



UNIVERSITI PUTRA MALAYSIA

***MORPHOLOGY IDENTIFICATION AND MOLECULAR
CHARACTERIZATION OF *Lasiodiplodia theobromae* CAUSING FRUIT
ROT DISEASE OF GUAVA (*Psidium guajava* L.) IN MALAYSIA***

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Morphology Identification and Molecular Characterization of *Lasiodiplodia theobromae* Causing Fruit Rot Disease of Guava (*Psidium guajava* L.) in
Malaysia

By

Rohaida binti Che Ludin

A final year project report submitted to the Faculty of Agriculture, Universiti Putra Malaysia in fulfilment of the requirement of PRT 4999 (PROJECT) for the award of the Degree of Bachelor of Horticultural Science

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CERTIFICATION

This project paper entitled, “Morphology Identification and Molecular Characterization of *Lasiodiplodia theobromae* Causing Fruit Rot Disease of Guava (*Psidium guajava* L.) in Malaysia” is prepared by Rohaida binti Che Ludin and submitted to the Faculty of Agriculture in partial fulfilment of the requirement of PRT 4999 (Project) for the award of the degree of Bachelor of Horticultural Science.

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ABSTRACT

Guava (*Psidium guajava* L.) belongs to the family of Myrtaceae is native to tropical America. Postharvest diseases, which cause serious problems during storage and transportation of guava fruits, are the major factors that limit the thriving guava industry in Malaysia. The fruit rot diseases reduce the fruit quality by altering the consistency, colour, taste and reduce shelf-life. Fruit samples from guava peel showing brownish black lesions of irregular shape and rapid spread of internal decay may appear. Few studies have investigated fruit rot disease on guava, however, only morphological characteristics were used and no molecular characterization was conducted to identify the pathogen identification. The objectives of this study were 1) to identify primary pathogen that caused guava fruit rot based on morphological and cultural characters as well as analyses of nucleotide sequences of the internal transcribed spacer (ITS) region of ribosomal DNA 2) to construct ITS phylogeny based on ITS sequences and their closest relatives by using Maximum Likelihood method. In order to identify the fruit rot-causing agent, 10 symptomatic fruits were collected from Seri Kembangan Selangor farmers market and fungi were isolated from fruit samples. One species, which is *Lasiodiplodia theobromae* was isolated frequently from the infected guava fruit. Colony morphology, and growth rates of the representative isolates were determined from the single conidium isolate obtained from the guava fruits. The pathogen identity was compared based on amplified ITS sequence using Basic Logical Alignment Search Tool (BLAST) nucleotide searches at GenBank. The findings of this study confirmed the true identity of the pathogen associated with fruit rot disease of guava in Malaysia was 99% matched with BLASTn search and ITS sequence was from *Lasiodiplodia theobromae*. Correct identification of this pathogen is helpful for more effective management of fruit rot disease of guava. Further screening of effective fungicides and understanding of the epidemiology of this fungal pathogen will help to reduce financial loss to the guava industry in Malaysia.

ABSTRAK

Jambu batu (*Psidium guajava* L.) adalah dari keluarga Myrtaceae yang berasal dari Amerika tropika. Penyakit pasca tuai, yang menyebabkan masalah yang sangat besar semasa penyimpanan dan penghantaran buah jambu batu, adalah faktor yang utama yang menghalang pengembangan industri jambu batu di Malaysia. Penyakit reput buah mengurangkan kualiti buah dengan mengubah struktur, warna, rasa dan mengurangkan jangka hayat. Sampel buah menunjukkan symptom reput buah yang berwarna coklat kehitaman dalam bentuk yang tidak teratur dan reput dalaman buah cepat merebak akan kelihatan. Terdapat beberapa kajian terhadap penyakit reput buah jambu batu, namun, hanya melalui kaedah pengenalpastian morfologi yang dijalankan dan tiada kaedah pencirian secara molekul dijalankan untuk mengenal pasti identiti patogen. Objektif kajian ini adalah untuk 1) mengenal pasti patogen yang menyebabkan reput buah jambu batu berdasarkan penegenalpastian morfologi dan kultura dan juga jujukan nukleotid bahagian *internal transcribed spacer* (ITS) pada ribosom DNA, 2) untuk membina filogeni ITS berdasarkan jujukan ITS dan spesies kulat berdekatan dengan menggunakan kaedah *Maximum Likelihood*. Untuk mengenalpasti agen penyakit reput buah, 10 biji buah yang menunjukkan simptom reput buah telah diambil dari pasar basah Selangor dan kulat-kulat tersebut telah diasingkan daripada sampel buah terinfeksi. Satu spesis *Lasiodiplodia theobromae* telah diasingkan beberapa kali daripada buah jambu batu yang dijangkiti. Morfologi koloni dan kadar pertumbuhan telah dikenalpasti dari wakil kulat yang telah diasingkan daripada satu konidium yang didapati daripada jambu batu. Identiti patogen telah dibandingkan berdasarkan jujukan ITS yang telah dibanyakkan menggunakan *Carian Alat Susunan Logik (BLAST)* nukleotida dari GenBank. Kajian ini membuktikan identiti sebenar patogen yang menyebabkan penyakit reput buah jambu batu di Malaysia adalah 99% sepadan dengan keputusan BLAST dan jujukan ITS adalah daripada spesis *Lasiodiplodia theobromae*. Pengenalan yang betul patogen ini sangat membantu untuk pengurusan yang

lebih berkesan terhadap penyakit reput buah jambu batu. Pemeriksaan lebih mendalam terhadap racun kulat yang efektif dan pemahaman terhadap epidemiologi kulat ini akan membantu dalam mengurangkan kerugian dalam industri jambu batu di Malaysia.



CHAPTER 1

Introduction

1.1 Background of Study

Psidium guajava or most known as guava is from family Mytaceae. Its generics name is determined from the Greek expression "psidion" name of the pomegranate, same time its scientific name is came from Spanish word "guayabe" implying guava tree, itself continuously determined from those Arawakan by means of the Cariban through those Tupian. Those Spaniards brought the fruits over those Pacific to the Philippines in 1526, named as "guayabas" or "bayabas" got to be great built (Morton, 1987). Starting with here those fruits spread through Southeast and East Asia and India by Portuguese voyagers in the early seventeenth century. Starting with India the fruit have been disseminated on her neighbors and to Arabia. Today the fruits could be discovered developing for more than fifty nations all around those tropics and subtropics including a part of the Mediterranean territories for example, inside the cutoff points about scope 35°N and 35°S of the equator (Lim & Khoo, 1990).

In Malaysia, guava is known as Jambu Batu. In spite of guava have been planted over Malaysia to quite a while it have been main in later a considerable length of time that the crop pulled in with great arrangement of consideration (Lim & Khoo, 1990). Those zone under guava might have been with the goal inconsequential that it did not actually value a posting under incidental harvests in the 1984 Annual Report of the Department of Agriculture, Peninsular Malaysia. Because of demand, guavas are now being grown or cultivated at warm and humid and also subtropical territory. Imperative guava-growing regions are placed in the states of Perak, Johor, Selangor, and Negeri Sembilan. Different states that about lesser importance are Malacca, Kelantan, Terengganu, Perlis, Kedah, Pahang, Sabah and Sarawak

(Anem, 2013. retrieved from <http://animhosnan.blogspot.my/2010/12/guava-clone-in-malaysia.html>).

In Malaysia guava is habitually planted likewise the sole crop. It is not, nonetheless unprecedented to figure the tree developed concerning illustration as inter-crop between rows of other tree grown foods products for example, such that durian, alternately in discrete pieces inside the blended crop orchards. The fruit appearance is round or ovary based on the species with its long from 3 on 14cm. The rough skin generally transformed from green to yellow when ripen. Guava basically have certain types of fragrance odor. Guava flesh might sweet or sour. Those flesh shade starting with white to profound pink with rely on upon species. In Malaysia there are several varieties that give economic importance, which are Gu4, Gu5, Gu7, Jambu Biji, Laknaw, Taiwan, Hongkong pink, Vietnamese and Red Malaysian. The guava skin contrast from pink to red. Guava species Gu4, Gu5, Gu7, Jambu biji and Hongkong pink with diameter from 4. 8 cm to 6. 7 cm and slender skin from 0. 5-1. 5cm will be littler and slender compared to guava Vietnamese, its skin breadth from 10 to 11cm and thickness from 1. 9 cm on 2. 5 cm. There are two top season for fruit collecting which is January until March and October to December (retrieved from <http://www.malaysiafruit.com/guava/guava.php>). The moderate yield of the seeded cultivars can extent to 50 MT/ha/year. By there, there are amount about issues in generation which influence expense and which in turn, influence the competitiveness of the guava both provincially and abroad.

The greater issues that may be posed is because of the pests and diseases. The tropical climate, described by high mean temperatures of 28-30°C, high relative moistness and overwhelming yearly rainfall about 2000 mm, may be favorable of the fast burgeoning and speedy pest proliferation, weeds and sicknesses. Under such climatic regime, those gainful productions for fruit yields necessitates costly control measures to decrease misfortunes. Ripe guavas simply wound and exceptionally perishable. Fruits for processing might be harvested

using mechanical tree-shakers or plastic nets. Specialists in Kurukshetra University, India, have demonstrated that treatment of harvested guavas with 100 ppm morphactin (chlorflurenol methyl ester 74050) surge shelf life of guavas by controlling contagious fungal decay, and lessening the color, weight, sugars, ascorbic acids and non-volatile organic acids (Morton, 1987). Joined together fungicidal and double-wax covering would extend the marketability by 30 days. Throughout the rainy season in India, and the area of Sancti Spiritus, Cuba, the fungus, *Phytophilous parasitica*, is answerable for much irresistible tree grown fruit rots. *Botryodiplodia* sp. and *Dothiorella* sp. caused stem-end decay in fruits injury during harvesting. *Macrophomina* sp. has been interfaced to fruit rot in Venezuela and *Gliocladium roseum* has been recognized looking into rotting fruits on the market in India (Morton, 1987).

1.2 Justification of the study

In Malaysia, researches on the guava postharvest diseases are very limited. The main focus in Malaysia majority are only on the main crops such as oil palm, rubber, paddy and cocoa. This limitation causing the importance of guava fruit in Malaysia's economic is being neglected.

1.3 Objectives of the study

- 1) To characterize and identify the causal agent causing guava fruit rot based on morphological and cultural characters
- 2) To construct internal transcribed spacer (ITS) phylogeny based on ITS sequence and their closest relatives by using Maximum Likelihood method

REFERENCES

- Abdollahzadeh, J., Javadi, A., Goltapeh, E. M., Zare, R., & Phillips, A. (2010).** Phylogeny and morphology of four new species of *Lasiodiplodia* from Iran. *Persoonia*, 25, 1–10.
- Alves, A., Correia, A., Luque, J., Phillips A. J. L. (2004).** *Botryosphaeria corticola* sp. nov. on *Quercus* species, with notes and description of *Botryosphaeria stevensii* and its anamorph *Diplodia mutila*. *Mycologia* 96:598–613.
- Alves, A., Correia, A., Phillips, A.J.L. (2006).** Multi-gene genealogies and morphological data support *Diplodia cupressi* sp. nov., previously recognized as *D. pinea* f. sp. *cupressi*, as a distinct species. *Fungal Diversity* 23: 1–15.
- Alves, A., Crous, P. W., Correia, A. C., & Phillips, A. J. L. (2008).** Morphological and molecular data reveal cryptic species in *Lasiodiplodia theobromae*. *Fungal diversity* . retrieved from <https://www.researchgate.net/publication/40095352>
- Alves, A., Crous, P.W., Correia, A. and Phillips, A.J.L. (2008).** Morphological and molecular data reveal cryptic speciation in *Lasiodiplodia theobromae*. *Fungal Diversity* 28: 1-13.
- Anem, M. (2013).** Guava Clone in Malaysia. Retrieved from <http://www.animhosnan.blogspot.my/2010/12/guava-clone-in-malaysia.html>
- Arbeit, R. D., Arthur, M. R. D. Dunn, C. Kim, R. K. Selander, and Goldstein, R. (1990).** Resolution of recent evolutionary divergence among *Escherichia coli* from related lineages: the application of pulsed field gel electrophoresis to molecular epidemiology. *J. Infect. Dis.* 161:230–235.
- Bagyalakshmi, R., K.L. Therese, S. Prasanna et al. (2008).** Newer emerging pathogens of ocular non-sporulating molds (NSM) identified by polymerase chain reaction (PCR)-based DNA sequencing technique targeting internal transcribed spacer (ITS) region. *Curr. Eye Res.* 33: 139-147.
- Bashir, H. A. and Abu-Goukh, A. A. (2003).** Compositional changes during guava fruit re-ripening. *Journal of Food Chemistry*, 80(4): 557-563.
- Binder M. & Hibbett D., (2003).** Hibbett lab protocols for DNA isolation, PCR, and DNA sequencing. PCR protocols, a guide to methods and applications. p282-287. Academic Press, San Diego, U.S.A.
- Borresen, A. L., E. Hovig, and A. Brogger. (1988).** Detection of base mutations in genomic DNA using denaturing gradient gel electrophoresis (DGGE) followed by transfer and hybridization with gene-specific probes. *Mutat. Res.* 202:77-83.
- Bridge, P.D., Spooner, B.M. and Roberts, P.J. (2005).** The impact of molecular data in fungal systematics. *Adv. Bot. Res.*, 42:33–67.
- Brown, B.I and Wills, R.B.H. (1983).** Post-harvest changes in guava fruit of different maturity. *Sci. Hortic.* 19:23-243
- Burgess T.I. Barber. P.A., Mohali, S., Pegg, G., de Beer, W., and Wingfield, M.J. (2006).** Three new *Lasiodiplodia* spp. from the tropics, recognized based on DNA sequence comparisons and morphology. *Mycologia*: 98(3), 2006, pp. 423–435. The Mycological Society of America, Lawrence, KS 66044-8897
- Cenis J.L., (1992).** Rapid extraction of fungal DNA for PCR amplification. *Nucleic Acids Research*, Vol. 20, No. 9.
- Chihlar, R.L., Sypherd, P.S. (1980).** The organization of the ribosomal RNA genes in the fungus *Mucor racemosus*. *Nucl. Acids Res.* 8: 793–804

- Comas, I., Moya, A., and Gonzales-Candelas, F. (2007).** From Phylogenetics to Phylogenomics: The Evolutionary Relationships of Insect Endosymbiotic γ -Proteobacteria as a Test Case. *Syst. Biol.* 56(1):1–16
- Cornell, M.J., Alam, I., Soanes, D.M., Wong, H.M., Hedeler, C., Paton, N.W., Rattray, M., Hubbard, S.J., Talbot, N.J., Oliver, S.G. (2007).** Comparative genome analysis across a kingdom of eukaryotic organisms: specialization and diversion in the fungi. *Genome Res.* 17:1809-822
- Crous, P. W., Slippers, B., Wingfield, M. J., Rheeder, J., Marasas, W. F. O., Phillips, A. J. L., Alves, A., Burgess, T., Barber, P., and Groenewald, J. Z. (2006).** Phylogenetic lineages in the *Botryosphaeriaceae*. *Stud. Mycol.* 55:235- 253.
- Cubeta, M.A., Echandi, E., Abernethy, T., Vilgalys, R. (1991).** Characterization of anastomosis groups of binucleate *Rhizoctonia* species using restriction analysis of an amplified ribosomal RNA. *Phytopathology* 81: 1395–1400.
- Delsuc, F., Brinkmann, H., Philippe, H., (2005).** Phylogenomic and the reconstruction of the tree of life. *Nature Reviews Genetics.* Vol.6: 361-373. : <https://www.researchgate.net/publication/7877411>
- Denman S, Crous PW, Taylor JE, Kang J-C, Pascoe I, Wingfield MJ. (2000).** An overview of the taxonomic history of *Botryosphaeria*, a re-evaluation of its anamorphs based on morphology and ITS rDNA phylogeny. *Stud Mycol* 45:129–140.
- Ebersberger, I., Simoes, R. D. M., Kupczok, A., Gube, M., Kothe, E., Voigt, K., von Haeseler, A. (2012).** A Consistent Phylogenetic Backbone for the Fungi. *Research article MBE Advance Access.* <http://mbe.oxfordjournals.org/>
- Erlich, H.A., Gelfand, D.H., Saiki, R.K. (1988).** Specific DNA amplification. *Nature*;331:4612.
- Finney, M. (1993).** Pulsed-field gel electrophoresis, p. 2.5.9–2.5.17. In F. M. Ausubel, R. Brent, R. E. Kingston, D. D. Moore, J. G. Seidman, J. A. Smith, and K. Struhl (ed.), *Current protocols in molecular biology*, vol. 1. Current Protocols, Greene-Wiley, New York
- Garber, R.C, Yoder, O.C. (1983).** Isolation of DNA from filamentous fungi and separation into nuclear, mitochondrial, ribosomal and plasmid components. *Anal. Biochem.* 135: 416–422.
- Gatesy, J. and Baker, R.H. (2005).** Hidden likelihood support in genomic data: can forty-five wrongs make a right? *Syst. Biol.* 54:583-492
- Gray, M., A. Charpentier, K. Walsh, P. Wu, and W. Bender. (1991).** Mapping point mutations in the *Drosophila rosy* locus using denaturing gradient gel blots. *Genetics* 127:139-149.
- Gupta, V.P., Tewari, S.K., Govindaiah, P. and Bajpai, A.K. (1999).** Ultrastructure of mycoparasitism of *Trichoderma*, *Gliocladium* and *Laetisania* sp. on *Botryodiplodia theobromae*. *Journal Phytopathology*, Vol.147, p19-24
- Hall, T.A. (1999).** BioEdit: a user-friendly biological sequence alignment editor and analysis program for windows 95/98/NT. Oxford University Press. *Nucleic Acid Symposium Series.* No.41;95-98
- Hashem M., and Alamri S. (2009).** The biocontrol of postharvest disease (*Botryodiplodia theobromae*) of guava (*Psidium guajava* L.) by the application of yeast strains. King Khalid University, Faculty of Science, Biological Science Department
- Henry, T., Iwen, P.C. and Hinrichs, S.H. (2000).** Identification of *Aspergillus* species using internal transcribed spacer regions 1 and 2. *J. Clin. Microbiol.*, 38:1510–5
- Hibbett, D. S., Binnder, M., Bischoff, J.F., Blackwell, M., Cannon, F. and Eriksson, O.E. (2007).** A higher-level phylogenetic classification of the Fungi. *Mycological Research* 111, 509-547. journal homepage: www.elsevier.com/locate/mycres

- Horton, T.R. and Bruns, T.D. (2001).** The molecular revolution in ectomycorrhizal ecology: peeking into the black-box. *Mol. Ecol.*, 10:1855–71.
- Iwen, P. C., Hinrichs, S. H. & Rupp, M. E. (2002).** Utilization of the internal transcribed spacer regions as molecular targets to detect and identify human fungal pathogens. Department of Pathology and Microbiology, University of Nebraska Medical Center, Omaha, Nebraska USA; Internal Medicine, University of Nebraska Medical Center, Omaha, Nebraska, USA. *Medical Mycology*, 40, 87-109
- Kumar S., Stecher G., and Tamura K. (2016).** MEGA7: Molecular Evolutionary Genetics Analysis version 7.0 for bigger datasets. *Molecular Biology and Evolution* 33:1870-1874.
- Larignon, P., Fulchic, R., Cere, L., and Dubos, B. (2001).** Observations of Black dead arm in French vineyards. *Phytopathol. Mediterr.* 40:336-342.
- Lehoczky, J. 1974.** Black dead arm disease of grapevine caused by *Botryosphaeria stevensii* infection. *Acta Phytopathol. Hung.* 9:319-327.
- Li, Q., Huang, S., Guo, T. and Mo, J. (2015).** Molecular phylogenetics of mango gummosis-causing agents in Guangxi.
- Lim, T. K. & Khoo, K. C. 1990.** Guava in Malaysia: Production, pest and diseases. Tropical Press Sdn. Bhd. Kuala Lumpur Malaysia. P11-188
- Liu, J.K., Phookamsak, R. Doilom, M. (2012).** Towards a natural classification of *Botryosphaeriales*. *Fung. Div.* 57: 149-210.
- Liu, Y., Steenkamp, E.T., Brinkmann, H., Forget, L., Philippe, H., and Lang, B.F. (2009).** Phylogenomic analyses predict sistergroup relationship of nucleariids and fungi and paraphyly of zygomycetes with significant support. *B.M.C. Evol. Biol.* 9:272
- Marcet-Houben, M., and Gabaldon, T. (2009).** The tree versus the forest: the fungal tree of life and the topological diversity within the yeast phylome. *PLoS One.* 4:e4357
- Marques, M.W., Michereff, S.J., Phillips, A.J.L. and Camara, M.P.S. (2012).** Species of *Lasiodiplodia* associated with mango in Brazil
- Marques, M.W., Lima, N.B., de Moraes, M.A. Jr., Barbosa, M.A.G., Souza, B.O., Michereff, S.J., Phillips, A.J.L. and Camara, M.P.S. (2014).** Species of *Lasiodiplodia* associated with mango in Brazil *Fungal Divers.* In press
- Martín, C., Moreno, C., and Gubler, W. D. (2006).** Occurrence of *Botryosphaeria obtusa*, *B. dothidea* and *B. parva* associated with grapevine trunk diseases in Castilla y León region, Spain. *Plant Dis.* 90:835. 48.
- Maslow, J. N., A. M. Slutsky, and R. D. Arbeit. (1993).** Application of pulsed-field gel electrophoresis to molecular epidemiology, p. 563–572. In D. H. Persing, T. F. Smith, F. C. Tenover, and T. J. White (ed.), *Diagnostic molecular microbiology: principles and applications.* American Society for Microbiology, Washington, D.C.
- Morton, J.F. (1987).** Fruits of warm climates. *Creative Resource Syst. Inc.* p356-363. Miami, FL.
- Motohashi, K., Inaba, S., Anzai, K., Takamatsu, S. and Nakashima, C. (2009).** Phylogenetic analyses of Japanese species of *Phyllosticta sensu stricto*
- Mullis, K.B., Faloona, F.A. (1987).** Specific synthesis of DNA in vitro via a polymerase-catalyzed chain reaction methods. *Enzymol;* 155:335–50.
- Muyzer, M., De Waal, E.C., Uitterlinden, A.G. (1992).** Profiling of Complex Microbial Populations by Denaturing Gradient Gel Electrophoresis Analysis of Polymerase Chain Reaction-Amplified Genes Coding for 16S rRNA. Applied And Environmental Microbiology, Mar. 1993, P. 695-700 0099-2240/93/030695-06\$02.00/0 Copyright C) 1993, American Society for Microbiology; Vol. 59, No. 3
- Netto, M.S.B., Correia, K.C., Michereff, S.J. and Camara, M.P.S. (2015).** Species of *Botryosphaeria* associated with *Anacardium* spp. in Brazil.

- Nunes, F.M., de Oliveira, M., Arriaga A.M.C., Lemos, T.L.G., Andrade-Neto, M., de Mattos, M.C., Mafezoli, J., Viana, F.M.P., Ferreira V.M., Rodrigues-Filho, E. and Ferreira, G. (2008). A New Eremophilane-type Sesquiterpene from the Phytopatogen Fungus *Lasiodiplodia theobromae* (Sphaeropsidaceae). *J. Braz. Chem. Soc.*, Vol. 19, No. 3, 478-482
- Oscar F. C., Crespo, A., Fatehi, J., and Bridge, P.D. (1999). DNA extraction and PCR amplification method suitable for fresh, herbarium-stored, lichenized, and other fungi. *Plant Systematics and Evolution* © Springer-Verlag 1999 Printed in Austria *Pl. Syst. Evol.* 216:243-24
- Pavlic, D., Slippers, B., Coutinho T.A., Gryzenhout, M., and Wingfield M.J. (2004). *Lasiodiplodia gonubiensis* sp. nov., a new *Botryosphaeria* anamorph from native *Syzygium cordatum* in South Africa. *Studies in Mycology*, 50: 313–322.
- Perez-Martinez, S. and Sosa del Castillo, D. (2015). Co-occurrence of pathogenic and not pathogenic *Fusarium decemcellulare* and *Lasiodiplodia theobromae* isolates within cushion
- Phillips, A. J. L. (2002). *Botryosphaeria* species associated with diseases of grapevines in Portugal. *Phytopathol. Mediterr.* 41:3-18.
- Phillips, A.J., Alves, A., Abdollahzadeh, J. (2013). The *Botryosphaeriaceae*: genera and species known from culture. *Stud. Mycol.* 76: 51-167.
- Phillips, A.J.L., Alves, A., Correia, A. and Luque, J. (2005). Two new species of *Botryosphaeria* with brown, 1-septate ascospores and *Dothiorella* anamorphs. *Mycologia* 97: 513-529.
- Phillips, A.J.L., Oudemans, P.V., Correia, A. and Alves, A. (2006). Characterisation and epitypification of *Botryosphaeria corticis*, the cause of blueberry cane canker. *Fungal Diversity* 21: 141-155.
- Ploetz, R.C. (2003). Anthracnose of mango; Management of the most important pre- and post-harvest disease. University of Florida, Department of Plant Pathology.
- Punithalingam, E. (1976). *Botryodiplodia theobromae*. Description of Pathogenic Fungi and Bacteria. *Commonwealth Mycological In- Plant Disease*. 2008. Institute, Kew, Surrey, England. pp519-529
- Punithalingam, E. (1980). Plant diseases attributed to *Botryodiplodia theobromae*. In: *Biblioteca Mycologica*. J. Cramer, Berlin
- Raeder, U., Broda, P. (1985). Rapid preparation of DNA from filamentous fungi. *Appl. Microbiol.* 1: 17–20.
- Rahman, M., Begum, K., Begum M. and Faruque, C.A.A. (2003). Correlation and path analysis in guava. *Bang. J. Agric. Res.*, 28: 93–98
- Razak, A.R., and Lim T.K. (1987). Occurrence of the Root-knot nematode *Meloidogyne incognita* on Guava in Malaysia. *Pertanika*: 10(3), 265-270
- Robbertse, B., Reeves, J.B., Schoch, C.L., Spatafora, J.W. (2006). A phylogenomic analysis of the Ascomycota. *Fungal Genetic Biol.* 43:715-725
- Rodriguez-Galvez, E., Guerrero, P., Barradas, C., Alves, A., and Crous, P.W. (2016). Phylogeny and pathogenicity of *Lasiodiplodia* species associated with dieback of mango in Peru. *Fungal Biol*
- Saiki, R.K., Bugawan, T.L., Horn, G.T., Mullis, K.B., Erlich, H.A. (1986). Analysis of enzymatically amplified beta-globin and HLA-DQ alpha DNA with allele-specific oligonucleotide probes. *Nature*; 324:163–6.
- Saiki, R.K., Gelfand, D.H., Stoffel, S. (1988). Primer-directed enzymatic amplification of DNA with a thermostable DNA polymerase. *Science*; 239:487–91.
- Saiki, R.K., Gelfand, D.H., Stoffel, S. (1988). Primer-directed enzymatic amplification of DNA with a thermostable DNA polymerase. *Science*; 239:487–91.

- Saiki, R.K., Scharf, S., Faloona, F. (1985).** Enzymatic amplification of β -globin genomic sequences and restriction site analysis for diagnosis of sickle cell anemia. *Science*; 230:1350–4.
- Sankoff D., Leduc, G., Antine, N., Paquin, B., Lang, B. F., and Cedergren, R. (1992).** Gene order comparisons for phylogenetic inference: Evolution of the mitochondrial genome. *Proc. Natl. Acad. Sci. USA* Vol. 89, pp. 6575-6579.
- Scharf, S.J., Horn G.T., Erlich, H.A. (1986).** Direct cloning and sequence analysis of enzymatically amplified genomic sequences. *Science*; 233:1076–8.
- Singh, S.P. and Pal, R.K. (2008b).** Response of climacteric-type guava (*Psidium guajava* L.) to postharvest treatment with 1-MCP. *Postharvest Biol. Technol.* 47:307–314.
- Slippers B, Wingfield M.J. (2007).** Botryosphaeriaceae as endophytes and latent pathogens of woody plants: diversity, ecology and impact. *Fungal Biology Reviews* 21: 90–106.
- Slippers, B., Crous, P.W., Denman, S., Coutinho, T.A., Wingfield, B.D. and Wingfield, M.J. (2004a).** Combined multiple gene genealogies and phenotypic characters differentiate several species previously identified as *Botryosphaeria dothidea*. *Mycologia* 96: 83-101.
- Slippers, B., Fourie, G., Crous, P.W. (2004b).** Speciation and distribution of *Botryosphaeria* spp. on native and introduced Eucalyptus trees in Australia and South Africa. *Studies in Mycology* 50: 343–358
- Stull, T. L., J. J. Li Puma, and Edlind, T. D. (1988).** A broad-spectrum probe for molecular epidemiology of bacteria: ribosomal RNA. *J. Infect. Dis.* 157:280–286.
- Sugita, T., Nishikawa, A., Ikeda, R. (1999).** Identification of medically relevant *Trichosporon* species based on sequences of internal transcribed spacer regions and construction of a database for *Trichosporon* identification. *J. Clin. Microbiol.*, 37:1985–93
- Tamura K. and Nei M. (1993).** Estimation of the number of nucleotide substitutions in the control region of mitochondrial DNA in humans and chimpanzees. *Molecular Biology and Evolution* 10:512-526.
- Taylor, A., Hardy, G. E. St. J., Wood, P., and Burgess, T. (2005).** Identification and pathogenicity of *Botryosphaeria* species associated with grapevine decline in Western Australia. *Aust. Plant Pathol.* 34:187-195
- Telford, M.J. (2007).** Phylogenomics. *Curr. Biol.* 17:R945–R946.
- Tenover, F. C. (1985).** Plasmid fingerprinting. A tool for bacterial strain identification and surveillance of nosocomial and community-acquired infections. *Clin. Lab. Med.* 5:413–436.
- Uitterlinden, A. G., P. E. Slagboom, D. L. Knook, and J. VUg. (1989).** Two-dimensional DNA fingerprinting of human individuals. *Proc. Natl. Acad. Sci. USA* 86:2742-2746.
- Úrbez-Torres J. R. (2008).** Identification and Pathogenicity of *Lasiodiplodia theobromae* and *Diplodia seriata*, the Causal Agents of Bot Canker Disease of Grapevines in Mexico. *Plant Dis.* 92:519-529
- Úrbez-Torres, J. R., Leavitt, G. M., Voegel, T., and Gubler W. D. (2006).** Identification and distribution of *Botryosphaeria* species associated with grapevines cankers in California. *Plant Dis.* 90:1490-1503.
- Van Belkum, A. (1994).** DNA fingerprinting of medically important microorganisms by use of PCR. *Clin. Microbiol. Rev.* 7:174–184.
- Van Niekerk, J. M., Crous, P. W., Groenewald, J. Z., Fourie, P. H., and Halleen, F. (2004).** DNA phylogeny, morphology and pathogenicity of *Botryosphaeria* species on grapevines. *Mycologia* 96:781-798.
- Von Arx, J.A. (1987).** Plant-pathogenic Fungi. J. Cramer, Berlin, Germany
- Von Arx, J.A., (1970).** A revision of the fungi classified as *Gloeosporium*. Lehre, Germany: J. Cramer.

- Von Arx, J.A., (1981).** The genera of fungi sporulating in pure culture, 3rd edn. Vaduz, Germany: *J. Cramer*.
- Wojciech J. J. & Lise K., (2002).** Biological control of postharvest diseases of fruits. *Annu. Rev. Phytopathol.* 2002. 40:411–41
- Zare, & Phillips, 2010.** Phylogeny and morphology of four new species of *Lasiodiplodia* from Iran. *Research Article Persoonia* 25, 2010: 1–10. Retrieved www.persoonia.org
- Zhou, S. and Stanosz, G.R. (2001).** Relationships among *Botryosphaeria* species and associated anamorphic fungi inferred from the analysis of the ITS and 5.8S rDNA sequences. *Mycologia* 93:516–527.

