 'Terubok' is mostly sold as salted fish in East Malaysia which is a very famous delicacy.

Overall, among the total 31.2 million Malaysian citizens, it has been recorded that each person consumes about 31.5 kg of fish in a year. This portrays the importance of fish as a frequently opted source of protein compared to meat which would usually be taken occasionally. Fish is believed to be one of the most important sources of protein for people around the world as 17% of intake of animal protein in 2013 was fish. Consumption of fish reduces the risk of heart disease, strokes, some cancer and other ailments (Sikorski, 1990). One of the highly desired nutrient in fish is the omega-3 polyunsaturated fatty acids (PUFAs) which has many benefits to human such as

reducing the risk of coronary heart diseases, mild hypertension, prevent cardiac arrhythmias and sudden death (Sidhu, 2003).

In the International Union of Conservation of Nature (IUCN) red list of threatened species, the status of both *T. toli* and *T. macrura* has yet to be assessed, but the status of *T. ilisha* is written as least concern (LC) and the population trend of this fish is stated as decreasing. Eventhough the status of *T. toli* has yet to be assessed it is highly recommended that preventive conservation related programme should be conducted in advance. In Sarawak, conservation and rehabilitation programme focuses more on the Big Mouth 'Terubok' or also known as *T. toli* as it is the species that is being over exploited. Terubok sungai is highly targeted by fishermen especially the ovaries of spawning females as it is highly valuable (Milton *et al.*, 1997; Willman *et al.*, 1989).

For the purpose of fish stock identification, one of the conventional methods utilized is by comparing morphological characters of fish. Examples of mostly used characters are number of scales in lateral series or relative body depth (Ferris and Berg, 1987). Due to various developmental stages of fish mainly larvae and juvenile where their morphological characteristics seems difficult to be used as a base for species identification, molecular marker based identification mainly Deoxyribonucleic acid (DNA) characters based identification is more appropriate to be utilized (Teletchea, 2009).

The gene content of mitochondria-genomes is highly conserved throughout the evolution of metazoan (Wolstenholme, 1992) which is one of the main criteria that qualifies mitochondrial DNA (mtDNA) marker's application in stock and species identification. Besides that, molecular markers such as mtDNA markers are also suitable to resolve phylogeography relationships and population structure as well as genetic variation analysis in fish species. Phylogeographic studies involve geographic and evolutionary processes which portrays the genetic divergence of the population. This is because, molecular sequences has a very simple structure where there are mainly four bases that are subjected to mutation, selection and drift (Kocher and Stepien, 1997) which makes it highly conserved. Mitochondrial DNA has been utilized to investigate significant genetic divergences among population across geographical regions as well as to conduct demographic study to identify any form of population expansion (Ma *et al.*, 2010).

In some research, nuclear and mitochondrial markers are used hand in hand to investigate the phylogeographic structure of a particular species (Hammer *et al.*, 2010). There are disprecancies on mtDNA biogeographical data compared to nuclear DNA (nuDNA) mainly due to adaptive introgression of mtDNA, demographic disparities and sex-biased asymmetries, hence the idea of combining both of this markers is believed to provide a much stronger and reliable molecular data for both phylogenetic and phylogeographic analysis (Toews and Brelsford, 2012). Thus, both mtDNA and nuDNA marker are suitable for utilization in molecular related studies.

Significance of the study

Currently there are not many studies which cover on the genetic attribute of T. toli eventhough there seem to be more genetic related studies on its congener, T. ilisha. Various studies such as population genetic study, allozyme and morphological variation, genetic variation and many more studies were conducted on Hilsa shad (Behera et al., 2015; Mazumder and Alam, 2009; Salini et al., 2004; Rahman and Naevdal, 2000). The decline in the population of *T. toli* calls for more genetic study for the purpose of conservation. One of the earliest study which focuses solely on T. toli is the study by Blaber et al., (1996). This study covers on the life history of this fish. This study could be acknowledged as the basis of various studies on T. toli. Toli shad is subjected to overfishing as well as climate changes which affects the population level. Another importance of this current study is that T. toli was subjected to overexploitation including other factors mainly environmental degradations and water pollutions. Hence the total catch landing of this fish has been reported to have depleted to a very low level due to overfishing activities (Blaber et al., 2005). It is also believed that the fish populations in Batang Saribas and Batang Lassa which are known as the core terubok areas in Sarawak has depleted to a low level (Khairul Adha et al., 2014).

Knowing the significance of *T. toli* as a foodfish as well as the characteristics of this fish as a protandric hermaphrodite which makes it unique, it is very important to identify the actual genetic makeup to know the current status of terubok fish population in Sarawak. The application of genetics in conservation is mainly to prioritize which particular population or species requires the allocated financial resources the most.

It is believed that *T. toli* in Sarawak shared the same estuarine habitat with more than about sixty other species. This lead to a rich biodiversity environment so any form of conservation programme implemented in the habitat of this fish would do a huge favour to all other species and increase the overall fish biomass production (Awang Alim *et al.*, 2012). Moreover Toli shad is placed at the lowest trophic (food) level in the pyramid so the stability of the system depends on the diversity and availability of this fish.

Objectives

Therefore, there were three main objectives in this study are,

- 1. To reconstruct phylogenetic tree inferred from cytochrome *b*, mitochondrial DNA in order to examine the systematic and evolutionary relationship among shad of genus *Tenualosa*.
- 2. To examine the population genetic structure and level of genetic diversity among and within populations of *Tenualosa toli* from different geographical groups using cytochrome *b*, mitochondrial DNA.
- 3. To examine the population genetic structure and level of genetic diversity among and within populations of *Tenualosa toli* from different geographical groups using microsatellite DNA.

REFERENCES

- Aziz, A. H. A., Tarmizi, Z., Ali, N. M., Ariffin, N. A., Abdullah, M. D. D., Lian, W. L., Giat, S. Y., Jaafar, T. N. A. M., Bolong, A. M. A. Kasim, A. A. A., and Sheriff, S. M. (2015). Mitochondrial DNA Diversity of Terubok (*Tenualosa toli*) from Daro and Mukah, Sarawak Inferred by Partial Cytochrome b (Cytb). Journal of Fisheries and Aquatic Science, 10(2):92-101.
- Allendorf, F. W., Leary, R. F., Spruell, P., and Wenburg, J. K. (2001). The problems with hybrids: setting conservation guidelines. *Trends in Ecology and Evolution*, 16(11):613-622.
- Arai, T., and Amalina, R. (2014). New record of a tropical shad *Tenualosa ilisha* (Teleostei: Clupeidae) in Malaysian waters. *Marine Biodiversity Records*, 7:1-4.
- Askari, G. H., Shabani, A., and Miandare, K. (2013). Application of molecular markers in fisheries and aquaculture. *Scientific Journal of Animal Science*, 2(4):82-88.
- Atan, Y., Jaafar, H., and Abdul Majid, A. R. (2010). *Ikan Laut Malaysia Glosari nama* sahih spesies ikan. Dewan Bahasa dan Pustaka.
- Avise, J. C. (2000). *Phylogeography: The history and formation of species*. Harvard University Press, Cambridge, M. A.
- Avise, J. C., Arnold, J., Ball, R. M., Bermingham, E., Lamb, T., Neigel, J., Reeb, C. A., and Saunders, N. C. (1987). Intraspecific phylogeography: the mitochondrial DNA bridge between population genetics and systematic. *Annual Review of Ecology and Systematic*. 18(1):489-522.
- Avise, J. C. (1994). *Molecular markers, natural history and evolution*. Chapman and Hall, New York.
- Awang A.A. K., Wong, P and Khairulniezawaymie. (2012). Proceedings from UMT 11th International annual symposium on sustainability science and management: *The terubok (Tenualosa spp.) rehabilitation integrated program (TRIP) and conservation in Sarawak.*
- Baker, R. J., and Bradley, R. D. (2001). A test of the genetic species concept: cytochrome-*b* sequences and mammals. *Journal of Mammalogy*. 82(4):960-973.
- Bandelt, H. J., Forster, P., and Rohl, A. (1999). Median-joining networks for inferring interspecific phylogenies. *Molecular Biology and Evolution*. 16(1):37-48.
- Barat, A., Ali, S., Sati, J., and Sivaraman, G. K. (2012). Phylogenetic analysis of fishes of the subfamily Schizothoracinae (Teleostei: Cyprinidae) from Indian Himalayas using cytochrome b gene. Indian Journal of Fisheries. 59(1):43-47.

- Behera, B. K., Singh, N. S., Paria, P., Sahoo. A. K., Panda, D. K., Meena, P., Das, S., Parakshi, S., Biswas, D. K., and Sharma, A. P. (2015). Population genetic structure of Indian shad, *Tenualosa ilisha* inferred from variation in mitochondrial DNA sequences. *Journal of Environmental Biology*, 36:1193-1197.
- Begg, G. A., and Waldman, J. R. (1999). An holistic approach to fish stock identification. *Fisheries Research*, 43:35-44.
- Blaber, S. J. M., Milton, D. A., Chenery, S. R., Fry, G. (2003). New insights into the life history of *Tenualosa ilisha* and fishery implications. *American Fisheries Society Symposium*, 35:223-240.
- Blaber, S. J. M., Fry, G., Milton, D., Van der Velde, T., Boon-Teck, O. Pang, J., and Wong, P. (2005). The life history of *Tenualosa macrura* in Sarawak, further notes on protandry in the genus and management strategies. *Fisheries Management and Ecology*, 12(3):201-210.
- Blaber, S. J. M, Brewer, D. T., Milton, D. A., Merta, G. S., Efizon, D., Fry, G., and Van der Velde, T. (1999). The life history of the protandrous tropical shad *Tenualosa macrura* (Alosinae: Clupeidae): Fishery implications. *Estuarine*, *Coastal and Shelf Science*, 49:689-701.
- Blaber, S. J. M., Milton, D. A., Pang, J., Wong, P., Boon-Teck, O., Nyigo L., and Lubim D. (1996). The life history of the tropical shad *Tenualosa toli* from Sarawak: first evidence of protandry in the Clupeiformes. *Environmental Biology of Fishes*, 46:225-242.
- Blaber, S. J. M. (2009). Can the ecosystem approach improve management of tropical estuarine fisheries for diadromous species. *American Fisheries Society Symposium*, 69:649-661.
- Bowen, B. R., Kreiser, B. R., Mickle, P. F., Schaefer, J. F., and Adams, S. B. (2008). Phylogenetic relationships among North Americal Alosa species (Clupeidae). Journal of Fish Biology, 72:1188-1201.
- Brahmane, M. P., Kundu, S. N., Das, M. K., and Sharma, A. P. (2013). Low genetic diversity and absence of population differentiation of hilsa (*Tenualosa ilisha*) revealed mitochondrial DNA cytochrome b region in Ganga and Hooghly rivers. African Journal of Biotechnology. 12(22): 3383-3389.
- Branicki, W., Kupiec, T., and Pawlowski, R. (2003). Validation of cytochrome b sequence analysis as a method of species identification. Journal of Forensic Science. 48(1):1-5.
- Brown, B., and Epifanio, J. (2003). Nuclear DNA. In: Hallerman, E. M. (Ed.), Population genetics: Principles and Applications for fisheries scientists. *American Fisheries Society*, Bethesda, Maryland. 101-123.

Brown, T. A. (2002). Genomes 2nd edition. Manchester, UK: Oxford Wiley-Liss.

- Bruno, W. J., Socci, N. D., and Halpern, A. L. (2000). Weighted neighbour joining: a likelihood-based approach to distance-based phylogeny reconstruction. *Molecular Biology and Evolution*. 917(1): 189-197.
- Bujang, N., Ismail, S., and Budin, H. (2013, April). Estimate of the maximum sustainable yield of Tenualosa toli in Sarawak. In A. Ishak, I. Hashim, E. S. Ismail, & R. Nazar (Eds), AIP Conference Proceedings (Vol. 1522, No. 1, pp. 798-803). AIP.
- Cantatore, P., Roberti, M., Pesole, G., Ludovico, A., Millela, F., Gadaleta, M. N., and Saccone, C. (1994). Evolutionary analysis of cytochrome *b* sequences in some Perciformes: Evidence for a slower rate of evolution in mammals. *Journal of Molecular Evolution*, 39:589-597.
- Carpenter, K. E. (1998). An introduction to the oceanography, geology, biogeography and fisheries of he tropical and subtropical waters and central Pacific. FAO species identification guide for fishery purpose: The living marine resources of the western central Pacific Rome, FAO:1-9.
- Cavalli-sforza, L. L., and Edwards, A. W. F. (1967). Phylogenetics analysis models and evolution procedures. American Journal of Human Genetics, 19(3): 233-257.
- Christiakov, D. A., Hellemans, B., and Volckaert. (2006). Microsatellites and their genomic distribution, evolution, function and applications: A review with special reference to fish genetics. *Aquaculture*, 255:1-29.
- Clisson, I., Lathuilliere, M., and Crouau-Ray, B. (2000). Conservation and evolution of microsatellite loci in primate taxa. *Americal Journal of Primatology*. 50(3):205-214.
- Connell, M. O., and Wright, J. M. (1997). Microsatellite DNA in fishes. *Reviews in Fish Biology and Fisheries*. 7:331-363.
- Dahle, G., Rahman, M., and Eriksen, A. G. (1997). RAPD fingerprinting used for discriminating among three populations of Hilsa shad (*Tenualosa ilisha*). Fisheries Research. 32: 263-269.
- Dereeper, A., Guignon, V., Blanc, G., Audic, S., Buffet, S., Chevenet, F., Dufayard, J.
 F., Guindon, S., Lefort, V., Lescot, M., Clevaerie, J. M., and Gascuel, O. (2008). Phylogeny.fr: robust phylogenetic analysis for the non-specialist. Nucleic Acids Research. 36: 465-469
- DeSalle, R., Giribet, G., and Wheeler, W. (Eds). (2013). *Techniques in molecular* systematic and evolution. Springer Science and Business Media.
- Durand, J. D., Tsigenopoulos, C. S., Unlu, E., and Berrebis, P. (2002). Phylogeny and biogeography of the family Cyprinidae in the middle east inferred from cytochrome b DNA- Evolutionary significance of this region. *Molecular Phylogenetics and Evolution*, 22(1):91-100.

- Ellegren, H. (2000). Microsatellite mutations in the germline: implications for the evolutionary inference. *Trends in Genetics*, 16(12):551-558.
- Esa, Y. B., Jeffrine, R. R. J., Khairul Adha, A. R., Siraj, S. S., Daud, S. K., Tan, S. G., and Sungan, S. (2012). Phylogenetic relationship among several freshwater fishes (Family: Cyprinidae) in Malaysia inferred from partial sequencing of the cytochrome *b* mitochondrial DNA (mtDNA) gene. *Pertanika Journal of Tropical Agricultural Science*, 35(2):307-318.
- Esa, Y. B., Siraj, S. S., Dud, S. K. Japning, J. R. R., Khairul Adha, A. R., and Tan, S. G. (2008). Molecular systematics of mahseers (Cyprinidae) in Malaysia inferred from sequencing of a mitochondrial cytochrome c oxidase I (COI) gene. *Pertanika Journal of Tropical Agricultural Science*, 31(2):263-269.
- Excoffier, L., and Lischer, H. E. L. (2010). Arlequin suite ver 3.5: A new series of programs to perform population and genetics analyses under Linux and Windows. *Molecular Ecology Resources*, 10:564-567.
- Faria, R., Wallner, B., Weiss, S., and Alexandrino, P. (2004). Isolation and characterization of eight dinucleotide microsatellite loci from two closely related clupeid species (*Alosa alosa and A. Fallax*). *Molecular Ecology Notes*, 4:586-588.
- Farias, I. P., Orti, G., Sampaio, I., Schneider, H., and Meyer, A. (2001). The cytochrome b gene as a phylogenetic marker: the limits of resolution for analyzing relationships among cichlid fishes. *Journal of Molecular Evolution*, 53:89-103.
- Felsenstein, J. (1985). Confidence limits on phylogenies: An approach using the bootstrap. *Evolution*, 39:783-791.
- Ferguson, A. (1994). Molecular genetics in fisheries: current and future perspectives. *Reviews in Fish Biology and Fiheries*, 4(3):379-383.
- Ferris, S. D., and Berg, W. J. (1987). The utility of mitochondrial DNA in fish genetics and fishery. Population genetics, Fishery management. University of Washington Press, Seattle, WA. 277-299.
- Fessler, J. L., and Westneat, M. W. (2007). Molecular phylogenetics of the butterflyfishes (Chaetodontidae): Taxonomy and biogeography of a global coral reef fish family. Molecular Phylogenetics and Evolution, 45:50-68.
- Fisheries Development Authority of Malaysia (LKIM). (2015). Retrieved from http://www.lkim.gov.my/en/ at 21st December 2016
- Froese, R., and Pauly, D. (2004). Fishbase. World Wide Web electronic publications. www.fishbase.org.

- Froukh, T., and Kochzius. (2007). Genetic population structure of the endemic fourline wrasse (*Larabicus quadrilineatus*) suggests limited larval dispersal distances in the Red Sea. *Molecular Ecology*, 16:1359-1367.
- Fu, Y. X. (1997). Statistical tests of neutrality of mutations against population growth, hitchhiking and background selection. *Genetics*. 157:915-925.
- Glenn, T. C., and Schable, N. A. (2005). Isolating microsatellite DNA loci. In: *Molecular Evolution: Producing the Biochemical Data, Part B* (eds Zimmer, E. A and Roalson, E). Academic Press, San Diego, USA. 202-222.
- Gonzalez, E. G., and Zardoya, R. (2007). Isolation and characterization of polymorphic microsatellites for the sardine *Sardina pilchardus* (Clupeiformes: Clupeidae). *Molecular Ecology Notes*. 7:519-521.
- Grant, W. S and Bowen, B. W. (1998). Shallow population histories in deep evolutionary lineages of marine fishes: Insights from sardines and anchovies and lessons for conservation. *Journal of Heredity*, 89(5):415-426
- Hammer, M. P., Unmack, P. J., Adams, M., Johnson, J. B., and Walker, K. F. (2010). Phylogeographic structure in the threatened Yarra pygmy perch *Nanoperca* obscura (Teleostei: Percichthyidae) has major implications for declining population. Conservation Genetics, 11(1):213-223.
- Harrison, R. G. (1989). Animal mitochondrial DNA as a genetic marker in population and evolutionary biology. *Trends in Ecology and Evolution*, 4(1):6-11.
- Hauser, L and Carvalho, G. R. (2008). Paradigm shifts in marine fisheries genetics: ugly hypothesis slain by beautiful facts. *Fish and fisheries*. 9: 333-362
- Hauser, L., and Seeb, J. E. (2008). Advances in molecular technology and their impact on fisheries genetics. *Fish and Fisheries*, 9:473-486.
- Hillis, D. M., and Huelsenbeck, J. P. (1992). Signal, noise, and reliability in molecular phylogenetic analyses. *Journal of Heredity*, 83:189-195.
- Holder, M., and Lewis, P. O. (2003). Phylogeny estimation: Traditiona and Bayesian approaches. *Nature Review Genetics*, 4:275-284.
- Hong Wan, Q., Wu, H., Fujihara, T., and Fang, S. G. (2004). Which genetic marker for conservation genetic issues. *Electrophoresis*, 25(14):2165-2176.
- Hsieh, H. M., Chiang, H. L., Tsai, L. C. Lai, S. Y., Huang, N. E., Linacre, A., and James, C. I. L. (2001). Cytochrome *b* gene for species identification of the conservation animals. *Forensic Science International*, 122(1):7-18.
- Hudson, R. R., Slatkin, M., and Maddison, W. P. (1992). Estimation of levels of gene flow from DNA sequence data. *Genetics*, 132(2):583-589.

- Huelsenbeck, J. P., and Ronquist, F. (2001). MRBAYES: Bayesian inference of phylogenetic inference of phylogenetic trees. *Bioinformatics*, 17(8):754-755.
- Irwin, D., Kocher, T., and Wilson, A. (1991). Evolution of the cytochrome *b* gene of mammals. *Journal of Molecular Evolution*, 32(2):128-144.
- Jawad, L., Al-Mamry, J., and Al-Mamary, D. (2011). First record of toli shad, *Tenualosa toli* (Valenciennes, 1847), from the Oman Sea (Gulf of Oman). *Journal of Applied Ichthyology*, 27:1379-1380.
- Jeffrine, R. J. R., and Esa, Y. (2006). Phylogenetic analysis of Hampala fishes (Subfamily Cyprinina) in Malaysia inferred from partial mitochondrial cytochrome *b* DNA sequences. *Zoological Science*, 23:893-901.
- Jukes, T. H., and Cantor, C. R. (1969). Evolution of protein molecules. In Mammalian protein metabolism'.(Ed. HN Munro.). 21-132.
- Julian, S. E., and Bartron, M. L. (2007). Microsatellite DNA markers for American shad (*Alosa sapidissima*) and cross-species amplification within the family Clupeidae. *Molecular Biology Notes*, 7:805-807.
- Kapuscinski, A. R., and Jacobson, L. D. (1987). Genetic guidelines for fisheries management (No 17). Duluth, Minnesota: Minnesota Sea Grant, University of Minnesota.
- Keith, P., Galewski, T., Cattaneo-Berrebi, G., Hoareau, T., and Berrebi, P. (2005). Ubiquity of *Sicyopterus lagocephalus* (Teleostei: Gobioidei) and phylogeography of the genus *Sicyopterus* in the Indo-Pacific area inferred from mitochondrial cytochrome b gene. *Molecular Phylogenetics and Evolution*, 37:721-732.
- Khairul Adha, A. R., Hambali Tumiran, M., Shabdin, M. L., Esa, Y. B., and Awang Husaini, A. A. S. (2014). The status of *Tenualosa toli* (Valenciennes, 1847) in the Southwest coast of Sarawak, Malaysia. *Kuroshio Science*, 8(1):9-44.
- Kimura, M. (1983). The Neutral Theory of Molecular Evolution, *Cambridge University Press.*
- Kimura, M. (1980). A simple method for estimating evolutionary rate of base substitutions through comparative studies of nucleotide sequences. *Journal of Molecular Evolution*, 16:111-120.
- Kocher, T. D. (2004). Adaptive evolution and explosive speciation: The cichlid fish model. *Nature Review Genetics*. 5: 288-298.
- Kocher, T. D., and Stepien, C. A. (1997). *Molecular Systematics of fishes*. London: Academic Press.
- Kuhner, M. K., and Felsenstein, J. (1994). A simulation comparison of phylogeny algorithms under equal and unequal evolutionary rates. *Molecular Biology and Evolution*, 11:459-468.

- .Kumar, S., Stecher, G., and Tamura, K. (2016). MEGA7: Molecular Evolutionary Genetics Analysis version 7.0 for bigger datasets. *Molecular Biology and Evolution*, 33:1870-1874.
- Krishnan, S., and Mishra, S. S. (2004). An Inventory of fish species described originally from fresh and coastal marine waters of Pondicherry. *Records of the Zoological Survey of India*, 102(3-4):65.
- Lakra, W., Verma, M., Goswami, M., Lal, K. K., Mohindra, V., Punia, P., Gopalakrishnan, A., Singh, K. V., Ward, R. D., and Hebert, P. (2011). DNA barcoding Indian marine fishes. *Molecular Ecology Resources*, 11(1):60-71.
- Larkin, M. A., Blackshields, G., Brown, N. P., Chenna, R., McGettigan, P. A., McWilliam, H., Valentin, F., Wallace, I. M., Wilm, A., Lopez, R., Thompson, J. D., Gibson, T. J., and Higgins, D. G. (2007). Clustal W and Clustal X version 2.0. *Bioinformatics*, 23:2947-2948.
- Lavoué, S., M. Miya, P., Musikasinthorn, P., Wei Jen, C and Mutsumi, N. (2013). Mitogenomic evidence for an Indo-West Pacific origin of the Clupeoidei (Teleostei: Clupeiformes). *PLos ONE*, 8(2):e56485.
- Lemey, P., Salemi, M., and Vandamme, A. M. (Eds). (2009). The Phylogenetic Handbook: A Practical Approach to Phylogenetic Analysis and Hypothesis Testing. Cambridge University Press.
- Librado, P., and Rozas, J. (2009). DnaSP v5: A software for comprehensive analysis of DNA polymorphism data. *Bioinformatics*, 25(11):1451-1452.
- Liu, J. X., Gao, T. X., Yokogawa, K., and Zhang, Y. P. (2006). Differential population structuring and demographic history of two closely related fish species, Japanese sea bass (*Lateolabrax japonicas*) and spoted sea bass (*Lateolabrax maculates*) in Northwestern Pacific. *Molecular Phylogenetics and Evolution*, 39(3):799-811.
- Lopez, P., Forterre, P., and Philippe, H., (1999). The root of the tree of life in the light of the covarion model. Molecular Biology and Evolution, 17:835-838.
- Lydeard, C. H. A. R. L. E. S., and Roe, K. J. (1997). The phylogenetics utility of the mitochondrial cytochrome b gene for inferring relationships among actinopterygian fishes. *Molecular Systematic of Fishes*, 285-303
- Ma, C., Cheng, Q., Zhang, Q., Zhuang, P., and Zhao, Y. (2010). Genetic variation of *Coilia ectenes* (Clupeiformes: Engraulidae) revealed by the complete cytochrome b sequences of mitochondrial DNA. *Journal of Experimental Marine Biology and Ecology*, 385:14-19.
- Madesis, P., Ganopoulos., and Tsaftaris, A. (2013). *Microsatellites: Evolution and contribution. Microsatellites methods and protocol.* Eds Kantartzi, S. K. New York: Humana press.

- Mazumder, S. K., and Alam, M. S. (2009). High levels of genetic variability and differentiation in hilsa shad, *Tenualosa ilisha* (Clupeidae, Clupeiformes) populations revealed by PCR-RFLP analysis of the mitochondrial DNA Dloop region. *Genetics and Molecular Biology*, 32(1):190-196.
- Meyer, A. (1994). Shortcomings of the cytochrome b gene as a molecular marker. TREE. 9:278-280.
- Miller, K. M., Laberee, K., Schulze, A. D., and Kaukinen, K. H. (2001). Development of microsatellite loci in Pacific herring (*Clupea pallasi*). *Molecular Ecology Notes*, 1:131-132.
- Milton, D. A., Chenery, S. R., Farmer, M. J., and Blaber, S. J. M. 1997. Identifying the spawning estuaries of the tropical shad, terubok *Tenualosa toli*, using otolith microchemistry. *Marine Ecology Progress Series*, 153:283-291.
- Nandeibam, S. S. Behera, B. K., and Sharma, A. P. (2013). Population structure of *Puntius sophore* inferred from variation in mitochondrial DNA sequences. *International Journal of Research in Fisheries and Aquaculture*, 3(3):112-115.
- Nguyen, T. T., Ingram, B., Sungan, S., Gooley, G., Sim, S. Y., Tinggi, D., and De Silva, S. S. (2006). Mitochondrial DNA diversity of broodstock of two indigenous mahseer species, *Tor tambroides* (Cyprinidae) cultured in Sarawak, Malaysia. *Aquaculture*, 253(1):259-269.
- Norouzi, M., and Samiei, M. H. (2015). Cross-species amplification of Clupeidae microsatellite DNA in common kilka, *Cluepeonella cultriventris* from the Caspian sea. *Caspian Journal of Environmental Science*, 13(2):167-171.
- Nurasyikin, A.R., and Jumat, S. (2006) Physicochemical characteristics of Terubok, *Tenualosa toli* Fish oil. *Malaysia Journal of Analytical Sciences*, 10(1):115-120.
- O'Brien, S. J. (1991). Molecular genome mapping: lessons and prospects. *Current Opinion in Genetic Development*, 1(1):105-111.
- Ogden, R. (2008). Fisheries forensics: the use of DNA tools for improving compliance, traceability and enforcement in the fishing industry. *Fish and Fisheries*, 9:462-472.
- Pääbo, S., and Wilson, A. C. (1988). Polymerase chain reaction reveals cloning artefacts. *Nature*, 334:387-388.
- Pang, S. C. (1991). Stock assessment of the *Hilsa toli* fishery and its management implications in the Batang Lupar estuary, Sarawak, Malaysia. M. Sc. Thesis, University of York, York.
- Parson, W., Pegoraro, K., Niederstatter, H, Foger, M., and Steinlechner, M. (2000). Species identification by means of the cytochrome *b* gene. *International Journal of Legal Medicine*, 114:23-28.

- Pepe, T., Trotta, M., Marco, I. D., Cennamo, P., Anastasio, A., and Cortesi, M. L. (2005). Mitochondrial cytochrome b DNA sequence variations: An approach to fish species identification in processed fish products. *Journal of food* protection, 68(2):421-425.
- Philippe, H., Forterre, P., (1999). The rooting of the universal tree of life is not reliable. *Journal of Molecular Evolution*, 49:509-523.
- Pritchard, D. W. (1967). Observations of circulation in coastal plain estuaries, In: Estuaries.
- Quinn, T. P. (1994). Shortcomings of the cytochrome b gene as a molecular marker. *Tree*, 9(8):278-280.
- Provost, M. M., and Jensen, O. P. (2012). Management and assessment of sex changing fishes. PMAFPS protogynous hermaphrodite modelling worlshop. Institute of Marine and Coastal Sciences, Rutgers University, New Brunswick.
- Rahman, M., and Naevdal, G. (2000). Population genetic studies of hilsa shad, *Tenualosa ilisha* (Hamilton), in Bangladesh waters: evidence for the existence of separate gene pools. *Fisheries Management and Ecology*, 7:401-411.
- Rogers, A. R., and Harpending, H. (1992). Population growth curves in the distribution of pairwise genetic differences. *Molecular Biology Evolution*. 9:552-569.
- Saitou, N., and Nei, T. (1987). The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Molecular Biology and Evolution*, 4(4):406-425.
- Salini, J. P., Milton, D. A., Rahman, M. J., and Hussain, M. G. (2004). Allozyme and morphological variation throughout the geographic range of the tropical shad, hilsa *Tenualosa ilisha*. *Fisheries Research*, 66(1):53-69.
- Santa Brigida, E. L., Erikson Luiz, D. B., Cunha, P. S., Rego, I., Sampaio, I., Schneider, H., and Vallinoto, M. (2007). Population analysis of Scomberomorus cavalla (Cuvier, 1829)(Perciformes, Scombridae) from the Northern and Northeastern coast of Brazil. *Brazilian Journal of Biology*, 67(4):919-924
- Saraswat, D., Lakra, W. S., Nautiyal, P., Goswami, M., Shyamakant, K., and Malakar, A. (2014). Genetic characterization of *Clupisoma garua* (Hamilton 1822) from six Indian populations using mtDNA cytochrome b gene. *Mitochondrial DNA*, 25(1):70-77.
- Sharma, M., Fomda, B. A., Mazta, S., Sehgal, R., Bagicha Singh, B., and Malla, N. (2013). Genetic diversity and population genetic structure analaysis of *Echinococcus granulosus* sensu strict Comples Based on Mitochondrial DNA signature. *PLOS One*, 8(12).

- Shifat, R., Begum, A.,and Khan, H. (2003). Use of RAPD fingerprinting for discriminating two populations of Hilsa shad (*Tenualosa ilisha* Ham.) from inland rivers of Bangladesh. Journal of Biochemistry and Molecular Biology, 36(5):462-467.
- Sidhu, K. S. (2003). Health benefits and potential risks related to consumption of fish or fish oil. *Regulatory Toxicology and Pharmacology*, 38:336-344.
- Sikorski, Z. E. (1990). *Seafood: Resources, Nutritional composition and preservation.* New York: CRC Press.
- Sivasundar, A., Birmingham, E and Orti, G. (2001). Population structure and biogeography of migratory freshwater fishes (Prochilodus: Characiformes) in major South American rivers. Molecular Ecology, 10:407-417.
- Slatkin, M. (1987). Gene flow and geographic structure of natural populations. *Science*, 236:787-792
- Srinivas, P. R. (2012). Introduction to protein electrophoresis. *Protein electrophoresis: Methods and Protocols*, 23-28.
- Tajima, F., and Nei, M. (1984). Estimation of evolutionary distance between nucleotide sequences. *Molecular Biology and Evolution*, 1(3):269-285.
- Tajima, F. (1989). Statistical method for testing the neutral mutation hypothesis by DNA polymorphism. *Genetics*, 123(3):585-595.
- Tamura, K., and Nei, M. (1993). Estimation of the number of nucleotide substitutions in the control region of mitochondrial DNA in humans and chimpanzees. *Molecular Biology and Evolution*, 10(3),512-536.
- Teletchea, F. (2009). Molecular identification methods of fish species: reassessment and possible applications. *Reviews in Fish Biology and Fisheries*, 19:265-293.
- The State of World Fisheries and Aquaculture: Opportunities and challenges 2014. (2016). Rome: Food and Agriculture Organization of the United Nations (FAO).223p
- The State of World Fisheries and Aquaculture: Opportunities and challenges 2003. (2003). Rome: Food and Agriculture Organization of the United Nations (FAO).223p
- Toews, D. P. L., and Brelsford, A. (2012). The biogeography of mitochondrial and nuclear discordance in animals. *Molecular Ecology*, 21:3907-3930.
- Van de Peer, Y., and De Wachter, R. (1994). TREECON for Windows: A software package for the construction and drawing and evolutionary tree for the Microsoft Windows environment. *Computer Applications in the Biosciences*, 10:569-570.

- Vidthayanon, C. (2005). Thailand red data: fishes. Bangkok, Thailand: Office of *Natural Resources and Environmental Policy and Plannning*.
- Ward, R. D. (2000). Genetics in fisheries management. Hydrobiologia, 420:191-201
- Ward, R. D., and Grewe, P. M. (1994). Appraisal of molecular genetic techniques in fisheries. *Reviews in Fish Biology and Fisheries*, 4:300-325.
- Ward, R. D., Zemlak, T. S., Innes, B. H., Last, P. R., and Hebert, P. D N. (2005). DNA barcoding Australia's fish species. *Philosophical Transactions of the Royal Society B*, 360:1847-1857.
- Whelan, S., Lio, P., and Goldma, N. (2001). Molecular phylogenetics: state-of-the-art methods for looking into the past. *Trends in Genetics*, 17(5):262-272.
- Whitehead, PJP. (1985). FAO species catalogue Vol.70. Clupeoid fishes of the world (suborder Clupeoidei). An annotated and illustrated catalogue of the herrings, sardines, pilchards, sprats, shads, anchovies and wolf herrings. Part1- Chirocentridae, Clupeidae and Pristigasteridae, FAO Fisheries Synopsis. 125(7/1):1-303.
- Wiley, E. O. (1978). The evolutionary species concept reconsidered. Systematic Zoology, 27:17-26
- Willman, R., Melvin, G., Jiram, S., Hadil, R., Yong A. H., and Gabriel G. (1989). Proposal for the management of the *Tenualosa toli* fishery in Sarawak. Techn Co-op Programme FAO-Malaysia. FAO-UN Rome F1: TCP/MAL/6751(1)
- Wolstenholme, D. R. (1992). Animal mitochondrial DNA: structure and evolution. *International Review of Cytology*, 141:173-216.
- Wright, J. M. (1993). DNA fingerprinting in fishes. Biochemistry and Molecular Biology of fishes, 2:58-91.
- Xia, X., (2000). Data analysis in molecular biology and evolution. Kluwer Academic Publishers, Boston.
- Xia, X., Xie, Z., Salemi, M., Chen, M and Wang, Y. (2003). An index of substitution saturation and its application. Molecular Phylogenetics and Evolution, 26:1-7.
- Xia, X. (2013). DAMBE5: A comprehensive software package for data analysis in molecular biology and evolution. *Molecular Biology and Evolution*, 30(7), 1720-1728.
- Zane, L., Bargelloni, L., and Patarnello, T. (2002). Strategies for microsatellite isolation: a review. *Molecular Biology*, 11:1-16.

BIODATA OF STUDENT

Puvaneswari a/p Puvanasundram was born on the 21st of January 1989 in Hospital Taiping, Perak. She attended her early education in SK (P) Treacher Methodist (1995 – 2001). She then furthered her secondary education in SMK (P) Treacher Methodist (2001 – 2006). She attended her pre university studies (Form 6) in SMK St George Institution in Taiping, Perak (2007-2008). The student then attained her degree in Animal Resources Science and Management at Universiti Malaysia Sarawak (2009-2012). Her final year project was on the Phylogeography analysis of Flycatcher, a bird belonging to the family Muscicapidae inferred from cytochrome oxidase I. Then she pursued her studies in Master of Science (Genetics and Breeding) in Universiti Putra Malaysia, Serdang, Selangor.



LIST OF PUBLICATIONS

Paper journal

Puvaneswari, P. S., Esa, Y., Khairul Adha, A. R and Nurul Amin, S. M. (2018). Phylogeography and Population Structure of *Tenualosa toli* inferred from Cytochrome b Mitochondrial DNA Fragment. *Journal of Environmental Biology*. (Accepted)

Proceedings

Puvaneswari, P. S., Esa, Y., Khairul Adha, A. R and Nurul Amin, S. M. (2018). Phylogeography and Population Structure of *Tenualosa toli* inferred from Cytochrome b Mitochondrial DNA Fragment. In proceedings of 11th Malaysia Genetic Congress: Focus on Genes: Maximise on impact, 12 – 13 Auguest 2015, Perdana University, Serdang, Selangor, Malaysia.



UNIVERSITI PUTRA MALAYSIA

STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

ACADEMIC SESSION :

TITLE OF THESIS / PROJECT REPORT :

PHYLOGEOGRAPHY OF GENUS Tenualosa AND POPULATION STRUCTURE OF

Tenualosa toli (Valenciennes, 1847) INFERRED FROM CYTOCHROME B

MITOCHONDRIAL DNA

NAME OF STUDENT: PUVANESWARI A/P PUVANASUNDRAM

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

1. This thesis/project report is the property of Universiti Putra Malaysia.

- 2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
- 3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as :

*Please tick (V)



(Contain confidential information under Official Secret Act 1972).

(Contains restricted information as specified by the organization/institution where research was done).

I agree that my thesis/project report to be published as hard copy or online open access.

This thesis is submitted for :

PATENT

Embargo from_____ until _____ (date) (date)

Approved by:

(Signature of Student) New IC No/ Passport No.: (Signature of Chairman of Supervisory Committee) Name:

Date :

Date :

[Note : If the thesis is CONFIDENTIAL or RESTRICTED, please attach with the letter from the organization/institution with period and reasons for confidentially or restricted.]