



UNIVERSITI PUTRA MALAYSIA

**SUPPRESSION OF *Botrytis cinerea*, CAUSATIVE AGENT OF GRAY
MOLD OF TOMATO BY ANTAGONISTIC BACTERIA**

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BY
NURUL HUDA BADARUNZAMAN

A project report submitted to Faculty of Agriculture, Universiti Putra Malaysia, in fulfillment of the requirement of PRT 4999 (Final Year Project) for the award of the degree of Bachelor of Agricultural Science

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ENDORSEMENT

This project report entitled “Suppression of *Botrytis Cinerea*, Causative Agent of Gray Mold of Tomato by Antagonistic Bacteria” is prepared by Nurul Huda bt Badarunzaman and submitted to the Faculty of Agriculture in fulfillment of the requirement of PRT 4999 (Final Year Project) for the award of the degree of Bachelor of Agricultural Science.

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LIST OF ABBREVIATIONS

ANOVA	Analysis of Variance
°C	Celcius
Cm	Centimetre
CRD	Completely Randomized Design
NA	Nutrient Agar
et al	et alia 'and others'
PCR	Polymerase Chain Reaction(PCR).
eDNA	Extracellular DNA
<i>B.cinerea</i>	<i>Botrytis cinerea</i>
LSD	Least significant differences
Hr	Hours
µl	Microlitre
µm	Micrometer
mg	Milligram
ml	Millilitre
<i>et al</i>	et alia 'and others'
PIRG	Percentage Inhibition of Radial Growth
PBS	Phosphate Buffered Saline
<i>B. brevis</i>	<i>Botrytis brevis</i>
PDA	Potato Dextrose Agar
NB	Nutrient Broth

Abstract

Botrytis cinerea is one of the most important fungal pathogen in tomato (*Lycopersicon esculentum* Mill.) worldwide. This fungal pathogen is also a major pathogen in other vegetables especially during postharvest stage. *Botrytis cinerea*, produces characteristic gray mold symptoms on fruits, leaves and stems within seven days following contamination. Thus, the objective of this study was to isolate and identify potential biocontrol agent (BCA) which have the ability to suppress growth of *B. cinerea in-vitro*. To achieve the objective, BCA was isolated from diseased and healthy tomato fruits. The *botrytis cinerea* was cultured in PDA medium and the bacteria was cultured in NA medium. The screening test was done via dual cultural test, culture filtrate test and spore suppression test. Finally, five potential BCA were identified in this test from bacteria isolate code BC4, BC23, BM4, BM11, and BM13 were found to be the most effective in suppressing growth of *B. cinerea*.

Abstrak

Botrytis cinerea, salah satu daripada patogen kulat yang paling penting dalam tomato (*Lycopersicon esculentum* Mill.) Hal Ini disebabkan kulat adalah patogen utama dalam sayur-sayuran lain di seluruh dunia terutama pada tanaman lepas tuai. *Botrytis cinerea*, menghasilkan ciri gejala acuan kelabu pada buah-buahan, daun dan batang dalam masa 7 hari selepas pencemaran. Oleh itu, objektif kajian ini adalah untuk mengasingkan dan mengenal pasti agen kawalan biologi yang berpotensi (BCA) yang mempunyai keupayaan untuk menyekat pertumbuhan *B. cinerea* dalam *in-vitro*. Untuk mencapai matlamat tersebut, BCA akan diasingkan daripada buah-buahan tomato berpenyakit dan sihat. Pathogen dari fungus *botrytis cinerea* dikultur dalam PDA media manakala bacteria dikultur di dalam NA media. Ujian saringan dilakukan melalui 'dual culture test', 'culture filtrate test', dan 'spore suppression test'. Akhirnya, lima berpotensi BCA telah dikenal pasti dalam ujian ini adalah bacteria kod bernombor BC4, BC23, BM4, BM11, dan BM13 menunjukkan potensi yang paling berkesan dalam menghalang dan mencegah pertumbuhan *Botrytis cinerea*.

CHAPTER 1

INTRODUCTION

Microorganisms such as bacteria, yeasts and fungi have gained attention recently as they were exhibited biological control ability that can help to reduce or contain certain diseases to be spread in crops. Generally, over the last two decades, the studied has been found that the biocontrol of *botrytis*-incited diseases with filamentous fungi, bacteria, and yeast capable to suppress the disease speared (Blakeman *et al.*, 1993). Biocontrol is the alternative to the conventional methods for the disease control because application of biocontrol control agents (BCAs) are the intensive way and environmental friendly while it also can reduce the mode of action the risk of resistance development. Since the world market for *Botrytis* control products increase by the year, the biocontrol product can help farmer to reduce their cost of pesticides either for conventional system or organic farming.

Newhook and wood (1951) discovered the common species of microorganism with antagonistic *Fusarium* spp. and *Penicillium claviforme* that originated from the same crop that can prevent primary progression of *B. cinerea*. They also investigated about saprophytic activity on dead lettuce tissue was found and it responsible to be control under the natural condition. The controlled of gray mold on glasshouse tomatoes by spraying method with the spore suspension of *Cladosporium herbarum* and *Penicillium* sp. on floral debris attached to the fruit. Besides that, the effective control of *B. cinerea* also can use *C. herbarum* as idea to control strawberry gray mold by protecting the flower under the fields condition (Bhatt and Vaughan,1962).

In addition, the research on chrysanthemum and beetroot leaves reported that there was the antagonistic effect of bacteria on *B. cinerea* (Blakeman and Brodie, 1976). Subsequently, *Trichoderma* spp. has been commonly used for control *B. cinerea* to the postharvest fruits such as grape, bean blossom, strawberry, and greenhouse crop (Eled *et al.*, 1995; O'Neill *et al.*,1996). The effectiveness in controlling *B. cinerea* using bacteria such as *Bacillus* and *Pseudomonas* has been proven to suppress the *Botrytis* conidiation (Redmond *et al.*,1987; Edward and Seddon,1992; Elad *et al.*,1994b)

Biocontrol agents has their strategy and ability to control the pathogens. For example *Botrytis* has behavior during the pre-penetration phase it can develop on healthy plant surface and the relative important of necrotic tissue within the crop as an inoculum source determine the ability plant to protect themselves with unfavourable condition. *Botrytis* was the strongly susceptible to competition condition because its dependence on an exogenous supply nutrients for germination, growth and infection on host. Sometime, *botrytis* conidia or germ-tube were sensitive to the effect of antibiotics and lytic enzymes that produced by microorganisms on plant surface. So, the antibiotic activity on plant that will inhibit germination to disease suppression may be based on reduction of pathogen saprophytic ability, spore dissemination or induced resistance in the host plant (Elad and Freeman,2002).

Microorganism such as bacteria was capable change the wettability of plant surface. Edward and Seddon(1992) discover the plant surface on Chinese cabbages applied with *bacillus brevis* can cause the water drops to spread and dry. Thus, decreasing the wetness and limit the period was suitable for *B. cinerea* germination or infection. It was common same with the effects when the formulation applied to plant surface for providing a secondary benefit from biocontrol research.

Antibiosis in biocontrol mechanism has been reported for many microorganisms. In natural ecosystem sometime hard to predict the importance inhibitory compound that was produced *in vitro* by antagonists. However, the ultimate prove for the role of these compounds in biocontrol was the loss of activity in non-producing mutants of the antagonist but in many case the mutant still operate as biocontrol activity because of the other mode of action. *B. cinerea* has been controlled by antibiotic producer of certain *bacillus* species in various hosts for example *B. brevis* secretes the antibiotic gramicidin S that was known to be effective against *B. cinerea* (Edward and seddon,1992).

In this study, *B. cinerea* pathogen are being tested *in vitro* by using non-identical bacteria that produce in tomato. Antagonistic bacteria were evaluated for their effectiveness in postharvest control of gray mould on fresh-market tomatoes. According to M. Mari *et.al*, (1996), Gray mould was reduced in fresh-market tomatoes treated with antagonists and artificially inoculated with *Botrytis cinerea* and stored at 20°C for at least 7 days. One strain, 5PVB (*Raciltus amyfoliquefaciens*) was particularly effective.

There was a lot of research carried out to investigate the biological control in *Botrytis cinerea* using bacteria. The reduction in inoculum production of *B. cinerea* especially in pathogen's ability will be suppressed and it also may build the positive effect over the other disease cycle in photosystem. Peng and Sutton (1991) found the several microorganisms have been reported to suppress sporulation of *B. cinerea* on strawberry. The same phenomenon was later exploited in various crops around the world especially postharvest crop. Due to its self-protective structure to defend itself, pathogen will resistance to the fungicides if we still continue using same chemical control. Thus, in this study, the bacterial as a biocontrol agent is used to test the inhibition the growth of *Botrytis cinerea* in tomato by antagonist bacteria. The antagonist bacteria are normally isolate from endophytic bacteria within tissue of healthy fruits (Pratella, Mari, Guizzardi and Folchi, 1993). The objective of this study was to isolate and identify BCA agents against *B. cinerea* and assess its antagonistic activity.

REFERENCES

- Y. Elad et al. (eds.), Microbial control of *Botrytis spp.*: Biology, Pathology and control, 223-241
- Newhook FJ (1951) Microbiological control *Botrytis cinerea* pers. II Antagonism by fungi and actinomycetes. *Annals of applied Biology* 35:185-202)
- Bhatt D.D., Vaughan (1962) E.K Preliminary investigation on biological control of grey mold (*Botrytis cinerea*) of strawberries. *Plant Disease Reporter*. 342–345.
- Elad Y (1994) Biological control of grape grey mould by *Trichoderma Harzianum* . *Crop Protection* 13: 35-38
- Blakeman JP and Brodie IDS (1976) Inhibition of pathogens by epiphytic bacteria on aerial plant surfaces. In: Dickson CH and Preece TF (eds) *Microbiology of Aerial Plant Surfaces*. (pp. 529-557)
- Redmond JC, Marois JJ and MacDonald JD (1987) Biological control of *Botrytis cinerea* with epiphytic microorganisms. *Plant Disease* 71: 799-802.
- Edwards SG and Seddon B (1992) *Bacillus brevis* as a biocontrol agent against *Botrytis cinerea* on protected Chinese cabbage. In: Verhoeff K, Malathrakis NE and Williamson B (eds) *Recent Advances in Botrytis Research*. (pp. 267-271) Pudoc Scientific Publishers, Wageningen, The Netherlands Elad.
- Elad Y and Freeman S (2002) Biological control of fungal plant pathogens. In: Kempken F (ed.) *The Mycota, A Comprehensive Treatise on Fungi as Experimental Systems for Basic and Applied Research*. Vol. XI. *Agricultural Applications*. (pp. 93-109)
- Mari, M., Leoni, O., Iori, R. and Marchi, A. (1996a) Bioassay of glucosinolate-derived isothiocyanates against postharvest pear pathogens. *Plant Pathology* **45**, 753–760
- Peng G and Sutton JC (1991) Evaluation of microorganisms for biocontrol of *Botrytis cinerea* in strawberry. *Canadian Journal of Plant Pathology* 13: 247-257
- Pratella, G.C., Mari M., Guizzardi, M. Folchi, A. (1993) Preliminary studies on the efficiency of endophytes in the biological control of the postharvest pathogens *Monilinia laxa* and *Rhizopus stolonifer* in the stone fruits. *Postharvest Biol. Technol.* 3, 361-368.

- Sigei, K. G., Ngeno, K. H., Kibe, M. A., Mwangi, M. M., and Mutai C. M. (2014). Challenges and strategies to improve tomato competitiveness along the tomato value chain in Kenya. *International Journal of Business and Management*, 9(9): 230-245.
- McGraw-Hill (1987). Encyclopedia of Science and Technology 6th Edition, Vol.18, McGraw-Hill Book Company, New York p.392. http://www.sciencepub.net/report/report0304/09_5168report0304_52_57.
- United State Department of Agriculture Economic Research Service Fresh Tomato Industry. <https://www.ers.usda.gov/topics/crops/vegetables-pulses/tomatoes>
- Wills, R. H., Lee, T. H., Graham, D., Mc glassom, W. B. and Hall, E. G. 1981. An Introduction to the physiology and handling of fruits and vegetables. London, 432-438.
- Stinson, E. E., Osman, S. F., Heisler, E. G., Sicihano, J. and Bill, D. D. 1981. Mycotoxin production in whole tomatoes, apples, oranges and lemons. *Journal of Agriculture and Food Chemistry*, 29:790-792.
- Moss, M. O. 2002. Mycotoxin Review *Journal on Aspergillus penicillium*. *Mycologist*, 16:116-119.
- Naika S, Juede J, Goffau M, Hilmi M, Dam V, (2005). “ Cultivation of Tomato” Production, processing and marketing, Agromisa/ CTA. Revised edition, 2005 Agro dok- series No 17.
- Lindy Coates and Greg Johnson (2003). Postharvest Diseases of Fruit and Vegetables, 535-539.
- Jarvis, W.R. 1977. *Botryotinia* and *Botrytis* species - Taxonomy, physiology and pathogenicity. A guide to the literature, Monograph no. 14, Ottawa, Research Branch, Canada Department of Agriculture..
- Staples, R.C. and Mayer, A.M. 1995. Putative virulence factors of *Botrytis cinerea* acting as a wound pathogen. *FEMS Microbiol. Letters* 134: 1-7.
- Prins, T.W., Tudzynski, P., Von Tiedemann, A., Tudzynski, B., Ten Have, A., Hansen, M.E., Tenberge, K. and Van Kan, J.A.L. 2000b. Infection strategies of *Botrytis cinerea* and related necrotrophic pathogens. pp.33-64 in J. Kronstad (ed.), *Fungal Pathology*, Kluwer Academic Publishers.
- .Doss RP, Potter SW, Soeldner AH, Christian JK and Fukunaga LE (1995) Adhesion of germlings of *Botrytis cinerea*. *Applied and Environmental Microbiology* 61: 260-265

- Coates LM, Johnson GI, Cooke AW (1993) Postharvest disease control in mangoes using high humidity hot air fungicide treatments. *Ann. Appl. Biol.* 123: 441 – 448
- T. A. Zitter, Department of Plant Pathology, Cornell University, Ithaca, NY (1986). *Botrytis* Gray Mold of Greenhouse & Field Tomato. http://vegetablemdonline.ppath.cornell.edu/factsheets/Tomato_Botrytis.htm
- Weinberger, K. and T.A. Lumpkin. 2005. Horticulture for poverty alleviation-the unfunded revolution. Shanhua, Taiwan: AVRDC-The World Vegetable Center, AVRDC Publication No. 05-613, Working paper no. 15. 20 p.
- Coley-Smith JR, Verhoeff K and Jarvis WR (1980). *The Biology of Botrytis*. Academic Press, London, UK
- Powelson, R.L., 1960. Initiation of strawberry fruit rot caused by *Botrytis cinerea*. *Phytopathology* 50, 491-494..
- Freeman, J. A. and Pepin, H. S. (1968). A comparison of two systemic fungicides with non-systemic for control of Fruit-rot and powdery mildew in strawberry. *Can. J. Pl. Surv.* 48 : 120-123.
- Gullino ML and Kuijpers LAM (1994) Social and political implications of managing plant diseases with restricted fungicides in Europe. *Annual Review of Phytopathology* 32: 559-579.
- Weller, D.M., Raaijmakers, J.M., McSpadden Gardener, B.B., Thomashow, L.S., (2002) Microbial population responsible for specific soil suppressiveness to plant pathogens. *Annual Review of Phytopathology* 40: 309-348.
- Welbaum, G., Sturz, A.V., Dong, Z., Nowak, J. (2004) Fertilizing soil microorganisms to improve productivity of agro-ecosystems. *Critical Reviews in Plant Sciences* 23: 175-193.
- Cook, R. J., and K. F. Baker. 1983. *The Nature and Practice of Biological Control of Plant Pathogens*. Amer. Phytopathol. Soc., St. Paul, Minnesota. 539 pp.
- Gerhardson, B. 2002. Biological substitutes for pesticides. *Trends Biotechnol.* 20: 338–343
- Johnston, A. E. (1986). "Soil organic-matter, effects on soils and crops". *Soil Use Management* 2, 97–105
- Arcury TA, Quandt SA (2003). Pesticides at work and at home: exposure of migrant farmworkers and their families. *Lancet.* 2003;362:2021.

- Alabouvette, C., Olivain, C. & Steinberg, C. Eur J Plant Pathol (2006). Biological Control of Plant Diseases: The European Situation 114: 329. doi:10.1007/s10658-005-0233-0
- Loper and Henkels (1997). Availability of iron to *pseudomonas fluorescens* in rhizosphere and bulk soil evaluated with with ice nucleation reported gene. Appl. Environ.Microbial.65: 5457-5363
- Haas, D. and Défago, G. (2005). Biological control of soil-borne pathogens by fluorescent pseudomonads. *Nat Rev Microbiol* 3, 307–319.
- Adams PB. 1990. The potential of mycoparasites for biological control of plant diseases. *Ann Rev Phytopathol* 28:59-72.
- Dunn TM, et al. (2000) Selection of yeast mutants in sphingolipid metabolism. *Methods Enzymol* 312:317-30
- Lugtenberg B, Chin-A-Woeng T, Bloemberg G (2002) Microbe-plant interactions: principles and mechanisms. *Antonie van Leeuwenhoek* 81:373–383
- Van Loon LC (2000) Systemic induced resistance. In: Slusarenko AJ, Fraser RSS, van Loon LC (eds) *Mechanism of resistance to plant diseases*. Kluwer, Dordrecht, pp 521–57.
- Whipps JM (2001) Microbial interactions and biocontrol in the *rhizosphere*. *J Exp Bot* 52:487–512
- Van Loon LC, Bakker PAHM, Pieterse CMJ (1998). Systemic resistance induced by *rhizosphere* bacteria. *Annu Rev Phytopathol.* ,page 453–83.
- Bakker, P. A. H. M., Ran, L. X., Pieterse, C. M. J., and Van Loon, L. C., 2003, Understanding the involvement of *rhizobacteria*-mediated induction of systemic resistance in biocontrol of plant diseases. *Can. J. Plant Pathol.* 25:5-9
- Pratella G.C., Mari M., Guizzardi F., Folchi A. (1993): Preliminary studies on the efficiency of endophytes in the biological control of the postharvest pathogens *Monilinia laxa* and *Rhizopus stolonifer* in stone fruit. – *Postharvest Biological Technology* 3: 361–368
- Chaurasia B., A. Pandey, L.M.S. Palni, P. Trivedi, B. Kumar and N. Colvin, (2005). Diffusible and volatile compounds produced by an antagonistic *Bacillus subtilis* strain cause structural deformations in pathogenic fungi in vitro. *Microbiological Research* 160, 75–81
- F. Fiume and G Fiume (2006). Biological Control of *botrytis* Gray Mould on Tomato cultivated in Greenhouse. *Comm.appl. Biol. Sci*, Ghent University, 71/3b,2006 Page 903-905.
- Cong-Jun Yang, Xin-Gang Zhang, Guan-Ying Shi, Hao-Yu Zhao, Long Chen, Ke Tao and Tai-Ping Hou1 (2010). Isolation and Identification of Endophytic

- David MW (1998). Biological control of soilborne plant pathogens in the *rhizosphere* with bacteria. *Ann. Rev. Phytopathol.*, 26: 379-407.
- Xu, S., Pan, X., Luo, J., Wu, J., Zhou, Z., Liang, X., et al. (2015). Effects of phenazine-1-carboxylic acid on the biology of the plant-pathogenic bacterium *Xanthomonas oryzae* pv. *oryzae*. *Pestic. Biochem. Physiol.* 117, 39–46.
- Sutton JC, Peng G (1993) Biocontrol of *Botrytis cinerea* in strawberry leaves. *Phytopathology* 83, 615–621
- Sutton JC, Li D-W, Peng G, Yu H, Zhang P (1997) *Gliocladium roseum*: a versatile adversary of *Botrytis cinerea* in crops. *Plant Disease* 81, 316–328.
- Esitken, A., H. Karlıdag., S. Ercisli and F. Sahin. 2002. Effects of foliar application of *Bacillus subtilis* Osu-142 on the yield, growth and control of shot-hole disease (*Coryneum* blight) of apricot. *Gartenbauwissenschaft*, 67: 139-142
- Altındag, M., M. Sahin, A. Esitken, S. Ercisli, M. Guleryuz, M.F. Donmez and F. Sahin. 2006. Biological control of brown root (*Moniliana laxa* Ehr.) on apricot (*Prunus armeniaca* L. cv. Hacıhaliloğlu) by *Bacillus*, *Burkholderia* and *Pseudomonas* application under In vitro and In vivo conditions. *Biological Control*, 38: 369-372.
- Köhl J, Moelhoek WML, van der Plas, CH, Fokkema NJ (1995a) Suppression of sporulation of *Botrytis* spp. as a valid biocontrol strategy. *European Journal of Plant Pathology* 101, 251–259.
- Kim, Y. S., and S. D. Kim, S.D. (1994). Antifungal mechanism and properties of antibiotic substance produced by *Bacillus subtilis* YB-70 as a biological control agent. *J. Microbiol. Biotech.* 4: 296-304
- Cherif, M., N. Benhamou, and R. R. Belanger (1992). Occurrence of cellulose and chitin in the hyphal cell walls of *Pythium ultimum*: a comparative study with other plant pathogenic fungi
- Essghaier, B., M. L. Fardeau, J. L. Cayol, M. R. Hajlaoui, A. Boudabous, H. Jijakli, and N. Sadfi-Zouaoui (2009). Biological control of grey mould in strawberry fruits by halophilic bacteria. *Journal of Applied Microbiology* 106: 833-846.
- Morandi, M.A.B., L. A. Maffia, E. S. G. Mizubuti, A. C. Alfenas, and J. G. Barbosa (2003). Suppression of *Botrytis cinerea* sporulation by *Clonostachys rosea* on rose debris: a valuable component in *Botrytis* blight management in commercial greenhouses. *Biological Control* 26: 311-317.

Rombouts F.M. and H.J. Phaff, (1976). Lysis of yeast cell walls. Lytic β -1,6-glucanase from *Bacillus circulans* WL-12. European Journal of Biochemistry 63, 109–120.

Manjula K. and A. Podile, 2005. Production of fungal cell wall degrading enzymes by a biocontrol strain of *Bacillus subtilis* AF1. Indian Journal of Experimental Biology 43 (10), 892–896.

Essghaier B., M. Bejji, H. Jijakli, A. Boudabous and N. Sadfi- Zouaoui,(2009). High salt-tolerant protease from a potential biocontrol agent *Bacillus pumilus* M3-16. Annals of Microbiology 59 (3), 553–558.

Weeds PL, Beever RE, Long PG (2000) Competition between aggressive and non-aggressive strains of *Botrytis cinerea* (*Botryotinia fuckeliana*) on French bean leaves. Australasian Plant Pathology 29, 200–204.