

UNIVERSITI PUTRA MALAYSIA

PCR DETECTION OF Ganoderma spp. WITH TRANSLATION ELONGATION FACTOR 1 - ALPHA (tef1-α) GENE

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FP 2017 54

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PCR DETECTION OF *Ganoderma* spp. WITH TRANSLATION ELONGATION FACTOR 1 - ALPHA (*tef1*-α) GENE

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A project report submitted to Faculty of Agriculture, Universiti Putra Malaysia, in fulfillment of the requirement of PRT 4999 (Final Year Project) for the award of the degree of Bachelor of Agricultural Science

Faculty of Agriculture University Putra Malaysia 2016/2017

ENDORSEMENT

This project entitled "PCR Detection of *Ganoderma* spp. with Translation Elongation Factor 1-alpha (*tef1-a*) gene" is prepared by Nur Sofnis Erina Binti Mohd Suhaimi and submitted to Faculty of Agriculture in fulfillment of the requirement of PRT 4999 (Final Year Project) for the award of the degree of Bachelor of Agricultural Science.

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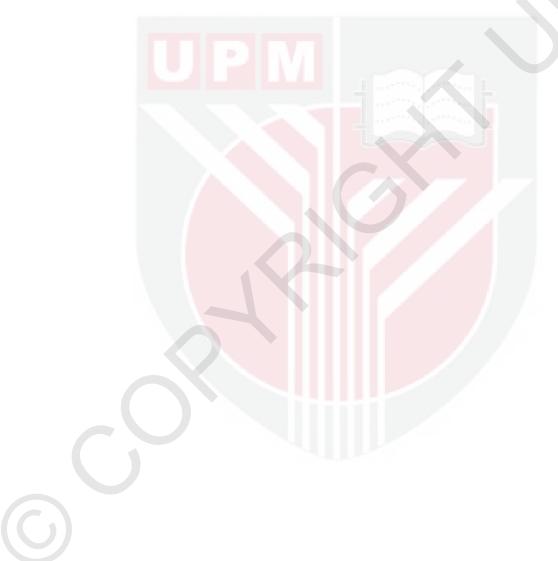
ACKNOWLEDGEMENT

First and above all, I praise God, the almighty for providing me this opportunity and granting me the capability to proceed this project. This thesis appears in its current form due to the assistance and guidance several people. Therefore, I would like to offer my sincere thanks to all of them. I would like to express my gratitude to my supervisor, Associate Professor Dr. Wong Mui Yun for accepting me as your Final Year Project student, your warm encouragement, thoughtful guidance, comment and correction of the thesis. I want to express my deep thanks to Dr. Kwan Yee Min for her willingness to advice on my lab works. I also want to deliver my thankful to Miss Siti Solehah Binti Kassim for the trust, and especially for your patience and guidance during the project. I would like to thank the members of staff, Madam Asmalina, Mr. Nazri and Mr. Johari for their excellent and friendly assistance in the lab with various problems all the time. I thank to my fellow labmate, Muhammad Adib for the insightful discussion, offering valuable advice, for your support during the whole period of the study, for the time we were working together and for all the fun we had. I also want to thank Muhamad Faiz for your support during the whole period of the study and offering valuable advice. Lastly, I would like to thank my family and friends for their support and encouragement throughout the period of project.

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LIST OF ABBREVIATIONS

BSR	Basal stem rot
BLAST	Basic Local Alignment Search Tools
PCR	Polymerase chain reaction
DNA	Deoxyribonucleic acid
tef1-a	Translation elongation factor 1-alpha
MEA	Malt extract agar
PDB	Potato dextrose broth
СТАВ	Centrimethyl ammonium bromide
Tris-HCl	Tris-Hydrochloric acid
PVP	Polivinylpyrrolidone
NaCl	Sodium Chloride
TE	Tris-EDTA
RNAse A	Ribonuclease A
TAE	Tris-Acetate-EDTA
bp	Base pair length
ITS	Internal Transcribe Spacer
Е	Expect value
ELISA	Enzyme-linked Immunosorbent Assay

C

ABSTRACT

The most serious disease of oil palm in Malaysia all the while is basal stem rot (BSR). The losses can be accounted to about 50-85% over the life cycle of oil palm planting. There are several species of Ganoderma that are associated with BSR or isolated from oil palm trees but the major pathogen of this disease is Ganoderma boninense. The identification of Ganoderma species is difficult using morphological method. This will lead to a problem in disease management. The objectives of this study were 1) to extract genomic DNA from four *Ganoderma* species and 2) to develop a polymerase chain reaction (PCR) protocol using translation elongation factor 1-alpha (tef1- α) gene. To achieve objective 1, centrimethyl ammonium bromide (CTAB) method will be used for DNA extraction. To achieve objective 2, universal primers of $tef1-\alpha$ will be used to amplify Ganoderma species, sequenced the PCR products and identify the DNA sequences using Basic Local Alignment Search Tool (BLAST). For objective 1, only one genomic DNA species was obtained successfully with 295.0 ng/µl (A260/280) and the purity were 1.8 (A260/280) and 1.9 (A260/230). For objective 2, PCR protocol successfully amplified tefl- α gene for one species of Ganoderma. Primer pair tefl- α and *ef1* could be used to amplify *tef1-a* gene from *Ganoderma* spp. even though it was successfully amplified only one species in this project which was Ganoderma boninense. PCR amplification will enable for better identification of Ganoderma species by improving DNA extraction and taking all the precautions.

ABSTRAK

Penyakit yang paling serius dalam industri kelapa sawit Malaysia adalah reput pangkal batang. Penyakit ini menyumbang kerugian di antara 50-85% sepanjang kitaran hidup pokok kelapa sawit. Sebenarnya, terdapat beberapa spesies Ganoderma yang menjadi punca penyakit reput pangkal batang ini tetapi patogen utamanya ialah Ganoderma boninense. Spesies-spesies Ganoderma sukar dikenalpasti dengan kaedah morfologi. Hal ini akan menjadi punca kepada masalah pengurusan penyakit. Jadi, tujuan kajian ini adalah untuk membangunkan satu prosedur untuk pengesanan dan pembezaan spesies Ganoderma yang tepat. Objektif kajian ini adalah 1) untuk mengekstrak genomik DNA dari empat spesies Ganoderma dan 2) untuk membangunkan sistem pengesahan PCR dengan menggunakan faktor gen terjemahan pemanjangan 1-alfa (tef1- α). Untuk mencapai objektif 1, kaedah centrimethyl ammonium bromide (CTAB) akan digunakan untuk pengekstrakan DNA. Untuk mencapai objektif 2, primer sejagat $tefl-\alpha$ akan digunakan untuk mengenalpasti spesies Ganoderma, disusun produk PCR dan mengenalpasti susunan DNA menggunakan Basic Local Alignment Search Tool (BLAST). Untuk objektif 1, hanya satu spesies DNA genomic telah diperolehi dengan 295.0 ng/µl (A260/280) dan kadar puriti adalah 1.8 (A260/280) dan 1.9 (A260/230). Untuk objektif 2, protocol PCR berjaya mengenalpasti gen *tef1-a* untuk satu species Ganoderma. Pasangan primer tef1- α dan ef1 boleh digunakan untuk mengenalpasti gen tef1-α dari Ganoderma spp. walaupun hanya berjaya mengenalpasti satu spesies sahaja dalam projek ini iaitu Ganoderma boninense. Penguatan PCR akan membolehkan pengenalpastian spesies Ganoderma lebih baik dengan meningkatkan pengekstrakan DNA dan mengambil semua langkah berjaga-jaga.

Chapter 1

INTRODUCTION

Oil palm (*Elaeis guineensis Jacq.*) is originated from West Africa and become one of the most important agricultural crops in Malaysia. Malaysia is one of the countries that contribute in world palm oil production. Oil palm trees can produce two types of oil which are from the flesh of fruit and palm kernel oil from seed or kernel. There are about 4.49 million hectares of land cover with oil palm plantation which can generate almost 17.73 million tonnes of oil palm and 2.13 tonnes of palm oil kernel. It is a monocious crop which has both male and female flowers on the same tree. The oil palm trees start to produce fruit 30 months after planting and its productivity can last for 20 to 30 years (Source:http://www.mpoc.org.my/Palm_Oil).

Basal stem rot (BSR) disease has becomes the most serious disease in oil palm industry. BSR disease can reduce the production of oil palm. This disease is caused by fungus *Ganoderma boninense*. Isolation of *Ganoderma* from diseased oil palm surround Peninsular Malaysia is concluded as a same species of *Ganoderma boninense* (Ho and Nawawi, 1985). There are four species that have been recorded recently which are *G. boninense*, *G. zonatum*, *G. miniatocinctum and G. tornatum* (Idris, 1999). Earlier stage of oil palms tend to easily can be infected, especially the young plantings of less than 5 years old. The earliest symptoms of this disease occur in the foliage of lower frond that become yellowing. A dry rotting of internal tissues at stem base or root bole is produces by the infection of *Ganoderma boninense*. The infection and distribution of *Ganoderma* spp. can be through root (Turner, 1981). In recent studies showed that BSR can also be spread through spores (Flood *et al.*, 2003; Flood and Hassan, 2004).

The invention of polymerase chain reaction (PCR) is by Kary Mullis and his colleagues in 1980s. Monoclonal and polyclonal antibodies had been used to detect pathogenic *Ganoderma* species. However, the assay of polyclonal antibodies showed cross reaction with saprophytic fungi commonly found on diseased oil palm roots and trunk although its percentage were lower (Utomo and Niepold, 2000; Idris and Rafidah, 2008). There is no distinction was found between pathogenic *Ganoderma* spp. isolated from oil palms and other pathogenic *Ganoderma* spp. that signified Enzyme-linked Immunosorbent Assay (ELISA) test cannot be used to distinguish among different *Ganoderma* species.

These conventional methods were time-consuming and their accuracy is not very high. For that reason, the availability of rapid, inexpensive and accurate diagnostic technique that is specific and readily adapted can give benefit decision-making for appropriate control (Utomo and Niepold, 2000). The identification of *Ganoderma* spp. can be detected using polymerase chain reaction (PCR). The objectives of this study were 1) to extract genomic DNA from four *Ganoderma* species and 2) to develop a polymerase chain reaction (PCR) protocol using translation elongation factor 1-alpha (*tef1-a*) gene

The DNA genomic for all of four species of *Ganoderma* is successfully obtained. Polymerase chain reaction (PCR) protocol is developed to detect each species using translation elongation factor 1-alpha (*tef1-a*) gene and specific enough

REFERENCES

- Al-Soud, W. A. and Rådström, P. (2001). Purification and characterization of PCRinhibitory components in blood cells. Journal of Clinical Microbiology; 39(2):485–93.
- Ariffin, D. and A.S. Idris, 1991. A selective medium for the isolation of Ganoderma from diseased tissue. Proceedings of the 1991 PIPOC International Palm Oil Conference-Progress, Prospects and Challenges Towards the 21st Century (Model I-Agriculture), MPOB, Malaysia, pp: 517-519.
- Ashwini, D., & Tiwari, S. P. (2015). Use of CTAB method for isolation of good quality and quantity of DNA. Journal of Pure and Applied Microbiology, 9(3), 2271-2275.
- Atkins, S. D., & Clark, I. M. (2004). Fungal molecular diagnostics: a mini review. Journal of Applied Genetic, 45(1), 3-15.
- Basiron, Y. 2007. Palm Oil Production Through Sustainable Plantations. European Journal of Lipid Science Technology, 109:289-295
- Bessetti. J. (2007). An introduction to PCR inhibitors. PCR inhibition. Promega Corporation.
- Chaverri, P. and Samuels G. J. (2003). *Hypocrea/Trichoderma* (Ascomycota, Hypocreales, Hypocreaceae): species with green ascospores. Study Mycology 48:1–116.
- Chen JL, Huang XL, Wu AD, et al. (2011). An efficient extraction method of pathogenic fungus DNA for PCR. Mycosystema 30: 147-149.
- Chen, Y. F., Chen, R. C., Chan, Y. K., Pan, R. H., Hseu, Y. C., Lin, E. (2009). Design of Multiplex PCR primers using heuristic algorithm for sequential deletion applications. Computational Biology and Chemistry 33, 181-188.
- Devi, K. D., Punyarani, K., Singh, N. S., Devi, H. S. (2013). An efficient protocol for total DNA extraction from the members of order Zingiberales- suitablefor diverse PCR based downstream applications. SpringerPlus 2:669
- Dendis, M., Horváth, R., Michálek, J., Růžička, F., Grijalva, M., Bartoš, M., & Benedík, J. (2003). PCR-RFLP detection and species identification of fungal pathogens in patients with febrile neutropenia. Clinical microbiology and infection, 9(12), 1191-1202.
- Dieffenbach, C., W., Lowe, T., M., J., and Dveksler, G., S. 1993. General Concepts for PCR Primer Design Manual Supplement. Cold Spring Harbor Laboratory Press., 3:S30-S37

- Flood, J., Hassan, Y and Foster H.L. 2003. *Ganoderma* disease of oil palm an interpretation from Bah Lias Research Station. Planter, 78:689-710
- Flood, J. and Hassan, Y. 2004. Basal Stem Rot taxonomy, biology, epidemiology, economic status and control in South East Asia and Pacific Islands, Proceedings of the Malaysian Palm Oil Board Conference. May 2004. 117-33. MPOB. Kuala Lumpur. Malaysia
- Hartley, C. W. S. 1967. The Oil Palm. Longmans, Green and Co Itd.
- Ho, Y.W. and A. Nawawi, 1985. *Ganoderma boninense* Pat. From Basal Stem Rot of oil palm (*Elaeis guineensis*) in Peninsular Malaysia. Pertanika, 8:425-428.
- Idris, A.S., S. Rajinder, A.Z. Madihah and M.B. Wahid, 2010. Multiplex PCR-DNA kit for early detection and identification of *Ganoderma* species in oil palm. MPOB Information Series 531, MPOB, Malaysia
- Idris, A.S. and A.R. Rafidah, 2008. Polyclonal antibody for detection of *Ganoderma*. MPOB Information Series 202, MPOB, Malaysia.
- Idris, A.S, Ariffin, D, Swinburne, T.R, and Watt, T.A. 2000. The Identity of *Ganoderma* Species Responsible for BSR Disease of Oil Palm in Malaysia Pathogenicity Test. Malaysian Palm Oil Board.
- Idris, A. S. (1999). Basal stem rot (BSR) of oil palm (*Elaeis guineensis Jacq.*) in Malaysia: factors associated with variation in disease severity. PhD Theses, Wye College, University of London, UK.
- Idris, A. S., Ariffin, D., Yamaoka., Hayakawa, M., Basri, S and Noorhashimah, M. W. PCR Technique for detection of *Ganoderma*. MPOB information series, ISSN 1511-7871 June 2003 No.188 (202).
- Jose, J. and Usha, R., 2000. Extraction of germiniviral DNA from a highly mucilaginous plant (*Abelmoschus esculentus*). Plant Moleccular Biology Reporter, 18: 349-355
- Karakousis, A., A., Tan, L., Ellis, D, Alexiou, H., Wormald, P. J. (2006). An assessment of the efficiency of fungal DNA extraction methods for maximizing the detection of medically important fungi using PCR. Journal of Microbiological Methods 65: 38–48
- Katcher, H.L. and Schwartz, I. (1994). A distinctive property of Tth DNA polymerase: Enzymatic amplification in the presence of phenol. BioTechniques 16, 84–92.
- Latifah, Z., Abdullah, F., Tan, S. G., Harikrishna, S. K and Ho, Y. W. 2002b. Morphological and Growth Characteristic and Somatic Incompability of *Ganoderma* from Infected Oil Palm and Coconut Stumps. Malaysian Applied Biology, 31:37-48

- Lievens, B., Brouwer, M., Vanachter, A. C., Cammue, B. P. A. and Thomma, B. P. H. J. 2006. Real-time PCR for Detection and Quantification of Fungal and Oomycetes Tomato Pathogens in Plant and Soil Samples. Plant Sciences, 171:155-165
- Lim, S., and Lee, K. T. (2012). Implementation of biofuels in Malaysian transportation sector towards sustainable development: A case study of international cooperation between Malaysia and Japan. Renewable and Sustainable Energy Reviews, 16(4), 1790-1800.
- Michiels, A., Van den Ende, W., Tucker, M., Van Riet, L., & Van Laere, A. (2003). Extraction of high-quality genomic DNA from latex-containing plants. Analytical biochemistry, 315(1), 85-89
- Mingot, J. M., Vega, S., Cano, A., Francisco Portillo, Nieto, M. A. (2013). eEF1A Mediates the Nuclear Export of SNAG-Containing Proteins via the Exportin5-Aminoacyl-tRNA Complex Cell Reports 5, 727–737
- Mo, C., and Rinkevich, B. (2001). A simple, reliable, and fast protocol for Thraustochytrid DNA extraction. Marine Biotechnology, 3(2), 100-102.
- Moncalvo, J. M., Wang, H. H and Hseu, R. S. 1995c. Phylogenetic relationships in *Ganoderma* inferred from the internal transcribed spacers and 25S ribosomal DNA sequences. Mycologia, 87 (2), 223-238
- Moncalvo, J. M., Wang, H. H and Hseu, R. S. 1995a. Gene Phylogeny of the *Ganoderma lucidum* complex based on ribosomal DNA sequences. Comparison with traditional taxonomic characters. Mycological Research, 99, 1489-1499.
- Moncalvo, J. M., Wang, H. H and Hseu, R. S. 1995c. Phylogenetic relationships in *Ganoderma* inferred from the internal transcribed spacers and ribosomal DNA sequences.
- Moťková, P., and Vytřasová, J. (2011). Comparison of Methods for Isolating Fungal DNA Volume 29, Special Issue: S76–S85
- Munshi, R., Kandl, K. A., Carr-Schmid, A., Whitacre, J. L., Adams, A. E., & Kinzy, T. G. (2001). Overexpression of translation elongation factor 1A affects the organization and function of the actin cytoskeleton in yeast. Genetics, 157(4), 1425-1436.
- Nanodrop. (2007). ND-1000 Spectrophotometer V3.5 user's manual. United Stated America: NanoDrop Technology Inc
- Nanodrop. 260/280 and 260/230 ratios. T009-Technical Bulettin. Nanodrop 1000 & 8000. Thermo Fisher Scientific. Wilmington, Delaware USA.

- Osuji, C., Abu bakar, S., Godwin, E. U., Mowobi, G., Nweke, O., & Onyenekwe, P. (2016). Optimization of rDNA sequence amplification of Nigerian *Ganoderma lucidum* using internal transcribed spacer primers for molecular analysis. Journal of Science and Technology Advances, 1(1), 25-32.
- Paterson, R. R., Sariah, M. M., Lima, N. (2013). How will climate change affect oil palm fungal diseases? Crop Protection 46: 113-120
- Porebski, S., Bailey, L. G., Baum, B. R. (1997). Modification of a CTAB DNA extraction protocol for plants containing high polysaccharide and polyphenol components. Plant Molecular Biology Reporter 15 (1): 8-15.
- Rådström P., Knutsson, R., Wolffs, P., Lovenklev, M., Löfström, C. (2004). Pre-PCR processing: strategies to generate PCR-compatible samples. Molecular Biotechnology 26, 133–146.
- Rahjoo, V., Zad, J., Javan-Nikkhah, M., Mirzadi Gohari, A., Okhovvat, S. M., Bihamta, M. R. (2008). Morphological and molecular identification of *Fusarium* isolated from maize ears. Iran Journal of Plant Pathology, 90:463–468.
- Rakib, M.R.M., C.F.J. Bong, A. Khairulmazmi and A.S. Idris, 2014. Genetic and morphological diversity of *Ganoderma* species isolated from infected oil palms (*Elaeis guineensis*). International. Journal of Agriculture and Biology, 16: 691-699.
- Rossen, L., Norskov, P., Holmstrom, K., Rasmussen, O. F. (1992). Inhibition of PCR by components of food samples, microbial diagnostic assays and DNA-extraction solutions. International Journal of Food Microbiology 17, 37–45.
- Samuels, G. J., Dodd, S. L., Gams, W., Castlebury, L. A., Petrini, O. (2002). Trichoderma species associated with the green mold epidemic of commercially grown Agaricus bisporus. Mycologia.;94(1):146-70.
- Samuels, G. J. (2006). *Trichoderma*: systematics, the sexual state, and ecology. Phytopathology 96, 195–206.
- Saulnier, P. and Andremont, A. (1992). Detection of genes in feces by booster polymerase chain reaction, Journal of Clinical Microbiology, 30: 2080–2083.
- Schrader, C., Schielke, A., Ellerbroek, L., Johne, R. (2012). PCR inhibitors–occurrence, properties and removal. Journal of Applied Microbiology. 113:1014–1026.
- Sharma, R., Mahla, H.R., Mohapatra, T., Bhargava, S.C. and Sharma, M.M., 2003. Isolating plant genomic DNA without liquid nitrogen. Plant Molecular Biology Reporter, 21: 43-50
- Singh, G. (1991). *Ganoderma*-the scourge of oil palm in coastal areas. The Planter, 67: 421–444

- Takahiro, A., Masako, S., Haruhisa, S. and Koji, K., 2009, Development of Multiplex to Detect Five Phytium Species Related to Turgrass Disease, Journal of Phytopathology, 158:609-615.
- Thompson, A. (1937). Observations on stem-rot of the oil palm. Federated Malay States Government Press.
- Turner, P.D. 1981. Oil Palm Disease and Disorders. 1st Edn. Oxford University Press, Oxford
- Utomo, C. and F. Niepold, 2000. Development of diagnostic methods for detecting *Ganoderma*-infected oil palms. Journal of Phytopathology, 148: 507-514. DOI: 10.1046/j.1439-0434.2000.00478.x
- Wang, Q., & Wang, X. (2012). Comparison of methods for DNA extraction from a single chironomid for PCR analysis. Pakistan Journal of Zoology, 44, 421-426.
- Weaver, R.F. 2008. Molecular Biology Fourth Edition. The Polymerase Chain Reaction, pp. 67-69. Singapore, New York.
- Williams, C. N. and Hsu, Y. C. 1970. Oil Palm Cultivation in Malaysia, Technical and Economic Aspect. University Malaya press
- Wilson, I. G. (1997). Inhibition and facilitation of nucleic acid amplification. Applied and environmental microbiology, 63(10), 3741.
- Wong, L. C., Bong, C.F. J. and Idris, A.S. 2012. *Ganoderma* Species Associated with Basal Stem Rot Disease of Oil Palm. American Journal of Applied Science, 9 (6): 879-885
- Zhang, J. and Stewart, J. M. (2000). Economical and rapid method for extracting cotton genomic DNA. The Journal of Cotton Science 4: 193-201.
- Zhang, Y. J., Zhang, S., Liu, X. Z., Wen, H. A., Wang, M. (2010). A simple method of genomic DNA extraction suitable for analysis of bulk fungal strains. The Society for Applied Microbiology, Letters in Applied Microbiology 51, 114– 118
- Zhao, X.M., Duszynski, D.W., and Loker, E.S., 2001. A simple method of DNA extraction for *Eimeria* species. Journal of Microbiology Methods, 44: 131-137
- Ministry Of Agriculture and Co-operatives. 1966. The oil palm in Malaya (Malaysia), pp. 1-16. Pulau Pinang: Sinaran Brothers Limited.

Malaysian Palm Oil Board (MPOB): Official Website. Retrieved 05 April 2016 from

http://www.mpob.gov.my/

Ganoderma boninense on a tree. Retrieved from 08 December 2016 from http://fnpsblog.blogspot.my/search?q=ganoderma+boninense

Malaysian Palm Oil Council (MPOC): Official Website. Retrieved 05 April 2016 from http://www.mpoc.org.my/Malaysian_Palm_Oil_Industry.aspx

Malaysian Palm Oil Council (MPOC): Official Website. Retrieved 04 April 2016 from http://www.mpoc.org.my/Palm_Oil.aspx

NCBI. Polymerase Chain Reaction (PCR). Retrieved from 17 March 2016 from http://www.ncbi.nlm.nih.gov/probe/docs/techpcr/

PCR Primer Design. Retrieved 04 April 2015 from

http://www.cybertory.org/exercises/primerDesign/

PCR Primer Design Guidelines. Retrieved 04 April 2016 from

http://www.premierbiosoft.com/tech_notes/PCR_Primer_Design.html

Plantwise Knowledge Bank. Retrieved 06 March 2016 from http://www.plantwise.org/KnowledgeBank/Datasheet.aspx?dsid=24924

Polymerase Chain Reaction: Benefits and Drawbacks. Retrieved 12 December 2016 from http://www.thehorse.com/articles/25338/polymerase-chain-reaction-benefits-and-drawbacks

The Oil Palm Tree. Retrieved from 08 December 2016 from http://www.palmoilhealth.org/what-is-palm-oil/the-oil-palm-tree/