



UNIVERSITI PUTRA MALAYSIA

***IN VITRO CATHARANTHUS ROSEUS (L.) G. DON CALLUS
RESPONSES TO CHITOSAN TREATMENTS***

NUR ATIKAH ISMAIL

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*IN VITRO CATHARANTHUS ROSEUS (L.) G. DON CALLUS RESPONSES
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By:

NUR ATIKAH BINTI ISMAIL

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NUR ATIKAH BINTI ISMAIL

FACULTY OF AGRICULTURE

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CERTIFICATION

This study report entitled “*IN VITRO CATHARANTHUS ROSEUS* (L.) G. DON CALLUS RESPONSES TO CHITOSAN TREATMENTS” is prepared by NurAtikahBinti Ismail and submitted to the Faculty of Agriculture in fulfillment of requirement of PRT 4999 (Project) for the award of degree of Bachelor of Horticultural Science.

Student's name:

NurAtikahBinti Ismail

Student's signature

Certified by:

(ASSOC. PROF. DR. SALEH KADZIMIN)

Project Supervisor,

Department of Crop Science,

Faculty of Agriculture,

Universiti Putra Malaysia

Date:

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LIST OF ABBREVIATIONS

ANOVA	Analysis of Variance
Df	Degree of freedom
<i>et al</i>	And others
g	Gram
mg/L	Milligram per litre
m	Milli
mm	Millimetre
µm	Micrometer
mL/min	Millilitre per minute
MS	Murashige and Skoog
LSD	Least Significant Difference
SAS	Statistical Analysis System
GCMS	Gas chromatography-mass spectrometry

ABSTRACT

The present study investigates the effects of chitosan on callus growth and development of *Catharanthus roseus* under light treatment. In the study, chitosan was supplemented in culture medium. Callus of *C. roseus*, previously initiated on MS medium before the onset of the experiment was cultured. Various levels of chitosan (0, 1, 2, 3, 4, 5 mg/L) were used. The experiment was conducted in a completely randomized design with 3 replications. Study parameters include dry weight of callus and boron level.

Data recorded that the control, 0 mg/L chitosan concentration gave the highest dry weight of callus 0.210 g. Treatment with 1 mg/L chitosan concentration increased boron level in callus of *C. roseus* to 0.097 mg/L. The study suggests that the application of chitosan in culture medium did not give significant increase in dry weight of callus compared to the control. Chitosan treatment did not give significant increase or decrease in boron level of callus.

The study concludes that chitosan had no significant effect on dry weight and boron level in callus of *C. roseus* under light treatment.

ABSTRAK

*Kajian ini menyiasat tentang kesan kitosan pada pertumbuhan kalus dan pembangunan *Catharanthus roseus* dibawah rawatan cahaya. Dalam kajian itu, kitosan telah ditambah dalam medium kultur. Kalus *C.roseus*, yang sebelum ini dimulakan pada medium MS sebelumbermulanya eksperimen ini dikulturkan. Pelbagai tahap kepekatan chitosan (0, 1, 2, 3, 4, 4, mg/L) telah digunakan. Eksperimen ini dijalankan dalam rekabentuk yang benar-benar rawak dengan ulangan. Parameter kajian termasuk berat kering kalus dan tahap boron.*

*Data direkodkan bahawa kawalan 0 mg/L kepekatan kitosan memberikan berat kering kalus yang tertinggi iaitu 0.210 g. rawatan dengan kepekatan kitosan 1 mg/L meningkatkan boron dalam kalus *C.roseus* iaitu 0.097 mg/L. kajian ini mencadangkan bahawa penggunaan kitosan dalam medium kultur memberi peningkatan yang tidak ketara dalam berat kering kalus berbanding dengan kawalan. Rawatan kitosan tidak memberikan peningkatan atau penurunan yang ketara dalam tahap boron di dalam kalus.*

*Kesimpulannya, kitosan tidak memberikan kesan yang ketara kepada berat kering dan tahap boron di dalam kalus *C.roseus* di bawah rawatan cahaya.*

CHAPTER 1

INTRODUCTION

Catharanthus roseus is an ornamental plant belonging to the family Apocynaceae and has been widely planted in several countries for the ornamental purposes. Besides its use as an ornamental plant, it is an important source of one hundred and thirty valuable alkaloids. These include alkaloids which are routinely used in the treatment of cancer, leukemia, and diabetes. However, the alkaloids are produced in minute amounts making them high in demand and cost. As a consequence numerous efforts to develop alternative strategies for their production have been documented.

The present study examines responses in growth and development of callus of *C. roseus* on chitosan treatments. Callus was previously initiated from culture of leaf explants on Murashige and Skoog medium. Resulting callus was cultured on same medium and supplemented with various levels of chitosan.

REFERENCES

- Aslam, J., Khan, S. H., Siddiqui, Z. H., Fatima, Z., Maqsood, M., Bhat, M. A. (2010). *Catharanthus roseus* (L.) G. Don. an important drug. Its application and production. *Pharmacie Globale*, 4: 104-110.
- Banos, S.B., Lauzardoa, A.N.H., Vallea, M.G.V.D., Lopeza, M.H., Barkab, E.A., Molinac, E.B., Wilson, C.L. (2006). Chitosan as a potential natural compound to control pre and postharvest diseases of horticultural commodities. *Crop Protection*, 25: 108–118.
- Benhamou, N., Kloepper, J.W., Tuzun, S. (1998). Induction of resistance against Fusarium wilt of tomato by combination of chitosan with an endophytic bacterial strain: ultrastructure and cytochemistry of the host response. *Planta*, 204 : 153–168.
- Blevins, D. G., and Lukaszewski, K. M. (1998). Boron in plant structure and function. *Annu. Rev. Plant Physiol. Plant Mol. Biol.*, 49: 481-500.
- Borax, (2012). Retrieved 18 October 2014, from <http://www.borax.com/docs/agronomy-notes/functionsofboroninplantnutrition-final-feb2012.pdf?sfvrsn=2>
- Cheng, X.Y., Zhou, H.Y., Cui, X., Ni, W., Liu, C.H. (2006). Improvement of phenylethanoid glycosides biosynthesis in *Cistanchedeserticola* cell suspension cultures by chitosan elicitor. *Journal of Biotechnology*, 121: 253–260.
- Dutta, A., Batra, J., Pandey-Rai, S., Singh, D., Kumar, S., Sen, J. (2004). Expression of terpenoidindole alkaloid biosynthetic pathway genes corresponds to accumulation of related alkaloids in *Catharanthus roseus* (L.) G. Don. *Planta* 220: 376-383.
- ETI products, (2014). Retrieved 18 October 2014, from <http://www.etiproducts.com/boron-in-agriculture-5s.htm>
- Etimine USA, Inc., (2014). Retrieved 12 October 2014, from <http://www.etimineusa.com/en/boric-acid>

- Fraser, R.S., Loening, U. E., and Yeoman, M. M. (1967).Effect of light on cell division in plant tissue cultures.*Nature*, 215:873.
- Freepons, D., 1991. Chitosan does it have a place in agriculture. *Proc. Plant Growth Regulators Soc. Am.*, pp.11–19.
- George, E.F. and Sherrington, P. D. (1984).Plant propagation by tissue culture.*Journal of Basic Microbiology*, 25: 475.
- Gilman, E.F., and Howe, T. (1999).*Catharanthus roseus* Periwinkle, Madagascar Periwinkle.University Florida IFAS Extension. Retrieved 13 July 2014, from <http://edis.ifas.ufl.edu/fp112>
- Goldbach, H. E., and Wimmer, M. (2007). Boron in plants and animals: Is there a role beyond cell-wall structure. *J. Plant Nutr. Soil Sci.*, 170: 39-48.
- Gupta, R. (1977). Periwinkle-produces anticancer drug.*Indian Farming* 7: 11-13.
- Hadrami, A. E., Adam, L. R., Hadrami, I. E., and Daayf, F. (2010).Chitosan in plant protection.*Marine Drug*, 8(4): 968-987).
- Hirano, S. (1988). The activation of plant cells and their self-defence function against pathogens in connection with chitosan (in Japanese with English summary). *Nippon Nogeikagaku Kaishi*, 62:293–295.
- Hisiger, S., and Jolicoeur, M. (2007).Analysis of *Catharanthus roseus* alkaloid by HPLC.*Phytochemistry Reviews*, 6: 207-234.
- Jaramillo, J., and Summers, W. L., (1991). Dark–Light Treatments Influence Induction of Tomato Anther Callus.*Hort Science*, 26(7): 915-916.
- Lata, B. (2007). Cultivation, mineral nutrition and seed production of *Catharanthus roseus*(L.) G. Don in the temperate climate zone. *Phytochemistry Reviews*, 6: 403-411.

- Kew, (2014). Retrieved 25 August 2014, from <http://www.kew.org/science-conservation/plants-fungi/catharanthus-roseus-madagascar-periwinkle>
- Mathias, R. J., Fukui, K., and Law, C. N. (1986). Cytoplasmic effects on the tissue culture response of wheat (*Triticum aestivum*) callus. *Theor Appl Genet*, 72: 70-75.
- Murashige, T., and Skoog, F. (1962). A revised medium for rapid growth and bio-assays with tobacco tissue cultures. *Physiol Plant*, 15(3): 473-497.
- National Tropical Botanical Garden, (2003). Retrieved 9 November 2014 from http://www.ntbg.org/plants/plant_details.php
- Nejat, N., Valdiani, A., Cahill, D., Tan, Y.H., Maziah, M., and Abiri, R. (2014). Ornamental exterior versus therapeutic interior of madagascar periwinkle (*Catharanthus roseus*): The two faces of a versatile herb. *The Scientific World Journal*. Retrieved from <http://www.hindawi.com/journals/tswj/aa/982412/>
- Nge, K. L., Nwe, N., Chandkrachang, S., Stevens, W. F. (2006). Chitosan as a growth stimulator in orchid tissue culture. *Plant Science*, 170: 1185–1190.
- Noth, M.H. and Abel, W. O. (1971). Zurentwicklung haploider pflanzen aus unreifen mikrosporen verschiedener *Nototiana* arten. *Z. Pflanzensucht.*, 65: 277-284.
- Ohta, K., Taniguchi, A., Konishi, N., and Hosoki, T. (1999). Chitosan treatment affects plant growth and flower quality in *Eustoma grandiflorum*. *Hort science*, 34(2):233–234.
- Orlita, A., Gorycka, M.S., Paszkiewicz, M., Malinski, E., Kumirska, J., Ewa M. Siedlecka, E.M., Lojkowska, E., and Stepnowski, P. (2010). Application of chitin

and chitosan as elicitors of coumarins and furoquinolone alkaloids in *Rutagraveolens* L. (common rue). *Biotechnol. Appl. Biochem.*, 51: 91–96.

Parrek, S.K., Singh, S., Srivastava, V.K., Mandal, S., Maheswari, M.L., and Gupta, R. (1981). Advances in periwinkle cultivation. *Indian Farming*, 31(6): 18-21.

Pinho, P.G.D., Goncalves, R.F., Valentao, P., Pereira, D.M., Seabra, R.M., Andrade, P.B., and Sottomayor, M. (2009). Volatile composition of *Catharanthus roseus* (L.) G. Don using solid-phase microextraction and gas chromatography/mass spectrometry. *Journal of Pharmaceutical and Biomedical Analysis*, 49: 674-685.

Preto, F. R., and Santarem, E. R. (2000). Callus formation and plant regeneration from *Hypericum perforatum* leaves. *Plant Cell, Tissue and Organ Culture*, 62: 107–113.

Rinaudo, M. (2006). Chitin and chitosan: Properties and applications. *Prog. Polym. Sci.*, 31, 603–632.

Rodriguez, M. B. H., Fontes, A. G., Rexach, J., Cristobal, J. J. C., Maldonado, J. M., and Gochicoa, M. T. N. (2010). Role of boron in vascular plants and response mechanisms to boron stress. *Plant Stress*, 4 (2): 115-122.

Saleem, M., Khanif, Y.M., Fauziah, I., Samsuri, A.W. and Hafeez, B. (2011). Importance of boron for agriculture productivity: A review. *International Research Journal of Agricultural Science and Soil Science*, Vol. 1(8) pp.293-300.

Shehata, S. A., Fawzy, Z. F., and El-Ramady, H. R. (2012). Response of cucumber plants to foliar application of chitosan and yeast under greenhouse conditions. *Australian Journal of Basic and Applied Sciences*, 6(4): 63-71.

Tariq, M., and C. J. B. Mott, C. J. B. (2006). Effect of boron supply on the uptake of Micronutrients by radish (*Raphanussativus* L.). *Journal of Agricultural and Biological Science*, 1(2): 1-8.

Tsugita, T., Takahashi, K., Muraoka, T., and Fukui, H.(1993).The application of chitin/chitosanfor agriculture (in Japanese).*7th Symposium on chitin and chitosan*, p. 21–22.

Vasconcelos, M. W. (2014). Chitosan and chitooligosaccharide utilization in phytoremediation and biofortification programs: current knowledge and future perspectives. *Front. Plant Sci.*, 5: 616.

Verma, A. K., Singh, R. R., and Singh, Seema.(2012). Improved alkaloid content in callus culture of *Catharanthusroseus*.*BotanicaSerbica*, 36(2): 123-130.

Wikipedia (2014). Retrieved 12 July 2014 from <http://en.wikipedia.org/wiki/Chitosan>

Zibbu, G., and Batra, A. (2011). GC-MS analysis of the desert plants of Apocynaceae family: *Nerium oleander* L. and *Thevetiaperuviana* (pers.) schum. *International Journal of Pharmaceutical Research and Development*, 3(10): 49 – 62.