



UNIVERSITI PUTRA MALAYSIA

**BIODEGRADATION OF WASTE AND PURE CANOLA OIL BY COLD-
ADAPTED *Rhodococcus* sp. AQ5-07 FROM ANTARCTICA**

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By

SALIHU IBRAHIM

**Thesis Submitted to the school of Graduate Studies, Universiti Putra Malaysia,
in Fulfillment of the Requirements for the degree of Doctor of Philosophy**

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DEDICATION

I hereby dedicate this thesis to the Almighty Allah who has been my help, sustainer, provider, guide, encouragement, keeper and my all in all throughout the course of my studies and also to my parents (Alhaji Ibrahim Umar and Hajiya Ramatu Ibrahim) whose prayers and support has kept me going. Finally to my brothers and sisters who are always by my side and ready to render any kind of support and assistance.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

BIODEGRADATION OF WASTE AND PURE CANOLA OIL BY COLD-ADAPTED *Rhodococcus* sp. AQ5-07 FROM ANTARCTICA

By

SALIHU IBRAHIM

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Chairperson: Siti Aqlima Ahmad, PhD

Faculty: Biotechnology and Biomolecular Sciences

With the increase in human activities in Antarctic region, the possibility of vegetable oil spillage becomes unavoidably high. The removal of oils including canola oil from the environment and wastewater using biological approaches is needed since the thermal process of oil degradation is ineffective and requires the use of high temperatures. More so, thermal degradation leads to the generation free radicals. Thus, in this present study, the potential canola oil-degrading ability of cold-adapted *Rhodococcus* sp. AQ5-07 from Antarctica was evaluated using one-factor-at-a-time (OFAT), and response surface method (RSM). The bacterial growth kinetics was examined and screened for biosurfactant production. The ability of *Rhodococcus* sp. AQ5-07 to degrade canola oil in the presence of different heavy metals and xenobiotics was also tested. Furthermore, canola oil degradation pathway was investigated via whole genome analysis. *Rhodococcus* sp. strain AQ5-07 was found to have a high canola oil-degrading ability over other strains tested. Considerable degradation (78.60% and 70.14%) of 3% waste and pure canola oil was achieved by this bacterium when incubated with 1.0 g/L ammonium sulphate, 0.3 g/L yeast extract, pH 7.5 and 10% inoculum at 10°C over 72 h incubation period. Optimisation of the medium conditions using response surface methodology (RSM) resulted in an increase in both oil degradations (87.61% and 86.34%). Three out of the 10 heavy metals tested namely mercury (Hg), cadmium (Cd) and silver (Ag) had a significant effect on canola oil degradation at 1 ppm. The IC₅₀ values of the waste canola oil (WCO) for Hg, Cd and Ag were found to be 0.3866, 0.4539 and 0.3217 ppm, respectively. Meanwhile, for the pure canola oil (PCO), the IC₅₀ was found to be Hg (0.2495), Cd (0.5452) and Ag (0.3088), respectively. This shows that the strain can also withstand 10 mg/L acrylamide, 50 mg/L phenol and 0.5% (v/v) diesel, respectively, for both oils. The bacterium was also found to produce high biosurfactant with beta haemolysis, high cellular hydrophobicity of 89.5% and 83.5% in hexadecane and tetradecane, respectively. The strain displayed 55 mm drop collapse with an oil displacement of 48 mm. Emulsification test (E₂₄) revealed the highest emulsification index of 92% in hexadecane. Haldane and Yano model best

describes the kinetics of the strain growth, the calculated constants for Haldane model such as maximum growth rate (μ_{max}), half saturation constant for maximal growth (K_s), and growth inhibition constant (K_i) tolerated were 0.142 h⁻¹, 7.743% (v/v) and 0.399% (v/v), respectively, whereas those for Yano model were 0.096 h⁻¹, 3.0% (v/v) and 4.527% (v/v), respectively. Lastly, the bacterium genome was sequenced using whole genome sequencing (WGS). The WGS was assembled into a draft genome of 6,753,902 bp with G+C content of 62.4% and 828.79x sequencing depth of coverage. It comprises 4 contigs with sizes of 81,655, 134,292, 214,483 and 6,323,472 bp. The smaller contigs could represent plasmids. The coding sequences (CDSs) contained a total of 6,350 annotated genes, 53 tRNA and 15 rRNA. The WGS showed that *Rhodococcus* sp. AQ5-07 was closely related to *Rhodococcus erythropolis* NBRC 15547 and *Rhodococcus qingshengii* JCM 15477, respectively. Analyses revealed the presence of lipase genes responsible for lipid degradation. This study therefore confirmed the possible use of cold-adapted *Rhodococcus* sp. AQ5-07 in biodegradation of canola oil-polluted Antarctic soils at low temperature, as it is disease free, eco-friendly, easier and cheaper.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**BIODEGRADASI SISA DAN MINYAK CANOLA TULEN DENGAN
DISESUAIKAN SEJUK *Rhodococcus* sp. AQ5-07 DARI ANTARCTICA**

Oleh

SALIHU IBRAHIM

Jun 2020

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Dengan peningkatan aktiviti manusia di kawasan Antartika, kemungkinan untuk berlaku pertumpahan minyak masak menjadi tinggi dan tidak dapat dielakkan. Penyingkiran minyak termasuk minyak canola dari persekitaran dan air sisa menggunakan pendekatan biologi diperlukan kerana proses termal degradasi minyak tidak berkesan dan memerlukan penggunaan suhu tinggi. Lebih-lebih lagi, degradasi termal membawa kepada penjana radikal bebas. Oleh itu, dalam kajian ini, keupayaan merendahkan minyak kanola *Rhodococcus* sp yang disesuaikan dengan sejuk. AQ5-07 dari Antartika dinilai menggunakan one-factor-at-a-time (OFAT), dan kaedah permukaan tindak balas (RSM). Kinetik pertumbuhan bakteria diperiksa dan disaring untuk pengeluaran biosurfaktan. Keupayaan *Rhodococcus* sp. AQ5-07 untuk menurunkan minyak kanola dengan adanya logam berat dan xenobiotik yang berbeza juga diuji. Selanjutnya, jalan degradasi minyak canola disiasat melalui analisis keseluruhan genom. *Rhodococcus* sp. AQ5-07 didapati mempunyai keupayaan yang tinggi untuk mengurai minyak kanola berbanding dengan isolat yang lain. Penguraian yang tinggi (78.60% dan 70.14%) adalah daripada 3% sisa dan minyak kanola tulen telah dicapai oleh bakteria ini apabila ianya diinkubasikan dengan 1.0 g/L amonium sulfat, 0.3 g/L ekstrak yis, pH 7.5 dan 10% inokulum pada 10°C dalam tempoh inkubasi 72 h. Pengoptimuman keadaan media menggunakan kaedah statistik gerak balas permukaan (RSM) menghasilkan peningkatan dalam kedua-dua penguraian minyak (87.61% and 86.34%). Tiga daripada 10 logam berat yang diuji iaitu merkuri (Hg), kadmium (Cd) dan perak (Ag) mempunyai kesan yang signifikan terhadap penguraian minyak canola pada 1 ppm. Oleh itu, kepekatan logam berat ini bervariasi daripada 0 - 1 ppm untuk dilihat di mana kepekatan setiap logam akan memberi kesan. IC₅₀ bagi sisa minyak kanola (WCO) Hg, Cd dan Ag didapati masing-masing 0.3866, 0.4539 dan 0.3217 ppm. Sementara itu, bagi minyak kanola tulen (PCO), IC₅₀ didapati Hg (0.2495), Cd (0.5452) dan Ag (0.3088). Ini menunjukkan bahawa strain ini juga mampu bertahan pada 10 mg/L akrilamida, 50 mg/L fenol dan 0.5% (v/v) diesel, masing-masing untuk kedua-dua jenis minyak. Bakterium ini juga didapati mampu menghasilkan biosurfaktan yang tinggi dengan hemolisis beta, hidrofobisiti selular yang tinggi 89.5% dan 83.5% dalam heksadekana

dan tetradekana masing-masing. Ini menunjukkan kejatuhan penurunan (drop collapse) yang positif dengan anjakan minyak (oil displacement) sebanyak 55 mm. Indeks emulsifikasi (E_{24}) mendedahkan indeks pengemulsian tertinggi pada 92% dalam heksadekana. Haldane dan Yano model terbaik menggambarkan kinetik pertumbuhan terikan. Pemalar yang dikira bagi model Haldane seperti kadar pertumbuhan maksima (μ_{max}), pemalar ketepuan tepu untuk pertumbuhan maksimum (K_s), dan kepekatan yang merencat pertumbuhan (K_i) masing-masing adalah pada 0.142 h^{-1} , 7.743% (v/v) dan 0.399% (v/v), manakala bagi model Yano masing-masing adalah 0.096 h^{-1} , 3.0% (v/v) dan 4.527% (v/v). Akhir sekali, bakteria yang diujuk menggunakan penjujukan keseluruhan genom. WGS telah disusun menjadi draf genom 6,753,902 bp dengan kandungan G+C 62.4% dan 828.79x penjujukan kedalaman liputan (sequencing depth of coverage). Ia terdiri daripada 4 contigs dengan saiz 81,655, 134,292, 214,483 dan 6,323,472 bp. Contigs yang lebih kecil boleh dikira sebagai plasmid. Jujukan pengekodan (coding sequences) (CDSs) mengandungi sejumlah 6,350 gen penjelasan (annotated genes), 53 tRNA dan 15 rRNA. WGS menunjukkan bahawa *Rhodococcus* sp. AQ5-07 berkait rapat dengan *Rhodococcus erythropolis* NBRC 15547 dan *Rhodococcus qingshengii* JCM 15477, masing-masing. Analisis mendedahkan bahawa kehadiran gen lipase yang bertanggungjawab terhadap penguraian lipid. Oleh itu, kajian ini mengesahkan kemungkinan penggunaan *Rhodococcus* sp. AQ5-07 dalam biodegradasi tanah Antartika tercemar minyak canola pada suhu rendah, kerana bebas penyakit, mesra alam, lebih mudah dan lebih murah.

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APPROVAL

I certify that a Thesis Examination Committee has met on (Date of viva voce) to conduct the final examination of Salihu Ibrahim on his Doctor of Philosophy thesis entitled “**Biodegradation of waste and pure canola oil by cold-adapted *Rhodococcus* sp. AQ5-07 from Antarctica**” in Accordance with the Universities and University College Act 1971 and the constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy Degree.

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REFERENCES

- Abatenh, E., Gizaw, B., Tsegaye, Z., and Wassie, M. (2017). Application of microorganisms in bioremediation-review. *Journal of Environmental Microbiology*, 1(1), 2–9.
- Abdel-Salam, A. M., Al-Dekheil, A., Babkr, A., Farahna, M., and Mousa, H. M. (2010). High fiber probiotic fermented mares milk reduces the toxic effects of mercury in rats. *North American Journal of Medical Sciences*, 2(12), 569–575. <https://doi.org/10.4297/najms.2010.2569>
- Abdulrasheed, M., Zakaria, N. N., Ahmad Roslee, A. F., Shukor, M. Y., Zulkharnain, A., Napis, S., Convey, P., Alias, S. A., Gonzalez-Rocha, G., and Ahmad, S. A. (2020). Biodegradation of diesel oil by cold-adapted bacterial strains of *Arthrobacter* spp. from Antarctica. *Antarctic Science*, 13, 1–13.
- Abeynayaka, A., and Visvanathan, C. (2011). Performance comparison of mesophilic and thermophilic aerobic sidestream membrane bioreactors treating high strength wastewater. *Bioresource Technology*, 102(9), 5345–5352.
- Abioye, O. P., Agamuthu, P., and Abdul Aziz, A. R. (2012). Biodegradation of used motor oil in soil using organic waste amendments. *Biotechnology Research International*, 2012, 1–8. <https://doi.org/10.1155/2012/587041>
- Abollino, O., Malandrino, M., Zelano, I., Giacomino, A., Buoso, S., and E. Mentasti. (2012). Characterization of the element content in lacustrine ecosystems in Terra Nova Bay, Antarctica. *Microchemical Journal*, 105, 142–151.
- Abou-Shanab, R. A. I., van Berkum, P., and Angle, J. S. (2007). Heavy metal resistance and genotypic analysis of metal resistance genes in gram-positive and gram-negative bacteria present in Ni-rich serpentine soil and in the rhizosphere of *Alyssum murale*. *Chemosphere*, 68(2), 360–367.
- Abouseoud, M., Maachi, R., Amrane, A., Boudergua, S., and Nabi, A. (2008). Evaluation of different carbon and nitrogen sources in production of biosurfactant by *Pseudomonas fluorescens*. *Desalination*, 223(1–3), 143–151.
- Abu- Ruwaida, A. S., Banat, I. M., Haditirto, S., Salem, A., and Kadri, M. (1991). Isolation of biosurfactant- producing bacteria, product characterization, and evaluation. *Acta Biotechnologica*, 11(4), 315–324.
- Abubakar, A., Muhammad, A., Shehu, D., Ya’u, M., Tanko, A. S., Ibrahim, H., Usman, S., Mohamat-yusuff, F., and Ibrahim, S. (2017). Modelling growth kinetics of *Klebsiella* sp. FIRD 2 on TBT-resistant containing lead. *Journal of Applied Science and Environmental Management*, 21(6), 1085–1091.
- Abubakar, M., Abdul Habib, N. M. S., Manogaran, M., Yasid, N. A., Alias, S. A., Ahmad, S. A., Smykla, J., Hassan, M. A., and Abd Shukor, M. Y. (2019). Response surface-based optimization of the biodegradation of a simulated vegetable oily ballast wastewater under temperate conditions using the antarctic bacterium *Rhodococcus erythropolis* ADL36. *Desalination and Water Treatment*, 144, 129–137. <https://doi.org/10.5004/dwt.2019.23529>
- Abusham, R. A., Rahman, R. N., Salleh, A. B., and Basri, M. (2009). Optimization of physical factors affecting the production of thermo-stable organic solvent-tolerant protease from a newly isolated halo tolerant *Bacillus subtilis* strain Rand. *Microbial Cell Factories*, 8, 20–28.
- Achir, N., Kara, W., Chipeaux, C., Trezzani, I., and Cuvelier, M. (2006). Effect of energy transfer conditions on the chemical degradation of frying oil. *European Journal of Lipid Science and Technology*, 12, 999–1006.

- Affandi, I. E., Suratman, N. H., Abdullah, S., Ahmad, W. A., and Zakaria, Z. A. (2014). Degradation of oil and grease from high-strength industrial effluents using locally isolated aerobic biosurfactant-producing bacteria. *International Biodeterioration and Biodegradation*, 95, 33–40. <https://doi.org/10.1016/j.ibiod.2014.04.009>
- Agarry, S. E., and Solomon, B. O. (2008). Kinetics of batch microbial degradation of phenols by indigenous *Pseudomonas fluorescens*. *International Journal of Environmental Science & Technology*, 5(2), 223–232.
- Agarwal, R., Mahanty, B., and Dasu, V. V. (2009). Modeling growth of *Cellulomonas cellulans* NRRL B 4567 under substrate inhibition during Cellulase production. *Chemical and Biochemical Engineering Quarterly*, 23(2), 213–218.
- Ahmad, S. A., Shamaan, N. A., Syed, M. A., Dahalan, F. A., Abdul Khalil, K., Ab Rahman, N. A., and Shukor, M. Y. (2017). Phenol degradation by *Acinetobacter* sp. in the presence of heavy metals. *Journal of the National Science Foundation of Sri Lanka*, 45(3), 247–253. <https://doi.org/10.4038/jnsfsr.v45i3.8189>
- Ahmad, S. A., Shukor, M. Y., Shamaan, N. A., Mac Cormack, W. P., and Syed, M. A. (2013). Molybdate reduction to molybdenum blue by an Antarctic bacterium. *BioMed Research International*, 2013, 1–10. <https://doi.org/10.1155/2013/871941>
- Ahmad, S. A., Asokan, G., Yasid, N. A., Nawawi, N. M., Subramaniam, K., Zakaria, N. N., and Shukor, M. Y. (2018). Effect of heavy metals on biodegradation of phenol by Antarctic: *Arthrobacter bamsusae* strain AQ5-003. *Malaysian Journal of Biochemistry and Molecular Biology*, 2, 47–51.
- Ahmad, S. A., Ibrahim, S., Shukor, M. Y., Johari, W. L. W. J., Rahman, N. A., and Syed, M. A. S. (2015). Biodegradation kinetics of caffeine by *Leifsonia* sp. strain SIU. *Journal of Chemical and Pharmaceutical Sciences*, 8(2), 312–316.
- Ahmad, S. A., KuAhamad, K. N. E., WanJohari, W. L., Halmi, M. I. E., Shukor, M. Y., and Yusof, M. T. (2014). Kinetics of diesel degradation by an acrylamide-degrading bacterium. *Rendiconti Lincei*, 25(4), 505–512. <https://doi.org/10.1007/s12210-014-0344-7>
- Ahmad, S. A., Shamaan, N. A., Arif, N. M., Koon, G. B., Shukor, M. Y. A., and Syed, M. A. (2012). Enhanced phenol degradation by immobilized *Acinetobacter* sp. strain AQ5NOL 1. *World Journal of Microbiology & Biotechnology*, 28(1), 347–352. <https://doi.org/10.1007/s11274-011-0826-z>
- Aiba, S., Shoda, M., and Nagalani, M. (1968). Kinetics of product inhibition in alcohol fermentation. *Biotechnology and Bioengineering*, 10(6), 845–864.
- Aislabie, J. M., Balks, M. R., Foght, J. M., and Waterhouse, E. J. (2004). Hydrocarbon spills on Antarctic soils: effects and management. *Environmental Science & Technology*, 38(5), 1265–1274.
- Ajoku, G. A. O. (2013). Kinetic model of pH effect on bioremediation of crude petroleum contaminated soil. 1. model development. *American Journal of Chemical Engineering*, 1(1), 6–10. <https://doi.org/10.11648/j.ajche.20130101.12>
- Akaike, H. (1974). New look at the statistical model identification. *IEEE Transactions on Automatic Control*, 19(6), 716–723.
- Aken, B. Van. (2009). Transgenic plants for enhanced phytoremediation of toxic explosives. *Current Opinion in Biotechnology*, 20(2), 231–236. <https://doi.org/10.1016/j.copbio.2009.01.011>
- Al-Darbi, M. M., Saeed, N. O., Islam, M. R., and Lee, K. (2005). Biodegradation of natural oils in seawater. *Energy Sources*, 27(1–2), 19–34.

- Altmaier, M., Herpers, U., Delisle, G., Merchel, S., and Ott, U. (2010). Glaciation history of Queen Maud Land (Antarctica) reconstructed from in-situ produced cosmogenic ^{10}Be , ^{26}Al and ^{21}Ne . *Polar Science*, 4(1), 42–61.
- Aluyor, E. O., Obahiagbon, K. O., and Ori-jesu, M. (2009). Biodegradation of vegetable oils: A review. *Scientific Research and Essay*, 4(6), 543–548. <https://doi.org/E-ISSN 0976-3945>
- Anonymous, K. (2020). I-80 near Colfax reopened after canola oil spill: <https://www.kolotv.com/2020/06/26/canola-oil-spill>.
- Aparna, A., Srinikethan, G., and Hedge, S. (2011). Effect of addition of biosurfactant produced by *Pseudomonas* sps. on biodegradation of crude oil. In *In: 2nd International Proceedings of Chemical, Biological & Environmental Engineering, vol. 61 ACSIT Press, Singapore*. pp. 71–75.
- Arażny, A., Kejna, M., and Sobota, I. (2013). Ground temperature at the Henryk Arctowski station (King George Island, Antarctic) – case study from the period January 2012 to February 2013. *Bulletin of Geography- Physical Geography Series*, 6(1), 59–80. <https://doi.org/10.2478/bgeo-2013-0004>
- Arisoy, K. (2008). Oxidative and thermal instability of biodiesel. Energy sources part A. *Recovery Utilization and Environmental Effects*, 30(16), 1516–1522.
- Ash, M. (2012). Soybeans & oil crops: USDA- United States, department of Agriculture 2012. Available <https://www.ers.usda.gov/topics/crops/soybeans-oil-crops/canola.aspx>.
- Asira, E. E. (2013). Factors that determine bioremediation of organic compounds in the soil. *Academic Journal of Interdisciplinary Studies*, 2(13), 125–128.
- Babák, L., Šupinová, P., and Burdychová, R. (2012). Growth models of *Thermus aquaticus* and *Thermus scotoductus*. *Acta Universitatis Agriculturae et Silviculturae Mendelianae Brunensis*, 60(5), 19–26.
- Banat, I. M., Makkar, R. S., and Cameotra, S. S. (2000). Potential commercial applications of microbial surfactants. *Applied Microbiology and Biotechnology*, 53(5), 495–508. <https://doi.org/10.1007/s002530051648>
- Baranyi, J. (1995). Mathematics of predictive food microbiology. *International Journal of Food Microbiology*, 26(2), 199–218. <https://doi.org/10.1016/0168-1>
- Bargagli, R. (2008). Environmental contamination in Antarctic ecosystems. *Science of the Total Environment*, 400(1–3), 212–226. <https://doi.org/10.1016/j.scitotenv.2>
- Bas, D., Ismail, H., and Boyaci, J. (2007). Modeling and optimization I: usability of response surface methodology. *Journal of Food Engineering*, 78(3), 836–845.
- Bassegoda, A., Pastor, F. I. J., and Diaz, P. (2012). *Rhodococcus* sp. strain CR-53 LipR , the first member of a new bacterial lipase family (Family X) displaying an unusual Y-Type oxyanion hole, similar to the *Candida antarctica* lipase clan. *Applied and Environmental Microbiology*, 78(6), 1724–1732. <https://doi.org/10.1128/AEM.0>
- Battersby, N. S. (2000). The biodegradability and microbial toxicity testing of lubricants – some recommendations. *Chemosphere*, 42, 1011–1027.
- Bednarski, W., Adamczak, M., Tomasik, J., and Paszczyk, M. (2004). Application of oil refinery waste in the biosynthesis of glycolipids by yeast. *Bioresource Technology*, 95, 15–18.
- Behera, S. K., Meena, H., Chakraborty, S., and Meikap, B. C. (2018). Application of response surface methodology (RSM) for optimization of leaching parameters for ash reduction from low-grade coal. *International Journal of Mining Science and Technology*, 28(4), 621–629. <https://doi.org/10.1016/j.ijmst.2018.04.014>

- Bentley, D. R., Balasubramanian, S., Swerdlow, H. P., Smith, G. P., Milton, J., Brown, C. G., Hall, K. P., Evers, D. J., Barnes, C. L., Helen, R. B., Jonathan, M. B., Jason, B., Richard, J. C., Keira, C., Anthony, J. C., Darren, J. E., Michael, R. F., Niall, A. G., Sean, J. H., Leslie, J. I., Mirian, S. K., Scott, M. K., Heng, L., Xiaohai, L., Klaus, S. M., Lisa, J. M., Bojan, O., Tobias, O., Michael, L. P., Mark, R. P., Isabelle, M. J. R., Mark, T. R., Roberto, R., Chiara, R., Mark, T. R., Andrea, S., Subramanian, V. S., A., S., Gary, P. S., Mark, E. S., Vincent, P. S., Anastassia, S., Peta, Torrance, E., Svilen, S. T., Eric H. V., Klaudia, W., Xiaolin, W., Lu, Z., Mohammed, D. A., Carole, A., Ify, C. A., David, M. D. B., Iain, R. B., Saibal, B., Selena, G. B., Primo, A. B., Vincent, A. B., Kevin, F. B., Claire, B., Phillip, J. B., Boodhun, A., Brennan, J. S., Bridgham, J. A., Brown, R. C., Brown, A. A., Buermann, D. H., Bundu, A. A., Burrows, J. C., Carter, N. P., Castillo, N., Catenazzi, M. C. E., Chang, S., Cooley, N. R., Crake, N. R., Dada, O. O., Diakoumakos, K. D., Dominguez-Fernandez, B., Earnshaw, D. J., Egbujor, U. C., Elmore, D. W., Etchin, S. S., Ewan, M. R., Fedurco, M., Fraser, L. J., Fajardo, K. V. F., Furey, W. S., George, D., Gietzen, K. J., Goddard, C. P., Golda, G. S., Granieri, P. A., Green, D. E., Gustafson, D. L., Hansen, N. F., Harnish, K., Haudenschild, C. D., Heyer, N. I., Hims, M. M., Ho, J. T., Horgan, A. M., Hoschler, K., Hurwitz, S., Ivanov, D. V., Johnson, M. Q., James, T., Jones, T. A. H., Kang, D., Kerelska, T. H., Kersey, A. D., Khrebtukova, I., Kindwall, A. P., Kingsbury, Z., Kokko-Gonzales, P. I., Kumar, A., Laurent, M. A., Lawley, C. T., Lee, S. E., Lee, X., Liao, A. K., Loch, A. K., Lok, M., Luo, S., Mammen, R. M., Martin, J. W., McCauley, P. G., McNitt, P., Mehta, P., Moon, K. W., Mullens, J. W., Newington, T., Ning, Z., Ng, B. L., Novo, S. M., O'Neill, M. J., Osborne, M. A., Osnowski, A., Ostadan, O., Paraschos, L. L., Pickering, L., Pike, A. C., Pike, A. C., Pinkard, D. C., Pliskin, D. P., Podhasky, J., Quijano, V. J., Raczy, C., Rae, V. H., Rawlings, S. R., Rodriguez, A. C., Roe, P. M., Rogers, J., Bacigalupo, M. C. R., Romanov, M., Romieu, A., Roth, R. K., Rourke, N. J., Ruediger, S. T., Rusman, E., Sanches-Kuiper, R. S., Schenker, M. R., Seoane, J. M., Shaw, R. J., Shiver, M. V., Short, S. W., Sizto, N. L., Sluis, J. P., Smith, M. A., Sohna, J. E. S., Spence, E. J., Stevens, E., Sutton, N., Szajkowski, L., Tregidgo, C. L., Turcatti, G., Vande-Vondele, S., Verhovskiy, Y., Virk, S. M., Wakelin, S., Walcott, G. C., Wang, J., Worsley, G. W., Yan, J., Yau, L., Zuerlein, M., Rogers, J., Mullikin, J. C., Hurles, M. E., McCooke, N. J., West, J. S., Oaks, F. L., Lundberg, P. L., Klenerman, D., Durbin, R., and Smith, A. J. (2008). Accurate whole human genome sequencing using reversible terminator chemistry. *Nature*, 456(7218), 53–59. <https://doi.org/10.1038/nature07517>
- Bergstrom, D. M., and Chown, S. L. (1999). Life at the front: history, ecology and change on southern ocean islands. *Trends in Ecology & Evolution*, 14(12), 472–477. [https://doi.org/10.1016/S0169-5347\(99\)01688-2](https://doi.org/10.1016/S0169-5347(99)01688-2)
- Bharathi, P., Elavarasi, N., and Mohanasundaram, S. (2012). Studies on rate of biodegradation of vegetable (Coconut) oil by using *Pseudomonas aureginosa*. *International Joournal of Environmental Biology*, 2(1), 12–19.
- Bhattacharya, M., Biswas, D., Sana, S., and Datta, S. (2015). Biodegradation of waste lubricants by a newly isolated *Ochrobactrum* sp. C1. *3 Biotech*, 5(5), 807–817. <https://doi.org/10.1007/s13205-015-0282-9>
- Bhumibhamon, O., Kopraserstak, A., and Funthong, S. (2002). Biotreatment of high fat and oil wastewater by lipase producing microorganisms. *Kasetsart Journal of Natural Sciences*, 36(3), 261–267.

- Bland, C., Ramsey, T. L., Sabree, F., Lowe, M., Brown, K., Kyrpides, N. C., and Hugenholtz, P. (2007). CRISPR Recognition Tool (CRT): A tool for automatic detection of clustered regularly interspaced palindromic repeats. *BMC Bioinformatics*, 8, 1–8. <https://doi.org/10.1186/1471-2105-8-209>
- Bodour, A. A., and Miller-Maier, R. M. (1998). Application of a modified drop-collapse technique for surfactant quantitation and screening of biosurfactant-producing microorganisms. *Journal of Microbiological Methods*, 32(3), 273–280. [https://doi.org/10.1016/S0167-7012\(98\)00031-1](https://doi.org/10.1016/S0167-7012(98)00031-1)
- Bofill, C., Prim, N., Mormeneo, M., Manresa, A., Javier Pastor, F. I., and Diaz, P. (2010). Differential behaviour of *Pseudomonas* sp. 42A2 LipC, a lipase showing greater versatility than its counterpart LipA. *Biochimie*, 92(3), 307–316.
- Boopathy, R. (2000). Factors limiting bioremediation technologies. *Bioremediation Technology*, 74, 63–67.
- Borghini, F., Grimalt, J. O., Sanchez-Hernandez, J. C., and Bargagli, R. (2005). Organochlorine pollutants in soils and mosses from victoria land (Antarctica). *Chemosphere*, 58(3), 271–278. [ht](https://doi.org/10.1016/j.chemosphere.2004.11.011)
- Borugadda, V. B., and Goud, V. V. (2015). Response surface methodology for optimization of bio-lubricant basestock synthesis from high free fatty acids castor oil. *Energy Science and Engineering*, 3(4), 371–383. <https://doi.org/10.1002/ese3.77>
- Buchanan, R. L. (1993). Predictive food microbiology. *Trends in Food Science and Technology*, 4(1), 6–11.
- Buchanan, R. L., Whiting, R. C., and Damert, W. C. (1997). When is simple good enough: A comparison of the Gompertz, Baranyi, and three-phase linear models for fitting bacterial growth curves. *Food Microbiology*, 14(4), 313–326. <https://doi.org/10.1006/fmic.1997.0125>
- Büdel, B., and Colesie, C. (2014). Biological soil crusts. In *Antarctic Terrestrial Microbiology*, 3, 131–161. <https://doi.org/10.1007/978-3-642-45213-0>
- Buranasilp, K., and Charoenpanich, J. (2011). Biodegradation of acrylamide by *Enterobacter aerogenes* isolated from wastewater in Thailand. *Journal of Environmental Sciences*, 23(3), 396–403. [https://doi.org/10.1016/S1001-](https://doi.org/10.1016/S1001-0745(11)00101-1)
- Burns, K. A., Codi, S., and Duke, N. C. (2000). Gladstone, Australia field studies: weathering and degradation of hydrocarbons in oiled mangrove and salt marsh sediments with and without the application of an experimental bioremediation protocol. *Marine Pollution Bulletin*, 41(7–12), 392–402.
- Cabrerizo, A., Dachs, J., Barcelo, D., and Jones, K. C. (2012). Influence of organic matter content and human activities on the occurrence of organic pollutants in Antarctic soils, lichens, grass, and mosses. *Environmental Science & Technology*, 46(3), 1396–1405.
- Callaghan, A. V., Morris, B. E. L., Pereira, I. A. C., McInerney, M. J., Austin, R. N., Groves, J. T., Kukor, J. J., Suflita, J. M., Young, L. Y., Zylstra, G. J., and Wawrik, B. (2012). The genome sequence of *Desulfatibacillum alkenivorans* AK-01: A blueprint for anaerobic alkane oxidation. *Environmental Microbiology*, 14(1), 101–113. <https://doi.org/10.1111/j.1462-2920.2011.02516.x>
- Campo, P., Zhao, Y., Suidan, M. T., Venosa, A. D., and Sorial, G. A. (2007). Biodegradation kinetics and toxicity of vegetable oil triacylglycerols under aerobic conditions. *Chemosphere*, 68(11), 2054–2062.
- Cao, J., Lai, Q., Yuan, J., and Shao, Z. (2015). Genomic and metabolic analysis of fluoranthene degradation pathway in *Celeribacter indicus* P73 T. *Scientific*

- Reports*, 5, 1–12. <https://doi.org/10.1038/srep07741>
- Cary, S. C., McDonald, I. R., Barrett, J. E., and Cowan, D. A. (2010). On the rocks: the microbiology of Antarctic dry valley soils. *Nature Reviews Microbiology*, 8(2), 129–138. <https://doi.org/10.1038/nrmicro2281>
- Čejková, A., Masák, J., Jirků, V., Veselý, M., Pátek, M., and Nešvera, J. (2005). Potential of *Rhodococcus erythropolis* as a bioremediation organism. *World Journal of Microbiology and Biotechnology*, 21(3), 317–321.
- Chan, Y. J., Chong, M. F., Law, C. L., and Hassell, D. G. (2009). A review on anaerobic-aerobic treatment of industrial and municipal wastewater. *Chemical Engineering Journal*, 155(1–2), 1–18. <https://doi.org/10.1016/j.cej.2009.06.041>
- Chandran, P., and Das, N. (2010). Biosurfactant production and diesel oil degradation by yeast species *Trichosporon asahii* isolated from petroleum hydrocarbon contaminated soil. *International Journal of Engineering Science and Technology*, 2(12), 6942–6953.
- Chen, S. Y., Lu, W. Bin, Wei, Y. H., Chen, W. M., and Chang, J. S. (2007). Improved production of biosurfactant with newly isolated *Pseudomonas aeruginosa* S2. *Biotechnology Progress*, 23(3), 661–666. <https://doi.org/10.1021/bp0700152>
- Chhetri, A., Watts, K., and Islam, M. (2008). Waste cooking oil as an alternate feedstock for biodiesel production. *Energies*, 1(1), 3–18. <https://doi.org/10.3390/en1010003>
- Choe, E., and Min, D. B. (2007). Chemistry of deep-fat frying oils. *Journal of Food Science*, 72(5), 1–10. <https://doi.org/10.1111/j.1750-3841.2007.00352.x>
- Chown, S. L., and Convey, P. (2007). Spatial and temporal variability across life's hierarchies in the terrestrial Antarctic. *Philosophical transactions of the royal society B: Biological sciences*, 362, 2307–2331.
- Chu, W., Dang, N., Kok, Y., Yap, K. I., Phang, S., and Convey, P. (2019). Heavy metal pollution in Antarctica and its potential impacts on algae. *Polar Science*, 20(1), 75–83. <https://doi.org/10.1016/j.polar.2018.10.004>
- Chudobova, D., Maskova, D., Nejdil, L., Kopel, P., Merlos-Rodrigo, M. A., Adam, V., and Kizek, R. (2013). The effect of silver ions and silver nanoparticles on *Staphylococcus aureus*. In *Microbial pathogens and strategies for combating them: science, technology and education* (A. Méndez-Vilas, Ed.) pp. 728–735.
- Chun, J., Grim, C. J., Hasan, N. A., Lee, J. H., Choi, S. Y., Haley, B. J., Taviani, E., Jeon, Y.-S., Kim, D. W., Lee, J.-H., Brettin, T. S., Bruce, D. C., Challacombe, J. F., Detter, J. C., Han, C. S., Munk, A. C., Chertkov, O., Meincke, L., Saunders, E., Walters, R. A., Huq, A., Nair, G. B., and Colwell, R. R. (2009). Comparative genomics reveals mechanism for short-term and long-term clonal transitions in pandemic *Vibrio cholerae*. *Proceedings of the National Academy of Sciences*, 106(36), 15442–15447. <https://doi.org/10.1073/pnas.0907787106>
- Cincinelli, A., Martellini, T., Del Bubba, M., Lepri, L., Corsolini, S., Borghesi, N., King, M. D., and Dickhut, R. M. (2009). Organochlorine pesticide air-water exchange and bioconcentration in krill in the ross sea. *Environmental Pollution*, 157(7), 2153–2158. <https://doi.org/10.1016/j.envpol.2009.02.010>
- Čipinytė, V., Grigiškis, S., and Baškys, E. (2009). Selection of fat-degrading microorganisms for the treatment of lipid-contaminated environment. *Biologija*, 55(3), 84–92. <https://doi.org/10.2478/v10054-009-0014-3>
- COMNAP. (2012). Council of Managers of National Antarctic Programs. Main Antarctic facilities operated by national Antarctic programs in the Antarctic Treaty area (south of 60° latitude south). <https://www.comnap>.

- aq/Information/SitePages/Home.aspx.
- Contesini, F. J., da Silva, V. C., Maciel, R. F., de Lima, R. J., Barros, F. F., and de Oliveira, C. P. (2009). Response surface analysis for the production of an enantioselective lipase from *Aspergillus niger* by solid-state fermentation. *The Journal of Microbiology*, 47(5), 563–571.
- Convey, P. (2007). Antarctic ecosystems. In: Levin, S.A. (Ed.), *Encyclopedia of Biodiversity*, 2nd ed. (online). Elsevier, San Diego, USA, <http://dx.doi.org/10.1016/B0-12-226865-2/00014-6>.
- Convey, P., Coulson, S. J., Worland, M. R., and Sjöblom, A. (2018). The importance of understanding annual and shorter-term temperature patterns and variation in the surface levels of polar soils for terrestrial biota. *Polar Biology*, 41(8), 1587–1605. <https://doi.org/10.1007/s00300-018-2299-0>
- Convey, P., Gibson, J., Hillenbrand, C. D., Hodgson, D. A., Pugh, P. J. A., Smellie, J. L., and Stevens, M. I. (2008). Antarctic terrestrial life – challenging the history of the frozen continent? *Biological Reviews*, 83, 103–17.
- Convey, Peter, Hughes, K. A., and Tin, T. (2012). Continental governance and environmental management mechanisms under the Antarctic Treaty system: Sufficient for the biodiversity challenges of this century? *Biodiversity*, 13(3–4), 234–248. <https://doi.org/10.1080/14888386.2012.703551>
- Dandare, S. U., Skvortsov, T., Arkhipova, K., and Allena, C. C. R. (2018). Draft genome sequence of *Rhodococcus* sp. strain NCIMB 12038, a Naphthalene-degrading bacterium. *Genome Announcements*, 6(1), 1–2.
- Danikuu, F. M., and Sowley, E. N. K. (2014). Biodegradation of shea nut cake by indigenous soil bacteria. *Journal of Medical and Biomedical Sciences*, 3(3), 9–15.
- Darajeh, N., Idris, A., Reza, H., Masoumi, F., Nourani, A., Rezanian, S., and Teknologi, U. (2017). Phytoremediation of palm oil mill secondary effluent (POMSE) by *Chrysopogon zizanioides* (L.) using artificial neural networks. *International Journal of Phytoremediation*, 19(5), 413–424.
- De Broyer, C., Koubbi, P., Griffiths, H. J., Raymond, B., Udekem d’Acoz, C. D., Van de Putte, A. P., Danis, B., David, B., Grant, S., Gutt, J., Held, C., Hosie, G., Huettmann, F., Post, A., and Ropert-Coudert, Y. (2016). Bio-geographic atlas of the southern ocean. Scientific committee on Antarctic research, Cambridge. *Scientia Marina*, 80(1), 135-136
- de Jesus, H. E., Peixoto, R. S., and Rosado, A. S. (2015). Bioremediation in Antarctic soils. *Journal of Petroleum & Environmental Biotechnology*, 06(06), 1–11. <https://doi.org/10.4172/2157-7463.1000248>
- Deng, M. C., Li, J., Liang, F. R., Yi, M., Xu, X. M., Yuan, J. P., Peng, J., Wu, C. F., and Wang, J. H. (2014). Isolation and characterization of a novel hydrocarbon-degrading bacterium *Achromobacter* sp. HZ01 from the crude oil-contaminated seawater at the Daya Bay, southern China. *Marine Pollution Bulletin*, 83(1), 79–86. <https://doi.org/10.1016/j.marpolbul.2014.04.018>
- Desai, J. D., and Banat, I. M. (1997). Microbial production of surfactants and their commercial potential. *Microbiology and Molecular Biology Reviews*, 61(1), 47–64. [https://doi.org/10.1016/s0140-6701\(97\)84559-6](https://doi.org/10.1016/s0140-6701(97)84559-6)
- Dias, R. L., Ruberto, L., Calabró, A., Balbo, A. Lo, Del Panno, M. T., and Mac Cormack, W. P. (2015). Hydrocarbon removal and bacterial community structure in on-site biostimulated biopile systems designed for bioremediation of diesel-contaminated Antarctic soil. *Polar Biology*, 38(5), 677–687. <https://doi.org/10.1007/s00300-014-1630-7>

- Dobrnjac, S., Dobrnjac, M., Skundric, J. P., Vasiljevic, L., Blagojevic, S., and Zvezdana S. (2019). Possibility for removing products of thermal degradation of edible oil by natural aluminosilicates. *Mitrovic*, 54, 59–71. <https://doi.org/10.1007/978-3-319-99620-2>
- Domínguez, A., Deive, F. J., Angeles Sanromán, M., and Longo, M. A. (2010). Biodegradation and utilization of waste cooking oil by *Yarrowia lipolytica* CECT 1240. *European Journal of Lipid Science and Technology*, 112(11), 1200–1208. <https://doi.org/10.1002/ejlt.201000049>
- Dors, G., Mendes, A. A., Pereira, E. B., de Castro, H. F., and Furigo, A. (2013). Simultaneous enzymatic hydrolysis and anaerobic biodegradation of lipid-rich wastewater from poultry industry. *Applied Water Science*, 3(1), 343–349. <https://doi.org/10.1007/s13201-012-0075-9>
- Edgar, R. C. (2007). PILER-CR: Fast and accurate identification of CRISPR repeats. *BMC Bioinformatics*, 8, 1–6. <https://doi.org/10.1186/1471-2105-8-18>
- Eid, J., Fehr, A., and Gray, J. (2009). Real-time DNA sequencing from single polymerase molecules. *Science*, 323, 133–138.
- El-Deeb, B., and Altalhi, A. D. (2009). Degradative plasmid and heavy metal resistance plasmid naturally coexist in phenol and cyanide assimilating bacteria. *American Journal of Biochemistry and Biotechnology*, 5(2), 84–93.
- El-Masry, M. H., El-Bestawy, E., and Nawal I. El-Adl. (2004). Bioremediation of vegetable oil and grease from polluted wastewater using a sand biofilm system. *World Journal of Microbiology & Biotechnology*, 20, 551–557.
- EMSA. (2009). Addressing Illegal discharges in the marine environment, 1–92.
- Fadile, A., El-Hassani, F. Z., Aissam, H., Merzouki, M., and Benlemlih, M. (2011). Aerobic treatment of lipid-rich wastewater by a bacterial consortium. *African Journal of Microbiology Research*, 5(30), 5333–5342.
- Falcocchio, S., Ruiz, C., Pastor, F. I. J., Saso, L., and Diaz, P. (2005). Identification of a carboxylesterase-producing *Rhodococcus* soil isolate. *Canadian Journal of Microbiology*, 51(9), 753–758. <https://doi.org/10.1139/W05-059>
- Farag, S., Soliman, N. A., and Abdel-Fattah, Y. R. (2018). Statistical optimization of crude oil bio-degradation by a local marine bacterium isolate *Pseudomonas* sp. sp48. *Journal of Genetic Engineering and Biotechnology*, 16(2), 409–420. <https://doi.org/10.1016/j.jgeb.2018.01.001>
- Farrajia, H., Qamaruz Zamana, N., Md Sa'at, S. K., and Fereidonian Dashti, A. (2017). Phytoremediation of suspended solids and turbidity of palm oil mill effluent (POME) by *Ipomea aquatica*. *Engineering Heritage Journal*, 1(1), 36–40. <https://doi.org/10.26480/gwk.01.2017.36.40>
- Federal, R. (1997). 40 CFR Part 112. oil pollution. prevention; non-transportation related onshore facilities. *Rules and Regulations*, 62(202), 54508–54543.
- Feng, L., Wang, W., Cheng, J., Ren, Y., Zhao, G., Gao, C., Tang, Y., Liu, X., Han, W., Peng, X., Liu, R., and Wang, L. (2007). Genome and proteome of long-chain alkane degrading *Geobacillus thermodenitrificans* NG80-2 isolated from a deep-subsurface oil reservoir. *Proceedings of the National Academy of Sciences of the United States of America*, 104(13), 5602–5607. <https://doi.org/10.1073/pnas.0609650104>
- Fingas, M. F. (2012). Vegetable oil spills: oil properties and behaviour. In proceedings of the 35th AMOP technical seminar on environmental contamination and response pp. 884–902. Retrieved from <http://www.scopus.com/inward/record.url?eid=2-s2.0-84865959215&partnerID=tZOtx3y1>

- Fitzpatrick, T. (2003). Vegetable oil spills hurt environment, too. <https://source.wustl.edu/2003/07/vegetable-oil-spi>.
- Focardi, S., Gaggi, C., Chemello, G., and Bacci, E. (1991). Organochlorine residues in Moss and Lichen samples from two Antarctic areas. *Polar Record*, 27(162), 241–244.
- Fox, N. J., and Stachowiak, G. W. (2007). Vegetable oil-based lubricants-A review of oxidation. *Tribology International*, 40(7), 1035–1046.
- Galaction, A. I., Cașcaval, D., Rotaru, R., Lupasteanu, A. M., and Turnea, M. (2012). Kinetic studies on biodegradation of lipids from olive oil mill wastewaters with free and immobilized *Bacillus* sp. cells. *Scientific Study and Research: Chemistry and Chemical Engineering, Biotechnology, Food Industry*, 13(1), 49–62.
- Gerdes, B., Brinkmeyer, R., Dieckmann, G., and Helmke, E. (2005). Influence of crude oil on changes of bacterial communities in Arctic sea-ice. *FEMS Microbiology Ecology*, 53(1), 129–139. <https://doi.org/10.1016/j.femsec.2004.11.010>
- Gesheva, V., Stackebrandt, E., and Vasileva-Tonkova, E. (2010). Biosurfactant production by halotolerant *Rhodococcus fascians* from Casey station, Wilkes Land, Antarctica. *Current Microbiology*, 61(2), 112–117. <https://doi.org/10.1007/s00284-010-9584-7>
- Ghosh, M., and Singh, S. P. (2005). A review on phytoremediation of heavy metals and utilization of it's by products. *Asian Journal on Energy and Environment*, 6(04), 214–231. <https://doi.org/10.1007/s10681-014-1088-2>
- Goepfert, S., and Poirier, Y. (2007). β -Oxidation in fatty acid degradation and beyond. *Current Opinion in Plant Biology*, 10(3), 245–251.
- Gompertz, B. (1825). On the nature of the function expressive of the law of human mortality, and on a new mode of determining the value of life contingencies. *Philosophical Transactions of the Royal Society of London*, 115, 513 – 585.
- Gorman, R., and Adley, C. C. (2004). An evaluation of five preservation techniques and conventional freezing temperatures of -20°C and -85°C for long-term preservation of *Campylobacter jejuni*. *Letters in Applied Microbiology*, 38(4), 306–310. <https://doi.org/10.1111/j.1472-765X.2004.01490.x>
- Gorret, N., Maubois, J. L., Engasser, J. M., and Ghoul, M. (2001). Study of the effects of temperature, pH and yeast extract on growth and exopolysaccharides production by *Propionibacterium acidipropionici* on milk microfiltrate using a response surface methodology. *Journal of Applied Microbiology*, 90(5), 788–796. <https://doi.org/10.1046/j.1365-2672.2001.01310.x>
- Grabowski, M. R., Coolbaugh, T., Dickins, D. F., Glenn, R., Lee, K., Majors, W., Myers, M. D., Norcross, B. L., Reed, M., Salerno, B., Suydam, R., Tiedje, J. M., Timmermans, M. L., and Wadhams, P. (2014). Responding to oil spills in the US Arctic marine environment. In *The National Academies*. pp. 1–196.
- Gran-Scheuch, A., Fuentes, E., Bravo, D. M., Jiménez, J. C., and Pérez-Donoso, J. M. (2017). Isolation and characterization of phenanthrene degrading bacteria from diesel fuel-contaminated Antarctic soils. *Frontiers in Microbiology*, 8, 1–12. <https://doi.org/10.3389/fmicb.2017.01634>
- Guerra, M. B. B., Neto, E. L., Prianti, M. T. A., Pereira-Filho, E. R., and Schaefer, C. E. G. R. (2013). Post-fire study of the Brazilian scientific Antarctic station: toxic element contamination and potential mobility on the surrounding environment. *Microchemical Journal*, 110, 21–27. <https://doi.org/10.1016/j.microc.2013.01.007>

- Habib, S., Ahmad, S. A., Johari, W. L. W., Shukor, M. Y. A., Alias, S. A., Khalil, K. A., and Yasid, N. A. (2018). Evaluation of conventional and response surface level optimisation of n-dodecane (n-C12) mineralisation by psychrotolerant strains isolated from pristine soil at Southern Victoria island, Antarctica. *Microbial Cell Factories*, 17(1), 1–21. <https://doi.org/10.1186/s12934-018-0889-8>
- Haldane, J. B. S. (1930). *Enzymes*, London, Longmans, Green.
- Halmi, M. I. E., Abdullah, S. R. S., Johari, W. L. W., Ali, M. S. M., Shaharuddin, N. A., Khalid, A., and Shukor, M. Y. (2016). Modelling the kinetics of hexavalent molybdenum (Mo 6+) reduction by the *Serratia* sp. strain MIE2 in batch culture. *Rendiconti Lincei*, 27(4), 653–663. <https://doi.org/10.1007/s12210-016-0545-3>
- Hamitouche, A. E., Bendjama, Z., Amrane, A., Kaouah, F., and Hamane, D. (2012). Relevance of the Luong model to describe the biodegradation of phenol by mixed culture in a batch reactor. *Annals of Microbiology*, 62(2), 581–6.
- Hamzah, A., Sabturani, N., and Radiman, S. (2013). Screening and optimization of biosurfactant production by the hydrocarbon-degrading bacteria. *Sains Malaysiana*, 42(5), 615–623. <https://doi.org/10.1016/j.fuel.2014.12.003>
- Hamzah, M. F., Yusof, N., and Alimon, H. (2016). Microbial assisted phytoremediation of palm oil final discharge (POMFD) wastewater. *Journal of Oil Palm Research*, 28, 320–330.
- Hassanzadeh-Tabrizi, S. A., and Taheri-Nassaj, E. (2011). Modeling and optimization of densification of nanocrystalline Al₂O₃ powder prepared by a sol-gel method using response surface methodology. *Journal of Sol-Gel Science and Technology*, 57(2), 212–220. <https://doi.org/10.1007/s10971-010-2344-0>
- Hazan, R., Que, Y. A., Maura, D., and Rahme, L. G. (2012). A method for high throughput determination of viable bacteria cell counts in 96-well plates. *BMC Microbiology*, 12(1), 1–7. <https://doi.org/10.1186/1471-2180-12-259>
- Hazen, T. C., Prince, R. C., and Mahmoudi, N. (2015). Marine oil biodegradation. *Environmental Science & Technology*, 1–22.
- Helmke, E., and Weyland, H. (2004). Psychrophilic versus psychrotolerant bacteria-occurrence and significance in polar and temperate marine habitats. *Cellular and Molecular Biology*, 50(5), 553–561. <https://doi.org/10.1170/T545>
- Hoffman, D. R., Okon, J. L., and Sandrin, T. R. (2005). Medium composition affects the degree and pattern of cadmium inhibition of naphthalene biodegradation. *Chemosphere*, 59(7), 919–927. <https://doi.org/10.1016/j.chemosphere.2004.11.057>
- Hong, S., Soyol-Erdene, T.-O., Hwang, H. J., Hong, S. B., Hur, S. D., and Motoyama, H. (2012). Evidence of global-scale As, Mo, Sb, and Tl atmospheric pollution in the Antarctic Snow. *Environmental Science and Technology*, 46, 11550–11557.
- Hong, Y. H., Ye, C. C., Zhou, Q. Z., Wu, X. Y., Yuan, J. P., Peng, J., Deng, H., and Wang, J. H. (2017). Genome sequencing reveals the potential of *Achromobacter* sp. HZ01 for bioremediation. *Frontiers in Microbiology*, 8, 1–14. <https://doi.org/10.3389/fmicb.2017.01507>
- Hosoyama, A., Noguchi, M., Numata, M., Tsuchikane, K., Hirakata, S., Uohara, A., Kitahashi, Y., Ohji, S., Ichikawa, N., Kimura, A., Yamazoe, A., and Fujita, N. (2019). *Rhodococcus erythropolis* NBRC 15567, whole genome shotgun sequencing project. NCBI Reference Sequence: NZ_BCRM000000000.1. https://www.ncbi.nlm.nih.gov/nuccore/NZ_BCRM000000.

- Huang, L. (2013). Optimization of a new mathematical model for bacterial growth. *Food Control*, 32(1), 283–288. <https://doi.org/10.1016/j.foodcont.2012.11.019>
- Huerta-Cepas, J., Szklarczyk, D., Forslund, K., Cook, H., Heller, D., Walter, M. C., Rattei, T., Mende, D. R., Sunagawa, S., Kuhn, M., Jensen, L. J., Von Mering, C., and Bork, P. (2016). EGGNOG 4.5: A hierarchical orthology framework with improved functional annotations for eukaryotic, prokaryotic and viral sequences. *Nucleic Acids Research*, 44, 286–293. <https://doi.org/10.1093/nar/gkv1248>
- Hughes, K. A., López-Martínez, J., Francis, J. E., Crame, J. A., Carcavilla, L., Shiraishi, K., Hokada, T., and Yamaguchi, A. (2016). Antarctic geoconservation: A review of current systems and practices. *Environmental Conservation*, 43(2), 97–108. <https://doi.org/10.1017/S0376892915000387>
- Hughes, K. A., and Pertierra, L. R. (2016). Evaluation of non-native species policy development and implementation within the Antarctic treaty area. *Biological Conservation*, 200, 149–159. <https://doi.org/10.1016/j.biocon.2016.03.011>
- Hughes, K. A., and Stallwood, B. (2005). Oil pollution in the antarctic terrestrial environment. *Polarforschung*, 75(2–3), 141–144.
- Hyatt, D., Chen, G. L., LoCascio, P. F., Land, M. L., Larimer, F. W., and Hauser, L. J. (2010). Prodigal: Prokaryotic gene recognition and translation initiation site identification. *BMC Bioinformatics*, 11, 1–11. <https://doi.org/10.1186/1471-2105-11-119>
- IAATO. Tourism Statistics. Available at: https://iaato.org/en_GB/tourism-statistics (2017).
- Ibrahim, H. M. M. (2016). Biodegradation of used engine oil by novel strains of *Ochrobactrum anthropi* HM-1 and *Citrobacter freundii* HM-2 isolated from oil-contaminated soil. *3 Biotech*, 6(2), 1–13. <https://doi.org/10.1007/s13205-016-0540-5>
- Ibrahim, M. L., Ijah, U. J. J., Manga, S. B., Bilbis, L. S., and Umar, S. (2013). Production and partial characterization of biosurfactant produced by crude oil degrading bacteria. *International Biodeterioration and Biodegradation*, 81, 28–34. <https://doi.org/10.1016/j.ibiod.2012.11.012>
- Ibrahim, S., Shukor, M. Y., Syed, M. A., Johari, W. L. W., Shamaan, N. A., Sabullah, M. K., and Ahmad, S. A. (2016a). Enhanced caffeine degradation by immobilised cells of *Leifsonia* sp. strain SIU. *Journal of General and Applied Microbiology*, 62, 18–24. <https://doi.org/10.2323/jgam.62.18>
- Ibrahim, S., Muhammad, A., Tanko, A. S., Abubakar, A., Ibrahim, H., Shukor, M. Y., and Ahmad, S. A. (2016b). Studies of action of heavy metals on caffeine degradation by immobilised *Leifsonia* sp. strain SIU. *Bayero Journal of Pure and Applied Sciences*, 8(2), 138–144. <https://doi.org/10.4314/bajopas.v8i2.24>
- Ibrahim, S., Shukor, M. Y., Khalil, K. A., Helmi, M. I. E., Syed, M. A., and Ahmad, S. A. (2015a). Application of response surface methodology for optimising caffeine-degrading parameters by *Leifsonia* sp. strain SIU. *Journal of Environmental Biology*, 36(5), 1215–1221.
- Ibrahim, S., Shukor, M. Y., Syed, M. A., Wan Johari, W. L., and Ahmad, S. A. (2015b). Characterisation and growth kinetics studies of caffeine-degrading bacterium *Leifsonia* sp. strain SIU. *Annals of Microbiology*, 66(1), 289–298. <https://doi.org/10.1007/s13213-015-1108-z>
- Ibrahim, S., Shukor, M. Y., Syed, M. A., Ab Rahman, N. A., Khalil, K. A., Khalid, A., and Ahmad, S. A. (2014). Bacterial degradation of caffeine: A review. *Asian Journal of Plant Biology*, 2(1), 24–33.

- Ilory, M., Adebusoye, E., and Ojo, A. (2008). Isolation and characterization of hydrocarbon-degrading and biosurfactant-producing yeast strains obtained from a polluted lagoon water. *World Journal of Microbiology and Biotechnology*, *24*, 2539–2545.
- Ismail, Z., Makmor, M., Hashim, R., Shah, R. M., Hanifah, N. A., and Ahmad, S. (2012). The role of Malaysia under the Antarctic treaty and Madrid protocol. *CHUSER 2012 - 2012 IEEE Colloquium on Humanities, Science and Engineering Research*, 42–47. <https://doi.org/10.1109/CHUSER.2012.6504278>
- Ito, K., Harada, M., Nakajima, N., Yamamura, S., Tomita, M., Suzuki, H., and Amachi, S. (2018). Genomic analysis of *Rhodococcus* sp. Br-6, a bromate reducing bacterium isolated from soil in chiba, Japan. *Journal of Genomics*, *6*, 122–126. <https://doi.org/10.7150/jgen.27741>
- Jayakumar, M. M. (2013). When and how to use Plackett-Burman experimental design pp. 1–8. Retrieved from <http://www.isixsigma.com/tools-templates/design-of-experiments-doe/w...>
- Kaakinen, J., Vähöja, P., Kuokkanen, T., and Roppola, K. (2007). Studies on the effects of certain soil properties on the biodegradation of oils determined by the manometric respirometric method. *Journal of Automated Methods and Management in Chemistry*, 1–7. <https://doi.org/10.1155/2007/34601>
- Kaczorek, E., Chrzanowski, L., Pijanowska, A., and Olszanoswski, A. (2008). Yeast and bacteria cell hydrophobicity and hydrocarbon biodegradation in the presence of natural surfactants: rhamnolipids and saponins. *Bioresource Technology*, *99*, 4285–4291.
- Kademi, A., Ait-Abdelkader, N., Fakhreddine, L., and Baratti, J. C. (1999). A thermostable esterase activity from newly isolated moderate thermophilic bacterial strains. *Enzyme and Microbial Technology*, *24*, 332–338.
- Kamble, P. P., Raut, A. A., and Gandhi, M. B. (2011). Isolation and characterization of vegetable oil degrading bacteria. *Journal of Pure and Applied Microbiology*, *5*(2), 887–890.
- Kanchanasuta, S., and Pisutpaisal, N. (2016). Waste utilization of palm oil decanter cake on biogas fermentation. *International Journal of Hydrogen Energy*, *41*(35), 15661–15666. <https://doi.org/10.1016/j.ijhydene.2016.04.129>
- Kanehisa, M., Goto, S., Sato, Y., Kawashima, M., Furumichi, M., and Tanabe, M. (2014). Data, information, knowledge and principle: Back to metabolism in KEGG. *Nucleic Acids Research*, *42*, 199–205. <https://doi.org/10.1093/nar/gkt1076>
- Kao, E. (2017). South China morning post, Hong Kong government struggles to clean up mess as palm oil spill spreads. *South China Morning Post*.
- Karamba, K. I., Ahmad, S. A., Zulkharnain, A., Syed, M. A., Khalil, K. A., Shamaan, N. A., Dahalan, F. A., and Shukor, M. Y. (2016). Optimisation of biodegradation conditions for cyanide removal by *Serratia marcescens* strain AQ07 using one-factor-at-a-time technique and response surface methodology. *Rendiconti Lincei*, *27*(3), 533–545. <https://doi.org/10.1007/s12210-016-0516-8>
- Karamba, K. I., Aqlima, S. A., Zulkharnain, A., Yasid, N. A., Ibrahim, S., and Shukor, M. Y. (2018). Batch growth kinetic studies of locally isolated cyanide - degrading *Serratia marcescens* strain AQ07. *3 Biotech*, *8*(1), 1–9.
- Karlapudi, A. P., Venkateswarulu, T. C., Tammineedi, J., Kanumuri, L., Ravuru, B. K., Dirisala, V. ramu, and Kodali, V. P. (2018). Role of biosurfactants in bioremediation of oil pollution-a review. *Petroleum*, *4*(3), 241–249.
- Kaviyani, M., Ghazi, N. M., Shariati, M. A., and Atarod, S. (2014). The study of

- frying oils properties. *International Journal of Advanced Engineering Research and Technology (IJAERT)*, 2, 90–96.
- Khalida, K., Naeem, M., Arshed, M. J., and Asif, M. (2006). Extraction and characterization of oil degrading bacteria. *Journal of Applied Sciences*, 10(6), 2302–2306.
- Kis, Á., Laczi, K., Zsíros, S., Rákhely, G., and Perei, K. (2015). Biodegradation of animal fats and vegetable oils by *Rhodococcus erythropolis* PR4. *International Biodeterioration and Biodegradation*, 105, 114–119.
- Klokočník, J., Kostelecký, J., and Bezděk, A. (2018). On the detection of the Wilkes Land impact crater. *Earth, Planets and Space*, 70(1), 1–12.
- Kodali, D. R. (2002). “High performance ester lubricants from natural oils”, industrial lubrication and tribology. *Industrial Lubrication and Tribology*, 54(4), 165–170. <https://doi.org/10.1108/00368790210431718>
- Kosaric, N. (2001). Biosurfactants and their application for soil bioremediation. *Food Technology and Biotechnology*, 39(4), 295–304.
- Koutsoumanis, K. P., and Sofos, J. N. (2005). Effect of inoculum size on the combined temperature, pH and aw limits for growth of *Listeria monocytogenes*. *International Journal of Food Microbiology*, 104(1), 83–91.
- Kumar, S., Mathur, A., Singh, V., Nandy, S., Khare, S. K., and Negi, S. (2012). Bioremediation of waste cooking oil using a novel lipase produced by *Penicillium chrysogenum* SNP5 grown in solid medium containing waste grease. *Bioresource Technology*, 120, 300–304. <https://doi.org/10.1016/j.biortech.2012.06.018>
- Kumari, B., Singh, S. N., and Singh, D. P. (2012). Characterization of two biosurfactant producing strains in crude oil degradation. *Process Biochemistry*, 47(12), 2463–2471. <https://doi.org/10.1016/j.procbio.2012.10.010>
- Kurtzman, C. P., Price, N. P. J., Ray, K. J., and Kuo, T. M. (2010). Production of sophorolipid biosurfactants by multiple species of the *Starmerella* (*Candida*) *bombicola* yeast clade. *FEMS Microbiology Letters*, 311(2), 140–146. <https://doi.org/10.1111/j.1574-6968.2010.02082.x>
- Kwapisz, E., Wszelaka, J., Marchut, O., and Bielecki, S. (2008). The effect of nitrate and ammonium ions on kinetics of diesel oil degradation by *Gordonia alkanivorans* S7. *International Biodeterioration and Biodegradation*, 61, 214–222.
- Lai, C. C. C., Huang, Y.-C. Y. C., Wei, Y. H. Y. H., and Chang, J. S. J. S. (2009). Biosurfactant-enhanced removal of total petroleum hydrocarbons from contaminated soil. *Journal of Hazardous Materials*, 167(1–3), 609–614. <https://doi.org/10.1016/j.jhazmat.2009.01.017>
- Lamilla, C., Braga, D., Castro, R., Guimarães, C., de Castilho, L. V. A., Freire, D. M. G., and Barrientos, L. (2018). *Streptomyces luridus* So3.2 from Antarctic soil as a novel producer of compounds with bioemulsification potential. *PLoS ONE*, 13(4), 1–20. <https://doi.org/10.1371/journal.pone.0196054>
- Lana, N. B., Berton, P., Covaci, A., Ciocco, N. F., Barrera-Oro, E., Atencio, A., and Altamirano, J. C. (2014). Fingerprint of persistent organic pollutants in tissues of Antarctic notothenioid fish. *Science of the Total Environment*, 499, 89–98. <https://doi.org/10.1016/j.scitotenv.2014.08.033>
- Larkin, M. J., Kulakov, L. A., and Allen, C. C. R. (2005). Biodegradation and *Rhodococcus* - Masters of catabolic versatility. *Current Opinion in Biotechnology*, 16(3), 282–290. <https://doi.org/10.1016/j.copbio.2005.04.007>
- Larkin, M. J., Kulakov, L. A., and Allen, C. C. R. (2006). Biodegradation by

- members of the genus *Rhodococcus*: Biochemistry, physiology, and genetic adaptation. *Advances in Applied Microbiology*, 59(06), 1–29. [https://doi.org/10.1016/S0065-2164\(06\)59001-X](https://doi.org/10.1016/S0065-2164(06)59001-X)
- Lee, G. L. Y., Ahmad, S. A., Yasid, N. A., Zulkharnain, A., Convey, P., Wan Johari, W. L., Alias, S. A., Gonzalez-Rocha, G., and Shukor, M. Y. (2018). Biodegradation of phenol by cold-adapted bacteria from Antarctic soils. *Polar Biology*, 41(3), 553–562. <https://doi.org/10.1007/s00300-017-2216-y>
- Lee, J.-Y., Lim, H. S., and Yoon, H. Il. (2016). Thermal characteristics of soil and water during summer at King Sejong station, King George Island, Antarctica. *Geosciences Journal*, 20(4), 503–516. <https://doi.org/10.1007/s12303-016-0026-9>
- Lefebvre, X., Paul, E., Mauret, M., Baptiste, P., and Capdeville, B. (1998). Kinetic characterization of saponified domestic lipid residues aerobic biodegradation. *Water Research*, 32, 3031–3038.
- Lemire, J. A., Harrison, J. J., and Turner, R. J. (2013). Antimicrobial activity of metals: Mechanisms, molecular targets and applications. *Nature Reviews Microbiology*, 11(6), 371–384. <https://doi.org/10.1038/nrmicro3028>
- Leong, X.-F., Ng, C.-Y., Jaarin, K., and Mustafa, M. (2015). Effects of repeated heating of cooking oils on antioxidant content and endothelial function. *Austin Journal of Pharmacology and Therapeutics*, 3(2), 1068.
- Letek, M., González, P., MacArthur, I., Rodríguez, H., Freeman, T. C., Valero-Rello, A., Blanco, M., Buckley, T., Cherevach, I., Fahey, R., Hapeshi, A., Holdstock, J., Leadon, D., Navas, J., Ocampo, A., Quail, M. A., Sanders, M., Scotti, M. M., Prescott, J. F., Fogarty, U., Meijer, W. G., Parkhill, J., Bentley, S. D., and Vázquez-Boland, J. A. (2010). The genome of a pathogenic *Rhodococcus*: Cooptive virulence underpinned by key gene acquisitions. *PLoS Genetics*, 6(9). <https://doi.org/10.1371/journal.pgen.1001145>
- Li, M., Liao, X., Zhang, D., Du, G., and Chen, J. (2011). Yeast extract promotes cell growth and induces production of polyvinyl alcohol-degrading enzymes. *Enzyme Research*, 2011(1), 1–8. <https://doi.org/10.4061/2011/179819>
- Li, Y. (2007). Principles and technology of fermentation engineering, Higher Education Press, Beijing, China.
- Li, Ying, Cui, T., Wang, Y., and Ge, X. (2018). Isolation and characterization of a novel bacterium: *Pseudomonas aeruginosa* for biofertilizer production from kitchen waste oil. *RSC Advances*, 8(73), 41966–41975. <https://doi.org/10.1039/c8ra09779h>
- Li, Z., and Wrenn, B. A. (2004). Effects of ferric hydroxide on the anaerobic biodegradation kinetics and toxicity of vegetable oil in freshwater sediments. *Water Research*, 38, 3859–3868.
- Li, Z., Wrenn, B. A., and Venosa, A. D. (2005). Anaerobic biodegradation of vegetable oil and its metabolic intermediates in oil-enriched freshwater sediments. *Biodegradation*, 16(4), 341–352. <https://doi.org/10.1007/s10532-004-2057-6>
- Lim, H. S., Han, M. J., C.Seo, D., Kim, J. H., Lee, J. I., Park, H., Hur, J. S., Cheong, Y. H., Heo, J. S., Yoon, H. I., and Cho, J. S. (2009). Heavy metal concentrations in the Fruticose Lichen *Usnea aurantiacoatra* from King George Island, South Shetland islands, west Antarctica. *Journal of the Korean Society for Applied Biological Chemistry*, 52(5), 503–508.
- Lima, C. J. B. De, Coelho, L. F., and Contiero, J. (2010). The use of response surface methodology in optimization of lactic acid production: focus on medium

- supplementation, temperature and pH control. *Food Technology and Biotechnology*, 48(2), 175–181.
- Lima e Silva, A. A., Ribeiro de Carvalho, M. A., Souza, S. A. L., Dias, P. M. T., Filho, R. G. da S., Saramago, C. S. de M., Bento, C. A. de M., and Hofer, E. (2012). Heavy metals tolerance (Cr, Ag and Hg) in bacteria isolated from sewage. *Brazilian Journal of Microbiology*, 1620–1631.
- Litova, K., Gerginova, M., Peneva, N., Manasiev, J., and Aelxieva, Z. (2016). Growth of Antarctic fungal strains on phenol at low temperatures. *Journal of BioScience and Biotechnology*, 43–46.
- Loganes, C., Ballali, S., and Minto, C. (2016). Main properties of canola oil components: A descriptive review of current knowledge. *The Open Agriculture Journal*, 10, 69–74. <https://doi.org/10.2174/1874331501610010069>
- López, S., Prieto, M., Dijkstra, J., Dhanoa, M. S., and France, J. (2004). Statistical evaluation of mathematical models for microbial growth. *International Journal of Food Microbiology*, 96(3), 289–300.
- Luong, J. H. T. (1987). Generalization of Monod kinetics for analysis of growth data with substrate inhibition. *Biotechnology and Bioengineering*, 29(2), 242–248.
- Ma, X., Zhang, H., Zhou, H., Na, G., Wang, Z., Chen, C., Chen, J., and Chen, J. (2014). Occurrence and gas/particle partitioning of short- and medium-chain chlorinated paraffins in the atmosphere of Fildes Peninsula of Antarctica. *Atmospheric Environment*, 90, 10–15. <https://doi.org/10.1016/j.atmosenv.2014>.
- Magi, E., and Tanwar, S. (2014). “Extreme mass spectrometry”: the role of mass spectrometry in the study of the Antarctic environment. *Journal of Mass Spectrometry*, 49(11), 1071–1085.
- Maharana, A. K., and Singh, S. M. (2018). Cold active lipases produced by *Cryptococcus* sp . Y-32 and *Rhodococcus erythropolis* N149 isolated from Nella Lake, Antarctica. *International Journal of Current Microbiology and Applied Sciences*, 7(03), 1910–1926.
- Majumder, A., and Jha, S. (2012). Hairy roots: A promising tool for phytoremediation. In *Microorganisms in Environmental Management: Microbes and Environment* (pp. 607–629). <https://doi.org/10.1007/978-94-007-2229-3>
- Malavenda, R., Rizzo, C., Michaud, L., Gerçe, B., Bruni, V., Sylđatk, C., Hausmann, R., and Lo Giudice, A. (2015). Biosurfactant production by Arctic and Antarctic bacteria growing on hydrocarbons. *Polar Biology*, 38(10), 1565–1574. <https://doi.org/10.1007/s00300-015-1717-9>
- Manan, F. A., Ibrahim, Z., and Shahir, S. (2014). Plants in antarctica: Current and future phytoremediation potential. *Jurnal Teknologi (Sciences and Engineering)*, 69(1), 59–65. <https://doi.org/10.11113/jt.v69.2413>
- Manogaran, M., Shukor, M. Y., Yasid, N. A., Khalil, K. A., and Ahmad, S. A. (2018). Optimisation of culture composition for glyphosate degradation by *Burkholderia vietnamiensis* strain AQ5-12. *3 Biotech*, 8(2), 1–13. <https://doi.org/10.1007/s13205-018-1123-4>
- Mansee, A. H., Montasser, M. R., and Shanab, A. S. A. (2004). Decontamination of pollutants in aquatic system: 1. biodegradation efficiency of isolated bacteria strains from certain contaminated areas. *Pakistan Journal of Biological Science*, 7(7), 1202–1207.
- Mar, C. C., Fan, Y., Li, F., and Hu, G. (2016). Bioremediation of wastewater from edible oil refinery factory using *Oleaginous Microalga*. *International Journal of Phytoremediation*, 1–28. <https://doi.org/10.1080/15226514.2016.1193466>
- Margesin, R. (2000). Potential of cold-adapted microorganisms for bioremediation of

- oil-polluted Alpine soils. *International Biodeterioration and Biodegradation*, 46(1), 3–10. [https://doi.org/10.1016/S0964-8305\(00\)00049-4](https://doi.org/10.1016/S0964-8305(00)00049-4)
- Margesin, R., and Schinner, F. (2001). Bioremediation (natural attenuation and biostimulation) of diesel-oil-contaminated soil in an alpine glacier skiing area. *Applied and Environmental Microbiology*, 67, 3127–3133.
- Maritime C. (2011). Palm oil spill in sea as barge sinks near Nagapattinam port. News Maritime connector. <http://maritime-connector.com/news/environment-and>.
- Martínková, L., Uhnáková, B., Pátek, M., Nesvera, J., Kren, V., Martínková, L., Uhnáková, B., Pátek, M., Ne, J., and Vladimír, K. (2009). Biodegradation potential of the genus *Rhodococcus*. *Environmental International*, 35(1), 162–177. <https://doi.org/10.1016/j.envint.2008.07.018>
- Martins, C. C., Aguiar, S. N., Bicego, M. C., and Montone, R. C. (2012). Sewage organic markers in surface sediments around the Brazilian Antarctic station: results from the 2009/10 austral summer and historical tendencies. *Marine Pollution Bulletin*, 64, 2867–2870.
- Matheson, B., Walker, K. Z., Taylor, D. M., Peterkin, R., Lugg, D., and Kerin O’Dea. (1996). Effect on serum lipids of monounsaturated oil and margarine in the diet of an Antarctic expedition. *The American Journal of Clinical Nutrition*, 63(6), 979–980. <https://doi.org/10.1093/ajcn/64.6.979>
- Matsumiya, Y., Wakita, D., Kimura, A., Sanpa, S., and Kubo, M. (2007). Isolation and characterization of a lipid-degrading bacterium and its application to lipid-containing wastewater treatment. *Journal of Bioscience and Bioengineering*, 103(4), 325–330. <https://doi.org/10.1263/jbb.103.325>
- McKelvey, R. W., Robertson, I., and Whitehead, P. E. (1980). Effect of non-petroleum spills on wintering birds near Vancouver. *Marine Pollution Bulletin*, 11(6), 169–171.
- McLeod, M. P., Warren, R. L., Hsiao, W. W. L., Araki, N., Myhre, M., Fernandes, C., Miyazawa, D., Wong, W., Lillquist, A. L., Wang, D., Dosanjh, M., Hara, H., Petrescu, A., Morin, R. D., Yang, G., Stott, J. M., Schein, J. E., Shin, H., Smailus, D., Siddiqui, A. S., Marra, M. A., Jones, S. J. M., Holt, R., Brinkman, F. S. L., Miyauchi, K., Fukuda, M., Davies, J. E., Mohn, W. W., and Eltis, L. D. (2006). The complete genome of *Rhodococcus* sp. RHA1 provides insights into a catabolic powerhouse. *Proceedings of the National Academy of Sciences*, 103(42), 15582–15587. <https://doi.org/10.1073/pnas.0607048103>
- Mercurio, P., Burns, K. A., and Negri, A. (2004). Testing the ecotoxicology of vegetable versus mineral based lubricating oils: 1. Degradation rates using tropical marine microbes. *Environmental Pollution*, 129(2), 165–173.
- Mobarak-Qamsari, E., Kasra-Kermanshahi, R., and Moosavi-Nejad, Z. (2011). Isolation and identification of a novel, lipase-producing bacterium, *Pseudomonas aeruginosa* KM110. *Iranian Journal of Microbiology*, 3(2), 92–98.
- Mongkolthararuka, W., and Dharmstithi, S. (2002). Biodegradation of lipid-rich wastewater by a mixed bacterial consortium. *International Biodeterioration & Biodegradation*, 50, 101–105. [https://doi.org/10.1016/S0964-8305\(02\)00057-4](https://doi.org/10.1016/S0964-8305(02)00057-4)
- Monod, J. (1949). The growth of bacterial cultures. *Annual Reviews in Microbiology*, 3, 371–394.
- Morais, I. M. C., Cordeiro, A. L., Teixeira, G. S., Domingues, V. S., Nardi, R. M. D., Monteiro, A. S., Alves, R. J., Siqueira, E. P., and Santos, V. L. (2017). Biological and physicochemical properties of biosurfactants produced by

- Lactobacillus jensenii* P6A and *Lactobacillus gasserii* P65. *Microbial Cell Factories*, 16(1), 1–15. <https://doi.org/10.1186/s12934-017-0769-7>
- Morikawa, M., Hirata, Y., and Imanaka, T. (2000). A study on the structure-function relationship of lipopeptide biosurfactants. *Biochimica et Biophysica Acta*, 1488, 211–218.
- Mudge, F. B. (1997). The development of the ‘greenhouse’ theory of global climate change from victorian times. *Weather*, 52(1), 13–17. <https://doi.org/10.1002/j.1477-8696.1997.tb06243.x>
- Mudge, G. H., Goldstein, S., Addonizio, L. J., Caplan, A., Mancini, D., Levine, T. B., Ritsch, M. E. J., and Stevenson, L. W. (1993). 24th Bethesda conference: Cardiac transplantation. Task Force 3: Recipient guidelines/prioritization. *Journal of the American College of Cardiology (JACC)*, 22(1), 21–31.
- Mudge, S. M. (1995). Deleterious effects from accidental spillages of vegetable oils. *Spill Science and Technology Bulletin*, 2(2–3), 187–191. [https://doi.org/10.1016/S1353-2561\(96\)00019-9](https://doi.org/10.1016/S1353-2561(96)00019-9)
- Mueller, B., Rock, S., Gowswami, D., and Ensley, D. (1999). Phytoremediation decision tree – prepared by - interstate technology and regulatory cooperation work group; (pp. 1–36).
- Mulchandani, A., Luong, J. H. T., and Groom, C. (1989). Substrate inhibition kinetics for microbial growth and synthesis of poly- β -hydroxybutyric acid by *Alcaligenes eutrophus* ATCC 17697. *Applied Microbiology and Biotechnology*, 30(1), 11–17. <https://doi.org/10.1007/BF00255990>
- Mulligan, C. N. (2005). Environmental applications for biosurfactants. *Environmental Pollution*, 133(2), 183–198. <https://doi.org/10.1016/j.envpol.2004.06.009>
- Muntari, B., Amid, A., Mel, M., Jami, M. S., and Salleh, H. M. (2012). Recombinant bromelain production in *Escherichia coli*: process optimization in shake flask culture by response surface methodology. *Springer Open Journal*, 1–9.
- Nagarajan, J., Nawawi, N. M., and Ibrahim, A. L. (2014). *Rhodococcus* UKMP-5M, an endogenous lipase producing actinomycete from Peninsular Malaysia. *Biologia (Poland)*, 69(2), 123–132. <https://doi.org/10.2478/s11756-013-0308-x>
- Nahar, A., Baker, A. L., Charleston, M. A., Bowman, J. P., and Britz, M. L. (2017). Draft genome sequences of three sub-Antarctic *Rhodococcus* spp., including two novel psychrophilic genomospecies. *Genome Announcements*, 5(36), 4–5.
- Nawaz, M. S., Billedeau, S. M., and Cerniglia, C. E. (1998). Influence of selected physical parameters on the biodegradation of acrylamide by immobilized cells of *Rhodococcus* sp. *Biodegradation*, 9(5), 381–387.
- Nawrocki, E. P., and Eddy, S. R. (2013). Infernal 1.1: 100-fold faster RNA homology searches. *Bioinformatics*, 29(22), 2933–2935.
- Negri, A., Burns, K., Boyle, S., Brinkman, D., and Webster, N. (2006). Contamination in sediments, bivalves and sponges of McMurdo sound, Antarctica. *Environmental Pollution*, 143(3), 456–467. <https://doi.org/10.1016/j.envpol.2005.12.005>
- Ng, Y. S., and Chan, D. J. C. (2017). Wastewater phytoremediation by *Salvinia molesta*. *Journal of Water Process Engineering*, 15, 107–115.
- Nickzad, A., Mogharei, A., Monazzami, A., Jamshidian, H., and Vahabzadeh, F. (2012). Biodegradation of phenol by *Ralstonia eutropha* in a Kissiris-immobilized cell bioreactor. *Water Environmental Research*, 84(8), 626–34.
- Nies, D. H. (1999). Microbial heavy-metal resistance. *Applied Microbiology and Biotechnology*, 51(6), 730–750.

- Nkeiruka, N. C., and Tagbo, N. J. (2014). Continuous process design model simulation for the anaerobic digestion of vegetable oil wastewater. *American Journal of Environmental Protection*, 3(5), 209–216.
- Nwaguma, I. V., Chikere, C. B., and Okpokwasili, G. C. (2016). Isolation, characterization, and application of biosurfactant by *Klebsiella pneumoniae* strain IVN51 isolated from hydrocarbon-polluted soil in ogoniland, Nigeria. *Bioresources and Bioprocessing*, 3(1), 40–53. <https://doi.org/10.1186/s40643-016-0118-4>
- Nwaichi, E. O., Frac, M., Nwoha, P. A., and Eragbor, P. (2015). Enhanced phytoremediation of crude oil-polluted soil by four plant species: effect of inorganic and organic bioaugmentation. *International Journal of Phytoremediation*, 17(12), 1253–1261. <https://doi.org/10.1080/15226514.2015.1058324>
- Nweke, C., Igbokwe, P., and Nwabanne, J. (2014). Kinetics of batch anaerobic digestion of vegetable oil wastewater. *Open Journal of Water Pollution and Treatment*, 2014(2), 1–10. <https://doi.org/10.15764/WPT.2014.02001>
- Nzila, A., Thukair, A., Sankara, S., and Abdur Razzak, S. (2017). Characterization of aerobic oil and grease-degrading bacteria in wastewater. *Environmental Technology*, 38(6), 661–670. <https://doi.org/10.1080/09593330.2016.1207712>
- Obayori, O. S., Ilori, M. O., Adebussaye, S. A., Oyetibo, G. O., Omoayo, A. E., and Amund, O. O. (2009). Degradation of hydrocarbons and biosurfactant production by *Pseudomonas* sp. strain LP1. *World Journal of Microbiology and Biotechnology*, 25, 1615–1623.
- Okonko, I. O., and Shittu, O. B. (2007). Bioremediation of wastewater and municipal water treatment using latex exudate from *Calotropis procera* (sodom apple). *Electronic Journal of Environmental, Agricultural and Food Chemistry*, 6(3), 1890–1904.
- Orro, A., Cappelletti, M., D’Ursi, P., Milanese, L., Di Canito, A., Zampolli, J., Collina, E., Decorosi, F., Viti, C., Fedi, S., Presentato, A., Zannoni, D., and Di Gennaro, P. (2015). Genome and phenotype microarray analyses of *Rhodococcus* sp. BCP1 and *Rhodococcus opacus* R7: Genetic determinants and metabolic abilities with environmental relevance. *PLoS ONE*, 10(10), 1–41.
- Othman, A. R., Bakar, N. A., Halmi, M. I. E., Johari, W. L. W., Ahmad, S. A., Jirangon, H., Syed, M. A., and Shukor, M. Y. (2013). Kinetics of molybdenum reduction to molybdenum blue by *Bacillus* sp. strain A.rzi. *BioMed Research International*, 2013, 1–9. <https://doi.org/10.1155/2013/371058>
- Overbeek, R., Begley, T., Butler, R. M., Choudhuri, J. V., Chuang, H. Y., Cohoon, M., de Crécy-Lagard, V., Diaz, N., Disz, T., Edwards, R., Fonstein, M., Frank, E. D., Gerdes, S., Glass, E. M., Goesmann, A., Hanson, A., Iwata-Reuyl, D., Jensen, R., Jamshidi, N., Krause, L., Kubal, M., Larsen, N., Linke, B., McHardy, A. C., Meyer, F., Neuweger, H., Olsen, G., Olson, R., Osterman, A., Portnoy, V., Pusch, G. D., Rodionov, D. A., Rülckert, C., Steiner, J., Stevens, R., Thiele, I., Vassieva, O., Ye, Y., Zagnitko, O., and Vonstein, V. (2005). The subsystems approach to genome annotation and its use in the project to annotate 1000 genomes. *Nucleic Acids Research*, 33(17), 5691–5702. <https://doi.org/10.1093/nar/gki866>
- Papanikolaou, S., and Aggelis, G. (2003). Selective uptake of fatty acids by the yeast *Yarrowia lipolytica*. *European Journal of Lipid Science and Technology*, 105, 651–655.
- Parhi, P., Jadhav, V. V., and Bhadekar, R. (2016). Increase in production of

- biosurfactant from *Oceanobacillus* sp. BRI 10 using low cost substrates. *Songklanakarinn Journal of Science and Technology*, 38(2), 207–211.
- Patowary, K., Patowary, R., Kalita, M. C., and Suresh Deka. (2017). Characterization of biosurfactant produced during degradation of hydrocarbons using crude oil as sole source of carbon. *Frontiers in Microbiology*, 8, 2–14.
- Peck, L. S., Convey, P., and Barnes, D. K. A. (2006). Environmental constraints on life histories in Antarctic ecosystems: tempos, timings and predictability. *Biology Reviews*, 81, 75–109. <https://doi.org/10.1017/S1464793105006871>
- Pereira, M. G., Mudge, S. M., and Latchford, J. (1998). Bacterial degradation of vegetable oils chemistry and ecology. *Chemistry and Ecology*, 14(3–4), 291–303.
- Pereira, M. G., Mudge, S. M., and Latchford, J. (2003). Vegetable oil spills on salt marsh sediments; comparison between sunflower and linseed oils. *Marine Environmental Research*, 56, 367–385.
- Phong, N. T., Duyan, N. T., and Diep, C. N. (2014). Isolation and characterization of lipid-degrading bacteria in wastewater of food processing plants and restaurants in Can Tho city, Vietnam. *American Journal of Life Sciences*, 2(6), 382–388. <https://doi.org/10.11648/j.ajls.20140206.18>
- Plackett, R. L., and Burman, J. P. (1946). The design of optimum multifactorial experiments. *Biometrika*, 33(4), 305–325.
- Poblet, A., Andrade, S., Scagliola, M., Vodopivec, C., Curtosi, A., Pucci, A., and Marcovecchio, J. (1997). The use of epilithic Antarctic lichens (*Usnea aurantiacoatra* and *U. antarctica*) to determine deposition patterns of heavy metals in the Shetland Islands, Antarctica. *Science of the Total Environment*, 207(2–3), 187–194. [https://doi.org/10.1016/S0048-9697\(97\)00265-9](https://doi.org/10.1016/S0048-9697(97)00265-9)
- Polak-Berecka, M., Waśko, A., Kordowska-Wiater, M., Podleśny, M., Targoński, Z., and Kubik-Komar, A. (2010). Optimization of medium composition for enhancing growth of *Lactobacillus rhamnosus* PEN using response surface methodology. *Polish Journal of Microbiology*, 59(2), 113–118. <https://doi.org/10.33073/pjm-2010-017>
- Pouillot, R., and Lubran, M. B. (2011). Predictive microbiology models vs. modeling microbial growth within *Listeria monocytogenes* risk assessment: What parameters matter and why. *Food Microbiology*, 28(4), 720–726. <https://doi.org/10.1016/j.fm.2010.06.002>
- Prakash, S. (2018). Beta (β)-oxidation of fatty acid and its associated disorders. *International Journal of Clinical Biochemistry*, 5(1), 158–172.
- Prasad, M. P., and Manjunath, K. (2011). Comparative study on biodegradation of lipid-rich wastewater using lipase producing bacterial species. *Indian Journal of Biotechnology*, 10(1), 121–124. <https://doi.org/0972-5849>
- Prus, W., Fabiańska, M. J., and Łabno, R. (2015). Geochemical markers of soil anthropogenic contaminants in polar scientific stations nearby (Antarctica, King George Island). *Science of the Total Environment*, 518–519, 266–279. <https://doi.org/10.1016/j.scitotenv.2015.02.096>
- Pugh, P. J. A., and Convey, P. (2008). Surviving out in the cold: Antarctic endemic invertebrates and their refugia. *Journal of Biogeography*, 35, 2176–86.
- Rack, U. (2015). “Polar Expeditions,” in *Exploring the Last Continent*, eds D. Liggett, B. Storey, Y. Cook, and Meduna V. (Cham: Springer), pp 307–325.
- Rajan, A., Kumar, S., and Nair, A. J. (2011). Isolation of novel alkaline lipase producing fungus *Aspergillus fumigatus* MTCC 9657 from aged and crude rice bran oil and quantification by HPLC. *International Journal of Biological*

- Chemistry*, 5(2), 116–126.
- Rajasekar, A., Babu, T. G., Pandian, S. T., Maruthamuthu, S., Palaniswamy, N., and Rajendran, A. (2007). Role of *Serratia marcescens* ACE2 on diesel degradation and its influence on corrosion. *Indian Journal of Microbiology and Biotechnology*, 34, 589–598.
- Rajendran, P., Muthukrishnan, J., and Gunasekaran, P. (2003). Microbes in heavy metal remediation. *Indian Journal of Experimental Biology*, 41, 935–944.
- Ramkrishna, S. (2010). Biosurfactants. Springer Science+Business Media, LLC Landes Bioscience. pp. 1–356.
- Ramos, M. J., Fernández, C. M., Casas, A., Rodríguez, L., and Pérez, Á. (2009). Influence of fatty acid composition of raw materials on biodiesel properties. *Bioresource Technology*, 100(1), 261–268.
- Richards, F. J. (1959). A flexible growth function for empirical use. *Journal of Experimental Botany*, 10, 290–300.
- Ricker, W. E. (1979). Growth rates and models. *Fish Physiology*, 8, 677–743.
- Roane, T. M., and Pepper, I. L. (2000). Microbial responses to environmentally toxic cadmium. *Microbial Ecology*, 38, 358–364.
- Robinson, J. B., and Tuovinen, O. H. (1984). Mechanisms of microbial resistance and detoxification of mercury and organomercury compounds: physiological, biochemical, and genetic analyses. *Microbiological Reviews*, 48(2), 95–124.
- Robles-González, I. V., Fava, F., and Poggi-Varaldo, H. M. (2008). A review on slurry bioreactors for bioremediation of soils and sediments. *Microbial Cell Factories*, 7(5), 1–16. <https://doi.org/10.1186/1475-2859-7-5>
- Rodrigues, L., Banat, I. M., Teixeira, J., and Oliveira, R. (2006). Biosurfactants: potential applications in medicine. *Journal of Antimicrobial Chemotherapy*, 57, 609–618. <https://doi.org/10.1093/jac/dkl024>
- Rodriguez-mateus, Z., Agualimpia, B., and Zafra, G. (2016). Isolation and molecular characterization of microorganisms with potential for the degradation of oil and grease from palm oil refinery wastes. *Chemical Engineering Transaction*, 49, 517–522. <https://doi.org/10.3303/CET1649087>
- Roman, O., Heyd, B., Broyart, B., Castillo, R., and Maillard, M. N. (2013). Oxidative reactivity of unsaturated fatty acids from sunflower, high oleic sunflower and rapeseed oils subjected to heat treatment, under controlled conditions. *LWT - Food Science and Technology*, 52(1), 49–59.
- Roosaare, M., Puustusmaa, M., Möls, M., Vaher, M., and Remm, M. (2018). PlasmidSeeker: identification of known plasmids from bacterial whole genome sequencing reads. *PeerJ*, 2018(4), 1–16. <https://doi.org/10.7717/peerj.4588>
- Rosenberg, M. (1984). Bacterial adherence to hydrocarbons: a useful technique for studying cell surface hydrophobicity. *FEMS Microbiology Letters*, 22, 289–295.
- Roslee, A. F. A., Zakaria, N. N., Convey, P., Zulkharnain, A., Lee, G. L. Y., Gomez-Fuentes, C., and Ahmad, S. A. (2020). Statistical optimisation of growth conditions and diesel degradation by the Antarctic bacterium, *Rhodococcus* sp. strain AQ5–07. *Extremophiles*, 24(2), 277–291. <https://doi.org/10.1007/s00792-019-01153-0>
- Ross, D. A. (2004). The toxicology of mercury. *The New England Journal of Medicine*, 350, 945–947.
- Ross, T., and McMeekin, T. A. (1994). Predictive microbiology. *International Journal of Food Microbiology*, 23, 241–264. <https://doi.org/10.1016/B978-1-78548-155-0.50001-0>
- Ruggieri, L., Artola, A., Gea, T., Sanchez, A., and Sánchez, A. (2008).

- Biodegradation of animal fats in a co-composting process with wastewater sludge. *International Biodeterioration & Biodegradation*, 62(3), 297–303.
- Russell, D. J., and Carlson, B. A. (1978). Edible oil pollution on fanning island. *Pacific Science*, 32(1), 1–16.
- Sadler, W. R., and Trudinger, P. A. (1967). The inhibition of microorganisms by heavy metals. *Mineralium Deposita*, 2(3), 158–168. <https://doi.org/10.1007/BF00201912>
- Sáenz-Marta, C. I., María de Lourdes Ballinas-Casarrubias, Rivera-Chavira, B. E., and Nevárez-Moorillón, G. V. (2015). Biosurfactants as useful tools in bioremediation. *Intech Open Science*, 2, 1–19. <https://doi.org/10.5772/32009>
- Salihu, A., and Alam, M. Z. (2012). Production and applications of microbial lipases: A review. *Scientific Research and Essays*, 7(30), 2667–2677. <https://doi.org/10.5897/SRE11.2023>
- Salihu, A., Bala, M., and Bala, S. M. (2013). Application of Plackett-Burman experimental design for lipase production by *Aspergillus niger* using shea butter cake. *BioMed Research International*, 2013, 1–5.
- Samah, A. A., Convey, P., Alias, S. A., and Smykla, J. (2013). Antarctica: Malaysia's journey to the ice. Kuala Lumpur: National Antarctic Research Centre (p. 275).
- Samah, A. A., and Rahman, N. A. (2007). Sustaining Malaysia's interest in Antarctica: from diplomacy to science. In *Proceedings of International Symposium Asian Collaboration in IPY 2007–2008*. www.researchgate.net/publication/268360430. pp. 1–4.
- Sandrin, T. R., and Maier, R. M. (2003). Impact of metals on the biodegradation of organic pollutants. *Environmental Health Perspectives*, 111(8), 1093–1101. <https://doi.org/10.1289/ehp.5840>
- Saravanan, P., Pakshirajan, K., and Saha, P. (2009). Batch growth kinetics of an indigenous mixed microbial culture utilizing m-cresol as the sole carbon source. *Journal of Hazardous Materials*, 162(1), 476–481. <https://doi.org/10.1016/j.jhazmat.2008.05.069>
- Sarwar, M., Ahmad, N., Bux, M., Ali, A., and Tofique, M. (2004). Response of various Brassica genotypes against aphids infestation under natural conditions. *Pakistan Journal of Zoology*, 36(1), 69–74.
- Sarwar, M., Ahmad, N., Siddiqui, Q. H., Rajput, A. A., and Tofique, M. (2003). Efficiency of different chemicals on canola strain Rainbow (*Brassica napus* L.) for aphids control. *Asian Journal of Plant Sciences*, 2(11), 831–833.
- Sathianathan, R. V., Kannapiran, A., Govindan, J., and Periasamy, N. (2014). Profiling of fatty acid compositional alterations in edible oils upon heating using gas chromatography. *Journal of Physical Science*, 25(2), 1–14.
- Satpute, S. K., Bhuyan, S. S., Pardesi, K. R., Mujumdar, S. S., Dhakephalkar, P. K., Shete, A. M., Chopade, B. A., Gvoduypobm, T., Boe, E., Tjhojgz, D., Jowpmwfnfou, U. I. F., and Qbsujdvms, P. G. (2010). Molecular genetics of biosurfactant synthesis in microorganisms. In *Biosurfactant* (pp. 14–41).
- Schattner, P., Brooks, A. N., and Lowe, T. M. (2005). The tRNAscan-SE, snoscan and snoGPS web servers for the detection of tRNAs and snoRNAs. *Nucleic Acids Research*, 33, 686–689. <https://doi.org/10.1093/nar/gki366>
- Schmitz, D., Putzke, J., de Albuquerque, M. P., Schünemann, A. L., Vieira, F. C. B., Victoria, F. de C., and Pereira, A. B. (2018). Description of plant communities on Half Moon Island, Antarctica. *Polar Research*, 37(1), 1–13. <https://doi.org/10.1080/17518369.2018.1523663>

- Scully, T. (2011). "The Development of the Antarctic Treaty system," in science diplomacy: Antarctica, science and the governance of international spaces, P. A. Berkman, M. A. Lang, D. W. H. Walton, and O. R. Young, Eds., ed Washington, DC: Smithsonian institution scholarly. pp. 10.
- Serikovna, S. Z., Serikovich, K. S., Sakenovna, A. S., Murzakhmetovich, S. S., and Khamitovich, A. K. (2013). Screening of lipid degrading microorganisms for wastewater treatment. *Malaysian Journal of Microbiology*, 9(3), 219–226. <https://doi.org/10.21161/mjm.49612>
- Shabtai, Y., and Daya-Mishne, N. (1992). Production, purification, and properties of a lipase from a bacterium (*Pseudomonas aeruginosa* YS-7) capable of growing in water-restricted environments. *Applied and Environmental Microbiology*, 58(1), 174–180.
- Shabudin, A. F. A., Said, N. A., Rahim, R. A., and Ng, T. F. (2016). Malaysia's EOC strategy in strengthening the science knowledge, awareness and national interest towards the polar regions. *International Journal of Environmental and Science Education*, 11(12), 5516–5532.
- Sharma, B., Singh, S., and Siddiqi, N. J. (2014). Biomedical implications of heavy metals induced imbalances in redox systems. *BioMed Research International*, 2014, 1–26. <https://doi.org/10.1155/2014/640754>
- Sharma, K., and Rathore, M. (2010). Characterisation of physical factors for optimum lipase activity from some bacterial isolates. *International Journal of Current Trends in Science and Technology*, 1(2), 51–62.
- Shimizu, K. (2013). Metabolic regulation of a bacterial cell system with emphasis on *Escherichia coli* metabolism. *ISRN Biochemistry*, 2013, 1–47. <https://doi.org/10.1155/2013/645983>
- Shoeb, E., Ahmed, N., Akhter, J., Badar, U., Siddiqui, K., Ansari, F. A., Waqar, M., Imtiaz, S., Akhtar, N., Shaikh, Q. ul A., Baig, R., Butt, S., Khan, S., Khan, S., Hussain, S., Ahmed, B., and Ansari, M. A. (2015). Screening and characterization of biosurfactant-producing bacteria isolated from the Arabian Sea coast of Karachi. *Turkish Journal of Biology*, 39, 210–216. <https://doi.org/10.3906/biy-1405-63>
- Shon, H. K., Tian, D., Kwon, D. Y., Jin, C. S., Lee, T. J., and Chung, W. J. (2002). Degradation of fat, oil, and grease (FOGs) by lipase-producing bacterium *Pseudomonas* sp. strain D2D3. *Journal of Microbiology and Biotechnology*, 12(4), 583–591.
- Shukor, M. Y., Gusmanizar, N., Ramli, J., Shamaan, N. A., MacCormack, W. P., and M A Syed. (2009a). Isolation and characterization of an acrylamide-degrading Antarctic bacterium. *Journal of Environmental Biology*, 30(1), 107–112. [https://doi.org/10.1016/s0378-1097\(97\)00409-6](https://doi.org/10.1016/s0378-1097(97)00409-6)
- Shukor, M. Y., Dahalan, F. A., Jusoh, A. Z., Muse, R., Shamaan, N. A., and Syed, M. A. (2009b). Characterization of diesel-degrading strain isolated from hydrocarbon-contaminated site. *Journal of Environmental Biology*, 30(1), 145–150.
- Sieber, J. R., Sims, D. R., Han, C., Kim, E., Lykidis, A., Lapidus, A. L., McDonnald, E., Rohlin, L., Culley, D. E., Gunsalus, R., and McInerney, M. J. (2010). The genome of *Syntrophomonas wolfei*: New insights into syntrophic metabolism and biohydrogen production. *Environmental Microbiology*, 12(8), 2289–2301. <https://doi.org/10.1111/j.1462-2920.2010.02237.x>
- Siegel, G., Ermilov, E., Pries, A. R., Winkler, K., Schmidt, A., Ringstad, L., Malmsten, and Lindman, B. (2014). The significance of lipid peroxidation in

- cardiovascular diseases. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 442, 173–180.
- Sieuwert, S., De Bok, F. A. M., Mols, E., De Vos, W. M., and Van Hylckama Vlieg, J. E. T. (2008). A simple and fast method for determining colony forming units. *Letters in Applied Microbiology*, 47(4), 275–278. <https://doi.org/10.1111/j.1472-765X.2008.02417.x>
- Sihag, S., and Pathak, H. (2016). Biodegradation of 2T engine oil using soil microbe and gravimetric analysis. *International Journal of Scientific & Engineering Research*, 7(2), 1286–1295. Retrieved from www.pelagiaresearchlibrary.com
- Simencio Otero, R. L., Canale, L. C. F., Totten, G. E., and Meekisho, L. (2017). Vegetable oils as metal quenchant: A comprehensive review. *Materials Performance and Characterization*, 6(1), 1–78.
- Singer, A. (2006). Phytoremediation and rhizoremediation: In Theoretical backgrounded. by M. Mackova, D. Dowling, T. Macek Springer, Dordrecht. pp. 5–21.
- Smith, D. W., and Herunter, S. M. (1989). Birds effected by a canola oil spill in Vancouver Harbour, February 1989. *Spill Technology Newsletter*, 14, 3–5.
- Sogani, M., Mathur, N., Sharma, P., and Bhatnagar, P. (2012). Comparison of immobilized whole resting cells in different matrices vis-A-vis free cells of *Bacillus megaterium* for acyltransferase activity. *Journal of Environmental Research and Development*, 6(3), 695–701.
- Souza, E. C., Vessoni-Penna, T. C., and De Souza Oliveira, R. P. (2014). Biosurfactant-enhanced hydrocarbon bioremediation: An overview. *International Biodeterioration and Biodegradation*, 89, 88–94. <https://doi.org/10.1016/j.ibiod.2014.01.007>
- Soyol-Erdene, T.-O., Huh, Y., Hong, S., and Hur, S. D. (2011). A 50-year record of platinum, iridium, and rhodium in Antarctic Snow: volcanic and anthropogenic sources. *Environmental Science and Technology*, 45(14), 5929–5935.
- Stebbing, A. R. D. (2002). Tolerance and hormesis - Increased resistance to copper in hydroids linked to hormesis. *Marine Environmental Research*, 54(3–5), 805–809. [https://doi.org/10.1016/S0141-1136\(02\)00119-8](https://doi.org/10.1016/S0141-1136(02)00119-8)
- Subramaniam, K., Mazuki, T. A. T., Shukor, M. Y., and Ahmad, S. A. (2019). Isolation and optimisation of phenol degradation by Antarctic isolate using in factor at time. *Malaysian Journal of Biochemistry & Molecular Biology*, 1, 79–86.
- Suhaila, Y. N., Ramanan, R. N., Rosfarizan, M., Latif, I. A., and Ariff, A. (2013). Optimization of parameters for improvement of phenol degradation by *Rhodococcus* UKMP-5M using response surface methodology. *Annual Reviews in Microbiology*, 63, 513–521. <https://doi.org/10.1007/s13213-012-0496-6>
- Suwansukho, P., Rukachisirikul, V., Kawai, F., and H-Kittikun, A. (2008). Production and applications of biosurfactant from *Bacillus subtilis* MUV4. *Songklanakarin Journal of Science and Technology*, 30(1), 87–93.
- Szopin´ska, M., Namies´nik, J., and Polkowska, Z. (2016). How important is research on pollution levels in Antarctica? historical approach, difficulties and current trends. *Reviews of Environmental Contamination and Toxicology*, 240, 1–78. <https://doi.org/10.1007/398>
- Tays, C., Guarnieri, M. T., Sauvageau, D., and Stein, L. Y. (2018). Combined effects of carbon and nitrogen source to optimize growth of proteobacterial methanotrophs. *Frontiers in Microbiology*, 9, 1–14. <https://doi.org/10.3389/fmicb.2018.02239>

- Teissier, G. (1942). Croissance des populations bactériennes et quantité d'aliment disponible (Growth of bacterial populations and the available substrate concentration). *Revue de Science*, 80, 209.
- Tepper, R., and Haward, M. (2005). The development of Malaysia's position on Antarctica: 1982 to 2004. *Polar Record*, 41(217), 113–124. <https://doi.org/10.1017/S0032247405004262>
- Terauds, A., Chown, S. L., Morgan, F., Peat, H. J., Watt, D., Keys, H., Convey, P., and Bergstrom, D. M. (2012). Conservation biogeography of the Antarctic. *Diversity and Distributions*, 18, 726–741.
- Thavasi, R., Sharma, S., and Jayalakshmi, S. (2011). Evaluation of screening methods for the isolation of biosurfactant producing marine bacteria. *Journal of Petroleum & Environmental Biotechnology*, 04(02), 1–6. <https://doi.org/10.4172/2157-7463.s1-001>
- Tin, T., Fleming, Z. L., Hughes, K. A., Ainley, D. G., Convey, P., Moreno, C. A., Pfeiffer, S., Scott, J., and Snape, I. (2009). Impacts of local human activities on the Antarctic environment. *Antarctic Science*, 21(1), 3–33.
- Tiwari, A. K. (2017). Environmental monitoring around Indian Antarctic stations. *Proceedings of the Indian National Science Academy*, 83(2), 399–413. <https://doi.org/10.16943/ptinsa/2017/48964>
- Tonini, R. M. C. W., de Rezende, C. E., and Grativol, A. D. (2010). Degradação e Biorremediação de Compostos do Petróleo por Bactérias: Revisão. *Oecologia Australis*, 14(4), 1025–1035. <https://doi.org/10.4257/oeco.2010.1404.12>
- Undabarrena, A., Salvà-Serra, F., Jaén-Luchoro, D., Castro-Nallar, E., Mendez, K. N., Valencia, R., Ugalde, J. A., Moore, E. R. B., Seeger, M., and Cámara, B. (2018). Complete genome sequence of the marine *Rhodococcus* sp. H-CA8f isolated from Comau fjord in Northern Patagonia, Chile. *Marine Genomics*, 40, 13–17. <https://doi.org/10.1016/j.margen.2018.01.004>
- Vecchiato, M., Argiriadis, E., Zambon, S., Barbante, C., Toscano, G., Gambaro, A., and Piazza, R. (2015). Persistent organic pollutants (POPs) in Antarctica: occurrence in continental and coastal surface snow. *Microchemical Journal*, 119, 75–82. <https://doi.org/10.1016/j.microc.2014.10.010>
- Volchenko, N. N., Karasev, S. G., Nimchenko, D. V., and Karaseva, E. V. (2007). Cell hydrophobicity as a criterion of selection of bacterial producers of biosurfactants. *Microbiology*, 76(1), 112–114. <https://doi.org/10.1134/s0026>
- Waller, C. L., Griffiths, H. J., Waluda, C. M., Thorpe, S. E., Loaiza, I., Moreno, B., Pachterres, C. O., and Hughes, K. A. (2017). Microplastics in the Antarctic marine system: An emerging area of research. *Science of the Total Environment*, 598, 220–227. <https://doi.org/10.1016/j.scitotenv.2017.03.283>
- Walzer, G., Rosenberg, E., and Ron, E. Z. (2006). The *Acinetobacter* outer membrane protein A (OmpA) is a secreted emulsifier. *Environmental Microbiology*, 8(6), 1026–1032. <https://doi.org/10.1111/j.1462-2920.2006.00994.x>
- Wang, Q., Shi, Y. H., Hu, J. X., Yao, Z. W., Fang, X. K., and Dong, Y. A. (2012). Determination of dioxin-like polychlorinated biphenyls in soil and moss from Fildes Peninsula, Antarctica. *Chinese Science Bulletin*, 57(9), 992–996. <https://doi.org/10.1007/s11434-011-4969-y>
- Wang, Y., Tian, Y., Han, B., Zhao, H., Bi, J., and Cai, B. (2007). Biodegradation of phenol by free and immobilized *Acinetobacter* sp. strain PD12. *Journal of Environmental Sciences*, 19(2), 222–225.
- Wayman, M., and Tseng, M. C. (1976). Inhibition threshold substrate concentrations.

- Biotechnology and Bioengineering*, 18(3), 383–387.
- Webb, J. L. (1963). "Enzymes and Metabolic Inhibitors." In *Boston: Academic Press*.
- Wei, I. T. A., Jamalib, N. S., and Ting, W. H. T. (2019). Phytoremediation of palm oil mill effluent (POME) using *Eichhornia crassipes*. *Journal of Applied Science & Process Engineering*, 6(1), 340–354. <https://doi.org/10.33736/jaspe.1349.2019>
- Wirén, N. von, and Merrick, M. (2004). Regulation and function of ammonium carriers in bacteria, fungi and plants. In *Topics in Current genetics*, 3, 95–120. <https://doi.org/10.1007/b95775>
- Yan, A., Wang, Y., Tan, S. N., Mohamed Lokman Mohd Yusof, Ghosh, S., and Chen, Z. (2020). Phytoremediation: a promising approach for revegetation of heavy metal-polluted land. *Frontiers in Plant Science*, 11, 1–15.
- Yang, S. Z., Jin, H. J., Wei, Z., He, R. X., Ji, Y. J., Li, X. M., and Yu, S. P. (2009). Bioremediation of oil spills in cold environments: A review. *Pedosphere*, 19(3), 371–381. <https://doi.org/10.1055/s-0032-1328410>
- Yano, T., and Koga, S. (1969). Dynamic behavior of the chemostat subject to substrate inhibition. *Biotechnology and Bioengineering*, 11(2), 139–153.
- Yogui, G. T., and Sericano, J. L. (2008). Polybrominated diphenyl ether flame retardants in lichens and mosses from King George Island, maritime Antarctica. *Chemosphere*, 73(10), 1589–1593. <https://doi.org/10.1016/j.chemosphere.2008.08.035>
- Yonebayashi, H., Yoshida, S., Ono, K., and Enomoto, H. (2000). Screening of microorganisms for microbial enhanced oil recovery processes. *Journal of The Japan Petroleum Institute*, 43(1), 59–69. <https://doi.org/10.1627/jpi1958.43.59>
- Youssef, N. H., Duncan, K. E., Nagle, D. P., Savage, K. N., Knapp, R. M., and McInerney, M. J. (2004). Comparison of methods to detect biosurfactant production by diverse microorganisms. *Journal of Microbiological Methods*, 56(3), 339–347. <https://doi.org/10.1016/j.mimet.2003.11.001>
- Yusuf, I., Aqlima, S., Phang, L. Y., Syed, M. A., Shamaan, N. A., Khalil, K. A., Dahaln, F. A., and Shukor, M. Y. (2016). Keratinase production and biodegradation of polluted secondary chicken feather wastes by a newly isolated multi heavy metal tolerant bacterium *Alcaligenes* sp. AQ05-001. *Journal of Environmental Management*, 183, 182–195. <https://doi.org/10.1016/j.jenvman.2016.08.059>
- Zakaria, N N, Roslee, A. F. A., Gomez-Fuentes, C., Zulkharnain, A., Abdulrasheed, M., Sabri, S., Ramírez-Moreno, N., Calisto-Ulloa, N., and Ahmad, S. A. (2020). Kinetic studies of marine psychrotolerant microorganisms capable of degrading diesel in the presence of heavy metals. *Revista Mexicana de Ingeniería Química*, 19(3), 1375–1388. Retrieved from <http://www.redalyc.org/articulo.oa?id=62029966013>
- Zakaria, Nur Nadhirah, Ahmad, S. A., Yasid, N. A., Lee, G. L., Manogaran, M., Subramaniam, K., Mazuki, T. T. A., Nawawi, N. M., and Shukor, M. Y. (2018). Biodegradation of Phenol by Antarctic bacterium *Rhodococcus baikonurensis* strain AQ5-001 in the presence of heavy metals. *Malaysian Journal of Biochemistry & Molecular Biology*, 3, 29–36.
- Zakaria, Nur Nadhirah, Roslee, A. F. A., Zulkharnain, A., Gomez-Fuentes, C., Abdulrasheed, M., Sabri, S., Calisto-Ulloa, N., and Ahmad, S. A. (2019). Bacterial growth and diesel biodegradation in the presence of As, Cu and Pb by Antarctic marine bacteria. *Malaysian Journal of Biochemistry and Molecular*

Biology, 22(3), 1–7.

- Zarei, O., Dastmalchi, S., and Hamzeh-Mivehroud, M. (2016). A simple and rapid protocol for producing yeast extract from *Saccharomyces cerevisiae* suitable for preparing bacterial culture media. *Iranian Journal of Pharmaceutical Research*, 15(4), 907–913. <https://doi.org/10.22037/ijpr.2016.1915>
- Zhang, Q., Wang, D., Li, M., Xiang, W. N., and Achal, V. (2014). Isolation and characterization of diesel degrading bacteria, *Sphingomonas* sp. and *Acinetobacter junii* from petroleum contaminated soil. *Frontiers of Earth Science*, 8(1), 58–63. <https://doi.org/10.1007/s11707-013-0415-6>
- Zhao, H., Tian, K., Qiu, Q., Wang, Y., Zhang, H., Ma, S., Jin, S., and Huo, H. (2018). Genome analysis of *Rhodococcus* Sp. DSSKP-R-001: A highly effective β -estradiol-degrading bacterium. *International Journal of Genomics*, 1–11. <https://doi.org/10.1155/2018/3505428>
- Zoueki, C. W., Tufenkji, N., and Ghoshal, S. (2010). A modified microbial adhesion to hydrocarbons assay to account for the presence of hydrocarbon droplets. *Journal of Colloid and Interface Science*, 344(2), 492–496.
- Zoun, P. E. F., Baars, A. J., and Boshuizen, R. S. (1991). A case of sea bird mortality in Netherlands caused by a spillage of nonylphenol and vegetable oils, winter 1988/1989. *Sula*, 5(3), 101–103.
- Zwietering, M. H., Jongenburger, I., Rombouts, F. M., and Van't Riet, K. (1990). Modeling of the bacterial growth curve. *Applied and Environmental Microbiology*, 56(6), 1875–1881.