



UNIVERSITI PUTRA MALAYSIA

**ASSESSMENT OF FUNGAL AND BACTERIAL POPULATIONS ON
SELECTED VEGETABLE CROPS USING THE VERTICAL
HYDROPONICS SYSTEM**

MOHD ALIF OMAR MUSTAFFA

FP 2016 31

**ASSESSMENT OF FUNGAL AND BACTERIAL POPULATIONS ON
SELECTED VEGETABLE CROPS USING THE VERTICAL HYDROPONICS
SYSTEM**



MOHD ALIF OMAR BIN MUSTAFFA

**FACULTY OF AGRICULTURE
UNIVERSITI PUTRA MALAYSIA
SERDANG, SELANGOR DARUL EHSAN**

2015/2016



ASSESSMENT OF FUNGAL AND BACTERIAL POPULATIONS ON SELECTED
VEGETABLE CROPS USING THE VERTICAL HYDROPONICS SYSTEM

BY
MOHD ALIF OMAR BIN MUSTAFFA

A project report submitted to Faculty of Agriculture, Universiti Putra Malaysia, in fulfillment of the requirement of PRT 4999 (Final Year Project) for the award of the degree in Bachelor of Agricultural Science

DEPARTMENT OF PLANT PROTECTION
FACULTY OF AGRICULTURE
UNIVERSITI PUTRA MALAYSIA

2015/2016

All material contained within the project report, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the project report for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia. Copyright © Universiti Putra Malaysia

CERTIFICATION

This project report entitled “*Assessment of Fungal and Bacterial Populations on Selected Vegetable Crops using the Vertical Hydroponics System*” is prepared by Mohd Alif Omar Bin Mustaffa and submitted to the Faculty of Agriculture in fulfillment of the requirement of PRT 4999 (Final Year Project) for the award of the degree in Bachelor of Agricultural Science.

Student's name:

MOHD ALIF OMAR BIN MUSTAFFA

Student's signature:

.....

Certified by:

.....
(DR. KHAIRULMAZMI BIN AHMAD)

Supervisor,

Department of Plant Protection,

Faculty of Agriculture,

Universiti Putra Malaysia

Date:.....

ACKNOWLEDGEMENTS

بِسْمِ اللَّهِ الرَّحْمَنِ الرَّحِيمِ

Alhamdulillah, all praises and thanks to Almighty Allah S.W.T for the strength and Him blessings that enable me to complete my final year project successfully.

Firstly, I would like to express my deepest and sincerest appreciations with a heartfelt gratitude to my supervisor, Dr. Khairulmazmi Bin Ahmad for his endless supervision and constant support. Also, for his invaluable advice, constructive criticism, support, encouragement, patience and understanding that made this study and preparation of this thesis possible. Not forgetting to Associate Professor Dr. Yahya Bin Awang from Department of Crop Science for their guidance, suggestions and invaluable advice and comments throughout the duration of this study especially during carry out this project in the urban agriculture unit.

Also with deep sense of honour, I wish to extend my sincere gratitude to Laboratory Assistant of the Department of Plant Protection, Mrs. Junaina Jaafar, post-graduate student, Mrs. Siti Nur Sarah Shafiei, Ms. Hazirah Mohd Din and especially other members of post-graduate student in the Bacteriology Laboratory for their time and assistance in the preparation of materials that required for my final year project. Not forgetting to post-graduate students from Department of Crop Science, Mrs. Nur Alyani Shakri and Ms. Siti Nadhirah Sidi Ahmad for always being helpful and sharing information for me during carry out this project in the urban agriculture unit.

Finally, special thanks to my family members and beloved friends because supportive from beginning until the end of my final year project and also who involved in accomplish this project directly or indirectly.

CONTENTS

<i>Title Page</i>	i
<i>Certification</i>	ii
<i>Acknowledgement</i>	iii
<i>Contents</i>	iv – vi
<i>List of Plates</i>	vii – x
<i>List of Figures</i>	xi
<i>List of Appendices</i>	xii – xiv
<i>List of Abbreviations</i>	xv
<i>Abstract</i>	xvi
<i>Abstrak</i>	xvii – xviii
Chapter 1 : Introduction	
1.1 Overview	1 – 3
Chapter 2 : Literature Review	
2.1 Definition of Urban Agriculture	4
2.2 Hydroponics Culture	4 – 5
2.3 History of Hydroponics	5
2.4 Hydroponics Culture in Glasshouse	6
2.5 Types of Hydroponics System	6 – 7
2.6 Vertical Hydroponics System	7 – 8
2.7 Requirements of Hydroponics System	9
2.8 Plant Nutrients used in Hydroponics System	9 – 10
2.9 Economic Importance of Vegetable Crops	10 – 11

2.10	Cultivation of Selected Crops	11 – 15
2.11	Microorganisms of Fungi and Bacteria	16 – 17
2.12	Cultivation of Fungi and Bacteria	17
2.13	Morphological Identification of Fungi and Bacteria	18
2.14	Identification of Fungi	18 – 19
2.15	Identification of Bacteria	19
2.16	Culture Media	20
2.17	Serial Dilutions	20 – 21
2.18	Enumeration Methods	21 – 22

Chapter 3 : Methodology

3.1	Experimental Areas	23 – 24
3.2	Planting Materials	25
3.3	Experiment Materials	25 – 26
3.4	Samples	26 – 30
3.5	Preparation of Plant Sample Homogenates	31 – 32
3.6	Media	33
3.7	Procedure for Enumeration	33 – 34
3.8	Colony Forming Unit Counts	35
3.9	Identification of Fungi and Bacteria	35 – 36
3.10	Statistical Analysis	36

Chapter 4 : Results

4.1	Fungi Population in Root	37 – 38
4.2	Fungi Population in Water	38 – 40

4.3	Bacteria Population in Root	40 – 43
4.4	Bacteria Population in Water	43 – 46
4.5	Fungi and Bacteria Identification	46 – 55

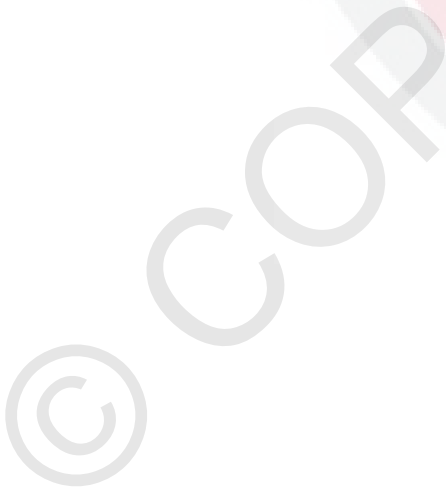
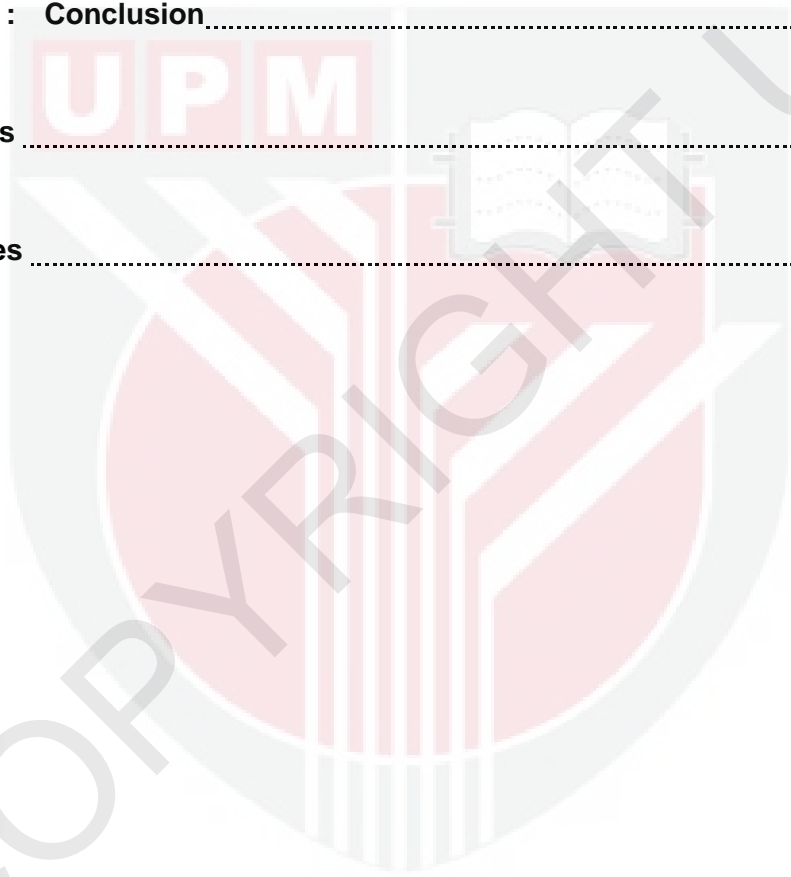
Chapter 5 : Discussion

5.1	Populations of Fungi and Bacteria on Selected Vegetable Crops	56 – 57
------------	------------------------------------------------------------------------	---------

Chapter 6 : Conclusion..... 58 – 59

References	60 – 68
-------------------------	---------

Appendices	69 – 86
-------------------------	---------



LIST OF PLATES

Plate	Title	Page
Plate 1	Vertical hydroponics structure with LED light	8
Plate 2	Vertical hydroponics system	8
Plate 3	<i>Ocimum americanum</i>	12
Plate 4	<i>Brassica oleracea</i> var. <i>alboglabra</i>	13
Plate 5	<i>Brassica chinensis</i> var. <i>parachinensis</i>	14
Plate 6	<i>Lactuca sativa</i>	15
Plate 7	Complex Agrobio, Faculty of Agriculture at Universiti Putra Malaysia (UPM)	23
Plate 8	Urban agriculture unit	24
Plate 9	Experimental laboratory	24
Plate 10	Samples of Lemon Basil, <i>Ocimum americanum</i>	27
Plate 11	Samples of Chinese Kale, <i>Brassica oleracea</i> var. <i>alboglabra</i>	28
Plate 12	Samples of Hong Kong Choy Sum, <i>Brassica chinensis</i> var. <i>parachinensis</i>	29
Plate 13	Samples of Lion Head Lettuce, <i>Lactuca sativa</i>	30
Plate 14	Preparation of crop samples homogenates by using root samples	31

Plate 15	Preparation of crop samples homogenates by using water samples	32
Plate 16	Observation of <i>Aspergillus fumigates</i> in PDA plate on root and water samples of <i>O. americanum</i> , <i>B. oleracea</i> var. <i>alboglabra</i> , <i>B. chinensis</i> var. <i>parachinensis</i> and <i>L. sativa</i> after 7 days incubation	47
Plate 17	Observation of <i>Aspergillus niger</i> in PDA plate on root and water samples of <i>O. americanum</i> , <i>B. oleracea</i> var. <i>alboglabra</i> , <i>B. chinensis</i> var. <i>parachinensis</i> and <i>L. sativa</i> after 7 days incubation	47
Plate 18	Observation of <i>Cladosporium herbarum</i> in PDA plate on root and water samples of <i>O. americanum</i> , <i>B. oleracea</i> var. <i>alboglabra</i> , <i>B. chinensis</i> var. <i>parachinensis</i> and <i>L. sativa</i> after 7 days incubation	48
Plate 19	Observation of <i>Curvularia lunata</i> in PDA plate on root and water samples of <i>O. americanum</i> , <i>B. oleracea</i> var. <i>alboglabra</i> , <i>B. chinensis</i> var. <i>parachinensis</i> and <i>L. sativa</i> after 7 days incubation	48
Plate 20	Observation of <i>Aspergillus fumigates</i> on root and water samples of <i>O. americanum</i> , <i>B. oleracea</i> var. <i>alboglabra</i> , <i>B. chinensis</i> var. <i>parachinensis</i> and <i>L. sativa</i> based on morphological identification method	49
Plate 21	Observation of <i>Aspergillus niger</i> on root and water samples of <i>O. americanum</i> , <i>B. oleracea</i> var. <i>alboglabra</i> , <i>B. chinensis</i> var. <i>parachinensis</i> and <i>L. sativa</i> based on morphological identification method	49

Plate 22	Observation of <i>Cladosporium herbarum</i> on root and water samples of <i>O. americanum</i> , <i>B. oleracea</i> var. <i>alboglabra</i> , <i>B. chinensis</i> var. <i>parachinensis</i> and <i>L. sativa</i> based on morphological identification method	50
Plate 23	Observation of <i>Curvularia lunata</i> on root and water samples of <i>O. americanum</i> , <i>B. oleracea</i> var. <i>alboglabra</i> , <i>B. chinensis</i> var. <i>parachinensis</i> and <i>L. sativa</i> based on morphological identification method	50
Plate 24	Observation colony pattern and subculture of <i>Bacillus subtilis</i> in new NA plate on root and water samples of <i>O. americanum</i> , <i>B. oleracea</i> var. <i>alboglabra</i> , <i>B. chinensis</i> var. <i>parachinensis</i> and <i>L. sativa</i> after 3 days incubation	51
Plate 25	Observation colony pattern and subculture of <i>Proteus vulgaris</i> in new NA plate on root and water samples of <i>O. americanum</i> , <i>B. oleracea</i> var. <i>alboglabra</i> , <i>B. chinensis</i> var. <i>parachinensis</i> and <i>L. sativa</i> after 3 days incubation	52
Plate 26	Observation colony pattern and subculture of <i>Staphylococcus aureus</i> in new NA plate on root and water samples of <i>O. americanum</i> , <i>B. oleracea</i> var. <i>alboglabra</i> , <i>B. chinensis</i> var. <i>parachinensis</i> and <i>L. sativa</i> after 3 days incubation	52
Plate 27	Observation colony pattern and subculture of <i>Streptococcus pyogenes</i> in new NA plate on root and water samples of <i>O. americanum</i> , <i>B. oleracea</i> var. <i>alboglabra</i> , <i>B. chinensis</i> var. <i>parachinensis</i> and <i>L. sativa</i> after 3 days incubation	53
Plate 28	Observation of <i>Bacillus subtilis</i> (Gram-positive) on root and water samples of <i>O. americanum</i> , <i>B. oleracea</i> var. <i>alboglabra</i> , <i>B. chinensis</i> var. <i>parachinensis</i> and <i>L. sativa</i> based on gram staining method	53

Plate 29	Observation of <i>Proteus vulgaris</i> (Gram-negative) on root and water samples of <i>O. americanum</i> , <i>B. oleracea</i> var. <i>alboglabra</i> , <i>B. chinensis</i> var. <i>parachinensis</i> and <i>L. sativa</i> based on gram staining method	54
Plate 30	Observation of <i>Staphylococcus aureus</i> (Gram-positive) on root and water samples of <i>O. americanum</i> , <i>B. oleracea</i> var. <i>alboglabra</i> , <i>B. chinensis</i> var. <i>parachinensis</i> and <i>L. sativa</i> based on gram staining method	54
Plate 31	Observation of <i>Streptococcus pyogenes</i> (Gram-positive) on root and water samples of <i>O. americanum</i> , <i>B. oleracea</i> var. <i>alboglabra</i> , <i>B. chinensis</i> var. <i>parachinensis</i> and <i>L. sativa</i> based on gram staining method	55

LIST OF FIGURES

Figure	Title	Page
Figure 1	Flow chart of fungi enumeration	34
Figure 2	Flow chart of bacteria enumeration	34
Figure 3	Assessment of fungi population using log ₁₀ CFU/mL in root samples for <i>O. americanum</i> , <i>B. oleracea</i> var. <i>alboglabra</i> , <i>B. chinensis</i> var. <i>parachinensis</i> and <i>L. sativa</i> within 4 weeks by surface plating method at 2, 4 and 7 days incubation	37 – 38
Figure 4	Assessment of fungi population using log ₁₀ CFU/mL in water samples for <i>O. americanum</i> , <i>B. oleracea</i> var. <i>alboglabra</i> , <i>B. chinensis</i> var. <i>parachinensis</i> and <i>L. sativa</i> within 4 weeks by surface plating method at 2, 4 and 7 days incubation	39 – 40
Figure 5	Assessment of bacteria population using log ₁₀ CFU/mL in root samples for <i>O. americanum</i> , <i>B. oleracea</i> var. <i>alboglabra</i> , <i>B. chinensis</i> var. <i>parachinensis</i> and <i>L. sativa</i> within 4 weeks by direct plating method at 1, 2, 3, 4, 5, 6 and 7 days incubation	41 – 43
Figure 6	Assessment of bacteria population using log ₁₀ CFU/mL in water samples for <i>O. americanum</i> , <i>B. oleracea</i> var. <i>alboglabra</i> , <i>B. chinensis</i> var. <i>parachinensis</i> and <i>L. sativa</i> within 4 weeks by direct plating method at 1, 2, 3, 4, 5, 6 and 7 days incubation	44 – 46

LIST OF APPENDICES

Annexure	Title	Page
Annexure 1	Assessment of fungi population using log ₁₀ CFU/mL in root samples for <i>Ocimum americanum</i> , <i>Brassica oleracea</i> var. <i>alboglabra</i> , <i>Brassica chinensis</i> var. <i>parachinensis</i> and <i>Lactuca sativa</i> within 4 weeks by surface plating method at 2, 4 and 7 days incubation	69
Annexure 2	Assessment of fungi population using log ₁₀ CFU/mL in water samples for <i>Ocimum americanum</i> , <i>Brassica oleracea</i> var. <i>alboglabra</i> , <i>Brassica chinensis</i> var. <i>parachinensis</i> and <i>Lactuca sativa</i> within 4 weeks by surface plating method at 2, 4 and 7 days incubation	70
Annexure 3	Assessment of bacteria population using log ₁₀ CFU/mL in root samples for <i>Ocimum americanum</i> , <i>Brassica oleracea</i> var. <i>alboglabra</i> , <i>Brassica chinensis</i> var. <i>parachinensis</i> and <i>Lactuca sativa</i> within 4 weeks by direct plating method at 1, 2, 3, 4, 5, 6 and 7 days incubation	71 – 72
Annexure 4	Assessment of bacteria population using log ₁₀ CFU/mL in water samples for <i>Ocimum americanum</i> , <i>Brassica oleracea</i> var. <i>alboglabra</i> , <i>Brassica chinensis</i> var. <i>parachinensis</i> and <i>Lactuca sativa</i> within 4 weeks by direct plating method at 1, 2, 3, 4, 5, 6 and 7 days incubation	73 – 74
Annexure 5	Results of ANOVA for assessment of fungi population in root samples on <i>Ocimum americanum</i> within 4 weeks by surface plating method	74
Annexure 6	Results of ANOVA for assessment of fungi population in root samples on <i>Brassica oleracea</i> var. <i>alboglabra</i> within 4 weeks by surface plating method	75

Annexure 7	Results of ANOVA for assessment of fungi population in root samples on <i>Brassica chinensis</i> var. <i>parachinensis</i> within 4 weeks by surface plating method	75
Annexure 8	Results of ANOVA for assessment of fungi population in root samples on <i>Lactuca sativa</i> within 4 weeks by surface plating method	75
Annexure 9	Results of ANOVA for assessment of fungi population in water samples on <i>Ocimum americanum</i> within 4 weeks by surface plating method	75
Annexure 10	Results of ANOVA for assessment of fungi population in water samples on <i>Brassica oleracea</i> var. <i>alboglabra</i> within 4 weeks by surface plating method	76
Annexure 11	Results of ANOVA for assessment of fungi population in water samples on <i>Brassica chinensis</i> var. <i>parachinensis</i> within 4 weeks by surface plating method	76
Annexure 12	Results of ANOVA for assessment of fungi population in water samples on <i>Lactuca sativa</i> within 4 weeks by surface plating method	76
Annexure 13	Results of ANOVA for assessment of bacteria population in root samples on <i>Ocimum americanum</i> within 4 weeks by direct plating method	76
Annexure 14	Results of ANOVA for assessment of bacteria population in root samples on <i>Brassica oleracea</i> var. <i>alboglabra</i> within 4 weeks by direct plating method	77
Annexure 15	Results of ANOVA for assessment of bacteria population in root samples on <i>Brassica chinensis</i> var. <i>parachinensis</i> within 4 weeks by direct plating method	77

Annexure 16	Results of ANOVA for assessment of bacteria population in root samples on <i>Lactuca sativa</i> within 4 weeks by direct plating method	77
Annexure 17	Results of ANOVA for assessment of bacteria population in water samples on <i>Ocimum americanum</i> within 4 weeks by direct plating method	77
Annexure 18	Results of ANOVA for assessment of bacteria population in water samples on <i>Brassica oleracea</i> var. <i>alboglabra</i> within 4 weeks by direct plating method	78
Annexure 19	Results of ANOVA for assessment of bacteria population in water samples on <i>Brassica chinensis</i> var. <i>parachinensis</i> within 4 weeks by direct plating method	78
Annexure 20	Results of ANOVA for assessment of bacteria population in water samples on <i>Lactuca sativa</i> within 4 weeks by direct plating method	78

LIST OF ABBREVIATIONS

UNDP	United Nations Development Programme
UN-HABITAT	United Nations Human Settlements Programme
FDTCP	Federal Department of Town and Country Planning Peninsular Malaysia
FAO	Food and Agriculture Organization
LED	Light-Emitting Diode
DEPI	Department of Environment and Primary Industry
PDA	Potato Dextrose Agar
NA	Nutrient Agar
PVC	Polyvinyl Chloride
°C	Degree Celsius
ml	Milliliter
cm	Centimeter
µm	Micrometer
CFU	Colony Forming Unit
cfu/mL	Colony forming unit per milliliter
Log ₁₀ cfu/mL	Colony forming unit per millilitre convert to log value
EC	Electrical Conductivity
LCB	Lactofenol cotton blue
SAS	Statistical Analysis System
ANOVA	Analysis of Variance
HSD	High Significant Different

ABSTRACT

The vertical hydroponics system is a method for plant cultivation that using the height of a hydroponics system. The role of microbes in plants growth is a very important. All plant surfaces have microbes on them, fungi and bacteria are among the microbes that successively colonize plants as they mature. Therefore, the objective of this study was to determine populations of fungi and bacteria on selected vegetable crops using vertical hydroponics system. The study was conducted to assess the high concentration on both populations of fungi and bacteria on Lemon Basil (*Ocimum americanum*), Chinese Kale (*Brassica oleracea* var. *alboglabra*), Hong Kong Choy Sum (*Brassica chinensis* var. *parachinensis*) and Lion Head Lettuce (*Lactuca sativa*). The experiment was conducted at the glasshouse in the urban agriculture unit at Faculty of Agriculture, Universiti Putra Malaysia. CFU/mL was be used in enumeration of fungi and bacteria populations using surface plating method and direct plating method from root and water sampling on selected vegetable crops within four week was assessed. Based on the results, the selected vegetable crops showed that there was significant different on fungi and bacteria populations. In fungi population, *B. oleracea* var. *alboglabra* and *L. sativa* has high concentration in root sampling and *O. americanum* also has high concentration in water sampling. For bacteria population, *L. sativa* has high concentration in the both of root and water sampling. The new finding in this study, the both populations of fungi and bacteria is more to give the beneficial microbes on selected vegetable crops and will be useful to plant growth in vertical hydroponics system. In conclusion, *Aspergillus fumigatus*, *Aspergillus niger*, *Cladosporium herbarum*, *Curvularia lunata*, *Bacillus subtilis*, *Proteus vulgaris*, *Staphylococcus aureus* and *Streptococcus pyogenes* were found from root and water sampling using morphological identification method.

ABSTRAK

Sistem hidroponik menegak adalah kaedah untuk penanaman tanaman yang menggunakan ketinggian sistem hidroponik. Peranan mikrob dalam pertumbuhan tanaman adalah sangat penting. Semua permukaan tanaman mempunyai mikrob, kulat dan bakteria adalah antara mikrob yang berturut-turut menjajah tanaman sehingga tanaman matang. Oleh itu, objektif kajian ini adalah untuk menentukan populasi kulat dan bakteria pada tanaman sayur-sayuran terpilih menggunakan sistem hidroponik menegak. Kajian ini telah dijalankan untuk menilai tumpuan populasi yang tinggi di kedua-dua populasi kulat dan bakteria pada tanaman basil (*Ocimum americanum*), kailan (*Brassica oleracea* var. *alboglabra*), sawi (*Brassica chinensis* var. *parachinensis*) dan salad (*Lactuca sativa*). Eksperimen ini telah dijalankan di rumah kaca dalam unit pertanian bandar di Fakulti Pertanian, Universiti Putra Malaysia. CFU/mL telah digunakan dalam penghitungan kulat dan bakteria dengan menggunakan kaedah penyaduran permukaan dan kaedah penyaduran terus dari persampelan akar dan air ke atas tanaman sayur-sayuran yang terpilih dalam tempoh empat minggu telah dinilai. Berdasarkan keputusan, tanaman sayur-sayuran terpilih menunjukkan bahawa terdapat perbezaan yang signifikan pada populasi kulat dan bakteria. Dalam populasi kulat, *B. oleracea* var. *alboglabra* dan *L. sativa* mempunyai tumpuan populasi yang tinggi dalam persampelan akar dan *O. americanum* pula mempunyai tumpuan populasi yang tinggi dalam persampelan air. Bagi populasi bakteria, *L. sativa* mempunyai tumpuan populasi yang tinggi dalam kedua-dua persampelan akar dan air. Penemuan baru dalam kajian ini, kedua-dua populasi bagi kulat dan bakteria lebih kepada memberi mikrob bermanfaat pada tanaman sayur-sayuran terpilih dan akan menjadi berguna kepada pertumbuhan tanaman dalam sistem hidroponik menegak. Kesimpulannya, *Aspergillus fumigatus*, *Aspergillus niger*,

Cladosporium herbarum, *Curvularia Lunata*, *subtilis Bacillus*, *Proteus vulgaris*, *Staphylococcus aureus* dan *Streptococcus pyogenes* ditemui dari persampelan akar dan air dengan menggunakan kaedah pengenalan morfologi.



CHAPTER 1

INTRODUCTION

1.1 Overview

According to UNDP (1996) reports that urban agriculture defined as an agriculture practice in activity of producing, processing and marketing of food and agricultural products in urban and suburban areas by using intensive production methods, natural resource and urban waste to yield a diversity of crops, fisheries and livestock. Besides, according to UN-HABITAT (1996) that the concept of urban agriculture has long existed if look at the development model of Machu Picchu in Peru around 1450 years ago, are designed to create a productive urban farm with farm houses in and around the city. Even, according to FDTCP (2012) also the urban agriculture as an important medium for continuously supply of local food, reduce urban poverty and improve the management of the urban environment (JPBD, 2012).

Today, the planting systems commonly used in urban agriculture are fertigation, hydroponics, aeroponics and roof-top farming. Through this system, planting vegetables on a small scale by individuals and communities can do to narrow the urban areas. In my project studies, I was selected vertical hydroponics system. According to Paraskevopoulou and co-workers (1995) assert that a vertical system is method for plant cultivation that using the height of a hydroponics system in addition to the ground space and this can increase the number of plants onwards enhance the yield. For example, vertical systems are mostly used in Japan, Australia, USA and European countries such as Italy (Shahla *et al.*, 2012).

The introduction of microorganisms population, according to Clements (1916) defined that the population which as a level of organization with have tight interactions between organisms that comprise a causal system and gives rise to emergence properties (Rangaswami, 1966). In addition, populations also are defined as multi-species assemblages. In which a contiguous environmental there are organisms that live together and interact with each other (Konopka, 2009). According to Pomeroy et al. (2007) and Findlay (2010) that they are mainly composed of fungi, bacteria, viruses, algae, protozoa and archaea (Liu *et al.*, 2013).

Furthermore, according to Paulitz (1997) and Utkhede *et al.* (2000) write that in the hydroponics system, fungi and bacteria are also very common carried in water supplies. Usually nutrients application and media provide an ideal breeding ground for many fungi and bacteria since they contain mineral elements, usually some moisture and are often warmed to just the right temperature for optimum microbial growth. The newly planted hydroponics system is very much like a soil garden and the mixtures of different microbes will compete with each other, just as they do in soil, and populations of different species will begin to dominate. But, many of the species are in fact beneficial microbes such as fungi and bacteria on the plant growth. Besides, many of the naturally occurring beneficial fungi and bacteria which colonise nutrient solutions are species of *Pseudomonas*, *Bacillus*, *Trichoderma*, and *Gliocladium virens* although a huge range of microbes may be in existence in different hydroponic systems. Because these types of species are known to have a natural suppressant action to many of the root diseases we encounter in hydroponics such as *Pythium*, *Fusarium* wilts and others (Morgan, 2008).

Presently, there is limited scientific researches have been done on populations of fungi and bacteria in the vertical hydroponics system. Therefore, this will be the first project report on assessment of fungal and bacterial populations on selected vegetable crops using vertical hydroponics system in Malaysia. The specific objective of this research was to determine populations of fungi and bacteria on selected vegetable crops using vertical hydroponics system. This study is to assess the high concentration on both populations of fungi and bacteria on Lemon Basil (*Ocimum americanum*), Chinese Kale (*Brassica oleracea* var. *alboglabra*), Hong Kong Choy Sum (*Brassica chinensis* var. *parachinensis*) and Lion Head Lettuce (*Lactuca sativa*).

REFERENCES

- Bhattarai, S.P.; Salvaudon, C. and Midmore, D.J. (2008). Oxygenation of the Rockwool Substrate for Hydroponics. Issue 49, Pages 29 – 30. Aquaponics Journal. <http://-acquire.cqu.edu.au:8080/vital/access/manager/Repository/cqu:4615>.
(accessed November 2015)
- Bokhtear, S.U. (2014). Medicinal Plants of Bangladesh: *Ocimum americanum* L. <http://www.mpbd.info/plants/ocimum-americanum.php>. (accessed January 2016)
- Bumbacco, A. (2013). Urban Agriculture: Improving Socio-Ecological Resilience and Wellbeing, Page 3. Senior Honours Thesis Project. https://uwaterloo.ca/environment_resource_studies/sites/ca.environment_resource_studies/files/uploads/file/Final%20Thesis%20Project%20%20Amy%20Bumbacco_2.pdf.
(accessed October 2015)
- Cavaglieria, L.; Orlando, J. and Etcheverry, M. (2007). Rhizosphere Microbial Community Structure at Different Maize Plant Growth Stages and Root Locations. Microbiological Research. Volume 164, Issue 4, 2009, Pages 391 – 399. <http://ac.els-cdn.com/S0944501307000596/1-s2.0-S0944501307000596-main.pdf> (accessed June 2016)
- DEPI (2013). Chinese flowering cabbage (Choy sum). Department of Environment and Primary Industries. ISSN 1329-8062. <http://agriculture.vic.gov.au/agriculture/horticulture/vegetables/vegetables-a-z/chinese-flowering-cabbage-choy-sum>.
(accessed January 2016)

- Dowsett, A.B. (2015). Introducing Microbes. Microbiology Online. <http://www.microbiologyonline.org.uk/about-microbiology/introducing-microbes>.
(accessed February 2016)
- Eraser, S. and Dimijian, G. (2015). Introduction of Fungi. Microbiology Online. <http://www.microbiologyonline.org.uk/about-microbiology/introducing-microbes/fungi>.
(accessed February 2016)
- Gottschal, J.C., Harder, W. and Prins, R.A. (2000). Principles of Enrichment, Isolation, Cultivation, and Preservation of Bacteria. Book Text View. <http://staff.unila.ac.id/mulyono/files/2011/11/Prinsip-Pengayaan-Isolasi-dan-Penyimpanan-Mikroba.pdf>.
(accessed April 2016)
- Gunstone, G. (1984). Growth Technology: Electrical Conductivity (EC). <http://www.growthtechnology.com/growtorial/what-is-conductivity/>. (accessed May 2016)
- Gunstone, G. (1984). Growth Technology: Target EC Ranges. <http://www.growthtechnology.com/growtorial/target-ec-ranges/>. (accessed May 2016)
- Haughton, J.R. (2015). Hydroponics Gardening Information. A brief Introduction to Hydroponics. Hydroponics Blog. <http://www.webring.org//rd?ring=verticalhydropon;id=15;url=http%3A%2F%2Fhydroponics-gardening-information.com%2FHydroponics.html>. (accessed October 2015)
- Heredia, N.A. (2014). Design, Construction, and Evaluation of a Vertical Hydroponic Tower. Page 2. Thesis Project. <http://digitalcommons.calpoly.edu/cgi/viewcontent.cgi?article=1108&context=braesp>. (accessed October 2015)

- Hillegass, C.K. (2012). Microbial Cultivation. CliffsNotes. <http://www.cliffsnotes.com/study-guides/biology/microbiology/microbial-cultivation-and-growth/microbial-cultivation>. (accessed February 2016)
- Hochmuth, G. and Hochmuth, R. (2011). Design Suggestions and Greenhouse Management for Vegetable Production in Perlite and Rockwool Media in Florida. Bulletin 327, Pages 2 – 13. UF/IFAS Extension, University of Florida. <http://edis.ifas.ufl.edu/pdffiles/CV/CV19500.pdf>. (accessed November 2015)
- Hoorweg, D. and Paul, M.F. (2008). Urban Agriculture for Sustainable Poverty Alleviation and Food Security, Page 10. Rome: Food and Agriculture Organization. http://www.fao.org/fileadmin/templates/FCIT/PDF/UPA_-WBpaper-Final_October_2008.pdf. (accessed October 2015)
- Hughey (2015). Principles of Epidemiology and Microbiology. Lesson 2: Public Health Microbiology. Section IV: Fungi. http://nursing411.org/Courses/MD0151_Principals_Epidem_Micro/2-18_Principals_Epidem_Micro.html. (accessed May 2016)
- Hunsinger, Kamp, B.A.; Kumala, C. and Bohm, R. (2005). Comparison of the Spread Plate Technique and the MPN-Technique for Determination of Variable Colony Counts in Quantitative Suspension Tests to Evaluate Bactericidal and Fungicidal Activity of Chemical Disinfectants. Journal, Volume 2, ISAH.
- Issarakraisila, M.; Ma, Q. and Turner, D.W. (2007). Photosynthetic and Growth Responses of Juvenile Chinese kale (*Brassica oleracea* var. *alboglabra*) and Caisin (*Brassica rapa* subsp. *parachinensis*) to Waterlogging and Water Deficit. Volume 111, Issue 2, Pages 107 – 113. <http://www.sciencedirect.com/science/article/pii/S0304423806004328>. (accessed January 2016)

- Jacqueline, C. (2012). Brief Summary: *Lactuca sativa* or Garden Lettuce. http://eol.org-/data_objects/20792742. (accessed January 2016)
- Jensen, M.H. (1997). Hydroponics Worldwide: a Technical Overview. Page 1, School of Agriculture, University of Arizona. <http://ag.arizona.edu/ceac/sites/ag.arizona.edu.ceac/files/Merle%20overview.pdf>. (accessed November 2015)
- Jensen, M.H. (2015). Hydroponics culture for the topics: Opportunities and alternatives. Food & Fertilizer Technology Center. <http://www.ffc.agnet.org/library.php?func=view&style=type&id=20110729175702>. (accessed October 2015)
- Jones, J.B. (1997). Hydroponics: A Practical Guide for the Soilless Grower. Second Edition, CRC Press. https://books.google.com.my/books?id=y_bKBQAAQBAJ&printsec=frontcover#v=onepage&q&f=false. (accessed November 2015)
- JPBD, Federal Department of Town and Country Planning Peninsular Malaysia (2012). Pertanian Bandar untuk Bandar Mampan. Blog Rasmi JPBD. <http://smp.townplanning.gov.my/blog/?tag=pertanian-bandar>. (accessed October 2015)
- Kamarudin, N.K.; Teh, C.B.S.; and Z.E.J. Hawa (2014). Modelling the Growth and Yield of Choy Sum (*Brassica chinensis* var. *parachinensis*) to include the Effects of Nitrogen and Water Stress. Malaysian Journal of Soil Science Vol. 18: 1 – 17. Malaysian Society of Soil Science. ISSN: 1394-7990. http://www.msss.com.my/-mjss/Full%20Text/vol18/1_Kamarudin.pdf. (accessed January 2016)
- Konopka, A. (2009). What is microbial community ecology? Vol. 3, pp 1223 – 1230. The ISME Journal. <https://www.pnl.gov/biology/research/mci/pubs/KonopkaISM-J.pdf>. (accessed October 2015)

- Kumar, A.M. (2011). CFU: Colony Forming Unit & Calculation. Bio-Resource: Technical Resources in Biotechnology. <http://technologyinscience.blogspot.my/2011/11/cfu-colony-forming-unit-calculation.html#.V0cyDCEtFzm>. (accessed April 2016)
- Larson, D.W. (2001). Plant Sciences: Economic Importance of Plants. <http://www.encyclopedia.com/doc/1G2-3408000121.html>. (accessed January 2016)
- Lee, S. and Lee, J. (2015). Beneficial Bacteria and Fungi in Hydroponic Systems: Types and Characteristics of Hydroponic Food Production Methods, Pages 207 – 213. Elsevier (Science Direct Article). <http://www.sciencedirect.com/science/article/pii/S0304423815301758>. (accessed October 2015)
- Liow, Y.K. (2013). Classification of Microbiology. Blogspot. <http://liowyingkee.blogspot.my/>. (accessed February 2016)
- Liu, L.; Yang, J.; Yu, X.; Chen, G. and Yu, Z. (2013). Patterns in the Composition of Microbial Communities from a Subtropical River: Effects of Environmental, Spatial and Temporal Factors. Volume 8, Issue 11, pp 1 – 10. PLoS ONE (Research Article). <http://journals.plos.org/plosone/article?id=10.1371/journal.pone.0081232>. (accessed October 2015)
- Liu, S. and Leung, B. (2010). Science Buddies: Interpreting Plates. http://www.sciencebuddies.org/science-fair-projects/project_ideas/MicroBio_Interpreting_Plates.-shtml. (accessed May 2016)
- Morales, P.J.P. (2007). Herbs and Leaf Crops: Cilantro, Broadleaf Ilantro and Vegetable Amarath. Soils, Plant Growth and Crop Production. <http://www.eolss.net/sample-chapters/c10/e1-05a-47.pdf>. (accessed January 2016)

Morgan, L. (2008). Nutrient Solution Problems. Article 5 – 3: Simply Hydroponics and Organics. http://www.simplyhydro.com/nutrient_solutions.htm.
(accessed October 2015)

Onanuga, A.O. (2013). A Hydroponic Approach to Evaluate Responses to Nutrients and Phytohormones in Cotton Plants (*Gossypium hirsutum* L.) Growth and Development. Pages 9 – 11, Online Thesis. <http://dalspace.library.dal.ca/bitstream/handle/10222/44678/Onanuga-Adebusoye-PHD-BIOL-Dec2013.pdf?sequence=1&isAllowed=y>. (accessed November 2015)

Pua, E.C. and Douglas, C.J. (2004). Biotechnology in Agriculture and Forestry: Brassica. Edited Edition, Pages 6 - 7, Springer. <https://books.google.com.my/books?id=66jvCAAQBAJ&printsec=frontcover#v=onepage&q&f=false>.
(accessed January 2016)

Rakow, G. (n.d.). Species Origin and Economic Importance of Brassica. http://www.springer.com/cda/content/document/cda_downloadaddocument/9783540202646-c1.-pdf?SGWID=0-0-45-115127-p23861264. (accessed January 2016)

Rangaswami, G. (1966). Agricultural Microbiology, pp 4 – 55. Asia Publishing House, London. ISBN 978-1-84407-668-0.

Redwood M. (2009). Agriculture in Urban Planning: Generating Livelihoods and Food Security, Pages 74. Earthscan and the International Development Research Centre (IDRC). ISBN 978-1-84407-668-0.

Resh, H.M. (2013). Hydroponic Food Production: a Definitive Guidebook for the Advanced Home Gardener and the Commercial Hydroponic Grower. Seventh Edition, CRC Press. <https://books.google.com.my/books?id=MbDMBQAAQBAJ-&printsec=frontcover#v=onepage&q&f=false>. (accessed November 2015)

Reynolds, J. (2009). Lab Manual: Identification of Fungi.

http://delrio.dcccd.edu/jreynolds/microbiology/2421/lab_manual/fungi_ID.pdf

(accessed June 2016)

Roberto, K. (2004). How to Hydroponics. Fourth Edition, Futuregarden Press.

<https://books.google.com.my/books?id=e5xLkJLTMygC&printsec=frontcover#v=onepage&q&f=false>. (accessed November 2015)

Rosberg, A.K. (2014). Dynamics of Root Microorganisms in Closed Hydroponic

Cropping Systems. Doctoral Thesis. http://pub.epsilon.slu.se/11393/1/rosberg_a-k_140804.pdf (accessed June 2016)

Shahla, M.; Mohsen, K.; Rouhangiz, N. and Toktam, S.T. (2012). Vertical Mobile

Planting System Consistent with the Pattern of Solar Radiation and Effects of System on Light Exposure and Growth of Gerbera Cut Flowers (*Gerbera jamesonii* cv. Antibes), in Greenhouse Culture. Vol. 8(4), pp 1461 – 1468, ISSN 1686-9141. Journal of Agricultural Technology. http://www.ijat-aatsea.com/pdf/v-8_n4_12_July/26_IJAT_201-2_-8_4_-Plant%20Science-_Shahla_Mahdavi.pdf.

(accessed October 2015)

Smith, A.C. and Hussey, M.A. (2013). Gram Stain Protocols. American Society for

Microbiology. <http://www.microbelibrary.org/component/resource/gram-stain/288-6-gram-stain-protocols> (accessed April 2016)

- Tellez, T.L.I and Merino, F.C.G (2012). Nutrient Solutions for Hydroponic Systems. Pages 1-2. <http://cdn.intechopen.com/pdfs-wm/33765.pdf>. (accessed February 2016)
- Tyson, R.; Hochmuth, R. and Cantliffe, D.J. (2009). Hydroponic Vegetable Production in Florida. Page 4. UF/IFAS Extension, University of Florida. <https://edis.ifas.ufl.edu/pdf/HS/HS40500.pdf>. (accessed November 2015)
- Vallance, J.; Deniel, F.; Le Floch, G.; Guerin-Dubrana, L.; Blancard, D. and Rey, P. (2011). Pathogenic and Beneficial Microorganisms in Soilless Cultures. Review Article, Pages 191 - 203. <https://hal.archives-ouvertes.fr/hal-00930473/document> (accessed June 2016)
- Vidaver, A.K. and Lambrecht, P.A. (2004). Bacteria as Plant Pathogens. The Plant Health Instructor. DOI: 10.1094/PHI-I-2004-0809-01. <http://www.apsnet.org/edcenter/intropp/PathogenGroups/Pages/Bacteria.aspx>. (accessed February 2016)
- Vlab.amrita.edu. (2011). Isolation and Identification of Two Bacterial Unknowns. vlab.amrita.edu/?sub=3&brch=76&sim=1109&cnt=1 (accessed June 2016)
- Vlab.amrita.edu. (2011). Gram Stain Technique. vlab.amrita.edu/?sub=3&brch=73&sim=208&cnt=2 (accessed June 2016)
- Vlab.amrita.edu. (2011). Slide Culture Technique for Fungi. vlab.amrita.edu/?sub=3&brch=76&sim=693&cnt=2 (accessed June 2016)
- Waggoner, B. and Brian R.S. (2006). Fungi: More on Morphology. <http://www.ucmp.berkeley.edu/fungi/fungimm.html> (accessed April 2016)

- Watanabe, M.; Tsutsumi, F.; Lee, K.; Sugita-Konishi, Y.; Kumagai, S.; Takatori, K.; Hara-Kudo, Y. and Konuma, H. (2010). Enumeration of Fungi in Fruits by the Most Probable Number Method. *Journal of Food Science*. Volume 75, Issue 9. <http://www.ncbi.nlm.nih.gov/pubmed/21535611>. (accessed April 2016)
- Yu, J.Q.; Lee, K.S. and Matsui, Y. (1993). Effect of the Addition of Activated Charcoal to the Nutrient Solution on the Growth of Tomato in Hydroponic Culture. Volume 39, Issue 1, Pages 13 – 22. *Soil Science and Plant Nutrition*. <http://www.tandfonline.com/doi/pdf/10.1080/00380768.1993.10416970>. (accessed November 2015)
- Yu, J. (2010). Identification of Fungi and Bacteria Associated With Internally Discolored Horseradish Roots. https://www.ideals.illinois.edu/bitstream/handle/2-142/16968/Yu_Junmyoung.pdf?sequence=3 (accessed April 2016)
- Zarna (2012). Enumeration. *Anti Essays*. <http://www.antiessays.com/free-essays/Enumeration-296344.html> (accessed April 2016)