



UNIVERSITI PUTRA MALAYSIA

**ENGINEERING INTEGRATIVE VECTORS BASED ON BACTERIOPHAGE
SITE SPECIFIC RECOMBINATION MECHANISM FOR HETEROLOGOUS
EXPRESSION OF SECRETED AND SURFACE-ANCHORED PROTEIN IN
*Lactococcus lactis***

INNANURDIANI KOKO

FBSB 2019 29



**ENGINEERING INTEGRATIVE VECTORS BASED ON BACTERIOPHAGE
SITE SPECIFIC RECOMBINATION MECHANISM FOR HETEROLOGOUS
EXPRESSION OF SECRETED AND SURFACE-ANCHORED PROTEIN IN
*Lactococcus lactis***

By

INNANURDIANI KOKO

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the
Degree of Master of Science**

July 2019

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in
fulfilment of the requirement for the degree of Master of Science

**ENGINEERING INTEGRATIVE VECTORS BASED ON BACTERIOPHAGE
SITE SPECIFIC RECOMBINATION MECHANISM FOR HETEROLOGOUS
EXPRESSION OF SECRETED AND SURFACE-ANCHORED PROTEIN IN
*Lactococcus lactis***

By

INNANURDIANI KOKO

July 2019

**Chairman : Raha Abdul Rahim, PhD
Faculty : Biotechnology and Biomolecular Sciences**

Bacterial integrating system allows integration of foreign DNA into bacterial host chromosome enabling stable expression of foreign gene for recombinant protein production. In this study, integrative expression vectors for secretion and surface display of heterologous protein in *Lactococcus lactis* have been successfully constructed based on the site-specific recombination mechanism, of temperate lactococcal phage TP901-1. Two variations of integrative vectors were constructed, denoted pS1-4, and pSD1-4, which allows the heterologous protein to be secreted into the extracellular environment or surface displayed on *L. lactis*, respectively. The integrative vectors composed of (i) P_{170} auto-inducible promoter or P_{nisA} inducible promoter, (ii) multiple cloning sites (MCS), (iii) TP901-1 bacteriophage attachment site, attP, (iv) signal peptide-encoding sequence USP45 fused with LEISSTCDA propeptide or SPK1 signal peptide for extracellular targeting, and (v) 344 amino acids of proteinase anchor domain (PrtP₃₄₄) for surface display application. A helper plasmid harbouring *int* gene, pNZint was also constructed to facilitate plasmid integration into the genome. A staphylococcal nuclease reporter gene was cloned into each of the constructed integrative vectors which were then successfully integrated into the *L. lactis* genome. Toluidine blue O-DNA assay and immunofluorescence microscopy data proved that the expressed nuclease was able to be secreted or anchored on the *L. lactis* cell wall. From the findings, signal peptide SPK1 was shown to be superior over USP45 in the secretion of Nuc, even though the USP45 was fused with LEISSTCDA propeptide which reportedly could enhance protein secretion. Meanwhile, the expression of Nuc showed that integrative vectors driven by P_{170} promoter have comparable strength to P_{nisA} promoter. However, the combination of P_{170} with USP45-LEISSTCDA fusion performed significantly worse than the other constructs for surface display of Nuc. Therefore, these newly constructed synthetic integrative vectors can be applied for the secretion and surface display of heterologous protein in *L. lactis*

for research or industrial purposes where strong and weak stable expression vectors are required.



© COPYRIGHT UPM

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk Ijazah Sarjana (Sains)

**PEMBINAAN VEKTOR INTEGRATIF BERDASARKAN MEKANISMA
REKOMBINASI KHUSUS BAKTERIOFAJ UNTUK EKSPRESI PROTEIN
HETEROLOG SECARA REMBESAN DAN PAPARAN PERMUKAAN DI
DALAM *Lactococcus lactis***

Oleh

INNANURDIANI KOKO

Julai 2019

Pengerusi : Raha Abdul Rahim, PhD
Fakulti : Bioteknologi dan Sains Biomolekul

Sistem integrasi bakteria membolehkan pengintegrasian DNA asing ke dalam kromosom hos bakteria yang membolehkan ekspresi gen asing dengan stabil untuk pembikinan protein rekombinan. Dalam kajian ini, vektor ungkapan terintegrasi untuk rembesan dan paparan permukaan protein heterolog dalam *Lactococcus lactis* telah berjaya dihasilkan berdasarkan mekanisma rekombinasi khusus bakteriofaj lactococcal TP901-1. Dua variasi vektor integratif telah dibina, dengan nama pS1-4, dan pSD1-4, yang membolehkan protein heterolog dirembes ke persekitaran ekstraselular atau diparparkan ke permukaan *L. lactis*. Vektor integratif yang dibina terdiri daripada (i) promoter teraruhkan auto, P_{170} atau promoter teraruh, P_{nisA} , (ii) tapak pengklonan berbilang, (MCS), (iii) tapak pelekat bakteriofaj TP901-1, attP, (iv) peptida isyarat dengan propeptida LEISSTCDA atau peptida isyarat SPK1 untuk pentargetan ekstraselular, dan (v) 344 asid amino pelekat domain protein, (PrtP₃₄₄) untuk aplikasi paparan permukaan sel. Plasmid pembantu yang membawa gen *int*, pNZint juga dibina untuk membantu integrasi plasmid ke dalam genom. Satu gen pelapor telah diklon ke dalam setiap vektor integratif yang dibina yang kemudiannya berjaya digabungkan ke dalam genom *L. Lactis*. Data yang diperoleh daripada asai toluidine blue O-DNA dan mikroskopi imunopendarfluor membuktikan bahawa Nuc yang terekspresi dapat dirembes atau berlabuh pada dinding sel *L. lactis*. Daripada hasil kajian ini, isyarat peptida SPK1 menunjukkan perembesan protein nuklease lebih unggul berbanding isyarat peptida USP45, walaupun USP45 telah digabungkan bersama propeptida LEISSTCDA yang dilaporkan dapat meningkatkan rembesan protein. Sementara itu, ungkapan Nuc menunjukkan bahawa vektor integratif yang didorong oleh promoter P_{170} mempunyai kekuatan setanding dengan promoter P_{nisA} . Walaubagaimanapun, Nuc yang diekspresikan oleh kombinasi P_{170} dengan gabungan USP45-LEISSTCDA jauh lebih buruk

daripada vektor-vektor lain yang telah dihasilkan untuk paparan permukaan Nuc. Oleh itu, vektor integratif sintetik yang baru dibina ini boleh digunakan untuk rembesan dan paparan permukaan protein heterolog bagi *L. lactis* untuk tujuan penyelidikan atau kegunaan industri di mana vektor ungkapan kuat dan lemah yang stabil adalah diperlukan.



ACKNOWLEDGEMENTS

Alhamdulillah, in the name of Allah, The Most Gracious, The Most Merciful, all the praises and thanks be to Allah who has graciously bestowed me the strength and guided me throughout this journey. I would like to take this opportunity to express my deepest gratitude to my supervisors, Prof. Dr. Raha Abdul Rahim and Dr. Adelene Song Ai Lian. They have been very supportive throughout my project by giving the guideline, encouragement and helpful advice in order to help me finish my lab work and this thesis preparation. My sincere appreciation also goes to Dr. Mas Jaffri Masarudin who served as my supervisory committee member and kindly provided valuable advice and suggestions for this work.

My sincere thanks and appreciation to my labmates in Bacterial Molecular Biology laboratory, Jeevan Nathan, Lee Chai Yan, Sarah Safwah, Nur Elina, Abdullah Munir, Aishah and Aqlili Riana for their helps and great cooperation. I would also like to express my gratitude to my family for their advice, encouragement, love, and understanding in all my life throughout these years. Last but not least, I would like to thank Ministry of Education and University Putra Malaysia for awarding me with MyMasters scholarship and Graduate Research Fellowship. A million thanks to all persons involved throughout this project and the thesis submission. Thanks to all, and only Allah will repay all of your kindness.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Raha Abdul Rahim, PhD

Professor

Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Chairman)

Adelene Song Ai Lian, PhD

Senior Lecturer

Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Member)

Mas Jaffri Masarudin, PhD

Associate Professor

Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature : _____ Date: _____

Name and Matric No.: _____

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____

Name of Chairman
of Supervisory
Committee: _____

Signature: _____

Name of Member of
Supervisory
Committee: _____

Signature: _____

Name of Member of
Supervisory
Committee: _____

REFERENCES

- Agarwal, P., Khatri, P., Billack, B., Low, W. K., & Shao, J. (2014). Oral delivery of Glucagon like peptide-1 by a recombinant *Lactococcus lactis*. *Pharmaceutical Research*, 31(12), 3404–3414.
- Almeida, J. F., Breyner, N. M., Mahi, M., Ahmed, B., Benbouziane, B., Boas, P. C. B. V., Chatel, J. M. (2016). Expression of fibronectin binding protein A (FnBPA) from *Staphylococcus aureus* at the cell surface of *Lactococcus lactis* improves its immunomodulatory properties when used as protein delivery vector. *Vaccine*, 34(10), 1312–1318.
- Asseldonk, M. Van, Vos, W. M. De, & Simons, G. (1993). Functional analysis of the *Lactococcus lactis* usp45 secretion signal in the secretion of a homologous proteinase and a heterologous a-amylase. *Molecular Genetics*, 1061, 428–434.
- Atlung, T., Nielsen, a, Rasmussen, L. J., Nellemann, L. J., & Holm, F. (1991). A versatile method for integration of genes and gene fusions into the lambda attachment site of *Escherichia coli*. *Gene*, 107(1), 11–17.
- Azevedo, M. S. P. De, Rocha, C. S., Pereira, V. B., & Junior, A. F. D. O. (2015). Prospective uses of recombinant *Lactococcus lactis* expressing both listeriolysin O and mutated internalin A from *Listeria monocytogenes* as a tool for DNA vaccination. *Genetics and Molecular Research : GMR*, 14(4), 18485–18493.
- Bahay-El-Din, M. (2012). *Lactococcus lactis*-based vaccines from laboratory bench to human use: an overview. *Vaccine*, 30(4), 685–690.
- Bahay-El-Din, M., Casey, P. G., Griffin, B. T., & Gahan, C. G. M. (2008). *Lactococcus lactis*-expressing listeriolysin O (LLO) provides protection and specific CD8(+) T cells against *Listeria monocytogenes* in the murine infection model. *Vaccine*, 26(41), 5304–5314.
- Baradaran, A., Sieo, C. C., Illias, R. M., Rahim, R. A., Yusoff, K., & Foo, H. L. (2013). Cloning and in silico characterization of two signal peptides from *Pediococcus pentosaceus* and their function for the secretion of heterologous protein in *Lactococcus lactis*. *Biotechnology Letters*, 35(2), 233–238.
- Berg, J. M., Tymoczko, J. L., Stryer, L. (2002). *Biochemistry*, 5th Edition. W. H. Freeman, New York.
- Bermúdez-Humarán, L. G., Cortes-Perez, N. G., L'Haridon, R., & Langella, P. (2008). Production of biological active murine IFN-gamma by recombinant *Lactococcus lactis*. *FEMS Microbiology Letters*, 280(2), 144–149.
- Bermudez-Humaran, L. G., Langella, P., Commissaire, J., Gilbert, S., Loir, Y., L'Haridon, R., & Corthier, G. (2003). Controlled intra- or extracellular production of staphylococcal nuclease and ovine omega interferon in *Lactococcus lactis*. *FEMS Microbiology Letters*, 224(2), 307–313.
- Bermúdez-Humaran, L. G., Motta, J. P., Aubry, C., Kharrat, P., Rous-Martin, L., Sallenave, J. M., Langella, P. (2015). Serine protease inhibitors protect better than IL-10 and TGF-β anti-inflammatory cytokines against mouse colitis when delivered by recombinant lactococci. *Microbial Cell Factories*, 14(1), 1–11.
- Blatny, J. M., Godager, L., Lunde, M., & Nes, I. F. (2004). Complete genome sequence of the *Lactococcus lactis* temperate phage phiLC3: comparative analysis of phiLC3 and its relatives in lactococci and streptococci. *Virology*, 318(1), 231–244.
- Bok, J.-D., Maharjan, S., Kim, J.-I., Piao, D., Lee, H.-B., Kang, S.-K., Cho, C.-S. (2015). Soluble RANKL expression in *Lactococcus lactis* and investigation of its potential

- as an oral vaccine adjuvant. *BMC Immunology*, 16(1), 1–11.
- Bolotin, A., Wincker, P., Mauger, S., Jaillon, O., Malarme, K., Weissenbach, J., Sorokin, A. (2001). The complete genome sequence of the lactic acid bacterium *Lactococcus lactis* ssp. *lactis* IL1403. *Genome Research*, 731–753.
- Bosma, T., Kannenga, R., Neef, J., Audouy, S. A. L., Roosmalen, M. L. Van, Steen, A., Leenhouts, K. (2006). Novel surface display system for proteins on non-genetically modified gram-positive bacteria. *Applied and Environmental Microbiology*, 72(1), 880–889.
- Boyce, J. D., Davidson, B. E., & Hillier, A. J. (1995). Spontaneous deletion mutants of the *Lactococcus lactis* temperate bacteriophage BK5-T and localization of the BK5-T attP site. *Applied and Environmental Microbiology*, 61(11), 4105.
- Breüner, A., Brøndsted, L., & Hammer, K. (2001). Resolvase-like recombination performed by the TP901-1 integrase. *Microbiology*, 147(Pt 8), 2051–2063.
- Brøndsted, L., & Hammer, K. (1999). Use of the integration elements encoded by the temperate lactococcal bacteriophage tp901-1 to obtain chromosomal single-copy transcriptional fusions in *Lactococcus lactis*. *Applied and Environmental Microbiology*, 65(2), 752.
- Buist, G., Steen, A., Kok, J., & Kuipers, O. P. (2008). LysM, a widely distributed protein motif for binding to (peptido)glycans. *Molecular Microbiology*, 68(4), 838–847.
- Caplice, E., & Fitzgerald, G. F. (1999). Food fermentations: role of microorganisms in food production and preservation. *International Journal of Food Microbiology*, 50(1–2), 131–149.
- Chamcha, V., Jones, A., Amara, R. R., Scott, J. R., & Quigley, B. R. (2015). Oral immunization with a recombinant *Lactococcus lactis* –expressing HIV-1 antigen on Group A *Streptococcus Pilus* induces strong mucosal immunity in the gut. *The Journal of Immunology*, 195(10), 5025–5034.
- Chapot-Chartier, M.-P., & Kulakauskas, S. (2014). Cell wall structure and function in lactic acid bacteria. *Microbial Cell Factories*, 13(Suppl 1), S9.
- Chien, L. J., & Lee, C. K. (2007). Hyaluronic acid production by recombinant *Lactococcus lactis*. *Applied Microbiology and Biotechnology*, 77(2), 339–346.
- Cho, H.-J., Shin, H.-J., Han, I.-K., Jung, W.-W., Kim, Y. B., Sul, D., & Oh, Y.-K. (2007). Induction of mucosal and systemic immune responses following oral immunization of mice with *Lactococcus lactis* expressing human papillomavirus type 16 L1. *Vaccine*, 25(47), 8049–8057.
- Cho, M.-H., Park, S.-E., Lee, M.-H., Ha, S.-J., Kim, H.-Y., Kim, M.-J., Park, C.-S. (2007). Extracellular secretion of a maltogenic amylase from *Lactobacillus gasseri* ATCC33323 in *Lactococcus lactis* MG1363 and its application on the production of branched maltooligosaccharides. *Journal of Microbiology and Biotechnology*, 17(9), 1521–1526.
- Chopin, M. C., Chopin, a, Rouault, a, & Galleron, N. (1989). Insertion and amplification of foreign genes in the *Lactococcus lactis* subsp. *lactis* chromosome. *Applied and Environmental Microbiology*, 55(7), 1769–1774.
- Christiansen, B., Johnsen, M. G., Stenby, E., Vogensen, F. K., & Hammer, K. (1994). Characterization of the lactococcal temperate phage TP901-1 and its site-specific integration. *Journal of Bacteriology*, 176(4), 1069–1076.
- Cone, J. L., Cusumano, C. L., Hiroshi, T., & Anfinsen, C. B. (1971). Staphylococcal nuclease (Foggi strain): II. THE AMINO ACID SEQUENCE. *Journal of Biological Chemistry*, 246(10), 3103–3110.

- Cortes-Perez, N. G., Bermúdez-Humarán, L. G., Loir, Y., Rodriguez-Padilla, C., Gruss, A., Saucedo-Cárdenas, O., Montes-de-Oca-Luna, R. (2003). Mice immunization with live lactococci displaying a surface anchored HPV-16 E7 oncoprotein. *FEMS Microbiology Letters*, 229(1), 37–42.
- Cotton, F. A., Hazen, E. E., & Legg, M. J. (2006). Staphylococcal nuclease: Proposed mechanism of action based on structure of enzyme--thymidine 3',5'-bisphosphate--calcium ion complex at 1.5-A resolution. *Proceedings of the National Academy of Sciences*, 76(6), 2551–2555.
- de Ruyter, P. G., Kuipers, O. P., Beertshuyzen, M. M., van Alen-Boerrigter, I., & de Vos, W. M. (1996). Functional analysis of promoters in the nisin gene cluster of *Lactococcus lactis*. *Journal of Bacteriology*, 178(12), 3434–3439.
- de Vos, W. M. (1999). Gene expression systems for lactic acid bacteria. *Ecology and Industrial Microbiology*, 2, 289–295.
- de Vos, Willem M. (2011). Systems solutions by lactic acid bacteria: from paradigms to practice. *Microbial Cell Factories*, 10 Suppl 1(Suppl 1), S2.
- Den Blaauwen, T., & Driessens, A. J. M. (1996). Sec-dependent preprotein translocation in bacteria. *Archives of Microbiology*, 165(1), 1–8.
- Derkx, P. M. F., Janzen, T., Sørensen, K. I., Christensen, J. E., Stuer-Lauridsen, B., & Johansen, E. (2014). The art of strain improvement of industrial lactic acid bacteria without the use of recombinant DNA technology. *Microbial Cell Factories*, 13(1), S5.
- Desvaux, M., Dumas, E., Chafsey, I., & Hébraud, M. (2006). Protein cell surface display in Gram-positive bacteria: From single protein to macromolecular protein structure. *FEMS Microbiology Letters*, 256(1), 1–15.
- Dieye, Y., Usai, S., Clier, F., Gruss, A., & Piard, J. C. (2001). Design of a protein-targeting system for lactic acid bacteria. *Journal of Bacteriology*, 183(14), 4157–4166.
- Duong, T., Miller, M. J., Barrangou, R., Azcarate-peril, M. A., & Klaenhammer, T. R. (2010). Construction of vectors for inducible and constitutive gene expression in *Lactobacillus*. *Microbial Biotechnology*, 4(3), 357–367.
- Eichenbaum, Z., Federle, M. J., Marra, D., de Vos, W. M., Kuipers, O. P., Kleerebezem, M., & Scott, J. R. (1998). Use of the lactococcal nisA promoter to regulate gene expression in gram-positive bacteria: comparison of induction level and promoter strength. *Applied and Environmental Microbiology*, 64(8), 2763–2769.
- F Lachica, R. V, Genigeorgis, C., & Hoeprich, P. D. (1971). TNase: Metachromatic agar-diffusion methods for detecting staphylococcal nuclease activity. *Applied Microbiology*, 21(4), 585–587.
- Fischetti, V. A., Pancholi, V., & Schneewind, O. (1990). Conservation of a hexapeptide sequence in the anchor region of surface proteins from Gram-positive cocci. *Molecular Microbiology*, 4(9), 1603–1605.
- Frantzen, C. A., Kleppen, H. P., & Holo, H. (2018). *Lactococcus lactis* diversity in undefined mixed dairy starter cultures as revealed by comparative genome analyses and targeted amplicon sequencing of epsD. *Applied and Environmental Microbiology*, 84(3), 1–15.
- Frossard, C. P., Steidler, L., & Eigenmann, P. A. (2007). Oral administration of an IL-10-secreting *Lactococcus lactis* strain prevents food-induced IgE sensitization. *Journal of Allergy and Clinical Immunology*, 119(4), 952–959.
- García-Fruitós, E. (2012). Lactic Acid Bacteria: a promising alternative for recombinant protein production. *Microbial Cell Factories*, 11, 157.

- Gasson, M. J. (1983). Plasmid complements of *Streptococcus lactis* NCDO 712 and other lactic streptococci after protoplast-induced curing. *Journal of Bacteriology*, 154(1), 1–9.
- Glick, B. R. (1995). Metabolic load and heterologous gene expression. *Biotechnology Advances*, 13(2), 247–261.
- GmbH, M. (2012). NICE ® Expression system for *Lactococcus lactis* The effective & easy to operate Nisin Controlled gene Expression system, 1–25.
- He, S., Gong, F., & Zhang, D. (2012). Food-grade selection markers in lactic acid bacteria. *TAF Preventive Medicine Bulletin*, 11(4), 499–510.
- Holo, H., & Nes, I. (1989). Transformation of *Lactococcus* by electroporation. *Methods in Molecular Biology*, 47, 195–199.
- Israelsen, H., Madsen, S. M., Vrang, A., Hansen, E. B., & Johansen, E. (1995). Cloning and partial characterization of regulated promoters from *Lactococcus lactis* Tn917-lacZ integrants with the new promoter probe vector, pAK80. *Applied and Environmental Microbiology*, 61(7), 2540–2547.
- Jannitire, L., Niaudet, B., Pierre, E., & Ehrlich, S. D. (1985). Stable gene amplification in the chromosome of *Bacillus subtilis*. *Gene*, 40, 47–55.
- Joan, S. S. X., Pui-Fong, J., Song, A. A. L., Chang, L. Y., Yusoff, K., AbuBakar, S., & Rahim, R. A. (2016). Oral vaccine of *Lactococcus lactis* harbouring pandemic H1N1 2009 haemagglutinin1 and nisP anchor fusion protein elevates anti-HA1 sIgA levels in mice. *Biotechnology Letters*, 38(5), 793–799.
- Jørgensen, C. M., Vrang, A., & Madsen, S. M. (2014). Recombinant protein expression in *Lactococcus lactis* using the P170 expression system. *FEMS Microbiology Letters*, 351(2), 170–178.
- Joseph, B. C., Pichaimuthu, S., Srimeenakshi, S., Murthy, M., Selvakumar, K., M, G., & Manjunath, S. R. (2015). An overview of the parameters for recombinant protein expression in *Escherichia coli*. *Journal of Cell Science and Therapy*, 6(5), 1000221.
- Kashket, E. R. (1987). Bioenergetics of lactic acid bacteria: cytoplasmic pH and osmotolerance. *FEMS Microbiology Letters*, 46(3), 233–244.
- Kim, D., Beck, B. R., Lee, S. M., Jeon, J., Lee, D. W., Lee, J. Il, & Song, S. K. (2016). Pellet feed adsorbed with the recombinant *Lactococcus lactis* BFE920 expressing SiMA antigen induced strong recall vaccine effects against *Streptococcus iniae* infection in olive flounder (*Paralichthys olivaceus*). *Fish and Shellfish Immunology*, 55, 374–383.
- Kleerebezem, M., Boekhorst, J., van Kranenburg, R., Molenaar, D., Kuipers, O. P., Leer, R., Siezen, R. J. (2003). Complete genome sequence of *Lactobacillus plantarum* WCFS1. *Proceedings of the National Academy of Sciences*, 100(4), 1990–1995.
- Kobierecka, P. A., Olech, B., Ksiazek, M., Derlatka, K., Adamska, I., Majewski, P. M., Wyszyńska, A. K. (2016). Cell wall anchoring of the Campylobacter antigens to *Lactococcus lactis*. *Frontiers in Microbiology*, 7(FEB), 1–18.
- Kolter, R. (1993). The stationary phase of the bacterial life cycle. *Annual Review of Microbiology*, 47(1), 855–874.
- Konings, W. N., Kok, J., Kuipers, O. P., & Poolman, B. (2000). Lactic acid bacteria: the bugs of the new millennium. *Current Opinion in Microbiology*, 3(3), 276–282.
- Kuipers, O. P., Beertshuyzen, M. M., de Ruyter, Pascale G. G. A. Luesink, E. J., & de Vos, W. M. (1995). Autoregulation of nisin biosynthesis in *Lactococcus lactis* by

- signal transduction. *Journal of Biological Chemistry*, 270(45), 27299–27304.
- Kuipers, O. P., de Ruyter, P. G., Kleerebezem, M., & de Vos, W. M. (1998). Quorum sensing-controlled gene expression in lactic acid bacteria. *Journal of Biotechnology*, 64(1), 15–21.
- Kunji, E. R. S., Slotboom, D-J., Poolman B. (2003). *Lactococcus lactis* as host for overproduction of functional membrane proteins. *Biochimica et Biophysica Acta (BBA) - Biomembranes*, 1610 (1), 97-108.
- Kylä-Nikkilä, K., Alakuijala, U., & Saris, P. E. J. (2010). Immobilization of *Lactococcus lactis* to cellulosic material by cellulose-binding domain of *Celvibrio japonicus*. *Journal of Applied Microbiology*, 109(4), 1274–1283.
- Laemmli, U. K. (1970). Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature*, 228(October 17th), 227–231.
- Le Loir, Y., Azevedo, V., Oliveira, S. C., Freitas, D. a, Miyoshi, A., Bermúdez-Humarán, L. G., Langella, P. (2005). Protein secretion in *Lactococcus lactis* : an efficient way to increase the overall heterologous protein production. *Microbial Cell Factories*, 4(1), 2.
- Leenhousts, K., Buist, G., & Kok, J. (1999). Anchoring of proteins to lactic acid bacteria. *Antonie van Leeuwenhoek*, 76, 367–376.
- Leenhousts, K. J., Kok, J. A. N., & Venema, G. (1989). Campbell-like integration of heterologous plasmid DNA into the chromosome of *Lactococcus lactis* subsp. *lactis*. *Applied and Environmental Microbiology*, 55(2), 394.
- Leenhousts, K. J., Kok, J., & Venema, G. (1990). Stability of integrated plasmids in the chromosome of *Lactococcus lactis*. *Applied and Environmental Microbiology*, 56(9), 2726–2735.
- Lei, H., Peng, X., Zhao, D., Ouyang, J., Jiao, H., & Shu, H. (2014). *Lactococcus lactis* displayed neuraminidase confers cross protective immunity against influenza A viruses in mice. *Virology*, 476(2015), 189–195.
- Liang, X., Zhang, L., Zhong, J., & Huan, L. (2007). Secretory expression of a heterologous nattokinase in *Lactococcus lactis*. *Applied Microbiology and Biotechnology*, 75(1), 95–101.
- Lim, S. H. E., Jahanshiri, F., Rahim, R. A., Sekawi, Z., & Yusoff, K. (2010). Surface display of respiratory syncytial virus glycoproteins in *Lactococcus lactis* NZ9000, (i), 658–664.
- Linares, D. M., Kok, J., & Poolman, B. (2010). Genome sequences of *Lactococcus lactis* MG1363 (revised) and NZ9000 and comparative physiological studies. *Journal of Bacteriology*, 192(21), 5806–5812.
- Lindholm, A., Smeds, A., & Palva, A. (2004). Receptor binding domain of *Escherichia coli* F18 fimbrial adhesin FedF can be both efficiently secreted and surface displayed in a functional form in *Lactococcus lactis*. *Applied and Environmental Microbiology*, 70(4), 2061–2071.
- Liu, K. F., Liu, X. R., Li, G. L., Lu, S. P., Jin, L., & Wu, J. (2016). Oral administration of *Lactococcus lactis*-expressing heat shock protein 65 and tandemly repeated IA2P2 prevents type 1 diabetes in NOD mice. *Immunology Letters*, 174, 28–36.
- Liu, S., Li, Y., Deng, B., & Xu, Z. (2016). Recombinant *Lactococcus lactis* expressing porcine insulin-like growth factor I ameliorates DSS-induced colitis in mice. *BMC Biotechnology*, 16(1), 1–8.
- Llull, D., & Poquet, I. (2004). New expression system tightly controlled by zinc availability in *Lactococcus lactis*. *Applied and Environmental Microbiology*, 70(9),

5398–5406.

- Loir, Y. L. E., Gruss, A., & Ehrlich, S. D. (1998). A nine-residue synthetic propeptide enhances secretion efficiency of heterologous proteins in *Lactococcus lactis*. *Journal of Bacteriology*, 180(7), 1895–1903.
- Loir, Y. L. E., Gruss, A., Ehrlich, S. D., & Langella, P. (1994). Direct screening of recombinants in gram-positive bacteria using the secreted staphylococcal nuclease as a reporter, 176(16), 5135–5139.
- Loir, Y. L. E., Nouaille, S., Commissaire, J., Bretigny, L., Gruss, A., & Langella, P. (2001). Signal peptide and propeptide optimization for heterologous protein secretion in *Lactococcus lactis*. *Applied and Environmental Microbiology*, 67(9), 4119–4127.
- Lupfer, C., Pastey, M., Johnson, R. C., Reese, K. A., Geller, B. L., Mitev, G. M., & Mullen, V. M. (2013). A novel lactococcal vaccine expressing a peptide from the M2 antigen of H5N2 highly pathogenic avian influenza a virus prolongs survival of vaccinated chickens. *Veterinary Medicine International*, 2013(February), 1–8.
- Madsen, S M, Arnau, J., Vrang, a, Givskov, M., & Israelsen, H. (1999). Molecular characterization of the pH-inducible and growth phase-dependent promoter P170 of *Lactococcus lactis*. *Molecular Microbiology*, 32(1), 75–87.
- Madsen, Søren M, Hindré, T., Le Pennec, J.-P., Israelsen, H., & Dufour, A. (2005). Two acid-inducible promoters from *Lactococcus lactis* require the cis-acting ACiD-box and the transcription regulator RcfB. *Molecular Microbiology*, 56(3), 735–746.
- Makarova, K., Slesarev, A., Wolf, Y., Sorokin, A., Mirkin, B., Koonin, E., Mills, D. (2006). Comparative genomics of the lactic acid bacteria. *Proceedings of the National Academy of Sciences of the United States of America*, 103(42), 15611–15616.
- Makrides, S. C. (1996). Strategied of achieveing high-level expression of genes in *Escherichia coli*. *Microbiological Reviews*, 60(3), 512–538.
- Mayo, B., Sinderen, D., & Ventura, M. (2008). Genome Analysis of Food Grade Lactic Acid-Producing Bacteria: From Basics to Applications. *Current Genomics*, 9(3), 169–183.
- McCann, M. P., Kidwell, J. P., & Matin, A. (1989). The putative sigma factor KatF has a central role in development of starvation-mediated general resistance in *Escherichia coli*. *Journal of Bacteriology*, 8(1), 3177–3185.
- Mckay, L. L. (1983). Functional properties of plasmids in lactic streptococci. *Antonie van Leeuwenhoek*, 49, 259–274.
- Michon, C., Langella, P., Eijsink, V. G. H., Mathiesen, G., & Chatel, J. M. (2016). Display of recombinant proteins at the surface of lactic acid bacteria: Strategies and applications. *Microbial Cell Factories*, 15(1), 1–16.
- Mierau, I., & Kleerebezem, M. (2005). 10 years of the nisin-controlled gene expression system (NICE) in *Lactococcus lactis*. *Applied Microbiology and Biotechnology*, 68(6), 705–717.
- Miller, W. G., & Lindow, S. E. (1997). An improved GFP cloning cassette designed for prokaryotic transcriptional fusions. *Gene*, 191, 149–153.
- Miyoshi, A., Poquet, I., Azevedo, V., Commissaire, J., Bermudez-Humaran, L., Domakova, E., Langella, P. (2002). Controlled production of stable heterologous proteins in *Lactococcus lactis*. *Applied and Environmental Microbiology*, 68(6), 3141–3146.
- Miyoshi, Anderson, Bermúdez-Humarán, L. G., Ribeiro, L. a, Le Loir, Y., Oliveira, S. C., Langella, P., & Azevedo, V. (2006). Heterologous expression of *Brucella abortus*

- GroEL heat-shock protein in *Lactococcus lactis*. *Microbial Cell Factories*, 5, 14.
- Morello, E., Bermúdez-Humarán, L. G., Llull, D., Solé, V., Miraglio, N., Langella, P., & Poquet, I. (2008). *Lactococcus lactis*, an efficient cell factory for recombinant protein production and secretion. *Journal of Molecular Microbiology and Biotechnology*, 14(1–3), 48–58.
- Mori, H., & Ito, K. (2001). The Sec protein-translocation pathway. *Trends in Microbiology*, 9(10), 494–500.
- Mu, D., Montalbán-López, M., Masuda, Y., & Kuipers, O. P. (2013). Zirex: A novel zinc-regulated expression system for *Lactococcus lactis*. *Applied and Environmental Microbiology*, 79(14), 4503–4508.
- Natale, P., Brüser, T., & Driessens, A. J. M. (2008). Sec- and Tat-mediated protein secretion across the bacterial cytoplasmic membrane-distinct translocases and mechanisms. *Biochimica et Biophysica Acta*, 1778(9), 1735–1756.
- Navarre, W. W., & Schneewind, O. (1999). Surface proteins of gram-positive bacteria and mechanisms of their targeting to the cell wall envelope. *Microbiology and Molecular Biology*, 63(1).
- Ng, D. T. W., & Sarkar, C. a. (2013). Engineering signal peptides for enhanced protein secretion from *Lactococcus lactis*. *Applied and Environmental Microbiology*, 79(1), 347–356.
- Nijland, R., Lindner, C., van Hartskamp, M., Hamoen, L. W., & Kuipers, O. P. (2007). Heterologous production and secretion of *Clostridium perfringens* beta-toxin in closely related Gram-positive hosts. *Journal of Biotechnology*, 127(3), 361–372.
- Pavan, S., Hols, P., Delcour, J., Geoffroy, M., Granette, C., Kleerebezem, M., & Mercenier, A. (2000). Adaptation of the nisin-controlled expression system in *Lactobacillus plantarum*: a tool to study in vivo biological effects. *Applied and Environmental Microbiology*, 66(10), 4427–4432.
- Perez, T., Balcazar, J. L., Peix, A., Valverde, A., Velazquez, E., de Blas, I., & Ruiz-Zarzuela, I. (2011). *Lactococcus lactis* subsp. *tructae* subsp. nov. isolated from the intestinal mucus of brown trout (*Salmo trutta*) and rainbow trout (*Oncorhynchus mykiss*). *International Journal Of Systematic And Evolutionary Microbiology*, 61(8), 1894–1898.
- Petersen, K. V., Martinussen, J., Jensen, P. R., & Solem, C. (2013). Repetitive, marker-free, site-specific integration as a novel tool for multiple chromosomal integration of DNA. *Applied and Environmental Microbiology*, 79(12), 3563–3569.
- Plaitbeuw, C., Simons, G., & Vos, W. M. D. E. (1994). Use of the *Escherichia coli* beta-glucuronidase (*gusA*) gene reporter gene for analyzing promoters in lactic acid bacteria. *Applied and Environmental Microbiology*, 60(2), 587–593.
- Pontes, D. S., de Azevedo, M. S. P., Chatel, J.-M., Langella, P., Azevedo, V., & Miyoshi, A. (2011). *Lactococcus lactis* as a live vector: heterologous protein production and DNA delivery systems. *Protein Expression and Purification*, 79(2), 165–175.
- Ravn, P., Jose' Arnau, Madsen, S. M., Vrang, A., & Israelsen, H. (2003). Optimization of signal peptide SP310 for heterologous protein production in *Lactococcus lactis*. *Microbiology*, 149(8), 2193–2201.
- Raya, R. R., Fremaux, C., Antoni, G. L. De, & Klaenhammer, T. R. (1992). Site-specific integration of the temperate bacteriophage phi-adh into the *Lactobacillus gasseri* chromosome and of the phage (*attP*) and bacterial (*attB*) attachment sites. *Journal of Bacteriology*, 174(17), 5584–5592.
- Robert, S., Van Huynegem, K., Gysemans, C., Mathieu, C., Rottiers, P., & Steidler, L. (2015). Trimming of two major type 1 diabetes driving antigens, GAD65 and IA-2,

- allows for successful expression in *Lactococcus lactis*. *Beneficial Microbes*, 6(4), 591–601.
- Rosano, G. L., & Ceccarelli, E. A. (2014). Recombinant protein expression in *Escherichia coli*: advances and challenges, 5(April), 1–17.
- Rupa, P., Monedero, V., & Wilkie, B. N. (2008). Expression of bioactive porcine interferon-gamma by recombinant *Lactococcus lactis*. *Veterinary Microbiology*, 129(1–2), 197–202.
- Sambrook, J., Fritsch, E.F. and Maniatis, T. (1989). Molecular cloning: A laboratory manual. Cold Spring Harbor Laboratory Press, New York.
- Schleifer, K. H. (1987). Recent changes in the taxonomy of lactic acid bacteria. *FEMS Microbiology Letters*, 46(3), 201–203.
- Schleifer, K. H., Kraus, J., Dvorak, C., Kilpper-Bälz, R., Collins, M. D., & Fischer, W. (1985). Transfer of *Streptococcus lactis* and related streptococci to the genus *Lactococcus* gen. nov. *Systematic and Applied Microbiology*, 6(2), 183–195.
- Schneewind, O., & Missiakas, D. M. (2012). Protein secretion and surface display in Gram-positive bacteria. *Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences*, 367(1592), 1123–1139.
- Schwab, C., Sørensen, K. I., & Gänzle, M. G. (2010). Heterologous expression of glycoside hydrolase family 2 and 42 β-galactosidases of lactic acid bacteria in *Lactococcus lactis*. *Systematic and Applied Microbiology*, 33(6), 300–307.
- Shortle, D. (1983). A genetic system for analysis of staphylococcal nuclease. *Gene*, 22, 181–189.
- Siezen, R. J. (1999). Multi-domain, cell-envelope proteinases of lactic acid bacteria. *Antonie van Leeuwenhoek, International Journal of General and Molecular Microbiology*, 76(1–4), 139–155.
- Sim, A. C. N., Lin, W., Tan, G. K. X., Sim, M. S. T., Chow, V. T. K., & Alonso, S. (2008). Induction of neutralizing antibodies against dengue virus type 2 upon mucosal administration of a recombinant *Lactococcus lactis* strain expressing envelope domain III antigen. *Vaccine*, 26, 1145–1154.
- Simşek, Ö., Sabanoğlu, S., Çon, A. H., Karasu, N., Akçelik, M., & Saris, P. E. J. (2013). Immobilization of nisin producer *Lactococcus lactis* strains to chitin with surface-displayed chitin-binding domain. *Applied Microbiology and Biotechnology*, 97(10), 4577–4587.
- Song, A. A. L., In, L. L. A., Lim, S. H. E., & Rahim, R. A. (2017). A review on *Lactococcus lactis*: From food to factory. *Microbial Cell Factories*, 16(1), 1–15.
- Song, D., & Gu, Q. (2009). Surface expression of *Helicobacter pylori* urease subunit B gene e fragment on *Lactococcus lactis* by means of the cell wall anchor of *Staphylococcus aureus* protein A. *Biotechnology Letters*, 31(7), 985–989.
- Steidler, L., Viaene, J., Fiers, W., & Remaut, E. (1998). Functional display of a heterologous protein on the surface of *Lactococcus lactis* by means of the cell wall anchor of *Staphylococcus aureus* protein A. *Applied and Environmental Microbiology*, 64(1), 342–345.
- Steidler, Lothar, Neirynck, S., Huyghebaert, N., Snoeck, V., Vermeire, A., Goddeeris, B., Remaut, E. (2003). Biological containment of genetically modified *Lactococcus lactis* for intestinal delivery of human interleukin 10. *Nature Biotechnology*, 21(7), 785–789.
- Stiles, M. E., & Holzapfel, W. H. (1997). Lactic acid bacteria of foods and their current taxonomy. *International Journal of Food Microbiology*, 36(1), 1–29.

- Subramaniam, M., Baradaran, A., Rosli, M. I., Rosfarizan, M., Khatijah, Y., & Raha, A. R. (2012). Effect of signal peptides on the secretion of β -cyclodextrin glucanotransferase in *Lactococcus lactis* NZ9000. *Journal of Molecular Microbiology and Biotechnology*, 22(6), 361–372.
- Terzaghi, B. E., & Sandine, A. W. E. (1975). Improved medium for lactic streptococci and their bacteriophages. *Applied Microbiology*, 29(6), 807–813.
- Theisen, M., Soe, S., Brunstedt, K., Follmann, F., Bredmose, L., Israelsen, H., Druilhe, P. (2004). A *Plasmodium falciparum* GLURP-MSP3 chimeric protein; expression in *Lactococcus lactis*, immunogenicity and induction of biologically active antibodies. *Vaccine*, 22(9–10), 1188–1198.
- Tjalsma, H., Antelmann, H., Jongbloed, J. D. H., Braun, P. G., Darmon, E., Dorenbos, R., van Dijken, J. M. (2004). Proteomics of protein secretion by *Bacillus subtilis*: Separating the “secrets” of the secretome. *Microbiology and Molecular Biology Reviews : MMBR*, 68(2), 207–233.
- van de Guchte, M., Daly, C., Fitzgerald, G. F., & Arendt, E. K. (1994). Identification of int and attP on the genome of lactococcal bacteriophage Tuc2009 and their use for site-specific plasmid integration in the chromosome of Tuc2009-resistant *Lactococcus lactis* MG1363. *Applied and Environmental Microbiology*, 60(7), 2324–2329.
- van de Guchte, M., van der Vossen, J. M., Kok, J., & Venema, G. (1989). Construction of a lactococcal expression vector: expression of hen egg white lysozyme in *Lactococcus lactis* subsp. *lactis*. *Applied and Environmental Microbiology*, 55(1), 224–228.
- van der Vossen, J. M., van der Lelie, D., & Venema, G. (1987). Isolation and characterization of *Streptococcus cremoris* Wg2-specific promoters. *Applied and Environmental Microbiology*, 53(10), 2452–2457.
- Vandenbroucke, K., Haard, H. De, Beirnaert, E., Dreier, T., Lauwereys, M., Huyck, L., Rottiers, P. (2009). Orally administered *L. lactis* secreting an anti-TNF Nanobody demonstrate efficacy in chronic colitis. *Mucosal Immunology*, 3(1), 49–56.
- Wang, M., Gao, Z., Zhang, Y., & Pan, L. (2016). Lactic acid bacteria as mucosal delivery vehicles: a realistic therapeutic option. *Applied Microbiology and Biotechnology*, 100(13), 5691–5701.
- Watson, N. E. D., Dunyak, D. S., Rosey, E. L., Slonczewski, J. L., & Olson, E. R. (1992). Identification of elements involved in transcriptional regulation of the *Escherichia coli* cad operon by external pH, 174(2), 530–540.
- Wegmann, U., O'Connell-Motherway, M., Zomer, A., Buist, G., Shearman, C., Canchaya, C., Kok, J. (2007). Complete genome sequence of the prototype lactic acid bacterium *Lactococcus lactis* subsp. *cremoris* MG1363. *Journal of Bacteriology*, 189(8), 3256–3270.
- Wernerus, H., & Ståhl, S. (2004). Biotechnological applications for surface-engineered bacteria. *Biotechnology and Applied Biochemistry*, 40(Pt 3), 209–228.
- Wu, C., Yang, G., & Bermu, L. G. (2006). Immunomodulatory effects of IL-12 secreted by *Lactococcus lactis* on Th1 / Th2 balance in ovalbumin (OVA)-induced asthma model mice, 6, 610–615.
- Wu, X., Zhao, C., Guo, Z., Hao, Y., Li, J., Shi, H., & Sun, Y. (2016). Genome Sequence of *Lactobacillus johnsonii* Strain W1, Isolated From Mice. *Genome Announcements*, 4(3), e00561-16.
- Xin, Y., Mu, Y., Kong, J., & Guo, T. (2019). Targeted and repetitive chromosomal integration enables high-level heterologous gene expression in *Lactobacillus*

- casei*. *Applied and Environmental Microbiology*, 85(9).
- Zeigler, D. R. (2002). Integration vectors for gram-positive bacteria. *Bacillus Genetic Stock Center, Catalog Strains, Seventh Edition*, 4.
- Zhang, Q., Zhong, J., & Huan, L. (2011). Expression of hepatitis B virus surface antigen determinants in *Lactococcus lactis* for oral vaccination. *Microbiological Research*, 166(2), 111–120.
- Zhang, R., Qiao, D., Peng, X., Duan, G., Shi, Q., Zhang, L., Fan, Q. (2018). A novel food-grade lactococcal expression system and its use for secretion and delivery of an oral vaccine antigen. *Journal of Chemical Technology and Biotechnology*, 93(6), 1655–1660.
- Zhu, D., Liu, F., Fu, Y., Xu, H., Saris, P. E. J., & Qiao, M. (2017). Enhanced heterologous protein productivity by genome reduction in *Lactococcus lactis* NZ9000. *Microbial Cell Factories*, 16(1), 1–13.
- Zhu, D., Liu, F., Xu, H., Bai, Y., Zhang, X., Saris, P. E. J., & Qiao, M. (2015). Isolation of strong constitutive promoters from *Lactococcus lactis* subsp. *lactis* N8. *FEMS Microbiology Letters*, 362(16), 1–6.
- Zhuang, Z., Wu, Z.-G., Chen, M., & Wang, P. G. (2008). Secretion of human interferon-beta 1b by recombinant *Lactococcus lactis*. *Biotechnology Letters*, 30(10), 1819–1823.