



**UNIVERSITI PUTRA MALAYSIA**

**DETECTION AND MOLECULAR IDENTIFICATION OF INFECTIOUS  
BRONCHITIS VIRUS ISOLATED IN MALAYSIA**

**MAIZAN BT HJ. MOHAMED**

**FSAS 2000 44**

**DETECTION AND MOLECULAR IDENTIFICATION OF INFECTIOUS  
BRONCHITIS VIRUS ISOLATED IN MALAYSIA**

**By**

**MAIZAN BT HJ. MOHAMED**

**Thesis Submitted in Fulfilment of the Requirements for the Degree of  
Master of Science in the Faculty of Science and Enviromental Studies  
Universiti Putra Malaysia**

**August 2000**



**Special dedication to my husband, my children and my parents.**



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirements for the degree of Master of Science.

**DETECTION AND MOLECULAR IDENTIFICATION OF AVIAN  
INFECTIOUS BRONCHITIS VIRUS ISOLATED IN MALAYSIA**

**By**

**MAIZAN BT HAJI MOHAMED**

**August 2000**

**Chairman : Associate Professor Dr. Khatijah Mohd. Yusoff**

**Faculty : Science and Environmental Studies**

Infectious bronchitis is a disease of economic importance in the poultry industry in Malaysia. Currently, natural outbreaks of IB are controlled through the use of vaccine. However, outbreaks still occur in vaccinated flocks. One of the major factors speculated for these outbreaks is the heterologous protection afforded by the current standard IB vaccines against different IB virus (IBV) serotypes and variants present in the country. Rapid and sensitive methods for the detection and identification of serotypes causing these outbreaks would therefore be valuable to the understanding and control of IB. In this study, reverse-transcription polymerase chain reaction (RT-PCR) using the published primers (C2U/C3L, S1oligo3'/S1oligo5', S1oligo3'/S1Newoligo5', IBP1+/IBRP2-, IBVN2+/IBVN1- and UTR2+/UTR1-) were used to detect 14 Malaysian IBV isolates. Only primers UTR2+/UTR1- and IBP1+/IBRP2- were found to be able to detect all of the 14



Malaysian isolates. These primers could therefore be used as universal primers for the detection of Malaysian IBV isolates.

The S1 gene of a Malaysian nephropathogenic IBV (MH5365/95) and the variable region in the S1 gene of the remaining 13 IBV isolates were RT-PCR using primers S1oligo5-1/S1oligo3' and TM897F/TM1328R respectively. The PCR products were cloned, sequenced and compared the sequences with published sequences of the S1 gene of other IBV strains. Based on this sequence comparison, MH5365/95 was found to be different from the other IBV strains. The remaining 13 isolates were classified into 2 general groups. The first group, that was designated as Group A, consists of three isolates and was closely related (more than 95% homology) to Massachusetts type (M41), the most commonly used vaccine strain in this country. The second group which consists of 10 isolates was genetically different from the published IBVs showing not more than 82% homology with the published sequence. These 10 isolates were further subdivided into Groups B and C, comprising 7 and 3 isolates respectively. The 3 isolates of Group C belong to the same group as MH5365/95. The Group B isolates, however, were more closely related to the M41 serotype (about 82% in homology) compared with that of Group C isolates (less than 80%). There was about 78% nucleic acid homology when Groups B and C were compared. It could therefore be concluded that the outbreaks in this country were caused by the new strains of IBV, which were different from the vaccine strains, thereby resulting in the vaccines becoming less efficacious to protect the chickens against the local IBVs.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia untuk memenuhi keperluan bagi mendapatkan ijazah Master Sains

**PENGESANAN DAN PENGENALPASTIAN MOLEKUL VIRUS BRONKITIS BERJANGKIT AVIAN YANG DIASINGKAN DI MALAYSIA**

Oleh

**MAIZAN BT. HAJI MOHAMED**

**August 2000**

**Pengerusi : Profesor Madya Dr. Khatijah Mohd Yusoff**

**Fakulti : Sains dan Pengajian Alam Sekitar**

Bronkitis berjangkit (IB) merupakan sejenis penyakit berkepentingan ekonomi dalam industri ayam di Malaysia. Semasa ini, wabak semulajadi IB dikawal melalui penggunaan vaksin. Walaubagaimanapun wabak ini masih berlaku di dalam kelompok ayam yang telah divaksinasi. Salah satu faktor penyebab yang dispekulasi adalah perlindungan separa yang diberikan oleh vaksin yang digunakan terhadap berbagai serotip dan varian virus IB (IBV) yang terdapat di negara ini. Oleh itu, kaedah yang cepat dan sensitif untuk pengesanan dan pencirian serotip yang menyebabkan wabak ini adalah penting untuk memahami dan mengesahkannya. Dalam pengkajian ini, tindak balas transkripsi berbalik rantai polimerase (RT-PCR) menggunakan primer rujukan (C2U/C3L, S1oligo3'/S1oligo5', S1oligo3'/S1New oligo5', IBP1+/IBRP2-, IBVN2+/IBVN1- dan UTR2+/UTR1-) telah digunakan dengan tujuan untuk mengesan kehadiran 14 isolat IBV tempatan. Hanya primer

UTR2+/UTR1- dan IBP1+/IBRP2- didapati boleh mengesan kehadiran kesemua 14 isolat ini. Oleh itu pasangan primer ini boleh dianggap sebagai primer universal untuk mengesan kehadiran isolat IBV tempatan.

Gen S1 virus IB isolat nepropatogenik tempatan (MH5365/95) dan bahagian yang berubah-ubah dalam gen tersebut untuk isolat virulen tempatan telah dijalankan tindak balas transkripsi terbalik rantai polimerase dengan menggunakan primer S1oligo5-1/S1oligo3' dan TM897F/TM1328R, di klonkan, dibaca jujukan nukleotidanya dan dibuat perbandingan dengan jujukan nukleotida gen S1 dari strain-strain yang lain. Daripada perbandingan ini didapati MH5365/95 mempunyai jujukan nukleotida yang berbeza daripada lain-lain strain IBV rujukan. Tiga belas isolat IBV telah dikelaskan kepada 2 kumpulan am. Kumpulan pertama yang dinamakan kumpulan A, mengandungi 3 isolat IBV dan didapati berkait-rapat (lebih daripada 95% homologi) dengan strain Massachusetts (M41), iaitu strain vaksin yang paling banyak digunakan di negara ini. Kumpulan kedua yang mengandungi 10 isolat IBV mempunyai jujukan nukleotida yang berlainan (kurang dari 82% homologi) daripada strain IBV rujukan. Sepuluh isolat IBV tempatan ini telah di bahagikan lagi kepada Kumpulan B dan C, masing-masing terdiri daripada 7 dan 3 isolat IBV. Tiga isolat ini berada di bawah kumpulan yang sama dengan MH5365/95. Kumpulan B bagaimanapun mempunyai homologi jujukan nukleotida yang lebih rapat dengan M41 (lebih kurang 82%) berbanding dengan kumpulan C (kurang 80%). Homologi jujukan nukleotida antara kumpulan B dan C adalah lebih kurang 78%. Kesimpulannya, wabak yang berlaku di negara ini adalah disebabkan

oleh IBV strain baru yang berbeza dari strain vaksin, menyebabkan vaksin menjadi kurang berkesan untuk memberi perlindungan kepada ayam terhadap infeksi IBV tempatan ini.



## ACKNOWLEDGEMENTS

I would like to express my thanks and deepest gratitude to the chairman of the supervisory committee, Associate Professor Dr. Khatijah Mohd Yusoff for her guidance, suggestions and comments on this project. I also wish to thank the members of the supervisory committee, Dr. Azri Adzhar, Dr. Siti Suri Arshad and Associate Professor Dr. Abdul Manaf Ali for their comments and helpful advice.

My special thanks to Dr. Takehara from Kitasato University, Japan for his very helpful guidance in the molecular work and to Dr. Sharifah Syed Hassan for her critical reading, editing and encouragement.

My greatest gratitude to the General Director of Veterinary Services for giving me approval to pursue this study and to the Director of VRI, Dr. Aziz Jamaluddin for his support throughout this project.

My appreciation goes to Mdm. Tan Lin Jee, Mdm. Ong Geok Huai, Mdm. Lim Siew Sam, Mdm. Cheah Ngan Yoke, Mdm. Ku Bi Di and Mdm. Rokiah Mustapha for their technical support.

This study was supported by IRPA Grant No. 01-02-04-0107.



# TABLE OF CONTENTS

	<b>Page</b>
DEDICATION	
ABSTRACT	ii
ABSTRAK	iv
ACKNOWLEDGEMENTS	vii
APPROVAL	viii
DECLARATION	x
LIST TABLES	xiii
LIST OF FIGURES	xiv
LIST OF ABBREVIATIONS	xvi
<b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	
1.1 Infectious Bronchitis	1
1.2 Objectives of This Study	4
<b>2 LITERATURE REVIEW</b>	
2.1 Incidence and Distribution	5
2.2 Etiology	6
2.2.1 Classification	6
2.2.2 Genome Structure	6
2.2.3 Chemical Properties	9
2.2.4 Antigenic and Immunogenic Properties of IBV	11
2.2.5 Location of Antigenic Sites	12
2.3 Antigenic Variation Within IBV Strains	14
2.4 Virus Replication	17
2.5 Sequence Variation Among IBV Isolates	19
2.6 Differentiation of IBV Strains	21
<b>3 MATERIALS AND METHODS</b>	
3.1 Viruses	25
3.2 Propagation of the Virus in Embryonated Chicken Eggs (ECE)	25
3.3 Primers	27
3.4 RNA Extraction	27
3.5 Reverse Transcription Process	32
3.6 PCR Amplification	32
3.7 Agarose Gel Electrophoresis	33
3.8 Elution and Purification of DNA Fragments Excised from an Agarose Gel	33



3.9	Cloning	34
3.9.1	Preparation of Competent <i>E. coli</i> Cells	35
3.9.2	Transformation	35
3.10	Small Scale Plasmid Preparation	36
3.11	Large Scale Plasmid Preparation	37
3.12	Nucleotide Sequencing	38
3.12.1	Column Purification of DNA Fragments	38
3.13	Nucleotide and Protein Analysis	39
4	RESULTS	
4.1	Analysis of Universal Primers for the Detection of Infectious Bronchitis Virus Isolated in Malaysia	40
4.1.1	PCR with Primers IBP1+/IBPR2-	42
4.1.2	PCR with Primers UTR2+/UTR1-	42
4.1.3	PCR with Primers C2U/C3L	42
4.1.4	PCR with Primers S1oligo3'/S1oligo5'	43
4.1.5	PCR with Primers S1oligo3'/S1Newoligo5'	43
4.1.6	PCR with Primers IBVN2+/I	43
4.2	Characterisation of a Nephropathogenic IBV Strain MH5365/95	50
4.2.1	Nucleotide Sequence of S1 Gene	52
4.3	Characterisation of Malaysian Isolates of Infectious Bronchitis Virus	57
4.3.1	Amplification of Desired Genes	58
4.3.2	Sequencing	60
4.3.3	Nucleotide Sequence Relationships	64
5	DISCUSSION	
5.1	Analysis of Universal Primers for the Detection of IBV Isolated in Malaysia	65
5.2	Characterisation of a Nephropathogenic IBV Strain MH5365/95	67
5.3	Characterisation of IBV Isolated in Malaysia	69
6	GENERAL DISCUSSION AND FUTURE WORK	71
7	CONCLUSION	73
	REFERENCES	74
	VITA	81

## LIST OF TABLES

<b>Table</b>		<b>Page</b>
<b>3.1</b>	List of Malaysian IBV isolates used in this study	26
<b>3.2a</b>	Sequences, location of primers and annealing temperatures used in PCR amplification of Malaysian IBVs	30
<b>3.2b</b>	List of primers used for sequencing of MH5365/95	31
<b>4.1</b>	Comparison of nucleotide identities (%) and protein similarities (%) between S1 sequences of IBV strains and Malaysian isolate (MH5365/95) using Genetyx-max Ver. 8 (Software Development Co.)	56



## LIST OF FIGURES

Figure		Page
2.1	Model of coronavirus structure	7
2.2	Genomic organisation of coronavirus	8
2.3	Antigenic regions of S1 and S2 glycoproteins	13
2.4	Model of coronavirus replication based on MHV	18
3.1a	Schematic diagram of the location of primers for RT-PCR on selected genes of IBV	28
3.1b	Location of the primers used for sequencing of MH5365/95	29
4.1	RT-PCR with primers IBP1+/IBRP2-	44
4.2	RT-PCR with primers UTR2+/UTR1-	45
4.3	RT-PCR with primers C2U/C3L	46
4.4	RT-PCR with primers S1oligo3'/S1oligo5'	47
4.5	RT-PCR with primers S1oligo3'/S1Newoligo5'	48
4.6	RT-PCR using primers IBVN2+/IBVN1-	49
4.7	PCR amplification of MH5365/95 using S1oligo3'/S1oligo5-1	53
4.8	Nucleotide and amino acid sequences of the S1 gene of MH5365/95	54
4.9	The phylogenetic tree showing the nucleotide sequence relationships between MH5365/95 and known IBV strains	55
4.10	Agarose gel electrophoresis of PCR products produced by amplifying the reference strain M41 and Malaysian isolates using TM897F and TM1328R primers	59
4.11	Sequences of cloned-DNA of 13 representative Malaysian isolates	61



<b>4.12</b>	<b>The phylogenetic tree showing the nucleotide sequence relationships of 13 IBV isolated in Malaysia to other IBV strains</b>	<b>62</b>
<b>4.13</b>	<b>S1 gene sequence homology (%) of American, European, Australian and Malaysian IBV strains</b>	<b>63</b>



## LIST OF ABBREVIATIONS

AF	:allantoic fluid
AGPT	:agar gel precipitin test
AGID	:agar gel immunodiffusion
B-ME	:β-mercapto-ethanol
°C	:degree Celcius
CaCl <sub>2</sub>	:calcium chlorida
cDNA	:complementary DNA
CFT	:complement fixation test
C <sub>2</sub> H <sub>3</sub> O <sub>2</sub> K	:potassium acetate
DNA	:deoxynucleotide acid
dNTP	:deoxynucleoside triphosphate
ECE	:embryonated chicken egg
ELISA	:enzyme-linked immunosorbent assay
g	:gravity
G+C	:Guanine+Cytosine
H	:hour
HA	:haemagglutination
HI	:haemagglutination inhibition
HVR	:hypervariable region
IB	:Infectious bronchitis
IBV	:Infectious bronchitis virus



kDa	:kilodalton
kb	:kilobase pair
KCl	:potassium chloride
KH <sub>2</sub> PO <sub>4</sub>	:di-potassium hydrogen phosphate
LB	:Luria Bertani
M	:membrane
Mabs	:monoclonal antibodies
MgCl <sub>2</sub>	:magnesium chloride
MHV	:mouse hepatitis virus
min	:minute
M <sub>w</sub>	:molecular weight
M41	:Massachusetts 41
μl	:microliter
N	:nucleocapsid
NaCl	:sodium chloride
NaOH	:sodium hydroxide
Na <sub>2</sub> PO <sub>4</sub>	:di-sodium hydrogen phosphate
ng	:nanogram
ORF	:open reading frame
PAGE	:polyacrylamide gel electrophoresis
PBS	:phosphate-buffered saline
PCR	:polymerase chain reaction
PCR-RFLP	PCR-Restriction Fragment Length Polymorphism





PCI	:phenol chloroform isoamyl
RNA	:ribonucleic acid
RT	:reverse transcription
RT-PCR	: reverse transcription- polymerase chain reaction
S	:spike
s	:second
SDS	:sodium dodecyl sulphate
Sec	:section
sM	:small membrane
SPF	:specific pathogen free
TBE	:Tris borate EDTA
TE	:Tris EDTA
TGEV	transmissible gastroenteritis virus
TOC	:tracheal organ culture
TSR	:template suspension reagent
U	:unit
UPM	:Universiti Putra Malaysia
UTR	:untranslated region
UV	:ultraviolet
VN	:virus neutralisation
VRI	:Veterinary Research Institute
v/v	:volume/volume
w/v	:weight/volume

# CHAPTER 1

## INTRODUCTION

### 1.1 INFECTIOUS BRONCHITIS

Infectious bronchitis (IB) is an acute highly contagious and infectious disease of the respiratory system in young chickens. Mortality in young chicks is usually 25-30% but in some outbreaks, it can be as high as 75%. In older birds, the disease often goes unnoticed, but in laying hens, there is a marked drop in egg production and quality (King and Cavanagh, 1991). Nephrosis is also observed in chicken infected with certain IB virus (IBV) strains. The severity of respiratory infections with IBV can be greatly enhanced by the presence of other pathogens of the respiratory tract such as mycoplasma and *Escherichia coli*. The disease was first reported in 1931 in the U.S.A (Beach and Schalm, 1936). IB is of economic importance because it is a cause of poor weight gain and feed efficiency, also a component of mixed infection that produce air-sacculitis resulting in condemnations of broilers at processing (King and Cavanagh, 1991).

In Malaysia, IBV was first reported as early as 1967 where the disease was mild and vaccination unwarranted (Chong and Omar, 1967). However, the situation changed dramatically ten years later when the IB cases increased in parallel with intensification of poultry farming. It was commonly believed for many years that most of the IBV isolates were of the Massachusetts (Mass) serotype since vaccine of



only this serotype was used. The increase in the incidence of IB occurred in flocks vaccinated with M41 vaccine. Nine out of 20 Malaysian isolates characterised by virus neutralisation test (VNT) at the Poultry Research Laboratory in Athens, Greece were found to be related to the Mass serotype and the other 11 isolates were considered as "local" strains (Opitz *et al.*, 1979). In 1995, a local nephropathogenic strain (MH5365/95) which caused more than 20% mortality in chickens was isolated and it was found not to be related to the Australian T and M41 strains by VNT (Aziz *et al.*, 1996). During the same time, an IBV Dutch strain was also present in this country (Azri *et al.*, 1997). Despite the widespread use of both live attenuated and inactivated IB vaccines, IB disease continues to be a major problem in Malaysia. Vaccine breakdown, lack of cross-protection between IBV vaccinal and field viruses, and the emergence of variant strains are some of the factors that resulted in the continued occurrence of IB outbreaks (Azri *et al.*, 1996). Variant strains may arise from the emergence of new antigenic variants and also to the inadvertent introduction of geographically exotic serotypes of the virus, of which protection is not covered by the vaccine currently used (Wang *et al.*, 1993). Other possibilities include mutations of the viruses or recombinations between strains due to genetic reassortments (Kusters *et al.*, 1990). Recovered birds may become carriers and shed the virus for variable periods of time, some are suspected of cycling recurrent infections within the flocks.

IBV infection in chickens is currently diagnosed by using embryonated chicken eggs (ECE) and tracheal organ cultures (TOC). Virus isolation can be a lengthy process,



because most field strains require adaptations and usually two or three passages are required before typical lesions are produced in the embryonated chicken eggs (ECE) and in the TOC. Other methods to identify the virus includes VNT test, haemagglutination (HA) test, enzyme-linked immunosorbent assay (ELISA) and electron microscopy (Lukert, 1980). These methods are expensive, labour intensive and often give inconclusive results, hence the development of an alternative method which is more sensitive, specific and rapid is needed.



## **1.2 OBJECTIVES OF THIS STUDY**

The objectives of this study were :

- i.** to identify universal primers that can detect all the IBV strains isolated in Malaysia.
  
- ii.** to sequence the complete S1 gene of the prototype Malaysian nephropathogenic strain of IBV (MH5365/95).
  
- iii.** to characterise 13 Malaysian IBV isolates by phylogenetic tree analysis using the variable region of the S1 gene sequence.

## CHAPTER 2

### LITERATURE REVIEW

#### 2.1 Incidence and distribution

The first isolate of IBV is of the Beaudette strain (Beaudette and Hudson, 1937). Later, the M41 strain was isolated in the North Dakota and was found to be serologically related to Beaudette strain (Bracewell, 1975). Strains of this serotype were among the first to be made into live vaccines, e.g. H120 (derived from a Dutch isolate of 1955). Strain M41 and its prototype; i.e. the Connecticut (Jungherr *et al.*, 1956), Holte and Gray strains have been widely used as a basis for comparison with other isolates mostly by the virus neutralisation (VN) test and these strains can cause problems and pathological changes to the kidney (Winterfield and Hitchner, 1962).

In Europe, only 30% of the isolates were serologically related to the known North American serotypes in the 1970s. The majority of the remainder strains were found to be related to four Dutch strains; D207, D212, D3128 and D3896 (Davelaar *et al.*, 1984) by VN test. Many of the British IBV strains isolated between 1981 to 1983 were closely related to the Dutch Strains; D207, D3128 and D3896 (Cook, 1984). Although these viruses caused respiratory disease and aberrant drop in egg production in layer flocks, they are not associated with poor egg quality (Cook, 1986).

The Australian 'T' strain was the first IBV strain isolated in Australia in 1962. It caused both nephritic and respiratory forms of IB with 10 to 15% mortality in

