



UNIVERSITI PUTRA MALAYSIA

***ISOLATION, IDENTIFICATION & PATHOGENICITY TEST OF CAUSAL
AGENT OF DIEBACK-LIKE SYMPTOM OF PLUMERIA SPP.***

NORSHAHADIB BIN ANUAR

FP 2017 60

**ISOLATION, IDENTIFICATION & PATHOGENICITY TEST OF CAUSAL
AGENT OF DIEBACK-LIKE SYMPTOM OF *PLUMERIA* SPP.**

NORSHAHADIB BIN ANUAR

DEPARTMENT OF PLANT PROTECTION

FACULTY OF AGRICULTURE

UNIVERSITI PUTRA MALAYSIA

NOVEMBER 2016

**ISOLATION, IDENTIFICATION & PATHOGENICITY TEST OF CAUSAL
AGENT OF DIEBACK-LIKE SYMPTOM OF *PLUMERIA* SPP.**

By

NORSHAHADIB BIN ANUAR

**A Project Report Submitted in Partial Fulfillment of Bachelor of Horticultural
Science in the Faculty of Agriculture, Universiti Putra Malaysia.**

DEPARTMENT OF PLANT PROTECTION

FACULTY OF AGRICULTURE

UNIVERSITI PUTRA MALAYSIA

NOVEMBER 2016

ENDORSEMENT

This project entitled “ Isolation, identification and pathogenicity test of causal agent of dieback-like symptom of *Plumeria* spp.” is prepared by Norshahadib Bin Anuar and submitted to Faculty of Agriculture in fulfillment of the requirement of PRT 4999 (Final Year Project) for the award of the degree of Bachelor of Horticultural Science.

Student's name

.....

Student's Signature

.....

Certified by:

.....

Supervisor's signature

Assoc. Prof. Dr. Wong Mui Yun

Department of Plant Protection

Date :

ACKNOWLEDGEMENTS

Alhamdulillah, I wish to express my sincere gratitude to Allah S.W.T that had guided me to finish this thesis completely. Not to forget, I am very appreciate to my supervisor, Assoc. Prof. Dr. Wong Mui Yun for her continuous supervision, intellectual advices and encouragement from the beginning of this research until the final review of the thesis. My sincere appreciation and thanks also extended to my co-supervisor Dr. Kwan Yee Min for her invaluable suggestions and guidance while carrying the research and experiments. Furthermore, I would like to express my honorable gratitude to Dr. Siti Izera Ismail and Dr. Dzarifah Zulperi for giving their time to guide and supports me during carrying this research. My special thanks also go to all the staff members in Department of Plant Protection, especially Mrs. Asmalina, Mr. Johari and Mr. Nazri, and also not to be forgotten all master students that always giving encouragement and guidance during the research which are Ms. Siti Solehah and Mr. Rahim. I am also indebted to my laboratories mate and friends, especially Ms. Nurmadhihah, Ms. Sofnis, Mr. Karthik, Mr. Mustaqim, Ms. Aziera, Ms. Rohaida and Ms. Atiqah for their caring, helping, guidance and their support during my research. Last but not least, I am very grateful to my parents and family members, Mr Anuar Othman, Halida Abu Bakar and my sisters whose encourage and supports that I will never forget.

TABLE OF CONTENTS

	Page
ENDORSEMENT	iii
ACKNOWLEDGEMENTS	iv
TABLE OF CONTENTS	v
LIST OF TABLE	viii
LIST OF FIGURES	ix
LIST OF ABBREVIATIONS	xi
ABSTRACT	xiii
ABSTRAK	xv
CHAPTER	
1.0 INTRODUCTION	
1.1 General introduction	1
1.2 Problem statement	2
1.3 Hypothesis	2
1.4 Objectives	2
2.0 LITERATURE REVIEW	
2.1 Apocynaceae	3
2.2 <i>Plumeria</i> spp.	3
2.2.1 <i>Plumeria acuminata</i>	5
2.2.2 <i>Plumeria rubra</i>	6

2.2.3 <i>Plumeria obtusa</i>	7
2.3 <i>Botryosphaeriaceae</i>	8
2.4 <i>Botrytis</i> sp.	10
2.5 Koch's Postulate	11
2.6 Polymerase Chain Reaction (PCR)	12
2.7 Internal Transcribed Spacer (ITS)	14
3.0 MATERIALS AND METHODS	
3.1 Study site and plant material	15
3.2 Fungal isolation	17
3.3 Subculture	17
3.4 Inoculation and pathogenicity test	18
3.5 Observations	18
3.6 CTAB buffer preparation	18
3.7 Wash buffer preparation	19
3.8 DNA extraction	19
3.9 Polymerase Chain Reaction (PCR) amplification	21
3.10 DNA sequence analysis	22
4.0 RESULTS AND DISCUSSION	
4.1 Isolation and identification of causal agent	23

4.2 Morphological analysis	25
4.2.1 Observations of grown culture	25
4.2.1 Observations of culture under microscope	25
4.3 Molecular analysis	26
4.3.1 Genomic DNA extraction	26
4.3.2 PCR amplification of Internal Transcribed Spacer 1 (ITS) gene	27
4.3.3 DNA sequence analysis	27
4.4 Pathogenicity test	32
5.0 CONCLUSION	36
REFERENCES	39
APPENDICES	
APPENDIX 1	48
APPENDIX 2	49

LIST OF TABLE

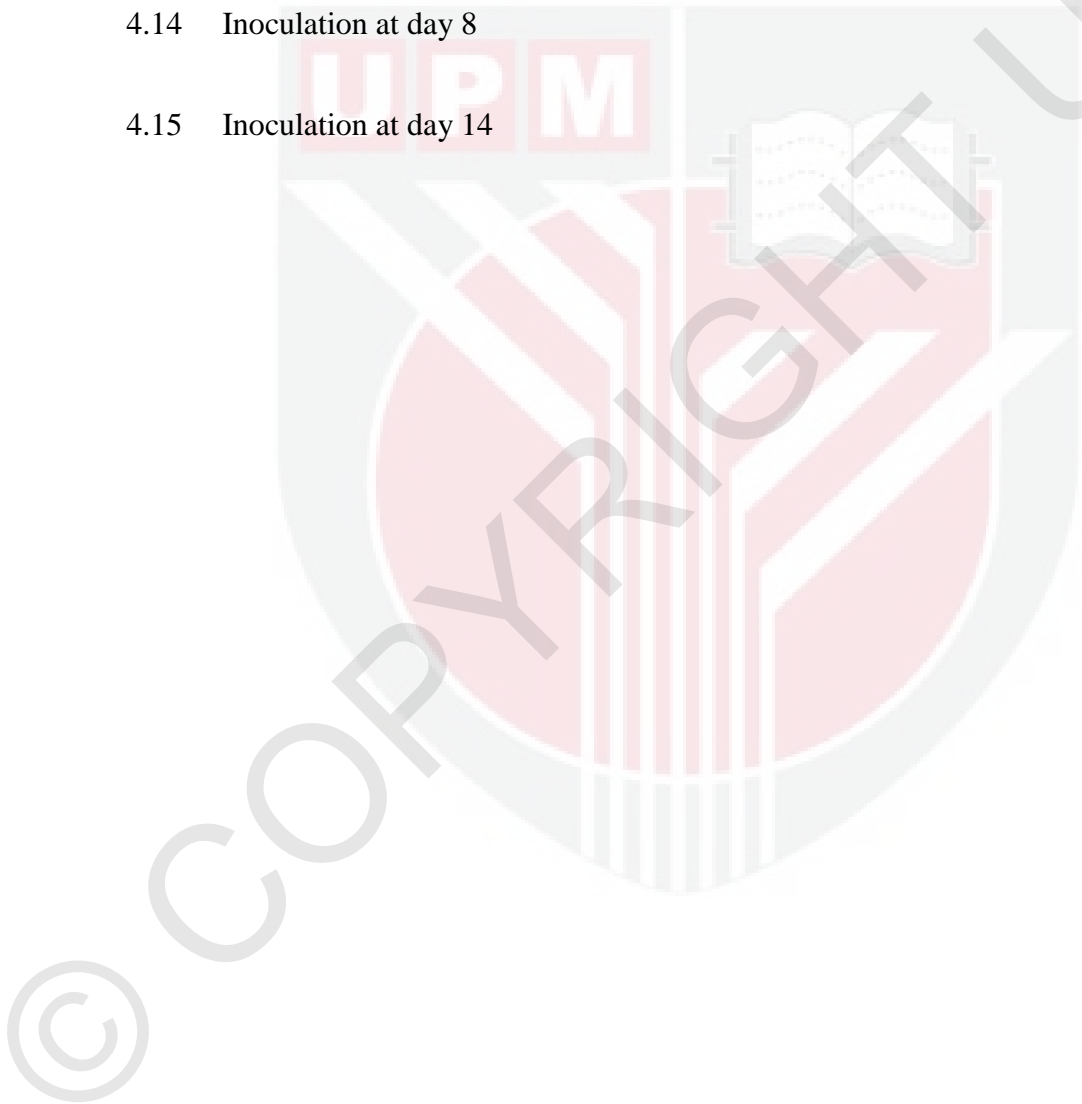
Table		Page
1	BLAST result of ITS gene of <i>Neoscytalidium dimidiatum</i> from NCBI Genbank.	28



LIST OF FIGURES

Figure	Page
2.1 <i>P. acuminata</i> morphology	5
2.2 <i>P. rubra</i> morphology	7
2.3 <i>P. obtusa</i> morphology	8
2.4 Light micrographs of conidia of six Botryosphaeriaceae species	10
3.1 Affected stage tree	16
4.1 Initial condition of the stems	23
4.2 Initial incubation of stem after 3 days	24
4.3 Observations under light microscope	26
4.4 PCR amplifications of unknown culture samples using ITS1 and ITS4	27
4.5 Graphical view of the <i>Neoscytalidium</i> isolate in fungal classification using BLAST	30
4.6 Inoculation at day 1	33
4.7 Inoculation at day 3	33
4.8 Inoculation at day 5	34
4.9 Inoculation at day 8	34

4.10	Inoculation at day 14	35
4.11	Inoculation at day 1	35
4.12	Inoculation at day 3	36
4.13	Inoculation at day 5	36
4.14	Inoculation at day 8	37
4.15	Inoculation at day 14	37



LIST OF ABBREVIATION

PDA	Potato Dextrose Agar
NA	Nutrient Agar
LCB	Lactophenol Cotton Blue
BLAST	Basic Local Search Alignment Tools
APHA	American Public Health Association
MUG	4-methylumbelliferyl- β -D-glucuronide
PCR	Polymerase Chain Reaction
DNA	Deoxyribonucleic Acid
RNA	Ribonucleic Acid
rDNA	ribosomal Deoxyribonucleic Acid
rRNA	ribosomal Ribonucleic Acid
$^{\circ}\text{C}$	Degree Celcius
ITS	Internal Transcribed Spacer
EF1- α	Elongation factor 1-alpha
IGS	Intergenic Spacer Region
CTAB	Cetyl trimethylammonium bromide

EDTA Ethylenediaminetetraacetic Acid

TE Tris EDTA

PVP Polyvinylpyrrolidone

NaCl Sodium Chloride

rpm Revolution per minute

UV Ultra-violet

kb kilobase

Rep Replications

MP Maximum Parsimony

Cm Centimeter

m meter

g grams

ml mililiter

inch inches

M Molar

mM milimolar

μl microliter

% Percentage

ABSTRACT

Plumeria spp. is an ornamental tree in the family Apocynaceae that can grow up to 9m tall and it is natively found in tropical America. The strong scent of their flowers is the main attraction in the landscape area. However, this ornamental tree can be infected by several diseases such as the stem rot, black tip rot, leaf rusts and dieback. Based on some related reports, dieback may be caused by fungus from Botryosphaeriaceae family. In Malaysia, there is no report found about the causal agent of dieback on *Plumeria* spp. even though the symptom has been observed and less case reported in other countries. In this research, the objectives were 1) to identify the causal agent of dieback on *Plumeria* spp and 2) to conduct pathogenicity test to verify the causal agent. The infected samples were collected from landscape area in Tropicana Height in Kajang, Selangor on May 2016 to July 2017. The pathogen was isolated and cultured on Potato Dextrose Agar (PDA) medium. The causal agent of dieback on *Plumeria* spp. was confirmed by Koch's Postulate method using 15 healthy stem cuttings of *Plumeria* spp at 4-6 weeks old by re-isolating the pathogen in PDA media. For objective 1, the fungal pathogen was identified through morphological and molecular characteristics. From morphological analysis, the fungus was grayish to black in color and turned out to be dimorphic in culture. The conidia were ellipsoid, translucent and mix with paraphyses ranging from 3.5-5 x 6.5-12 μm in size and arranged in arthric chains. Meanwhile, for molecular analysis the culture samples were successfully extracted and amplified by PCR using ITS1 and ITS4 primers to produce about 1000 basepairs (bp). The PCR products were sent for sequencing and identified by using

Basic Local Alignment Tool (BLAST). From BLAST analysis, it has about 971 nucleotide sequences for ITS1. The ITS1 gene has 99% of identity and 95% of the query covered for the sequence. Based on morphological characteristics and ITS sequence region, the pathogen was identified as *Neoscytalidium dimidiatum*. For objective 2, pathogenicity test was successfully done and shows that both plant species of *P. rubra* and *P. obtusa* shows the same severity when inoculated with this pathogen in duration of two weeks. It can be concluded that this fungus is the main pathogen that cause dieback-like symptom on this plant species. Generally, this is the first report of dieback disease on *Plumeria* spp. caused by *Neoscytalidium dimidiatum* in Malaysia.

ABSTRAK

Plumeria spp. adalah sejenis pokok hiasan yang tergolong di dalam keluarga Apocynaceae yang boleh membesar setinggi 9m dan ia secara asalnya dijumpai di kawasan tropikal Amerika. Bau bunganya yang kuat menjadi tarikan utama di kawasan landskap. Walaubagaimanapun, pokok hiasan ini boleh dijangkiti oleh beberapa penyakit seperti reput batang, reput pangkal hitam, mati rosot dan karat daun. Berdasarkan beberapa laporan yang berkaitan, penyakit mati rosot (*dieback*) mungkin disebabkan oleh kulat daripada keluarga *Botryosphaeriaceae*. Di Malaysia, tiada laporan yang menyatakan ejen penyebab penyakit dieback pada *Plumeria* spp. walaupun simptom telah banyak diperhatikan dan kurang kes dilaporkan di negara-negara lain. Dalam kajian ini, objektif utama adalah 1) untuk mengenalpasti ejen penyebab *dieback* pada *Plumeria* spp. dan 2) menjalankan ujian patogenesisiti untuk mengesahkan ejen penyebab tersebut. Sampel yang ada simptom jangkitan telah dikumpulkan di kawasan taman lanskap sekitar Tropicana Heights di Kajang, Selangor pada Mei 2016 – Julai 2017. Kulat pathogen dipencil dan dibiakkan di atas medium Potato Dextrose Agar (PDA). Agen penyebab *dieback* pada *Plumeria* spp. telah disahkan dengan kaedah Koch's Postulate menggunakan 15 keratan batang *Plumeria* spp di usia 4-6 minggu dengan mengasingkan semula pathogen di atas medium PDA. Untuk mencapai objektif 1, patogen telah dikenalpasti dengan kaedah morfologi dan molecular. Melalui analisis secara morfologi, kulat ini adalah berwarna kelabu kehitaman dan adalah dimorfik dalam kultur. Konidia berbentuk ellipsoid, lutsinar dan bercampur dengan paraphyses yang bersaiz antara 3,5-5 x 6,5-12 μm dan tersusun

secara dalam rantaian *arthric*. Sementara itu, untuk analisis molekular, sampel telah berjaya diekstrak dan diamplifikasi oleh PCR menggunakan ITS1 dan ITS4 primers dan menghasilkan kira-kira 1000 basepairs (bp). Produk PCR telah dihantar untuk penjujukan dan dikenalpasti dengan menggunakan *Basic Local Alignment Tool* (BLAST). Menurut analisis BLAST, ia mempunyai kira-kira 971 urutan nukleotida untuk ITS1. Gen ITS1 mempunyai 99% daripada identiti dan 95% daripada pertanyaan meliputi untuk urutan nukleotida. Berdasarkan ciri-ciri morfologi dan rantauan urutan ITS, patogen ini telah dikenalpasti sebagai *Neoscytalidium dimidiatum*. Untuk objektif ke-2, ujian patogenisiti telah berjaya dilaksanakan dan menunjukkan bahawa kedua-dua spesis pokok, *P. rubra* dan *P. obtusa* terjangkit dengan serius dalam masa dua minggu. Kesimpulannya, kulat ini adalah merupakan patogen utama yang menyebabkan simptom seakan *dieback* pada spesis pokok ini. Umumnya, ini adalah laporan pertama penyakit *dieback* pada *Plumeria* spp. yang disebabkan oleh *Neoscytalidium dimidiatum* di Malaysia.

CHAPTER 1

INTRODUCTION

1.1 General Introduction

Plumeria spp. has a wide contribution and importance to economy as it's provides medicinal values and culturally can be used to create drums, bowls and furniture from its wood (Eggenberger and Eggenberger 2005). For example, the latex produced by the tree is mix with coconut oil can be utilized to treat itch, rheumatism, and gum troubles (Farooque et al. 2012). However, this plant species can also infected by several diseases that can affect the quality of the plant itself such as stem rot, black tip rot, leave rusts and dieback.

During May 2016 to July 2016, stem cutting samples of *Plumeria* spp. with dieback-like symptoms were randomly sampled in Tropicana Heights, Kajang located in Selangor, Malaysia with disease incidence exceeding 90% in some severely affected landscape areas reported by Nerissa Chu, staff of the landscape area. The dieback symptom was observed in the infected plants including the rusts of leaves but, in this study dieback of *Plumeria* spp. was observed as primary observation. Since there is no research has been done regarding the causal of dieback on *Plumeria* spp. in Malaysia,

therefore, a pathogenicity test and observation for dieback of *Plumeria* spp. were conducted in the laboratory by taking infected samples to be examined.

1.2 Problem statement

In Malaysia, there is no report found about the causal agent of dieback on *Plumeria* spp. even though the symptom has been observed and less case reported in other countries.

1.3 Hypothesis

1. The dieback of *Plumeria* spp. may be caused by fungus in family of Botryosphaeriaceae because many of these fungi cause canker and dieback diseases on wide range of plant hosts over the world (von Arx, 1987; Burgess et al., 2006; Slippers and Wingfield, 2007).

1.4 Objectives

1. To identify the causal agent of dieback on *Plumeria* spp.
2. To conduct pathogenicity test to verify the causal agent.

References

Burkill I.H. 1935, A Dictionary Of The Economic Products Of The Malay Peninsula, Crown Agents for the Colonies, London, 2(I-Z), pp 1776–1778.

Burgess T.I, Barber P.A., Mohali S., Pegg G., De Beer W., Wingfield M.J. 2006, Three new *Lasiodiplodia* spp. From the trop-ics, recoguized based on DNA sequence comparisons and morphology. *Mycologia*, 98:423– 435.

Barr M.E. 1987, Prodrumus to class Loculoascomycetes, Published by the author, Amherst, Massachusetts, US.

Boundless. 2015, Koch's Postulates: *Boundless Microbiology*. Retrieved 4th May, 2016 from

<https://www.boundless.com/microbiology/textbooks/boundless-microbiology-textbook/epidemiology-10/principles-of-epidemiology-130/koch-s-postulates-668-5418/>

Corner E.J.H. 1952, Wayside Trees of Malaya, The Malayan Nature Society, pp 147-148.

Crous P.W., Slippers B., Wingfield M.J., Rheeder J., Marasas W.F.O et al. 2006, Phylogenetic lineages in the *Botryosphaeriaceae*, *Studies in Mycology*, 55: 235–253.

Chinn J.T and Criley R.A. 1983, *Plumeria cultivars in Hawaii*, Research Bulletin 158, University of Hawaii, p 48.

Chuang M.F., Ni H.F., Yang H.R., Shu S.L., and Lai S.Y. 2012, First Report of Stem Canker Disease of Pitaya (*Hylocereus undatus* and *H. polyrhizus*) Caused by *Neoscytalidium dimidiatum* in Taiwan, 96(6): p 906.

Doyle J.J., and Doyle J.L. 1987, A Rapid DNA Isolation Procedure for Small Quantities of Fresh Leaf Tissue, *Phytochemical Bulletin*, 19, 11-15.

Department of Agriculture and Fisheries, Queensland. 2012, A-Z horticultural diseases and disorders, Retrieved 27th March, 2016 from <https://www.daf.qld.gov.au/plants/fruit-and-vegetables/a-z-list-of-horticultural-diseases-and-disorders/botrytis-grey-mould>

Edward F., Gilman D.G. 1994, *Plumeria rubra* Middleaged Frangipani, Fact Sheet ST-491. Watson, 1-4.

Endress M.E., Bruyns P.V. 2000, A revised classification of the Apocynaceae, *The Botanical Review*, 66 (1):1–56.

Eriksson O.E. 1981, The families of bitunicate ascomycetes, *Opera Botanica*, 60: 1–220.

Elshafie A.E., Ba-Omar T. 2001, First report of *Albizia lebbeck* dieback caused by *Scytalidium dimidiatum* in Oman, *Mycopathologia*, 154, 37–40.

Eggenberger R. and Eggenberger M. 2005, *The Handbook on Plumeria Culture*, Fourth Rev. ed. and Expanded. Golden Bridge Publications.

Farooque Ashraf M.D., Mazumder A., Shambhawe S. and Mazumder R. 2012, Review on *Plumeria acuminata*, *International Journal of Research in Pharmacy and Chemistry*, 2(2), p 468.

Frohlich and Michael. 2016, CTAB Extraction Method to Obtain DNA for Genomic Library Preparation, The Floral Genome Project, Pennsylvania State University, n.d.

Farr D.F., Bills G.F., Chamuris G.P, Rossman A.Y. 1989, Fungi on plants and plant products in the United States, APS Press: Saint Paul, MN, US.

Garibyan L. and Nidhi A. 2013, Research techniques made simple: Polymerase chain reaction (PCR), National Institute of Health.

Gardes M., Bruns T.D. 1993, ITS primers with enhanced specificity for basidiomycetes: application to the identification of mycorrhiza and rusts, *Molecular Ecology*, 2 (2): 113–118.

Gupta V.K., Tuohy M.G., Ayyachamy M., Turner K.M., O'Donovan A. 2012, *Laboratory Protocols in Fungal Biology : Current methods in fungal biology*, Springer science and Business media, p 121.

Hsieh W.H, Chen C.Y. 1994, *Sivanesania*, a new botryosphaeriaceous ascomycete genus on *Rubus* from Taiwan, *Mycological Research*, 98: 44–46.

Hawksworth D.L, Kirk P.M, Sutton B.C, Pegler D.N. 1995, *Ainsworth and Bisby's Dictionary of the Fungi*, 8th Edition CAB International, Wallingford.

Joshi M. and Deshpande J.D. 2001, Polymerase Chain Reaction: Methods, principles and application, International Journal of Biomedical Research.

Kamariah A.S., Linda B.L. Lim K.H.C., Baser T., Ozek and Demirci B. 1999, Composition of the Essential Oil of *Plumeria obtusa* L. *Flav. Fragr. J.*, 14, 237–240.

Khare C.P. 2007, *Indian Medicinal Plants*, Springer, New York, p 502-509.

Könemann. 2004, *Botanica: The Illustrated AZ of over 10000 garden plants and how to cultivate them*, p 691.

Lafontaine D.L.J., Tollervey D. 2001, The function and synthesis of ribosomes, *Nature Reviews Molecular Cell Biology*, 2 (7): 514

Menninger E.A. 1962, *Flowering trees of the world*. Hearshide Press Inc., New York. p 336.

Mullis K.B. 1990, The unusual origin of the polymerase chain reaction, *Scientific American*, 262(4) :56–61. 64–5.

Moss S.T. 1986, *The Biology of Marine Fungi*. Cambridge, UK: Cambridge University Press, p 76.

Nandkarni K.M. 1976, *Indian Materia Medica*, Popular Prakashan, Bombay, p 993.

Nazar N., David J.G., Clarkson J.J., Mahmood T. and Chase M.W. 2013, The taxonomy and systematics of *Apocynaceae*: Where we stand in 2012, *Bot. J. Linnean Soc.*, 171(3, March), pp 482–490.

Pavlic D., Slippers B., Coutinho T.A., Wingfield M.J. 2007, Botryosphaeriaceae occurring on native *Syzygium cordatum* in South Africa and their potential threat to *Eucalyptus*, *Plant Pathology*, 56 : p 624-636.

Peay K.G., Kennedy P.G., Bruns T.D. 2008, Fungal community ecology: a hybrid beast with a molecular master, *BioScience*, 58: 799–810.

Phillips A. J. L., Alves A., Abdollahzadeh J., Slippers B., Wingfield M. J., Groenewald J. Z., and Crous P.W. 2013, The *Botryosphaeriaceae*: genera and species known from culture, *Studies in Mycology*, 76(1), 51–167.

Pino J.A, Ferrer A., Alvarez D., and Rosado A. 1991, Volatile alcoholic extract of flowers from *Plumeria rubra* L. var. *acutifolia*, Flavour Fragr. I, 9: 343-345.

Pollizi G., Aiello D., Vitale A. 2009, First report of shoot blight, canker and gummosis caused by *Neoscytalidium dimidiatum* on citrus in Italy. 93(11), p. 1215.

Phillips A.J.L., Alves A., Pennycook S.R., Johnston P.R., Ramaley A., et al. 2008, Resolving the phylogenetic and taxonomic status of dark-spored teleomorph genera in the *Botryosphaeriaceae*, Persoonia. 21: 29–55

Stuartxchange: Philipine Medicinal Plants. 2013, Retrieved 13th December 2016, from <http://www.stuartxchange.org/Kalachuchi.html>

Richard A. 1998, Ornamental flowers: *Plumeria*. Cooperative Extension Service, C/T/A/H/R. Department of Horticulture, College of Tropical Agriculture and Human Resource

Ray J.D., Burgess T., Lanoiselet V.M. 2010, First record of *Neoscytalidium dimidiatum* and *N. novaehollandiae* on *Mangifera indica* and *N. dimidiatum* on *Ficus carica* in Australia, 5, 48-50.

Sanahuja G., Lopez P., and Palmateer A.J. 2016, First report of *Neoscytalidium dimidiatum* causing stem and fruit canker of *Hylocereus undatus* in Florida. 100(7): p 1499.

Schoch C.L., Seifert K.A., Huhndorf S., Robert V., Spouge J.L., Levesque C.A., Chen W., Bolchacova E., Voigt K., Crous P.W. et al. 2012, Nuclear Ribosomal Internal Transcribed Spacer (ITS) Region as a Universal DNA Barcode Marker for Fungi, *PNAS*, 109 (16): 6241–6246

Slippers B. and Wingfield M.J. 2007, Botryosphaeriaceae as endophytes and latent pathogens of woody plants: diversity, ecology and impact. *Fung Biol Rev*, 21: 90-106.

Sivanesan A. 1984, The bitunicate Ascomycetes and their anamorphs.

Staples G.W, Herbst D.R. 2005, A Tropical Garden Flora: Plants cultivated in the Hawaiian Islands and other tropical places, Bishop Museum Press, Honolulu, Hawaii

The University of Adelaide: Mycology Online.2016, Retrieved 23rd November, 2016

from

http://www.mycology.adelaide.edu.au/Fungal_Descriptions/Coelomycetes/Neoscytalidium/

University Of Maryland. 2004, General microbiology. Retrieved 5th May, 2016 from

<http://www.life.umd.edu/classroom/bsci424/BSCI223WebSiteFiles/KochsPostulates.htm>

m

Von Arx J.A and Müller E. 1954, Die Gattungen der amersporen Pyrenomyceten, Beiträge zur Kryptogamenflora der Schweiz, 11(1): 1–434.

Weier H.U, Gray J.W. 1988, A programmable system to perform the polymerase chain reaction, DNA, 7 (6):441–7.

Zhang, Y. J., Zhang, S., Liu, X. Z., Wen, H. A., Wang, M. 2010, A simple method of genomic DNA extraction suitable for analysis of bulk fungal strains. The Society for Applied Microbiology, Letters in Applied Microbiology 51, 114–118