

PHYTOCHEMICAL OF CHROMOLAENA ODORATA

MALA A/P GANESAN

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PHYTOCHEMICAL OF CHROMOLAENA ODORATA



By

MALA A/P GANESAN



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DEDICATION



My father, Ganesan a/I Muniandi and Mother, Mariammah a/p Ratnem

Lecturers, Family member and dear friends.

Thanks for giving such a love, support, patient and understanding.

ABSTRACT

C.odorata is commonly consumed as traditional medicine in some countries, including Malaysia. This study was aimed to identify the extractive values in C.odorata on a different type of solvents and parts of plants and analyzed its total phenolic contents (TPC) and phytochemical using Gas chromatography-mass spectrometry (GC-MS). In comparison with different solvents in parts which are leaf and stem extracts of C.odorata had the significantly different in yield of extraction and TPC, respectively. Among that, the leaf of this species had a great yield than the stem. The hot water extract on the leaf was found has the highest yield with hot water (27.96%) followed by methanol (26.56%), ethanol (26.33%), cold water (24.96%), and petroleum ether (5.54%) respectively. In the stem, the most extractive value was found in hot water (18.53%), followed by methanol (15.06%), cold water (13.46%), ethanol (12.40%), and petroleum ether (0.79%) respectively. In TPC, the leaf of this species had highest amount of phenolic compounds with hot water (331.80 ± 5.92 mg GAE/g DW) followed by methanol (318.71 ± 1.67 mg GAE/g DW) and petroleum ether (125.13 ± 8.84 mg GAE/g DW) and in stem the highest amount of phenolic compound was found in hot water(315.73± 2.37 mg GAE/g DW) followed by methanol (221.67 ± 0.01 mg GAE/g DW) and petroleum ether (12.84 ± 4.66 mg GAE/g DW). GCMS analysis of the leaf showed the presence of fatty acids, glycerides, sugar alcohols, alcohols, alkanes and chlorine compounds, and for the stem presence of alkanes, which contribute to its antioxidants agents.

ABSTRAK

C.odorata biasanya digunakan sebagai ubat tradisional di beberapa negara, termasuk Malaysia. Kajian ini bertujuan untuk mengenal pasti nilai-nilai ekstraktif dalam C.odorata pada jenis pelarut dan bahagian tumbuhan yang berlainan untuk menganalisis dengan jumlah kandungan penolik (TPC) dan fitokimia menggunakan spektrometri massa-kromatografi Gas (GC-MS). C.odorata mempunyai perbezaan dalam hasil pengekstrakan dan TPC. Di antara itu, daun spesies ini mempunyai hasil yang tinggi daripada batang. Ekstrak air panas pada daun didapati mempunyai hasil tertinggi dengan air panas (27.96%) diikuti oleh metanol (26.56%), etanol (26.33%), air sejuk (24.96%) dan petroleum eter (5.54%) masing-masing. Dalam batang, nilai yang paling ekstraktif didapati dalam air panas (18.53%), diikuti oleh metanol (15.06%), air sejuk (13.46%), etanol (12.40%), dan petroleum eter (0.79%) masing-masing. Dalam TPC, daun spesies ini mempunyai sebatian penolik yang tinggi dengan air panas (331.80 ± 5.92 mg GAE / g DW) diikuti oleh metanol (318.71 ± 1.67 mg GAE / g DW) dan ether petroleum (125.13 ± 8.84 mg GAE / g DW) dan dalam batang jumlah yang paling banyak terdapat sebatian penolik yang terdapat dalam air panas (315.73 ± 2.37 mg GAE / g DW) diikuti oleh metanol (221.67 ± 0.01 mg GAE / g DW) dan petroleum ether (12.84 ± 4.66 mg GAE / g DW). Analisis GCMS daun menunjukkan kehadiran asid lemak, gliserida, alkohol gula, alkohol, alkana dan sebatian klorin, dan untuk kehadiran batang alkana, yang menyumbang kepada agen-agen antioksidannya.

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APPROVAL SHEET

I certify that this research project entitles "Phytochemical of *Chromolaena Odorata*" by "Mala a/p Ganesan" has been examined and approved as a partial fulfillment of the requirements for the degree of Bachelor of Wood Science and Technology in the Faculty of Forestry, University Putra Malaysia.

Assoc .Prof. Dr. Rasmina Halis Faculty of Forestry Universiti Putra Malaysia (Supervisor)

Prof. Dr. Mohamed Zakaria Hussin Dean Faculty of Forestry Universiti Putra Malaysia

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LIST OF ABBREVIATIONS

ANOVA	Analysis of variance
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CH3OH Methanol

cm centimetre

DW Dry weight

G

GAE Gallic acid equivalent

hour

meter

gram

hr

m

ml millilitre

min minit

nm nanometre

NaOH Sodium hydroxide

Na₂SO₃ Sodium carbonate NaCl Sodium chloride

pH potential of hydrogen

Percentage

C

%

CHAPTER 1

INTRODUCTION

1.1 Background

Phytochemicals are defined as chemicals generate by plants. It improves the health condition of organisms, and those attributed to macronutrients and micronutrients. Phytochemicals are generally used to thrive the predators or pathogens in human (Bhanu *et al.*, 2011). There is wide evidence to support the health benefits of the diet in the form of fruits, vegetable, legumes, whole grains and nuts (Doss *et al.*, 2009).

Moreover, some plants also have medicinal elements such a shrub and herbs that provide colour, aroma, and flavour, and inviting attention from potential consumers. It also makes many chemical compounds for biological functions. Some Phytochemicals compounds that can be found in plants are carbohydrates, proteins, alkaloids, flavonoids, steroids, saponins, tannins and so on (Paulseit, 2010).

Apart from that phytochemical in some plants have been used as animal feed and others as traditional medicine. The leaves of some plants are also used to scent aromatic baths and used as emergency medication and prevention of chronic diseases (Farnidah, 2009). Therefore, the present study carried out for finding phytochemicals in species of *Chromleana odorata* with the

botanical aspect and to provide some information on the phytochemical constituents.

1.2 Problem statement and justification

The fresh leaves and extract of *C. odorata* are used as traditional herbal treatment and medicinal uses in some developing countries. This plant was uses for burns, soft tissue wounds and skin infections (Vijayaraghavan *et al.*, 2017). Leaves of *C.odorata* helps to reduce the appetite for smoking, cures fever, coughing, and stomachache (Defilipps, 2004). Although Malaysia is relatively abundant of *C. odorata*, the use of *C. odorata* is still limited. This plant even can use as animal feed, traditional herbal and as medicinal use. In previous studies, the research on *C. odorata* very scanty and quantification on its phytochemical not yet published. The species of *C. odorata* choose because of its sustainability abundant, and fast spread species and sustainability in medicinal. However, in Malaysia, *C.odorata* usage is very low compared to other Asian countries. For these reasons, a research must be conducted persistently to know what are the phytochemicals can be found in *C. odorata* and quantify phytochemical that has medicinal values and other values through GCMS.

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1.3 Objectives

General objective

 To quantify the phytochemical of *C. odorata* that available in idle areas in Malaysia.

Specific objectives

- To identify the extractive yield in *C.odarata* on a different type of solvents and parts of the plant.
- To analyse the phytochemicals of *C.odarata* on total phenolic compounds and GCMS

REFERENCES

Abegun, S. M. (2015). Comparison of efficiency of different solvents used for the extraction of phytochemicals from the Leaf, Seed and Stem Bark of Calotropis Procera. *Journal Article*, *4*(7).

Agostinis, P., Donella-Deana, A., Cuveele, J., Vandenbogaerde, A., Sarno, S., Merlevede, W. & Patrick D. (2011). Development of a simple, rapid and reproducible HPLC assay for the simultaneous determination of hypericins and stabilized hyperform in commercial St. John's Wort preparations. *Journal of Chromatography A*, 6(2), 1–8.

Aja, P. M., Enechi, O. C., Agu, K. A., Ikechukwu, A. A., & Nweke, O. L. (2016). Phytochemical Composition, Gas Chromatography-Mass Spectrometric (GC-MS) Analysis and Anti-Bacterial Activity of Ethanol Leaf-Extract of Ageratum conyzoides Department of Medical Biochemistry, Faculty of Basic Medicine, 8(1), 34–40.

Apori, S. O. (1992). Chromolaena Odorata : a Multipurpose Shrub, 558, 31–32.

Bairi Raju, C., & Vijaya, A. R. (2011). Elementary analysis of thespesia populnea fruits. *International Journal of Phytopharmacology*, *2*, *1*–6.

Bamisaye, F. A., Ajani, E. O., Nurain, I.O., & Minari, J. B. (2014). Medicobotanical investigation of siam weed (*Chromolaena odorata*) used among the " ljebu " people of Ogun state, Nigeria. *Journal of Medicine and Medical Sciences*, *5* (1), 20-24.

Bengal, W.(2014). Ethnobotanical Study in a Remote District of west Bengal,India. Department of Pharmaceutical Technology, Kolkata : Jadavpur University.

Bhanu, S., Vandana, S., & Archana, S. (2011). Vitro Study of Entamoeba Histolytica Causative Agent of Amoebiasis With Lemon Juice At Different

Concentration Showed Antiamoebic Properties. International Research Journal of Pharmacy, 2(9), 88–90.

Bruni, R.,& Sacchetti, G. (2009). Factors affecting polyphenol biosynthesis in wild and field grown St. John's Wort (Hypericum perforatum L. Hypericaceae/Guttiferae). *Journal of Molecules*, *14(2)*, *682–725*.

Bryan, N., & Loscalzo, J. (2011). Nitrite and nitrate in human health and disease. Nutrition and Health Series Adrianne Bendich, PhD, FACN, Series Editor.

Burritt, E. A., Malechek, J. C., & Provenza, F. D. (1987). Changes in concentrations of tannins, total phenolics, crude protein, and in vitro digestibility of browse due to mastication and insalivation by cattle. *Journal of Range Management*, 40, 409–411.

Dai, J., & Mumper, R. J. (2010). Plant phenolics: Extraction, analysis and their antioxidant and anticancer properties. *Journal of Molecules*, 15(10), 7313–7352.

DeFilipps, R., Maina, S., & Crepin, J. (2004). Medicinal plants of the Guianas (Guyana, Surinam, French Guiana). Department of Botany National Museum of Natural Smithsonian Institution Washington, D.C.300013-7012

Doss, A, Sc, M., & Phil, M. (2009). Preliminary phytochemical screening of some Indian Medicinal Plants. *International Journal of Pharmacy and Pharmaceutical Sciences* 29(2), 12–16.

Doughari, J. H. (2009). Phytochemicals : Extraction Methods , Basic Structures and Mode of Action as Potential Chemotherapeutic Agents. *A Global Perspective of Their Role in Nutrition and Health*, 1–33.

Edeoga, H. O., Okwu, D. E., & Mbaebie, B. O. (2005). Phytochemical constituents of some Nigerian medicinal plants. *Journal of Biotechnology, 4(7), 685–688.*

Edrah, S. M., Aljenkawi, A., Omeman, A., & Alafid, F. (2016). Qualitative and quantities analysis of phytochemicals of various extract for Ephedra altissima from Libya. *Journal of Medicinal Plants Studies*, 4(3), 119–121.

Farnidah H.J.(2009). *Biological Activities And Chemical Constituents Of Chromolaena odorata (L.) King & Robinson*. Master thesis, Faculty of Science, Universiti of Malaya Kuala Lumpur.

Guo, Z. (2017). The modification of natural products for medical use. *Acta Pharmaceutica Sinica B, 7(2), 119–136.*

Hanh, T. T. H., Hang, D. T. T., Van Minh, C., & Dat, N. T. (2011). Antiinflammatory effects of fatty acids isolated from Chromolaena odorata. *Asian Pacific Journal of Tropical Medicine*, 4(10), 760–763.

Iqbal, E., Salim, K. A., & Lim, L. B. L. (2015). Phytochemical screening, total phenolics and antioxidant activities of bark and leaf extracts of Goniothalamus velutinus (Airy Shaw) from Brunei Darussalam. *Journal of King Saud University - Science*, 27(3), 224–232.

Kayne, S. B. (2009). Introduction to traditional medicine. *Journal of Traditional Medicine: A Global Perspective,* 1–24.

Kennedy, D. O., & Wightman, E. L. (2011). Herbal extracts and phytochemicals: plant secondary metabolites and the enhancement of human brain function. *Advances in Nutrition*, 2(1), 32–50.

Kong, H. W., & Malaysia, F. R. I. (2009). Current Status of biomass utilization in Malaysia, Forest Research Institute Malaysia ,1–15.

Kostovlrenaa, Ojala, T., Lacy, A.,O'Kennedy, R., Widelski, J., Melliou, E., & Butler, M. S. (2010). Natural Product Chemistry For Drug Discovery. *Journal of Natural Products*, *5*(*8*), *440*.

Kouamé, P. B., Jacques, C., Bedi, G., Silvestre, V., Loquet, D., Barillé-nion, S., & Tea, I. (2013). Phytochemicals Isolated from Leaves of Chromolaena odorata Impact on Viability and Clonogenicity of Cancer Cell Lines. *Journal of Advances in Microbiology*, *835–840*

Kumar, H., Choudhary, N., Kumar, N., & Seth, R. (2014). Phenolic compounds and their health benefits. *Journal of food research and technology* 2(2), 46–59.

Kurmukov, A. G. (2013). Phytochemistry of medicinal plants. *Medicinal Plants of Central Asia: Uzbekistan and Kyrgyzstan. Journal of Pharmacognosy and Phytochemistry Phytochemistry 6, 13–14.*

L.Y. Ng, Y.K. Ang, H.E., & Khoo, H. S. Y. (2009). Influence of different extraction parameters on antioxidant properties of carica papaya peel and seed. *Research Journal of Phytochemistry*, *3*(2), *101–112*.

Li, L. (2000). Opportunity and challenge of traditional Chinese medicine in face of the entrance to WTO (World Trade Organization). *Chinese Journal of Information on Traditional Chinese Medicine*, *7*, *7*-8.

Luwum, P. (2002). Control of Invasive Chromolaena odorata Control of Invasive Chromolaena odorata, Degree thesis, Master thesis, Faculty of Science, Master thesis, Faculty of Science, Universiti of Pretoria.

Martins, S., Mussatto, S. I., Martínez-Avila, G., Montañez-Saenz, J., Aguilar, C. N., & Teixeira, J. A. (2011). Bioactive phenolic compounds: Production and extraction by solid-state fermentation, A review. *Biotechnology Advances*, 29(3), 365–373.

C

Milatos, G. I. (2010). Contributions of Systems - *Theory to the Understanding of Therapy and Health.* Publications about systems thinking in medicine since 2000.

Motaleb, M. A. (2011). Selcted Medicinal plants of chittagong hill tracts. Journal of International Union for conservation for nature,116.

Mullaicharam, A.R. (2014). A review on medicinal properties of Camel milk. *World Journal of Pharmaceutical Sciences, 1-10.*

Nweke, O. L., Nwachukwu, N., Aja, P. M., Agbafor, K. N., Nwaka, A. C., & Uchenna Ezeilo, R. (2015). Phytochemical and Gas Chromatograpy-Mass Spectrophotometeric (GC-MS) Analyses of Vitex doniana Leaf from Abakaliki, Ebonyi State. *IOSR Journal of Pharmacy and Biological Sciences Ver. III*, 10(5), 2319–7676.

O, O. K. (2015). Studies on the insecticidal properties of Chromolaena odorata (Asteraceae) against adult stage of Periplaneta americana. *Journal of Entomology and Zoology Studies*, 318(31), 318–321.

Offor, C. E., Ugwu, P. C., Aja, P. M., & Igwenyi, I. O. (2015). Proximate and Phytochemical Analyses of Terminalia catappa Leaves. *European Journal of Applied Sciences 7 (1): 09-11.*

Omar, M. H. (2013). Analysis of phytochemical in Malaysian medicinal plant and the bioavailability of dietary hyroxycinnamates. PhD thesis, Veterinary and Life Sciences for the degree of Doctor of Philosophy 1–196.

Palaniswamy, S., Mayilvanan, M., & Dharamani, A. S. (2009). Classification of functional groups of a phytochemical: Gymnemic acid using fuzzy logic. *Rasayan Journal of Chemistry*, *2*(2), 475–487.

Pandey, K. B., & Rizvi, S. I. (2009). Plant Polyphenols as Dietary Antioxidants in Human Health and Disease. *Oxidative Medicine and Cellular Longevity*, *2*(5), 270–278.

Paulseit Smestadn, B. (2010). Highlights through the history of plant medicine. Bioactive compounds in plants - benefits and risks for man and animals . National Veterinary Institute, Oslo, Norway, and Committee for Information and Research in Geomedicine, The Norwegian Academy of Science and Letters, Oslo, Norway. Prashant Tiwari, B., Kumar, M. K., & Gurpreet Kaur, H. K. (2011). Phytochemical screening and extraction A review. *Internationale Pharmaceutica Sciencia*, *1*(*1*), *98*–*106*.

Ramachandramoorthy, T., George, M. S., Balasubramaniyan, S., Rajasekar, K., Palanivelan, L., Govindharaju, R., Nadu, T. (2016). Phytochemical Screening and Antibacterial Activity of Ethanolic Extract of Terminalia Catappaflowers, *Asian Pacific Journal of tropical Biomedicine*, *6*(2), 345–349.

Rice-Evans, C. A., Miller, N. J., & Paganga, G. (1996). Structure-antioxidant activity relationships of flavonoids and phenolic acids. *Free Radical Biology and Medicine*, *20*(7), 933–956.

Rohman, A., Riyanto, S., Yuniarti, N., Saputra, W. R., Utami, R., & Mulatsih, W. (2010). Antioxidant activity, total phenolic, and total flavaonoid of extracts and fractions of red fruit (Pandanus conoideus Lam). *International Food Research Journal, 17(1), 97–106.*

Samshuddin, S., Kamath, S. D., Arunkumar, D., Avinash, N. G., & Samshuddin, S. (2015). Determination of total phenolic content and total antioxidant activity in locally consumed food stuffs in Moodbidri, Karnataka, India. *Advances in Applied Science Research*, *6*(*6*), *99*–102.

Scalbert, A., Manach, C., Morand, C., Rémésy, C., & Jiménez, L. (2005). Dietary polyphenols and the prevention of diseases. *Critical Reviews in Food Science and Nutrition*, *45*(*4*), 287–306.

Surh, Y. (2003). Phytochemicals. Nature Reviews Cancer, 8.pages

Swami Handa, S., Singh Khanuja, S. P., Longo, G., & Dutt Rakesh, D. (2008). Extraction techniques of medicinal plants. *Extraction Technologies for Medicinal and Aromatic Plants*, *1–10.*

Tsibranska, I., & Tylkowski, B. (2016). Solid-liquid extraction of bioactive compounds: Effect of polydispersity and particle size evolution. *Journal of Chemical Technology and Metallurgy*, *51(5)*, *489*–499.

Udobi, C. E., & Onaolapo, J. A. (2009). Phytochemical analysis and antibacterial evaluation of the leaf stem bark and root of the African locust bean (Parkia biglobosa). *Journal of medicinal plant research*, 3(5), 338–344.

Van De Wier, B., Koek, G. H., Bast, A., & Haenen, G. R. M. M. (2017). The potential of flavonoids in the treatment of non-alcoholic fatty liver disease. *Critical Reviews in Food Science and Nutrition*, *57*(*4*), *834–855*.

Vijayaraghavan, K., Rajkumar, J., & Seyed, M. (2017). Efficacy of Chromolaena odorata leaf extracts for the healing of rat excision wounds. *Veterinární Medicína*, 62 (10), 565–578.

Wangchuk, P. (2014). Phytochemical analysis, bioassays and the identification of drug lead compounds from seven Bhutanese medicinal plants. University of Wollongong Thesis Collection, 1–238. Unpublished.

World Health Organisation (WHO). (2013).WHO Traditional Medicine Strategy 2014-2023, 1–76.

Zhang, S., Bi, H., & Liu, C. (2007). Extraction of bio-active components from Rhodiola sachalinensisunder ultrahigh hydrostatic pressure. Separation and Purification Technology. *Journal of analytical science*, *57*, *275–280*.

Zheng, R., Su, S., Li, J., Zhao, Z., Wei, J., Fu, X., & Liu, R. H. (2017). Recovery of phenolics from the ethanolic extract of sugarcane (Saccharum officinarum L.) baggase and evaluation of the antioxidant and antiproliferative activities. *Industrial Crops and Products, 107, 360–369.*