

ISOLATION AND CLONING OF THE PROMOTER SEQUENCE OF VITAMIN E BIOSYNTHESIS GENE (HOMOGENTISATE PHYTYLTRANSFERASE) FROM OIL PALM

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BY MOHD SAIFUL BIN YAMBA

A project submitted to Faculty of Agriculture, Universiti Putra Malaysia, in fulfillment of the requirement of PRT 4999 (Final Year Project) for the award of the degree of

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CERTIFICATION

This project paper entitled **"Isolation and Cloning of The Promoter Sequence of Vitamin E Biosynthesis Gene (Homogentisate Phytyltransferase) from Oil Palm"** is prepared by Mohd Saiful Bin Yamba and submitted to the Faculty of Agriculture in fulfillment of the requirement of PRT 4999 (Final Year Project) for the award of the degree of Bachelor of Horticultural Science.



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ABBREVATIONS

CI	Chloroform isoamyl
CI	Chloroform isoamylalcohol
СТАВ	Cetyltrimethyl Ammonium Bromide
DNA	Deoxyribonucleic acid
FAO	Food and Agriculture Organization of the
	United Nations
GFP	Gentisate Phytyltranferase
НРТ	Homophytyltranferase
PCI	Phenol chloroform isoamyl
PCR	Polymerase Chain Reaction
РЕ	Phycoerythrin
SDSC	San Diego Supercomputer Centre
ТЕ	Tris-EDTA

ABSTRACT

The oil palm tissue contains vitamin E in the form of tocotrienols and tocopherols. The combination of prenyldiphosphate and homogentisic acid (HGA) is the first committed step in vitamin E production. The reaction is catalyzed by homogentisate phytyltransferase (HPT). The oil palm genomic sequence containing the HPT gene is used to study the expression of HPT gene in oil palm tissues. The objective of this study is to isolate and clone the HPT promoter sequence. To obtain the result from this study, some molecular work was done, including DNA extraction by DNA extraction kit and the cetyltrimethylammonium bromide (CTAB) method, designing the primer for HPT promoter isolation, running the polymerase chain reaction (PCR) to get the HPT promoter sequences. After obtaining the amplified HPT promoter sequence, the HPT promoter was introduced into an entry vector and then to destination vector forming the expression clone.

ABSTRAK

Tisu kelapa sawit mengandungi vitamin E dalam bentuk tokotrienol dan tokoferol. Gabungan asid prenyldiphosphate dan homogentisic acid (HGA) adalah langkah pertama yang berlaku dalam pengeluaran vitamin E. Tindak balas ini dimangkinkan oleh homogentisate phytyltransferase (HPT). Genom kelapa sawit mengandungi urutan gen HPT. Setiap gen dikawal oleh urutan promoter. Promoter gen HPT akan digunakan untuk mengkaji pengekspresan HPT dalam tisu kelapa sawit. Objektif kajian ini adalah untuk mengasingkan dan mengklonkan urutan promoter HPT. Untuk mendapatkan hasil dari kajian ini, kaedah biologi molekul telah digunakan, termasuk pengekstrakan DNA oleh kit pengekstrakan DNA atau dengan kaedah cetyltrimethylammonium bromida (CTAB), mereka bentuk pencetus untuk pemencilan HPT promoter, menjalankan tindak balas rantai polymerase (PCR) untuk mendapatkan urutan promoter HPT dan penulinan gel agarose serpihan DNA promoter. Selepas mendapat urutan promoter HPT, promoter HPT yang telah diamplifikasikan dimasukan ke dalam vektor dan seterusnya dipindahkan ke vektor destinasi untuk membentuk klon pengekspresan.

CHAPTER 1

INTRODUCTION

Oil palm originated from Africa. There are two well-known species of oil palm, *Elaeis guineensis* and *Elaeis oleifera* (Arunachalam, 2012). Oil palm also has different types of variety. The tenera variety from *Elaeis guineensis* is the well known commercial variety. It is now planted in large hectarage in certain countries in South East Asia. Oil palm industries continue to expand in the South East Asian countries, especially in Indonesia and Malaysia. According to Food and Agriculture Organization of the United Nation (FAO) in 2013, Indonesia is the leader in oil palm fruit production in the world, followed by Malaysia. Increasing progress in world oil palm industry causes increases in the production of oil palm-based edible oils.

Vitamin E is synthesized through plant secondary metabolic pathway (Eitenmiller & Lee, 2005). Plant vitamin E consists of tocotrienols and tocopherols (Hofius and Sonnewald, 2003). Vitamin E has specific and various functions in the human body and plant tissues. It was first discovered during research on leafy vegetables (Musa, 2012). Vitamin E can be synthesized naturally in plant and also by chemical synthesis to form artificial vitamin E. The research on vitamin E focuses on extraction from vegetable oils such as from soy bean and oil palm. They are several steps in the biosynthesis pathway to produce vitamin E (Collakova & DellaPenna, 2003b). The pathway contains several biosynthesis enzymes (Collakova & DellaPenna, 2003b).

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Plant biotechnology research is advancing aggressively including in the field of plant genetic engineering. Genetic engineering is the way to improve the plant characteristics by modification of the gene of interest and expressing the introduced foreign gene in plant to form transgenic plant (Izadfard, 2009). Genetic engineering can produce hybrid and new cultivar without having to wait for a long time unlike the traditional breeding approach (Izadfard, 2009). Transient gene expression and promoter reporter gene analysis are part of plant genetic engineering.

Promoter sequences of the gene is located in front of gene sequence. Promoter used in this study is homogentisate phytyltranferase (HPT) promoter. HPT is one of the enzyme involved in the biosynthesis of tocopherol. In this study, there are some experimental work including DNA extraction, agarose gel electrophoresis, designing of primer, PCR and gel extraction in order to isolate the promoter sequence.

After the HPT promoter sequence is isolated from oil palm, the promoter will be cloned to form expression clone by a Gateway Cloning strategy. Gateway Cloning basically has two reactions which are BP reaction, where a the promoter sequence and a donor vector is combined and introduced into an entry clone. The second is LR reaction, where the entry clone is mixed with a destination vector to form expression clone.

The objectives of this study are:

- 1. To isolate HPT promoter sequence from oil palm DNA.
- 2. To clone HPT promoter using a Gateway Cloning strategy.

REFERENCES

Arunachalam, V. (2012). Oil Palm. Genomics of Cultivated Palms, 29-48.

- Basu, C., Kausch, A. P., Luo, H., & Chandlee, J. M. (2003). Promoter analysis in transient assays using a GUS reporter gene construct in creeping bentgrass (Agrostis palustris). *Journal of Plant Physiology*, 160(10), 1233–1239.
- Collakova, E., & DellaPenna, D. (2003a). Homogentisate phytyltransferase activity is limiting for tocopherol biosynthesis in Arabidopsis. *Plant Physiology*, *131*(2), 632–642.
- Collakova, E., & DellaPenna, D. (2003b). The role of homogentisate phytyltransferase and other tocopherol pathway enzymes in the regulation of tocopherol synthesis during abiotic stress. *Plant Physiology*, *133*(2), 930–940.
- Dabrowska, G., Mierek-Adamska, A., & Goc, A. (2012). Plant metallothioneins: Putative functions identified by promoter analysis in silico. *Acta Biologica Cracoviensia Series Botanica*, 54(2), 109–120.
- Eitenmiller, R., & Lee, J. (2005). *Vitamin E Food Chemistry*, New York: Marcel Dekker, Inc.
- Elbing, K., & Brent, R. (2002). CHAPTER 1 Escherichia coli, Plasmids, and Bacteriophages. *Current Protocols in Molecular Biology*, 59–79.
- Hanan, F. (2012). Determination of the genomic structure of homogentisate geranylgeranyl transferase gene from different oil palm germplasm material. Universiti Putra Malaysia.
- Herbers, K. (2003). Vitamin production in transgenic plants. *Journal of Plant Physiology*, *160*(7), 821–829.
- Hernandez-Garcia, C. M., & Finer, J. J. (2014). Identification and validation of promoters and cis-acting regulatory elements. *Plant Science*, 217-218, 109–119.
- Izadfard, A. (2009). Universiti Putra Malaysia Functional Analysis of the Oil Palm Metallothioninelike Gene Promoter Using Transient Expression Assay.
- Kong, S. L., Abdullah, S. N. A., & Ho, C. L. (2013). Molecular Cloning, Characterization and In Silico Promoter Analysis of Tocopherol Biosynthetic Genes from the Oil Palm. Universiti Putra Malaysia.

- Mène-Saffrané, L., & DellaPenna, D. (2010). Biosynthesis, regulation and functions of tocochromanols in plants. *Plant Physiology and Biochemistry*, 48(5), 301–309.
- Musa, I. (2012). Protective effects of palm vitamin E on Glutamate-induced injury of astrocytes.
- Nakagawa, T., Ishiguro, S., & Kimura, T. (2009). Gateway vectors for plant transformation. *Plant Biotechnology*.
- Puah, C. W., Choo, Y. M., Ma, A. N., & Chuah, C. H. (2007). The effect of physical refining on palm vitamin E (tocopherol, tocotrienol and tocomonoenol). *American Journal of Applied Sciences*, 4(6), 374–377.
- Seong, L. M. (2011). CDNA Cloning And Expression Analysis Of Tocopherol Cyclase Gene From Various Oil Palm Tissues (Elaeis guineensis and Elaeis oleifera).
- Smith, D. S., Maxwell, P. W., De Boer, S. H., Food, C., Agency, I., Smith, D. S., ... De Boer, S. H. (2005). Comparison of several methods for the extraction of DNA from potatoes and potato-derived products. *Journal of Agricultural and Food Chemistry*, 53(26), 9848–9859.