

# **UNIVERSITI PUTRA MALAYSIA**

# EFFECT OF ANTIMICROBIAL EXTRACTS OF PIPER BETLE AND CLINACANTHUS NUTANS ON IN VITRO GROWTH OF SELECTED FUNGAL PATHOGENS

NUR MALIYA RAHIM

FP 2016 37

## EFFECT OF ANTIMICROBIAL EXTRACTS OF

## PIPER BETLE AND CLINACANTHUS NUTANS

## ON IN VITRO GROWTH OF SELECTED FUNGAL PATHOGENS

By

NUR MALIYA BT RAHIM

A project report submitted to Faculty of Agriculture, Universiti Putra Malaysia in fulfilment of the requirement of PRT4999 (Final Year Project) for the award of the degree in Bachelor of Agricultural Science

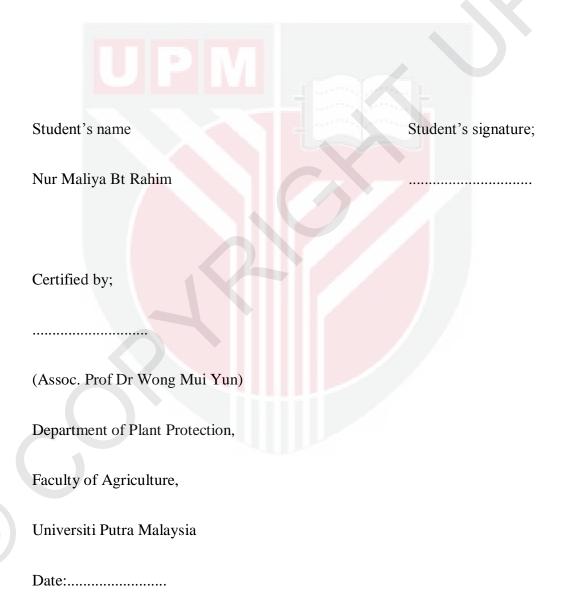
Faculty of Agriculture

Universiti Putra Malaysia

2015/2016

### CERTIFICATION

This project report entitled 'Effect of Antimicrobial Extracts of *Piper betle* and *Clinacanthus nutans* on *in vitro* growth of selected fungal pathogens' is prepared by Nur Maliya Bt Rahim and submitted to the Faculty of Agriculture in fulfilment of the requirement of PRT4999(Final Year Project) for the award of the degree in Bachelor of Agricultural Science.



#### ACKNOWLEDGEMENTS

First and foremost, I wish to express my thankfulness to God for His kindness and guidance. I would like to extend my great appreciation to my final year project supervisor, Assoc. Prof Dr Wong Mui Yun, for all the valuable guidance, advices, comments and encouragements in completing my final year project.

In addition, I would like to thank all Postgraduate students (Kai Wei, Solehah and Shila) and Laboratory Assistance of Makmal A (Mr Johari, Mr Nazri, Mrs. Asmalina, Mr. Sanin) and Makmal B (Mrs. Junaidah and Mr. Razali) from Department of Plant Protection, Faculty of Agriculture, UPM. Not forgotten my friends (Siti Zurain, Nur Liza Azah, Nur Shazmine, Nur Asifa, Intan, Maisarah and Siti Fairuz). I am particularly grateful for the assistance and guidance given by them. May He shower all of you with happiness and grant you success in all your works.

I would like to extend my special thanks to my beloved family especially my parents, Mr Rahim and Mrs Sharifah Zurhaini for their support, understanding and endless love throughout my study and upon completing this final year project. Not forgetting, all my friends and everybody who had offered valuable assistance, moral support and also guidance.

I am also taking this opportunity to sincerely apologize for any inconvenience while completing my final year project. May God keep us under His care and love.

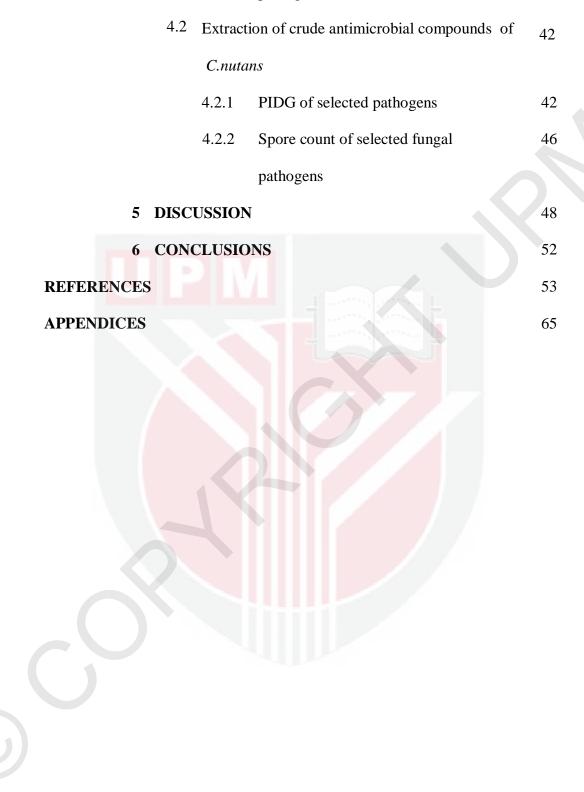
i

## TABLE OF CONTENTS

ACKNOWLEDGEMENTS	i					
TABLE OF CONTENTS	ii					
LIST OF APPENDICES	V					
LIST OF ABBREVIATIONS	vi					
ABSTRACT	vii					
ABSTRAK	ix					
CHAPTER 1 INTRODUCTION						
1.1 Sustainable agriculture	1					
1.2 Problem statements of the study	3					
1.3 Introduction of herbal plants	5					
1.4 Objectives of the study	6					
1.5 Hypothesis of the study	6					
2 LITERATURE REVIEW						
2.1 Method in controlling plant pathogen						
2.1.1 Cultural method	7					
2.1.2 Physical method	8					
2.1.3 Chemical method	8					
2.1.4 Biological control method	9					
2.2 Piper betle						
2.2.1 Taxonomy classification	10					
2.2.2 Distribution	11					
2.2.3 Morphological characteristic	11					
2.2.4 Uses	12					

	2.2.5	Chemical constituents	13
	2.2.6	Antimicrobial activity of <i>P. betle</i>	14
2.3	3 Clinac	Clinacanthus nutans	
	2.3.1	Taxonomy classification	16
	2.3.3	Morphological characteristic	17
	2.3.4	Uses	18
	2.3.5	Chemical constituents	19
	2.3.6	Antimicrobial activity of <i>C. nutans</i>	20
2.4	Plant f	ungal pathogens	
	2.4.1	Exserohilum rostratum	22
	2.4.2	Rhizoctonia solani	24
	2.4.3	Fusarium oxysporum f.sp cubense	27
	2.4.4	Ganoderma boninense	30
3 MATE	RIALS A	AND METHODS	
3.1	l Plant r	naterials	33
3.2	2 Fungal	cultures	33
3.3	8 Prepar	ation of plant materials	33
3.4	Antifu	ngal activity of <i>P. betle</i> and <i>C. nutans</i>	34
	agains	t selected pathogens	
3.5	5 Experi	mental design	35
4 RESUL	LTS		
4.1	Extrac	tion of crude antimicrobial	36
	compo	unds of <i>P.betle</i>	
	4.1.1	PIDG of selected pathogens	36
	4.1.2	Spore count of selected fungal	40

## pathogens



## LIST OF APPENDICES

Appendix		Page
1	Plate incubated with P. betle extract	62
2	Plate incubated with C. nutans extract	63
3	Spores of F.o.c and E. rostratum	64
4	SAS Data of G. boninense in P. betle extract	65
5	SAS Data of <i>E. rostratum</i> in <i>P. betle</i> extract	66
6	SAS Data of F.o.c in P. betle extract	67
7	SAS Data of <i>R. solani</i> in <i>P. betle</i> extract	68
8	SAS Data of <i>G. boninense</i> in <i>C. nutans</i> extract	69
9	SAS Data of <i>E. rostratum</i> in <i>C. nutans</i> extract	70
10	SAS Data of F.o.c in C. nutans extract	71
11	SAS Data of <i>R. solani</i> in <i>C. nutans</i> extract	72
12	SAS Data of number of spores of <i>F.o.c</i> in <i>P. betle</i> extract	73
13	SAS Data of number of spores of <i>E. rostratum</i> in <i>P. betle</i>	74
	extract	
14	SAS Data of number of spores of F.o.c in C. nutans	75
	extract	
15	SAS Data of number of spores of <i>E. rostratum</i> in <i>C</i> .	76
	nutans extract	

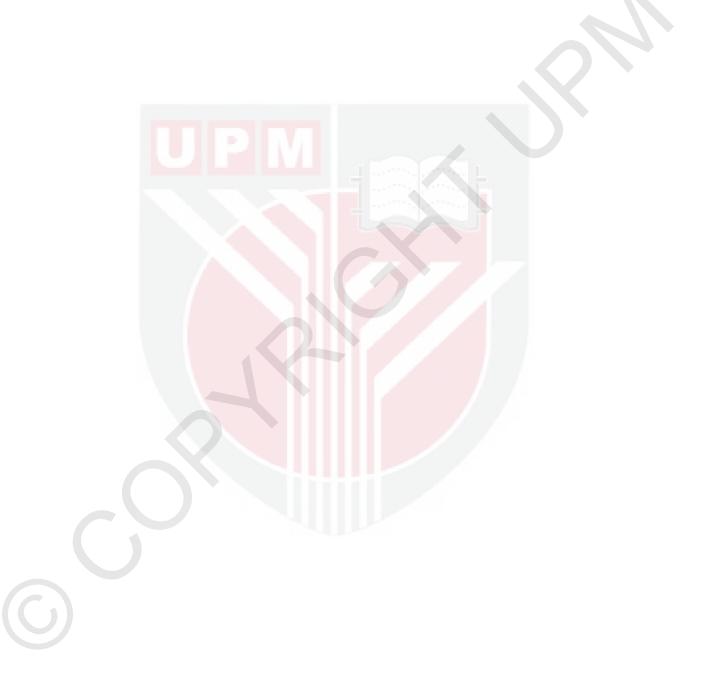
## LIST OF ABBREVIATIONS

	YHRC	:	Yellow Head Rhibdovirus
	DPPH	:	2,2-diphenyl-1-picrylhydrazyl
	NMR	:	Nuclear Magnetic Resonance
	IR	:	Infrared spectroscopy
	TBARS	:	Thiobarbituric acid reactive substances
	MIC	:	Minimal Inhibitory Concentration
	MFC	P	Minimal Fungicidal Concentration
	cm	:	centimetres
	mm		millimetre
	%		Percentage
	PIDG	:	Percentage of Inhibition Diameter Growth
	ml	:	millilitres
	L	:	Litre
	>	i	more than
	<		less than
	°C	÷	degree Celsius
	rpm	:	rate per minute
	h	:	Hour
	g	:	Gram
	kg	:	kilogram
	:	:	ratio of

#### ABSTRACT

Plant pathogenic fungi have been identified as the causal agent of various types of plant diseases in agricultural crops in Malaysia. Due to this problem, many methods of control had been introduced. The most common method is by using chemical control. However, application of chemical fungicides causes a major destruction to the environment because of high synthetic toxicity elements. The use of Piper betle and Clinacanthus nutans extracts as biopesticides is a good application towards sustainable agriculture. In view of this, this research was conducted 1) to extract crude antimicrobial compounds from P.betle and C. nutans and 2) to investigate the effectiveness of different concentrations of crude antimicrobial extracts on the *in vitro* growth of selected plant fungal pathogens. The selected fungal pathogens were Rhizoctonia solani, Ganoderma boninense, Fusarium oxysporum f sp cubense and Exserohilum rostratum. The leaves were washed, air dried at room temperature and ground using a blender. The leaf powder were soaked in methanol and loaded in orbital shaker. The mixture was filtered and the filtrate was concentrated using rotary evaporator to obtain dried extract. There were four concentrations (5, 10, 15 and 20%) used and compared with control (0%). Poisoned agar plates with P. betle extract showed inhibition on the mycelial growth and number of spores. The percent inhibition of diameter growth (PIDG) of F.o.c was 94.04% at 12<sup>th</sup> day, G. boninense 89.42% at 12<sup>th</sup> day, R. solani 82.84% at 4<sup>th</sup> day and *E. rostratum* 43.74% at 8<sup>th</sup> day. The number of spores produced  $(4x10^6)$  in 20% by F.o.c in P.betle extract is 969 while for R.solani is 178. Poisoned agar plates with C. nutans extract showed PIDG of E. rostratum was 64.42% at 8<sup>th</sup> day R. solani 58.86% at 4<sup>th</sup> day, G. boninense 27.48% at 12<sup>th</sup> day and F.o.c 10.36% at 12<sup>th</sup> day. The

number of spore produced  $(4x10^6)$  in 20% by *F.o.c* in *C.nutans* extract is 1723 while for *R.solani* is 38. Thus, this showed that the extracts of *P. betle* and *C. nutans* have the potential to be used as bio-fungicides as a safe alternative to synthetic fungicides.

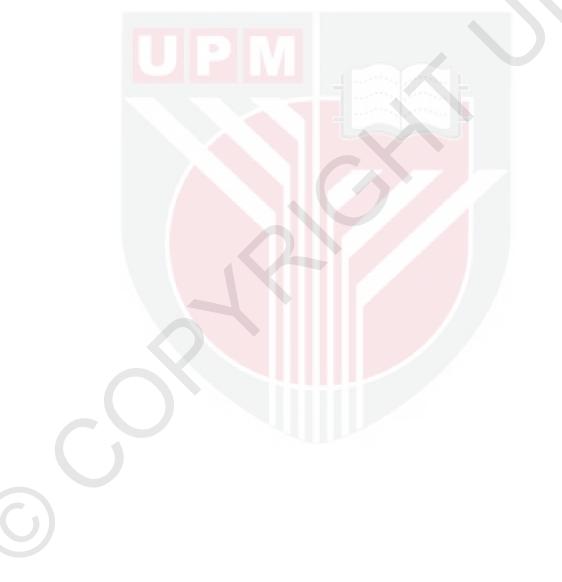


#### ABSTRAK

Kulat patogen tumbuhan telah dikenalpasti sebagai salah satu ajen yang menjadi penyebab kepada kepelbagaian penyakit bagi tumbuhan dalam sektor pertanian di Malaysia. Disebabkan permasalahan ini, terdapat banyak cara pengawalan penyakit yang telah diperkenalkan. Antara cara pengawalan yang selalu digunakan adalah dengan menggunakan bahan kimia .Walaubagaimanapun, teknik menggunakan racun kulat ke atas tanaman mengakibatkan kemusnahan kepada alam sekitar disebabkan oleh kandungan toksik yang sangat tinggi. Oleh itu, penggunaan ekstrak Sireh dan Belalai Gajah sebagai salah satu biopesticides adalah salah satu langkah yang terbaik dalam usaha kearah melestarikan sektor pertanian. Untuk itu, kajian ini dibuat adalalah untuk 1) mengekstrak komponen antimikrob daripada Sireh dan Belalai Gajah dan 2) mengkaji keberkesanan kepekatan ekstrak yang berlainan terhadap pertumbuhan kulat. Kulat yang dipilih bagi kajian ini ialah Rhizoctonia solani, Ganoderma boninense, Fusarium oxysporum f.sp cubense dan juga Exserohilum rostratum. Daun yang diambil telah dibersihkan dan dikeringkan pada suhu bilik dan dikisar menggunakan pengisar. Serbuk daun tersebut direndam didalam methanol dan diletakkan di atas orbital shaker. Larutan itu kemudiannya ditapis dan diletakkan pada rotary evaporator untuk mendapatkan ekstrak. Terdapat 4 kepekatan yang digunakan iaitu 5,10.15 dan 20% dan dibandingkan dengan 0% kepekatan (tanpa ekstrak). Agar yang telah diletakkan ekstrak Sireh menunjukkan pengurangan dalam pertumbuhan mycelia dan bilangan spora. Pengurangan dalam pertumbuhan mycelia F.o.c mencapai 94.04% pada hari ke-12, G. boninense 89.42% pada hari ke-12, R. solani 82.84% pada hari ke-4 and E. rostratum 43.74% pada hari ke-8. Bilangan spora  $(4x10^6)$  yang dihasilkan pada kepekatan 20% pula ialah 969 bagi F.o.c dan

C

178 bagi *R.solani*. Begitu juga dengan ekstrak Belalai Gajah. Pengurangan dalam pertumbuhan mycelia *E.rostratum* mencapai 64.42% pada hari ke-8, *R. solani* 58.86% pada hari ke-4 *F.o.c* 10.36% pada hari ke-12 dan *G. boninense* 27.48% pada hari ke-12. Bilangan spora (4x10<sup>6</sup>) yang dihasilkan pada kepekatan 20% pula ialah 1723 bagi *F.o.c* dan 38 bagi *R.solani*. Ini menunjukkan bahawa ekstrak Sireh dan Belalai Gajah boleh dijadikan sebagai bio-fungicide sebagai salah satu alternatif kepada racun kulat kimia sintetik



# CHAPTER ONE

#### **INTRODUCTION**

### **1.1** Sustainable agriculture

The term 'sustainability' is currently widely used in many aspects of our lives, and especially in agriculture because of the effect that certain crop production methods have on the environment (Hanson *et al.*, 2007). Sustainable agriculture is the management and the utilization of the agricultural ecosystem in a way that maintains its biological diversity, productivity, regeneration capacity, vitality and ability to function, so that it can fulfill today and in the future significant ecological, economic and social functions at the local, national and also global levels in a way that it does not give harm to the ecosystem (Lewandowski *et al.*, 1999).

The sustainability in agriculture has faced some of the most important challenges in recent years (Hanson *et al.*, 2007). The first major challenge is the increasing of human population. When this happen, increased demand for agricultural land and resources will occur. Next, over dependence on fossil energy and the increased monetary and also environmental costs of nonrenewable resources. Lastly, global climate change (Brown 2006; Diamond 2005), and globalization (Hanson *et al.*, 2007). These issues are challenging agriculturists to develop more sustainable management systems like no other time in history. To meet the food and nutritional needs of a growing population, agriculture sector will need to move beyond the past emphasis on productivity to improved public health, social wellbeing and a good environment (Hanson *et al.*, 2007). Also, it is crucial to find alternative way to control plant diseases which do not harm the environment and at the same time increase yield and also improve product quality (Batish *et al.*, 2006).

In recent years, the importance of sustainable agriculture has risen to become one of the most important issues in agriculture. With the occurrence of plant disease in agriculture sector, it limits the agricultural production. Controlling plant disease using classical pesticide will increase serious concern about the quality of environment, level of toxicity in products and also pesticide resistance in which becoming a factor for the need of alternative pest management techniques.



### **1.2** Problem statement of the study

Losses of agricultural products due to plant disease are increasing from day to day. It is recorded that in 2002, losses that were contributed by disease caused by pathogens and also other environmental factor was estimated about \$220 billion (Agrios, 2004). Increasing of losses by agriculture product from time to time will eventually not going to feed all the population in this world that will increase up to 8,500 million in 2020 (Agrios, 2004).

There is various disease that lead to the crop damage every year and among them fungal diseases are very common (Naidu *et al.*, 1981). Although the use of synthetic fungicides in plant disease control has shown to reduce the amount of plant disease in agriculture, several of these have been found to display side-effects in the form of carcinogenicity, detrimental effects and other residual toxicities. It is believed that antimicrobial agents are present in higher plants. Some recent researches on the antifungal activity of extracts of several higher plants have shown the possibility of this extract as natural antifungal agents to control plant diseases (Qureshi SM *et al.*, 1997).

The use of systemic fungicides simplifies that not many systemic fungicide is practical to be used. Furthermore, there is also the rise of concerns for the problems of fungicide insensitivity, residues on edible product and for tree crops (Bailey and Jeger, 1992). One approach might be to test the plants that are traditionally used for their antifungal activities as potential sources in a drug development (Lucy *et al.*, 1999)

The plant disease that is caused by fungal pathogen is usually controlled by application of chemical based antifungal compounds. For example, *P. azadirachtae* 

3

is sensitive to bavistin (Satesh MK, 1998). The chemical based pesticides are nonbiodegradable and extremely toxic (Yadav SK, 2010; Lakshmeesha TR *et al.*, 2013). They have various drawbacks in terms of genotoxicity (Bolognesi C, 2003), hepatotoxicity (Cecchi A *et al.*, 2012), reproductive disorders (Richard S *et al.*, 2005) and immunosuppression (Agarwal *et al.*, 2012). Frequent use of fungicides has led to the emergence of resistant strains. Furthermore, environmental pollution caused by excessive use of agrochemicals, has increased the public pressure to reduce the use of synthetic fungicides in agriculture (Manasi K *et al.*, 2014). Hence, the need for effective and safe alternative has increased. Concerns have been raised about both the environmental impact, and the effect to human health when using these synthetic compounds (Abad MJ *et al.*, 2007; Lakshmeesha Tr *et al.*, 2013).

Synthetic fungicides have been used for almost 20 years for the control of post-harvest diseases of tropical and subtropical fruit. Many microbial pathogens, however, have begun to develop resistance to the most widely used chemicals, so there is a need to develop new fungicides with improved performance and less potential environmental impact (Suhaila *et al.*, 1996).

### **1.3** Introduction of herbal plants

To date, many researchers had been working on identifying antimicrobial compound in herbal plants as it is free from the side effect caused by the synthetic chemicals. The use of synthetic fungicide has been found to display side-effects in the form of carcinogenicity, detrimental effects and other residual toxicities. The alternative choice, therefore would be the use of botanical fungicides, which are advocated to be largely non-phytotoxic, systematic and easily biodegradable in nature (Fawcett CH et al., 1970). Moreover, plant extracts could be an alternative to toxic fungicides for controlling plant pathogens since they are composed of various bioactive compounds such as alkaloids, flavonoids, glycosides, phenols, saponins, sterols and etc. (Lakshmeesha et al., 2013). Huge efforts are being made to isolate bioactive products from medicinal plants for their possible utility in development of plant based biopesticides (Helton LR, 1996). The antimicrobial activity of different plant extracts has been reported by many studies. It is believed that antimicrobial agents are present in higher plants. Some recent researches on antifungal activity of extracts of several higher plants have indicated the possibility of their exploitation as natural antifungal agents for control of plant diseases. Many herbal plants are able to inhibit microbial growth since they contain active compounds (Begum J et al., 2007). In the present study, we have evaluated the antimicrobial effects of the extract of two herbs, *Piper betle* and *Clinacanthus nutans* to control selected plant pathogen.

## **1.4 Objectives of the study**

The general objective of this research was to investigate the effect of *P. betle* and *C. nutans* extract on the growth of selected plant pathogen *in vitro* 

The specific objectives are:

- 1) To extract crude antimicrobial compounds from *P. betle* and *C. nutans*.
- 2) To investigate the effectiveness of different concentrations of crude antimicrobial extracts on the *in vitro* growth of selected plant fungal pathogens.

## **1.5** Hypothesis of the study

The hypothesis of this research was *P. betle* and *C. nutans* produce an antifungal compound which inhibits the growth of selected plant fungal pathogen.

#### REFERENCES

- A. Shahzad, A. Syarhabil, M. Awang Soh. 2014. A Review on Phytochemical Constituents and Pharmacological Activities of Clinacanthus nutans. International Journal of Pharmaceutical Sciences. 7(2)
- A.Sangeetha, R.Prabu, T.Naadeirmuthu ans K.C Clara .2014. In vitro screening of cycotoxic, antimicrobial and antioxidant activities of Clinacanthus nutans (Acanthaceae) leaf extract. Tropical Journal of Pharmaceutical Research (9); 1455-1461
- Abad MJ, Ansuategui M, Bermejo P. 2007. Active antifungal substances from natural sources. Arkivoc 7:116-145.
- Agarwal T., Singh R., Shukla A.D., Waris I., Gujrati A. 2012. Comparative analysis of antibacterial activity of four Piper betel varieties. Advances in Applied Science Research, 3 (2), pp. 698-705.
- Agrios, G.N. 2004. Losses caused by plant diseases. Plant Pathology. Elsevier, Oxford, UK.
- Ali H and Nadrajah K .2014. Evaluating the efficacy of Trichoderma spp and Bacillus substilis as biocontrol agents agains Magnaporthe grisea in rice.
   Australian Journal of Crop Science, 1324-1335.
- Asha M. Brunings. (24 September 2009). Exserohilum Leaf Spot on Tiger Grass. Plant Health Progress. Retrieved from:
- Bailey, J.A. & Jeger, M.J. (eds). 1992. Colletotrichum: Biology, Pathology and Control. CAB International., Wallingford UK.

- Bajpai V., Sharma D., Kumar B., Madhusudanan K.P. 2010. Profiling of Piper betle Linn. Cultivars by direct analysis in real time mass spectrometric technique.
  Biomedical Chromatography, 24(12), pp. 1283-1286.
- Batish, D.R., Arora, K., Singh, H.P., Kohli, R.K., 2006. Potential utilization of dried powder of Tagetes minuta as a natural herbicide for managing rice weeds. Crop Prot., in press, <u>doi:10.1016/j.cropro.2006.05.008</u>.
- Begum J, Yusuf M, Chowdhury JU, Saifulla K, Nural AM. 1996. Antifungal activity of forty higher plants against phytopathogenic fungi. Bangladesh J.Microbiol. 24:76-78
- Bissa S., Bohra A., Songara D., 2007. Traditions in oral hygiene: Chewing of betel (Piper betle L.) Leaves. Scientific correspondence. 92(1), pp. 24-28.
- Bolognesi C. 2003. Genotoxicity of pesticides: a review of human biomonitoring studies Mutat Res. 543: 251-272. Botany University of Karachi 75270
  Pakistan. 111 p.
- Brunings, S.A. 2009. Plant disease cause by Exserohilum rostratum. Plant Fungal Pathogens. 12:43-50
- C. Devjani and S. Barkha. 2011. Antimicrobial, Antioxidative and Anti-Hemolytic Activity of Piper betle Lead Extracts. International Journal of Pharmacy and Pharmaceutical Sciences
- Canwei S, S Sun, C. Jieling, C. Jianyi and E Zhou. 2014. Comparison of different methods or total RNA extraction from sclerotia of Rhizoctonia solani. Department of Plant Pathology. 17; 50-54
- Cecchi A, Rovedatti MG, Sabino G, Magnarelli GG. 2012. Environmental exposure to organophosphate pesticides: Assessment of endocrine disruption and hepatotoxicity in pregnant women. Ecotox Environ Safe. 80:280-287.

- Chandra T., Sadique J., Somasundaram S. 1987. *Effect of Elipta alba on inflammation and liver injury*. Fitoterapia, 58, pp.23-31.
- Chandra V., Tripathi S., Verma N.K., Singh D.P., Chaudhary S.K., Roshan A. 2011. *Piper betle: phytochemistry, traditional use & pharmacological activity-a review*, IJPRD. 4(4), pp. 216 -223.
- Chuah. 22<sup>nd</sup> October 2007. Ganoderma boninense. Retrieved on: mushroomobserver.org/name/show\_name/5087. [Accessed 16<sup>th</sup> August 2015]
- Cowan, M.M. 1999. *Plant products as antimicrobial agents*. Clinical Microbiology Reviews 10:564-582.
- D. Pradhan, Dr. K.A Suri, Dr.D.K Pradhan, P.Biswasroy .2013. Golden Heart of the Nature: Piper betle L. Journal of Pharmacognosy and Phytochemistry. 1 (6) 147-167
- Dampawan P. 1976. Studies of the chemical constituents of Clinacanthus nutans (Acanthaceae) and Zingiber cassumunar Roxb (Doctoral dissertation, Master thesis, Mahidol University, Bangkok, Thailand
- Dampawan Pimchit, Huntrakul Charus, Reutrakul Vichai, Raston CL, White A H. 1977. Constituents of Clinacanthus nutans and the crystal structure of LUP-20 (29)-ene-3-one. Journal Science Social Thailand;3:14-26.
- Davicino, R., M.A. Mattar, Y.A. Casali, S. Graciela, E. Margarita, and B. Micalizzi.
  2007. Antifungal activity of plant extracts used in folk medicine in Argentina.
  Revista Peruana de Biología 14:247-251.
- Demo, M.S., and M. Oliva. 2008. Antimicrobial activity of medicinal plants from South America. p. 152-164. In Watson, R.R., and V.R. Preedy (eds.) Botanical medicine in clinical practice. CABI International, Wallingford, UK.

diArk.*Fusarium oxysporum f.sp cubense*. A researcher for eukaryotic genome research. Retrieved on:

www.diark.org/diark/species\_list/Fusarium\_oxysporum\_f\_sp\_cubense\_tropi cal\_race\_4\_54006.[ Accessed 22<sup>nd</sup> August 2015]

Dr. J. Raja. Rhozoctonia Solani on rice. APS Publication. Retrieved from:

- El-Mahmood, A.M., J.H. Doughari, and N. Ladan. 2008. Antimicrobial screening of stem bark extracts of Vitellaria paradoxa against some enteric pathogenic microorganisms. African Journal of Pharmacy and Pharmacology 2:89-94
- F.R Sanderson .2005. An insight into spore dispersal of Ganoderma boninese in oil palm. Mycopathologia. 159;139-141
- Fawcett CH and Spencer DM. 1970. *Plant chemotherapy with natural products*. Annu Rev Phytopathol 8: 403-418
- Garret SD. 1965. *Toward biological control of soil-borne plant pathogens*. Pp14-16.
  In: Baker KF and Synder WC (Eds.), Ecology of Soil-borne Plant Pathogens.
  University of California Press, Berkeley and Los Angeles, 571.
- Ghaffar, A. 1988 . *Soil borne disease research centre*. Final Research report. Department of Botany University of Karachi 75270 Pakistan, 111 p
- Goldsberg,Persky and White. (17 October 2012). Quick Facts. Attorney at Law. Retrieved from:<u>http://www.gpwlaw.com/practice/contaminated-steroid-shots/</u>. [Accessed 16<sup>th</sup> August 2015]
- Guha P. 2006. *Betel Leaf: The Neglected Green Gold of India*, J. Hum. Ecol. 19(2), pp. 87-93.
- H. Xinqi, Chen L, Ran W, Shen Q, Yang X. 2011. Trichoderma harzianum strain SQR-T37 and its bio-organic fertilizer could control Rhizoctonia solani

*damping-off disease in cucumber seedling mainly by the mycoparasitism.* Appl Microbial Biotechnol 91;741-755.

- H.P Lim & Y.K Fong .2005. Possible genetic resistance in oil palm (Elais guineenssis Jacq.) to basal stem rot caused by Ganoderma boninese-prospect for future breeding. Mycopathologia 159;93-100
- Hanson J.D., Liebig M.A., Merrill S.D., Tanaka D.L., Krupingsky J.M., Scott D.E. 2007. Dynamic cropping systems; increasing adaptability amid and uncertain future. Agron.J. 99,939-943
- Helton LR. 1996. Folk medicine and health beliefs: an Appalachian perspective. J Cult Divers, 3: 123-128.
- Himratul Aznita W.H, Mohd Al Faisal N. and Fathilah A.R. 2011. Determination of the percentage inhibition of diameter growth (PIDG) of Piper betle crude aqueous extract against oral Candida species. Journal of Medicinal Plant Research 5(6); 878-884

http://www.apsnet.org/publications/imageresources/Pages/Rhizoctoniasolanion rice.aspx. [Accessed 22<sup>nd</sup> August 2015]

https://owl.english.purdue.edu/owl/resource/560/10/, [Accessed 21<sup>st</sup> August 2015]

Hushiarian R., Yusof N.A, Dutse S.W. 2013. Detection and control of Ganoderma boninense ; strategies and perspectives. SpringerPlus 2:555

J.Lucy, Y.Umi Kalsom, R.Nulit. 2011. Antifungal activity of selected plant leaves crude extract against a pepper anthracnose fungus, Collectotrichum capsici (Sydow) butler and bisby (Ascomycota: Phyllachorales). African Journal of Biotechnology 10(20); 4157-4165

- Kurucheve, V., Gerard, E.J. and Jayaraj, J. 1997. Screening of higher plant for fungitoxicity against Rhizoctonia solani in vitro. Indian Phytopathology 50 (2); 235-241
- Lakshmeesha TR, Sateesh MK, Vedashree S, Sofi MS, Umesha S. 2013. *Efficacy of botanicals on soybean seed-borne Fusarium equiseti*. VCFL Sciences 3:10-16.
- Lakshmeesha TR, Sateesh MK, Vedashree S, Sofi MS. 2013. Antifungal activity of some medicinal plants on Soybean seed -borne Macrophomina phaseolina. J Appl Pharma Science. 3: 84-87.
- Leina Mary Joseph, Tan Teck Koon, Wong Sek Man. 1998. Antifungal effects of hydrogen peroxide and preoxidase on spore germination and mycelia growth of Pseudocercospora species. Canadian Journal of Botany. 76(12), 2119-2124
- Lewandowski, I., Härdtlein, M., & Kaltschmitt, M. 1999. Sustainable crop production: definition and methodological approach for assessing and implementing sustainability. Crop Sciences (39), 184-193
- Lim H, Fong Y. 2005. Research on basal stem rot of ornamental palms caused by basidiospores from Ganoderma boninense. Mycopathologia. 159 (1);171-179
- Lucy H, Edgar JD. 1999. *Medicinal plants: a reemerging health aid*. Electronic Journal of Biotechnology 2, 56-70

Luo, R.A. 2012. Exserohilum rostratum. Plant pathogens. 12:24-25

- M.P Mithali, M.Kar and R.K Sahu. 2012. *Bioefficacy of some plant extracts on* growth parameters and control of disease in Lycopersicum esculentum. Pelagic Research Library, Asian Journal of Plant Sciences and Research; 2(2);129-142
- Manasa M, Vivek M.N, Yashoda Kambar, Ramesh Kumar K.A, Prashith Kekuda T.R. *Mineral content, Antimicrobial and Radical Scavenging Potential of*

*Caesalpinia mimosoides Lamk. (Caesalpiniaceae).* World Journal of Pharmaceutical Research. Volume 3,Issue 4; 1047-1063

- Manasi K. Bhagwat, Ajit G. Datar. 2014. *Antifungal activity of herbal extracts* against plant pathogenic fungi. Arch Phytopathology Plant Protect.
- Mandana Bimakr, Russly Abdul Rahmana, Farah Saleena Taipa, Ali Ganjloo, Liza Md Salleha, Jinap Selamat, Azizah Hamid, I.S.M. Zaidul. 2011. *Comparison of different extraction methods for the extraction of major bioactive flavonoid compounds from spearmint (Mentha spicata L.) leaves*. Food and bioproducts processing.67–72
- Mc Ginnis, M.R.,M.G. Rinaldi and R.E. Winn. 1986. Emerging agents of Phaeohyphomycosis; pathogenic species of Bipolaris and Exserohilum. J. Clin.Microbiol. 24;250-259
- Morrisey, J.P., and A. Osbourn. 1999. *Fungal resistance to plant antibiotics as a mechanism of pathogenesis*. Microbiology and Molecular Biology Reviews 63:708-724.
- Muhammad Shahzad Aslam, Muhammad Syarhabil Ahmad, Awang Soh Mamat.
   2015. A review on phytochemical constituents and pharmacological activities of Clinacanthus nutans. International Journal of Pharmacy and Pharmaceutical Sciences. Issue 2
- N A Anderson. 1982. The Genetics and Pathology of Rhizoctonia Solani Annual Review of Phytopathology Vol. 20: 329-347
- N.Vanketswarlu,T.Vijaya,S.Suresh Bhargav, K.Chandra mouli,D.Pragathi, D.Anitha, Vasu N.Reddy,A.Sreeramulu.2013. *In vitro inhibitory effects of medicinal plant extracts on sclerotium oryzae-a fungi causing stem root disease in paddy*. IJPBS. Volume 3 Issue 3

- Naidu AD & John VT. 1981. In vitro inhibition of rice fungal pathogens by extracts from higher plants. Int Rice Res Newsl 6 (5): 12-14
- Nur Ain Izzati M.Z., Abdullah F. 2008. Disease suppression in Ganoderma-infected oil palm seedling treated with Trichoderma harzianum. Plant Protection Science 101-107
- Periyanayagam K., Jagadeesan M., Kavimani S., Vetriselvan T., 2012. Pharmacognostical and Phyto-physicochemical profile of the leaves of Piper betle L. var Pachaikodi (Piperaceae)-Valuable assessment of its quality. Asian Pacific journal of tropical biomedicine; 2(2),pp. S506-S510.
- Pinelo, M., M.J. Manzocco, M.J. Nunez, and M.C. Nicoli. 2004. Solvent effect on quercetin antioxidant capacity. Food Chemistry 88:201-207.
- Ploetz RC, Pegg KG. 2000. Fungal diseases of root, corm and pseudostem. In: Jones DR, ed. Diseases of Banana, Abacá and Enset. Wallingford, UK: CAB International Publishing. 143–72.
- Ploetz, R.C., Haynes, J.L., Vazquez, A. 1999. *Responses of newbanana accessions in South Florida to panama disease*. Crop Prot. 18, 445–449.
- Prabhu M.S., Platel K., Saraswathi G., Srinivasan K., 1995. Effect of orally administered betel leaf (Piper betle Linn.) on digestive enzymes of pancreas and intestinal mucosa and on bile production in rats. Indian J Exp Biol., , 33(10), pp.752-756
- Qureshi SM, Rai K & Agrawal SC. 1997. In vitro evaluation of inhibitory nature of extracts of 18 plant species of chhindwara against 3 keratinophilic fungi.
  Hindustan Antibiot Bull. 39(1-4): 56-60.

Rahim, Maliya." Clinacanthus nutans. "JPEG file.

Rahim, Maliya." Piper betle. "JPEG file.

- Rahman MS & Anwar MN. 2006. Antifungal and cytotoxic activity of conessine isolated from the stem bark of Holarrhena antidysenterica. Bangladesh J Med Sci. 12(2): 116-119.
- Rahman S. A. 2009. Anti -ulcer effects of Piper betel, Solanum nigrum and Zingibercassumunar on ulceration induced by selected ulcerogens in rats. Master's thesis, University Putra Malaysia, pp. 4.
- Richard S, Moslemi S, Sipahutar H, Benachour N, Gilles -Eric Seraline. 2005. Differenential effects of glyphosate and roundup on human placental cells and aromatase. Environ Health Persp.113:716-720.
- Roozbeh Hushiarian. Nor Azah Yusof and Sabo Wada Dutse. 2013. Detection and Control of Ganoderma boninense; strategies and perspectives. Springer Plus.2;555
- S Arullappan, P Rajamanickam, N Thevar, CC Kodimani .2014. In Vitro Screening of Cytotoxic, Antimicrobial and Antioxidant Activities of Clinacanthus nutans (Acanthaceae) leaf extract. Tropica; Jpurnal of Pharmaceutical Research. Vol 13, No 9
- S. Direkbusarakom, L. Ruangpan, Y. Ezura, M. Yoshimizu (1998). Protective Efficacy of Clinacanthus nutans on yellow-head disease in black tiger shrimp (Penaeus monodon.) Fish Pathology, 33 (4)), pp. 401–404
- S. Direkbusarakom, Y. Ezura, M. Yoshimizu, A. Herunsalee. 1998. Efficacy of Thai Traditional Herb Extracts against Fish and Shrimp Pathogenic Bacteria. J.
   Fish Pathology;(4) 437- 441
- S.C Hwang and W.H Ko. Fusarium wilt of Giant Cavendish banana. Retrieved on: <u>www.apsnet.org/publications/imageresources/Pages/Jun 88-6-1.aspx</u>. [Accessed 22<sup>nd</sup> August 2015]

- Sakdarat S, Shuyprom A, Pientong C, Ekalaksananan T, Thongchai S. 2009. Bioactive constituents from the leaves of Clinacanthus nutans Lindau. Bioorg Med Chem;17:1857-60.
- Sateesh MK Vedashree S, Lakshmeesha TR, Sofi MS, Vedamurthy AB. 2013. Screening and assay of extracellular enzymes in Phomopsis azadirachtae causing die-back disease of neem. J Agricultural Technol. 9 :915-927
- Sateesh MK. 1998. *Microbiological investigations on die-back disease of neem* (*Azadirachta indica* A. Juss. PhD thesis, Mysore: University of Mysore
- Selitrennikoff, C.P. 2001. Antifungal proteins. Applied and Environmental Microbiology 67:2883-2894
- Senny O. Anti-cancer and Anti-inflammation herbs Sabah snake grass, 2009. Retrieve from: <u>http://sennyong.blogspot.com/2009/11/anti-cancer-herbs-and-anti-inflammation.html.</u> [Accessed 13<sup>th</sup> August. 2015]
- Sharma K. K., Saikia R., Kotoky J., Kalita J. C. and Das J. 2011. Evaluation of Antidermatophytic activity of Piper betle, Allamanda cathertica and their combination: An in vitro and in vivo stud. International Journal of PharmTech. Research, 3(2), pp. 644-651.
- Stone A.G, Scheurell SJ, Darby HD. 2004. Suppression of soilborne disease in field agriculture systems, organic matter management, cover cropping and other cultural practices In. Magdoff F. Weil RR(eds). Soil organic matter in sustainable agriculture. CRC, London
- Suhaila, M., S. Sizama, S.H.E.Sharkawy, A.M. Ali and S. Muid. 1996. Antimycotic screening of 58 Malaysian plants against plant pathogens. Pesticide science, 43(3): 259-264.

- Susanto A, Sudharto PS, Purba RY .2005. Enhancing biological control of basal stem rot disease (Ganoderma boninense) in oil palm plantations. PubMed 159(1);153-7
- Tangonan NG, Quebral FC. 1992. *Host Index of Plant diseases in the Philippines*, 2<sup>nd</sup> edn. Los Banos, Phillipines; University of the Phillipines at Los Banos.
- Trakranrungsie N., Chatchawanchonteera A., Khunkitti W. 2006. *Antidermatophytic Activity of Piper betle Cream*. Thai J Pharmacol.,28(3), pp. 16-20
- Tropicos. org. Tropicos | Name-Clinacanthus nutans (Burm. f.) Lindau. [online] Available From: http://www.tropicos.org/Name/50058310[Accessed 12<sup>th</sup> August 2015].
- Tuntiwachwuttikul P, Pootaeng-on Y, Phansa P, Taylor WC. 2004. Cerebrosides and a monoacylmonogalactosyglycerol from Clinacanthus nutans. Chem Pharm Bull;52(1):27-32.
- Tuntiwachwuttikul P, Pootaeng-on Y, Phansa P, Taylor WC. 2004. Cerebrosides and a monoacylmonogalactosyglycerol from Clinacanthus nutans. Chem Pharm Bull. 52(1):27-32.
- Usmanghani K., Saeed A., Alam M.T. 1997. Piper betle In, Indusyunic Medicine, (University of Karachi Press), Karachi. pp. 340-341.
- W.W Mc Call. 1981. *Treatment for control of soil organisms*. Hawaii Cooperative Extension Services. General Home Garden Series No.28
- Wallwork H. 1996. Cereal root and crown diseases, p.14-16. Perth, Australia;Kondinin Group
- Yadav SK. 2010. Pesticide applications-Threat to ecosystems. J Hum Ecol. 32; 37-45.

- Yash Paul Singh and Geeta Sumbali. 2011. Aflatoxin B, contamination in commercial varieties of apple and pear fruits infected with Aspergillus flavus
   Link ex Fries. Indian Phytopath 64(1):100-101
- Yoke Keong Yong, Jun Jie Tan, Soek Sin Teh. 2013. *Clinacanthus nutans Extracts Are Antioxidant with Antiproliferative Effect on Cultured Human Cancer Cell Lines*. Evidence-Based Complementary and Alternative Medicine, vol. 2013, Article ID 462751

