



UNIVERSITI PUTRA MALAYSIA

**RESPONSE OF BROILER CHICKENS TO DIETARY
SUPPLEMENTATION OF L-GLUTAMINE AND L-GLUTAMATE UNDER
STRESSFUL CONDITIONS**

MAJID SHAKERI

ITA 2014 5



**RESPONSE OF BROILER CHICKENS TO DIETARY
SUPPLEMENTATION OF L-GLUTAMINE AND L-GLUTAMATE
UNDER STRESSFUL CONDITIONS**

By

MAJID SHAKERI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirement for the Degree of Master of Science**

June 2014

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

I dedicate this thesis to my beloved parents and my brothers for their unconditional love and support



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in
fulfilment of the requirement for the degree of Master of Science

**RESPONSE OF BROILER CHICKENS TO DIETARY
SUPPLEMENTATION OF L-GLUTAMINE AND L-GLUTAMATE
UNDER STRESSFUL CONDITIONS**

By

MAJID SHAKERI

June 2014

Chairman: Professor Zulkifli Idrus, PhD

Institute: Institute of Tropical Agriculture

High stocking density (HSD) and delay in placement are practices that may impair growth performance and well-being of broiler chickens. Dietary supplementation with L-glutamine and L-glutamate (LG) has been suggested to improve poultry performance under stressful conditions. Thus, this research was conducted to evaluate the effect of LG supplementation on performance, gut morphology, physiology and well-being of broiler chickens subjected to different stocking densities and posthatch holding times. In Chapter 3, one-day-old broiler chicks were stocked either at 10 birds/m² or 15 birds/m² and assigned to four dietary treatments namely (i) basal diet (ii) basal diet supplemented with 0.5% LG from 1 -21 days of age (LG3), (iii) basal diet supplemented with 0.5% LG for from 1 – 42 days of age (LG6), (iv) and basal diet supplemented with virginiamycin from 1 – 42 days of age (VM). The results indicated that neither LG3 nor LG6 had beneficial effects on growth performance, duodenal villi and crypt depth, gut *E. coli* and *Lactobacilli counts*, and incidence of foot pad dermatitis. VM improved growth performance, villi length and decreased the population of *E. coli*. Irrespective of LG and VM supplementation, the high stocking density impaired growth performance, and villi length, and increased incidence of foot pad dermatitis. The high stocking density elevated serum levels of ceruloplasmin (CP), α 1 acid glycoprotein (AGP), corticosterone (CORT), and heterophil / lymphocyte ratios (HLR) as compared to the low stocking density. In Chapter 4, broiler chicks were subjected to either immediate placement or delayed placement (24 hours after arrival). Chicks from each placement time were supplemented with LG at 1% from 1 -21 day of age or unsupplemented diet. Chicks subjected to 24 hours of delayed placement had larger yolk sac, and poorer early weight gain (during 1 -21 days of age) than their control counterparts. However, the final weight gain and feed conversion ratios (FCR) of the chicks subjected to immediate and delayed placement were not significantly different. Delayed placement had no significant effect on gut morphology, CP, AGP, OVT, and duodenal heat shock protein (HSP) 70 density at 21 days of age.

Supplementing birds with LG increased duodenal villi length and crypt depth at 21 days of age, and improved final weight gain, FCR and survivability when compared to controls. The CP, AGP, OVT and HSP 70 density of chicks subjected to delayed placement were elevated when compared to controls. The present findings suggested that LG supplementation at 1% from day 1 – 21 days of age can improve weight gain, FCR, and gut morphology of chicks subjected to 24 hours of delayed placement. Supplementing chicks with LG may also aid chickens to cope with stresses through elevation in APP and HSP 70.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai
memenuhi keperluan untuk ijazah Master Sains

**RESPONS AYAM PEDAGING KEPADA PEMAKANAN TAMBAHAN
L- GLUTAMINA DAN L- GLUTAMAT DALAM KEADAAN TERTEKAN**

Oleh

MAJID SHAKERI

Jun 2014

Pengerusi: Professor Zulkifli Idrus, PhD

Institut: Institut Pertanian Tropika

Tahap kepadatan tinggi (HSD) dan kelewatan dalam penempatan anak ayam adalah dua faktor penting yang boleh menyebabkan tindak balas tekanan fisiologi dan menjelaskan prestasi pertumbuhan dan fungsi imun bagi ayam . Baru-baru ini , L- glutamina dan L- glutamat (LG) telah dicadangkan untuk meningkatkan prestasi ayam daging di bawah keadaan tekanan. Oleh itu, kajian ini dijalankan untuk menilai dua tahap pemakanan LG iaitu 0.5% dan 1% ke atas prestasi pembesaran , morfologi usus, pengeluaran antibodi, duodenum , heat shock protein (HSP) 70 penyataan, dan parameter darah tahap serum protein fasa kritikal (ovotransferrin , ceruloplasmin dan asid Glycoprotein alfa 1), kepekatan plasma corticosterone dan nisbah heterophil / limfosit) dalam ayam daging tertakluk kepada kepadatan bekalan dan penempatan tertangguh selama satu hari umur. Dalam percubaan pertama , anak ayam berumur satu hari diberikan kepada 32 kurungan (floor pen) (1.7m × 1.4m) sebanyak 10 anak ayam/m² dan 15 anak ayam/ m² dan diberikan kepada empat rawatan pemakanan iaitu (i) suplemen diet asas (ii) suplemen diet asas dibekalkan sebanyak 0.5 % LG pada tiga minggu pertama , (iii) suplemen diet asas ditambah sebanyak 0.5% LG selama 6 minggu, (iv) dan suplemen diet asas ditambah sebanyak 0.02 % virginiamycin selama 6 minggu. Pengambilan makanan , nisbah pertukaran makanan (FCR) dan berat badan badan direkodkan setiap minggu. Sampel darah dan duodenum diambil pada hari ke-42. Keputusan eksperimen pertama menunjukkan bahawa pemberian suplemen LG sebanyak 0.5% selama 3 minggu pertama dan 6 minggu berikutnya tidak menjelaskan prestasi pertumbuhan, FCR, parameter darah dan tempoh penyerapan ($p >0.05$) berbanding dengan suplemen diet asas dalam kepadatan stok yang sama. Walau bagaimanapun, dengan memasukkan Virginiamycin didapati signifikan dengan bertambah baiknya parameter tersebut ($p <0.05$). Ketumpatan stok tinggi menjelaskan berat badan, FCR , tempoh penyerapan , tindak balas antibodi dan peningkatan paras fasa kritikal protein serum, nisbah

heterophil / limfosit dan plasma corticosterone ($p <0.05$) berbanding dengan kepadatan stok rendah. Dalam kajian yang kedua , anak ayam telah diberikan kepada 20 kurungan ($2.5m \times 1.4m$) sebanyak 25 burung /kurungan selama 42 hari dan dibekalkan dengan diet asas atau suplemen diet asas dengan 1% LG untuk tiga minggu pertama. Anak ayam umur sehari daripada setiap kumpulan diet yang berlainan tertakluk kepada penempatan serta-merta atau penempatan yang bertangguh (24 jam selepas ketibaan). Pengambilan makanan , FCR dan berat badan badan direkodkan pada setiap minggu. Sampel darah dan duodenum diambil pada hari yang ke-21. Gabungan 1% LG mempunyai kesan positif ke atas tempoh serapan , tahap fasa kritisik serum protein dan HSP 70 ($p < 0.05$) berbanding dengan kumpulan yang menerima diet asas selama 21 hari . Begitu juga, FCR dan nilai berat badan sebanyak 1% LG mempunyai signifikan yang jauh lebih baik daripada yang diberi pemakanan diet *asas* pada akhir eksperimen. penempatan tertangguh anak ayam tidak mempunyai kesan yang ketara ke atas prestasi pertumbuhan , morfologi usus dan parameter darah di kedua-dua kumpulan pemakanan. Oleh yang demikian, hasil kajian ini menunjukkan bahawa suplemen 0.5% LG dalam diet tidak berfungsi di bawah tekanan kepadatan stok tinggi dan 1% LG meningkatkan kesihatan ayam daging dan meningkatkan prestasi di bawah tekanan penempatan tertangguh.

ACKNOWLEDGEMENTS

First of all, thanks God for giving me another opportunity to know myself by living in Malaysia. I wish to express my utmost thanks and deepest gratitude to the chairman of the supervisory committee, Professor Zulkifli Idrus, PhD, for his supervision, invaluable guidance and advice and encouragement throughout this study and for his critical analysis and helpful suggestions during the preparation of the thesis.

I would also like to express my appreciation to Professor Dr. Mohd Hair Bejo and Dr. Faez Firdaus Jesse Abdullah, who are members of the supervisory committee, for their guidance, advices and kind assistance, which guide me through the proper direction and my friend Dr. Reza Soleimani Farjam and Pardis Najafi who helped me during my study.

I would like to express my special thanks to my parents and brothers for their continuous support, encouragement and prayers which have inspired and motivated me throughout the course of this study. I will always be grateful for their constant love and encouragement that they gave me along the way.

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xii
LIST OF FIGURES	xiii
LIST OF ABBREVIATIONS	xiv
 CHAPTER	
1 GENERAL INTRODUCTION	1
2 LITERATURE REVIEW	
2.1 Glutamine and Glutamate	3
2.2 Intestinal morphology	4
2.3 Stocking density	5
2.4 Delayed placement	5
2.5 Acute phase protein	6
2.6 Corticosterone	7
2.7 Heterophil to lymphocyte ratio	7
2.8 Heat shock protein	7
2.9 Gut microflora	8
2.10 Summary	8
 3 EFFECT OF DIETARY L-GLUTAMINE, L-GLUTAMATE AND ANTIBIOTIC SUPPLEMENTATIONS ON INTESTINAL STRUCTURE MICROFLORA, BLOOD PARAMETERS AND PERFORMANCE IN BROILER CHICKEN STOCKED AT DEFERENT DENSITIES UNDER TROPICAL CLIMATE CONDITION	
3.1 Introduction	9
3.2 Materials and methods	11
3.2.1 Birds, husbandry and housing	11
3.2.2 Experimental treatment	11
3.2.3 Growth performance	13
3.2.4 Blood parameters	13
3.2.5 Intestinal microbial populations and morphology	14
3.2.6 Foot pad dermatitis	14
3.2.7 Statistical analysis	15
3.3 Results	16
3.3.1 Performance	16
3.3.2 Blood parameters	18
3.3.3 Morphometric variable	18

3.3.4	Enumeration of bacteria	20
3.3.5	Foot pad dermatitis	20
3.4	Discussion	23
3.5	Conclusion	25
4	RESPONSE TO DIETARY SUPPLEMENTATION OF L-GLUTAMINE AND L-GLUTAMATE IN BROILER CHICKENS SUBJECTED TO DELAYED PLACEMENT	
4.1	Introduction	27
4.2	Materials and methods	29
4.2.1	Chickens, Housing, Management and experimental treatment	29
4.2.2	Sample collection	30
4.2.3	Acute phase protein measurement	30
4.2.4	Intestinal morphology	31
4.2.5	Heat shock protein density analysis	31
4.2.6	Statistical analysis	32
4.3	Results	33
4.3.1	Growth performance	33
4.3.2	Acute phase protein and heat shock protein	33
4.3.3	Villi length and crypt depth	33
4.3.4	Yolk sac	33
4.4	Discussion	37
4.5	Conclusion	39
5	GENERAL DISCUSSION AND CONCLUSIONS	
5.1	General discussion	41
5.2	General conclusion	43
5.3	General recommendation	43
REFERENCES		44
BIODATA OF STUDENT		61
LIST OF PUBLICATIONS		62

LIST OF TABLES

Table	Page
3.1 AminoGut composition.	12
3.2 Nutrient composition of experimental diets.	12
3.3 Mean (\pm SEM) weight gains (g), feed intake (g), feed conversion ratios (FCR) and mortality rate by diet and stocking density.	17
3.4 Mean (\pm SEM) serum ceruloplasmin (CER), α -1 acid glycoprotein (AGP), ovotransferrin (OVT), corticosterone (CORT) concentrations, and heterophil to lymphocyte ratios (HLR) by diet and stocking density.	19
3.5 Mean (\pm SEM) villi lengths (μm) and crypt depths (μm) by diet and stocking density.	20
3.6 Mean (\pm SEM) intestinal <i>E.coli</i> and <i>lactobacilli</i> counts and foot pad dermatitis (FPD) scores by diet and stocking density.	21
3.7 Mean intestinal <i>E.coli</i> counts where diet \times density interactions were significant in broiler chickens.	22
4.1 Nutrient composition of experimental diets.	30
4.2 Mean (\pm SEM) body weights, feed intake, feed conversion ratio and mortality rate by diet and placement time.	34
4.3 Mean (\pm SEM) serum ceruloplasmin (CER), α -1 acid glycoprotein (AGP), ovotransferrin (OVT) and heat shock protein (HSP) by diet and placement time.	35
4.4 Mean (\pm SEM) villi lengths (μm) and crypt depths (μm) by diet and placement time.	36

LIST OF FIGURES

Figure		Page
2.1	Chemical composition of glutamine and glutamate. Glutamine transforms to the glutamate (Tapiero <i>et al.</i> , 2002).	3
2.2	The acute phase response (Murata <i>et al.</i> , 2004) .	6

LIST OF ABBREVIATIONS

LG	Mixture of L- glutamine and L-glutamate
AGP	Acid glycoprotein
APP	Acute phase protein
CP	Ceruloplasmin
CORT	Corticosterone
FCR	Feed conversion ratio
FPD	Foot pad dermatitis
g	Gram
GIT	Gastrointestinal tract
GLN	Glutamine
GLU	Glutamate
h	Hour
HLR	Heterophil to Lamphocyte
HSD	High Stocking Density
HSP	Heat shock protein
LSD	Low stocking density
mg	Mili gram
ml	Mili litter
nm	Nano meter
OVT	Ovotransferin
VM	Virginiamycin
µl	Micro litter

CHAPTER 1

GENERAL INTRODUCTION

Over the past several decades the poultry meat industry has been growing rapidly, particularly in developing countries (Taha, 2003). In 2008, approximately 71.4 million metric tons of poultry meat was produced and this was 5 % more compared to 2007 productions (US Department of Agriculture, 2009). With the estimation of 2.6% annual growth, poultry meat production will reach 67 million metric tons by 2013 globally. The current strategy of the industry is to increase production efficiency through reducing the growing time period. Accordingly, researchers are constantly looking for alternatives and solutions to achieve this target. One of the areas among the factors and conditions that are of increasing interest to researchers and industry is animal welfare. Animal welfare is directly related to the housing system (Hubrecht and Kirkwood, 2010), nutrition and environment conditions such as stocking density (Broom, 1993).

Stocking density is defined as the number of chicks per specific amount of area per bird. High stocking density can reduce animal welfare by decreasing the quality of the environment and increasing competition for access to water and feed (Knowles *et al.*, 2008). Optimum stocking density for broiler chicken is 10 birds per square meter (Thomas *et al.*, 2004). Generally, high stocking density (HSD) may negatively affect performance, liveability, litterer moisture, feed efficiency (Sanotra *et al.*, 2001; Dozier *et al.*, 2005; Estevez, 2007; Skrbic and Lukic, 2009) and decrease the amount of production due to economic losses (Braun *et al.*, 2010). Moreover, it increases microbial activity and promotes higher incidences of dermatitis among chickens through increase moisture of litterer (Bessei, 2006). However, from the economic viewpoints of the market and producers, apart from the negative effects of high stocking density on the welfare of chickens (Hall, 2001), produce more meat is more considered than the weight of each chicks (Bilgili and Hess, 1995), which cause higher profitability (Shanawany, 1988).

The handling of chicken after hatching is a crucial matter which is directly connected with optimum growth performance and good welfare. Under commercial procedures, chicks are subjected to various processes such as sorting, vaccination, preparation and transportation which can physiologically affect them. Transportation of animals to the farm has been considered as an important factor which can be related to food quality and animal welfare (Keeling, 2005; Marahrens *et al.*, 2011) and also cause delay placement which is a negative factor in chickens performance. Delayed placement has been shown to cause dehydration and depletion of yolk sac reserves (Vieira *et al.* 2005). Earlier studies have shown a relation between delayed placement and depression in growth performance of chickens (Mahmoud and Edens, 2012; Peng and Guo, 2010). In fact, delivery to farms has been known to occur delays in chicken placement for as long as 24-48 h.

As a result, it is common in the poultry industry to supplement chickens with antibiotics to eliminate the negative effects of high stocking density on growth

performance and gut health. However, the concern on antibiotic resistance - particularly in human – has led the poultry industry and researchers to look for the consumer-farmer friendly alternatives. Among the several alternatives to fortify the diets with dietary nutrition, glutamine supplementation is considered as an important factor in promoting the well-being and increasing the health of chickens. As shown by Andrews and Griffiths, (2002), Yi *et al.* (2001) and Aledo, (2004) glutamine supplementation can increase villa length and act as an energy source for the intestinal tissue and immune cells. During stress incidence, the damage to the intestinal epithelium may increase, and therefore, it may enhance the pathogens chances to enter into the body (Adjei *et al.*, 1994). Glutamine may function to decrease the incidence of infection by improving the gut health and epithelial permeability (Andrews and Griffiths, 2002; Medina, 2001).

Thus, the main objective of the this study was to investigate the effects of L- glutamine and L-glutamate (LG) supplementation on factors affecting performance of broiler chickens which were subjected to stressful conditions.

Therefore, the objectives of this study were:

- (1) To investigate the effects of L- glutamine and L-glutamate and antibiotic dietary supplements on growth performance, intestinal morphology and microflora, and blood parameters in broiler chickens stocked at different densities under the hot tropical environment.
- (2) To evaluate the effects of dietary L- glutamine and L-glutamate supplementation, and delayed placement on the growth performance, intestinal morphologyand physiological stress responses in broiler chickens under the hot tropical environment.

REFERENCES

- Abelson, S. P., Adem, B., Royo, F., Carlsson, H., and Hau, J. (2005). High plasma corticosterone levels persist during frequent automatic blood sampling in rats. *In Vivo*, 19(5), 815-819.
- Adjei, A., Matsumoto, Y., Oku, T., Hiroi, Y., and Yamamoto, S. (1994). Dietary agrinine and glutamine combination improves survival in septic mice. *Nutrition Research*, 14(10), 1591-1599.
- Ahmadi, F. (2011). The Effect of Different Levels of Virginiamycin on Performance, Immune organs and Blood Metabolite of Broiler Chickens. *Annals of Biological Research*, 2(5).
- Al-Aqil, A., and Zulkifli, I. (2009). Changes in heat shock protein 70 expression and blood characteristics in transported broiler chickens as affected by housing and early age feed restriction. *Poultry Science*, 88(7), 1358-1364.
- Aledo, J. C. (2004). Glutamine breakdown in rapidly dividing cells: waste or investment? *Bioessays*, 26(7), 778-785.
- Allain, V., Mirabito, L., Arnould, C., Colas, M., Le Bouquin, S., Lupo, C., and Michel, V. (2009). Skin lesions in broiler chickens measured at the slaughterhouse: relationships between lesions and between their prevalence and rearing factors. *British Poultry Science*, 50(4), 407-417.
- Andrews, F. J., and Griffiths, R. D. (2002). Glutamine: essential for immune nutrition in the critically ill. *British Journal of Nutrition*, 87(1), 3-8.
- Arthington, J. D., Eicher, S. D., Kunkle, W . E., and Martin, F. G. (2003). Effect of transportation and commingling on the acute-phase protein response, growth, and feed intake of newly weaned beef calves. *Journal of Animal Science*, 81(5), 1120-1125.
- Askanazi, J., Carpentier, Y. A., Michelsen, C. B., Elwyn, D. H., Furst, P., Kantrowitz, L. R., Kinney, J. M. (1980). Muscle and plasma amino acids following injury: Influence of intercurrent infection. *Annals of Surgery*, 192(1), 78.
- Awad, W., Ghareeb, K., and Böhm, J. (2008). Intestinal structure and function of broiler chickens on diets supplemented with a probiotic containing

Enterococcus faecium and oligosaccharides. *International Journal of Molecular Sciences*, 9(11), 2205-2216.

Baddeley, A. J., Gundersen, H. G., and Cruz-Orive, L. (1986). Estimation of surface area from vertical sections. *Journal of Microscopy*, 142(3), 259-276.

Baghshani, H., Nazifi, S., Saeb, M., and Saeb, S. (2010). Influence of road transportation on plasma concentrations of acute phase proteins, including fibrinogen, haptoglobin, serum amyloid A, and ceruloplasmin, in dromedary camels (*Camelus dromedarius*). *Comparative Clinical Pathology*, 19(2), 193-198.

Bancroft, J. D., Gamble MBancroft, J. D., and Gamble, M. (2002). Connective tissue stains Theory and practice of histological techniques: Horcourt publishers limited. London. New York-Toronto.

Bartell, S. M., and Batal, A. B. (2007). The effect of supplemental glutamine on growth performance, development of the gastrointestinal tract, and humoral immune response of broilers. *Poultry Science*, 86(9), 1940-1947.

Beloor, J., Kang, H. K., Kim, Y. J., Subramani, V. K., Jang, I. S., Sohn, S. H., and Moon, Y. S. (2010). The effect of stocking density on stress related genes and telomeric length in broiler chickens. *Asian-Aust. J. Anim. Sci.*, 23, 437-443.

Bessei, W. (2006). Welfare of broilers: a review. *World's Poultry Science Journal*, 62(3), 455-466.

Beuving, G., and Vonder, G. M. A. (1978). Effect of stressing factors on corticosterone levels in the plasma of laying hens. *General and Comparative Endocrinology*, 35(2), 153-159.

Bhanja, S. K., Devi, C. A., Panda, A. K., and Sunder, G. S. (2009). Effect of post hatch feed deprivation on yolk-sac utilization and performance of young broiler chickens. *Asian-Australasian Journal of Animal Sciences*, 22(8), 1174-1179.

Bigot, K., Mignon-Grasteau, S., Picard, M., and Tesseraud, S. (2003). Effects of delayed feed intake on body, intestine, and muscle development in neonate broilers. *Poultry Science*, 82(5), 781-788.

- Bilgili, S. F., and Hess, J. B. (1995). Placement density influences broiler carcass grade and meat yields. *The Journal of Applied Poultry Research*, 4(4), 384-389.
- Braun, N., de Lima, R., Baldisserotto, B., Dafre, A. L., and de Oliveira Nuñez, A. P. (2010). Growth, biochemical and physiological responses of *Salminus brasiliensis* with different stocking densities and handling. *Aquaculture*, 301(1), 22-30.
- Broom, D. M. (1993). *Stress and Animal Welfare*: Springer.
- Burkholder, K. M., Thompson, K. L., Einstein, M. E., Applegate, T. J., and Patterson, J. A. (2008). Influence of stressors on normal intestinal microbiota, intestinal morphology, and susceptibility to *Salmonella enteritidis* colonization in broilers. *Poultry Science*, 87(9), 1734-1741.
- Butaye, P., Devriese, L. A., and Haesebrouck, F. (2003). Antimicrobial growth promoters used in animal feed: effects of less well known antibiotics on gram-positive bacteria. *Clinical Microbiology Reviews*, 16(2), 175-188.
- Buyse, J., Swennen, Q., Niewold, T. A., Klasing, K. C., Janssens, G. P. J., Baumgartner, M., and Goddeeris, B. M. (2007). Dietary L-carnitine supplementation enhances the lipopolysaccharide-induced acute phase protein response in broiler chickens. *Veterinary Immunology and Immunopathology*, 118(1), 154-159.
- Cabrera, R. A., Usry, J. L., Arrellano, C., Nogueira, E. T., Kutschenko, M., Moeser, A. J., and Odle, J. (2013). Effects of creep feeding and supplemental glutamine or glutamine plus glutamate (Aminogut) on pre- and post-weaning growth performance and intestinal health of piglets. *Journal of Animal Science and Biotechnology*, 4(1), 1-13.
- Calder, P. C., and Yaqoob, P. (1999). Glutamine and the immune system. *Amino Acids*, 17(3), 227-241.
- Chamblee, T. N., Brake, J. D., Schultz, C. D., and Thaxton, J. P. (1992). Yolk sac absorption and initiation of growth in broilers. *Poultry Science*, 71(11), 1811-1816.

- Chow, A., and Zhang, R. (1998). Glutamine reduces heat shock-induced cell death in rat intestinal epithelial cells. *The Journal of Nutrition*, 128(8), 1296-1301.
- Cockrem, J. F. (2007). Stress, corticosterone responses and avian personalities. *Journal of Ornithology*, 148(2), 169-178.
- Colley, C. M., Fleck, A., Goode, A. W., Muller, B. R., and Myers, M. A. (1983). Early time course of the acute phase protein response in man. *Journal of Clinical Pathology*, 36(2), 203-207.
- Corless, A. B., and Sell, J. L. (1999). The effects of delayed access to feed and water on the physical and functional development of the digestive system of young turkeys. *Poultry Science*, 78(8), 1158-1169.
- Craig, E. A., and Gross, C. A. (1991). Is hsp70 the cellular thermometer? *Trends in Biochemical Sciences*, 16, 135-140.
- Craig, E. A., and Schlesinger, M. J. (1985). The Heat Shock Response. *Critical Reviews in Biochemistry and Molecular Biology*, 18(3), 239-280.
- Cravener, T. L., Roush, W. B., and Mashaly, M. M. (1992). Broiler production under varying population densities. *Poultry Science*, 71(3), 427-433.
- Dai, S. F., Wang, L. K., Wen, A. Y., Wang, L. X., and Jin, G. M. (2009). Dietary glutamine supplementation improves growth performance, meat quality and colour stability of broilers under heat stress. *British Poultry Science*, 50(3), 333-340.
- Daniele, B., Perrone, F., Gallo, C., Pignata, S., De Martino, S., De Vivo, R., D'Agostino, L. (2001). Oral glutamine in the prevention of fluorouracil induced intestinal toxicity: a double blind, placebo controlled, randomised trial. *Gut*, 48(1), 28-33.
- Dawkins, M. S. (2003). Behaviour as a tool in the assessment of animal welfare. *Zoology*, 106(4), 383-387.
- Dawkins, M. S., Donnelly, C. A., and Jones, T. A. (2004). Chicken welfare is influenced more by housing conditions than by stocking density. *Nature*, 427(6972), 342-344.

- De Graaf, T. W., Van der Stelt, M. E., Anbergen, M. G., and Van Dijk, W. (1993). Inflammation-induced expression of sialyl Lewis X-containing glycan structures on alpha 1-acid glycoprotein (orosomucoid) in human sera. *The Journal of Experimental Medicine*, 177(3), 657-666.
- DeMarco, V. G., Li, N., Thomas, J., West, C. M., and Neu, J. (2003). Glutamine and barrier function in cultured Caco-2 epithelial cell monolayers. *The Journal of Nutrition*, 133(7), 2176-2179.
- Dibner, J. J., Knight, C. D., Kitchell, M. L., Atwell, C. A., Downs, A. C., and Ivey, F. J. (1998). Early feeding and development of the immune system in neonatal poultry. *The Journal of Applied Poultry Research*, 7(4), 425-436.
- Dibner, J. J., and Richards, J. D. (2004). The digestive system: Challenges and opportunities. *The Journal of Applied Poultry Research*, 13(1), 86-93.
- Dibner, J. J., Ivey, F. J., and Knight, C. D. (1999). Nutrient formulation and process for enhancing the health, livability, cumulative weight gain or feed efficiency in poultry and other animals: Google Patents.
- Dozier, W. A., Thaxton, J. P., Branton, S. L., Morgan, G. W., Miles, D. M., Roush, W. B., Vizzier-Thaxton, Y. (2005). Stocking density effects on growth performance and processing yields of heavy broilers. *Poultry Science*, 84(8), 1332-1338.
- Dozier, W. A., Thaxton, J. P., Purswell, J. L., Olanrewaju, H. A., Branton, S. L., and Roush, W. B. (2006). Stocking density effects on male broilers grown to 1.8 kilograms of body weight. *Poultry Science*, 85(2), 344-351.
- Eckersall, P. D. (2004). The time is right for acute phase protein assays. *The Veterinary Journal*, 168(1), 3-5.
- Eckersall, P. D., Young, F. J., Nolan, A. M., Knight, C. H., McComb, C., Waterston, M. M., Fitzpatrick, J. L. (2006). Acute Phase Proteins in Bovine Milk in an Experimental Model of *Staphylococcus aureus* Subclinical Mastitis. *Journal of Dairy Science*, 89(5), 1488-1501.
- Ehrenfried, J. A., Chen, J., Li, J., and Mark Evers, B. (1995). Glutamine-mediated regulation of heat shock protein expression in intestinal cells. *Surgery*, 118(2), 352-357.

- Erbil, Y., Berber, E., Ozarmagan, S., Seven, R., Eminoglu, L., Calis, A., and Gürler, N. (1998). The effects of sodium deoxycholate, lactulose and glutamine on bacterial translocation in common bile duct ligated rats. *Hepato-Gastroenterology*, 46(29), 2791-2795.
- Estevez, I. (2007). Density allowances for broilers: where to set the limits? *Poultry Science*, 86(6), 1265-1272.
- Fanguy, R. C., Misra, L. K., Vo, K. V., Blohowiak, C. C., and Krueger, W. F. (1980). Effect of delayed placement on mortality and growth performance of commercial broilers. *Poultry Science*, 59(6), 1215-1220.
- Feddes, J. J., Emmanuel, E. J., and Zuidhoft, M. J. (2002). Broiler performance, body weight variance, feed and water intake, and carcass quality at different stocking densities. *Poultry Science*, 81(6), 774-779.
- Franco, J. R. G., Murakami, A. E., Natali, M. R. M., Garcia, E. R. M., and Furlan, A. C. (2006). Influence of delayed placement and dietary lysine levels on small intestine morphometrics and performance of broilers. *Revista Brasileira de Ciência Avícola*, 8(4), 233-241.
- Georgieva, T. M., Koinarski, V. N., Urumova, V. S., Marutsov, P. D., Christov, T. T., Nikolov, J., Georgiev, I. P. (2010). Effects of Escherichia coli infection and Eimeria tenella invasion on blood concentrations of some positive acute phase proteins (haptoglobin (PIT 54), fibrinogen and ceruloplasmin) in chickens. *Revue De Medecine Veterinaire*, 161(2), 84.
- Gerner, E. W., and Schneider, M. J. (1975). Induced thermal resistance in HeLa cells. *Nature*, 256(5517), 500-502.
- Grant, J. P., and Snyder, P. J. (1988). Use of L-glutamine in total parenteral nutrition. *Journal of Surgical Research*, 44(5), 506-513.
- Gross, W. B., and Siegel, H. S. (1983). Evaluation of the heterophil/lymphocyte ratio as a measure of stress in chickens. *Avian Diseases*, 972-979.
- Gruys, E., Toussaint, M. J. M., Upagarin, N., Van Ederen, A. M., Adewuyi, A. A., Candiani, D., Sabeckiene, J. (2005). Acute phase reactants, challenge in the near future of animal production and veterinary medicine. *Journal of Zhejiang University Science B*, 6(10), 941-947.

- Gu, X. H., Hao, Y., and Wang, X. L. (2012). Overexpression of heat shock protein 70 and its relationship to intestine under acute heat stress in broilers: 2. Intestinal oxidative stress. *Poultry Science*, 91(4), 790-799.
- Halevy, O., Geyra, A., Barak, M., Uni, Z., and Sklan, D. (2000). Early posthatch starvation decreases satellite cell proliferation and skeletal muscle growth in chicks. *The Journal of Nutrition*, 130(4), 858-864.
- Hall, A. L. (2001). The effect of stocking density on the welfare and behaviour of broiler chickens reared commercially. *Animal Welfare*, 10(1), 23-40.
- Heckert, R. A., Estevez, I., Russek-Cohen, E., and Pettit-Riley, R. (2002). Effects of density and perch availability on the immune status of broilers. *Poultry Science*, 81(4), 451-457.
- Holt, P. S., and Gast, R. K. (2002). Comparison of the effects of infection with *Salmonella enteritidis*, in combination with an induced molt, on serum levels of the acute phase protein, alpha₁ acid glycoprotein, in hens. *Poultry Science*, 81(9), 1295-1300.
- Houshmand, M., Azhar, K., Zulkifli, I., Bejo, M. H., and Kamyab, A. (2012). Effects of prebiotic, protein level, and stocking density on performance, immunity, and stress indicators of broilers. *Poultry Science*, 91(2), 393-401.
- Hubrecht, R., and Kirkwood, J. (2010). *The UFAW handbook on the care and management of laboratory and other research animals*: John Wiley & Sons.
- Imaeda, N. (2000). Influence of the stocking density and rearing season on incidence of sudden death syndrome in broiler chickens. *Poultry Science*, 79(2), 201-204.
- Inoue, Y., Grant, J. P., and Snyder, P. J. (1993). Effect of glutamine-supplemented total parenteral nutrition on recovery of the small intestine after starvation atrophy. *Journal of Parenteral and Enteral Nutrition*, 17(2), 165-170.
- Joerger, R. D. (2003). Alternatives to antibiotics: bacteriocins, antimicrobial peptides and bacteriophages. *Poultry Science*, 82(4), 640-647.

- Johnson, I. R., Ball, R. O., Baracos, V. E., and Field, C. J. (2006). Glutamine supplementation influences immune development in the newly weaned piglet. *Developmental and Comparative Immunology*, 30(12), 1191-1202.
- Keeling, D. (2005). The Economics of Migrant Transport between Europe and the United States, 1900-1914.
- Khan, J., Iiboshi, Y., Cui, L., Wasa, M., Sando, K., Takagi, Y., and Okada, A. (1999). Alanyl-glutamine-supplemented parenteral nutrition increases luminal mucus gel and decreases permeability in the rat small intestine. *Journal of Parenteral and Enteral Nutrition*, 23(1), 24-31.
- Khodambashi Emami, N., Samie, A., Rahmani, H. R., and Ruiz-Feria, C. A. (2012). The effect of peppermint essential oil and fructooligosaccharides, as alternatives to virginiamycin, on growth performance, digestibility, gut morphology and immune response of male broilers. *Animal Feed Science and Technology*, 175(1), 57-64.
- Klasing, K. C. (1988). Nutritional aspects of leukocytic cytokines. *The Journal of Nutrition*, 118(12), 1436.
- Knowles, T. G., Kestin, S. C., Haslam, S. M., Brown, S. N., Green, L. E., Butterworth, A., Nicol, C. J. (2008). Leg disorders in broiler chickens: prevalence, risk factors and prevention. *PLoS One*, 3(2), e1545.
- Kregel, K. C. (2002). Invited review: heat shock proteins: modifying factors in physiological stress responses and acquired thermotolerance. *Journal of Applied Physiology*, 92(5), 2177-2186.
- Lavoinne, A., Husson, A., and Quillard, M. (1998). *Glutamine et cellule hépatique: métabolisme, propriétés et concept de régulation du métabolisme par le gonflement cellulaire*. Paper presented at the Annales de Biologie Clinique.
- Mahmoud, K. Z., and Edens, F. W. (2012). Breeder age affects small intestine development of broiler chicks with immediate or delayed access to feed. *British Poultry Science*, 53(1), 32-41.
- Maiorka, A., Silva, A. V. F., Santin, E., Borges, S. A., Boleli, I. C., and Macari, M. (2000). Influence of glutamine supplementation on performance and intestinal villous and crypt development in broiler chickens. *Arquivo Brasileiro de Medicina Veterinaria e Zootecnia*, 52(5), 487-490.

- Makarova, K., Slesarev, A., Wolf, Y., Sorokin, A., Mirkin, B., Koonin, E., Polouchine, N. (2006). Comparative genomics of the lactic acid bacteria. *Proceedings of the National Academy of Sciences*, 103(42), 15611-15616.
- Mancini, G., Carbonara, A. O. T., and Heremans, J. F. (1965). Immunochemical quantitation of antigens by single radial immunodiffusion. *Immunochemistry*, 2(3), 235-IN236.
- Marahrens, M., Kleinschmidt, N., Di Nardo, A., Velarde, A., Fuentes, C., Truar, A., and Villa, P. D. (2011). Risk assessment in animal welfare—Especially referring to animal transport. *Preventive Veterinary Medicine*, 102(2), 157-163.
- Maxwell, M. H. (1993). Avian blood leucocyte responses to stress. *World's Poultry Science Journal*, 49(01), 34-43.
- Meddings, J. B., and Swain, M. G. (2000). Environmental stress-induced gastrointestinal permeability is mediated by endogenous glucocorticoids in the rat. *Gastroenterology*, 119(4), 1019-1028.
- Medina, M. A. (2001). Glutamine and cancer. *The Journal of Nutrition*, 131(9), 2539S-2542S.
- Miles, D. M., Branton, S. L., and Lott, B. D. (2004). Atmospheric ammonia is detrimental to the performance of modern commercial broilers. *Poultry Science*, 83(10), 1650-1654.
- Miles, R. D., Butcher, G. D., Henry, P. R., and Littell, R. C. (2006). Effect of antibiotic growth promoters on broiler performance, intestinal growth parameters, and quantitative morphology. *Poultry Science*, 85(3), 476-485.
- Mitchell, M. A., and Carlisle, A. J. (1992). The effects of chronic exposure to elevated environmental temperature on intestinal morphology and nutrient absorption in the domestic fowl (*Gallus domesticus*). *Comparative Biochemistry and Physiology Part A: Physiology*, 101(1), 137-142.
- Morris, M. P. (1993). National survey of leg problems. *Pigs and Poultry*, 6, 016.
- Müller, C., Jenni-Eiermann, S., and Jenni, L. (2011). Heterophils/Lymphocytes-ratio and circulating corticosterone do not indicate the same stress

imposed on Eurasian kestrel nestlings. *Functional Ecology*, 25(3), 566-576.

Murakami, A. E., Sakamoto, M. I., Natali, M. R. M., Souza, L. M. G., and Franco, J. R. G. (2007). Supplementation of glutamine and vitamin E on the morphometry of the intestinal mucosa in broiler chickens. *Poultry Science*, 86(3), 488-495.

Murata, H., Shimada, N., and Yoshioka, M. (2004). Current research on acute phase proteins in veterinary diagnosis: an overview. *The Veterinary Journal*, 168(1), 28-40.

Murata, H. (2007). Stress and acute phase protein response: an inconspicuous but essential linkage. *The Veterinary Journal*, 173(3), 473-474.

Newsholme, E., Crabtree, B., and Ardawi, M. (1985). The role of high rates of glycolysis and glutamine utilization in rapidly dividing cells. *Bioscience Reports*, 5, 393-400.

Newsholme, P. (2001). Why is L-glutamine metabolism important to cells of the immune system in health, postinjury, surgery or infection? *The Journal of Nutrition*, 131(9), 2515S-2522S.

Newsholme, P., and Newsholme, E. A. (1989). Rates of utilization of glucose, glutamine and oleate and formation of end-products by mouse peritoneal macrophages in culture. *Biochem. J.*, 261, 211-218.

Newsholme, P., Procopio, J., Lima, M. M. R., Pithon-Curi, T. C., and Curi, R. (2003). Glutamine and glutamate-their central role in cell metabolism and function. *Cell Biochemistry and Function*, 21(1), 1-9.

Niewold, T. A., Toussaint, M. J. M., and Gruijs, E. (2003). *Monitoring health by acute phase proteins*. Paper presented at the Animal Welfare and Acute Phase Proteins. Proceedings of Fourth European Colloquium on Acute Phase Proteins. Segovia, Spain.

Nir, I., and Levanon, M. (1993). Research note: effect of posthatch holding time on performance and on residual yolk and liver composition. *Poultry Science*, 72(10), 1994-1997.

Noy, Y., Uni, Z., and Sklan, D. (1996). Routes of yolk utilisation in the newly-hatched chick. *British Poultry Science*, 37(5), 987-996.

O'Reilly, E. L., Burchmore, R. J., Sandilands, V., Sparks, N. H., Walls, C., and Eckersall, P. D. (2012). The plasma proteome and acute phase proteins of broiler chickens with gait abnormalities *Farm Animal Proteomics* (pp. 177-180): Springer.

Obled, C. (2003). Amino acid requirements in inflammatory states. *Canadian Journal of Animal Science*, 83(3), 365-373.

Panda, A. K., Rama R., Savaram S., Raju, M. V. L. N., and Sharma, S. S. (2008). Effect of probiotic (*Lactobacillus sporogenes*) feeding on egg production and quality, yolk cholesterol and humoral immune response of White Leghorn layer breeders. *Journal of the Science of Food and Agriculture*, 88(1), 43-47.

Parsaie, S., Shariatmadari, F., Zamiri, M. J., and Khajeh, K. (2007). Influence of wheat-based diets supplemented with xylanase, bile acid and antibiotics on performance, digestive tract measurements and gut morphology of broilers compared with a maize-based diet. *British Poultry Science*, 48(5), 594-600.

Peng, C., Sun, H., and Guo, J. (2010). Multi-objective optimal PMU placement using a non-dominated sorting differential evolution algorithm. *International Journal of Electrical Power & Energy Systems*, 32(8), 886-892.

Pettit-Riley, R., and Estevez, I. (2001). Effects of density on perching behavior of broiler chickens. *Applied Animal Behaviour Science*, 71(2), 127-140.

Pinchasov, Y., and Noy, Y. (1993). Comparison of post-hatch holding time and subsequent early performance of broiler chicks and Turkey poult. *British Poultry Science*, 34(1), 111-120.

Pineiro, M., Pineiro, C., Carpintero, R., Morales, J., Campbell, F. M., Eckersall, P. D., Lampreave, F. (2007). Characterisation of the pig acute phase protein response to road transport. *The Veterinary Journal*, 173(3), 669-674.

Potturi, P. V., Patterson, J. A., and Applegate, T. J. (2005). Effects of delayed placement on intestinal characteristics in turkey pouls. *Poultry Science*, 84(5), 816-824.

Qiu, X., Arthington, J. D., Riley, D. G., Chase, C. C., Phillips, W. A., Coleman, S. W., and Olson, T. A. (2007). Genetic effects on acute phase protein response to the stresses of weaning and transportation in beef calves. *Journal of Animal Science*, 85(10), 2367-2374.

Reeds, P. J., Fjeld, C. R., and Jahoor, F. (1994). Do the differences between the amino acid compositions of acute-phase and muscle proteins have a bearing on nitrogen loss in traumatic states? *The Journal of Nutrition*, 124(6), 906-910.

Russo, T. A., and Johnson, J. R. (2003). Medical and economic impact of extraintestinal infections due to Escherichia coli: focus on an increasingly important endemic problem. *Microbes and Infection*, 5(5), 449-456.

Sakamoto, M. I., Murakami, A. E., Silveira, T. G. V., Fernandes, J. I. M., and Oliveira, C. A. L. (2006). Influence of glutamine and vitamin E on the performance and the immune responses of broiler chickens. *Revista Brasileira de Ciência Avícola*, 8(4), 243-249.

Sanders, M. E. (1993). Summary of conclusions from a consensus panel of experts on health attributes of lactic cultures: significance to fluid milk products containing cultures. *Journal of Dairy Science*, 76(7), 1819-1828.

Sandine, W. E., Muralidhara, K. S., Elliker, P. R., and England, D. C. (1972). Lactic acid bacteria in food and health: a review with special reference to enteropathogenic Escherichia coli as well as certain enteric diseases and their treatment with antibiotics and lactobacilli. *Journal of Milk and Food Technology*.

Sanotra, G. S., Lawson, L. G., Vestergaard, K. S., and Thomsen, M. G. (2001). Influence of stocking density on tonic immobility, lameness, and tibial dyschondroplasia in broilers. *Journal of Applied Animal Welfare Science*, 4(1), 71-87.

Sapolsky, R. M., Romero, L. M., and Munck, A. U. (2000). How do glucocorticoids influence stress responses? Integrating permissive, suppressive, stimulatory, and preparative actions. *Endocrine Reviews*, 21(1), 55-89.

- Segura, J. C., Feddes, J. J. R., and Zuidhof, M. J. (2006). Midday and nighttime cooling of broiler chickens. *The Journal of Applied Poultry Research*, 15(1), 28-39.
- Shanawany, M. M. (1988). Broiler performance under high stocking densities. *British Poultry Science*, 29(1), 43-52.
- Sikora, A., and Grzesiuk, E. (2007). Heat shock response in gastrointestinal tract. *Journal of Physiology and Pharmacology*, 58, 43.
- Singleton, K. D., and Wischmeyer, P. E. (2006a). Effects of HSP70. 1/3 gene knockout on acute respiratory distress syndrome and the inflammatory response following sepsis. *American Journal of Physiology-Lung Cellular and Molecular Physiology*, 290(5), L956-L961.
- Singleton, K. D., and Wischmeyer, P. E. (2006b). Oral glutamine enhances heat shock protein expression and improves survival following hyperthermia. *Shock*, 25(3), 295-299.
- Sklan, D., and Noy, Y. (2000). Hydrolysis and absorption in the small intestines of posthatch chicks. *Poultry Science*, 79(9), 1306-1310.
- Sklan, D. (2001). Development of the digestive tract of poultry. *World's Poultry Science Journal*, 57(04), 415-428.
- Škrbić, Z., Pavlovski, Z., and Lukić, M. (2009). Stocking density: Factor of production performance, quality and broiler welfare. *Biotechnology in Animal Husbandry*, 25(5-6-1), 359-372.
- Soleimani, A. F., Zulkifli, I., Hair-Bejo, M., Omar, A. R., and Raha, A. R. (2012). The role of heat shock protein 70 in resistance to *Salmonella enteritidis* in broiler chickens subjected to neonatal feed restriction and thermal stress. *Poultry Science*, 91(2), 340-345.
- Soltan, M. A. (2009). Influence of dietary glutamine supplementation on growth performance, small intestinal morphology, immune response and some blood parameters of broiler chickens. *International Journal of Poultry Science*, 8(1), 60-68.
- Souba, W. W. (1993). Glutamine and cancer. *Annals of Surgery*, 218(6), 715.

- Souba, W. W., and Pacitti, A. J. (1992). Review: How Amino Acids Get Into Cells: Mechanisms, Models, Menus, and Mediators. *Journal of Parenteral and Enteral Nutrition*, 16(6), 569-578.
- Sunderman, F. W., and Nomoto, S. (1970). Measurement of human serum ceruloplasmin by its p-phenylenediamine oxidase activity. *Clinical Chemistry*, 16(11), 903-910.
- Taha, F. A. (2003). *Poultry Sector in Middle-income Countries and Its Feed Requirements: The Case of Egypt*: United States Department of Agriculture, Economic Research Service.
- Tanguay, R. M. (1983). Genetic regulation during heat shock and function of heat-shock proteins: a review. *Canadian Journal of Biochemistry and Cell Biology*, 61(6), 387-394.
- Tapiero, H., Mathe, G., Couvreur, P., and Tew, K. D. (2002). II. Glutamine and glutamate. *Biomedicine and Pharmacotherapy*, 56(9), 446-457.
- Thaxton, J. P., Dozier, W. A., Branton, S. L., Morgan, G. W., Miles, D. W., Roush, W. B., Vizzier-Thaxton, Y. (2006). Stocking density and physiological adaptive responses of broilers. *Poultry Science*, 85(5), 819-824.
- Tilg, H., Vannier, E., Vachino, G., Dinarello, C. A., and Mier, J. W. (1993). Antiinflammatory properties of hepatic acute phase proteins: preferential induction of interleukin 1 (IL-1) receptor antagonist over IL-1 beta synthesis by human peripheral blood mononuclear cells. *The Journal of Experimental Medicine*, 178(5), 1629-1636.
- Tissiéres, A., Mitchell, H. K., and Tracy, U. M. (1974). Protein synthesis in salivary glands of *Drosophila melanogaster*: Relation to chromosome puffs. *Journal of Molecular Biology*, 84(3), 389-398.
- Uni, Z., Noy, Y., and Sklan, D. (1999). Posthatch development of small intestinal function in the poult. *Poultry Science*, 78(2), 215-222.
- US Department of Agriculture, Natural Resources Conservation Service. (2009). The Plants database: National Plant Data Center Baton Rouge, LA 70874-4490 USA.

- Vachon, P., and Moreau, J. P. (2001). Serum corticosterone and blood glucose in rats after two jugular vein blood sampling methods: comparison of the stress response. *Journal of the American Association for Laboratory Animal Science*, 40(5), 22-24.
- Van Horne, P. L. M., and Achterbosch, T. J. (2008). Animal welfare in poultry production systems: impact of EU standards on world trade. *World's Poultry Science Journal*, 64(01), 40-52.
- Vieira, S. L., Almeida, J. G., Lima, A. R., Conde, O. R. A., and Olmos, A. R. (2005). Hatching distribution of eggs varying in weight and breeder age. *Revista Brasileira de Ciência Avícola*, 7(2), 73-78.
- Waldroup, P. W., Spencer, G. K., Waibel, P. E., Quarles, C. L., and Grant, R. J. (1985). The use of bambermycins (Flavomycin®) and halofuginone (Stenorol®) in diets for growing turkeys. *Poultry Science*, 64(7), 1296-1301.
- Wischmeyer, P. E. (2002). Glutamine and heat shock protein expression. *Nutrition*, 18(3), 225-228.
- Wischmeyer, P. E., Kahana, M., Wolfson, R., Ren, H., Musch, M. M., and Chang, E. B. (2001). Glutamine induces heat shock protein and protects against endotoxin shock in the rat. *Journal of Applied Physiology*, 90(6), 2403-2410.
- Wu, G., Bazer, F. W., Davis, T. A., Jaeger, L. A., Johnson, G. A., Kim, S. W., Yin, Y. L. (2007). Important roles for the arginine family of amino acids in swine nutrition and production. *Livestock Science*, 112(1), 8-22.
- Wu, Y. M., Tang, J., Zhao, P., Chen, Z. N., and Jiang, J. L. (2009). Morphological changes and molecular expressions of hepatocellular carcinoma cells in three-dimensional culture model. *Experimental and Molecular Pathology*, 87(2), 133-140.
- Yahav, S. (2009). Alleviating heat stress in domestic fowl: different strategies. *World's Poultry Science Journal*, 65(04), 719-732.
- Yamauchi, K. (2002). Review on chicken intestinal villus histological alterations related with intestinal function. *The Journal of Poultry Science*, 39(4), 229-242.

- Yeh, S. L., Yeh, C. L., Lin, M. T., Lo, P. N., and Chen, W. J. (2001). Effects of glutamine-supplemented total parenteral nutrition on cytokine production and T cell population in septic rats. *Journal of Parenteral and Enteral Nutrition*, 25(5), 269-274.
- Yi, G. F., Allee, G. L., Frank, J. W., Spencer, J. D., and Touchette, K. J. (2001). Impact of glutamine, menhaden fish meal and spray-dried plasma on the growth and intestinal morphology of broilers. *Poultry Sci*, 80(Suppl 1).
- Yi, G. F., Allee, G. L., Knight, C. D., and Dibner, J. J. (2005). Impact of glutamine and oasis hatchling supplement on growth performance, small intestinal morphology, and immune response of broilers vaccinated and challenged with *Eimeria maxima*. *Poultry Science*, 84(2), 283-293.
- Yin, Y., Huang, R., Li, T., Ruan, Z., Xie, M., Deng, Z., Wu, G. (2010). Amino acid metabolism in the portal-drained viscera of young pigs: effects of dietary supplementation with chitosan and pea hull. *Amino Acids*, 39(5), 1581-1587.
- Zhang, X. P., and Glaser, E. (2002). Interaction of plant mitochondrial and chloroplast signal peptides with the Hsp70 molecular chaperone. *Trends in Plant Science*, 7(1), 14-21.
- Zulkifli, I., Dunnington, E. A., Gross, W. B., and Siegel, P. B. (1994). Inhibition of adrenal steroidogenesis, food restriction and acclimation to high ambient temperatures in chickens. *British Poultry Science*, 35(3), 417-426.
- Zulkifli, I., and Siegel, P. B. (1995). Is there a positive side to stress?. *World's Poultry Science Journal*, 51(01), 63-76.
- Zulkifli, I. (1999). Heterophil/lymphocyte response and performance of feed and water restricted broiler chickens under tropical conditions. *Asian Australasian Journal of Animal Sciences*, 12, 951-955.
- Zulkifli, I., Norma, M. C., Israf, D. A., and Omar, A. R. (2000). The effect of early age feed restriction on subsequent response to high environmental temperatures in female broiler chickens. *Poultry Science*, 79(10), 1401-1407.

- Zulkifli, I., Liew, P. K., Israf, D. A., Omar, A. R., and Hair-Bejo, M. (2003). Effects of early age feed restriction and heat conditioning on heterophil/lymphocyte ratios, heat shock protein 70 expression and body temperature of heat-stressed broiler chickens. *Journal of Thermal Biology*, 28(3), 217-222.
- Zulkifli, I., and Siti Nor Azah, A. (2004). Fear and stress reactions, and the performance of commercial broiler chickens subjected to regular pleasant and unpleasant contacts with human being. *Applied Animal Behaviour Science*, 88(1), 77-87.
- Zulkifli, I., Htin, N. N., Alimon, A. R., Loh, T. C., and Hair-Bejo, M. (2007). Dietary selection of fat by heat-stressed broiler chickens. *Asian Australasian Journal of Animal Sciences*, 20(2), 245.
- Zulkifli, I. (2008). The influence of contact with humans on bird-to-bird pecking, fear-related behaviour, stress response, and growth in commercial broiler chickens and red jungle fowl when reared separately or intermingled. *Archiv Fur Geflugelkunde*, 72(6), 250-255.
- Zulkifli, I., Al-Aqil, A., Omar, A. R., Sazili, A. Q., and Rajion, M. A. (2009). Crating and heat stress influence blood parameters and heat shock protein 70 expression in broiler chickens showing short or long tonic immobility reactions. *Poultry Science*, 88(3), 471-476.

BIODATA OF STUDENT

Majid Shakeri was born in Mashhad, Iran, on 21 March 1985. He received his Bachelor of Science degree (Animal science) in University Iran, Birjand from 2006 to 2008. In 2011, he pursued his Master of Science degree majoring in Poultry Reproduction at the Institute of Tropical Agriculture, Universiti Putra Malaysia. He had participated in many research projects during his master degree. He is currently working with some companies related to his subject in Iran.



LIST OF PUBLICATIONS

Journal

1. M. Shakeri, I. Zulkifli, A. F. Soleimani, P.D. Eckersall, A. Anna Aryani, S. Kumari, and Faez Firdaus Jesse Abdullah. (2014). Response to dietary supplementation of L-glutamine and L-glutamate in broiler chickens raised at different stocking densities under the hot, humid tropical conditions. (Accepted by Poultry Science Journal).