



UNIVERSITI PUTRA MALAYSIA

**BIOSYNTHESIS OF ZINC OXIDE NANOPARTICLES BY  
*Pichia kudriavzevii* GY1 AND THEIR BIOMEDICAL PROPERTIES**

**BOROUMANDMOGHADDAM AMIN**

**FBSB 2018 22**



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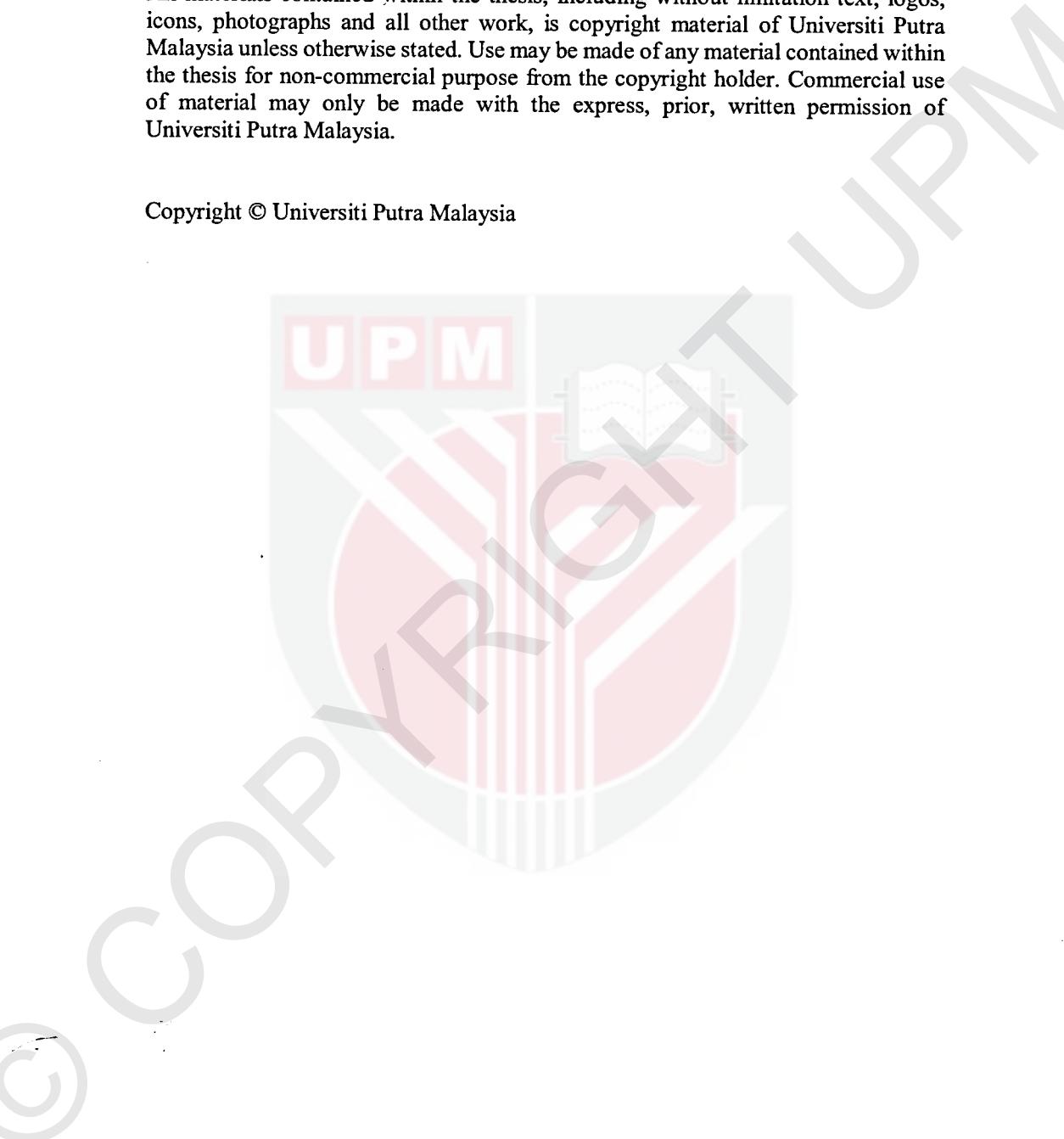
Thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of Philosophy

June 2018

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**June 2018**

**Chairman : Professor Rosfarizan Mohamad, PhD**  
**Faculty : Biotechnology and Biomolecular Sciences**

In the field of nanotechnology, the use of various biological materials instead of toxic chemicals for the synthesis of nanoparticles, has merited substantial attention. The use of toxic compounds limits their applications especially in biomedical field. To overcome the problem of toxicity in synthesis, safe eco-friendly green procedures have a major role for producing nanoparticles. Among the many possible bio resources, biologically active products from fungi and yeast represent high potential for this purpose. The present research aims to study the potential ability of a new yeast strain, *Pichia kudriavzevii*, in the synthesis of zinc oxide nanoparticles (ZnO-NPs) in order to evaluate their physical and biomedical properties.

Zinc oxide nanoparticles (ZnO-NPs) were synthesized by a green method using simple precursor from the solution consisting of zinc acetate and the fungal cell-free filtrate of *Pichia kudriavzevii* GY1. The effect of reaction time (12, 24 and 36 h) on the structure of the resulting ZnO nanoparticles was investigated. The samples were characterized using Fourier transform infrared (FTIR), ultraviolet-visible (UV-vis), x-ray diffraction (XRD), transmission electron microscope (TEM), field emission scanning electron microscope (FESEM), Photoluminescence (PL) and biomedical tests as free radical scavenging activity, cytotoxicity, antibacterial and anticancer activities.

From the XRD and TEM results, the ZnO-NPs with a hexagonal wurtzite structure and a particle crystal size of ~10–61 nm were formed at different reaction times. The most favourable nanosized structure with the lowest defect concentration was obtained at intermediate duration (24 h). The ZnO-NPs showed strong DPPH free

radical scavenging and a dose dependent toxicity with non-toxic effects on Vero cells for concentrations below 190  $\mu\text{g}/\text{mL}$ . The antibacterial tests reveal a favourable antibacterial effect for those ZnO-NPs prepared at intermediate duration.

The anticancer activity of biosynthesized ZnO-NPs on breast cancer cells (MCF-7) was determined using MTT assay. Cell cycle analysis and the mode of cell death were evaluated using a flow cytometry instrument. Quantitative real-time-PCR (qRT-PCR) was employed to investigate the expression of apoptosis in MCF-7 cells. ZnO NPs were cytotoxic to the MCF-7 cells in a dose-dependent manner. The 50% growth inhibition concentration ( $\text{IC}_{50}$ ) of ZnO NPs at 24 h was 121  $\mu\text{g}/\text{mL}$ . Cell cycle analysis revealed that ZnO NPs induced sub-G<sub>1</sub> phase (apoptosis), with values of 1.87% at 0  $\mu\text{g}/\text{mL}$  (control), 71.49% at  $\text{IC}_{25}$ , 98.91% at  $\text{IC}_{50}$ , and 99.44% at  $\text{IC}_{75}$ . Annexin V/propidium iodide (PI) flow cytometry analysis confirmed that ZnO NPs induce apoptosis in MCF-7 cells. The pro-apoptotic genes *p53*, *p21*, *Bax*, and *JNK* were upregulated, whereas anti-apoptotic genes *Bcl-2*, *AKT1*, and *ERK1/2* were downregulated in a dose-dependent manner. The arrest and apoptosis of MCF-7 cells were induced by ZnO NPs through several signalling pathways. Generally, the study has successfully contributed to the knowledge on the discovery of using fast-growing microorganism as a novel green approach in producing nanoparticles as new materials for the current innovation of bionanotechnology field particularly for medical applications.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**BIOSINTESIS NANOPARTIKEL ZINK OKSIDA OLEH *Pichia Kudriavzevii*  
GY1 DENGAN PENCIRIAN BIOPERUBUATAN**

Oleh

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**Jun 2018**

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**Fakulti : Bioteknologi dan Sains Biomolekul**

Dalam bidang nanoteknologi penggunaan pelbagai bahan biologi dan bukannya bahan kimia toksik bagi sintesis nanopartikel telah mendapat perhatian besar. Penggunaan sebatian toksik mengehadkan aplikasi mereka terutama dalam bidang bioperubatan. Bagi mengatasi masalah keracunan dalam sintesis, prosedur yang selamat dan mesra alam berperanan penting bagi penghasilan nanopartikel. Di antara kepelbagaiannya sumber biologi, produk bioaktif daripada kulat dan ragi mempunyai potensi tinggi untuk tujuan ini. Penyelidikan ini bertujuan untuk mengkaji potensi strain yis baru, *Pichia kudriavzevii*, dalam sintesis nanopartikel zink oksida (ZnO-NPs) bagi menilai sifat fizikal dan bioperubatan.

Nanopartikel zink oksida (ZnO-NPs) disintesis melalui kaedah mesra alam iaitu menggunakan prekursor mudah dari larutan yang terdiri daripada zink asetat dan filtrat bebas-kulat filtrat *Pichia kudriavzevii* GY1. Kesan masa tindakbalas (12, 24 dan 36 jam) pada struktur nanopartikel ZnO yang dihasilkan telah dikaji. Sampel dianalisis menggunakan Fourier transform infrared (FTIR), ultraviolet-visible (UV-vis), x-ray diffraction (XRD), transmission electron microscope (TEM), field emission scanning electron microscope (FESEM), Photoluminescence (PL) dan ujian biomedikal iaitu pengujian penangkapan radikal bebas, sitotoksik, aktiviti antibakteria dan antikanser.

Daripada hasil XRD dan TEM, ZnO-NP dengan struktur wurtzit heksagon dan ukuran kristal partikel di antara ~ 10-61 nm terbentuk pada masa tindakbalas yang berbeza. Struktur nanosaiz yang paling baik dengan kepekatan kecacatan terendah didapatkan pada tempoh pertengahan (24 jam). ZnO-NPs memperlihatkan penangkapan

radikal bebas DPPH dan ketoksikan yang bergantung kepada dos dengan kesan tidak toksik kepada sel Vero untuk kepekatan di bawah  $190 \mu\text{g} / \text{mL}$ . Ujian antibakteria menunjukkan kesan antibakteria yang baik untuk ZnO-NPs adalah pada tempoh pertengahan.

Aktiviti antikanser oleh ZnO-NPs pada sel-sel kanser payudara (MCF-7) ditentukan menggunakan assay MTT. Analisis kitaran sel dan mod kematian sel telah dinilai menggunakan instrumen aliran cytometry. *Quantitative real-time-PCR* (qRT-PCR) digunakan untuk menyiasat ekspresi apoptosis dalam sel-sel MCF-7. ZnO-NPs adalah sitotoksik kepada sel MCF-7 di mana bergantung kepada nilai dos. Nilai Kepekatan perencutan 50% pertumbuhan (IC50) oleh ZnO-NPs pada 24 jam adalah  $121 \mu\text{g} / \text{mL}$ . Analisis kitaran sel menunjukkan bahawa ZnO-NPs menggalakkan fasa sub-G1 (apoptosis), dengan nilai 1.87% pada  $0 \mu\text{g} / \text{mL}$  (kawalan), 71.49% pada IC25, 98.91% pada IC50, dan 99.44% di IC75. Analisa aliran sitometri Annexin V / propidium iodide (PI) mengesahkan bahawa ZnO-NPs telah menggalakkan apoptosis di dalam sel MCF-7. Gen-gen pro-apoptotik p53, p21, Bax, dan JNK dikawal selaras, manakala gen anti-apoptotik Bcl-2, AKT1, dan ERK1/2 dikurangkan dengan cara yang bergantung kepada dos. Penangkapan dan apoptosis sel MCF-7 yang digalakkan oleh ZnO-NPs dilaksanakan melalui beberapa laluan isyarat sel. Secara umumnya, kajian ini telah berjaya menyumbang kepada pengetahuan dalam penemuan menggunakan mikroorganisma yang tumbuh secara cepat sebagai kaedah hijau yang baharu bagi penghasilan partikel nano sebagai bahan baharu untuk inovasi terkini bidang nanobiotehnologi secara khususnya untuk aplikasi perubatan.

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## LIST OF ABBREVIATIONS

DMSO	Dimethyl sulfoxide
NPS	Nanoparticles
ZnO	Zinc Oxide
h	Hour
PDA	Potato dextrose agar
PDB	Potato dextrose broth
mg	Milligram
min	Minutes
mL	Milliliter
PI	Propidium IODID
ROS	Reactive oxygen species
rpm	Rotation per minutes
TEM	Transmission electron microscopy
FESEM	Field emission scanning electron microscopy
ICP-AES	Inductively Coupled Plasma-atomic Emission Spectroscopy
XRD	X-ray Diffraction Pattern
UV	Ultraviolet
MTT	3-(4, 5-dimethylthiazolyl-2)-2,5-diphenyltetrazolium bromide
DPPH	2,2-diphenyl-1-picrylhydrazyl
SDS	sodium dodecyl sulphate
°C	Degree Celsius
µ	Micro
%	Percentage
ATCC	American Type Culture Collection
FITC	Fluorescein isothiocyanate
IC <sub>50</sub>	Median lethal concentration
µg	Microgram
FBS	Fetal bovine serum
PBS	Phosphate buffered saline
DMEM	Dulbecco's Modification Eagle's Medium
PCR	Polymerase Chain Reaction

## CHAPTER 1

### INTRODUCTION

Nanotechnology provides many possibilities in various scientific and technological fields (Adibkia et al., 2007). Researchers are becoming increasingly interested in pharmacological nanotechnology due to its widespread applications. Nanoparticles can be produced by either chemical, physical and biotic processes (Alani et al., 2012). Green synthesis is an eco-friendly method for the production of nontoxic and biocompatible nanoparticles. These methods of NP synthesis are simple and efficient, and the products are safe for medical applications (Gopalakrishnan et al., 2012). In addition, molecular components of biological materials have great affinity for the surface of nano-structures, stabilize and prevent aggregation while improving the biological effects of NPs (Shabestarian et al., 2017). Biological procedures for nanoparticle synthesis using microorganisms (Taraftdar & Raliya, 2013; Gericke & Pinches, 2006; Liu et al., 2014; Ahmad et al., 2003), plants (Sadeghi & Gholamhoseinpoor, 2015), and alga (Singaravelu et al., 2007) have been proposed as feasible eco-friendly alternatives to chemical and physical approaches, because these methods are hazardous and costly (Sinha et al., 2009).

The potential use of yeasts in NP biosynthesis is promising because of the ease of handling yeasts under laboratory conditions, their abundant enzyme synthesis and rapid growth without the need of complex nutrients (Kumar et al., 2011; Yan et al., 2009). Economic longevity and capacity for employing biomass is yet another advantage of using the green approach employing yeasts to produce metallic NPs. Furthermore, numerous yeasts species grow at accelerated rates and therefore producing cultures as well as their storage in the laboratory is very practical (Dameron et al., 1989). Yeast biomasses are capable of producing metal nanoparticles and nanostructures through the reduction of proteins in enzymes either intracellularly or extracellularly (Castro, Blázquez et al., 2014). The reductive properties of proteins are related to many functional groups such as  $-\text{NH}_2$ ,  $-\text{OH}$ ,  $\text{C}=\text{O}$ ,  $\text{C}-\text{O}-\text{C}$ ,  $\text{C}-\text{N}-\text{C}$ , and  $-\text{S}-\text{S}-$ , which enable the donation of electrons to form NP from metal ions.

The biosynthesized ZnO-NPs are suitable to be used as drug carriers, cosmetics, and fillings in medicinal materials (Nagarajan & Kuppusamy, 2013). Zinc oxide (ZnO) shows biocompatibility compared to different materials (Arakelova et al., 2014). ZnO is an inorganic compound listed as “Generally Recognized as Safe” (GRAS) by the USA Food and Drug Administration (FDA) (21CFR182.8991) (Espitia et al., 2012).

Breast cancer is one of the most frequent types of cancers in women, and overall it is the second most common form of cancer worldwide. As many as 25% of cases of cancer (1.67 million cases) and 15% of deaths in women (522,000 individuals) were caused by forms of breast cancer, as reported by GLOBOCAN (Wolff et al., 2013).

The balance lost between the proliferated cells and apoptosis is a hallmark that intensifies the failure of damaged cells to be wiped out via apoptosis. Activating apoptotic paths in cells affected by tumour is a crucial approach to cancer therapy. Several natural products that are considered potentially powerful sources of anticancer drugs apply anti-tumour effects by inducing apoptosis (Elkady et al., 2012). Additionally, the deregulations that cause the initiation and development of cancers represent hundreds of genes or signalling cascades (Teiten et al., 2010). Furthermore, numerous genes like *p53*, *p21*, *JNK*, *Bax*, *Bcl-2*, *AKT*, and *ERK1/2* engage in apoptotic pathways.

ZnO nanoparticles with sizes less than 100 nm has been reported to possess good antimicrobial (Sirelkhatim et al., 2015), antioxidant (Nagajyothi et al., 2015) and antitumor (Selvakumari et al., 2015) activities. The technique for synthesizing ZnO nano-sized materials is a challenge for attaining standard medical therapies. Green and eco-friendly processes for the synthesis of ZnO-NPs are believed to be safe, nontoxic, and bio-compatible, for pharmaceutical and bio-medical applications as compared to chemical and physical methods which are hazardous and expensive. Moreover, the biologically active molecules involved in the green synthesis of NPs act as functionalizing ligands leads to the occurrence of succeeding different surface effects, controlling the size and structure of nanoparticles making these NPs more suitable for biomedical applications (Lu et al., 2007).

The main purpose of this study was used a new yeast strain, *Pichia kudriavzevii*, in the synthesis of ZnO-NPs through a green method for biomedical applications. In this study, it demonstrated a simple one-step green process to synthesize ZnO-NPs at different reaction durations using a newly isolated yeast strain, *Pichia kudriavzevii* GY1. *Pichia kudriavzevii* is synonymously known as *Issatchenka orientalis* and is an anamorph of *Candida krusei* (Carlotti et al., 1996; Kurtzman et al., 1980). This yeast strain has been isolated from food and fruit sources, such as sourdoughs (Meroth et al., 2003), fermented butter-like products (Ongol & Asano, 2009) and the starter culture of Tanzanian fermented togwa (Mugula et al., 2003).

To our knowledge, there is no report on the green synthesis of ZnO-NPs using *P. kudriavzevii* GY1 in order to evaluate their antibacterial, antioxidant, and antitumor activities. As a result, a better insight into the anticancer activities of ZnO NPs and the cytotoxic effects of their constituents can contribute to facilitating the improvement of auspicious cancer therapeutics for use in nanomedicine. Nanomedicine has recently emerged as a better choice for treating some common cancers, resulting in many nanoparticles being used as treatment in cancer cell lines.

Hence, the specific objectives were as follows:

1. To isolate and characterize the yeast strain of *Pichia kudriavzevii* GY1.
2. To determine the effect of biosynthesis reaction on the characterization of ZnO-NPs.
3. To evaluate the antibacterial, antioxidant and anticancer activity of biosynthesized ZnO-NPs.
4. To evaluate the gene regulation of biosynthesized ZnO-NPs on the MCF-7 cell line.

## REFERENCES

- Adibkia, K., Omidi, Y., Siah, M. R., Javadzadeh, A. R., Barzegar-Jalali, M., Barar, J., Nokhodchi, A. (2007). Inhibition of endotoxin-induced uveitis by methylprednisolone acetate nanosuspension in rabbits. *Journal of Ocular Pharmacology and Therapeutics*, 23(5), 421–432.
- Afifi, M., Almaghrabi, O. A., & Kadasa, N. M. (2015). Ameliorative effect of zinc oxide nanoparticles on antioxidants and sperm characteristics in streptozotocin-induced diabetic rat testes. *BioMed Research International*, 2015, 1-6.
- Agarwal, H., Kumar, S. V., & Rajeshkumar, S. (2017). A review on green synthesis of zinc oxide nanoparticles—An eco-friendly approach. *Resource-Efficient Technologies*. 3 (2017), 406-413
- Ahmad, A., Mukherjee, P., Mandal, D., Senapati, S., Khan, M. I., Kumar, R., & Sastry, M. (2002). Enzyme mediated extracellular synthesis of CdS nanoparticles by the fungus, *Fusarium oxysporum*. *Journal of the American Chemical Society*, 124(41), 12108–12109.
- Ahmad, A., Mukherjee, P., Senapati, S., Mandal, D., Khan, M. I., Kumar, R., & Sastry, M. (2003). Extracellular biosynthesis of silver nanoparticles using the fungus *Fusarium oxysporum*. *Colloids and Surfaces B: Biointerfaces*, 28(4), 313–318.
- Ahmad, A., Senapati, S., Khan, M. I., Kumar, R., & Sastry, M. (2003). Extracellular Biosynthesis of Monodisperse Gold Nanoparticles by a Novel Extremophilic *Actinomycete*, *Thermomonospora sp.* *Langmuir*, 19(8), 3550–3553.
- Ahmad, A., Senapati, S., Khan, M. I., Kumar, R., Ramani, R., Srinivac, V., and Sastry, M. (2003). Intracellular synthesis of gold nanoparticles by a novel alkalotolerant actinomycete, *Rhodococcus* species. *Nanotechnology*, 14(7), 824.
- Ahmadian, S., Barar, J., Saei, A. A., Fakhree, M. A. A., & Omidi, Y. (2009). Cellular toxicity of nanogenomedicine in MCF-7 cell line: MTT assay. *Journal of Visualized Experiments: JoVE*, (26) 1-2.
- Ahsan, H., Ali, A., & Ali, R. (2003). Oxygen free radicals and systemic autoimmunity. *Clinical & Experimental Immunology*, 131(3), 398–404.
- Aladpoosh, R., & Montazer, M. (2015). The role of cellulosic chains of cotton in biosynthesis of ZnO nanorods producing multifunctional properties: Mechanism, characterizations and features. *Carbohydrate Polymers*, 126, 122–129.
- Ali, K., Dwivedi, S., Azam, A., Saquib, Q., Al-Said, M. S., Alkhedhairy, A. A., & Musarrat, J. (2016). Aloe Vera extract functionalized zinc oxide nanoparticles

- as nanoantibiotics against multi-drug resistant clinical bacterial isolates. *Journal of colloid and interface science*, 472, 145-156.
- Al-Shabib, N. A., Husain, F. M., Ahmed, F., Khan, R. A., Ahmad, I., Alsharaeh, E., Yusuf, M. (2016). Biogenic synthesis of Zinc oxide nanostructures from *Nigella sativa* seed: Prospective role as food packaging material inhibiting broad-spectrum quorum sensing and biofilm. *Scientific Reports*, 6, 36761.
- Ambika, S., & Sundrarajan, M. (2015a). Antibacterial behaviour of *Vitex negundo* extract assisted ZnO nanoparticles against pathogenic bacteria. *Journal of Photochemistry and Photobiology B: Biology*, 146, 52-57.
- Ambika, S., & Sundrarajan, M. (2015). Green biosynthesis of ZnO nanoparticles using *Vitex negundo* L. extract: spectroscopic investigation of interaction between ZnO nanoparticles and human serum albumin. *Journal of photochemistry and photobiology B: biology*, 149, 143-148.
- Anbuvannan, M., Ramesh, M., Viruthagiri, G., Shanmugam, N., & Kannadasan, N. (2015). *Anisochilus carnosus* leaf extract mediated synthesis of zinc oxide nanoparticles for antibacterial and photocatalytic activities. *Materials Science in Semiconductor Processing*, 39, 621-628.
- Alani, F., Moo-Young, M., & Anderson, W. (2012). Biosynthesis of silver nanoparticles by a new strain of *Streptomyces* sp. compared with *Aspergillusfumigatus*. *World Journal of Microbiology and Biotechnology*, 28(3), 1081-1086.
- Alkhalfaf, M., & El-Mowafy, A. M. (2003). Overexpression of wild-type p53 gene renders MCF-7 breast cancer cells more sensitive to the antiproliferative effect of progesterone. *Journal of Endocrinology*, 179(1), 55-62.
- Alkaladi, A., Abdelazim, A. M., & Afifi, M. (2014). Antidiabetic activity of zinc oxide and silver nanoparticles on streptozotocin-induced diabetic rats. *International Journal of Molecular Sciences*, 15(2), 2015-2023.
- Ali, K., Dwivedi, S., Azam, A., Saquib, Q., Al-Said, M. S., Alkhedhairy, A. A., & Musarrat, J. (2016). *Aloe vera* extract functionalized zinc oxide nanoparticles as nanoantibiotics against multi-drug resistant clinical bacterial isolates. *Journal of Colloid and Interface Science*, 472, 145-156.
- Anker, J. N., Hall, W. P., Lyandres, O., Shah, N. C., Zhao, J., & Van Duyne, R. P. (2008). Biosensing with plasmonic nanosensors. *Nat Mater*, 7(6), 442-453.
- Aruoma, O. I., Halliwell, B., & Dizdaroglu, M. (1989). Iron ion-dependent modification of bases in DNA by the superoxide radical-generating system hypoxanthine/xanthine oxidase. *Journal of Biological Chemistry*, 264(22), 13024-13028.

- Asmathunisha, N., & Kathiresan, K. (2013). A review on biosynthesis of nanoparticles by marine organisms. *Colloids and Surfaces B: Biointerfaces*, 103, 283–287.
- Atkinson, A., & Winge, D. R. (2009). Metal acquisition and availability in the mitochondria. *Chemical Reviews*, 109(10), 4708–4721.
- Ambika, S., & Sundrarajan, M. (2015). Green biosynthesis of ZnO nanoparticles using *Vitex negundo L.* extract: spectroscopic investigation of interaction between ZnO nanoparticles and human serum albumin. *Journal of Photochemistry and Photobiology B: Biology*, 149, 143–148.
- Azam, A., Ahmed, A. S., Oves, M., Khan, M. S., Habib, S. S., & Memic, A. (2012). Antimicrobial activity of metal oxide nanoparticles against Gram-positive and Gram-negative bacteria: a comparative study. *IJNM*, (7(7)), 6003–6009.
- Azizi, S., Ahmad, M. B., Namvar, F., & Mohamad, R. (2014). Green biosynthesis and characterization of zinc oxide nanoparticles using brown marine macroalga *Sargassum muticum* aqueous extract. *Materials Letters*, 116, 275–277.
- Azizi, S., Ahmad, M., Mahdavi, M., & Abdolmohammadi, S. (2013). Preparation, characterization, and antimicrobial activities of ZnO nanoparticles/cellulose nanocrystal nanocomposites. *BioResources*, 8(2), 1841–1851.
- Azizi, S., Mohamad, R., Bahadoran, A., Bayat, S., Rahim, R. A., Ariff, A., & Saad, W. Z. (2016). Effect of annealing temperature on antimicrobial and structural properties of bio-synthesized zinc oxide nanoparticles using flower extract of *Anchusa italicica*. *Journal of Photochemistry and Photobiology B: Biology*, 161, 441–449.
- Azizi, S., Mohamad, R., & Mahdavi Shahri, M. (2017). Green Microwave-Assisted Combustion Synthesis of Zinc Oxide Nanoparticles with *Citrullus colocynthis* (L.) Schrad: Characterization and Biomedical Applications. *Molecules*, 22(2), 301.
- Azizi, S., Mahdavi Shahri, M., & Mohamad, R. (2017). Green Synthesis of Zinc Oxide Nanoparticles for Enhanced Adsorption of Lead Ions from Aqueous Solutions: Equilibrium, Kinetic and Thermodynamic Studies. *Molecules*, 22(6), 831.
- Azizi, S., Mohamad, R., Rahim, R. A., Moghaddam, A. B., Moniri, M., Ariff, A., Namvab, F. (2016). ZnO-Ag core shell nanocomposite formed by green method using essential oil of wild ginger and their bactericidal and cytotoxic effects. *Applied Surface Science*, 384, 517–524.
- Balavandy, S. K., Shameli, K., Biak, D. R. B. A., & Abidin, Z. Z. (2014). Stirring time effect of silver nanoparticles prepared in glutathione mediated by green method. *Chemistry Central Journal*, 8(1), 11.

- Bandow, J. E., Brötz, H., & Hecker, M. (2002). *Bacillus subtilis* tolerance of moderate concentrations of rifampin involves the σB-dependent general and multiple stress response. *Journal of Bacteriology*, 184(2), 459–467.
- Beech, F. W., & Davenport, R. R. (1971). Purification and maintenance of yeasts. In *Methods in microbiology*, 4, 153–182
- Bertram, J. S. (2000). The molecular biology of cancer. *Molecular Aspects of Medicine*, 21(6), 167–223.
- Beyermann, D. (2002). Homeostasis and cellular functions of zinc. *Materialwissenschaft Und Werkstofftechnik*, 33(12), 764–769.
- Bhattacharjee, S. (2016). DLS and zeta potential—What they are and what they are not? *Journal of Controlled Release*, 235, 337–351.
- Bhuyan, T., Mishra, K., Khanuja, M., Prasad, R., & Varma, A. (2015). Biosynthesis of zinc oxide nanoparticles from *Azadirachta indica* for antibacterial and photocatalytic applications. *Materials Science in Semiconductor Processing*, 32, 55–61.
- Bhardwaj, A., Rautaray, D., Bansal, V., Ahmad, A., Sarkar, I., Yusuf, S. M., ... Sastry, M. (2006). Extracellular biosynthesis of magnetite using fungi. *Small*, 2(1), 135–141.
- Bird, S. M., El-Zubir, O., Rawlings, A. E., Leggett, G. J., & Staniland, S. S. (2016). A novel design strategy for nanoparticles on nanopatterns: interferometric lithographic patterning of Mms6 biotemplated magnetic nanoparticles. *Journal of Materials Chemistry C*, 4(18), 3948–3955.
- Binupriya, A. R., Sathishkumar, M., & Yun, S.-I. (2010). Biocrystallization of silver and gold ions by inactive cell filtrate of *Rhizopus stolonifer*. *Colloids and Surfaces B: Biointerfaces*, 79(2), 531–534.
- Biosciences, B. D. (2011). Detection of Apoptosis Using the BD Annexin V FITC Assay on the BD FACSVers<sup>TM</sup> System.
- Birbrair, A., Zhang, T., Wang, Z. M., Messi, M. L., Olson, J. D., Mintz, A., & Delbono, O. (2014). Type-2 pericytes participate in normal and tumoral angiogenesis. *American Journal of Physiology-Cell Physiology*, 307(1), C25–C38.
- Birla, S. S., Tiwari, V. V., Gade, A. K., Ingle, A. P., Yadav, A. P., & Rai, M. K. (2009). Fabrication of silver nanoparticles by *Phoma glomerata* and its combined effect against *Escherichia coli*, *Pseudomonas aeruginosa* and *Staphylococcus aureus*. *Letters in Applied Microbiology*, 48(2), 173–179.
- Blois, M. S. (1958). Antioxidant determinations by the use of a stable free radical. *Nature*, 181(4617), 1199–1200.

- Botham, K. M., & Mayes, P. A. (2006). Biologic oxidation. *Harper's Illustrated Biochemistry*. London: Lange-McGraw Hill.
- Brayner, R. (2008). The toxicological impact of nanoparticles. *Nano Today*, 3(1), 48–55.
- Brayner, R., Ferrari-Iliou, R., Brivois, N., Djediat, S., Benedetti, M. F., & Fiévet, F. (2006). Toxicological impact studies based on *Escherichia coli* bacteria in ultrafine ZnO nanoparticles colloidal medium. *Nano Letters*, 6(4), 866–870.
- Brigger, I., Dubernet, C., & Couvreur, P. (2002). Nanoparticles in cancer therapy and diagnosis. *Advanced Drug Delivery Reviews*, 54(5), 631–651.
- Brunner, T. J., Wick, P., Manser, P., Spohn, P., Grass, R. N., Limbach, L. K., Stark, W. J. (2006). In vitro cytotoxicity of oxide nanoparticles: comparison to asbestos, silica, and the effect of particle solubility. *Environmental Science & Technology*, 40(14), 4374–4381.
- Burgess, R. (2009). Medical applications of nanoparticles and nanomaterials. *Stud Health Technol Inform*, 149, 257–283.
- Burlacu, A. (2003). Regulation of apoptosis by Bcl-2 family proteins. *Journal of Cellular and Molecular Medicine*, 7(3), 249–257.
- Buzea, C., Pacheco, I. I., & Robbie, K. (2007). Nanomaterials and nanoparticles: sources and toxicity. *Biointerphases*, 2(4), MR17-MR71.
- Cai, R., Kubota, Y., Shuin, T., Sakai, H., Hashimoto, K., & Fujishima, A. (1992). Induction of cytotoxicity by photoexcited TiO<sub>2</sub> particles. *Cancer Research*, 52(8), 2346–2348.
- Carlotti, A., Couble, A., Domingo, J., Miroy, K., & Villard, J. (1996). Species-specific identification of *Candida krusei* by hybridization with the CkF1, 2 DNA probe. *Journal of Clinical Microbiology*, 34(7), 1726–1731.
- Carnero, A. (2002). Targeting the cell cycle for cancer therapy. *British Journal of Cancer*, 87(2), 129.
- Castro, L., Blázquez, M. L., González, F. G., & Ballester, A. (2014). Mechanism and applications of metal nanoparticles prepared by bio-mediated process. *Reviews in Advanced Sciences and Engineering*, 3(3), 199–216.
- Castro-Longoria, E., Moreno-Velázquez, S. D., Vilchis-Nestor, A. R., Arenas-Berumen, E., & Avalos-Borja, M. (2012). Production of platinum nanoparticles and nanoaggregates using *Neurospora crassa*. *Journal of Microbiology and Biotechnology*, 22(7), 1000–4.
- Castro-Longoria, E., Vilchis-Nestor, A. R., & Avalos-Borja, M. (2011). Biosynthesis of silver, gold and bimetallic nanoparticles using the filamentous fungus

*Neurospora crassa. Colloids and Surfaces B: Biointerfaces*, 83(1), 42–48.

- Chaudhary, P., Sharma, R., Sharma, A., Vatsyayan, R., Yadav, S., Singhal, S. S., ... Awasthi, Y. C. (2010). Mechanisms of 4-hydroxy-2-nonenal induced pro-and anti-apoptotic signaling. *Biochemistry*, 49(29), 6263–6275.
- Chauhan, R., Reddy, A., & Abraham, J. (2015). Biosynthesis of silver and zinc oxide nanoparticles using *Pichia fermentans* JA2 and their antimicrobial property. *Applied Nanoscience*, 5(1), 63–71.
- Chaudhuri, S. K., & Malodia, L. (2017). Biosynthesis of zinc oxide nanoparticles using leaf extract of *Calotropis gigantea*: characterization and its evaluation on tree seedling growth in nursery stage. *Applied Nanoscience*, 7(8), 501–512.
- Chandrasekaran, R., Gnanasekar, S., Seetharaman, P., Keppanan, R., Arockiaswamy, W., & Sivaperumal, S. (2016). Formulation of *Carica papaya* latex-functionalized zinc oxide nanoparticles for its improved antibacterial and anticancer applications. *Journal of Molecular Liquids*, 219, 232–238.
- Chen, S. J., Liu, Y. C., Shao, C. L., Mu, R., Lu, Y. M., Zhang, J. Y., Fan, X. W. (2005). Structural and optical properties of uniform ZnO nanosheets. *Advanced Materials*, 17(5), 586–590.
- Chen, J., Liu, X., Wang, C., Yin, S.-S., Li, X.-L., Hu, W.-J., Chu, C.-C. (2015). Nitric oxide ameliorates zinc oxide nanoparticles-induced phytotoxicity in rice seedlings. *Journal of Hazardous Materials*, 297, 173–182.
- Chen, Y., & Pepin, A. (2001). Nanofabrication: Conventional and nonconventional methods. *Electrophoresis*, 22(2), 187–207.
- Cheng, B., & Samulski, E. T. (2004). Hydrothermal synthesis of one-dimensional ZnO nanostructures with different aspect ratios. *Chemical Communications*, (8), 986–987.
- Chin, Y. R., & Toker, A. (2010). The actin-bundling protein palladin is an Akt1-specific substrate that regulates breast cancer cell migration. *Molecular Cell*, 38(3), 333–344.
- Cho, Y., Gorina, S., Jeffrey, P. D., & Pavletich, N. P. (1994). Crystal structure of a p53 tumor suppressor-DNA complex: understanding tumorigenic mutations. *Science-AAAS-Weekly Paper Edition*, 265(5170), 346–355.
- Choi, I. (2013). Reactive oxygen species and cancer. *Hanyang Medical Reviews*, 33(2), 118–122.
- Constantinou, C., Papas, K. A., & Constantinou, A. I. (2009). Caspase-independent pathways of programmed cell death: the unraveling of new targets of cancer therapy? *Current Cancer Drug Targets*, 9(6), 717–728.

- Contrerasguzman, E. S., & Strong, F. C. (1982). Determination of tocopherols (vitamin-E) by reduction of cupric ion. *Journal of the Association of Official Analytical Chemists*.
- Cregg, J. M., Vedvick, T. S., & Raschke, W. C. (1993). Recent advances in the expression of foreign genes in *Pichia pastoris*. *Nature Biotechnology*, 11(8), 905–910.
- Cuzick, J. (2002). First results from the International Breast Cancer Intervention Study (IBIS-I): a randomised prevention trial. *The Lancet*, 360(9336), 817.
- Dameron, C. T., Reese, R. N., Mehra, R. K., Kortan, A. R., Carroll, P. J., Steigerwald, M. L., Winge, D. R. (1989). Biosynthesis of cadmium sulphide quantum semiconductor crystallites. *Nature*, 338(6216), 596–597.
- Damaso, M. C. T., Almeida, M. S., Kurtenbach, E., Martins, O. B., Pereira, N., Andrade, C. M. M. C., & Albano, R. M. (2003). Optimized expression of a thermostable xylanase from *Thermomyces lanuginosus* in *Pichia pastoris*. *Applied and Environmental Microbiology*, 69(10), 6064–6072.
- Das, D., Nath, B. C., Phukon, P., Kalita, A., & Dolui, S. K. (2013). Synthesis of ZnO nanoparticles and evaluation of antioxidant and cytotoxic activity. *Colloids and Surfaces B: Biointerfaces*, 111, 556–560.
- Das, S. K., Das, A. R., & Guha, A. K. (2009). Gold nanoparticles: microbial synthesis and application in water hygiene management. *Langmuir*, 25(14), 8192–8199.
- Das, S. K., Das, A. R., & Guha, A. K. (2010). Microbial synthesis of multishaped gold nanostructures. *Small*, 6(9), 1012–1021.
- De Dios, M., Barroso, F., Tojo, C., Blanco, M. C., & Lopez-Quintela, M. A. (2005). Effects of the reaction rate on the size control of nanoparticles synthesized in microemulsions. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 270, 83–87.
- Deng, X., Luan, Q., Chen, W., Wang, Y., Wu, M., Zhang, H., & Jiao, Z. (2009). Nanosized zinc oxide particles induce neural stem cell apoptosis. *Nanotechnology*, 20(11), 115101.
- Dhanasekaran, D. N., & Reddy, E. P. (2008). JNK signaling in apoptosis. *Oncogene*, 27(48), 6245.
- Dhandapani, P., Siddarth, A. S., Kamalasekaran, S., Maruthamuthu, S., & Rajagopal, G. (2014). Bio-approach: ureolytic bacteria mediated synthesis of ZnO nanocrystals on cotton fabric and evaluation of their antibacterial properties. *Carbohydrate Polymers*, 103, 448–455.

- Dhawan, D. K., & Chadha, V. D. (2010). Zinc: a promising agent in dietary chemoprevention of cancer. *Indian Journal of Medical Research*, 132(6), 676.
- Dineley, K. E., Votyakova, T. V., & Reynolds, I. J. (2003). Zinc inhibition of cellular energy production: implications for mitochondria and neurodegeneration. *Journal of Neurochemistry*, 85(3), 563–570.
- Dive, C., Gregory, C. D., Phipps, D. J., Evans, D. L., Milner, A. E., & Wyllie, A. H. (1992). Analysis and discrimination of necrosis and apoptosis (programmed cell death) by multiparameter flow cytometry. *Biochimica et Biophysica Acta (BBA)-Molecular Cell Research*, 1133(3), 275–285.
- Du, L., Suo, S., Wang, G., Jia, H., Liu, K. J., Zhao, B., & Liu, Y. (2013). Mechanism and Cellular Kinetic Studies of the Enhancement of Antioxidant Activity by Using Surface-Functionalized Gold Nanoparticles. *Chemistry—A European Journal*, 19(4), 1281–1287.
- Durán, N., Marcato, P. D., Alves, O. L., De Souza, G. I. H., & Esposito, E. (2005). Mechanistic aspects of biosynthesis of silver nanoparticles by several *Fusarium oxysporum* strains. *J Nanobiotechnol*, 3(8), 1–7.
- Durán, N., Marcato, P. D., Durán, M., Yadav, A., Gade, A., & Rai, M. (2011). Mechanistic aspects in the biogenic synthesis of extracellular metal nanoparticles by peptides, bacteria, fungi, and plants. *Applied Microbiology and Biotechnology*, 90(5), 1609–1624.
- Dzhambazov, B., Daskalova, S., Monteva, A., & Popov, N. (2002). In vitro screening for antitumour activity of *Clinopodium vulgare* L.(Lamiaceae) extracts. *Biological and Pharmaceutical Bulletin*, 25(4), 499-504.
- Elahi, A., & Rehman, A. (2018). Bioconversion of hemicellulosic materials into ethanol by yeast, *Pichia kudriavzevii* 2-KLP1, isolated from industrial waste. *Revista Argentina de Microbiología*, 1-9.
- Elmore, S. (2007). Apoptosis: a review of programmed cell death. *Toxicologic Pathology*, 35(4), 495–516.
- Elumalai, K., Velmurugan, S., Ravi, S., Kathiravan, V., & Ashokkumar, S. (2015). RETRACTED: Green synthesis of zinc oxide nanoparticles using *Moringa oleifera* leaf extract and evaluation of its antimicrobial activity. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 143, 158-164.
- Emami-Karvani, Z., & Chehrazi, P. (2011). Antibacterial activity of ZnO nanoparticle on gram-positive and gram-negative bacteria. *African Journal of Microbiology Research*, 5(12), 1368-1373.
- Elkady, A. I., Abuzinadah, O. A., Baeshen, N. A., & Rahmy, T. R. (2012). Differential control of growth, apoptotic activity, and gene expression in human breast cancer cells by extracts derived from medicinal herbs *Zingiber officinale*.

- Espitia, P. J. P., Soares, N. de F. F., dos Reis Coimbra, J. S., de Andrade, N. J., Cruz, R. S., & Medeiros, E. A. A. (2012). Zinc oxide nanoparticles: synthesis, antimicrobial activity and food packaging applications. *Food and Bioprocess Technology*, 5(5), 1447–1464.
- Fabian, C. J., & Kimler, B. F. (2005). Selective estrogen-receptor modulators for primary prevention of breast cancer. *Journal of Clinical Oncology*, 23(8), 1644–1655.
- Fadok, V. A., Voelker, D. R., Campbell, P. A., Cohen, J. J., Bratton, D. L., & Henson, P. M. (1992). Exposure of phosphatidylserine on the surface of apoptotic lymphocytes triggers specific recognition and removal by macrophages. *The Journal of Immunology*, 148(7), 2207–2216.
- Feris, K., Otto, C., Tinker, J., Wingett, D., Punnoose, A., Thurber, A., ... Hanna, C. (2009). Electrostatic interactions affect nanoparticle-mediated toxicity to gram-negative bacterium *Pseudomonas aeruginosa* PAO1. *Langmuir*, 26(6), 4429–4436.
- Fink, S. L., & Cookson, B. T. (2005). Apoptosis, pyroptosis, and necrosis: mechanistic description of dead and dying eukaryotic cells. *Infection and Immunity*, 73(4), 1907–1916.
- Fisher, B., Costantino, J. P., Wickerham, D. L., Redmond, C. K., Kavanah, M., Cronin, W. M., ... Atkins, J. (1998). Tamoxifen for prevention of breast cancer: report of the National Surgical Adjuvant Breast and Bowel Project P-1 Study. *JNCI: Journal of the National Cancer Institute*, 90(18), 1371–1388.
- Franklin, N. M., Rogers, N. J., Apte, S. C., Batley, G. E., Gadd, G. E., & Casey, P. S. (2007). Comparative toxicity of nanoparticulate ZnO, bulk ZnO, and ZnCl<sub>2</sub> to a freshwater microalga (*Pseudokirchneriella subcapitata*): the importance of particle solubility. *Environmental Science & Technology*, 41(24), 8484–8490.
- Franklin, R. B., & Costello, L. C. (2009). The important role of the apoptotic effects of zinc in the development of cancers. *Journal of Cellular Biochemistry*, 106(5), 750–757.
- Fridman, J. S., & Lowe, S. W. (2003). Control of apoptosis by p53. *Oncogene*, 22(56), 9030.
- Fulda, S. (2009). Tumor resistance to apoptosis. *International Journal of Cancer*, 124(3), 511–515.
- Fu, L., & Fu, Z. (2015). *Plectranthus amboinicus* leaf extract-assisted biosynthesis of ZnO nanoparticles and their photocatalytic activity. *Ceramics International*, 41(2), 2492–2496.

- Gadaga, T. H., Mutukumira, A. N., & Narvhus, J. A. (2000). Enumeration and identification of yeasts isolated from Zimbabwean traditional fermented milk. *International Dairy Journal*, 10(7), 459–466.
- Gao, F., Ma, N., Zhou, H., Wang, Q., Zhang, H., Wang, P., ... Li, L. (2016). Zinc oxide nanoparticles-induced epigenetic change and G2/M arrest are associated with apoptosis in human epidermal keratinocytes. *International Journal of Nanomedicine*, 11, 3859.
- Gartel, A. L., & Tyner, A. L. (2002). The role of the cyclin-dependent kinase inhibitor p21 in apoptosis supported in part by NIH grant R01 DK56283 (to ALT) for the p21 research and Campus Research Board and Illinois Department of Public Health Penny Severns Breast and Cervical Cancer grant. *Molecular Cancer Therapeutics*, 1(8), 639–649.
- George, S., Pokhrel, S., Xia, T., Gilbert, B., Ji, Z., Schowalter, M., Mädler, L. (2009). Use of a rapid cytotoxicity screening approach to engineer a safer zinc oxide nanoparticle through iron doping. *ACS Nano*, 4(1), 15–29.
- Gericke, M., & Pinches, A. (2006). Biological synthesis of metal nanoparticles. *Hydrometallurgy*, 83(1), 132–140.
- Germain, M., Affar, E. B., D'Amours, D., Dixit, V. M., Salvesen, G. S., & Poirier, G. G. (1999). Cleavage of automodified poly (ADP-ribose) polymerase during apoptosis evidence for involvement of caspase-7. *Journal of Biological Chemistry*, 274(40), 28379–28384.
- Gharagozlu, M., Baradaran, Z., & Bayati, R. (2015). A green chemical method for synthesis of ZnO nanoparticles from solid-state decomposition of Schiff-bases derived from amino acid alanine complexes. *Ceramics International*, 41(7), 8382–8387.
- Ghodake, G., & Lee, D. S. (2011). Biological synthesis of gold nanoparticles using the aqueous extract of the brown algae *Laminaria japonica*. *Journal of Nanoelectronics and Optoelectronics*, 6(3), 268–271.
- Gillet, C. E., & Barnes, D. M. (2000). The Molecular Basis of Cell Cycle and Growth Control.: Stein GS, Baserga R, Giordano A, et al, eds.(£ 45.50.) Wiley, 1998. ISBN 0 471 15706 6. BMJ Group.
- Gopalakrishnan, K., Ramesh, C., Ragunathan, V., & Thamilselvan, M. (2012). Antibacterial activity of Cu<sub>2</sub>O nanoparticles on E. coli synthesized from Tridax procumbens leaf extract and surface coating with polyaniline. *Dig J Nanomater Bios*, 7(2), 833–839.
- Govender, Y., Riddin, T., Gericke, M., & Whiteley, C. G. (2009). Bioreduction of platinum salts into nanoparticles: a mechanistic perspective. *Biotechnology Letters*, 31(1), 95–100.

- Greppi, A., Saubade, F., Botta, C., Humblot, C., Guyot, J.-P., & Cocolin, L. (2017). Potential probiotic *Pichia kudriavzevii* strains and their ability to enhance folate content of traditional cereal-based African fermented food. *Food Microbiology*, 62, 169–177.
- Group, E. B. C. T. C. (1998). Tamoxifen for early breast cancer: an overview of the randomised trials. *The Lancet*, 351(9114), 1451–1467.
- Guerriero, J. L., Ditsworth, D., Fan, Y., Zhao, F., Crawford, H. C., & Zong, W.-X. (2008). Chemotherapy induces tumor clearance independent of apoptosis. *Cancer Research*, 68(23), 9595–9600.
- Hajipour, M. J., Fromm, K. M., Ashkarran, A. A., de Aberasturi, D. J., de Larramendi, I. R., Rojo, T., Mahmoudi, M. (2012). Antibacterial properties of nanoparticles. *Trends in Biotechnology*, 30(10), 499–511.
- Hameed, A. S. H., Karthikeyan, C., Ahamed, A. P., Thajuddin, N., Alharbi, N. S., Alharbi, S. A., & Ravi, G. (2016). In vitro antibacterial activity of ZnO and Nd doped ZnO nanoparticles against ESBL producing *Escherichia coli* and *Klebsiella pneumoniae*. *Scientific Reports*, 6.
- Hanley, C., Layne, J., Punnoose, A., Reddy, K. M., Coombs, I., Coombs, A., ... Wingett, D. (2008). Preferential killing of cancer cells and activated human T cells using ZnO nanoparticles. *Nanotechnology*, 19(29), 295103.
- Hanley, C., Thurber, A., Hanna, C., Punnoose, A., Zhang, J., & Wingett, D. G. (2009). The influences of cell type and ZnO nanoparticle size on immune cell cytotoxicity and cytokine induction. *Nanoscale Research Letters*, 4(12), 1409.
- Hassani Sangani, M., Nakhaei Moghaddam, M., & Forghanifard, M. M. (2015). Inhibitory effect of zinc oxide nanoparticles on *pseudomonas aeruginosa* biofilm formation. *Nanomedicine Journal*, 2(2), 121–128.
- Hashem, M., Hesham, A. E.-L., Alrumanan, S. A., Alamri, S. A., & Moustafa, M. F. M. (2014). Indigenous yeasts of the rotten Date fruits and their potentiality in bioethanol and single-cell protein production. *International Journal of Agriculture & Biology*, 16(4), 752–758.
- He, J. H., Lao, C. S., Chen, L. J., Davidovic, D., & Wang, Z. L. (2005). Large-scale Ni-doped ZnO nanowire arrays and electrical and optical properties. *Journal of the American Chemical Society*, 127(47), 16376–16377.
- Heinlaan, M., Ivask, A., Blinova, I., Dubourguier, H.-C., & Kahru, A. (2008). Toxicity of nanosized and bulk ZnO, CuO and TiO<sub>2</sub> to bacteria *Vibrio fischeri* and crustaceans *Daphnia magna* and *Thamnocephalus platyurus*. *Chemosphere*, 71(7), 1308–1316.
- Hellström, A. M., Almgren, A., Carlsson, N.-G., Svanberg, U., & Andlid, T. A. (2012). Degradation of phytate by *Pichia kudriavzevii* TY13 and

*Hanseniaspora guilliermondii* TY14 in Tanzanian togwa. *International Journal of Food Microbiology*, 153(1), 73–77.

Ho, E. (2004). Zinc deficiency, DNA damage and cancer risk. *The Journal of Nutritional Biochemistry*, 15(10), 572–578.

Hu, W., Chen, S., Zhou, B., & Wang, H. (2010). Facile synthesis of ZnO nanoparticles based on bacterial cellulose. *Materials Science and Engineering: B*, 170(1), 88–92.

Huang, X., Neretina, S., & El-Sayed, M. A. (2009). Gold nanorods: from synthesis and properties to biological and biomedical applications. *Advanced Materials*, 21(48), 4880–4910.

Hulley, S. B., Cummings, S. R., Browner, W. S., Grady, D. G., & Newman, T. B. (2013). Designing clinical research. *Lippincott Williams & Wilkins*.

Iravani, S. (2011). Green synthesis of metal nanoparticles using plants. *Green Chemistry*, 13(10), 2638–2650.

Iwamoto, M., Kuroda, K., Kanzow, J., Hayashi, S., & Faupel, F. (2005). Size evolution effect of the reduction rate on the synthesis of gold nanoparticles. *Advanced Powder Technology*, 16(2), 137–144.

Jaidev, L. R., & Narasimha, G. (2010). Fungal mediated biosynthesis of silver nanoparticles, characterization and antimicrobial activity. *Colloids and Surfaces B: Biointerfaces*, 81(2), 430–433.

Jain, K. K. (2007). Applications of nanobiotechnology in clinical diagnostics. *Clinical Chemistry*, 53(11), 2002–2009.

Jamdagni, P., Khatri, P., & Rana, J. S. (2016). Green synthesis of zinc oxide nanoparticles using flower extract of Nyctanthes arbor-tristis and their antifungal activity. *Journal of King Saud University-Science*.

Jansen, J., Karges, W., & Rink, L. (2009). Zinc and diabetes—clinical links and molecular mechanisms. *The Journal of Nutritional Biochemistry*, 20(6), 399–417.

Jayaseelan, C., Rahuman, A. A., Kirthi, A. V., Marimuthu, S., Santhoshkumar, T., Bagavan, A., Rao, K. V. B. (2012). Novel microbial route to synthesize ZnO nanoparticles using Aeromonas hydrophila and their activity against pathogenic bacteria and fungi. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 90, 78–84.

Jha, A. K., Prasad, K., & Prasad, K. (2009). A green low-cost biosynthesis of Sb<sub>2</sub>O<sub>3</sub> nanoparticles. *Biochemical Engineering Journal*, 43(3), 303–306.

- Jin, T., Sun, D., Su, J. Y., Zhang, H., & Sue, H. (2009). Antimicrobial efficacy of zinc oxide quantum dots against *Listeria monocytogenes*, *Salmonella Enteritidis*, and *Escherichia coli* O157: H7. *Journal of Food Science*, 74(1), M46–M52.
- Jones, N., Ray, B., Ranjit, T.K. and Manna, C.A. Antibacterial activity of ZnO nanoparticle suspensions on a broad spectrum of microorganisms. *FEMS Microbiology Letters* 2008; 279(1): 71-76.
- Ju-Nam, Y., & Lead, J. R. (2008). Manufactured nanoparticles: an overview of their chemistry, interactions and potential environmental implications. *Science of the Total Environment*, 400(1), 396–414.
- Kao, Y.-Y., Chen, Y.-C., Cheng, T.-J., Chiung, Y.-M., & Liu, P.-S. (2011). Zinc oxide nanoparticles interfere with zinc ion homeostasis to cause cytotoxicity. *Toxicological Sciences*, 125(2), 462–472.
- Karp, G. (2007). *Cell and molecular biology*. John Wiley & Sons Incorporated.
- Kasibhatla, S., & Tseng, B. (2003). Why target apoptosis in cancer treatment? *Molecular Cancer Therapeutics*, 2(6), 573–580.
- Kathiresan, K., Alikunhi, N. M., Pathmanaban, S., Nabikhan, A., & Kandasamy, S. (2010). Analysis of antimicrobial silver nanoparticles synthesized by coastal strains of *Escherichia coli* and *Aspergillus niger*. *Canadian Journal of Microbiology*, 56(12), 1050–1059.
- Kavithaa, K., Paulpandi, M., Ponraj, T., Murugan, K., & Sumathi, S. (2016). Induction of intrinsic apoptotic pathway in human breast cancer (MCF-7) cells through facile biosynthesized zinc oxide nanorods. *Karbala International Journal of Modern Science*, 2(1), 46–55.
- Kerr, J. F. R., Winterford, C. M., & Harmon, B. V. (1994). Apoptosis. Its significance in cancer and cancer therapy. *Cancer*, 73(8), 2013–2026.
- Kim, Y.-C., Park, N.-C., Shin, J.-S., Lee, S. R., Lee, Y. J., & Moon, D. J. (2003). Partial oxidation of ethylene to ethylene oxide over nanosized Ag/ $\alpha$ -Al<sub>2</sub>O<sub>3</sub> catalysts. *Catalysis Today*, 87(1), 153–162.
- King, R. J. B., & Robins, M. W. (2006). *Cancer biology*. Pearson Education.
- Klaus, T., Joerger, R., Olsson, E., & Granqvist, C.-G. (1999). Silver-based crystalline nanoparticles, microbially fabricated. *Proceedings of the National Academy of Sciences*, 96(24), 13611–13614.
- Knight, J. A. (1995). Diseases related to oxygen-derived free radicals. *Annals of Clinical & Laboratory Science*, 25(2), 111–121.

- Kooti, M., & Naghdi Sedeh, A. (2012). Microwave-assisted combustion synthesis of ZnO nanoparticles. *Journal of Chemistry*, 28(1), 167-172.
- Kowshik, M., Deshmukh, N., Vogel, W., Urban, J., Kulkarni, S. K., & Paknikar, K. M. (2002). Microbial synthesis of semiconductor CdS nanoparticles, their characterization, and their use in the fabrication of an ideal diode. *Biotechnology and Bioengineering*, 78(5), 583-588.
- Kowshik, M., Vogel, W., Urban, J., Kulkarni, S. K., & Paknikar, K. M. (2002). Microbial synthesis of semiconductor PbS nanocrystallites. *Advanced Materials*, 14(11), 815.
- Krumov, N., Oder, S., Perner-Nochta, I., Angelov, A., & Posten, C. (2007). Accumulation of CdS nanoparticles by yeasts in a fed-batch bioprocess. *Journal of Biotechnology*, 132(4), 481-486.
- Krupa, A. N. D., & Vimala, R. (2016). Evaluation of tetraethoxysilane (TEOS) sol-gel coatings, modified with green synthesized zinc oxide nanoparticles for combating microfouling. *Materials Science and Engineering: C*, 61, 728-735.
- Kubota, Y., Shuin, T., Kawasaki, C., Hosaka, M., Kitamura, H., Cai, R., Fujishima, A. (1994). Photokilling of T-24 human bladder cancer cells with titanium dioxide. *British Journal of Cancer*, 70(6), 1107.
- Kulandaivelu, B., & Gothandam, K. M. (2016). Cytotoxic effect on cancerous cell lines by biologically synthesized silver nanoparticles. *Brazilian Archives of Biology and Technology*, 59, 1-7.
- Kumar, A., Najafzadeh, M., Jacob, B. K., Dhawan, A., & Anderson, D. (2014). Zinc oxide nanoparticles affect the expression of p53, Ras p21 and JNKs: an ex vivo/in vitro exposure study in respiratory disease patients. *Mutagenesis*, 30(2), 237-245.
- Kumar, B., Smita, K., Seqqat, R., Benalcazar, K., Grijalva, M., & Cumbal, L. (2016). In vitro evaluation of silver nanoparticles cytotoxicity on Hepatic cancer (Hep-G2) cell line and their antioxidant activity: Green approach for fabrication and application. *Journal of Photochemistry and Photobiology B: Biology*, 159, 8-13.
- Kumar, D., Karthik, L., Kumar, G., & Roa, K. B. (2011). Biosynthesis of Silver nanoparticles from Marine Yeast and Their Antimicrobial Activity Against Multidrug Resistant Pathogens. *Pharmacologyonline*, 3, 1100-1111.
- Kumar, K. D., Narayana, K., & Rao, K. A. (2018). Green Synthesis of Zinc Oxide Nanoparticles using Extracts of *Ocimum Tenuiflorum* and its Characterization. *Journal of Nanoscience Nanoengineering and Applications*, 8(1), 9-19.
- Kumar, S. A., Ansary, A. A., Ahmad, A., & Khan, M. I. (2007). Extracellular biosynthesis of CdSe quantum dots by the fungus, *Fusarium oxysporum*.

*Journal of Biomedical Nanotechnology*, 3(2), 190–194.

Kumar, V., Abbas, A. K., & Aster, J. C. (2013). *Robbins basic pathology*. Elsevier Health Sciences.

Kumar, D., Karthik, L., Kumar, G., & V, B. R. K. (2011). Biosynthesis of Silver nanoparticles from Marine Yeast and Their Antimicrobial Activity Against Multidrug Resistant Pathogens. *Pharmacologyonline*, 3, 1100–1111.

Kumari, B., Sharma, S., Singh, N., Verma, A., Satsangi, V. R., Dass, S., & Shrivastav, R. (2014). ZnO thin films, surface embedded with biologically derived Ag/Au nanoparticles, for efficient photoelectrochemical splitting of water. *International Journal of Hydrogen Energy*, 39(32), 18216–18229.

Kundu, D., Hazra, C., Chatterjee, A., Chaudhari, A., & Mishra, S. (2014). Extracellular biosynthesis of zinc oxide nanoparticles using *Rhodococcus pyridinivorans* NT2: multifunctional textile finishing, biosafety evaluation and in vitro drug delivery in colon carcinoma. *Journal of Photochemistry and Photobiology B: Biology*, 140, 194–204.

Kunze, G., Kang, H. A., & Gellissen, G. (2009). *Hansenula polymorpha (Pichia angusta)*: biology and applications. In *Yeast Biotechnology: Diversity and Applications* (pp. 47–64). Springer.

Kupchan, S. M. (1976). Novel plant-derived tumor inhibitors and their mechanisms of action. *Cancer Treatment Reports*, 60(8), 1115–1126.

Kuppusamy, P., Yusoff, M. M., Maniam, G. P., & Govindan, N. (2016). Biosynthesis of metallic nanoparticles using plant derivatives and their new avenues in pharmacological applications—An updated report. *Saudi Pharmaceutical Journal*, 24(4), 473–484.

Kurtzman, C. P., & Fell, J. W. (2006). Yeast systematics and phylogeny—implications of molecular identification methods for studies in ecology. In *Biodiversity and Ecophysiology of Yeasts* (pp. 11–30). Springer.

Kurtzman, C. P., Smiley, M. J., & Johnson, C. J. (1980). Emendation of the genus *Issatchenkia* Kudriavzev and comparison of species by deoxyribonucleic acid reassociation, mating reaction, and ascospore ultrastructure. *International Journal of Systematic and Evolutionary Microbiology*, 30(2), 503–513.

Kwolek-Mirek, M., & Zadrag-Tecza, R. (2014). Comparison of methods used for assessing the viability and vitality of yeast cells. *FEMS Yeast Research*, 14(7), 1068–1079.

Lany, S., Osorio-Guillén, J., & Zunger, A. (2007). Origins of the doping asymmetry in oxides: Hole doping in NiO versus electron doping in ZnO. *Physical Review B*, 75(24), 241203.

- Lewinski, N., Colvin, V., & Drezek, R. (2008). Cytotoxicity of nanoparticles. *Small*, 4(1), 26–49.
- Lindahl, M., Leanderson, P., & Tagesson, C. (1998). Novel aspect on metal fume fever: zinc stimulates oxygen radical formation in human neutrophils. *Human & Experimental Toxicology*, 17(2), 105–110.
- Lin, B., Fu, Z., & Jia, Y. (2001). Green luminescent center in undoped zinc oxide films deposited on silicon substrates. *Applied Physics Letters*, 79(7), 943–945.
- Lincz, L. F. (1998). Deciphering the apoptotic pathway: all roads lead to death. *Immunology and Cell Biology*, 76(1), 1–19.
- Liu, J., Huang, X., Li, Y., Duan, J., & Ai, H. (2006). Large-scale synthesis of flower-like ZnO structures by a surfactant-free and low-temperature process. *Materials Chemistry and Physics*, 98(2), 523–527.
- Liu, L., Liu, T., Tade, M., Wang, S., Li, X., & Liu, S. (2014). Less is more, greener microbial synthesis of silver nanoparticles. *Enzyme and Microbial Technology*, 67(1), 53–58.
- Liu, Y., He, L., Mustapha, A., Li, H., Hu, Z. Q., & Lin, M. (2009). Antibacterial activities of zinc oxide nanoparticles against *Escherichia coli* O157: H7. *Journal of Applied Microbiology*, 107(4), 1193–1201.
- Lu, A., Salabas, E. emsp14L, & Schüth, F. (2007). Magnetic nanoparticles: synthesis, protection, functionalization, and application. *Angewandte Chemie International Edition*, 46(8), 1222–1244.
- Luk, S. C.-W., Siu, S. W.-F., Lai, C.-K., Wu, Y.-J., & Pang, S.-F. (2005). Cell cycle arrest by a natural product via G2/M checkpoint. *International Journal of Medical Sciences*, 2(2), 64.
- Ma, H., Williams, P. L., & Diamond, S. A. (2013). Ecotoxicity of manufactured ZnO nanoparticles—a review. *Environmental Pollution*, 172, 76–85.
- Ma, X.-Y., & Zhang, W.-D. (2009). Effects of flower-like ZnO nanowiskers on the mechanical, thermal and antibacterial properties of waterborne polyurethane. *Polymer Degradation and Stability*, 94(7), 1103–1109.
- Madan, H. R., Sharma, S. C., Suresh, D., Vidya, Y. S., Nagabhushana, H., Rajanaik, H., Maiya, P. S. (2016). Facile green fabrication of nanostructure ZnO plates, bullets, flower, prismatic tip, closed pine cone: their antibacterial, antioxidant, photoluminescent and photocatalytic properties. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 152, 404–416.
- Mahanta, S., Prathap, S., Ban, D. K., & Paul, S. (2017). Protein functionalization of ZnO nanostructure exhibits selective and enhanced toxicity to breast cancer cells through oxidative stress-based cell death mechanism. *Journal of*

- Mahdavi, M., Namvar, F., Ahmad, M. Bin, & Mohamad, R. (2013). Green biosynthesis and characterization of magnetic iron oxide ( $\text{Fe}_3\text{O}_4$ ) nanoparticles using seaweed (*Sargassum muticum*) aqueous extract. *Molecules*, 18(5), 5954–5964.
- Mal, S., Nori, S., Jin, C., Narayan, J., Nellutla, S., Smirnov, A. I., & Prater, J. T. (2010). Reversible room temperature ferromagnetism in undoped zinc oxide: Correlation between defects and physical properties. *Journal of Applied Physics*, 108(7), 73510.
- Mandal, D., Bolander, M. E., Mukhopadhyay, D., Sarkar, G., & Mukherjee, P. (2006). The use of microorganisms for the formation of metal nanoparticles and their application. *Applied Microbiology and Biotechnology*, 69(5), 485–492.
- Manning, B. D., & Cantley, L. C. (2007). AKT/PKB signaling: navigating downstream. *Cell*, 129(7), 1261–1274.
- Marsalek, R. (2014). Particle size and zeta potential of  $\text{ZnO}$ . *APCBEE Procedia*, 9, 13–17.
- Mattson, M. P., & Bazan, N. G. (2006). Apoptosis and necrosis. *Basic Neurochemistry*, 7th Edn. Elsevier, Boston, 603–615.
- Meroth, C. B., Hammes, W. P., & Hertel, C. (2003). Identification and population dynamics of yeasts in sourdough fermentation processes by PCR-denaturing gradient gel electrophoresis. *Applied and Environmental Microbiology*, 69(12), 7453–7461.
- Mirzaei, H., & Darroudi, M. (2017). Zinc oxide nanoparticles: biological synthesis and biomedical applications. *Ceramics International*, 43(1), 907–914.
- Mitra, S., Patra, P., Pradhan, S., Debnath, N., Dey, K. K., Sarkar, S., Goswami, A. (2015). Microwave synthesis of  $\text{ZnO}@\text{mSiO}_2$  for detailed antifungal mode of action study: Understanding the insights into oxidative stress. *Journal of Colloid and Interface Science*, 444, 97–108.
- Mohammadi, G., Valizadeh, H., Barzegar-Jalali, M., Lotfipour, F., Adibkia, K., Milani, M., Nokhodchi, A. (2010). Development of azithromycin–PLGA nanoparticles: Physicochemical characterization and antibacterial effect against *Salmonella typhi*. *Colloids and Surfaces B: Biointerfaces*, 80(1), 34–39.
- Mohanpuria, P., Rana, N. K., & Yadav, S. K. (2008). Biosynthesis of nanoparticles: technological concepts and future applications. *Journal of Nanoparticle Research*, 10(3), 507–517.
- Mohan, S., Abdelwahab, S. I., Kamalidehghan, B., Syam, S., May, K. S., Harmal, N. S. M., Rahmani, M. (2012). Involvement of NF- $\kappa$ B and Bcl2/Bax signaling

- pathways in the apoptosis of MCF7 cells induced by a xanthone compound Pyranocycloartobiloxanthone A. *Phytomedicine*, 19(11), 1007–1015.
- Moslehi-Jenabian, S., Lindegaard, L., & Jespersen, L. (2010). Beneficial effects of probiotic and food borne yeasts on human health. *Nutrients*, 2(4), 449–473.
- Mosmann, T. (1983). Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *Journal of Immunological Methods*, 65(1–2), 55–63.
- MubarakAli, D., Gopinath, V., Rameshbabu, N., & Thajuddin, N. (2012). Synthesis and characterization of CdS nanoparticles using C-phycoerythrin from the marine cyanobacteria. *Materials Letters*, 74, 8–11.
- Mugula, J. K., Narvhus, J. A., & Sørhaug, T. (2003). Use of starter cultures of lactic acid bacteria and yeasts in the preparation of togwa, a Tanzanian fermented food. *International Journal of Food Microbiology*, 83(3), 307–318.
- Mukherjee, P., Ahmad, A., Mandal, D., Senapati, S., Sainkar, S. R., Khan, M. I., Alam, M. (2001). Bioreduction of AuCl<sub>4</sub><sup>-</sup> ions by the fungus, *Verticillium* sp. and surface trapping of the gold nanoparticles formed. *Angewandte Chemie International Edition*, 40(19), 3585–3588.
- Mukherjee, P., Ahmad, A., Mandal, D., Senapati, S., Sainkar, S. R., Khan, M. I., Kumar, R. (2001). Fungus-mediated synthesis of silver nanoparticles and their immobilization in the mycelial matrix: a novel biological approach to nanoparticle synthesis. *Nano Letters*, 1(10), 515–519.
- Mukherjee, P., Senapati, S., Mandal, D., Ahmad, A., Khan, M. I., Kumar, R., & Sastry, M. (2002). Extracellular synthesis of gold nanoparticles by the fungus *Fusarium oxysporum*. *Chembiochem : A European Journal of Chemical Biology*, 3(5), 461–463.
- Musa, I., Qamhieh, N., & Mahmoud, S. T. (2017). Synthesis and length dependent photoluminescence property of zinc oxide nanorods. *Results in Physics*, 7, 3552–3556.
- Musarrat, J., Dwivedi, S., Singh, B. R., Al-Khedhairy, A. A., Azam, A., & Naqvi, A. (2010). Production of antimicrobial silver nanoparticles in water extracts of the fungus *Amylomyces rouxii* strain KSU-09. *Bioresource Technology*, 101(22), 8772–8776.
- Nagajyothi, P. C., Cha, S. J., Yang, I. J., Sreekanth, T. V. M., Kim, K. J., & Shin, H. M. (2015). Antioxidant and anti-inflammatory activities of zinc oxide nanoparticles synthesized using *Polygala tenuifolia* root extract. *Journal of Photochemistry and Photobiology B: Biology*, 146, 10–17.
- Nagajyothi, P. C., & Lee, K. D. (2011). Synthesis of plant-mediated silver nanoparticles using *Dioscorea batatas* rhizome extract and evaluation of their

- antimicrobial activities. *Journal of Nanomaterials*, 2011, 1–7.
- Nagarajan, S., & Kuppusamy, K. A. (2013). Extracellular synthesis of zinc oxide nanoparticle using seaweeds of gulf of Mannar, India. *Journal of Nanobiotechnology*, 11(1), 39.
- Nair, S., Sasidharan, A., Rani, V. V. D., Menon, D., Nair, S., Manzoor, K., & Raina, S. (2009). Role of size scale of ZnO nanoparticles and microparticles on toxicity toward bacteria and osteoblast cancer cells. *Journal of Materials Science: Materials in Medicine*, 20(1), 235–241.
- Nel, A., Xia, T., Meng, H., Wang, X., Lin, S., Ji, Z., & Zhang, H. (2012). Nanomaterial toxicity testing in the 21st century: use of a predictive toxicological approach and high-throughput screening. *Accounts of Chemical Research*, 46(3), 607–621.
- Newman, M. D., Stotland, M., & Ellis, J. I. (2009). The safety of nanosized particles in titanium dioxide-and zinc oxide-based sunscreens. *Journal of the American Academy of Dermatology*, 61(4), 685–692.
- Ng, K. W., Khoo, S. P. K., Heng, B. C., Setyawati, M. I., Tan, E. C., Zhao, X., Loo, J. S. C. (2011). The role of the tumor suppressor p53 pathway in the cellular DNA damage response to zinc oxide nanoparticles. *Biomaterials*, 32(32), 8218–8225.
- Ng, W. K., Yazan, L. S., & Ismail, M. (2011). Thymoquinone from Nigella sativa was more potent than cisplatin in eliminating of SiHa cells via apoptosis with down-regulation of Bcl-2 protein. *Toxicology in Vitro*, 25(7), 1392–1398.
- Ng, L. Y., Mohammad, A. W., Leo, C. P., & Hilal, N. (2013). Polymeric membranes incorporated with metal/metal oxide nanoparticles: a comprehensive review. *Desalination*, 308, 15–33.
- Nogi, K., Hosokawa, M., Naito, M., & Yokoyama, T. (2012). *Nanoparticle technology handbook*. Elsevier.
- Nohl, H., Kozlov, A. V., Gille, L., & Staniek, K. (2003). Cell respiration and formation of reactive oxygen species: facts and artefacts. Portland Press Limited.
- Nohynek, G. J., Dufour, E. K., & Roberts, M. S. (2008). Nanotechnology, Cosmetics and the Skin: Is There a Health Risk? *Skin Pharmacology and Physiology*, 21(3), 136–149.
- Nohynek, G. J., Lademann, J., Ribaud, C., & Roberts, M. S. (2007). Grey goo on the skin? Nanotechnology, cosmetic and sunscreen safety. *Critical Reviews in Toxicology*, 37(3), 251–277.
- Novak, D. G. (2000). Handout on Health: Systemic Lupus Erythematosus.

- Obasi, B. C., Whong, C. M. Z., Ado, S. A., & Abdullahi, I. O. (2014). Isolation and Identification of yeast associated with fermented orange juice. *Int J Eng*, 3, 64–69.
- Ochieng, P. E., Iwuoha, E., Michira, I., Masikini, M., Ondiek, J., Githira, P., & Kamau, G. N. (2015). Green route synthesis and characterization of ZnO nanoparticles using *Spathodea campanulata*. *International Journal of Bio. Chemi. Physics*, 23, 53–61.
- Ogston, A. (1984). On abscesses. *Reviews of Infectious Diseases*, 6(1), 122–128.
- Ongol, M. P., & Asano, K. (2009). Main microorganisms involved in the fermentation of Ugandan ghee. *International Journal of Food Microbiology*, 133(3), 286–291.
- Orazizadeh, M., Khodadadi, A., Bayati, V., Saremy, S., Farasat, M., & Khorsandi, L. (2015). In vitro toxic effects of zinc oxide nanoparticles on rat adipose tissue-derived mesenchymal stem cells. *Cell Journal (Yakhteh)*, 17(3), 412.
- Ostrakhovitch, E. A., & Cherian, M. G. (2005). Inhibition of extracellular signal regulated kinase (ERK) leads to apoptosis inducing factor (AIF) mediated apoptosis in epithelial breast cancer cells: the lack of effect of ERK in p53 mediated copper induced apoptosis. *Journal of Cellular Biochemistry*, 95(6), 1120–1134.
- Ostrovsky, S., Kazimirsky, G., Gedanken, A., & Brodie, C. (2009). Selective cytotoxic effect of ZnO nanoparticles on glioma cells. *Nano Research*, 2(11), 882–890.
- Otari, S. V., Patil, R. M., Nadaf, N. H., Ghosh, S. J., & Pawar, S. H. (2012). Green biosynthesis of silver nanoparticles from an actinobacteria *Rhodococcus* sp. *Materials Letters*, 72, 92–94.
- Otsuki, Y., Li, Z., & Shibata, M.-A. (2003). Apoptotic detection methods—from morphology to gene. *Progress in Histochemistry and Cytochemistry*, 38(3), 275–339.
- Padmavathy, N., & Vijayaraghavan, R. (2008). Enhanced bioactivity of ZnO nanoparticles—an antimicrobial study. *Science and Technology of Advanced Materials*.
- Paknikar, M. K. and S. A. and S. K. and W. V. and J. U. and S. K. K. and K. M. (2003). Extracellular synthesis of silver nanoparticles by a silver-tolerant yeast strain MKY3. *Nanotechnology*, 14(1), 95.
- Patil, B. N., & Taranath, T. C. (2016). *Limonia acidissima* L. leaf mediated synthesis of zinc oxide nanoparticles: A potent tool against *Mycobacterium tuberculosis*. *International Journal of Mycobacteriology*, 5(2), 197–204.

- Perez, C., Pauli, M., & Bazerque, P. (1990). An antibiotic assay by the agar well diffusion method. *Acta Biol Med Exp*, 15(1), 113–115.
- Péter, G., Tornai-Lehoczki, J., Suzuki, M., & Dlauchy, D. (2005). *Metschnikowia viticola* sp. nov., a new yeast species from grape. *Antonie van Leeuwenhoek*, 87(2), 155–160.
- Prescott, L. M., Harley, J. P., & Klein, D. A. (2008). Microbiology 7th edition. McGram-Hill companies. Inc, New York, 32–34.
- Piccolo, M. T., & Crispi, S. (2012). The dual role played by p21 may influence the apoptotic or anti-apoptotic fate in cancer. *Journal of Cancer Research Updates*, 1(2), 189–202.
- Pimprikar, P. S., Joshi, S. S., Kumar, A. R., Zinjarde, S. S., & Kulkarni, S. K. (2009). Influence of biomass and gold salt concentration on nanoparticle synthesis by the tropical marine yeast *Yarrowia lipolytica* NCIM 3589. *Colloids and Surfaces B: Biointerfaces*, 74(1), 309–316.
- Piner, R. D., Zhu, J., Xu, F., Hong, S., & Mirkin, C. A. (1999). “ Dip-pen” nanolithography. *Science*, 283(5402), 661–663.
- Pinna, N., & Niederberger, M. (2008). Oxide Synthesis as Cornerstone of Nanoscience. *European Journal of Inorganic Chemistry*, 2008(6), 825.
- Prashanth, G. K., Prashanth, P. A., Bora, U., Gadewar, M., Nagabhushana, B. M., Ananda, S., Sathyanaanda, H. M. (2015). In vitro antibacterial and cytotoxicity studies of ZnO nanopowders prepared by combustion assisted facile green synthesis. *Karbala International Journal of Modern Science*, 1(2), 67–77.
- Prakash, D., Upadhyay, G., Singh, B. N., & Singh, H. B. (2007). Antioxidant and free radical-scavenging activities of seeds and agri-wastes of some varieties of soybean (*Glycine max*). *Food Chemistry*, 104(2), 783–790.
- Prasad, A. S. (2008). Zinc in human health: effect of zinc on immune cells. *Molecular Medicine*, 14(5–6), 353.
- Prashanth, G. K., Prashanth, P. A., Bora, U., Gadewar, M., Nagabhushana, B. M., Ananda, S., Sathyanaanda, H. M. (2015). In vitro antibacterial and cytotoxicity studies of ZnO nanopowders prepared by combustion assisted facile green synthesis. *Karbala International Journal of Modern Science*, 1(2), 67–77.
- Premanathan, M., Karthikeyan, K., Jeyasubramanian, K., & Manivannan, G. (2011). Selective toxicity of ZnO nanoparticles toward Gram-positive bacteria and cancer cells by apoptosis through lipid peroxidation. *Nanomedicine: Nanotechnology, Biology and Medicine*, 7(2), 184–192.
- Pulit-Prociak, J., Chwastowski, J., Kucharski, A., & Banach, M. (2016). Functionalization of textiles with silver and zinc oxide nanoparticles. *Applied*

*Surface Science*, 385, 543–553.

- Puri, P. L., MacLachlan, T. K., Levrero, M., & Giordano, A. (1999). The intrinsic cell cycle: from yeast to mammals. *The Molecular Basis of Cell Cycle and Growth Control*, 1, 15–79.
- Qian, Y., Yao, J., Russel, M., Chen, K., & Wang, X. (2015). Characterization of green synthesized nano-formulation (ZnO-A. vera) and their antibacterial activity against pathogens. *Environmental Toxicology and Pharmacology*, 39(2), 736–746.
- Qvist, L., Vorontsov, E., Veide Vilg, J., & Andlid, T. (2016). Strain improvement of *Pichia kudriavzevii* TY13 for raised phytase production and reduced phosphate repression. *Microbial Biotechnology*.
- Qu, J., Luo, C., & Hou, J. (2011). Synthesis of ZnO nanoparticles from Zn-hyperaccumulator (*Sedum alfredii* Hance) plants. *Micro & Nano Letters*, 6(3), 174–176.
- Qu, J., Yuan, X., Wang, X., & Shao, P. (2011). Zinc accumulation and synthesis of ZnO nanoparticles using *Physalis alkekengi* L. *Environmental Pollution*, 159(7), 1783–1788.
- Raghupathi, K. R., Koodali, R. T., & Manna, A. C. (2011). Size-dependent bacterial growth inhibition and mechanism of antibacterial activity of zinc oxide nanoparticles. *Langmuir*, 27(7), 4020–4028.
- Rahil-Khazen, R., Bolann, B. J., Myking, A., & Ulvik, R. J. (2002). Multi-element analysis of trace element levels in human autopsy tissues by using inductively coupled atomic emission spectrometry technique (ICP-AES). *Journal of Trace Elements in Medicine and Biology*, 16(1), 15–25.
- Rainey, F. a., Ward-Rainey, N., Kroppenstedt, R. M., & Stackebrandt, E. (1996). The genus *Nocardiopsis* represents a phylogenetically coherent taxon and a distinct actinomycete lineage: proposal of *Nocardiopsaceae* fam. nov. *International Journal of Systematic Bacteriology*, 46(4), 1088–1092.
- Rajakumar, G., Rahuman, A. A., Roopan, S. M., Khanna, V. G., Elango, G., Kamaraj, C., Velayutham, K. (2012). Fungus-mediated biosynthesis and characterization of TiO<sub>2</sub> nanoparticles and their activity against pathogenic bacteria. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 91, 23–29.
- Rajiv, P., Rajeshwari, S., & Venkatesh, R. (2013). Bio-Fabrication of zinc oxide nanoparticles using leaf extract of *Parthenium hysterophorus* L. and its size-dependent antifungal activity against plant fungal pathogens. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 112, 384–387.

- Rajeshkumar, S., Malarkodi, C., Vanaja, M., & Annadurai, G. (2016). Anticancer and enhanced antimicrobial activity of biosynthesized silver nanoparticles against clinical pathogens. *Journal of Molecular Structure*, 1116, 165–173.
- Raliya, R., & Tarafdar, J. C. (2013). ZnO nanoparticle biosynthesis and its effect on phosphorous-mobilizing enzyme secretion and gum contents in Clusterbean (*Cyamopsis tetragonoloba L.*). *Agricultural Research*, 2(1), 48-57.
- Rao, M. D., & Gautam, P. (2016). Synthesis and characterization of ZnO nanoflowers using *Chlamydomonas reinhardtii*: A green approach. *Environmental Progress & Sustainable Energy*, 35(4), 1020-1026.
- Rauwel, P., Salumaa, M., Aasna, A., Galeckas, A., & Rauwel, E. (2016). A Review of the Synthesis and Photoluminescence Properties of Hybrid ZnO and Carbon Nanomaterials. *Journal of Nanomaterials*, 2016, 19.
- Ravikumar, S., Gokulakrishnan, R., & Boomi, P. (2012). In vitro antibacterial activity of the metal oxide nanoparticles against urinary tract infectious bacterial pathogens. *Asian Pacific Journal of Tropical Disease*, 2(2), 85–89.
- Reddy, A. J., Kokila, M. K., Nagabhushana, H., Rao, J. L., Shivakumara, C., Nagabhushana, B. M., & Chakradhar, R. P. S. (2011). Combustion synthesis, characterization and Raman studies of ZnO nanopowders. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 81(1), 53–58.
- Riddin, T. L., Gericke, M., & Whiteley, C. G. (2006). Analysis of the inter-and extracellular formation of platinum nanoparticles by *Fusarium oxysporum* f. sp. *lycopersici* using response surface methodology. *Nanotechnology*, 17(14), 3482.
- Roberts, P. J., & Der, C. J. (2007). Targeting the Raf-MEK-ERK mitogen-activated protein kinase cascade for the treatment of cancer. *Oncogene*, 26(22), 3291.
- Rowe, M. K., & Chuang, D.-M. (2004). An overview of pathways regulating apoptosis. *Expert Rev Mol Med*, 6(21), 1–18.
- Roy, S. K., Srivastava, R. K., & Shankar, S. (2010). Inhibition of PI3K/AKT and MAPK/ERK pathways causes activation of FOXO transcription factor, leading to cell cycle arrest and apoptosis in pancreatic cancer. *Journal of Molecular Signaling*, 5(1), 10.
- Saber, A., Alipour, B., Faghfoori, Z., & Khosroushahi, A. Y. (2017). Secretion metabolites of probiotic yeast, *Pichia kudriavzevii* AS-12, induces apoptosis pathways in human colorectal cancer cell lines. *Nutrition Research*.
- Sadayandi, K. (2016). Effect of temperature on structural, optical and photoluminescence studies on ZnO nanoparticles synthesized by the standard co-precipitation method. *Physica B: Condensed Matter*, 487(1), 1–7.

- Salata, O. (2004). Applications of nanoparticles in biology and medicine. *Journal of Nanobiotechnology*, 2(1), 3.
- Salam, H. A., Sivaraj, R., & Venkatesh, R. (2014). Green synthesis and characterization of zinc oxide nanoparticles from Ocimum basilicum L. var. purpurascens Benth-Lamiaceae leaf extract. *Materials Letters*, 131, 16–18.
- Samanta, P. K., & Chaudhuri, P. R. (2011). Substrate effect on morphology and photoluminescence from ZnO monopods and bipods. *Frontiers of Optoelectronics in China*, 4(2), 130–136.
- Sastry, M., Ahmad, A., Islam Khan, M., & Kumar, R. (2003). Biosynthesis of metal nanoparticles using fungi and actinomycete. *Current Science*, 85(2), 162–170.
- Sadeghi, B., & Gholamhoseinpoor, F. (2015). A study on the stability and green synthesis of silver nanoparticles using Ziziphora tenuior (Zt) extract at room temperature. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 134, 310–315.
- Seil, J. T., & Webster, T. J. (2012). Antimicrobial applications of nanotechnology: methods and literature. *International Journal of Nanomedicine*, 7, 2767.
- Selvakumari, D., Deepa, R., Mahalakshmi, V., Subhashini, P., & Lakshminarayan, N. (2015). Anti cancer activity of ZnO nanoparticles on MCF7 (breast cancer cell) and A549 (lung cancer cell). *Asian Research Publishing Network Journal of Engineering and Applied Sciences*, 10, 5418–5421.
- Senapati, S., Ahmad, A., Khan, M. I., Sastry, M., & Kumar, R. (2005). Extracellular biosynthesis of bimetallic Au–Ag alloy nanoparticles. *Small*, 1(5), 517–520.
- Setyawati, M. I., Tay, C. Y., & Leong, D. T. (2015). Mechanistic investigation of the biological effects of SiO<sub>2</sub>, TiO<sub>2</sub>, and ZnO nanoparticles on intestinal cells. *Small*, 11(28), 3458–3468.
- Shabestarian, H., Homayouni-Tabrizi, M., Soltani, M., Namvar, F., Azizi, S., Mohamad, R., & Shabestarian, H. (2017). Green synthesis of gold nanoparticles using Sumac aqueous extract and their antioxidant activity. *Materials Research*, 20(1), 264–270.
- Shankar, S. S., Ahmad, A., Pasricha, R., Khan, M. I., Kumar, R., & Sastry, M. (2004). Immobilization of biogenic gold nanoparticles in thermally evaporated fatty acid and amine thin films. *Journal of Colloid and Interface Science*, 274(1), 69–75.
- Shankar, S. S., Ahmad, A., Pasricha, R., & Sastry, M. (2003). Bioreduction of chloroaurate ions by geranium leaves and its endophytic fungus yields gold nanoparticles of different shapes. *Journal of Materials Chemistry*, 13(7), 1822–1826.

- Sharma, N. C., Sahi, S. V., Nath, S., Parsons, J. G., Gardea-Torresde, J. L., & Pal, T. (2007). Synthesis of plant-mediated gold nanoparticles and catalytic role of biomatrix-embedded nanomaterials. *Environmental Science & Technology*, 41(14), 5137–5142.
- Sharma, V., Anderson, D., & Dhawan, A. (2012). Zinc oxide nanoparticles induce oxidative DNA damage and ROS-triggered mitochondria mediated apoptosis in human liver cells (HepG2). *Apoptosis*, 17(8), 852–870.
- Shen, L., Bao, N., Yanagisawa, K., Domen, K., Gupta, A., & Grimes, C. A. (2006). Direct synthesis of ZnO nanoparticles by a solution-free mechanochemical reaction. *Nanotechnology*, 17(20), 5117.
- Singh, N., Zaidi, D., Shyam, H., Sharma, R., & Balapure, A. K. (2012). Polyphenols sensitization potentiates susceptibility of MCF-7 and MDA MB-231 cells to Centchroman. *PloS One*, 7(6), e37736.
- Silva, M. T. (2010). Secondary necrosis: the natural outcome of the complete apoptotic program. *FEBS Letters*, 584(22), 4491–4499.
- Singaravelu, G., Arockiamary, J. S., Kumar, V. G., & Govindaraju, K. (2007). A novel extracellular synthesis of monodisperse gold nanoparticles using marine alga, *Sargassum wightii Greville*. *Colloids and Surfaces B: Biointerfaces*, 57(1), 97–101.
- Singer, B., & Grunberger, D. (2012). *Molecular biology of mutagens and carcinogens*. Springer Science & Business Media.
- Singh, B. N., Singh, B. R., Sarma, B. K., & Singh, H. B. (2009). Potential chemoprevention of N-nitrosodiethylamine-induced hepatocarcinogenesis by polyphenolics from *Acacia nilotica* bark. *Chemico-Biological Interactions*, 181(1), 20–28.
- Singh, B. N., Singh, B. R., Singh, R. L., Prakash, D., Dhakarey, R., Upadhyay, G., & Singh, H. B. (2009). Oxidative DNA damage protective activity, antioxidant and anti-quorum sensing potentials of *Moringa oleifera*. *Food and Chemical Toxicology*, 47(6), 1109–1116.
- Singh, B. N., Rawat, A. K. S., Khan, W., Naqvi, A. H., & Singh, B. R. (2014). Biosynthesis of stable antioxidant ZnO nanoparticles by *Pseudomonas aeruginosa rhammolipids*. *PLoS One*, 9(9), 106937.
- Singh, B. N., Singh, B. R., Singh, R. L., Prakash, D., Sarma, B. K., & Singh, H. B. (2009). Antioxidant and anti-quorum sensing activities of green pod of *Acacia nilotica* L. *Food and Chemical Toxicology*, 47(4), 778–786.
- Sinha, S., Pan, I., Chanda, P., & Sen, S. K. (2009). Nanoparticles fabrication using ambient biological resources. *J Appl Biosci*, 19, 1113–1130.

- Sivaraj, R., Rahman, P. K. S. M., Rajiv, P., & Venkatesh, R. (2014). Biogenic zinc oxide nanoparticles synthesis using *Tabernaemontana Divaricata* leaf extract and its anticancer activity against MCF-7 breast cancer cell Lines. *International Conference on Advances in Agricultural, Biological & Environmental Sciences*, 1(1), 83–85.
- Sirelkhatim, A., Mahmud, S., Seen, A., Kaus, N. H. M., Ann, L. C., Bakhori, S. K. M., Mohamad, D. (2015). Review on zinc oxide nanoparticles: antibacterial activity and toxicity mechanism. *Nano-Micro Letters*, 7(3), 219–242.
- Slocik, J. M., & Wright, D. W. (2003). Biomimetic mineralization of noble metal nanoclusters. *Biomacromolecules*, 4(5), 1135–1141.
- Society, A. C. (2013). Cancer facts and figures 2013. American Cancer Society Atlanta.
- Sotiropoulou, S., Sierra-Sastre, Y., Mark, S. S., & Batt, C. A. (2008). Biotemplated nanostructured materials. *Chemistry of Materials*, 20(3), 821–834.
- Sriram, M. I., Kalishwaralal, K., & Gurunathan, S. (2012). Biosynthesis of silver and gold nanoparticles using *Bacillus licheniformis*. *Nanoparticles in Biology and Medicine: Methods and Protocols*, 33–43.
- Stanner, S. A., Hughes, J., Kelly, C. N. M., & Buttriss, J. (2004). A review of the epidemiological evidence for the “antioxidant hypothesis.” *Public Health Nutrition*, 7(3), 407–422.
- Stegenga, R. W., Al-Azawi, S., Bandyopadhyay, D., & Bandyopadhyay, K. (2011). Biosynthesis of gold nanoparticles by *Saccharomyces cerevisiae*. *The FASEB Journal*, 25(1 Supplement), 726–728.
- Stevenson, D. S. (2017). The Nature of Life and Its Potential to Survive. *Springer International Publishing AG*, 2017.
- Stoimenov, P. K., Klinger, R. L., Marchin, G. L., & Klabunde, K. J. (2002). Metal oxide nanoparticles as bactericidal agents. *Langmuir*, 18(17), 6679–6686.
- Sun, J.-H., Dong, S.-Y., Wang, Y.-K., & Sun, S.-P. (2009). Preparation and photocatalytic property of a novel dumbbell-shaped ZnO microcrystal photocatalyst. *Journal of Hazardous Materials*, 172(2), 1520–1526.
- Sun, Q., Van Dam, R. M., Willett, W. C., & Hu, F. B. (2009). Prospective study of zinc intake and risk of type 2 diabetes in women. *Diabetes Care*, 32(4), 629–634.
- Sun, Y., & Xia, Y. (2002). Shape-controlled synthesis of gold and silver nanoparticles. *Science*, 298(5601), 2176–2179.

- Sundrarajan, M., Ambika, S., & Bharathi, K. (2015). Plant-extract mediated synthesis of ZnO nanoparticles using Pongamia pinnata and their activity against pathogenic bacteria. *Advanced Powder Technology*, 26(5), 1294–1299.
- Sutradhar, P., & Saha, M. (2016). Green synthesis of zinc oxide nanoparticles using tomato (*Lycopersicon esculentum*) extract and its photovoltaic application. *Journal of Experimental Nanoscience*, 11(5), 314–327.
- Sutherland, R. L., Hall, R. E., & Taylor, I. W. (1983). Cell proliferation kinetics of MCF-7 human mammary carcinoma cells in culture and effects of tamoxifen on exponentially growing and plateau-phase cells. *Cancer Research*, 43(9), 3998–4006.
- Taccolla, L., Raffa, V., Riggio, C., Vittorio, O., Iorio, M. C., Vanacore, R., Cuschieri, A. (2011). Zinc oxide nanoparticles as selective killers of proliferating cells. *International Journal of Nanomedicine*, 6, 1129-1140.
- Tada-Oikawa, S., Ichihara, G., Suzuki, Y., Izuoka, K., Wu, W., Yamada, Y., Ichihara, S. (2015). Zn (II) released from zinc oxide nano/micro particles suppresses vasculogenesis in human endothelial colony-forming cells. *Toxicology Reports*, 2, 692–701.
- Talam, S., Karumuri, S. R., & Gunnam, N. (2012). Synthesis, characterization, and spectroscopic properties of ZnO nanoparticles. *ISRN Nanotechnology*, 2012, 1–6.
- Tao, J., Chen, X., Sun, Y., Shen, Y., & Dai, N. (2008). Controllable preparation of ZnO hollow microspheres by self-assembled block copolymer. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 330(1), 67–71.
- Tarafdar, J. C., & Raliya, R. (2013). Rapid, Low-Cost, and Ecofriendly Approach for Iron Nanoparticle Synthesis Using *Aspergillus oryzae* TFR9. *Journal of Nanoparticles*, 2013(1), 1–4.
- Taylor, E. N., & Webster, T. J. (2009). The use of superparamagnetic nanoparticles for prosthetic biofilm prevention. *International Journal of Nanomedicine*, 4, 145–152.
- Taylor, W. R., & Grabovich, A. (2009). Targeting the cell cycle to kill cancer cells. *Pharmacology: Principles and Practice*. Academic Press, Burlington, MA, 429–453.
- Teiten, M.-H., Eifes, S., Dicato, M., & Diederich, M. (2010). Curcumin—the paradigm of a multi-target natural compound with applications in cancer prevention and treatment. *Toxins*, 2(1), 128–162.
- Thakkar, K. N., Mhatre, S. S., & Parikh, R. Y. (2010). Biological synthesis of metallic nanoparticles. *Nanomedicine: Nanotechnology, Biology and Medicine*, 6(2), 257–262.

- Thema, F. T., Manikandan, E., Dhlamini, M. S., & Maaza, M. (2015). Green synthesis of ZnO nanoparticles via *Agathosma betulina* natural extract. *Materials Letters*, 161, 124–127.
- Tian, X., He, W., Cui, J., Zhang, X., Zhou, W., Yan, S., ... Yue, Y. (2010). Mesoporous zirconium phosphate from yeast biotemplate. *Journal of Colloid and Interface Science*, 343(1), 344–349.
- Tiffin, N., Adeyemo, A., & Okpechi, I. (2013). A diverse array of genetic factors contribute to the pathogenesis of systemic lupus erythematosus. *Orphanet journal of rare diseases*, 8(1), 2.
- Umrani, R. D., & Paknikar, K. M. (2014). Zinc oxide nanoparticles show antidiabetic activity in streptozotocin-induced Type 1 and 2 diabetic rats. *Nanomedicine*, 9(1), 89–104.
- Vainio, E. J., & Hantula, J. (2000). Direct analysis of wood-inhabiting fungi using denaturing gradient gel electrophoresis of amplified ribosomal DNA. *Mycological Research*, 104(8), 927–936.
- Valdiglesias, V., Costa, C., Kılıç, G., Costa, S., Pásaro, E., Laffon, B., & Teixeira, J. P. (2013). Neuronal cytotoxicity and genotoxicity induced by zinc oxide nanoparticles. *Environment International*, 55, 92–100.
- Vallee, B. L., & Auld, D. S. (1990). Zinc coordination, function, and structure of zinc enzymes and other proteins. *Biochemistry*, 29(24), 5647–5659.
- Van Engeland, M., Nieland, L. J. W., Ramaekers, F. C. S., Schutte, B., & Reutelingsperger, C. P. M. (1998). Annexin V-affinity assay: a review on an apoptosis detection system based on phosphatidylserine exposure. *Cytometry*, 31(1), 1–9.
- Vermes, I., Haanen, C., Steffens-Nakken, H., & Reutelingsperger, C. (1995). A novel assay for apoptosis flow cytometric detection of phosphatidylserine expression on early apoptotic cells using fluorescein labelled annexin V. *Journal of Immunological Methods*, 184(1), 39–51.
- Vidya, R., Ravindran, P., Fjellvåg, H., Svensson, B. G., Monakhov, E., Ganchenkova, M., & Nieminen, R. M. (2011). Energetics of intrinsic defects and their complexes in ZnO investigated by density functional calculations. *Physical Review B*, 83(4), 45206.
- Vidya, C., Hiremath, S., Chandraprabha, M. N., Antonyraj, M. A. L., Gopal, I. V., Jain, A., & Bansal, K. (2013). Green synthesis of ZnO nanoparticles by *Calotropis gigantea*. *Int J Curr Eng Technol*, 1, 118–120.
- Vigneshwaran, N., Kathe, A. A., Varadarajan, P. V., Nachane, R. P., & Balasubramanya, R. H. (2006). Biomimetics of silver nanoparticles by white rot fungus, *Phaenerochaete chrysosporium*. *Colloids and Surfaces B*:

*Biointerfaces*, 53(1), 55–59.

- Vijayakumar, S., & Ganesan, S. (2012). In vitro cytotoxicity assay on gold nanoparticles with different stabilizing agents. *Journal of Nanomaterials*, 2012, 14.
- Voldman, J., Gray, M. L., & Schmidt, M. A. (1999). Microfabrication in biology and medicine. *Annual Review of Biomedical Engineering*, 1(1), 401–425.
- Volesky, B., & Holan, Z. R. (1995). Biosorption of heavy metals. *Biotechnology Progress*, 11(3), 235–250.
- Wahab, R., Mishra, A., Yun, S.-I., Hwang, I. H., Mussarat, J., Al-Khedhairy, A. A., Shin, H.-S. (2012). Fabrication, growth mechanism and antibacterial activity of ZnO micro-spheres prepared via solution process. *Biomass and Bioenergy*, 39, 227–236.
- Wahab, R., Mishra, A., Yun, S.-I., Kim, Y.-S., & Shin, H.-S. (2010). Antibacterial activity of ZnO nanoparticles prepared via non-hydrolytic solution route. *Applied Microbiology and Biotechnology*, 87(5), 1917–1925.
- Wahab, R., Siddiqui, M. A., Saquib, Q., Dwivedi, S., Ahmad, J., Musarrat, J., Shin, H.-S. (2014). ZnO nanoparticles induced oxidative stress and apoptosis in HepG2 and MCF-7 cancer cells and their antibacterial activity. *Colloids and Surfaces B: Biointerfaces*, 117, 267–276.
- Wang, Y., Zhang, Q., Zhang, C., & Li, P. (2012). Characterisation and cooperative antimicrobial properties of chitosan/nano-ZnO composite nanofibrous membranes. *Food Chemistry*, 132(1), 419–427.
- Wang, Z. L. (2008). Splendid one-dimensional nanostructures of zinc oxide: a new nanomaterial family for nanotechnology. *Ac Nano*, 2(10), 1987–1992.
- Wang, Z., Qian, X., Yin, J., & Zhu, Z. (2004). Large-scale fabrication of tower-like, flower-like, and tube-like ZnO arrays by a simple chemical solution route. *Langmuir*, 20(8), 3441–3448.
- Waris, G., & Alam, K. (1998). Attenuated antigenicity of ribonucleoproteins modified by reactive oxygen species. *IUBMB Life*, 45(1), 33–45.
- Weinberg, R. A. (1996). How cancer arises. *Scientific American*, 275(3), 62–71.
- Wolff, A. C., Hammond, M. E. H., Hicks, D. G., Dowsett, M., McShane, L. M., Allison, K. H., Fitzgibbons, P. (2013). Recommendations for human epidermal growth factor receptor 2 testing in breast cancer: American Society of Clinical Oncology/College of American Pathologists clinical practice guideline update. *Archives of Pathology and Laboratory Medicine*, 138(2), 241–256.

- Wyllie, A. H., Kerr, J. F. R., & Currie, A. R. (1980). Cell death: the significance of apoptosis. *International Review of Cytology*, 68, 251–306.
- Xia, M., Knezevic, D., & Vassilev, L. T. (2011). p21 does not protect cancer cells from apoptosis induced by nongenotoxic p53 activation. *Oncogene*, 30(3), 346.
- Xia, T., Kovochich, M., Brant, J., Hotze, M., Sempf, J., Oberley, T., Nel, A. E. (2006). Comparison of the abilities of ambient and manufactured nanoparticles to induce cellular toxicity according to an oxidative stress paradigm. *Nano Letters*, 6(8), 1794–1807.
- Xie, J., Lee, J. Y., Wang, D. I. C., & Ting, Y. P. (2007). Silver nanoplates: from biological to biomimetic synthesis. *ACS Nano*, 1(5), 429–439.
- Xie, Y., He, Y., Irwin, P. L., Jin, T., & Shi, X. (2011). Antibacterial activity and mechanism of action of zinc oxide nanoparticles against *Campylobacter jejuni*. *Applied and Environmental Microbiology*, 77(7), 2325–2331.
- Xu, S., & Wang, Z. L. (2011). One-dimensional ZnO nanostructures: solution growth and functional properties. *Nano Research*, 4(11), 1013–1098.
- Xu, A.-W., Ma, Y., & Cölfen, H. (2007). Biomimetic mineralization. *Journal of Materials Chemistry*, 17(5), 415–449.
- Yamada, Y., Okada, T., Ueshima, O., & Kondo, K. (1973). Coenzyme Q System In The Classification Of The Ascosporogenous Yeast Genera *Hansenula* And *Pichia*. *The Journal of General and Applied Microbiology*, 19(3), 189–208.
- Yan, S., He, W., Sun, C., Zhang, X., Zhao, H., Li, Z., Han, X. (2009). The biomimetic synthesis of zinc phosphate nanoparticles. *Dyes and Pigments*, 80(2), 254–258.
- Yu, J., Li, C., & Liu, S. (2008). Effect of PSS on morphology and optical properties of ZnO. *Journal of Colloid and Interface Science*, 326(2), 433–438.
- Yuvakkumar, R., Suresh, J., Saravanakumar, B., Nathanael, A. J., Hong, S. I., & Rajendran, V. (2015). Rambutan peels promoted biomimetic synthesis of bioinspired zinc oxide nanochains for biomedical applications. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 137, 250–258.
- Yuvakkumar, R., Suresh, J., Nathanael, A. J., Sundrarajan, M., & Hong, S. I. (2014). Novel green synthetic strategy to prepare ZnO nanocrystals using rambutan (*Nephelium lappaceum* L.) peel extract and its antibacterial applications. *Materials Science and Engineering: C*, 41, 17–27.
- Zak, A. K., Abrishami, M. E., Majid, W. H. A., Yousefi, R., & Hosseini, S. M. (2011). Effects of annealing temperature on some structural and optical properties of ZnO nanoparticles prepared by a modified sol-gel combustion method. *Ceramics International*, 37(1), 393–398.

- Zak, A. K., Majid, W. H. A., Darroudi, M., & Yousefi, R. (2011). Synthesis and characterization of ZnO nanoparticles prepared in gelatin media. *Materials Letters*, 65(1), 70–73.
- Zak, A. K., Razali, R., Majid, W. H. A., & Darroudi, M. (2011). Synthesis and characterization of a narrow size distribution of zinc oxide nanoparticles. *International Journal of Nanomedicine*, 6, 1399-1403.
- Zak, A. K., Majid, W. H. A., Abrishami, M. E., & Yousefi, R. (2011). X-ray analysis of ZnO nanoparticles by Williamson-Hall and size-strain plot methods. *Solid State Sciences*, 13(1), 251–256.
- Zelechowska, K. (2014). Methods of ZnO nanoparticles synthesis. BioTechnologia. *Journal of Biotechnology Computational Biology and Bionanotechnology*, 95(2), 150-159.
- Zhang, F., & Yang, J. (2009). Preparation of nano-ZnO and its application to the textile on antistatic finishing. *International Journal of Chemistry*, 1(1), 18-22.
- Zhang, L., Ding, Y., Povey, M., & York, D. (2008). ZnO nanofluids—A potential antibacterial agent. *Progress in Natural Science*, 18(8), 939–944.
- Zhang, L., Gu, F. X., Chan, J. M., Wang, A. Z., Langer, R. S., & Farokhzad, O. C. (2008). Nanoparticles in medicine: therapeutic applications and developments. *Clinical Pharmacology and Therapeutics*, 83(5), 761–769.
- Zhang, L., Jiang, Y., Ding, Y., Povey, M., & York, D. (2007). Investigation into the antibacterial behaviour of suspensions of ZnO nanoparticles (ZnO nanofluids). *Journal of Nanoparticle Research*, 9(3), 479–489.
- Zheng, Y., Yu, X., Xu, X., Jin, D., & Yue, L. (2010). Preparation of ZnO particle with novel nut-like morphology by ultrasonic pretreatment and its luminescence property. *Ultrasonics Sonochemistry*, 17(1), 7–10.

## **BIODATA OF STUDENT**

Amin Boroumand Moghaddam, was born in Sabzevar, Iran on September 21<sup>st</sup>, 1988. He received his early education at Sharyati School. Later, he continued his secondary education at Valiasr School and Shahed High School in Sabzevar. In 2007, he enrolled at the Islamic Azad University of Tonekabon (Iran) as an undergraduate student and graduated in 2011 with a Bachelor of Microbiology degree. In 2011, he was admitted to the Master of Science in Cell and Molecular Biology in Guilan University of Gilan province, Iran and graduated in 2013. In September 2014, he started his PhD programme at Universiti Putra Malaysia. His research interests include the biosynthesis of nanoparticles for biomedical properties.

## LIST OF PUBLICATIONS

- Boroumand Moghaddam, A., Namvar, F., Moniri, M., Md Tahir, P., Azizi, S., & Mohamad, R. (2015). Nanoparticles biosynthesized by fungi and yeast: a review of their preparation, properties, and medical applications. *Molecules*, 20(9), 16540-16565.
- Boroumand Moghaddam, A., Moniri, M., Azizi, S., Rahim, R. A., Ariff, A. B., Saad, W. Z., & Mohamad, R. (2017). Biosynthesis of ZnO nanoparticles by a new *Pichia kudriavzevii* yeast strain and evaluation of their antimicrobial and antioxidant activities. *Molecules*, 22(6), 872.
- Moniri, M., Boroumand Moghaddam, A., Azizi, S., Rahim, R. A., Ariff, A. B., Saad, W. Z., & Mohamad, R. (2017) Production and status of bacterial cellulose in biomedical engineering. *Nanomaterials*, 7(9), 257.
- Azizi, S., Mohamad, R., Rahim, R. A., Boroumand Moghaddam, A., Moniri, M., Ariff, A., & Namvab, F. (2016). ZnO-Ag core shell nanocomposite formed by green method using essential oil of wild ginger and their bactericidal and cytotoxic effects. *Applied Surface Science*, 384, 517-524.
- Ghorbani, P., Soltani, M., Homayouni-Tabrizi, M., Namvar, F., Azizi, S., Mohammad, R., & Boroumand Moghaddam, A. (2015). Sumac silver novel biodegradable nano composite for bio-medical application: Antibacterial activity. *Molecules*, 20(7), 12946-12958.
- Namvar, F., Azizi, S., Mohamad, R., Moghaddam, A. B., Soltani, M., & Moshfegh, F. (2016). Nanosized silver-palm pollen nanocomposite, green synthesis, characterization and antimicrobial activity. *Research on Chemical Intermediates*, 42(3), 1571-1581.